

# 1 Systems Biology Theory Clarification of a Controversy in

## 2 Pancreatic Beta Cell Regeneration

3 Haoran Cai<sup>1,3</sup>, Runtan Cheng<sup>1</sup>, Ruoshi Yuan<sup>2</sup>, Xiaomei Zhu<sup>5\*</sup>, Ping  
4 Ao<sup>1,2,4,5\*</sup>

5 <sup>1</sup> Key Laboratory of Systems Biomedicine (Ministry of Education), Shanghai  
6 Center for Systems Biomedicine, Shanghai Jiao Tong University, 800  
7 Dongchuan Road, Shanghai 200240, China

8 <sup>2</sup> Department of Systems Biology, Harvard Medical School, Boston, MA 02115,  
9 USA

10 <sup>3</sup> Department of Biostatistics and Computational Biology, Dana Farber Cancer  
11 Institute, Boston, MA 02115, USA

12 <sup>4</sup> State Key Laboratory for Oncogenes and Related Genes, Shanghai Cancer  
13 Institute, Shanghai Jiao Tong University School of Medicine, Shanghai 200240,  
14 China

15 <sup>5</sup> Shanghai Center for Quantitative Life Sciences and Physics Department,  
16 Shanghai University, Shanghai 200444, China

17 \* Co-senior author

18 Corresponding author: [aoping@sjtu.edu.cn](mailto:aoping@sjtu.edu.cn)

19

20 Keywords: systems biology; cell replacement; dynamical system; endogenous

21 network

22

23

24

25

26

## 27 **ABSTRACT**

28 Whether new pancreatic beta-cells arise via pre-existing beta-cells or from  
 29 differentiation of precursor cells – a question of fundamental importance for diabetic  
 30 therapy – has long been debated. Recent experiments suggest that multipotent  
 31 precursors from adult mouse pancreas, that give rise to beta-cells, do exist. However,  
 32 such a finding is at odds with prior evidence that beta-cell expansions occurs  
 33 exclusively through self-replication. Here we show that these two observations can be  
 34 partially compatible. We use a systems biology approach to analyze the dynamics of  
 35 the endogenous molecular-cellular network in the pancreas. Our results show that self-  
 36 replicating ‘beta-cells’ can themselves be multipotent precursors. In addition, our model  
 37 predicts heterogeneity in beta-cell regeneration and suggests various differentiation  
 38 paths of precursors. This work therefore provides a means of reconciling an apparent  
 39 contradiction in the field, but also sheds light on possible paths of beta-cell regeneration  
 40 from a systems biology perspective.

41

## Introduction

Diabetes has brought great public health burden as well as economic costs[1, 2], which is caused by the body's lack of proper response to insulin production (Insulin resistance) or the pancreas's inability to produce enough insulin. Both  $\beta$ -cell dysfunction and decreased  $\beta$ -cell mass account for insulin deficiency. It is now recognized that beta-cell loss is a common theme of type 1 diabetes and type 2 diabetes. For example, in patients with type 2 diabetes, the beta-cell mass was found to be reduced by 50%[3]. Thus, many studies have tried to elucidate the mechanisms that control beta cell formation and replacement in order to design regenerative therapy of diabetes[4, 5].

Yet, one highly controversial issue remains unsettled on the postnatal origins of beta cells: whether new pancreatic beta-cells arise via pre-existing beta-cells or differentiation of precursor cells. It has been found that adult beta-cell retains a small capacity for proliferation[6–8]. Surprisingly, a seminal lineage-tracing study found that the fraction of labeled beta-cells remained unchanged over a one-year chase period, suggesting that beta-cell expansion was driven by self-replication without any

58 contribution of precursor cell differentiation[9], which can be further amplified by  
 59 subsequent studies[10, 11]. On the other hand, several studies have supported the idea  
 60 that the formation of new endocrine cells is from the pancreatic stem cells since the  
 61 late 19<sup>th</sup>-century [12–14], Recently, several studies have found that multipotent  
 62 precursors do exist. A report argued that the beta-cells labeled in the lineage-tracing  
 63 study may not necessarily be mature beta-cell and can be insulin-expressing  
 64 precursors that give rise to endocrine cell types[15]. Another group also identified  
 65 non-insulin-expressing cells in islets that could give rise to new insulin+ cells as a  
 66 slow renewal for beta-cells.

