

1 **MBIR: A Cryo-electron Tomography 3D Reconstruction Method that Effectively
2 Minimizes Missing Wedge Artifacts and Restores Missing Information**

3 Rui Yan^a, Singanallur V. Venkatakrishnan^b, Jun Liu^c, Charles A. Bouman^d, Wen Jiang^{a*}

4

5 ^aMarkey Center for Structural Biology, Department of Biological Sciences, Purdue University,
6 West Lafayette, IN 47907, USA

7 ^bOak Ridge National Laboratory, Oak Ridge, TN 37831, USA

8 ^cDepartment of Microbial Pathogenesis, Yale University School of Medicine, West
9 Haven, CT 06516, USA

10 ^dSchool of Electrical and Computer Engineering, Purdue University, West Lafayette, IN 47907,
11 USA

12

13 *Corresponding author: Wen Jiang

14 Email: jiang12@purdue.edu

15 Phone: 765-496-8436

16

17

18 **Abstract**

19 Cryo-Electron Tomography (cryo-ET) has become an essential technique in revealing cellular
20 and macromolecular assembly structures in their native states. However, due to radiation
21 damage and the limited tilt range, cryo-ET suffers from low contrast and missing wedge
22 artifacts, which limits the tomograms to low resolution and hinders further biological
23 interpretation. In this study, we applied the Model-Based Iterative Reconstruction (MBIR)
24 method to obtain tomographic 3D reconstructions of experimental cryo-ET datasets and
25 demonstrated the advantages of MBIR in contrast improvement, missing wedge artifacts
26 reduction, and missing information restoration compared with other reconstruction approaches.
27 Considering the outstanding reconstruction quality, MBIR has a great potential in the
28 determination of high resolution biological structures with cryo-ET.

29

30 **Introduction**

31 Cryo-electron tomography (cryo-ET) has emerged as a promising technique that allows us to
32 comprehensively explore macromolecular complexes and cellular architecture in near-native
33 states ¹. Using cryo-ET, the 3D tomogram of the biological sample can be reconstructed from
34 a 2D tilt series collected by sequentially tilting the sample at different projection angles around
35 a tilt axis ². In practice, the quality of reconstruction with cryo-ET remains limited by several
36 challenges in the data acquisition and reconstruction process.

37 The extremely poor signal-to-noise ratio (SNR) of cryo-ET is the first major challenge in
38 improving cryo-ET resolution ³. To prevent significant radiation damage to biological samples
39 by the electron beam, the total dose used for a cryo-ET tilt series is typically less than 100 e/Å².
40 This low-dose imaging strategy in combination with the increment of sample thickness during
41 tilting results in very noisy, low contrast 2D projections, which poses a challenge in subsequent
42 2D tilt series alignments and deteriorates the resolution of cryo-ET 3D reconstruction ^{4,5}.

43 The second major challenge of cryo-ET is the missing wedge artifacts caused by the limited tilt
44 angle range during data collection ⁶. Since more electrons are lost to inelastic scattering as the
45 effective sample thickness increases when the sample is tilted ³, the maximal tilt range of cryo-
46 ET is typically restricted within $\pm 70^\circ$ to ensure enough electrons can traverse through the
47 sample, generate elastic scattering, and form reliable images ⁷. Consequently, the absence of
48 the high tilt angles ($-90^\circ \sim -70^\circ$ and $+70^\circ \sim +90^\circ$) becomes a “missing wedge” of un-sampled
49 information in Fourier space, leading to severe ray artifacts, structural elongation, and distortion
50 effects in the final reconstruction ⁸. The missing wedge artifacts dramatically weaken the
51 interpretability of the reconstructed tomogram and limit the achievable resolution of cryo-ET ¹.

52 To address these challenges of cryo-ET, we introduce the Model-Based Iterative
53 Reconstruction (MBIR) method ⁹ for tomographic reconstruction and benchmark the tomogram
54 quality with the state-of-the art algorithms, including Back Projection (BP), Simultaneous
55 Iterative Reconstruction Technique (SIRT), and Iterative Compressed-sensing Optimized Non-
56 uniform fast Fourier transform reconstruction (ICON) ¹⁰. In MBIR framework, the

57 reconstruction is formulated as the maximum a posterior (MAP) estimate of the unknowns,
58 given the measurements

$$59 (\hat{f}, \hat{\emptyset}) = \underset{f, \emptyset}{\operatorname{argmax}} \{ \log p(f, \emptyset | g) \} = \underset{f, \emptyset}{\operatorname{argmin}} \{ -\log p(g | f, \emptyset) - \log p(f) \} \quad (1)$$

60 where g represents the data obtained from an imaging system (e.g. cryo-ET tilt series), f
61 represents the unknown 3D structure to be discovered, \emptyset represents the unknown nuisance
62 parameters of the system such as beam intensity fluctuations and noise characteristics.
63 $p(g | f, \emptyset)$ is the likelihood function that models how the observations are related to the
64 unknowns, $p(f)$ is the assumed prior distribution of the unknown structure. Here $p(g | f, \emptyset)$
65 and $p(f)$ indicate the forward model of image formation and prior model of the tomogram in
66 MBIR algorithm, respectively ⁹. Currently, the forward model accounts for the decay of
67 electron beam intensity following Beer-Lambert Law and combines with the estimation of
68 detector noise. The prior model uses a Gaussian Markov Random Field to account for diffuse
69 or sharp interfaces between structural features and encourage smoothness in the solution. The
70 goal of MBIR will be to compute a final estimate \hat{f} that represents a balance between fitting
71 the measurements based on the system forward model $p(g | f, \emptyset)$ while remaining consistent
72 with the prior model $p(f)$. Fig. 1 illustrates a general framework of MBIR for solving inverse
73 problems in imaging applications.

74 MBIR method has been previously shown to generate better quality tomograms when applied
75 to tomography applications like CT scan, X-Ray tomography, positron emission tomography
76 (PET), optical diffusion tomography (ODT), and atomic resolution electron tomography of
77 radiation-resistant material specimens ¹¹. MBIR combines a forward model for image formation
78 with a prior model for the unknown structure to reconstruct tomograms ⁹. In this study, tests
79 with both plastic embedded ET dataset and ice embedded cryo-ET datasets have shown that
80 MBIR can significantly improve the reconstruction quality with enhanced contrast, reduced
81 missing wedge artifacts, and partially restored information in the un-sampled angular region.

82

83 **Results**

84 **Missing wedge assessments using gold markers**

85 We first evaluated MBIR using one cryo-ET dataset (EMPIAR-10045) by visually examining
86 the missing wedge artifacts of gold markers in different slice views of the tomograms. Due to
87 the missing wedge problem, the gold markers become elongated along the direction of the
88 missing wedge and suffer from halos and streaking artifacts in the adjacent region. Fig. 2
89 compares slice views of the reconstructions generated by the four methods using gold markers
90 as an indicator of quality. In each block, three planes represent the XY-slice (middle plane),
91 XZ-slice (top plane) and YZ-slice (right plane) of the tomogram, respectively, intersecting at
92 the same gold marker. The zoomed-in view of the gold markers pointed by white arrows in the
93 three planes are placed at the corner of the corresponding planes. From the XY-slices of
94 tomograms, it is clear MBIR (XY-slice in Fig. 2d) has eliminated the halos artifacts and displays
95 more round, sharp-edged gold markers than other methods. In the XZ and YZ-slices, MBIR
96 (Fig. 2d) significantly reduced the elongation and ray artifacts of gold markers with improved
97 contrast of the biological structures, compared with the tomograms reconstructed by other
98 methods. Hence, MBIR-reconstructed tomograms show less artifacts from the missing wedge
99 problem, better contrast in cryo specimen, and clearer background.

100 To further examine the performance of MBIR, we applied it to one cryo-ET dataset acquired
101 with VPP (EMPIAR-10064 in Fig. 3a), two cryo-ET datasets without VPP (EMPIAR-10037
102 and EMPIAR-10110 in Fig. 3b and c), and one plastic embedded ET dataset (IMOD tutorial
103 dataset in Fig. 3d). Fig. 3 shows the slice views of these four datasets in which each row
104 represents the results of one dataset reconstructed by the four methods and each column
105 represents the results of one method applied to different datasets. In Fig. 3b and d, XY-slices
106 are mainly used to reveal the reconstruction quality of sample areas without targeting at a gold
107 marker because the sample and markers are not on the same XY plane. For a challenging dataset
108 shown in Fig. 3b, it is clear that BP reconstruction quality is too poor to make the biological
109 sample visible. SIRT and ICON reconstructions contain phantoms of gold markers at the upper

110 left corners in XY-slice (circled by dash lines in Fig. 3b) which is caused by the missing wedge
111 artifacts and should not appear here since gold markers are located in different Z sections of the
112 sample. In stark contrast, MBIR in Fig. 3b is able to drastically reduce the missing wedge
113 problem in XZ-slice and YZ-slice, completely suppress the gold marker phantoms in XY-slice
114 and considerably enhance the contrast of biological samples. In addition, MBIR provides better
115 quality of tomogram in other datasets of Fig. 3, which is in a good agreement with the results
116 shown in Fig. 2. In summary, the comparison of slice views among different methods in Fig. 3
117 and Fig. 2 gives a clear impression that MBIR has superior performance in boosting contrast of
118 biological specimens, eliminating halos and streaking artifacts, retaining sharp features, and
119 reducing noise. The superior performance of MBIR is evident in both cryo-ET (Fig. 2 and Fig.
120 3a-c) and plastic-embedded ET (Fig. 3d) datasets.

121

122 **Power spectra evaluation**

123 To quantitatively evaluate MBIR's ability in restoring missing information, we calculated the
124 log-scaled power spectrum of the central XZ-slice and used it as a measurement of information
125 restoration in 3D reconstruction. As depicted in Fig. 4, four plots of power spectra correspond
126 to the central XZ-slices of the tomograms reconstructed by the four methods shown in Fig. 2.
127 It is noted that MBIR can fill more un-sampled region in Fourier space than other methods, not
128 only in the region of the missing wedge but also the empty space between two adjacent tilts,
129 suggesting better performance of MBIR in restoring missing information. It is worth noting that
130 the lines at the corners of BP (Fig. 4a) and SIRT (Fig. 4b) power spectra are due to the aliasing
131 issue. To check if such aliasing issues are unique to our results, we downloaded another four
132 3D tomograms from EMDB ¹², calculated their central XZ-slices power spectra, and plotted
133 them in Supplementary Fig. 1. The results in Supplementary Fig. 1 suggest that this aliasing
134 issue is a general phenomenon in the cryo-ET field since it is observed in a variety of data,
135 including data from multiple research groups, varying TEM facilities and imaging conditions,
136 a diverse range of samples, and different reconstruction software.

137 We next examined the central XZ-slice power spectra of the datasets displayed in Fig. 3 and
138 compared them in Supplementary Fig. 2. In general, MBIR and ICON yield more non-zero
139 values in the missing wedge region than BP and SIRT, except for one challenging dataset
140 (Supplementary Fig. 2b). However, power spectrum may not be a reliable and complete
141 assessment for the information restoration because it only conveys the amplitude information
142 without considering the phase information. What's more, varying filters can be internally
143 applied to tomograms in different methods to balance the non-uniform sampling in Fourier
144 space ¹³. As a result, further validation is still needed to confirm the advantage of MBIR in
145 restoring not only amplitude but also phase information.

