

1 Area 2 of primary somatosensory cortex 2 encodes kinematics of the whole arm

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11 ABSTRACT

12 Proprioception, the sense of body position, movement, and associated forces, remains poorly
13 understood, despite its critical role in movement. Most studies of area 2, a proprioceptive area of
14 somatosensory cortex, have simply compared neurons' activities to the movement of the hand
15 through space. By using motion tracking, we sought to elaborate this relationship by
16 characterizing how area 2 activity relates to whole arm movements. We found that a whole-arm
17 model, unlike classic models, successfully predicted how features of neural activity changed as
18 monkeys reached to targets in two workspaces. However, when we then evaluated this whole-
19 arm model across active and passive movements, we found that many neurons did not
20 consistently represent the whole arm over both conditions. These results suggest that 1) neural
21 activity in area 2 includes representation of the whole arm during reaching and 2) many of these
22 neurons represented limb state differently during active and passive movements.

23 1 INTRODUCTION

24 Moving in an uncontrolled environment is a remarkably complex feat. In addition to the
25 necessary computations on the efferent side to generate movement, an important aspect of
26 sensorimotor control is processing the afferent information we receive from our limbs, essential
27 both for movement planning and for the feedback it provides during movement. Of the relevant
28 sensory modalities, proprioception, or the sense of body position, movement and associated
29 forces, is arguably the most critical for making coordinated movements (Ghez and Sainburg
30 1995; Gordon et al. 1995; Sainburg et al. 1995; Sainburg et al. 1993; Sanes et al. 1984).
31 However, despite its importance, few studies have explicitly addressed how proprioception is
32 represented in the brain during natural movement; touch, vision, and the motor areas of the brain
33 have received far more attention.

34 One brain area likely important for mediating reach-related proprioception is the proximal arm
35 representation within area 2 of primary somatosensory cortex (S1) (Jennings et al. 1983; Kaas et
36 al. 1979; London and Miller 2013). Though this area receives a combination of muscle and
37 cutaneous inputs (Hyvärinen and Poranen 1978; Padberg et al. 2018; Pons et al. 1985), the few
38 studies examining it during reaching have found that a model involving simply the translation of
39 the hand approximates neural activity quite well (London and Miller 2013; London et al. 2011;
40 Prud'homme and Kalaska 1994; Weber et al. 2011). Interestingly, this finding fits with
41 psychophysical data showing that humans are better at estimating the location of the hand than
42 joint angles (Fuentes and Bastian 2010), as well as our conscious experience of reaching to
43 objects, which typically focuses on the hand. However, recent computational studies have shown
44 that while neural activity may appear to be tuned to the state of a limb's endpoint, features of this
45 tuning might be a direct consequence of the biomechanics of the limb (Chowdhury et al. 2017;
46 Lillicrap and Scott 2013). Consistent with those results, we have recently observed, using
47 artificial neural networks, that muscle lengths were better predictors of area 2 activity than
48 were hand kinematics (Lucas et al. 2019).

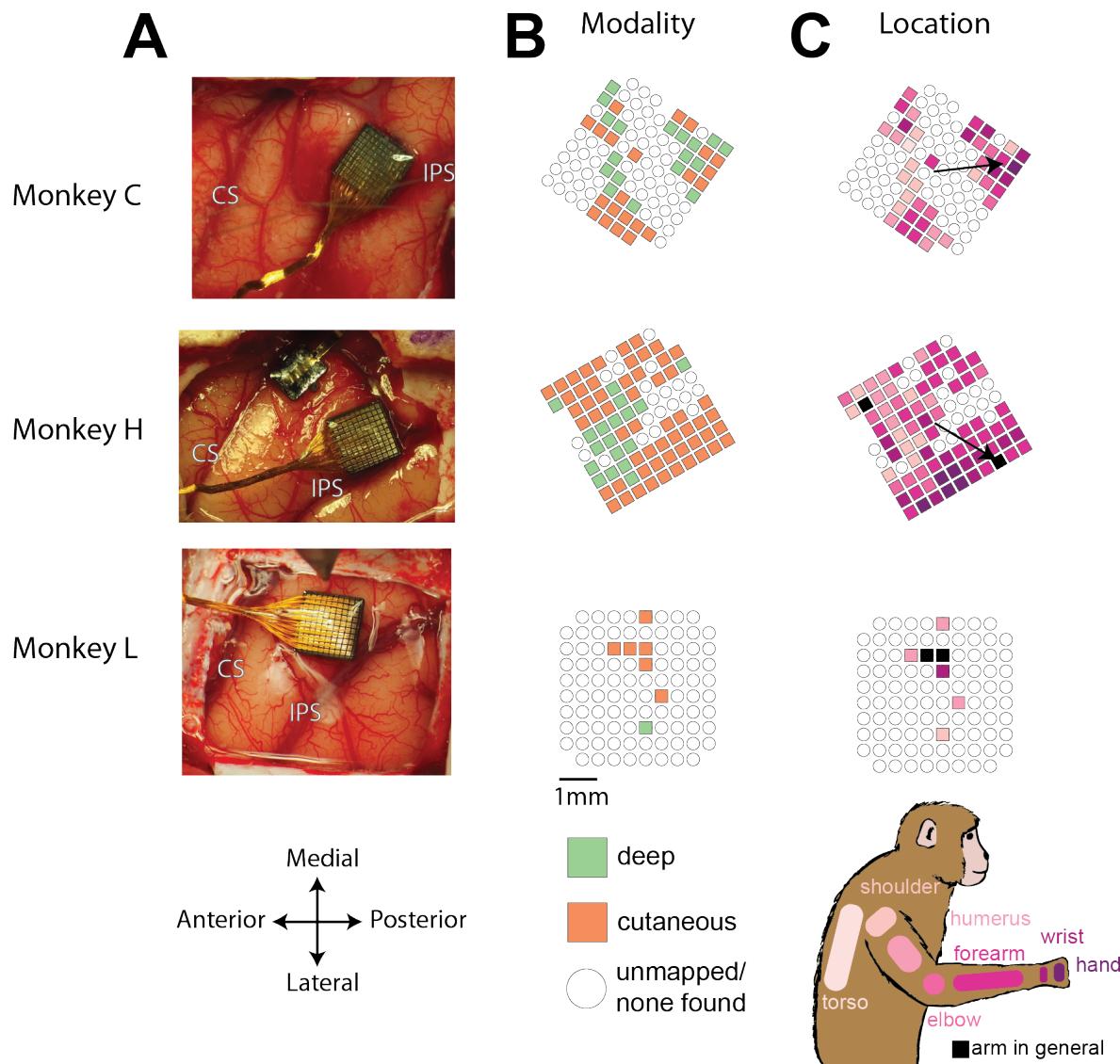
49 As in the classic reaching studies of M1 (Caminiti et al. 1991; Georgopoulos et al. 1982;
50 Georgopoulos et al. 1986), the appeal of the hand-based model of area 2 neural activity is its
51 reasonable accuracy despite its simplicity. However, the recent emphasis on studying less
52 constrained, more natural movements (Mazurek et al. 2018) is pushing the limits of such simple
53 models (Berger and Gail 2018; Hasson et al. 2012; Sharon and Nisky 2017). As in the motor
54 system, it is increasingly important to characterize proprioceptive regions' responses to reaching
55 more fully. Here, we used two experiments that altered the relationship between hand and whole-
56 arm kinematics. In the first experiment, we found that neurons in area 2 have a consistent
57 relationship with whole-arm kinematics during active reaching within two disjoint workspaces.
58 Whole-arm kinematics predicted neural activity significantly better than the hand-only model,
59 and were able to effectively explain neural activity changes across workspaces. In the second
60 experiment, we compared area 2 responses to active reaching and passive perturbations of the
61 hand. While some neurons were predicted well with only kinematic inputs, others were not,
62 adding to the evidence that area 2 may receive efferent information from motor areas of the brain
63 (London and Miller 2013; Nelson 1987).

64 2 RESULTS

65 For the experiments detailed in this paper, we recorded neural signals from three Rhesus
66 macaques (Monkeys C, H, and L) using Utah multi-electrode arrays (Blackrock Microsystems)
67 implanted in the arm representation of Brodmann's area 2 of S1 (Figure 1). After implantation,
68 we mapped sensory receptive fields of each neuron, to examine how the multi-unit activity on
69 each electrode responded to sensory stimulation, noting the modality (deep or cutaneous) and
70 location of each field. We classified an electrode as "cutaneous" if we could find a receptive
71 field on the arm or torso in which brushing the skin caused an increase in activity. "Deep"
72 electrodes were those that responded to joint movement or muscle palpation and did not appear
73 to have a cutaneous receptive field. With these criteria, it is likely that some of the electrodes we

74 marked cutaneous actually responded to both deep and cutaneous stimuli. However, as we were
75 most interested in the distribution of receptive field types over the array, we did not test for such
76 mixed modality neurons.

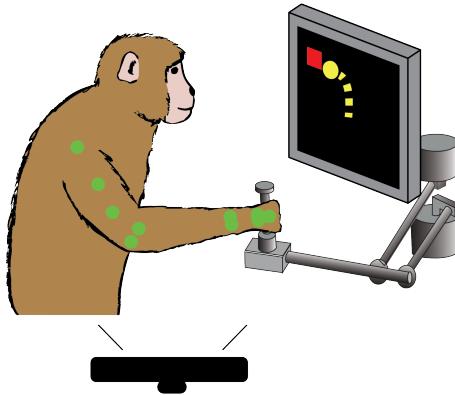
77 Figure 1 shows the resulting sensory maps from the mapping session for each monkey in which
78 we were able to test the most electrodes. We found a mix of deep and cutaneous receptive fields
79 across each array, largely matching the description of area 2 from previous studies (Hyvärinen
80 and Poranen 1978; Pons et al. 1985; Seelke et al. 2011). Compared to the two bordering regions,
81 area 1 tends to have a higher fraction of cutaneous responses, and area 5 tends to have a higher
82 fraction of deep responses (Seelke et al. 2011), suggesting that our arrays were implanted largely
83 in area 2. For Monkeys C and H, we found a rough proximal to distal arm gradient, running from
84 anterior to posterior across the array (Figure 1, black arrows), consistent with the somatotopy
85 found by (Pons et al. 1985). There were too few well-mapped neurons from Monkey L to
86 determine a meaningful gradient.



87

88 *Figure 1: Array locations and receptive field maps from one mapping session for each monkey. A –*
89 *locations of Utah arrays implanted in area 2 of Monkeys C, H, and L. IPS, intraparietal sulcus; CS*
90 *central sulcus. B – map of the receptive field modality (deep, cutaneous, or mixed) for each electrode. C –*
91 *map of receptive field location (see legend on bottom right). Open circles indicate both untested*
92 *electrodes and tested electrodes with no receptive field found. Black arrows on maps in C show*
93 *significant gradient across array of proximal to distal receptive fields (see Methods).*

94 We trained each of these monkeys to grasp a two-link planar manipulandum and make reaching
95 movements to targets presented on a screen in front of them (Figure 2). During these sessions,
96 we collected interface force from a six degree of freedom load cell attached to the manipulandum
97 handle. We also tracked the locations of markers on the monkey's arm using a custom motion
98 tracking system based on a Microsoft Kinect. Our experiments included two components: one
99 comparing reaching movements in two different workspaces and one comparing active and
100 passive movements.



101

102 *Figure 2: Behavioral task. Monkey controls a cursor on screen (yellow) with a two link manipulandum to*
103 *reach to visually presented targets (red). We track the locations of markers (see Methods) on the*
104 *monkey's arm (green) during the task, using a Microsoft Kinect.*

105 2.1 SOMATOSENSORY AREA 2 REPRESENTS THE MOVEMENT OF THE WHOLE ARM DURING REACHING

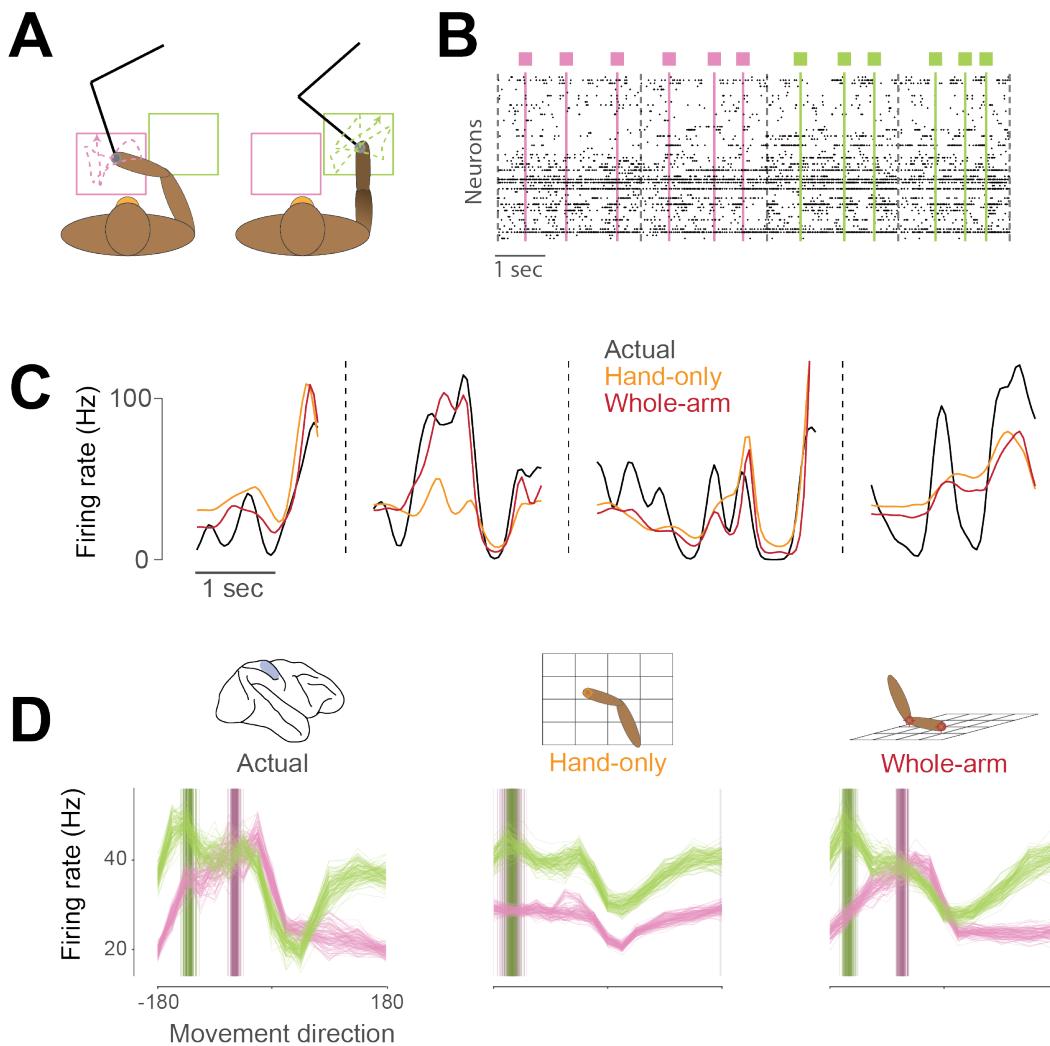
106 Previous literature has characterized area 2 primarily in terms of the hand trajectory through
107 space (London and Miller 2013; Prud'homme and Kalaska 1994; Weber et al. 2011), likely in
108 part due to the difficulty of tracking the motion of the full arm, and the then recent finding that
109 motor cortex could be well explained simply by the direction of hand movement (Caminiti et al.
110 1991; Georgopoulos et al. 1982). Given advances in motion tracking capability and subsequent
111 observations of the dependence of M1 on arm posture (Morrow et al. 2007; Scott and Kalaska
112 1995), we set out to characterize more fully, how neural activity in area 2 corresponds to
113 reaching movements.

114 In particular, we aimed to characterize how much could be gained by using models incorporating
115 the movement of the whole arm, as opposed to just the hand. A challenge in comparing these
116 models is that for the typical, center-out reaching task in a small workspace, the behavioral
117 signals used in our models are highly correlated. Because a high correlation means that a linear
118 transform can accurately convert one set of signals into another, all models would make very
119 similar predictions of neural activity.