67  
 68 Here we used a systems biology method, the endogenous molecular-cellular network  
 69 theory, to clarify the contrasting observations from a systematic view. The theory is  
 70 intended to analyze the complex biological process via the dynamics of the network  
 71 system based on the fundamental properties of the biological system[16, 17]. The  
 72 endogenous molecular-cellular network is composed of essential modules (specified by  
 73 a set of nodes representing key proteins or signaling pathway) and crosstalk between

74 these modules, and then we analyze the dynamics on the network. We assume cell  
75 phenotypes are endogenous attractors underlying the endogenous molecular–cellular  
76 network. One of the most prominent predictions of endogenous network theory is the  
77 existence of quantitative functional landscape, where locally stable states are  
78 interconnected with each other through the transition states. The natural and qualitative  
79 consequence of the mathematical setup implies that natural conversion between two  
80 cell states may have patterns[17], and the topological structure of the functional  
81 landscape can be a natural framework for beta-cell regeneration. We sought to analyze  
82 the natural mechanisms at a network level by which beta cells are formed, where the  
83 observations of beta-cell expansion can be integrated into a single model.

## 84 **Methods**

### 85 **Construction of an endogenous network of pancreas development**

86 In this work, we chose a set of essential proteins to depict pancreatic core regulatory  
87 structures (detailed description in the unpublished paper). These proteins and their  
88 causal interactions form the core endogenous network of the pancreas.

## Quantitative description and analysis

A set of ordinary differential equations (ODEs) were obtained to quantify the core dynamics on the pancreatic endogenous network. The dynamics of the activation/expression level of each protein  $x$  is governed by

$$\frac{d[x]}{dt} = V_{\max} * \frac{k * (\sum[\text{activator}]^n)}{1 + k * (\sum[\text{activator}]^n)} * \frac{1}{1 + k * (\sum[\text{inhibitor}]^n)} - \tau * [x] \quad (1)$$

where  $V_{\max}$  represents the maximal production rate of protein  $x$ ,  $n$  represents Hill coefficients, and  $k$  represents dissociation constant. Specifically, the relative expression level of each protein was normalized to range from 0 to 1. The maximal production rate  $V_{\max}$  and degradation rate  $\tau$  were taken as 1. Here the values of  $n$  and  $k$  were 3 and 10 while we conduct multiple simulation varying  $n$  and  $k$  within a reasonable range to grasp the key feature of activation or inhibition. The threshold of the sigmoid-shaped function, at which the value of  $x$  was expected to be half maximal. Two independent algorithms, random sampling and Newton's method, were adopted to calculate the robust fixed points of the dynamical system (see Supplementary Materials).

In the dynamic system (Eqn. 1), we perturbed the system with small random noise when

it stayed at an unstable state (transition state or hyper-transition state) to obtain the topological structure of landscape. We utilized random-perturbed states as the initial value of the system and let the dynamical systems iterate at the constraints and tracked routes of system evolution. We obtained the trajectories from each unstable state to it connected stable states and recorded the unstable states that each trajectory passed through.

Independent datasets[18] are used to validate the model results. Firstly, four pathways expression levels are denoted by the average of targeted proteins (pathways). We averaged expression level by cell type annotation. Then Z-score normalization is conducted respectively for single cell transcriptome and modeling results (12 stable states and 23 transition states) over each protein (pathways). Eventually, we do linearly rescale for all the expression value to 0 – 1.

## Results

We obtained 12 stable states and 23 transition states in the simulations of our model (see in Table S2, Table S3). Each state is depicted by the combinational expression level of endogenous network nodes. We assumed pancreatic phenotypes are robust stable states of the endogenous network, thus biological meanings were linked to the stable states emerging from the endogenous network (see in Figure 2, Figure S2). The interconnections among phenotypic states reveal the lineage conversion routes in the pancreas, making it possible to understand the maintenance of pancreas homeostasis from a systematic view. In our work, a beta cell proliferation landscape could be obtained describing how states connect with each other, part of the whole pancreatic landscape (Figure 3).

