

1 **Effects of dietary NFC/NDF on rumen microbiomes**  
2 **of Karakul sheep based on Three Generations of**  
3 **Full-length Amplifiers sequencing**

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14 The research was supported by the National Natural Science Foundation Project (31760680) and  
15 Xinjiang Production and Construction Group with the young and middle-aged innovation talents  
16 fund (No. 2016BC001).

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26 **Abstract**

27 An study was was conducted to investigate the effects of dietary(non fibrous carbohydrate)  
28 NFC/(neutral detergent fiber)NDF on ruminal bacteria in Karakul sheep. Twelve Karakul sheep  
29 were assigned randomly to four dietary treatments of NFC/NDF (0.78, 1.23, 1.61 and 2.00  
30 respectively) as group 1, 2, 3 to 4. The experiment lasted for four periods, period I (1~18 d), II  
31 (19~36 d), III (37~54 d) and IV (55~72 d). Ruminal digesta were collected consecutively for three  
32 days to measure pH and bacteria per period. The results indicated that the average ruminal pH and  
33 amounts of OTUs were decreased with the increase of dietary NFC/NDF for four periods. At  
34 phylum level, Bacteroidetes and Firmicutes were the predominant bacteria of four periods,  
35 Bacteroidetes were decreased, while the relative abundance of Firmicutes was increased with  
36 dietary NFC/NDF for four periods, but the difference wasn't significant ( $P>0.05$ ). At genus level,  
37 the most relative abundance genus was unidentified-Lachnospiraceae which reached the highest in  
38 group 3 for four periods, but the difference wasn't significant ( $P>0.05$ ). Conclusion: ruminal pH  
39 and bacteria were decreased with the increase of dietary NFC/NDF and the most dominant  
40 bacteria were not change with dietary NFC/NDF and periods in Karakul sheep.

41 **Introduction**

42 Rumen plays an important role in the growth and production of ruminants and it contains a large  
43 number of rumen microorganisms (*Bacteria*, *Protozoa*, *Eukarya* and *Archaea*[1-3]). Rumen  
44 microorganisms breakdown food to provide volatile fatty acids, bacterial protein, and so on to the  
45 host animal[4]. Yang et al.[5] confirmed that 80% of the starch, 50% of the fibre and 60% of the  
46 organic matter in the diet were fermented in the rumen to provide energy for the host. Meanwhile,  
47 after a long-term selection and evolution, rumen microbiomes and the host have formed a  
48 symbiotic relationship to maintain the host's health[6]. Dietary regulation has an important effect  
49 on rumen fermentation[7-9] and microbiome[10-11]. Wei et al.[12] showed that with the increase  
50 of dietary NFC/NDF, ruminal pH decreased significantly, furthermore the composition of rumen  
51 microbial flora also changed, and the total number of rumen bacteria decreased in goats. However,  
52 there are scarce studies on rumen microbiome structure changes with dietary NFC/NDF in  
53 Karakul sheep and the rumen is very complex in which microbiome may change again with  
54 prolong of periods. NDF plays an important role in dry matter intake (DMI) and feed  
55 digestibility[13-14], and NFC in diets is another factor that affect DMI, Hall et al. reported that  
56 NFC would be degraded rapidly in the rumen[14]. The development of technology makes it more  
57 accurate to study on rumen microbiomes, Three Generation of Full-length Amplifiers sequencing  
58 is one of them, which can improve the resolution of species identification, and improve the  
59 accuracy of microbial species composition identification in the samples[15-16]. In this  
60 experiment, ruminal pH and microbiome were measured for four periods to investigate effects of  
61 dietary NFC/NDF on ruminal microbiome in Karakul sheep.

62 **Materials and Methods**

## 63 **Animals and Dietary composition**

64 All experimental procedures were approved by Tarim University Animal Care and Use  
65 Committee, and humane animal care were followed throughout the experiment. Twelve Karakul  
66 sheep with similar age and weight ( $35.3 \pm 3.3$  kg) were fitted with permanent fistula and were  
67 randomly assigned into four dietary treatments of NFC/NDF (0.78, 1.23, 1.61, 2.00 respectively)  
68 as group 1, 2, 3 and group 4, each group with three replicates. They all received vaccines for  
69 parasites before the adaption period, and were fed meeting the standards for raising meat and  
70 sheep in the People's Republic of China[17]. All sheep were housed individually in metabolic  
71 cages ( $1.2\text{ m} \times 1.5\text{ m}$ ) and fed the experimental diet individually twice a day at 9:00 a.m. and 8:00  
72 p.m with free access to water. The ingredients and nutrient level of the diet were shown in S1  
73 Table.

## 74 **The experimental design and sample collection**

75 The experiment lasted for 72 d including four periods, including period I (1~18 d), II (19~36 d),  
76 III (37~54 d) and IV (55~72 d). Each period lasted for 18 d. The first 15 d for adaption and 3 d for  
77 samples, feed intake and defecation rule were studied in the adaption period. The ruminal digesta  
78 were sampled consecutively before morning feeding for three days, the sheep of one group were  
79 collected together for 50 mL, and then, were stored in -80 °C to investigate on rumen  
80 microorganisms. Meanwhile, the ruminal pH was measured after feeding of 0, 1, 3, 6 and 9 h  
81 using pH meter (FE22).

## 82 **DNA Extraction, PCR and Pacio sequencing**

83 The total genetic DNA was extracted using QIAamp Fast DNA Stool Mini Kit (QIAGEN,  
84 Shanghai) according to the illustration and the extracted DNA was detected by 1% agarose gel.  
85 The V1-V9 regions of 16S rDNA were amplified by PCR from the extracted DNA using the  
86 universal primers: F, 5'-AGAGTTGATCCTGGCTCAG-3'; R,  
87 5'-GNTACCTTGTACGACTT-3' (synthesized by Biological engineering co., Ltd). PCR was  
88 carried out in triplicate 50- $\mu$ L reactions which containing 2  $\mu$ L Primer Mix (1uM), 5 ng gDNA, 1  
89  $\mu$ L Trans Fastpfu, 10  $\mu$ L 5 $\times$  Buffer, 5  $\mu$ L 5 $\times$  StimuLate, 5  $\mu$ L dNTPs (2.5mM each), 27  $\mu$ L NFW.  
90 Thermocycling parameters were as follows: 2 min predenaturation at 95 °C; 35 cycles of  
91 denaturation at 95 °C for 30 s, Annealing at 60 °C for 40 s, extension at 72 °C for 90 s; and a Final  
92 extension at 72 °C for 10 min. The production was detected by 2% agarose gel. PCR products  
93 was purified with Gel Extraction Kit (QIAGEN, Shanghai), and the productions were sequenced  
94 on PacBio platform.

## 95 **The sequence analysis**

96 The 16S rDNA reads were firstly processed to get clean reads by discarding the reads that are  
97 shorter than 1340 bp, longer than 1640 bp, and not matching the expected barcodes. Using Uparse  
98 software [18] to cluster all Clean Reads of all samples. Operational taxonomic units (OTUs) were  
99 formed at the similarity of 97%[19]. The OTUs were annotated by the Mothur and SILVA  
100 (<http://www.arb-silva.de/>) [20] according to the reference taxonomy provided by SSUrRNA  
101 database[21]. The OTUs were analyzed by Qiime pipeline (Version 1.9.1) to calculate the richness  
102 and diversity indices i e. observed OTUs, Chao1, Shannon, Simpson, ACE.