120 To deal with this problem, we trained the monkeys to reach to randomly-generated targets
121 presented in two different workspaces (Figure 3). This had two important effects. First, the
122 random locations of the targets lessened the stereotypy of the movements, allowing for the
123 collection of more varied movement data than from a center-out paradigm. Second, the average
124 postures in the two workspaces were quite different, such that while the signals of different
125 models were still correlated within a given workspace, this correlation (and the mapping between
126 sets of behavioral signals) changed significantly between workspaces. This forced the models to
127 make different predictions of neural activity across the two workspaces. By comparing modeled
128 and observed changes in neural activity, we could more reliably discriminate between models.

129 This idea is exemplified in Figure 3D. When tested in the two workspaces, this example neuron
130 changed both its tuning curve and the direction in which it fired maximally (its preferred

131 direction, or PD), as did many neurons we recorded. The corresponding predictions of the hand-
132 only and whole-arm models differed, which allowed us to compare the accuracy of the two
133 models. We recorded three of these two-workspace sessions with each of Monkeys C and H and
134 two sessions with Monkey L.



135

136 *Figure 3: Example neural activity for two-workspace task. A – Two-workspace behavior. On each trial,*
137 *monkey reaches with manipulandum (black) to randomly placed targets in one of two workspaces: one*
138 *close to the body and contralateral to the reaching hand (pink) and the other distant and ipsilateral*
139 *(green). Trials in the two workspaces were interleaved randomly. B – Example neural raster plot from*
140 *one session for two randomly drawn trials in each workspace. Dots in each row represent activity for one*
141 *of the simultaneously recorded neurons. Black dashed lines indicate starts and ends of trials, and colored*
142 *lines and boxes indicate times of target presentation, with color indicating the workspace for the trial. C*
143 *– Firing rate plot for an example neuron during four randomly drawn trials from the distal (green)*
144 *workspace. Black trace represents neural firing rate, smoothed with a 50 ms Gaussian kernel. Colored*
145 *traces represent unsmoothed firing rates predicted by hand-only (orange), and whole-arm (red) models.*
146 *D – Actual and predicted tuning curves and preferred directions (PDs) computed in the two workspaces*
147 *for an example neuron. Each trace represents the tuning curve or PD calculated for one cross-validation*
148 *fold (see Methods). Leftmost plot shows actual tuning curves and PDs, while other plots show curves and*

149 *PDs for activity predicted by each of the models. Each panel shows mean movement-related firing rate*
150 *plotted against direction of hand movement for both workspaces. Darker vertical bars indicate preferred*
151 *directions.*

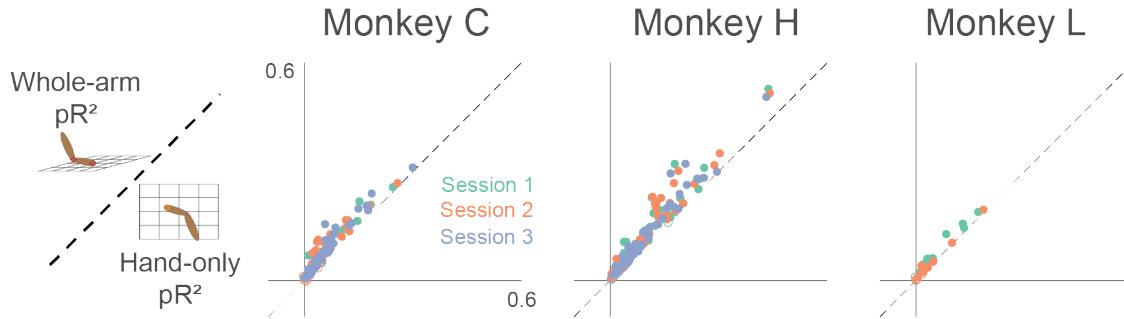
152 **2.1.1 Model overview**

153 We tested several kinematic models of area 2 activity that could be divided into hand-only and
154 whole-arm models (see Methods for a full description of all the models). We've chosen to
155 represent the two sets with two of the models, which we termed, for simplicity, the "hand-only"
156 and "whole-arm" models. The hand-only model stems from classic, endpoint models of limb
157 movement-related neural activity (Bosco et al. 2000; Georgopoulos et al. 1982; Prud'homme and
158 Kalaska 1994). It assumes neurons relate only to the Cartesian coordinates of hand position and
159 velocity. The whole-arm model builds on the hand-only model by adding the Cartesian
160 kinematics (position and velocity) of the elbow, in order to account more fully for movement of
161 the whole arm. Surprisingly, the performance of this representation of the whole arm was similar
162 to, or even better than more complicated biomechanical models based on the 7 degree-of-
163 freedom joint kinematics or musculotendon lengths (see Supplementary Info). We aimed to test
164 how well the hand-only and whole-arm models predicted features of neural activity during
165 reaching, in order to determine the importance of whole-arm kinematics for explaining neural
166 activity.

167 For us to consider the whole-arm model to be an effective one for area 2, it should satisfy three
168 main criteria. First and most direct, it should explain the variance of neural firing rates across the
169 two workspaces better than the hand-only model, as is the case in the example in Figure 3C.
170 Second, the mapping between neural activity and whole-arm kinematics should not change
171 between the individual workspaces, meaning that the accuracy of a model trained over both
172 workspaces be similar to that trained in a single workspace. Last, the model should be able to
173 capture features of neural activity that it was not explicitly trained on, for example, the changes
174 in directional tuning shown in Figure 3D.

175 **2.1.2 Whole-arm model explains more variance of area 2 neural activity than hand-only model**

176 To assess how well our models fit area 2 neural activity, we used repeated k-fold cross-validation
177 (see Methods for more details). Goodness-of-fit metrics like R^2 or variance-accounted-for (VAF)
178 are ill-suited to the Poisson-like statistics of neural activity; instead, we used the likelihood-based
179 pseudo- R^2 (Cameron and Windmeijer 1997; 1996; McFadden 1977). Like VAF, pseudo- R^2 has a
180 maximum value of 1, but it can be negative for models that fail even to predict the mean firing
181 rate during cross-validation. In general, the values corresponding to a good fit are lower for pR^2
182 than for either R^2 or VAF, with a value of 0.2 usually considered a "good" fit (McFadden 1977).
183 We found that for this measure, the whole-arm model out-performed the hand-only model
184 (Figure 4). Of the 288 neurons recorded across the 8 sessions, 238 were significantly better
185 predicted by the whole-arm model than the hand-only model, and for the other 50, there was no
186 significant difference (using $p < 0.05$; see Methods for more details).



187

188 *Figure 4: Goodness-of-fit comparison analysis. Scatter plots compare the pseudo- R^2 (pR^2) of the whole-
189 arm model to that of the hand-only model for each monkey. Each point in the scatter plot represents the
190 pR^2 values of one neuron, with whole-arm pR^2 on the vertical axis and hand-only pR^2 on the horizontal.
191 Different colors represent neurons recorded during different sessions. Filled circles represent neurons for
192 which one model's pR^2 was significantly higher than that of the other model. In this comparison, all filled
193 circles lie above the dashed unity line, indicating that the whole-arm model performed better than the
194 hand-only model for every neuron in which there was a significant difference.*

195 **2.1.3 Whole-arm model captures a consistent relationship between area 2 and arm kinematics**
196 A reasonable benchmark of how well the whole-arm model fits the two-workspace data is its
197 ability to match the accuracy of models trained in the individual workspaces. It is possible to
198 imagine a scenario in which a model might achieve a good fit by capturing a global relation
199 across the two workspaces without capturing the information local to either workspace. This
200 situation is akin to fitting a line to data distributed along an exponential curve. In this analogy,
201 we would expect a piecewise linear fit to each half of the data to achieve significantly better
202 goodness-of-fit.

203 We tested this scenario by training whole-arm models on the individual workspaces, and
204 comparing the resulting pR^2 with that calculated from the model fit to data from both
205 workspaces. The symbols lying very close to the unity line in each panel of Figure 5 indicate that
206 the full model explained just as much neural variance as did the individual models. This suggests
207 that the whole-arm model describes a consistent, generalizable relationship between neural
208 activity and arm kinematics across the two workspaces.

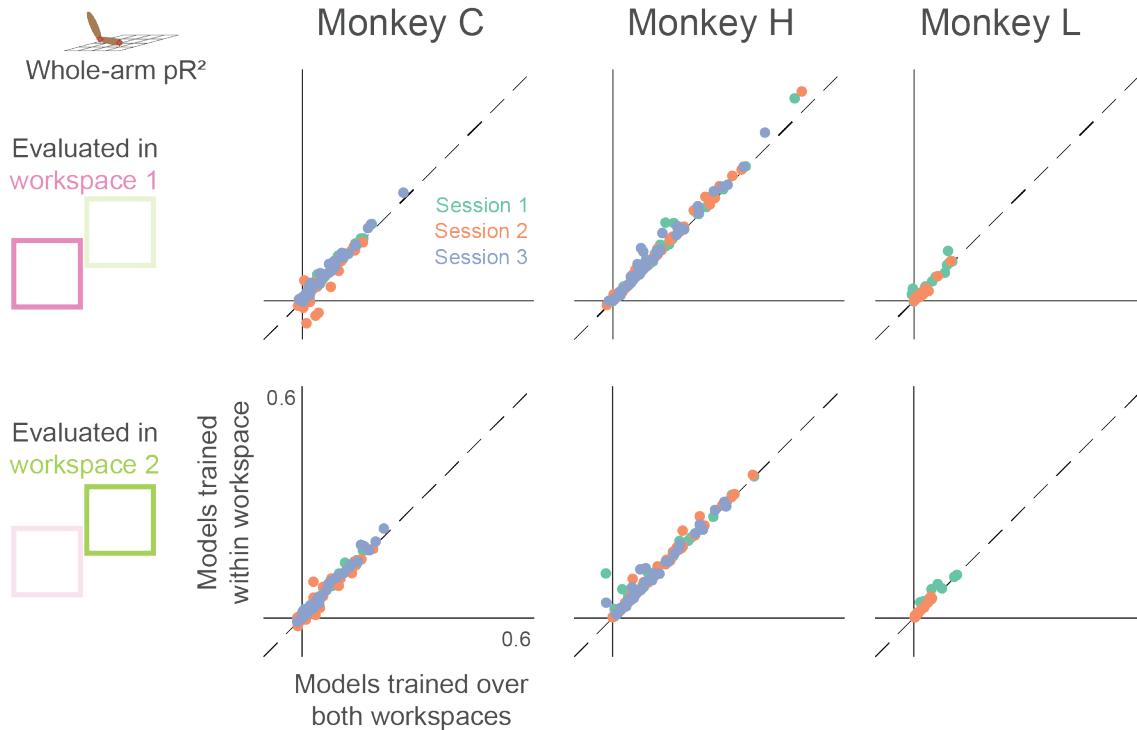
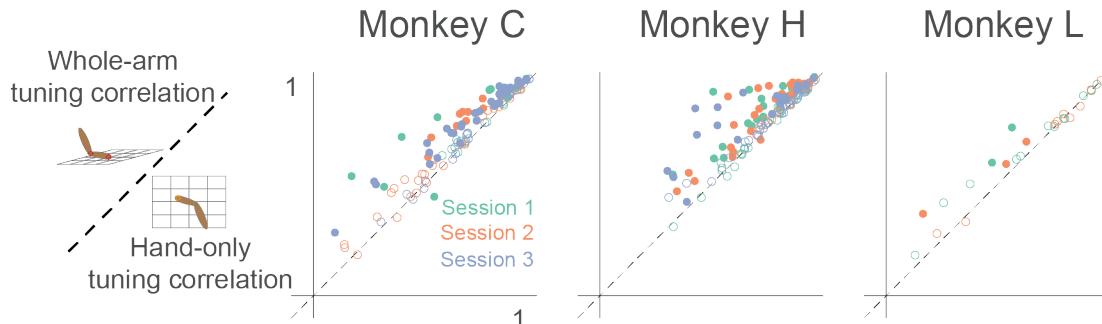


Figure 5: Dependence of whole-arm model accuracy on workspace location of training data. Each panel compares a model trained and tested in the same workspace (either near or far) to a model trained on data from both workspaces. Each dot corresponds to a single neuron, where color indicates the recording session. Dashed line is the unity line.

2.1.4 Whole-arm model captures changes in area 2 directional tuning between workspaces

From previous studies of area 2, we know that at least within a single workspace, neural activity is tuned approximately sinusoidally to the direction of hand movement (London and Miller 2013; Prud'homme and Kalaska 1994; Weber et al. 2011). Figure 3D shows the directional tuning curves for an example neuron, along with the tuning curves predicted by both models. Because we trained each model on data from both workspaces, they needed to capture a single relationship between movement and neural activity. As shown in the example in Figure 3D, the hand-only model predicted essentially the same tuning curve for both workspaces, with the exception of a baseline shift due to the position component. In contrast, the whole-arm model predicted altered tuning curves, which matched the actual ones well.

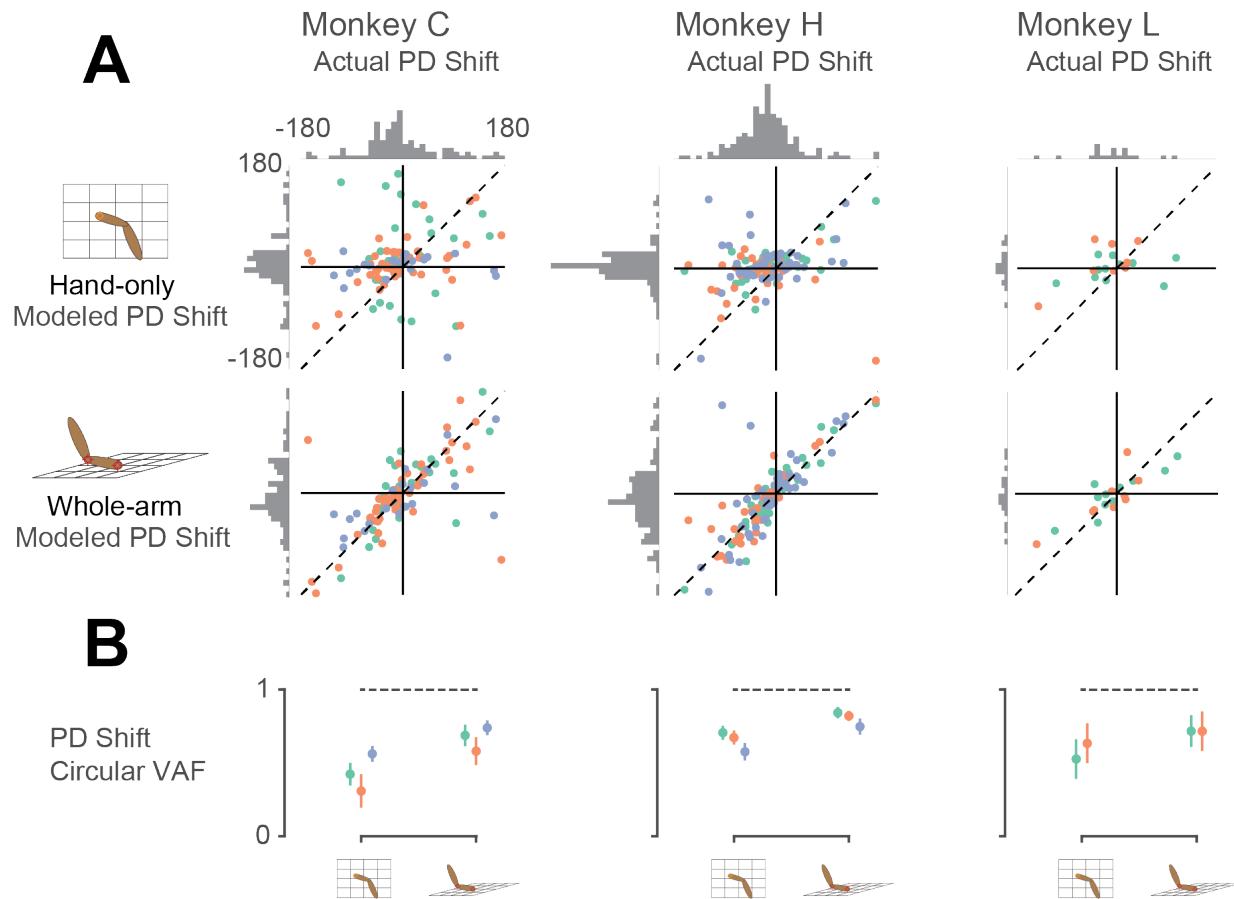
To quantify this model accuracy over all neurons, we calculated the correlation between the model-predicted and actual tuning curves in the two workspaces. With this measure, the whole-arm model once again won most of the pairwise comparisons (Figure 6). Only two out of 288 neurons were significantly better predicted by the hand-only model (using $p < 0.05$), while 138 of 288 neurons were significantly better predicted by the whole-arm model.