By state-connected graph, multiple potential sources including 1 stable state and 11 transition states for expansion can be identified (Figure 3). By examining the expression pattern, U23, S3, U2 states are found to be insulin-producing cells that could give rise to formation of new beta-cells and other phenotypic of states (See in Figure S1)



136 according to our modeling results, as markers that activate Ins are highly expressed at  
137 these states (Figure 4). This can be validated by the report that identified an multipotent  
138 insulin-expressing population as the precursor of beta-cells[15]. While states such as  
139 U11, U12, U20 that do not express a high level of Pdx1, MafA (Figure 4) are insulin-  
140 negative states in our model. And they maintained the capacity to give rise to new beta-  
141 cells as well (Figure 3), consistent with the postulations from [19] that Insulin-negative  
142 precursors participate in the renewal of the beta-cell mass during aging. In this way our  
143 model not only explained two evidences straightforwardly but integrated the evidences  
144 of existence of precursor cells into a single framework from a systematic view.

145

146 Furthermore, our model suggested that the sources of beta-cells regeneration may even  
147 be more heterogeneous than expected. For example, some of the precursor states are  
148 unipotent to beta-cells according to the complete landscape such as U2, U16, U23,  
149 suggesting a capacity in vivo that might be exploited to exclusively generating beta-  
150 cells for therapeutic cell replacement. Furthermore, as the newly differentiated beta-  
151 cells are hard to be distinguished from pre-existing population in many experimental

models, the expression characteristics of multiple possible pre-beta states can be obtained (see in Table S2, Table S3) through our modeling to enable future experimental validations for post-natal origin of beta-cells.

## Discussion

Whether new beta cells arise from differentiation or by the proliferation of existing beta cells has remained a highly controversial issue for decades. Dor's study which used an mice model to trace the fraction of insulin-expressing cells suggested that self-replication was the major source of beta-cell regeneration exclusively[9]. The existence of progenitor-like insulin-positive states in our model, distinct from mature beta-cells, was probably the partial reason why they found the new insulin-positive cells are differentiated from existing insulin-expressing cells. Because the precursor cells can express insulin, lineage labelling of beta cells by the insulin expression could not discriminate between whether it is self-replication of pre-existing mature beta cells or differentiation from stem cells that give rise to the formation of new beta-cells.

166 However, two evidences demonstrated that the multipotent insulin expressing  
167 precursors could not provide total reconciliation with the work of [9]. First, insulin-  
168 negative states are possible to contributed to new insulin+ cells replacement both  
169 theoretically and experimentally. Secondly, the precursor beta cells identified before  
170 are multipotent[15] and there is no unipotent insulin expressing state in our landscape  
171 while Dor and his colleagues found the labeled cells only give rise to beta cells. This  
172 could be attributed by techniques: One reason related is that the staining method that  
173 Dor took probably hindered the visualization of labeled cells giving rise to states other  
174 than beta cells. In addition, it may overlook the cases that the pre-existing mature beta  
175 cells retain the capacity to dedifferentiate to an unipotent progenitor state and re-  
176 differentiate back, which indeed has been reported by several studies[20, 21]. Since  
177 these precursor cells are quite similar with the mature beta cells, it might be hard to  
178 discriminate them in experimental models. Another reason might concern with the  
179 small fraction of progenitor states: the lineage tracing results merely reflect the  
180 behaviors of mature beta-cells.

181 Enlightened by pancreatic development endogenous network, we constructed the  
 182 quantitative landscape of pancreatic development without prior knowledge of beta-  
 183 cell expansion. Emerged stable states are linked to phenotypic states within the  
 184 pancreas, reproducing the core features of phenotypic states within pancreas to better  
 185 understand the beta cell replacement. We conducted perturbation analysis to generate  
 186 topological graph describing interconnection among states, serving as the guideline to  
 187 understand the beta-cell replacement. The roadmap of beta cell regeneration can be  
 188 established: Various possible states retain the capacity to give rise to new beta cells  
 189 either during development, aging or even under injury. Our results supported that the  
 190 new post-natal beta cells can originate from the unmatured pre-beta cells. The  
 191 precursors can be rather heterogenous, characterized by combinational expression  
 192 level of genes(proteins) in our landscape. Besides, we showed that pre-existing beta-  
 193 cells could transiently dedifferentiate to a progenitor-like state and facilitate the beta-  
 194 cell replacement, which can be the case hindered by the experimental techniques. The  
 195 observations were integrated into a single model, and an explanation of the beta-cell  
 196 origins within adult pancreas has been obtained from a systems biology theory.