146

147 **Cross validation of projections using the leave-one-out FRC method**

148 We used the leave-one-out Fourier ring correlation (FRC) method ¹⁴ to explore the correctness
149 of the information restored by MBIR and compare it with the performance of other
150 reconstruction methods. In this test, the FRC is calculated for the raw tilt image X and the
151 corresponding reprojection \tilde{X}^- from a tomogram computed from all other tilts without tilt X .
152 Here the tilde sign represents the reprojection from a tomogram, and the minus sign represents
153 the tomogram used for reprojection is calculated by omitting the tilt X from the original tilt
154 series to avoid bias. We first excluded a raw image X at a certain tilt angle and utilized the
155 remaining images of the tilt series to generate a tomogram. Next, we re-projected this tomogram
156 at the angle of tilt X to obtain a reprojection \tilde{X}^- . Finally, we calculated the FRC curve between
157 the excluded raw image X and the reprojection \tilde{X}^- , and used this FRC curve as a quantitative
158 evaluation of phase information recovery. As shown in Fig. 5a, the first row and the second
159 row are the raw images X (the first image in each row) and the reprojecions \tilde{X}^- corresponding
160 to different reconstruction methods at a smaller tilt angle 0° and a larger tilt angle 45° ,
161 respectively. The gold markers indicated by white arrows are zoomed in and placed at the lower
162 left corners of each images. It is evident that the gold marker in MBIR reprojecion is circular
163 without discernible distortion or blurring, which is nearly identical to the original tilted image,

164 even at a high tilt angle. In contrast, the gold markers in the reprojections of other methods
165 clearly suffer from missing wedge artifacts including elongation, white halos, and blurring.
166 Furthermore, such visual assessments are verified quantitatively by the FRC (Fig. 5b and c) of
167 the raw tilt images and reprojections shown in Fig. 5a. As shown in Fig. 5b and c, the quick
168 drop of BP (blue curve), SIRT (red curve) and ICON (green curve) FRC curves implies that
169 only low resolution information is reliably restored in the non-sampled angular regions.
170 However, the FRC curve of MBIR exhibits a significantly higher correlation between the
171 reprojection and the original tilt image, confirming the successful restoration of the missing
172 information.

173 To further substantiate the capability of MBIR in restoring missing information, we performed
174 the same analysis as described in Fig. 5 on more datasets and summarized the comparisons of
175 raw images and reprojections in Supplementary Fig. 3-4 and FRC comparisons in Fig. 6. As
176 can be seen from Supplementary Fig. 3-4, MBIR preserved the round shape of gold markers in
177 the leave-one-out reprojections at low (Supplementary Fig. 3) and high (Supplementary Fig. 4)
178 tilt angles in both cryo (Supplementary Fig. 3a-c, Supplementary Fig. 4a-c) and plastic
179 embedded datasets (Supplementary Fig. 3d, Supplementary Fig. 4d), which is consistent with
180 the results shown in Fig. 5a. Fig. 6 shows the FRC comparisons of different methods when 0°
181 (Fig. 6a, c, e, g) and 45° (Fig. 6b, d, f, h) tilts were excluded in the leave-one-out tests,
182 respectively. The FRC curve of MBIR (yellow curve) in Fig. 6 is typically higher than that of
183 other methods, which suggests the superior quality of MBIR in recovering authentic
184 information of biological samples in 3D tomographic reconstructions. As demonstrated in Fig.
185 6a-b, VPP used in this dataset boosts the signal-to-noise ratio of cryo-ET images and improves
186 the low frequency signal in FRC curve compared with Fig. 5b-c, leading to a smaller difference
187 among the results of the four reconstruction methods than the case shown in Fig. 5. However,
188 the local missing wedge artifacts around the gold markers remained in the tomograms
189 reconstructed by the other three methods but not by MBIR for this cryo-ET dataset with VPP
190 as shown in the corresponding slice views (Fig. 3a) and reprojections (Supplementary Fig. 3a

191 and Supplementary Fig. 4a), emphasizing the advantages of MBIR method. Therefore, all the
192 analyses above validate MBIR's capability to partially restore the missing information in both
193 cryo-ET and plastic embedded datasets.

194

195

196 **Discussion**

197 As a widely explored 2D/3D reconstruction method, MBIR has a growing impact on the
198 medical, industrial, and scientific imaging fields. In the present work, we introduced the MBIR
199 method into biological ET and corroborated the substantial advantages of MBIR over current,
200 state-of-the-art reconstruction methods for both cryo and plastic embedded data. MBIR
201 employs a model of the image formation process and combines it with a prior model of the 3D
202 object to formulate a MAP estimation cost function which rejects measurements that do not fit
203 the model. MBIR is finding a fit that balances between generating a reconstruction that matches
204 the data and constraining it to have some properties that any real world object would
205 have. Results on experimental data have effectively demonstrated the excellent performance of
206 MBIR in contrast enhancement, missing wedge artifacts reduction, and missing information
207 restoration, generating visually and quantitatively accurate tomograms.

208 Cryo-ET tomographic reconstruction usually suffers from problems such as high level of noise,
209 poor contrast, artifacts caused by the missing wedge issue and unreliable restoration of missing
210 information, which poses significant challenges to subsequent analysis of the tomograms. The
211 clear benefits of MBIR should not only help achieve better quality reconstruction as shown in
212 this work, but also facilitate further visualization and computational tasks, such as biological
213 feature interpretation, structure segmentation, subtomogram averaging, and ultimately help
214 advance cryo-ET to higher resolution.

215 While MBIR significantly improves tomography quality, the extensive computational load
216 makes its speed slower compared to other approaches (Supplementary Table 1) and restricts
217 the application of MBIR to large datasets. Recently, a computationally optimized algorithm
218 termed Non-Uniform Parallel Super-Voxel (NU-PSV) has been developed for MBIR 3D
219 reconstruction of CT images which enables rapid and massively parallel reconstruction while
220 ensuring fast convergence ¹⁵. Thus, it is desirable to implement this powerful parallel algorithm
221 into cryo-ET MBIR reconstruction in the future, using either GPU or multicore CPUs on
222 multiple computer nodes. Furthermore, MBIR should be generalized to support tomographic

223 reconstruction using double-tilt geometry and incorporate the objective lens contrast transfer
224 function (e.g. defocus, astigmatism, Volta phase shift) into its forward image formation model
225 during its iterative reconstruction process.

226

227 **Methods**

228 **Implementation of MBIR**

229 The MBIR algorithm was implemented as a standalone program using the C++ language by Dr.
230 Charles Bouman's group at Purdue University. The implementation is cross-platform portable
231 and works on Linux, Windows and Mac OS X operating systems. The MBIR software package
232 used for ET is freely available in the form of binary executables and source codes from Dr.
233 Bouman's website (<https://engineering.purdue.edu/~bouman/OpenMBIR/bf-em>). A tutorial of
234 MBIR can be found in the Supplementary Note.

235

236 **Test datasets**

237 We evaluated the performance of MBIR method on both plastic embedded ET dataset and cryo-
238 ET datasets by comparing its results with three reconstruction techniques used in the cryo-ET
239 community, including BP and SIRT available in IMOD ¹⁶, and ICON ¹⁰. The plastic embedded
240 ET dataset obtained from IMOD tutorial website ¹⁶ was originally provided for dual axes
241 reconstruction, but we only used the first tilt series (BBa.st) in our study. Four published
242 experimental cryo-ET datasets (EMPIAR-10037, EMPIAR-10045, EMPIAR-10064 and
243 EMPIAR-10110) were downloaded from the public database EMPIAR ¹⁷. EMPIAR-10064
244 dataset was collected with the Volta phase plate (VPP). These tilt series were aligned based on
245 fiducial gold markers using IMOD and then the same aligned tilt series were reconstructed by
246 the four reconstruction techniques, respectively. The details of these datasets are summarized
247 in Supplementary Table 2 including data type, biological sample, instrument, defocus, tilt
248 scheme, total dose, and data collection software. In this study, the figures used for comparing
249 the performance of different methods are contrast-normalized to avoid subjectivity of
250 observations and to ensure the reliability of comparison.

251

252 **Acknowledgements**

253 This work was supported in part by Showalter Faculty Scholar grant. We thank Ms. Brenda
254 Gonzalez for her assistance in preparation of the manuscript.

255

256 **Author Contributions**

257 R.Y. and W.J. conceived the project and designed the research, analyzed and interpreted data,
258 drafted, revised, and completed final approval of the manuscript. S.V. and C.B. developed the
259 algorithm and implemented the code. J.L. provided some test data. All authors reviewed the
260 manuscript, agreed to all the contents and agreed the submission.

261

262 **Competing Interests**

263 The authors declare no competing financial interests.

264

265

266 **References**

267 1. Lučić, V., Förster, F. & Baumeister, W. STRUCTURAL STUDIES BY ELECTRON
268 TOMOGRAPHY: From Cells to Molecules. *Annu. Rev. Biochem.* **74**, 833–865 (2005).

269 2. Herman, G. T. *Fundamentals of Computerized Tomography: Image Reconstruction from*
270 *Projections*. (Springer, 2009).

271 3. Pilhofer, M., Ladinsky, M. S., McDowall, A. W. & Jensen, G. J. Bacterial TEM: new
272 insights from cryo-microscopy. *Methods Cell Biol.* **96**, 21–45 (2010).

273 4. Vanhecke, D. *et al.* Cryo-electron tomography: methodology, developments and
274 biological applications. *J. Microsc.* **242**, 221–227 (2011).

275 5. Turoňová, B., Marsalek, L., Davidovič, T. & Slusallek, P. Progressive Stochastic
276 Reconstruction Technique (PSRT) for cryo electron tomography. *J. Struct. Biol.* **189**,
277 195–206 (2015).

278 6. Orlova, E. V. & Saibil, H. R. Structural Analysis of Macromolecular Assemblies by
279 Electron Microscopy. *Chem. Rev.* **111**, 7710–7748 (2011).

280 7. Penczek, P., Marko, M., Buttle, K. & Frank, J. Double-tilt electron tomography.
281 *Ultramicroscopy* **60**, 393–410 (1995).

282 8. Chen, Y. *et al.* FIRT: Filtered iterative reconstruction technique with information
283 restoration. *J. Struct. Biol.* **195**, 49–61 (2016).

284 9. Venkatakrishnan, S. V. *et al.* Model-Based Iterative Reconstruction for Bright-Field
285 Electron Tomography. *IEEE Transactions on Computational Imaging* **1**, 1–15 (2015).

286 10. Deng, Y. *et al.* ICON: 3D reconstruction with ‘missing-information’ restoration in
287 biological electron tomography. *J. Struct. Biol.* **195**, 100–112 (2016).

288 11. Venkatakrishnan, S. V. *et al.* A model based iterative reconstruction algorithm for high
289 angle annular dark field-scanning transmission electron microscope (HAADF-STEM)
290 tomography. *IEEE Trans. Image Process.* **22**, 4532–4544 (2013).

291 12. Tagari, M., Newman, R., Chagoyen, M., Carazo, J. M. & Henrick, K. New electron
292 microscopy database and deposition system. *Trends Biochem. Sci.* **27**, 589 (2002).

293 13. Wei, Y., Wang, G. & Hsieh, J. An intuitive discussion on the ideal ramp filter in
294 computed tomography (I). *Comput. Math. Appl.* **49**, 731–740 (2005).

295 14. Cardone, G., Grünewald, K. & Steven, A. C. A resolution criterion for electron
296 tomography based on cross-validation. *J. Struct. Biol.* **151**, 117–129 (2005).

297 15. Wang, X. *et al.* Massively parallel 3D image reconstruction. in *Proceedings of the*
298 *International Conference for High Performance Computing, Networking, Storage and*
299 *Analysis on - SC '17* (2017). doi:10.1145/3126908.3126911

300 16. Kremer, J. R., Mastronarde, D. N. & McIntosh, J. R. Computer visualization of three-
301 dimensional image data using IMOD. *J. Struct. Biol.* **116**, 71–76 (1996).

302 17. Iudin, A., Korir, P. K., Salavert-Torres, J., Kleywegt, G. J. & Patwardhan, A. EMPIAR:
303 a public archive for raw electron microscopy image data. *Nat. Methods* **13**, 387–388
304 (2016).

305

306 **Figure Legends**

307 **Fig. 1. Graphical scheme of the MBIR algorithm.** g denotes the tilt series from cryo-ET, f
308 denotes the unknown structure, and \emptyset denotes unknown nuisance parameters of the system (e.g.
309 noise characteristics) which needs to be determined in the inverse process. $p(\cdot)$ denotes the
310 probability density function and $p(g|f, \emptyset)$ and $p(f)$ are the forward model and prior model in
311 the MBIR algorithm, respectively. \hat{f} and $\hat{\emptyset}$ denote the estimate of f and \emptyset , respectively.