229

230 *Figure 6: Tuning curve shape correlation analysis. Scatter plot compares tuning curve shape correlation*
231 *between whole-arm and hand-only models. Filled circles indicate neurons significantly above or below*
232 *the dashed unity line. As for pR^2 , most filled circles lie above the dashed line of unity, indicating that the*
233 *whole-arm model was better at predicting tuning curve shape than the hand-only model.*

234 Of the 288 recorded neurons, 260 were significantly tuned to movement direction in both
235 workspaces. Thus, in addition to the tuning curve correlation analysis, we also examined the PD
236 in the two workspaces. For many neurons, the PD changed significantly between workspaces, as
237 in the leftmost panel of Figure 3D. Figure 7A shows the actual PD shifts for all neurons plotted
238 against the PD shifts predicted by each model. The large changes in PD, shown on the horizontal
239 axes of the scatter plots, are a clue that the hand-only model does not fully account for area 2
240 neural activity; if it had, the PD changes should have been insignificant (in principle, zero), as
241 shown by the generally small hand-only model-predicted changes (first row of Figure 7A).
242 Additionally, and perhaps counterintuitively, the actual changes included both clockwise and
243 counter-clockwise rotations. However, we found that the whole-arm model predicted both types
244 of PD changes quite well, indicated by a clustering of the scatter plot points in Figure 7A along
245 the dashed diagonal line. Based on the circular VAF (cVAF; see Methods for details) of the
246 predicted PD changes, Figure 7B shows that the whole-arm model once again out-performed the
247 hand-only model, with an average cVAF over all neurons of 0.75 compared to 0.57. We made
248 pairwise comparisons between models for each session. In every session but one, the whole-arm
249 model out-performed the hand-only model. In the remaining session, the difference between the
250 two models was not significant ($p > 0.05$). These results lead to the same conclusion as the pR^2
251 and tuning curve correlation analyses: the kinematics of the whole-arm are important predictors
252 of area 2 activity, and can explain differences between activity in the two workspaces that classic
253 models cannot.



254

255 *Figure 7: Model predictions of PD shift. A – Scatter plots of model-predicted PD shifts plotted against*
256 *actual PD shifts. Each dot represents the actual and modeled PD shifts of a single neuron, where*
257 *different colors correspond to neurons recorded during different sessions. Dashed diagonal line shows*
258 *perfect prediction. Horizontal histograms indicate distributions of actual PD shifts for each monkey.*
259 *Vertical histograms indicate distributions of modeled shifts. Note that both horizontal and vertical axes*
260 *are circular, meaning that opposing edges of the plots (top/bottom, left/right) are the same. Horizontal*
261 *histograms show that the distribution of actual PD shifts included both clockwise and counter-clockwise*
262 *shifts. Clustering of scatter plot points on the diagonal line for the whole-arm model indicates that it was*
263 *more predictive of PD shift. B – plot showing circular VAF (cVAF) of scatter plots in A, an indicator of*
264 *how well clustered points are around the diagonal line (see Methods for details). Each point corresponds*
265 *to the average cVAF for a model in a given session (indicated by color), and the horizontal dashed lines*
266 *indicate the cVAF for perfect prediction. Error bars show 95% confidence intervals (derived from cross-*
267 *validation – see Methods). Pairwise comparisons between model cVAFs showed that the whole-arm*
268 *model out-performed the hand-only model in all but one session, in which the two models were not*
269 *significantly different.*

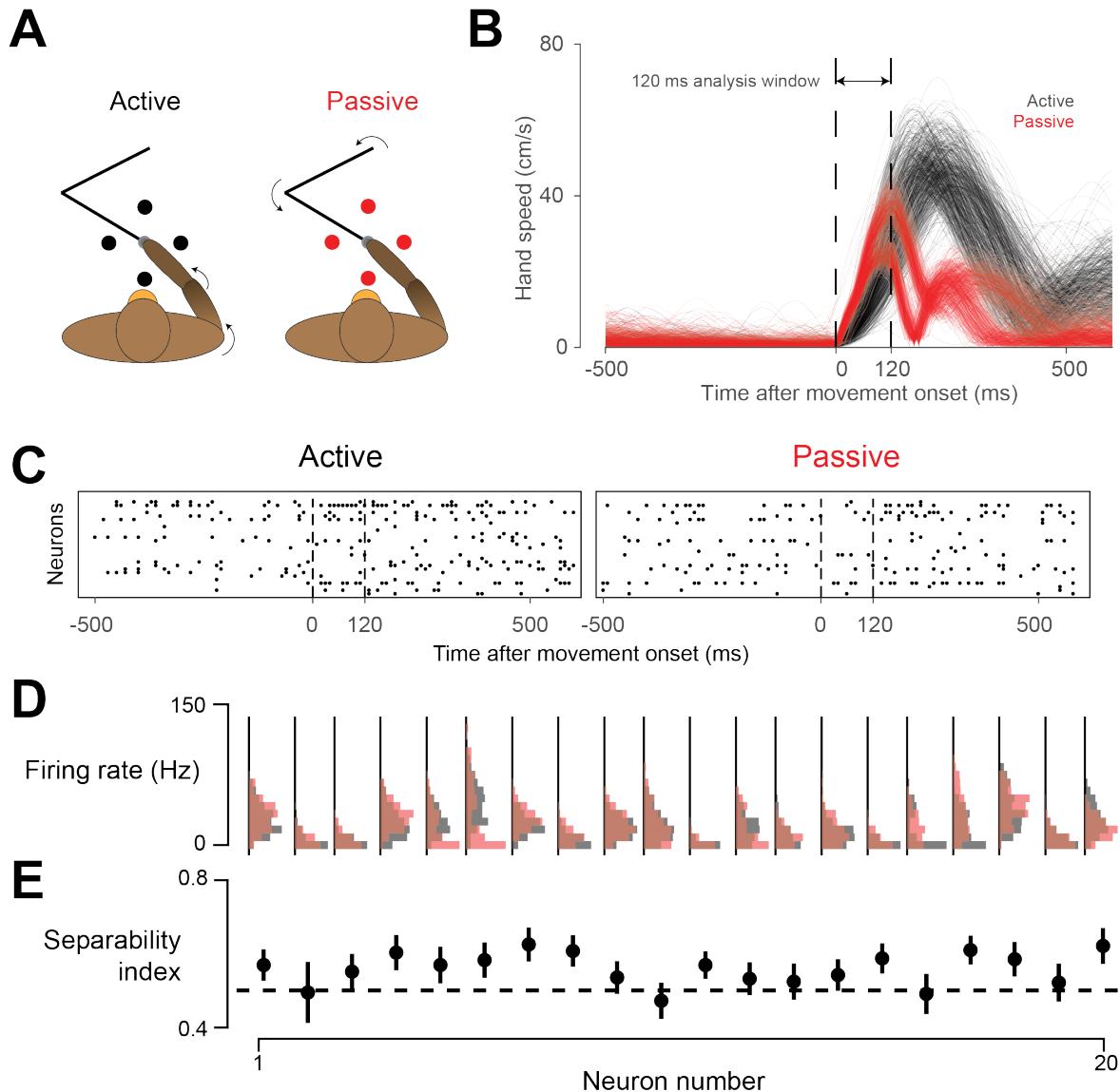
270 **2.2 AREA 2 REPRESENTS PASSIVE MOVEMENTS DIFFERENTLY FROM ACTIVE REACHES**

271 Given our success at modeling neural activity across workspaces with the whole-arm model, we
272 set out to examine its effectiveness in a task that compared area 2 activity during active reaches
273 and passive limb perturbations.

274 For this experiment, the monkey performed a center-out reaching task to four targets. On half of
275 these trials, the monkey's hand was bumped by the manipulandum during the center-hold period
276 in one of the four target directions (Figure 8A; see Methods section for task details). This
277 experiment included two sessions with each of Monkeys C and H. As in the earlier study
278 performed by our group (London and Miller 2013), we analyzed behavior and neural activity
279 only during the 120 ms after movement onset for which the kinematics of the hand were similar
280 in active and passive trials (Figure 8B and C). This is also the time period in which we can
281 reasonably expect there not to be a voluntary reaction to the bumps in the passive trials.

282 Despite the similar hand kinematics in the active and passive movements, we found that whole-
283 arm kinematics were quite different between the two conditions. Averaged over the sessions, a
284 linear discriminant analysis (LDA) classifier could predict the movement type 89% of the time,
285 using only the whole-arm kinematics in the analysis window, meaning that these whole-arm
286 kinematics were highly separable based on movement condition. Considering our results from
287 the two-workspace experiment, we would thus expect that the activity of area 2 neurons would
288 also be highly separable.

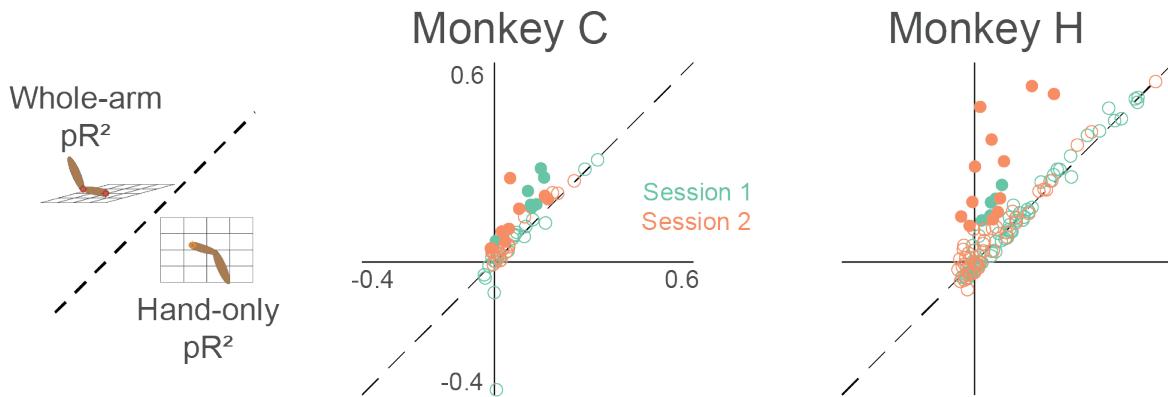
289 As reported earlier, area 2 neurons had a wide range of sensitivities to active and passive hand
290 movements (London and Miller 2013). Figure 8D shows this difference for the neurons recorded
291 during one session from Monkey C. As with our separability analysis for arm kinematics, we
292 used LDA to classify movement type based on individual neurons, calling this prediction rate the
293 neuron's "separability index" (Figure 8E). We found that many neurons had an above chance
294 separability index, as we would expect from neurons representing whole-arm kinematics.



295

296 *Figure 8: Active vs. passive behavior. A – Schematic of task. On active trials (black), monkey reaches*
 297 *from center target to a target presented in one of four directions, indicated by the black circles. On*
 298 *passive trials, manipulandum bumps monkey's hand in one of the four target directions (red circles). B –*
 299 *Speed of hand during active (black) and passive (red) trials, plotted against time, for one session. Starting*
 300 *around 120 ms after movement onset, a bimodal distribution in passive movement speed emerges. This*
 301 *bimodality reflects differences in the impedance of the arm for different directions of movement.*
 302 *Perturbations towards and away from the body tended to result in a shorter overall movement than those*
 303 *to the left or right. However, average movement speed was similar between active and passive trials in*
 304 *this 120 ms window, which we used for data analysis. C – Neural raster plots for example active and*
 305 *passive trials for rightward movements. In each plot, rows indicate spikes recorded from different*
 306 *neurons, plotted against time. Vertical dashed lines delimit the analysis window. D – Histograms of firing*
 307 *rates during active (black) and passive (red) movements for 20 example neurons from one session with*
 308 *Monkey H. E – Separability index for each neuron during the session, found by testing how well linear*
 309 *discriminant analysis (LDA) could predict movement type from the neuron's average firing rate on a*
 310 *given trial. Black dashed line indicates chance level separability. Error bars indicate 95% confidence*
 311 *interval of separability index.*

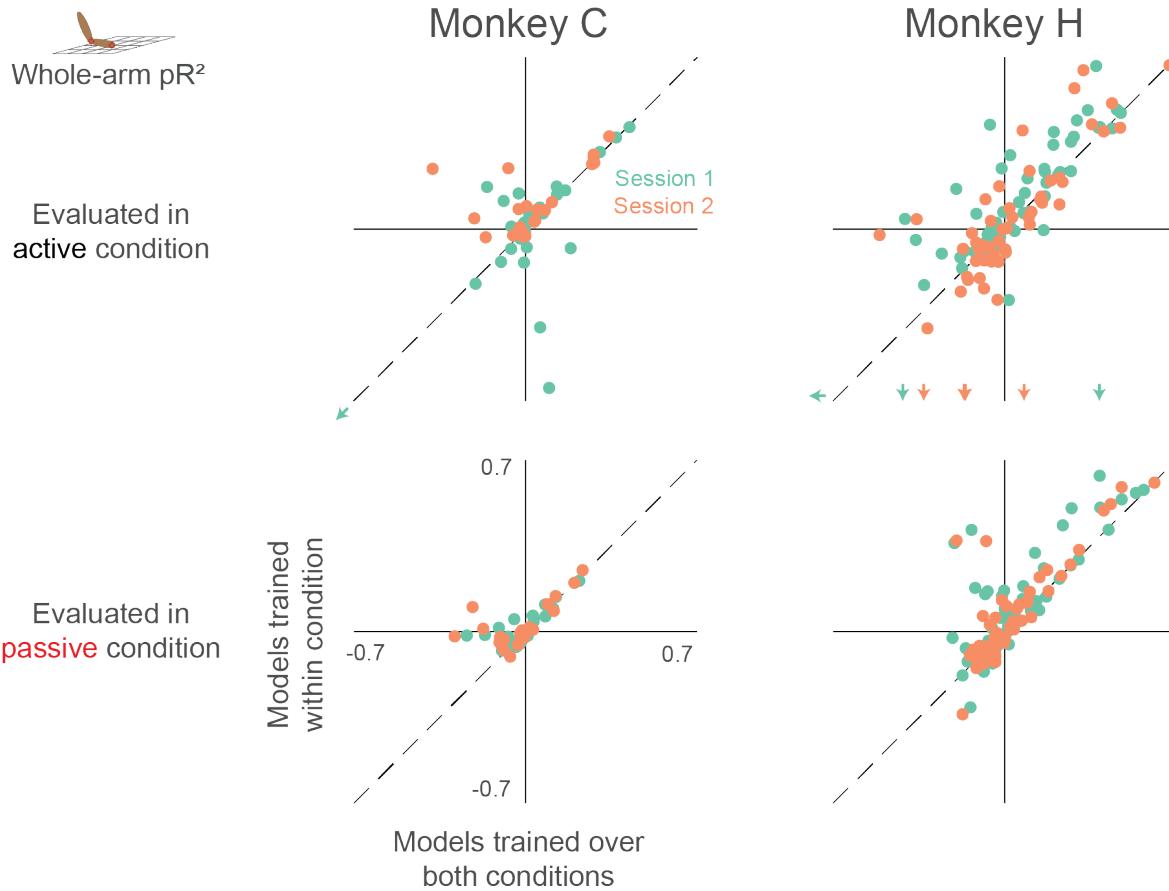
312 There is thus a clear analogy between this experiment and the two-workspace experiment—both
313 have two conditions which altered both the kinematics of the arm and the neural responses.
314 Continuing the analogy, we asked how well our two models could predict neural activity across
315 active / passive conditions. As with the two-workspace experiment, we fit both the hand-only
316 and whole-arm models to neural activity during both active and passive movements, and found
317 that the whole-arm model again tended to out-perform the hand-only model (Filled circles above
318 the dashed unity line in Figure 9). However, there were many more neurons (open circles) for
319 which the difference between models was insignificant compared to the two-workspace
320 experiment (Figure 4).



