

Mapping molecular HLA typing data to UNOS antigen equivalents for improved virtual crossmatch

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Disclosure: The authors declare no conflict of interest

Abbreviations

AFA, African American

ALLAN, ALLele to ANtigen

API, Asia / Pacific Islander

CAU, Caucasians

CPRA, calculated panel reactive antibodies

GL, genotype list

HIS, Hispanics

HLA, human leukocyte antigen

HML, Histoimmunogenetics Markup Language

HSCT, hematopoietic stem cell transplantation

IHIWS, International HLA and Immunogenetics Workshop

L/Q, Low expression or Questionable expression alleles

MAC, multiple allele code

NGS, next-generation sequencing

NMDP, National Marrow Donor Program

OPO, organ procurement organization

OPTN, Organ Procurement and Transplantation Network

PCR, polymerase chain reaction

REST, representational state transfer

SSOP, sequence-specific oligonucleotide probes

SSP, sequence-specific primers

UNOS, United Network for Organ Sharing

WHO, World Health Organization

WMDA, World Marrow Donor Association

Abstract

Background: Virtual crossmatch utilizes HLA typing and antibody screen assay data as a part of organ offers in deceased donor allocation systems. Histocompatibility labs must convert molecular HLA typings to antigen equivalencies for entry into the United Network for Organ Sharing (UNOS) UNet system. While an Organ Procurement and Transplantation Network (OPTN) policy document provides general guidelines for conversion, the process is complex because no antigen mapping table is available. We present a UNOS antigen equivalency table for all IMGT/HLA alleles at the A, B, C, DRB1, DRB3/4/5, DQA1, and DQB1 loci.

Methods: An automated script was developed to generate a UNOS antigen equivalency table. Data sources used in the conversion algorithm included the World Marrow Donor Association (WMDA) antigen table, the HLA Dictionary, and UNOS-provided tables. To validate antigen mappings, we converted National Marrow Donor Program (NMDP) high resolution allele frequencies to antigen equivalents and compared with the UNOS Calculated Panel Reactive Antibodies (CPRA) reference panel.

Results: Normalized frequency similarity scores between independent NMDP and UNOS panels for 4 US population categories (Caucasian, Hispanic, African American and Asian/Pacific Islander) ranged from 0.85 to 0.97, indicating correct antigen mapping. An open source web application (ALLEle to ANtigen (“ALLAN”)) and web services were also developed to map unambiguous and ambiguous HLA typing data to UNOS antigen equivalents based on NMDP population-specific allele frequencies (<http://www.transplanttoolbox.org>).

Conclusions: This tool sets a foundation for using molecular HLA typing to compute the virtual crossmatch and may aid in reducing typing discrepancies in UNet.

Introduction

Human leukocyte antigen (HLA) specificities were originally distinguished by allosera from multiparous women or individuals with multiple blood transfusions using cell-based testing strategies. HLA typing is currently performed by more accurate and specific molecular methods that assay nucleotide sequences. The IMGT/HLA database was developed to catalog the growing number of known HLA gene sequences¹. While the naming of HLA sequences has its roots in the antigen nomenclature from serologic typing², new HLA alleles are named based on sequence similarity to known alleles.

The United Network for Organ Sharing (UNOS) organ allocation system UNet utilizes donor and recipient HLA typing information and a list of unacceptable HLA antigens determined from antibody assays to perform virtual crossmatch as part of organ offers^{3,4}. While the more specific IMGT/HLA allele nomenclature is used to interpret DNA-based typing data, only serologic antigen equivalents are accepted by the UNet⁵. Antigen groups are used to represent unacceptable HLA specificities because a single allo-HLA antibody may react similarly against several distinct HLA alleles within the same antigen group.

Histocompatibility labs face a data management challenge in mapping molecular-level HLA data to UNOS antigen equivalents for entry into UNet. While the Organ Procurement and Transplantation Network (OPTN) published a policy document in 2003 with general guidelines for mapping molecular data to antigen equivalents⁶, there is a need to automate the mapping process with a standardized mapping table that includes

every IMGT/HLA allele. UNOS deemed it infeasible to maintain such a table at the time. Because of the difficulty in synthesizing antigen equivalency information from multiple reference data sources, there is likely to be some variability in reporting of HLA typing results into UNOS systems, even among commercial typing platforms.

A rapid turnaround for HLA typing is necessary for limiting cold ischemic time, therefore deceased donors are typed by sequence-specific primer (SSP), sequence-specific oligonucleotide probe (SSOP), or real-time polymerase chain reaction (PCR) based methods rather than next-generation sequencing (NGS). These methods rarely achieve allele-level specificity. Hence, there is also a need to interpret ambiguous HLA typings in the context of population HLA frequencies, as OPTN guidelines call for reporting UNOS antigens based on the most common HLA allele.

