

Crowdsourced identification of multi-target kinase inhibitors for RET- and TAU-based disease: the Multi-Targeting Drug DREAM Challenge

Short title: Crowdsourced identification of multi-target kinase inhibitors

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Abstract

A continuing challenge in modern medicine is the identification of safer and more efficacious drugs. Precision therapeutics, which have one molecular target, have been long promised to be safer and more effective than traditional therapies. This approach has proven to be challenging for multiple reasons including lack of efficacy, rapidly acquired drug resistance, and narrow patient eligibility criteria. An alternative approach is the development of drugs that address the overall disease network by targeting multiple biological targets ('polypharmacology'). Rational development of these molecules will require improved methods for predicting single chemical structures that target multiple drug targets. To address this need, we developed the Multi-Targeting Drug DREAM Challenge, in which we challenged participants to predict single chemical entities that target pro-targets but avoid anti-targets for two unrelated diseases: RET-based tumors and a common form of inherited Tauopathy. Here, we report the results of this DREAM Challenge and the development of two neural network-based machine learning approaches that were applied to the challenge of rational polypharmacology. Together, these platforms provide a potentially useful first step towards developing lead therapeutic compounds that address disease complexity through rational polypharmacology.

Author Summary

Many modern drugs are developed with the goal of modulating a single cellular pathway or target. However, many drugs are, in fact, 'dirty;' they target multiple cellular pathways or targets. This phenomenon is known as multi-targeting or polypharmacology. While

some strive to develop ‘cleaner’ therapeutics that eliminate secondary targets, recent work has shown that multi-targeting therapeutics have key advantages for a variety of diseases. However, while multi-targeting drugs that affect a precisely-defined profile of targets may be more effective, it is difficult to computationally predict which molecules have desirable target profiles. Here, we report the results of a competitive crowdsourcing project (the Multi-Targeting Drug DREAM Challenge), where we challenged participants to predict chemicals that have desired target profiles for cancer and neurodegenerative disease.

Introduction

Despite important advances in drug development, many diseases remain partly or wholly resistant to drug-based treatments. In recent decades, the field has attempted to address this by developing precision therapeutics with the goal of targeting critical nodes in disease networks. However, this approach has proven to be challenging. Most targeted therapeutics do not progress past preclinical research or clinical trials due to poor efficacy or unacceptable toxicity [1,2]. Additional hurdles are encountered even after clinical approval. For example, initial efficacy against melanoma by the BRAF inhibitors dabrafenib or vemurafenib as single-agent therapeutics is generally followed by emergent tumor resistance [3]. Further, based on biomarkers only a small number of patients with metastatic tumors are eligible for these target-driven precision therapies, and fewer still show sustained response [4].

An alternative to developing drugs with single targets is the development of polypharmacology (multi-targeting) drugs. Many clinically approved drugs bind multiple

targets (<http://ruben.ucsd.edu/dnet>) [5,6], and in some cases the drug is improved by these secondary activities. For example, vandetanib, a drug used in RET-dependent medullary thyroid cancer (MTC), has a broad range of kinase targets that include RET, VEGFR, etc. [7]; pre-clinical studies suggest that some of these secondary targets can contribute to drug efficacy (e.g., [8,9]). Developing optimized polypharmacology-based drugs remains a challenge, however, as there are few roadmaps for predicting structures that combine beneficial targets ('pro-targets') while avoiding liabilities ('anti-targets') in a single molecule.

Current approaches to predict multi-target compounds range from 'physics-based' methods that directly predict binding affinities, to 'omics-based' approaches utilizing large datasets such as those including side-effects or gene expression profiles [10–14]. Notably, recent advances in machine learning architectures have allowed the development of more effective methods relying on both physics-based and omics-based approaches [15,16]. For example, a generative tensorial reinforcement learning (GENTRL) model was used to design a small-molecule with optimal biological activity for the discoidin domain receptor 1 (DDR1) with desirable pharmacokinetic properties in just 21 days [17]. However, several limitations remain: current methods often (i) identify frequent binders or obvious compounds, (ii) fail to discriminate activities among close analogs, or (iii) identify small molecule inhibitors of 'promiscuous kinases' (e.g., DDR1 [18]) that are inherently easier to target. Notably, exploration of the full chemical space—estimated between 10^{24} and 10^{60} structures—still remains a tremendous challenge: known binders of, e.g., the kinome together address a limited set of chemical scaffolds [19].

Rational polypharmacology will require an improved ability to predict chemical structures that specifically target optimized target sets. To identify better computational strategies for this problem, we developed the Multi-Targeting Drug DREAM Challenge [20]. In this challenge, we challenged participants to predict single chemical entities that target pro-targets but avoid anti-targets for two unrelated diseases: RET-based tumors and an inherited form of Tauopathy. Exploring each disease, we used a *Drosophila* ‘dominant genetic modifier screen’ to identify mediators of *Drosophila* RET^{M955T}-mediated transformation and of Tau-mediated dysfunction [21,22]. The resulting functional suppressors and enhancers of RET transformation provide a ‘roadmap’ for the ideal profile of a lead polypharmacological therapeutic.

These data provide therapeutic roadmaps of suppressors (candidate ‘pro-targets’) and enhancers (candidate ‘anti-targets’) for RET- and TAU-based disease. We publicly posted this data, challenging the community to develop computational approaches with improved ability to predict single chemical structures that successfully inhibit pro-targets while avoiding anti-targets. Here, we report the development of two artificial neural network-based machine learning approaches that were applied to the challenge of rational polypharmacology. Together, these platforms provide a potentially useful first step towards developing lead therapeutic compounds that address disease complexity through polypharmacology.

Results

Rational development of drugs that act through polypharmacology will require identifying (i) pro-targets that, as a group, are predicted to be effective in addressing a disease network, and (ii) anti-targets that are predicted to act as liabilities either due to reduced disease efficacy or increased whole body toxicity. Our work in *Drosophila* [21–27] identified whole body pro-targets and anti-targets for RET-associated cancer and Tauopathy. To leverage this data, we established the Multi-targeting Drug DREAM challenge (www.doi.org/10.7303/syn8404040) designed to promote and assess novel rational approaches to polypharmacology (Figure 1).

Figure 1 - Multi-targeting drug DREAM Challenge overview. This challenge sought to identify new algorithms for the identification of multi-targeting drugs that bind disease-relevant targets and avoid disease-specific anti-targets. Participating teams used a broad array of different datasets and computational approaches to develop these methods. They were then asked to use these methods to predict small molecules that would bind to RET-driven cancer targets and anti-targets (Problem 1, P1) as well as Tauopathy targets and anti-targets (Problem 2, P2). A shortlist of these predicted molecules were then experimentally validated (Kd determination) as binders or non-binders of P1 and P2 proteins. Finally, the submitted compounds were scored using a combination of criteria including selectivity, novelty, patentability, and drug-like properties to identify the top performing teams.