321

322 *Figure 9: Goodness-of-fit comparison analysis for active/passive experiment (same format as figure 4).
323 Each dot represents a single neuron, with color indicating the recording session. Filled circles indicate
324 neurons that are significantly far away from the dashed unity line.*

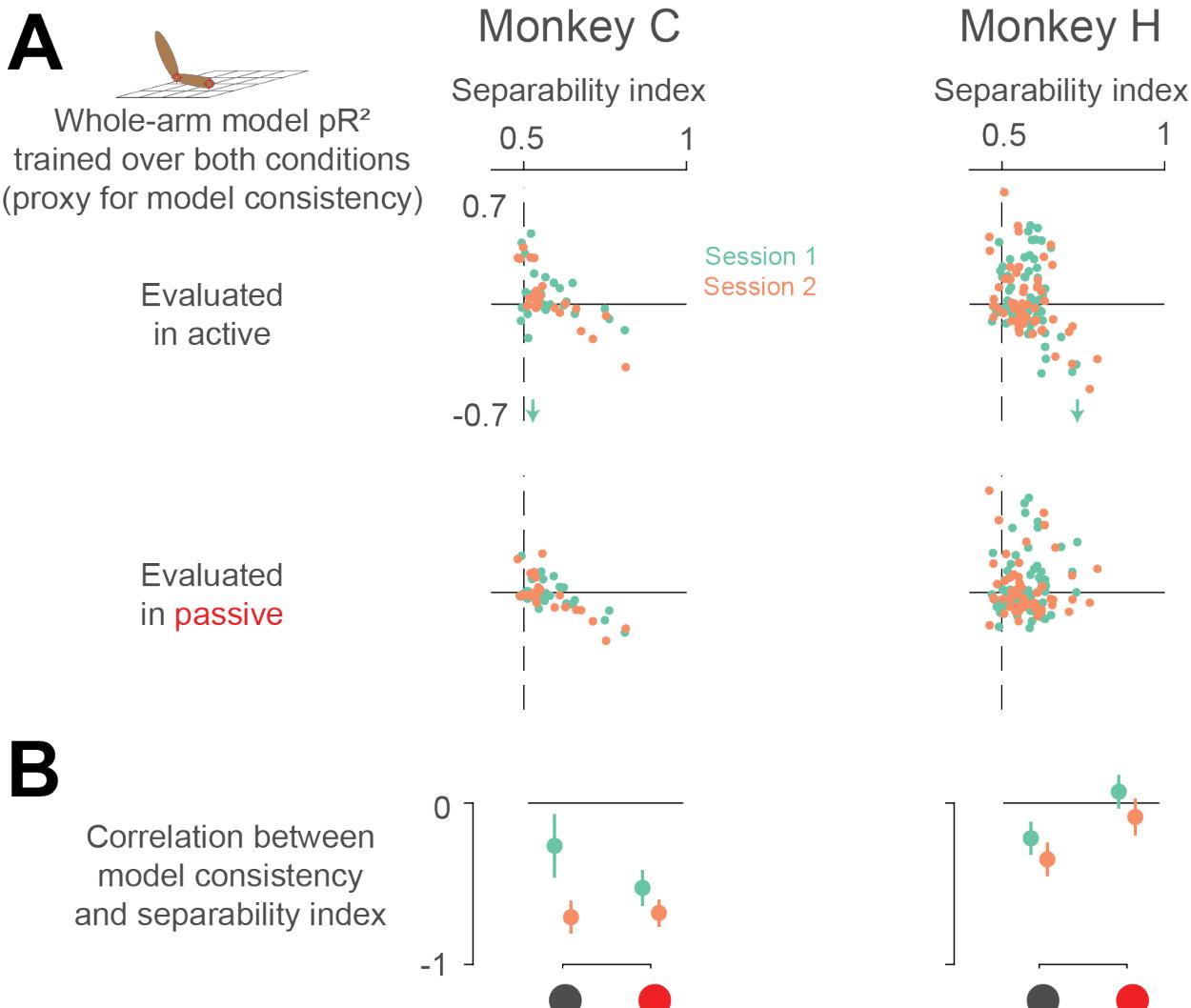
325 As in the two-workspace experiment, we compared models trained within an individual (active
326 or passive) condition, to those trained in both conditions (Figure 10). A number of neurons had
327 consistent relationships with arm kinematics, indicated by the dots with positive pR^2 values lying
328 close to the unity line. Surprisingly however, unlike our results from the two-workspace
329 experiment (see Figure 5), many neurons in the active/passive task did not have this consistent
330 relationship, indicated by the many neurons with negative pR^2 values for the model trained over
331 both conditions.



332

333 *Figure 10: Dependence of whole-arm model accuracy on active and passive training data (same format*
334 *as figure 5). Plots in the upper row contain colored arrows at the edges indicating neurons with pR²*
335 *value beyond the axis range, which we omitted for clarity.*

336 The initial question of this experiment remains, however: does the neural separability index stem
337 simply from arm kinematics? If this were true, then neurons with high separability index should
338 have a consistent relationship to arm kinematics. To test this, we compared each neuron's pR²
339 value when trained on both conditions (our proxy for model consistency) against its separability
340 index (Figure 11). Interestingly, we found the opposite result—model consistency actually
341 correlated negatively with the separability index. Essentially, this means that neurons responding
342 to active and passive movements differently are likely not drawing this distinction based on arm
343 kinematics, as those are the neurons for which we could not find a consistent whole-arm model.
344 Instead, this suggests that neurons in area 2 distinguish active and passive movements by some
345 other means, perhaps an efference copy signal from motor areas of the brain (Bell 1981; London
346 and Miller 2013; Nelson 1987).



347

348 *Figure 11: Neural separability index predicts whole-arm model inconsistency. A – Scatter plots*
349 *comparing the consistency of the whole-arm model against the separability index. Conventions are the*
350 *same as in figure 10. B – correlation between model consistency and separability index. Each dot*
351 *represents the correlation between model consistency and separability index for a given session, with*
352 *error bars representing the 95% confidence intervals.*

353 **3 DISCUSSION**

354 **3.1 SUMMARY**

355 In this study, we explored, in two separate experiments, how somatosensory area 2 represents
356 arm movements. In the first experiment, a monkey reached to targets in two separate
357 workspaces. We found that a model incorporating whole-arm kinematics explained area 2 neural
358 activity well, especially when compared to the hand-only model typically used to explain these
359 neurons' responses. Our results from the experiment thus suggest that area 2 represents the state

360 of the whole arm during reaching. In the second experiment, we tested the whole-arm model's
361 ability to explain area 2 neural activity both during reaching, and when the hand was
362 unexpectedly displaced passively. As in the first experiment, these two conditions differed both
363 kinematically and in the neural responses to movement. However, we found that while some
364 neurons maintained a consistent relationship with arm kinematics across the two conditions,
365 many others did not. Furthermore, those neurons most sensitive to movement type were also
366 those most poorly modeled across conditions. The results from this second experiment suggest
367 that for some neurons, area 2 relates to arm kinematics differently for active and passive
368 movements.

369 3.2 MODEL COMPLEXITY

370 A significant difference between the hand and whole-arm models is their number of parameters,
371 which make the whole-arm models more complex and expressible. There are two concerns with
372 testing models of differing complexity, the first dealing with model training and evaluation, and
373 the second with interpretation of the results.

374 In training and evaluating our models, we had to make sure that the complex models did not
375 overfit the data, resulting in artificially high performance on the training dataset but low
376 generalizability to new data. However, because we found through cross-validation that the more
377 complex models generalized to test data better than the simpler models, they were not
378 overfitting. Consequently, the hand-based models are clearly impoverished compared to the
379 whole-arm models.

380 The second concern is in interpreting what it means when the more complex models perform
381 better. One interpretation is that this is an obvious result; if the added degrees of freedom have
382 anything at all to do with area 2 neural activity, then the more complex models should perform
383 better. In fact, our main goal was primarily to improve our understanding of this area of S1 by
384 exploring how incorporating measurements of whole-arm kinematics could help explain its
385 function. As a result, we found that the whole-arm model not only out-performed the hand-only
386 model, but it also predicted changes in PD across the two workspaces well in its own right.
387 Furthermore, as demonstrated by the findings from our second experiment, the more complex
388 model does not necessarily lead to a satisfactory fit. Despite its increased complexity and its
389 success in the two-workspace task, the whole-arm model could not find a consistent fit for many
390 neurons over both active and passive movements. As such, the active/passive experiment serves
391 as a useful control for the two-workspace findings.

392 3.3 COORDINATE FRAME VS. INFORMATIONAL CONTENT

393 Because of their differing dimensionality, the signals from the hand-only model and those from
394 whole-arm model do not have a one-to-one relationship: there are many different arm
395 configurations that result in a given hand position. Thus, a comparison between the hand-only
396 and whole-arm models is mainly a question of information content (do area 2 neurons have
397 information about more than just the hand?). In contrast, signals predicted by the various whole-
398 arm models (see Supplementary Information) do have a one-to-one (albeit nonlinear)

399 relationship to each other. Knowledge of the hand and elbow position should completely
400 determine estimated joint angles and musculotendon lengths, indicating that these models should
401 have the same informational content. As such, a comparison between these models (as in the
402 Supplementary Information) is purely one of coordinate frame. While the interpretation for a
403 comparison of information content is straightforward, interpreting the results of a comparison
404 between coordinate frames is not. One major issue is that these comparisons only make sense
405 when using linear models to relate neural activity to behavior. Once nonlinear models are
406 considered, as in our study with artificial neural networks (Lucas et al. 2019), coordinate frames
407 with one-to-one correspondence become nearly equivalent, and much more difficult to compare
408 meaningfully.

409 Clear parallels exist between this and earlier studies seeking to find a unique representation of
410 movement in motor areas. Over the last few decades, a controversy involving the exact nature of
411 the neural representation of movement has played itself out in the literature surrounding motor
412 cortex, with some advocating a hand-based representation of motor control (Georgopoulos et al.
413 1982; Georgopoulos et al. 1986; Moran and Schwartz 1999) and others a muscle-based
414 representation (Evarts 1968; Fetz et al. 1989; Morrow et al. 2007; Oby et al. 2012). Recently, the
415 motor control field started turning away from questions of coordinate frame and towards
416 questions of neural population dynamics and information processing (Churchland et al. 2010;
417 Elsayed et al. 2016; Gallego et al. 2017; Kaufman et al. 2014; Perich et al. 2018; Russo et al.
418 2018; Sussillo et al. 2015). Part of the motivation for this pivot in viewpoint is that it became
419 increasingly clear that a “pure” coordinate frame of movement representation is unlikely to exist
420 (Fetz 1992; Kakei et al. 1999). Further, studies tended to use correlation between neural activity
421 and behavioral variables as evidence that the neurons represent movements in a particular
422 coordinate frame. However, as noted above, these correlations could often be explained by
423 multiple coordinate frames, casting doubt on the conclusiveness of the exact coordinate frame of
424 representation (Mussa-Ivaldi 1988). Consequently, in our study, we put aside the question of the
425 coordinate frame of area 2, focusing instead on what we can gain by modeling area 2 in terms of
426 whole-arm kinematics.

427 A major question this study leaves open is that of how information about reaching is processed
428 by different areas of the proprioceptive neuraxis. While we might expect a muscle spindle-like
429 representation at the level of the dorsal root ganglia (DRG) or the cuneate nucleus, removed from
430 the receptors by one and two synapses, respectively, this representation likely changes as the
431 signals propagate through thalamus and into S1. Even different areas of S1 may have different
432 representations. Area 3a, which receives input mostly from muscle afferents (Heath et al. 1976;
433 Kaas et al. 1979; Phillips et al. 1971; Yamada et al. 2016), seems more likely to retain a muscle-
434 like representation than is area 2, which integrates muscle afferent input with that from
435 cutaneous receptors (Hyvärinen and Poranen 1978; Padberg et al. 2018; Pons et al. 1985).
436 Likewise, area 5 may have an even higher-level representation, as it receives input from both
437 somatosensory (Mountcastle et al. 1975) and motor cortices (Padberg et al. 2018), and appears to
438 depend on attention (Chapman et al. 1984; Omrani et al. 2016). As it becomes increasingly
439 feasible to record from several of these areas simultaneously (Richardson et al. 2016; Suresh et
440 al. 2017; Weber et al. 2006), future experiments could examine how these areas project

441 information to each other, as has been explored in motor and premotor cortices (Churchland et
442 al. 2010; Elsayed et al. 2016; Kaufman et al. 2014; Perich et al. 2018), without modeling the
443 more complex cortical areas explicitly in terms of particular behavioral variables “encoded” by
444 single neurons.

445 3.4 POSSIBLE EVIDENCE OF EFFERENCE COPY IN AREA 2

446 Our inability to find a consistent model across conditions suggests a difference between neural
447 activity during active and passive movements that can't be captured by our whole-arm model.
448 One possible explanation for this is that area 2 may represent arm kinematics nonlinearly.
449 Because we modeled area 2 activity with a generalized linear model (GLM; see methods), we
450 implicitly discounted this possibility. The fact that the whole-arm kinematics for the two
451 conditions are highly discriminable (89% separable on average) means that the different
452 conditions correspond to different zones of kinematic space. Following the analogy of fitting a
453 line to data distributed on an exponential curve, it is possible that the neurons with inconsistent
454 linear relationships to arm kinematics may simply reflect a single nonlinear relationship, with
455 different linear approximations in the two zones. Indeed, several of these neurons had high pR^2
456 for models trained within condition (top left quadrants of Figure 10).

457 Another possible explanation for this finding is that voluntary movements may change the
458 afferent activity from the moving limb. This could be caused by altered descending gamma drive
459 to muscle spindles that changes their sensitivity (Loeb et al. 1985; Prochazka and Wand 1981;
460 Prochazka et al. 1976). Another possibility is that of an efference copy signal sent to the
461 brainstem or S1 from motor areas during active movements (Bell 1981; London and Miller 2013;
462 Nelson 1987). Many studies suggest that we use internal forward models of our bodies and
463 environment to coordinate our movements and predict their sensory consequences (Shadmehr
464 and Mussa-Ivaldi 1994; Wolpert et al. 1995). A key piece of this framework is comparing the
465 actual feedback received following movement with the feedback predicted by the internal model,
466 which generates a sensory prediction error. Recent studies suggest that S1 is important for
467 updating the internal model using a sensory prediction error (Mathis et al. 2017; Nasir et al.
468 2013). Thus, one potential avenue to study the effect of efference copy in S1 would be to
469 examine how motor areas communicate with area 2 during active and passive movements.