To meet these challenges in HLA data reporting for virtual crossmatch, we were inspired by matching systems for hematopoietic stem cell transplantation (HSCT) that have the capability to represent HLA typing data in IMGT/HLA allele nomenclature and match to donors whose HLA is represented in antigen nomenclature⁷. The World Health Organization (WHO) Nomenclature Committee for Factors of the HLA System maintains a list of official WHO antigen equivalencies for common HLA alleles. For HSCT registry matching, the IMGT/HLA database also maintains a World Marrow Donor Association (WMDA) antigen equivalency table⁸ that is updated quarterly with each database release⁹. No such mapping table exists for UNOS antigen equivalents. UNOS antigen equivalents differ from WHO and WMDA antigens for some alleles and we found these

differences can be derived from interpreting UNOS/OPTN guidelines and using UNOS-provided tables that list IMGT/HLA alleles that are exceptions to these guidelines.

To improve fidelity in the communication of HLA data from histocompatibility labs to the UNOS allocation system, we describe a mapping table to convert IMGT/HLA alleles to UNOS antigen equivalents. We have also made available open source informatics tools to implement the aforementioned UNOS/OPTN guidelines, to keep the mapping table current with each IMGT/HLA database release, and to perform conversion of both ambiguous and unambiguous molecular HLA typing to UNOS antigens. We further describe how these tools will enable molecular HLA typing data to be used directly to compute the virtual crossmatch.

Materials and Methods

Data Sources

UNOS/OPTN Policy Documents for “Interpretation of HLA Typing Results for Entry into UNet”: We have summarized the rules for histocompatibility labs to assign UNOS antigens for IMGT/HLA alleles in Table 1. These rules were developed based on interpretation of guidelines published in 2003 in an OPTN policy document⁶. The OPTN documents^{5,6} contain tables of UNOS antigen equivalencies for some IMGT/HLA alleles that differ from WHO-assigned antigens. These tables include some 4-digit antigens representing alleles that may be readily identified by low resolution molecular testing as well as antigens that were not recognized by WHO serologic nomenclature at the time

of policy implementation. The UNOS Histocompatibility Committee issued updates in 2017 that specified antigen assignments for some additional IMGT/HLA alleles¹⁰.

HLA Dictionary 2008: To standardize HLA antigen assignments for stem cell donor registries, the WMDA maintains an HLA Dictionary¹¹. The 2008 version of the HLA Dictionary describes serologic equivalents of alleles for 6 loci (A, B, C, DRB1, DRB3/4/5, and DQB1). The data sources used to compile the dictionary include the following: WHO serological assignments as described in the Nomenclature report of 2004¹², data submitted by individual laboratories to WHO committee for factors of HLA system, the international cell exchange program that has 200 laboratories participating in evaluating the standardized sera, NMDP-assigned serological equivalents, and data submitted for 13th International HLA and Immunogenetics Workshop (IHIWS). Besides these sources that assigned serotypes based on the testing of cells with sera, the dictionary also lists antigens that were computationally assigned by a neural network¹³. Final consensus antigen assignments are listed as an expert-assigned type.

WHO and WMDA Antigen Equivalents Maintained by IMGT/HLA: The quarterly updated IMGT/HLA database curates the sequences of HLA alleles. The current release (v.3.32.0) includes 16,886 alleles for A, B, C, DRB1, DQA1, DQB1 and DRB3/4/5 loci. Each release includes a file with the WHO and WMDA antigen assignments for every IMGT/HLA allele⁸ (rel_ser_dna.txt, referred as the “WMDA File” in this paper). There are several categories of antigen equivalencies in this file based on the amount of information available from serologic testing. Many of the common

IMGT/HLA alleles have WHO antigen assignments with unambiguous serology. WHO assignments were either made by the nomenclature committee when the allele sequence was submitted or were based on serologic testing data from the HLA Dictionary¹¹. Possible serological antigens are given when there is no serologic testing data and are separated by a slash (“/”) when there is more than one possibility. Assumed antigens are typically based on first field of the IMGT/HLA allele name. WMDA expert-assigned antigens come from search determinants assigned by stem cell donor registries. UNOS antigen assignments for new IMGT/HLA alleles can thus rely upon updates to this WMDA file.

NMDP Allele Frequencies for Assigning Antigens for Ambiguous HLA typing: We used population-specific high resolution allele frequencies to calculate the frequencies of all possible UNOS antigens and provide the most probable antigen as the best assignment. High resolution allele frequencies for 4 broad US race/ethnic groups (Caucasian (CAU), African American (AFA), Hispanic (HIS), and Asian/Pacific Islander (API)) for the A, B, C, DRB3/4/5 DRB1, and DQB1 loci were obtained from NMDP¹⁴. These frequencies have been derived from 6.59 million volunteer donors from the Be The Match registry for HSCT. The NMDP US frequencies also include the Native American broad race and 21 detailed race/ethnic subcategories that could potentially be selected to make more precise antigen assignments.