312 **Fig. 2. Comparison of tomograms from an experimental cryo-ET dataset (EMPIAR-
313 10045) reconstructed by BP(a), SIRT (b), ICON (c) and MBIR (d) methods.** The three
314 planes for each method represent the XY-slice (middle plane), XZ-slice (top plane) and YZ-
315 slice (right plane) of the tomogram intersecting at the same gold marker. In each plane, the gold
316 marker is indicated by a white arrow with corresponding zoomed-in view showing the missing
317 wedge artifacts.

318 **Fig. 3. Comparison of tomograms from multiple experimental ET datasets reconstructed
319 by different reconstruction techniques.** Each row indicates the reconstructions from the same
320 dataset using different methods. Each column indicates the reconstructions from the same
321 method applied to different datasets. The data type and EMPIAR ID are denoted at the right
322 side of each row. The method of comparison in each dataset is the same as described in Fig. 2.
323 Note that the XY-slices of the dataset shown in (b) and (d) are used to show the biological
324 sample area and not targeted at the gold markers since the sample and gold markers are located
325 in different Z sections.

326 **Fig. 4. Comparison of the central XZ-slice power spectra from the tomograms shown in
327 Fig. 2.** The tomograms were reconstructed by BP (a), SIRT (b), ICON (c) and MBIR (d)
328 methods, respectively.

329 **Fig. 5. Comparison of missing information restoration from an experimental cryo-ET
330 data (EMPIAR-10045) reconstructed by different reconstruction techniques using the
331 leave-one-out FRC method.** (a) Comparison of reprojections at two tilt angles (0° in the first

332 row and 45° in the second row) using the tomograms generated without the corresponding tilt.
333 The images in the first column are extracted from the tilt series, serving as the ground truth for
334 comparison. In each plane, the gold marker indicted by a white arrow is displayed with
335 corresponding zoomed-in view. (b) and (c) are comparisons of the FRC curves of reprojections
336 against the ground truth as depicted in (a) when 0° and 45° tilt is excluded in the leave-one-out
337 test, respectively.

338 **Fig. 6. Comparisons of FRC curves from multiple experimental ET datasets reconstructed**
339 **by different reconstruction techniques using the leave-one-out FRC method.** Each row
340 represents the comparison of FRC curves from the same dataset when 0° (left plot) and 45°
341 (right plot) tilt is excluded in the leave-one-out test, respectively. The details of the
342 corresponding reprojections and ground truths are shown in Supplementary Fig. 3 (0° excluded)
343 and Supplementary Fig. 4 (45° excluded), respectively.

344 **Supplementary Fig. 1. Observations of aliasing issue in the central XZ-slice power spectra**
345 **of cryo-ET tomograms downloaded from EMDB.** The EMDB ID is marked at the bottom of
346 each image.

347 **Supplementary Fig. 2. Comparison of the central XZ-slice power spectra from the**
348 **tomograms reconstructed by different reconstruction techniques.** Each row shows the
349 power spectra of central XZ-slices from the same dataset using different methods. Each column
350 shows the power spectra from the same method applied to different datasets. The data type and
351 EMPIAR ID are denoted at the right side of each row.

352 **Supplementary Fig. 3. Comparison of reprojections at 0° when the tomograms are**
353 **generated using different reconstruction techniques without the corresponding tilt.** Each
354 row shows the reprojections from the same dataset using different methods. Each column shows
355 the reprojections from the same method applied to different datasets. The data type and
356 EMPIAR ID are denoted at the right side of each row. In each image, the gold marker indicted
357 by a white arrow is displayed with corresponding zoomed-in view.

358 **Supplementary Fig. 4. Comparison of reprojections at 45° when the tomograms are**
359 **generated using different reconstruction techniques without the corresponding tilt.** The
360 details are the same as described in Supplementary Fig. 3.

361 **Supplementary Table 1. Computational speeds of different reconstruction techniques.**
362 The input data size is 400 pixels × 400 pixels × 61 tilts and the output tomogram size is 400
363 pixels × 400 pixels × 128 pixels. The CPU model is Intel(R) Core(TM) i7-6900K @ 3.20GHz.
364 The GPU model is NVIDIA GeForce GTX 1080.

365 **Supplementary Table 2. A summary of datasets used in this study.**

Fig. 1

$$(\hat{f}, \hat{\emptyset}) = \underset{f, \emptyset}{\operatorname{argmax}} \{ \log p(f, \emptyset | g) \} = \underset{f, \emptyset}{\operatorname{argmin}} \{ -\log p(g | f, \emptyset) - \log p(f) \}$$

