

# Interacting phenotypes and the coevolutionary process: Interspecific indirect genetic effects alter coevolutionary dynamics

Running title: Trait interactions and coevolution

Stephen P. De Lisle<sup>1,2</sup>, Daniel I. Bolnick<sup>1</sup>, Edmund D. Brodie III<sup>3</sup>, Allen J. Moore<sup>4</sup>, and Joel W. McGlothlin<sup>5</sup>

Corresponding author: Stephen P. De Lisle      Email: [stephen.de\\_lisle@biol.lu.se](mailto:stephen.de_lisle@biol.lu.se)

1. Department of Ecology & Evolutionary Biology  
University of Connecticut  
75 N. Eagleville Road  
Storrs, Connecticut, USA 06269
2. Present address:  
Evolutionary Ecology Unit, Department of Biology  
Lund University  
Solvegatan 37, Lund, Sweden
3. Department of Biology and Mountain Lake Biological Station  
University of Virginia  
485 McCormick Road  
Charlottesville, VA 22904 USA
4. Department of Entomology  
University of Georgia  
Athens, GA 30602 USA
5. Department of Biological Sciences  
Virginia Tech  
2125 Derring Hall, 926 West Campus Drive  
Blacksburg, Virginia, USA 24060

**Author Contributions** All authors contributed to all aspects of the manuscript.

**Acknowledgements** Funding was provided by grants from the Royal Swedish Academy of Sciences and Swedish Research Council to S. De Lisle (VR registration number 2019-03706), the University of Connecticut and the NIAID (1R01AI123659-01A1) to D. Bolnick, and the National Science Foundation (DEB 1457463) to J. McGlothlin.

**Data Accessibility** No data to be archived

1 **Interacting phenotypes and the coevolutionary process: Interspecific**  
2 **indirect genetic effects alter coevolutionary dynamics**

3

4

5 **Abstract** Coevolution occurs when species interact to influence one another's fitness,  
6 resulting in reciprocal evolutionary change. In many coevolving lineages, trait expression in  
7 one species is modified by the genotypes and phenotypes of the other, forming feedback  
8 loops reminiscent of models of intraspecific social evolution. Here, we adapt the theory of  
9 within-species social evolution, characterized by indirect genetic effects and social selection  
10 imposed by interacting individuals, to the case of interspecific interactions. In a trait-based  
11 model, we derive general expressions for multivariate evolutionary change in two species and  
12 the expected between-species covariance in evolutionary change when selection varies across  
13 space. We show that reciprocal interspecific indirect genetic effects can dominate the  
14 coevolutionary process and drive patterns of correlated evolution beyond what is expected  
15 from direct selection alone. In extreme cases, interspecific indirect genetic effects can lead to  
16 coevolution when selection does not covary between species or even when one species lacks  
17 genetic variance. Moreover, our model indicates that interspecific indirect genetic effects  
18 may interact in complex ways with cross-species selection to determine the course of  
19 coevolution. Importantly, our model makes empirically testable predictions for how different  
20 forms of reciprocal interactions contribute to the coevolutionary process.

21

22 **Key words:** Coevolution, cross-species selection, interspecific indirect genetic effects,  
23 quantitative genetics, species interactions

## 24 **Introduction**

25 Coevolution occurs when interacting lineages evolve reciprocally in response to one another  
26 (Janzen 1980, Thompson 1982). Although the concept of coevolution may be applied to  
27 lineages that share genes, such as males and females of the same species (Arnqvist and Rowe  
28 2002), it was originally invoked to explain patterns of correlated evolution between  
29 interacting species (Ehrlich and Raven 1964). In some cases, coevolution can result in tightly  
30 integrated mutualisms or spectacular arms races that drive the evolution of exceptional  
31 phenotypes (Brodie et al. 2002, Pellmyr 2003, Johnson and Anderson 2010). Yet even  
32 beyond these striking cases, coevolution is likely important for a wide range of interacting  
33 lineages, including consumers and their resources, hosts and their pathogens, competitors,  
34 and mutualists (Thompson 1982, 1994). Although coevolution has clearly played a major  
35 role in the origins of diversity, much is still unknown about when and how species  
36 interactions generate reciprocal evolutionary change.

37       Theoretical models of coevolution typically focus on the fitness effects of trait  
38 interactions between coevolving species and how selection imposed by one species manifests  
39 evolutionary change in an interacting species (Nuismer 2017). The interaction between  
40 species in coevolving lineages is often intimate, with one species spending a greater part of  
41 its life cycle in close contact with the other. Thus, coevolution bears a striking resemblance  
42 to intraspecific social evolution (Stearns 2012). Like social evolution, coevolution often  
43 includes interacting or extended phenotypes, which arise when traits can only be understood  
44 within the context of interactions with others (Dawkins 1982, Moore et al. 1997).

45       In quantitative genetic models of intraspecific social evolution, social interactants  
46 influence one another via two pathways, each of which has a counterpart in coevolutionary  
47 theory. First, the phenotype of one individual may cause fitness effects in a social partner,  
48 leading to a form of selection known as social selection (West-Eberhard 1979, West-

49 Eberhard 1983, West-Eberhard 1984, Wolf et al. 1999). At the heart of all coevolutionary  
50 models is a form of reciprocal fitness interaction that resembles social selection, where the  
51 fitness of individuals in one species is influenced by traits in an interacting species (Brodie  
52 and Ridenhour 2003, Ridenhour 2005, Nuismer 2017). Second, models of social evolution  
53 may also include indirect genetic effects, which occur when the phenotype of one individual  
54 depends on the genotype of an interacting partner (Moore et al. 1997, Wolf et al. 1998). A  
55 classic example of indirect genetic effects is maternal effects, in which offspring phenotype is  
56 a function of both their own genes (a direct genetic effect) and maternal phenotypes such as  
57 litter size and provisioning (an indirect genetic effect) (Kirkpatrick and Lande 1989,  
58 Mousseau and Fox 1998, McAdam et al. 2002). When indirect genetic effects are reciprocal,  
59 feedback effects may inflate the genetic variance available for response to selection,  
60 drastically accelerating the rate of evolution (Moore et al. 1997). Although most models of  
61 indirect genetic effects do not extend beyond species boundaries, indirect genetic effects may  
62 also be common in species interactions between mutualists, competitors, parasites and hosts,  
63 and predators and prey. Phenotypic plasticity in response to an interacting species is  
64 common (Agrawal 2001), and when these influences on trait expression have a genetic basis  
65 they may represent interspecific indirect genetic effects (IIGEs; Shuster et al. 2006).  
66 Although IIGEs have received some attention in the context of community genetics (Shuster  
67 et al. 2006, Witham et al. 2020), their potential role in driving trait coevolution has been  
68 mostly unexplored.

