

Determining the stability of genome-wide factors in BMI between ages 40 to 69 years.

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## Abstract

Genome-wide association studies (GWAS) have successfully identified common variants associated with BMI. However, the stability of genetic variation influencing BMI from midlife and beyond is unknown. By analyzing BMI data collected from 165,717 men and 193,073 women from the UKBiobank, we performed BMI GWAS on six independent five-year age intervals between 40 and 73 years. We then applied genomic structural equation modeling (gSEM) to test competing hypotheses regarding the stability of genetic effects for BMI. LSDR genetic correlations between BMI assessed between ages 40 to 73 were all very high and ranged 0.89 to 1.00. Genomic structural equation modeling revealed that genetic variance in BMI at each age interval could not be explained by the accumulation of any age-specific genetic influences or autoregressive processes. Instead, a common set of stable genetic influences appears to underpin variation in BMI from middle to early old age in men and women alike.

## Introduction

Body mass index (BMI) is a commonly assessed trait in population studies and often used to index underweight, normal weight, overweight, or obese individuals. An individual's BMI and how it changes can differentially predict numerous health (e.g., cardiometabolic, diabetes, dementia<sup>1,2</sup>) and mortality outcomes<sup>3</sup>. Despite the known limitations of BMI<sup>4-6</sup> and value added by considering additional adiposity indices such as waist-to-hip ratio and waist circumference for predicting cardio-metabolic diseases<sup>7</sup> and mortality<sup>8</sup>, increased BMI still remains a significant predictor of dementia<sup>2,9,10</sup> and all-cause mortality<sup>11</sup>, including more recently, COVID19-related hospitalization and death<sup>12</sup>. Understanding the genetic and environmental etiology of BMI is therefore a public health priority. Among the many endeavors aimed at identifying the loci underpinning variation in complex traits<sup>13</sup>, genome-wide association scan (GWAS) analyses of adult BMI<sup>14</sup> are some of the most successful. For instance, BMI meta-analytic GWAS results can now account for a significant proportion of BMI heritability<sup>15,16</sup>. A limitation however, of these results is their reliance on aggregated data, which

include different cohorts sampled from varying geographic and economic regions, comprising multiple birth cohorts, and age distributions. Here, we examine the last caveat because it remains to be empirically determined if genome-wide variation associated with adult BMI is age-invariant or age-specific.

#### *BMI heritability & longitudinal genetic correlations*

Despite moderate stability across time <sup>1,17-20</sup>, average adult BMI increases from age 20 to age 65 at which time it levels off until age 80 <sup>1</sup> when it begins to decline. Such changes might be attributed to variable contributions of genetic and environmental risks across the lifespan. Apart from birth cohort differences, Dahl et al. <sup>1</sup>, found that factors such as an obesity genetic risk score, type-2 diabetes mellitus, cardiovascular disease, substance use, and educational attainment were all differentially predictive of both average BMI and changes in BMI before and after age 65. In contrast, many of these risks were no longer predictive after age 65.

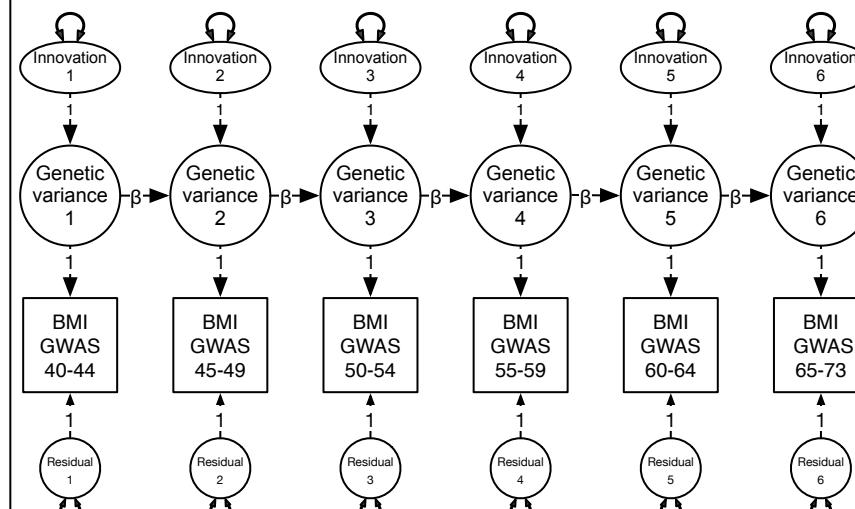
Whereas average lifetime BMI heritability is 0.75 <sup>16</sup> with estimates range from 0.47 - 0.90 and from 0.24 - 0.81 in twin studies and family studies respectively <sup>16,21</sup> - heritability actually increases throughout infancy and adolescence <sup>22</sup> before decreasing during adulthood <sup>16</sup>. Twin studies have shown that genetic influences in BMI are correlated across time from infancy to adolescence <sup>17-19,22</sup> and from early adulthood to midlife <sup>20,23</sup> and sometimes very highly <sup>24</sup>, which indicates continuous expression of the same genetic influences <sup>25</sup>. However, longitudinal genetic correlations for BMI never reach unity. Indeed, there is considerable variability in longitudinal genetic correlations ( $r_g$ =~.55-.95). This is consistent with age-specific genetic influences, which could be obscured in GWAS meta-analyses that model age as a covariate.

