

A Comparison Of Robust Mendelian Randomization Methods Using Summary Data

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Abstract

The number of Mendelian randomization analyses including large numbers of genetic variants is rapidly increasing. This is due to the proliferation of genome-wide association studies, and the desire to obtain more precise estimates of causal effects. Since it is unlikely that all genetic variants will be valid instrumental variables, several robust methods have been proposed. We compare nine robust methods for Mendelian randomization based on summary data that can be implemented using standard statistical software. Methods were compared in three ways: by reviewing their theoretical properties, in an extensive simulation study, and in an empirical example to investigate the effect of body mass index on coronary artery disease risk. In the simulation study, the overall best methods, judged by mean squared error, were the contamination mixture method and the mode based estimation method. These methods generally had well-controlled Type 1 error rates with up to 50% invalid instruments across a range of scenarios. Outlier-robust methods such as MR-Lasso, MR-Robust, and MR-PRESSO, had the narrowest confidence intervals in the empirical example. They performed well when most variants were valid instruments with a few outliers, but less well with several invalid instruments. With isolated exceptions, all methods performed badly when over 50% of the variants were invalid instruments. Our recommendation for investigators is to perform a variety of robust methods that operate in different ways and rely on different assumptions for valid inferences to assess the reliability of Mendelian randomization analyses.

Keywords: Mendelian randomization, instrumental variables, causal inference, pleiotropy, robust estimation, summary statistics, genome-wide association study.

Introduction

Mendelian randomization (MR) uses genetic variants as instrumental variables (IV) to determine whether an observational association between a risk factor and an outcome is consistent with a causal effect [1, 2]. This approach is less vulnerable to traditional problems of epidemiological studies such as confounding and reverse causality. With the increasing availability of genome-wide association studies that find robust associations between genetic variants and exposures of interest [3, 4], the potential of this approach is rapidly evolving. A genetic variant is a valid IV if (i) it is associated with the exposure, (ii) it has no direct effect on the outcome, and (iii) there are no associations between the variant and any potential confounders.

There has been much discussion on the potentials and limitations of MR, as the IV assumptions cannot be fully tested [1, 5, 6]. Violation of the IV assumptions can lead to invalid conclusions in applied investigations. In practice, the exclusion restriction assumption that the proposed instruments (genetic variants) should not have a direct effect on the outcome of interest is debatable, particularly if the biological roles of the genetic variants are insufficiently understood [5, 7].

Some genetic variants are associated with multiple phenotypic variables [8, 9]. This is referred to as pleiotropy. There are two types of pleiotropy. Vertical pleiotropy occurs when a variant is directly associated with the exposure and another phenotype on the same biological pathway. This does not lead to violation of the IV assumptions provided the only causal pathway from the genetic variant to the outcome passes via the exposure. Horizontal pleiotropy occurs when the second phenotype is on a different biological pathway, and so there may exist different causal pathways from the variant to the outcome. This would violate the exclusion restriction assumption. To solve the problems that arise due to horizontal pleiotropy, several robust methods for MR have been developed that can provide reliable inferences when some genetic variants violate the IV assumptions, or when genetic variants violate the IV assumptions in a particular way. To our knowledge, a comprehensive review and simulation study to compare the statistical performance of these different methods has not been performed.

To focus our simulation study and compare the most relevant robust methods for applied practice, we concentrate on methods that satisfy two criteria. First, the method requires only summary data on estimates (beta-coefficients and standard errors) of genetic variant–exposure and genetic variant–outcome associations. We exclude methods that require individual participant data [10–13], and those that require data on additional variants not associated with the risk factor [14, 15]. This is because the sharing of individual participant data is often impractical, so that many empirical researchers only have access to summary data, and for fairness, to ensure that all methods are using the same information to make inferences. Secondly, the method must be performed using standard statistical software packages. We exclude methods requiring specific computational tools that are unlikely to be accessible to the majority of epidemiologists [16] or are computationally infeasible for large numbers of variants in a reasonable running time [17].

In this article, we review nine robust methods for MR from a theoretical perspective, and evaluate their performance in a simulation study set in a two-sample summary data setting. The methods differ in how they estimate a causal effect of the exposure on the outcome, as well as in the assump-

tions required for consistent estimation. We consider the weighted median, mode based estimation, MR-PRESSO, MR-Robust, MR-Lasso, MR-Egger, contamination mixture, MR-Mix, and MR-RAPS methods. Some methods take a summarized measure of the variant-specific causal estimates as the overall causal effect estimate (weighted median, and mode based estimation), whereas others remove or downweight outliers (MR-PRESSO, MR-Lasso, MR-Robust), or attempt to model the distribution of the estimates from invalid IVs (MR-Egger, contamination mixture, MR-Mix, and MR-RAPS). We also consider the performance of the methods in an empirical example to evaluate the causal effect of body mass index on coronary artery disease risk.

This paper is organized as follows. First, we give an overview of the robust methods and compare their theoretical properties. Then, we introduce the simulation framework and applied example to compare their properties in practice. Finally, we discuss the implications of this work for applied practice.

Methods

Modelling assumptions and summary data

We consider a model as previously described [18, 19] for J genetic variants $G_1, G_2, \dots, G_J$ that are independent in their distributions, a modifiable exposure X , an outcome variable Y , and a confounder U . We assume that all relationships between variables are linear and homogeneous without effect modification, meaning that the same causal effect is estimated by any valid IV [20]. A visual representation of the model is shown in Figure 1.

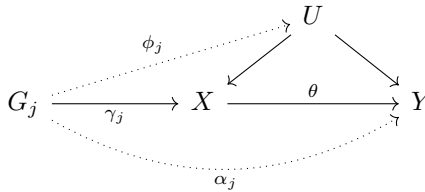


Fig. 1 Illustrative diagram showing the model assumed for genetic variant G_j , with effect ϕ_j on the unobserved confounder U , effect γ_j on exposure X , and direct effect α_j on outcome Y . The causal effect of the exposure on the outcome is θ . Dotted lines represent possible ways the instrumental variable assumptions could be violated.

We assume that summary data are available on genetic associations with the exposure (beta-coefficient $\hat{\beta}_{X_j}$ and standard error σ_{X_j}) and with the outcome (beta-coefficient $\hat{\beta}_{Y_j}$ and standard error σ_{Y_j}) for each variant G_j .

Inverse-variance weighted method

The causal effect of the exposure on the outcome can be estimated using a single genetic variant G_j by the ratio method:

$$\hat{\theta}_{R_j} = \frac{\hat{\beta}_{Y_j}}{\hat{\beta}_{X_j}}. \quad (1)$$

The ratio estimate $\hat{\theta}_{R_j}$ is a consistent estimate of the causal effect if variant G_j satisfies the IV assumptions [20]. If the uncertainty in the genetic association with the exposure is low, then the standard error of the ratio estimate σ_{R_j} can be approximated as [21]:

$$\sigma_{R_j} = \left| \frac{\sigma_{Y_j}}{\hat{\beta}_{X_j}} \right|. \quad (2)$$

The individual ratio estimates can be combined to obtain a single more efficient estimate. The optimally-efficient combination of the ratio estimates is referred to as the inverse-variance weighted (IVW) estimate [22]:

$$\hat{\beta}_{\text{IVW}} = \frac{\sum_{j=1}^J \hat{\theta}_{R_j} \sigma_{R_j}^{-2}}{\sum_{j=1}^J \sigma_{R_j}^{-2}} = \frac{\sum_{j=1}^J \hat{\beta}_{X_j} \hat{\beta}_{Y_j} \sigma_{Y_j}^{-2}}{\sum_{j=1}^J \hat{\beta}_{X_j}^2 \sigma_{Y_j}^{-2}}. \quad (3)$$

The IVW estimate is equal to the estimate from the two-stage least squares method that is performed using individual participant data [23]. It is a weighted mean of the ratio estimates, where the weights are the inverse-variances of the ratio estimates. The IVW estimate can also be obtained by weighted regression of the genetic associations with the outcome on the genetic associations with the exposure:

$$\hat{\beta}_{Y_j} = \theta \hat{\beta}_{X_j} + \varepsilon_j, \quad \varepsilon_j \sim \mathcal{N}(0, \sigma_{Y_j}^2). \quad (4)$$

However, the IVW method has a 0% breakdown point, meaning that if only one genetic variant is not a valid IV, then the estimator is typically biased [24]. Bias will be present unless the pleiotropic effects of genetic variants average to zero (balanced pleiotropy) and the pleiotropic effects are independent of the genetic variant–exposure associations (see MR-Egger method below) [19]. With the increasing number of variants used in MR investigations, it is increasingly unlikely that all variants are valid IVs. Hence, it is crucial to consider robust estimation methods despite their lower statistical efficiency (that is, lower power to detect a causal effect).

We proceed to introduce the different robust methods we consider in this study in three categories: consensus methods, outlier-robust methods, and modelling methods.

Consensus methods

A consensus method is one that takes its causal estimate as a summary measure of the distribution of the ratio estimates. The most straightforward consensus method is the median method. Rather than taking a weighted mean of the ratio estimates as in the IVW method, we take the median of the ratio estimates. The median estimator is consistent (that is, unbiased in large samples) even if up to 50% of the variants are invalid [24]. We consider a weighted version of the median method, where the median is taken from a distribution of the ratio estimates in which genetic variants with more precise ratio estimates receive more weight. Here, an unbiased estimate will be obtained if up to 50% of the weight comes from variants that are valid IVs. We refer to this as the ‘majority valid’ assumption.

A related assumption is the ‘plurality valid’ assumption [11]. In large samples, while ratio estimates for all valid IVs should equal the true causal effect, ratio estimates for invalid IVs will take different values. The ‘plurality valid’ assumption is that, out of all the different values taken by ratio estimates in large samples (we term these the ratio estimands), the true causal effect is the value taken for the largest number of genetic variants (that is, the modal ratio estimand). For example, the plurality assumption would be satisfied if only 40% of the genetic variants are valid instruments, provided that out of the remaining 60% invalid instruments, no larger group with the same ratio estimand exists. This assumption is also referred to as the Zero Modal Pleiotropy Assumption (ZEMPA) [25].

This assumption is exploited by the mode based estimation (MBE) method [25]. As no two ratio estimates will be identical in finite samples, it is not possible to take the mode of the ratio estimates directly. In the MBE method, a normal density is drawn for each genetic variant centered at its ratio estimate. The spread of this density depends on a bandwidth parameter, and (for the weighted version of the MBE method) the precision of the ratio estimate. A smoothed density function is then constructed by summing these normal densities. The maximum of this distribution is the causal estimate.

As these consensus methods take the median or mode of the ratio estimate distribution as the causal estimate, they are naturally robust to outliers, as the median and mode of a distribution are unaffected by the magnitude of extreme values. However, they are still influenced by outliers, as these variants still contribute to determining the location of the median or mode of a distribution. These methods can also be sensitive to changes in the ratio estimates for variants that contribute to the median or mode, and to the addition and removal of variants from the analysis. Additionally, the methods may not be as efficient as those that base their estimates on all the genetic variants.

Outlier-robust methods

Next, we present three outlier-robust methods. These methods either downweight or remove genetic variants from the analysis that have outlying ratio estimates. They provide consistent estimates under the same assumptions as the IVW method for the set of genetic variants that are not identified as outliers.

In the MR-Pleiotropy Residual Sum and Outlier (MR-PRESSO) method [26], the IVW method is implemented by regression using all the genetic variants, and the residual sum of squares (RSS) is calculated from the regression equation. The RSS is a heterogeneity measure for the ratio estimates.

Then, the IVW method is performed omitting each genetic variant from the analysis in turn. If the RSS decreases substantially compared to a simulated expected distribution, then that variant is removed from the analysis. This procedure is repeated until no further variants are removed from the analysis. The causal estimate is then obtained by the IVW method using the remaining genetic variants.

In MR-Robust, the IVW method is performed by regression, except that instead of using ordinary least squares regression, MM-estimation is used combined with Tukey's biweight loss function [27]. MM-estimation provides robustness against influential points and Tukey's loss function provides robustness against outliers. Tukey's loss function is a truncated quadratic function, meaning that there is a limit in the degree to which an outlier contributes to the analysis [28]. This contrasts with the quadratic loss function used in ordinary least squares regression, which is unbounded, meaning that a single outlier can have an unlimited effect on the IVW estimate.

In MR-Lasso, the IVW regression model is augmented by adding an intercept term for each genetic variant [27]. The IVW estimate is the value of θ that minimizes:

$$\sum_{j=1}^J \sigma_{Y_j}^{-2} \left(\hat{\beta}_{Y_j} - \theta \hat{\beta}_{X_j} \right)^2. \quad (5)$$

In MR-Lasso, we minimize:

$$\sum_{j=1}^J \sigma_{Y_j}^{-2} \left(\hat{\beta}_{Y_j} - \theta_{0j} - \theta \hat{\beta}_{X_j} \right)^2 + \lambda \sum_{j=1}^J |\theta_{0j}|, \quad (6)$$

where λ is a tuning parameter. As the regression equation contains more parameters than there are genetic variants, a lasso penalty term is added for identification [29]. The intercept term θ_{0j} represents the direct (pleiotropic) effect on the outcome, and should be zero for a valid IV, but will be non-zero for an invalid IV. The causal estimate is then obtained by the IVW method using the genetic variants that had $\theta_{0j} = 0$ in equation (6). A heterogeneity criterion is used to determine the value of λ . Increasing λ means that more of the pleiotropy parameters equal zero and so the corresponding variants are included in the analysis; we increase λ step-by-step until one step before there is more heterogeneity in the ratio estimates for variants included in the analysis than expected by chance alone.