Pro-targets and anti-targets

To ensure whole animal relevance, we used data from two *Drosophila* models. To model RET-dependent oncogenic transformation, we used data from previous work that screened *Drosophila* *GMR-RET^{M955T}* and *ptc>RET^{M955T}* cancer models [21,22]. Broad genetic screening identified multiple key ‘pro-targets’ defined as ‘dominant genetic suppressors’ of *GMR-RET^{M955T}*-mediated transformation including *Drosophila* orthologs of BRAF, SRC, and S6K. Candidate ‘anti-targets’—genetic loci that, when reduced, acted as ‘dominant genetic enhancers’ of *RET^{M955T}*-mediated transformation—include MKNK1, TTK, ERK8, PDK1, and PAK3. Given its central role, we also included RET as a pro-target.

To provide a second independent test for compound predictions, we used data recently generated in a screen of a previously-described inherited *Drosophila* Tauopathy model [23–27]. Expressing the human disease isoform TAU^{R406W} exogenously in a transgenic *Drosophila* line led to nervous system defects and animal lethality [23–27]. Genetic screening of most of the *Drosophila* kinome identified several dominant genetic suppressors of TAU^{R406W}. For example, reducing the activity of the *Drosophila* orthologs of AURKA, PAK1, FGFR1, or STK11 suppressed TAU^{R406W}-mediated fly lethality; these loci represent candidate therapeutic targets. Conversely, reducing activity of PAK3, MAP3K7, or PIK3CA enhanced TAU^{R406W}-mediated lethality (*unpublished data*), indicating they should be avoided by candidate therapeutics.

We used these two sets of data as benchmarks for computation-based compound predictions: ideal therapeutic leads would inhibit most or all pro-targets while exhibiting minimum activity against anti-targets.

Participation and Performance

The challenge had 190 registered participants. Of these registrants, 34 submitted predictions, either as a part of one of 5 teams or as individuals. 10 prediction sets were submitted for Problem 1, and 8 prediction sets were submitted for Problem 2. Each prediction was scored using the rubric described in Table 1 (see Methods for additional details).

Criteria	Points Awarded
Problem 1	
Binds RET[M918T]	5
Binds BRAF, SRC, S6K	1, 3 or 9 if binding 1, 2 or 3 of these targets
Avoids MKNK1*	3
Avoids TTK, ERK8, PDK, PAK3	1, 2, 3, or 4 if it avoids 1, 2, 3, or 4 of these targets
Problem 2	
Binds AURKA*	5
Binds PAK1*	5
Binds FGFR1, LKB	1 or 3 if binding 1 or 2 of these targets
Avoids PAK3*	3
Avoids MAP3K7*	3
Avoids PIK3CA	1
Predicted to enter CNS	3
Both Problems	
Novelty	2
Ability to Patent	2
Drug-like molecule	3

Table 1 - Scoring guidelines for Problems 1 and 2. Starred targets indicate that binding or avoiding that target was a requirement to receive target-based points.

Problem 1: Prediction of multi-targeting compounds for RET-driven cancers

Problem 1 asked participants to predict molecules that bound the RET^{M955T}-associated cancer pro-targets RET[M918T], BRAF, SRC, S6K, and did not bind the anti-targets MKNK1, TTK, ERK8, PDK1, and PAK3. Of the 10 prediction files submitted, two of the predictions had the same top-ranked molecule and, therefore, 9 compounds were selected by the challenge organizers for experimental evaluation. Supplemental Figure 1A shows that while most of the submitted compounds had no similar molecule submitted by another team (Tanimoto similarity >0.5), the few similar submitted molecules were mostly predicted by the same method/team.

Binding evaluation revealed that the majority of predicted compounds (7/9) did not bind any of the pro-targets at 10 μ M or any of the anti-targets at 30 μ M (Supplemental Table 1). Two compounds were observed to significantly bind one or more pro-targets: ZINC98209221 and ZINC40900273 (Figure 2, Table 2). ZINC98209221 was observed to significantly bind all 4 pro-targets, while avoiding 2 of the 5 anti-targets. Of the remaining 3 'bound' anti-targets, the K_d was larger than that of the pro-targets, indicating that ZINC98209221 displayed a lower binding affinity for the Problem 1 anti-targets as compared to the pro-targets. The other compound in Table 2, ZINC40900273, only significantly bound one of 4 pro-targets (BRAF) but avoided all 5 anti-targets.

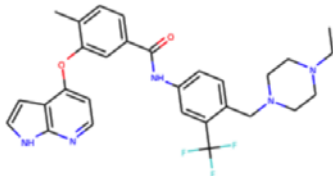
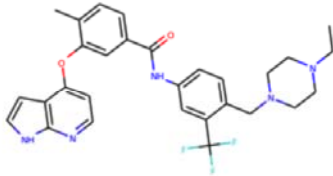
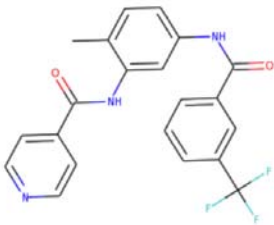
Submitter or Team	Chemical ID	Structure	InChIKey
Zhaoping Xiong	ZINC98209221		SMPGEBOIKULBCT-UHFFFAOYSA-N
DMIS_MTD	ZINC98209221		SMPGEBOIKULBCT-UHFFFAOYSA-N
UM-BISBII	ZINC40900273		KUQAFPIBXZKZON-UHFFFAOYSA-N

Table 2 - Compounds with activity against the Problem 1 targets. “Submitter or Team” indicates the team that identified the compound as a solution to Problem 1. “Chemical ID” is the ZINC identifier for the molecule. “Structure” is a two-dimensional representation of the molecule. “InChIKey” is the hashed InChI for the molecule. Strikingly, two teams identified the same molecule as a solution for Problem 1.

Figure 2 - K_d values of experimentally tested candidates from Problem 1. A heatmap indicating the K_ds of all compounds tested in Problem 1. The best binder of the pro-targets (ZINC98209221) was predicted by two different teams.

Table 3 - Compounds with activity against the Problem 2 targets. “Submitter or Team” indicates the team that identified the compound as a solution to Problem 2. “Chemical ID” is the ZINC identifier for the molecule. “Structure” is a two-dimensional representation of the molecule. “InChIKey” is the hashed InChI for the molecule.

Figure 3 - K_d values of experimentally tested candidates from Problem 2. A heatmap indicating the K_ds of all compounds tested in Problem 1. ZINC3938668 and ZINC538658 bind a subset of the pro-targets, and also both bind 2 of the three anti-targets.

The molecule predicted by Zhaoping Xiong, ZINC3938668, bound 3 of 4 pro-targets at 10 μM: AURKA, PAK1, and STK11. However, this molecule also bound 2 of 3 anti-targets at 10 μM: PAK3 and MAP3K7. Similarly, the predicted molecule from team DMIS-MTD, ZINC538658, bound 3 of 4 pro-targets at 10 μM: AURKA, FGFR1, and STK11, and 2 of 3 anti-targets at 10 μM: PAK3 and MAP3K7.