## 103 **Statistical Analyses**

104 The results of pH were expressed as means using SPSS 17.0. Comparisons between groups were  
105 performed with ANOVA followed by Duncan test. The difference of ruminal bacteria was  
106 measured and expressed using SPSS 17.0. As well, and the differences were considered to be  
107 significant at  $P<0.05$ .

108 **Results**

109 **pH**

110 The average pH of 0, 1, 3, 6, 9 h after feeding for four periods were shown in Table 1, the pH was  
111 that: group 1> group 2> group 3> group 4 for four periods. There was no significant difference  
112 between four groups in period I ( $P>0.05$ ), while there was significant difference between four  
113 groups in period II, III and IV ( $P<0.05$ ), which showed that the ruminal pH decreased  
114 significantly as the dietary NFC/NDF increased.

115 **Table 1. Effects of dietary NFC/NDF on ruminal pH**

116 **Extraction DNA of rumen bacteria**

117 The DNA extraction results were shown in Fig. 1. The main band was clear and there was no  
118 concentrated band below 500 bp, which indicated that the purity of DNA was well and it could  
119 meet the requirement of sequencing.

120 **Fig. 1. Extraction DNA of rumen bacteria**

121 **The analysis of basic sequencing data**

122 The OTU number of each group in one period was shown in Venn graph as Fig. 2. The result

123 showed that the OTUs was that: group 1> group 2 > group 3> group 4 for four periods, which  
124 showed that the diversity of rumen microbiome decreased with the increase of NFC/NDF, and the  
125 number of each group became more stable with prolong of periods

126 **Fig. 2. Vene graph of OTUs in Karakur sheep**

127 **The analysis of OTU Alpha diversity**

128 The Alpha diversity is a kind of analysis in the diversity of microbiome, which involves the  
129 abundance index of Chao1[22] and ACE[23] and the diversity index of Shannon and Simpson[24].  
130 Before diversity analysis, the dilution curve was drawn by R software (Version 2.15.3) to detect  
131 whether the obtained data could fully reflect the distribution of rumen fluid flora in Karakul sheep.

132 **OTU dilution curve**

133 As showed in Fig. 3, the dilution curve of each group keep increasing with the increase of the  
134 depth of sequencing, which is indicated that new bacteria had been found. The results showed that  
135 the sequencing quantity of each sample could be used to analyze the diversity of flora.

136 **Fig. 3. OTU dilution curve of bacteria in the rumen of Karakul sheep**

137 **Sample diversity index**

138 The results of Alpha diversity were shown in Table 2. The results showed that there was some  
139 difference in the number, richness and diversity of rumen bacterial species in different dietary of  
140 NFC/NDF.

141 **Table 2. Analysis of Alpha diversity of rumen liquid samples at 0.03 distance**

## 142 Structural analysis of rumen microbiome

143 At phyla level, seventeen different phyla were detected. The S2 table and Fig. 4 showed that the  
144 main phylum microbiome didn't change with the dietary NFC/NDF and periods, and  
145 Bacteroidetes (52%~72%) and Firmicutes (19%~46%) was the main microbiome for four periods.  
146 The relative abundance of Tenericutes was 3% to 8% and the others was less than 1%. Fig. 5 was  
147 composed by four main dominant phylum to investigate effects of NFC/NDF on their relative  
148 abundance, the abundance of Bacteroidets and Proteobacteria were that: group 1> group 2> group  
149 3> group 4 for four periods, but the difference wasn't significant( $P>0.05$ ). While the abundance of  
150 Firmicutes was that : group 4> group 3> group 2> group 1, the difference wasn't  
151 significant( $P>0.05$ ) as well. The abundance of Tenericutes reached the highest in group 4 for four  
152 periods.

153 **Fig. 4. The column chart of the main dominant phylum in Karakul sheep**

154 **Fig. 5(a~d) . Effects of dietary NFC/NDF on relative abundance (% reads) of rumen phylum**  
155 **in Karakul Sheep**

156 At the genus level, a total of 77 genera were obtained from sequence alignment. It can be seen  
157 from S3 Table and Fig. 6. The main genus microbiome didn't change with the dietary NFC/NDF  
158 and periods. The highest relative abundance of genus was a kind of semi-cellulose degrading  
159 bacteria, *unidentified-Lachnospiraceae* (1.86%~16.68%). The following relative abundance of  
160 genus was *Succinivlasticum* (0.12%~17.03%). Fig. 7 was composed by four main dominant genus  
161 to investigate effects of dietary NFC/NDF on their relative abundance, the abundance of  
162 *Succinivlasticum* was that: group 2> group 1> group 3> group 4, and the difference wasn't

163 significant ( $P>0.05$ ). The relative abundance of *unidentified-Lachnospiraceae*, *Anaeroplasma* and  
164 *unidentified-Bacteroidales* reached the highest in group 3 for four periods, and the difference  
165 wasn't significant ( $P>0.05$ ) as well.

166 **Fig. 6. The column chart of the main dominant genus in Karakul sheep**

167 **Fig. 7(a~d) . Effects of dietary NFC/NDF on relative abundance (% reads) of rumen genus**  
168 **in**

## 169 **Discussion**

### 170 **Effects of dietary NFC/NDF on ruminal pH**

171 pH is the directest index affecting rumen fermentation[25] and diets are key factors affecting pH.  
172 The results showed that the ruminal average pH decreased with the increase of NFC/NDF for four  
173 periods, which was approved with T.Ma et al.[26]. Agle et al. [27] and Pina et al. [28] also  
174 reported that with adding of concentration, ruminal pH decreased. This is mainly due to that the  
175 increase content of NFC resulted in the increase of VFA, while the low content of NDF resulted in  
176 the decreased rumination in sheep and decreased saliva to the rumen. Thus, the average pH of  
177 group IV was significantly lower than the other groups. The results were consistent with other  
178 studies[29-30]. Yang et al [31] pointed out that when pH was lower than 6.0 for a long time, the  
179 sheep would be in a long-term pathological state. The results showed that the average pH in group  
180 IV was near to 6.0 , it may have negative effects on Karakul sheep, and needs to be further  
181 verified.

### 182 **Effects of dietary NFC/NDF on rumen bacteria in Karakul**

183 **sheep**

184 Diets have crucial effects on rumen microorganisms, Jin et al. [32] showed that the number and  
185 diversity of rumen bacteria in goats feed under high grain (71.5%) diet were lower than those with  
186 high forage diet (0% grain). In this study, the results showed that the rumen bacteria diversity in  
187 Karakul sheep decreased with the increase of dietary NFC/NDF, which was consistent with the  
188 results of Liu [33], However Yong et al. [34] showed that there was no significant difference in  
189 the number of rumen bacteria when the sheep were fed with different ratio of forage to  
190 concentration diets, which might be caused by the difference of diets and species.

191 Roughage is the main feed source for ruminants, and rumen bacteria play a crucial role in the  
192 utilization of roughage. Research showed that diets with easily fermentable carbohydrates would  
193 decrease fiber degradation[35], resulting the imbalance of cellulolytic bacterial species.

