

# **Amitosis confers benefits of sex in the absence of sex to *Tetrahymena***

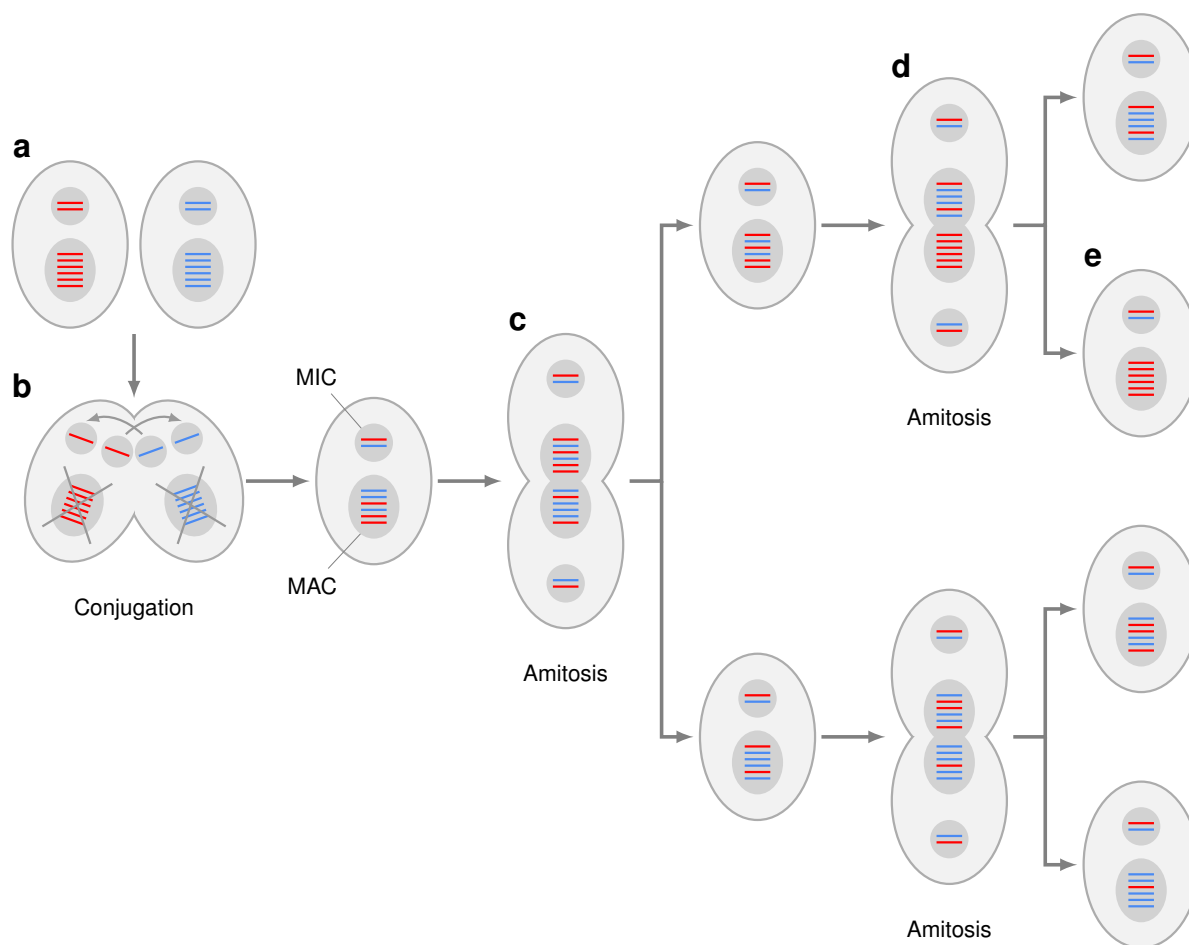
Hao Zhang<sup>1</sup>, Joe A. West<sup>1</sup>, Rebecca A. Zufall<sup>1</sup> & Ricardo B. R. Azevedo<sup>1</sup>

<sup>1</sup>Department of Biology and Biochemistry, University of Houston, Houston, TX 77204-5001, USA

