

# 1   **Effects of dietary NFC/NDF on rumen microbiomes** 2   **of Karakul sheep based on Three Generations of** 3   **Full-length Amplifiers sequencing**

4   Xuanxuan Pu<sup>1</sup>, Xuefeng Guo<sup>1,2\*</sup>, Chenyu Jiang<sup>1</sup>, Junfeng Liu<sup>1,2</sup>, Xiuping Zhang<sup>1,2</sup>, Sujiang  
5   Zhang<sup>1,2</sup>, Long Cheng<sup>3</sup>, Anshan Shan<sup>4</sup>

6   1 College of Animal Science, Tarim University, Alar 843300, Xinjiang, PR China

7   2 Key Laboratory of Tarim Animal Husbandry Science and Technology of Xinjiang Production  
8   and Construction Group, Alar 843300, Xinjiang, PR China

9   3 Faculty of Veterinary & Agricultural Sciences, Dookie Campus, The University of Melbourne,  
10   Victoria 3647, Australia

11   4 Institute of Animal Nutrition, Northeast Agricultural University, Harbin 150030, PR China

12

13

14   The research was supported by the National Natural Science Foundation Project (31760680) and  
15   Xinjiang Production and Construction Group with the young and middle-aged innovation talents  
16   fund (No. 2016BC001).

17

18

19   \* Corresponding authors: gxfdky@126.com.

20

21

22

23

24

25

## 26 **Abstract**

27 An study was was conducted to investigate the effects of dietary(non fibrous carbohydrate)  
 28 NFC/(neutral detergent fiber)NDF on ruminal bacteria in Karakul sheep. Twelve Karakul sheep  
 29 were assigned randomly to four dietary treatments of NFC/NDF (0.78, 1.23, 1.61 and 2.00  
 30 respectively) as group 1, 2, 3 to 4. The experiment lasted for four periods, period I (1~18 d), II  
 31 (19~36 d), III (37~54 d) and IV (55~72 d). Ruminal digesta were collected consecutively for three  
 32 days to measure pH and bacteria per period. The results indicated that the average ruminal pH and  
 33 amounts of OTUs were decreased with the increase of dietary NFC/NDF for four periods. At  
 34 phylum level, Bacteroidetes and Firmicutes were the predominant bacteria of four periods,  
 35 Bacteroidetes were decreased, while the relative abundance of Firmicutes was increased with  
 36 dietary NFC/NDF for four periods, but the difference wasn't significant ( $P>0.05$ ). At genus level,  
 37 the most relative abundance genus was unidentified-Lachnospiraceae which reached the highest in  
 38 group 3 for four periods, but the difference wasn't significant ( $P>0.05$ ). Conclusion: ruminal pH  
 39 and bacteria were decreased with the increase of dietary NFC/NDF and the most dominant  
 40 bacteria were not change with dietary NFC/NDF and periods in Karakul sheep.

## 41 **Introduction**

Rumen plays an important role in the growth and production of ruminants and it contains a large number of rumen microorganisms (*Bacteria*, *Protozoa*, *Eukarya* and *Archaea*[1-3]). Rumen microorganisms breakdown food to provide volatile fatty acids, bacterial protein, and so on to the host animal[4]. Yang et al.[5] confirmed that 80% of the starch, 50% of the fibre and 60% of the organic matter in the diet were fermented in the rumen to provide energy for the host. Meanwhile, after a long-term selection and evolution, rumen microbiomes and the host have formed a symbiotic relationship to maintain the host's health[6]. Dietary regulation has an important effect on rumen fermentation[7-9] and microbiome[10-11]. Wei et al.[12] showed that with the increase of dietary NFC/NDF, ruminal pH decreased significantly, furthermore the composition of rumen microbial flora also changed, and the total number of rumen bacteria decreased in goats. However, there are scarce studies on rumen microbiome structure changes with dietary NFC/NDF in Karakul sheep and the rumen is very complex in which microbiome may change again with prolong of periods. NDF plays an important role in dry matter intake (DMI) and feed digestibility[13-14], and NFC in diets is another factor that affect DMI, Hall et al. reported that NFC would be degraded rapidly in the rumen[14]. The development of technology makes it more accurate to study on rumen microbiomes, Three Generation of Full-length Amplifiers sequencing is one of them, which can improve the resolution of species identification, and improve the accuracy of microbial species composition identification in the samples[15-16]. In this experiment, ruminal pH and microbiome were measured for four periods to investigate effects of dietary NFC/NDF on ruminal microbiome in Karakul sheep.

## Materials and Methods

## **Animals and Dietary composition**

All experimental procedures were approved by Tarim University Animal Care and Use Committee, and humane animal care were followed throughout the experiment. Twelve Karakul sheep with similar age and weight ( $35.3 \pm 3.3$  kg) were fitted with permanent fistula and were randomly assigned into four dietary treatments of NFC/NDF (0.78, 1.23, 1.61, 2.00 respectively) as group 1, 2, 3 and group 4, each group with three replicates. They all received vaccines for parasites before the adaption period, and were fed meeting the standards for raising meat and sheep in the People's Republic of China[17]. All sheep were housed individually in metabolic cages (1.2 m  $\times$  1.5 m) and fed the experimental diet individually twice a day at 9:00 a.m. and 8:00 p.m with free access to water. The ingredients and nutrient level of the diet were shown in S1 Table.

## **The experimental design and sample collection**

The experiment lasted for 72 d including four periods, including period I (1~18 d), II (19~36 d), III (37~54 d) and IV (55~72 d). Each period lasted for 18 d. The first 15 d for adaption and 3 d for samples, feed intake and defecation rule were studied in the adaption period. The ruminal digesta were sampled consecutively before morning feeding for three days, the sheep of one group were collected together for 50 mL, and then, were stored in -80 °C to investigate on rumen microorganisms. Meanwhile, the ruminal pH was measured after feeding of 0, 1, 3, 6 and 9 h using pH meter (FE22).

## **DNA Extraction, PCR and Pacio sequencing**

The total genetic DNA was extracted using QIAamp Fast DNA Stool Mini Kit (QIAGEN, Shanghai) according to the illustration and the extracted DNA was detected by 1% agarose gel. The V1-V9 regions of 16S rDNA were amplified by PCR from the extracted DNA using the universal primers: F, 5'-AGAGTTTGATCCTGGCTCAG-3'; R, 5'-GNTACCTTGTTACGACTT-3' (synthesized by Biological engineering co., Ltd). PCR was carried out in triplicate 50-μL reactions which containing 2 μL Primer Mix (1uM), 5 ng gDNA, 1 μL Trans Fastpfu, 10 μL 5× Buffer, 5 μL 5× StimuLate, 5 μL dNTPs (2.5mM each), 27 μL NFW. Thermocycling parameters were as follows: 2 min predenaturation at 95 °C; 35 cycles of denaturation at 95 °C for 30 s, Annealing at 60 °C for 40 s, extension at 72 °C for 90 s; and a Final extension at 72 °C for 10 min. The production was detected by 2% agarose gel. PCR products was purified with Gel Extraction Kit (QIAGEN, Shanghai), and the productions were sequenced on PacBio platform.

## The sequence analysis

The 16S rDNA reads were firstly processed to get clean reads by discarding the reads that are shorter than 1340 bp, longer than 1640 bp, and not matching the expected barcodes. Using Uparse software [18] to cluster all Clean Reads of all samples. Operational taxonomic units (OUTs) were formed at the similarity of 97%[19]. The OUTs were annotated by the Mothur and SILVA ( <http://www.arb-silva.de/>) [20] according to the reference taxonomy provided by SSUrRNA database[21]. The OTUs were analyzed by Qiime pipeline (Version 1.9.1) to calculate the richness and diversity indices i.e. observed OTUs, Chao1, Shannon, Simpson, ACE.

## Statistical Analyses

The results of pH were expressed as means using SPSS 17.0. Comparisons between groups were performed with ANOVA followed by Duncan test. The difference of ruminal bacteria was measured and expressed using SPSS 17.0. As well, and the differences were considered to be significant at  $P < 0.05$ .

## Results

### pH

The average pH of 0, 1, 3, 6, 9 h after feeding for four periods were shown in Table 1, the pH was that: group 1 > group 2 > group 3 > group 4 for four periods. There was no significant difference between four groups in period I ( $P > 0.05$ ), while there was significant difference between four groups in period II, III and IV ( $P < 0.05$ ), which showed that the ruminal pH decreased significantly as the dietary NFC/NDF increased.

**Table 1. Effects of dietary NFC/NDF on ruminal pH**

### Extraction DNA of rumen bacteria

The DNA extraction results were shown in Fig. 1. The main band was clear and there was no concentrated band below 500 bp, which indicated that the purity of DNA was well and it could meet the requirement of sequencing.