194 Bacteroidetes play an important role in the degradation of non-fiber substances and Firmicutes  
195 mainly degrade fiber substances. A large number of studies have shown that Bacteroidetes and  
196 Firmicutes are the most dominant flora in the gastrointestinal tract of mammals[36-39]. Li et al.

197 [40] showed that when the calves were fed with two kinds of NFC/NDF diets, Bacteroidetes and  
198 Firmicutes were still the main dominant flora. In this study, the results showed that the relative  
199 abundance of Bacteroidetes and Firmicutes in different dietary NFC/NDF were still the main

200 dominant phylum. Some results showed that the abundance of bacteria in the same sample would  
201 be different if the gene region was sequenced different[41], In our experiment, the region of  
202 V1-V9 was sequenced and the results showed that the relative abundance of rumen bacteroidetes

203 decreased with the increase of dietary NFD/NDF in Karakul sheep, which was consistent with  
204 Ellison[42]. However, when Kim et al. [43] researched on the content of Bacteroidetes in beef

205 cattle by sequencing V1-V3 region, the content of Bacteroidetes in high forage group was  
206 significantly lower than that of high proportion cereal group, which may be due to the difference  
207 of species differences and the sequence regions measured. In addition, the degradation rate of dry  
208 matter and organic matter was higher in group 3, 4 than which in group 1, 2 (the results were  
209 found previously by our team) so other bacteria except Bacteroidetes in Karakul sheep might have  
210 digested non-fiber substances and needs to be further studied.

211 Improving the fiber degradation rate is very important for ruminants. Bacteria and fungi play a  
212 crucial role in the decomposition and utilization of cellulose. In this study, the relative abundance  
213 of Firmicutes reached the highest when the dietary of NFC/NDF were 1.61 and 2.00, which was  
214 consistent with the results that had done before (The result was that: the NDF degradation rate in  
215 dietary NFC/NDF of 1.61 was the highest). *Unidentified-Lachnospiraceae* was the most dominant  
216 genus and its relative abundance reached the highest in group 3 for four periods, which further  
217 identified the results that: NDF degradation rate in dietary NFC/NDF of 1.61 was the highest.

218 In addition, there were many unidentified bacteria in the rumen of Karakul sheep, which might  
219 mean that there were some new species in Karakul sheep and needs to be further studied.

## 220 **Conclusions**

221 The ruminal pH and total diversity of rumen bacteria decreased with the increase of dietary  
222 NFC/NDF. The most dominant phylum, genus and species didn't change with dietary NFC/NDF  
223 and the ruminal bacteria became more stable with prolong of periods in Karakul sheep.

## 224 **Acknowledgments**

225 We are grateful for other tutors and classmates in our department that helped in the experiment.

226 **Author contributions**

227 Contributed reagents/materials/analysis/ tools: X.G.; Performed the experiments: X.P; Analyzed  
228 the data: X.P; Writing- original draft: X.P.; Writing–review editing: X.P., X.G., C.J., J.L., X.Z.,  
229 S.Z., C.L., and A.S..

230 **Supporting information**

231 **S1 Table. The ingredients and nutrient composition of the diet (% of DM).** ①The premix  
232 provided the following per kg of diets: VA 1800 IU, VD3 600 IU, VE 30 mg, Fe 65 mg, Se 0.15  
233 mg, I 0.6 mg, Cu 10 mg, Mn 28 mg, Zn 45 mg, Cu 12 mg. ②Nutrition level was a calculated  
234 value. ③NFC= (1NDFCPFatAsh) × 100%.

235 **S2 Table Effects of dietary NFC/NDF on relative abundance of phylum in Karakul sheep**

236 Period I (1~18 d), II (19~36 d), III (37~54 d) and IV (55~72 d) and Group 1, 2, 3, 4 means four  
237 groups of sheep treated with four dietary levels of NFC/NDF (0.78, 1.23, 1.61, 2.00 respectively).  
238 The same as below.

239 **S3 Table Effects of dietary NFC/NDF on relative abundance of genus in Karakul sheep**

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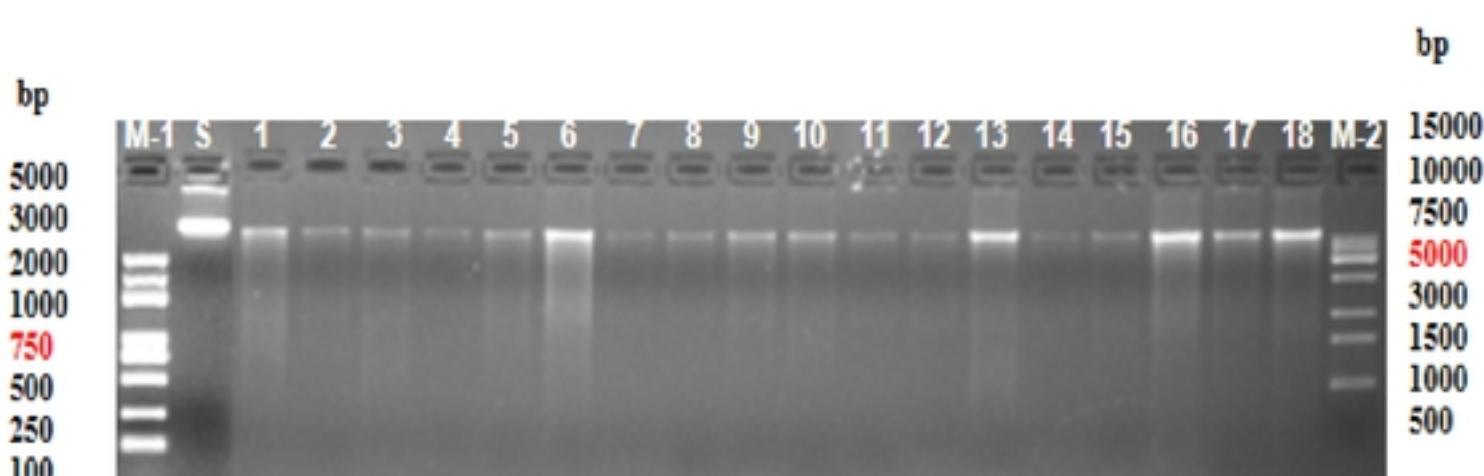
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1 **Table 1. Effects of different NFC/NDF on rumen fluid pH**

| Items      | pH    |        |        |       | SEM  | <i>P</i> -value |
|------------|-------|--------|--------|-------|------|-----------------|
|            | 1     | 2      | 3      | 4     |      |                 |
| Period I   | 6.36  | 6.32   | 6.26   | 6.05  | 0.07 | 0.507           |
| Period II  | 6.42a | 6.28ab | 6.03bc | 6.01c | 0.06 | 0.015           |
| Period III | 6.68a | 6.48ab | 6.34bc | 6.19c | 0.06 | 0.012           |
| Period IV  | 6.52a | 6.27ab | 6.16b  | 6.05b | 0.06 | 0.016           |