Sex appears to be the most successful reproductive strategy in eukaryotes despite its many costs<sup>1-3</sup>. While a complete explanation for sex's success remains elusive, several evolutionary benefits of sex have been identified<sup>4,5</sup>, such as, the purging of deleterious mutations<sup>6,7</sup>, the accumulation of beneficial mutations<sup>8,9</sup>, and an advantage in biotic interactions<sup>3,10,11</sup>. It is predicted that, by forgoing these benefits, asexual lineages are evolutionary dead-ends<sup>2,12</sup> due to genetic deterioration and/or an inability to adapt to environmental changes. Consistent with this prediction, many asexual lineages show signs of accelerated accumulation of deleterious mutations compared to their sexual relatives<sup>13-18</sup>. Despite these low expectations, some asexual eukaryotic lineages appear to be successful, including the ciliate *Tetrahymena*<sup>19</sup>. Here, we show that the mechanism of somatic nuclear division in *Tetrahymena*, termed amitosis, provides benefits similar to sex, allowing for the long-term success of asexual lineages. We found that, when compared to mitosis, amitosis with chromosome copy number control reduces mutation load deterministically, slows the accumulation of deleterious mutations under genetic drift, and accelerates adaptation. These benefits arise because, like sex, amitosis can generate substantial genetic variation in fitness among (asexual) progeny. Our results indicate that the ability of *Tetrahymena* to persist in the absence of sex may depend on non-

**sexual genetic mechanisms conferring benefits typically provided by sex, as has been found in other asexual lineages<sup>20–23</sup>.**

Although rare throughout ciliates, obligately asexual lineages are abundant, and possibly ancient, in the genus *Tetrahymena*<sup>19</sup>. The reason for this abundance is unknown. One possibility is that the peculiar genomic architecture of *Tetrahymena* allows it to avoid some of the negative consequences of asexuality<sup>19,24</sup>. Ciliates are microbial eukaryotes characterized by the separation of germline and somatic functions into two distinct types of nuclei within a single cell. The somatic macronucleus (MAC) is the site of all transcription during growth and asexual reproduction, and the germline micronucleus (MIC) is responsible for the transmission of genetic material during sexual conjugation (Fig. 1). Following conjugation, a zygotic nucleus divides and differentiates into the two types of nuclei (Fig. 1a,b). During this differentiation, the macronuclear genome undergoes massive rearrangements resulting in a genome with many small, highly polyploid, acentromeric chromosomes<sup>25</sup>. This genome structure results in amitotic macronuclear division (Fig. 1c,d). Amitosis generates variation among individuals in the number of each allele at a locus. In most ciliates, amitosis results in differing numbers of chromosomes among progeny, which eventually leads to senescence and death<sup>26</sup>. However, *Tetrahymena* have an unknown mechanism to control chromosome copy number during amitosis that results in roughly constant ploidy<sup>27</sup>. 25% of 2,609 *Tetrahymena*-like wild isolates lacked a MIC and were, therefore, asexual<sup>19</sup>. To test whether amitosis with chromosome copy number control can account for the relative success of asexual *Tetrahymena*, we examined the evolutionary consequences of various forms of reproduction, nuclear division, and ploidy.



**Figure 1:** Amitosis with chromosome copy number control generates variation among individuals.

See next page for full legend.

**Figure 1: Amitosis with chromosome copy number control generates variation among individuals.** Schematic of sexual conjugation followed by two rounds of asexual division. For simplicity, only one chromosome is shown: it occurs in two copies in the micronucleus (MIC) and six copies in the macronucleus (MAC) (in reality, each chromosome occurs in 45 copies in the *Tetrahymena thermophila* MAC). **a**, During sexual reproduction (conjugation), the diploid MIC undergoes meiosis<sup>27,28</sup>. **b**, Two cells can fuse transiently and exchange haploid meiotic products. A resident meiotic product then fuses with the transferred meiotic product to produce a new diploid zygotic nucleus, which divides to generate the new MIC and MAC (the old MAC is destroyed). During asexual reproduction (**c, d**), the MIC divides by mitosis while the MAC divides by amitosis. Amitosis allows the random segregation of parental chromosomes among daughter cells generating variation among individuals. Ultimately, this results in phenotypic assortment, in which individual chromosomes in the MAC become completely homozygous within several generations<sup>29</sup> (**e**). *T. thermophila*, has an unknown copy number control mechanism that results in an approximately equal number of homologous chromosomes in each daughter cell<sup>27</sup>.