69 To date, most explorations of IIGEs have been studies providing empirical support for  
70 their likely existence and their contribution to trait variation. Examples of interspecific  
71 phenotypic manipulation are common in nature (Table 1), and many of these cases can be  
72 argued to be putative cases of IIGEs. One possible example occurs in arbuscular  
73 mycorrhizae, where the genotypes of fungal mutualists can alter root traits in the plants they

**Table 1.** Coevolutionary interactions where IIGEs may be prevalent.

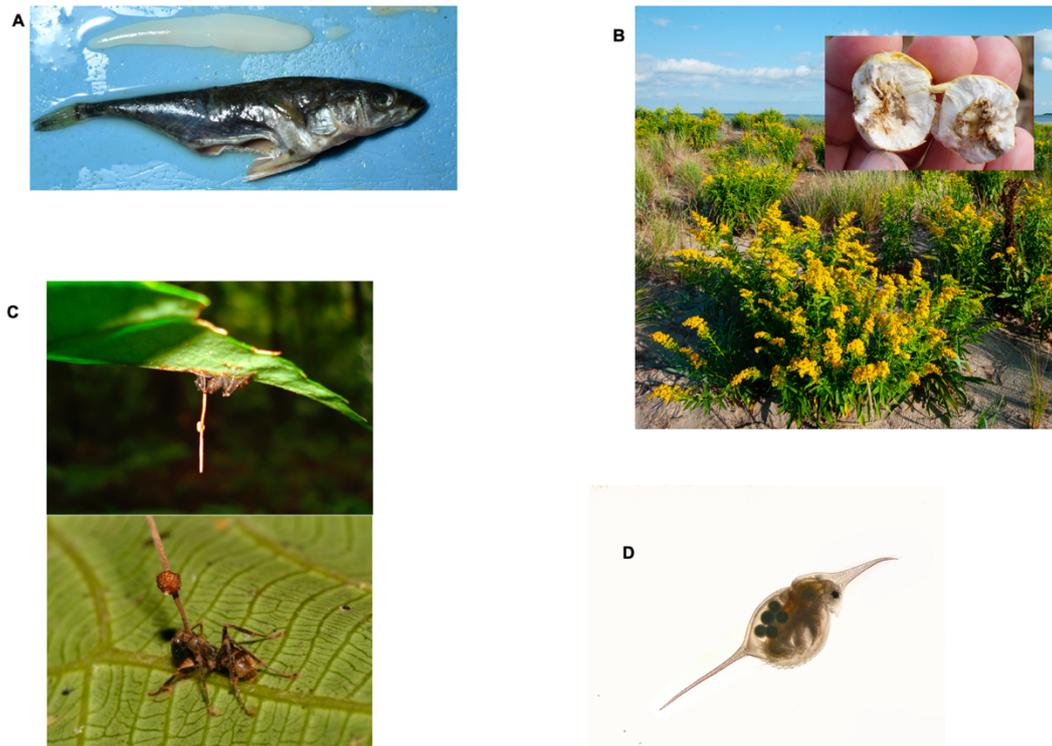
<b>Interaction</b>	<b>IIGE synonym</b>	<b>Diffuse/individual interaction</b>	<b>Example References</b>
host-parasite	host manipulation	both	(Thomas et al. 2012)
predator-prey	trait-mediated indirect interaction	both	(Peacor and Werner 2001)
plant-animal	inducible direct defense host-plant manipulations	both	(Weis and Abrahamson 1986, Chen 2008)
plant-microbe	joint trait microbially mediated trait	diffuse	(Friesen et al. 2011, O'Brien et al. 2021)
host-microbiome	host control host-microbiome interaction	diffuse	(Stappenbeck and Virgin 2016, Foster et al. 2017)
consumer-resource	toxin sequestration	both	(Züst et al. 2018)

74 inhabit (Gianinazzi-Pearson et al. 2007). In host-parasite systems, parasite manipulation of  
75 host traits (such as behavior) and reciprocal host manipulation of parasite traits (such as  
76 growth rate, via immune response) are key features of species interactions (Thomas et al.  
77 2012). Importantly, in many host-parasite systems, both the host and the parasite experience  
78 sustained interactions with a small number of individuals of the other species, often over key  
79 periods of the life history. For example, helminth parasites excrete a variety of  
80 immunomodulatory products that suppress or misdirect the immune system of their  
81 individual host (Damian 1997, Schmid-Hempel 2008, Oladiran and Belosevic 2012). Thus,  
82 host immune response to infection is controlled by the genotype of both the host and parasite  
83 (e.g., Barribeau 2014). As a specific example, some populations of threespine stickleback  
84 (*Gasterosteus aculeatus*) initiate a strong immune response to infection by the cestode  
85 *Schistocephalus solidus* (Fig. 1A) involving granulocyte proliferation and fibrosis, which

86 effectively suppress cestode growth and viability (Weber et al. 2017). Other stickleback  
87 populations do not exhibit this response and allow rapid cestode growth, perhaps representing  
88 a tolerance strategy. Cestode growth is thus an indirect genetic effect of its host's genotype.  
89 Conversely, the cestode has been shown to secrete compounds that suppress this host  
90 response (Scharsack et al. 2004, 2007, 2013) and down-regulate sticklebacks' pro-fibrotic  
91 gene expression (Fuess et al. 2020), suggesting reciprocal indirect genetic effects.

92 In goldenrod (*Solidago*; Fig. 1B), size of galls produced by the gall fly *Eurosta* is  
93 determined by genotypes of both fly and plant, and evolution of gall size is influenced in part  
94 by cross-species selection imposed on *Eurosta* larvae by species at other trophic levels (Weis  
95 and Abrahamson 1986, Weis et al. 1992, Abrahamson and Weis 1997). This type of  
96 interaction is common across gall-forming insects and their plant hosts; in *Hormaphis* aphids,  
97 variation the *bicycle* gene has been linked to variation in gall size (Korgaonkar et al. 2021).  
98 In many herbivore-plant interactions, physical damage to leaves induces upregulation of  
99 defensive compounds to deter further herbivory, which can be countered by matching  
100 physiological changes in the herbivore (Ohgushi 2005). For example, *Littorina* snail  
101 herbivory changes foliar chemistry of the brown seaweed *Ascophyllum nodosa* (increased  
102 phlorotannin concentrations), which in turn reduces snail movement and consumption rates  
103 (Borell et al. 2004).