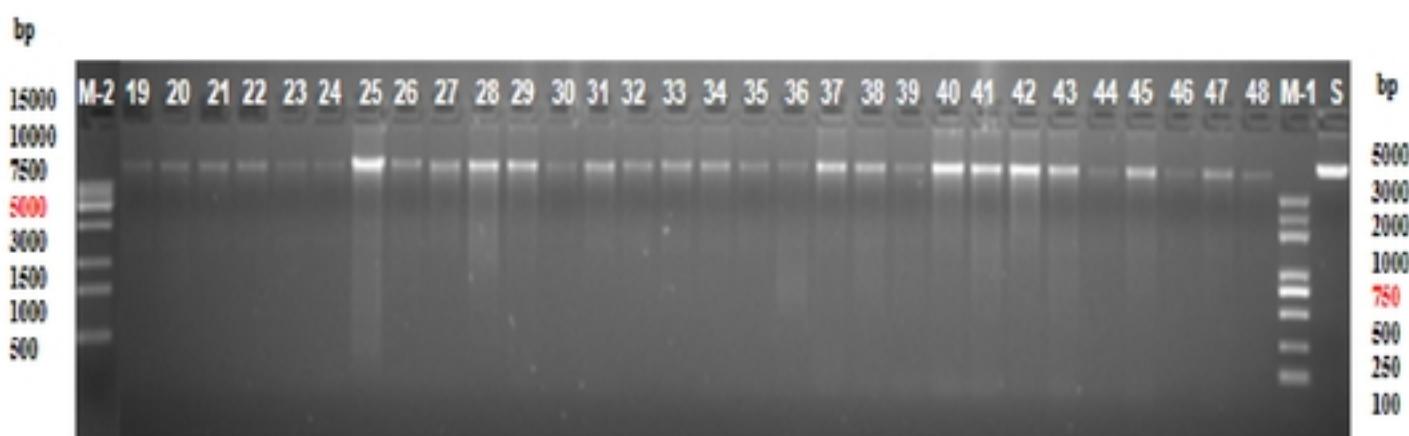
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2 Note: In the same row, values with no letter or the same letter superscripts mean no significant  
 3 difference (*P*>0.05), while with different small letter superscripts mean significant difference (*P*  
 4 <0.05). Period I (1~18 d), II (19~36 d), III (37~54 d) and IV (55~72 d) and Group 1, 2, 3, 4  
 5 means four groups of sheep treated with four dietary levels of NFC/NDF (0.78, 1.23, 1.61, 2.00  
 6 respectively). The same as Table 3.



7

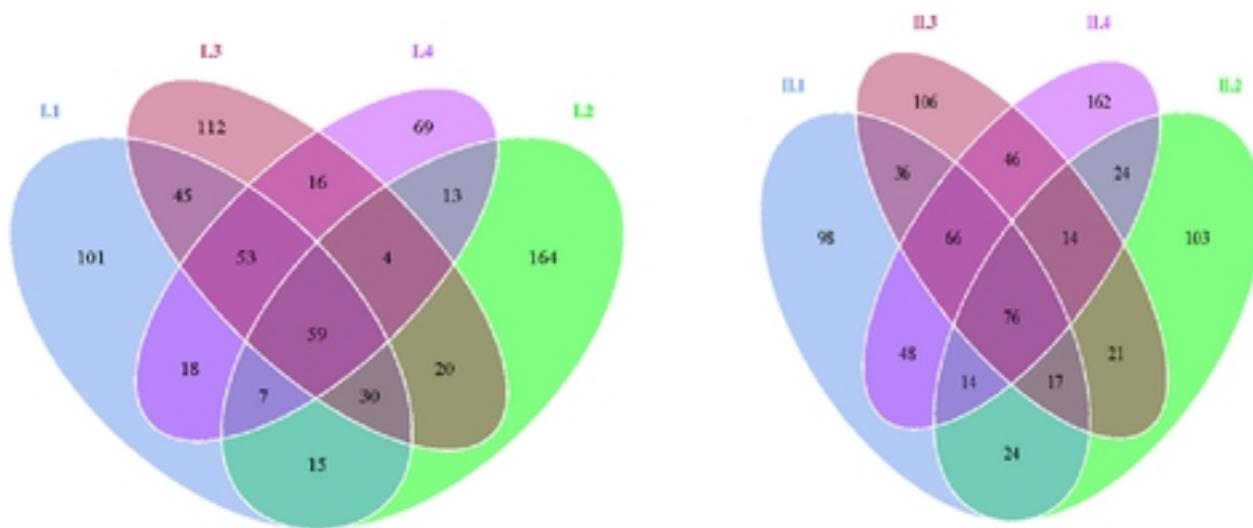
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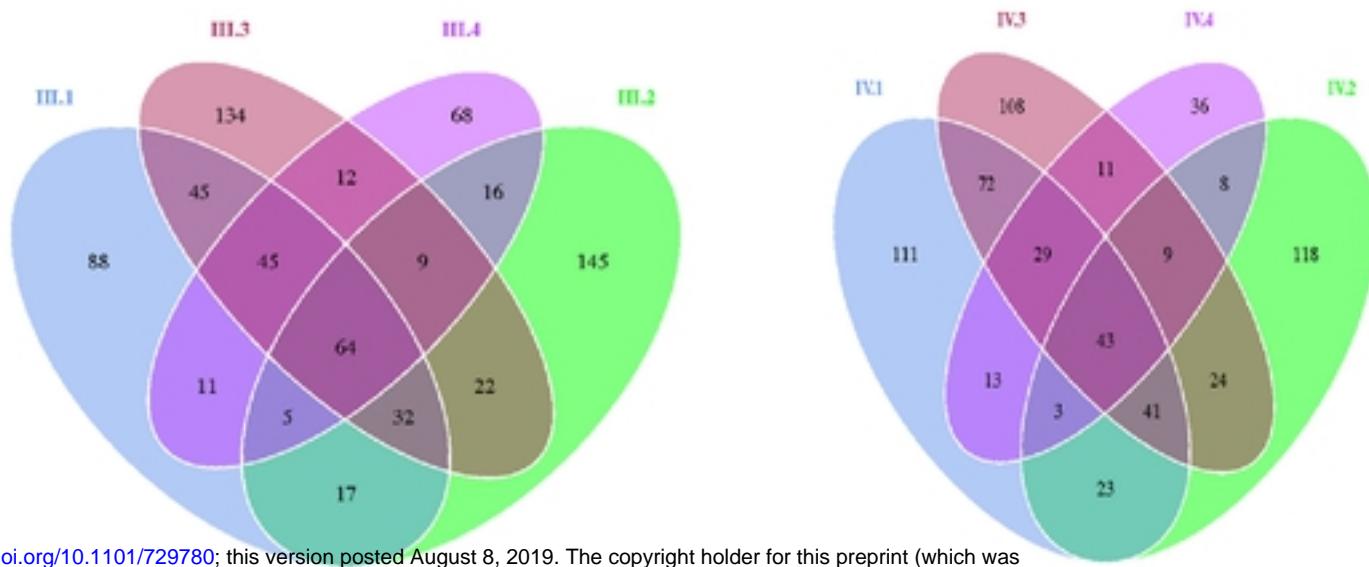
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10 **Fig. 1. Extraction of rumen bacterial DNA.** The DNA of forty-eight samples from four periods  
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11 samples were extracted, the order of 1~3, 4~6, 7~9 and 10~12 means samples in group 1, 2, 3 and  
 12 group 4 of period I respectively, each group with three replicates. The following order are as  
 13 period I.