Most mutations with effects on fitness are deleterious but natural selection cannot remove all of them from populations. As a result, many individuals carry deleterious mutations that reduce their fitness, which leads to a reduction in the mean fitness of populations, or mutation load. We begin by investigating the extent to which amitosis with chromosome copy number control affects mutation load. A population of asexual diploids that reproduces by mitosis is expected to show the following mean fitness at equilibrium<sup>30–33</sup>:

$$\hat{W}_{\text{mit}} = \exp(-U_d) \quad (1)$$

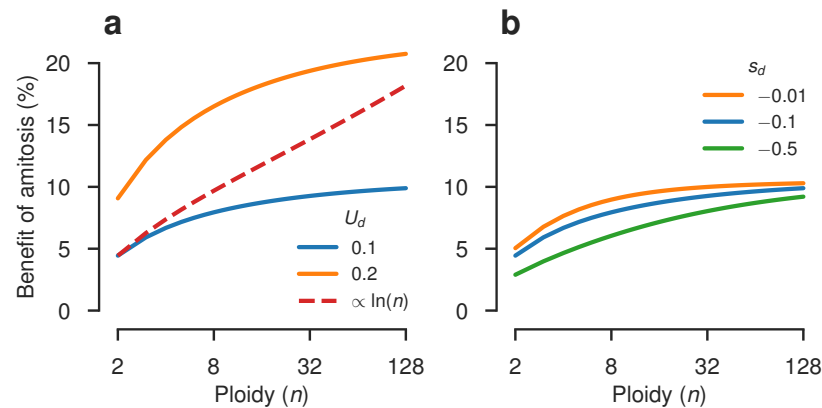
where  $U_d = 2L\mu_d$  is the deleterious mutation rate per diploid genome per generation,  $L$  is the number of loci influencing fitness, and  $\mu_d$  is the deleterious mutation rate per locus per generation (see Supplementary Information). In contrast, if an asexual diploid population reproduces by amitosis, its mean fitness at equilibrium is given by

$$\hat{W}_{\text{amit}} = \exp \left[ -U_d \left( \frac{1 - 3s_d}{2 - 3s_d} \right) \right] \quad (2)$$

where  $s_d < 0$  is the effect on fitness of a deleterious mutation in a homozygous state (see Supplementary Information). This scenario is purely theoretical because no diploid nucleus is known to reproduce amitotically. Equations 1 and 2 rely on several assumptions: (i) population size is very large, so we can ignore genetic drift; (ii) mutations are irreversible;  $\mu_d$  is (iii) low and (iv) equal across loci; (v) there is linkage equilibrium among fitness loci; all mutations (vi) have the same deleterious effect  $s_d$ , and contribute to fitness (vii) additively within loci (i.e., are codominant) and (viii) multiplicatively among loci (i.e., do not interact epistatically). Equations 1 and 2 show that amitosis can reduce mutation load compared to mitosis in diploid populations. For

example, if  $U_d = 0.1$  and  $s_d = -0.1$ , the mean fitness at equilibrium is  $\widehat{W}_{\text{mit}} = 0.905$  under mitosis and  $\widehat{W}_{\text{amit}} = 0.945$  under amitosis. Thus, amitosis has a selective advantage over mitosis of  $\widehat{W}_{\text{amit}}/\widehat{W}_{\text{mit}} - 1 = 4.4\%$ . The deleterious mutation rate,  $U_d$ , has a large effect on the benefit of amitosis: doubling the value of  $U_d$  more than doubles the advantage of amitosis to 9.1% (Fig. 2a). The selection coefficient of a deleterious mutation,  $s_d$ , however, has a comparatively small effect on the benefit of amitosis: making mutations one tenth as deleterious ( $s_d = -0.01$ ) causes the advantage of amitosis to increase to only 5.0% (Fig. 2b).

Amitosis with copy number control is observed in the genus *Tetrahymena*, which have high ploidy in their macronuclear genome (e.g., *T. thermophila* are 45-ploid). Interestingly, the benefit of amitosis relative to a mitotically reproducing organism with the same ploidy increases with ploidy (Fig. 2). For example, if  $U_d = 0.1$  and  $s_d = -0.1$ , the benefit of amitosis increases to 6.7% in tetraploids, 7.9% in octoploids, 8.7% in 16-ploids, and so on. Further increases in ploidy cause diminishing returns in the benefit of amitosis. These expected benefits are conservative because they assume that the deleterious mutation rate,  $U_d$ , is constant across ploidies. If, for example, doubling ploidy causes an increase of 10% in  $U_d$ , a substantially greater benefit of amitosis would be achieved at high ploidies (Fig. 2a, dashed line). A mutation accumulation study estimated that *T. thermophila* has a deleterious mutation rate in the MIC of  $U_d^{(\text{MIC})} = 0.0094$  per genome per generation and that mutations have an expected deleterious effect of  $s_d^{(\text{MIC})} = -0.11$  in a homozygous state<sup>34</sup>. If we assume that the MAC genome has  $U_d^{(\text{MAC})} = (45/2) \times U_d^{(\text{MIC})} = 0.2115$  and  $s_d^{(\text{MAC})} = s_d^{(\text{MIC})}$ , we estimate that amitosis has a benefit of 21.0% relative to mitosis in this species.

