1 **Genome sequence and description of *Blautia brookingsii* str SG772 nov., a new species of**

2 **anaerobic bacterium isolated from healthy human gut.**

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21 **Abstract**

22 Strain SG-772 is a Gram positive, strictly anoxic bacterium isolated from the feces of a healthy

23 human fecal donor. Based on 16S rRNA gene sequence, the strain showed maximum similarity

24 (94.39%) with *Blautia stercoris* GAM6-1 in EZ-Taxon server and thus assigned the genus

25 *Blautia*. The scanning electron micrograph of the bacterium revealed the characteristic

26 coccobacillus shape as well as the complete absence of flagellum, suggesting its non-motile

27 phenotype. This strain was found to utilize 27 substrates based on Biolog AN plates assay, with

28 maximum preference for D-mannitol. Additionally, the strain was found to be resistant to

29 tetracycline and streptomycin. Genome sequencing and analysis revealed an overall genome size

30 of 3.49 Mbp and GC content of 43.97%. Based on RAST annotation server, the closest neighbor

31 was *Blautia hansenii* DSM20583. Average Nucleotide Identity (ANI) of these strains were

32 81.69%, suggesting a high level of genomic variation. The comparative genome analysis of

33 strain SG772 with *B. hansenii* DSM20583 revealed a total of 411 orthologous genes coding for

34 basic metabolic functions. Furthermore, the genomes were functionally distinct based on COG

35 categories. Thus, based on all these differences, we propose a novel species of genus *Blautia*

36 named as *Blautia brookingsii* SG772.

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41 **Keywords:** *Blautia, brookingsii* SG772, Human gut, Genome, Anaerobe

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Introduction

44 The members of family *Lachnospiraceae* are commensal bacteria inhabiting the human

45 gastrointestinal tract (1). These group of bacteria degrade complex polysaccharides to produce

46 short chain fatty acids. Acetate, butyrate and propionate produced as the result of digestion of

47 polysaccharides can be used as an energy source for the host (2). *Lachnospiraceae* in human gut

48 microbiota plays important role in maintaining health and disease balance (3-7). Recently, a new

49 genus *Blautia* has been formed under *Lachnospiraceae* family by reclassifying few members of

50 genus *Clostridium* and *Ruminococcus* (8). *Blautia* has been reported to provide colonization

51 resistance against *Enterococcus faecium* (9) and *Clostridium difficile* (5).

52 The members of genus *Blautia* are gram- positive, non-motile, coccoid shaped; obligate

53 anaerobes which can produce a wide variety of compounds as the by-product of fermentation.

54 Most of the members of this genus were either isolated from the human or mammalian fecal

55 samples (8, 10). During the culturomics of human feces, a novel *Blautia*-like strain was isolated

56 from Brain Heart Infusion (BHI) based medium and named as strain SG772. Based on 16S

57 rRNA gene sequence phylogeny, it was taxonomically clustered with the genus *Blautia*.

58 Additionally, using taxo-genomics approach, the bacterium SG772 showed distinct

59 morphological, physiological and genomic attribute as compared to its closest neighbor. Based

60 on all these differences, we propose a novel species of genus *Blautia* named as *Blautia*

61 *brookingsii* SG772.

62 **Organism information**

63 **Growth conditions and biochemical properties**

64   During culturomics of healthy human fecal sample, strain SG772 was isolated from a healthy

65   human fecal sample using Brain Heart Infusion (BHI) agar medium anaerobically at 37^0C.

66   Morphologically, colonies were round, whitish, convex and smooth after 48 hours incubation on

67   BHI agar. Further, cellular morphology of strain SG772 was examined using a scanning electron

68   microscope. Microscopic study revealed the complete absence of flagellum, indicating that strain

69   SG772 is non-motile. The size of the bacterium varied between 0.5-0.8 x 1.8-2.5 µm (Figure 1).

70   The Gram-stain test was performed using a Gram staining kit (HiMedia) and the strain was

71   found to be Gram-positive. Optimum temperature and pH for growth of the bacteria was found to

72   be 37^0C and 6.8 respectively.