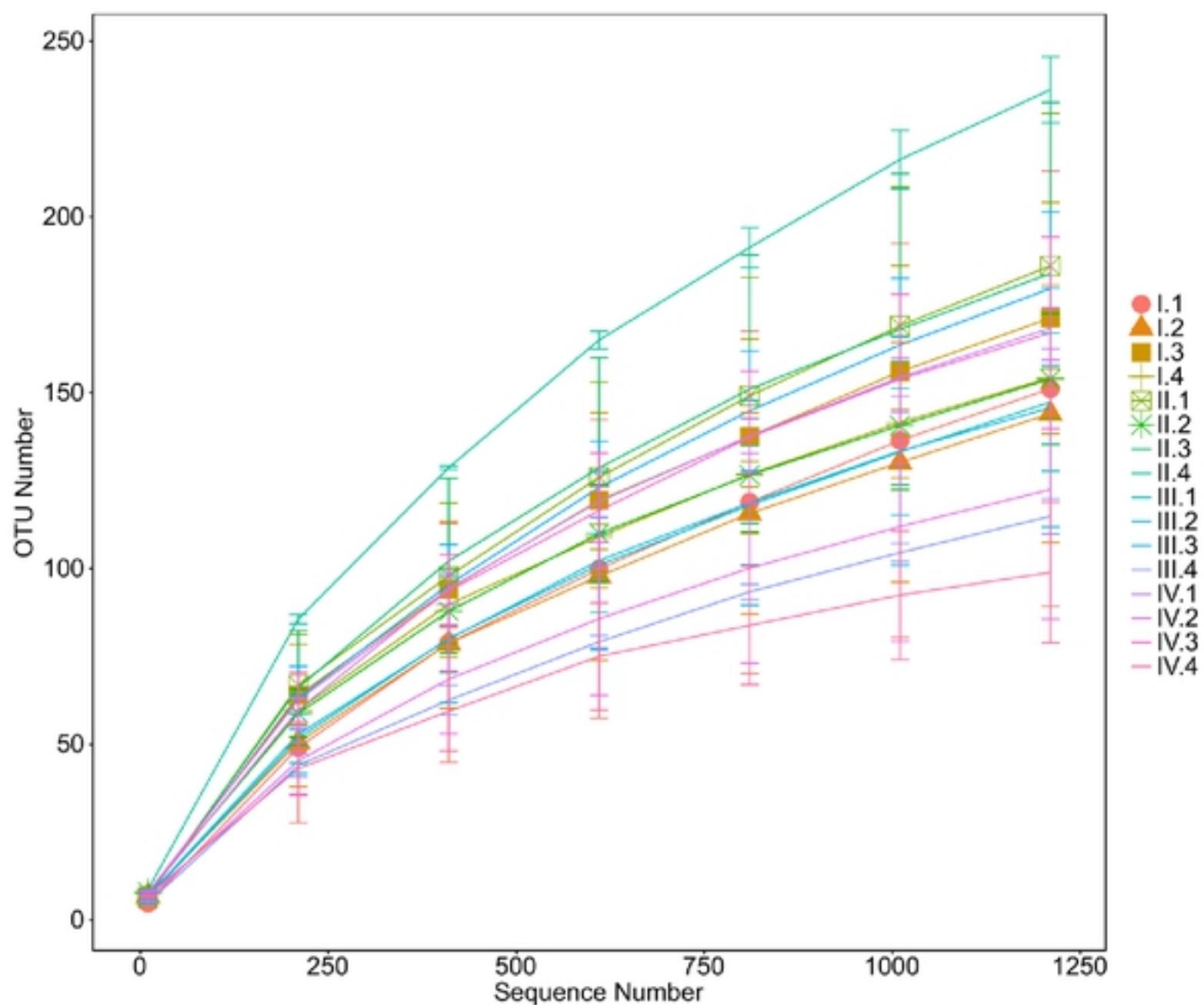
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15

16 **Fig. 2. Vene graph of microflora in rumen fluid of Karakul sheep.** The amounts of OTUs in  
17 each group were shown and four groups of one period were formed in one Vene graph.



18

19 **Fig. 3. OTU dilution curve of bacteria in the rumen of Karakul sheep.** Rarefaction curves of  
20 OTUs clustered at 97% sequence identity across different samples.

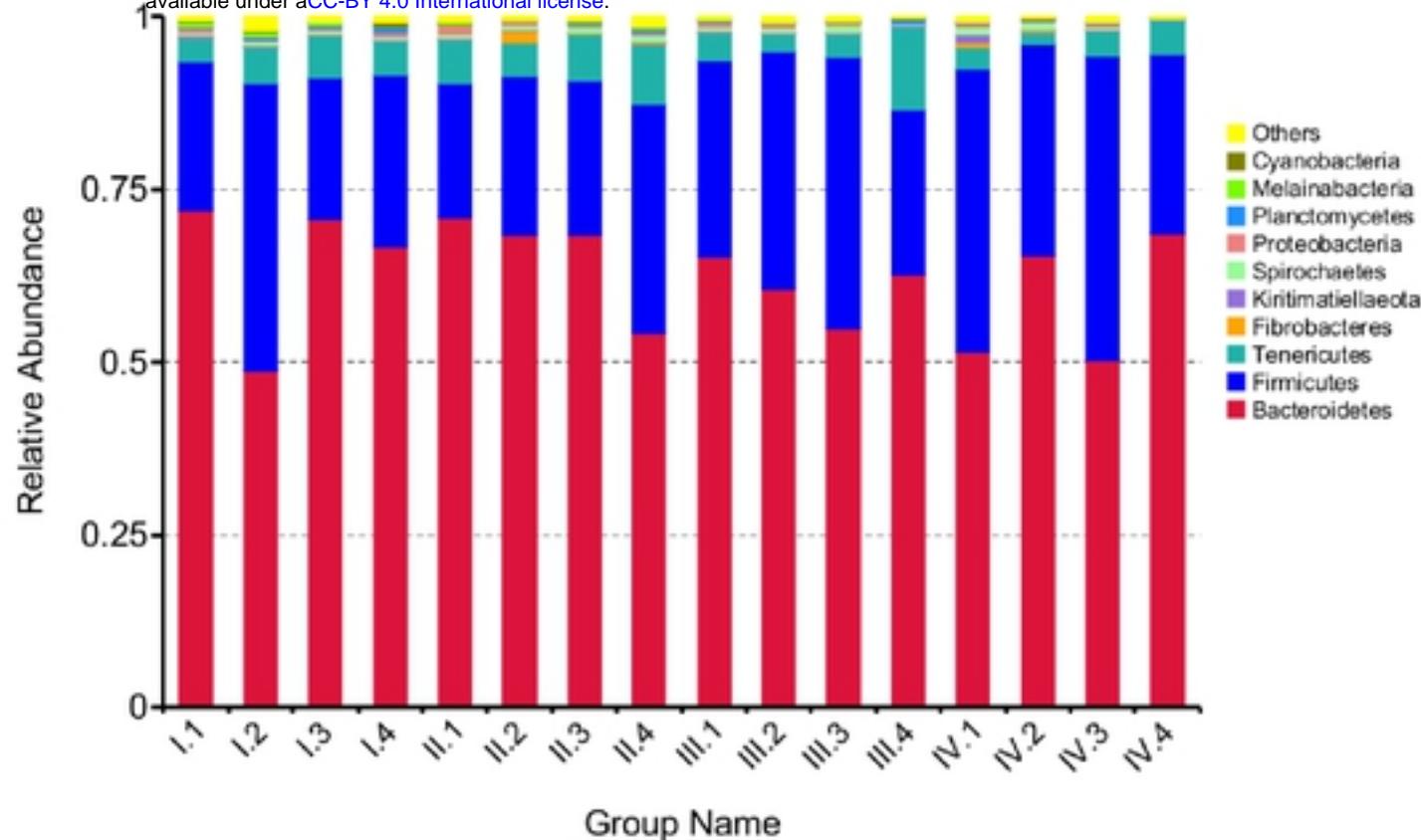
21 **Table 2. Analysis of Alpha diversity of rumen liquid samples at 0.03 distance**