The MR-PRESSO and MR-Lasso methods remove variants from the analysis, whereas MR-Robust downweights variants. These methods will be valuable when there is a small number of genetic variants with heterogeneous ratio estimates, as they will be removed from the analysis or heavily down-weighted, and so will not influence the overall estimate. In such a case, these methods are likely to be efficient, as they are based on the IVW method. The methods are less likely to be valuable when there is a larger number of genetic variants that are pleiotropic, particularly if the pleiotropic effects are small in magnitude, and when the average pleiotropic effect of non-outliers is not zero.

Modelling methods

Finally, we present four methods that attempt to model the distribution of estimates from invalid IVs or make a specific assumption about the way in which the IV assumptions are violated. The MR-

Egger method is performed similarly to the IVW method, except that the regression model contains an intercept term θ_0 :

$$\hat{\beta}_{Y_j} = \theta_0 + \theta \hat{\beta}_{X_j} + \varepsilon_j, \quad \varepsilon_j \sim \mathcal{N}(0, \sigma_{Y_j}^2). \quad (7)$$

This differs from the MR-Lasso method, as there is only one intercept term, which represents the average pleiotropic effect. The MR-Egger method gives consistent estimates of the causal effect under the Instrument Strength Independent of Direct Effect (InSIDE) assumption, which states that pleiotropic effects of genetic variants must be uncorrelated with genetic variant–exposure association. As the regression model is no longer symmetric to changes in the signs of the genetic association estimates (which result from switching the reference and effect alleles), we first re-orientate the genetic associations before performing the regression by fixing all genetic associations with the exposure to be positive, and correspondingly changing the signs of the genetic associations with the outcome if necessary. The intercept in MR-Egger also provides a test of the IV assumptions. The intercept will differ from zero when either the average pleiotropic effect is not zero, or the InSIDE assumption is violated. These are precisely the conditions required for the IVW estimate to be unbiased.

The contamination mixture method assumes that only some of the genetic variants are valid IVs [30]. We construct a likelihood function from the ratio estimates. If a variant is a valid instrument, then its ratio estimate is assumed to be normally distributed about the true causal effect θ with variance $\sigma_{R_j}^2$. If a variant is not a valid instrument, then its ratio estimate is assumed to be normally distributed about zero with variance $\psi^2 + \sigma_{R_j}^2$, where ψ^2 represents the variance of the estimands from invalid IVs. This parameter is specified by the analyst. We then maximize the likelihood over different values of the causal effect θ and different configurations of valid and invalid IVs. Maximization is performed in linear time by first constructing a profile likelihood as a function of θ , and then maximizing this function with respect to θ . The value of θ that maximizes the profile likelihood is the causal estimate.

The MR-Mix method [31] is similar to the contamination mixture method, except that rather than dividing the genetic variants into valid and invalid IVs, the method divides variants into four categories: 1) variants that directly influence the exposure only (valid instruments), and 2) variants that influence the exposure and outcome, 3) that influence the outcome only, and 4) that neither influence the exposure or outcome (invalid instruments). This allows for more flexibility in modelling genetic variants, although potentially leads to more uncertainty in assigning genetic variants to categories.

The MR-Robust Adjusted Profile Score (RAPS) [32] method models the pleiotropic effects of genetic variants directly using a random-effects distribution. The pleiotropic effects are assumed to be normally distributed about zero with unknown variance. Estimates are obtained using a profile likelihood function for the causal effect and the variance of the pleiotropic effect distribution. To provide further robustness to outliers, either Tukey’s biweight loss function or Huber’s loss function [28] can be used.

Modelling methods are likely to be valuable when the modelling assumptions are correct, but not when the assumptions are incorrect. For example, the MR-Egger method requires the InSIDE assumption to be satisfied to give a consistent estimate. The MR-RAPS method is likely to perform

well when pleiotropic effects truly are normally distributed about zero, but less well when they are not. The MR-Mix method is likely to require large numbers of genetic variants in order to correct classify variants into the different categories. The contamination mixture method is less likely to be affected by modelling assumptions as it does not make such strict assumptions, but it is likely to be sensitive to specification of the variance parameter.

Simulation study

To compare the performance of these methods in a realistic setting, we perform a simulation study. Full details of the simulation study are given in the Supplementary Material. In brief, we consider three scenarios:

1. balanced pleiotropy, InSIDE satisfied – invalid IVs have direct effects on the outcome generated from a normal distribution centered at zero;
2. directional pleiotropy, InSIDE satisfied – invalid IVs have direct effects on the outcome generated from a normal distribution centered away from zero;
3. directional pleiotropy, InSIDE violated – invalid IVs have direct effects on the outcome generated from a normal distribution centered away from zero, and indirect effects on the outcome via the confounder.

We simulated data on $J = 10, 30$, and 100 genetic variants. A portion of the genetic variants were invalid IVs (30%, 50% and 70%), and the direct effects of the variants explain 10% of the variance in the exposure. Summary genetic associations were calculated for the exposure and the outcome on non-overlapping sets of individuals, each consisting of 10 000 individuals [33]. This situation is often referred to as two-sample summary data MR [34]. We considered situations with a null causal effect ($\theta = 0$) and a positive causal effect ($\theta = 0.2$). In total, 10 000 datasets were generated in each scenario. We additionally considered scenarios with 500 genetic variants and a wider range of proportions of invalid IVs (additionally 1%, 5%, and 10% invalid).

Empirical example: the effect of body mass index on coronary artery disease risk

We also compare the methods in an empirical example considering the effect of body mass index (BMI) on coronary artery disease (CAD) risk. Since BMI is influenced by several biological mechanisms [35], it is likely that the exclusion restriction is not satisfied for all associated genetic variants. Hence it is necessary to use robust methods to analyse these data. Additionally, we consider methods that detect outliers (MR-Presso, MR-Robust, MR-Lasso, contamination mixture, MR-Mix, and MR-RAPS), and compare whether the same outliers are detected in each of these methods.

We take 97 genome-wide significant variants associated with BMI from the GIANT consortium [36]. Associations with BMI are estimated in up to 339,224 participants from this consortium. Associations with coronary artery disease risk are estimated in up to 60,801 CAD cases and 123,504 controls

from the CARDIoGRAMplusC4D Consortium [37]. Association estimates for CAD were available for 94 of these variants.

The scatter plot of the genetic associations with BMI and CAD risk is shown in Figure 2. While most variants seem to suggest a harmful effect of increased BMI on CAD risk, there is substantial heterogeneity in the plot. This suggests that some of the variants violate the IV assumptions.

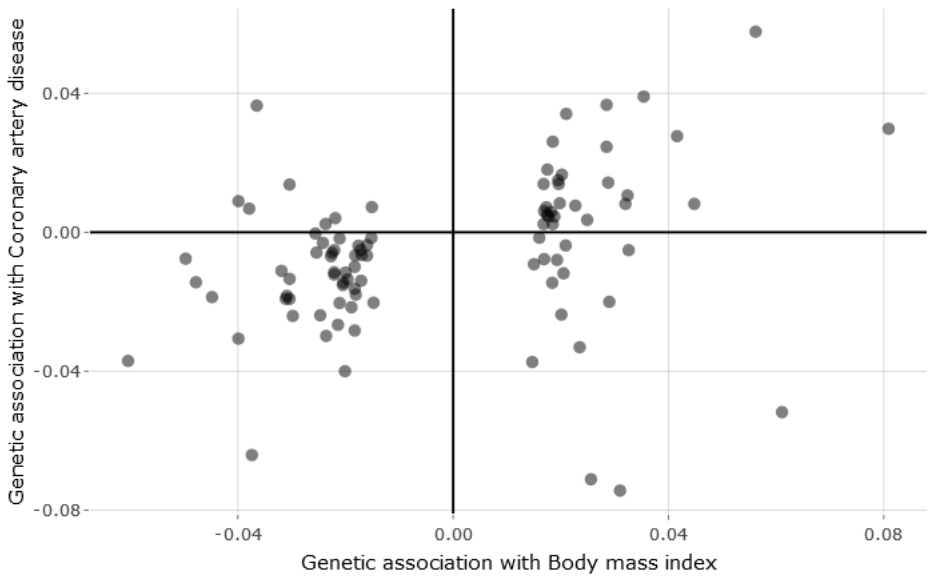


Fig. 2 Scatter plot of genetic associations with BMI (standard deviation units) and coronary artery disease risk (log odds ratios) for 94 variants taken from the GIANT and CARDIoGRAMplusC4D consortia respectively.

Results

Simulation study

Results of the simulation study are presented in Table 1 (10 variants), Table 2 (30 variants), and Table 3 (100 variants). For each scenario, we present the mean, median, and standard deviation of estimates across simulations, and the empirical power of the 95% confidence interval. With a null causal effect, the empirical power is the Type 1 error rate, and should be close to 0.05. The mean squared error across simulations for the different methods with a null causal effect is presented in Figure 3 (Scenario 2), and Figure 4 (Scenario 3) for 30 variants. The corresponding plots for 10 variants (Supplementary Figures 5 and 6) and 100 variants (Supplementary Figures 7 and 8) were broadly similar, as were results with 500 variants (Supplementary Tables 6 and 7).

Overall, judging by mean squared error, the contamination mixture and MBE methods performed best. The contamination mixture method performed slightly better with 30% and 50% invalid variants, and the MBE method performed better with 70% invalid variants. However, with some isolated exceptions, all the methods performed badly with 70% invalid instruments. Between these two methods,

MBE tended to be more conservative, whereas the contamination mixture method had slightly lower standard deviation of estimates and increased power to detect a causal effect. Neither method consistently dominated the other in terms of Type 1 error rate. Several other methods performed well in particular scenarios.

Amongst consensus methods, estimates from the MBE method were less biased than those from the weighted median method, with lower Type 1 errors. The weighted median method had slightly higher power to detect a causal effect, although comparisons of power lose much of their value when a method has inflated Type 1 error rates. Amongst outlier-robust methods, estimates were similar amongst the methods, with the MR-Lasso method generally having the lowest bias, but MR-Robust having the lowest Type 1 error rates. None of the methods dominated in terms of power to detect a causal effect.

The modelling methods performed well in some scenarios, but less well in others. This is unsurprising, as in some scenarios, consistency assumptions for the methods were satisfied, and in others they were not. The MR-Egger method performed well in terms of Type 1 error rate in Scenarios 1 and 2, where the InSIDE assumption was satisfied. Estimates from the method were generally imprecise with low power. However, power in the MR-Egger method depends on the genetic associations with the exposure varying substantially between variants, which was not the case in the simulation study [38]. The contamination mixture method performed well with 30% and 50% valid instruments, with low bias and Type 1 error rates at or below 10% with 10 variants, 12% with 30 variants, and 20% with 100 variants. The MR-Mix method performed badly throughout, with highly inflated Type 1 error rates in almost all scenarios and comparatively low power to detect a causal effect. It performed slightly better with more genetic variants, although its performance was still worse than other methods. The MR-RAPS method performed well in Scenario 1, where its consistency assumption was satisfied, but less well in other scenarios with highly inflated Type 1 error rates.

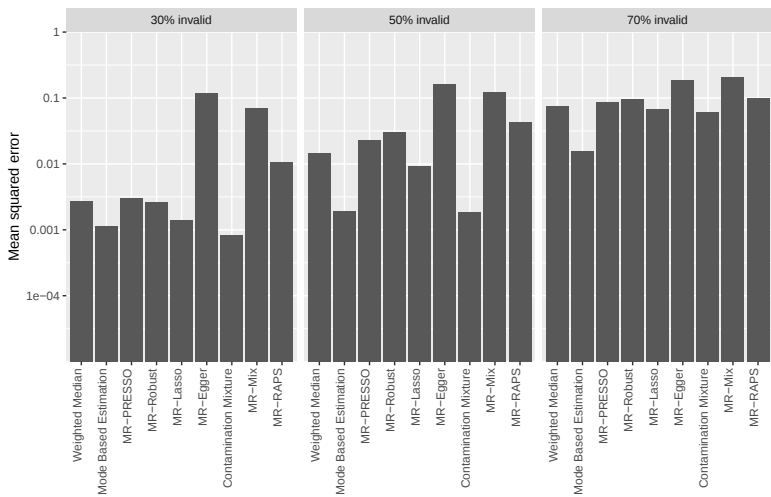


Fig. 3 Mean squared errors for the different methods in scenario 2 (directional pleiotropy, InSIDE satisfied) with a null causal effect for 30 variants. Note the vertical axis is on a logarithmic scale.

Table 1 Mean, median, standard deviation (SD) of estimates, and empirical power with 10 genetic variants.