73   As the strain SG772 exhibited no significant scores on Matrix-assisted desorption-ionization

74   time-of-flight (MALDI-ToF) (Bruker Daltonics, Germany) (11), the genes encoding for 16S

75   rRNA gene was amplified, sequenced using Applied Biosystems 3500xL, Genetic analyser

76   (Applied Biosystems, MA, USA) at Department of Veterinary and Biomedical Sciences, South

77   Dakota State University. The sequence was assembled using Sequencing Analysis version 5.1.1

78   Genious 10.2.3 (NJ, US). A continuous stretch of 1440 bp of the 16S rRNA gene of strain

79   SG772 was obtained and this sequence was subjected to similarity search against EzTaxon- e-

80   server (12). Analysis of 16S rRNA gene revealed highest sequence similarity (94.39%) to

81   *Blautia stercoris* GAM6-1. Thereafter, the phylogenetic tree was constructed based on 16S

82   rRNA gene sequence, together with top 30 taxonomically characterized strains.   The

83   evolutionary distance matrix was calculated using the distance model of Jukes & Cantor (1969)

84   and an evolutionary tree was reconstructed using the neighbor joining method. The sequence of

85   *Atobobium minutum* NCFB2751 was used as an outgroup (Figure 2). The resultant tree topology

86   was evaluated by bootstrap analysis based on 1000 replicates (Felesenstein J, 1985) in Mega

87 version 5.2.2. Strain SG772 falls in the clade containing members belonging to the genus

88 *Blautia*. (Figure. 2)

89 The growth pattern for the strain SG772 was determined using growth curve assay (Figure 3A).

90 The biphasic growth pattern indicate its ability to use simple sugars in media initially with

91 utilization of complex carbohydrates when simple sugars are depleted. Additionally, in order to

92 elucidate the pattern of substrate oxidation, Biolog assay was performed using Biolog AN

93 MicroPlate^{TM} (Biolog Catalog # 70007) anaerobically. For this, 100 µl of overnight grown

94 culture was plated onto 150 mm BHI agar plate and incubated for 48 hours anaerobically at 37^0

95 C. Grown colonies were picked by a sterile cotton swab and inoculated to AN inoculating fluid

96 (Biolog Catalog # 72007) until OD_{650} reached 0.3. From this suspension, 100µl was pipetted into

97 each well of 96 well biolog AN microplate in triplicate and incubated at 37^0C anaerobically.

98 OD_{650} readings were taken at 0 hour and 24 hours post inoculation and results were analyzed and

99 compared against *Blautia stercoris* GAM6-1 (Table 1). The strain was found to utilize 27

100 different substrates with maximum preference for D-mannitol (Figure 3B).

101 Antimicrobial susceptibility of the strain was tested using agar diffusion method in BHI agar

102 medium. One hundred microliters of overnight grown culture of the strain was plated in BHI

103 agar medium and antibiotic discs were placed. Zone of inhibition by antibiotics was measured

104 after 24 hours of anaerobic incubation to determine the antibiotic sensitivity of the strain. The

105 strain was found to be resistant to tetracycline and streptomycin. In contrast, the strain was

106 susceptible to chloramphenicol, ampicillin, erythromycin and novobiocin.

107 **Genome sequencing information**

108 **DNA isolation**

109   Strain SG772 was sub-cultured on BHI agar and incubated at 37^0C anaerobically for 48 hours.

110   Further, it was cultured in 5 ml of BHI broth for 24 hours. Isolation of DNA was performed

111   following E.Z.N.A DNA isolation kit (Omega, Biotek) protocol. Initially, 500µl of bacterial

112   broth was pelleted by centrifuging at 10,000×g for 1 min. The pellet obtained was lysed in tris

113   EDTA buffer using lysozyme for 1 hour and protein was digested using proteinase K overnight

114   at 55^0C. Subsequently, the mixture was pelleted by centrifuging at 8,000×g for 1 min and

115   supernatant was spun in spin column followed by two washes of wash buffer. Finally, DNA was

116   eluted using 50 µl of nuclease free water and kept at -20^0C until use.

117   **Genome sequencing, assembly and annotation**

118   The genome was sequenced using llumina MiSeq (Illumina Inc, CA) usong 2x 250 paired end

119   chemistry. Further, it was assembled using SPAdes 3.9.0 (13) and validated using Quast (14).

