

Heteromeric GABA_A receptor structures in positively-modulated active states

Paul S. Miller^{1,12*}, Simonas Masiulis^{1,2,12*}, Tomas Malinauskas¹, Abhay Kotecha^{1,13}, Shanlin Rao³, Sreenivas Chavali², Luigi De Colibus¹, Els Pardon^{4,5}, Saad Hannan⁶, Suzanne Scott^{1,2}, Zhaoyang Sun^{1,2}, Brandon Frenz⁹, Gianni Klesse^{3,7}, Sai Li¹, Jonathan M. Diprose¹⁰, C. Alistair Siebert¹¹, Robert M. Esnouf¹⁰, Frank DiMaio⁹, Stephen J. Tucker^{7,12}, Trevor G. Smart⁶, Jan Steyaert^{4,5}, M. Madan Babu², Mark S. P. Sansom^{3,12}, Juha T. Huiskonen^{1,8}, A. Radu Aricescu^{1,2,12}

¹Division of Structural Biology, Wellcome Centre for Human Genetics, University of Oxford, Oxford OX3 7BN, United Kingdom.

²MRC Laboratory of Molecular Biology, Francis Crick Avenue, Cambridge Biomedical Campus, Cambridge CB2 0QH, United Kingdom.

³Department of Biochemistry, University of Oxford, Oxford OX1 3QU, United Kingdom.

⁴Structural Biology Brussels, Vrije Universiteit Brussel (VUB), Brussels, Belgium.

⁵VIB-VUB Center for Structural Biology, VIB, Brussels, Belgium.

⁶Department of Neuroscience, Physiology and Pharmacology, University College London, London WC1E 6BT, United Kingdom.

⁷Clarendon Laboratory, Department of Physics, University of Oxford, Oxford OX1 3PU, United Kingdom.

⁸Helsinki Institute of Life Science and Molecular and Integrative Biosciences Research Programme, Faculty of Biological and Environmental Sciences.

⁹Department of Biochemistry, University of Washington, Seattle, Washington, USA.

¹⁰Wellcome Centre for Human Genetics, University of Oxford, Oxford OX3 7BN, United Kingdom.

¹¹Electron Bio-imaging Centre, Diamond Light Source, Didcot OX11 0DE, United Kingdom.

¹²OXION Initiative in Ion Channels and Disease, University of Oxford, Oxford OX1 3PT, United Kingdom.

¹³Current address: Materials and Structural Analysis, Thermo Fisher Scientific, Eindhoven, Netherlands.

*These authors contributed equally to this work.

Correspondence and requests for materials should be addressed to A.R.A. (radu@mrc-lmb.cam.ac.uk), P.S.M. (paul@strubi.ox.ac.uk) or J.T.H. (juha@strubi.ox.ac.uk).

Type-A γ -aminobutyric acid (GABA_A) receptors are pentameric ligand-gated ion channels (pLGICs), typically consisting of $\alpha/\beta/\gamma$ subunit combinations. They are the principal mediators of inhibitory neurotransmission throughout the central nervous system and targets of major clinical drugs, such as benzodiazepines (BZDs) used to treat epilepsy, insomnia, anxiety, panic disorder and muscle spasm. However, the structures of heteromeric receptors and the molecular basis of BZD operation remain unknown. Here we report the cryo-EM structure of a human $\alpha 1\beta 3\gamma 2$ GABA_AR in complex with GABA and a nanobody that acts as a novel positive allosteric modulator (PAM). The receptor subunits assume a unified quaternary activated conformation around an open pore. We also present crystal structures of engineered $\alpha 5$ and $\alpha 5\gamma 2$ GABA_AR constructs, revealing the interfacial site for allosteric modulation by BZDs, including the binding modes and the conformational impact of the potent anxiolytic and partial PAM, bretazenil, and the BZD antagonist, flumazenil. These findings provide the foundation for understanding the mechanistic basis of GABA_AR activation.

The common architecture of eukaryotic pLGICs is now established, with at least one crystal or cryo-EM structure determined for the cation-selective nicotinic acetylcholine (nAChR) and serotonin type 3 receptors (5HT₃R), and the anion-selective GABA_A, glycine (GlyR), and GluCl receptors¹⁻⁹. Each subunit extracellular domain (ECD) of 200-250 amino acids comprises an N-terminal α -helix and ten β -strands forming a twisted β -sandwich. Each TMD comprises an α -helical M1-M4 bundle, the M3 and M4 helices being connected by a large intracellular loop of 85-255 residues¹⁰. The two orthosteric neurotransmitter binding sites are situated between ECD principal (P) and complementary (C) faces, which intercalate around a water-filled vestibule that funnels into the ion channel transmembrane pore lined by five M2 helices. The structures have brought considerable understanding of the molecular basis of

receptor function in homomeric receptor formats. However, heteromeric pLGIC structures have only been solved for nAChRs^{1,2}. This is a key limitation given that the vast majority of mammalian pLGICs are heteromers possessing heightened complexity both molecularly and pharmacologically.

The GABA_AR family comprises 19 different subunit subtypes: α 1-6, β 1-3, γ 1-3, δ , ϵ , θ , π and ρ 1-3¹¹. Selective neuronal expression of particular subtypes, along with preferential assembly rules, ensure that the majority of GABA_ARs in the human brain comprise 2 α -subunits, 2 β -subunits and 1 γ -subunit, with α 1, β 2, β 3, and γ 2 subtypes exhibiting widespread overlapping expression profiles^{12,13}. Alternative subtypes perform more specialised roles, for example, the α 5 subtype influences cognition¹⁴ and moderation of α 5 β 2/3 γ 2 receptors ameliorates animal model diseases of autism and Down syndrome, associated with cognitive deficits^{15,16}. Modulation of α 5 β 2/3 γ 2 receptors also improves recovery after stroke in a rodent model¹⁷. The homomeric GABA_AR structures so far solved, GABA_AR β 3 and chimaeras that include α subunit TMDs, possess five-fold symmetry and will possess five identical copies of agonist and anaesthetic binding pockets^{18,19}. In contrast, $\alpha\beta\gamma$ GABA_ARs possess two orthosteric GABA binding sites at β - α interfaces between the ECDs, two non-GABA binding α - β and γ - β interfaces, and one BZD binding site located at the α - γ interface²⁰. Similarly, within the TMDs, $\alpha\beta\gamma$ GABA_ARs have at least three non-equivalent potential anaesthetic binding pockets. In the case of GABA_AR β 3, solved in a desensitized state, each subunit is bound by agonist and the subunit conformations are indistinguishable⁵. However, it is not known if different subunits within a heteromer adopt equivalent conformations in the activated state.

PAMs, such as BZDs, the intravenous general anaesthetics propofol and etomidate, barbiturates, endogenous neurosteroids, and alcohol (ethanol), bind GABA_ARs to promote their

activation and channel opening, and thereby enhance GABAergic signalling¹⁸⁻²⁵. This makes BZDs essential treatments for hyper-excitability disorders such as anxiety, insomnia and epilepsy²⁴. However, these agents lack receptor subtype selectivity and cause unwanted sedation, addiction, and motor and cognitive impairment²⁶. Agents with improved selectivity can ameliorate these side effects and allow for more selective targeting to treat other neurological disorders, including autism, Down syndrome, neuropathic pain, schizophrenia and stroke^{15-17,27,28}. Structures of heteromeric GABA_ARs are essential to understand the molecular basis of inhibitory neurotransmission and drug subtype selectivity, and are anticipated to expedite the delivery of selective therapeutics against these disorders.

RESULTS

Structures of GABA_AR constructs

Our initial goal was to obtain high resolution structures of the α and γ human GABA_AR subunits. In addition to the β subunit structure we previously reported⁵, these are the essential constituents of the principal heteromeric receptor subtypes in the CNS²⁹. Moreover, the α/γ interfaces harbour the BZD allosteric site³⁰⁻³⁷, which is of major pharmacological importance. Iterative rounds of screening and engineering of $\alpha 5$ subunits for pentamer monodispersity and yield identified a construct with 12 residue swaps from the $\beta 3$ subunit, which readily forms homopentamers⁵, and 11 residue swaps from the BZD site complementary (C) face of the $\gamma 2$ subunit. This construct recapitulated 100 % residue identity within the BZD site to the wild type $\alpha 5$ - $\gamma 2$ subunit interface (construct designated $\alpha 5_{\text{HOM}}$; site designated BZD_{HOM}; **Supplementary Fig. 1a-d**)³⁵⁻³⁷. Of particular note, an Asn114Gly substitution ($\beta 3$ subunit glycine) to remove an N-linked glycosylation site that faces the extracellular vestibule and restricts homomeric assembly of α -subunits (observed in the cryo-EM construct – discussed below) was crucial to achieving monodispersity and pentamerisation (**Supplementary Fig.**

1e). We solved its structure to 2.6 Å resolution in complex with the drug, flumazenil³⁸ (Anexate), a BZD site antagonist used to treat overdose (Fig. 1a, b and Table. 1). Notably, although $\alpha 5_{\text{HOM}}$ bound the BZD radioligands ³H-flunitrazepam and ³H-flumazenil, the affinity was 60-fold and 150-fold lower than for wild-type $\alpha 5\beta 3\gamma 2$ receptors (Supplementary Fig. 2a-e). To solve the structure of a BZD site with higher affinity we performed further screening and engineering on this background for heteromeric combinations containing $\gamma 2$ subunits. These were transfected at 9:1 ratios of $\alpha 5$ to $\gamma 2$ DNAs to favour one $\gamma 2$ per pentamer. This identified a construct formed from $\alpha 5_{\text{HOM}}$ subunits containing 4 additional residue swaps from the $\beta 3$ subunit, and a chimera subunit comprising the $\gamma 2$ ECD and $\alpha 1$ TMD, designated $\alpha 5\gamma 2_{\text{HET}}$. We crystallised this construct in complex with the BZD partial PAM, bretazenil³⁹ and solved its structure to 2.5 Å resolution (Fig. 1c, d; Table 1). In contrast to $\alpha 5_{\text{HOM}}$, inclusion of the chimeric $\gamma 2$ subunit resulted in an additional higher apparent affinity site, designated BZD_{HET} (as well as the lower affinity BZD_{HOM} sites), which had only 2-fold, 5-fold and 15-fold lower affinity for ³H-flunitrazepam, ³H-flumazenil and bretazenil respectively, compared to wild-type receptors (Supplementary Fig. 2a-f).

Attempts to crystallize tri-heteromeric GABA_ARs were unsuccessful, therefore we generated new constructs for single-particle cryo-electron microscopy. We designed $\alpha 1$ and $\gamma 2$ subunit constructs in which only the M3-M4 intracellular loops were substituted by a short linker, SQPARAA⁴⁰, and $\beta 3$ subunits in which the M3-M4 intracellular loop was substituted by a thermostabilised apocytochrome b562RIL (BRIL)⁴¹ domain for unambiguous subunit identification. These constructs were co-expressed and assembled into functional $\alpha 1\beta 3\gamma 2$ GABA_ARs, designated $\alpha 1\beta 3\gamma 2_{\text{EM}}$ (Supplementary Fig. 3). Only the $\gamma 2$ subunit possessed a purification tag, in order to guarantee its presence in the pentamers. To facilitate particle

alignment, we raised nanobodies against GABA_ARs and selected one of these, Nb38, that binds to $\alpha 1$ subunits. We obtained a 5.17 Å (FSC=0.143) cryo-EM map of $\alpha 1\beta 3\gamma 2_{EM}$, in the presence of a saturating 1 mM concentration of GABA (**Fig. 1e-f** and **Supplementary Figs. 4 and 5; Table 2**). Two Nb38 molecules were bound between non-adjacent ECD interfaces, consisting of $\alpha 1(P)$ faces and neighbouring $\beta 3(C)$ and $\gamma 2(C)$ faces, respectively, validating $\alpha 1$ subunit inclusion and receptor stoichiometry. At this resolution, the TMD α -helices of $\alpha 1$ and $\beta 3$ subunits showed helical turns and side-chain densities for several large hydrophobic amino acids, although β -sheets were not fully separated into strands (**Supplementary Fig. 6**). The N-linked glycans served as markers that further allowed unambiguous subunit identification. Mannose branching could be clearly seen, for example, at $\alpha 1$ Asn111 and $\beta 3$ Asn149. The detergent belt, BRILs and nanobody edges were largely disordered (**Supplementary Fig. 5g**). The $\gamma 2$ subunit TMD was also less ordered, but otherwise the local resolution for the rest of the pentamer was consistent throughout (**Supplementary Figs. 5g and 6**). Viewed from above, the cryo-EM map confirms a clockwise subunit arrangement of $\alpha 1$ -A, $\beta 3$ -B, $\gamma 2$ -C, $\alpha 1$ -D, $\beta 3$ -E, consistent with previous indirect studies^{42,43}. Despite the low resolution of this map, availability of high resolution crystal structures of individual subunits from $\alpha 5\gamma 2_{HET}$ and the previously solved GABA_AR- $\beta 3_{cryst}$ ⁵ allowed us to build a model of the heteromeric $\alpha 1\beta 3\gamma 2_{EM}$ receptor (**Fig. 1g; Table 2**). Inclusion of BRIL domains between the M3-M4 helices of the $\beta 3$ subunit did not distort these helices (RMSD = 0.98 Å between $\alpha 1\beta 3\gamma 2_{EM}$ $\beta 3$ and GABA_AR $\beta 3_{cryst}$ across 54 M3-M4 equivalent C $_{\alpha}$ positions).