197 One remaining question is that to which extent the beta-cell expanded by self-  
 198 replication or from precursor cells. It is also of interests to know conditions that a  
 199 certain phenotypic state of cells will occur and contribute more to the beta-cell  
 200 expansion. We acknowledge that the network has been greatly simplified. It is expected  
 201 that a more comprehensive network can reproduce more detailed features through the  
 202 inclusion of more modules, for example, cell cycle. These issues require an explicit  
 203 inclusion of stochastic effects, where the potential energy landscape can be used to  
 204 explore more detailed issues[22, 23].

205

206 Apart from elucidating the controversy in this work, the dynamical network system  
 207 we built may have other applications: Our model does not exclude the possibility that  
 208 other terminally differentiated phenotypes of cells trans-differentiate into beta-cells  
 209 for their expansion. In the light of our hypothesis, these trans-differentiation behaviors  
 210 could correspond to the states traveling in the landscape (Figure 3) as well. Indeed,  
 211 stem cells have been observed in multiple experiment models[24–26]. Hence, it is  
 212 possible that other multiple routes can generate new beta-cells. Our model provides a

213 framework to understand the interconversion of cell states during aging or embryonic  
 214 development. The patterns and preferred routes that our model implies can be further  
 215 studied to predict potential target genes and develop successful therapies for beta cell  
 216 regeneration in the treatment of diabetes.

217

# 218 **Competing Interest**

219 The authors declare no competing interests.

# 220 **Data availability.**

221 The published data sets used in this manuscript are available through the following accession numbers: SMART-seq2  
 222 platform pancreas data by Segerstolpe *et al.* [18], ArrayExpress [E-MTAB-5061](#).