Implementation

Conversion Table Program and Functions: The program to generate the IMGT/HLA allele to UNOS antigen conversion table was written in Python. The conversion table includes all current IMGT/HLA alleles at A, C, B, DRB1, DRB3/4/5, DQA1, DQB1 loci and their respective UNOS antigen equivalents according to UNOS guidelines shown in Table 1. A command line Python conversion script supports four different input forms of HLA typing results: unambiguous HLA alleles (either a single allele or a list of alleles) and ambiguous multi-locus HLA typing (represented as a genotype list string (GL string)¹⁵ or as NMDP multiple allele codes (MACs)¹⁶). For ambiguous HLA typings, a US population must be selected, as the most probable antigens are calculated given the population-specific allele frequencies and the antigen mappings for the alleles. An up-to-date list of MACs is accessed via the NMDP MAC web service (<https://hml.nmdp.org/mac/>). A function for reverse mapping of an unacceptable UNOS antigen to a list of IMGT/HLA alleles is also provided. DQA1 antigen mapping for ambiguous typing is not included in this release, as interpretation requires population-specific allele frequencies that are under development by NMDP.

Command Line Tool, Web Application, and Web Services: We developed a web application named ALLele to ANtigen (“ALLAN”) for users to enter molecular HLA typings and perform UNOS antigen conversion. The tool is available at <http://www.transplanttoolbox.org>. This web tool was implemented using the Django web framework¹⁷. As a programming interface, Representational State Transfer (RESTful)¹⁸ enabled web services available at http://www.transplanttoolbox.org/tool_services were

developed using Django REST Framework¹⁹. An example Python script that illustrates how to access the web services is also provided in our GitHub source code repository (<https://github.com/lgragert/hla-who-to-unos>). For advanced users who wish to run the conversion tool locally, a Python command line tool and pip installable package “transplanttoolbox-allan” are also available (<https://pypi.org/project/transplanttoolbox-allan/>).

Antigen Frequencies and Similarity Index Calculation

Reference antigen frequencies for 4 broad US race/ethnic categories (CAU, AFA, HIS, and API) were generated by mapping NMDP haplotype frequencies to UNOS antigen equivalencies. To assess accuracy in antigen assignments we used a normalized similarity index, “ I_F ”²⁰, to compare the converted NMDP antigen frequencies to an independent antigen-level frequency dataset of previous UNOS donors²¹ that is currently used operationally as the reference panel for measuring CPRA values.

Results

Mapping Serological Specificities for IMGT/HLA Alleles Based on UNOS

Guidelines

An algorithm was designed to map IMGT/HLA alleles to antigens following the UNOS guidelines. Our interpretation of the rule precedence for antigen mapping guidelines is shown as a schematic diagram in Figure 1. The antigen assignments found in tables from UNOS histocompatibility committee updates, UNOS antigen equivalency tables for specific alleles (Table 1: Suggested UNOS serology equivalents for molecular types

(2003))⁶, and 4-digit antigens defined by UNOS were given the highest precedence⁵.

For the next level of precedence, WHO-assigned antigens from the WMDA file were assigned (see Rule 3 in Table 1) followed by antigens from HLA Dictionary 2008¹¹. No antigens are assigned for DQA1 locus alleles in the WMDA file, however, UNOS has added 2nd-field allele digits as DQA1 antigen equivalencies in their latest update (e.g. DQA01:01 for DQA1*01:01:01:01).

If antigens could not be assigned from the data sources listed above, then possible and assumed antigens from the WMDA file were used. Cases where we fell back to assigning 2-digit allele equivalencies when information from all other data sources was lacking were mostly restricted to B and C locus alleles. When in doubt, we erred on the side of assigning a broader antigen category rather than assuming a split antigen.

A complete conversion table of UNOS antigen equivalencies for the IMGT/HLA alleles in release v.3.32.0 is available as Supplementary Table 1. We also provide a table that lists the IMGT/HLA alleles that correspond to an unacceptable antigen as Supplementary Table 2. Future conversion tables will be available on <http://www.transplanttoolbox.org> in tandem with quarterly updates to IMGT/HLA.

Precedence Order when Multiple Rules for Assigning Antigen Specificity may Apply

Multiple guidelines for antigen assignments may apply to a single IMGT/HLA allele, therefore an order of precedence for these guidelines must be set. When the UNOS

antigen assignment listed in the UNOS-provided table conflicted with the WHO assignment, we chose the UNOS table. For example, for allele B*15:29, the WHO assignment is B15, whereas the UNOS assignment is B70. It is not always correct to simply reduce allele nomenclature to 2-digits to get antigen specificity because different alleles from the same allele group can have different antigens. This is most evident in the B*15 group, where alleles may have antigen equivalents of B15, B62, B63, B70, B71, B72, B75, B76, or B77. Rule precedence resolves cases where different data sources suggest differing antigen assignments, as we illustrate with several examples in Table 2.

Most Probable Antigen for Ambiguous HLA Typing

As per OPTN guidelines, for a group of possible alleles in an ambiguous HLA typing, the serologic equivalent of the most common antigen should be entered. We apply reference population-specific allele frequencies from the NMDP population categories to make the assignment. We show that for a particular ambiguous HLA typing, the most probable antigen for the B locus may vary depending on the broad race/ethnic category selected (Table 3).