Method	30% invalid				50% invalid				70% invalid			
	Mean	Median	SD	Power	Mean	Median	SD	Power	Mean	Median	SD	Power
Null causal effect: $\theta = 0$												
Scenario 1: Balanced pleiotropy, InSIDE satisfied												
Weighted Median	0.000	0.000	0.050	0.079	0.000	0.001	0.079	0.167	0.001	0.001	0.132	0.317
Mode Based Estimation	0.000	0.001	0.048	0.039	0.001	0.001	0.069	0.098	0.001	-0.001	0.119	0.271
MR-PRESSO	0.000	0.000	0.054	0.094	0.000	0.000	0.093	0.120	0.003	0.002	0.137	0.157
MR-Robust	-0.001	0.000	0.048	0.067	0.000	0.001	0.099	0.061	0.001	0.001	0.150	0.082
MR-Lasso	-0.001	-0.001	0.055	0.035	0.000	0.001	0.076	0.068	0.000	0.000	0.128	0.126
MR-Egger	-0.007	0.002	0.553	0.087	0.002	0.000	0.708	0.093	-0.015	-0.013	0.823	0.084
Contamination Mixture	0.000	0.000	0.042	0.061	0.000	0.001	0.069	0.088	0.002	0.001	0.214	0.152
MR-Mix	-0.002	-0.006	0.377	0.604	-0.003	-0.002	0.401	0.588	0.009	0.010	0.432	0.578
MR-RAPS	-0.001	-0.002	0.077	0.034	0.000	-0.001	0.120	0.054	0.002	0.002	0.156	0.076
Scenario 2: Directional pleiotropy, InSIDE satisfied												
Weighted Median	0.078	0.065	0.073	0.317	0.245	0.232	0.143	0.837	0.382	0.381	0.135	0.983
Mode Based Estimation	0.019	0.008	0.073	0.068	0.119	0.062	0.157	0.325	0.285	0.267	0.187	0.765
MR-PRESSO	0.086	0.074	0.071	0.232	0.253	0.244	0.126	0.539	0.386	0.384	0.120	0.841
MR-Robust	0.053	0.044	0.063	0.071	0.252	0.257	0.108	0.516	0.377	0.377	0.097	0.919
MR-Lasso	0.056	0.052	0.056	0.142	0.180	0.153	0.130	0.607	0.365	0.356	0.159	0.947
MR-Egger	0.000	-0.032	0.441	0.085	-0.007	-0.029	0.507	0.081	-0.005	-0.014	0.536	0.086
Contamination Mixture	0.006	0.005	0.043	0.055	0.053	0.016	0.150	0.101	0.361	0.378	0.282	0.452
MR-Mix	0.360	0.346	0.169	0.808	0.423	0.405	0.161	0.842	0.484	0.466	0.164	0.832
MR-RAPS	0.154	0.148	0.071	0.333	0.277	0.274	0.087	0.844	0.379	0.377	0.092	0.981
Scenario 3: Directional pleiotropy, InSIDE violated												
Weighted Median	0.037	0.034	0.049	0.138	0.104	0.087	0.092	0.455	0.225	0.213	0.134	0.813
Mode Based Estimation	0.009	0.005	0.051	0.046	0.042	0.028	0.081	0.141	0.136	0.101	0.140	0.455
MR-PRESSO	0.043	0.038	0.051	0.132	0.134	0.120	0.095	0.337	0.257	0.248	0.125	0.670
MR-Robust	0.040	0.036	0.051	0.070	0.148	0.145	0.089	0.244	0.259	0.257	0.101	0.654
MR-Lasso	0.037	0.035	0.048	0.097	0.090	0.079	0.080	0.315	0.221	0.196	0.146	0.719
MR-Egger	0.454	0.406	0.475	0.249	0.531	0.505	0.516	0.262	0.490	0.475	0.517	0.229
Contamination Mixture	0.007	0.006	0.043	0.064	0.031	0.015	0.096	0.104	0.206	0.107	0.249	0.319
MR-Mix	0.321	0.303	0.195	0.719	0.370	0.353	0.182	0.747	0.427	0.408	0.189	0.756
MR-RAPS	0.087	0.083	0.058	0.115	0.175	0.171	0.078	0.429	0.265	0.261	0.095	0.758
Positive causal effect: $\theta = +0.2$												
Scenario 1: Balanced pleiotropy, InSIDE satisfied												
Weighted Median	0.198	0.198	0.057	0.962	0.198	0.198	0.084	0.898	0.195	0.195	0.133	0.799
Mode Based Estimation	0.193	0.194	0.058	0.906	0.189	0.192	0.080	0.840	0.174	0.181	0.129	0.706
MR-PRESSO	0.199	0.199	0.058	0.947	0.198	0.198	0.092	0.829	0.194	0.194	0.137	0.633
MR-Robust	0.199	0.199	0.055	0.840	0.199	0.199	0.102	0.520	0.196	0.196	0.150	0.329
MR-Lasso	0.199	0.199	0.061	0.920	0.200	0.200	0.082	0.849	0.197	0.196	0.134	0.704
MR-Egger	0.174	0.172	0.559	0.120	0.172	0.178	0.709	0.101	0.165	0.167	0.826	0.094
Contamination Mixture	0.200	0.199	0.050	0.965	0.202	0.200	0.083	0.882	0.219	0.202	0.233	0.669
MR-Mix	0.278	0.268	0.368	0.579	0.304	0.296	0.373	0.578	0.321	0.320	0.401	0.601
MR-RAPS	0.201	0.200	0.079	0.692	0.200	0.200	0.119	0.461	0.197	0.197	0.157	0.315
Scenario 2: Directional pleiotropy, InSIDE satisfied												
Weighted Median	0.282	0.270	0.079	1.000	0.444	0.433	0.141	1.000	0.579	0.578	0.134	1.000
Mode Based Estimation	0.213	0.203	0.073	0.930	0.305	0.257	0.150	0.924	0.471	0.448	0.187	0.969
MR-PRESSO	0.283	0.275	0.072	0.981	0.442	0.434	0.119	0.917	0.578	0.575	0.122	0.968
MR-Robust	0.265	0.258	0.072	0.730	0.452	0.458	0.105	0.846	0.576	0.575	0.097	0.987
MR-Lasso	0.259	0.255	0.063	0.998	0.384	0.362	0.127	0.999	0.559	0.548	0.155	1.000
MR-Egger	0.167	0.144	0.442	0.122	0.168	0.141	0.521	0.110	0.171	0.160	0.538	0.109
Contamination Mixture	0.208	0.205	0.054	0.979	0.289	0.229	0.189	0.947	0.597	0.614	0.274	0.960
MR-Mix	0.584	0.566	0.178	0.874	0.645	0.628	0.163	0.881	0.699	0.681	0.161	0.855
MR-RAPS	0.356	0.351	0.074	1.000	0.477	0.475	0.086	1.000	0.579	0.577	0.094	1.000
Scenario 3: Directional pleiotropy, InSIDE violated												
Weighted Median	0.240	0.237	0.056	0.998	0.308	0.292	0.096	0.999	0.424	0.412	0.133	0.999
Mode Based Estimation	0.205	0.201	0.057	0.941	0.238	0.225	0.083	0.943	0.322	0.292	0.138	0.946
MR-PRESSO	0.243	0.239	0.054	0.997	0.330	0.319	0.094	0.969	0.448	0.437	0.123	0.966
MR-Robust	0.247	0.243	0.057	0.873	0.352	0.350	0.091	0.844	0.459	0.456	0.101	0.961
MR-Lasso	0.241	0.238	0.054	0.997	0.299	0.288	0.086	0.997	0.422	0.400	0.141	0.997
MR-Egger	0.633	0.582	0.496	0.350	0.708	0.682	0.511	0.375	0.675	0.658	0.528	0.335
Contamination Mixture	0.209	0.207	0.051	0.981	0.248	0.223	0.125	0.955	0.430	0.344	0.253	0.941
MR-Mix	0.547	0.526	0.210	0.828	0.604	0.583	0.189	0.833	0.653	0.637	0.184	0.816
MR-RAPS	0.288	0.284	0.060	0.996	0.377	0.374	0.081	0.997	0.466	0.461	0.095	0.998

Table 2 Mean, median, standard deviation (SD) of estimates, and empirical power with 30 genetic variants.

Method	30% invalid				50% invalid				70% invalid			
	Mean	Median	SD	Power	Mean	Median	SD	Power	Mean	Median	SD	Power
Null causal effect: $\theta = 0$												
Scenario 1: Balanced pleiotropy, InSIDE satisfied												
Weighted Median	0.000	0.000	0.032	0.058	0.000	0.000	0.044	0.112	0.000	0.000	0.068	0.208
Mode Based Estimation	0.000	0.000	0.035	0.014	0.000	0.001	0.038	0.033	0.000	0.000	0.058	0.111
MR-PRESSO	0.000	0.000	0.029	0.090	0.001	0.001	0.046	0.152	0.000	0.000	0.072	0.233
MR-Robust	0.000	0.000	0.030	0.047	0.001	0.001	0.059	0.034	-0.001	-0.002	0.098	0.052
MR-Lasso	0.000	0.000	0.029	0.045	0.000	0.001	0.041	0.072	0.000	0.000	0.067	0.123
MR-Egger	0.004	0.004	0.451	0.067	0.005	0.010	0.563	0.060	0.008	0.011	0.673	0.063
Contamination Mixture	0.000	0.000	0.028	0.067	0.000	0.001	0.038	0.094	0.000	0.000	0.071	0.153
MR-Mix	-0.004	-0.002	0.206	0.202	0.002	0.000	0.307	0.235	0.002	0.000	0.411	0.302
MR-RAPS	0.000	0.000	0.047	0.018	0.001	0.000	0.077	0.041	-0.001	-0.001	0.104	0.052
Scenario 2: Directional pleiotropy, InSIDE satisfied												
Weighted Median	0.085	0.082	0.042	0.653	0.293	0.289	0.106	0.994	0.452	0.452	0.089	1.000
Mode Based Estimation	0.006	0.005	0.034	0.019	0.053	0.037	0.082	0.120	0.302	0.285	0.189	0.742
MR-PRESSO	0.093	0.090	0.044	0.749	0.278	0.273	0.077	0.999	0.438	0.437	0.078	1.000
MR-Robust	0.049	0.045	0.041	0.044	0.289	0.294	0.075	0.691	0.433	0.433	0.059	0.999
MR-Lasso	0.037	0.036	0.030	0.292	0.183	0.171	0.084	0.963	0.419	0.417	0.106	1.000
MR-Egger	0.000	-0.021	0.343	0.058	0.000	-0.013	0.405	0.062	0.004	-0.010	0.432	0.065
Contamination Mixture	0.005	0.005	0.028	0.070	0.017	0.013	0.051	0.106	0.373	0.432	0.290	0.592
MR-Mix	0.254	0.236	0.192	0.455	0.384	0.367	0.208	0.548	0.513	0.511	0.190	0.634
MR-RAPS	0.175	0.174	0.047	0.950	0.321	0.320	0.056	1.000	0.435	0.434	0.058	1.000
Scenario 3: Directional pleiotropy, InSIDE violated												
Weighted Median	0.042	0.041	0.031	0.243	0.106	0.101	0.055	0.726	0.256	0.250	0.096	0.980
Mode Based Estimation	0.004	0.003	0.034	0.014	0.021	0.018	0.039	0.048	0.089	0.072	0.088	0.331
MR-PRESSO	0.045	0.044	0.032	0.335	0.140	0.136	0.056	0.893	0.281	0.277	0.082	0.997
MR-Robust	0.039	0.038	0.033	0.071	0.163	0.163	0.063	0.409	0.301	0.301	0.065	0.968
MR-Lasso	0.025	0.024	0.028	0.178	0.082	0.078	0.048	0.658	0.238	0.228	0.100	0.980
MR-Egger	0.838	0.817	0.375	0.754	0.919	0.904	0.365	0.789	0.845	0.830	0.359	0.719
Contamination Mixture	0.006	0.006	0.028	0.072	0.017	0.015	0.040	0.123	0.139	0.061	0.203	0.344
MR-Mix	0.205	0.184	0.166	0.360	0.299	0.283	0.178	0.436	0.409	0.395	0.191	0.516
MR-RAPS	0.097	0.096	0.037	0.499	0.202	0.201	0.052	0.963	0.307	0.306	0.061	0.999
Positive causal effect: $\theta = +0.2$												
Scenario 1: Balanced pleiotropy, InSIDE satisfied												
Weighted Median	0.196	0.195	0.036	1.000	0.195	0.194	0.048	0.991	0.197	0.196	0.073	0.942
Mode Based Estimation	0.189	0.189	0.041	0.976	0.187	0.187	0.044	0.964	0.180	0.181	0.067	0.853
MR-PRESSO	0.198	0.197	0.033	1.000	0.197	0.197	0.049	0.989	0.199	0.197	0.075	0.929
MR-Robust	0.198	0.197	0.035	0.995	0.197	0.197	0.061	0.824	0.199	0.198	0.098	0.518
MR-Lasso	0.198	0.198	0.033	1.000	0.197	0.197	0.045	0.996	0.199	0.199	0.073	0.939
MR-Egger	0.113	0.120	0.453	0.076	0.118	0.123	0.566	0.073	0.116	0.121	0.666	0.064
Contamination Mixture	0.199	0.199	0.033	1.000	0.199	0.198	0.043	0.993	0.206	0.201	0.084	0.933
MR-Mix	0.255	0.225	0.213	0.378	0.276	0.232	0.283	0.367	0.323	0.254	0.385	0.386
MR-RAPS	0.200	0.200	0.049	0.942	0.200	0.200	0.077	0.708	0.201	0.200	0.103	0.502
Scenario 2: Directional pleiotropy, InSIDE satisfied												
Weighted Median	0.293	0.290	0.048	1.000	0.493	0.488	0.104	1.000	0.646	0.647	0.091	1.000
Mode Based Estimation	0.198	0.196	0.040	0.987	0.246	0.232	0.081	0.940	0.472	0.434	0.188	0.938
MR-PRESSO	0.294	0.292	0.045	1.000	0.465	0.461	0.072	1.000	0.624	0.622	0.080	1.000
MR-Robust	0.265	0.262	0.049	0.941	0.495	0.500	0.073	0.958	0.632	0.632	0.061	1.000
MR-Lasso	0.243	0.242	0.036	1.000	0.396	0.386	0.084	1.000	0.614	0.610	0.104	1.000
MR-Egger	0.110	0.096	0.352	0.081	0.110	0.100	0.401	0.076	0.111	0.103	0.429	0.077
Contamination Mixture	0.206	0.204	0.034	1.000	0.233	0.218	0.093	0.999	0.648	0.707	0.270	0.999
MR-Mix	0.504	0.490	0.171	0.647	0.614	0.601	0.173	0.672	0.718	0.708	0.166	0.708
MR-RAPS	0.378	0.377	0.048	1.000	0.522	0.522	0.057	1.000	0.636	0.636	0.060	1.000
Scenario 3: Directional pleiotropy, InSIDE violated												
Weighted Median	0.242	0.241	0.036	1.000	0.312	0.307	0.058	1.000	0.453	0.448	0.095	1.000
Mode Based Estimation	0.195	0.193	0.039	0.994	0.213	0.211	0.043	0.995	0.277	0.263	0.086	0.966
MR-PRESSO	0.247	0.246	0.035	1.000	0.341	0.339	0.056	1.000	0.473	0.469	0.079	1.000
MR-Robust	0.246	0.245	0.038	0.997	0.370	0.370	0.062	0.982	0.500	0.500	0.066	0.999
MR-Lasso	0.228	0.227	0.032	1.000	0.294	0.290	0.054	1.000	0.441	0.433	0.095	1.000
MR-Egger	0.987	0.964	0.375	0.859	1.081	1.063	0.368	0.890	0.993	0.975	0.354	0.832
Contamination Mixture	0.207	0.206	0.034	1.000	0.225	0.221	0.054	0.999	0.399	0.302	0.233	0.997
MR-Mix	0.455	0.434	0.169	0.592	0.549	0.535	0.168	0.602	0.637	0.623	0.171	0.637
MR-RAPS	0.298	0.297	0.039	1.000	0.403	0.402	0.053	1.000	0.508	0.507	0.063	1.000