**Fig. 1. Extraction DNA of rumen bacteria**

### The analysis of basic sequencing data

The OTU number of each group in one period was shown in Venn graph as Fig. 2. The result

showed that the OTUs was that: group 1> group 2 > group 3> group 4 for four periods, which showed that the diversity of rumen microbiome decreased with the increase of NFC/NDF, and the number of each group became more stable with prolong of periods

**Fig. 2. Vene graph of OTUs in Karakur sheep**

## **The analysis of OTU Alpha diversity**

The Alpha diversity is a kind of analysis in the diversity of microbiome, which involves the abundance index of Chao1[22] and ACE[23] and the diversity index of Shannon and Simpson[24]. Before diversity analysis, the dilution curve was drawn by R software (Version 2.15.3) to detect whether the obtained data could fully reflect the distribution of rumen fluid flora in Karakul sheep.

## **OTU dilution curve**

As showed in Fig. 3, the dilution curve of each group keep increasing with the increase of the depth of sequencing, which is indicated that new bacteria had been found. The results showed that the sequencing quantity of each sample could be used to analyze the diversity of flora.

**Fig. 3. OTU dilution curve of bacteria in the rumen of Karakul sheep**

## **Sample diversity index**

The results of Alpha diversity were shown in Table 2. The results showed that there was some difference in the number, richness and diversity of rumen bacterial species in different dietary of NFC/NDF.

**Table 2. Analysis of Alpha diversity of rumen liquid samples at 0.03 distance**

## Structural analysis of rumen microbiome

At phyla level, seventeen different phyla were detected. The S2 table and Fig. 4 showed that the main phylum microbiome didn't change with the dietary NFC/NDF and periods, and Bacteroidetes (52%~72%) and Firmicutes (19%~46%) was the main microbiome for four periods. The relative abundance of Tenericutes was 3% to 8% and the others was less than 1%. Fig. 5 was composed by four main dominant phylum to investigate effects of NFC/NDF on their relative abundance, the abundance of Bacteroidetes and Proteobacteria were that: group 1> group 2> group 3> group 4 for four periods, but the difference wasn't significant( $P>0.05$ ). While the abundance of Firmicutes was that : group 4> group 3> group 2> group 1, the difference wasn't significant( $P>0.05$ ) as well. The abundance of Tenericutes reached the highest in group 4 for four periods.

**Fig. 4. The column chart of the main dominant phylum in Karakul sheep**

**Fig. 5(a~d) . Effects of dietary NFC/NDF on relative abundance (% reads) of rumen phylum in Karakul Sheep**

At the genus level, a total of 77 genera were obtained from sequence alignment. It can be seen from S3 Table and Fig. 6. The main genus microbiome didn't change with the dietary NFC/NDF and periods. The highest relative abundance of genus was a kind of semi-cellulose degrading bacteria, *unidentified-Lachnospiraceae* (1.86%~16.68%). The following relative abundance of genus was *Succiniclasticum* (0.12%~17.03%). Fig. 7 was composed by four main dominant genus to investigate effects of dietary NFC/NDF on their relative abundance, the abundance of *Succiniclasticum* was that: group 2> group 1> group 3> group 4, and the difference wasn't



significant ( $P>0.05$ ). The relative abundance of *unidentified-Lachnospiraceae*, *Anaeroplasma* and *unidentified-Bacteroidales* reached the highest in group 3 for four periods, and the difference wasn't significant ( $P>0.05$ ) as well.

**Fig. 6. The column chart of the main dominant genus in Karakul sheep**

**Fig. 7(a~d) . Effects of dietary NFC/NDF on relative abundance (% reads) of rumen genus in**

## **Discussion**

### **Effects of dietary NFC/NDF on ruminal pH**

pH is the directest index affecting rumen fermentation[25] and diets are key factors affecting pH. The results showed that the ruminal average pH decreased with the increase of NFC/NDF for four periods, which was approved with T.Ma et al.[26]. Agle et al. [27] and Pina et al. [28] also reported that with adding of concentration, ruminal pH decreased. This is mainly due to that the increase content of NFC resulted in the increase of VFA, while the low content of NDF resulted in the decreased rumination in sheep and decreased saliva to the rumen. Thus, the average pH of group IV was significantly lower than the other groups. The results were consistent with other studies[29-30]. Yang et al [31] pointed out that when pH was lower than 6.0 for a long time, the sheep would be in a long-term pathological state. The results showed that the average pH in group IV was near to 6.0 , it may have negative effects on Karakul sheep, and needs to be further verified.

### **Effects of dietary NFC/NDF on rumen bacteria in Karakul**

## sheep

Diets have crucial effects on rumen microorganisms, Jin et al. [32] showed that the number and diversity of rumen bacteria in goats feed under high grain (71.5%) diet were lower than those with high forage diet (0% grain). In this study, the results showed that the rumen bacteria diversity in Karakul sheep decreased with the increase of dietary NFC/NDF, which was consistent with the results of Liu [33], However Yong et al. [34] showed that there was no significant difference in the number of rumen bacteria when the sheep were fed with different ratio of forage to concentration diets, which might be caused by the difference of diets and species.

Roughage is the main feed source for ruminants, and rumen bacteria play a crucial role in the utilization of roughage. Research showed that diets with easily fermentable carbohydrates would decrease fiber degradation[35], resulting the imbalance of cellulolytic bacterial species. Bacteroidetes play an important role in the degradation of non-fiber substances and Firmicutes mainly degrade fiber substances. A large number of studies have shown that Bacteroidetes and Firmicutes are the most dominant flora in the gastrointestinal tract of mammals[36-39]. Li et al. [40] showed that when the calves were fed with two kinds of NFC/NDF diets, Bacteroidetes and Firmicutes were still the main dominant flora. In this study, the results showed that the relative abundance of Bacteroidetes and Firmicutes in different dietary NFC/NDF were still the main dominant phylum. Some results showed that the abundance of bacteria in the same sample would be different if the gene region was sequenced different[41], In our experiment, the region of V1-V9 was sequenced and the results showed that the relative abundance of rumen bacteroidetes decreased with the increase of dietary NFD/NDF in Karakul sheep, which was consistent with Ellison[42]. However, when Kim et al. [43] researched on the content of Bacteroidetes in beef

cattle by sequencing V1-V3 region, the content of Bacteroidetes in high forage group was significantly lower than that of high proportion cereal group, which may be due to the difference of species differences and the sequence regions measured. In addition, the degradation rate of dry matter and organic matter was higher in group 3, 4 than which in group 1, 2 (the results were found previously by our team) so other bacteria except Bacteroidetes in Karakul sheep might have digested non-fiber substances and needs to be further studied.

Improving the fiber degradation rate is very important for ruminants. Bacteria and fungi play a crucial role in the decomposition and utilization of cellulose. In this study, the relative abundance of Firmicutes reached the highest when the dietary of NFC/NDF were 1.61 and 2.00, which was consistent with the results that had done before (The result was that: the NDF degradation rate in dietary NFC/NDF of 1.61 was the highest). *Unidentified-Lachnospiraceae* was the most dominant genus and its relative abundance reached the highest in group 3 for four periods, which further identified the results that: NDF degradation rate in dietary NFC/NDF of 1.61 was the highest.

In addition, there were many unidentified bacteria in the rumen of Karakul sheep, which might mean that there were some new species in Karakul sheep and needs to be further studied.

## Conclusions

The ruminal pH and total diversity of rumen bacteria decreased with the increase of dietary NFC/NDF. The most dominant phylum, genus and species didn't change with dietary NFC/NDF and the ruminal bacteria became more stable with prolong of periods in Karakul sheep.

## Acknowledgments

We are grateful for other tutors and classmates in our department that helped in the experiment.

## Author contributions

Contributed reagents/materials/analysis/ tools: X.G.; Performed the experiments: X.P; Analyzed the data: X.P; Writing- original draft: X.P.; Writing–review editing: X.P., X.G., C.J., J.L., X.Z., S.Z., C.L., and A.S..

## Supporting information

**S1 Table. The ingredients and nutrient composition of the diet (% of DM).** ①The premix provided the following per kg of diets: VA 1800 IU, VD3 600 IU, VE 30 mg, Fe 65 mg, Se 0.15 mg, I 0.6 mg, Cu 10 mg, Mn 28 mg, Zn 45 mg, Cu 12 mg. ②Nutrition level was a calculated value. ③NFC=  $(1 - \text{NDF} - \text{CP} - \text{Fat} - \text{Ash}) \times 100\%$ .

### **S2 Table Effects of dietary NFC/NDF on relative abundance of phylum in Karakul sheep**

Period I (1~18 d), II (19~36 d), III (37~54 d) and IV (55~72 d) and Group 1, 2, 3, 4 means four groups of sheep treated with four dietary levels of NFC/NDF (0.78, 1.23, 1.61, 2.00 respectively). The same as below.