“ALLAN”: IMGT/HLA ALLele to UNOS ANtigen conversion tool

The conversion functions described above are available via the ALLAN web tool at <http://www.transplanttoolbox.org>. When the HLA typing is unambiguous, users may select to enter either a single allele or a list of alleles. When the HLA typing contains typing ambiguity, users may enter the typing either in GL string or MAC format (Table

3). The output shows the UNOS antigen assignments and Bw4/6 epitopes. Entry of ambiguous HLA typings requires the selection of a US race/ethnic group and returns the probability distribution of all the possible antigens considering the allele frequencies in the selected race/ethnic group. A complete user guide for the tool is available on the website. For advanced users, we also developed a Python script as a command line tool with similar functionality, which can be accessed via our GitHub repository (<https://github.com/lgragert/hla-who-to-unos>). We have also developed web services as a programming interface for researchers and laboratory information management systems. Client applications such as HLA typing software platforms or other lab information systems can send HLA typing and race/ethnicity information to the service, and UNOS antigens (including probabilities) are returned. The endpoint URLs and commands for these services are shown in Table 4.

Antigen Mappings in Commercially Available Software Programs

Most histocompatibility labs rely on commercial software to analyze HLA typing results and perform antigen conversion. Currently, the software platforms from leading vendors in this domain, i.e. One Lambda (HLA Fusion) and Immucor (LIFECODES® MATCH IT!®), perform antigen mapping for a limited number of IMGT/HLA alleles but would benefit from a complete standardized table. “HLA Fusion” does mapping for 4,927 alleles and “MATCH IT!” does mapping for 4,439 of the 16,886 alleles. Compliance with IMGT database updates for new releases of HLA alleles and UNOS updates represents a major maintenance burden for these software vendors. Vendors must rely on available sources such as the HLA Dictionary, which has not been updated since 2008.

The WMDA does provide quarterly updates, but again vendors are left with choosing between “Unambiguous associations,” “Possible associations,” “Assumed associations,” and “Expert assigned exceptions,” and have to reconcile these options with customer specific requests. As a result, some antigen mappings made by these software packages differ from those recommended by UNOS. A list of discrepancies from apparent UNOS guidelines, apart from the missing mappings, is shown in Table 5. The commercial software vendors are in agreement that the issues arising from independent maintenance and interpretation of antigen conversion rules will be alleviated by having a consensus set of mappings.

Reference Antigen Frequencies for Six HLA Loci in 4 Broad US Races

To validate the antigen assignments, we converted published NMDP high resolution haplotype frequencies¹⁴ into antigen equivalents and compared antigen frequencies at each HLA locus to the UNOS CPRA HLA reference panel. Antigen-level NMDP frequency tables for 4 US broad race/ethnic categories are available as Supplementary Table 3. The normalized “ I_F ” similarity index for these races ranged from 0.85 to 0.97, indicating accuracy in antigen assignments (Table 6). One cause of disparity between panels was that the UNOS DQ typing data was more often reported as broad DQ1 antigens rather than split DQ5 and DQ6 antigens. Meanwhile, the only high resolution IMGT/HLA alleles in the NMDP panel that mapped to broad DQ1 were the relatively uncommon alleles DQB1*06:11 and DQB1*06:12.

Discussion

The OPTN guidelines for HLA allele to antigen mapping require that histocompatibility labs refer to several different data sources which as we have shown here can lead to differences in antigen assignment. To streamline this process, we present a complete mapping table between all current IMGT/HLA alleles and UNOS antigens that was developed based on published OPTN guidelines. This conversion table report gives a standardized source for antigen mapping that can be updated in tandem with every new IMGT/HLA database release.

Our web tool, ALLAN, offers a convenient mapping of HLA typing data to UNOS antigens for entry into UNet, including resolving HLA typing ambiguities in antigen assignments as shown in Table 3. This will aid in streamlining HLA typing that is performed for deceased donors, which has limited time for experimentally resolving ambiguities. Molecular methods such as SSP may not always resolve null expression variants in time, however, our tool helps in predicting the probability of null alleles and therefore a blank antigen for a specified population.

One potential cause of unexpected positive tissue crossmatch²² post organ shipment is inaccuracy in manual HLA data entry, as the accuracy of prediction of negative virtual crossmatch relies solely on HLA data mapped and entered. A report from 22 organ procurement organizations (OPOs) describing the allocation practices of kidneys in high PRA patients indicates that a substantial number of kidneys shipped were transplanted in unintended recipients (19.2%) or discarded (3.5%) because of a positive crossmatch

²³. An analysis by the Discrepant HLA Typing Subcommittee of the UNOS Histocompatibility Committee showed that among 18,719 deceased donors whose HLA data were entered into UNet from 2015-2016, 6,670 (35.6%) had some discrepancy in their typing including allele vs antigen resolution, broad/split antigen nomenclature differences, or incorrect Bw4/6 or DRB3/4/5 assignments. 2% had a critical antigen-level discrepancy that would have affected the match run or allocation (Personal communication – Peter Lalli ²⁴). Automated management of HLA typing data, including the use of a standardized antigen mapping table or direct use of molecular typing data to compute the virtual crossmatch, has the potential of reducing such HLA discrepancies and the need for repeat match runs, thus reducing the cold ischemia time. Further improvement in accuracy of typing data in UNet could be achieved via adoption of the Histoimmunogenetics Markup Language (HML) data standard that is used today for automated transfer of HLA typing data from laboratories into the NMDP stem cell donor registry²⁵.