223

224

225

226

227

228

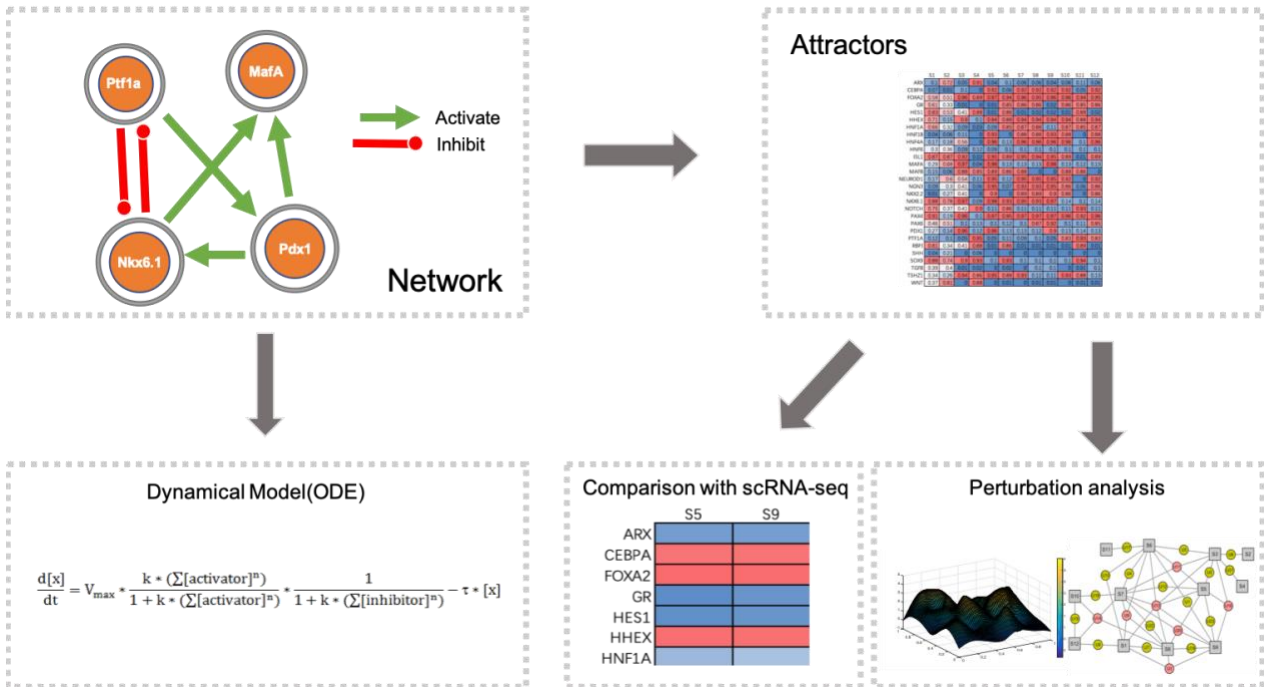


Figure 1

**Figure 1. Schematics of endogenous molecular-cellular network modelling.** The interactions were collected from the literature. Ordinary differential equations (described in Methods and Supplementary Material) were used to compute the attractors generated by the constructed pancreatic developmental network structure. Two algorithms (see Supplementary Materials) were performed, demonstrating robustness of the simulation results. Comparison of gene expression levels predicted by the attractors with single cell RNA-seq data validated our scientific simulation results. Multiple phenotypes within pancreas corresponded to the attractors of network dynamics.

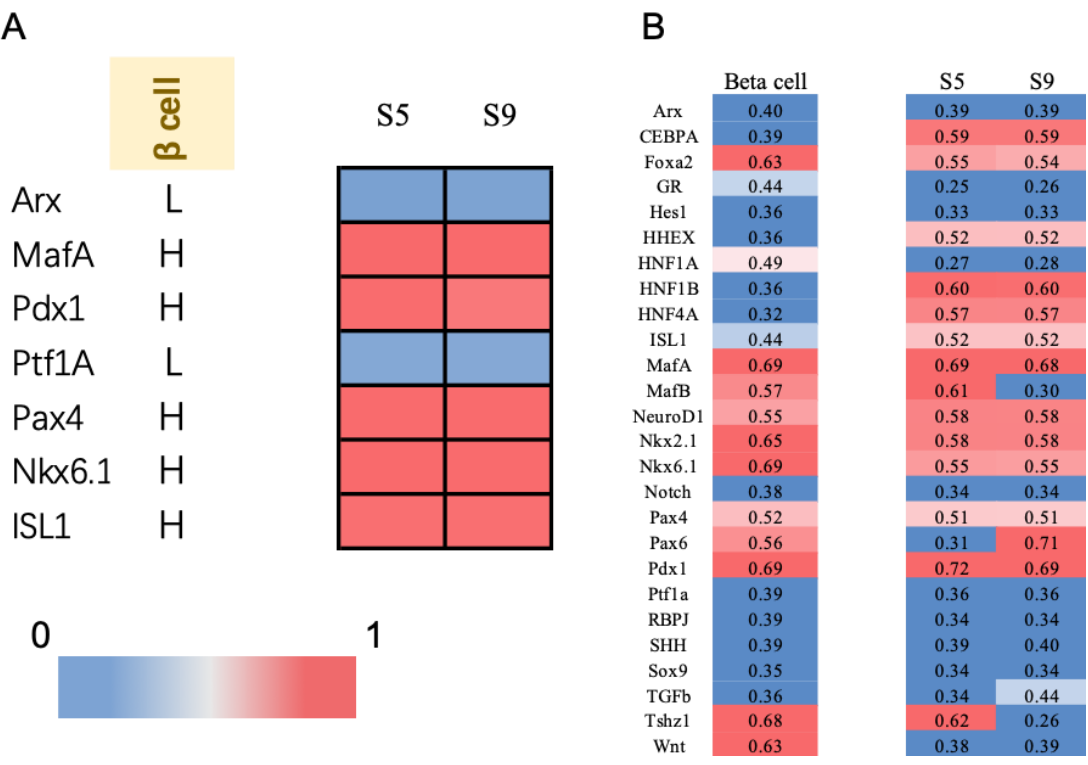


Figure.2

**Figure 2. Linking biological meaning to stable states and its validation comparing with RNA-Seq.** **A.** We used the known markers (L denotes Low expression, H denotes High expression), we could easily link the cell type to corresponding stable states generated from the endogenous network (See Figure S2). **B** The biological meanings of beta cell states were validated at the molecular level. We selected the relevant expression data in a published dataset[18] and set a threshold to find out the high or low expressed status of each gene. When we set the threshold as 0.5, the agreement ratio was 71.2%