Table 3 Mean, median, standard deviation (SD) of estimates, and empirical power with 100 genetic variants.

Method	30% invalid				50% invalid				70% invalid			
	Mean	Median	SD	Power	Mean	Median	SD	Power	Mean	Median	SD	Power
Null causal effect: $\theta = 0$												
Scenario 1: Balanced pleiotropy, InSIDE satisfied												
Weighted Median	0.000	0.000	0.019	0.057	0.000	0.000	0.026	0.098	0.000	0.000	0.038	0.176
Mode Based Estimation	0.000	0.000	0.026	0.005	0.000	0.000	0.026	0.010	0.000	0.000	0.033	0.036
MR-PRESSO	0.000	0.000	0.017	0.092	0.000	0.000	0.027	0.151	0.000	0.000	0.041	0.229
MR-Robust	0.000	0.000	0.017	0.044	0.000	0.000	0.033	0.035	0.000	0.000	0.055	0.044
MR-Lasso	0.000	0.000	0.016	0.047	0.000	0.000	0.022	0.080	0.000	0.000	0.034	0.130
MR-Egger	0.000	0.000	0.257	0.054	0.000	0.003	0.327	0.055	-0.002	-0.001	0.385	0.055
Contamination Mixture	0.000	0.000	0.017	0.070	0.000	0.000	0.023	0.100	0.000	0.001	0.037	0.164
MR-Mix	0.000	0.000	0.062	0.042	0.000	0.000	0.068	0.035	0.000	0.000	0.085	0.037
MR-RAPS	0.000	0.000	0.027	0.018	0.000	0.000	0.044	0.035	0.000	0.000	0.059	0.046
Scenario 2: Directional pleiotropy, InSIDE satisfied												
Weighted Median	0.092	0.091	0.024	0.990	0.306	0.305	0.063	1.000	0.471	0.471	0.050	1.000
Mode Based Estimation	0.005	0.004	0.024	0.008	0.028	0.027	0.026	0.043	0.246	0.178	0.177	0.590
MR-PRESSO	0.095	0.094	0.024	0.995	0.269	0.268	0.040	1.000	0.437	0.436	0.044	1.000
MR-Robust	0.053	0.052	0.024	0.178	0.307	0.308	0.041	0.990	0.450	0.450	0.034	1.000
MR-Lasso	0.042	0.041	0.019	0.767	0.206	0.201	0.051	1.000	0.446	0.447	0.056	1.000
MR-Egger	-0.002	-0.002	0.195	0.049	-0.003	-0.005	0.228	0.052	0.002	0.003	0.244	0.054
Contamination Mixture	0.006	0.006	0.017	0.087	0.017	0.016	0.025	0.179	0.391	0.492	0.277	0.793
MR-Mix	0.083	0.030	0.163	0.134	0.235	0.098	0.272	0.273	0.468	0.532	0.282	0.452
MR-RAPS	0.183	0.182	0.027	1.000	0.335	0.335	0.032	1.000	0.454	0.453	0.034	1.000
Scenario 3: Directional pleiotropy, InSIDE violated												
Weighted Median	0.044	0.044	0.018	0.651	0.109	0.108	0.029	0.993	0.265	0.264	0.056	1.000
Mode Based Estimation	0.003	0.003	0.024	0.006	0.014	0.013	0.023	0.022	0.056	0.053	0.037	0.242
MR-PRESSO	0.048	0.048	0.018	0.801	0.140	0.139	0.030	1.000	0.280	0.279	0.044	1.000
MR-Robust	0.040	0.040	0.019	0.313	0.170	0.169	0.036	0.945	0.315	0.315	0.037	1.000
MR-Lasso	0.026	0.026	0.017	0.485	0.090	0.088	0.028	0.991	0.258	0.255	0.059	1.000
MR-Egger	0.902	0.902	0.193	0.998	0.980	0.980	0.185	1.000	0.882	0.881	0.181	0.998
Contamination Mixture	0.007	0.006	0.016	0.096	0.018	0.017	0.023	0.202	0.071	0.049	0.105	0.481
MR-Mix	0.064	0.035	0.119	0.122	0.163	0.080	0.207	0.202	0.325	0.315	0.262	0.323
MR-RAPS	0.100	0.099	0.021	0.993	0.211	0.210	0.030	1.000	0.322	0.322	0.035	1.000
Positive causal effect: $\theta = +0.2$												
Scenario 1: Balanced pleiotropy, InSIDE satisfied												
Weighted Median	0.192	0.192	0.022	1.000	0.193	0.192	0.029	1.000	0.193	0.193	0.041	0.999
Mode Based Estimation	0.180	0.180	0.031	0.999	0.180	0.180	0.030	0.999	0.175	0.175	0.038	0.986
MR-PRESSO	0.196	0.195	0.020	1.000	0.197	0.196	0.030	1.000	0.197	0.196	0.045	0.999
MR-Robust	0.196	0.196	0.020	1.000	0.196	0.196	0.035	0.998	0.197	0.197	0.057	0.925
MR-Lasso	0.196	0.196	0.019	1.000	0.196	0.197	0.025	1.000	0.196	0.196	0.038	1.000
MR-Egger	0.047	0.048	0.256	0.062	0.051	0.054	0.329	0.062	0.045	0.049	0.386	0.055
Contamination Mixture	0.196	0.196	0.020	1.000	0.198	0.198	0.027	1.000	0.199	0.198	0.043	0.999
MR-Mix	0.210	0.208	0.076	0.293	0.211	0.200	0.095	0.401	0.215	0.200	0.134	0.626
MR-RAPS	0.200	0.200	0.028	1.000	0.200	0.199	0.044	0.989	0.201	0.201	0.060	0.910
Scenario 2: Directional pleiotropy, InSIDE satisfied												
Weighted Median	0.301	0.300	0.029	1.000	0.505	0.503	0.060	1.000	0.662	0.662	0.052	1.000
Mode Based Estimation	0.189	0.187	0.028	1.000	0.217	0.215	0.034	0.984	0.425	0.370	0.168	0.892
MR-PRESSO	0.305	0.305	0.027	1.000	0.471	0.471	0.040	1.000	0.627	0.626	0.043	1.000
MR-Robust	0.274	0.273	0.031	1.000	0.512	0.513	0.039	1.000	0.646	0.646	0.035	1.000
MR-Lasso	0.251	0.251	0.023	1.000	0.421	0.418	0.051	1.000	0.636	0.636	0.056	1.000
MR-Egger	0.053	0.050	0.201	0.068	0.053	0.052	0.231	0.061	0.046	0.047	0.247	0.058
Contamination Mixture	0.206	0.205	0.022	1.000	0.226	0.223	0.037	1.000	0.702	0.761	0.217	1.000
MR-Mix	0.354	0.302	0.184	0.460	0.540	0.525	0.251	0.544	0.730	0.753	0.220	0.605
MR-RAPS	0.386	0.386	0.029	1.000	0.536	0.536	0.033	1.000	0.654	0.654	0.036	1.000
Scenario 3: Directional pleiotropy, InSIDE violated												
Weighted Median	0.244	0.243	0.022	1.000	0.316	0.315	0.034	1.000	0.461	0.459	0.055	1.000
Mode Based Estimation	0.187	0.186	0.028	1.000	0.201	0.199	0.028	1.000	0.244	0.240	0.044	0.997
MR-PRESSO	0.255	0.255	0.022	1.000	0.349	0.348	0.032	1.000	0.478	0.477	0.043	1.000
MR-Robust	0.249	0.249	0.023	1.000	0.377	0.377	0.037	1.000	0.511	0.512	0.038	1.000
MR-Lasso	0.229	0.229	0.020	1.000	0.302	0.301	0.032	1.000	0.458	0.456	0.056	1.000
MR-Egger	1.015	1.015	0.198	1.000	1.098	1.097	0.190	1.000	0.996	0.994	0.182	1.000
Contamination Mixture	0.207	0.207	0.020	1.000	0.224	0.223	0.030	1.000	0.340	0.284	0.164	1.000
MR-Mix	0.316	0.284	0.138	0.445	0.443	0.410	0.209	0.475	0.605	0.604	0.223	0.535
MR-RAPS	0.303	0.302	0.023	1.000	0.412	0.412	0.032	1.000	0.523	0.522	0.037	1.000

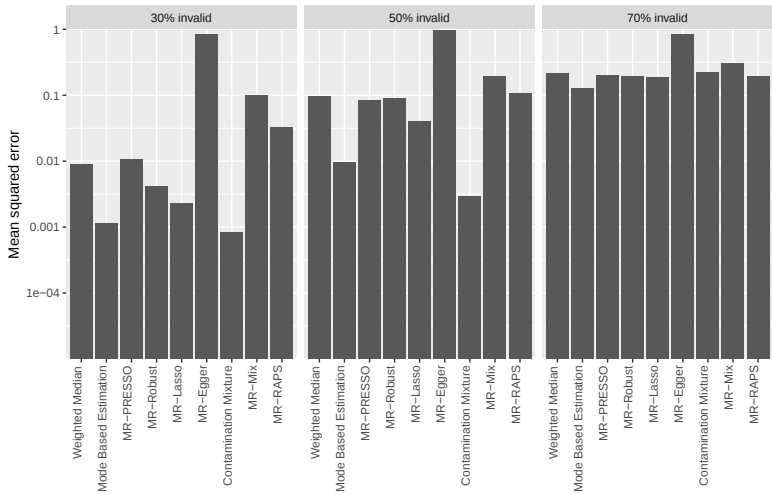


Fig. 4 Mean squared errors for the different methods in scenario 3 (directional pleiotropy, InSIDE violated) with a null causal effect for 30 variants. Note the vertical axis is on a logarithmic scale.

Empirical example: The effect of body mass index on coronary artery disease

Results from the empirical example are shown in Table 4. All methods agree that there is a positive effect of BMI on CAD risk, except for the MR-Mix method which gives a wide confidence interval that includes the null. The narrowest confidence intervals are for the outlier-robust methods (MR-Lasso, MR-Robust, MR-PRESSO), followed by the modelling methods except MR-Mix and MR-Egger (contamination mixture, MR-RAPS), then the consensus methods (weighted median, mode based estimation), and finally MR-Egger and MR-Mix.

Table 4 Estimates and 95% confidence intervals (CI) for the effect of BMI on coronary artery disease risk from robust methods. Estimates represent log odds ratios for CAD risk per 1 kg/m² increase in BMI.