In addition to biometrical genetics or longitudinal twin designs, molecular methods such as linkage disequilibrium score regression (LDSR) <sup>26</sup> can leverage GWAS summary statistics and population reference panels to estimate correlations ( $r_G$ ) across unrelated and independent samples. Extending this approach to cohort studies with different ages can address the question of whether or not genetic risks in BMI are correlated across time. Currently, there is a paucity of reports examining BMI  $r_G$  across time using GWAS data. Studies to date have implemented a variety of approaches and produced mixed results. For instance, Trzaskowski et al. <sup>27</sup> used LDSR to report a genetic correlation ( $r_g$ =0.86) between BMI assessed at ages 11 and 65. Winkler et. al. <sup>28</sup> estimated Spearman rank genetic correlations between BMI assessed in populations above and below age 50, which revealed a much smaller correlations ( $r_g$ = 0.12). Notwithstanding the need for greater precision regarding longitudinal genetic correlations, such correlations are descriptive and provide no insight regarding competing theories underlying developmental processes in BMI.

We propose that at least two theoretical mechanisms <sup>29</sup> can explain the observed continuity in genetic correlations. The first is a growth process whereby genetic or environmental factors determine the levels and rates of change in BMI over time. In this model, variances and covariances between longitudinal measures of BMI depend on individual genetic or environmental differences in growth patterns unfolding with age

(“random growth curves”) <sup>30-34</sup>. We are aware of three twin studies that have applied genetically informative growth models to longitudinal BMI data <sup>35-37</sup>. Unfortunately, random growth curves do not determine the extent to which stability or changes in BMI are governed by time-invariant versus age-specific genetic influences. To address this question, the second mechanism predicts that variances and covariances are determined by random, time-specific genetic and environmental effects, which are more or less persistent over time i.e., autoregressive effects<sup>38-40</sup>. Illustrated in Figure 1, this approach predicts a causal hypothesis of inertial effects, whereby genetic factors contributing to BMI at one time causally affect BMI at the next. We have applied this approach to personality <sup>41</sup>, anxiety and depression <sup>42,43</sup>, substance use <sup>44</sup> and brain aging <sup>45</sup>. We are aware of two reports that have tested the fit of autoregression models to BMI data <sup>18,25</sup>. In addition to age-invariant genetic risks, Cornes et al. <sup>18</sup> also found evidence of distinct, age-specific genetic influences on BMI at ages 12, 14 and 16. To our knowledge, autoregressive effects have not been tested in adult BMI, especially across a wide window comprising narrow age intervals in adults. Fortunately, the recent, innovative application of structural equation modelling (SEM) to LDSR genetic correlations <sup>46</sup> based on available GWAS results can now address the aforementioned gaps.

Figure 1. Autoregression model depicting genome-wide variation in BMI at each age interval in terms of time-specific variation or “innovations” & the causal contribution of genome-wide variation from previous age intervals. Model also includes residual genetic variation not otherwise explained by the autoregression.



By applying genomic SEM or “gSEM” to BMI GWAS data from the UK Biobank <sup>47</sup>, our aim was to determine if genetic influences across middle age were best explained by age-dependent versus age-invariant processes. We also tested if alternative, more parsimonious theoretical explanations i.e., common factor models, whereby covariance between genetic influences across time could be captured by a single latent factor <sup>48</sup>, provided

a better fit to the data. Given that standardized estimates of BMI heritability for men and women are statistically equal <sup>16</sup> and that there appear to be no sex differences in terms of the observed adult decline in heritability <sup>49</sup>, we hypothesized that developmental processes governing changes in heritability over time likewise ought to be comparable across sex.