470 3.5 RELEVANCE FOR BCI

471 One motivation for this work is its potential to augment brain-computer interfaces (BCI) for
472 restoring movement to persons with spinal cord injury or limb amputation. As BCI for motor
473 control gets more advanced (Collinger et al. 2013; Ethier et al. 2012; Kao et al. 2015; Young et
474 al. 2018), it will become more necessary to develop a method to provide feedback about
475 movements to the brain, potentially using intracortical microstimulation (ICMS) to activate
476 somatosensory areas. While ICMS in S1 has seen some success in providing feedback about
477 touch (Flesher et al. 2016; Romo et al. 1998; Salas et al. 2018; Tabot et al. 2013), the path
478 towards providing proprioceptive feedback remains relatively unexplored. At least one study did
479 use electrical stimulation in S1 for feedback during movement, using the stimulation to specify

480 target direction with respect to the evolving hand position (Dadarlat et al. 2015). In that study,
481 monkeys used the ICMS to reach to targets, even in the absence of visual feedback. However,
482 target-location information is very different from the information normally encoded by S1, and
483 the monkeys required several months to learn to use it. To our knowledge, no study has yet
484 shown a way to use ICMS to provide more biomimetic proprioceptive feedback during reaching.
485 Previously, our lab attempted to address this gap by stimulating a small number of electrodes in
486 area 2 based on neural activity recorded from them during normal reaching movements. In that
487 experiment, the monkey reported the direction of a mechanical bump to his arm that occurred
488 simultaneously with the ICMS. The ICMS biased one monkey's reports of the mechanical bump
489 direction toward the PDs of the stimulated electrodes. Key to this finding was the fact that any
490 bias in reporting actually decreased the reward rate, suggesting that the ICMS was
491 indistinguishable from the perception of the bump itself (Tomlinson and Miller 2016).
492 Unfortunately, the result could not be replicated in other monkeys; while the ICMS often biased
493 their reports, the direction of the bias could not be explained by the PDs of the stimulated
494 electrodes. One potential reason may be that the stimulation paradigm in those experiments was
495 derived from the classic, hand-based model and the assumption that area 2 represents active and
496 passive movements similarly. As this paper has shown, both of these assumptions have important
497 caveats. It is possible that a stimulation paradigm based on a whole-arm model may be more
498 successful, due to its greater accuracy at predicting neural activity (Figure 7). It is also possible
499 that the stimulus model would need to include information about forces in addition to
500 kinematics. Regardless of the exact model, prospects for stimulating S1 to create natural
501 proprioceptive sensations would likely improve given a more accurate generative model of S1
502 activity.

503 In addition to developing better models for S1 activity, it will be important to consider the
504 implications of the difference between sensation for perception versus action. These two broad
505 purposes for sensation are thought to involve distinct pathways in both vision and touch
506 (Dijkerman and De Haan 2007; Mishkin and Ungerleider 1982; Sedda and Scarpina 2012). It is
507 quite plausible that this distinction exists for proprioception as well (Dijkerman and De Haan
508 2007). However, studies of the effects of ICMS in S1 tend to use perceptual reporting to test the
509 effect of stimulation (Salas et al. 2018; Tomlinson and Miller 2016; Zaaimi et al. 2013), thereby
510 not directly addressing how effectively ICMS can be used as feedback for action. Even in the
511 study conducted by Dadarlat et al., movements guided by ICMS were slower and contained more
512 submovements than those guided by even a noisy visual signal, suggesting that monkeys used the
513 ICMS as a learned sensory substitute, rather than as a biomimetic replacement for
514 proprioception. As such, that study was also likely a cognitive one, engaging the perceptual
515 stream rather than the action stream of proprioception (see (Deroy and Auvray 2012; Elli et al.
516 2014) for discussion of the limits of sensory substitution). As we better characterize how S1
517 represents movements, we hope to develop a stimulation paradigm in which we can engage both
518 streams, to enable users of a BCI both to perceive their limb, and to respond rapidly to
519 movement perturbations.

520 **4 CONCLUSION**

521 Our goal in conducting this study was to improve our understanding of how area 2 neural activity
522 represents arm movements. We began by asking what we would learn about area 2 when we
523 tracked the movement of the whole arm, rather than just the hand. The results of our first
524 experiment showed that a model built on these whole-arm kinematics was highly predictive of
525 area 2 neural activity, suggesting that it indeed represents the kinematic state of the whole arm
526 during reaching. In our second experiment, we sought to extend these findings to similar
527 movements when the limb is passively displaced. There, we found that while some neurons
528 consistently represented arm kinematics, others did not, suggesting that the area may process
529 active and passive movements differently, possibly with the addition of efference copy inputs.

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538 **6 METHODS AND MATERIALS**

Key Resources Table				
Reagent type (species) or resource	Designation	Source or reference	Identifiers	Additional information
software, algorithm	MATLAB	MathWorks	RRID:SCR_001622	All code developed for this paper available on GitHub (See relevant sections of Methods)

539 All surgical and experimental procedures were fully consistent with the guide for the care and
540 use of laboratory animals and approved by the institutional animal care and use committee of
541 Northwestern University under protocol #IS00000367.

542 **6.1 BEHAVIOR**

543 We recorded data from a monkey while it used a manipulandum to reach for targets presented on
544 a screen within a 20 cm x 20 cm workspace. After each successful reaching trial, the monkey
545 received a pulse of juice or water as a reward. We recorded the position of the handle using
546 encoders on the manipulandum joints. We also recorded the interaction forces between the
547 monkey's hand and the handle using a six-axis load cell mounted underneath the handle.

548 For the two-workspace experiment, we partitioned the full workspace into four 10cm x 10cm
549 quadrants. Of these four quadrants, we chose the far ipsilateral one and the near contralateral one
550 in which to compare neural representations of movement. Before each trial, we chose one of the
551 two workspaces randomly, within which the monkey reached to a short sequence of targets
552 randomly positioned in the workspace. For this experiment, we only analyzed the portion of data
553 from the end of the center-hold period to the end of the trial.

554 For the active vs. passive experiment, we had the monkey perform a classic center-out (CO)
555 reaching task, as described in (London and Miller 2013). Briefly, the monkey held in a target at
556 the center of the full workspace for a random amount of time, after which one of four outer
557 targets was presented. The trial ended in success once the monkey reached to the outer target. On
558 50% of the trials (deemed “passive” trials), during the center hold period, we used motors on the
559 manipulandum to deliver a 2 N perturbation to the monkey's hand in one of the four target
560 directions. After the bump, the monkey returned to the center target, after which the trial
561 proceeded like an active trial. From only the successful passive and active trials, we analyzed the
562 first 120 ms after movement onset. Movement onset was determined by looking for the peak in
563 handle acceleration either after the motor pulse (in the passive condition) or after 200 ms post-go
564 cue (in the active condition) and sweeping backwards in time until the acceleration was less than
565 10% of the peak.

566 **6.2 MOTION TRACKING**

567 Before each reaching experiment, we painted 10 markers on the outside of the monkey's arm,
568 marking bony landmarks and a few points in between, a la (Chan and Moran 2006). Using a
569 custom motion tracking system built from a Microsoft Kinect, we recorded the 3D locations of
570 these markers with respect to the camera, synced in time to the other behavioral recordings. We
571 then aligned the Kinect-measured marker locations to the lab frame by aligning location of the
572 Kinect hand marker to the location of the handle in the manipulandum coordinate frame. Code
573 for motion tracking can be found at <https://github.com/limblab/KinectTracking.git>.

574 **6.3 NEURAL RECORDINGS**

575 We implanted 100-electrode arrays (Blackrock Microsystems) into the arm representation of area
576 2 of S1 in these monkeys. For more details on surgical techniques, see (Weber et al. 2011). In
577 surgery, we roughly mapped the postcentral gyrus by recording from the cortical surface while
578 manipulating the arm and hand to localize their representations. To record neural data for our
579 experiments, we used a Cerebus recording system (Blackrock). This recording system sampled
580 signals from each of the 96 electrodes at 30 kHz. To conserve data storage space, the system

581 detected spikes online using a threshold set at -5x signal RMS, and only wrote to disk a time
582 stamp and the 1.6 ms snippet of signal surrounding the threshold crossing. After data collection,
583 we used Plexon Offline Sorter to manually sort these snippets into putative single units, using
584 features like waveform shape and inter-spike interval.

585 **6.4 SENSORY MAPPINGS**

586 In addition to recording sessions, we also occasionally performed sensory mapping sessions to
587 identify the neural receptive fields. For each electrode we tested, we routed the corresponding
588 recording channel to a speaker and listened to multi-unit neural activity while manipulating the
589 monkey's arm. We noted both the modality (deep or cutaneous) and the location of the receptive
590 field (torso, shoulder, humerus, elbow, forearm, wrist, hand, or arm in general). We classified an
591 electrode as cutaneous if we found an area of the skin, which when brushed or stretched, resulted
592 in an increase in multi-unit activity. We classified an electrode as deep if we found activity to be
593 responsive to joint movements and/or muscle palpation but could not find a cutaneous field. As
594 neurons on the same electrode tend to have similar properties (Weber et al. 2011), we usually did
595 not separate neurons on individual electrodes during mapping. However, when we did, we
596 usually found them to have similar receptive field modality and location.

597 In Monkeys C and H, we found a gradient of receptive field location across the array,
598 corresponding to a somatotopy from proximal to distal. To quantify this gradient, we assigned
599 each receptive field location a score from 1 to 7 (with 1 being the torso and 7 being the hand),
600 and we fit a simple linear model relating this location on the limb to the x and y coordinates of
601 electrodes on the array. We show the calculated gradients for Monkeys C and H as black arrows
602 in Figure 1 (both significant linear fits with $p < 0.05$). Monkey L's array had too few neurons to
603 calculate a significant linear model.

604 **6.5 NEURAL ANALYSIS**

605 Code for the following neural analyses can be found at [https://github.com/raeedcho/s1-
606 kinematics.git](https://github.com/raeedcho/s1-kinematics.git).

607 **6.5.1 Preferred directions**

608 We used a simple bootstrapping procedure to calculate PDs for each neuron. On each bootstrap
609 iteration, we randomly drew timepoints from the reaching data, making sure that the distribution
610 of movement directions was uniform to mitigate the effects of any potential bias. Then, as in
611 (Georgopoulos et al. 1982), we fit a cosine tuning function to the neural activity with respect to
612 the movement direction, using equations 1a-b.

613
$$f_i(\tau) = b_0 + b_1 * \sin(\theta_m(\tau)) + b_2 * \cos(\theta_m(\tau)) \quad (1a)$$

614
$$= b_0 + r_i * \cos(\theta_m(\tau) - PD_i) \quad (1b)$$

615 where

616
$$PD_i = \text{atan2}(b_1, b_2) \text{ and } r_i = \sqrt{b_1^2 + b_2^2}$$

617 Here, $f_i(\tau)$ is the average firing rate of neuron i for a given time point τ , and $\theta_m(\tau)$ is the
618 corresponding movement direction, which for the active/passive task was the target or bump
619 direction, and for the two-workspace experiment was the average movement direction over a
620 time bin. We took the circular mean of PD_i and mean of r_i over all bootstrap iterations to
621 determine the preferred direction and the modulation depth respectively, for each neuron.

622 As the PD analysis is meaningless for neurons that don't have a preferred direction of movement,
623 we only analyzed the PDs of neurons that were significantly tuned. We assessed tuning through a
624 separate bootstrapping procedure, described in (Dekleva et al. 2018). Briefly, we randomly
625 sampled the timepoints from reaching data, again ensuring a uniform distribution of movement
626 directions, but this time also randomly shuffled the corresponding neural activity. We calculated
627 the r_i for this shuffled data on each bootstrap iteration, thereby creating a null distribution of
628 modulation depths. We considered a neuron to be tuned if the true r_i was greater than the 95th
629 percentile of the null distribution.

630 **6.5.2 Models of neural activity**

631 For the two-workspace analyses, both behavioral variables and neural firing rate were averaged
632 over 50 ms bins. For the active/passive analyses, we averaged behavioral variables and neural
633 firing rates over the 120 ms period following movement onset in each trial. We modeled neural
634 activity with respect to the behavior using Poisson generalized linear models (outline in
635 (Truccolo et al. 2005)) shown in equation 2a, below.

636
$$f \sim \text{Poisson}(\lambda), \lambda = \exp(X\beta) \quad (2a)$$

637 In this equation, f is a T (number of time points) x N (number of neurons) matrix of average
638 firing rates, X is a $T \times P$ (number of behavioral covariates, explained below) matrix of behavioral
639 correlates, and β is a $P \times N$ matrix of model parameters. We fit these GLMs by finding
640 maximum likelihood estimation of the parameters, $\hat{\beta}$. With these fitted models, we predicted
641 firing rates (\hat{f}) on data not used for training, shown in equation 2b, below.

642
$$\hat{f} = \exp(X\hat{\beta}) \quad (2b)$$

643 We tested six firing rate encoding models, detailed below. Of these six models, the first two
644 (hand-only and whole-arm) were the ones shown in the main text, with results from the other
645 models detailed in Supplementary Information. Note that each model also includes an offset
646 term, increasing the number of parameters, P , by one.

- 647 • Hand-only: behavioral covariates were position and velocity of the hand, estimated by using
648 the location of one of the hand markers, in three-dimensional Cartesian space, with origin at
649 the shoulder ($P = 7$).
- 650 • Whole-arm: behavior covariates were position and velocity of both the hand and elbow
651 markers in three-dimensional Cartesian space, with origin at the shoulder. This is the
652 simplest extension of the extrinsic model that incorporates information about the
653 configuration of the whole arm ($P = 13$)

654 • Hand kinematics+force: behavioral covariates were position and velocity of the hand, as
 655 well as forces and torques on the manipulandum handle, in three-dimensional Cartesian
 656 space ($P = 13$).
 657 • Egocentric: behavior covariates were position and velocity of the hand marker in spherical
 658 coordinates (θ , ϕ , and ρ), with origin at the shoulder ($P = 7$).
 659 • Joint kinematics: behavioral covariates were the 7 joint angles (shoulder
 660 flexion/abduction/rotation, elbow flexion, wrist flexion/deviation/pronation) and
 661 corresponding joint angular velocities ($P = 15$).
 662 • Muscle kinematics: behavioral covariates were derived from the length of the 39 modeled
 663 muscles (Chan and Moran 2006) and their time derivatives. However, because this would
 664 result in almost 78 (highly correlated) covariates, we used PCA to extract 5-dimensional
 665 orthogonal basis sets for both the lengths and their derivatives. On average, five
 666 components explained 99 and 96 percent of the total variance of lengths and length
 667 derivatives, respectively. Behavioral covariates of this model were the projections of the
 668 muscle variables into these spaces during behavior ($P = 11$).

669 We used repeated 5-fold cross-validation to evaluate our models of neural activity, given that the
 670 models had different numbers of parameters, P . On each repeat, we randomly split trials into five
 671 groups (folds) and trained the models on four of them. We used these trained models to predict
 672 neural firing rates (\hat{f}_i) in the fifth fold. We then compared the predicted firing rates from each
 673 model to the actual firing rates in that test fold, using analyses described in the following
 674 sections. This process (including random splitting) was repeated 20 times, resulting in $n=100$
 675 sample size for each analysis result. Thus, if a more expressive model with more parameters
 676 performs better than a simpler model, it would suggest that the extra parameters do provide
 677 relevant information about the neural activity not accounted for by the simpler models.

678 **6.5.3 Statistical tests and confidence intervals**

679 To perform statistical tests on the output of repeated 5-fold cross-validation, we used a corrected
 680 resampled t-test, outlined in (Ernst 2017) and (Nadeau and Bengio 2003). Here, sample mean
 681 and variance are calculated as in a normal t-test, but a correction factor needs to be applied to the
 682 standard error, depending on the nature of the cross-validation. Equation 3a-c shows a general
 683 case of this correction for R repeats of K -fold cross-validation of some analysis result d_{kr} .

684
$$\hat{\mu}_d = \frac{1}{K \times R} \sum_{k=1}^K \sum_{r=1}^R d_{kr} \quad (3a)$$

685
$$\hat{\sigma}_d^2 = \frac{1}{(K \times R) - 1} \sum_{k=1}^K \sum_{r=1}^R (d_{kr} - \hat{\mu}_d)^2 \quad (3b)$$

686
$$t_{stat} = \frac{\hat{\mu}_d}{\sqrt{\left(\frac{1}{K \times R} + \frac{1/K}{1 - 1/K}\right) \hat{\sigma}_d^2}} \quad (3c)$$

687 We then compare the t-statistic here (t_{stat}) to a t-distribution with $K \times R - 1$ degrees of
688 freedom. Note that the correction applied is an extra term (i.e., $\frac{1/K}{1-1/K}$) under the square root,
689 compared to the typical standard error calculation. Note that we performed all statistical tests
690 within individual sessions or for individual neurons, never across sessions or monkeys.