120   Coding sequences were predicted using Glimmer 3.0 (15) and annotated with RAST 2.0 server

121   (16) and the NCBI Prokaryotic Genomes Annotation Pipeline (PGAP) version 4.4. For core

122   genome analysis, GenBank files were retrieved for strains SG772 along with its RAST neighbor

123   *Blautia hansenii* DSM 20583 and were used for core genome analysis using OrthoMCL

124   clustering algorithm (17) at 75% query coverage and sequence similarity (18). For functional

125   annotation, the amino acid sequences were searched against the COG database to predict the

126   abundance of COG gene families and heat map was constructed using Pearson correlation

127   method and hierarchical clustering.

128   **Genomic attributes**

129   Assembly of the *B. brookingsii* SG772 genome produced 55 contigs with the genome size of

130   3.46 Mbp (N50 = 179,176) and 43.97% GC content (Table 2). The largest contig was of 401,827

131   bp while the smallest was of 624 bps. The genome contains 57 tRNA genes, 3343 coding

132   sequences, a phage element and a CRISPR element (Table 2).

Insights from genome sequence

Insight into the genetic repertoire

135   In order to compare the newly sequenced genome of strain SG772 to its closest RAST neighbor

136   *B. hansenii* DSM 20583 (8), we used BLAST Ring Image Generator (BRIG) (19) as a tool to

137   show genome wide sequence similarity using *B. hansenii* DSM 20583 as a reference genome

138   (Figure 4). Based on average nucleotide identity, these two genomes were 81.69% identical (19).

139   Furthermore, this comparison revealed several differences in genome size, GC content and RNA

140   copies (Table 2). For functional annotation, the amino acid sequences were searched against the

141   COG database to predict the abundance of COG gene families and were compared against *B.*

142   *hansenii* DSM 20583 (21). The gene families for energy production and conversion (C), amino

143   acid transport and metabolism (E), carbohydrate transport and metabolism (G), transcription,

144   ribosomal structure and biogenesis (J), transcription (K), replication, recombination and repair

145   (L), cell wall/membrane/envelope biogenesis (M), inorganic ion transport and metabolism (P),

146   defense mechanism (V) and unknown functions (S) were differentially enriched among these two

147   strains (Figure 3C). Core genome analysis revealed a total of 411 core genes coding for basic

148   metabolic functions. A total of 4,880 COG functions were annotated out of which only 70%

149   were assigned to known categories. Additionally, resistance to cadmium, tetracycline,

150   vancomycin, beta lactamase and fluoroquinolones along with dormancy and sporulation genes

151   were relatively abundant in strain SG772. Furthermore, the genome was devoid of genes for

152   motility and chemotaxis suggesting its non-motile phenotype.

Conclusions

This study presents the genome sequence for the strain SG772, with marked physiological and genomic differences from its neighbors *i.e*., *B. stercoris* GAM6-1 and *B. hansenii* DSM20583. Based on the differences, we propose a novel species of genus *Blautia* named as *Blautia brookingsii* SG772.

Taxonomic and nomenclatural proposals

Blautia brookingsii (*brookingsii* referring to the isolation site of the type strain from Brookings, SD, USA). The cells are Gram stain positive, non-motile, coccobacillus, and 1.8-2.5 µm in length and 0.5-0.8 µm in diameter. Colonies were whitish, circular, smooth, and convex after 48 h of incubation in BHI agar. Optimal growth was observed at 37^0C and pH of 6.8. It assimilates D-Mannitol, D-Melezitose, Adonitol, D-Glucosaminic acid, D-Arabitol, D- Sorbitol, D-Trehalose etc. Furthermore, the strain was resistant to tetracycline and streptomycin antibiotics whereas, it was susceptible to chloramphenicol, erythromycin, novobiocin and ampicillin. Genome sequence revealed the genome size of the strain SG772 to be 3.46Mbp with G+C content of 43.97 %. The type strain SG772 isolated from hlthy human fecal sample collected at Brookings, SD, USA and has been deposited in the Microbial Culture Collection at the National Centre for Cell Science, India.

Declarations

Authors declare no competing interests.