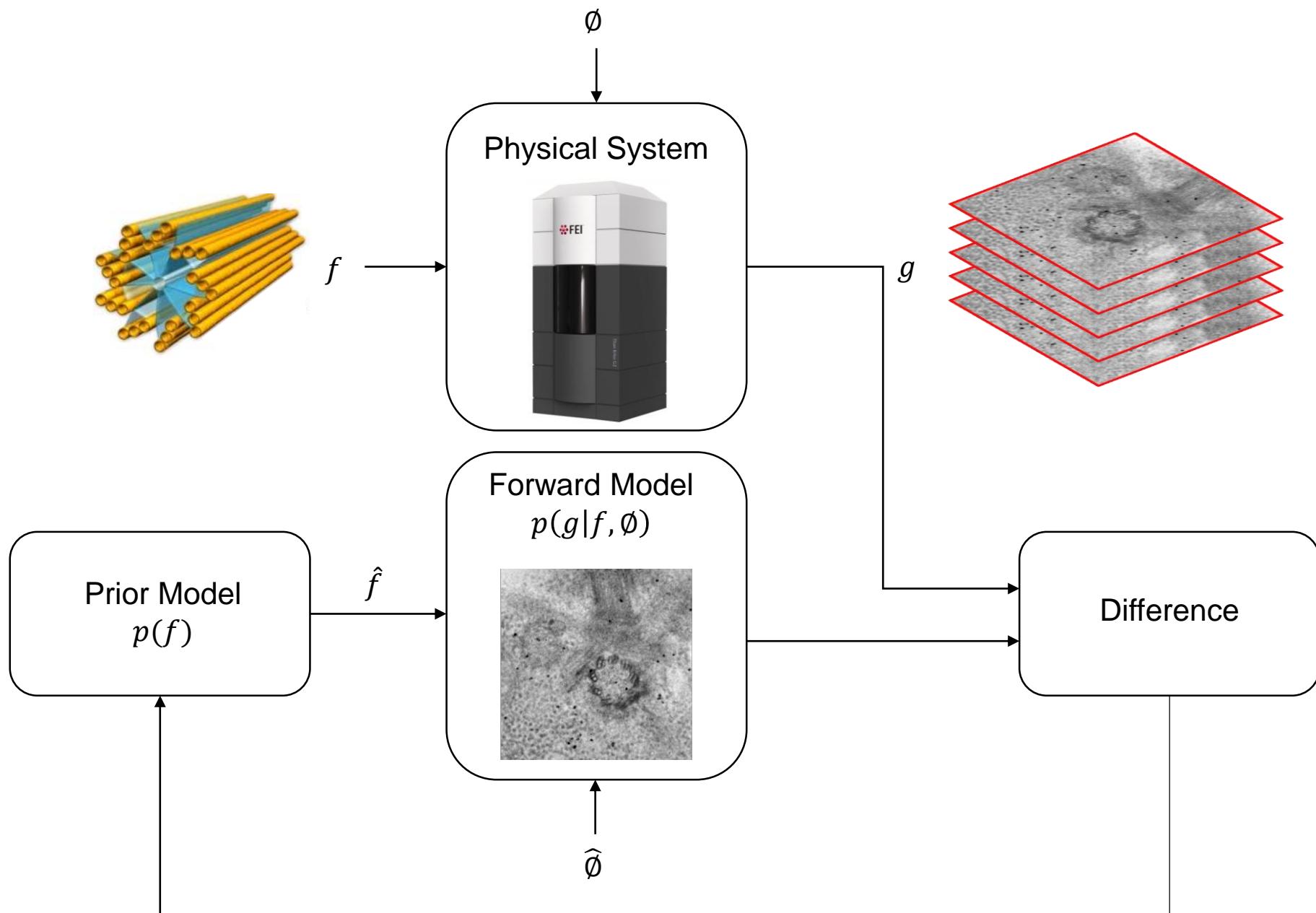


Fig. 2

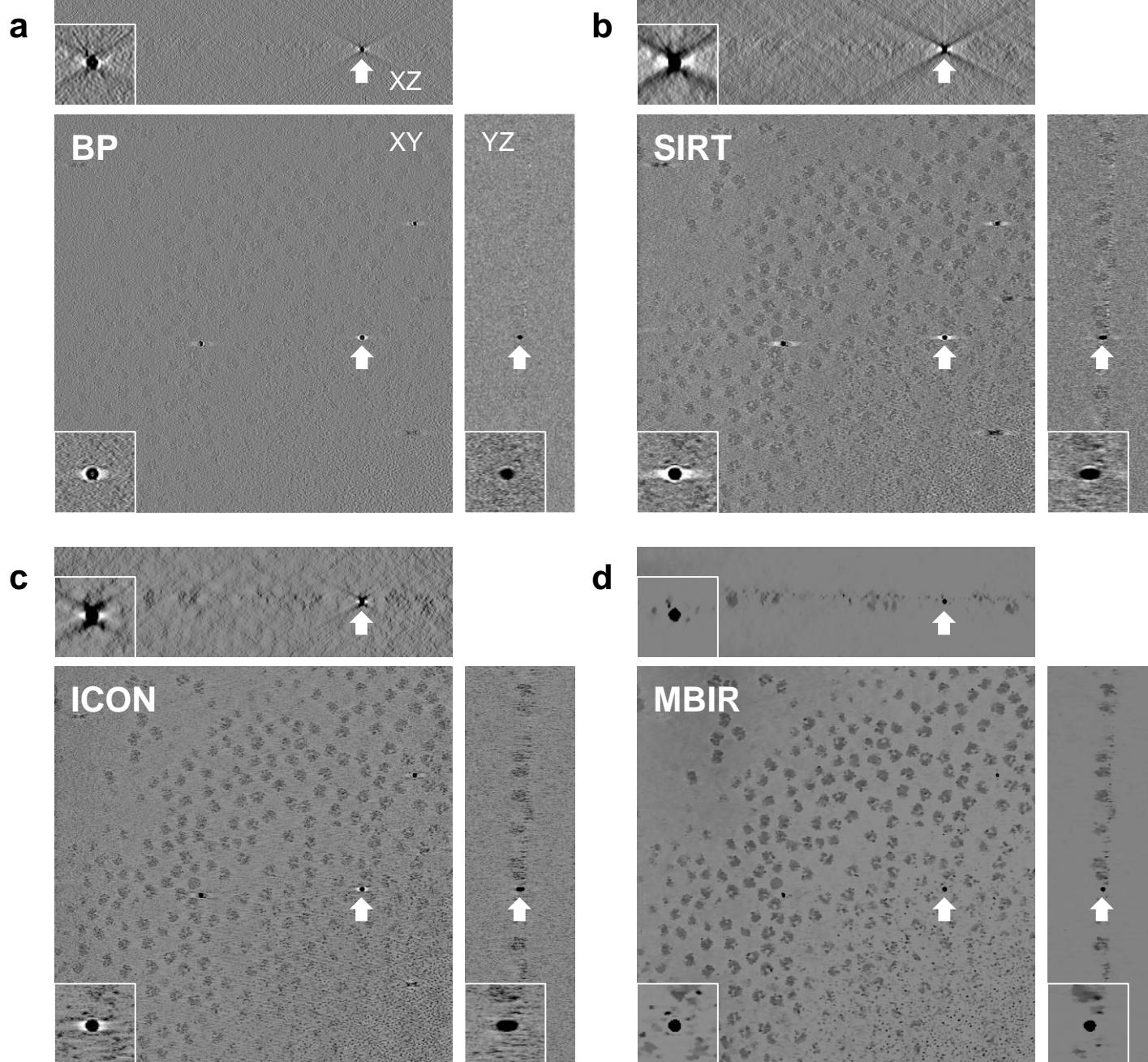


Fig. 3

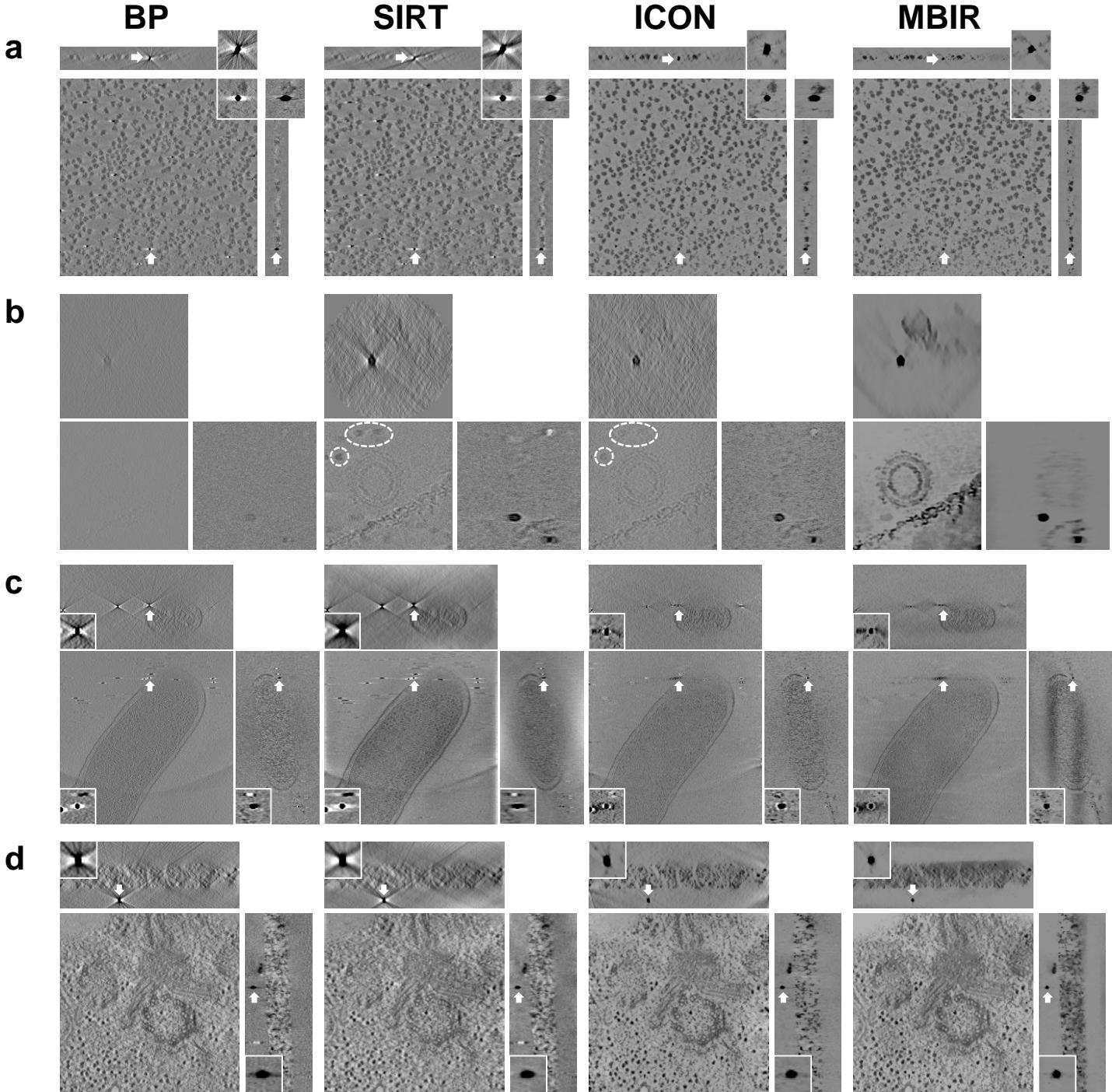


Fig. 4

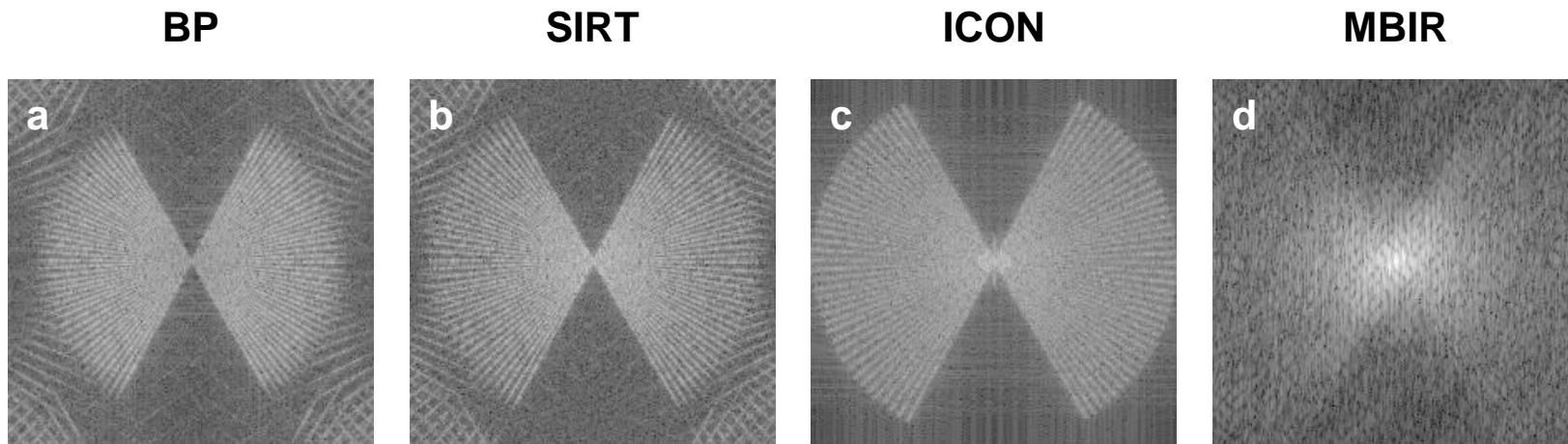


Fig. 5

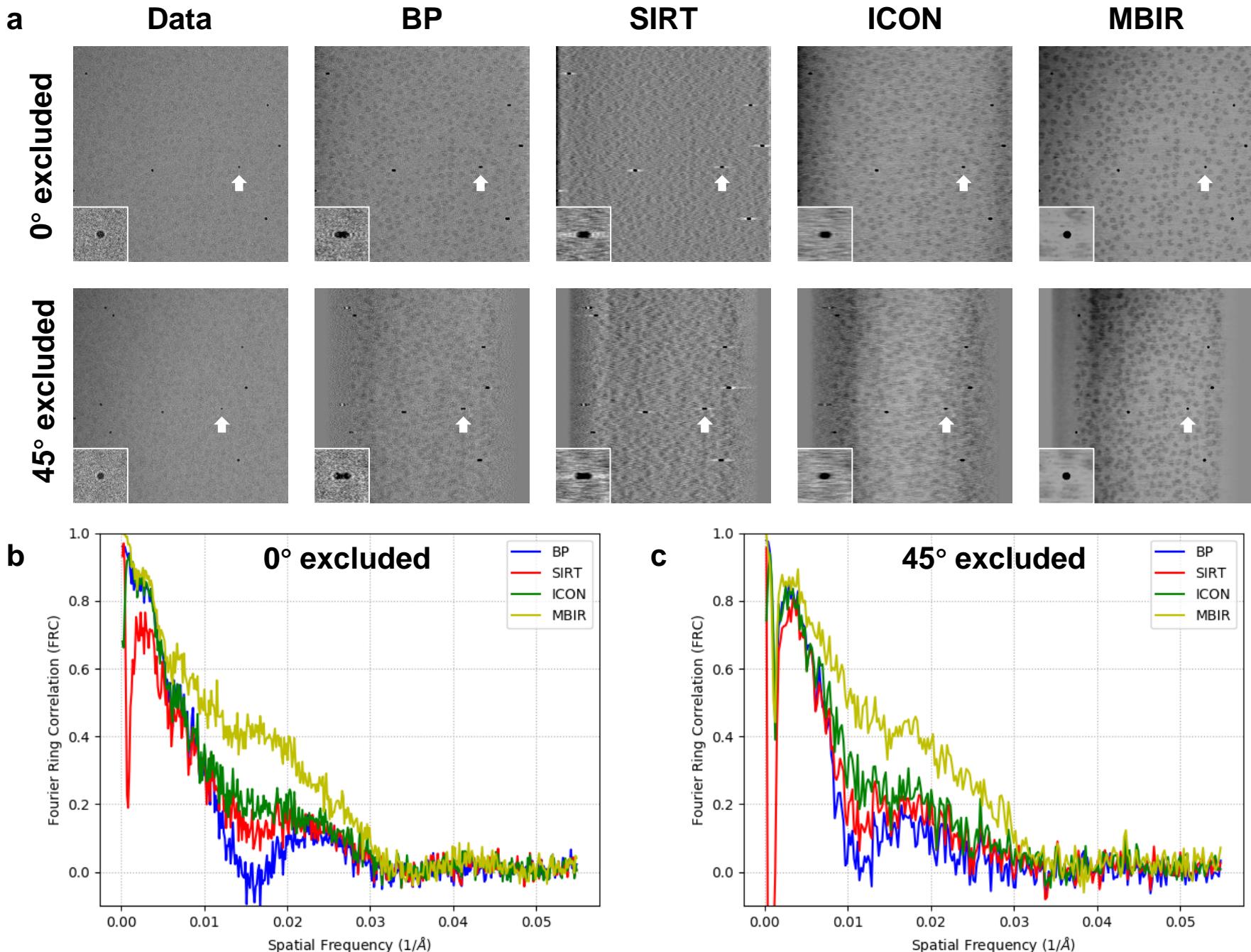
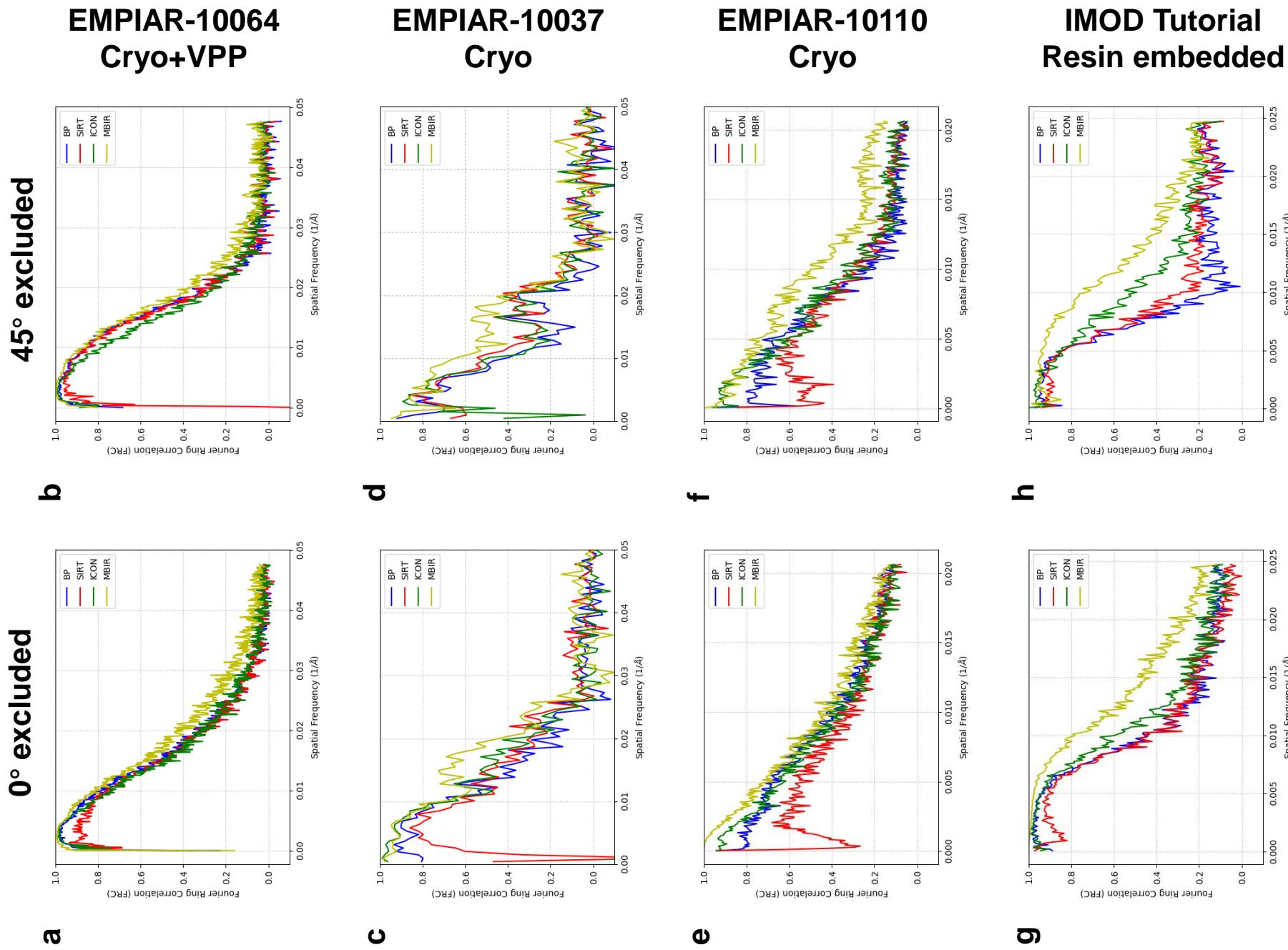