### **S3 Table Effects of dietary NFC/NDF on relative abundance of genus in Karakul sheep**

## References

1. Elizabeth M Ross, Peter J Moate, Carolyn R Bath, et al. (2012) High throughput whole rumen metagenome profiling using untargeted massively parallel sequencing[J]. BMC Genetics. 53.
2. Kumar, S, Indugu N, Vecchiarelli B, Pitta, DW (2015) Associative patterns among anaerobic fungi, methanogenic archaea, and bacterial communities in response to changes in diet and

245 age in the rumen of dairy cows[J]. FRONTIERS IN MICROBIOLOGY..

246 3. Hespell RB, Akin DE, Dehority BA (1997). In: Mackie RI, White BA, Isaacson R (eds)

247 Gastrointestinal microbiology, vol 2. New York: Chapman and Hall. pp. 59–186

248 4. Deng W, Xi DM, Mao H, et al. (2008) The use of molecular techniques based on ribosomal

249 RNA and DNA for rumen microbial ecosystem studies:a review[J].Molecular Biology

250 Reports. Vol. 35(No.2): 265-274.

251 5. Shuo Yang (2018) Effects of Feeding Types and Breeds on Rumen Methanogens and Related

252 Microflora in Inner Mongolia Cashmere Goats[D]. Inner Mongolia Agriculture University.

253 6. Rawls JF, Samuel BS, Gordon JI (2004) Gnotobiotic zebrafish reveal evolutionarily

254 conserved responses to the gut microbiota[J]. Proceedings of the National Academy of

255 Science of the United States of America. Vol. 101(No.13): 4596-4601.

256 7. ZHANG Jianxun, LIU Jiangbo, XUE Bai (2013) Forage to Concentrate Ratio: Effects on

257 Rumen Fermentation in Nanjiang Brown Goats in Vitro[J]. Journal of animal nutrition,

258 25(04): 870-877.

259 8. Corley R N, Murphy M R (2004) An in vitro technique for measuring the production rate of

260 volatile fatty acids in the rumen under dynamic conditions[J]. Small Ruminant Res, 54(3):

261 219-225.

262 9. Song SD, Chen GJ, Guo CH, Rao KQ, Gao YH, Peng ZL, et al. (2018) Effects of exogenous

263 fibrolytic enzyme supplementation to diets with different NFC/NDF ratios on the growth

264 performance, nutrient digestibility and ruminal fermentation in Chinese domesticated black

265 goats[J].Animal Feed Science and Technology, 170-177.

266 10. Kumar S, Indugu N, Vecchiarelli B, Pitta DW (2015) Associative patterns among anaerobic  
267 fungi, methanogenic archaea, and bacterial communities in response to changes in diet and  
268 age in the rumen of dairy cows[J]. FRONTIERS IN MICROBIOLOGY.

269 11. Han X, Yang Y, Yan H, Wang X, Qu L, Chen Y (2015) Rumen Bacterial Diversity of 80 to  
270 110-Day-Old Goats Using 16S rRNA Sequencing[J]. PLoS ONE.2015, Vol. 10(No.2):  
271 e0117811.

272 12. Wei Deyong, Zhu Weiyun, Mao Shengyong (2012) Effects of dietary NFC/NDF ratios on  
273 rumen fermentation and rumen microbial flora in goats [J]. China Agricultural Science, 45  
274 (07): 1392/1398.

275 13. Bowman J G P, Sowell B F, Surber L M M, Daniels T K (2004) Nonstructural carbohydrate  
276 supplementation of yearling heifers and range beef caws[J]. Anim. Sci. 82, 2724–2733.

277 14. Hall M B (2003) Challenges with nonfiber carbohydrate methods[J]. Journal of Animal  
278 Science. Vol. 81(NO.12): 3226-3232.

279 15. DeSantis T Z, et al. (2006) NAST: a multiple sequence alignment server for comparative  
280 analysis of 16S rRNA genes. Nucleic acids research 34. suppl 2: W394-W399.

281 16. Ondov Brian D, Nicholas H, Bergman and Adam M. Phillippy (2011) Interactive  
282 metagenomic visualization in a Web browser. BMC bioinformatics 12. 1: 385.

283 17. Agricultural Industry Standard of the people's Republic of China-Meat Sheep feeding

284 Standard (NY/T816-2004)[J]. Hunan feed ,2006 (06): 9-15。

285 18. Edgar Robert C (2013) UPARSE: highly accurate OTU sequences from microbial  
286 amplicon reads. Nature methods 10.10 (2013): 996-998.

287 19. SIMPSON J M, MCCracken V J, WHITE B A, et al. (1999) Application of  
288 denaturant gradient gel electrophoresis for the analysis of the porcine gastrointestinal  
289 microbiota[J]. Journal of Microbiological Methods, 36: 167-179.

290 20. Wang Qiong et al. (2007) Naive Bayesian classifier for rapid assignment of rRNA sequences  
291 into the new bacterial taxonomy. Applied and environmental microbiology 73.16:  
292 5261-5267.

293 21. Quast C, Pruesse E, et al. (2013) The SILVA ribosomal RNA gene database project: improved  
294 data processing and web-based tools. Nucl. Acids Res: D590-D596.

295 22. Chao A (1984) Nonparametric-estimation of the number of classes in a population.  
296 Scand. J. Stat. 11:265–270.

297 23. Chao A, Lee SM (1992) Estimating the number of classes via sample coverage. J. Am.  
298 Stat. Assoc. 87: 210–217.

299 24. Sar C, Santoso B, Mwenya B, Gamo Y, Kobayashi T, Morikawa R, Kimura K, et al.  
300 (2004) Manipulation of rumen methanogenesis by the combination of nitrate with  $\beta$  1-4  
301 galacto-oligosaccharides or nisin in sheep[J]. Animal Feed Science and Technology.  
302 Vol.115(No.1-2): 129-142.

303 25. Seon-Ho Kim, Lovelia L Mamuad, Eun-Joong Kim, Ha-Guyn, Sung, Gui-Seck Bae,

304 Kwang-Keun Cho, et al. (2018) Effect of different concentrate diet levels on rumen fluid  
305 inoculum used for determination of in vitro rumen fermentation, methane concentration, and  
306 methanogen abundance and diversity[J]. Italian Journal of Animal Science. Vol.17(No.2):  
307 359-367.

308 26. Ma T, Tu Y, Zhang NF, Deng KD, Diao QY (2015) Effect of the Ratio of Non-fibrous  
309 Carbohydrates to Neutral Detergent Fiber and Protein Structure on Intake, Digestibility,  
310 Rumen Fermentation, and Nitrogen Metabolism in Lambs[J]. ASIAN-AUSTRALASIAN  
311 JOURNAL OF ANIMAL SCIENCES. Vol.28(No.10): 1419-1426.

312 27. Agle M, Hristov A N, Zaman S, Schneider C, Ndegwa P M, Vaddella V K (2010) Effect  
313 of dietary concentrate on rumen fermentation, digestibility, and nitrogen losses in dairy  
314 cows[J]. Journal of Dairy Science. Vol.93(No.9): 4211-4222.

315 28. Pina D S, S C Valadares Filho, L O Tedeschi, A M Barbosa, and R F D Valadares (2009)  
316 Influence of different levels of concentrate and ruminally undegraded protein on  
317 digestive variables in beef heifers[J]. Anim. Sci. 87: 1058-1067.

318 29. Bargo F, Muller LD, Delahoy JE, Cassidy TW (2002) Milk response to concentrate  
319 supplementation of high producing dairy cows grazing at two pasture allowances[J]. Journal  
320 of Dairy Science. Vol.85(NO.7): 1777-1792.

321 30. Na R, Dong H, Zhu Z, et al. (2013) Effects of Forage Type and Dietary Concentrate to  
322 Forage Ratio on Methane Emissions and Rumen Fermentation Characteristics of Dairy  
323 Cows in China[J]. Transactions of the ASABE. Vol. 56(No.3): 1115-1122.