The BZD binding site and BZD binding modes

The ECD pockets between $\alpha 5_{HOM}$ and $\alpha 5\gamma 2_{HET}$ subunits contained large positive peaks in the *Fo*–*Fc* electron density maps, which were unambiguously assigned to the co-crystallisation

ligands flumazenil and bretazenil, respectively (**Supplementary Fig. 2i-l**). For $\alpha 5\gamma 2_{\text{HET}}$ the electron density map revealed inclusion of a single $\gamma 2$ chimera subunit per pentamer, distinguishable by its unique side chains and glycosylation sites. This confirms the transfection ratio of 9 $\alpha 5$:1 $\gamma 2$ ensured a pure stoichiometric population, as required to enable crystallization, and for radioligand binding studies. The lower apparent affinity of the BZD_{HOM} sites did not affect the ligand binding mode which was the same as at the BZD_{HET} site (**Supplementary Fig. 2m, n**). Furthermore, the binding modes of both flumazenil (observed only in BZD_{HOM} sites in $\alpha 5_{\text{HOM}}$) and bretazenil are similar (**Figure 2a-h**). BZDs such as flunitrazepam (which lack the imidazo C(3)-linked ester moiety and instead possess a diazepine C(6) phenyl ring) exhibit a distinct pharmacology from flumazenil and bretazenil because they require a histidine residue in the BZD site of $\alpha 1/2/3/5$ subtypes for binding^{32,44}. Radioligand binding on a modified $\alpha 5\gamma 2_{\text{HET}}$ construct, $\alpha 5\gamma 2_{\text{HET}\Delta}$, with BZD_{HOM} sites removed (by reversing the $\gamma 2$ substitutions in the $\alpha 5$ subunits), so that only the BZD_{HET} site remained, revealed that a His105Arg substitution ablated ³H-flunitrazepam binding but retained the same apparent affinity for flumazenil (**Supplementary Fig. 2g, h**). Thus, the differential impact of the His to Arg substitution on the binding of distinct classes of BZDs to wild type receptors is reproduced in this construct.

Intermolecular binding contacts between the ligands and receptor occur predominantly through van der Waals (vdW) interactions, which span the interface between subunits. From the α -subunit (P)-face, five hydrophobic residues make vdW contacts with the ligand benzene A-rings: Phe103 and His105 from the $\beta 4$ strand (historically named loop A¹⁰); Tyr163 from the $\beta 7$ - $\beta 8$ loop (loop B); Ile206 and Tyr213 from the $\beta 9$ -10 hairpin (loop C) (**Fig. 2 a-h** and **Supplementary Fig. 1**). Of particular importance are Tyr163, which forms T-shaped π -stacking interactions with the A and I ligand rings, and Tyr213, forming parallel π -stacking

interactions with the A rings. Consistent with such interactions, Tyr163Phe and Tyr213Phe substitutions retain BZD potentiation whilst Ser substitutions reduce sensitivity at least 10-fold³³. The structures also explain why moving the flumazenil C(8) fluorine to C(9) or C(10) chlorine or adding a C(1) methyl substituent (Compounds 1-3 in **Supplementary Fig. 7a-d**) reduce binding by over 100-fold⁴⁵⁻⁴⁷. These substituents clash with loop B Tyr163, as shown by *in silico* docking studies (**Supplementary Fig. 7a-d**). The lowest free energy binding mode of flumazenil overlays the one observed experimentally in $\alpha 5_{\text{HOM}}$, whereas compounds 1-3 are displaced by ~ 2 Å away from Tyr163, impairing the T-shaped π -stacking interactions. In contrast, the C8 azide derivative Ro15-4513⁴⁸, a competitive BZD antagonist developed as an antidote to alcohol and more recently used as a PET ligand⁴⁹, docked similarly to flumazenil, with the azide positioned under the loop C Tyr213, which it chemically photolabels⁵⁰ (**Supplementary Fig. 7e**).

The loop-A His105 undertakes a ligand-dependent reorientation within the site (**Fig. 2b-d, f-h**), its side chain rotating about the C $_{\alpha}$ -C $_{\beta}$ bond to accommodate either the bretazenil bromine or the flumazenil chlorine atoms (at adjacent positions on the A rings, **Fig. 2a, e**). As shown by the radioligand data (**Supplementary Fig. 6g, h**), a His105Arg substitution does not affect flumazenil binding even though it ablates flunitrazepam binding. The explanation for this is observed in the $\gamma\text{P}/\alpha\text{C}$ site in $\alpha 5\gamma 2_{\text{HET}}$, where the equivalent residue to His105 is $\gamma 2$ Arg114, which is well accommodated under the bretazenil ligand (**Supplementary Fig. 7i, j**).

From the (C)-face, the $\beta 2$ strand (loop D) phenylalanine side chain ($\gamma 2$ Phe77 in the BZD $_{\text{HET}}$ site, $\gamma 2^*$ Phe68 in the BZD $_{\text{HOM}}$ site) forms π -stacking interactions with the ligand I rings (**Fig. 2c, g**). These explain why a Phe77Ile substitution reduces flumazenil affinity 1000-fold and why GABA $_{\text{A}}$ receptors containing the $\gamma 1$ subunit, where an Ile residue occupies the equivalent

position, are much less responsive to BZDs³⁶. Above Phe77, an alanine residue ($\gamma 2$ Ala79 in the BZD_{HET} site, $\gamma 2^*$ Ala70 in the BZD_{HOM} site) demarcates the top of the BZD site and faces the I ring C(3) substituent, consistent with a previous mutagenesis study predicting close apposition of these two elements⁵¹ (**Fig. 2d, h**). The neighbouring $\beta 6$ strand (loop E) threonine ($\gamma 2$ Thr142 in the BZD_{HET} site, $\gamma 2^*$ Thr133 in the BZD_{HOM} site) forms putative hydrogen bonds with both the imidazole nitrogen and ester carbonyl of the ligands, consistent with original pharmacophore models proposing a dual H-bond contribution from these moieties⁵² (**Fig. 2d, h**). The ester carbonyl group is essential for flumazenil binding versus ketone or ether functionalities⁴⁶, whilst other derivatives that maintain the isosteric constraints retain high affinity binding^{52,53}.

Patients on BZDs can experience a variety of adverse events, including cognitive and psychomotor effects, tolerance and, in some cases, paradoxical behaviours such as disinhibition leading to aggression²⁶. To explore whether there is genetic variation within the BZD site, which may contribute to response variability in individuals, we mapped allelic variants of 138,632 unrelated healthy humans (gnomAD database⁵⁴) for $\alpha 1$ -6 and $\gamma 1$ -3 subunit residues within 5 Å of ligand bound to $\alpha 5\gamma 2_{\text{HET}}$ and $\alpha 5_{\text{HOM}}$ (**Fig. 3a-c**). Although no common allelic variants were found, many rare allelic variants (ranging from 1 in 1,000 to 1 in 100,000 people) were identified, spanning the four major physiological BZD site subtypes, $\alpha 1\gamma 2$, $\alpha 2\gamma 2$, $\alpha 3\gamma 2$ and $\alpha 5\gamma 2$. The $\alpha 5$ subunit Gly161Val African variant is predicted to sterically disrupt the folding of loop B reducing ligand-binding affinity, as shown previously for the orthosteric GABA site reducing agonist binding 400-fold⁵⁵. Several African, East Asian and European Non-Finnish variants cluster at loop A His105, including $\alpha 1$ subunit tyrosine or arginine substitutions and $\alpha 3$ subunit tyrosine or asparagine substitutions, all of which have been shown to reduce sensitivity to classical benzodiazepines 10-20-fold⁵⁶. Finally, the loop C tip (Ser209-

Thr210-Gly211 in the $\alpha 5$ subunit) experiences the highest incidence of variation in all four BDZ binding $\alpha 1/2/3/5$ subunits. Substitutions within this region have previously been shown to reduce sensitivity to potentiation by classical BZDs and the sedative zolpidem, a non-BZD that binds the BZD site, by 5-10-fold^{34,57}. Whilst it might be expected that reduced binding will result in a suboptimal patient response to BZDs, for some of these allelic variants the responses to anxiety or epilepsy treatment might actually be improved. The GABA_AR $\alpha 1$ subunit is linked to unwanted sedative and addictive side effects⁵⁸ so individuals possessing $\alpha 1$ His102 substitutions might, intriguingly, experience less side effects.

Nb38 is a novel positive allosteric modulator

While the cryo-EM map resolution limits our ability to define precise side-chain interactions, it is clear that major receptor-Nb38 contact points exist between: the CDR2/3 loops and $\beta 9/\beta 10$ strands and $\beta 9$ - $\beta 10$ hairpin (loop C), with CDR2 inserting into the pocket under loop C; CDR1-3 loops and the $\beta 6/\beta 7$ strands and $\beta 6$ - $\beta 7$ loop (Cys-loop); CDR1/2 loops and the (C)-face $\beta 1/\beta 2$ strands and $\beta 8/\beta 9$ loop (loop F) (**Fig. 4a, b** and **Supplementary Fig. 8a, b**). Nb38 binding affinity (K_D) for detergent-solubilised $\alpha 1\beta 3\gamma 2_{EM}$ receptors, as determined by surface plasmon resonance (SPR), was increased 6.5-fold in the presence of 1 mM GABA, from 1.61 nM to 248 pM, suggesting that it favours binding and stabilises an activated receptor conformation (**Supplementary Fig. 8c, d**). Whole cell patch-clamp recording confirmed this. 10 μ M Nb38 strongly potentiated EC₁₀ GABA currents by 480 ± 30 % ($n = 7$) for $\alpha 1\beta 3\gamma 2_{EM}$ and 290 ± 20 % ($n = 7$) for wild-type (WT) $\alpha 1\beta 3\gamma 2$ (**Fig. 4c**), greater than achieved by the BZD diazepam, 180 ± 20 % ($n = 12$) for $\alpha 1\beta 3\gamma 2_{EM}$ and 130 ± 20 % ($n = 9$) for $\alpha 1\beta 3\gamma 2_{WT}$ (**Supplementary Fig. 3b**). Application of Nb38 alone at concentrations up to 10 μ M had only a weak direct agonist effect (**Fig. 4c**). Importantly, Nb38 was $\alpha 1$ subunit selective, eliciting no potentiation of wild-

type $\alpha 2$ - $\alpha 6\beta 3\gamma 2$ receptors (**Fig. 4d**). Thus, Nb38 is a novel pharmacological tool with superior efficacy and selectivity for $\alpha 1$ subunit receptors over benzodiazepines. The Nb38 ability to potentiate GABA_ARs was also observed for spontaneous inhibitory post-synaptic currents (sIPSCs) from dentate gyrus granule cells (DGGCs) in acute hippocampal slices, which primarily stem from $\alpha 1$ -subunit containing receptors⁵⁹. Without significantly affecting sIPSC frequency or rise-time, 2 μ M Nb38 increased amplitude $37 \pm 9 \%$ ($p < 0.05$) and prolonged decay times by $96 \pm 19 \%$ ($p < 0.05$) to increase GABAergic drive (in terms of charge transfer calculated as the average sIPSC surface area) by $94 \pm 14 \%$, $n = 4$ ($p < 0.01$) (**Fig. 4e, f**). By comparison, 500 nM diazepam also significantly prolonged sIPSC decay time, although to a lesser extent ($39 \pm 10 \%$, $p < 0.01 \%$), and did not significantly increase GABAergic drive.

Impact of N-linked glycosylation on pentamerization

The $\alpha 1\beta 3\gamma 2_{EM}$ cryo-EM map revealed multiple N-linked glycans attached to the $\alpha 1$ (Asn111, $\beta 5$ - $\beta 5'$ loop), $\beta 3$ (Asn80, $\beta 3$ -strand and Asn149, $\beta 7$ -strand) and $\gamma 2$ (Asn208, $\beta 9$ -strand) subunits (**Fig. 1c**). Of particular note, the two $\alpha 1$ Asn111 glycans occupy the ECD vestibule and, unexpectedly, adopt well-ordered conformations (**Fig. 5a**). The $\alpha 1$ -D glycan projects upwards into the extracellular space (**Fig. 5b**). In contrast, the $\alpha 1$ -A glycan projects horizontally to form putative CH- π interactions between the pyranose ring of a mannose moiety and the apposing Trp123 side chain from the $\gamma 2$ $\beta 5$ - $\beta 5'$ loop (**Fig. 5c, d**). This glycosylation site is conserved across all α -subunits ($\alpha 1$ -6) and Trp123 is conserved across all γ -subunits ($\gamma 1$ -3), but not β -subunits (**Fig. 5e**). Thus, this glycan pairing is expected to exist in all $\alpha\beta\gamma$ GABA_ARs in the human brain, a feature absent from all previously solved pLGIC structures¹⁰. Enzymatic glycosylation of $\alpha 1$ Asn111 precedes exit from the endoplasmic reticulum (ER)⁶⁰ and requires access to the inner face of $\alpha 1$, meaning that it must precede assembly and closure

of the pentameric ring. The non-random orientation induced by the $\alpha 1$ -A glycan interaction with $\gamma 2$ Trp123 suggests that pentamerization comes from pre-assembled $\alpha\beta\gamma$ trimers with a horizontal glycan, followed by the addition of a second $\alpha\beta$ dimeric unit (**Fig. 5f**). Furthermore, the Asn111 glycosylation represents a stoichiometric control mechanism, preventing the inclusion of more than two α -subunits via steric hindrance. Accordingly, we could only obtain the pentameric $\alpha 5_{\text{HOM}}$ and $\alpha 5\gamma 2_{\text{HET}}$ constructs described earlier after mutating the Asn111 residue (**Supplementary Fig. 1e**).