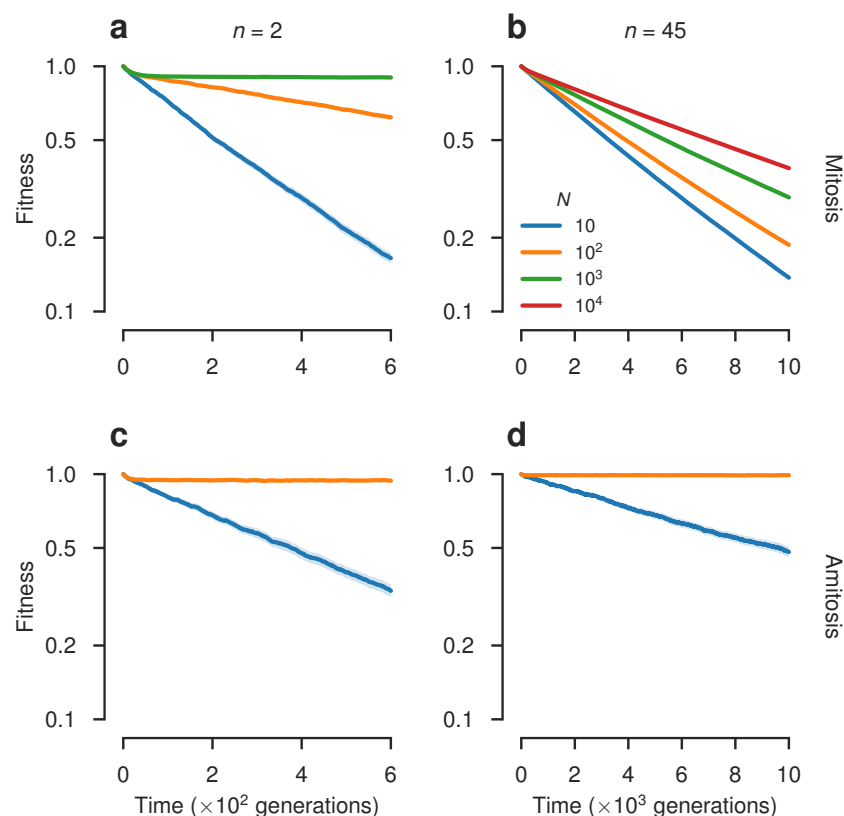


**Figure 2: Amitosis with chromosome copy number control reduces mutation load relative to mitosis in large populations.** Values show the selective advantage of amitosis over mitosis,  $\widehat{W}_{\text{amit}}/\widehat{W}_{\text{mit}} - 1$ , at different ploidies ( $\widehat{W}_X$  is the mean fitness at equilibrium of a population of individuals following reproductive strategy  $X$  for a certain ploidy). **a**, Effect of the genomic deleterious mutation rate,  $U_d$ . Solid lines show selective benefits corresponding to constant values of  $U_d$  at all ploidies. The dashed line assumes that a doubling of the ploidy results in a 10% increase in  $U_d$ . Mutations have a deleterious effect of  $s_d = -0.1$  at all ploidies. **b**, Effect of the selection coefficient of a deleterious mutation,  $s_d$ . We set  $U_d = 0.1$  at all ploidies. In both **a** and **b** we assumed that there were  $L = 100$  fitness loci. Note that ploidy is shown in a log scale.

The analyses so far have ignored the effect of genetic drift. Drift can cause a population to accumulate deleterious mutations stochastically, further increasing genetic load, or drift load<sup>32,35,36</sup>. In asexuals this phenomenon is known as Muller's ratchet<sup>6,37,38</sup>. We now evaluate the extent to which amitosis with copy number control can slow down the accumulation of drift load. Populations of  $N = 10$  or 100 diploid mitotic individuals experience strong Muller's ratchet when  $U_d = 0.1$  and  $s_d = -0.1$  (Fig. 3a). Increasing population size to  $N = 10^3$  individuals causes the ratchet to slow down considerably, allowing populations to achieve mutation-selection equilibrium (Fig. 3a). Reproduction through amitosis makes populations less susceptible to Muller's ratchet. The accumulation of drift load slows down by 39% (95% confidence interval, CI: 31%, 46%) in diploid populations of  $N = 10$  individuals, and effectively halts in populations of  $N = 100$  individuals (Fig. 3c).