- 324 31. Yang W Z, Beauchemin K A (2006) Effects of physically effective fiber on chewing  
325 activity and ruminal pH of dairy cows fed diets based on bar-leys silage[J]. Journal of  
326 Dairy Science, 89 ( 1 ): 217—228.
- 327 32. Jin Wei, Li Yin, Cheng Yanfen, Mao Shengyong, Zhu Weiyun (2018) The bacterial and  
328 archaeal community structures and methanogenic ptotential of the cecal microbiota of goats  
329 fed with hay and high-grain diets[J]. Antonie van Leeuwenhoek.
- 330 33. Liu J, Xu T, Zhu W, Mao S (2014) High-grain feeding alters caecal bacterial microbiota  
331 composition and fermentation and results in caecal mucosal injury in goats. Br J  
332 Nutr112(3): 416–427.
- 333 34. Rong Yong, Xianghong Jiang (2018) Effects of Different Ratios of Concentrates and  
334 Roughages on Rumen Microbial Protein. Sichuan Animal Husbandry and Veterinary  
335 Surgeons, 2018, 45(11): 28-30.
- 336 35. P Mosoni<sup>1</sup>, F Chaucheyras-Durand, C. Béra-Maillet<sup>1</sup>, E. Forano<sup>1</sup> (2007) Quantification by  
337 real-time PCR of cellulolytic bacteria in the rumen of sheep after supplementation of a forage  
338 diet with readily fermentable carbohydrates: effect of a yeast additive[J].Journal of applied  
339 microbiology. Vol.103(No.6): 2676-2685.
- 340 36. Singh K M, V B Ahir, A K Tripathi, U V Ramani, M Sajnani, P G Koringa, et al. (2012)  
341 Metagenomic analysis of Surti buffalo (Bubalus bubalis) rumen: a preliminary  
342 study. Molecular biology reports, 39(4): 4841-4848.
- 343 37. Ley R E, Lozupone C A, Hamady M, Knight R, Gordon J I (2008) Worlds within worlds:

344 Evolution of the vertebrate gut microbiota(Article)[J]. Nature Reviews Microbiology.  
345 Vol.6(No.10): 776-788.

346 38. N Borruel, C Manichanh, K S Burgdorf, M Arumugam, JJ Raes, RQ Li, et al. (2010) A  
347 human gut microbial gene catalogue established by metagenomic sequencing[J]. Nature,  
348 2010, 464 (7285): 59-65.2010

349 39. OLIVEIRA M L S, AREAS A P M, CAMPOS I B, et al. (2006) Induction of systemic  
350 and mucosal im mune response and decrease in Streptococcus pneumoniae colonization  
351 by nasal inoculation of m ice with recom binant lactic acid bacteria expressing pneum  
352 ococcal surface antigen A[J]. M icrobes and Infection, 8(4): 1016—1024.

353 40. Lanjie Li, Shuru Cheng, Qiyu Diao, Tong Fu, Yanliang Bi, Ansi Wang, et al. (2017)  
354 Effects of diets with different levels of NFC/NDF on rumen fermentation parameters  
355 and bacterial community in male calves.Journal of Livestock and Veterinary Medicine,  
356 48(12): 2347-2357.

357 41. Pitta D, S Kumar, B Veiccharelli, N Parmar, B Reddy and C Joshi. (2014) Bacterial  
358 diversity associated with feeding dry forage at different dietary concentrations in the  
359 rumen contents of Mehshana buffalo (Bubalus bubalis) using 16S pyrotags. Anaerobe,  
360 25: 31-41.

361 42. Ellison M J, G C Conant, R. R. Cockrum, K J Austin, H Truong, M Becchi, et al. (2014)  
362 Diet alters both the structure and taxonomy of the ovine gut microbial ecosystem. DNA  
363 research, 21(2): 115-125.

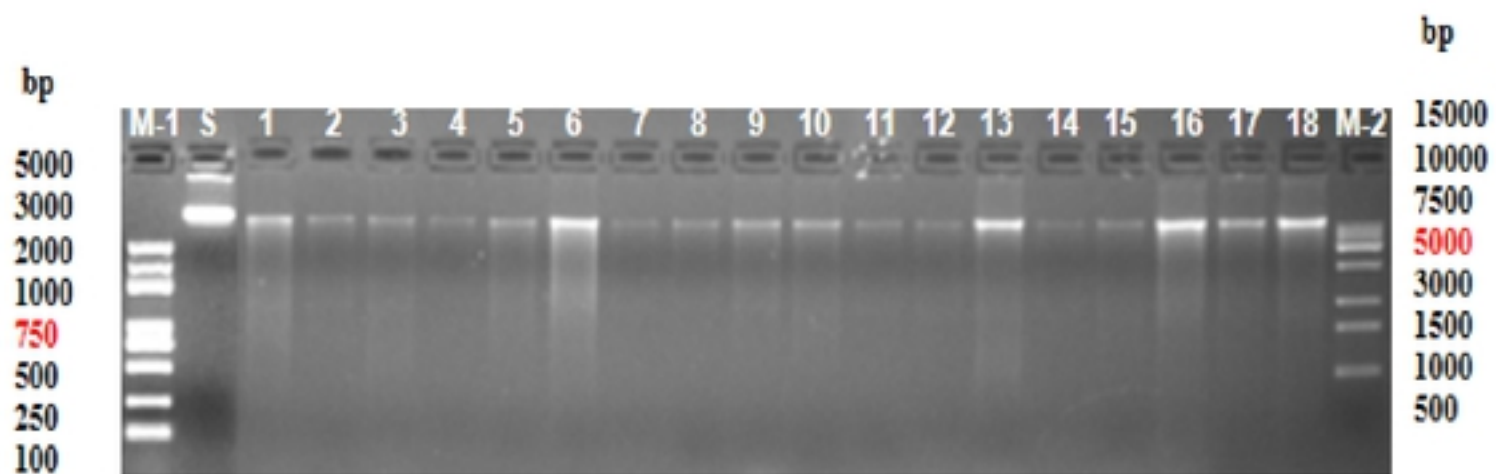
- 364           43. Kim M, J Kim, L Kuehn, J Bono, E Berry, N Kalchayanand, et al. (2014) Investigation of  
365           bacterial diversity in the feces of cattle fed different diets. Journal of animal science,  
366           92(2): 683-694.

1 **Table 1. Effects of different NFC/NDF on rumen fluid pH**

Items	pH				SEM	P-value
	1	2	3	4		
Period I	6.36	6.32	6.26	6.05	0.07	0.507
Period II	6.42a	6.28ab	6.03bc	6.01c	0.06	0.015
Period III	6.68a	6.48ab	6.34bc	6.19c	0.06	0.012
Period IV	6.52a	6.27ab	6.16b	6.05b	0.06	0.016

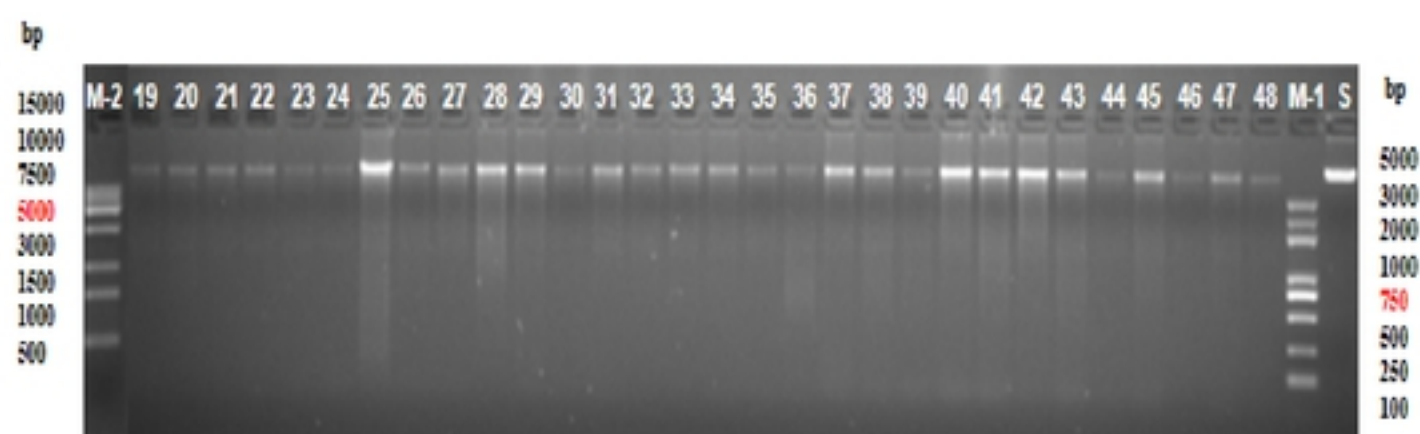
bioRxiv preprint doi: <https://doi.org/10.1101/729780>; this version posted August 9, 2019. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

2 Note: In the same row, values with no letter or the same letter superscripts mean no significant  
 3 difference ( $P > 0.05$ ), while with different small letter superscripts mean significant difference ( $P$   
 4  $< 0.05$ ). Period I (1~18 d), II (19~36 d), III (37~54 d) and IV (55~72 d) and Group 1, 2, 3, 4  
 5 means four groups of sheep treated with four dietary levels of NFC/NDF (0.78, 1.23, 1.61, 2.00  
 6 respectively). The same as Table 3.



