

1 **KCND3 is a novel susceptibility locus for early repolarization**

2 Alexander Teumer^{1,2*}, Teresa Trenkwalder^{3*}, Thorsten Kessler³, Yalda Jamshidi⁴, Marten E. van den
3 Berg⁵, Bernhard Kaess⁶, Christopher P. Nelson^{7,8}, Rachel Bastiaenen⁹, Marzia De Bortoli¹⁰,
4 Alessandra Rossini¹⁰, Isabel Deisenhofer³, Klaus Stark¹¹, Solmaz Assa¹², Peter S. Braund^{7,8}, Claudia
5 Cabrera^{13,14,15}, Anna F. Dominiczak¹⁶, Martin Gögele¹⁰, Leanne M. Hall^{7,8}, M. Arfan Ikram⁵, Maryam
6 Kavousi⁵, Karl J. Lackner^{17,18}, Lifelines Cohort Study¹⁹, Christian Müller²⁰, Thomas Münzel^{18,21},
7 Matthias Nauck^{2,22}, Sandosh Padmanabhan¹⁶, Norbert Pfeiffer²³, Tim D. Spector²⁴, Andre G.
8 Uitterlinden⁵, Niek Verweij¹², Uwe Völker^{2,25}, Helen R. Warren^{13,14}, Mobeen Zafar¹², Stephan B.
9 Felix^{2,26}, Jan A. Kors²⁷, Harold Snieder²⁸, Patricia B. Munroe^{13,14}, Cristian Pattaro¹⁰, Christian
10 Fuchsberger¹⁰, Georg Schmidt^{29,30}, Ilja M. Nolte²⁸, Heribert Schunkert^{3,30}, Peter Pramstaller¹⁰, Philipp
11 S. Wild³¹, Pim van der Harst¹², Bruno H. Stricker⁵, Renate B. Schnabel²⁰, Nilesh J. Samani^{7,8},
12 Christian Hengstenberg³², Marcus Dörner^{2,26}, Elijah R. Behr³³, Wibke Reinhard³

13
14
15 1 Institute for Community Medicine, University Medicine Greifswald, Germany
16 2 DZHK (German Center for Cardiovascular Research), partner site Greifswald, Greifswald, Germany
17 3 Klinik für Herz- und Kreislauferkrankungen, Deutsches Herzzentrum München, School of Medicine,
18 Technical University of Munich, Munich, Germany
19 4 Genetics Research Centre, Institute of Molecular and Clinical Sciences, St George's University of
20 London, United Kingdom
21 5 Department of Epidemiology, Erasmus MC - University Medical Center Rotterdam, The Netherlands
22 6 Medizinische Klinik I, St. Josefs-Hospital, Wiesbaden, Germany
23 7 Department of Cardiovascular Sciences, Cardiovascular Research Centre, Leicester, Leicester,
24 United Kingdom
25 8 NIHR Leicester Biomedical Research Centre University of Leicester, Leicester, United Kingdom
26 9 Cardiology Clinical Academic Group, Institute of Molecular and Clinical Sciences, St George's,
27 University of London, United Kingdom
28 10 Institute for Biomedicine, Eurac Research, affiliated with the University of Lübeck, Bolzano, Italy
29 11 Department of Genetic Epidemiology, University Regensburg, Germany
30 12 Department of Cardiology, University of Groningen, University Medical Center Groningen, The
31 Netherlands
32 13 Clinical Pharmacology, William Harvey Research Institute, Barts and The London, Queen Mary
33 University of London, London, UK
34 14 NIHR Barts Cardiovascular Biomedical Research Centre, Barts and The London School of
35 Medicine and Dentistry, Queen Mary University of London, London, UK
36 15 Centre for Translational Bioinformatics, William Harvey Research Institute, Barts and the London
37 School of Medicine and Dentistry, Charterhouse Square, London, UK
38 16 Institute of Cardiovascular and Medical Sciences, College of Medical, Veterinary and Life Sciences,
39 University of Glasgow, Glasgow, Scotland, UK
40 17 Institute of Clinical Chemistry and Laboratory Medicine, University Medical Center of the Johannes
41 Gutenberg-University Mainz, Mainz, Germany
42 18 German Center for Cardiovascular Research (DZHK), partner site RhineMain, Mainz, Germany
43 19 The list of group authors is provided in the study acknowledgments
44 20 Department of General and Interventional Cardiology, University Heart Center Hamburg-
45 Eppendorf, Germany, DZHK (German Center for Cardiovascular Research), partner site
46 Hamburg/Kiel/Lübeck, Germany
47 21 Center for Cardiology - Cardiology I, University Medical Center of the Johannes Gutenberg-
48 University Mainz, Mainz, Germany
49 22 Institute of Clinical Chemistry and Laboratory Medicine, University Medicine Greifswald,
50 Greifswald, Germany
51 23 Department of Ophthalmology, University Medical Center of the Johannes Gutenberg-University
52 Mainz, Mainz, Germany
53 24 Department of Twin Research and Genetic Epidemiology, King's College London, London, UK
54 25 Interfaculty Institute for Genetics and Functional Genomics, University Medicine Greifswald,
55 Greifswald, Germany
56 26 Department of Internal Medicine B, University Medicine Greifswald, Greifswald, Germany
57 27 Department of Medical Informatics, Erasmus MC - University Medical Center Rotterdam, The
58 Netherlands

59 28 Department of Epidemiology, University of Groningen, University Medical Center Groningen,
60 Groningen, The Netherlands
61 29 Innere Medizin I, Klinikum rechts der Isar, Technical University Munich, Munich, Germany
62 30 Deutsches Zentrum für Herz- und Kreislauf-Forschung (DZHK) e.V. (German Center for
63 Cardiovascular Research), Partner Site Munich Heart Alliance, Munich, Germany
64 31 Preventive Cardiology and Preventive Medicine, Center for Cardiology, University Medical Center
65 of the Johannes Gutenberg-University Mainz, Mainz, Germany; Center for Thrombosis and
66 Hemostasis, University Medical Center of the Johannes Gutenberg-University Mainz, Mainz,
67 Germany; DZHK (German Center for Cardiovascular Research), partner site Rhine-Main, Mainz,
68 Germany
69 32 Division of Cardiology, Department of Internal Medicine II, Medical University of Vienna, Vienna,
70 Austria
71 33 Cardiology Clinical Academic Group, Institute of Molecular and Clinical Sciences, St George's,
72 University of London, United Kingdom AND St George's University Hospitals NHS Foundation Trust,
73 London, United Kingdom
74

75 * These authors contributed equally to this work.
76

77 Corresponding Author:
78 PD Dr. med. Wibke Reinhard
79 Klinik für Herz- und Kreislauferkrankungen
80 Deutsches Herzzentrum München, Technische Universität München
81 Lazarettstrasse 36
82 80636 München
83 Germany
84 +49 89 1218 4005
85
86

87 **Abstract**

88 The presence of an early repolarization pattern (ERP) on the surface electrocardiogram (ECG) is
 89 associated with risk of ventricular fibrillation and sudden cardiac death. Family studies have shown
 90 that ERP is a highly heritable trait but molecular genetic determinants are unknown. We assessed the
 91 ERP in 12-lead ECGs of 39,456 individuals and conducted a two-stage meta-analysis of genome-wide
 92 association studies (GWAS). In the discovery phase, we included 2,181 cases and 23,641 controls
 93 from eight European ancestry studies and identified 19 genome-wide significant ($p < 5 \times 10^{-8}$) variants in
 94 the *KCND3* (potassium voltage gated channel subfamily D member 3) gene with a p-value of 4.6×10^{-10} .
 95 Replication of two loci in four additional studies including 1,124 cases and 12,510 controls confirmed
 96 the association at the *KCND3* gene locus with a pooled odds ratio of 0.82, $p = 7.7 \times 10^{-12}$ (rs1545300
 97 minor allele T). A subsequent GWAS meta-analysis combining all samples did not reveal additional
 98 loci. The lead SNP of the discovery stage (rs12090194) was in strong linkage disequilibrium with
 99 rs1545300 ($r^2 = 0.96$, $D' = 1$). Summary statistics based conditional analysis did not reveal any
 100 secondary signals. Co-localization analyses indicate causal effects of *KCND3* gene expression levels
 101 on ERP in both the left ventricle of the heart and in tibial artery.
 102 In this study we identified for the first time a genome-wide significant association of a genetic variant
 103 with ERP. Our findings of a locus in the *KCND3* gene not only provide insights into the genetic
 104 determinants but also into the pathophysiological mechanism of ERP, revealing a promising candidate
 105 for functional studies.