Fig. 6



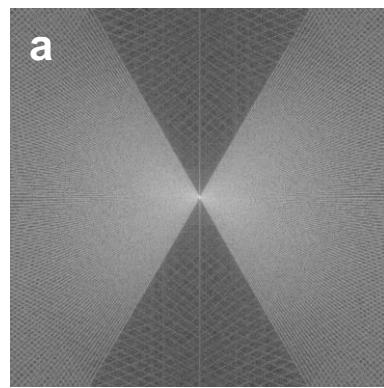
Supplementary

Supplementary Figures

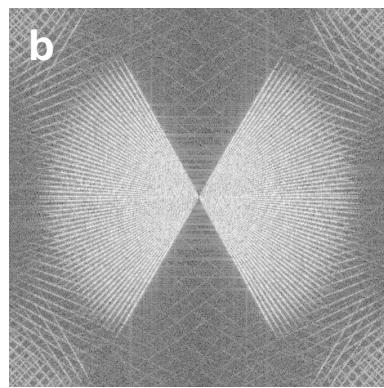
Supplementary Tables

Supplementary Note

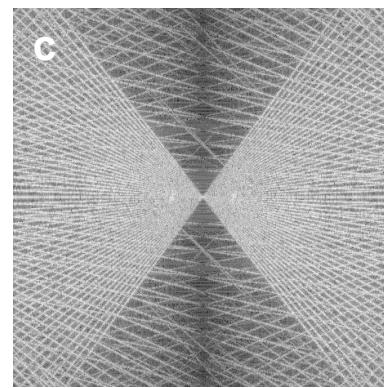
Supplementary Fig. 1



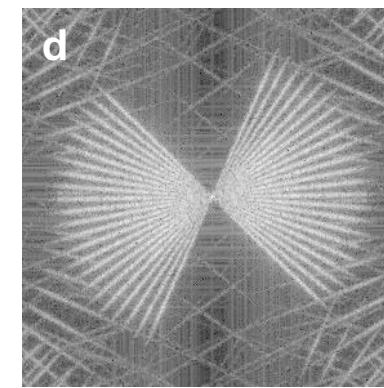
emd-3792



emd-3967

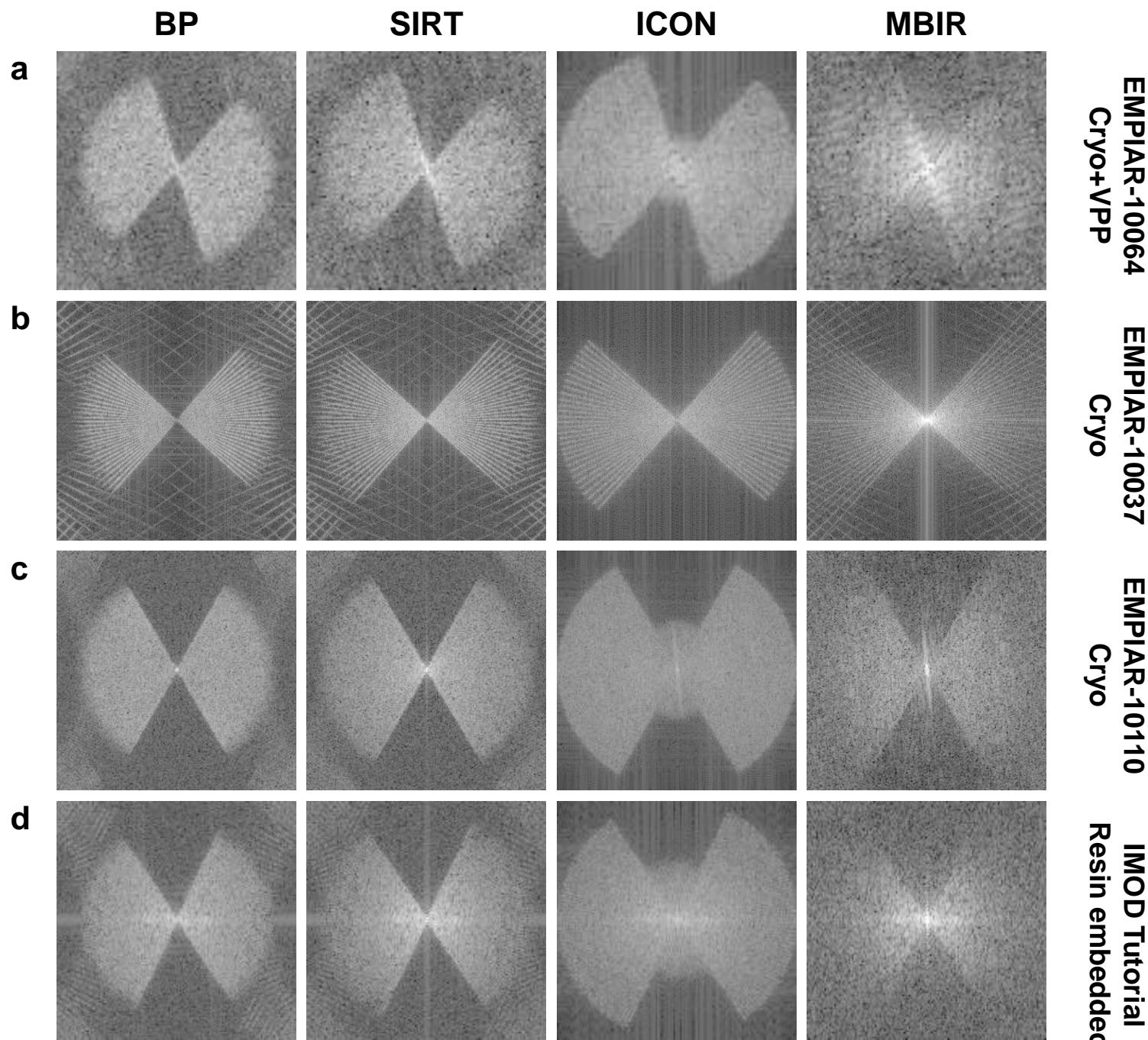


emd-3977

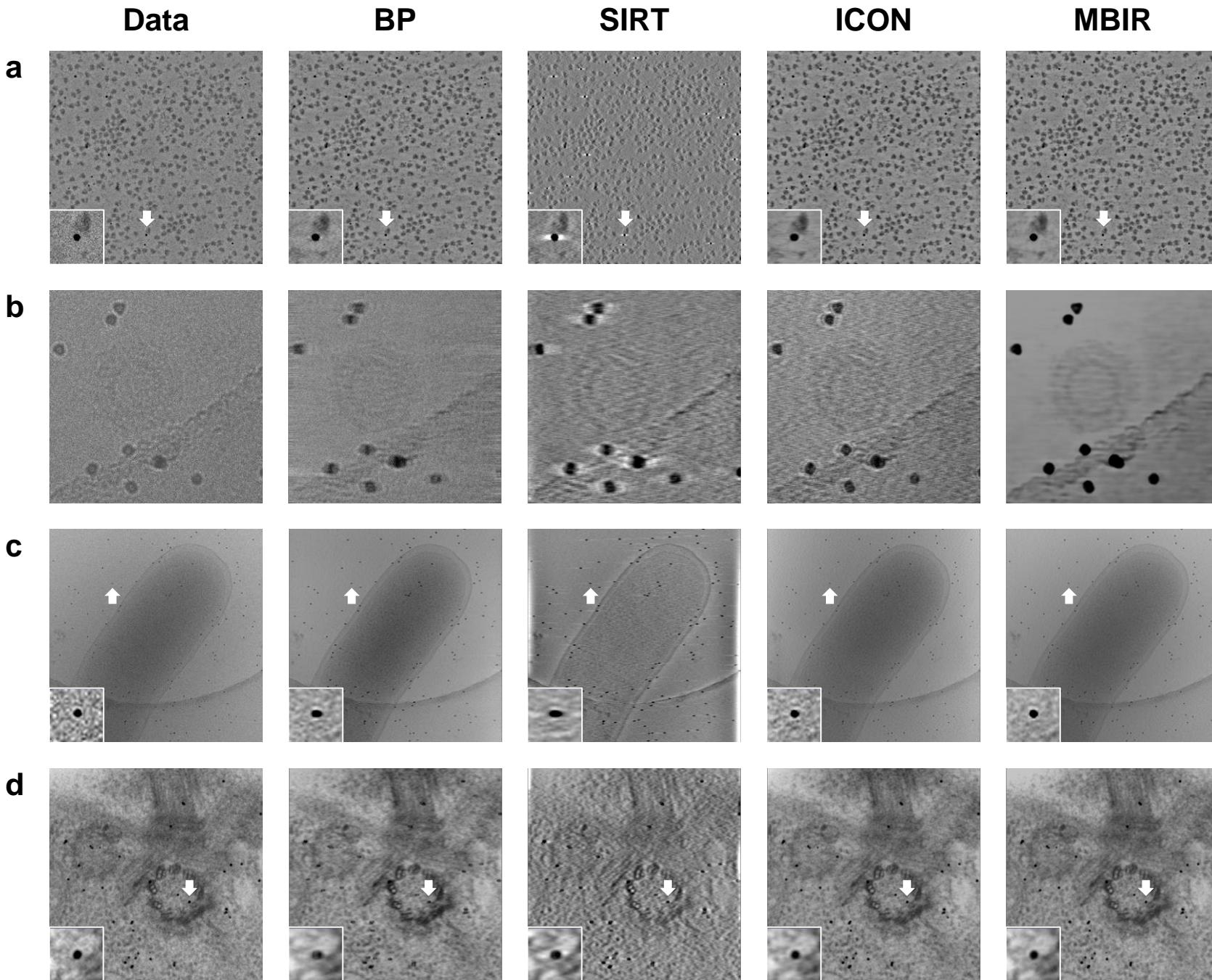


emd-5887

Supplementary Fig. 2



Supplementary Fig. 3



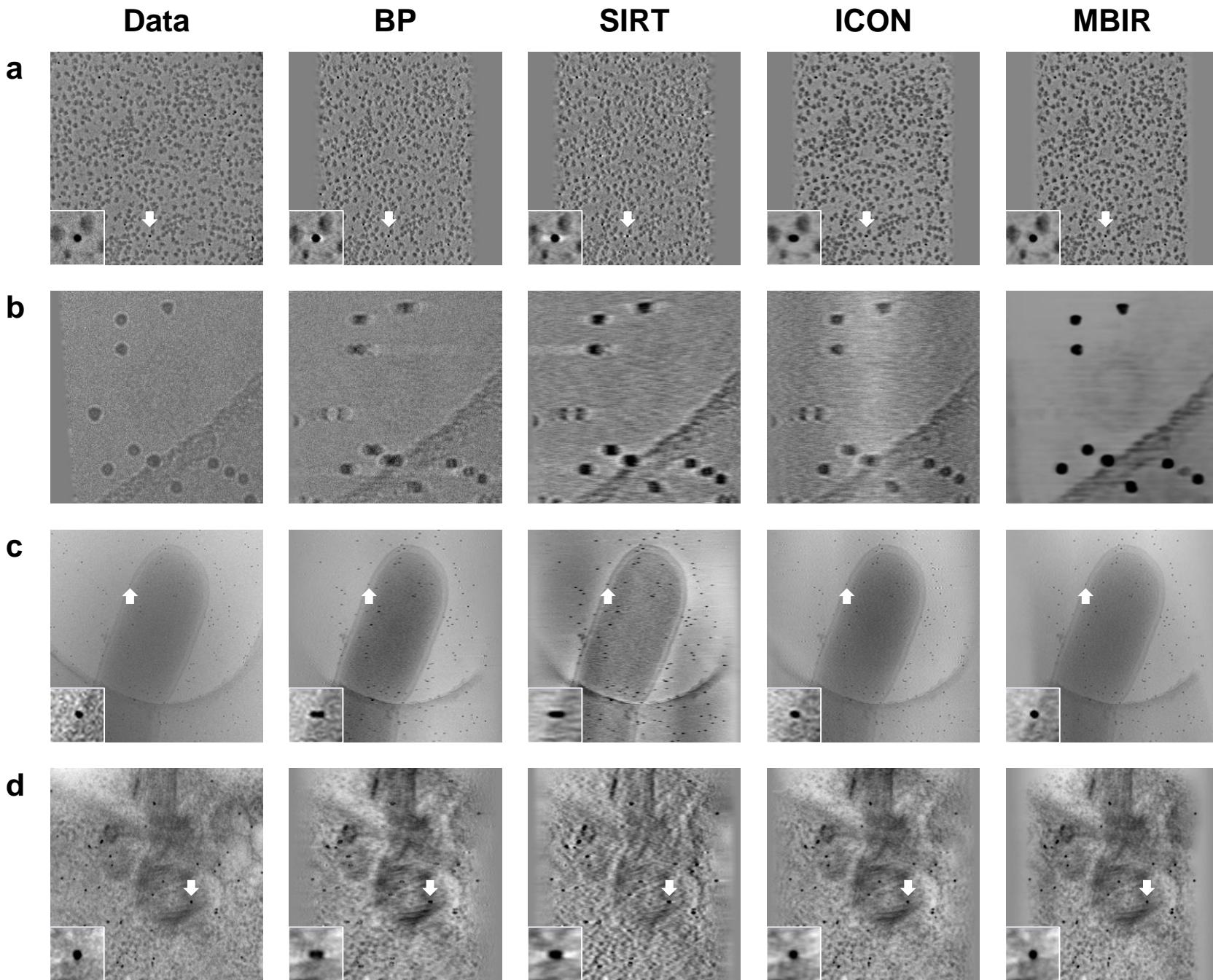
EMPIAR-10064
Cryo+VPP

EMPIAR-10037
Cryo

EMPIAR-10110
Cryo

IMOD Tutorial
Resin embedded

Supplementary Fig. 4



EMPIAR-10064
Cryo+VPP

EMPIAR-10037
Cryo

EMPIAR-10110
Cryo

IMOD Tutorial
Resin embedded

Supplementary Table 1

Method	CPU/GPU	Time (seconds)
BP	1 CPU core	17 s
SIRT	1 CPU core	186 s
ICON	1 GPU	246 s
MBIR	8 CPU cores	1325 s

Supplementary Table 2

	EMPIAR-10045	EMPIAR-10064	EMPIAR-10037	EMPIAR-10110	Imod Tutorial Dataset
Data Type	Cryo	Cryo+VPP	Cryo	Cryo	Plastic embedded
Biological Sample	<i>S. cervisiae</i> 80 S ribosome	80S ribosome	Immature-like Rous-Sarcoma Virus Gag particles	Toxin-coregulated pilus machine	Centriole
Instrument	FEI Titan Krios 300KV; Gatan Quantum Energy Filter; Gatan K2 Summit DDD	FEI Titan Krios 300KV; Gatan K2 Summit DDD	FEI Titan Krios 200KV; GIF2002 Energy Filter; Gatan MultiScan 795 CCD	FEI Polar 300KV; Gatan Energy Filter; Gatan K2 Summit DDD	Gatan Camera on TF30