Overall performance and methods summaries

To identify the top participants, we used the scoring algorithm described in Table 1. This algorithm considered multiple factors including binding of targets and avoiding anti-targets as well as fulfilling novelty, patentability, and drug-likeness criteria to derive a numerical score. The top teams for both Problem 1 and Problem 2 were identical: Zhaoping Xiong and DMIS_MTD, with 16 points each for Problem 1, and 11 and 9 points for Problem 2, respectively (Table 4). The remaining teams received points for

novelty, patentability, and drug-likeness criteria, but the compounds predicted by these teams failed to meet binding criteria to receive points. Therefore overall top-performers for the challenge were Zhaoping Xiong and DMIS_MTD, with overall scores of 28 and 25, respectively (Table 4).

Submitter or Team	Problem 1	Problem 2	Interactome	Final Points
Zhaoping Xiong	16	11	TRUE	28
DMIS_MTD	16	9	FALSE	25
Lasige-BioISI - Multi-target Drug Designing	7	7	TRUE	15
SuperModels	7	7	FALSE	14
Huiyuan Chen	7	7	FALSE	14
Masahiro Mochizuki	7	7	FALSE	14
David Koes	7	7	FALSE	14
Gregory Koytiger	7	7	FALSE	14
UM-BISBII	8		FALSE	8
Stratified	7		FALSE	7

Table 4 - Challenge scoreboard. “Submitter or Team” indicates which team was scored. The “Problem 1” and “Problem 2” scores indicate each team’s performance in the two problems as scored using the criteria described in Table 1. “Interactome” indicates whether the team participated in a text-mining exercise to facilitate team interaction for bonus points. “Final Points” indicates the final score for each team.

Across the 10 participating teams a variety of approaches were employed to arrive at a set of predicted compounds for Problems 1 and 2 (Table 5 and Supplemental Methods). The majority of these methods utilized multiple steps including (i) combining structure- and ligand-based approaches, (ii) applying one or more machine learning/classification algorithms, (iii) use of other computational approaches such as filtering, ligand docking, or creation and application of confidence scoring metrics to identify compounds for the two problems, or (iv) manual selection of the top candidates. Importantly, some teams used fully automated methods, while others used approaches that required several manual curation steps. While the majority of teams utilized one or more machine learning approaches, the two top-performing teams specifically relied on neural network-based approaches to identify the best compounds for Problem 1 and 2 (Table 2 & 3). We therefore examined these two approaches more closely to identify key similarities and differences.

Team	Computational Approach Used	Manual Selection of Top Candidates	Method Data Description
David Koes	neural network	yes	PDB
Team Stratified	ensemble gradient boosting	yes	pIC50
DMIS-MTD	neural network	yes	Cmap scores
UM-BISBII	dataset filtering		SEA-TC algorithm
Lasige-BioISI - Multi-target Drug Designing	random forest, support vector machine	yes	ChEMBL
Gregory Koytiger	neural network		ChEMBL, SwissProt
SuperModels	random forest, logistic regression	yes	ChEMBL, Klaeger et al, Eidogen, ExCAPE-DB
Zhaoping Xiong	neural network	yes	Merget and Fulle et al, Uniprot
Masahiro Mochizuki	random forest, logistic regression		unspecified kinase inhibition assay dataset
Huiyuan Chen	similarity calculations		Drugbank

Table 5 - Methods summary table. “Computational Approach Used” describes the methods that were mentioned in each team’s writeup (see Supplemental Methods). “Manual Selection of Top Candidates” indicates whether a team described using a manual curation step to produce their final set of predicted compounds. “Method Data Description” highlights the data sources and data types used by each team.

Top performing teams

The two teams that were most effective at predicting binding of pro-targets and avoidance of anti-targets both used artificial neural network approaches. Method 1 (Zhaoping Xiong) used features derived from the protein and small molecules. Method 2 (Team DMIS_MTD) relied on transcriptional responses in combination with chemical features of the drug, with a post processing step that included docking.

Method 1 used a graph convolutional neural network deep learning approach [28,29] to predict candidate bioactive compounds (Figure 4). Ligand-based models are constrained when testing is done with just a few bioactive compounds. To build a more generalizable and useful model, this approach incorporated key features of each target kinase by converting the amino acid sequence into a ‘word embedding’ vector, then trained together with graph convolutional neural fingerprints. In developing this method, different combinations were used of extended connectivity fingerprints (ECFP4) of compounds with known bioactivity data, neural fingerprints, and protein embeddings. Testing on a 10% internal hold-out test set demonstrated that the combination of neural fingerprints and protein embeddings performed the best, with an RMSE of 0.414 (Supplemental Table 3).

295

296 **Figure 4** - A schematic summarizing Zhaoping Xiong's method. The molecule is
 297 embedded with graph neural fingerprints and the target proteins are represented as
 298 sequences and pretrained with a language model (doc2vec) model. These two
 299 embeddings are fused together to classify the binding affinity as inactive, weak, or
 300 potent.

301

302 The Method 1 researcher also made the observation that the majority of $-\log_{10} \text{IC}_{50}$
 303 (pIC₅₀) training data was centered around a pIC₅₀ of 5, making prediction of more
 304 extreme compound-target pIC₅₀ values more challenging. To address this, the method
 305 was developed as a classification problem in which the bioactivity of training data was
 306 classified as "inactive" (pIC₅₀ < 5), "weak" (5 ≤ pIC₅₀ < 6) or "potent" (pIC₅₀ ≥ 6). A
 307 classification model trained on these data was then used to predict compound classes
 308 for the targets/anti-targets posed by the Challenge. Finally, the top predictions were
 309 manually selected based on predicted classes for the query targets and anti-targets.
 310 Overall, this model performed best in the Challenge (Supplemental Figure 3).

311 Team DMIS-MTD based Method 2 on ReSimNet [30], a Siamese neural network
 312 model that receives the structural information of two drugs and predicts their
 313 transcriptional response similarity. ReSimNet was trained on the Connectivity Map
 314 (CMap) dataset [31], which provides CMap scores of around 260,000 drug pairs from
 315 2,400 unique drugs. The CMap score of a pair of drugs is computed based on the
 316 similarity of the observed drug-induced gene expression profiles of the two drugs.

Trained on these CMap scores of observed drug pairs, ReSimNet predicts the CMap scores of any drug pair that includes novel compounds.

Before utilizing ReSimNet to identify novel drug candidates, Team DMIS-MTD first identified several prototype drugs with kinase inhibition profiles that satisfied most of the Challenge's pro-target and anti-target conditions. They used the kinase inhibition profiles from the *KInase Experiment Omnibus* (KIEO) database (<http://kieo.tanlab.org>), which was constructed from experimental data on kinase inhibitors. The data was curated from more than 600 published articles (Figure 5A). ReSimNet, trained on the CMap dataset (Figure 5B), was used to predict the transcriptional response similarity between a prototype drug and a novel compound from ZINC15 [32] (Figure 5C). Finally, the top-ranking ZINC15 compounds were aggregated and filtered by Lipinski's rule, a patent search through PubChem, and a protein-ligand docking algorithm (Figure 5D).