7

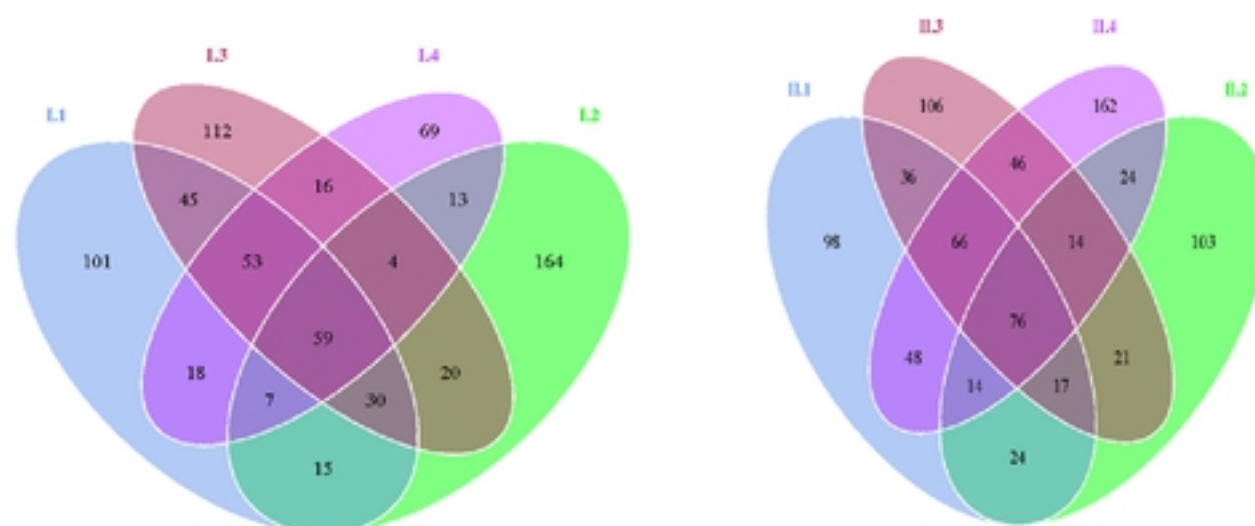
8



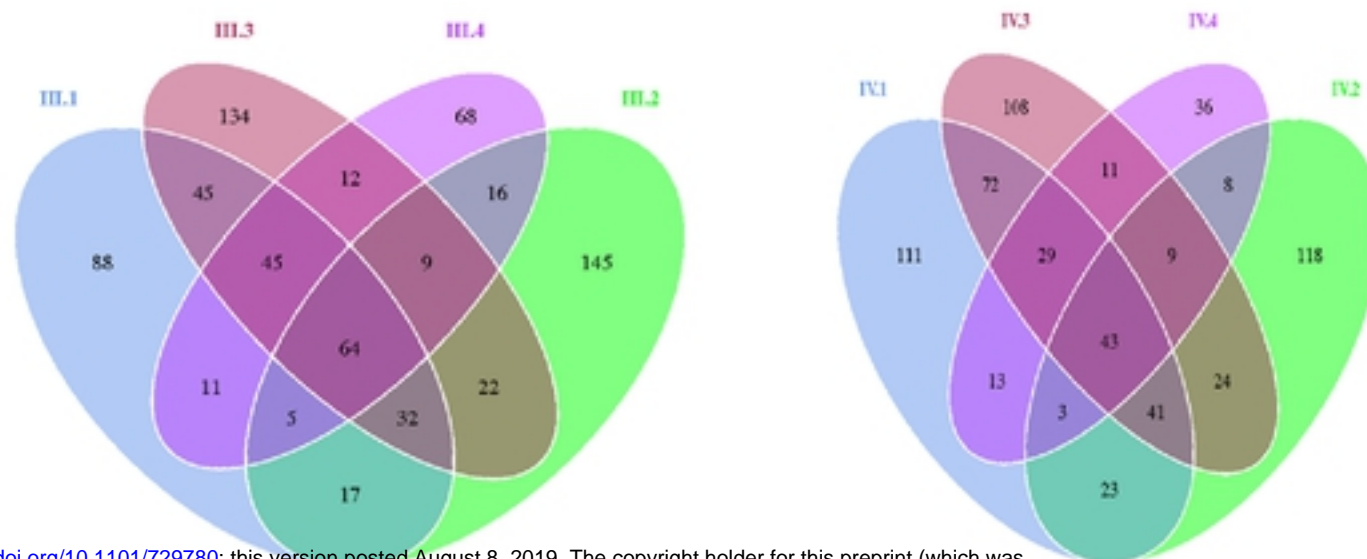
9

10 **Fig. 1. Extraction of rumen bacterial DNA.** The DNA of forty-eight samples from four periods  
 bioRxiv preprint doi: <https://doi.org/10.1101/729780>; this version posted August 8, 2019. The copyright holder for this preprint (which was  
 not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made  
 available under aCC-BY 4.0 International license.

11 samples were extracted, the order of 1~3, 4~6, 7~9 and 10~12 means samples in group 1, 2, 3 and  
 12 group 4 of period I respectively, each group with three replicates. The following order are as  
 13 period I.



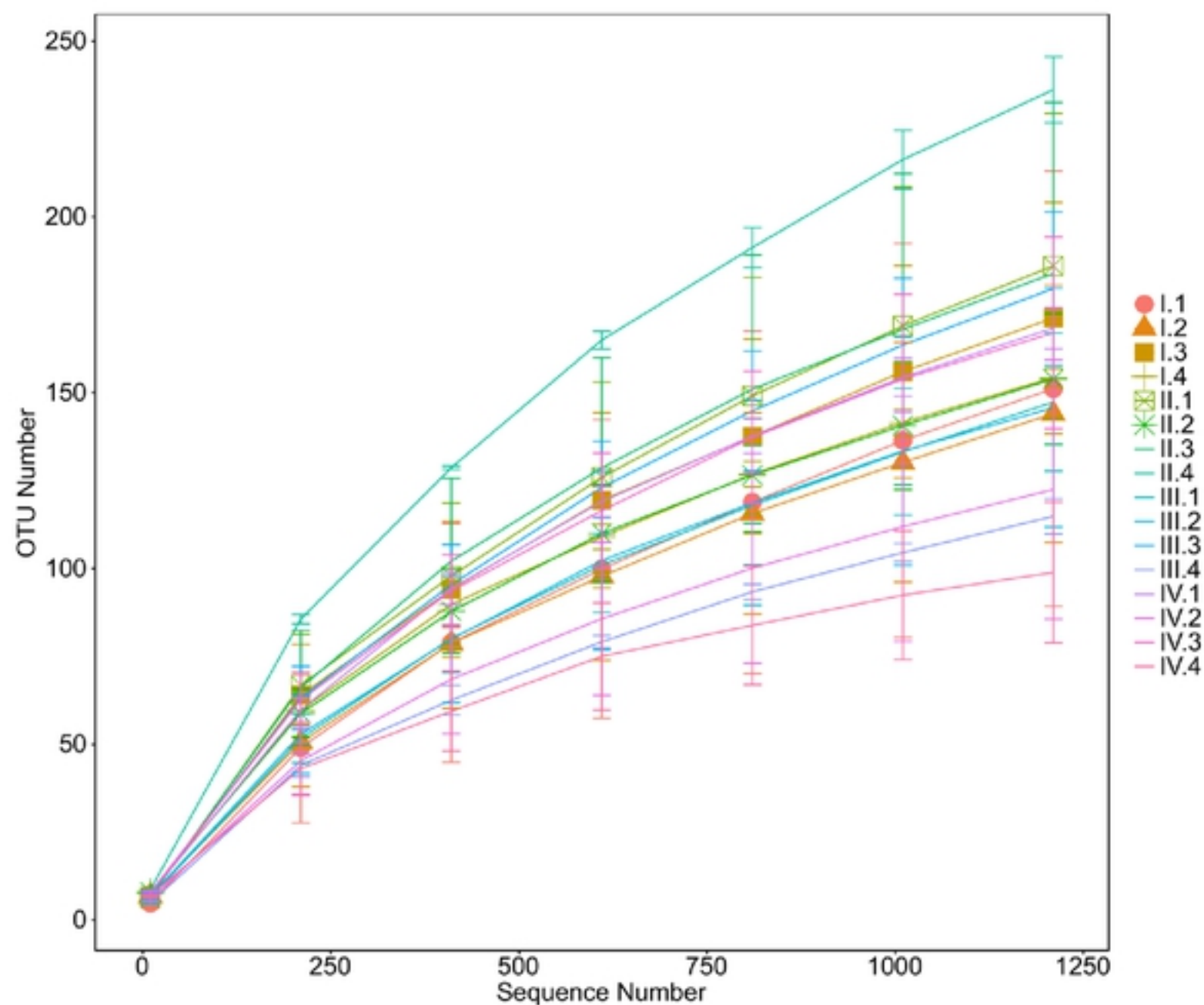
14



bioRxiv preprint doi: <https://doi.org/10.1101/729780>; this version posted August 8, 2019. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

15

16 **Fig. 2. Vene graph of microflora in rumen fluid of Karakul sheep. The amounts of OTUs in**  
 17 **each group were shown and four groups of one period were formed in one Vene graph.**