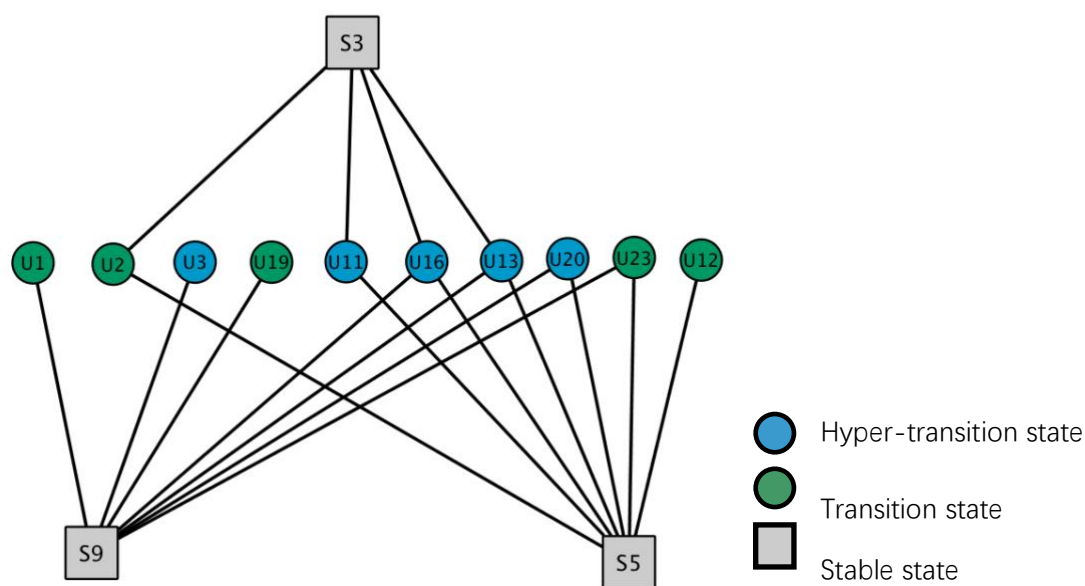


Figure 3

**Figure 3. The network dynamics incorporated the seemingly conflicted beta-cell expansion**

**models.** This state connection graph is a part of the whole landscape (See Table S1) through perturbation analysis. Each state is depicted by the expression level of a set of molecular we selected to form the endogenous network. S5, S9 represented beta cell states while all the other phenotypic states including stable states S3 and transition state/hyper-transition state retain the potential to differentiate into the beta cell. Stable state: all the eigenvalues of the Jacobian matrix of dynamical system at this state were negative; Transition state: one eigenvalue of the Jacobian matrix at this state was positive while the others were negative; Hyper-transition state: more than one positive eigenvalues of the Jacobian matrix at this state were positive.