104 Host-parasite, host-parasitoid, and some plant-herbivore interactions can entail  
105 intimate long-term associations between individuals. In contrast, predator-prey interactions  
106 tend to be more diffuse (Brodie and Brodie 1999). Prey sense predation risk through  
107 chemical, auditory, or visual cues and change their morphology, physiology, or behavior in



**Figure 1.** Examples of traits potentially mediated by IIGEs. IIGEs are a common feature of species interactions, particularly in host-parasite systems where prolonged contact between single individuals of both species determines fitness and trait expression for both. Here we highlight four examples where such IIGEs are likely contributing to trait expression and possibly patterns of among-population (co)variation. Panel A shows threespine stickleback fish (*Gasterosteus aculeatus*) and their cestode parasite *Schistocephalus*. Freshwater stickleback are the intermediate host for *Schistocephalus*, and each cestode acquires most of its lifetime resource pool while living inside a single host fish. *Schistocephalus* impose substantial reproductive and survival costs on hosts, and hosts have evolved an inducible (by the cestode) defense that suppresses cestode growth. Substantial among population-variation in this trait and in infection rates suggest these IIGEs may be mediating coevolution. Panel B shows goldenrod (*Solidago*) and a gall induced by the larvae (visible inside the gall) of the specialist gall-forming fly *Eurosta*. Gall size is induced by the genotype of the *Eurosta* larvae, and past work has shown that gall expression is a complex interaction between plant and fly genotype. Panel C shows the fungus *Ophiocordyceps unilateralis*, which manipulates behavior of its ant host prior to emergence of its fruiting body (Anderson et al. 2009). Panel D shows the water flea *Daphnia lumholtzi*, which induces growth of protective spines in response to chemical cues released by predatory fish (Agrawal 2001). Panel A and B main photos: S. De Lisle. Inset (B) photo: SriMesh / CC BY-SA (<https://creativecommons.org/licenses/by-sa/3.0/>); C photo: "File:Ophiocordyceps unilateralis.png" by David P. Hughes, Maj-Britt Pontoppidan / CC BY 2.5; Panel D: "Water flea (*Daphnia lumholtzi*)" by Frupus / CC BY-NC 2.0.

108 ways that mitigate their risk of predator encounter (Werner and Peacor 2003, Preisser et al.  
109 2005). Perhaps the most prominent example is the tendency of some *Daphnia* genotypes to  
110 grow spines (Fig. 1D), or to migrate to other depths, when they detect scent cues  
111 (kairomones) from predatory fish (Weber and Declerck 1997, Boersma et al. 1998). These  
112 antipredator responses lead to systemic changes in gene expression and morphology (Tams et  
113 al. 2019), which are controlled in part by fish traits (e.g., production of a scent cue). These  
114 scents can themselves be variable and genetic, as illustrated by differences in how *Daphnia*  
115 respond to cues from landlocked versus anadromous alewife (Walsh and Post 2011). The  
116 *Daphnia* traits can thus be described resulting from IIGEs controlled by both *Daphnia* and  
117 fish genotypes. Unlike the intimate host-parasite interactions, however, prey may be  
118 responding to diffuse cues from a predator population as a whole.

119         Although IIGEs would appear to play a central role in trait interactions between many  
120 coevolving species, little is known about how these effects influence the dynamics of  
121 coevolution (Scheiner et al. 2015). Past approaches, which have included variance-  
122 partitioning models of community assembly (Shuster et al. 2006, Whitham et al. 2020) and  
123 models of “joint traits” expressed together by interacting species (Queller 2014, O’Brien et  
124 al. 2021), are suggestive of an important role for IIGEs in species interactions. However, we  
125 lack a general understanding of how reciprocal IIGEs may affect the coevolution of  
126 interacting phenotypes. The fact that indirect genetic effects within a species can create  
127 feedback loops and other complex evolutionary dynamics, including in the context of within-  
128 species coevolution (Drown and Wade 2014), suggests that IIGEs could play a major role in  
129 mediating trait coevolution between species. Coopting concepts from intraspecific social  
130 evolution theory, where trait-based IIGEs are well developed, thus provides a natural way to  
131 understand the trait interactions that drive coevolution. Importantly, this trait-based approach  
132 allows the contribution of indirect genetic effects to the coevolutionary process to be fully

133 explored. Our goal in this paper is to develop such models to provide a comprehensive  
134 theoretical assessment of how IIGEs contribute to coevolution between species.

135 Here we adapt the trait-based theory of intraspecific social evolution (Moore et al.  
136 1997, Wolf et al. 1999, McGlothlin et al. 2010) to the case of two interacting species. Our  
137 model applies to both pairwise interactions between individuals (e.g., stickleback and  
138 cestodes) as well as diffuse interactions between species mean values (e.g., predatory fish and  
139 *Daphnia*). Importantly, our model accommodates both IIGEs, where genes in individuals of  
140 one species influence trait expression in individuals of another species, and cross-species  
141 selection, where phenotypes of individuals in one species influence fitness of individuals of  
142 another species (Figs. 1, 2). In addition to describing the contribution of these interspecific  
143 interactions to evolutionary change, we develop expressions for the among-population  
144 evolutionary covariance between traits of two interacting species; that is, the expected  
145 covariance in trait means across populations between two coevolving species. In the process,  
146 we formalize an interspecific analog of Zeng's (1988) quantitative genetic model of among-  
147 population trait covariation, which we expand to incorporate interacting phenotypes. Our  
148 analysis shows that IIGEs may have a central role in driving and mediating coevolution.

149

## 150 **Reciprocal evolutionary change in interacting species**

151 To model coevolution in two interacting species, we first decompose trait expression into  
152 three components: direct genetic effects, environmental effects, and indirect effects mediated  
153 by the phenotype of an interacting species. The phenotypic interface of coevolution involves  
154 traits with interacting effects across individuals of two species  $x$  and  $y$  and can be written as

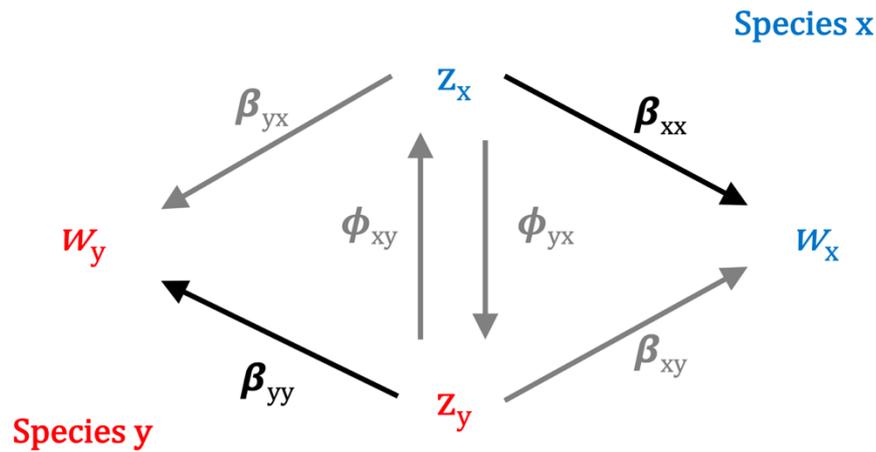
155

$$\mathbf{z}_x = \mathbf{a}_x + \mathbf{e}_x + \Phi_{xy}\mathbf{z}'_y \quad (1a)$$

$$\mathbf{z}_y = \mathbf{a}_y + \mathbf{e}_y + \Phi_{yx}\mathbf{z}'_x, \quad (1b)$$