| Items |   | observed_s<br>pecies | Shannon | Simpson | Chao1   | ACE     |
|-------|---|----------------------|---------|---------|---------|---------|
| <hr/> |   |                      |         |         |         |         |
|       | 1 | 151                  | 3.488   | 0.631   | 295.35  | 309.08  |
| I     | 2 | 144                  | 4.447   | 0.853   | 259.414 | 271.994 |
|       | 3 | 171                  | 4.594   | 0.799   | 332.167 | 340.381 |
|       | 4 | 154                  | 4.946   | 0.897   | 232.5   | 252.125 |
| <hr/> |   |                      |         |         |         |         |
|       | 1 | 186                  | 4.536   | 0.787   | 321.413 | 357.069 |
| II    | 2 | 154                  | 5.088   | 0.927   | 246.385 | 274.676 |
|       | 3 | 184                  | 4.865   | 0.838   | 315.519 | 348.306 |
|       | 4 | 236                  | 6.127   | 0.961   | 399.36  | 417.33  |
| <hr/> |   |                      |         |         |         |         |
|       | 1 | 147                  | 4.465   | 0.863   | 243.969 | 295.035 |
| III   | 2 | 146                  | 4.545   | 0.874   | 233.092 | 263.23  |
|       | 3 | 180                  | 5.024   | 0.906   | 275.943 | 318.957 |
|       | 4 | 115                  | 3.786   | 0.798   | 198.268 | 208.16  |
| <hr/> |   |                      |         |         |         |         |

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|    |   |     |       |       |         |         |
|----|---|-----|-------|-------|---------|---------|
|    | 1 | 168 | 4.94  | 0.899 | 246.109 | 288.862 |
| IV | 2 | 122 | 3.992 | 0.81  | 338.777 | 248.709 |
|    | 3 | 167 | 5.08  | 0.912 | 250.554 | 293.258 |
|    | 4 | 99  | 4.36  | 0.879 | 139.488 | 144.216 |

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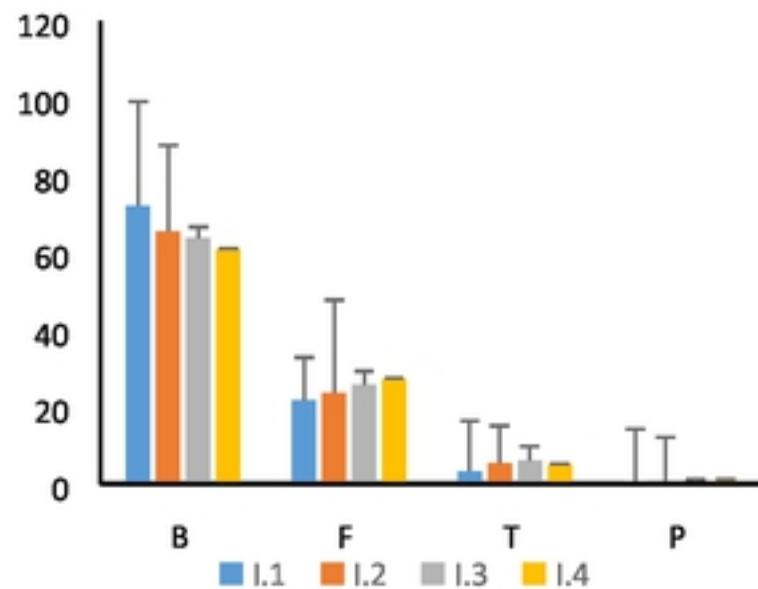


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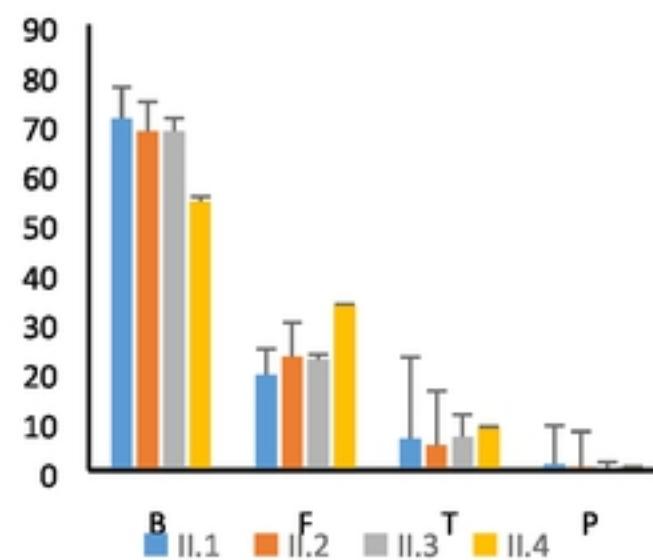
23 **Fig. 4. The column chart of the main dominant phylum in Karakul sheep fed with different**  
 24 **NFC/NDF diets.** A color-coded bar plot showing the average bacterial phylum distribution across  
 25 the different age groups that were sampled.