## Methods

All BMI and GWAS data came from the UK Biobank, which is accessible to the scientific community, which is a large and detailed prospective study of over 500,000 subjects aged 40–73 years recruited from 2006 to 2010 of predominantly European ancestry <sup>47</sup>.

### *BMI data*

Described in detail elsewhere <sup>50</sup>, weight was collected from subjects using a Tanita BC418MA body composition analyzer. Standing and sitting height measurements were collected from subjects using a Seca 240cm height measure. Weight and height were amalgamated into mass index (BMI) calculated as weight divided by height squared (kg/m<sup>2</sup>). We divided the BMI data into six age intervals: 40-44; 45-49; 50-54; 55-59; 60-64; and 65-73 years based on age and BMI at baseline assessment. The range was based on available data whereas the number of age tranches was selected to maximize our power to choose between competing longitudinal and multivariate models without minimizing the statistical power of the GWAS analyses at each interval. The number of subjects with complete BMI and GWAS data are shown in Supplementary Table S1.

### *Genotypic data*

Genotype data were filtered according to the Neale Lab pipeline, using filtration parameters and scripts publicly available from the lab's GitHub <sup>51</sup> repository. Samples were filtered to retain only unrelated subjects of British ancestry (n=359,980.) Imputed variants <sup>52</sup> were filtered based on INFO scores > 0.8, MAF > 0.001, and HWE p-value > 1e-10.

### *GWAS analyses*

This is proof-of-principle illustration of the application of structural equation modelling (SEM) to GWAS summary data to test longitudinal hypotheses. While a subset of UKB subjects has repeated BMI measures, the data freeze used here contained insufficient subjects with a minimum of three BMI assessments required to fit autoregression models. To maximize power and subject inclusion, we divided the sample into 5-year age intervals, which yielded 6 age tranches for GWAS and the subsequent gSEM pseudo-longitudinal analyses. Separate GWAS analyses were conducted for each 5-year age interval using BGENIE (version 1.3.) <sup>52</sup>. The first 10 ancestry principal components were included as covariates in all models and sex was also included as a covariate in the non-sex stratified models.

### *Genomic structural equation modelling*

We then applied the gSEM software package <sup>46</sup> in R (version 4.0.3) <sup>53</sup> to estimate the genetic variance-covariance (S) and asymptotic sampling covariance 'weight' (V) matrices based on the 6 BMI GWAS results combined across, and second, based on the separate GWAS results by sex. Estimation of the S and V matrices is a 3-step process. In step 1, the raw GWAS summary statistics were manipulated using the gSEM *munge* option to remove all SNPs with MAF < 1%, information scores < 0.9, and SNPs in the MHC region. In step 2, we used the gSEM *ldsc* option to run multivariate LD score regression <sup>46</sup> to estimate the covariance (S) and asymptotically weighted covariance (V) matrices between the GWAS summary statistics. In step 3, we then used

the lavaan (version 0.6-7)<sup>54</sup> in R (version 4.0.3)<sup>53</sup> to fit and compare competing longitudinal and multivariate models to the S and V matrices.

The autoregression model predicts that time-specific random genetic or environmental effects “innovations” are more or less persistent over time (autoregressive effects)<sup>38</sup>. As described by Eaves and others<sup>38-40</sup> and illustrated in Figure 1, genetic variance at each occasion is a function of i) new random effects “innovations” on the phenotype as well as ii) the linear contribution of genetic differences expressed at the preceding time. We assume that cross-temporal correlations arise because the innovations have a more or less persistent effect over time and may, under some circumstances accumulate, potentially giving rise to developmental increases in genetic variance and increased correlations between adjacent measures. One consequence of the autoregressive model is the tendency of cross-temporal correlations to decay as a function of increasing lag-time. Depending on the magnitude of an innovation and its relative persistence, the observed variances and cross-temporal covariances may increase towards a stable asymptotic value. We tested this autoregression model by fitting genetic innovations at the 45-49, 50-54, 55-59, 60-64 and 65-73 age-intervals, which we then successively dropped. We then specified an autoregression model that included a single source of genetic variance at the 40-44 age interval, which accounted for genetic variance at all subsequent age intervals. Finally, we fitted an exploratory factor analysis comprising a single factor.