The benefit of amitosis in slowing down the accumulation of drift load, like the deterministic benefit, increases with ploidy. Muller's ratchet operates in populations as large as  $N = 10^4$  mitotic 45-ploid individuals (Fig. 3b). Amitosis is able to halt the accumulation of drift load in populations with as few as  $N = 100$  45-ploid individuals (Fig. 3d). Even when amitotic populations are small enough to accumulate drift load, they do so more slowly than mitotic ones. For example, populations of  $N = 10$  amitotic 45-ploid individuals accumulate drift load 64% (95% CI: 59%, 68%) more slowly than mitotic populations of the same size (Fig. 3b,d).





**Figure 3: Amitosis with chromosome copy number control slows down the accumulation of drift load relative to mitosis.** Evolutionary responses of mean fitness in populations of different sizes ( $N$ ) and ploidy ( $n$ ), following different reproductive strategies. Lines show the means of stochastic simulations of 100 populations; shaded regions represent 95% CIs. **a**, Mitosis in diploids ( $n = 2$ ). **b**, Mitosis with a ploidy of  $n = 45$ . **c**, Amitosis in diploids ( $n = 2$ ). **d**, Amitosis with a ploidy of  $n = 45$ . We assumed  $L = 100$  fitness loci, a genomic deleterious mutation rate of  $U_d = 0.1$  per generation, that mutations have a deleterious effect of  $s_d = -0.1$  in a homozygous state, and that, initially, all individuals are unmutated. Note that fitness is shown in a log scale.

98 The benefits of amitosis over mitosis identified so far are analogous to benefits of sexual  
99 over asexual reproduction. In diploids, sexual reproduction by selfing confers a deterministic ad-  
100 vantage over mitosis almost identical to that of asexual amitosis shown in Equations 1 and 2 (see  
101 Supplementary Information). Unlike amitosis, sex with random mating in diploids only confers a  
102 deterministic advantage over asexual reproduction if there is negative epistasis between deleteri-  
103 ous mutations<sup>7,39</sup>, or if deleterious mutations are partially recessive<sup>40,41</sup>. Sex can also counteract  
104 Muller's ratchet<sup>6,37</sup>, much like amitosis (Fig. 3a,c). Are the benefits of asexual amitosis also sim-  
105 ilar to those of sexual reproduction when ploidy is high? We investigated this question in popu-  
106 lations of  $N = 20$  individuals of a 45-ploid organism like *T. thermophila* experiencing  $U_d = 0.1$   
107 and  $s_d = -0.1$ . Amitosis slows down the accumulation of drift load relative to mitosis by 90%  
108 (95% CI: 88%, 92%; Figure 4a). An organism like *T. thermophila* but reproducing sexually, with  
109 outcrossing, every generation (i.e., obligate sex with no amitosis) and then generating a 45-ploid  
110 macronucleus from the recombinant diploid micronucleus (see Fig. 1a,b) would slow down the  
111 accumulation of drift load by 92% (95% CI: 90%, 94%;  $\tau = 1$ , Fig. 4a). However, *T. thermophila*  
112 cannot reproduce sexually every generation; rather, it requires  $\sim 100$  asexual cell divisions to  
113 reach sexual maturity<sup>42,43</sup>. Facultative sex every  $\tau = 100$  generations slows down the ratchet by  
114 only 68% (95% CI: 64%, 72%; measured based on fitness in the generation immediately before  
115 the population reproduces sexually), much less than amitosis (Fig. 4a). The benefit of amitosis  
116 is also comparable to that of sex in larger populations in the presence of beneficial mutations. In  
117 an evolutionary scenario under which asexual populations are not able to adapt, both amitosis and  
118 obligate sex every generation ( $\tau = 1$ ) allow populations to adapt, and more rapidly than facultative

sex every  $\tau = 100$  generations (Fig. 4b).

The results shown in Fig. 4 raise the intriguing possibility that amitosis is actually evolutionarily superior to facultative sex in *T. thermophila* and its relatives, which have  $\tau \approx 100$ . If true, this would lead to the prediction that asexual lineages should outcompete sexual ones in *Tetrahymena*. This could explain why obligately asexual lineages are abundant in *Tetrahymena*<sup>19</sup>. If this explanation is correct, we would expect that asexual lineages of *Tetrahymena* do not show the typical signs of accelerated accumulation of deleterious mutations compared to their sexual relatives<sup>13–18</sup>.