18



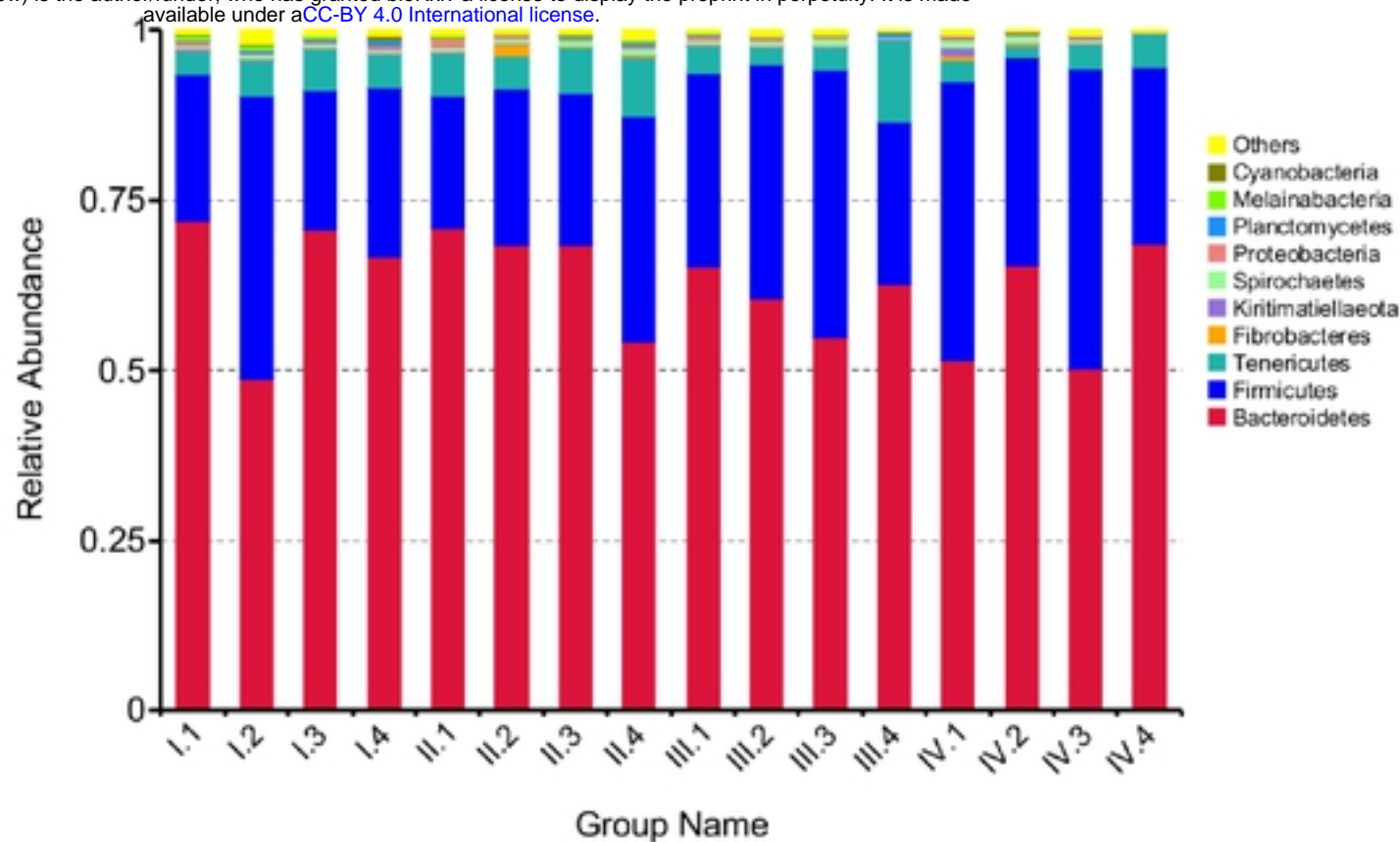
**Fig. 3. OTU dilution curve of bacteria in the rumen of Karakul sheep.** Rarefaction curves of OTUs clustered at 97% sequence identity across different samples.

**Table 2. Analysis of Alpha diversity of rumen liquid samples at 0.03 distance**

Items		observed_s pecies	Shannon	Simpson	Chao1	ACE
I	1	151	3.488	0.631	295.35	309.08
	2	144	4.447	0.853	259.414	271.994
	3	171	4.594	0.799	332.167	340.381
	4	154	4.946	0.897	232.5	252.125
II	1	186	4.536	0.787	321.413	357.069
	2	154	5.088	0.927	246.385	274.676
	3	184	4.865	0.838	315.519	348.306
	4	236	6.127	0.961	399.36	417.33
III	1	147	4.465	0.863	243.969	295.035
	2	146	4.545	0.874	233.092	263.23
	3	180	5.024	0.906	275.943	318.957
	4	115	3.786	0.798	198.268	208.16

	1	168	4.94	0.899	246.109	288.862
IV	2	122	3.992	0.81	338.777	248.709
	3	167	5.08	0.912	250.554	293.258
	4	99	4.36	0.879	139.488	144.216

bioRxiv preprint doi: <https://doi.org/10.1101/729780>; this version posted August 8, 2019. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.



22

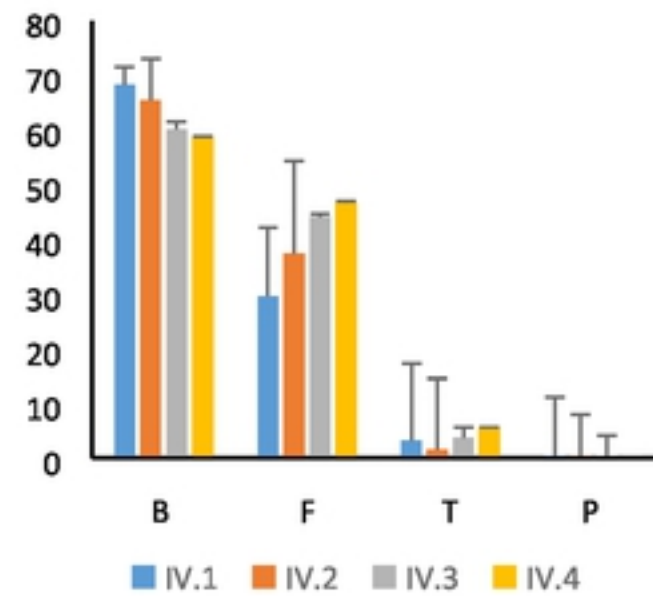
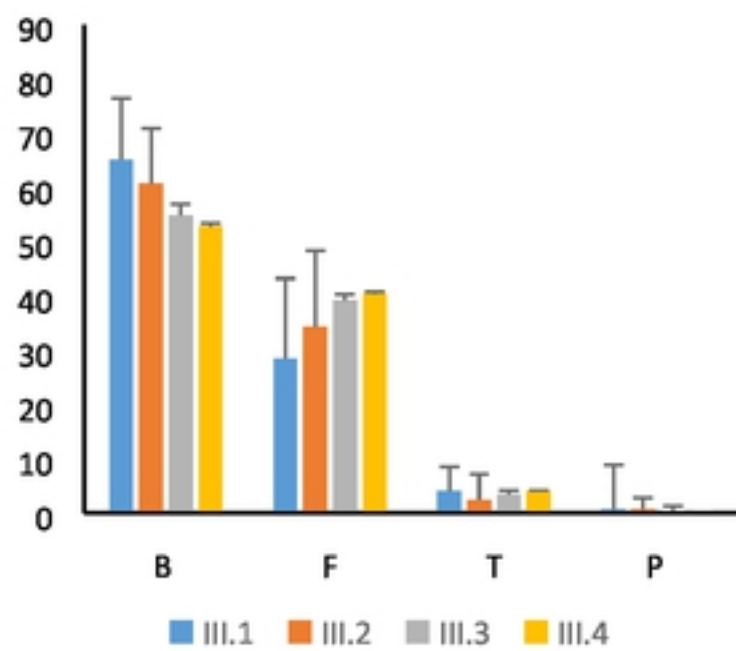
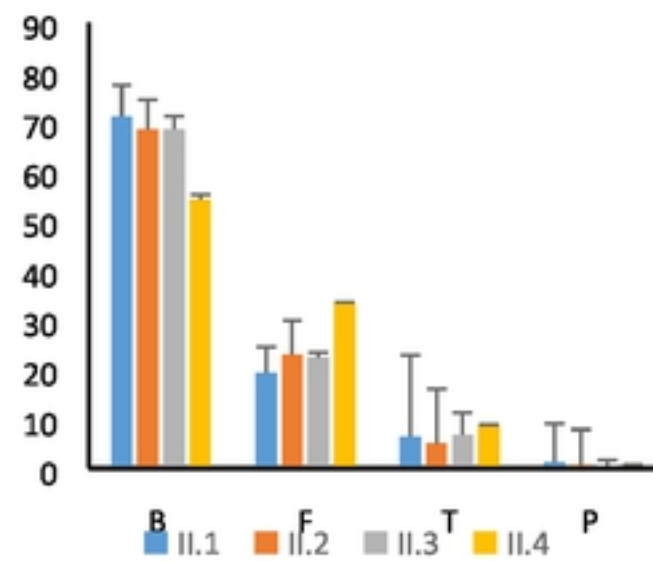
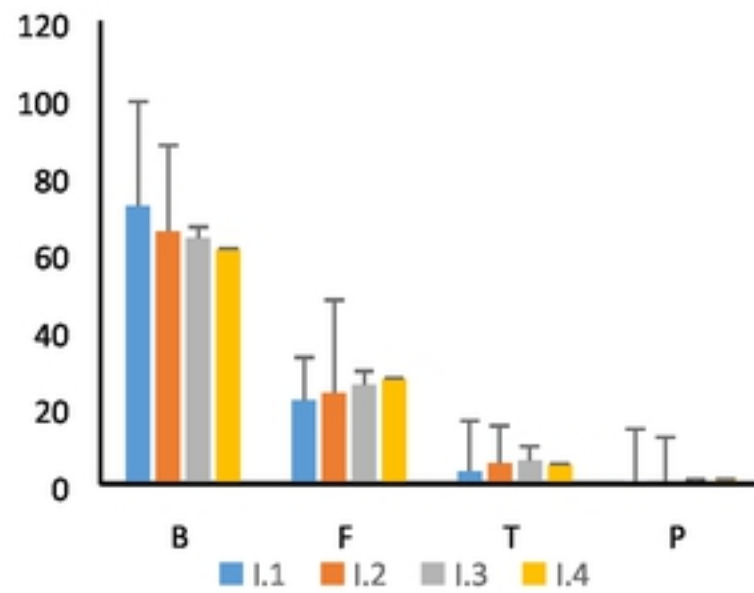
23 **Fig. 4. The column chart of the main dominant phylum in Karakul sheep fed with different**  
 24 **NFC/NDF diets. A color-coded bar plot showing the average bacterial phylum distribution across**  
 25 **the different age groups that were sampled.**