Defocus	3~5 um	0~200 nm	1.5~5 um	~6 um	Unknown
Tilt Scheme	-60°~+60° Stepsize 3°	-60°~+60° Stepsize 2° First +20°~−60°, then +22°~+60°	-45°~+45° Stepsize 3° First 0°~−45°; then +3°~+45°	-60°~+60° Stepsize 1°	-60°~+60° Stepsize 2°
Total Dose (e/Å ²)	60	90~100	24~34	160	Unknown
Data Collection Software	Serial EM	Serial EM	FEI Tomography Software	UCSF Tomography Software	Serial EM

MBIR tutorial

Step 1: Download MBIR.

<https://engineering.purdue.edu/~bouman/OpenMBIR/bf-em/index.html>

There are two Youtube application tutorials (Basic and Advanced) on MBIR website using one simulated dataset. It is recommended to watch these tutorials first. This protocol is similar to the tutorials and used to show the application of MBIR on cryo-ET dataset based on Linux system.

Step 2: Decompress the downloaded package and activate the GUI.

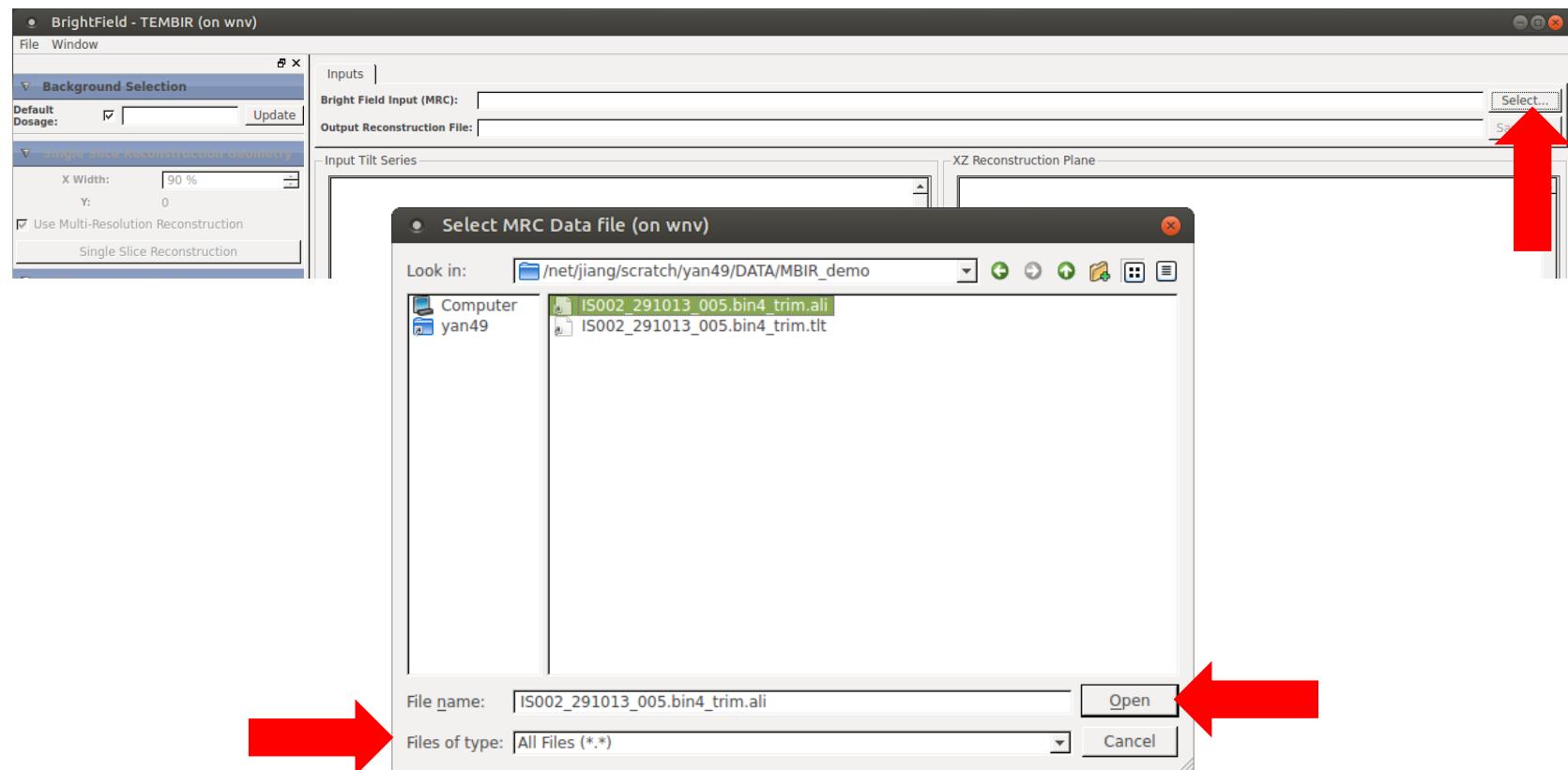
`./OpenMBIR-v2.35-RHEL6-x86_64/bin/BrightFieldGui.sh`

Step 3: Load your aligned tilt series as input.

This software supports aligned input files stored in FEI MRC format.

Click **Select** to load your aligned tilt series. If the file suffix is not default, change **Files of type** to **All files** to access your file.

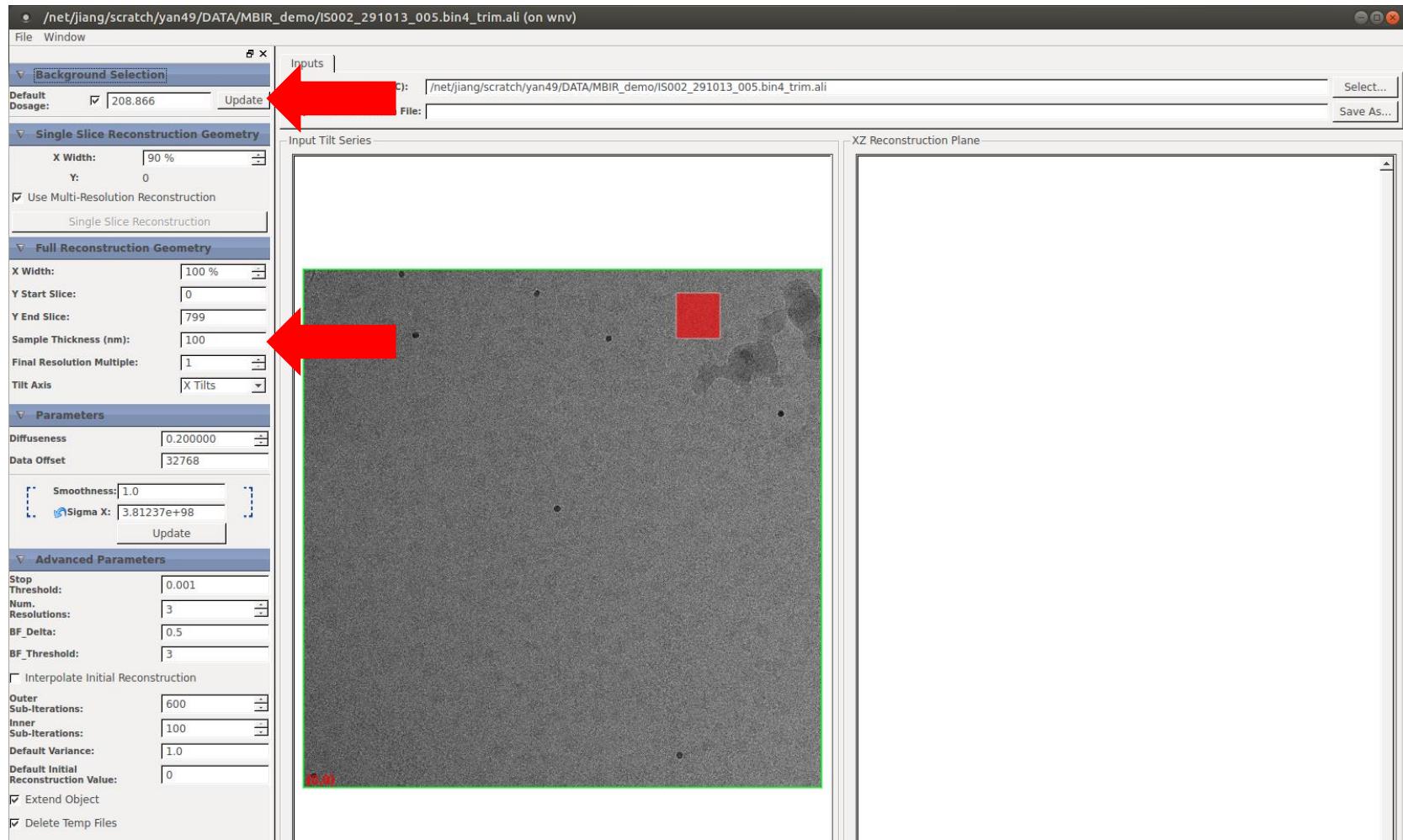
Click **Open** to load your aligned tilt series.