The hypothesis outlined in the previous paragraph may be invalid for two reasons. First, our analysis may overestimate the benefit of amitosis relative to facultative sex. Our hypothesis assumes that chromosome copy number control during amitosis is perfect, or at least, highly precise on an evolutionary time-scale. However, the precision of copy number control is unknown even in *T. thermophila*. Control of chromosome copy number could be less precise than we have assumed and, therefore, confer a smaller benefit to *Tetrahymena*. Second, our analysis may underestimate the benefit of facultative sex relative to amitosis. We have considered only two possible benefits of sex, both “mutational” in nature<sup>4</sup>. Other benefits of sex are not guaranteed to show the same pattern. For example, we have not considered the potential benefits of sex in the face of biotic interactions<sup>3,10,11</sup>. Even if our hypothesis is correct, it is also conceivable that there are additional factors contributing to the relative success of asexual *Tetrahymena*. For example, it has been proposed that high ploidy alone may inhibit the accumulation of deleterious mutations through gene conversion<sup>23</sup>. However, this proposed advantage has not been modelled, and therefore it is difficult

to evaluate.

What is the mechanistic basis of the benefits of amitosis identified here? The main difference between the two types of nuclear division is that amitosis, like sex, can generate more genetic variation in fitness than mitosis. For example, an  $n$ -ploid individual (we assume  $n$  is even for simplicity) with  $n/2$  wild-type alleles and  $n/2$  deleterious alleles will have a fitness of  $W = 1 - s_d/2$ . Mutation will generate a variance in fitness of

$$V_{\text{mut}} = \frac{(u_d - u_d^2) s_d^2}{n^2} \quad (3)$$

every generation, where  $u_d = n\mu_d$  is the deleterious mutation rate at the locus per generation.

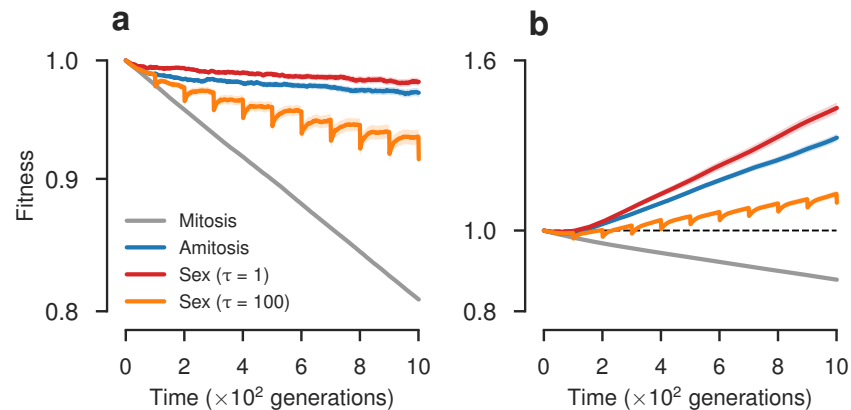
Mitosis is not expected to generate any variance in fitness in addition to mutation (i.e.,  $V_{\text{mit}} = V_{\text{mut}}$ ).

Amitosis will, however, increase the variance in fitness further

$$V_{\text{amit}} = V_{\text{mut}} + \frac{s_d^2}{8n - 4} \quad (4)$$

every generation<sup>44</sup>. Since  $u_d$  is likely to be low, amitosis is expected to increase the variance in fitness to a much greater extent than mutation, and therefore mitosis ( $V_{\text{amit}} \gg V_{\text{mit}}$ ).

We propose that amitosis causes an increase in the additive genetic variance in fitness, therefore making natural selection more efficient—an analog of Weismann’s hypothesis for the advantage of sex<sup>1,4,5</sup>. Consistent with this idea, the variance in fitness generated by amitosis relative to mitosis increases approximately linearly with ploidy ( $V_{\text{amit}}/V_{\text{mit}} \approx n/(8u_d)$ ), which explains why the benefit of amitosis relative to mitosis increases with ploidy. We conclude that amitosis with chromosome copy number control confers benefits of sex in the absence of sex and can account for the high incidence of obligately asexual lineages in *Tetrahymena*<sup>19</sup>.



**Figure 4: The benefit of amitosis with chromosome copy number control is similar to that of sex.** Evolutionary responses of population mean fitness under different reproductive strategies. Lines show the means of stochastic simulations of 500 populations; shaded regions represent 95% CIs. **a**, Populations of  $N = 20$  individuals with a deleterious mutation rate of  $U_d = 0.1$  per genome per generation. All mutations are deleterious and have a selection coefficient of  $s_d = -0.1$  in a homozygous state. **b**, Populations of  $N = 10^3$  individuals with a genomic mutation rate of  $U = 0.1$  per generation; 99% of mutations are deleterious and 1% are beneficial with selection coefficients of  $s_d = -0.1$  and  $s_b = 0.1$ , respectively. We assumed that individuals have a MAC ploidy of  $n = 45$  with  $L = 100$  fitness loci, and that, initially, they carry no mutations. Sexual reproduction takes place with random mating and free recombination every  $\tau$  generations. Note that fitness is shown in a log scale.