The extracellular region conformations

Superpositions of $\alpha 5_{\text{HOM}}$, $\alpha 5\gamma 2_{\text{HET}}$ and GABA_AR- $\beta 3_{\text{cryst}}$ ⁵ single ECDs, reveal that all three subunits comprising the major synaptic heteromer format ($2\alpha_n-2\beta_n-1\gamma_n$) adopt highly similar β -sandwich arrangements (RMSD in the 0.59-0.76 Å range between different chains over 208 equivalent C α positions), with a notable divergence in the flexible $\beta 8$ - $\beta 8'$ and $\beta 8'$ - $\beta 9$ loops (loop F) (**Supplementary Fig. 8e**). Global superposition of the $\alpha 5_{\text{HOM}}$, $\alpha 5\gamma 2_{\text{HET}}$ and $\alpha 1\beta 3\gamma 2_{\text{EM}}$ ECD pentamers reveal that all the subunits have adopted positions relative to the pore axis similar to agonist-bound GABA_AR- $\beta 3_{\text{cryst}}$ and the homologous agonist-bound glycine receptor⁶ (GlyR), rather than antagonist-bound GlyR (RMSD in the 0.7-1 Å range versus GABA_AR- $\beta 3$; 1.0-1.4 Å versus agonist-bound GlyR; 2.1-2.3 Å versus antagonist-bound GlyRs, over 208 equivalent C α positions) (**Supplementary Fig. 8f-j**). The activated ECD bases swing out from the pore allowing it to widen while the loop C tips close inwards. Of note, the same assumed ECD positions between subunits means the $\alpha 1\beta 3\gamma 2_{\text{EM}}$ BZD site is highly similar to the BZD sites of $\alpha 5_{\text{HOM}}$ and $\alpha 5\gamma 2_{\text{HET}}$ (**Fig. 6a-c**). Consistent with this, *in silico* docked flumazenil and bretazenil assumed the same binding modes in $\alpha 1\beta 3\gamma 2_{\text{EM}}$ (**Fig. 6d-e**). In the case of $\alpha 5_{\text{HOM}}$ and $\alpha 5\gamma 2_{\text{HET}}$ this means the polar Thr208-Ser209-Thr210 residues at the tip of loop C sterically

trap BZD in the site (**Fig. 2b, f**). The tight closure is incompatible with the inverted concave tetracyclic ring series of the (*R*)-bretazenil enantiomer⁴⁷, as confirmed by *in silico* docking in which the lowest energy binding mode of (*S*)-bretazenil recaptures the binding mode in $\alpha 5\gamma 2_{\text{HET}}$ whereas (*R*)-bretazenil is pushed out of the pocket (**Supplementary Fig. 7f, g**). The activated ECD conformations of $\alpha 5\gamma 2_{\text{HET}}$ and $\alpha 1\beta 3\gamma 2_{\text{EM}}$ are consistent with them being bound by PAMs, i.e. bretazenil and Nb38, respectively. That all five subunits of $\alpha 1\beta 3\gamma 2_{\text{EM}}$ adopt equivalent ECD conformations despite non-equivalent occupancy of the inter-subunit pockets (two $\beta 3\text{P}/\alpha 1\text{C}$ orthosteric sites presumed to be bound by GABA; one $\alpha 1\text{P}/\beta 3\text{C}$ and one $\alpha 1\text{P}/\gamma 2\text{C}$ site bound by Nb38; one $\alpha 1\text{P}/\beta 3\text{C}$ site presumed empty) holds to the Monod-Wyman-Changeux (MWC) theory that postulates transitions between states (e.g. closed-to-open states) preserve symmetry of the oligomer⁶¹. In the case of $\alpha 5_{\text{HOM}}$, the bound ligand, flumazenil, is a neutral BZD antagonist with no preference between activated or resting states⁵³, which will bind either equally well. In this instance $\alpha 5_{\text{HOM}}$ has adopted the activated conformation as is the case for $\alpha 5\gamma 2_{\text{HET}}$, which might reflect an intrinsic preference of the construct ECDs to favour the activated conformation, as previously observed for $\beta 3$ ECDs in the absence of bound ligand⁶². Interestingly, increasing the size of the C(3)-linked ester substituent of flumazenil analogues correlates with PAM activity⁵³. A flumazenil analogue where the ester is substituted by an alkyne of the same length was a neutral modulator (antagonist), whilst addition of a *tert*-butyl group, as is present in bretazenil, conferred PAM activity⁴⁶. *In silico* docking of this PAM analogue (compound 4) into $\alpha 5\gamma 2_{\text{HET}}$ resulted in the lowest free energy binding mode directly overlapping with bretazenil (**Supplementary Fig. 7h**). In both cases, the *tert*-butyl groups are precisely positioned to bridge the gap between loop C Thr208-Ser209 and the adjacent subunit at $\beta 1$ -strand Asp56-Tyr58 (**Fig. 2b** and **Supplementary Fig. 7h**). Closure of loop C is primarily a consequence of the quaternary motion undertaken by the pLGIC ECDs during activation^{6,8,10,20}. This suggests that, by bridging the subunits at this critical juncture, bretazenil

and other BZDs with appropriately large C(3) substituents stabilise the integrally linked process of loop C closure and ECD quaternary activation via their larger contact surfaces compared to flumazenil.

Ion permeation pathway and pore conformation

$\alpha 1\beta 3\gamma 2_{EM}$ possesses two positively charged rings, both previously observed in $GABA_A R-\beta 3_{crist}$, halfway down the vestibule and at the intracellular end of the pore (**Supplementary Fig. 9a-c**), and previously proposed as ion selectivity filters in pLGICs⁶³⁻⁶⁵. In the crowded environment of the glycan-filled vestibule, the ion permeation pathway is restricted to $\sim 5\text{ \AA}$ diameter (**Fig. 7a**), less than a fully hydrated Cl^- ion ($6.1\text{ \AA}$)⁶⁶ but larger than a dehydrated Cl^- ion (Pauling radius of $1.8\text{ \AA}$). However, the polar, semi-flexible glycan chains are not expected to impede the Cl^- flux significantly. The membrane spanning pore, lined by five M2 helices, one from each subunit, narrows from greater than $8\text{ \AA}$ diameter on the extracellular side (excluding the uppermost concentric residue ring, designated 20', which comprises flexible polar side chains that will not impede conductance), to $\sim 5.6\text{ \AA}$ on the intracellular side, at the -2' ring ($\alpha 1$ Pro253, $\beta 3$ Ala248, $\gamma 2$ Pro263; **Fig. 7b**). Although slightly narrower than a fully hydrated Cl^- ion, this is wider than the closed -2' desensitization gate of $GABA_A R-\beta 3_{crist}$ ^{5,67} ($3.2\text{ \AA}$), and also dehydrated Cl^- ions, and is consistent with an open-channel state, within the $5.5\text{-}6\text{ \AA}$ range previously estimated on the basis of different-size anion permeability⁶⁸. It is important to state that although the M2 helices (lining the pore) of $\alpha 1$ and $\beta 3$ subunits are well ordered in the cryo-EM map, the $\gamma 2$ subunit TMD region is mobile. This appears to be an artefact caused by detergent solubilisation, as we observed a reduction of this motion upon addition of lipids (**Supplementary Fig. 4a**). Iterative rounds of particle classification led to a map in which the $\gamma 2$ subunit TMD model could be unambiguously placed, but not accurately refined. Therefore, to ascertain the conductance state, we performed 100 ns molecular

dynamics simulations⁶⁹ on the TMD pore embedded in a POPC membrane with water and 0.15 M NaCl on each side and a + 0.12 V transmembrane potential (where the sign of the voltage refers to that of the cytoplasmic face). Water molecules occupied the length of the pore, indicating there was no significant hydrophobic barrier leading to de-wetting of the channel⁷⁰. There was an influx of multiple Cl⁻ ions, equivalent to an estimated conductance of the order of 100 pS, sufficient to account for physiological conductances (25-28 pS⁷¹) (**Fig. 7c** and **Supplementary Fig. 9d-i**). In contrast, Na⁺ ions failed to traverse the pore despite having a smaller radius (1.2 Å), as expected for an anion selective pore. Consistent with the pore assuming an open state that is narrowest at the intracellular side, single channel recordings of GABA_ARs in membranes show reduced conductance upon mutation of the only polar side chain in the lowest two residue rings, γ2 2' Ser, to non-polar alanine or valine⁷². Furthermore, the α1β3γ2_{EM} pore topology differs from the closed conformation of the related GlyR^{6,7}, where the M2 helices were drawn together at the midpoint 9' leucine residue ring to create a hydrophobic conductance barrier (2.8 Å diameter) (**Fig. 7b**). Nevertheless, the GABA_AR open pore we observe is narrower than the pore of glycine-activated GlyR (8.8 Å diameter⁶) which, fittingly, possesses a higher conductance (60-90 pS⁷³). From previous single channel studies wild type α1β2γ2 receptors in HEK cell membranes in the presence of saturating GABA concentrations occupy the open state 25 % of the time relative to 75 % for the desensitised state⁷⁴. Presumably, the different experimental conditions imposed here, such as extraction from membranes into detergent micelles along with freezing in vitreous ice, have shifted the equilibrium in favour of the open state, resulting in its observation rather than the desensitised state.

pLGICs in which proposed closed and open conformations of the same receptor have been determined, nAChR⁷⁵ and GlyR⁶, reveal that pore opening arises from a relative 'rocking'

motion between the ECD and the TMD regions. For $\alpha 1\beta 3\gamma 2_{EM}$, the activated ECD base loops, in particular the $\beta 1$ -2 loop and $\beta 6$ -7 loop (Cys-loop), are displaced away from the pore axis similarly to those of agonist-bound GlyR versus closed GlyR (**Supplementary Fig. 8n, o**). These base loops form contacts along the M2-M3 loop to concomitantly withdraw the TMD. Superposition of single ECDs from antagonist (strychnine), agonist (glycine) and glycine/ivermectin bound GlyRs on the $\alpha 1\beta 3\gamma 2_{EM}$ $\beta 3$ subunit reveal the impact of this ‘rocking’. Relative to the pore axis, closed GlyR M2 is over-straightened by $\theta = -3.3^\circ$ and rotated clockwise by $\phi = 17.6^\circ$, which contrasts with the $\theta = 7.1^\circ$ recline of the $\alpha 1\beta 3\gamma 2_{EM}$ $\beta 3$ subunit (**Fig. 7e**). As expected for the agonist-bound GlyR M2s, these assume similar reclines to $\alpha 1\beta 3\gamma 2_{EM}$ $\beta 3$, $\theta = 10.8^\circ$ and 9.9° , respectively, with glycine alone bound GlyR also rotated anti-clockwise by $\phi = 20.7^\circ$ (**Fig. 7f, g**).

To evaluate the possible transition motions from the $\alpha 1\beta 3\gamma 2_{EM}$ open pore to the desensitized state^{5,67}, we superposed an ECD from the desensitized GABA_AR- $\beta 3_{crist}$ ⁵ onto either $\alpha 1$, $\beta 3$ or $\gamma 2$ ECDs from the open pore $\alpha 1\beta 3\gamma 2_{EM}$. This analysis revealed that the desensitized conformation also results from a combined tilting and rotation of subunit TMDs relative to the ECDs: the lower portions of the TMDs tilt 10 - 15° and rotate 9 - 11° relative to the $\alpha 1\beta 3\gamma 2_{EM}$ subunits, moving towards the pore axis and closing the channel at the -2’ ring (**Fig. 7h-j**). A similar motion is observed for GlyR, in which the glycine plus ivermectin activated conformation undertakes an inward motion of the lower portions of the TMDs relative to the glycine bound alone activated state⁶.

DISCUSSION

The $\alpha 5_{HOM}$ and $\alpha 5\gamma 2_{HET}$ crystal structures reported here reveal the binding modes of the neutral ligand and BZD antagonist, flumazenil, and the partial PAM, bretazenil. These BZD sites retain

100 % residue identity to the wild type receptor site. Thus, these engineered templates serve as potent tools for future studies to unambiguously determine the binding modes of many of the small molecules previously found to bind the BZD site²⁴. Of note, the binding affinities are in the range of one or two orders of magnitude lower for the BZD_{HET} and BZD_{HOM} sites, respectively. Nevertheless, the binding modes for flumazenil and bretazenil are maintained regardless of the site and are consistent with the previously established functional data discussed throughout the results. Importantly, introduction of a His to Arg substitution into the BZD_{HET} site reproduced the same differential impact on flunitrazepam versus flumazenil binding as previously described for wild-type GABA_A receptors^{32,44}. Furthermore, superposition of these sites over the BZD site in $\alpha 1\beta 3\gamma 2_{EM}$ reveal highly similar positioning of the secondary structure elements and relative appositions of the inter-subunit (P) and (C) faces. Thus, the lower affinities are not due to the sites being distorted or belonging to artefactual structures. Instead, differences likely reflect differing receptor energetics to assume favourable bound conformations for $\alpha 5_{HOM}$ and $\alpha 5\gamma 2_{HET}$ versus wild type receptors in which two of the α -subunits are replaced by β -subunits.

Here we report activated homomeric and heteromeric GABA_AR states bound by a BZD antagonist, and BZD and nanobody PAMs, respectively. These structures reveal fundamental insights into the activation and allosteric modulation processes, and lay the foundation to understand important aspects of GABA_AR inhibitory neurotransmission. Furthermore, although BZDs are widely used to treat epilepsy, insomnia, anxiety, panic disorder and muscle spasm, they lack subtype selectivity and cause unwanted sedation, addiction, and motor and cognitive impairment²⁶. Subtype selective drugs against the BZD site will ameliorate side effects and broaden the therapeutic repertoire to include treatments for autism, Down syndrome, neuropathic pain, schizophrenia and stroke^{15-17,27,28}. We anticipate that the

mechanistic insights and crystallographic platforms described here will expedite the arrival of structure-guided design of therapeutics to treat these disorders.

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AUTHOR CONTRIBUTIONS

Molecular biology, protein expression, purification and crystallization, radioligand binding assays, whole cell electrophysiology: PSM; X-ray data collection: A.R.A., P.S.M., T.M.; X-ray data processing: A.R.A., P.S.M.; cryo-EM data collection: S.M., A.K., Z.S., P.S.M., J.T.H.; cryo-EM data processing: S.M., J.T.H.; cryo-EM model building: S.M., L.D.C., S.L., B.F., F.D.M.; small-molecule docking: T.M.; human genome bioinformatics: S.C., M.M.B.; molecular dynamics simulations: S.R., G.K., S.J.T., M.S.P.S; software and hardware: J.M.D, A.S., R.M.E.; nanobody generation: E.P., J.S.; SPR: S.M.; slice electrophysiology: S.H., T.G.S. The manuscript was written by P.S.M, S.M. T.M. and A.R.A, with input from all co-authors.

FIGURE LEGENDS

Figure 1 Architecture of GABA_ARs bound by BZDs and a Nb38 PAM.