Out of 16,886 current IMGT/HLA alleles mapped to UNOS antigen equivalents, 12,489 had clear assignments where only one of the OPTN mapping guidelines would apply, while the remaining required us to interpret rule precedence. The high similarity between independent UNOS and NMDP derived antigen frequencies increases confidence in the accuracy of the mapping table. A lower similarity between API samples compared to other populations is likely due to a shallow sampling of only 333 Asian individuals in the UNOS data.

In recent updates, UNOS increased the number of 4-digit unacceptable antigens that can be reported. Because many of these 4-digit antigens are not included in the UNOS antigen frequencies for CPRA²⁶ and do not increase CPRA values when selected as unacceptable antigens, we chose to map to broader antigens to ensure compatibility with the current UNOS system.

Bw4/Bw6 epitope categories are determined based on amino acid sequence for positions 77 through 83 of HLA-B, and there has not been serologic testing of some of the rare sequence motifs. The OPTN policy document⁶ does not include assignments for all Bw4/Bw6-defining motifs among IMGT/HLA alleles, therefore we used NMDP assignments, some of which were manually curated⁷. For the rare Bw4/Bw6 motifs, there were some discrepancies with the commercial HLA typing software programs. This highlights the need for an improved curation process that would lead to the availability of public Bw4 and Bw6 epitope assignment resources.

Few low (L) or questionable (Q) expression alleles were assigned antigens in the HLA dictionary and we have assigned antigens to all these alleles as if they were expressed. While L alleles have been confirmed to have weak surface expression that can elicit alloimmunity, little information is available for Q alleles²⁷. The current version of IMGT/HLA database includes 75 Q alleles, yet only 8 of these have been observed unambiguously in the NMDP registry donor file. Further examination of mRNA transcripts, cell surface expression, and tests of alloimmunity are needed to make definitive assignments for these alleles, but we prefer to err on the side of caution. The

assigning of antigens to L/Q alleles intends to avoid cases of false negative virtual crossmatch at a potential cost of increasing positive virtual crossmatch predictions.

OPTN guidelines do not call for histocompatibility laboratories to analyze HLA sequences. Therefore, decisions on antigen assignments are best maintained in the reference data sources such as the IMGT/HLA database, and the HLA dictionary. These resources synthesize information from individual reports, such as describing B64, B65 antigen splits of the broad B14 antigen defined by Street and Darke^{28,29} for B*14:01 and B*14:02. While B*14:03 had a shared amino acid motif with B*14:02 that could define the epitope, B*14:03 could be deemed a B65 based on sequence analysis. However, both WHO and the HLA dictionary call B*14:03 a B14. As another example, in a recent report³⁰ these authors describe the serological specificity of B*15:33 as a B62. OPTN guidelines call for the WHO assignment, which is B15³¹. If the allele lacked a WHO assignment, the 2008 HLA dictionary would be used to assign a B62. For the UNOS assignment to become B62, the WHO antigen would need to change or UNOS would need to specify an exception to the rule. In summary, these complex cases cannot all be sorted out here. We believe such decisions are best left to UNOS, WHO, or the next HLA Dictionary.

Standardized antigen assignments for molecular typing are necessary for the UNOS organ allocation system to evolve to incorporate DNA-based HLA assignments and epitope-based matching strategies³². We anticipate future updates to our tool to support changes in the UNOS CPRA to include antigens for the DQA1, DPA1, and DPB1 loci.

These loci together account for >60% of the unacceptable antigens in candidates with CPRA > 99%^{33,34}. We intend to develop additional tools that would allow the NMDP high resolution frequency panel to be used for allele-level CPRA calculations incorporating these additional loci. Imputation tools based on NMDP frequency data are also useful for estimating amino acid assignments from antigen-level data to perform epitope analysis with tools such as PIRCHE: predicted indirectly recognizable HLA epitopes³⁵. The antigen mapping tools we developed here will take UNOS one step closer to their long-term goal of maximally utilizing HLA molecular typing data in solid organ allocation systems.

Acknowledgments:

We thank Anat Tambur and Malek Kamoun for their advice and review of early drafts of this manuscript. We also thank the UNOS Histocompatibility Committee for their support of this project.

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Table 1: Rules for Mapping Molecular HLA typing to UNOS Antigen equivalents.