106

Introduction

The early repolarization pattern (ERP) is a common ECG finding characterized by an elevation at the QRS-ST junction (J-point) of at least 0.1 mV in two adjacent ECG leads. The prevalence of ERP in the general population ranges from 2 to 13% being more common in young athletic men(1–5). The classical notion of ERP being a benign ECG phenotype was challenged in 2008 by the landmark study of Haissaguerre and colleagues showing an association of ERP with increased risk of ventricular fibrillation and sudden cardiac death(6): the Early Repolarization Syndrome (ERS)(7). Since then several studies demonstrated an elevated risk of cardiovascular and all-cause mortality in individuals with ERP underscoring its arrhythmogenic potential(2, 8, 9). Although the mechanistic basis for malignant arrhythmias in ERS is unclear, it has been suggested that they occur as a result of an augmented transmural electrical dispersion of repolarization. Ex vivo studies point towards a central role of the cardiac transient outward potassium current (I_{to}) in the development of both, ERP and ERS(10). Furthermore, case descriptions of ERS identified genetic variations in genes encoding proteins for cardiac ion channels(11–13). Studies among relatives of sudden arrhythmic death syndrome show that ERP is more prevalent in the relatives than in controls indicating that ERP is an important potentially inheritable pro-arrhythmic trait(14, 15). Moreover, in family studies the heritability estimate for the presence of ERP was $h^2=0.49$ (16). However, estimates for common SNP heritability from unrelated individuals are lower(17). This may explain why the only available genome-wide association study (GWAS) on ERP failed to identify genetic variants reaching genome-wide significance(18), and indicates the need for GWAS with more power by including a larger number of ERP cases.

In order to identify genetic variations that convey susceptibility to ERP we performed a GWAS and meta-analysis in European ancestry individuals, using a combined two-stage GWAS approach with a discovery phase of 2,181 ERP cases and 23,641 controls from eight cohorts, and replicated the results in 1,124 cases and 12,510 controls from four additional cohorts.

Results

Clinical characteristics of the study cohorts are depicted in **Table 1**. The proportion of ERP based on the definition by Haisaguerre and Macfarlane(6, 19) ranged from 6% to 14% which is in line with previously reported prevalence in the general population(2–4).

Novel variants associated with ERP

We performed a GWAS meta-analysis in up to 2,181 cases and 23,641 controls from eight discovery cohorts. In total, 6,976,246 SNPs passed quality control (see Methods). We identified 19 variants spanning 49 kb in *KCND3* (Potassium Voltage-Gated Channel Subfamily D Member 3) as well as rs139772527 (effect allele frequency [EAF] 1.4%, OR=2.57, $p=2.0E-8$) near *HBZ* (Hemoglobin Subunit Zeta) to be genome-wide significantly associated ($p<5E-8$) with ERP. The SNP with the lowest p-value of the region (lead SNP) at *KCND3* was the intronic rs12090194 (EAF 32.5%, OR=0.80, $p=4.6E-10$), and was replicated in an independent sample of 1,124 cases and 12,510 controls from four additional cohorts ($p_{\text{replication}}=2.5E-3$, $p_{\text{combined}}=9.3E-12$, **Table 2**). The SNP rs139772527 near *HBZ* did not fulfil the criteria for replication ($p_{\text{replication}}=0.28$, $p_{\text{combined}}=1.4E-6$, **Table 2**) as described in the Methods. The combined meta-analysis of all 12 cohorts including up to 39,456 individuals revealed only the locus at *KCND3* to be genome-wide significantly associated with ERP (**Supplementary Figure 1**). The lead SNP was rs1545300 (EAF 31.9%, OR=0.82, $p=7.7E-12$), followed by the discovery stage lead SNP rs12090194 being in strong linkage disequilibrium with rs1545300 ($r^2=0.96$, $D'=1$) (**Figure 1**). Both SNPs were imputed at very high confidence (imputation quality score >0.97) in all cohorts. The quantile-quantile plots did not show any inflation (individual study λ_{GC} between 0.81 and 1.03, median: 0.91), and overall meta-analysis $\lambda_{GC}=1.02$ (linkage disequilibrium [LD] score regression intercept: 1.01, see Methods) (**Supplementary Figure 2**). Summary statistics based conditional analysis to select independent hits did not reveal any secondary signals.

Statistical finemapping of the associated locus

All significantly associated SNPs were located within *KCND3*, the potassium voltage-gated channel subfamily D member 3 gene and were intronic. We used the discovery and replication stage combined GWAS results to assess whether a single SNP or set of variants drive the association signal in *KCND3* (credible set). The 99% credible set was computed based on Approximate Bayes Factors for each SNP, resulting for each in a set of SNPs that with 99% posterior probability contained the variant(s) driving the association signal. For the associated locus at *KCND3* the credible set spanned 49 kb, and contained 19 variants. The two lead SNPs rs1545300 and rs12090194 had a posterior probability of 21% and 19%, respectively, whereas the former candidate SNP rs17029069(18) had a posterior probability of 2% (**Supplementary Table 2**).

To test whether the association in *KCND3* might be driven by heart rate or RR interval, we performed a sensitivity analysis in the 1,253 ERP cases and 11,463 controls of the Lifelines cohort adjusting the

genetic association of rs1545300 additionally for these two traits in separate models. The effect estimates were virtually unchanged ($OR=0.78$) with $p=1.2E-7$ for both adjustments. In addition, we assessed whether the association of rs1545300 might be related to a specific ERP subtype i.e ST segment or ERP localization. In all subtype-stratified analyses the 95% confidence intervals of the effect sizes overlapped with the overall results not pointing to a subtype driven signal (**Supplementary Table 3**).

eQTL and co-localization

We searched the Genotype-Tissue Expression (GTEx) project database(20) to look for tissue-specific eQTLs including all genes in vicinity of $\pm 1Mb$ of the lead SNP rs1545300 and found an association with *KCND3* expression levels in tibial artery ($p=3.0E-6$, $n=388$). Two additional eQTL associations of rs1545300 at a false discovery rate (FDR) <0.2 across the 48 tissues tested were found with *KCND3* (ENSG00000171385.5) in the left ventricle ($p=2.9E-4$, $n=272$) of the human heart, and with *CEPT1* (ENSG00000134255.9) in the minor salivary gland ($p=3.4E-4$, $n=97$) (**Supplementary Table 4**).

Subsequent co-localization analyses of rs1545300 in these three tissues revealed also a significant correlation of gene expression pattern with ERP ($p_{SMR} \leq 0.01$) (**Figure 2, Supplementary Table 5**), where for the left ventricle the correlation seems to be attributable to the same underlying causative variant ($p_{HEIDI} \geq 0.05$), and for tibial artery the test was close to nominal significance ($p_{HEIDI}=0.05$). However, the significant $p_{HEIDI}=1.7E-3$ of *CEPT1* in the minor salivary gland points rather towards a pleiotropic effect of rs1545300 than to a causal effect of gene expression on ERP in this tissue. For all three tissues, an increased gene expression level was associated with a higher risk of ERP (**Supplementary Table 5**).

Pleiotropic effects of the lead SNPs

To assess pleiotropic effects of the *KCND3* lead SNP rs1545300 or its proxies ($r^2 > 0.8$), we looked for genome-wide significant associations in the NHGRI-EBI Catalog of published genome-wide association studies(21) (accessed: 05/03/2019). Pleiotropic associations were found for P-wave terminal force (rs12090194 and rs4839185)(22) and for atrial fibrillation (rs1545300 and rs1443926)(23, 24). All these SNPs were in strong linkage disequilibrium ($r^2 > 0.97$) with the lead SNP. In addition, variants in low to moderate LD with rs1545300 were associated with P-wave duration (rs2798334, $r^2=0.26$)(25) and ST-T-wave amplitudes (rs12145374, $r^2=0.60$)(26).

Discussion

In this GWAS meta-analysis comprising 3,305 cases and 36,151 controls including independent replication samples, we describe an association of ERP with a locus on chromosome 1 in the *KCND3* gene. This is the first study identifying a robust genome-wide significant association between genetic variants and ERP. Our findings form the genetic basis for further functional studies examining the pathophysiological mechanism of ERP and potentially ERS. The *KCND3* gene encodes the main pore-forming α subunit of the voltage-gated rapidly inactivating A-type potassium channel. In the cardiac ventricle *KCND3* contributes to the fast cardiac transient outward potassium current (I_{to}), which plays a major role in the early repolarization phase 1 of the cardiac action potential (AP).

To date, two competing theories explain the presence of J waves and ERP: the repolarization and the depolarization theory, both involving the I_{to} channel. On the basis of animal models evidence for the former is more compelling. Thus, J waves result from a transmural voltage gradient created by a more prominent epicardial phase 1 AP notch relative to the endocardial AP notch(10, 27). The I_{to} current notably influences the degree of the transmural heterogeneity of the phase 1 AP notch and consecutively the magnitude of the J wave(10, 27). Pharmacological inhibition of the I_{to} current with 4-aminopyridine results in a reduction of the J wave amplitude(10). The depolarization theory is based on clinical overlap of ERP with Brugada syndrome, which has led to the suggestion of Brugada syndrome being a right ventricular variant of the ERP(28). In theory, deviation from the sequential activation of cardiac currents I_{Na} , I_{to} , and I_{CaL} can lead to regional conduction slowing and appearance of inferior and/or lateral ERP(27, 29). In patients with ERS, distinct phenotypes of both delayed depolarization and early repolarization have been identified(30).