(a) Schematic top-down view of the subunit make-up of $\alpha 5_{\text{HOM}}$. The BZD_{HOM} site is formed from homomeric $\alpha 5$ subunits (red) with the complementary (C)-face of the pocket engineered to contain $\gamma 2$ residues (yellow). Loop C in cyan. (b) Crystal structure of $\alpha 5_{\text{HOM}}$ pentamer shown in top-down and side-on views with flumazenil bound at BZD_{HOM} sites shown with carbon atoms in orange (oxygen – red, nitrogen – blue, fluorine – cyan), inside dashed box for side view. A single M2 helix of the five pore-lining helices is labelled in the top-down view. N-linked glycans are shown as small orange spheres and sticks coloured by atom type (carbon atoms in orange, oxygen atoms in red). The principal (P) and complementary (C) faces between subunits in the side-on view are shown, boxed, for the BZD site between the foremost two subunits. (c) Schematic of $\alpha 5\gamma 2_{\text{HET}}$ showing a BZD_{HOM} site replaced by a BZD_{HET} site. (d) Equivalent views to (b) for $\alpha 5\gamma 2_{\text{HET}}$. Bretazenil shown with carbon atoms in magenta (oxygen

– red, nitrogen – blue, bromine – brown). (e) Schematic of $\alpha 1\beta 3\gamma 2_{EM}$ indicating interfaces bound by Nb38. (f) The 5.17 Å resolution cryo-EM map, top and side views. $\alpha 1$ subunits, red, $\beta 3$ with thermostabilised apocytochrome b562RIL (BRIL) inserted in intracellular loop, blue, $\gamma 2$, yellow, Nb38, green, detergent belt, white, N-linked glycans, orange except $\alpha 1$ -A Asn111-linked glycan, pink. (g) $\alpha 1\beta 3\gamma 2_{EM}$ atomic model. N-linked glycans are in ball-and-stick representation, outlined in yellow.

Figure 2 Binding modes of bretazenil and flumazenil at the BZD_{HET} and BZD_{HOM} sites. (a) Structural formula of flumazenil. Carbon atoms referred to in the text are numbered. Imidazole (I) and benzene rings (A) are labelled. (b) Surface representation of the BZD_{HOM} binding pocket ($\alpha 5$ subunit, red; loop-C of $\alpha 5$ subunit, cyan; the complementary (C)-face containing $\gamma 2$ residues, yellow, designated $\gamma 2^*$) with flumazenil shown as sticks (carbon, orange; oxygen, red; nitrogen, blue; fluorine, cyan). (c), (d) Side and top views of flumazenil bound to this BZD site. e, Structural formula of (S)-bretazenil. (f-h), Bretazenil bound to the BZD_{HET} site (carbon, magenta; oxygen, red; nitrogen, blue; bromine, brown). $\gamma 2$ subunit, yellow. Side chains of amino acid residues contacting the ligand are numbered and shown as sticks. Distances (Å) between selected atoms and aromatic rings are shown as grey and green dashed lines, respectively.

Figure 3 Sequence variability at the BZD binding pocket in the human population.

(a) and (b) Heat maps of allelic variants from GABA_AR α and γ subunits respectively, for residues within 5 Å of bound BZD in $\alpha 5\gamma 2_{HET}$ and $\alpha 5_{HOM}$, across ethnic groups from the gnomAD database⁵⁴ of 138,632 unrelated healthy individuals. Boxes are colour coded by frequency, with intensity indicators underneath. Text colour corresponds to residue location:

red – α -subunit loops A or B; blue – α -subunit loop C; yellow – γ -subunit. Residue numberings are for mature sequences (Uniprot: $\alpha 1$ P14867, $\alpha 2$ P47869, $\alpha 3$ P34903, $\alpha 4$ P48169, $\alpha 5$ P31644, $\alpha 1$ Q16455, $\gamma 2$ P18507, $\gamma 3$ Q99928) with $\alpha 5\gamma 2_{\text{HET}}$ numbering in parentheses where appropriate. (c) Asteroid plots of ligand-residue contacts between bretazenil and $\alpha 5$ P-face and $\gamma 2$ C-face residues. Circle size reflects contact number (size indicator provided). Circle outline colour corresponds to residue location (same as for text in, a). Circles are filled for residues with allelic variants, and coloured according to frequency (same as in, a).

Figure 4 Nb38 binding and modulation of $\alpha 1\beta 3\gamma 2$ receptors.

(a) ‘Open book’ view of the Nb38 interaction surface with $\alpha 1\beta 3\gamma 2_{\text{EM}}$ at the $\alpha 1$ -A- $\beta 3$ -E and $\alpha 1$ -D- $\gamma 2$ -C (just under the BZD site) interfaces. Contact regions for: CDR1, purple; CDR2, cyan; CDR3, yellow. (b) Detailed view of Nb38 binding to the $\alpha 1$ subunit. CDR3 loop binding to $\beta 9$ and $\beta 10$ strands and alternate view rotated 184° degrees around the receptor longitudinal axis to show CDR1 and CDR2 interactions with the Cys-loop, loop-C, and $\beta 7$ strand. Amino acids in $\alpha 1$ subunit which are predicted to contribute to Nb38 binding are highlighted. (c) Nb38 concentration-response curves for potentiation of GABA EC_{10} responses (WT^{PO} , EM^{PO}) and direct activation (WT^{ACT} , EM^{ACT}), respectively, measured from whole-cell patch-clamp recordings of HEK293T cells expressing either wild-type (WT) $\alpha 1\beta 3\gamma 2$ GABA_ARs or $\alpha 1\beta 3\gamma 2_{\text{EM}}$. $n = 7$, EC_{50}s are accrued from different cells. (d) Bar chart demonstrating $\alpha 1$ -subunit selectivity of Nb38 compared to $\alpha n\beta 3\gamma 2$ GABA_ARs ($n = 2$ -6). Note, no potentiation at other subtypes. $n = 3$ -4, each data point comes from different cells. (e) Effect of Nb38 (2 μM) and diazepam (DZPM, 500 nM) relative to control ACSF on sIPSC frequency (freq) in dentate gyrus granule cells, amplitude (amp), rise-time (RT), decay time (τ) and charge transfer. $*p < 0.05$ and $**p < 0.01$ significance compared to ACSF (paired t-test). Nb38 and DZPM were

tested on 4 and 5 cells respectively. (f) Average sIPSC waveforms after incubation with Nb38 and DZPM (grey) superimposed over control ACSF (black) before incubation with ligand.

Figure 5 N-linked glycosylation of $\alpha 1\beta 3\gamma 2_{EM}$ $\alpha 1$ subunits.

(a) Side view of $\alpha 1\beta 3\gamma 2_{EM}$ map showing $\alpha 1$ N-linked glycans filling the channel vestibule. $\beta 3$ -B and $\gamma 2$ -C subunits were hidden to visualise the vestibule. (b) and (c) Close-ups of $\alpha 1$ -D and $\alpha 1$ -A Asn111-linked glycans. Atomic model and cryo-EM maps are shown in orange and pink, respectively. (d) Putative CH- π interaction between the Trp123 side-chain and the pyranose ring of a mannose moiety from the $\alpha 1$ -A Asn111-linked glycan. Distances between the centre of mass of the Trp123 indole and hydrogens of the interacting mannose residue are shown. (e) Sequence alignment of $\beta 5$ - $\beta 5'$ loops. All α subunits contain N-linked glycosylation sites, and all γ subunits contain Trp residues at the equivalent position (boxed). Sequence numbering shown is based on $\beta 3$ subunit (mature isoform 1). (f) Proposed assembly sequence for $\alpha\beta\gamma$ GABA_ARs. $\alpha 1$ and $\beta 3$ subunits produced in the ER first form dimers through the $\beta 3(P)$ - $\alpha 1(C)$ interface. An $\alpha 1\beta 3$ dimer binds to the $\gamma 2$ subunit through the $\gamma 2(P)$ - $\beta 3(C)$ interface with the Asn111-linked glycan on the $\alpha 1$ subunit interacting with $\gamma 2$ Trp123 to stabilise the horizontal glycan conformer. Pentamerization occurs through $\beta 3(P)$ - $\alpha 1(C)$ and $\alpha 1(P)$ - $\gamma 2(C)$ interfaces between an $\alpha 1\beta 3\gamma 2$ trimer and an $\alpha 1\beta 3$ dimer in which its Asn111-linked glycan adopts an upward trajectory.

Figure 6 $\alpha 5_{HOM}$, $\alpha 5\gamma 2_{HET}$ and $\alpha 1\beta 3\gamma 2_{EM}$ ECD activated conformations and BZD binding modes.

Side-on views of single ECDs from globally superposed pentamers comparing quaternary conformations relative to the pore axis (not shown – right of ECDs), shown looking into loop

C. (a) $\alpha 5_{\text{HOM}}$ (light grey) versus $\alpha 1\beta 3\gamma 2_{\text{EM}}$ $\alpha 1$ subunit, (b) $\alpha 5\gamma 2_{\text{HET}}$ $\alpha 5$ subunit (light grey) versus $\alpha 1\beta 3\gamma 2_{\text{EM}}$ $\alpha 1$ subunit, (c) $\alpha 5\gamma 2_{\text{HET}}$ $\gamma 2$ subunit (light grey) versus $\alpha 1\beta 3\gamma 2_{\text{EM}}$ $\gamma 2$ subunit. Agonist binding C-loop and the ECD base loops ($\beta 1$ -2 loop, $\beta 6$ -7 loop – Cys-loop, $\beta 8$ -9 loop) are also shown for activated $\beta 3_{\text{cryst}}$ (PDB IDs: 4COF – cyan), agonist-bound (activated) glycine receptor (PDB ID: 5JAE – light green) and resting closed GlyR (PDB ID: 5JAD – dark green). The $\alpha 1\beta 3\gamma 2_{\text{EM}}$ $\alpha 1$ and $\gamma 2$ ECDs, and $\alpha 5_{\text{HOM}}$ and $\alpha 5\gamma 2_{\text{HET}}$ $\alpha 5$ or $\gamma 2$ subunits all occupy the activated conformation with loop-C closed and base loops tilted away from the pore axis, similar to previously solved activated $\beta 3_{\text{cryst}}$ and GlyR structures, and different from resting state GlyR. Computational docking of flumazenil (d, estimated free energy of binding, -8.9 kcal/mol) and bretazenil (e, -7.8 kcal/mol) to $\alpha 1\beta 3\gamma 2_{\text{EM}}$. Docked ligands, $\alpha 1$ and $\gamma 2$ subunits from $\alpha 1\beta 3\gamma 2_{\text{EM}}$ are colored, crystal structures of GABA_AR in complex with flumazenil (d) and bretazenil (e) are shown in light grey.

Figure 7 $\alpha 1\beta 3\gamma 2_{\text{EM}}$ ion permeation pathway, open pore, and subunit conformations.

(a) Solvent accessible tunnel through the vestibule between the $\alpha 1$ subunit Asn111-linked glycans and pore of $\alpha 1\beta 3\gamma 2_{\text{EM}}$. $\beta 3$ -B and $\gamma 2$ -C subunits were hidden to visualise permeation pathway. The $\alpha 1$ Asn111-linked glycans are shown in ball and stick representation. (b) Plot of pore radii for $\alpha 1\beta 3\gamma 2_{\text{EM}}$, desensitized GABA_AR- $\beta 3_{\text{cryst}}$ (PDB ID: 4COF), glycine-bound GlyR (PDB ID: 5JAE), glycine/ivermectin-bound GlyR (PDB ID: 5JAF) and antagonist-bound GlyR (PDB ID: 5JAD). (c) Snapshot from a molecular dynamics simulation showing that the $\alpha 1\beta 3\gamma 2_{\text{EM}}$ pore is fully hydrated and may be permeated by chloride ions. Pore viewed parallel to the membrane between two $\beta 3$ subunits (represented as surface, white outlines are TMD helices). Chloride (green) and sodium (orange) ions are shown as van der Waals spheres whereas water molecules (white-red) are represented in bonds-only format. (d) Plot showing

how the tilt (θ) and rotation (ϕ) angles are determined relative to the channel pore (z axis). M2 helices are defined as vectors between -2' C $_{\alpha}$ and 9' C $_{\alpha}$. Rotation angles for $\alpha 1\beta 3\gamma 2_{EM}$ $\alpha 1$, $\beta 3$ and $\gamma 2$ subunits are set to zero. **(e-g)** Superposition between single subunits from GlyR in antagonist-bound, glycine-bound and glycine/ivermectin-bound conformations, respectively, and the $\alpha 1\beta 3\gamma 2_{EM}$ $\beta 3$ subunit. The ECD regions were used for structural alignment. A vector line between the M2 -2' C $_{\alpha}$ and 9'C $_{\alpha}$ is drawn for each subunit. Tilt and rotation angles are shown for each pair. **(h-j)** Superposition between GABA $_A$ R- $\beta 3_{cryst}$ and $\alpha 1\beta 3\gamma 2_{EM}$ $\alpha 1$, $\beta 3$ and $\gamma 2$ subunits, respectively, for TMD tilt and rotation analyses.

METHODS

Construct design, $\alpha 5_{HOM}$ and $\alpha 5\gamma 2_{HET}$. Details of the $\alpha 5$ subunit constructs design, including protein sequences, are shown in **Supplementary Figure 1**. The chimeric $\gamma 2$ -ECD: $\alpha 1$ -TMD subunit of $\alpha 5\gamma 2_{HET}$ comprises mature sequence (Uniprot P18507) $\gamma 2$ residues 39 to 232 (QKSDD...DLSRR) appended to $\alpha 1$ (Uniprot P62813) from 223 to 455 (IGYFVI...PTPHQ) with a single $\beta 3$ substitution, ($\alpha 1$ P280A). The $\alpha 5$ intracellular M3-M4 loop amino acids 316-392 (RGWA...NSIS) (Uniprot P31644) and the $\alpha 1$ intracellular M3-M4 loop amino acids 313-390 (RGYA...NSVS) were substituted by the SQPARAA sequence^{5,40} to enhance the recombinant protein yield and facilitate crystallisation. Constructs were cloned into the pHLsec vector⁷⁶, between the N-terminal secretion signal sequence and either a double stop codon or a C-terminal 1D4 purification tag derived from bovine rhodopsin (TETSQVAPA) that is recognised by the Rho-1D4 monoclonal antibody (University of British Columbia)^{77,78}.