156

157 where  $\mathbf{z}_i$  is a column vector of  $m$  traits expressed in an individual of species  $i = x$  or  $y$  ( $m$  is  
158 not necessarily equal in each species),  $\mathbf{a}_i$  is the corresponding column vector of direct genetic  
159 effects, and  $\mathbf{e}_i$  is an uncorrelated vector of residual environmental effects; primes denote traits  
160 of interacting individuals of another species. The matrix  $\Phi_{xy}$  quantifies the effect of traits in  
161 an interacting individual of species  $y$  ( $\mathbf{z}'_y$ ) on the expression of traits in a focal individual of  
162 species  $x$  (Fig. 2, Table 2), while the matrix  $\Phi_{yx}$  quantifies such effects in the opposite  
163 direction. Thus,  $\Phi_{ij}$  is an interspecific analog of the matrix of conspecific indirect genetic  
164 effects,  $\Psi$  (Moore et al. 1997). The individual elements of  $\Phi_{ij}$ , which we write as  $\phi_{ij}^{kl}$ ,  
165 represent partial regression coefficients of trait  $l$  in species  $j$  on trait  $k$  in species  $i$ . When  
166 traits are standardized to the same scale, these coefficients will typically be limited to a range  
167 of -1 to 1. Although we focus on interactions between pairs of individuals for simplicity of  
168 notation, this model could be easily extended to describe interactions with multiple  
169 individuals (cf. McGlothlin et al. 2010). For more diffuse interactions (e.g., alewife and  
170 *Daphnia*), the elements of  $\Phi_{ij}$  could represent a weighted average of effects from integrating  
171 across the phenotype distribution of the interacting population.



**Figure 2.** Path model of selection when traits and fitness are determined by interactions with heterospecifics. Individual trait values of two interacting species, x (blue) and y (red), are represented by  $z$ . Traits directly influence individual relative fitness ( $w$ ) of the species that express them via natural selection ( $\beta_{xx}$  and  $\beta_{yy}$ , black arrows). Traits can also reciprocally influence expression of traits of heterospecific social partners, via interspecific indirect genetic effects  $\Phi$ . Traits can also influence fitness of heterospecific social partners, via cross-species selection ( $\beta_{xy}$  and  $\beta_{yx}$ , grey arrows). Social effects are illustrated in grey arrows, direct effects in black.

**Table 2.** Definition of key parameters and expressions.

Expression / parameter	Biological definition
$\mathbf{z}_x, \mathbf{z}_y$	Vectors of individual trait values for species $x$ and $y$ , with a prime when present denoting partner traits.
$\mathbf{a}_x, \mathbf{a}_y$	Vectors of individual breeding values for species $x$ and $y$ .
$\Phi_{xy}, \Phi_{yx}$	Interspecific indirect effects (in matrix form). The effect of traits in individuals of species $y$ on trait expression in species $x$ , and vice-versa.
$\beta_{xx}, \beta_{yy}$	Vectors of within-species natural selection gradients for each species.
$\beta_{xy}, \beta_{yx}$	Vectors of cross-species selection gradients for each species. The effects of traits in species $y$ on fitness of individual species $x$ , and vice-versa.
$\mathbf{G}_{xx}, \mathbf{G}_{yy}$	Genetic covariance matrices of traits for species $x$ and $y$ .
$\mathbf{G}_{xy}, \mathbf{G}_{yx}$	Covariance matrices for breeding values of interacting individuals of species $x$ and $y$ within a population.
$\text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y)$	Coevolutionary covariance, defined as the covariance in evolutionary change between interacting species, across populations. When standardized by the variances in evolutionary response for each species, becomes a scale-free correlation.
$\text{Cov}(\beta_{xx}, \beta_{yy}),$	Between-species, among-population covariance in selection gradients (shown here for within-species selection)

172 Assuming that the mean residual environmental effect is zero, the population mean  
173 phenotype vector for each species is

174

$$\bar{\mathbf{z}}_x = \bar{\mathbf{a}}_x + \Phi_{xy} \bar{\mathbf{z}}_y \quad (2a)$$

$$\bar{\mathbf{z}}_y = \bar{\mathbf{a}}_y + \Phi_{yx} \bar{\mathbf{z}}_x. \quad (2b)$$

175

176 Noting the change in the mean additive genetic value in each species is  $\Delta \bar{\mathbf{a}} = \text{Cov}(\mathbf{a}, w)$

177 (Robertson 1966, Price 1970, 1972) where  $w$  is relative fitness, we can now define

178 evolutionary change in the multivariate mean phenotype as

179

$$\Delta \bar{\mathbf{z}}_x = \text{Cov}(\mathbf{a}_x, w_x) + \Phi_{xy} \Delta \bar{\mathbf{z}}_y \quad (3a)$$

$$\Delta \bar{\mathbf{z}}_y = \text{Cov}(\mathbf{a}_y, w_y) + \Phi_{yx} \Delta \bar{\mathbf{z}}_x. \quad (3b)$$

180

181 In equation (3), the first term on each right-hand side describes direct change due to natural

182 selection on the focal species, and the second term describes indirect change due to the

183 product of IIGEs and phenotypic change in the interacting species. This second term is

184 analogous to transmission bias (Fisher and McAdam 2019), and in this case, specifically

185 describes changes in one species that are induced by evolution in an interacting species. It

186 can be seen from equation (3) that species will coevolve whenever there are IIGEs in both

187 species, because the change in phenotypic mean in species  $x$  depends upon the change of

188 mean in species  $y$  and vice versa whenever both  $\Phi_{ij} \neq \mathbf{0}$ . Expanding equation (3) by

189 substitution results in explicit equations for evolutionary change in each species,

190

$$\Delta \bar{\mathbf{z}}_x = (\mathbf{I}_x - \Phi_{xy} \Phi_{yx})^{-1} [\text{Cov}(\mathbf{a}_x, w_x) + \Phi_{xy} \text{Cov}(\mathbf{a}_y, w_y)] \quad (4a)$$

$$\Delta \bar{\mathbf{z}}_y = (\mathbf{I}_y - \Phi_{yx} \Phi_{xy})^{-1} [\text{Cov}(\mathbf{a}_y, w_y) + \Phi_{yx} \text{Cov}(\mathbf{a}_x, w_x)]. \quad (4b)$$

191

192 where  $\mathbf{I}_x$  and  $\mathbf{I}_y$  are identity matrices with dimensionality equal to the number of traits in the  
 193 two species and the multiplier  $(\mathbf{I} - \Phi_{ij} \Phi_{ji})^{-1}$  quantifies the feedback effect of reciprocal  
 194 IIGEs. Equation 4 illustrates that total evolutionary change in a species is determined both  
 195 by change in the breeding value of that species ( $\text{Cov}(\mathbf{a}_i, w_i)$ ) and by change in the breeding  
 196 value of the interacting species, mediated by IIGEs ( $\Phi_{ij} \text{Cov}(\mathbf{a}_j, w_j)$ ). Whenever IIGEs occur  
 197 in both species such that  $\Phi_{ij} \Phi_{ji} \neq \mathbf{0}$ , this multiplier alters the total amount of evolutionary  
 198 change in both species. In order for such an effect to arise, there must be a feedback loop in  
 199 phenotypic expression. The simplest of these arises when there are two traits with reciprocal  
 200 IIGEs, such that trait  $k$  in species  $i$  affects trait  $l$  in species  $j$  and trait  $l$  in turn influences the  
 201 expression of trait  $k$ . In general, when  $\phi_{ij}^{kl}$  and  $\phi_{ji}^{lk}$  are of the same sign, the magnitude of  
 202 evolutionary change will be enhanced, and when they are of opposite signs, evolutionary  
 203 change will be diminished (Fig. 3).