ERP is a highly heritable trait within families(3, 16), however limited heritability can be attributed to common SNPs in unrelated individuals(17). This might be a reason why the only GWAS to date which included 452 cases failed to replicate any genome-wide significant loci(18). In our study, which includes 3,334 cases, we discovered and replicated variants in the *KCND3* gene. Interestingly, one of these variants (rs17029069), which is in moderate LD ($r^2=0.18$, $D'=-1$) with our lead SNP rs1545300 (**Supplementary Figure 3**) was reported as a candidate in the earlier GWAS meta-analysis(18). However, this variant did not replicate in their study, which the authors attributed to limited power based on the small sample size and/or heterogeneous phenotyping. In our study, experienced cardiologists centrally adjudicated more than 39,000 ECGs with high reproducibility ensuring a very

high phenotyping quality(17). The resulting homogenously assessed phenotype and the substantially increased number of cases are two aspects that elevated the statistical power of our GWAS meta-analysis. All detected variants cluster in intronic regions of the *KCND3* gene, without significant allelic heterogeneity. The annotation of the locus does not point to a direct pathogenic effect, i.e. a protein altering mutation, and also the statistical finemapping revealed no single SNP with a substantial posterior probability (e.g. >80%) of being causal. However, the latter approach has limitations of detecting rare causal variants due to imputation uncertainty and minimum minor allele frequency (MAF). Nevertheless, eQTL analysis suggested that the detected variants may affect gene expression of *KCND3*. Potential mechanisms include modification of gene expression via altered binding of transcription factors at *cis*-elements. This is supported by the results of the test for co-localization showing an increase of ERP risk due to increased gene expression levels of *KCND3* in tissues of the human heart and tibial artery. Similar, pharmacological *ex vivo* data predict gain of function mutations in the I_{to} current to increase the overall transmural outward shift, leading to an increased epicardial AP notch and thereby inducing ERP in the surface ECG(27). Additionally, in close proximity to the lead SNP rs1545300 a long non-coding RNA (lncRNA), *KCND3* antisense RNA 1 (*KCND3-AS1*) is described. lncRNAs have been shown to physiologically influence gene regulation through various mechanism e.g. chromatin remodeling, control of transcription initiation and post-transcriptional processing(31, 32). On the other hand, dysregulation of lncRNA control circuits can potentially impact development of disease(33): a very prominent example in cardiovascular diseases is the lncRNA *ANRIL*, which is a key effector of *9p21* in atherosclerotic risk and cardiovascular events(33–35).

Given the high prevalence of ERP in the general population and a high MAF of the identified genetic variants in our study the key question remains why only a very small subset of individuals develops severe ventricular arrhythmias and ERS. The fine interplay of a genetic predisposition and specific precipitating conditions might lead to an electrically vulnerable cardiac state. Insights into the potential origin of ventricular arrhythmias in ERS come from animal models and highlight the role of different ion channels including I_{to} (36). A pharmacological model of ERS in canine wedges from the inferior and lateral ventricular wall showed marked regional dispersion of repolarization (loss of phase 2 AP dome and AP shortening in some epicardial regions but not others). Presence of transmural repolarization heterogeneity allowed local re-excitation in form of closely coupled extrasystolic activity (phase 2 re-entry). The combination of an arrhythmogenic substrate, represented by regional electrical instability,

and triggering premature ventricular beats resulted in ventricular fibrillation(36). Human data in ERS patients suggest that in a subgroup, the ERP is due to a pure repolarization phenotype and arrhythmia(30) is triggered by Purkinje fiber ectopic beats.

Genetic variants in various ion channel genes have been associated with ERS(37) including the *KCNJ8* and *ABCC9* genes encoding the Kir6.1 and ATP-sensing subunits of the K_{ATP} channel(6, 11, 38, 39). The commonly implicated variant *KCNJ8*-p.S422L has a population frequency not consistent with ERS, and is predicted to be benign by multiple in silico algorithms according to the ClinVar database(40). A recent study by Chaveau *et al.* has, however, identified a de novo duplication of the *KCND3* gene in a patient who survived sudden cardiac death and in his 2-year-old daughter(12). Both exhibited marked ERP in the inferolateral leads that was augmented by bradycardia and pauses in heart rhythm, in keeping with a repolarization mechanism underlying the ERS phenotype. Studies have suggested that the inferior region of the left ventricle has a higher density of *KCND3* expression and higher intrinsic levels of I_{to} (36). This may explain the higher vulnerability of this region for the development of ERS in the setting of a genetically mediated gain-of-function in the I_{to} current. The findings of our study therefore suggest that common variation may play a role in the expression of *KCND3* and the I_{to} current and that it is likely to be relevant in ERS as well. This may in part explain the minimal yield of pathogenic variants in ERS cases. Further GWAS in large collaborative cohorts of ERS patients are therefore necessary to determine the importance of polygenic risk. A systematic evaluation of pleiotropic effects demonstrated known associations of the identified *KCND3* SNPs with ECG phenotypes only. The lead SNP rs1545300 or its proxies in strong LD ($r^2 > 0.97$) were found to be related with P-wave terminal force(22) and atrial fibrillation(23). This highlights the sharing of underlying mechanisms between *KCND3* variation and cardiac electrical activity.

Our study has some limitations, which need to be acknowledged. Presence of ERP in the ECG can be variable, as it has been described to be dependent on age, heart rate, vagal activity and medication, although our findings were valid after adjusting for some of these factors. Therefore, we cannot exclude that we have missed some individuals with ERP. Second, the tissue-specific gene expression data used for the co-localization analysis is based on a limited sample size. A larger gene expression sample or functional studies are needed to validate the revealed effect of *KCND3* expression on the ERP.

In conclusion, we show for the first time, a robust association of genetic variants with the ERP in a large GWAS of individuals of European ancestry. The locus in the *KCND3* ion channel gene is an intuitive candidate and supports the theory that at least a proportion of ERS is a pure channelopathy. Intensive future research will be needed to extend the discovery of ERP susceptibility loci to individuals of non-European ancestry, and to improve identification and risk stratification of the subset of individuals with the ERP who are at highest risk for potentially lethal ventricular arrhythmias.

Methods

Study cohorts and SNP genotyping

The discovery stage included 25,822 subjects (2,181 ERP cases) from eight independent cohorts: the British Genetics of Hypertension (BRIGHT) study, the Gutenberg Health Study (GHS1, GHS2), the Genetic Regulation of Arterial Pressure In humans in the Community (GRAPHIC) study, the Lifelines Cohort Study (Lifelines), the Study of Health in Pomerania (SHIP, SHIP-Trend), and TwinsUK. Additional 13,634 subjects (1,124 ERP cases) from four cohorts (Rotterdam Study I, II, III, and CHRIS) were used as independent replication: the Rotterdam Study (Rotterdam Study I, II, III), and the Cooperative Health Research In South Tyrol (CHRIS) study. The included subjects of all cohorts were of European ancestry, and all cohorts but BRIGHT (which sampled hypertensive cases) were population based (**Supplementary Table 1**). All subjects gave written informed consent and the studies were approved by the local ethics committees.

Electrocardiogram analysis and ERP evaluation

12-lead ECGs of all 12 studies were analyzed manually by experienced and specifically trained cardiologists for the presence of ERP according to the established definition by Haissaguerre and Macfarlane(6, 19). In case of a QRS duration of >120 ms or rhythm other than sinus rhythm (e.g. atrial fibrillation, pacemaker stimulation) ECGs were excluded from the analysis. The methodology employed and robustness of inter-observer correlations have been presented elsewhere(17).

In detail, ERP was defined as elevation of the J-point above the level of QRS onset of ≥ 0.1 mV in at least two corresponding leads. To avoid confusion or overlap with Brugada syndrome or arrhythmogenic right ventricular dysplasia, leads V1 to V3 were excluded from ERP scoring. In case of presence of ERP, region, either inferior (leads II, III, aVF), antero-lateral (leads I, aVL, V₄-V₆), or both, and the maximum amplitude of J-point elevation was documented. Further, the morphology of ERP

was assessed as either notching, slurring or both as well as the ST segment according to Tikkanen and colleagues(41) as either concave/rapidly ascending (>0.1 mV elevation 100 ms after J-point peak or persistently elevated ST segment >0.1 mV) or horizontal/descending (≤ 0.1 mV elevation within 100 ms after J-point peak)(19, 41).

GWAS in individual studies

The GWAS in each study for both the discovery and replication stage was performed on autosomal imputed SNP genotypes using study-specific quality control protocols which are provided in detail in **Supplementary Table 1**. Association analyses were performed using logistic regression for ERP status as outcome and an additive genetic model on SNP dosages, thus taking genotype uncertainties of imputed SNPs into account. The analyses were adjusted for age, sex, and relevant study-specific covariates such as principal components for population stratification (**Supplementary Table 1**).

Statistical methods for meta-analysis

The result files from individual studies GWAS underwent extensive quality control before meta-analysis using the `gwasqc()` function of the `GWAToolbox` package v2.2.4(42). The quality control included file format checks as well as plausibility and distributions of association results including effect sizes, standard errors, allele frequencies and imputation quality of the SNPs.