Construct design, $\alpha 1\beta 3\gamma 2_{EM}$. The protein sequences used were: human GABA $_A$ R $\alpha 1$ (mature polypeptide numbering 1-416, QPSL...TPHQ; Uniprot P14867), human GABA $_A$ R $\beta 3$ (mature

polypeptide numbering 1-447, QSVN...YYVN; Uniprot P28472), human GABA_AR γ 2 (mature polypeptide numbering 1-427, QKSD...YLYL; Uniprot P18507). These constructs were cloned into the pHLsec vector⁷⁶, after the N-terminal secretion signal sequence and before a double stop codon unless stated otherwise below. The α 1 intracellular M3-M4 loop amino acids 313-391 (RGYA...NSVS) were substituted by the SQPARAA sequence^{5,40}. Four β 3 ECD residues were replaced by β 2 subunit residues (Gly171Asp, Lys173Asn, Glu179Thr, Arg180Lys), which block homomer assembly⁷⁹, and the β 3 intracellular M3-M4 loop amino acids 308-423 (GRGP...TDVN) were substituted by a modified SQPARAA sequence containing the *E.coli* soluble cytochrome B562RIL⁴¹ (BRIL, amino acids 23-130, ADLE...QKYL, Uniprot P0ABE7) to give an M3-M4 loop of sequence SQPAGT-BRIL-TGAA. The γ 2 intracellular M3-M4 loop amino acids 323-400 (NRKP...IRIA) were substituted by the SQPARAA sequence, and a C-terminal GTGGT linker followed by a 1D4 purification tag derived from bovine rhodopsin (TETSQVAPA) that is recognised by the Rho-1D4 monoclonal antibody (University of British Columbia)^{77,78}. Nb38 was identified from a previously described nanobody library²⁰.

Large-scale expression and purification of α 5_{HOM}, α 5 γ 2_{HET} and α 1 β 3 γ 2_{EM}. Twenty-litre batches of HEK293S-GnTI⁻ cells (which yield proteins with truncated N-linked glycans, Man₅GlcNAc₂^{80,81}) were grown in suspension to densities of 2×10^6 cells ml⁻¹ in Protein Expression Media (PEM, Invitrogen) supplemented with L-glutamine, non-essential amino-acids (Gibco) and 1% foetal calf serum (Sigma-Aldrich). Typical culture volumes were 200 ml, in 600 ml recycled media bottles, with lids loose, shaking at 130 rpm, 37°C, 8 % CO₂. For transient transfection, cells from 1 litre of culture were collected by centrifugation (200 g for 5 min) and resuspended in 150 ml Freestyle medium (Invitrogen) containing 3 mg PEI Max (Polysciences) and 1 mg plasmid DNA, followed by a 4 h shaker-incubation in a 2 litre conical

flask at 160 rpm. For $\alpha 5\gamma 2_{\text{HET}}$ DNA plasmids were transfected at 9:1 ratio (i.e. 0.9:0.1 mg) $\alpha 5$ construct DNA without a 1D4 tag to the chimera $\gamma 2\text{-ECD}:\alpha 1\text{-TMD}$ with a 1D4 purification tag. For $\alpha 1\beta 3\gamma 2_{\text{EM}}$ DNA plasmid ratios were 1:1:0.5, respectively. Subsequently, culture media were topped up to 1 litre with PEM containing 1 mM valproic acid and returned to empty bottles. Typically, 40-70 % transfection efficiencies were achieved, as assessed by control transfections with a monoVenus-expressing plasmid^{82,83}. 72 h post-transfection cell pellets were collected, snap-frozen in liquid N₂ and stored at -80 °C.

Cell pellets (approx. 200g) were solubilised in 600 ml buffer containing 20 mM HEPES pH 7.2, 300 mM NaCl, 1 % (v/v) mammalian protease inhibitor cocktail (Sigma-Aldrich, cat. P8340) and 1.5 % (w/v) dodecyl 1-thio- β -maltoside (DDTM, Anatrace) for $\alpha 5_{\text{HOM}}$ and $\alpha 1\beta 3\gamma 2_{\text{EM}}$ or 1.5 % (w/v) decyl β -maltoside (DM, Anatrace) for $\alpha 5\gamma 2_{\text{HET}}$, for 2 hours at 4 °C. Insoluble material was removed by centrifugation (10,000 g, 15 min). The supernatant was diluted 2-fold in a buffer containing 20 mM HEPES pH 7.2, 300 mM NaCl and incubated for 2 hr at 4 °C with 10 ml CNBr-activated sepharose beads (GE Healthcare) pre-coated with 50 mg Rho-1D4 antibody (3.3 g dry powdered beads expand during antibody coupling to approximately 10 ml). Affinity-bound samples were washed slowly by gravity flow over 2 hours at 4 °C with 200 ml buffer containing 20 mM HEPES pH 7.2, 300 mM NaCl, and either 0.1 % (w/v) DDTM (approximately 20 x CMC) for $\alpha 5_{\text{HOM}}$, or 0.2 % (w/v) DM (approximately 3 x CMC) $\alpha 5\gamma 2_{\text{HET}}$, or for $\alpha 1\beta 3\gamma 2_{\text{EM}}$ 0.1 % (w/v) DDTM and 0.01 % (w/v) porcine brain polar lipid extract (141101C Avanti; chloroform was evaporated under argon then 100 mg lipid film was dissolved in 10 ml 10 % (w/v) DDTM (1000 mg) in water and stored at -80 C until needed). Beads were then washed in a second round of buffer: 20 mM HEPES pH 7.2, 300 mM NaCl, and either 0.01 % (w/v) DDTM (approximately 3 x CMC) for $\alpha 5_{\text{HOM}}$ or 0.2 % (w/v) DM (approximately 3 x CMC) for $\alpha 5\gamma 2_{\text{HET}}$, or for $\alpha 1\beta 3\gamma 2_{\text{EM}}$, 1mM GABA, 0.01 % (w/v) DDTM

(approximately 4 x CMC), 0.001 % (w/v) porcine brain polar lipid extract. Protein samples were eluted overnight in 15 ml buffer containing 15 mM HEPES pH 7.2, 225 mM NaCl, 500 μ M TETSQVAPA peptide (Genscript), and corresponding detergents. The eluate was centrifuged (30,000g, 15 min) and the supernatant was concentrated by ultrafiltration to 1-2 ml at 1-5 mg/ml using 100-kDa cut-off membranes (Millipore). The concentrated sample was centrifuged (30,000 g, 15 min) and the supernatant was aliquoted in 0.5-1.5 mg protein per 0.7 ml aliquots and either snap-frozen for storage at -80 °C or gel filtrated as appropriate. A single aliquot was loaded onto a Superose 6 10/300 Increase gel filtration column (GE Healthcare) equilibrated in 10 mM HEPES pH 7.2, 150 mM NaCl, and either: 0.007 % (w/v) DDTM, 50 μ M flumazenil, 50 μ M pregnanolone for α 5_{HOM} or 0.2 % (w/v) DM, 50 μ M bretazenil for α 5 γ 2_{HET}, or for α 1 β 3 γ 2_{EM}, 1 mM GABA, 0.007 % (w/v) DDTM. The peak fractions were approximately 0.5 mg/ml. The fractionated protein was concentrated by ultrafiltration to 3-5 mg/ml, using 100 kDa cut-off membranes (Millipore), for crystallisation trials. Typical final yields were 0.1-0.2 mg protein per litre of cells grown in suspension (10 g cell pellet). In the case of α 1 β 3 γ 2_{EM}, Nb38 was added to α 1 β 3 γ 2 at 4-fold molar excess and the complex was concentrated by ultrafiltration to 2.5 mg/ml, using 100 kDa cut-off membranes (Millipore).

Nb38 purification. Nb38 was produced and purified in milligram quantities from WK6su⁻ *E. coli* bacteria⁸⁴. Bacteria were transformed with ~200 ng of nanobody expression plasmid pMESy4 containing the nanobody of interest and selected on Lysogeny broth (LB)-agar plates containing 2% glucose and 100 μ g/ml ampicillin. 2-3 colonies were used for preparing a pre-culture, which was used to inoculate 0.5 L Terrific broth (TB) cultures supplemented with 0.1 % glucose, 2 mM MgCl₂ and 100 μ g/mL ampicillin. Cultures were grown at 37 °C until their OD₆₀₀ reached 0.7, at which point Nb38 expression was induced with 1 mM IPTG. After induction, cells were grown at 28 °C overnight and harvested by centrifugation (20 min, 5000

g). Nanobodies were released from the bacterial periplasm by incubating cell pellets with an osmotic shock buffer containing 0.2 M Tris pH 8.0, 0.5 mM EDTA, and 0.5 M sucrose. The C-terminally His6-tagged Nb38 was purified using nickel affinity chromatography (binding buffer: 50 mM HEPES pH 7.6, 1 M NaCl, 10 mM imidazole; elution buffer: 50 mM HEPES pH 7.6, 0.2 M NaCl, 0.5 M imidazole), followed by size-exclusion chromatography on a Superdex 75 16/600 column (GE Healthcare) in 10 mM Hepes pH 7.6, 150 mM NaCl. Nb38 stocks were concentrated to 5-10 mg/mL, snap-frozen in liquid nitrogen and stored at -80 °C.

Crystallization and data collection. $\alpha 5\gamma 2_{\text{HET}}$ and $\alpha 5_{\text{HOM}}$ contain 15 N-linked glycosylation sites each, bringing a considerable extra volume, flexibility and potential occupancy heterogeneity. Therefore, prior to crystallization, concentrated protein samples (6 mg/ml $\alpha 5\gamma 2_{\text{HET}}$ and 4 mg/ml $\alpha 5_{\text{HOM}}$) were incubated with 0.01 mg ml⁻¹ endoglycosidase F1⁸⁵ for 2h at room temperature. Sitting drop vapour diffusion crystallization trials were performed in 96-well Swissci 3 well crystallisation plates (Hampton Research), at three ratios: 200 nl protein plus 100 nl reservoir, 100 nl protein plus 100 nl reservoir, 100 nl protein plus 200 nl reservoir. Drops were dispensed by a Cartesian Technologies robot⁸⁶, and plates were maintained at 6.5 °C in a Formulatrix storage and imaging system. In the case of $\alpha 5\gamma 2_{\text{HET}}$, crystals appeared in a range of conditions⁸⁷ within 1-28 days, with the best diffracting crystals (to ~2.5 Å resolution) taking 4 weeks to grow in: 22 % poly-ethylene (PEG) 400, 0.37 M potassium nitrate, 0.1 molar 2-(N-morpholino)ethanesulfonic acid (MES) pH 6.5. For $\alpha 5_{\text{HOM}}$, crystals also grew in a range of conditions, typically within 2 weeks, and in the first instance diffracted up to intermediate resolution (>5 Å). Following additive-based optimization (MemAdvantage, Molecular Dimensions), crystals diffracting to ~2.6 Å resolution were identified, grown in: 19 % PEG 1000, 0.1 M sodium chloride, 0.15 M ammonium sulphate, 0.1 M MES pH 6.5, 2.5 mM sucrose monodecanoate (sucrose monocaprate). Crystals were cryoprotected by soaking in reservoir

solution supplemented with 30 % ethylene glycol, and then cryocooled in liquid nitrogen. Diffraction images were collected at the Diamond Light Source beamline I04, $\lambda=0.9795$ Å, 0.1° oscillation (bretazenil-bound $\alpha 5\gamma 2_{\text{HET}}$) and 0.2° oscillation (flumazenil-bound $\alpha 5_{\text{HOM}}$), on a Pilatus 6M-F detector. X-ray data were indexed, integrated and scaled using the HKL2000 package⁸⁸. Diffraction from both $\alpha 5\gamma 2_{\text{HET}}$ and $\alpha 5_{\text{HOM}}$ crystals was severely anisotropic, therefore scaled but unmerged data were processed with STARANISO⁸⁹, allowing for the anisotropic diffraction cut-offs to be applied before merging with Aimless^{90,91}, within the autoPROC toolbox⁹². Upon ellipsoidal truncation, resolution limits were 2.33 Å, 3.15 Å and 3.73 Å (in the $-0.022 a^* + c^*$, b^* and $0.945 a^* - 0.327 c^*$ directions, respectively) for $\alpha 5\gamma 2_{\text{HET}}$, and 2.49 Å, 3.13 Å and 4.63 Å (in the $0.872 a^* - 0.490 c^*$, b^* and $0.842 a^* + 0.540 c^*$ directions, respectively) for $\alpha 5_{\text{HOM}}$. Data collection and merging statistics are detailed in the **Table 1**.

Structure determination, refinement and analysis. $\alpha 5\gamma 2_{\text{HET}}$ and $\alpha 5_{\text{HOM}}$ structures were solved by molecular replacement using the human GABA_AR- $\beta 3_{\text{cryst}}$ homopentamer⁵ (PDB ID: 4COF) as a search model in Phaser⁹³. Polypeptide chains were traced using iterative rounds of manual model building in Coot⁹⁴ and refinement in BUSTER-TNT⁹⁵, Refmac⁹⁶ and Phenix^{97,98}. Automated X-ray and atomic displacement parameter (ADP) weight optimisation, and torsion angle non-crystallographic symmetry (NCS) restraints, were applied. Ligand coordinates and geometry restraints were generated using the grade server⁹⁹. The $\alpha 5\gamma 2_{\text{HET}}$ and $\alpha 5_{\text{HOM}}$ models contain one homopentamer per asymmetric unit. Crystal packing impaired map quality in regions where ECD from certain subunits were near of detergent micelles of neighbouring molecules. Nevertheless, complete polypeptide chains could be built, with the exception of 14 N-terminal $\alpha 5$ residues (QMPTSSVKDETNDN), 22 N-terminal $\gamma 2$ residues

(QKSDDYEDYTSNKTWVLTPKV) and the C-terminal purification tags, presumably disordered. Strong additional electron density peaks were clearly visible in the BZD_{HOM} and BZD sites, that could be unambiguously assigned to flumazenil in $\alpha 5_{\text{HOM}}$ and bretazenil in $\alpha 5\gamma 2_{\text{HET}}$, respectively, based on shape, coordination and refinement statistics. Furthermore, electron density corresponding to five pregnanolone molecules, one per inter-subunit interface, could be observed at the TMD interfaces of $\alpha 5_{\text{HOM}}$, as previously described²⁰, and five well-ordered detergent molecules (decyl β -maltoside) could be built inside the $\alpha 5\gamma 2_{\text{HET}}$ pore region. The $\alpha 5$ and $\gamma 2$ extracellular regions have three N-linked glycosylation sites each, and we could observe clear electron density for six NAG moieties in $\alpha 5_{\text{HOM}}$ and five in $\alpha 5\gamma 2_{\text{HET}}$, the others being disordered. Stereochemical properties of the models were assessed in Coot⁹⁴ and Molprobit¹⁰⁰. Refinement statistics are provided in the **Table 1**. Protein geometry analysis revealed no Ramachandran outliers, 97.96% residues in favoured regions and 2.04% residues in allowed regions for $\alpha 5_{\text{HOM}}$, and one Ramachandran outlier (0.06%), 95.97% residues in favoured regions and 4.03% residues in allowed regions for $\alpha 5\gamma 2_{\text{HET}}$. Molprobit clash scores after adding hydrogen atoms is 9.16 (99th percentile) for $\alpha 5_{\text{HOM}}$ and 11.06 (94th percentile) for $\alpha 5\gamma 2_{\text{HET}}$. Overall Molprobit scores are 1.50 (100th percentile) for $\alpha 5_{\text{HOM}}$ and 1.84 (98th percentile) for $\alpha 5\gamma 2_{\text{HET}}$. Structural alignments were performed in PyMOL using the align function. Structural figures were prepared with the PyMOL Molecular Graphics System, Version 1.8, Schrödinger, LLC., and the UCSF Chimera package, developed by the Resource for Biocomputing, Visualization, and Informatics at the University of California, San Francisco¹⁰¹.