Acknowledgements

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197   hansenii comb. nov., Blautia hydrogenotrophica comb. nov., Blautia luti comb. nov., Blautia
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231   **Table 1:** Physiological features of strain SG-772 compared to phylogenetic neighbor *Blautia*

232   *stercoris* GAM6-1.

Characteristics	SG772	*Blautia stercoris* GAM6-1*
Habitat	Human gut	Human gut
Classification	Domain: Bacteria	Domain: Bacteria
	Phylum: Firmicutes	Phylum: Firmicutes
	Class: Clostridia	Class: Clostridia
	Order: Clostridiales	Order: Clostridiales
	Family: Lachnospiraceae	Family: Lachnospiraceae
	Genus: *Blautia*	Genus: *Blautia*
	Species: *brookingsii*	Species: *stercoris*
Optimum temperature	37^0C	37^0C
Shape	Coccobacilli	Cocci
Optimum pH	6.8	6.2
a-Galactosidase activity	-	+
β-Galactosidase activity	-	+
a-Glucosidase	-	+
β –Glucosidase	-	+
N-Acetyl-D-glucosamine	-	+
Turanose	-	-
Erythritol	-	+
Salicin	-	+
L-Glutamine	-	+
a-methyl-D-glucoside	-	NA
Glycyl-L- Proline	-	NA

Glycyl-L- Glutamine	-	-
Dextrin	+	NA
Arbutin	+	NA
D-Saccharic Acid	+	NA
a-D-Glucose	+	NA
L-Valine	+	NA
D-Cellobiose	+	NA
L-Rhamnose	+	NA
Succinic Acid	+	NA
D-Galactose	+	-
D-Mannose	+	NA
Palatinose	+	NA
Uridine-5'- Mono- phosphate	+	-
L-Fucose	+	NA
D-Glucose- 6-Phosphate	+	NA
Lactulose	+	+
D-Trehalose	+	NA
Succinic Acid Mono-Methyl Ester	+	NA
b-Cyclodextrin	+	NA
D-Sorbitol	+	NA
3-Methyl-D- Glucose	+	NA
D-Arabitol	+	NA
D-Glucosaminic Acid	+	NA
Adonitol	+	+
D-Melezitose	+	NA
D-Mannitol	+	NA

233   (*Data adopted from Park et al., 2012)

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235   **Table 2**: General attributes of *Blautia brookingsii* SG772 with *Blautia hansenii* DSM20583

Characteristics	***Blautia brookingsii* SG772**	***Blautia hansenii* DSM20583**
Genome size (bp)	3,459,071	3,065,946
Number of contigs	55	1
Coding sequences	3343	3073
%GC content	43.97	39.04
tRNA	57	65
rRNAs (5S, 16S, 23S)	1, 8, 7	5, 5, 5