Method	Causal estimate (95% CI)	CI width
Weighted Median	0.376 (0.206, 0.546)	0.340
Mode Based Estimation	0.382 (0.181, 0.583)	0.402
MR-PRESSO	0.410 (0.309, 0.511)	0.202
MR-Robust	0.425 (0.325, 0.526)	0.201
MR-Lasso	0.442 (0.354, 0.530)	0.176
MR-Egger	0.481 (0.165, 0.796)	0.631
(intercept)	-0.003 (-0.011, 0.005)	
Contamination Mixture	0.490 (0.372, 0.602)	0.230
MR-Mix	0.425 (-0.283, 1.133)	1.416
MR-RAPS	0.390 (0.308, 0.546)	0.238

While the methods that detect outliers varied in terms of how lenient or strictly they identified outliers, they agreed on the order of outliers (Supplementary Table 8). The MR-Robust method was the most lenient, downweighting two variants as outliers. Each subsequent method in order of strictness identified all previously identified variants as outliers. MR-PRESSO excluded the two variants identified by MR-Robust plus an additional three variants. MR-RAPS identified these five plus an additional two variants. MR-Lasso identified an additional three variants, 10 in total. The contamination mixture

method identified an additional 14 variants, 24 in total. MR-Mix identified an additional 21 variants, 45 in total. This suggests that any difference between results from outlier-robust methods are likely due to the strictness of outlier detection, rather than due to intrinsic differences in how the different methods select outliers. In several methods, the threshold at which outliers are detected can be varied by the analyst (for example, by varying the penalization parameter λ in MR-Lasso, or the significance threshold in MR-PRESSO). In practice, rather than performing different outlier-robust methods, it may be better to concentrate on one method, but vary this threshold.

Discussion

In this paper, we have provided a review of robust methods for MR, focusing on methods that can be performed using summary data and implemented using standard statistical software. We have divided methods into three categories: consensus methods, outlier-robust methods, and modelling methods. Methods were compared in three ways: by their theoretical properties, including the assumptions required for the method to give a consistent estimate, in an extensive simulation study, and in an empirical investigation. A summary table comparing the methods is presented as Table 5.

Table 5 Summary comparison of methods.

Method	Consistency assumption	Strengths and/or weaknesses
Weighted Median	Majority valid	Robust to outliers, sensitive to additional/removal of genetic variants, may be less efficient
Mode Based Estimation	Plurality valid	Robust to outliers, sensitive to bandwidth parameter and addition/removal of genetic variants, generally conservative
MR-PRESSO	Outlier-robust	Removes outliers, efficient with valid IVs, very high false positive rate with several invalid IVs
MR-Robust	Outlier-robust	Downweights outliers, efficient with valid IVs, high false positive rate with several invalid IVs
MR-Lasso	Outlier-robust	Removes outliers, efficient with valid IVs, high false positive rate with several invalid IVs
MR-Egger	InSIDE	Sensitive to outliers, sensitive to violations of InSIDE assumption, InSIDE assumption often not plausible, may be less efficient
Contamination Mixture	Plurality valid	Robust to outliers, sensitive to variance parameter and addition/removal of genetic variants, less conservative than MBE
MR-Mix	Plurality valid	Robust to outliers, requires large numbers of genetic variants, very high false positive rate in several scenarios
MR-RAPS	Pleiotropic effects (except outliers) normally distributed about zero	Downweights outliers, sensitive to violations of balanced pleiotropy assumption

While the use of robust methods for MR analyses with multiple genetic variants is highly recommended, it is not practical or desirable to perform and report results from every single robust method that has been proposed. Guidance is therefore needed as to which robust methods should be performed in practice. As an example, if an investigator performed the MR-PRESSO, MR-Robust, and MR-Lasso

methods, they would have assessed robustness of the result to outliers, but they would not have not assessed other potential violations of the IV assumptions. The categorization of methods proposed here is not the only possible division of methods, but we hope it is practically useful. For instance, the contamination mixture and MR-Mix methods make the same ‘plurality valid’ assumption as the MBE method, and so could have been placed in the same category.

The similarity and ubiquity of the ‘outlier-robust’ and ‘majority/plurality valid’ assumptions should encourage investigators to consider methods that make alternative assumptions, such as the MR-Egger method. While the InSIDE assumption is often not plausible [38], the MR-Egger method and the intercept test have value in providing a different route to testing the validity of an MR study. Another potential choice is the constrained IV method, which uses information on measured confounders to construct a composite IV that is not associated with these confounders [12]. This method was not considered in the simulation study, as it requires additional data on confounders and individual participant data. Further methods development is needed to develop robust methods for summary data that make different consistency assumptions.

We encourage researchers to perform robust methods from different categories, and that make varied consistency assumptions. For example, an investigator could perform the weighted median method (majority valid assumption), the MBE and/or contamination mixture methods (both plurality valid assumption, MBE is more conservative), and the MR-Egger method (InSIDE assumption). If there are a few clear outliers in the data, then an outlier-robust method such as MR-PRESSO or MR-Robust could also be performed. While we are hesitant to make a definitive recommendation as each method has its own strengths and weaknesses, this set of methods would be a reasonable compromise between performing too few methods and not adequately assessing the IV assumptions, and performing so many methods that clarity is obscured. Another danger of the use of large numbers of methods is the possibility to cherry-pick results, either by an investigator seeking to present their results in a more positive light, or a reader picking the one method that gives a different result (such as the MR-Mix method in our empirical example).

One important limitation of these methods is the assumption that all valid IVs estimate the same causal effect. Particularly for complex risk factors such as BMI, it is possible that different genetic variants have different ratio estimates not because they are invalid IVs, but because there are different ways of intervening on BMI that lead to different effects on the outcome. This can be remedied somewhat in methods based on the IVW method by using a random-effects model [19], or in the contamination mixture method, where causal effects evidenced by different sets of variants will lead to a multimodal likelihood function, and potentially a confidence interval that consists of more than one region.

In summary, while robust methods for MR do not provide a perfect solution to violations of the IV assumptions, they are able to detect such violations and help investigators make more reliable causal inferences. Investigators should perform a range of robust methods that operate in different ways and make different assumptions to assess the robustness of findings from a MR investigation.

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References

- [1] George Davey Smith and Shah Ebrahim. ‘Mendelian randomization’: can genetic epidemiology contribute to understanding environmental determinants of disease? *International Journal of Epidemiology*, 32(1):1–22, 2003.
- [2] George Davey Smith and Shah Ebrahim. Mendelian randomization: prospects, potentials, and limitations. *International Journal of Epidemiology*, 33(1):30–42, 2004.
- [3] Jie Zheng, A Mesut Erzurumluoglu, Benjamin L Elsworth, John P Kemp, Laurence Howe, Philip C Haycock, Gibran Hemani, Katherine Tansey, Charles Laurin, Early Genetics, Lifecourse Epidemiology (EAGLE) Eczema Consortium, Beate St Pourcain, Nicole M Warrington, Hilary K Finucane, Alkes L Price, Brendan K Bulik-Sullivan, Verner Anttila, Lavinia Paternoster, Tom R Gaunt, David M Evans, and Benjamin M Neale. LD Hub: a centralized database and web interface to perform ld score regression that maximizes the potential of summary level GWAS data for SNP heritability and genetic correlation analysis. *Bioinformatics*, 33(2):272–279, 2017.
- [4] Danielle Welter, Jacqueline MacArthur, Joannella Morales, Tony Burdett, Peggy Hall, Heather Junkins, Alan Klemm, Paul Flicek, Teri Manolio, Lucia Hindorff, and Helen Parkinson. The NHGRI GWAS catalog, a curated resource of SNP-trait associations. *Nucleic Acids Research*, 42(D1):D1001–D1006, 2014.
- [5] M Maria Glymour, Eric J Tchetgen Tchetgen, and James M Robins. Credible Mendelian randomization studies: approaches for evaluating the instrumental variable assumptions. *American Journal of Epidemiology*, 175(4):332–339, 2012.
- [6] Tyler J VanderWeele, Eric J Tchetgen Tchetgen, Marilyn Cornelis, and Peter Kraft. Methodological challenges in Mendelian randomization. *Epidemiology*, 25(3):427–435, 2014.
- [7] Stephanie von Hinke, George Davey Smith, Debbie A Lawlor, Carol Propper, and Frank Windmeijer. Genetic markers as instrumental variables. *Journal of Health Economics*, 45:131–148, 2016.
- [8] Shanya Sivakumaran, Felix Agakov, Evropi Theodoratou, James G Prendergast, Lina Zgaga, Teri Manolio, Igor Rudan, Paul McKeigue, James F Wilson, and Harry Campbell. Abundant

- pleiotropy in human complex diseases and traits. *The American Journal of Human Genetics*, 89(5):607–618, 2011.
- [9] Nadia Solovieff, Chris Cotsapas, Phil H Lee, Shaun M Purcell, and Jordan W Smoller. Pleiotropy in complex traits: challenges and strategies. *Nature Reviews Genetics*, 14(7):483–495, 2013.
- [10] Hyunseung Kang, Anru Zhang, T Tony Cai, and Dylan S Small. Instrumental variables estimation with some invalid instruments and its application to Mendelian randomization. *Journal of the American Statistical Association*, 111(513):132–144, 2016.
- [11] Zijian Guo, Hyunseung Kang, T. Tony Cai, and Dylan S. Small. Confidence intervals for causal effects with invalid instruments by using two-stage hard thresholding with voting. *Journal of the Royal Statistical Society: Series B (Statistical Methodology)*, 80(4):793–815, 2018.
- [12] Lai Jiang, Karim Oualkacha, Vanessa Didelez, Antonio Ciampi, Pedro Rosa, Andrea L. Benedet, Sulantha S. Mathotaarachchi, Brent Richards, and Celia M.T. Greenwood. Constrained instruments and their application to mendelian randomization with pleiotropy. *bioRxiv* 22754. <https://doi.org/10.1101/227454>, 2017.
- [13] Eric J. Tchetgen Tchetgen, BaoLuo Sun, and Stefan Walter. The GENIUS Approach to Robust Mendelian Randomization Inference. *arXiv:1709.07779 [stat.ME]*, 2017.
- [14] Luke J. O’Connor and Alkes L. Price. Distinguishing genetic correlation from causation across 52 diseases and complex traits. *Nature Genetics*, 50(12):1728–1734, 2018.
- [15] Thomas A. DiPrete, Casper A. P. Burik, and Philipp D. Koellinger. Genetic instrumental variable regression: Explaining socioeconomic and health outcomes in nonexperimental data. *Proceedings of the National Academy of Sciences*, 115(22):E4970–E4979, 2018.
- [16] Carlo Berzuini, Hui Guo, Stephen Burgess, and Luisa Bernardinelli. A bayesian approach to mendelian randomization with multiple pleiotropic variants. *Biostatistics*, 2018.
- [17] Stephen Burgess, Verena Zuber, Apostolos Gkatzionis, and Christopher N Foley. Modal-based estimation via heterogeneity-penalized weighting: model averaging for consistent and efficient estimation in Mendelian randomization when a plurality of candidate instruments are valid. *International Journal of Epidemiology*, 47(4):1242–1254, 2018.
- [18] TM Palmer, JR Thompson, MD Tobin, NA Sheehan, and PR Burton. Adjusting for bias and unmeasured confounding in Mendelian randomization studies with binary responses. *International Journal of Epidemiology*, 37(5):1161–1168, 2008.
- [19] Jack Bowden, Fabiola Del Greco M, Cosetta Minelli, George Davey Smith, Nuala Sheehan, and John Thompson. A framework for the investigation of pleiotropy in two-sample summary data Mendelian randomization. *Statistics in Medicine*, 36(11):1783–1802, 2017.
- [20] V Didelez and N Sheehan. Mendelian randomization as an instrumental variable approach to causal inference. *Statistical Methods in Medical Research*, 16(4):309–330, 2007.

- [21] D.C. Thomas, D.A. Lawlor, and J.R. Thompson. Re: Estimation of bias in nongenetic observational studies using “Mendelian triangulation” by Bautista et al. *Annals of Epidemiology*, 17(7):511–513, 2007.
- [22] S. Burgess, A S Butterworth, and S G Thompson. Mendelian randomization analysis with multiple genetic variants using summarized data. *Genetic Epidemiology*, 37(7):658–665, 2013.
- [23] S Burgess, F Dudbridge, and S G Thompson. Combining information on multiple instrumental variables in Mendelian randomization: comparison of allele score and summarized data methods. *Statistics in Medicine*, 35(11):1880–1906, 2016.
- [24] Jack Bowden, George Davey Smith, Philip C Haycock, and Stephen Burgess. Consistent estimation in Mendelian randomization with some invalid instruments using a weighted median estimator. *Genetic Epidemiology*, 40(4):304–314, 2016.
- [25] Fernando P Hartwig, George Davey Smith, and Jack Bowden. Robust inference in summary data Mendelian randomization via the zero modal pleiotropy assumption. *International Journal of Epidemiology*, 46(6):1985–1998, 2017.
- [26] Marie Verbanck, Chia-Yen Chen, Benjamin Neale, and Ron Do. Detection of widespread horizontal pleiotropy in causal relationships inferred from Mendelian randomization between complex traits and diseases. *Nature Genetics*, 50(5):693–698, 2018.
- [27] Stephen Burgess, Jack Bowden, Frank Dudbridge, and Simon G Thompson. Robust instrumental variable methods using multiple candidate instruments with application to Mendelian randomization. *arXiv:1606.03729*, 2016.
- [28] Frederick Mosteller and John Wilder Tukey. *Data analysis and regression: a second course in statistics*. Addison-Wesley Series in Behavioral Science: Quantitative Methods, 1977.
- [29] Frank Windmeijer, Helmut Farbmacher, Neil Davies, and George Davey Smith. On the use of the lasso for instrumental variables estimation with some invalid instruments. Technical Report Discussion Paper 16/674, University of Bristol, 2016.
- [30] Stephen Burgess, Christopher N Foley, Elias Allara, James R Staley, and Joanna MM Howson. A robust and efficient method for mendelian randomization with hundreds of genetic variants: unravelling mechanisms linking hdl-cholesterol and coronary heart disease. *bioRxiv*, 2019.
- [31] Guanghao Qi and Nilanjan Chatterjee. Mendelian randomization analysis using mixture models (mrmix) for genetic effect-size-distribution leads to robust estimation of causal effects. *bioRxiv* 367821. <https://doi.org/10.1101/367821>, 2018.
- [32] Qingyuan Zhao, Jingshu Wang, Jack Bowden, and Dylan S Small. Statistical inference in two-sample summary-data Mendelian randomization using robust adjusted profile score. *arXiv:1801.09652*, 2018.