Electron microscopy and image processing. CryoEM samples were prepared using C-flatTM Holey Carbon grids (R2/1, 200 mesh, 53% of collected images) and UltraAuFoilTM grids

(R1.2/1.3, 200 mesh, 47% of collected images). Carbon substrate grids were glow discharged for 10 s, then 3.5 μL of protein sample (4.2 mg/mL) was applied for 15 s. The sample was blotted for 3 s using VitroBot Mark IV (FEI) and flash-frozen in liquid ethane. Gold substrate grids were glow discharged for 15 s and 2.5 μL sample (2.5 mg/mL) was applied for 30s, which was followed by 7.5 s blotting. In both cases, blotting was performed at room temperature and 90-100% humidity. All cryo-EM data were collected using a Tecnai F30 Polara electron microscope (FEI) operating at 300 kV, fitted with a K2 Summit direct electron detector (Gatan) and a GIF-Quantum energy filter (Gatan). SerialEM was used to manually record zero energy-loss (20 eV slit) images at a calibrated magnification of 37,000x (1.35Å/pix) in counting mode. Images were recorded as movies consisting of 47 frames with total dose of 38 $\text{e}^-/\text{\AA}^2$ and exposure of 14.1s. Nominal defocus values ranged from -2.0 to -3.5 μm .

Classification of $\alpha 1\beta 3\gamma 2_{\text{EM}}$ particles in the decylmaltoside neopentylglycol, (DMNG) detergent revealed that the particles had a ‘collapsed’ TMD region, with the $\gamma 2$ TMD occupying the central pore (**Supplementary Fig. 4a**). We discarded this sample, considering it to be a sample preparation artefact, presumably caused by extraction of the receptor from its native cell bilayer into DMNG alone. Preparation of a second sample in a different detergent, dodecyl 1-thio- β -maltoside (DDTM) and in the presence of porcine brain polar lipid extract (see methods section on Large-scale expression and purification) yielded the predominant particle population with TMDs pseudo-symmetrically arranged around the pore, i.e. the $\gamma 2$ TMD was no longer ‘collapsed’ into central pore. This sample was used for all subsequent processing. A total of 8,548 movies were motion-corrected at micrograph level with UCSF MotionCorr¹⁰² and CTFFIND4¹⁰³ was used to estimate the contrast function parameters. Data processing was performed using RELION software package¹⁰⁴. First, ~5,000 particles were picked using EMAN2¹⁰⁵, grouped in RELION using reference-free 2D classification and class averages were

used as references for RELION automated particle picking in the same software. False-positives were manually removed leading to a dataset of 525,124 particles. Reference-free 2D classification was performed and particles belonging to classes showing GABA_AR features were selected (436,320 particles). An initial 3D model was generated from the GABA_AR- β 3_{cryst} structure⁵ (PDB ID 4COF) and low-pass filtered to 40 Å resolution. Particles were first oriented in 3D by imposing C5 symmetry, and further classified in 3D imposing no symmetry (C1) by only allowing rotation around the symmetry axis. The best class showing features of two BRILs and two nanobodies was selected and used as a new initial reference model. Particles selected from 2D classes were 3D-classified into 10 classes using C1 symmetry. Particles assigned to the best 3D classes (186,786 particles) were used for particle ‘polishing’ step in RELION, where particle motion and radiation damage for particles from each movie frame was estimated. ‘Gold standard’ refinement of the resulting ‘shiny’ particles resulted in a 5.65 Å map. Movie motion correction with MotionCor2¹⁰⁶ using 25 patches and dose-weighting scheme improved the resolution of the refined map to 5.25 Å. These particles were further 3D-classified into 10 volumes. Particles assigned to the best 9 classes (165,621 particles) were combined to yield a map of 5.17 Å. The resolution was estimated using *relion post-process* with the FSC criteria of 0.143. The final unsharpened and unfiltered map was globally auto-sharpened using *phenix.auto_sharpen*¹⁰⁷ (5.17 Å high-resolution cut-off) to maximise the map detail while maintaining the connectivity of the map.

Model building and refinement of α 1 β 3 γ 2_{EM}. Crystal structures (2.9 Å β 3_{cryst} (PDB ID 4COF), 2.5 Å α 5 γ 2_{HET} (PDB ID XXXX) and 3.2 Å α 5_{TMD}²⁰ (PDB ID 5O8F)) were used to build the GABA_AR α 1 β 3 γ 2_{EM} heteromer and Nb38 models into the 5.17 Å map. First, the α 5 subunit and Nb25²⁰ (5O8F) structures were manually mutated to match the α 1 and Nb38 sequences, respectively. TMD of the γ 2 subunit was prepared by mutating α 5 TMD (5O8F)

residues to match the $\gamma 2$ sequence. The Nb38 model and the corresponding ECDs and TMDs of $\alpha 1$, $\beta 3$ and $\gamma 2$ subunits were docked into the cryo-EM map using Chimera. Map was cut around each subunit and nanobody (4 Å radius) using Chimera. Rosetta-CM¹⁰⁸ was used to refine the models into the resulting cryo-EM densities to improve model geometry and fitting in the density. N-linked glycan models from $\alpha 5_{TMD}$ crystal structure (5O8F) were docked into the cryo-EM map and added to the model. Rotamers were manually adjusted to match the high resolution structures; where no prior information was available, the most common rotamers were chosen. The model was further optimised by rounds of manual correction in Coot⁹⁴ and iterative refinement in real space with *phenix_real_space_refine*¹⁰⁹ using secondary structure and NCS restraints. The final model contains $\alpha 1$ subunit residues 12-312 (TTVF...YFTK) and 391-414 (KIDR...YWAT); $\beta 3$ subunit residues 10-307 (SFVK...YIFF) and 422-447 (AIDR...LYYV); $\gamma 2$ subunit residues 27-322 (VTVI...YFVS) and 401-424 (KMDS...YWVS) and Nb38 residues 1-123 (QVQL...TVSS). For model validation, the final model coordinates were randomly displaced by 0.2 Å and then this model was refined with *phenix_real_space_refine* against one of the half-maps produced by RELION¹⁰⁴. FSC curves were then calculated between the refined model and half-map used for refinement ('work') and between the second half-map, not used for refinement, ('free'). No significant separation between FSC_{work} and FSC_{free} curves was observed, indicating the model was not over-refined. The stereochemistry of the final model was evaluated using MolProbity¹¹⁰. Figures were prepared using UCSF Chimera package, developed by the Resource for Biocomputing, Visualization, and Informatics at the University of California, San Francisco¹⁰¹. Structural alignments were performed in SHP¹¹¹. Pore dimensions were analysed using the Coot implementation of Hole¹¹² and with CAVER¹¹³ PyMOL plugin with a probe radius of 2.5 Å.

Small molecule docking. Molecular docking of small molecules to $\alpha 5\gamma 2_{\text{HET}}$ and $\alpha 5_{\text{HOM}}$ crystal structures was performed using AutoDock Vina¹¹⁴. Stereochemistry of small molecules was optimised using Grade webserver¹¹⁵. Structures of the receptor were kept rigid during docking. The region selected for docking encompassed whole benzamidine-binding pocket.

Molecular dynamics simulations. The protein portion of the receptor structure, either full length (for uniform electric field simulations) or with only the transmembrane domain included (for water equilibrium simulations), was embedded within a POPC lipid bilayer, in a simulation box containing an aqueous solution of Cl^- and Na^+ ions at 0.5 M or 0.15 M concentration, respectively, following a previously established protocol¹¹⁶ and employing the TIP4P water model¹¹⁷. Simulations were performed using GROMACS version 5.1^{118,119}, and the OPLS united-atom force field¹²⁰. The temperature and pressure were maintained at 37 °C and 1 bar, respectively, using the velocity-rescale thermostat¹²¹ in combination with a semi-isotropic Parrinello and Rahman barostat¹²², with coupling constants of $\tau_T = 0.1$ ps and $\tau_P = 1$ ps. A Verlet cut-off scheme was applied, and long-range electrostatic interactions were measured using the Particle Mesh Ewald method¹²³. Bonds were constrained using the LINCS algorithm¹²⁴, and an additional harmonic restraint at a force constant of $1000 \text{ kJ mol}^{-1} \text{ nm}^{-2}$ was placed on protein backbone atoms to preserve the conformational state of the original, experimentally determined structure. The integration time-step was 2 fs. For the water distribution and free energy profiles: 50 ns repeats were each initiated from an independently assembled simulation system. Free energy profiles for water permeation along the channel pore were derived (using the CHAP channel annotation package, <http://www.channotation.org/>) from the equilibrium distribution of water molecules upon simulation through an inverse Boltzmann calculation-based method⁷⁰. Pore radius profiles (i.e. radii at positions along the pore axis) were estimated using the programme HOLE¹¹². To determine the behaviour of ions

a +0.12 V potential difference was imposed across the membrane during 100 ns simulations by setting a linear potential difference of 0.5 V across the z-length of the simulation box. The conductance was estimated based on the number of Cl⁻ ions passing through the channel.

Radioligand binding experiments. GABA_AR constructs containing a single BZD site ($\alpha 1\beta 3\gamma 2_{EM}$, $\alpha 5\gamma 2_{HET\Delta}$ and $\alpha 5\beta 3\gamma 2_{WT}$) at 2 nM and five sites ($\alpha 5\gamma 2_{HET}$ and $\alpha 5_{HOM}$) at 0.4 nM, were used, in 10 mM HEPES pH 7.2, 150 mM NaCl, and: 0.05 % (w/v) DDTM, 0.005 % (w/v) porcine brain polar lipid extract; 0.2 % (w/v) DM detergent for $\alpha 5\gamma 2_{HET}$; 0.05 % (w/v) detergent (decylmaltoside neopentylglycol (DMNG) 5:1 (molar ratio) cholesterol hemisuccinate (CHS) for $\alpha 5\beta 3\gamma 2_{WT}$; 0.05 % n-dodecyl-beta-maltoside (DDM) for $\alpha 5\gamma 2_{HET\Delta}$ and $\alpha 5_{HOM}$. Samples were incubated with WGA YSI beads (bind N-linked glycans, beads at 2 mg/ml, Perkin Elmer) for 30 minutes at 4 °C under slow rotation. 50 µl aliquots of the GABA_A receptor-bead mix were added to 50 µl aliquots of 2 x radioligand ([³H]-flunitrazepam or [³H]-flumazenil) concentrations ranging from 0.06-2000 nM (Perkin Elmer) in Serocluster 96-Well ‘U’ Bottom plates (Corning) and incubated for 60 minutes at room temperature (20-22 °C) and [³H] cpm were determined by scintillation proximity assay using a Microbeta TriLUX 1450 LSC. The same ligand binding assay was performed in the presence of 50 µM flumazenil to ascertain the non-specific binding (NSB), which was subtracted from the total radioligand cpm to obtain the specific binding values. [³H]-flunitrazepam binding affinity (K_d) was calculated in OriginPro2015 using the one-site binding curve fit equation ($y = B_{max} * x / (kI + x)$), or two-site binding curve fit equation ($y = B_{max1} * x / (kI + x) + B_{max2} * x / (k2 + x)$), or using the Hill equation ($y = B_{max} * x^n / (kI^n + x^n)$) where B_{max} values are maximal binding for each site and kI and $k2$ are K_d for each site, n is Hill slope, x is ligand concentration, y is proportion of binding. Displacement curves were performed by adding ligand (bretazenil or diazepam) over the concentration range 1-50000 nM to aliquots of GABA_A receptor-bead mix for 30 minutes, then

adding this to aliquots of radioligand ($[^3\text{H}]$ -flumazenil or $[^3\text{H}]$ -flunitrazepam, respectively) at final concentrations corresponding to approximately $10 \times K_d$. Diazepam displacement curves were plotted on log concentration axis and fitted using the logistic equation ($y = A2 + (A2 - A1) / (1 + (x/x0)^p)$) where $A2$ and $A1$ are maximal and minimal binding respectively, $x0$ is IC_{50} and p is the Hill coefficient. IC_{50} values of displacement curves were converted to K_i values according to the Cheng-Prusoff equation¹²⁵, $K_i = IC_{50} / (1 + ([L] / K_d))$ referring to the $[^3\text{H}]$ -flumazenil K_d and the bretazenil IC_{50} , and where L is the concentration of $[^3\text{H}]$ -flumazenil used in the displacement assay.