| ncRNAs	4	4
Pseudogenes	152	68
CRISPR elements	1	2
Phage	1	0

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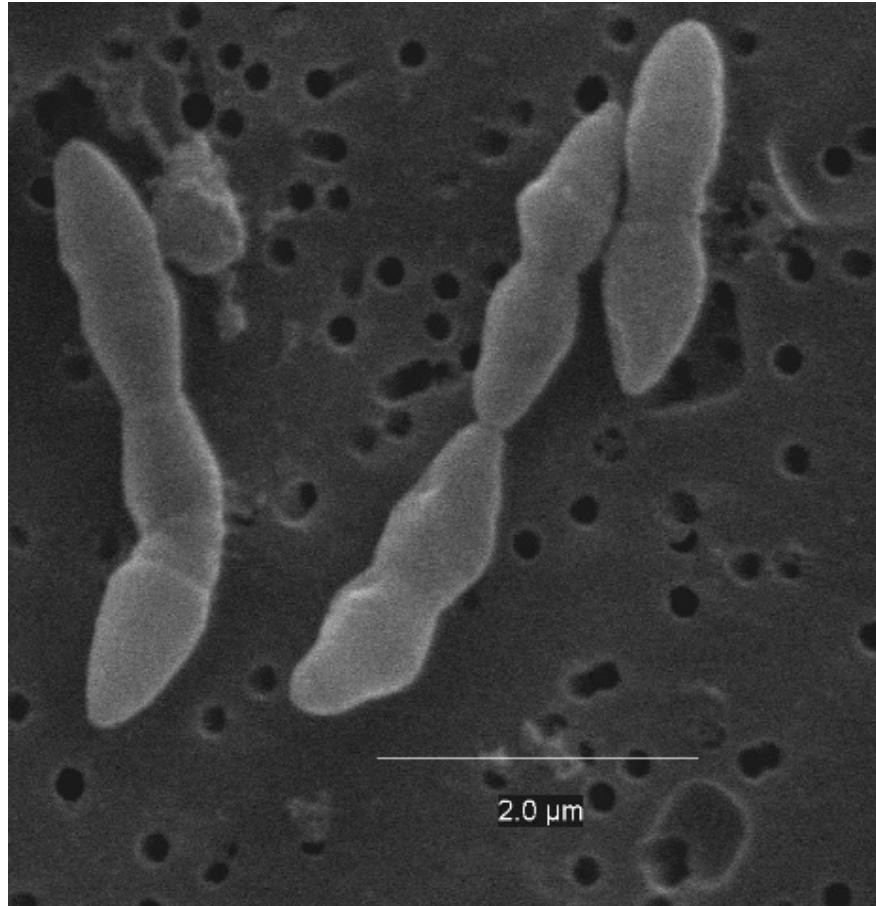
Figure 1: Scanning electron micrograph of strain SG772 grown on BHI agar at 37^0C for 24

hours. Bar, 2.0 µm

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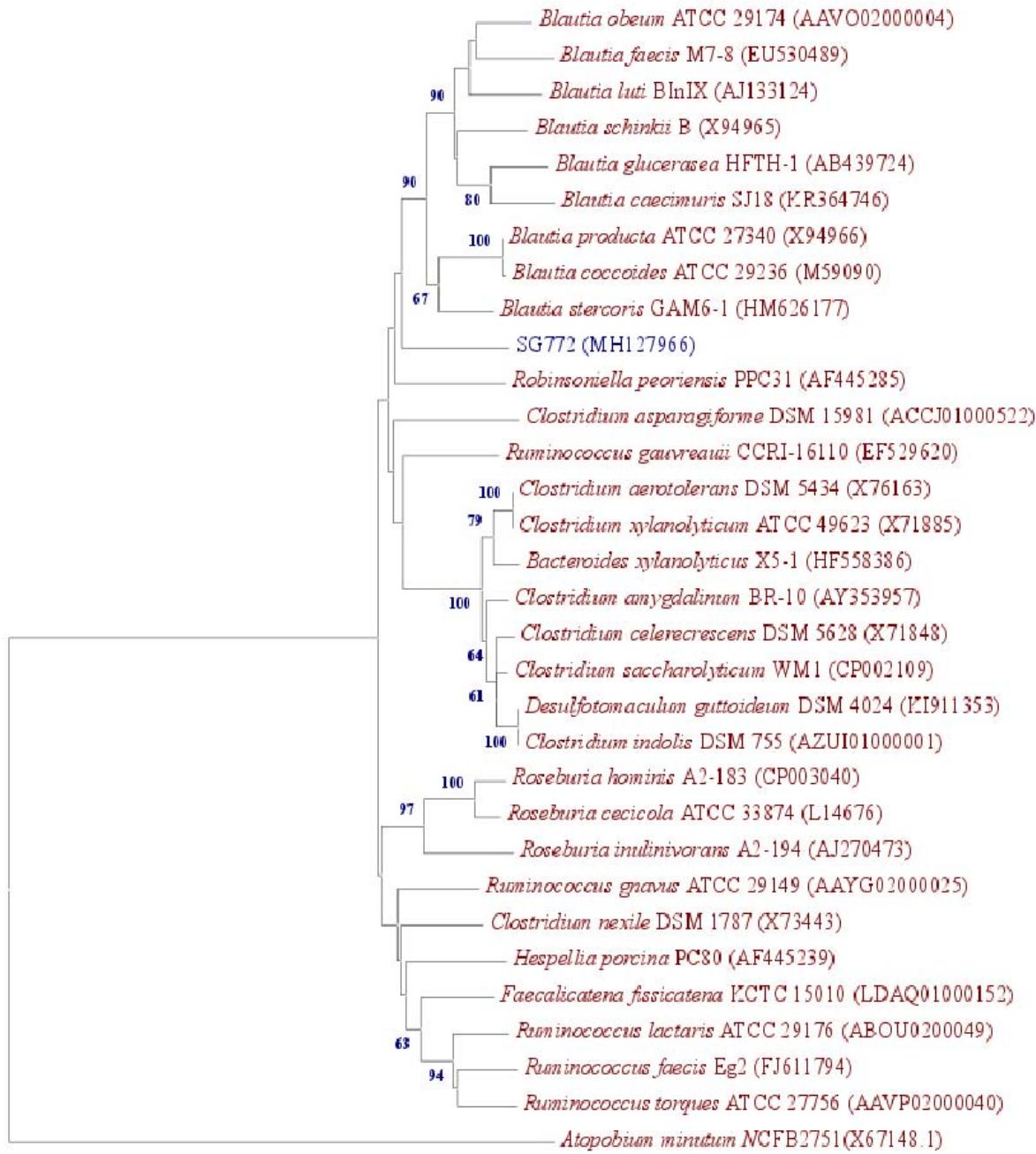
Figure 2: 16s rRNA sequence based phylogenetic tree constructed using neighbor-joining method. The 16S rRNA gene sequence was compared against the EZ-taxon, which showed the