- [33] Philip C Haycock, Stephen Burgess, Kaitlin H Wade, Jack Bowden, Caroline Relton, and George Davey Smith. Best (but oft-forgotten) practices: the design, analysis, and interpretation of Mendelian randomization studies. *The American Journal of Clinical Nutrition*, 103(4):965–978, 2016.
- [34] Brandon L. Pierce and Stephen Burgess. Efficient design for Mendelian randomization studies: Subsample and 2-sample instrumental variable estimators. *American Journal of Epidemiology*, 178(7):1177–1184, 2013.
- [35] Claire Monnereau, Suzanne Vogelezang, Claudia J Kruithof, Vincent WV Jaddoe, and Janine F Felix. Associations of genetic risk scores based on adult adiposity pathways with childhood growth and adiposity measures. *BMC Genetics*, 17(1):120, 2016.
- [36] Adam E Locke, Bratati Kahali, Sonja I Berndt, Anne E Justice, Tune H Pers, Felix R Day, Corey Powell, Sailaja Vedantam, Martin L Buchkovich, Jian Yang, et al. Genetic studies of body mass index yield new insights for obesity biology. *Nature*, 518(7538):197–206, 2015.
- [37] Majid Nikpay, Anuj Goel, Hong-Hee Won, Leanne M Hall, Christina Willenborg, Stavroula Kanoni, Danish Saleheen, Theodosios Kyriakou, Christopher P Nelson, Jemma C Hopewell, et al. A comprehensive 1000 genomes–based genome-wide association meta-analysis of coronary artery disease. *Nature Genetics*, 47(10):1121–1130, 2015.
- [38] Stephen Burgess and Simon G Thompson. Interpreting findings from Mendelian randomization using the MR-Egger method. *European Journal of Epidemiology*, 2017. Available online before print. doi: 10.1007/s10654-017-0255-x.

Supporting Information

S1 Details of simulation study

For each participant i , we simulate data on J genetic variants $G_{i1}, G_{i2}, \dots, G_{iJ}$, a modifiable exposure X_i , an outcome variable Y_i , and a confounder U_i (assumed unknown). The confounder is a linear function of the genetic variants and an independent error term ε_i^U . The effect of variant j on the confounder is represented by coefficient ϕ_j . The exposure is linear in the genetic variants, the confounder and an independent error term ε_i^X . The effect of variant j on the exposure is represented by coefficient γ_j (this is zero for a valid IV). The outcome is linear in the genetic variants, exposure, confounders and an independent error term ε_i^Y . The effect of variant j on the outcome is represented by coefficient α_j (again, this is zero for a valid IV). The effect of the exposure on the outcome is represented by θ . The genetic variants are modelled as single nucleotide polymorphisms (SNPs) with minor allele frequency 30%, and take values 0, 1 or 2. The error terms ε_i^U , ε_i^X and ε_i^Y each follow an independent normal distribution with mean 0 and unit variance.

We can represent the model mathematically as:

$$U_i = \sum_{j=1}^J \phi_j G_{ij} + \varepsilon_i^U, \quad (8)$$

$$X_i = \sum_{j=1}^J \gamma_j G_{ij} + U_i + \varepsilon_i^X, \quad (9)$$

$$Y_i = \sum_{j=1}^J \alpha_j G_{ij} + \theta X_i + U_i + \varepsilon_i^Y, \quad (10)$$

$$G_{ij} \sim \text{Binomial}(2, 0.3) \text{ independently,} \quad (11)$$

$$\varepsilon_i^U, \varepsilon_i^X, \varepsilon_i^Y \sim \mathcal{N}(0, 1) \text{ independently.} \quad (12)$$

The causal effect of the exposure on the outcome was either taken as null ($\theta = 0$) or positive ($\theta = 0.2$). Genetic associations with the exposure γ_j are drawn from a truncated normal distribution. Parameters are chosen such that the total proportion of variance explained in the exposure by direct effects of the genetic variants is 10%. In scenario 3, the overall proportion of variance explained in the exposure by genetic variants is slightly larger, as there is an additional effect of the invalid IVs on the exposure via their effect on the confounder.

For valid IVs, $\phi_j = 0$ and $\alpha_j = 0$. For invalid IVs, in scenario 1 (balanced pleiotropy, InSIDE satisfied), the effects of the genetic variants on the outcome are generated from a normal distribution centered at zero ($\alpha_j \sim \mathcal{N}(0, 0.15)$) and genetic effects on the confounder are zero ($\phi_j = 0$). In scenario 2 (directional pleiotropy, InSIDE satisfied), the effects of the genetic variants on the outcome are generated from a normal distribution centered away from zero ($\alpha_j \sim \mathcal{N}(0.1, 0.075)$) and genetic effects on the confounder are zero ($\phi_j = 0$). In scenario 3 (directional pleiotropy, InSIDE violated), the direct effects of the genetic variants on the outcome are generated from a normal distribution centered away from zero ($\alpha_j \sim \mathcal{N}(0.1, 0.075)$) and genetic effects on the confounder are generated from a

uniform distribution ($\phi_j \sim \mathcal{U}(0, 0.1)$).

Summary genetic association data are calculated by regression of the risk factor on each genetic variant in turn, and the outcome on each genetic variant in turn. Individual participant data are generated for 20 000 individuals: the exposure regressions are performed on the first 10 000 individuals, and the outcome regressions on the remaining 10 000 individuals. This represents a two-sample Mendelian randomization study. We generated 10 000 simulated datasets for each scenario, and for null and positive causal effects.

Each method is performed using the default options suggested by the authors of the method, either in the corresponding publication, or in the software code recommended by the authors. The weighted median method is performed using inverse-variance weights. The mode based estimation method is performed using inverse-variance weights, the ‘no measurement error’ assumption, and the default bandwidth setting ($\phi = 1$). The MR-PRESSO method is performed using a significance cut-off of $p < 0.05$ for determining outliers. The MR-Lasso method is performed using the heterogeneity criterion for selecting the lasso penalty parameter. The contamination mixture method is performed using the standard deviation of the ratio estimates multiplied by 1.5 for the variance parameter. For MR-Mix, we choose an initial value of the probability mass at the null component as 0.6 and the initial value of the variance of the non-null component as 1×10^{-5} . As the method performs a grid search, these decisions should not influence the results. For MR-RAPS, we use the overdispersed robust version with the Huber loss function. All regression models use random-effects.

The mean squared errors of the different methods are presented in Supplementary Figure 5 (10 variants, scenario 2), Supplementary Figure 6 (10 variants, scenario 3), Supplementary Figure 7 (100 variants, scenario 2), and Supplementary Figure 8 (100 variants, scenario 3). Note that in each case the vertical axis is on a logarithmic scale. Findings are similar to before among the different scenarios. We observe again that the performance of the mode based estimator is the best for the consensus based approach, MR-Robust gets the best result among the outlier-robust methods, and the contamination mixture approach has the best performance among the modelling methods.

In addition to the scenarios presented in the main paper, we also performed simulations with 500 genetic variants, and a wider range of proportions of invalid IVs (1%, 5%, and 10%). Due to computational burden, only 1000 simulated datasets were generated in each case.

The results of these additional simulations can be found in Table 6 (invalidness 1%, 5%, and 10%), and 7 (invalidness 30%, 50% and 70%).

With few invalid IVs, most methods had reasonable behaviour. An exception was the MR-Egger method, which had inflated Type 1 error rates in Scenario 3 even with only 1% of variants invalid. Although we would expect the outlier-removal methods to behave best with few invalid IVs, in fact most methods have some mechanism for providing robustness to outliers, and so it was difficult to differentiate between the methods.

Table 6 Mean, median, standard deviation (SD) of estimates, and empirical power (%) for scenarios with 500 genetic variants.

Method	1% invalid				5% invalid				10% invalid			
	Mean	Median	SD	Power	Mean	Median	SD	Power	Mean	Median	SD	Power
Null causal effect: $\beta = 0$												
Scenario 1: Balanced pleiotropy, InSIDE satisfied												
Weighted Median	0.000	0.000	0.006	0.038	0.000	0.000	0.006	0.014	0.000	0.000	0.007	0.034
Mode Based Estimation	0.000	-0.001	0.017	0.001	0.000	0.000	0.017	0.003	0.000	0.000	0.018	0.004
MR-PRESSO	0.000	0.000	0.005	0.049	0.000	0.000	0.005	0.046	0.000	0.000	0.006	0.059
MR-Robust	0.000	0.000	0.005	0.048	0.000	0.000	0.005	0.046	0.000	0.000	0.006	0.046
MR-Lasso	0.000	0.000	0.005	0.023	0.000	0.000	0.005	0.027	0.000	0.000	0.006	0.037
MR-Egger	0.000	0.000	0.023	0.049	-0.001	0.000	0.036	0.047	0.000	0.000	0.047	0.054
Contamination Mixture	0.000	0.000	0.005	0.055	0.000	0.000	0.006	0.050	0.000	0.001	0.006	0.052
MR-Mix	0.000	0.000	0.023	0.007	-0.001	0.000	0.020	0.004	-0.001	0.000	0.021	0.005
MR-RAPS	0.000	0.000	0.005	0.047	0.000	0.000	0.006	0.016	0.000	0.000	0.007	0.020
Scenario 2: Directional pleiotropy, InSIDE satisfied												
Weighted Median	0.001	0.001	0.006	0.021	0.005	0.005	0.006	0.073	0.011	0.011	0.007	0.289
Mode Based Estimation	0.000	0.000	0.016	0.003	0.000	0.000	0.017	0.004	-0.001	0.000	0.037	0.000
MR-PRESSO	0.001	0.001	0.005	0.049	0.004	0.004	0.005	0.114	0.010	0.010	0.006	0.392
MR-Robust	0.000	0.000	0.005	0.041	0.002	0.002	0.005	0.072	0.006	0.006	0.006	0.152
MR-Lasso	0.001	0.001	0.005	0.046	0.003	0.003	0.005	0.078	0.005	0.006	0.006	0.184
MR-Egger	0.000	0.000	0.022	0.052	0.000	0.000	0.032	0.052	0.001	0.001	0.039	0.049
Contamination Mixture	0.000	0.000	0.005	0.048	0.001	0.001	0.006	0.061	0.002	0.002	0.006	0.067
MR-Mix	0.000	0.000	0.022	0.007	0.003	0.001	0.019	0.002	0.003	0.001	0.021	0.007
MR-RAPS	0.001	0.001	0.005	0.050	0.008	0.008	0.006	0.239	0.020	0.020	0.006	0.783
Scenario 3: Directional pleiotropy, InSIDE violated												
Weighted Median	0.002	0.002	0.006	0.030	0.010	0.010	0.007	0.232	0.022	0.022	0.007	0.778
Mode Based Estimation	0.000	0.000	0.016	0.004	0.000	0.001	0.018	0.004	0.001	0.001	0.017	0.003
MR-PRESSO	0.001	0.000	0.005	0.059	0.004	0.004	0.005	0.130	0.013	0.013	0.007	0.582
MR-Robust	0.000	0.000	0.005	0.048	0.002	0.002	0.005	0.064	0.006	0.006	0.006	0.148
MR-Lasso	0.001	0.001	0.005	0.058	0.003	0.003	0.005	0.102	0.007	0.007	0.006	0.230
MR-Egger	0.031	0.030	0.027	0.269	0.140	0.139	0.043	0.956	0.250	0.249	0.051	0.999
Contamination Mixture	0.000	0.000	0.005	0.043	0.001	0.001	0.006	0.058	0.002	0.002	0.006	0.059
MR-Mix	0.002	0.000	0.018	0.007	0.004	0.001	0.018	0.003	0.005	0.000	0.018	0.002
MR-RAPS	0.002	0.002	0.005	0.063	0.014	0.014	0.006	0.487	0.035	0.035	0.006	0.998
Positive causal effect: $\theta = +0.2$												
Scenario 1: Balanced pleiotropy, InSIDE satisfied												
Weighted Median	0.179	0.179	0.008	1.000	0.179	0.179	0.008	1.000	0.180	0.179	0.009	1.000
Mode Based Estimation	0.154	0.154	0.020	0.989	0.153	0.153	0.021	0.992	0.154	0.153	0.020	0.995
MR-PRESSO	0.188	0.188	0.007	1.000	0.188	0.188	0.007	1.000	0.189	0.189	0.008	1.000
MR-Robust	0.188	0.188	0.007	1.000	0.188	0.188	0.007	1.000	0.189	0.189	0.007	1.000
MR-Lasso	0.188	0.188	0.007	1.000	0.188	0.188	0.007	1.000	0.189	0.188	0.008	1.000
MR-Egger	0.010	0.009	0.027	0.073	0.009	0.009	0.038	0.057	0.010	0.009	0.049	0.055
Contamination Mixture	0.187	0.187	0.007	1.000	0.187	0.187	0.008	1.000	0.187	0.187	0.008	1.000
MR-Mix	0.060	0.055	0.034	0.076	0.057	0.052	0.033	0.012	0.048	0.045	0.033	0.002
MR-RAPS	0.200	0.200	0.007	1.000	0.199	0.199	0.008	1.000	0.200	0.199	0.008	1.000
Scenario 2: Directional pleiotropy, InSIDE satisfied												
Weighted Median	0.181	0.181	0.008	1.000	0.186	0.186	0.009	1.000	0.193	0.193	0.009	1.000
Mode Based Estimation	0.155	0.154	0.020	0.993	0.155	0.155	0.022	0.992	0.156	0.156	0.021	0.991
MR-PRESSO	0.190	0.190	0.007	1.000	0.195	0.195	0.007	1.000	0.204	0.203	0.008	1.000
MR-Robust	0.189	0.189	0.007	1.000	0.192	0.192	0.007	1.000	0.198	0.197	0.008	1.000
MR-Lasso	0.189	0.189	0.007	1.000	0.192	0.192	0.007	1.000	0.196	0.196	0.008	1.000
MR-Egger	0.010	0.010	0.026	0.070	0.010	0.010	0.034	0.062	0.010	0.010	0.042	0.062
Contamination Mixture	0.187	0.187	0.008	1.000	0.188	0.189	0.008	1.000	0.189	0.190	0.009	1.000
MR-Mix	0.063	0.061	0.035	0.061	0.066	0.064	0.033	0.018	0.064	0.060	0.034	0.017
MR-RAPS	0.202	0.202	0.007	1.000	0.210	0.210	0.008	1.000	0.222	0.222	0.009	1.000
Scenario 3: Directional pleiotropy, InSIDE violated												
Weighted Median	0.182	0.182	0.008	1.000	0.191	0.191	0.009	1.000	0.206	0.207	0.010	1.000
Mode Based Estimation	0.154	0.153	0.020	0.997	0.154	0.154	0.022	0.987	0.152	0.155	0.057	0.988
MR-PRESSO	0.190	0.189	0.007	1.000	0.195	0.195	0.008	1.000	0.211	0.210	0.009	1.000
MR-Robust	0.189	0.189	0.007	1.000	0.192	0.191	0.007	1.000	0.199	0.199	0.008	1.000
MR-Lasso	0.189	0.189	0.007	1.000	0.192	0.192	0.008	1.000	0.199	0.199	0.009	1.000
MR-Egger	0.044	0.043	0.031	0.360	0.161	0.160	0.046	0.965	0.278	0.277	0.054	1.000
Contamination Mixture	0.187	0.187	0.008	1.000	0.188	0.188	0.008	1.000	0.190	0.190	0.009	1.000
MR-Mix	0.060	0.058	0.033	0.057	0.053	0.049	0.031	0.016	0.043	0.040	0.028	0.008
MR-RAPS	0.203	0.202	0.007	1.000	0.215	0.215	0.008	1.000	0.240	0.240	0.009	1.000