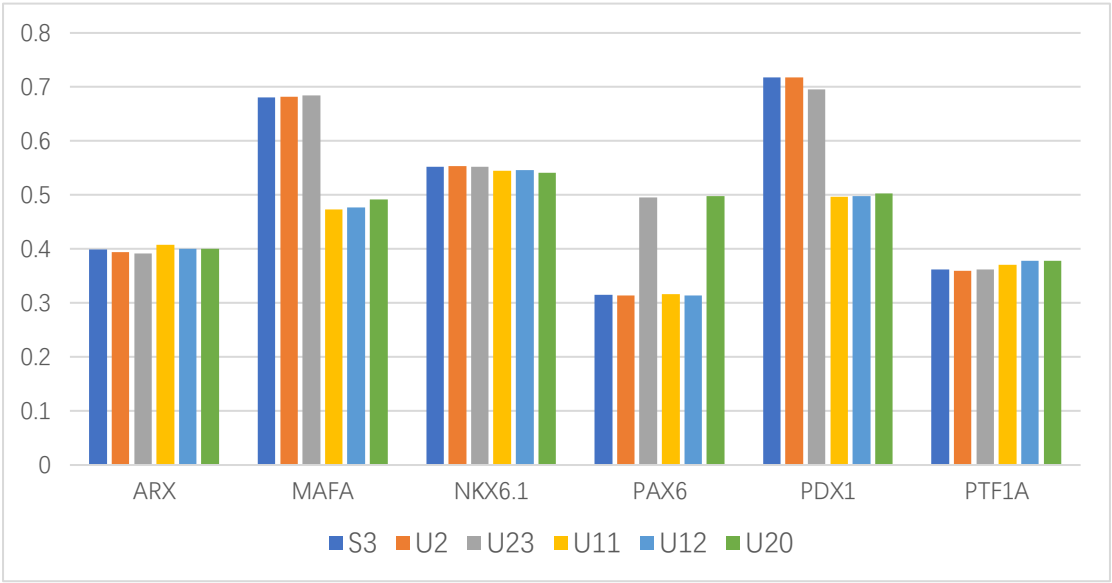


Figure 4

**Figure 4. Modeling results characterized the insulin expression features of putative beta cell precursor through a quantitative expression level of proteins.** States that highly express MafA, Nkx6.1, Pax6, Pdx1 and low express Arx, Ptf1a are more likely to express insulin[28–31]. State S3, U2, U23 were assumed to be insulin-positive. Due to the lack of expression of MafA, Pdx1, U11, U12, U20 are probably insulin-negative cells.

## Reference

1. Dieren S Van, Beulens JWJ, Schouw YT Van Der, Grobbee DE, Neal B. The global burden of diabetes and its complications : an emerging pandemic. 2016.
2. Statements ADA. Hyperglycemic Crises in Adult Patients. 2009;32.
3. Rhodes CJ. Type 2 Diabetes-a Matter of {beta}-Cell Life and Death? Science (80- ). 2005.
4. Cheng CW, Villani V, Buono R, Wei M, Kumar S, Yilmaz OH, et al. Fasting-Mimicking Diet Promotes Ngn3-Driven  $\beta$ -Cell Regeneration to Reverse Diabetes. Cell. 2017.
5. Zeng C, Mulas F, Sui Y, Guan T, Miller N, Tan Y, et al. Pseudotemporal Ordering of Single Cells Reveals Metabolic Control of Postnatal  $\beta$  Cell Proliferation. Cell Metab. 2017;25:1160–1175.e11.
6. Messier B, Leblond CP. Cell proliferation and migration as revealed by radioautography after injection of thymidine-<sup>3</sup>H into male rats and mice. Am J Anat. 1960.

268 7. Kassem SA, Ariel I, Thornton PS, Scheimberg I, Glaser B.  $\beta$ -Cell proliferation and  
269 apoptosis in the developing normal human pancreas and in hyperinsulinism of infancy.  
270 Diabetes. 2000.

271 8. Acini MP. Demonstration of Expanding Cell Populations in Mouse Pancreatic Acini  
272 and Islets. 1987;115.

273 9. Dor Y, Brown J, Martinez OI, Melton DA. Adult pancreatic  $\beta$ -cells are formed by  
274 self-duplication rather than stem-cell differentiation. Nature. 2004;429:41–6.  
275 doi:10.1038/nature02520.

276 10. Nir T, Melton D a, Dor Y. Recovery from diabetes in mice by beta cell regeneration.  
277 J Clin Invest. 2007;117:2553–61.

278 11. Salpeter SJ, Klein AM, Huangfu D, Grimsby J, Dor Y. Glucose and aging control  
279 the quiescence period that follows pancreatic beta cell replication. Development.  
280 2010;137:3205–13. doi:10.1242/dev.054304.

281 12. Bensley RR. Studies on the pancreas of the guinea pig. Am J Anat. 1911;12:297–  
282 388.

283 13. Bonner-Weir S, Weir GC. New sources of pancreatic beta-cells. Nat Biotechnol.

284 2005;23:857–61.

285 14. Endocrine P, Zulewski H, Abraham EJ, Gerlach MJ, Daniel PB, Moritz W, et al.

286 Multipotential Nestin-Positive Stem Cells Isolated From. 1:521–33.

287 15. Smukler SR, Arntfield ME, Razavi R, Bikopoulos G, Karpowicz P, Seaberg R, et

288 al. The adult mouse and human pancreas contain rare multipotent stem cells that express

289 insulin. *Cell Stem Cell*. 2011;8:281–93.