691 **6.5.4 Bonferroni corrections**

692 At the beginning of this project, we set out to compare three of these six models: hand-only,
693 egocentric, and muscle kinematics. In making pairwise comparisons between these models, we
694 used $\alpha = 0.05$ and a Bonferroni correction of 3, for the three original comparisons. In this
695 analysis, we found that the muscle model performed best. As we developed this project,
696 however, we tried the three other models to see if they could outperform the muscle kinematics
697 model, eventually finding that the whole-arm model, built on Cartesian kinematics of the hand
698 and elbow outperformed it. As this appeared to be primarily due to modeling and measurement
699 error in the muscle model (see Supplementary Information), we decided to focus on the hand-
700 only and whole-arm model. Despite only making one pairwise comparison in the main text, we
701 chose to use a Bonferroni correction factor of 6: three for the original three pairwise comparisons
702 and one more for each additional model we tested, which were compared against the best model
703 at the time, and could have changed the end result of this project.

704 **6.5.5 Goodness-of-fit (pseudo- R^2)**

705 We evaluated goodness-of-fit of these models for each neuron by using a pseudo- R^2 (pR^2)
706 metric. We used a formulation of pseudo- R^2 based on a comparison between the deviance of the
707 full model and the deviance of a “null” model, i.e., a model that only predicts the overall mean
708 firing rate (Cameron and Windmeijer 1997; 1996; Heinzl and Mittlböck 2003; Perich et al.
709 2018).

710
$$pR^2 = 1 - \frac{D(f_i; \hat{f}_i)}{D(f_i; \bar{f}_i)} \quad (4a)$$

711
$$= 1 - \frac{\log L(f_i) - \log L(\hat{f}_i)}{\log L(f_i) - \log L(\bar{f}_i)} \quad (4b)$$

712 When computing the likelihood of a Poisson statistic, this is:

713
$$= 1 - \frac{\sum_{\tau=1}^T f_i(\tau) \log\left(\frac{f_i(\tau)}{\hat{f}_i(\tau)}\right) - (f_i(\tau) - \hat{f}_i(\tau))}{\sum_{\tau=1}^T f_i(\tau) \log\left(\frac{f_i(\tau)}{\bar{f}_i}\right) - (f_i(\tau) - \bar{f}_i)} \quad (4c)$$

714 This pR^2 metric ranges from $-\infty$ to 1, with a value of 1 corresponding to a perfectly fit model
715 and a value of 0 corresponding to a model that only fits as well as the “null” model. In contrast
716 with the general intuition for regular R^2 , a pR^2 of ~ 0.2 is considered a “good” fit (McFadden
717 1977).

718 6.5.6 Tuning curves

719 We binned the trajectory into 16 bins, each 22.5 degrees wide, based on the mean direction
720 across 50 ms of hand motion. For each directional bin, we calculated the sample mean and 95%
721 confidence interval of the mean. In figures, we plotted this mean firing rate against the center-
722 point of the bin.

723 6.5.7 Preferred direction shift

724 We calculated PDs for each neuron in each workspace and found the predicted change in PD
725 from the contralateral workspace to the ipsilateral workspace, given each model. We compared
726 these changes to those observed for each neuron. The values of these PD shifts are shown in
727 Figure 7 for all neurons tuned to movements in both workspaces, averaged over all 100 test
728 folds.

729 We computed a variance-accounted-for (VAF) metric, here called the “circular VAF” (cVAF)
730 for each neuron (i) in each fold as:

731 $cVAF_i = \cos(\Delta\theta_{PD,i} - \Delta\hat{\theta}_{PD,i})$ (5)

732 As the cVAF metric is essentially the inner product of unit vectors with direction $\Delta\theta_{PD,i}$ and
733 $\Delta\hat{\theta}_{PD,i}$, it accounts for the circular domain of the PD shifts. Like regular VAF, the cVAF has a
734 maximum value of 1 when $\Delta\theta_{PD,i}$ and $\Delta\hat{\theta}_{PD,i}$ are the same, and decreases in proportion to the
735 squared difference between $\Delta\theta_{PD,i}$ and $\Delta\hat{\theta}_{PD,i}$. We took the average cVAF over all neurons as
736 the cVAF for the fold. In total, given the 20 repeats of 5-fold cross-validation, this gave us 100-
737 samples of the cVAF for each model in a given session.

738 6.5.8 Separability index

739 In the active/passive experiment, we calculated the separability index for each neuron by fitting a
740 linear discriminant analysis (LDA) classifier, predicting trial type (active or passive) from the
741 neuron’s average activity in the 120 ms after movement onset. As with the other neural analyses,
742 we fit and evaluated each LDA classifier using our repeated 5-fold cross-validation scheme,
743 calling the average test set classification percentage the neuron’s separability index.

744 Our procedure for calculating the separability of the whole-arm kinematics was similar, simply
745 substituting the whole-arm kinematics for the neural activity when training and testing the LDA
746 classifier.

747 **7 SUPPLEMENTARY INFORMATION**

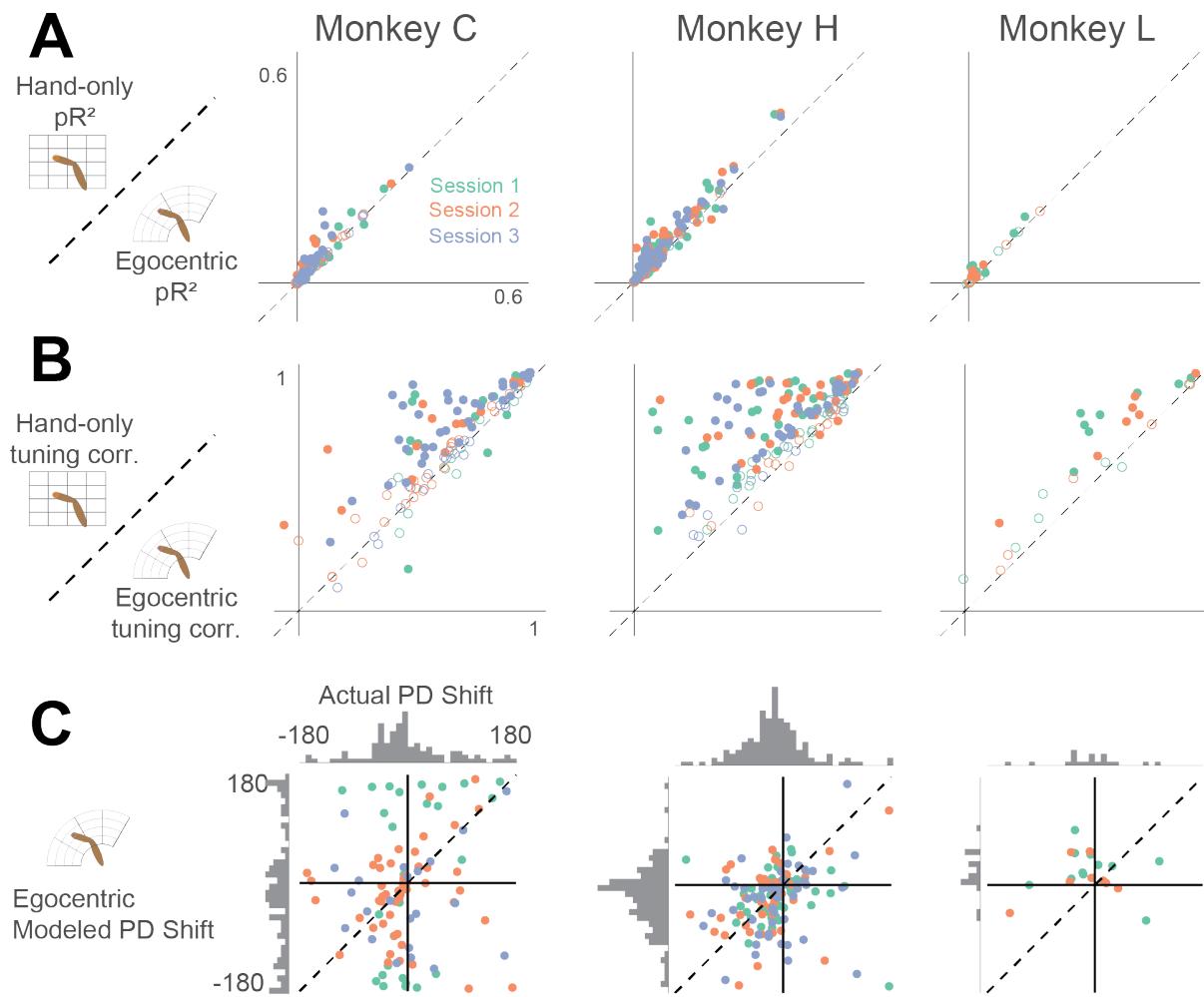
748 **7.1 WITHIN CLASS MODEL COMPARISONS**

749 Over the course of this project, we analyzed several different models of area 2 activity. We
750 categorized these models into two classes based on whether they contained information about the
751 hand or the arm in different coordinate frames. Of these models, we picked the hand-only and
752 whole-arm models to represent the two model classes in the main paper, as we found that the

753 other within-class models offered little additional insight into area 2 activity. For completeness,
754 however, this section expands on the comparisons between within-class models.

755 **7.1.1 Hand model comparison**

756 Two of our models used the kinematics of hand movement as behavioral covariates for area 2
757 neural activity: the hand-only model in the main paper and the egocentric model, which
758 represents hand kinematics in a spherical coordinate frame with origin at the shoulder. While the
759 egocentric model, or a model like it, has been proposed as a possible coordinate frame for
760 representation of the limb (Bosco et al. 1996; Caminiti et al. 1990), we found that it performed
761 rather poorly at explaining neural activity in area 2 from the two-workspace task. Figure 7 –
762 figure supplement 1A and B show comparisons between the hand-only model and the egocentric
763 model in terms of pR^2 and tuning curve correlation, as in the main paper. These comparisons
764 show that the hand-only model tended to out-perform the egocentric model. Further, the
765 egocentric model predicted large shifts in PD between the two workspaces (Figure 7 – figure
766 supplement 1C) that did not match up at all to the actual PD shifts.



767

768 *Figure 7 – figure supplement 1: Comparison between hand-only model and egocentric model. A – pR^2*
769 *comparison, as in Figure 4. B – tuning curve correlation comparison, as in Figure 6. C – Modeled PD*
770 *shift compared to actual PD shift for egocentric model, as in Figure 7A.*

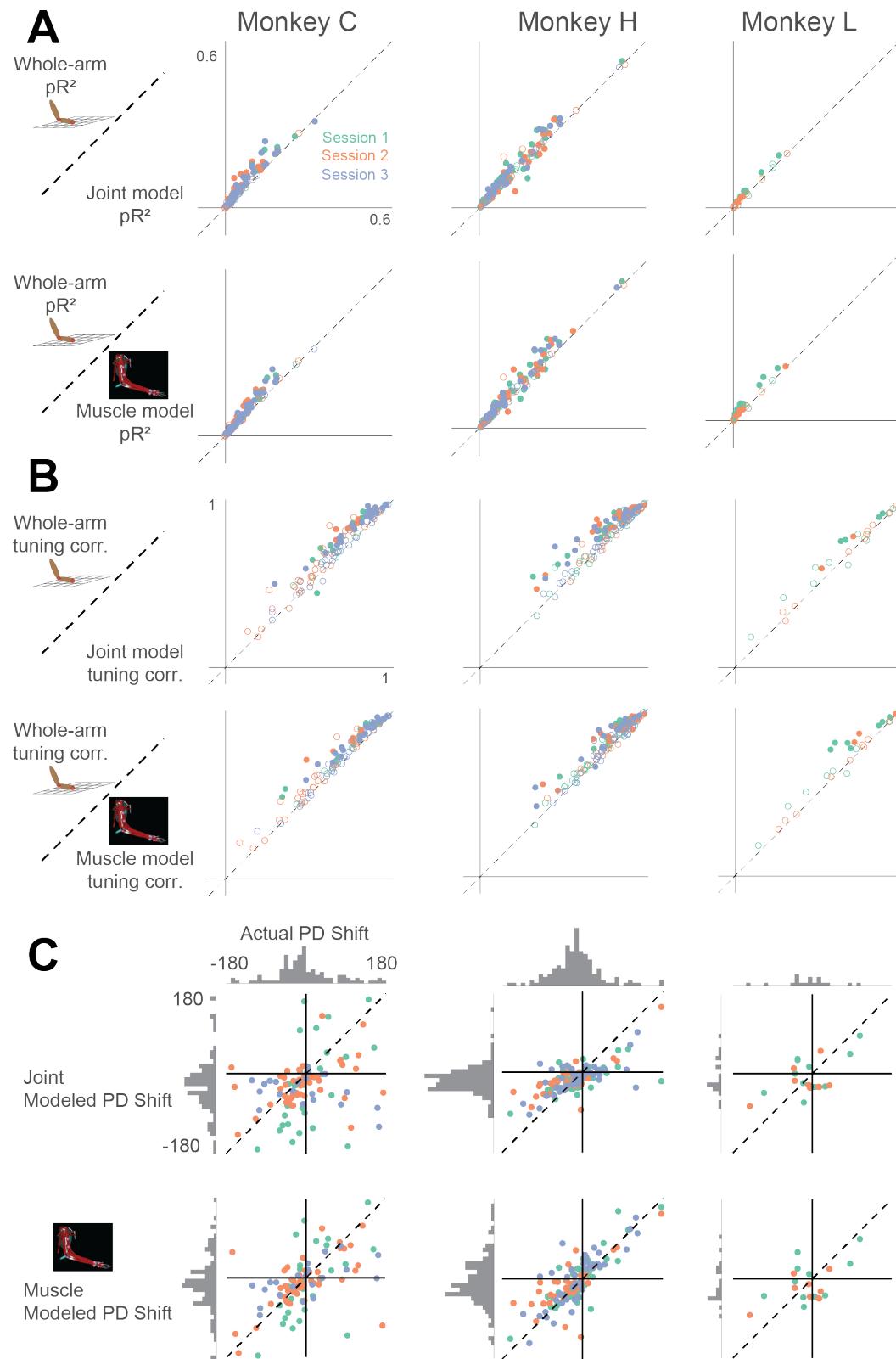
771 7.1.2 Arm model comparison

772 In addition to the whole-arm model detailed in the main paper, we tested two models of area 2
773 activity based on biomechanics: one based on joint kinematics and the other based on
774 musculotendon lengths. To find these behavioral covariates, we registered these marker locations
775 to a monkey arm musculoskeletal model in OpenSim (SimTK), based on a model of the macaque
776 arm published by (Chan and Moran 2006), and which can be found at
777 <https://github.com/limblab/monkeyArmModel.git>. After scaling the limb segments of the model to
778 match those of each monkey, we used the inverse kinematics analysis tool provided by OpenSim
779 to estimate the joint angles (and corresponding muscle lengths) required to match the model's
780 virtual marker positions to the positions of the actual recorded markers. Previously, Chan and
781 Moran used this model to analyze the joint and muscle kinematics as a monkey performs a center

782 out task (Chan and Moran 2006). Here, we use the musculoskeletal model to predict neural
783 activity.

784 Figure 7 – figure supplement 2A and B show comparisons of pR^2 and tuning curve correlation
785 between the whole-arm model detailed in the paper and these two biomechanical models. We
786 found that the three models provided similar predictions, but surprisingly, the whole-arm model
787 generally outperformed the biomechanical models. Figure 7 – figure supplement 2C shows the
788 predicted PD shifts from these models, as in Figure 7A. We found that neither biomechanical
789 model could predict PD shifts as well as the whole-arm model, though the muscle model in
790 particular appeared to perform well.