Rule Description	OPTN Policy Guidelines (Verbatim)	Data Source	Rule Notation*
Assign split antigens when typing serology (not applicable to this paper)	<i>“For serologic typing, enter all identified splits”</i> [□]	(OPTN policy document)	1_Sero_typing
Assign Bw4 and Bw6 epitope for all HLA-B alleles	<i>“For both serologic and molecular typing, enter Bw4 and Bw6 for all B locus antigens. Do not enter Bw4 or Bw6 for A locus antigens that bear those epitopes”</i> [□]	NMDP registry Bw4/6 epitope table ⁷	2_Bw4_6
Assign WHO antigen when designated.	<i>“For molecular typing, if an allele is identified, and the WHO has designated a serologic type for an allele, enter that type”</i> [□]	WMDA file (rel_dna_ser.txt from IMGT/HLA database) ⁸	3_WHO_overrule

For ambiguous HLA typing, assign probable antigens based on population allele frequency.	<i>“For a group of alleles, the serologic equivalent (according to the HLA Dictionary) of the most common allele(s) should be entered into the UNOS computer”</i> □	NMDP allele ¹⁴ frequencies	4_ambig
Assign expert-assigned antigen if listed in HLA dictionary 2008	<i>“For specific alleles, if there is no WHO designation, but there are data available in the HLA dictionary or other valid reference, the likely serologic type from the available data should be used”</i>	HLA dictionary ¹¹	5b_hla_dict
Assign possible antigens if listed WMDA file		WMDA file	5ap_wmda_file
Assign assumed antigen if listed in WMDA file		WMDA file	5aa_wmda_file
Assign expert-assigned antigen if listed in WMDA file		WMDA file	5ea_wmda_file
Assign broad antigens if multiple split antigens are listed but no probability information based on serologic testing available	<i>“For specific alleles, if there is no serologic WHO designation for an allele, and there are data in the HLA dictionary that indicate the type is within a broad antigen grouping but no clear split is</i>	WMDA file	6_broad_ag

	<i>identified, the allele should usually be listed as the broad antigen”</i> <input type="checkbox"/>		
Assign first-field 2-digit allele family as antigen when no information on serology is available in HLA dictionary or WMDA file.	<i>“For specific alleles, if there is no WHO designation, and the serologic data are absent or inconclusive (but consistent with the 2-digit type), then the 2-digit allele designation should be used (Ex. A*0219, B*0813). For alleles whose 2-digit type is a broad antigen (Ex. B*15), conversion to the 2-digit type should be carefully considered due to the consequences”</i> <input type="checkbox"/>	WMDA file	7_allele_2_digit
Assign blank antigen for null alleles	<i>“Confirmed null alleles should be entered as a blank”</i> <input type="checkbox"/>	WMDA file	8a_null_allele
Assign antigens for low or questionable expression (L/Q) as they are not confirmed null alleles	Not explicitly addressed by OPTN.	WMDA file	8b_lq_allele
Do not assign DR53 antigen using DRB4 allele has been typed at high		(N/A) 2003 OPTN policy document	9_DR3-4-5_agr

resolution.	<i>"When low resolution molecular typings on donors give the type DRB1*07, DQB1*03 (DQ9), DRB4 positive, DR53 should not be entered as present based only on low resolution molecular typing for DRB4"</i>		
Assign antigen equivalents provided by OPTN for some alleles	<i>Table 1. Suggested UNOS Serology Equivalents For Molecular Types</i>	2003 OPTN policy document ⁶	10_UNOS_table
OPTN document with 4 digit equivalency for alleles	<i>Tables in Section 4:10. Suggested allelic UNOS Serology Equivalents for alleles that may not have WHO approved serologic specificity</i>	OPTN policies ⁵	11_OPTN_4_digit
Assign antigen equivalents provided by OPTN for additional alleles	See table in UNOS 2017 updates.	2017 UNOS Histocompatibility Committee Updates ¹⁰	12_UNOS_update

*Rule notation is used in Figure 1 as well as supplementary Table 1 for easy review

Table 2: Illustration of Disparate Antigen Assignments Among Data Sources. The UNOS Antigen Equivalent column gives the final call given by the conversion tool. Disparities are resolved by the rule precedence in the flowchart in Figure 1. Shaded column indicates which data source was used to make the final call for few of the IMGT/HLA alleles.

IMGT/HLA Allele	UNOS Antigen Equivalent	WHO Antigen (3_WHO_overrule)	HLA Dictionary (5b_hla_dict) or Possible or Assumed or Expert Assigned Antigen from WMDA file (5ap/aa/ea_wmda_file)	UNOS 2003 Antigen Equivalency Table (10_UNOS_table)	UNOS 2017 Antigen Equivalency Table (12_UNOS_update)	UNOS 4 digit Allele Equivalencies (11_OPTN_4_digit)	2-digit Allele (7_allele_2_digit)
A*66:01:03	A*6601	A66	A26/34(WMDA)	NA	NA	A*6601	A66
A*66:02	A*6602	A66	A34(WMDA)	NA	NA	A*6602	A66
B*13:04	B*1304	NA	B15/21 (Dictionary), B49/B15 (WMDA)	NA	NA	B*1304	B13
B*15:19	B76	B76	B62 (WMDA)	B76	NA		B15
B*15:20	B62	B62	NA	B62	NA	NA	B15
B*15:21	B75	B75	NA	B75 B51	NA	NA	B15
B*15:23	B51	NA	B70/B5 (Dictionary), B15/5 (WMDA)		NA	NA	B15
B*15:24:01	B62	B62	B77(WMDA)	B62	NA	NA	B15
B*15:36	B13	NA	B15/70(WMDA)	B13	NA	NA	B15
B*15:80	B70	B70	B15(WMDA)	NA	NA	NA	B15
B*15:87	B15	NA	B15 (Dictionary), B15/70	NA	NA	NA	B15