1. Weismann, A. On the signification of the polar globules. *Nature* **36**, 607–609 (1887).
2. Maynard Smith, J. *The Evolution of Sex* (Cambridge University Press, Cambridge, 1978).
3. Bell, G. *The Masterpiece of Nature: The Evolution and Genetics of Sexuality* (University of California Press, Berkeley, 1982).
4. Kondrashov, A. S. Classification of hypotheses on the advantage of amphimixis. *J. Hered.* **84**, 372–387 (1993).
5. Burt, A. Perspective: Sex, recombination, and the efficacy of selection—Was Weismann right? *Evolution* **54**, 337–351 (2000).
6. Muller, H. J. The relation of recombination to mutational advance. *Mutat. Res.* **1**, 2–9 (1964).
7. Kondrashov, A. S. Deleterious mutations and the evolution of sexual reproduction. *Nature* **336**, 435–440 (1988).
8. Fisher, R. A. *The Genetical Theory of Natural Selection* (Clarendon Press, Oxford, 1930).
9. Muller, H. J. Some genetic aspects of sex. *Am. Nat.* **66**, 118–138 (1932).
10. Hamilton, W. D., Axelrod, R. & Tanese, R. Sexual reproduction as an adaptation to resist parasites (a review). *Proc. Natl. Acad. Sci. U. S. A.* **87**, 3566–3573 (1990).
11. Otto, S. P. & Nuismer, S. L. Species interactions and the evolution of sex. *Science* **304**, 1018–1020 (2004).

12. Stebbins, G. L. Self fertilization and population variability in the higher plants. *Am. Nat.* **91**, 337–354 (1957).
13. Paland, S. & Lynch, M. Transitions to asexuality result in excess amino acid substitutions. *Science* **311**, 990–992 (2006).
14. Johnson, S. G. & Howard, R. S. Contrasting patterns of synonymous and nonsynonymous sequence evolution in asexual and sexual freshwater snail lineages. *Evolution* **61**, 2728–2735 (2007).
15. Neiman, M., Hehman, G., Miller, J. T., Logsdon, J. M. & Taylor, D. R. Accelerated mutation accumulation in asexual lineages of a freshwater snail. *Mol. Biol. Evol.* **27**, 954–963 (2010).
16. Henry, L., Schwander, T. & Crespi, B. J. Deleterious mutation accumulation in asexual *Timema* stick insects. *Mol. Biol. Evol.* **29**, 401–408 (2012).
17. Tucker, A. E., Ackerman, M. S., Eads, B. D., Xu, S. & Lynch, M. Population-genomic insights into the evolutionary origin and fate of obligately asexual *Daphnia pulex*. *Proc. Natl. Acad. Sci. U. S. A.* **110**, 15740–15745 (2013).
18. Hollister, J. D. *et al.* Recurrent loss of sex is associated with accumulation of deleterious mutations in *Oenothera*. *Mol. Biol. Evol.* **32**, 896–905 (2015).
19. Doerder, F. P. Abandoning sex: multiple origins of asexuality in the ciliate *Tetrahymena*. *BMC Evol. Biol.* **14**, 112 (2014).

20. Gladyshev, E. A., Meselson, M. & Arkhipova, I. R. Massive horizontal gene transfer in bdelloid rotifers. *Science* **320**, 1210–1213 (2008).
21. Flot, J.-F. *et al.* Genomic evidence for ameiotic evolution in the bdelloid rotifer *Adineta vaga*. *Nature* **500**, 453–457 (2013).
22. Seidl, M. F. & Thomma, B. P. H. J. Sex or no sex: Evolutionary adaptation occurs regardless. *BioEssays* **36**, 335–345 (2014).
23. Maciver, S. K. Asexual amoebae escape Muller’s ratchet through polyploidy. *Tr. Parasitol.* **32**, 855–862 (2016).
24. Zufall, R. A. Mating systems and reproductive strategies in *Tetrahymena*. In Witzany, G. & Nowacki, M. (eds.) *Biocommunication of Ciliates*, 221–233 (Springer, Cham, 2016).
25. Chalker, D. L. Dynamic nuclear reorganization during genome remodeling of *Tetrahymena*. *Biochim. Biophys. Acta* **1783**, 2130–2136 (2008).
26. Bell, G. *Sex and Death in Protozoa: The History of an Obsession* (Cambridge University Press, Cambridge, U.K., 1988).
27. Orias, E., Cervantes, M. D. & Hamilton, E. P. *Tetrahymena thermophila*, a unicellular eukaryote with separate germline and somatic genomes. *Res. Microbiol.* **162**, 578–586 (2011).
28. Jahn, C. L. & Klobutcher, L. A. Genome remodeling in ciliated protozoa. *Annu. Rev. Microbiol.* **56**, 489–520 (2002).