204 To expand equation (4), we define the fitness of each interacting species using the  
 205 linear equations

206

$$w_x = \alpha_x + \mathbf{z}_x^T \boldsymbol{\beta}_{xx} + \mathbf{z}'_y{}^T \boldsymbol{\beta}_{xy} + \varepsilon_x \quad (5a)$$

$$w_y = \alpha_y + \mathbf{z}_y^T \boldsymbol{\beta}_{yy} + \mathbf{z}'_x{}^T \boldsymbol{\beta}_{yx} + \varepsilon_y \quad (5b)$$

207

208 where  $w$  is individual relative fitness,  $\alpha$  is an intercept,  $\varepsilon$  is an error term, and the superscript  
 209 T denotes transposition. Directional selection in each species is partitioned into two selection  
 210 gradients. First, the within-species selection gradients ( $\boldsymbol{\beta}_{xx}$  and  $\boldsymbol{\beta}_{yy}$ ) describe the direct  
 211 effects of an individual's traits on its own fitness (Fig. 2; Lande and Arnold 1983). The

212 cross-species selection gradients ( $\beta_{xy}$  and  $\beta_{yx}$ ) relate the fitness of a focal individual of one  
 213 species ( $x$  or  $y$ ) to the traits of individuals of the other coevolving species ( $y$  or  $x$ ) (Fig. 2).  
 214 Both of these are linear components of selection; we consider nonlinear terms (interactions  
 215 for fitness between focal and partner traits) below (see Incorporating Specific Fitness  
 216 Models). The cross-species selection gradient is analogous to the directional social selection  
 217 gradient in within-species models (Wolf et al. 1999) and if pairs or groups of interacting  
 218 individuals can be identified in a natural population, can be estimated in a similar way (cf.  
 219 Ridenhour 2005).

220 We now substitute our definitions of fitness into equation (4) and expand, yielding  
 221 predictive equations for evolutionary change in each species:

222

$$\Delta \bar{z}_x = (\mathbf{I}_x - \Phi_{xy} \Phi_{yx})^{-1} [\text{Cov}(\mathbf{a}_x, \mathbf{z}_x^T) \beta_{xx} + \text{Cov}(\mathbf{a}_x, \mathbf{z}'_y^T) \beta_{xy} + \Phi_{xy} \text{Cov}(\mathbf{a}_y, \mathbf{z}_y^T) \beta_{yy} + \Phi_{xy} \text{Cov}(\mathbf{a}_y, \mathbf{z}'_x^T) \beta_{yx}] \quad (6a)$$

223

$$\Delta \bar{z}_y = (\mathbf{I}_y - \Phi_{yx} \Phi_{xy})^{-1} [\text{Cov}(\mathbf{a}_y, \mathbf{z}_y^T) \beta_{yy} + \text{Cov}(\mathbf{a}_x, \mathbf{z}_x^T) \beta_{yx} + \Phi_{yx} \text{Cov}(\mathbf{a}_x, \mathbf{z}_x^T) \beta_{xx} + \Phi_{yx} \text{Cov}(\mathbf{a}_x, \mathbf{z}'_y^T) \beta_{xy}]. \quad (6b)$$

224

225 Each of these equations consists of four terms representing four components of the total  
 226 response to selection. The first term ( $\text{Cov}(\mathbf{a}_x, \mathbf{z}_x^T) \beta_{xx}$ ) represents response to within-species  
 227 selection ( $\beta_{xx}$  or  $\beta_{yy}$ ), the second ( $\text{Cov}(\mathbf{a}_x, \mathbf{z}'_y^T) \beta_{xy}$ ) represents response to cross-species  
 228 linear selection ( $\beta_{xy}$  or  $\beta_{yx}$ ), and the last two terms ( $\Phi_{xy} \text{Cov}(\mathbf{a}_y, \mathbf{z}_y^T) \beta_{yy} +$   
 229  $\Phi_{xy} \text{Cov}(\mathbf{a}_y, \mathbf{z}'_x^T) \beta_{yx}$ ) represent the component of change caused by the change in mean of  
 230 the interacting species. Note that the third and fourth terms of the first part of equation (6a)  
 231 equal the first two terms of (6b) multiplied by the IIGE coefficient  $\Phi_{xy}$ . Equation (6) also

232 shows that the change in response to within-species selection depends on the covariance of  
 233 additive genetic value with the phenotype of the same species, while change in response to  
 234 cross-species selection depends on the covariance of additive genetic value with the  
 235 phenotype of the opposite species.

236 To determine the components that give rise to these covariances, we expand our  
 237 definition of the phenotypes in equation (2) and rearrange, yielding

$$\begin{aligned}
 239 \quad \Delta \bar{z}_x = & (\mathbf{I}_x - \Phi_{xy} \Phi_{yx})^{-1} [(\mathbf{G}_{xx} + \mathbf{G}_{xy} \Phi_{xy}^T)(\mathbf{I}_x - \Phi_{yx}^T \Phi_{xy}^T)^{-1} \boldsymbol{\beta}_{xx} + & (7a) \\
 240 & (\mathbf{G}_{xy} + \mathbf{G}_{xx} \Phi_{yx}^T)(\mathbf{I}_y - \Phi_{xy}^T \Phi_{yx}^T)^{-1} \boldsymbol{\beta}_{xy} + \\
 241 & \Phi_{xy} (\mathbf{G}_{yy} + \mathbf{G}_{yx} \Phi_{yx}^T)(\mathbf{I}_y - \Phi_{xy}^T \Phi_{yx}^T)^{-1} \boldsymbol{\beta}_{yy} + \\
 242 & \Phi_{xy} (\mathbf{G}_{yx} + \mathbf{G}_{yy} \Phi_{xy}^T)(\mathbf{I}_x - \Phi_{yx}^T \Phi_{xy}^T)^{-1} \boldsymbol{\beta}_{yx}]
 \end{aligned}$$