Figure 5 - A schematic summarizing DMIS-MTD's method. The method was based on ReSimNet [30], a Siamese neural network model that predicts their transcriptional response similarity of two compounds. (A) Several prototype drugs whose kinase inhibition profiles satisfy most of the pro-target and anti-target conditions of the challenge problem were identified using the *KInase Experiment Omnibus* (KIEO) database. (B-C) ReSimNet, trained on the CMap dataset, was used to predict the transcriptional response similarity between a prototype drug and a novel compound from ZINC15. (D) Finally, the top-ranking ZINC15 compounds are aggregated and

filtered for compounds that fulfill Lipinski's rules, a patent search through PubChem, and a protein-ligand docking algorithm.

Translation of hits to in vivo model systems

After identifying the best-binders in the competitive phase of the Challenge, we assessed the *in vivo* translatability of the *in silico* predicted binders from Problem 1 that were also found to bind the P1 targets *in vitro* (Table 2, Figure 2A). We obtained and tested the compounds for their ability to rescue viability of transgenic flies that expressed the oncogenic form of Drosophila RET in multiple tissues (*ptc>RET^{M955T}*) as previously described [22,33].

Control transgenic *ptc>RET^{M955T}* (RET2B) flies fail to eclose to adulthood due to expression of the transgene (0% adult viability; Supplemental Figure 4, Supplemental Table 4). Feeding *ptc>RET^{M955T}* flies with food containing AD80 or APS6-45 (50 μ M)—experimental multi-targeting kinase inhibitors that reduce *ptc>RET^{M955T}* lethality [22,33]—significantly increased survival to 27% and 22% mean eclosure, respectively (Supplemental Figure 4, Supplemental Table 4). Feeding with ZINC98209221 or ZINC40900273 (1, 10, or 50 μ M final food concentrations) rescued to 1-1.5% of animals, respectively (Supplemental Figure 4, Supplemental Table 4), a number that failed to rise to statistical significance.

Ensemble modeling

As a proof-of-concept to identify candidates for future study, we devised a “wisdom of the crowds” approach to identify consensus Problem 1 predictions of the

top-performing models by identifying the nexus of chemical space predicted by these methods. Specifically, we obtained a larger list of top predictions from the two top-performing teams using the same method and then determined the pairwise Tanimoto similarity (Supplemental Figure 2A) between these two sets. Using this data, we performed network analysis (Supplemental Figure 2B), which allowed us to identify subclusters of structurally similar predicted molecules within the full set of top predicted molecules. We further profiled these molecules using computationally generated absorption, distribution, metabolism and excretion (ADME) metrics (Supplemental Table 5) to facilitate future prioritization of these molecules for medicinal chemistry and *in vivo* screening.

Discussion

One approach to overcoming the limitations of precision medicine approaches in drug development is to develop new drugs with a goal of multi-targeting drugs that target multiple nodes in a disease network. Developing these drugs is a challenge, as both desired targets as well as toxic targets must be considered in the design process. One possible framework for identifying these drugs involves the use of predictive computational modeling strategies to perform virtual screening and identify compounds with a desirable target profile. Here, we described our efforts to identify better computational approaches for this pharmacologic problem.

To provide a disease context to this challenge, we used a set of previously identified pro-targets and anti-targets to design two challenge problems. Problem 1 asked teams to predict molecules that bound the RET^{M955T}-associated cancer targets RET[M918T], BRAF, SRC, S6K, and avoided binding to MKNK1, TTK, ERK8, PDK1, and PAK3. Problem 2 asked participants to predict compounds that bound the Tauopathy pro-targets, AURKA, PAK1, FGFR1, and STK11 and did not bind PAK3, MAP3K7, and PIK3CA. The teams collectively identified 3 compounds for Problem 1 and 2 compounds for Problem 2 that were observed to have any binding potency to the pro-targets. Furthermore, the same set of teams in both problems were successful, indicating that their methods were consistently best-performing. Both of the top-performing teams used neural networks based learning approaches. However, there were other teams that also employed a neural network based approach but did not successfully identify any binding compounds, suggesting that the specific approach may be critical for success. Other approaches (solely manual curation, random forest,

logistic regression) were unsuccessful in identifying binders, suggesting that these methods may not be well-poised to perform well in this type of problem.

Another key finding was that most of the submitted methods, including the top-performing methods, required a manual curation/selection step to select the top performing compounds (Table 5). The decisions and criteria that are used in manual selection can be difficult to define or reproduce using a decision tree or other algorithm. A clear direction for future studies is to better define the nature and rationale for these manual curation steps and devise strategies to automate these steps. The benefits of such future work would likely be two-fold: it would increase the reproducibility of these methods, and it could yield ensemble methods that are more successful in identifying optimal compounds for a given set of targets and anti-targets.

The predominant limitation of this challenge was a relatively small participant pool, which likely resulted in constrained space of methodology. Remarkably, the top-performing methods predicted the identical compound (ZINC98209221) as the best candidate for solving Problem 1, which is surprising given the size of the ZINC15 compound library. Furthermore, previous DREAM Challenges have consistently demonstrated that a variety of methodologic approaches can perform well in many different applications; therefore, it may be the case that methodologic approaches beyond neural networks may indeed be applicable to the problems defined in this challenge, but that the implementation may need to be altered to be successful.

The other key limitation of this study was the difficulty in translating computationally predicted best binders to activity in an *in vivo* system. We tested the top prediction from Problem 1 in the Drosophila RET^{M955} model system of RET-driven cancer but did not

observe bioactivity. There are multiple possible explanations for this, such as the standard ADME challenges faced in drug development, as well as the differences between the fly and human targets and networks [22,34]. To ameliorate this, improvements could include increasing the scale of experimentally tested compounds to identify a larger number of best binders, performing more stringent computational filtering to eliminate compounds with poorer predicted bioavailability characteristics, and performing standard medicinal chemistry and structure-activity relationship studies to identify analogues with better *in vivo* performance.

In the Results, we described a proof-of-concept approach to use these methods to identify molecular candidates for future experimental validation. By leveraging a “wisdom of the crowds” approach, we identified consensus Problem 1 predictions of the top-performing models by identifying the nexus of chemical space predicted by these methods. We subsequently evaluated these molecules using computationally ADME metrics to facilitate future prioritization of these molecules for medicinal chemistry and *in vivo* screening. With this in mind, employing a tandem approach where that leverages ensembled computational models to identify a general region of chemical space with an appropriate polypharmacological profile, followed by more traditional drug optimization approaches, may be a worthwhile future avenue to accelerate the development of multi-targeting drugs.

Materials and Methods

Challenge infrastructure

Multi-targeting Drug DREAM Challenge participants were required to submit a prediction file following a pre-formatted YAML template that included each team's top 5 compound predictions for each of the two problems (RET^{M955T1}- and TAU^{R406W}-mediated phenotypes) (Figure 1). Each prediction included a compound name, vendor name and ID, a methods description, a rationale explaining why the approach was innovative, how the approach is generalizable, and why top predicted compounds are chemically novel. Participants were restricted to compounds that were available through a vendor listed on ZINC15 or other vendors for no more than \$250 per mg. The full set of predictions and methods descriptions provided by the Multi-targeting Drug DREAM Challenge community is available as Supplementary Methods.