#### *Model fit indices & comparisons*

In gSEM analyses there is no one sample size to speak of. This is because GWAS studies from which the summary statistics are derived can vary in size and subject overlap. Thus, potentially, a different (effective) sample size may apply to each element of S. We were therefore limited to fit indices that do not explicitly depend upon sample size: the pseudo Akaike Information Criterion (pseudoAIC); Comparative Fit Index (CFI); Tucker Lewis Index (TFI); and the Standardized Root Mean Square Residual (SRMR) to judge the best-fitting model. Both the CFI and TFI are incremental fit indices that penalize models with increasing complexity. The SRMR is an absolute measure of fit based on the difference between the observed and predicted correlations under each model, such that a value of zero indicates a perfect fit. The pseudoAIC is a comparative fit index, whereby the model with the lowest AIC values is interpreted as the best-fitting.

**Table 1.** Sample sizes, estimates of SNP-based heritability (including standard errors along diagonal) & linkage disequilibrium score regression genetic correlations between the six age intervals based on the combined male & female GWAS summary statistics.

	Sample size	1.	2.	3.	4.	5.	6.
1. BMI GWAS 40-44 yrs	34,001	0.23 (0.02)					
2. BMI GWAS 45-49 yrs	45,294	1.00	0.26 (0.02)				
3. BMI GWAS 50-54 yrs	53,602	0.99	1.00	0.26 (0.02)			
4. BMI GWAS 55-59 yrs	64,891	0.93	0.93	0.95	0.29 (0.01)		
5. BMI GWAS 60-64 yrs	89,824	0.95	0.94	0.93	0.90	0.24 (0.01)	
6. BMI GWAS 65-73 yrs	71,178	0.97	0.96	0.95	0.93	1.00	0.22 (0.01)

## Results

### Combined male & female analyses

The LDSR-based genome-wide genetic correlations between the six GWAS summary statistics, including GWAS sample sizes and SNP-based heritability at each age interval, are shown in Table 1. The correlations do not continue to decline with increasing time intervals, which would be indicative of a simplex structure best explained by autoregression models. For example, the LDSR genetic correlation ( $r_g$ ) between BMI at ages 40-45 and 66-73 years was higher than the  $r_g$  between BMI at ages 40-45 and 56-60 years ( $r_g=0.97$  vs 0.93). Overall, the genetic correlations were very high and ranged from 0.93 to 1.00.

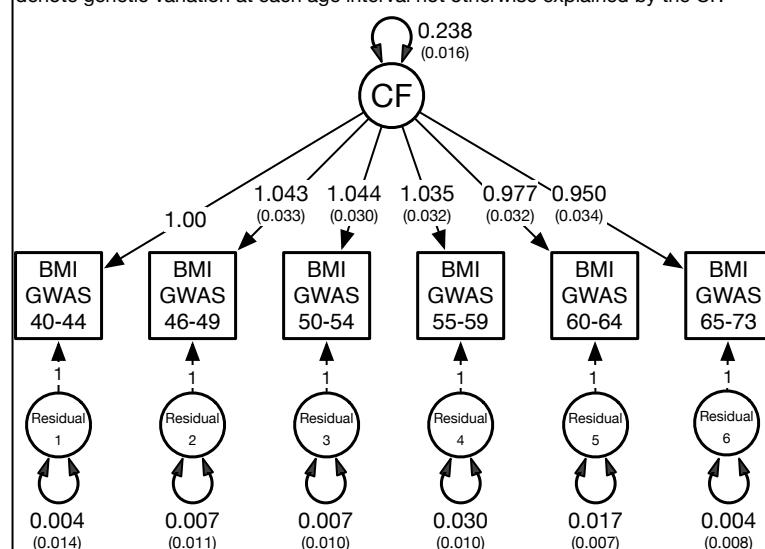
**Table 2.** Multivariate modeling fitting comparisons based on the combined male & female GWAS summary statistics.

Models	Chi-square(df)	p	pseudoAIC	CFI	TLI	SRMR
Full auto-regression (AutoReg)	21.113 <sub>(13)</sub>	0.071	55.113	0.999	0.999	0.039
AutoReg: genetic innovation at 65-73 yrs dropped	22.419 <sub>(14)</sub>	0.070	54.419	1.000	0.999	0.039
AutoReg: genetic innovation at 60-64 yrs dropped	20.872 <sub>(14)</sub>	0.105	52.872	1.000	0.999	0.039
AutoReg: genetic innovation at 55-59 yrs dropped	25.403 <sub>(14)</sub>	0.031	57.403	0.999	0.999	0.041
AutoReg: genetic innovation at 50-54 yrs dropped	21.768 <sub>(14)</sub>	0.084	53.768	0.999	0.999	0.040
AutoReg: genetic innovation at 45-49 yrs dropped	34.073 <sub>(14)</sub>	0.002	66.073	0.998	0.998	0.051
AutoReg: genetic innovations at 45-73 yrs dropped	46.133 <sub>(14)</sub>	0.000	70.133	0.998	0.998	0.056
Exploratory factor analysis - 1 factor	13.005 <sub>(9)</sub>	0.162	37.005	1.000	1.000	0.016