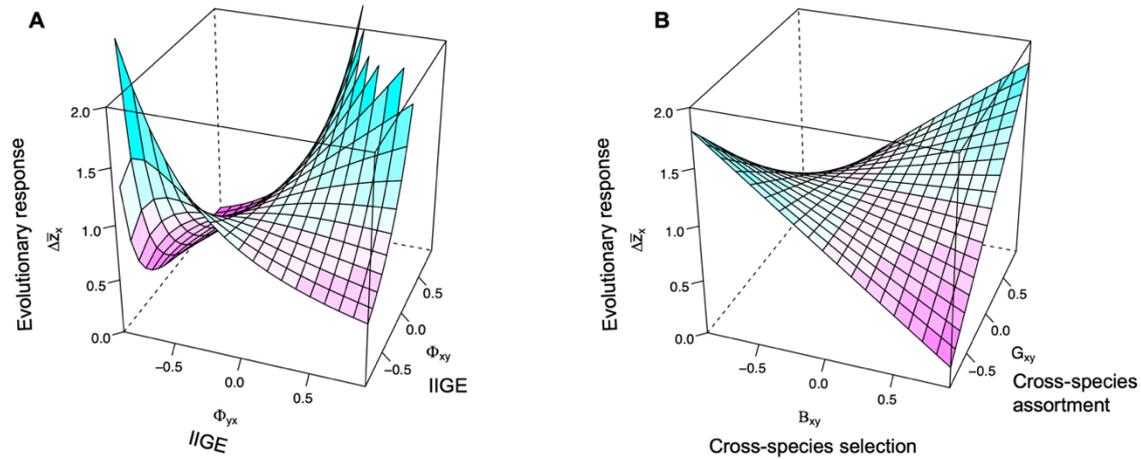
$$\begin{aligned}
 244 \quad \Delta \bar{z}_y = & (\mathbf{I}_y - \Phi_{yx} \Phi_{xy})^{-1} [(\mathbf{G}_{yy} + \mathbf{G}_{yx} \Phi_{yx}^T)(\mathbf{I}_y - \Phi_{xy}^T \Phi_{yx}^T)^{-1} \boldsymbol{\beta}_{yy} + & (7b) \\
 245 & (\mathbf{G}_{yx} + \mathbf{G}_{yy} \Phi_{xy}^T)(\mathbf{I}_x - \Phi_{yx}^T \Phi_{xy}^T)^{-1} \boldsymbol{\beta}_{yx} + \\
 246 & \Phi_{yx} (\mathbf{G}_{xx} + \mathbf{G}_{xy} \Phi_{xy}^T)(\mathbf{I}_x - \Phi_{yx}^T \Phi_{xy}^T)^{-1} \boldsymbol{\beta}_{xx} + \\
 247 & \Phi_{yx} (\mathbf{G}_{xy} + \mathbf{G}_{xx} \Phi_{yx}^T)(\mathbf{I}_y - \Phi_{xy}^T \Phi_{yx}^T)^{-1} \boldsymbol{\beta}_{xy}],
 \end{aligned}$$

248  
 249 where  $\mathbf{G}_{xx}$  and  $\mathbf{G}_{yy}$  represent within-species genetic (co)variance matrices and  $\mathbf{G}_{xy}$  and  $\mathbf{G}_{yx}$   
 250 represent cross-species genetic covariance. Note that in a model of pairwise interactions,  
 251  $\mathbf{G}_{yx} = \mathbf{G}_{xy}^T$ .

252 The cross-species genetic covariance represents a covariance of breeding values  
 253 between the species within the population of interest. This covariance may arise via any  
 254 mechanism that leads to nonrandom genetic assortment between species  $x$  and  $y$  with respect  
 255 to traits at the phenotypic interface of coevolution. A variety of phenomena could lead to

256 such assortment including behavioral preference for certain traits in heterospecific partners,  
257 fine-scale population structure, habitat preference, and vertical transmission of symbionts.  
258 We elaborate on the contribution of  $\mathbf{G}_{xy}$  and  $\Phi$  to phenotypic assortment between individuals  
259 of interacting species ( $C_{xy}$ ) in equation (A1). However, there is currently limited evidence of  
260 this type of interspecific genetic assortment, as few investigators seemed to have attempted to  
261 measure such a covariance. We therefore investigate dynamics in both the presence and  
262 absence of such assortment in our model development below.

263         Response to within-species selection depends on the sum of within-species genetic  
264 variance,  $\mathbf{G}_{yy}$  or  $\mathbf{G}_{xx}$ , and  $\mathbf{G}_{xy}\Phi_{xy}^T$  or  $\mathbf{G}_{yx}\Phi_{yx}^T$ , which represent an interaction between IIGEs  
265 and cross-species genetic covariance. Response to cross-species selection depends on the  
266 sum of cross-species genetic variance and  $\mathbf{G}_{xx}\Phi_{yx}^T$  or  $\mathbf{G}_{yy}\Phi_{xy}^T$ , which describes the genetic  
267 variance created by IIGEs. Each term in equation (7) also contains an additional feedback  
268 multiplier,  $(\mathbf{I}_y - \Phi_{xy}^T\Phi_{yx}^T)^{-1}$ , which further enhances the response to selection when there  
269 are reciprocal IIGEs. Because of this additional multiplier, when IIGEs are reciprocal and of  
270 the same sign, their effects on can be massive, mirroring the effects observed for within-  
271 species IIGEs (Moore et al. 1997, McGlothlin et al. 2010). As in previous equations, the last  
272 two terms in equation (7) represent a sort of evolutionary feedback that occurs across  
273 generations and is only present when there are IIGEs. These effects of  $\Phi$  and cross-species  
274 selection on evolutionary response are illustrated in Fig. 3.



**Figure 3.** Reciprocal IIGEs and cross-species selection change evolutionary response in a single species. Panels show the separate effects on evolutionary response in species x of indirect genetic effects (Panel A) and cross-species selection with genetic assortment (Panel B). Panel A shows the effects of reciprocal IIGEs holding all other evolutionary parameters constant, and assuming no cross-species selection. Panel B shows the effects of cross-species selection imposed by species y on species x, in combination with the genetic assortment between interactants of the different species, and assuming IIGEs are absent. In both panels,  $G_x = G_y = 1$ ,  $\beta_{yy} = \beta_{xx} = 1$ .

### 275 **Correlated evolution between interacting species: the coevolutionary covariance**

276 A key feature of coevolving lineages is correlated evolution across populations subject to  
 277 varying ecological conditions. Here, we seek to understand how the equations of selection  
 278 response can be used to understand how selection and IIGEs contribute to this shared among-  
 279 population divergence. We can explore the contribution of interspecific social effects to  
 280 correlated evolution between interacting species by solving for the covariance in evolutionary  
 281 change between interacting species,  $\text{Cov}(\Delta \bar{z}_x, \Delta \bar{z}_y)$ , which we call the coevolutionary  
 282 covariance (with the caveat that this covariance also reflects changes due in part to effects of  
 283 IIGEs). Such a covariance represents the expected pattern of reciprocal phenotypic change  
 284 through time in a single pair of populations of two species experiencing fluctuating selection,  
 285 or perhaps more importantly, across a set of populations in space under varying selection  
 286 pressures. Although mathematically equivalent, we focus on the latter scenario for its  
 287 relevance to understanding geographic variation among populations.