247 highest similarity (94.39%) with *Blautia stercoris* GAM-61. The sequence of *Atobobium*

248 *minutum* NCFB2751 was used as an outgroup.Numbers at nodes indicate bootstrap values

249 expressed as percentages of 1000 replications. Bar, 0.02 accumulated changes per nucleotide.

250 GenBank accession numbers are shown in parentheses.
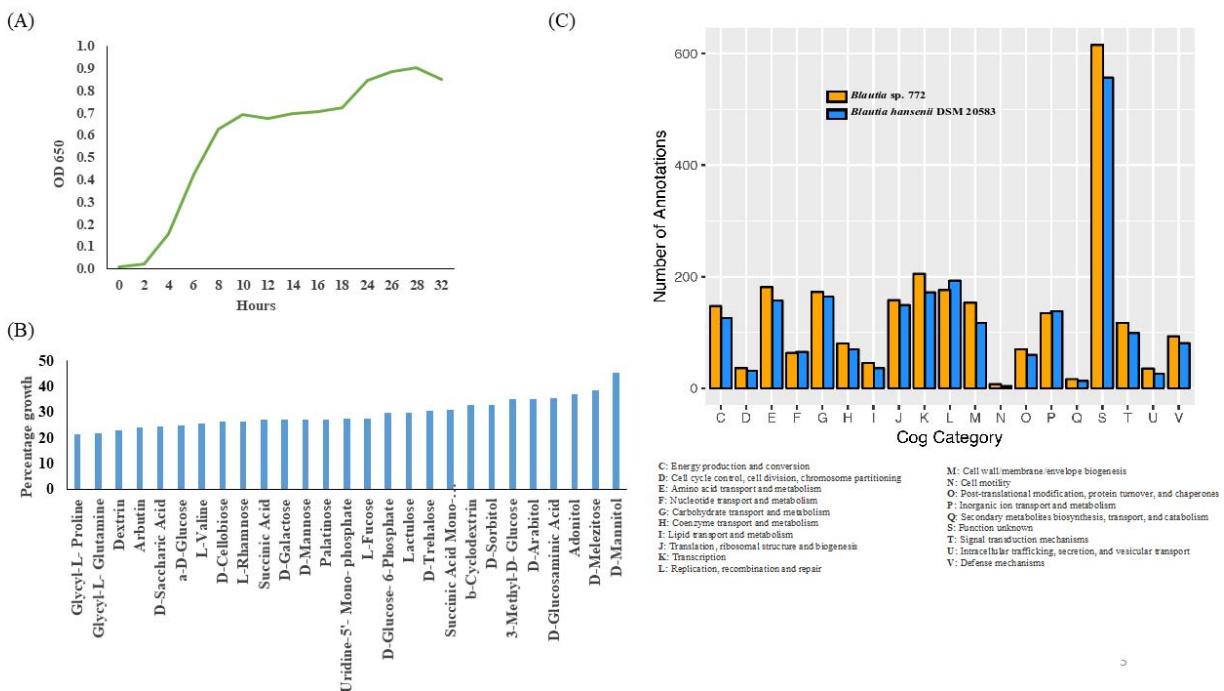
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252 **Figure 3**: **Phenotypic and genotypic profiling of *Blautia sp.* SG772-** (A) Growth curve of the