791 As a control for errors introduced into the muscle model by processing marker data with
792 OpenSim, we performed the cVAF analysis on a whole-arm model where hand and elbow
793 kinematics were derived from joint angles of the musculoskeletal model, rather than directly
794 from the marker locations captured by the motion tracking system. We re-ran the model
795 prediction analysis for only the muscle model, marker-derived whole-arm model, and OpenSim-
796 based whole-arm model. Unsurprisingly, we found average cVAFs similar to those from the
797 main analysis for the marker-derived whole-arm model (0.75). However, the cVAF for the
798 OpenSim-based whole-arm model (0.67) dropped to that for the muscle model (0.67). This
799 suggests that the difference in predictive capability between the muscle and whole-arm models
800 stems at least in part from errors introduced in OpenSim modeling, rather than from the whole-
801 arm model necessarily being the better model for area 2 neural activity.



802

803 *Figure 7 – figure supplement 2: Comparison between whole-arm model and biomechanical models (joint*
804 *kinematics and musculotendon length). Same arrangement as in Figure 7 – figure supplement 1.*

805 7.1.3 Discussion of arm model comparisons

806 As proprioceptive signals originate in the muscles, arising from muscle spindles and Golgi
807 tendon organs, we expected to find that the muscle model would outperform the other models.
808 However, there are several potential reasons why this was not so. The most important ones can
809 be divided into two categories loosely tied to 1) errors in estimating the musculotendon lengths,
810 through motion tracking and musculoskeletal modeling, and 2) the fidelity of the muscle model
811 to the actual signals sent by the proprioceptors.

812 In the first category, the main issue is that of error propagation. The extra stages of analysis
813 required to compute musculotendon lengths (registering markers to a musculoskeletal model,
814 performing inverse kinematics to find joint angles, and using modeled moment arms to estimate
815 musculotendon lengths) introduce errors not present when simply using the positions of markers
816 on the arm. As a control, we ran the whole-arm model through two of these extra steps by
817 computing the hand and elbow positions from the joint angles of the scaled model, estimated
818 from inverse kinematics. The results of this analysis showed that the performance of the whole-
819 arm model with added noise dropped to that of the muscle model, indicating that there are, in
820 fact, errors introduced in even this portion of the processing chain.

821 The other potential source of error in this processing chain stems from the modeled moment
822 arms, which might not accurately reflect those of the actual muscles. In developing their
823 musculoskeletal model, Chan and Moran collected muscle origin and insertion point
824 measurements from both cadaveric studies and existing literature (Chan and Moran 2006).
825 However, due to the complexity of some joints, along with ambiguity of how the muscle wraps
826 around bones and other surfaces, determining moment arms purely by bone and muscle geometry
827 is a difficult problem (An et al. 1984). Because moment arms are irrelevant for determining hand
828 and elbow kinematics, we could not subject the whole-arm model to the error introduced by this
829 step.

830 In addition to the questions of error propagation and musculoskeletal model accuracy is the
831 question of whether our muscle model was truly representative of the signals sensed by the
832 proprioceptors. The central complication is that spindles sense the state of the intrafusal fibers in
833 which they reside, and have a complex, nonlinear relation to the musculotendon length that we
834 used in our muscle model. Factors like load-dependent fiber pennation angle (Azizi et al. 2008),
835 or tendon elasticity (Rack and Westbury 1984) can decouple muscle fiber length from
836 musculotendon length. Additionally, intrafusal fibers receive motor drive from gamma motor
837 neurons, which continuously alters muscle spindle sensitivity (Loeb et al. 1985; Prochazka and
838 Wand 1981; Prochazka et al. 1976) and spindle activity also depends on the history of strain on
839 the fibers (Haftel et al. 2004; Proske and Stuart 1985). Altogether, this means that while the
840 musculotendon lengths we computed provide a reasonably good approximation of what the arm
841 is doing, they may not be a good representation of the spindle responses themselves. Spindle
842 activity might be more accurately modeled when given enough information about the
843 musculotendon physiology. However, to model the effects of gamma drive, we would either
844 have to record directly from gamma motor neurons or make assumptions of how gamma drive
845 changes over the course of reaching. In developing models of neural activity, one must carefully

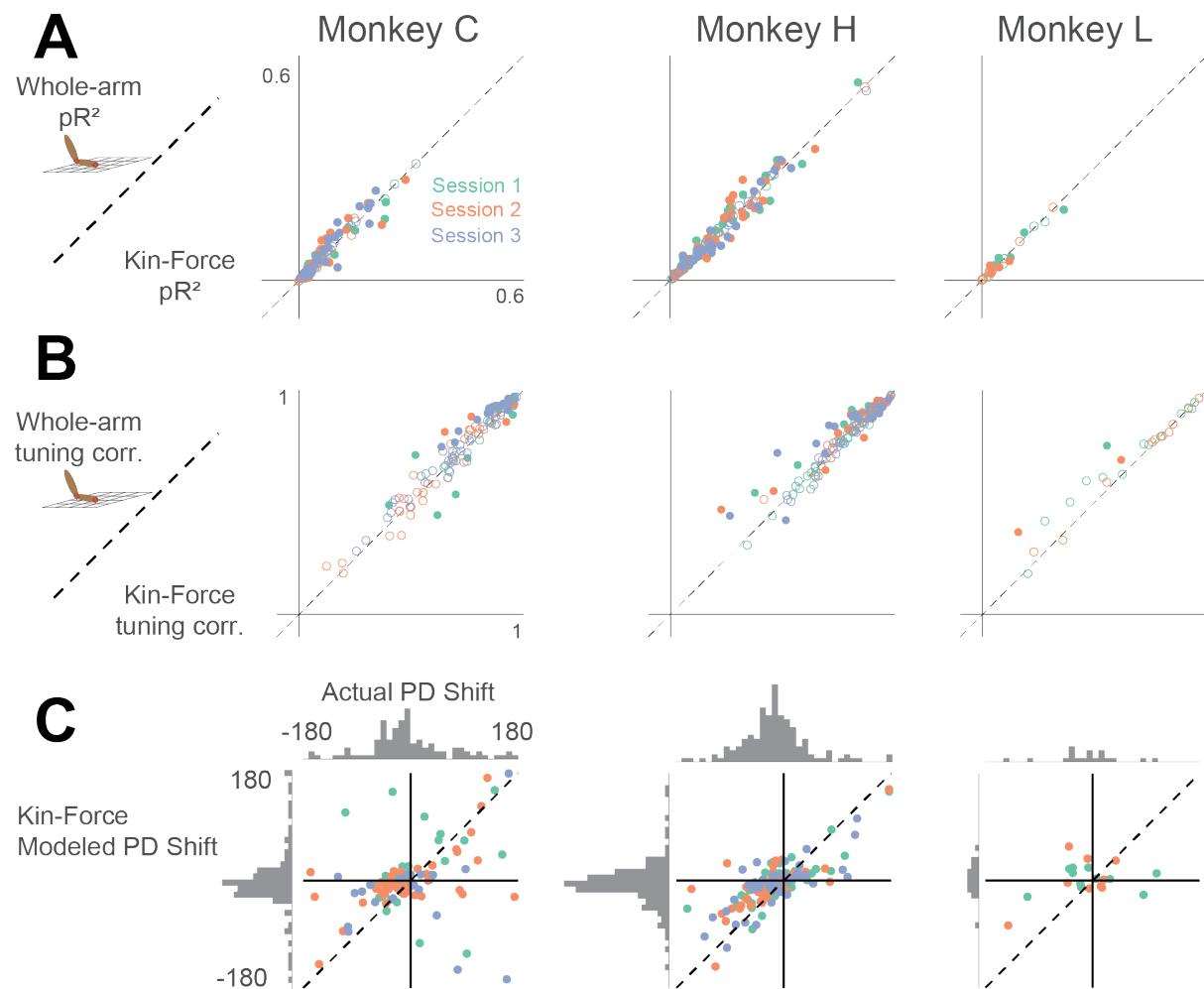
846 consider the tradeoff between increased model complexity and the extra error introduced by
847 propagating through the additional requisite measurement and analysis steps. Given our data
848 obtained by measuring the kinematics of the arm with motion tracking, it seems that the
849 coordinate frame with which to best explain area 2 neural activity is simply the one with the
850 most information about the arm kinematics and the fewest steps in processing. However, this
851 does not rule out the idea that area 2 more nearly represents a different whole-arm model that
852 may be less abstracted from physiology, like musculotendon length or muscle spindle activity.

853 Still, this model comparison shows that even after proprioceptive signals reach area 2, neural
854 activity can still be predicted well by a convergence of muscle-like signals, even though the
855 signals have been processed by several sensory areas along the way. One potential explanation
856 for this is that at each stage of processing, neurons simply spatially integrate information from
857 many neurons of the previous stage, progressively creating more complex response properties.
858 This idea of hierarchical processing was first used to explain how features like edge detection
859 and orientation tuning might develop within the visual system from spatial integration of the
860 simpler photoreceptor responses (Felleman and Van Essen 1991; Hubel and Wiesel 1959; 1962).
861 This inspired the design of deep convolutional artificial neural networks, now the state of the art
862 in machine learning for image classification (Krizhevsky et al. 2012). Unlike previous image
863 recognition methods, these feedforward neural networks are not designed to extract specific,
864 human-defined features of images. Instead, intermediate layers learn to integrate spatially
865 patterned information from earlier layers to build a library of feature detectors. In the
866 proprioceptive system, such integration, without explicit transformation to some intermediate
867 movement representation, might allow neurons in area 2 to serve as a general-purpose library of
868 limb-state features, whose activity is read out in different ways for either perception or use in
869 motor control.

870 7.1.4 Hand kinematic-force model

871 Overall, our main results showed that the whole-arm model better captures firing rates and
872 features of the neural activity than does the hand-only model. One consideration in interpreting
873 these results is the fact that the whole-arm model is almost twice as expressive as the hand-only
874 model, due to its greater number of parameters. While we took care to make sure the models
875 were not overfitting (see Methods for details on cross-validation), a concern remains that any
876 signal related to the behavior may improve the fits, simply because it provides more information.
877 To address this concern, we would ideally compare these results with those from a model with
878 the same number of parameters, but with behavioral signals uncorrelated with elbow kinematics,
879 e.g., kinematics of the other hand. Unfortunately, due to experimental constraints, we only
880 collected tracking information from the reaching arm. As a substitute, we also tested a model we
881 titled “hand kinematic-force”, which builds on the hand-only kinematic model by adding the
882 forces and torques on the manipulandum handle. This model is similar to one proposed by
883 (Prud’homme and Kalaska 1994) and has the same number of parameters as the whole-arm
884 model. While the handle forces and torques are likely correlated with the elbow kinematics, this
885 model serves as a reasonable control to explore the particular importance of whole-arm
886 kinematics to area 2.

887 Figure 7 – figure supplement 3 shows comparisons between the whole-arm model and the hand
888 kinematic-force model on the three metrics we used. We found that the pR^2 and the tuning curve
889 correlation values for both models were comparable, with some neurons better described by the
890 whole-arm model and others by the kinematic-force model. However, we also found that the
891 hand kinematic-force model often could not predict large changes in PD as well as the whole-
892 arm model could (Figure S3C and 7). In four out of eight sessions, the whole-arm model had a
893 significantly higher cVAF than the hand kinematic-force model. In the other sessions, there was
894 no significant difference. While the two models made similar activity predictions, the better PD
895 shift predictions suggest that the whole-arm model is a better model for area 2 neural activity.



896

897 *Figure 7 – figure supplement 3: Comparison between whole-arm model and hand kinematic-force model*
898 (*shortened as “Kin-Force”*). *Same format as Figure 7 – figure supplements 1 and 2.*

899 **8 REFERENCES**

900 **An K, Takahashi K, Harrigan T, and Chao E.** Determination of muscle orientations and moment arms.
901 *Journal of biomechanical engineering* 106: 280-282, 1984.

902 **Azizi E, Brainerd EL, and Roberts TJ.** Variable gearing in pennate muscles. *Proceedings of the National*
903 *Academy of Sciences* 105: 1745-1750, 2008.

904 **Bell CC.** An efference copy which is modified by reafferent input. *Science* 214: 450-453, 1981.

905 **Berger M, and Gail A.** The Reach Cage environment for wireless neural recordings during structured
906 goal-directed behavior of unrestrained monkeys. *bioRxiv* 2018.

907 **Bosco G, Poppele RE, and Eian J.** Reference frames for spinal proprioception: limb endpoint based or
908 joint-level based? *J Neurophysiol* 83: 2931-2945, 2000.

909 **Bosco G, Rankin A, and Poppele R.** Representation of passive hindlimb postures in cat spinocerebellar
910 activity. *J Neurophysiol* 76: 715-726, 1996.

911 **Cameron AC, and Windmeijer FA.** An R-squared measure of goodness of fit for some common nonlinear
912 regression models. *Journal of econometrics* 77: 329-342, 1997.

913 **Cameron AC, and Windmeijer FA.** R-squared measures for count data regression models with
914 applications to health-care utilization. *Journal of Business & Economic Statistics* 14: 209-220, 1996.

915 **Caminiti R, Johnson PB, Galli C, Ferraina S, and Burnod Y.** Making arm movements within different
916 parts of space: the premotor and motor cortical representation of a coordinate system for reaching to
917 visual targets. *J Neurosci* 11: 1182-1197, 1991.

918 **Caminiti R, Johnson PB, and Urbano A.** Making arm movements within different parts of space:
919 Dynamic aspects in the primate motor cortex. *J Neurosci* 10: 2039-2058, 1990.

920 **Chan SS, and Moran DW.** Computational model of a primate arm: from hand position to joint angles,
921 joint torques and muscle forces. *Journal of neural engineering* 3: 327, 2006.

922 **Chapman C, Spidalieri G, and Lamarre Y.** Discharge properties of area 5 neurones during arm
923 movements triggered by sensory stimuli in the monkey. *Brain Research* 309: 63-77, 1984.

924 **Chowdhury RH, Tresch MC, and Miller LE.** Musculoskeletal geometry accounts for apparent extrinsic
925 representation of paw position in dorsal spinocerebellar tract. *Journal of neurophysiology* 2017.

926 **Churchland MM, Cunningham JP, Kaufman MT, Ryu SI, and Shenoy KV.** Cortical Preparatory Activity:
927 Representation of Movement or First Cog in a Dynamical Machine? *Neuron* 68: 387-400, 2010.

928 **Collinger JL, Wodlinger B, Downey JE, Wang W, Tyler-Kabara EC, Weber DJ, McMorland AJC, Velliste
929 M, Boninger ML, and Schwartz AB.** High-performance neuroprosthetic control by an individual with
930 tetraplegia. *The Lancet* 381: 557-564, 2013.

931 **Dadarlat MC, O'Doherty JE, and Sabes PN.** A learning-based approach to artificial sensory feedback
932 leads to optimal integration. *Nat Neurosci* 18: 138-144, 2015.

933 **Dekleva BM, Kording KP, and Miller LE.** Single reach plans in dorsal premotor cortex during a two-target
934 task. *Nature communications* 9: 3556, 2018.

935 **Deroy O, and Auvray M.** Reading the world through the skin and ears: A new perspective on sensory
936 substitution. *Frontiers in psychology* 3: 457, 2012.

937 **Dijkerman HC, and De Haan EH.** Somatosensory processing subserving perception and action:
938 Dissociations, interactions, and integration. *Behavioral and brain sciences* 30: 224-230, 2007.

939 **Elli GV, Benetti S, and Collignon O.** Is There a Future for Sensory Substitution Outside Academic
940 Laboratories? 27: 271, 2014.

941 **Elsayed GF, Lara AH, Kaufman MT, Churchland MM, and Cunningham JP.** Reorganization between
942 preparatory and movement population responses in motor cortex. *Nature communications* 7: 13239,
943 2016.

944 **Ernst D.** Get CI and p-values for cross validated performance measures. Cross Validated: 2017.

945 **Ethier C, Oby ER, Bauman MJ, and Miller LE.** Restoration of grasp following paralysis through brain-
946 controlled stimulation of muscles. *Nature* 485: 368-371, 2012.

947 **Evarts EV.** Relation of pyramidal tract activity to force exerted during voluntary movement. *J*
948 *Neurophysiol* 31: 14-27, 1968.

949 **Felleman DJ, and Van Essen DC.** Distributed hierarchical processing in the primate cerebral cortex.
950 *Cereb Cortex* 1: 1-47, 1991.