26 a

b



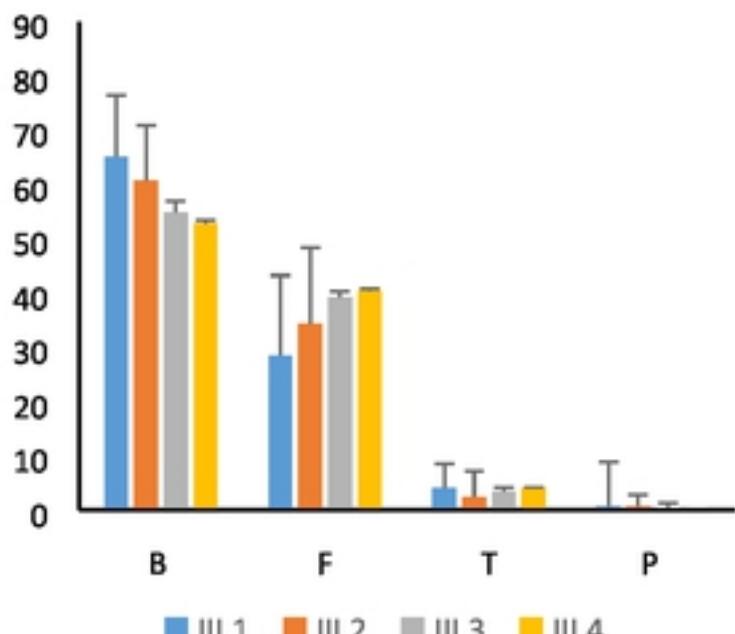
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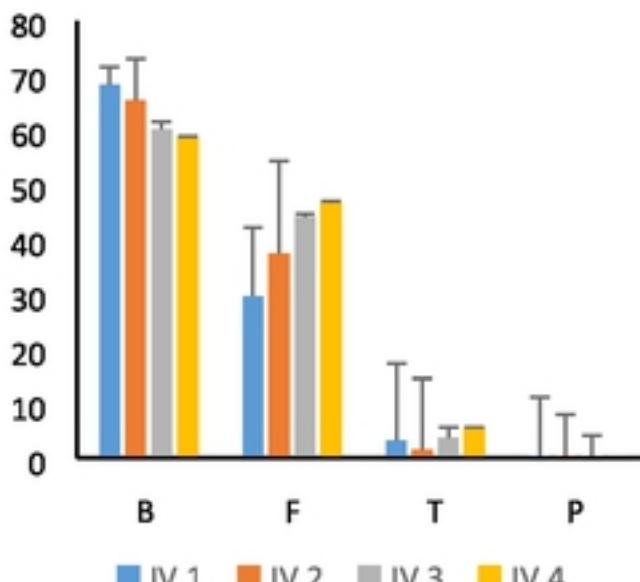
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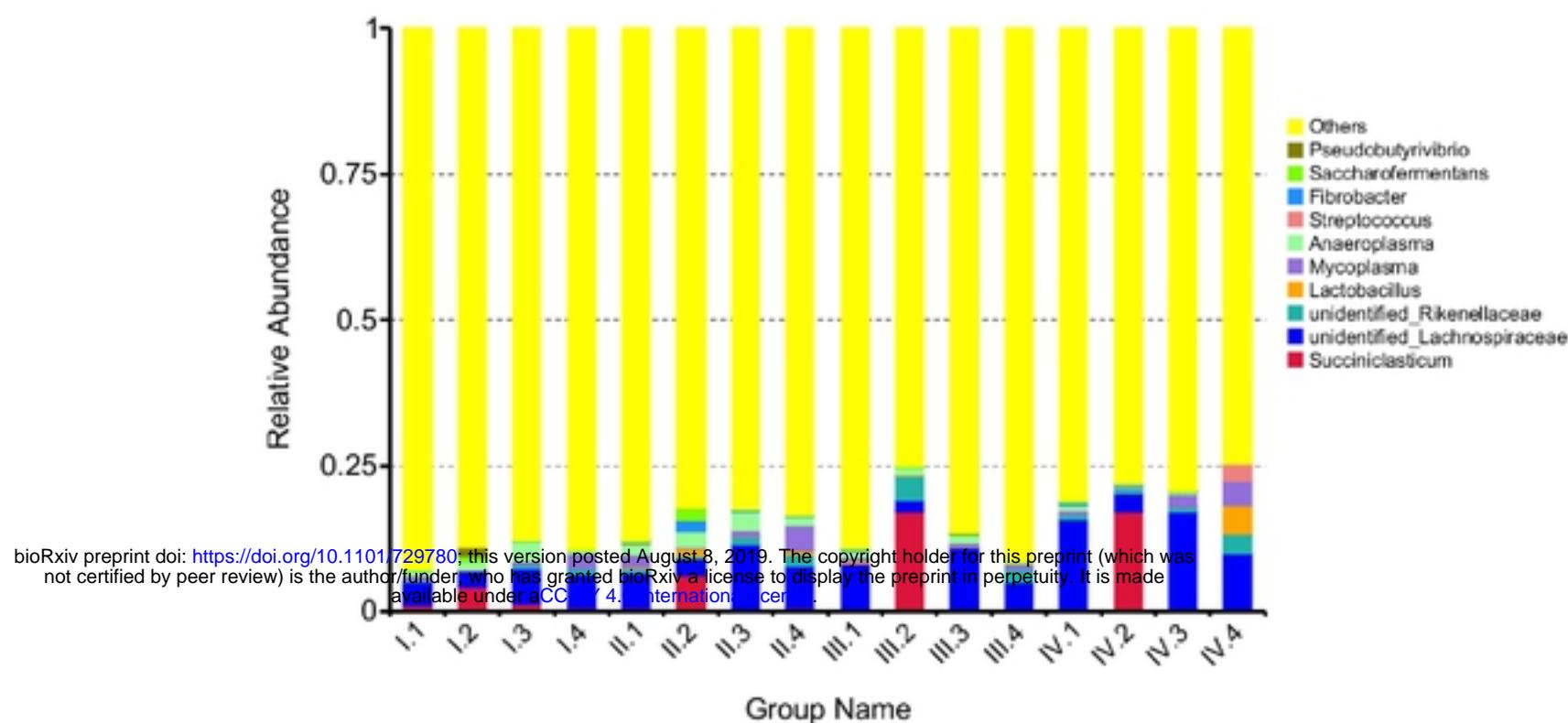
30 Fig. 5(a-d) . Effects of different NFC/NDF diets on relative abundance (% reads) of rumen