In vitro testing

Compounds were purchased and delivered to a contract research organization for experimental testing of binding to the target and anti-target kinases defined for Problems 1 (Figure 2) and 2 (Figure 3). Binding was evaluated using the DiscoverX KinomeScan profiling service. In brief, this method evaluated the ability of the predicted compounds to compete with a bead-immobilized ligand for the active sites of DNA-tagged kinases from Problem 1 and Problem 2. Kinases bound to the immobilized ligand were exposed to a broad range of concentrations of the prediction compounds (11 doses, 3-fold serial dilutions starting at 30 μ M of compound). At each dose, the amount of drug bound to kinase was measured by eluting the non-bead-bound DNA-

tagged kinase and performing quantification by quantitative polymerase chain reaction (qPCR).

Scoring Algorithm

Submissions were assessed using a point-based approach (Table 1). Points were awarded for meeting multiple criteria, in which “binding” of a pro-target was a positive hit at 10 μ M and “non-binding” of an anti-target was defined as a negative result at 30 μ M (Supplemental Tables 1,2).

For problem 1, each of the compounds submitted were awarded points for binding RET[M918T], BRAF, SRC, and/or S6K. Compounds were also awarded points for avoiding MKNK1, TTK, ERK8, PDK1, and/or PAK3. For problem 2: E, each of the compounds submitted were awarded points for binding AURKA, FGFR1, LKB1, and PAK1 and avoiding the anti-targets PAK3, MAP3K7 (TAK1), and PIK3CA. In addition, points were awarded for predictions for meeting other criteria including: novelty (ECFP6-based Tanimoto coefficient <0.4 against all other ChEMBL compounds known to be active against these targets), ability to patent (<2 hits in SciFinder database), and drug-like features (adheres to $\frac{3}{4}$ of Lipinski’s rule of 5). In addition, for Problem 2, predictions predicted to enter the central nervous system (polar surface area <75) were awarded additional points.

Chemical similarity calculations

Chemical similarity calculations and visualizations were performed in the R programming language using rcdk, fingerprint, pheatmap and visNetwork packages.

SMILES strings for each structure were provided by each team, and then converted to a binary fingerprint using the “standard” method provided by the fingerprint package (a 1024-bit “path based, hashed fingerprint”). These fingerprints were then used to generate pairwise similarity matrices using the `fingerprint::fp.sim.matrix` with the method parameter set to ‘tanimoto’. Full R notebooks describing these analyses can be found at <https://github.com/Sage-Bionetworks-Challenges/Multi-Target-Challenge>.

Ensemble method

Ensemble predictions were performed by generating lists of top predictions for each top-performing team and assessing structural similarity as described above. We then used the SwissADME [35] webapp to calculate the number of Lipinski violations, Ghose violations, Veber violations, Egan violations, Muegge violations, PAINS alerts, Brenk alerts, and Lead-likeness violations (Supplemental Table 5). For each compound and we assigned a score of 1 for each value without a violation or alert, or a score of 0 otherwise. We then summed these values to create a composite score where a larger score indicates fewer applicable violations/alerts to prioritize compounds for in vivo screening.

In vivo validation

Female *UAS-RET^{M955T}* virgins were mated to *ptc-GAL4* males to generate *ptc>RET^{M955T}* progeny. Progeny were grown at 25°C and compounds were tested by feeding larvae at three doses: 10 µM, 5 µM, and 1 µM final food concentration. DMSO alone served as a

negative control and the experimental compounds AD80 and APS6-45 [22,33] served as positive controls. Using survival to adulthood (eclosure), each condition was done in quadruplicate, with 41-110 animals tested for each drug concentration.

Code availability

Zhaoping Xiong's top-performing method is available in a Github repository: <https://github.com/xiongzhp/FusedEmbedding>. Instructions for running the method can be found here:

<https://github.com/xiongzhp/FusedEmbedding/blob/master/REPRODUCING.md>.

Team DMIS-MTD's top-performing method is available at:

<https://github.com/dmis-lab/ReSimNet>

<https://www.synapse.org/#!Synapse:syn20545440>

Instructions for running this method can be found at:

<https://github.com/dmis-lab/ReSimNet/blob/master/README.md>.

A detailed description of the implementation and results of these two methods as well as all other submitted methods can be found in the Supplemental Methods. Post challenge analysis notebooks can be retrieved from:

<https://github.com/Sage-Bionetworks-Challenges/Multi-Target-Challenge>.

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Contributions

RJA, JG, XG, NS, MJ, AT, HAC, RA, AS, ZX, HJ, MZ, AS, and RC wrote and edited the manuscript. JG, XG, NS, KD, MJ, KJ, DP, JL, HJ, MK, ACT, AT, HAC, RA, AS, and RC designed and implemented or competed in the competitive phase of the challenge. RJA, XG, KD, RA, CH, TK, AS, and RC designed and implemented post-challenge experiments and analyses. JG, KD, AS, RC were responsible for procuring funding for this study and supervising the work.

545 **Conflict of interest statement**

546 Avner Schlessinger is co-founder of Alchemy, LLC. RC is a consultant to My Personal
547 Therapeutics/ Vivan.

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References

1. Barton P, Riley RJ. A new paradigm for navigating compound property related drug attrition. *Drug Discovery Today*. 2016. pp. 72–81. doi:10.1016/j.drudis.2015.09.010
2. Empfield JR, Clark MP. *Reducing Drug Attrition*. Springer; 2014.
3. Sullivan RJ, Flaherty KT. Resistance to BRAF-targeted therapy in melanoma. *Eur J Cancer*. 2013;49: 1297–1304.
4. Marquart J, Chen EY, Prasad V. Estimation of the Percentage of US Patients With Cancer Who Benefit From Genome-Driven Oncology. *JAMA Oncol*. 2018;4: 1093–1098.
5. Allaway RJ, La Rosa S, Guinney J, Gosline SJC. Probing the chemical–biological relationship space with the Drug Target Explorer. *Journal of Cheminformatics*. 2018. doi:10.1186/s13321-018-0297-4
6. Shi D, Khan F, Abagyan R. Extended Multitarget Pharmacology of Anticancer Drugs. *J Chem Inf Model*. 2019;59: 3006–3017.
7. Davis MI, Hunt JP, Herrgard S, Ciceri P, Wodicka LM, Pallares G, et al. Comprehensive analysis of kinase inhibitor selectivity. *Nat Biotechnol*. 2011;29: 1046–1051.
8. Vidal M, Wells S, Ryan A, Cagan R. ZD6474 suppresses oncogenic RET isoforms in a Drosophila model for type 2 multiple endocrine neoplasia syndromes and papillary thyroid carcinoma. *Cancer Res*. 2005;65: 3538–3541.
9. Vitagliano D, De Falco V, Tamburrino A, Coluzzi S, Troncone G, Chiappetta G, et al. The tyrosine kinase inhibitor ZD6474 blocks proliferation of RET mutant medullary thyroid carcinoma cells. *Endocr Relat Cancer*. 2011;18: 1–11.
10. Szalai B, Subramanian V, Holland CH, Alföldi R, Puskás LG, Saez-Rodriguez J. Signatures of cell death and proliferation in perturbation transcriptomics data-from confounding factor to effective prediction. *Nucleic Acids Res*. 2019;47: 10010–10026.
11. Cichonska A, Ravikumar B, Allaway RJ, Park S, Wan F, Isayev O, et al. Crowdsourced mapping of unexplored target space of kinase inhibitors. *bioRxiv*. 2020. p. 2019.12.31.891812. doi:10.1101/2019.12.31.891812
12. Thafar M, Raies AB, Albaradei S, Essack M, Bajic VB. Comparison Study of Computational Prediction Tools for Drug-Target Binding Affinities. *Front Chem*. 2019;7: 782.
13. Pabon NA, Xia Y, Estabrooks SK, Ye Z, Herbrand AK, Süß E, et al. Predicting