253 strain *Blautia sp.* SG772, (B) Bar plot depicting different patterns of substrate utilization, (C) Bar

254 graph depicting comparative abundance of COG gene families between the two genomes.

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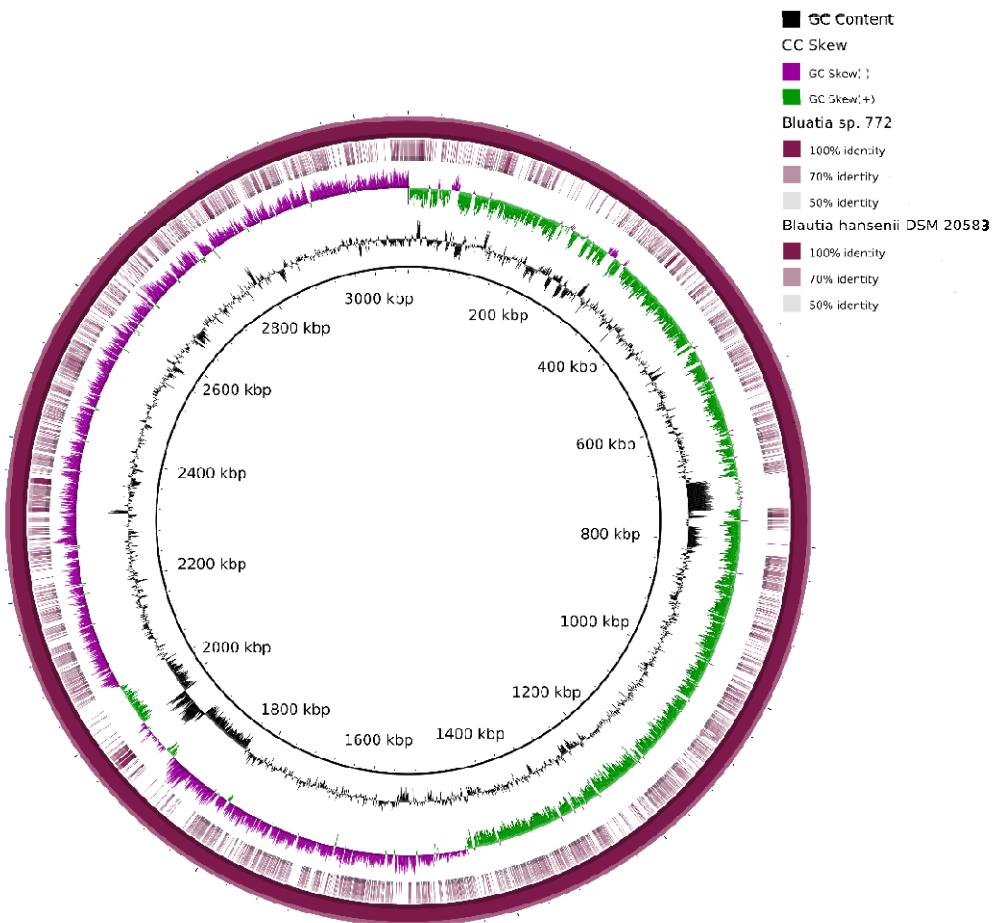
Figure 4: **Comparative genome map of *Blautia sp.* SG772 using *B. hansenii* DSM 20583 as a reference**. From the inside out, circle 1 represents the mean centered G+C content; circle 2 shows GC skew calculated as $(G - C) / (G + C)$; circle 3 represents the genome sequence similarity of *Blautia sp.* SG772 based on color gradient against the reference genome of *B. hansenii* DSM 20583; while circle 4 represents the genome of *B. hansenii* DSM 20583.

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