Table 7 Mean, median, standard deviation (SD) of estimates, and empirical power (%) for scenarios with 500 genetic variants.

Method	30% invalid				50% invalid				70% invalid			
	Mean	Median	SD	Power	Mean	Median	SD	Power	Mean	Median	SD	Power
Null causal effect: $\theta = 0$												
Scenario 1: Balanced pleiotropy, InSIDE satisfied												
Weighted Median	0.000	-0.001	0.010	0.054	-0.001	0.000	0.015	0.099	0.000	-0.001	0.021	0.164
Mode Based Estimation	0.003	-0.001	0.101	0.005	-0.002	-0.001	0.020	0.004	-0.004	-0.001	0.112	0.008
MR-PRESSO	0.000	0.000	0.010	0.080	-0.001	0.000	0.016	0.150	0.000	0.000	0.023	0.187
MR-Robust	0.000	-0.001	0.009	0.043	0.000	0.000	0.018	0.059	-0.001	-0.001	0.026	0.048
MR-Lasso	0.000	0.000	0.009	0.058	-0.001	-0.001	0.013	0.085	0.000	-0.001	0.019	0.136
MR-Egger	-0.001	0.000	0.076	0.049	-0.001	-0.001	0.097	0.053	0.001	0.000	0.115	0.052
Contamination Mixture	0.000	0.000	0.010	0.085	-0.001	-0.002	0.015	0.131	0.000	0.000	0.025	0.197
MR-Mix	0.001	0.000	0.016	0.002	0.000	0.000	0.020	0.000	0.000	0.000	0.015	0.000
MR-RAPS	0.000	0.000	0.013	0.018	0.000	0.001	0.021	0.040	-0.001	-0.001	0.028	0.050
Scenario 2: Directional pleiotropy, InSIDE satisfied												
Weighted Median	0.105	0.105	0.014	1.000	0.294	0.292	0.027	1.000	0.440	0.440	0.025	1.000
Mode Based Estimation	0.006	0.006	0.016	0.002	0.035	0.035	0.021	0.052	0.207	0.183	0.100	0.762
MR-Robust	0.086	0.086	0.014	0.999	0.313	0.313	0.018	1.000	0.438	0.438	0.018	1.000
MR-Lasso	0.061	0.061	0.011	1.000	0.233	0.233	0.023	1.000	0.426	0.426	0.025	1.000
MR-PRESSO	0.121	0.120	0.014	1.000	0.285	0.284	0.019	1.000	0.424	0.424	0.020	1.000
MR-Egger	0.000	0.000	0.059	0.050	0.000	0.000	0.069	0.052	0.000	0.000	0.072	0.044
Contamination Mixture	0.011	0.011	0.010	0.254	0.042	0.041	0.020	0.797	0.502	0.527	0.120	1.000
MR-Mix	0.004	0.000	0.011	0.000	0.004	0.000	0.011	0.001	0.012	0.000	0.020	0.000
MR-RAPS	0.188	0.188	0.014	1.000	0.341	0.340	0.017	1.000	0.460	0.460	0.019	1.000
Scenario 3: Directional pleiotropy, InSIDE violated												
Weighted Median	0.048	0.048	0.010	1.000	0.121	0.120	0.016	1.000	0.256	0.257	0.024	1.000
Mode Based Estimation	0.003	0.003	0.016	0.004	0.016	0.015	0.016	0.018	0.054	0.055	0.022	0.313
MR-PRESSO	0.063	0.062	0.010	1.000	0.161	0.160	0.015	1.000	0.284	0.285	0.019	1.000
MR-Robust	0.051	0.050	0.010	0.999	0.180	0.180	0.016	1.000	0.309	0.309	0.017	1.000
MR-Lasso	0.033	0.033	0.009	0.990	0.109	0.109	0.015	1.000	0.260	0.260	0.024	1.000
MR-Egger	0.485	0.485	0.061	1.000	0.537	0.537	0.061	1.000	0.473	0.473	0.062	1.000
Contamination Mixture	0.010	0.010	0.009	0.257	0.030	0.030	0.014	0.738	0.095	0.092	0.036	0.993
MR-Mix	0.003	0.000	0.020	0.003	0.006	0.000	0.026	0.003	0.013	0.000	0.038	0.004
MR-RAPS	0.103	0.103	0.011	1.000	0.216	0.215	0.015	1.000	0.328	0.328	0.017	1.000
Positive causal effect: $\theta = +0.2$												
Scenario 1: Balanced pleiotropy, InSIDE satisfied												
Weighted Median	0.180	0.179	0.013	1.000	0.179	0.179	0.016	1.000	0.180	0.179	0.023	1.000
Mode Based Estimation	0.154	0.154	0.022	0.987	0.152	0.151	0.023	0.983	0.146	0.145	0.029	0.978
MR-PRESSO	0.189	0.189	0.012	1.000	0.188	0.188	0.018	1.000	0.190	0.189	0.025	1.000
MR-Robust	0.189	0.189	0.011	1.000	0.188	0.188	0.018	1.000	0.189	0.188	0.026	1.000
MR-Lasso	0.189	0.189	0.011	1.000	0.188	0.188	0.015	1.000	0.190	0.189	0.022	1.000
MR-Egger	0.010	0.010	0.078	0.056	0.011	0.009	0.098	0.056	0.011	0.012	0.115	0.051
Contamination Mixture	0.188	0.188	0.012	1.000	0.190	0.190	0.017	1.000	0.196	0.194	0.028	1.000
MR-Mix	0.021	0.000	0.028	0.001	0.006	0.000	0.019	0.000	0.002	0.000	0.015	0.000
MR-RAPS	0.200	0.200	0.014	1.000	0.200	0.200	0.022	1.000	0.201	0.201	0.028	1.000
Scenario 2: Directional pleiotropy, InSIDE satisfied												
Weighted Median	0.307	0.307	0.018	1.000	0.488	0.487	0.026	1.000	0.619	0.618	0.026	1.000
Mode Based Estimation	0.164	0.163	0.022	0.986	0.201	0.202	0.105	0.971	0.395	0.384	0.097	0.983
MR-PRESSO	0.343	0.343	0.017	1.000	0.500	0.501	0.019	1.000	0.622	0.623	0.020	1.000
MR-Robust	0.314	0.314	0.018	1.000	0.510	0.510	0.018	1.000	0.627	0.627	0.019	1.000
MR-Lasso	0.273	0.273	0.015	1.000	0.449	0.448	0.024	1.000	0.613	0.614	0.025	1.000
MR-Egger	0.010	0.010	0.061	0.051	0.010	0.009	0.071	0.053	0.010	0.010	0.076	0.056
Contamination Mixture	0.207	0.207	0.015	1.000	0.288	0.280	0.051	1.000	0.717	0.721	0.081	1.000
MR-Mix	0.022	0.019	0.022	0.000	0.020	0.012	0.025	0.007	0.039	0.030	0.037	0.011
MR-RAPS	0.394	0.394	0.016	1.000	0.545	0.544	0.018	1.000	0.662	0.662	0.020	1.000
Scenario 3: Directional pleiotropy, InSIDE violated												
Weighted Median	0.238	0.238	0.013	1.000	0.319	0.319	0.019	1.000	0.445	0.445	0.025	1.000
Mode Based Estimation	0.156	0.161	0.137	0.987	0.179	0.176	0.053	0.990	0.223	0.221	0.032	0.990
MR-PRESSO	0.273	0.273	0.013	1.000	0.375	0.374	0.018	1.000	0.490	0.489	0.020	1.000
MR-Robust	0.259	0.259	0.013	1.000	0.382	0.382	0.018	1.000	0.500	0.500	0.019	1.000
MR-Lasso	0.234	0.234	0.012	1.000	0.321	0.321	0.018	1.000	0.461	0.461	0.025	1.000
MR-Egger	0.535	0.535	0.064	1.000	0.592	0.591	0.065	1.000	0.527	0.526	0.066	1.000
Contamination Mixture	0.204	0.204	0.012	1.000	0.238	0.237	0.020	1.000	0.386	0.368	0.087	1.000
MR-Mix	0.039	0.036	0.034	0.009	0.034	0.016	0.048	0.011	0.052	0.027	0.069	0.036
MR-RAPS	0.308	0.308	0.013	1.000	0.419	0.418	0.018	1.000	0.530	0.531	0.020	1.000

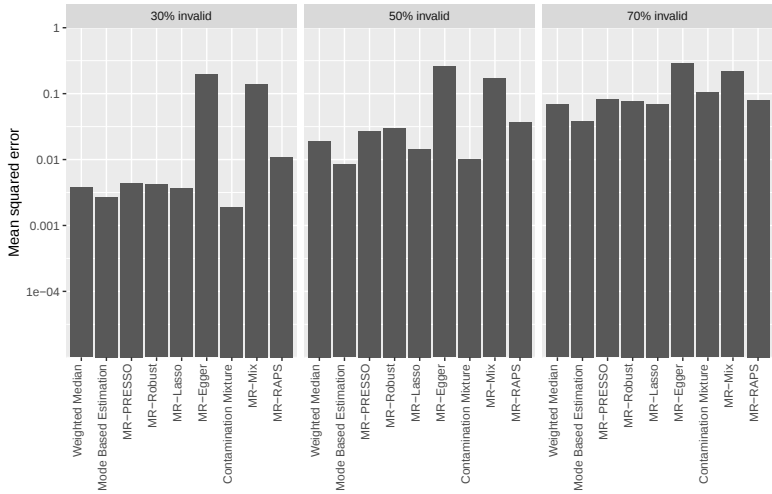


Fig. 5 Mean squared error for the different methods in scenario 2 for 10 000 simulations, with direc-tional pleiotropy and InSIDE satisfied with 10 variants.