Note: AIC = Akaike Information Criterion, CFI = comparative fit indice, TLI = Tucker Lewis Indice, SRMR = (Standardized) Root Mean Square Residual

Formal model fitting comparisons are shown in Table 2. We began with a fully saturated autoregression model comprising unique genetic influences or innovations at each age interval (Figure 1). This provided a reasonable fit to the data as judged by the non-significant chi-square, very high CFI and TLI values and very low SRMR. Autoregression sub-models in which the genetic innovations at ages 65-73, 60-64, and

Figure 2. Best-fitting common factor (CF) model based on the combined male and female GWAS data. This model best explains the covariation between the six GWAS summary statistics based on five-year age intervals between ages 40-73 years. To identify this model, the factor loading on the BMI GWAS at 40-45 years was constrained to one. The double-headed arrow on the CF denotes the standardized variance, or SNP-based heritability, for BMI. Double-headed arrows on the residuals denote genetic variation at each age interval not otherwise explained by the CF.



65-73, 60-64, 55-59, 50-54 and 45-49 years were each successively removed provided only marginal improvements in terms of their pseudoAIC values. In contrast, the model with a single factor provided the overall best fit in terms of the smallest chi-square, lowest pseudoAIC and lowest SRMR. Under this model (see Figure 2), genetic variance at each five-year age interval was best explained by a single factor with

a genome-wide SNP heritability of 24%.

#### Sex specific analyses

An identical pattern emerged when the model fitting was repeated by sex. Male and female sample sizes at each age interval are shown in Supplementary Table S1. Table 3 shows the LDSR genetic correlations for men and women. Varying only slightly, the separate male and female genetic correlations were again high and ranged from  $r_g=0.88$  to  $r_g=1.00$ . Supplementary Table S1 also shows the SNP-based heritability estimates by sex, which were not only very similar at each age interval, but were largely unchanged when 40 ancestry principal components were included as covariates.

**Table 3.** Linkage disequilibrium score regression genetic correlations based on the male (below diagonal) & female (above diagonal *italics*) GWAS summary statistics at six age intervals.

	1.	2.	3.	4.	5.	6.
1. BMI GWAS 40-44 yrs	1	0.99	0.99	0.91	0.95	0.92
2. BMI GWAS 45-49 yrs	0.98	1	1.00	0.96	0.93	0.93
3. BMI GWAS 50-54 yrs	1.00	0.99	1	0.95	0.91	0.90
4. BMI GWAS 55-59 yrs	0.93	0.89	0.93	1	0.88	0.93
5. BMI GWAS 60-64 yrs	0.97	0.90	0.95	0.88	1	0.98
6. BMI GWAS 65-73 yrs	0.97	0.90	0.96	0.93	0.99	1