Step 4: Calculate the default dosage.

Move the red box to a background area without sample, adjust its size, then click **Update** in **Background Selection**.

Step 5: Input Sample Thickness (nm) in **Full Reconstruction Geometry**.



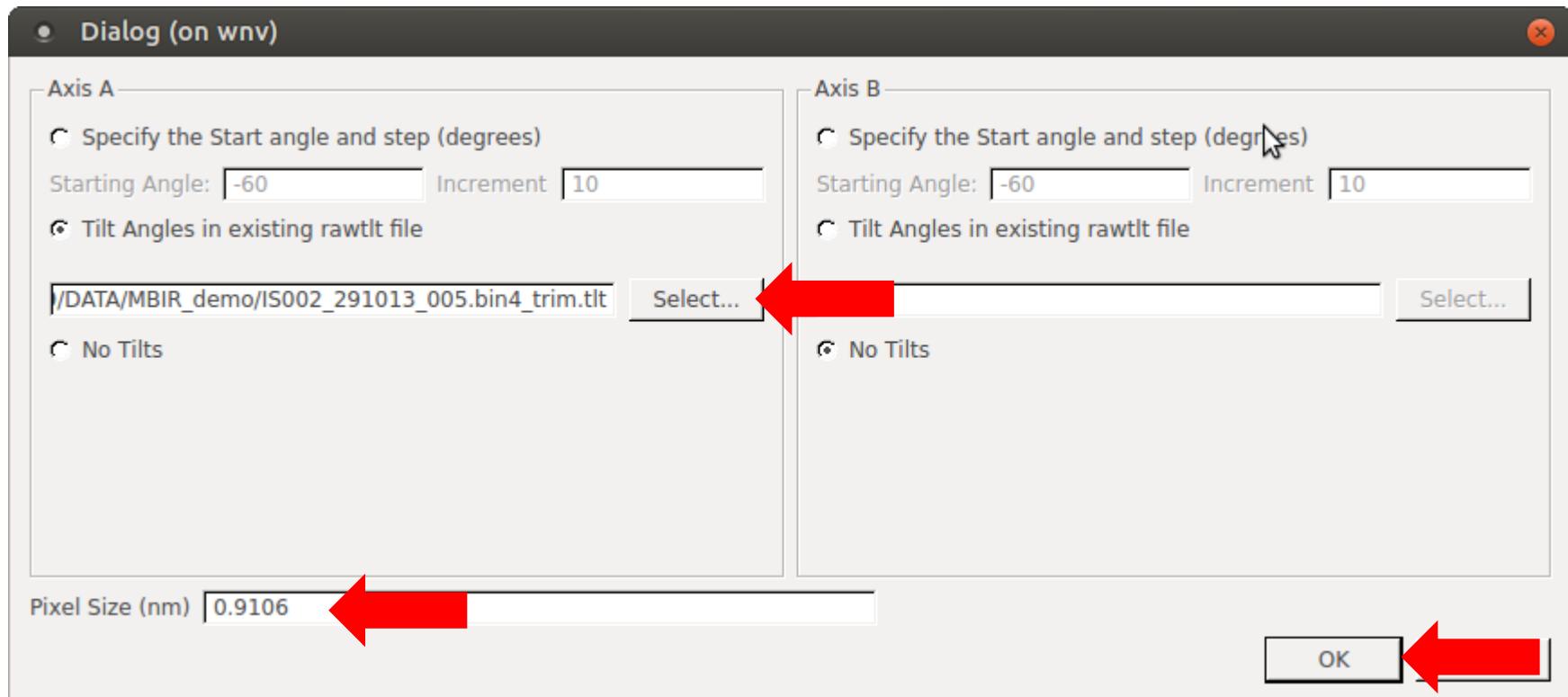
Step 6: Load your tilt angles file and input apix.

Go to upper left corner, select File → Load Tilt Values

In the dialog, click **Select** to load your tilt angle file. If the file suffix is not default, change **Files of type** to **All files** to access your file.

Input **Pixel Size (nm)**.

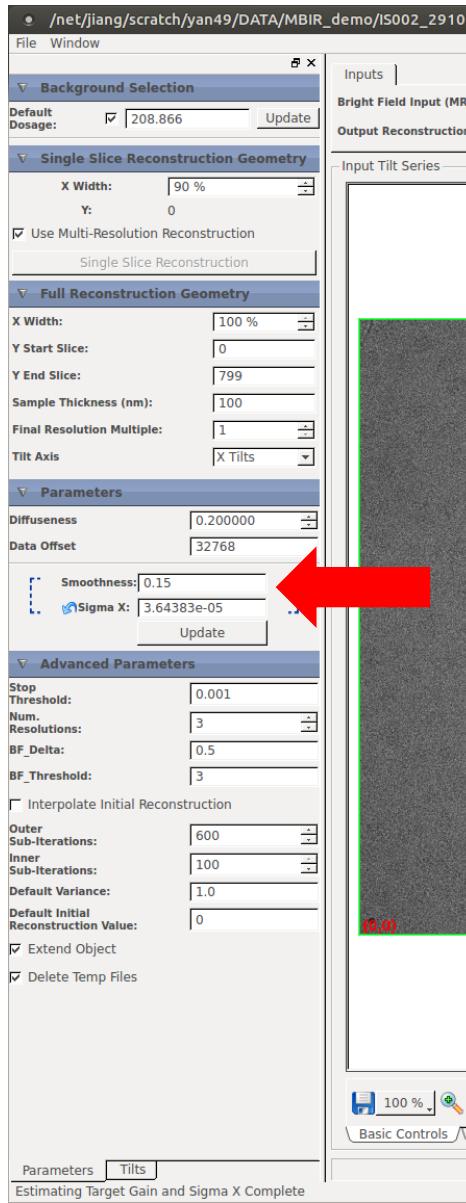
Click **OK** to complete this step.



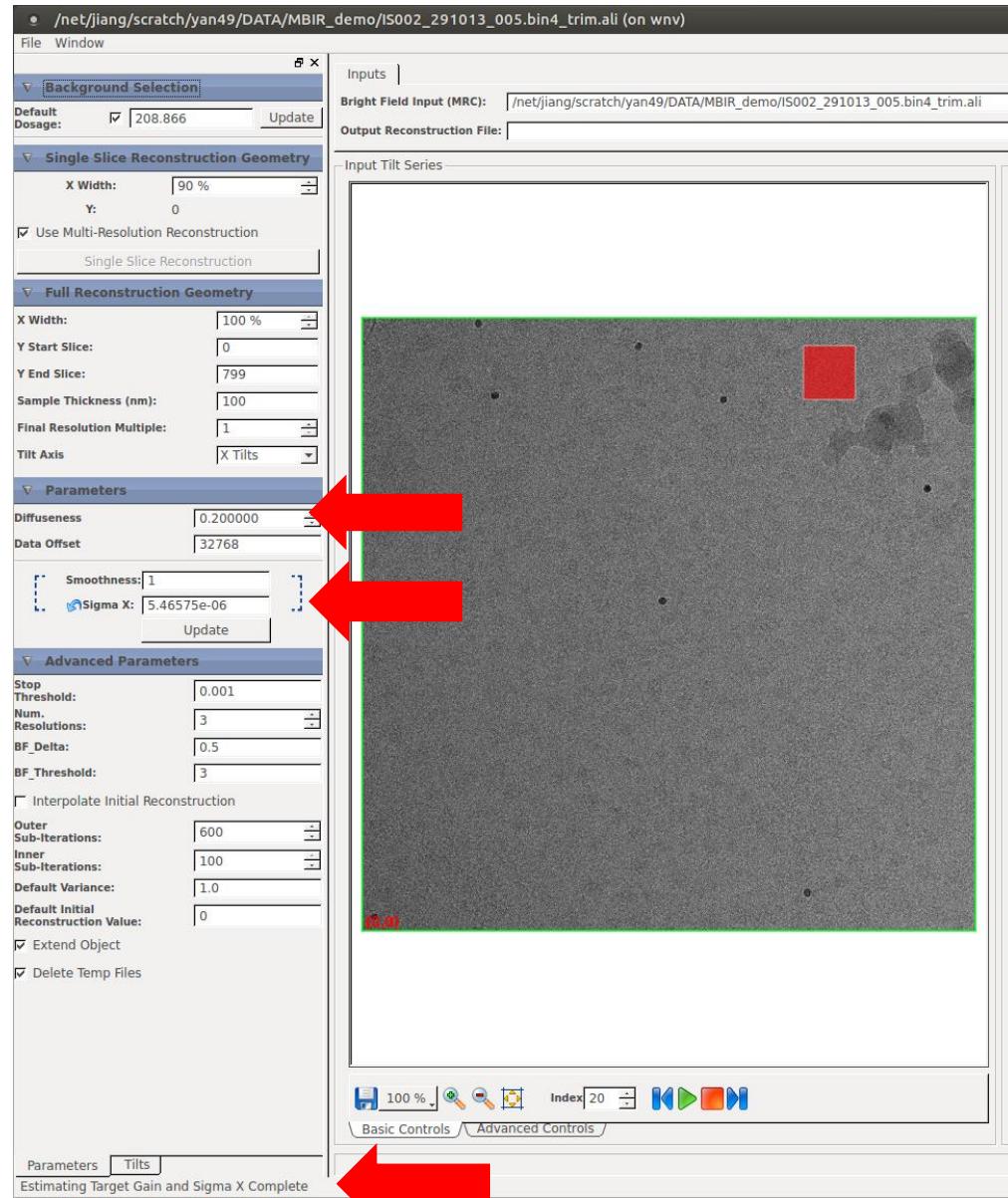
Step 7: Smoothness and diffuseness.

After loading tilt values in step 6, the software will automatically calculate **Sigma X** in **Parameters**. Wait a few seconds until **Estimating Target Gain and Sigma X Complete** appears at the lower left corner of the GUI window. This is equivalent to click **Update** in **Parameters**. If you change the area of interest (e.g. X Width, Y start slice and Y end slice, Sample thickness), you may need to click **Update** in **Parameters** to recalculate the value of **Sigma X**.

In **Parameters**, you will see a **Sigma X** value corresponding to **smoothness 1**. Our empirical value of **smoothness** is 0.15~0.35 for cryo dataset. Here we use 0.15 for this dataset. And the default value of **diffuseness** 0.2 is good for cryo dataset.