The meta-analyses were conducted using a fixed-effect inverse variance weighting as implemented in `Metal`(43). Monomorphic SNPs, SNPs with implausible association results (i.e. $p \leq 0$, $SE \leq 0$, $|\log(OR)| \geq 1000$), and SNPs with an imputation quality score ≤ 0.4 were excluded prior to the meta-analyses resulting in a median of 12,839,202 SNPs per cohort (IQR: 10,756,073-13,184,807). During the meta-analysis, the study-specific results were corrected by their specific λ_{GC} if >1 . Results were checked for possible errors like use of incorrect association model by plotting the association p-values of the analyses against those from a z-score based meta-analysis for verifying overall concordance. SNPs that were present in $<75\%$ of the total sample size contributing to the respective meta-analysis or with a MAF ≤ 0.01 were excluded from subsequent analyses. Finally, data for up to 6,976,246 SNPs were available after the meta-analysis.

Quantile-quantile plots of the meta-analysis results are provided in **Supplementary Figure 2**. To assess whether there was an inflation of p-values in the meta-analysis results attributed to reasons other than polygenicity, we performed LD score regression(44). The LD score corrected λ_{GC} value of the discovery and replication combined meta-analysis was 1.01, supporting the absence of

unaccounted population stratification. Genome-wide significance was defined as a p-value $<5E-8$, corresponding to a Bonferroni correction of one million independent tests. Unless stated otherwise, all reported p-values are two-sided. The I^2 statistic was used to evaluate between-study heterogeneity(45).

To evaluate the presence of allelic heterogeneity within each locus, the GCTA stepwise model selection procedure (cojo-slc algorithm) was used to identify independent variants employing a step-wise forward selection approach(46). We used the genotype information of 4,081 SHIP individuals for LD estimation, and set the significance threshold for independent SNPs to $5E-8$.

All loci were named according to the nearest gene of the lead SNP. Genomic positions correspond to build 37 (GRCh37).

Replication analysis

To minimize the burden for multiple testing correction and thus maximizing the power for replication, the lead SNPs of genome-wide significant loci in the discovery stage were taken forward to the replication stage in independent samples (**Table 1**). SNPs were considered as replicated if the p-value of a one-sided association test was <0.025 which corresponds to a Bonferroni correction for the two lead SNPs tested at 5% significance level.

Finally, the GWAS results from the discovery and replication studies were meta-analyzed to search for additional genome-wide significant loci by maximizing the statistical power for locus discovery.

Gene expression based analyses

The lead SNP rs1545300 of the *KCND3* locus of the combined discovery and replication GWAS meta-analysis was tested for *cis* eQTLs (± 1 Mb window around the transcription start site) in 48 tissues available in the GTEx v7 database that included at least 70 samples. Significant associations were selected based on a Bonferroni corrected p-value $<3.0E-5$ for the number of genes and tissues tested. Subsequently, the SNP rs1545300 was tested and plotted for co-localization in the three tissues with an eQTL $FDR < 0.2$ by applying the SMR method(47) using the GWAS and GTEx eQTL summary statistics. The method includes a test whether the effect on expression observed at a SNP or at its proxies is independent of the signal observed in the GWAS, i.e. that gene expression and y are associated only because of a latent non-genetic confounding variable (SMR test), and a second test that evaluates if the eQTL and GWAS associations can be attributable to the same causative variant (HEIDI test). Significance for co-localization of the gene expression and the GWAS signals was

defined by $p_{SMR} < 0.01$, where additionally a $p_{HEIDI} \geq 0.05$ indicates the same underlying causal variant(47).

Acknowledgments

Detailed acknowledgments are provided in the Supplementary Information.

Author Contributions

Project design and analysis: W.R., T.T., A.T., M.D, E.R.B. Management of individual study: A.F.D., C.F., C.P., E.R.B., K.J.L., M.A.I., M.D., M.G., M.Z., N.P., N.V., P.B.M., P.P., P.v.d.H., S.A., S.B.F., T.D.S., T.M., T.T., W.R., Y.J. Recruitment of individual study subjects: A.F.D., G.S., H.S., I.D., M.D., M.Z., N.V., P.v.d.H., S.A., S.B.F. Drafting of the manuscript: A.R., A.T., B.H.S., E.R.B., M.D.B., M.E.v.d.B., T.T., W.R. Statistical methods and analysis of individual study: A.T., C.C., C.F., H.R.W., H.S., I.M.N., K.S., M.E.v.d.B., S.P. Genotyping of individual study: A.G.U., C.F., M.N., P.B.M., U.V. Interpretation of the results: A.R., A.T., B.K., C.H., E.R.B., M.D., M.D.B., T.K., T.T., W.R. Critical review of the manuscript: all authors.

Competing Financial Interests

The authors declare no competing financial interests.

Data availability

Summary genetic association results have been submitted for full download to the CHARGE dbGaP website under accession phs000930 [<https://www.ncbi.nlm.nih.gov/gap>].

Figure Legends

Figure 1. GWAS results of the *KCND3* locus

The results of the combined ERP GWAS results for the *KCND3* locus are shown for the replicated discovery stage lead SNP rs12090194 (A and B), and for the combined GWAS lead SNP rs1545300 (C and D). The regional association plots (A and C) show the association results in a ± 500 kb region around the lead SNP. SNPs are plotted on the x-axis according to their chromosomal position with the $-\log_{10}(p\text{-value})$ of the GWAS association on the y-axis. Correlation with the lead SNP (purple) is

estimated based on the 1000 Genomes reference samples. Plots were generated using the website of LocusZoom (Pruim, R. J. *et al.* Bioinformatics, 2010). Genetic positions refer to GRCh37/hg19 coordinates. Forest plots of the respective lead SNPs are provided in (B) and (D), with odds ratios and their 95% confidence intervals plotted on the x-axis. I^2 is the percentage of total variation across studies that is due to heterogeneity.

Figure 2. Co-localization results

Illustration of the SMR test for ERP risk and expression QTLs at the rs1545300 locus at chromosome 1p13.2 for (A) left ventricle of the heart, (B) tibial artery, and (C) minor salivary gland tissue. In each panel, the upper box shows the GWAS regional association plot with ERP risk, with level of significance of the SMR test (y-axis) for each transcript in the locus indicated by a diamond positioned at the center of the transcript. A significant SMR test represented by a purple diamond indicates an association of the transcript level of the respective genes (purple label) with the trait. For all three tissues, an increased gene expression level of a significant SMR test was associated with a higher risk of ERP. A filled purple diamond indicates a HEIDI test p-value >0.05, thus a likely co-localization. The lower box shows the regional association distribution with changes in expression of the highlighted (purple) gene transcript in the respective tissue. In both boxes, the x-axis refers to GRCh37/hg19 genomic coordinates.

References

1. Reinhard W *et al.* The early repolarization pattern: Echocardiographic characteristics in elite athletes.. *Ann. Noninvasive Electrocardiol.* 2018;e12617.
2. Sinner MF *et al.* Association of early repolarization pattern on ECG with risk of cardiac and all-cause mortality: a population-based prospective cohort study (MONICA/KORA).. *PLoS Med.* 2010;7(7):e1000314.
3. Noseworthy PA *et al.* The Early Repolarization Pattern in the General Population. *J. Am. Coll. Cardiol.* 2011;57(22):2284–2289.
4. Ueberoi A *et al.* Early repolarization in an ambulatory clinical population.. *Circulation* 2011;124(20):2208–14.
5. Trenkwalder T *et al.* Left ventricular geometry and function in early repolarization: results from the population-based Gutenberg Health Study.. *Clin. Res. Cardiol.* [published online ahead of print: February 28, 2019]; doi:10.1007/s00392-019-01445-7
6. Haïssaguerre M *et al.* Sudden cardiac arrest associated with early repolarization.. *N. Engl. J. Med.* 2008;358(19):2016–23.
7. Priori SG *et al.* HRS/EHRA/APHRS expert consensus statement on the diagnosis and management of patients with inherited primary arrhythmia syndromes: document endorsed by HRS, EHRA, and APHRS in May 2013 and by ACCF, AHA, PACES, and AEPC in June 2013.. *Hear. Rhythm* 2013;10(12):1932–63.
8. Rosso R *et al.* J-point elevation in survivors of primary ventricular fibrillation and matched control subjects: incidence and clinical significance.. *J. Am. Coll. Cardiol.* 2008;52(15):1231–8.
9. Tikkanen JT *et al.* Long-term outcome associated with early repolarization on electrocardiography..