288           The coevolutionary covariance reflects the degree to which divergence among  
289 population means in two species has occurred jointly. High absolute values of the  
290 coevolutionary covariance indicate tightly-coupled coevolutionary change between the two  
291 species, whereas values around zero indicate that evolutionary change occurs independently.  
292 When scaled by the total amount of population divergence (variances in  $\Delta\bar{z}_x$  and  $\Delta\bar{z}_y$ ), this  
293 becomes a scale-free correlation describing the proportion of total divergence shared between  
294 the two species. Importantly, we expect the coevolutionary covariance to be often be related  
295 to the covariance in population means,  $\text{Cov}(\bar{z}_x, \bar{z}_y) \propto \text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y)$  because, for example,  
296  $\text{Cov}(\bar{z}_x, \bar{z}_y)^{t+1} = \text{Cov}(\bar{z}_x, \bar{z}_y)^t + \text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y)$  (under the assumption  $\text{Cov}(\Delta\bar{z}_x, \bar{z}_y) +$   
297  $\text{Cov}(\bar{z}_x, \Delta\bar{z}_y) = 0$ ). Thus, expanding  $\text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y)$  allows the possibility to assess how  
298 selection and IIGEs contribute to patterns of geographic variation in species mean  
299 phenotypes.

300           To simplify our analysis, we use models of a single trait in each species. Developing  
301 a full equation including all sources of covariance quickly becomes cumbersome, so we focus  
302 on three instructive special cases that illustrate the explicit impacts of considering cross-  
303 species selection and interspecific interacting phenotypes. The simplest case occurs when  
304 there are no IIGEs ( $\phi_{xy} = \phi_{yx} = 0$ ) and heterospecific interactions occur at random within  
305 populations ( $G_{xy} = G_{yx} = 0$ ). If we make the simplifying assumption that genetic variance  
306 does not differ among populations, the only source of covariance in selection response is  
307 covariance in direct within-species selection, or

308

$$\text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y) = G_{xx}G_{yy}\text{Cov}(\beta_{xx}, \beta_{yy}). \quad (8)$$

309

310 Equation (8) represents what is usually thought of as one source of correlated evolution  
311 between interacting species: predictable variation in the type of selection occurring across

312 space. Empirical data suggest that selection varies substantially in magnitude across space  
313 (Siepielski et al. 2013) and that spatially autocorrelated biotic selection plays a substantial  
314 role in driving divergence in trait means (Urban et al. 2011). Importantly, not just any  
315 variance in selection will do. In order to create covariance in evolutionary change, selection  
316 must covary between the interacting species. Cross-species selection, however, does not play  
317 a role in equation (8) because it does not contribute to an evolutionary response to selection  
318 in the absence of IIGEs and cross-species genetic covariance. Equation (8) is analogous to  
319 the results of Zeng's (1988) model of correlated trait evolution under directional selection,  
320 although for the case of traits expressed in different species.

321 To see an effect of cross-species selection, we first add cross-species genetic  
322 covariance but no IIGEs. For a one-trait (per species) model,  $G_{xy} = G_{yx}$ . Adding this effect  
323 leads to three new sources of covariance among populations:

324

$$\begin{aligned} \text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y) = & G_{xx}G_{yy}\text{Cov}(\beta_{xx}, \beta_{yy}) + G_{xy}^2\text{Cov}(\beta_{yx}, \beta_{xy}) + \\ & G_{xx}G_{xy}\text{Cov}(\beta_{xx}, \beta_{yx}) + G_{yy}G_{xy}\text{Cov}(\beta_{yy}, \beta_{xy}). \end{aligned} \quad (9)$$

325

326 The second term in equation (9) ( $G_{xy}^2\text{Cov}(\beta_{yx}, \beta_{xy})$ ) shows that cross-species selection will  
327 contribute to the coevolutionary covariance only when there is cross-species genetic  
328 covariance for the traits that affect fitness. Such covariances occur if, for example, parasite  
329 and host genotypes associate nonrandomly for the traits that influence their partner's fitness.

330 The third and fourth terms ( $G_{xx}G_{xy}\text{Cov}(\beta_{xx}, \beta_{yx}) + G_{yy}G_{xy}\text{Cov}(\beta_{yy}, \beta_{xy})$ ) represent a  
331 relationship between the effect of species on itself and on its heterospecific partner. These  
332 will be nonzero if populations with strong within-species selection also exhibit strong cross-  
333 species selection.

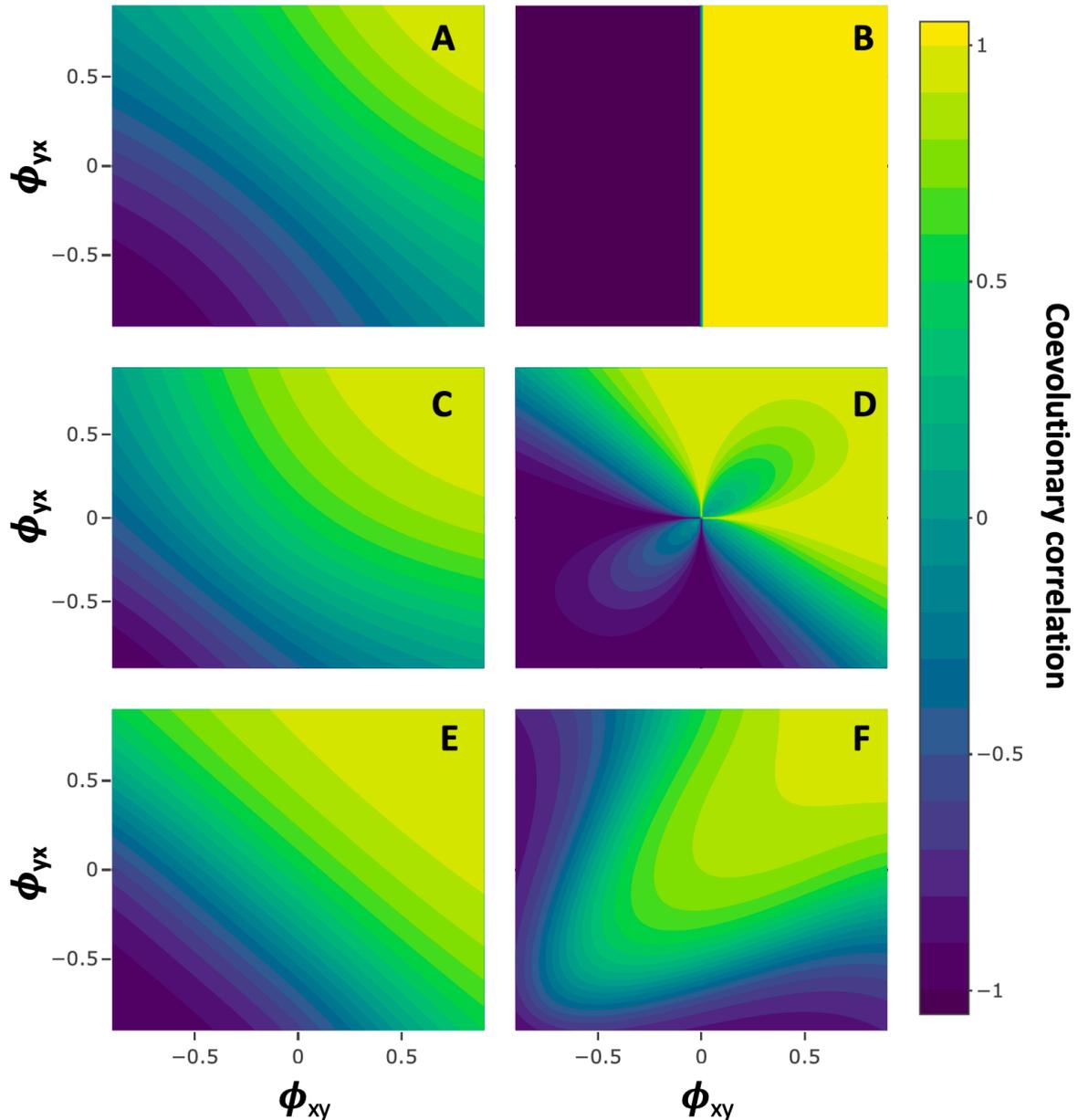
334 The most interesting effects on the coevolutionary covariance occur when we add  
 335 IIGEs ( $\phi_{xy}$  and  $\phi_{yx}$ ). With  $G_{xy} = 0$ , an appropriate simplifying assumption given that the  
 336 prevalence of  $G_{xy}$  is uncertain and is likely to be transient through time, the covariance  
 337 among populations in evolutionary response to selection is  
 338