- 583 protein targets for drug-like compounds using transcriptomics. PLoS Comput Biol.
584 2018;14: e1006651.
- 585 14. Douglass EF, Allaway RJ, Szalai B, Wang W, Tian T, Fernández-Torras A, et al. A
586 Community Challenge for Pancancer Drug Mechanism of Action Inference from
587 Perturbational Profile Data. bioRxiv. 2020. p. 2020.12.21.423514.
588 doi:10.1101/2020.12.21.423514
- 589 15. Lim H, Xie L. Omics Data Integration and Analysis for Systems Pharmacology.
590 Methods Mol Biol. 2019;1939: 199–214.
- 591 16. Schneider P, Walters WP, Plowright AT, Sieroka N, Listgarten J, Goodnow RA Jr,
592 et al. Rethinking drug design in the artificial intelligence era. Nat Rev Drug Discov.
593 2020;19: 353–364.
- 594 17. Zhavoronkov A, Ivanenkov YA, Aliper A, Veselov MS, Aladinskiy VA, Aladinskaya
595 AV, et al. Deep learning enables rapid identification of potent DDR1 kinase
596 inhibitors. Nat Biotechnol. 2019;37: 1038–1040.
- 597 18. Hanson SM, Georgiou G, Thakur MK, Miller WT, Rest JS, Chodera JD, et al.
598 What Makes a Kinase Promiscuous for Inhibitors? Cell Chem Biol. 2019;26: 390–
599 399.e5.
- 600 19. Rahman R, Ung PM-U, Schlessinger A. KinaMetrix: a web resource to investigate
601 kinase conformations and inhibitor space. Nucleic Acids Res. 2019;47: D361–
602 D366.
- 603 20. Schlessinger A, Abagyan R, Carlson HA, Dang KK, Guinney J, Cagan RL. Multi-
604 targeting Drug Community Challenge. Cell Chem Biol. 2017;24: 1434–1435.
- 605 21. Read RD, Goodfellow PJ, Mardis ER, Novak N, Armstrong JR, Cagan RL. A
606 Drosophila model of multiple endocrine neoplasia type 2. Genetics. 2005;171:
607 1057–1081.
- 608 22. Sonoshita M, Scompton AP, Ung PMU, Murray MA, Silber L, Maldonado AY, et al. A
609 whole-animal platform to advance a clinical kinase inhibitor into new disease space.
610 Nat Chem Biol. 2018. doi:10.1038/nchembio.2556
- 611 23. Beharry C, Alaniz ME, Alonso ADC. Expression of Alzheimer-like pathological
612 human tau induces a behavioral motor and olfactory learning deficit in Drosophila
613 melanogaster. J Alzheimers Dis. 2013;37: 539–550.
- 614 24. Blard O, Feuillette S, Bou J, Chaumette B, Frébourg T, Campion D, et al.
615 Cytoskeleton proteins are modulators of mutant tau-induced neurodegeneration in
616 Drosophila. Hum Mol Genet. 2007;16: 555–566.
- 617 25. Grammenoudi S, Anezaki M, Kosmidis S, Skoulakis EMC. Modelling cell and
618 isoform type specificity of tauopathies in Drosophila. SEB Exp Biol Ser. 2008;60:

- 619 39–56.
- 620 26. Pflieger CM, Wang J, Friedman L, Vittorino R, Conley LM, Ho L, et al. Grape-seed
621 polyphenolic extract improves the eye phenotype in a *Drosophila* model of
622 tauopathy. *Int J Alzheimers Dis*. 2010;2010. doi:10.4061/2010/576357
- 623 27. Williams DW, Tyrer M, Shepherd D. Tau and tau reporters disrupt central
624 projections of sensory neurons in *Drosophila*. *J Comp Neurol*. 2000;428: 630–640.
- 625 28. Li X, Li Z, Wu X, Xiong Z, Yang T, Fu Z, et al. Deep Learning Enhancing Kinome-
626 Wide Polypharmacology Profiling: Model Construction and Experiment Validation. *J*
627 *Med Chem*. 2019. doi:10.1021/acs.jmedchem.9b00855
- 628 29. Xiong Z, Wang D, Liu X, Zhong F, Wan X, Li X, et al. Pushing the Boundaries of
629 Molecular Representation for Drug Discovery with the Graph Attention Mechanism.
630 *J Med Chem*. 2019. doi:10.1021/acs.jmedchem.9b00959
- 631 30. Jeon M, Park D, Lee J, Jeon H, Ko M, Kim S, et al. ReSimNet: drug response
632 similarity prediction using Siamese neural networks. *Bioinformatics*. 2019;35:
633 5249–5256.
- 634 31. Subramanian A, Narayan R, Corsello SM, Peck DD, Natoli TE, Lu X, et al. A Next
635 Generation Connectivity Map: L1000 Platform and the First 1,000,000 Profiles. *Cell*.
636 2017;171: 1437–1452.e17.
- 637 32. Sterling T, Irwin JJ. ZINC 15--Ligand Discovery for Everyone. *J Chem Inf Model*.
638 2015;55: 2324–2337.
- 639 33. Dar AC, Das TK, Shokat KM, Cagan RL. Chemical genetic discovery of targets and
640 anti-targets for cancer polypharmacology. *Nature*. 2012;486: 80–84.
- 641 34. Ung PMU, Sonoshita M, Scopton AP, Dar AC, Cagan RL, Schlessinger A.
642 Integrated computational and *Drosophila* cancer model platform captures
643 previously unappreciated chemicals perturbing a kinase network. *PLoS Comput*
644 *Biol*. 2019;15: e1006878.
- 645 35. Daina A, Michielin O, Zoete V. SwissADME: a free web tool to evaluate
646 pharmacokinetics, drug-likeness and medicinal chemistry friendliness of small
647 molecules. *Sci Rep*. 2017;7: 42717.