- 438 *N. Engl. J. Med.* 2009;361(26):2529–37.
- 439 10. Yan GX, Antzelevitch C. Cellular basis for the electrocardiographic J wave.. *Circulation*
- 440 1996;93(2):372–9.
- 441 11. Haïssaguerre M et al. Ventricular fibrillation with prominent early repolarization associated with a
- 442 rare variant of KCNJ8/KATP channel.. *J. Cardiovasc. Electrophysiol.* 2009;20(1):93–8.
- 443 12. Chauveau S et al. Early repolarization syndrome caused by de novo duplication of KCND3
- 444 detected by next-generation sequencing.. *Hear. case reports* 2017;3(12):574–578.
- 445 13. Yao H et al. SCN1B β mutations that affect their association with Kv4.3 underlie early
- 446 repolarization syndrome.. *J. Cell. Mol. Med.* 2018;22(11):5639–5647.
- 447 14. Nunn LM et al. Prevalence of J-point elevation in sudden arrhythmic death syndrome families.. *J.*
- 448 *Am. Coll. Cardiol.* 2011;58(3):286–90.
- 449 15. Mellor G et al. The Prevalence and Significance of the Early Repolarization Pattern in Sudden
- 450 Arrhythmic Death Syndrome Families.. *Circ. Arrhythm. Electrophysiol.* 2016;9(6).
- 451 doi:10.1161/CIRCEP.116.003960
- 452 16. Reinhard W et al. Heritability of early repolarization: a population-based study.. *Circ. Cardiovasc.*
- 453 *Genet.* 2011;4(2):134–8.
- 454 17. Bastiaenen R et al. The narrow-sense and common single nucleotide polymorphism heritability of
- 455 early repolarization.. *Int. J. Cardiol.* [published online ahead of print: October 4, 2018];
- 456 doi:10.1016/j.ijcard.2018.09.119
- 457 18. Sinner MF et al. A meta-analysis of genome-wide association studies of the electrocardiographic
- 458 early repolarization pattern.. *Hear. Rhythm* 2012;9(10):1627–34.
- 459 19. Macfarlane PW et al. The Early Repolarization Pattern: A Consensus Paper.. *J. Am. Coll. Cardiol.*
- 460 2015;66(4):470–7.
- 461 20. GTEx Consortium et al. Genetic effects on gene expression across human tissues.. *Nature*
- 462 2017;550(7675):204–213.
- 463 21. MacArthur J et al. The new NHGRI-EBI Catalog of published genome-wide association studies
- 464 (GWAS Catalog). *Nucleic Acids Res.* 2017;45(D1):D896–D901.
- 465 22. Christophersen IE et al. Fifteen Genetic Loci Associated With the Electrocardiographic P Wave..
- 466 *Circ. Cardiovasc. Genet.* 2017;10(4). doi:10.1161/CIRCGENETICS.116.001667
- 467 23. Roselli C et al. Multi-ethnic genome-wide association study for atrial fibrillation.. *Nat. Genet.*
- 468 2018;50(9):1225–1233.
- 469 24. Nielsen JB et al. Biobank-driven genomic discovery yields new insight into atrial fibrillation
- 470 biology.. *Nat. Genet.* 2018;50(9):1234–1239.
- 471 25. Verweij N et al. Genetic Determinants of P Wave Duration and PR Segment. *Circ. Cardiovasc.*
- 472 *Genet.* 2014;7(4):475–481.
- 473 26. Verweij N et al. Twenty-eight genetic loci associated with ST-T-wave amplitudes of the
- 474 electrocardiogram.. *Hum. Mol. Genet.* 2016;25(10):2093–2103.
- 475 27. Mercer BN et al. Early Repolarization Syndrome; Mechanistic Theories and Clinical Correlates..
- 476 *Front. Physiol.* 2016;7:266.
- 477 28. Haïssaguerre M et al. Characteristics of recurrent ventricular fibrillation associated with
- 478 inferolateral early repolarization role of drug therapy.. *J. Am. Coll. Cardiol.* 2009;53(7):612–9.
- 479 29. Mavragani-Tsipidou P, Scouras ZG, Haralampidis K, Lavrentiadou S, Kastritsis CD. The polytene
- 480 chromosomes of *Drosophila triauraria* and *D. quadraria*, sibling species of *D. auraria*.. *Genome*
- 481 1992;35(2):318–26.
- 482 30. Haïssaguerre M et al. Depolarization versus repolarization abnormality underlying inferolateral J-
- 483 wave syndromes: New concepts in sudden cardiac death with apparently normal hearts.. *Hear.*
- 484 *Rhythm* [published online ahead of print: November 2, 2018]; doi:10.1016/j.hrthm.2018.10.040
- 485 31. Bonasio R, Shiekhhattar R. Regulation of transcription by long noncoding RNAs.. *Annu. Rev.*
- 486 *Genet.* 2014;48(1):433–55.
- 487 32. Ulitsky I. Evolution to the rescue: using comparative genomics to understand long non-coding
- 488 RNAs.. *Nat. Rev. Genet.* 2016;17(10):601–14.
- 489 33. Sallam T, Sandhu J, Tontonoz P. Long Noncoding RNA Discovery in Cardiovascular Disease:
- 490 Decoding Form to Function.. *Circ. Res.* 2018;122(1):155–166.
- 491 34. Harismendy O et al. 9p21 DNA variants associated with coronary artery disease impair interferon-
- 492 γ signalling response.. *Nature* 2011;470(7333):264–8.
- 493 35. Broadbent HM et al. Susceptibility to coronary artery disease and diabetes is encoded by distinct,
- 494 tightly linked SNPs in the ANRIL locus on chromosome 9p.. *Hum. Mol. Genet.* 2008;17(6):806–14.
- 495 36. Koncz I et al. Mechanisms underlying the development of the electrocardiographic and arrhythmic
- 496 manifestations of early repolarization syndrome.. *J. Mol. Cell. Cardiol.* 2014;68:20–8.
- 497 37. Antzelevitch C et al. J-Wave syndromes expert consensus conference report: Emerging concepts
- 498 and gaps in knowledge.. *Europace* 2017;19(4):665–694.

499 38. Medeiros-Domingo A et al. Gain-of-function mutation S422L in the KCNJ8-encoded cardiac
500 K(ATP) channel Kir6.1 as a pathogenic substrate for J-wave syndromes.. *Hear. Rhythm*
501 2010;7(10):1466–71.
502 39. Barajas-Martínez H et al. Molecular genetic and functional association of Brugada and early
503 repolarization syndromes with S422L missense mutation in KCNJ8. *Hear. Rhythm* 2012;9(4):548–555.
504 40. Landrum MJ et al. ClinVar: public archive of interpretations of clinically relevant variants.. *Nucleic*
505 *Acids Res.* 2016;44(D1):D862-8.
506 41. Tikkanen JT et al. Early repolarization: electrocardiographic phenotypes associated with favorable
507 long-term outcome.. *Circulation* 2011;123(23):2666–73.
508 42. Fuchsberger C, Taliun D, Pramstaller PP, Pattaro C, CKDGen consortium. GWAtoolbox: an R
509 package for fast quality control and handling of genome-wide association studies meta-analysis data..
510 *Bioinformatics* 2012;28(3):444–445.
511 43. Willer CJ, Li Y, Abecasis GR. METAL: fast and efficient meta-analysis of genomewide association
512 scans.. *Bioinformatics* 2010;26(17):2190–1.
513 44. Bulik-Sullivan BK et al. LD Score regression distinguishes confounding from polygenicity in
514 genome-wide association studies.. *Nat. Genet.* 2015;47(3):291–5.
515 45. Higgins JPT, Thompson SG, Deeks JJ, Altman DG. Measuring inconsistency in meta-analyses..
516 *BMJ* 2003;327(7414):557–560.
517 46. Yang J, Lee SH, Goddard ME, Visscher PM. GCTA: a tool for genome-wide complex trait
518 analysis.. *Am. J. Hum. Genet.* 2011;88(1):76–82.
519 47. Zhu Z et al. Integration of summary data from GWAS and eQTL studies predicts complex trait
520 gene targets.. *Nat. Genet.* 2016;48(5):481–7.
521