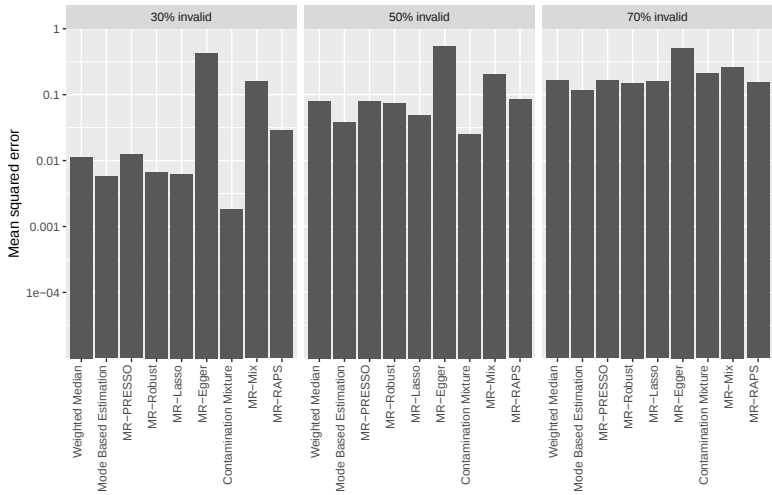


Fig. 6 Mean squared error for the different methods in scenario 3 for 10 000 simulations, with direc-tional pleiotropy and InSIDE violated with 10 variants.

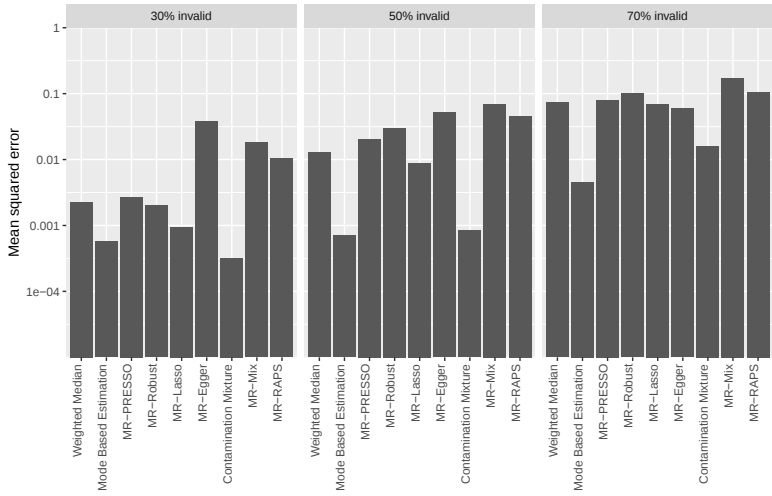


Fig. 7 Mean squared error for the different methods in scenario 2 for 10 000 simulations, with direc-tional pleiotropy and InSIDE satisfied with 100 variants.

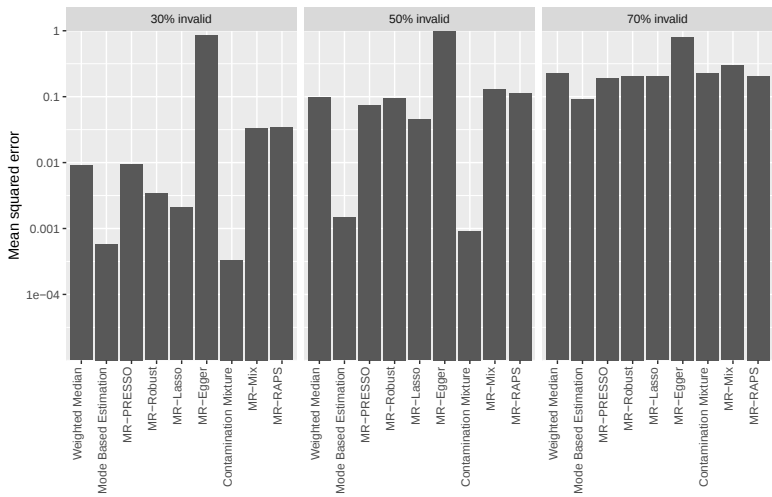


Fig. 8 Mean squared error for the different methods in scenario 3 for 10 000 simulations, with direc-tional pleiotropy and InSIDE violated with 100 variants.

S2 Outliers according to different methods

Table 8 Genetic variants identified as outliers by the different methods in the Mendelian Randomization study of the effect of BMI on cardiovascular disease risk.

Variant	MR-Robust	MR-PRESSO	MR-RAPS	MR-Lasso	Contamination mixture	MR-Mix
rs11191560	✓	✓	✓	✓	✓	✓
rs2075650	✓	✓	✓	✓	✓	✓
rs2176040		✓	✓	✓	✓	✓
rs6567160		✓	✓	✓	✓	✓
rs7903146		✓	✓	✓	✓	✓
rs11727676			✓	✓	✓	✓
rs17024393			✓	✓	✓	✓
rs11126666				✓	✓	✓
rs13078960				✓	✓	✓
rs9914578				✓	✓	✓
rs1000940					✓	✓
rs11057405					✓	✓
rs11847697					✓	✓
rs12446632					✓	✓
rs12566985					✓	✓
rs16907751					✓	✓
rs205262					✓	✓
rs2650492					✓	✓
rs2836754					✓	✓
rs3849570					✓	✓
rs4787491					✓	✓
rs492400					✓	✓
rs7243357					✓	✓
rs9641123					✓	✓
rs10938397						✓
rs10968576						✓
rs11030104						✓
rs11688816						✓
rs12016871						✓
rs13021737						✓
rs13191362						✓
rs13201877						✓
rs1460676						✓
rs1516725						✓
rs1528435						✓
rs17203016						✓
rs2176598						✓
rs2287019						✓
rs2820292						✓
rs3810291						✓
rs3817334						✓
rs543874						✓
rs7164727						✓
rs7599312						✓
rs7899106						✓

S3 Software code

This section includes the code to run the robust methods used in this paper. Please note that the MR-Mix package is not publicly available, please contact the authors for the package ¹.

```
#install required packages
if (!require("MendelianRandomization")) {install.packages("
  MendelianRandomization")} else {}
if (!require("mr.raps")) {install.packages("mr.raps")} else {}
if (!require("devtools")) { install.packages("devtools") } else {}
if (!require("penalized")) {install.packages("penalized")} else {}

library("devtools")
devtools::install_github("rondolab/MR-PRESSO")

#load packages
library("MendelianRandomization")
library("mr.raps")
library("MRMix")
library("MRPRESSO")
library("penalized")

#create dataframe and object by different methods
mr_frame<-as.data.frame(cbind(ldlc,ldlcse,chdlodds,chdloddsse))
names(mr_frame)<-c("ldlc","ldlcse","chdlodds","chdloddsse")
mr_object<-mr_input(bx = ldlc, bxse = ldlcse, by = chdlodds, byse =
  chdloddsse)#create used by methods from MendelianRandomization
  package

#perform weighted median
mr_median(mr_object,weighting = "weighted", iterations = 10000)

#perform Mode based estimation
mr_mbe(mr_object, weighting = "weighted", stderror = "delta", phi =
  1,
  seed = 19940407, iterations = 10000, distribution = "normal",
  alpha = 0.05)

#perform MR-PRESSO
mr_presso(BetaOutcome = "chdlodds", BetaExposure = "ldlc",
```

¹current maintainer of the package is Guanghao Qi (gqi1@jhu.edu).

```

SdOutcome = "chdloddsse", SdExposure = "ldlcse", OUTLIERTest =
TRUE, DISTORTIONtest = TRUE, data = mr_**frame**, NbDistribution =
1000, SignifThreshold = 0.05)

#perform MR-Robust
mr_ivw(mr_object,"random", robust = TRUE)

#define function for MR-Lasso with heterogeneity criterion
MR_lasso<-function (betaYG,betaXG,sebetaYG) {

  betaYGw = betaYG/sebetaYG # dividing the association estimates by
    sebetaYG is equivalent
  betaXGw = betaXG/sebetaYG # to weighting by sebetaYG^-2
  pleio = diag(rep(1, length(betaXG)))
  llgrid = c(seq(from=0.1, to=5, by=0.1), seq(from=5.2, to=10, by
    =0.2))
  # values of lambda for grid search
  llgrid_rse = NULL; llgrid_**length** = NULL; llgrid_**beta** = NULL;
    llgrid_**se** = NULL
  for (i in 1:length(llgrid)) {
    llgrid_**which** = which(attributes(penalized(betaYGw, pleio,
      betaXGw, lambda1=llgrid[i], trace=FALSE))$penalized==0)
    llgrid_rse[i] = summary(lm(betaYG[llgrid_**which**]~betaXG[llgrid_
      **which**]-1, weights=sebetaYG[llgrid_**which**]^2))$sigma
    llgrid_**length**[i] = length(llgrid_**which**)
    llgrid_**beta**[i] = lm(betaYG[llgrid_**which**]~betaXG[llgrid_**which**]-1,
      weights=sebetaYG[llgrid_**which**]^2)$coef[1]
    llgrid_**se**[i] = summary(lm(betaYG[llgrid_**which**]~betaXG[llgrid_
      **which**]-1, weights=sebetaYG[llgrid_**which**]^2))$coef[1,2]/min(
      summary(lm(betaYG[llgrid_**which**]~betaXG[llgrid_**which**]-1,
        weights=sebetaYG[llgrid_**which**]^2))$sigma, 1)
  }
  llwhich_hetero = c(which(llgrid_rse[1:(length(llgrid)-1)]>1& diff(
    llgrid_rse)>qchisq(0.95, df=1)/llgrid_**length**[2:length(llgrid)]
    , length(llgrid))[1]
  # heterogeneity criterion for choosing lambda

  llhetero_**beta** = llgrid_**beta**[llwhich_hetero]

```

```

llhetero_se = llgrid_se[llwhich_hetero]
list(ThetaEstimate=llhetero_beta, ThetaSE=llhetero_se )

}

#perform MR-Lasso
MR_lasso(mr_frame$chdlodds, mr_frame$ldlc, mr_frame$chdloddsse)

#perform MR-Egger
mr_egger(mr_object)

#define function for contamination mixture
contaminationmixture<-function(by,bx,byse){
  iters = 2001; theta = seq(from=-3, to=3, by=2/(iters-1))
  # if the causal estimate (and confidence interval) is not expected
    to lie between -1 and 1 then change from and to (and maybe
    increase iters)
  ratio = by/bx; ratio.se = abs(byse/bx); psi = 1.5*sd(ratio)
  lik=NULL
  for (j1 in 1:iters) {
    lik.inc = exp(-(theta[j1]-ratio)^2/2/ratio.se^2) /sqrt(2*pi*
      ratio.se^2)
    lik.exc = exp(-ratio^2/2/(psi^2+ratio.se^2)) /(sqrt(2*pi*(psi^2+
      ratio.se^2)))
    valid = (lik.inc>lik.exc)*1
    lik[j1] = prod(c(lik.inc[valid==1], lik.exc[valid==0]))
    if (which.max(lik)==length(lik)) { valid.best = valid }
  }
  phi = ifelse(sum(valid.best)<1.5, 1, max(sqrt(sum((ratio[valid.
    best==1]-weighted.mean(ratio[valid.best==1] , ratio.se[valid.
    best==1]^2)) ^2 * ratio.se[valid.best==1]^2)) /(sum(valid.best
    )-1)), 1))
  loglik = log(lik)
  whichin = which(2*loglik>(2*max(loglik)-qchisq(0.95, df=1)*phi^2))
  theta[which.max(loglik)] # estimate
  theta[whichin[1]] # lower limit of CI

```

```

theta[whichin[length(whichin)]] # upper limit of CI
list(ThetaEstimate=theta[which.max(loglik)], ThetaLower=theta[
  whichin[1]] , ThetaUpper= theta[whichin[length(whichin)]] )
}

#perform contamination mixture, note we removed the 27th variable
  due to having a ratio to close to infty.
contaminationmixture(mr_frame$chdlodds[-27],mr_frame$ldlc[-27],mr_
frame$chdloddsse[-27])

#perform MR-Mix
estMix = MRMix(mr_frame$chdlodds, mr_frame$ldlc, mr_frame$
  chdloddsse^2, mr_frame$ldlcse^2)
se = MRMix_se(mr_frame$chdlodds, mr_frame$ldlc, mr_frame$chdloddsse
  ^2, mr_frame$ldlcse^2, estMix$theta, estMix$pi0, estMix$sigma2)

#perform MR-RAPS with Huber loss function

mr.raps.overdispersed.robust(mr_frame$chdlodds, mr_frame$ldlc, mr_
frame$chdloddsse, mr_frame$ldlcse,
  loss.function = "huber", k = 1.345,
  initialization = c("l2"), suppress.warning
    = FALSE, diagnosis = FALSE, niter = 20,
  tol = .Machine$double.eps^0.5)

```