29. Doerder, F. P., Deak, J. C. & Lief, J. H. Rate of phenotypic assortment in *Tetrahymena thermophila*. *Dev. Genet.* **13**, 126–132 (1992).
30. Haldane, J. B. S. A mathematical theory of natural and artificial selection, Part V: Selection and mutation. *Math. Proc. Camb. Phil. Soc.* **23**, 838–844 (1927).
31. Kimura, M. & Maruyama, T. The mutational load with epistatic gene interactions in fitness. *Genetics* **54**, 1337–1351 (1966).
32. Crow, J. F. Genetic loads and the cost of natural selection. In Kojima, K.-i. (ed.) *Mathematical Topics in Population Genetics*, 128–177 (Springer, Berlin, 1970).
33. Kondrashov, A. S. & Crow, J. F. King’s formula for the mutation load with epistasis. *Genetics* **120**, 853–856 (1988).
34. Long, H. *et al.* Antibiotic treatment enhances the genome-wide mutation rate of target cells. *Proc. Natl. Acad. Sci. U. S. A.* **113**, E2498–E2505 (2016).
35. Kimura, M., Maruyama, T. & Crow, J. F. The mutation load in small populations. *Genetics* **48**, 1303–1312 (1963).
36. Poon, A. & Otto, S. P. Compensating for our load of mutations: Freezing the meltdown of small populations. *Evolution* **54**, 1467–1479 (2000).
37. Felsenstein, J. The evolutionary advantage of recombination. *Genetics* **78**, 737–756 (1974).
38. Haigh, J. The accumulation of deleterious genes in a population—Muller’s ratchet. *Theor. Popul. Biol.* **14**, 251–267 (1978).

39. Otto, S. P. & Feldman, M. W. Deleterious mutations, variable epistatic interactions, and the evolution of recombination. *Theor. Popul. Biol.* **51**, 134–147 (1997).
40. Chasnov, J. R. Mutation-selection balance, dominance and the maintenance of sex. *Genetics* **156**, 1419–1425 (2000).
41. Otto, S. P. The advantages of segregation and the evolution of sex. *Genetics* **164**, 1099–1118 (2003).
42. Doerder, F. P., Gates, M. A., Eberhardt, F. P. & Arslanyolu, M. High frequency of sex and equal frequencies of mating types in natural populations of the ciliate *Tetrahymena thermophila*. *Proc. Natl. Acad. Sci. U. S. A.* **92**, 8715–8718 (1995).
43. Nanney, D. L., Caughey, P. A. & Tefankjian, A. The genetic control of mating type potentialities in *Tetrahymena pyriformis*. *Genetics* **40**, 668–680 (1955).
44. Schensted, I. V. Appendix model of subnuclear segregation in the macronucleus of ciliates. *Am. Nat.* **92**, 161–170 (1958).

**Acknowledgements** We thank P. Doerder, M. Orive, T. Paixão, and E. Kelleher for discussions. R.A.Z. and R.B.R.A. acknowledge support from grant R01GM101352 from the National Institutes of Health. R.A.Z. acknowledges support from grant DEB-1911449 from the National Science Foundation. R.B.R.A. acknowledges support from grant DEB-1354952 from the National Science Foundation.

**Author contributions** R.A.Z. and R.B.R.A. conceived the study; H.Z., J.A.W., and R.B.R.A. wrote code and conducted simulations; H.Z. and R.B.R.A. analyzed the data; R.B.R.A. conducted mathematical analyses; H.Z., R.A.Z., and R.B.R.A. wrote the manuscript; J.A.W. contributed to editing the manuscript.

**Competing interests** The authors declare no competing interests.

**Supplementary information** is available for this paper paper at <https://doi.org/>...

**Reprints and permissions information** is available at [www.nature.com/reprints](http://www.nature.com/reprints).

**Correspondence and requests for materials** should be addressed to R.B.R.A.