31 phylum in Karakul Sheep

32 Note: B means Bacteroidetes, F means Firmicutes, T means Tenericutes, P means

33 Proteobacteria; a, b, c, d represents the experiment period of I, II, III, IV respectively.

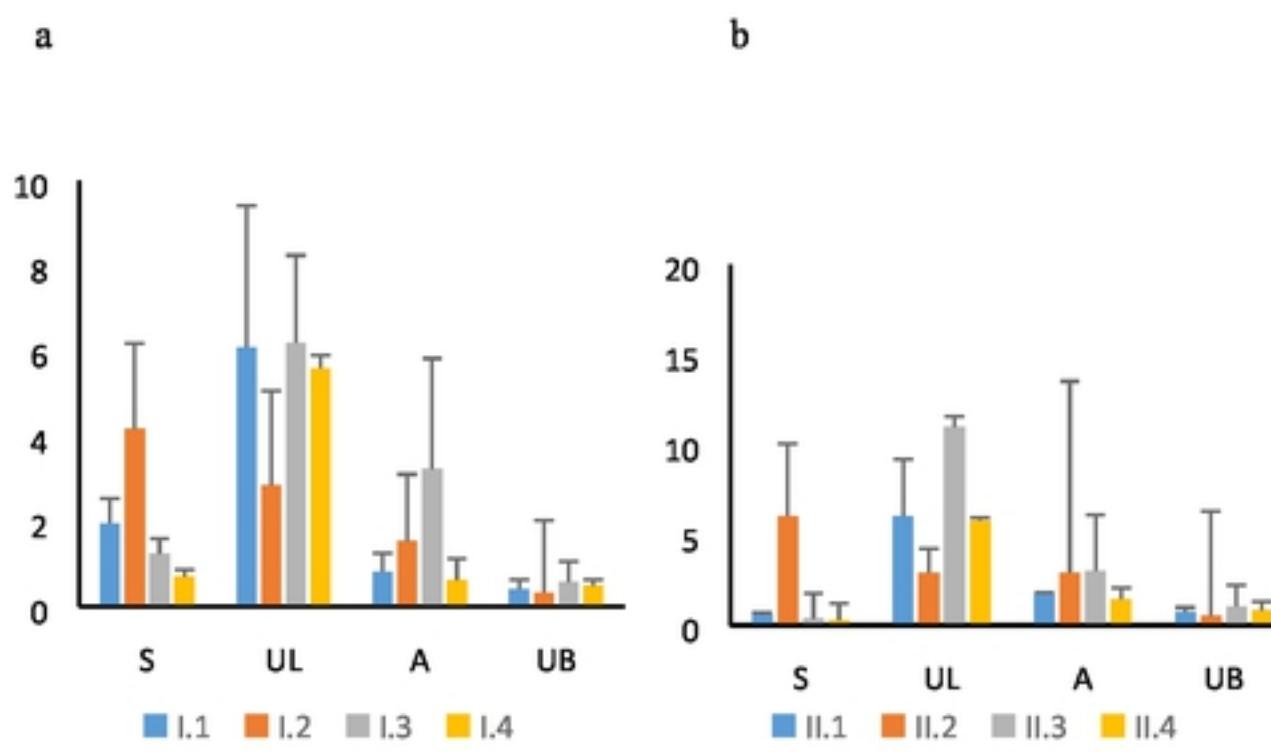
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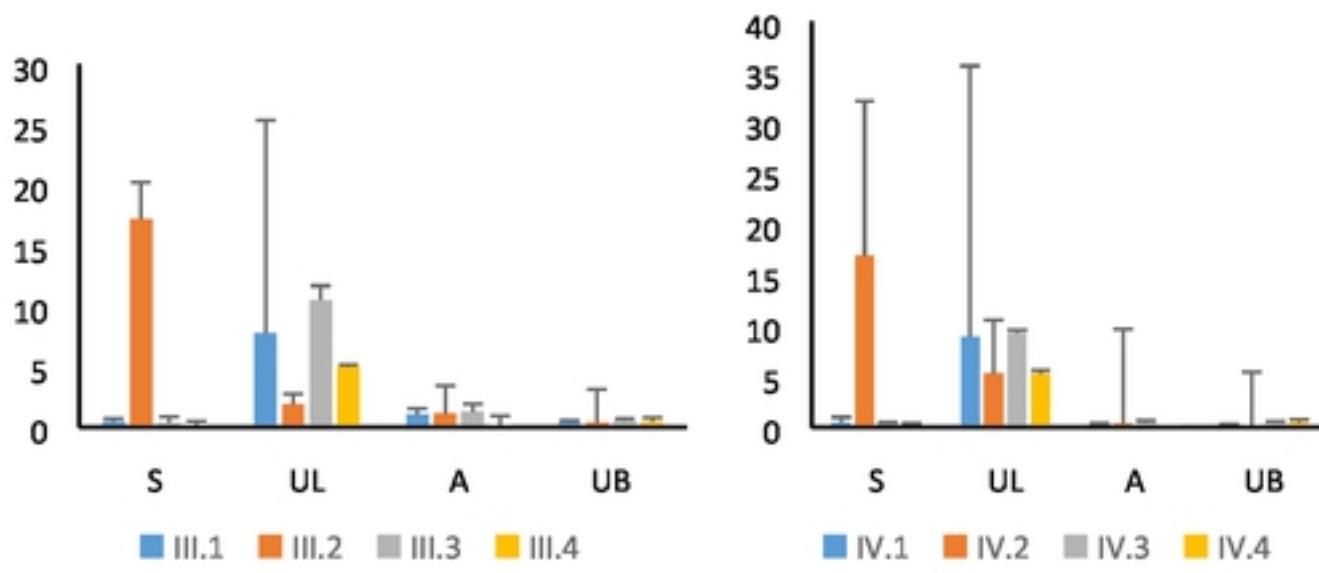
36 **Fig. 6. The column chart of the main dominant genus in Karakul sheep fed with different**  
 37 **NFC/NDF diets. A color-coded bar plot showing the average bacterial genera distribution across**  
 38 **the different age groups that were sampled**

39



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43 **Fig. 7 (a-d): Effects of different AFC/NDF diets on relative abundance (% reads) of**

44 **rumen genus in Karakul Sheep.** Note: S means *Succinivibacter*, UL means  
 45 *unidentified-Lachnospiraceae*, A means *Anaeroplasma*, and UB means *unidentified*  
 46 *Bacteroidales*, a, b, c, d represents the experiment period of I, II, III, IV respectively.

47

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48

49 Relative abundance of community (%)

50 **Fig. 8. Heat map of the rumen bacteria composition at species level.** The heat map indicates

51 the relative percentage of each species for the different NFC/NDF group sampled.

52 **Table 3. Effects of different NFC/NDF on the relative abundance (%) of cellulose-degrading**

## Period I

| Species                          | 1      | 2      | 3      | 4      | SEM   | P-value |
|----------------------------------|--------|--------|--------|--------|-------|---------|
| <i>Butyrivibrio-fibrisolvens</i> | 2.664a | 0.466b | 2.990a | 2.123a | 0.371 | 0.035   |
| <i>Fibrobacter-sp-UWCM</i>       | 0.027  | 0.027  | 0.027  | -      | 0.006 | 0.441   |
| <i>Ruminococcus-slavefaciens</i> | 0.027  | 0.054  | 0.051  | -      | 0.038 | 0.216   |
| <i>Ruminococcus-albus</i>        | -      | 0.055  | -      | -      | 0.009 | 0.052   |

## Period II

| Species                          | 1      | 2      | 3      | 4      | SEM   | P-value |
|----------------------------------|--------|--------|--------|--------|-------|---------|
| <i>Butyrivibrio-fibrisolvens</i> | 2.689a | 0.548b | 3.123a | 2.406a | 1.132 | 0.027   |
| <i>Fibrobacter-sp-UWCM</i>       | 0.082  | 0.411  | 0.027  | -      | 0.350 | 0.294   |
| <i>Ruminococcus-slavefaciens</i> | -      | 0.055  | 0.050  | 0.027  | 0.036 | 0.290   |
| <i>Ruminococcus-albus</i>        | 0.027  | 0.055  | -      | -      | 0.01  | 0.561   |

## Period III

| Species                          | 1      | 2      | 3      | 4      | SEM   | P-value |
|----------------------------------|--------|--------|--------|--------|-------|---------|
| <i>Butyrivibrio-fibrisolvens</i> | 6.785a | 0.657c | 7.687a | 3.292b | 1.143 | <0.01   |

|                                  |       |       |       |       |       |       |
|----------------------------------|-------|-------|-------|-------|-------|-------|
| <i>Fibrobacter-sp-UWCM</i>       | 0.025 | 0.027 | -     | -     | 0.009 | 0.596 |
| <i>Ruminococcus-slavefaciens</i> | 0.027 | 0.082 | 0.055 | 0.027 | 0.021 | 0.627 |
| <i>Ruminococcus-albus</i>        | -     | -     | -     | -     | -     | -     |

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Period IV

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| Species                          | 1      | 2      | 3      | 4      | SEM   | P-value |
|----------------------------------|--------|--------|--------|--------|-------|---------|
| <i>Butyrivibrio-fibrisolvens</i> | 7.234a | 2.301c | 8.694a | 5.975b | 1.581 | <0.01   |
| <i>Fibrobacter-sp-UWCM</i>       | -      | 0.082  | -      | -      | 0.07  | 0.441   |
| <i>Ruminococcus-slavefaciens</i> | -      | 0.082  | 0.079  | -      | 0.016 | 0.052   |
| <i>Ruminococcus-albus</i>        | -      | 0.085  | -      | -      | 0.013 | 0.063   |

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