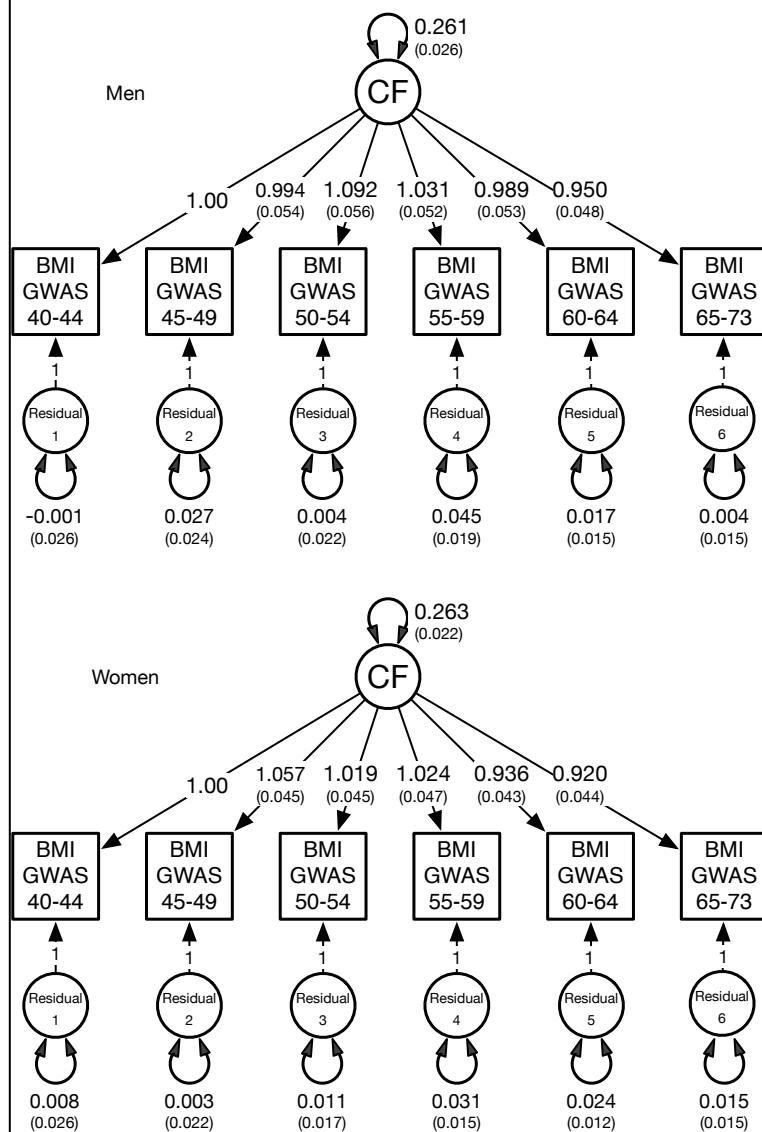
As shown in Table 4, the genetic innovation parameters at ages 46-73 years for each sex could each be dropped from the full autoregression model as judged by the non-significant chi-square value. Overall, however, a model with a single common factor again provided the best fit to the data for each sex in terms of the lowest chi-square, pseudoAIC and SRMR values (see Figure 3). This suggests that there is no evidence of age-specific genetic variation in BMI for either men or women.

**Table 4.** Multivariate modeling fitting comparisons based on the combined MALE GWAS summary statistics at six age intervals.

	Women	ChiSquare <sub>df</sub>	p	pseudoAIC	CFI	TLI	SRMR
Full auto-regression (AutoReg)		15.019 <sub>(13)</sub>	0.306	49.019	1.000	1.000	0.043
AutoReg: genetic innovation at 65-73 yrs dropped		14.866 <sub>(14)</sub>	0.387	46.866	1.000	1.000	0.043
AutoReg: genetic innovation at 60-64 yrs dropped		14.883 <sub>(14)</sub>	0.386	46.883	1.000	1.000	0.043
AutoReg: genetic innovation at 55-59 yrs dropped		16.813 <sub>(14)</sub>	0.266	48.813	0.999	0.999	0.046
AutoReg: genetic innovation at 50-54 yrs dropped		14.213 <sub>(14)</sub>	0.434	46.213	1.000	1.000	0.043
AutoReg: genetic innovation at 45-49 yrs dropped		21.482 <sub>(14)</sub>	0.090	53.482	0.998	0.998	0.057
AutoReg: genetic innovations at 45-73 yrs dropped		25.617 <sub>(14)</sub>	0.109	49.617	0.998	0.999	0.059
Exploratory factor analysis - 1 factor		8.832 <sub>(9)</sub>	0.453	32.832	1.000	1.000	0.023
	Men						
Full auto-regression (AutoReg)		11.858 <sub>(13)</sub>	0.539	45.858	1.000	1.000	0.054
AutoReg: genetic innovation at 65-73 yrs dropped		12.814 <sub>(14)</sub>	0.541	44.814	1.000	1.000	0.054
AutoReg: genetic innovation at 60-64 yrs dropped		12.085 <sub>(14)</sub>	0.599	44.085	1.000	1.000	0.053
AutoReg: genetic innovation at 55-59 yrs dropped		11.889 <sub>(14)</sub>	0.615	43.889	1.000	1.000	0.053
AutoReg: genetic innovation at 50-54 yrs dropped		21.826 <sub>(14)</sub>	0.082	53.826	0.998	0.998	0.064
AutoReg: genetic innovation at 45-49 yrs dropped		12.718 <sub>(14)</sub>	0.549	44.718	1.000	1.000	0.059
AutoReg: genetic innovations at 45-73 yrs dropped		1628.983 <sub>(14)</sub>	0.000	1652.983	0.168	0.001	0.803
Exploratory factor analysis - 1 factor		4.398 <sub>(9)</sub>	0.883	28.398	1.001	1.000	0.018

Note: AIC = Akaike Information Criterion, CFI = comparative fit indice, TLI = Tucker Lewis Indice, SRMR = (Standardized) Root Mean Square Residual

Figure 3. Best-fitting common factor (CF) models based on separate male and female GWAS data. To identify these models, the factor loadings on the BMI GWAS at 40-45 years were constrained to one. The double-headed arrow on the CF denotes the standardized variance, or SNP-based heritability, for BMI. Double-headed arrows on the residuals denote genetic variation at each age interval not otherwise explained by the CF.



and the other age intervals were all equally high. Thus, the genetic variance at age 60-64 does not appear to be linked to any age-specific or distinct genetic processes occurring around this time.