290 16. Yuan R, Zhu X, Wang G, Li S, Ao P. Cancer as robust intrinsic state shaped by

291 evolution: a key issues review. *Reports Prog Phys*. 2017;80:042701. doi:10.1088/1361-

292 6633/aa538e.

293 17. Ao P, Galas D, Hood L, Zhu X. Cancer as robust intrinsic state of endogenous

294 molecular-cellular network shaped by evolution. *Med Hypotheses*. 2008;70:678–84.

295 18. Segerstolpe Å, Palasantza A, Eliasson P, Andersson E-M, Andréasson A-C, Sun X,

296 et al. Single-Cell Transcriptome Profiling of Human Pancreatic Islets in Health and

297 Type 2 Diabetes. *Cell Metab*. 2016;24:593–607. doi:10.1016/j.cmet.2016.08.020.

298 19. Liu H, Guz Y, Kedeas MH, Winkler J, Teitelman G. Precursor cells in mouse islets

299 generate new  $\beta$ -cells in vivo during aging and after islet injury. *Endocrinology*.

2010;151:520–8.

20. Weinberg N, Ouziel-Yahalom L, Knoller S, Efrat S, Dor Y. Lineage tracing evidence for in vitro dedifferentiation but rare proliferation of mouse pancreatic  $\beta$ -cells. *Diabetes*. 2007.

21. Ouziel-Yahalom L, Zalzman M, Anker-Kitai L, Knoller S, Bar Y, Glandt M, et al. Expansion and redifferentiation of adult human pancreatic islet cells. *Biochem Biophys Res Commun*. 2006.

22. Zhu XM, Yin L, Hood L, Ao P. Calculating biological behaviors of epigenetic states in the phage  $\gamma$  life cycle. *Funct Integr Genomics*. 2004;4:188–95.

23. Yuan R, Ao P. Beyond Itô versus Stratonovich. *J Stat Mech Theory Exp*. 2012;2012.

24. Chera S, Baronnier D, Ghila L, Cigliola V, Jensen JN, Gu G, et al. Diabetes recovery by age-dependent conversion of pancreatic  $\delta$ -cells into insulin producers. *Nature*. 2014;514:503–7. doi:10.1038/nature13633.

25. Kopp JL, Grompe M, Sander M. Stem cells versus plasticity in liver and pancreas regeneration. *Nat Cell Biol*. 2016;18:238–45. doi:10.1038/ncb3309.

26. Katsuta H, Akashi T, Katsuta R, Nagaya M, Kim D, Arinobu Y, et al. Single

316 pancreatic beta cells co-express multiple islet hormone genes in mice. *Diabetologia*.  
317 2010;53:128–38.

318 27. Kang J, Nathan E, Xu SM, Tzahor E, Black BL. Isl1 is a direct transcriptional target  
319 of Forkhead transcription factors in second heart field-derived mesoderm. *Dev Biol*.  
320 2009;334:513–22.

321 28. Tang DQ, Lu S, Sun YP, Rodrigues E, Chou W, Yang C, et al. Reprogramming liver-  
322 stem WB cells into functional insulin-producing cells by persistent expression of Pdx1-  
323 and Pdx1-VP16 mediated by lentiviral vectors. *Lab Investig*. 2006;86:83–93.

324 29. Xu L, Xu C, Zhou S, Liu X, Wang J, Liu X, et al. PAX4 promotes PDX1-induced  
325 differentiation of mesenchymal stem cells into insulin-secreting cells. *Am J Transl Res*.  
326 2017;9:874–86.

327 30. Grzeskowiak R, Amin J, Oetjen E, Knepel W. Insulin responsiveness of the  
328 glucagon gene conferred by interactions between proximal promoter and more distal  
329 enhancer-like elements involving the paired-domain transcription factor Pax6. *J Biol*  
330 *Chem*. 2000;275:30037–45.

331 31. Aguayo-Mazzucato C, Koh A, El Khattabi I, Li WC, Toschi E, Jermendy A, et al.

332 Mafa expression enhances glucose-responsive insulin secretion in neonatal rat beta

333 cells. Diabetologia. 2011;54:583–93.

334