Table 1: Baseline characteristics of the study populations

Study (discovery stage)	BRIGHT		GHS1		GHS2		GRAPHIC	
	ERP+	ERP-	ERP+	ERP-	ERP+	ERP-	ERP+	ERP-
Number of samples (n)	189	1173	182	2628	70	1028	57	893
Females (n)	105	747	60	1358	26	536	18	457
Age in years (mean±SD)	57.6±12.1	59.4±12.3	54.5±10.0	55.6±10.9	54.0±10.2	54.9±10.9	52.3±3.9	52.8±4.5
Heart rate in bpm (mean±SD)	61.7±9.9	63.7±11.2	67.6±11.5	69.1±10.8	67.1±11.4	68.7±10.8	63.5±8.0	64.1±9.8
BMI (mean±SD)	27.7±3.4	27.4±3.8	26.8±4.4	27.1±4.7	27.5±5.5	27.2±4.9	27.4±4.0	27.4±4.3
	Lifelines		SHIP		SHIP-Trend		TwinsUK	
	ERP+	ERP-	ERP+	ERP-	ERP+	ERP-	ERP+	ERP-
Number of samples (n)	1253	11463	173	2835	86	848	171	2773
Females (n)	639	6902	79	1508	38	494	150	2651
Age in years (mean±SD)	48.0±11.5	47.9±11.3	46.6±16.1	48.5±15.8	49.8±14.5	49.7±13.4	51.7±13.2	52.7±12.4
Heart rate in bpm (mean±SD)	66.3±10.9	68.4±11.5	70.5±11.6	73.7±11.6	64.4±8.9	65.9±9.6	64.1±10.3	66.8±10.4
BMI (mean±SD)	25.7±3.8	26.4±4.3	25.9±4.2	27.3±4.9	26.9±4.4	27.3±4.6	25.3±4.4	25.7±4.6
Study (replication stage)	CHRIS		Rotterdam Study I		Rotterdam Study II		Rotterdam Study III	
	ERP+	ERP-	ERP+	ERP-	ERP+	ERP-	ERP+	ERP-
Number of samples (n)	427	3953	308	4438	164	1476	225	2643
Females (n)	159	2318	182	2739	84	825	116	1541
Age in years (mean±SD)	45.2±16.3	45.7±16.1	66.4±7.6	66.3±7.7	64.1±7.3	64.4±7.5	56.7±5.6	57.0±6.7
Heart rate in bpm (mean±SD)	60.3±8.9	62.5±8.8	68.7±11.6	69.2±11.9	67.5±10.6	68.8±10.8	69.0±11.7	69.6±10.5
BMI (mean±SD)	25.4±4.2	25.6±4.6	27.5±7.4	27.1±6.9	27.5±4.1	27.5±4.1	27.6±4.9	27.5±5.0

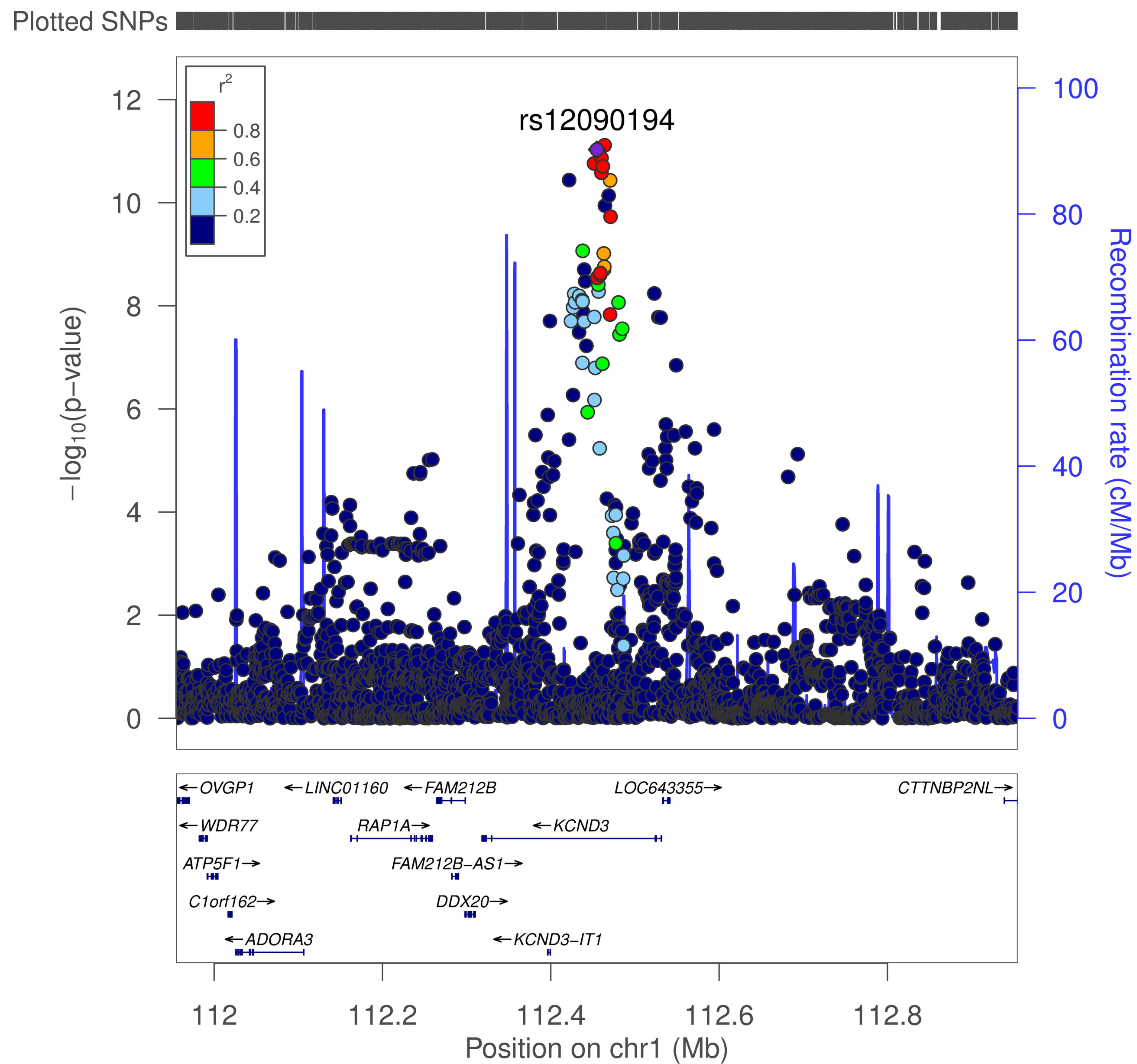
ERP+: cases with early repolarization pattern; ERP-: controls; SD: standard deviation; bpm: beats per minute; BMI: body mass index

Table 2: Lead SNPs of the GWAS association results

variant information				discovery					replication				combined			
SNP	chr:position	A1/A2	nearest gene	AF1	OR	P	I ²	N	OR	P	I ²	N	OR	P	I ²	N
rs12090194	1:112,454,822	t/c	KCND3	0.32	0.80	4.6E-10	34	25177	0.86	2.5E-03	39	13634	0.82	9.3E-12	35	38811
					[0.75-0.86]				[0.79-0.95]				[0.78-0.87]			
rs1545300	1:112,464,004	t/c	KCND3	0.32	0.81	1.4E-09	41	25172	0.85	9.4E-04	56	13634	0.82	7.7E-12	43	38806
					[0.75-0.86]				[0.77-0.94]				[0.78-0.87]			
rs139772527	16:208,761	t/c	HBZ	0.01	2.57	2.0E-08	0	21495	1.21	2.8E-01	0	13634	1.81	1.4E-06	11	35129
					[1.85-3.58]				[0.85-1.73]				[1.42-2.31]			

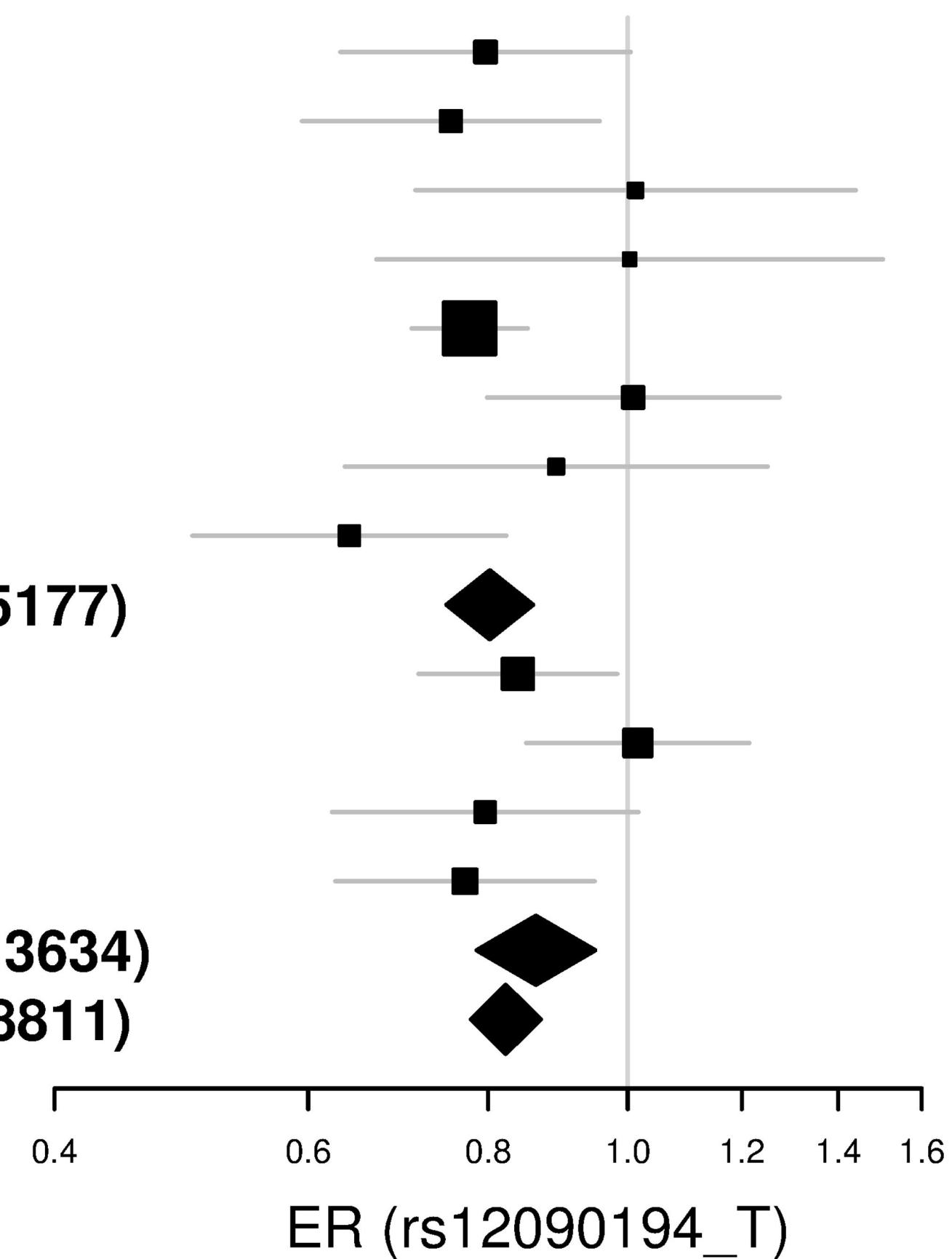
A1: effect allele; AF1: allele frequency of A1; OR: odds ratio of A1 [95% confidence interval]; P: association p-value; I²: percentage of total variation across studies that is due to heterogeneity; N: sample size. Bold values indicate the lead SNP (lowest p-value) of a significantly associated locus in the corresponding meta-analysis stage.