26

a

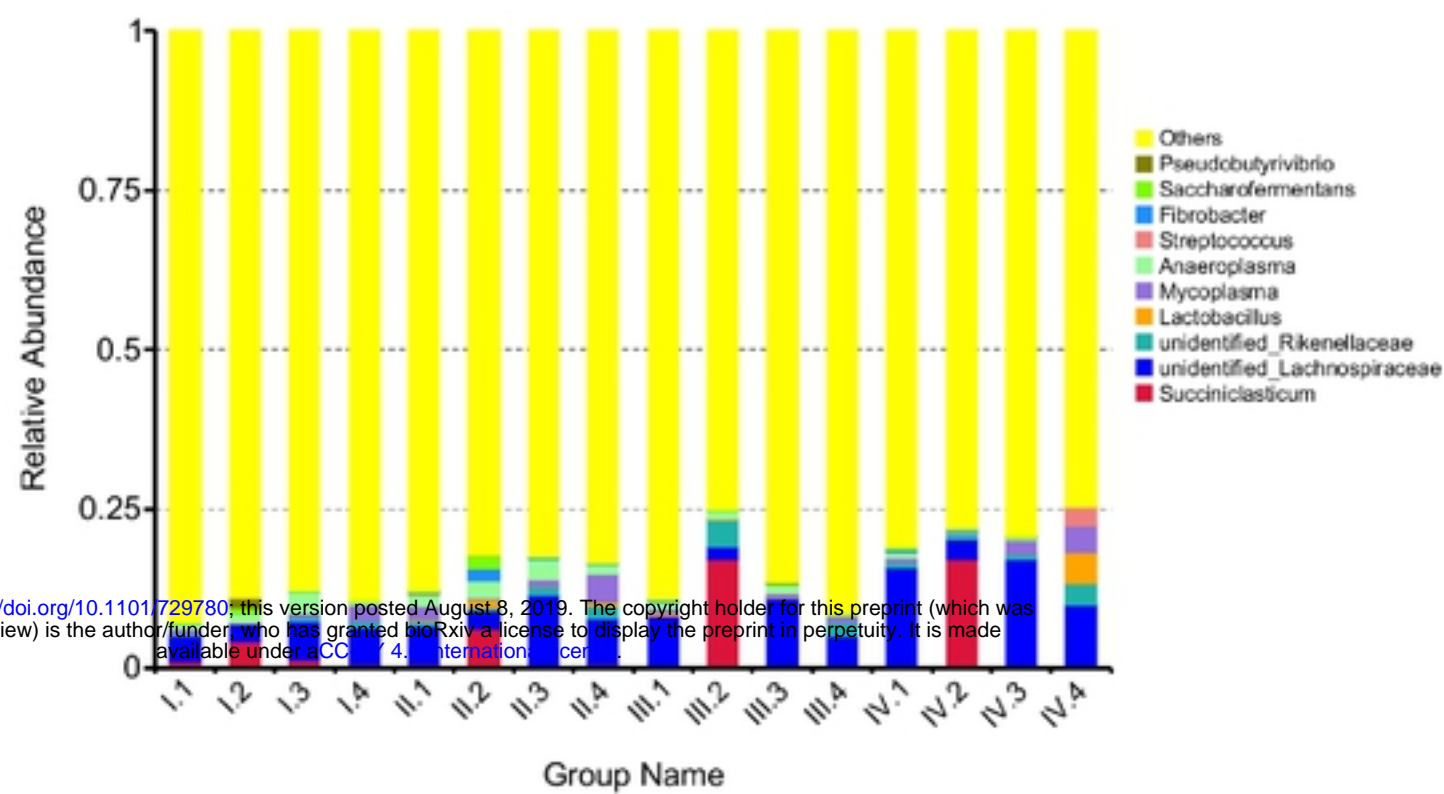
b





**Fig. 5(a~d) . Effects of different NFC/NDF diets on relative abundance (% reads) of rumen phylum in Karakul Sheep**

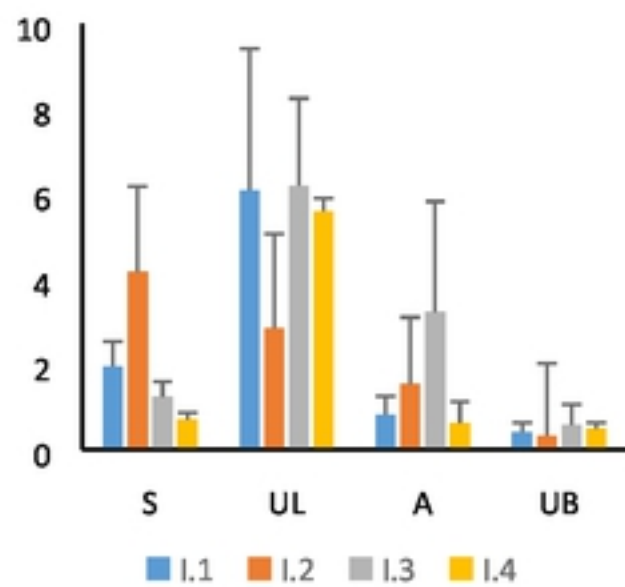
Note: B means Bacteroidetes, F means Firmicutes, T means Tenericutes, P means Proteobacteria; a, b, c, d represents the experiment period of I, II, III, IV respectively.



35

36 **Fig. 6. The column chart of the main dominant genus in Karakul sheep fed with different**  
 37 **NFC/NDF diets. A color-coded bar plot showing the average bacterial genera distribution across**  
 38 **the different age groups that were sampled**