			(WMDA)				
B*58:08:01	B17	B17	B5 (WMDA)	NA	NA	NA	B58
C*17:02	C17	C?	NA	NA	NA	NA	C17
C*17:03:01:01	C07	C?	C7 (Dictionary, WMDA)	NA	C07	NA	C17
DRB1*03:09	DR3	NA	DR3 (Dictionary, WMDA)	NA	NA	NA	DR3
DRB1*03:10	DR17	DR17	NA	NA	NA	NA	DR3
DRB1*04:22	DR4	DR4	DR3 (WMDA)	NA	NA	NA	DR4
DRB1*08:31	DR11		DR8 (WMDA)	NA	NA	NA	DR8
DRB1*13:54	DR14	DR14	DR13 (WMDA)	NA	NA	NA	DR13
DRB1*14:15	DR8		DR14 (WMDA)	NA	NA	NA	DR14
DRB1*14:53	DR13	DR13	DR14 (WMDA)	NA	NA	NA	DR14

Table 3: Antigen Mapping for Unambiguous and Ambiguous HLA Typing by The Conversion Tool “ALLAN”

HLA typing data input		Input example	Population selection	Antigen assignment by tool (antigen probabilities mentioned in parenthesis)
Unambiguous typing	Allele	B*13:01:01:01	Not required	B13, Bw4
	Allele Lists	A*02:301N B*42:08 C*03:04:01:08, DQB1*03:241 DRB5*01:07	Not required	A0, B42, C10, DQ3, DR51, Bw6
Ambiguous typing	Genotype List String	A*02:01+A*26:01g^C*05:01g+C*01:0 2g ^B*15:01/B*15:02/B*15:03/B*15:04+ B*39:01/B*39:02/B*39:04/B*39:05	African	A2 + A26 (1.0), C05 + C01 (1.0), B72 + B3901 (0.77), Bw6
			Asian/Pacific Islander	A2 + A26 (1.0), C05 + C01 (1.0), B75 + B3901 (0.52), Bw6
			Caucasian	A2 + A26 (1.0), C05 + C01 (1.0), B62 + B3901 (0.96), Bw6

			Hispanic	A2 + A26 (1.0), C05 + C01 (1.0), B62 + B39 (0.47), Bw6
			African American	A1 + A2 (0.50), C04 + C05 (0.99), B8 + B13 (1.0), DR4 + DR7 (1.0), DQ7 + DQ3 (0.82), Bw4, Bw6
NMDP	A*01:AABJE A*02:HBMC B*08:NMTJ		Asian/Pacific	A1 + A2 (0.50), C04 + C05 (0.90), B8 + B13 (1.0),
Multiple	B*13:GR C*04:CYMD C*05:YDYE		Islander	DR4 + DR7 (1.0), DQ7 + DQ3 (0.82), Bw4, Bw6
Allele Codes	DRB1*04:AMR DRB1*07:GC			A1 + A2 (0.49), C04 + C05 (0.99), B8 + B13 (1.0), DR4 + DR7 (1.0), DQ7 + DQ3 (0.85), Bw4, Bw6
	DQB1*03:AG DQB1*03:AFYYJ		Caucasian	

			Hispanic	A1 + A2 (0.80), C04 + C05 (0.99) B8 + B13 (1.0), DR4 + DR7 (1.0), DQ7 + DQ3 (0.82), Bw4, Bw6
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Table 4: “ALLAN” Web Services for UNOS Antigen Assignments and Reverse Mapping