A

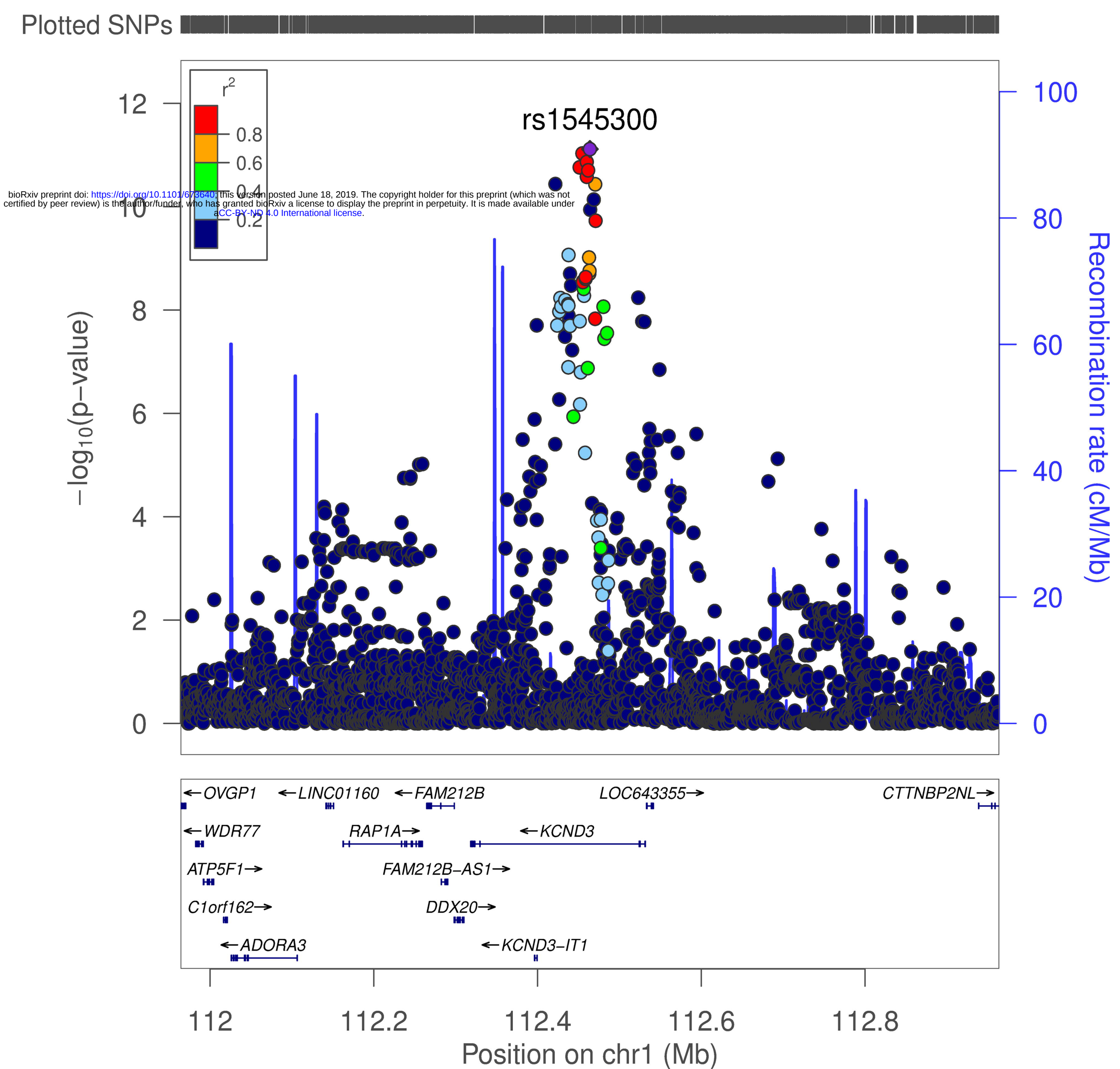


B

BRIGHT (n=738)
GHS1 (n=2810)
GHS2 (n=1098)
GRAPHIC (n=929)
LifeLines (n=12716)
SHIP (n=3008)
SHIP-Trend (n=934)
TwinsUK (n=2944)
discovery: $I^2=34.1$ (n=25177)
CHRIS (n=4380)
RS1 (n=4746)
RS2 (n=1640)
RS3 (n=2868)
replication: $I^2=38.6$ (n=13634)
combined: $I^2=35.3$ (n=38811)