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Supplemental Figure Legends

Supplemental Figure 1 - Chemical similarity comparison of all submitted Problem 1 and Problem 2 solutions.

Supplemental Figure 2 - (A) Similarity heatmap of top predicted compounds for Problem 1 provided by the top-performing teams. Columns correspond to predictions from Zhaoping Xiong, while rows correspond to predictions from DMIS-MTD. The majority of compounds are relatively dissimilar from one another. (B) The similarity matrix was converted into a network, where nodes are individual predicted compounds and edges encode compound-compound Tanimoto similarities. Edge thickness represents similarity (thicker edges = greater similarity). Edges representing similarity below 0.4 were filtered out, and cluster subnetworks were identified (with each color representing an individual subnetwork).

Supplemental Figure 3 - Recapitulating Zhaoping Xiong's three-class classification model used for screening. (A) The distribution of pIC50; (B) The distribution of classes; (C) The accuracy of the model across train, valid and test sets

Supplemental Figure 4 - RET2B and control *D. melanogaster* were treated with DMSO (negative control), two candidate RET-targeting compounds identified using top performing methods (ZINC4020 and ZINC9820) and two positive control multi-targeting RET inhibitors previously demonstrated to rescue the RET2B model (AD80 [33], APS6-45 [22]). Bars show the mean percent survival and error bars show the standard deviation of four replicates per condition. A Mann-Whitney test was used to assess the significance of changes to percent survival in the varying conditions. The RET2B Adult panel (bottom right) indicates that higher concentrations of AD80 or APS6-45

673 significantly ($p < 0.05$) rescue the RET2B model, while the two candidate ZINC molecules
674 have a minimal and non statistically-significant effect on percent survival to adulthood.

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Supplemental Table Legends

Supplemental Table 1 - Problem 1 Results. “Submitter or Team” indicates which team was scored. “Chemical ID” indicates the ZINC or MolPort identifier for each predicted compound. “InChIKey” indicates the InChIKey for each predicted compound. Columns 4-12 indicate the experimentally-validated binding constant (K_d , nanomolar) for the pro-target or anti-target listed in the column header (>30000 nM indicates it was above the detection limit for the assay). Columns 13-16 indicate the status of the compound with respect to the challenge rules for novelty, drug-like properties, and likelihood to pass the CNS as defined in the challenge scoring rules. The final column indicates the number of points the prediction was given based on the defined scoring criteria.

Supplemental Table 2 - Problem 2 Results. “Submitter or Team” indicates which team was scored. “Chemical ID” indicates the ZINC or MolPort identifier for each predicted compound. “InChIKey” indicates the InChIKey for each predicted compound. Columns 4-10 indicate the experimentally-validated binding constant (K_d , nanomolar) for the pro-target or anti-target listed in the column header (>30000 nM indicates it was above the detection limit for the assay). Columns 11-14 indicate the status of the compound with respect to the challenge rules for novelty, drug-like properties, and likelihood to pass the CNS as defined in the challenge scoring rules. The final column indicates the number of points the prediction was given based on the defined scoring criteria.

Supplemental Table 3 - Zhaoping Xiong’s method predictive performance on regression models. A summary of the performance of various model architectures tested by top performer Zhaoping Xiong using root-mean-square-error calculated using true pIC50 values.

Supplemental Table 4 - Top Problem 1 predictions fail to rescue the RET2B fly

model. The RET2B model was treated with two positive controls expected to rescue this model (AD80 and APS6-45), a vehicle control (DMSO), and two test compounds (ZINC4090, ZINC9820) predicted by the challenge results to rescue the RET2B model. “tx”: the treatment (compound and concentration) used; “genotype”: WT or RET2B flies; stage: class of counted animal - adult or pupae; “mean_percent_surv”: the mean percent survival across 4 replicates; “std_dev_survival”: the standard deviation of the percent survival across 4 replicates.

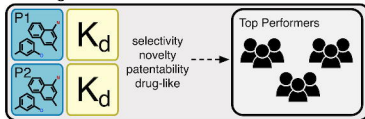
Supplemental Table 5 - *in silico* chemical modeling of Problem 1 predictions from

top performers. Additional hits from top performing teams were characterized using SwissADME to identify compounds with preferred absorption, distribution, metabolism, excretion, and other medicinal chemistry properties.

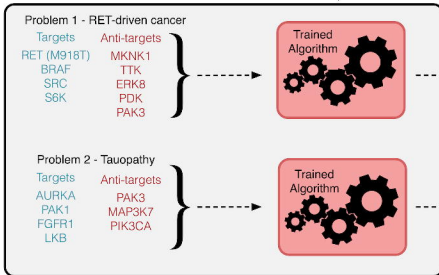
Method Development



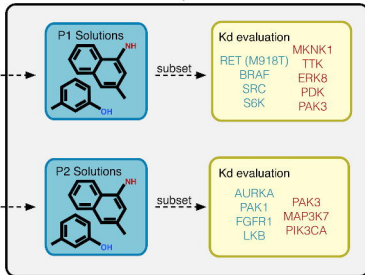
Scoring



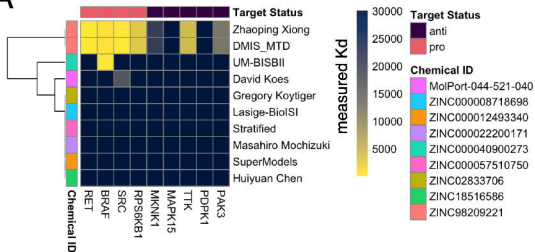
Predictions



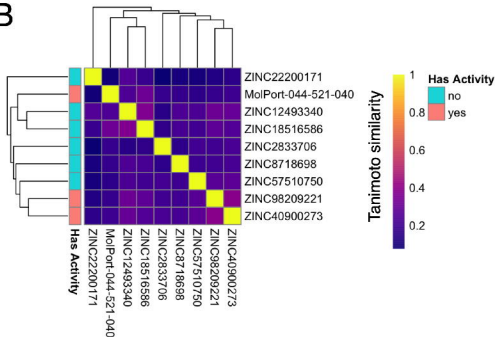
Validation



A



B



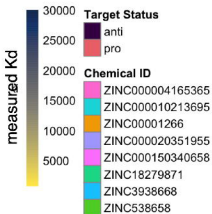
A

Target Status

Gregory Koytiger
David Koes
Lasige-BioISI
Masahiro Mochizuki
SuperModels
Huiyuan Chen
Zhaoping Xiong
DMIS_MTD

Chemical ID

AURKA
PAK1
FGFR1
STK11
PAK3
MAP3K7
PIK3CA



B

Heatmap showing activity levels for various ligands (rows) across different targets (columns). The color scale ranges from dark blue (low activity) to yellow (high activity). The 'Has Activity' column is highlighted in red.