951 **Fetz EE.** Are movement parameters recognizably coded in activity of single neurons? *Behavioral and*
952 *Brain Sciences* 15: 679-690, 1992.

953 **Fetz EE, Cheney PD, Mewes K, and Palmer S.** Control of forelimb muscle activity by populations of
954 corticomotoneuronal and rubromotoneuronal cells. In: *Progress in Brain Research: Afferent Control of*
955 *Posture and Locomotion*, edited by Allum JHJ, and Hulliger M. Amsterdam: Elvsevier Science, 1989, p.
956 437-449.

957 **Flesher SN, Collinger JL, Foldes ST, Weiss JM, Downey JE, Tyler-Kabara EC, Bensmaia SJ, Schwartz AB,**
958 **Boninger ML, and Gaunt RA.** Intracortical microstimulation of human somatosensory cortex. *Science*
959 *translational medicine* 8: 361ra141-361ra141, 2016.

960 **Fuentes CT, and Bastian AJ.** Where Is Your Arm? Variations in Proprioception Across Space and Tasks.
961 *Journal of neurophysiology* 103: 164-171, 2010.

962 **Gallego JA, Perich MG, Miller LE, and Solla SA.** Neural Manifolds for the Control of Movement. *Neuron*
963 94: 978-984, 2017.

964 **Georgopoulos AP, Kalaska JF, Caminiti R, and Massey JT.** On the relations between the direction of two-
965 dimensional arm movements and cell discharge in primate motor cortex. *J Neurosci* 2: 1527-1537, 1982.

966 **Georgopoulos AP, Schwartz AB, and Kettner RE.** Neuronal population coding of movement direction.
967 *Science* 233: 1416-1419, 1986.

968 **Ghez C, and Sainburg R.** Proprioceptive control of interjoint coordination. *Can J Physiol Pharmacol* 73:
969 273-284, 1995.

970 **Gordon J, Ghilardi MF, and Ghez C.** Impairments of reaching movements in patients without
971 proprioception. I. Spatial Errors. *Journal of neurophysiology* 73: 347-360, 1995.

972 **Haftel VK, Bichler EK, Nichols TR, Pinter MJ, and Cope TC.** Movement reduces the dynamic response of
973 muscle spindle afferents and motoneuron synaptic potentials in rat. *Journal of neurophysiology* 91:
974 2164-2171, 2004.

975 **Hasson CJ, Shen T, and Sternad D.** Energy margins in dynamic object manipulation. *Journal of*
976 *neurophysiology* 108: 1349-1365, 2012.

977 **Heath C, Hore J, and Phillips C.** Inputs from low threshold muscle and cutaneous afferents of hand and
978 forearm to areas 3a and 3b of baboon's cerebral cortex. *The Journal of physiology* 257: 199-227, 1976.

979 **Heinzl H, and Mittlböck M.** Pseudo R-squared measures for Poisson regression models with over- or
980 underdispersion. *Computational statistics & data analysis* 44: 253-271, 2003.

981 **Hubel DH, and Wiesel TN.** Receptive fields of single neurones in the cat's striate cortex. *J Physiol* 148:
982 574-591, 1959.

983 **Hubel DH, and Wiesel TN.** Receptive fields, binocular interaction and functional architecture in the cat's
984 visual cortex. *J Physiol* 160: 106-154, 1962.

985 **Hyvärinen J, and Poranen A.** Receptive field integration and submodality convergence in the hand area
986 of the post-central gyrus of the alert monkey. *The Journal of Physiology* 283: 539, 1978.

987 **Jennings VA, Lamour Y, Solis H, and Fromm C.** Somatosensory cortex activity related to position and
988 force. *J Neurophysiol* 49: 1216-1229, 1983.

989 **Kaas JH, Nelson RJ, Sur M, Lin CS, and Merzenich MM.** Multiple representations of the body within the
990 primary somatosensory cortex of primates. *Science* 204: 521-523, 1979.

991 **Kakei S, Hoffman DS, and Strick PL.** Muscle and movement representations in the primary motor cortex.
992 *Science* 285: 2136-2139, 1999.

993 **Kao JC, Nuyujukian P, Ryu SI, Churchland MM, Cunningham JP, and Shenoy KV.** Single-trial dynamics of
994 motor cortex and their applications to brain-machine interfaces. *Nat Commun* 6: 2015.

995 **Kaufman MT, Churchland MM, Ryu SI, and Shenoy KV.** Cortical activity in the null space: permitting
996 preparation without movement. *Nat Neurosci* 17: 440-448, 2014.

997 **Krizhevsky A, Sutskever I, and Hinton GE.** Imagenet classification with deep convolutional neural
998 networks. In: *Advances in neural information processing systems* 2012, p. 1097-1105.

999 **Lillicrap TP, and Scott SH.** Preference distributions of primary motor cortex neurons reflect control
1000 solutions optimized for limb biomechanics. *Neuron* 77: 168-179, 2013.

1001 **Loeb GE, Hoffer JA, and Marks WB.** Activity of spindle afferents from cat anterior thigh muscles. III.
1002 Effects of external stimuli. *J Neurophysiol* 54: 578-591, 1985.

1003 **London BM, and Miller LE.** Responses of somatosensory area 2 neurons to actively and passively
1004 generated limb movements. *J Neurophysiol* 109: 1505-1513, 2013.

1005 **London BM, Ruiz-Torres R, Slutzky MW, and Miller LE.** Designing stimulation patterns for an afferent
1006 BMI: Representation of kinetics in somatosensory cortex. In: *Engineering in Medicine and Biology
1007 Society, EMBC, 2011 Annual International Conference of the IEEE* 2011, p. 7521-7524.

1008 **Lucas A, Tomlinson T, Rohani N, Chowdhury RH, Solla SA, Katsaggelos AK, and Miller LE.** Deep Neural
1009 Networks for Modeling Neural Spiking in S1 Cortex. *Frontiers in Systems Neuroscience* 13: 13, 2019.

1010 **Mathis MW, Mathis A, and Uchida N.** Somatosensory cortex plays an essential role in forelimb motor
1011 adaptation in mice. *Neuron* 93: 1493-1503. e1496, 2017.

1012 **Mazurek K, Berger M, Bollu T, Chowdhury R, Elangovan N, Kuling I, and Sohn M.** Highlights from the
1013 28th Annual Meeting of the Society for the Neural Control of Movement: American Physiological Society
1014 Bethesda. MD, 2018.

1015 **McFadden D.** *Quantitative methods for analyzing travel behavior of individuals: some recent
1016 developments.* Institute of Transportation Studies, University of California Berkeley, 1977.

1017 **Mishkin M, and Ungerleider LG.** Contribution of striate inputs to the visuospatial functions of parieto-
1018 preoccipital cortex in monkeys. *Behavioural brain research* 6: 57-77, 1982.

1019 **Moran DW, and Schwartz AB.** Motor cortical representation of speed and direction during reaching. *J
1020 Neurophysiol* 82: 2676-2692, 1999.

1021 **Morrow MM, Jordan LR, and Miller LE.** Direct comparison of the task-dependent discharge of M1 in
1022 hand space and muscle space. *J Neurophysiol* 97: 1786-1798, 2007.

1023 **Mountcastle VB, Lynch JC, Georgopoulos A, Sakata H, and Acuna C.** Posterior parietal association
1024 cortex of the monkey: command functions for operations within extrapersonal space. *J Neurophysiol* 38:
1025 871-908, 1975.

1026 **Mussa-Ivaldi FA.** Do neurons in the motor cortex encode movement direction? An alternative
1027 hypothesis. *Neurosci Lett* 91: 106-111, 1988.

1028 **Nadeau C, and Bengio Y.** Inference for the Generalization Error. *Machine Learning* 52: 239-281, 2003.

1029 **Nasir SM, Darainy M, and Ostry DJ.** Sensorimotor adaptation changes the neural coding of
1030 somatosensory stimuli. *J Neurophysiol* 109: 2077-2085, 2013.

1031 **Nelson RJ.** Activity of monkey primary somatosensory cortical neurons changes prior to active
1032 movement. *Brain Research* 406: 402-407, 1987.

1033 **Oby ER, Ethier C, and Miller LE.** Movement representation in the primary motor cortex and its
1034 contribution to generalizable EMG predictions. *J Neurophysiol* 109: 666-678, 2012.

1035 **Omran M, Murnaghan CD, Pruszynski JA, and Scott SH.** Distributed task-specific processing of
1036 somatosensory feedback for voluntary motor control. *Elife* 5: e13141, 2016.

1037 **Padberg J, Cooke DF, Cerkevich CM, Kaas JH, and Krubitzer L.** Cortical connections of area 2 and
1038 posterior parietal area 5 in macaque monkeys. *Journal of Comparative Neurology* 2018.

1039 **Perich MG, Gallego JA, and Miller LE.** A Neural Population Mechanism for Rapid Learning. *Neuron* 100:
1040 964-976.e967, 2018.

1041 **Phillips CG, Powell TP, and Wiesendanger M.** Projection from low-threshold muscle afferents of hand
1042 and forearm to area 3a of baboon's cortex. *J Physiol* 217: 419-446, 1971.

1043 **Pons TP, Garraghty PE, Cusick CG, and Kaas JH.** The somatotopic organization of area 2 in macaque
1044 monkeys. *J Comp Neurol* 241: 445-466, 1985.

1045 **Prochazka A, and Wand P.** Independence of fusimotor and skeletomotor systems during voluntary
1046 movement. In: *Muscle receptors and movement*, edited by A. Taylor A. ProchazkaMacMillan London,
1047 1981.

1048 **Prochazka A, Westerman R, and Ziccone S.** Discharges of single hindlimb afferents in the freely moving
1049 cat. *Journal of neurophysiology* 39: 1090-1104, 1976.

1050 **Proske U, and Stuart G.** The initial burst of impulses in responses of toad muscle spindles during stretch.
1051 *The Journal of physiology* 368: 1-17, 1985.

1052 **Prud'homme MJL, and Kalaska JF.** Proprioceptive activity in primate primary somatosensory cortex
1053 during active arm reaching movements. *J Neurophysiol* 72: 2280-2301, 1994.

1054 **Rack PM, and Westbury DR.** Elastic properties of the cat soleus tendon and their functional importance.
1055 *The Journal of Physiology* 347: 479-495, 1984.

1056 **Richardson AG, Weigand PK, Sritharan SY, and Lucas TH.** A chronic neural interface to the macaque
1057 dorsal column nuclei. *Journal of neurophysiology* 115: 2255-2264, 2016.

1058 **Romo R, Hernandez A, Zainos A, and Salinas E.** Somatosensory discrimination based on cortical
1059 microstimulation. *Nature* 392: 387-390, 1998.

1060 **Russo AA, Bittner SR, Perkins SM, Seely JS, London BM, Lara AH, Miri A, Marshall NJ, Kohn A, Jessell
1061 TM, Abbott LF, Cunningham JP, and Churchland MM.** Motor Cortex Embeds Muscle-like Commands in
1062 an Untangled Population Response. *Neuron* 2018.

1063 **Sainburg RL, Ghilardi MF, Poizner H, and Ghez C.** Control of limb dynamics in normal subjects and
1064 patients without proprioception. *JNeurophysiol* 73: 820-835, 1995.

1065 **Sainburg RL, Poizner H, and Ghez C.** Loss of proprioception produces deficits in interjoint coordination. *J
1066 Neurophysiol* 70: 2136-2147, 1993.

1067 **Salas MA, Bashford L, Kellis S, Jafari M, Jo H, Kramer D, Shanfield K, Pejsa K, Lee B, Liu CY, and
1068 Andersen RA.** Proprioceptive and cutaneous sensations in humans elicited by intracortical
1069 microstimulation. *eLife* 7: e32904, 2018.

1070 **Sanes JN, Mauritz KH, Evarts EV, Dalakas MC, and Chu A.** Motor deficits in patients with large-fiber
1071 sensory neuropathy. *Proc Natl Acad Sci U S A* 81: 979-982, 1984.

1072 **Scott SH, and Kalaska JF.** Changes in motor cortex activity during reaching movements with similar hand
1073 paths but different arm postures. *Journal of neurophysiology* 73: 2563-2567, 1995.

1074 **Sedda A, and Scarpina F.** Dorsal and ventral streams across sensory modalities. *Neuroscience Bulletin*
1075 28: 291-300, 2012.

1076 **Seelke AM, Padberg JJ, Disbrow E, Purnell SM, Recanzone G, and Krubitzer L.** Topographic maps within
1077 Brodmann's area 5 of macaque monkeys. *Cerebral Cortex* 22: 1834-1850, 2011.

1078 **Shadmehr R, and Mussa-Ivaldi FA.** Adaptive representation of dynamics during learning of a motor task.
1079 *J Neurosci* 14: 3208-3224., 1994.

1080 **Sharon Y, and Nisky I.** What Can Spatiotemporal Characteristics of Movements in RAMIS Tell Us? *arXiv
1081 preprint arXiv:171005818* 2017.

1082 **Suresh AK, Winberry J, Versteeg C, Chowdhury RH, Tomlinson T, Rosenow JM, Miller LE, and Bensmaia
1083 SJ.** Methodological considerations for a chronic neural interface with the cuneate nucleus of macaques.
1084 *Journal of neurophysiology* jn. 00436.02017, 2017.

1085 **Sussillo D, Churchland MM, Kaufman MT, and Shenoy KV.** A neural network that finds a naturalistic
1086 solution for the production of muscle activity. *Nat Neurosci* 18: 1025-1033, 2015.

1087 **Tabot GA, Dammann JF, Berg JA, Tenore FV, Boback JL, Vogelstein RJ, and Bensmaia SJ.** Restoring the
1088 sense of touch with a prosthetic hand through a brain interface. *Proceedings of the National Academy of*
1089 *Sciences* 110: 18279-18284, 2013.

1090 **Tomlinson T, and Miller LE.** Toward a proprioceptive neural interface that mimics natural cortical
1091 activity. In: *Progress in Motor Control: Theories and Translations*, edited by Laczko J, and Latash
1092 MLSpringer, 2016.

1093 **Truccolo W, Eden UT, Fellows MR, Donoghue JP, and Brown EN.** A point process framework for relating
1094 neural spiking activity to spiking history, neural ensemble, and extrinsic covariate effects. *J Neurophysiol*
1095 93: 1074-1089, 2005.

1096 **Weber DJ, London BM, Hokanson JA, Ayers CA, Gaunt RA, Ruiz-Torres R, Zaaimi B, and Miller LE.** Limb-
1097 state information encoded by peripheral and central somatosensory neurons: Implications for an
1098 afferent interface. *Neural Systems and Rehabilitation Engineering, IEEE Transactions on* 19: 501-513,
1099 2011.

1100 **Weber DJ, Stein RB, Everaert DG, and Prochazka A.** Decoding sensory feedback from firing rates of
1101 afferent ensembles recorded in cat dorsal root ganglia in normal locomotion. *IEEE Trans Neural Syst*
1102 *Rehabil Eng* 14: 240-243, 2006.

1103 **Wolpert DM, Ghahramani Z, and Jordan MI.** An internal model for sensorimotor integration. *Science*
1104 269: 1880-1882., 1995.

1105 **Yamada H, Yaguchi H, Tomatsu S, Takei T, Oya T, and Seki K.** Representation of Afferent Signals from
1106 Forearm Muscle and Cutaneous Nerves in the Primary Somatosensory Cortex of the Macaque Monkey.
1107 *PLoS ONE* 11: e0163948, 2016.

1108 **Young D, Willett F, Memberg WD, Murphy BA, Rezaii P, Walter B, Sweet JA, Miller J, Shenoy KV, and**
1109 **Hochberg L.** Closed-loop cortical control of virtual reach and posture using Cartesian and joint velocity
1110 commands. *Journal of neural engineering* 2018.

1111 **Zaaimi B, Ruiz-Torres R, Solla SA, and Miller LE.** Multi-electrode stimulation in somatosensory cortex
1112 increases probability of detection. *Journal of neural engineering* 10: 056013, 2013.