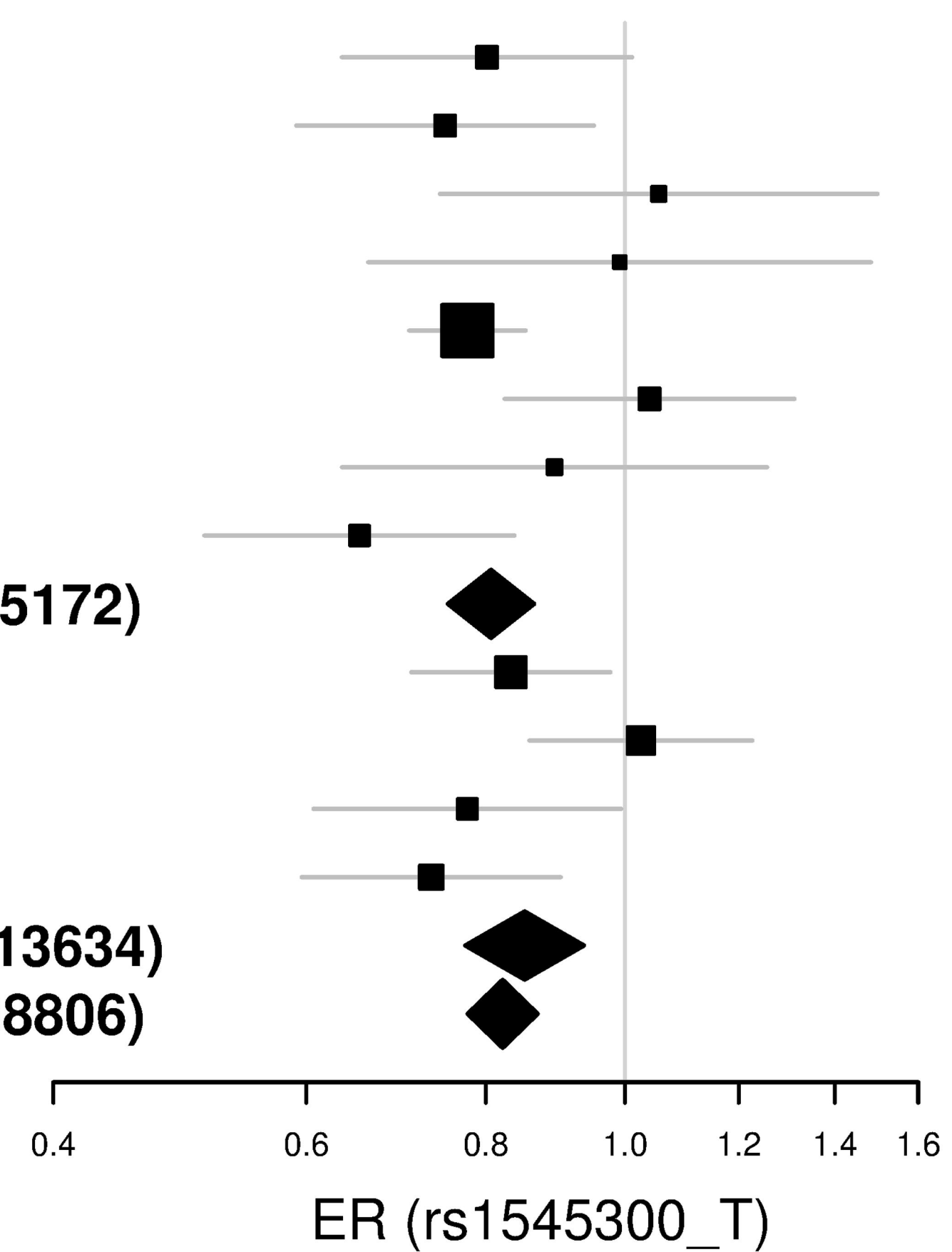


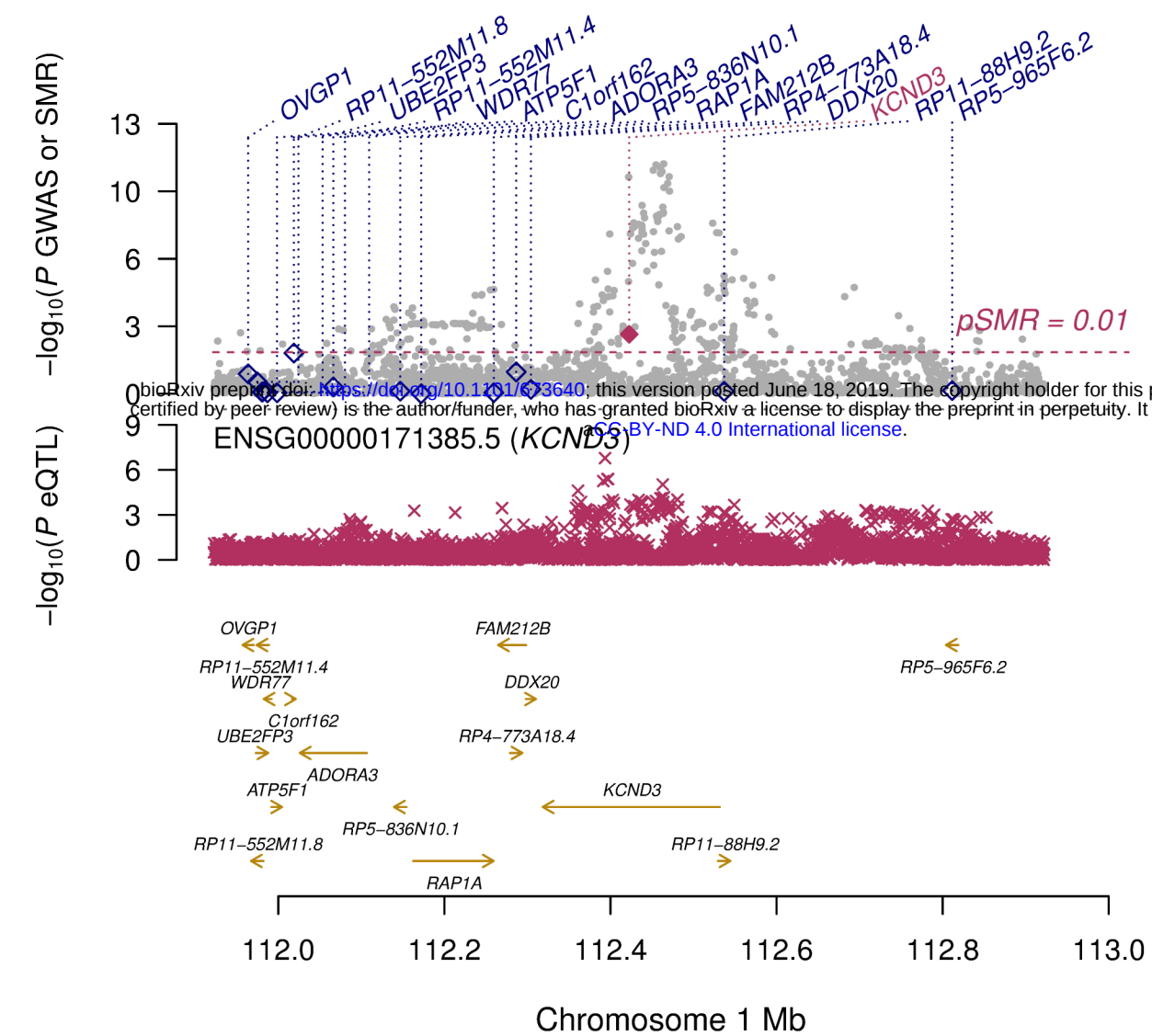
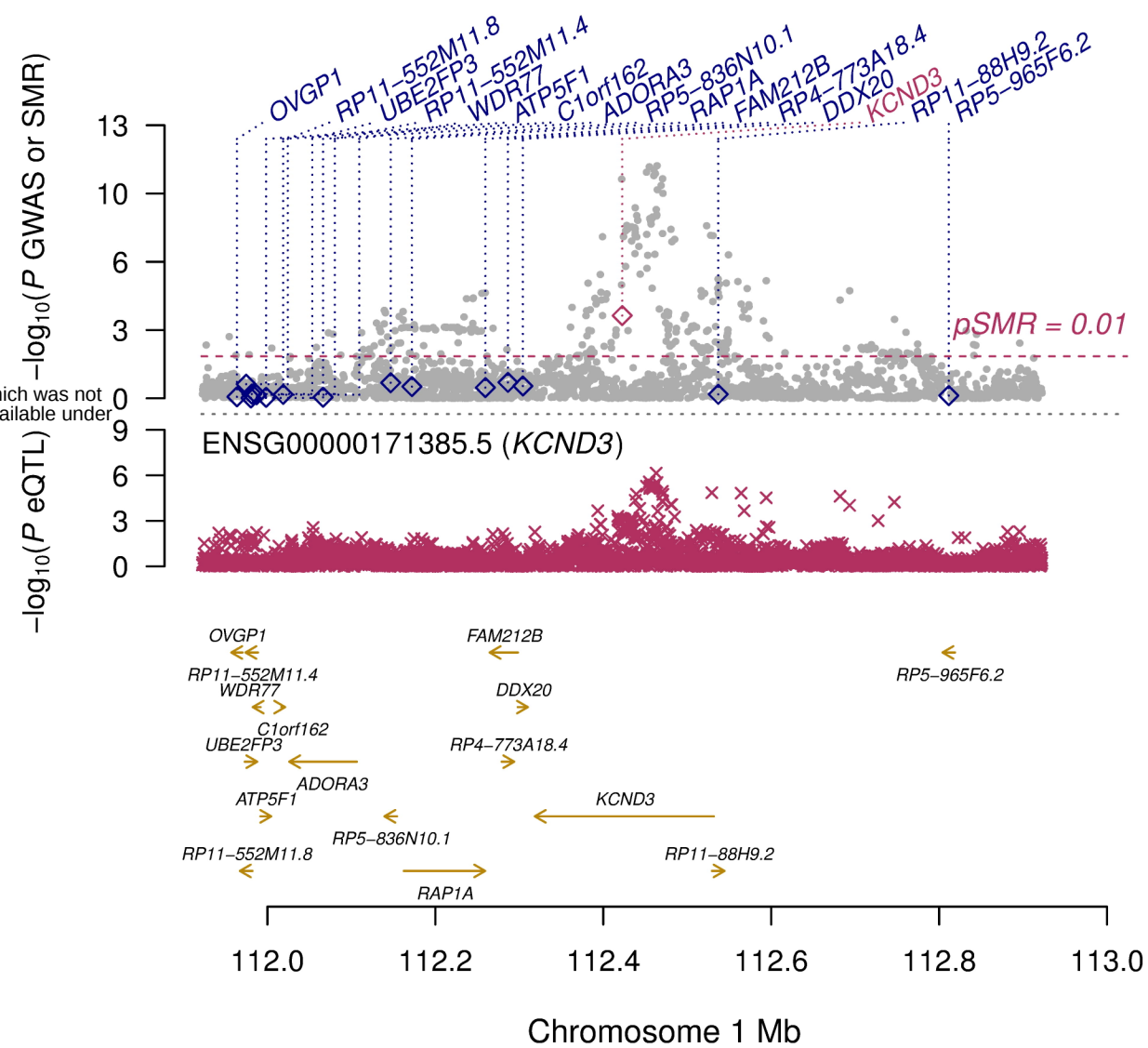
C



D

BRIGHT (n=735)
GHS1 (n=2810)
GHS2 (n=1098)
GRAPHIC (n=929)
LifeLines (n=12716)
SHIP (n=3008)
SHIP-Trend (n=934)
TwinsUK (n=2942)
discovery: $I^2=41.0$ (n=25172)
CHRIS (n=4380)
RS1 (n=4746)
RS2 (n=1640)
RS3 (n=2868)
replication: $I^2=55.7$ (n=13634)
combined: $I^2=43.4$ (n=38806)



A**B****C**