Thermostability binding experiments. Information regarding the thermostability of a detergent-solubilised protein can be determined by heating protein samples over a range of temperatures for equal time periods and then measuring the reduction in the intensity of the monodisperse SEC profile for each protein sample¹²⁶. With increasing temperature an increased proportion of protein is denatured, aggregates and is lost from the monodisperse peak when the protein is subsequently run on SEC. A measure of protein stability can then be obtained by plotting the decay in peak UV absorbance against increasing temperature, for example to obtain a 50 % melting temperature (T_m). Purified $\alpha 1\beta 3\gamma 2_{EM}$ at 0.02 mg ml^{-1} (100 nM) in 150 mM NaCl, 10 mM HEPES pH 7.2, 0.007 % DDTM (w/v) was separated into 50 μl aliquots in PCR tubes, and heated at a range of temperatures from 30-80 °C for 1 hour. Samples were then run on a high-performance liquid chromatography system with automated micro-volume loader (Shimadzu) through a Superdex 200 Increase 3.2/300 column (GE Healthcare) maintained in 300 mM NaCl, 10 mM HEPES pH 7.2, 0.007 % DDTM (w/v). Monodisperse peak reduction with increasing temperature was measured relative to an unheated control sample maintained at 4 °C.

Importantly, because some drugs when bound thermostabilise detergent-solubilised protein¹²⁶, the thermostability assay offers an efficient strategy to measure protein sensitivity to drugs in the detergent-solubilised environment. Purified GABA_AR Cryo-EM construct was separated into PCR tubes, supplemented with picrotoxin at a range of concentrations and heated at T_m30 % (the temperature at which the monodisperse peak was reduced by 70 %) for 1 hour. Afterwards samples were run on a high-performance liquid chromatography system with automated micro-volume loader (Shimadzu) through a Superdex 200 3.2/300 column (GE Healthcare) maintained in 300 mM NaCl, 10 mM HEPES pH 7.2, 0.007 % DDTM. Drug dose-response curves were generated by plotting UV absorbance against drug concentration.

Surface plasmon resonance analysis (SPR). SPR measurements were performed using a Biacore T200 (GE Healthcare) machine at 25 °C. All reagents and consumables for SPR were purchased from GE Healthcare. The carboxyl groups on CM5 chip flow channels were activated with a 10-minute injection of a 1:1 mixture of 0.1 M N-hydroxysuccinimide (NHS) and 0.4 M 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (EDC). Streptavidin (Sigma-Aldrich) was covalently bound by a 5-minute injection until an immobilization level of 5000 RU was reached. Free activated carboxyl groups were quenched with a 10-minute injection of 1 M ethanolamine-HCl pH 8.5. The working flow-cell was functionalized by injecting GABA_A receptor electron cryo-microscopy construct containing a biotinylated C-terminus containing a biotin ligase recognition sequence, until 350-375 RU were reached, whereas the reference cell containing streptavidin was left unmodified. Running buffer contained 10 mM HEPES pH 7.5, 150 mM NaCl, 0.007% DMNG:CHS (5:1, w/w). For experiments testing GABA effect on Nb38 binding, 1 mM GABA was supplemented to the running buffer. Measurements were performed by injecting nanobodies at concentrations ranging from 2.5 to 40 nM (two-fold serial dilutions) during a single cycle. For reliable single cycle kinetics (SCK) data fitting, the final

dissociation phase was set to 15 minutes. Biacore T200 evaluation software was used to analyse all the SCK data. A 1:1 binding model was used to fit the experimental results.

HEK cell preparation and electrophysiology. One day prior to experiments, 8 ml of Dulbecco's Modified Eagle Medium (DMEM) was pre-incubated for 10 min at room temperature with 96 μ l lipofectamine2000 (Thermofisher) and 48 μ g plasmid DNA, then added to a single T175 cm² flask containing HEK293T cells (30-50 % confluency) and 2 ml DMEM (supplemented with 10 % fetal calf serum, L-Gln and non-essential amino acids). After 3 hrs this media was removed and replaced by DMEM supplemented with 10 % fetal calf serum. For GABA_AR heteromers, pHLsec plasmids containing human cDNA constructs were mixed in 1:1:2 ratio (α : β : γ), supplemented with 3% plasmid encoding enhanced green fluorescent protein (EGFP) to assess transfection efficiency. The electron cryo-microscopy α 1 β 3 γ 2 construct used was as described in 'Construct design'. The wild-type cDNA inserts used for heteromeric receptor expression were as follows: human GABA_AR α 1 mature protein sequence (α 1 Uniprot P14867 entry, Gln28 is Gln1, 1-429 QPSL...PTPHQ) and human β 3 mature protein sequence (β 3 Uniprot P28472 entry isoform 1, Gln26 is Gln1, 1-448 QVSN...LYYVN) cloned into the pHLsec vector³⁴ between the N-terminal secretion signal sequence and a double stop codon; Human GABA_AR γ 2 mature protein sequence (γ 2 Uniprot P18507 isoform 1 entry Gln40 is Gln1 1-428 QKSDD...SYLYL) cloned into the pHLsec vector³⁴ between the N-terminal secretion signal sequence followed by streptavidin binding protein (MDEKTTGWRGGHVVEGLAGELEQLRARLEHHPQGQREP) and a C-terminal 1D4 purification tag. Transfection efficiencies were typically 50-80 % (cells expressing EGFP, as estimated by fluorescence microscopy). Eighteen to twenty-four hours later cells were washed with phosphate buffered saline, incubated in 4 ml TrypLE (Gibco) for 7 min at 37 °C, suspended in 21 ml DMEM supplemented with 10 % fetal calf serum and L-Gln, centrifuged

at 100 g for 1.5 min, then suspended in 50 ml Freestyle 293 Expression Medium (Gibco) and placed in a shaking incubator (130 rpm, 37°C, 8 % CO₂) for 30 min. 25 ml cell suspension was then centrifuged at 100 g for 1.5 min, and suspended in 4 ml external recording solution. This solution contained (mM): 137 NaCl, 4 KCl, 1 MgCl₂, 1.8 CaCl₂, 10 HEPES, and 10 D-Glucose, pH 7.4 (≈ 305 mOsm). The internal recording solution contained (mM): 140 CsCl, 5 NaCl, 1 MgCl₂, 10 HEPES, 5 EGTA, 0.2 ATP, pH 7.35 (≈ 295 mOsm). Electrophysiological recordings were performed at room temperature using an Ionflux16 (Molecular Devices) in ensemble mode, with series resistance compensation set at 80 % and cells held at -60 mV. Diazepam and picrotoxin (Sigma-Aldrich) were dissolved in DMSO as 100 mM and 1 M stocks respectively prior to dilution in external recording solution. Diazepam and Nb38, or picrotoxin, were co-applied with EC₁₀₋₁₅ GABA or EC₇₀ GABA doses respectively to generate dose response curves. Expression of heteromeric receptors as assemblies of $\alpha\beta\gamma$ subunits was confirmed by response to GABA, which requires co-assembly of $\alpha 1$ and $\beta 3$ subunits, and efficient inclusion of the $\gamma 2$ subunit into $\alpha\beta\gamma$ heteromers was verified by measuring low sensitivity to 100 μ M Zn²⁺ inhibition, defined as less than 50 % inhibition of an EC₅₀ GABA response¹²⁷.

Brain slice preparation and electrophysiology. All work on animals was carried out in accordance with the UK Animals (Scientific Procedures) Act 1986 under project and personal licenses granted by the UK Home Office. 250 μ m thick transverse acute brain slices were prepared from adult (P67-84) male C57BL/6J mice using a Leica VT1200S vibroslicer. After terminal anaesthesia with isoflurane the brain was rapidly removed and sliced in an ice-cold slicing solution composed of (mM): 85 NaCl, 2.5 KCl, 1 CaCl₂, 4 MgCl₂, 1.25 NaH₂PO₄, 26 NaHCO₃, 75 sucrose, and 25 glucose, 2 kynurenic acid, pH - 7.4; bubbled with 95% O₂ and 5% CO₂. The slicing solution was exchanged at 37 °C for 60 min under gravity flow with a recording solution or artificial cerebrospinal fluid (ACSF) containing (mM): 125 NaCl, 2.5

KCl, 2 CaCl₂, 1 MgCl₂, 1.25 NaH₂PO₄, 26 NaHCO₃, 2 kynurenic acid, and 25 glucose, pH - 7.4.

Spontaneous inhibitory postsynaptic currents (sIPSCs) were recorded from hippocampal dentate gyrus granule cells (DGGCs) using patch electrodes of 4-5 MΩ resistance filled with an internal solution containing (mM): 120 CsCl 1 MgCl₂, 11 EGTA, 30 KOH, 10 HEPES, 1 CaCl₂, and 2 K₂ATP; pH - 7.2. sIPSCs were recorded at -60 mV using 5 kHz filtering and optimal series resistance and whole-cell capacitance compensation.

sIPSCs were analysed using WinEDR and WinWCP (John Dempster, University of Strathclyde, UK). Percentage change in IPSC amplitudes, frequencies, rise times, decay times and charge transfers were calculated in the presence of nanobody or diazepam in comparison to controls in normal recording solution (aCSF). The statistical significance of sIPSC parameters in the presence or absence of treatments in each cell was assessed using a paired t-test in Graphpad Instat.

Natural genetic variation in ligand-binding residues. To investigate the extent of natural genetic variation in the BZD binding site, ligand-binding residues were identified using in-house written Perl scripts, available upon request from authors. Residues making atomic contacts with the ligands within 5Å distance were classified as ligand-binding residues. The number of such contacts were also calculated and are provided in the asteroid plots¹²⁸. Conservation profiles for ligand binding positions across all human GBRA and GBRG subunits were then generated. We retrieved genetic variation data in different GBRA and GBRG receptors in 138,632 unrelated healthy humans from gnomAD database⁵⁴. Genetic variation data was retrieved only for non-engineered positions and those that have identical residues as that of GBRA5 or GBRG2.

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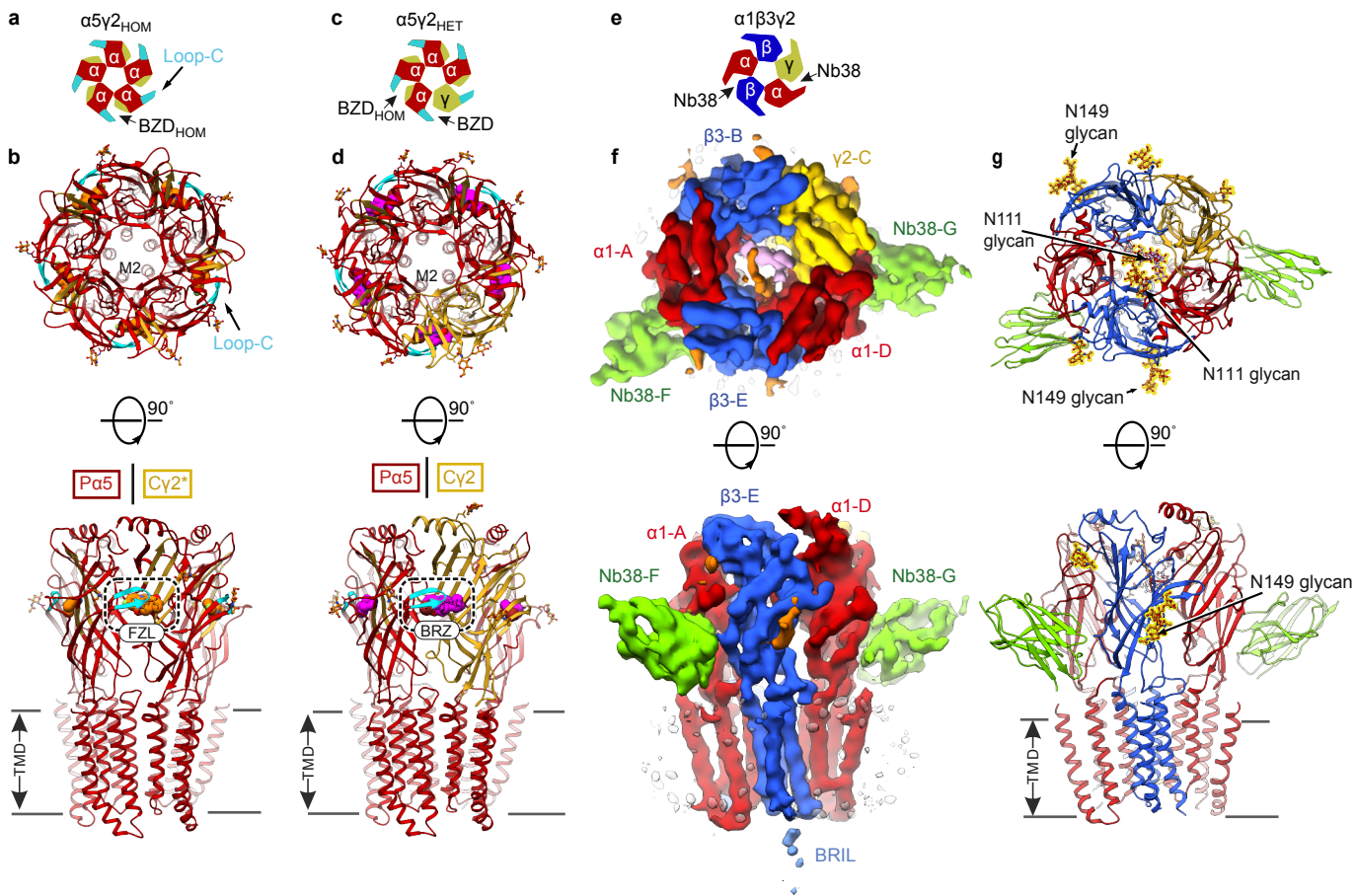