Web Service URL	HLA Typing format	POST request example (using Unix HTTPie or curl command line tools)
http://transplanttoolbox.org/single_allele/	Single allele	<ul style="list-style-type: none"> • http -f POST http://transplanttoolbox.org/single_allele/ allele=B*07:02 • curl -X POST http://transplanttoolbox.org/single_allele/ -d '{"allele": "B*07:02"}' -H 'Content-Type: application/json'
http://transplanttoolbox.org/array/	Alleles List	<ul style="list-style-type: none"> • http -f POST http://transplanttoolbox.org/array/ allele_list="B*07:02 A*02:01 DRB1*03:01" • curl -X POST http://transplanttoolbox.org/array/ -d '{"allele_list": "B*07:02 A*02:01 DRB1*03:01"}' -H 'Content-Type: application/json'
http://transplanttoolbox.org/gls/	Genotype List Strings	<ul style="list-style-type: none"> • http -f POST http://transplanttoolbox.org/gls/ gl_string="A*02:01g+A*26:01g A*02:55+A*26:07^C*05:01g+C*01:02g^B*15:01/B*15:02/B*15:03/B*15:04+B*27:05g^DRB1*12:01g+DRB1*01:01^DQB1*03:01g+DQ B1*05:01" pop=CAU • curl -X POST http://transplanttoolbox.org/gls/ -d '{"gl_string": "A*02:01g+A26:01g A*02:55+A*26:07^C*05:01g+C*01:02g^B*15:01/B*15:02/B*15:03/B*15:04+B*27:05g^DRB1*12:01g+DRB1*01:01^DQB1*03:01g+DQ B1*05:01", "pop": "CAU"}' -H 'Content-Type: application/json'
http://transplanttoolbox.org/macs/	NMDP Multiple Allele Codes	<ul style="list-style-type: none"> • http -f POST http://transplanttoolbox.org/macs/ allele_codes_list="A*01:ABJE A*02:HBMC B*08:NMTJ B*13:GR C*04:CYMD C*05:YDYE DRB1*04:AMR DRB1*07:GC DQB1*03:AG DQB1*03:AFYYJ"

		<p>pop=CAU</p> <ul style="list-style-type: none">curl -X POST http://transplanttoolbox.org/macrs/ -d '{"allele_codes_list": "A*01:AABJE A*02:HBMC B*08:NMTJ B*13:GR C*04:CYMD C*05:YDYE DRB1*04:AMR DRB1*07:GC DQB1*03:AG DQB1*03:AFYYJ", "pop": "CAU"}' -H 'Content-Type: application/json'
http://transplanttoolbox.org/reverse_mapping	UNOS Antigen Equivalent	<ul style="list-style-type: none">http -f POST http://transplanttoolbox.org/reverse_mapping/ antigen=A1curl -X POST http://transplanttoolbox.org/reverse_mapping/ -d'{"antigen":"A1"}' -H 'Content-Type: application/json'

Table 5: Discrepancies in UNOS Antigen Assignments between the Conversion Tool and Commercial HLA Typing Software Packages

Allele	UNOS Antigen Equivalency From ALLAN	One	Immucor
		Lambda	LifeCodes
A*02:10 ^a	A2	A210	A210
A*66:01:01:01	A*6601	A66	A66
A*66:01:01:02	A*6601	A66	A66
A*66:01:02	A*6601	A66	A66
A*66:01:03	A*6601	A66	A66
A*66:02	A*6602	A66	A66
B*07:03 ^a	B7	B703	B703
B*15:29	B70	B15	B15
B*15:46	B50	B72	B72
B*15:58	B62	B62	B15
B*27:08	B2708	B2708	B27
B*35:76	B35	B35/22	B35/22
B*39:01:01:02L	B3901	B39"Low"	Low B3901
B*47:02	B60	B47	B47
B*51:03	B51	B5103	B5103
B*82:01	B*8201	B82	B82

^aNew antigen equivalency provided by 2017 UNOS updates

Table 6: Similarity between UNOS Antigen Frequencies and NMDP High Resolution Allele

Frequencies after Conversion to Antigen Equivalents. Normalized similarity index “ I_F ” values

compared NMDP-derived antigen frequencies to the UNOS CPRA reference panel for A, B, C, DR, DQ loci for four broad US race/ethnic categories: Caucasian, African American, Hispanic and Asian/Pacific Islander.

Locus	Caucasian	African American	Hispanic	Asian/Pacific Islander
A	0.97	0.95	0.94	0.87
B	0.93	0.91	0.88	0.85
C	0.90	0.86	0.87	0.86
DR	0.96	0.96	0.95	0.88
DQ	0.89	0.90	0.90	0.89

Figure Legends:

Figure 1: Flow diagram depicting defined rule precedence for UNOS histocompatibility committee guidelines. Highest precedence for antigen assignment to IMGT/HLA alleles was given to UNOS resources, followed by WHO antigens, the HLA dictionary, and the WMDA antigen file. If an antigen could not be assigned to an allele from these antigen equivalency reference sources, then the first-field of the IMGT/HLA allele name was assigned as the antigen.

Supplemental digital content captions:

Supplementary Table 1: Reference table for HLA antigen mapping as per UNOS guidelines for 16,886 IMGT/HLA alleles encompassing A, C, B, DRB1, DRB3/4/5, DQA1 and DQB1 loci.

Supplementary Table 2: Reference table for the list of IMGT/HLA alleles that correspond to each unacceptable UNOS antigen. The assignments utilize the unacceptable antigen equivalency tables specified in the OPTN Policies Section 4.10 - Reference Tables of HLA Antigen Values and Split Equivalences, Tables 4-5 through 4-11.

Supplementary Table 3: NMDP antigen-level HLA frequencies for the A, B, C, DR, and DQ loci for 4 US broad ethnic groups: Caucasian, African American, Hispanic and Asian/Pacific Islander. Frequency data was converted from previously published high resolution allele frequencies.