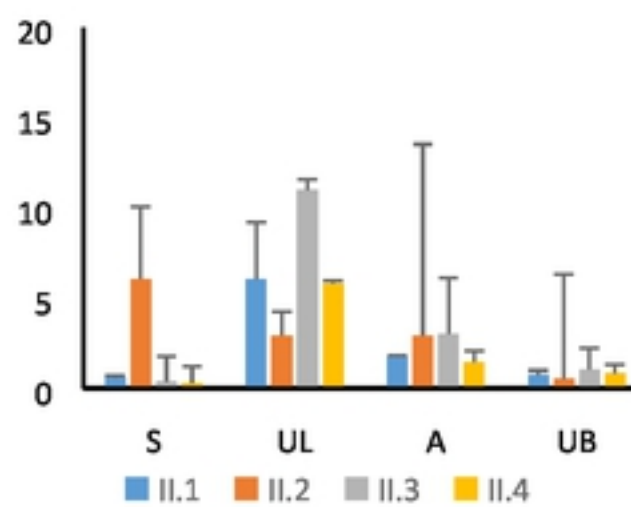
39 a



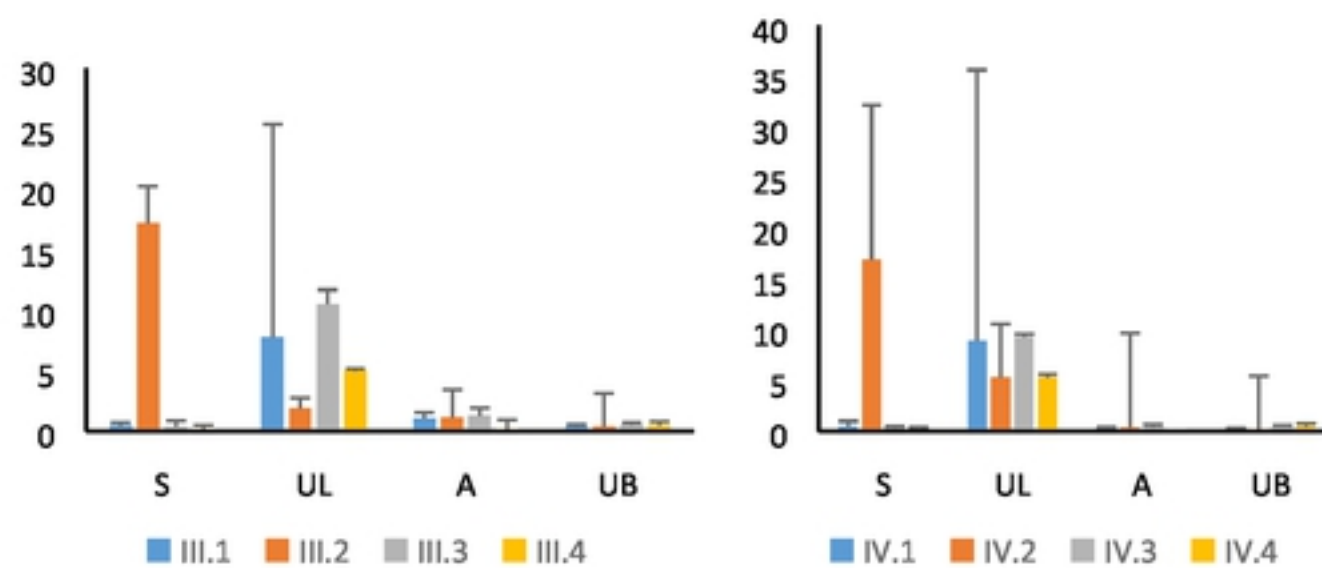
40

41 c

b



d

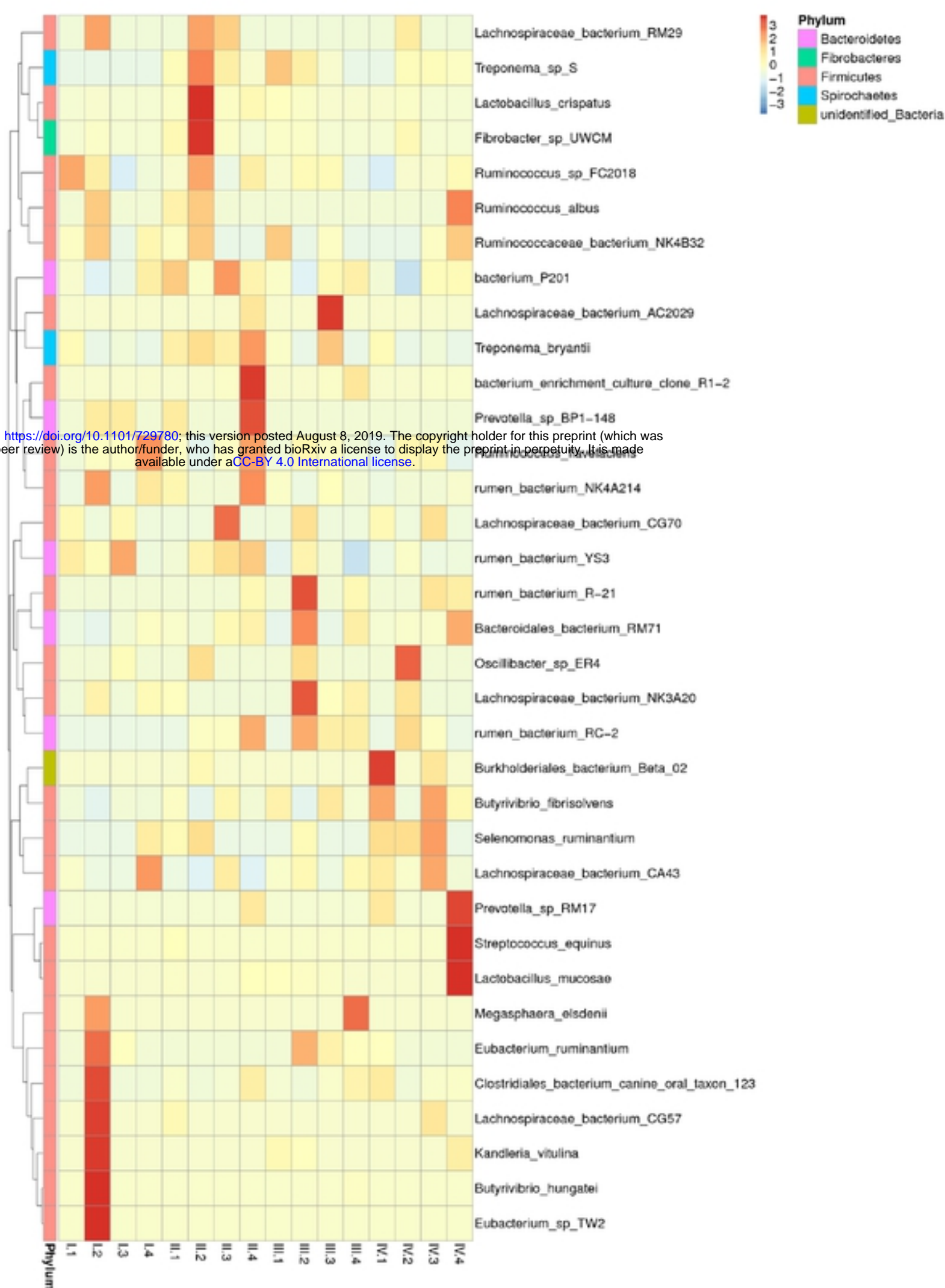


42

bioRxiv preprint doi: <https://doi.org/10.1101/729780>; this version posted August 8, 2019. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

43 **Fig. 7 (a~d) : Effects of different NFC/NDF diets on relative abundance (% reads) of**  
 44 **rumen genus in Karakul Sheep. Note: S means *Succiniclasticum*, UL means**  
 45 ***unidentified-Lachnospiraceae*, A means *Anaeroplasma*, and UB means *unidentified***  
 46 ***Bacteroidales*, a, b, c, d represents the experiment period of I, II, III, IV respectively.**

47



Relative abundance of community (%)

**Fig. 8. Heat map of the rumen bacteria composition at species level.** The heat map indicates the relative percentage of each species for the different dietary NFC/NDF group sampled.

**Table 3. Effects of different NFC/NDF on the relative abundance (%) of cellulose-degrading**

Period I						
Species	1	2	3	4	SEM	P-value
<i>Butyrivibrio-fibrisolvens</i>	2.664a	0.466b	2.990a	2.123a	0.371	0.035
<i>Fibrobacter-sp-UWCM</i>	-	0.027	-	-	0.006	0.441
<i>Ruminococcus-flavefaciens</i>	0.027	0.054	0.051	-	0.038	0.216
<i>Ruminococcus-albus</i>	-	0.055	-	-	0.009	0.052
Period II						
Species	1	2	3	4	SEM	P-value
<i>Butyrivibrio-fibrisolvens</i>	2.689a	0.548b	3.123a	2.406a	1.132	0.027
<i>Fibrobacter-sp-UWCM</i>	0.082	0.411	0.027	-	0.350	0.294
<i>Ruminococcus-flavefaciens</i>	-	0.055	0.050	0.027	0.036	0.290
<i>Ruminococcus-albus</i>	0.027	0.055	-	-	0.01	0.561
Period III						
Species	1	2	3	4	SEM	P-value
<i>Butyrivibrio-fibrisolvens</i>	6.785a	0.657c	7.687a	3.292b	1.143	<0.01

<i>Fibrobacter-sp-UWCM</i>	0.025	0.027	-	-	0.009	0.596
<i>Ruminococcus-flavefaciens</i>	0.027	0.082	0.055	0.027	0.021	0.627
<i>Ruminococcus-albus</i>	-	-	-	-	-	-

---

Period IV

Species	1	2	3	4	SEM	P-value
<i>Butyrivibrio-fibrisolvens</i>	7.234a	2.301c	8.694a	5.975b	1.581	<0.01
<i>Fibrobacter-sp-UWCM</i>	-	0.082	-	-	0.07	0.441
<i>Ruminococcus-flavefaciens</i>	-	0.082	0.079	-	0.016	0.052
<i>Ruminococcus-albus</i>	-	0.085	-	-	0.013	0.063

54

55