Figure 1

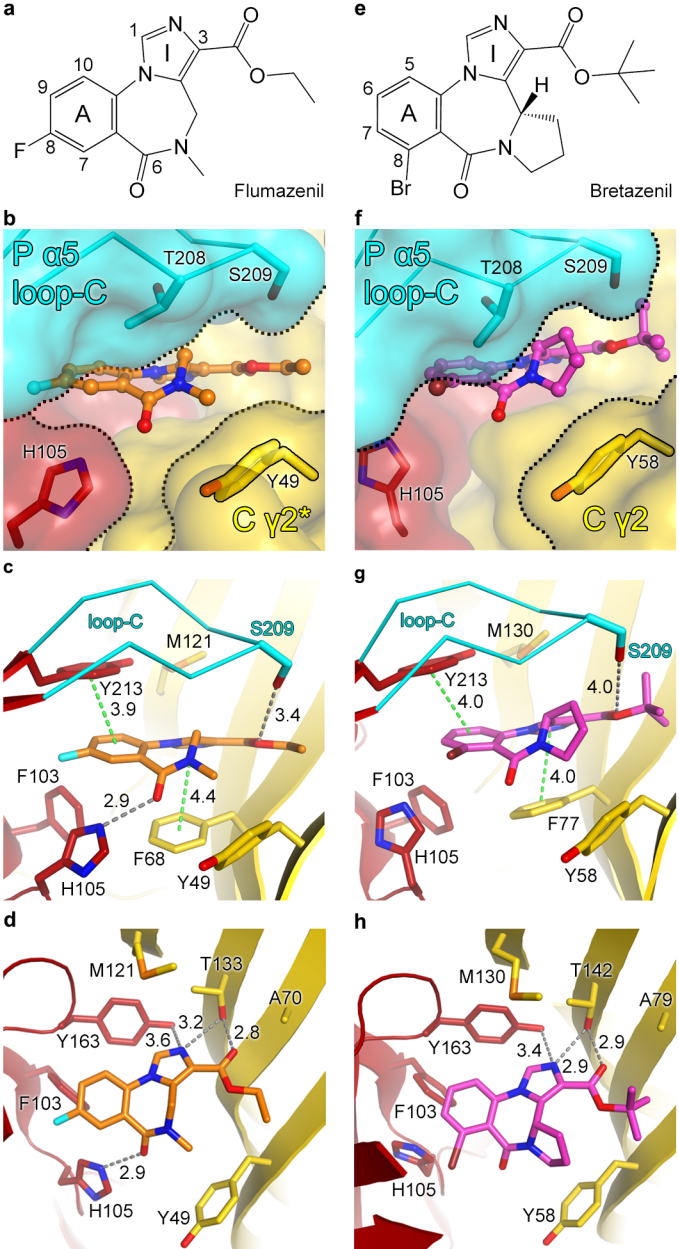


Figure 2

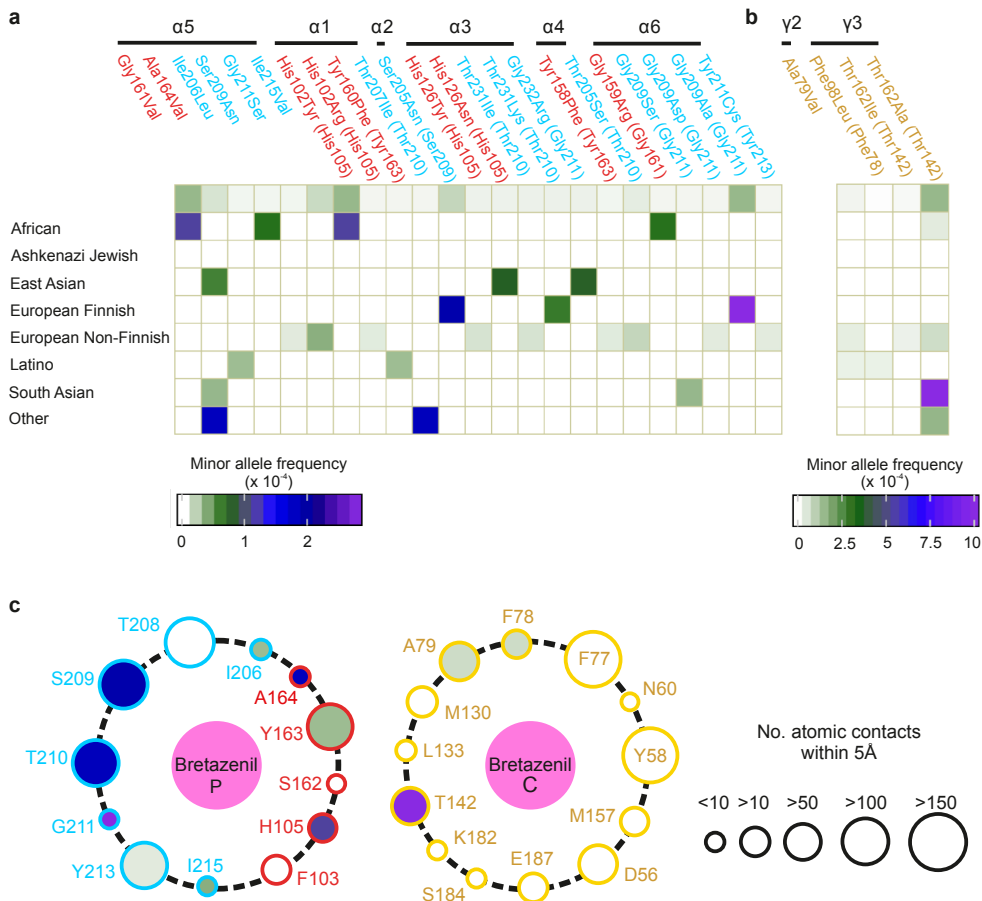


Figure 3

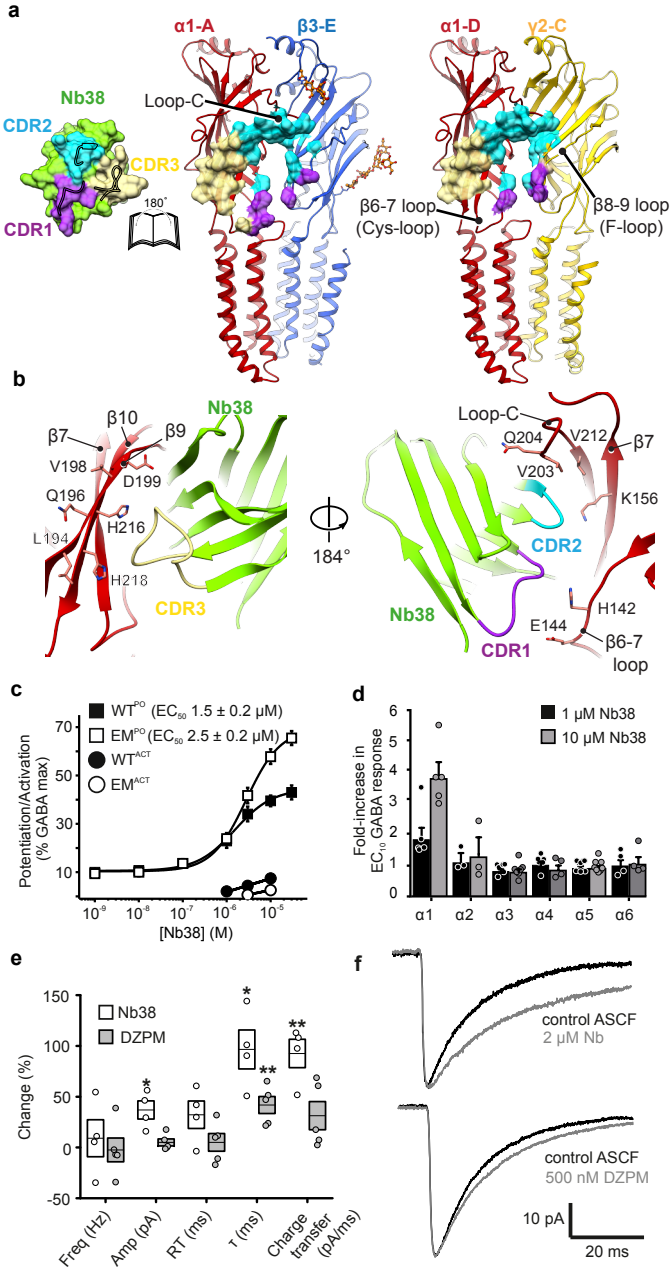


Figure 4

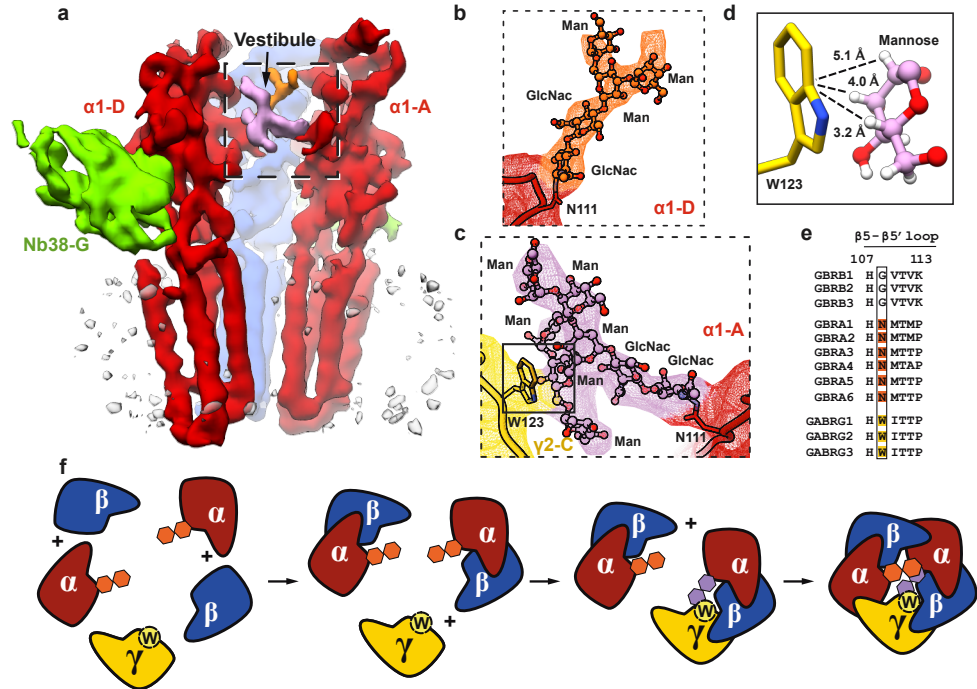


Figure 5

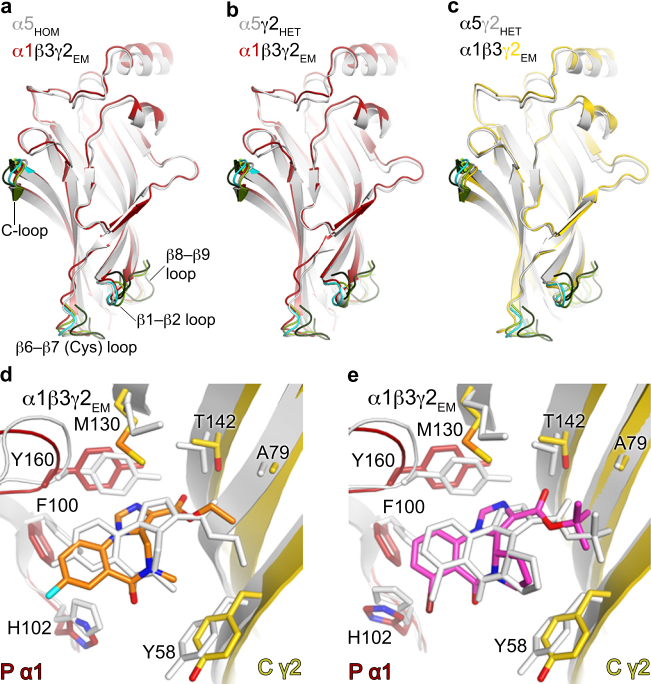


Figure 6

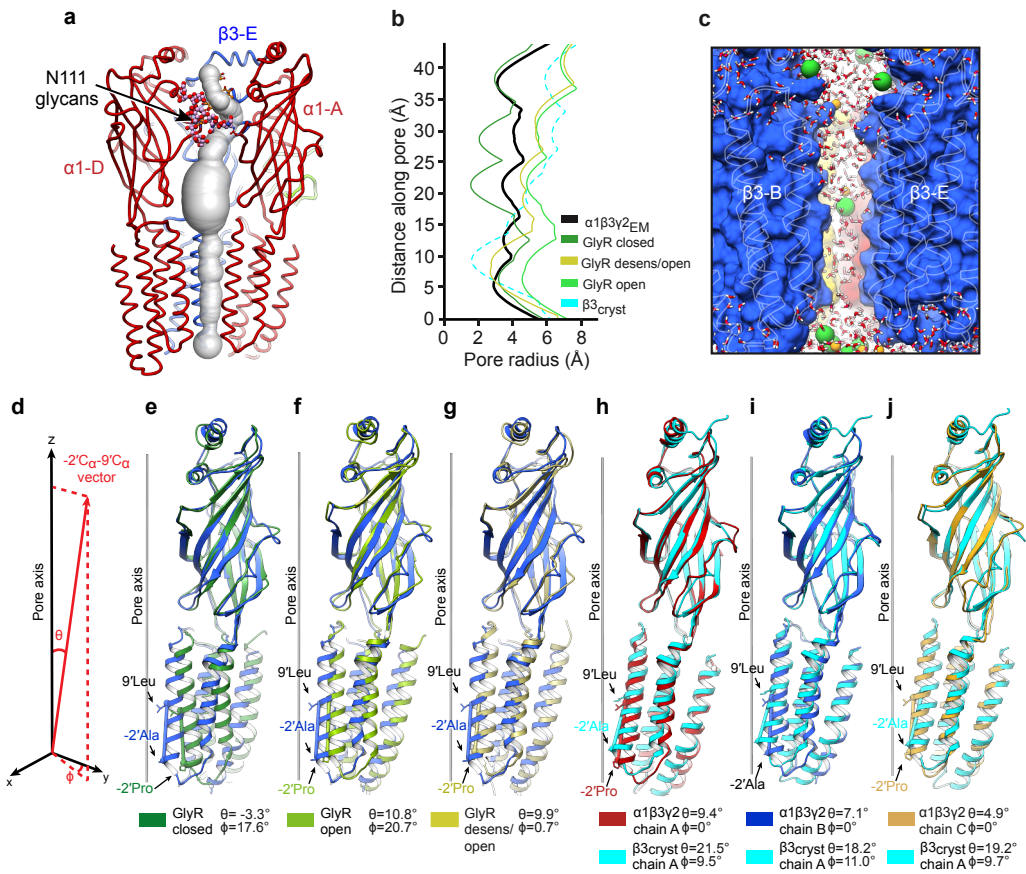


Figure 7