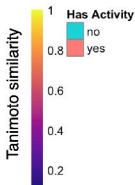
Ligands (rows):

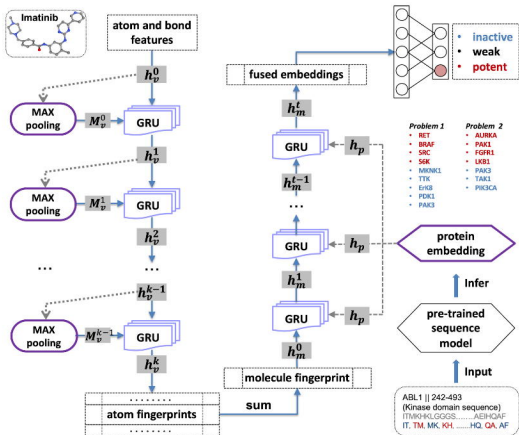
- ZINC18279871
- ZINC20351955
- ZINC3938668
- ZINC538658
- ZINC1266
- ZINC10213695
- ZINC4165365
- ZINC150340658

Targets (columns):

- ZINC150340658
- ZINC4165365
- ZINC10213695
- ZINC1266
- ZINC538658
- ZINC3938668
- ZINC20351955
- ZINC18279871

Has Activity

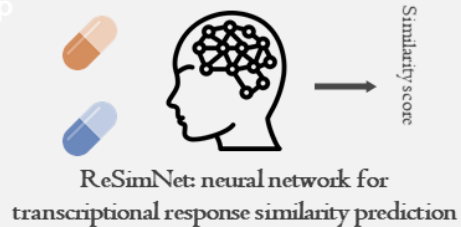




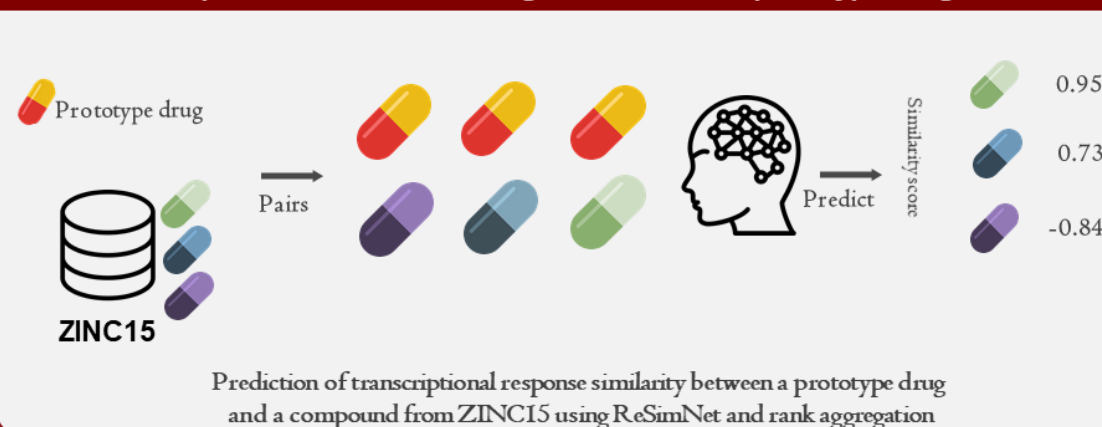
A. Selection of prototype drugs from KIEO



B. Training ReSimNet on the Connectivity Map



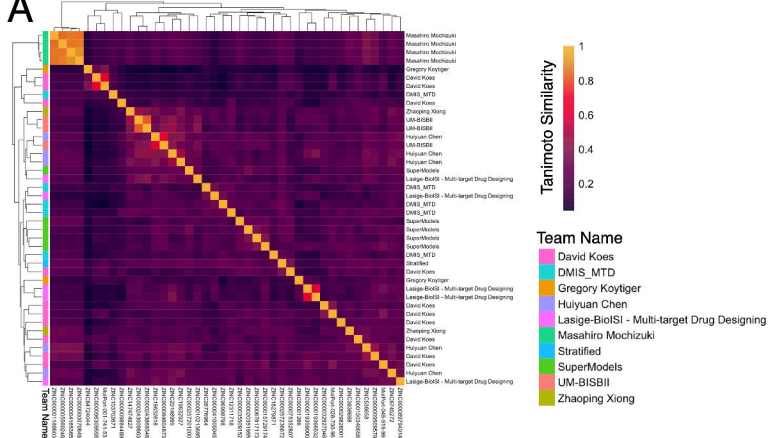
C. Rank compounds from ZINC using ReSimNet and prototype drugs



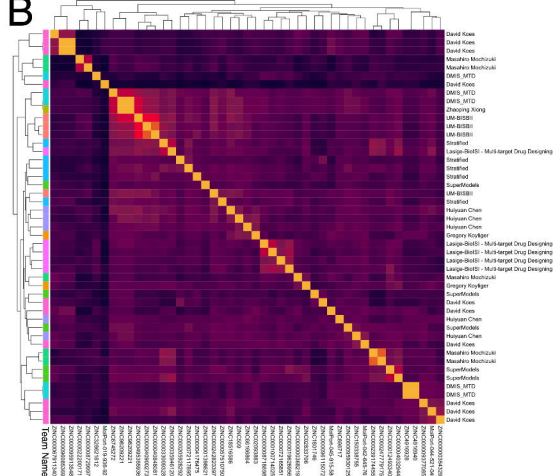
D. Post processing

- ✓ Lipinski's rules
- ✓ Patent search
- ✓ Docking algorithm

A



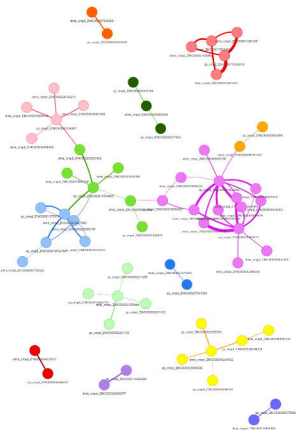
B

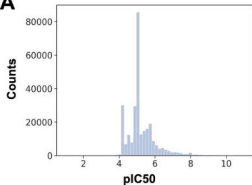
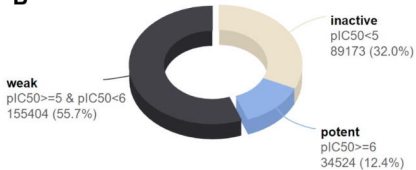
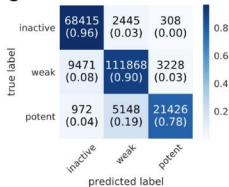
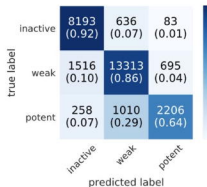
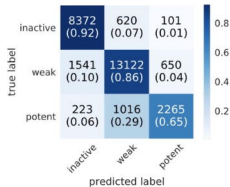


A

Heatmap A displays gene expression data across various samples. The color scale ranges from blue (low expression) to red (high expression). The heatmap is divided into two main clusters by a horizontal line. The top cluster shows higher expression levels, while the bottom cluster shows lower expression levels. The dendrogram on the left indicates the hierarchical clustering of samples.

B



A**B****C****(80%) Train accuracy: 0.903****(10%) Valid accuracy: 0.849****(10%) Test accuracy: 0.851**