The extent to which our observed pattern of homogenous and stable genetic influences is replicable at younger ages remains to be determined. Longitudinal genetic correlations assessed at intervals spanning infancy, adolescence and young adult range from moderate to high<sup>17,19,55,56</sup>. Felix et al.<sup>57</sup> estimated a high genetic correlation between childhood and adult BMI of 0.73. Typically, however, higher correlations are observed between shorter intervals. For example, Silventoinen et al.<sup>17</sup> reported an  $r_g$  of

## Discussion

This is the first study to test a genetic developmental theory of BMI heritability using molecular data and structural equation modeling. From ages 40 and 73, changes in BMI heritability could not be explained by age-specific genetic factors or an accumulation of new sources of genetic variance over time. Instead, individual differences in BMI genetic variance across this time span were best explained by a single or common set of stable, genetic influences that are observable in early midlife. This pattern was observed in both men and women.

Dahl et al.'s<sup>1</sup> analysis of Swedish twin data revealed that for men and women, BMI increases across midlife, before leveling off at 65 years and declining at approximately age 80. The extent to which the inflection at age 65 is characterized by genetic innovations was unsupported by our results. In fact, we found that the genetic correlation between the GWAS based on BMI assessed at ages 60-64

0.31 between BMI at ages 1 and 18 that increased to 0.93 between BMI at ages 17 and 18 years. Haberstick et al. <sup>19</sup> reported a similarly high genetic correlation ( $r_g = 0.97$ ) between BMI at ages 17 and 22 years. The autoregressive modeling reported by Cornes et al. <sup>18</sup> found evidence of distinct, age-specific genetic influences on BMI at ages 12, 14 and 16. However, the authors also found large transmission coefficients in both sexes, which means that same genetic factors were largely responsible for the observed variation across the three measurement occasions. In older samples, longitudinal genetic correlations between BMI assessed at age 20 and when subjects were in their mid-forties range from 0.60 to 0.69 <sup>20,23</sup>. In terms of GWAS findings, many adult BMI variants have also been observed in early growth traits in childhood, but not infancy <sup>58</sup>. We note that Sovio et al. identified one the clearest examples using molecular data of age-specific genetic effects, whereby the effect of the FTO locus on BMI reverses between adiposity peak adiposity rebound during infancy <sup>59</sup>. In terms of LDSR findings, Trzaskowski et al. <sup>27</sup> reported a genetic correlation of 0.86 between BMI at ages 11 and 65. The same authors also found that the adult PRS for BMI explained at most 10% of the phenotypic variance in childhood BMI. Combined, the moderate to high longitudinal genetic correlations based on twin and molecular data suggest that individual differences in heritability spanning infancy, adolescence and early adult are likely to be explained by a combination of mostly age-invariant, and age-specific genetic influences.

### *Limitations*

Our findings should be interpreted in the context of four limitations.

First, the BMI data were not based on repeated measures. Consequently, our approach relied on between-subject analyses that assumed no year of birth or cohort effects. Analyses of Danish and Swedish twin data have shown that increases in mean BMI in successively younger cohorts have been accompanied by larger standardized estimates of heritability <sup>60,61</sup>. To examine if cohort effects exist, we inspected the LDSR genetic correlations between the youngest and oldest age intervals i.e., two maximally age-discrepant samples of unrelated individuals. The genetic correlation was  $r_g = 0.97$  (see Table 1), which suggests that the impact of cohort-related genetic heterogeneity, if present, was likely to be minimal.

Second, the UKB recruitment process did not represent a random sample of the UK population <sup>62</sup>. Subjects were predominately European ancestry, more likely to be older, female, to live in less socioeconomically deprived areas than nonparticipants, and when compared with the general population, were also less likely to be obese, to smoke, and to drink alcohol daily while reporting fewer self-reported health conditions <sup>63,64</sup>. Although Silventoinen et al.'s meta-analysis of twin data reported only minor differences in BMI heritability across divergent cultural-geographic regions <sup>65,66</sup>, the extent to which the molecular-based genetic covariance structure observed here generalizes to non-European populations remains to be determined.