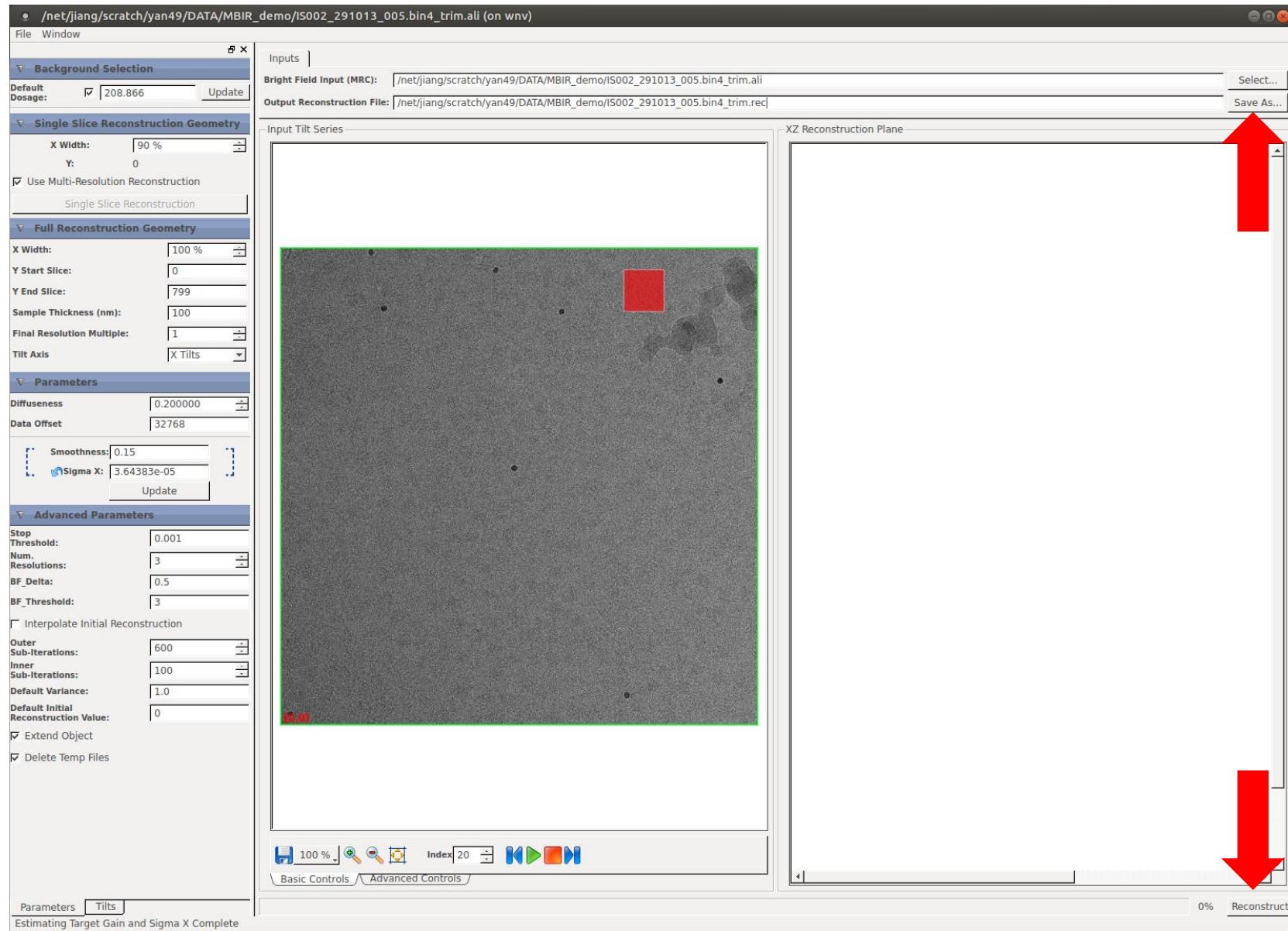
Smoothness 0.15
for cryo-ET



Step 8: Reconstruct your tomogram.

Click **Save As** to save your output tomogram.

Click **Reconstruct** at the lower right corner to start the reconstruction.



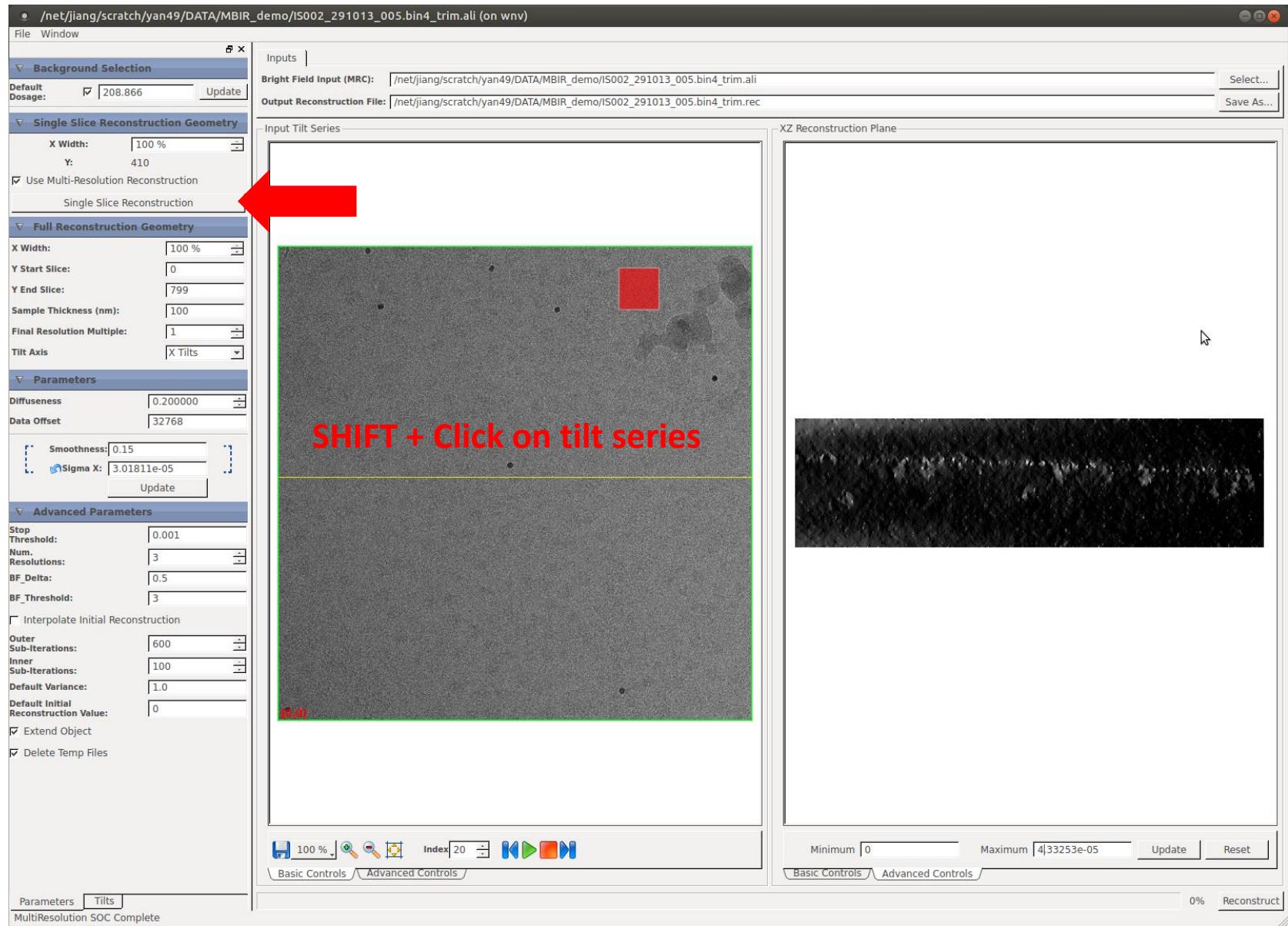
Tips:

If you do not know which smoothness is optimal for your dataset, you can either use **Single Slice Reconstruction Geometry** or reconstruct a subarea of your tilt series.

If you use **Single Slice Reconstruction Geometry**, hold SHIFT key and click somewhere on your input file, you will see a yellow line, indicating where you want to make a single XZ slice reconstruction.

Click **Single Slice Reconstruction** in **Single Slice Reconstruction Geometry** and wait several seconds. You will see a single XZ slice reconstruction shown in the right window. At this time, you can only view this single XZ slice here but not save it.

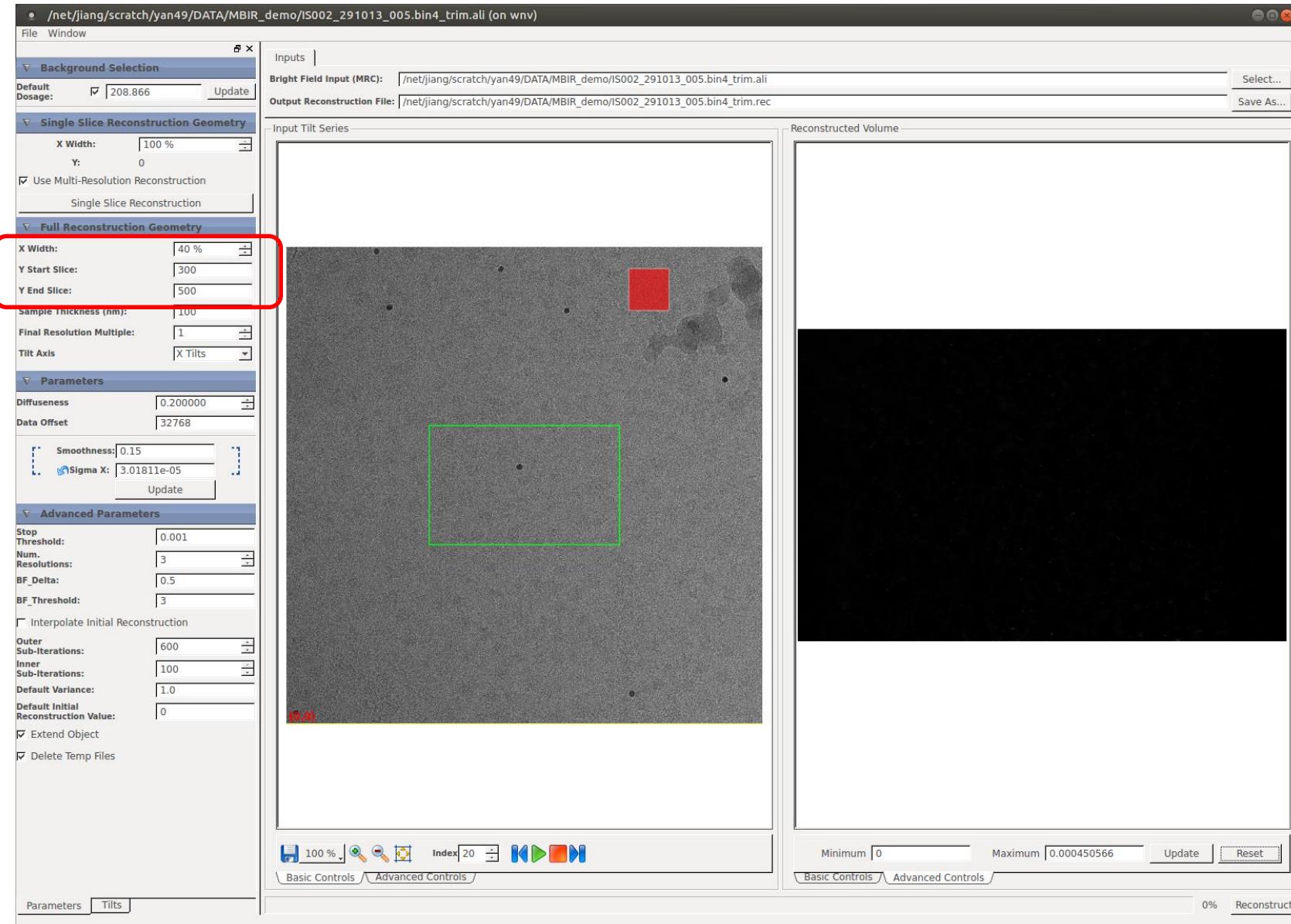
You can adjust the **smoothness** value to reconstruct different single slices and determine the optimal **smoothness** for your dataset.



It is also recommended to reconstruct a small area to determine the optimal **smoothness**. You can adjust the top and bottom edges of the outer green box to find the **Y start slice** and **Y end slice** of your target area. When you adjust the top and bottom edges of the green box, you will see the corresponding change of the numbers in **Y start slice** and **Y end slice** in **Full Reconstruction Geometry**. However, the range of X coordinate can only be adjusted using **X Width** in **Full Reconstruction Geometry** in order to guarantee the selected area is symmetric to the tilt axis.

When you change the area of interest (e.g. X Width, Y start slice and Y end slice, Sample thickness), you may need to click **Update** in **Parameters** to recalculate the value of **Sigma X**. Click **Save As** to save your trial tomogram.

Click **Reconstruct** at the lower right corner to start the reconstruction.



Here your trial tomogram is saved to the provided directory. You can use IMOD or other visualization software to open it and examine the quality of reconstruction.