Third, while our results illustrate the flexibility of SEM in terms of its application to GWAS data to test a theory of longitudinal change, our modeling was not exhaustive.

For instance, we were unable to test the hypothesis that changes in heritability could be better explained either by latent growth processes or mixture distributions<sup>67,68</sup> capable of discerning distinct genetic trajectories. We emphasize that the current method is limited to the analysis of summary variance-covariance matrices derived from the GWAS analysis of common variants. GenomicSEM does not model observed phenotypic information. Consequently, sample means could not be generated with which to model latent growth or mixture distributions. We also did not test hypotheses regarding sex differences other than to report results by sex. Dubois' meta-analysis of 23 twin birth-cohorts found evidence of sex-limitation in terms of greater genetic variance in boys in early infancy through to 19 years<sup>69</sup>. In contrast, Elks et al.'s meta-regression of 88 twin-bases estimates of BMI heritability found no evidence of sex effects<sup>16</sup>. It remains to be determined if the observed minor differences in the genetic covariances and the ultimate, best fitting single-factor structure are empirically equivalent across sex.

Finally, our genomic modelling was based on aggregated GWAS summary data and so was entirely independent of environmental risks, which are known to be significant in the etiology of complex traits<sup>70</sup>. Consequently, our current approach precludes modeling the contribution of environmental influences with increasing age<sup>71</sup> or making allowances for phenomenon such as genetic control of sensitivity to the environment i.e., GxE interaction<sup>65</sup>, which may explain the previously observed decreases in heritability during adulthood<sup>16</sup>. In this regard, methods capable of simultaneously modelling the joint effect of genes and environment are likely to prove more informative. For instance, innovative approaches capable of applying genomic-relatedness based restricted maximum-likelihood<sup>72</sup> to structural equation modeling software packages such as OpenMx<sup>73</sup> have the potential to analyze individual GWAS and phenotypic data and hold promise.

### **Conclusion**

In absence of true longitudinal repeated measures, SEM based analyses of genetic covariances derived from independent age restricted GWAS can be used to investigate questions regarding the stability and independence of genetic influences across time. Applying this framework to BMI across middle to late adulthood and testing two competing hypotheses, revealed that differences in BMI across age could not be explained by the accumulation of age-specific genetic influences or autoregressive processes. Instead, a common set of stable genetic influences appears to underpin genome-wide variation in BMI from middle to early old age in both men and women.

### **Data Availability**

All BMI and GWAS data used here are publicly available from the UK Biobank (<https://www.ukbiobank.ac.uk>).

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**Supplementary Table S1.** Number of men & women with complete BMI & GWAS data at each age interval, as well as genomic inflation ( $\lambda$ ) & SNP-based heritability ( $h^2$ ) for the GWAS analyses comprising 10 versus 40 principal components (PCs).

	Women	Sample size	10 PCs			40 PCs		
			$\lambda$	$h^2$	SE	$\lambda$	$h^2$	SE
1.	BMI GWAS 40-44 yrs	18,110	1.08	0.26	0.03	1.08	0.26	0.03
2.	BMI GWAS 45-49 yrs	24,975	1.12	0.28	0.03	1.12	0.27	0.03
3.	BMI GWAS 50-54 yrs	30,149	1.12	0.27	0.02	1.12	0.26	0.02
4.	BMI GWAS 55-59 yrs	36,010	1.18	0.30	0.02	1.17	0.30	0.02
5.	BMI GWAS 60-64 yrs	48,369	1.21	0.25	0.01	1.21	0.25	0.01
6.	BMI GWAS 65-73 yrs	35,460	1.16	0.23	0.02	1.15	0.23	0.02
Men								
1.	BMI GWAS 40-44 yrs	15,891	1.08	0.24	0.03	1.08	0.29	0.03
2.	BMI GWAS 45-49 yrs	20,319	1.09	0.28	0.03	1.08	0.17	0.03
3.	BMI GWAS 50-54 yrs	23,453	1.12	0.31	0.03	1.09	0.24	0.02
4.	BMI GWAS 55-59 yrs	28,881	1.16	0.32	0.02	1.12	0.25	0.02
5.	BMI GWAS 60-64 yrs	41,455	1.19	0.27	0.02	1.16	0.22	0.01
6.	BMI GWAS 65-73 yrs	35,718	1.16	0.24	0.02	1.18	0.31	0.02