$$\begin{aligned}
 \text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y) = & \mathcal{U}[G_{xx}G_{yy}\text{Cov}(\beta_{xx}, \beta_{yy}) + \\
 & G_{xx}^2\phi_{yx}\text{Var}(\beta_{xx}) + G_{yy}^2\phi_{xy}\text{Var}(\beta_{yy}) + \\
 & G_{xx}G_{yy}\phi_{xy}\phi_{yx}\text{Cov}(\beta_{xx}, \beta_{yy}) + G_{xx}G_{yy}\phi_{xy}\phi_{yx}\text{Cov}(\beta_{xy}, \beta_{yx}) + \\
 & G_{xx}G_{yy}\phi_{xy}\text{Cov}(\beta_{xx}, \beta_{yx}) + G_{xx}G_{yy}\phi_{yx}\text{Cov}(\beta_{yy}, \beta_{xy}) + \\
 & 2G_{xx}^2\phi_{yx}^2\text{Cov}(\beta_{xx}, \beta_{xy}) + 2G_{yy}^2\phi_{xy}^2\text{Cov}(\beta_{yy}, \beta_{yx}) + \mathcal{V}],
 \end{aligned} \tag{10}$$

339  
 340 where feedback effects of IIGEs are described by  $\mathcal{U} = (1 - \phi_{xy}\phi_{yx})^{-4}$  and  $\mathcal{V}$  collects  
 341 negligible third- and fourth-order  $\phi$  terms:  
 342

$$\begin{aligned}
 \mathcal{V} = & G_{xx}^2\phi_{yx}^3\text{Var}(\beta_{xy}) + G_{yy}^2\phi_{xy}^3\text{Var}(\beta_{yx}) + \\
 & G_{xx}G_{yy}\phi_{xy}\phi_{yx}^2\text{Cov}(\beta_{yy}, \beta_{xy}) + G_{xx}G_{yy}\phi_{xy}^2\phi_{yx}\text{Cov}(\beta_{yx}, \beta_{xx}) + \\
 & G_{xx}G_{yy}\phi_{xy}^2\phi_{yx}^2\text{Cov}(\beta_{yx}, \beta_{xy}).
 \end{aligned} \tag{11}$$

343  
 344 The first term in equation (10) is identical to equation (8) and represents covariance in direct  
 345 within-species selection. The second ( $G_{xx}^2\phi_{yx}\text{Var}(\beta_{xx})$ ) and third ( $G_{yy}^2\phi_{xy}\text{Var}(\beta_{yy})$ ) terms



**Figure 4.** IIGEs drive and mediate coevolution between interacting species. Panels show the coevolutionary correlation between two interacting species as a function of the indirect genetic effect parameters  $\phi$ , from equation (10) standardized by the evolutionary rates. In the absence of social interaction effects, correlated evolution (coevolution) between species is driven entirely by covariance in natural selection between the species (see text). Reciprocal IIGEs can generate coevolution even when there is not covariance in natural selection (Panel A), and can even drive coevolution even when one species lacks genetic variance (Panel B). IIGEs modify observed coevolutionary patterns when natural selection does covary ( $\text{Cov}(\beta_{xx}, \beta_{yy}) = 0.5$ ; Panel C). When cross-species selection and IIGEs act together, coevolutionary patterns are a complex third order polynomial (Panel D;  $\text{Cov}(\beta_{xy}, \beta_{yx}) = 0.5$ ,  $\text{Var}(\beta_{xy}) = \text{Var}(\beta_{yx}) = 1$ ). When natural and cross-species selection both act ( $\text{Var}(\beta_{xy}) = \text{Var}(\beta_{yx}) = \text{Var}(\beta_{xx}) = \text{Var}(\beta_{yy}) = 1$ ) and covary positively (0.5), (Panel E), effects of IIGEs become stronger in comparison to the case of (co)variance in natural selection alone. When covariance between natural and cross-species selection is negative (-0.8) a ridge is observed (Panel F).

347 show that in the presence of IIGEs, simple variance in within-species selection across  
348 populations can generate a coevolutionary covariance (or correlation, Fig. 4A; corresponding  
349 evolutionary rates and covariances are plotted in Figs. S1 and S2, respectively). This  
350 covariance in evolutionary response occurs as a necessary consequence of the dependence of  
351 the trait mean of one species on that of the other. Thus, in the presence of IIGEs, a  
352 coevolutionary covariance may occur even when selection is uncorrelated between the  
353 species, and in extreme cases, when only one species varies in selection or when only one  
354 species has genetic variation (Fig. 4B).

355         The fourth and fifth terms ( $G_{xx}G_{yy}\phi_{xy}\phi_{yx}\text{Cov}(\beta_{xx}, \beta_{yy}) +$   
356 ( $G_{xx}G_{yy}\phi_{xy}\phi_{yx}\text{Cov}(\beta_{xy}, \beta_{yx})$ ) show the effects of reciprocal IIGEs. When traits in the two  
357 species exist in a feedback loop, the effect of covariance in within-species selection is  
358 amplified if  $\phi_{xy}$  and  $\phi_{yx}$  are of the same sign and diminished if they are of opposite signs.  
359 The combined effects of the second, third, and fourth terms lead to a complex relationship  
360 between IIGEs and the coevolutionary covariance when within-species selection varies (Fig.  
361 4C). In some cases, IIGEs can even reverse the sign of the correlation that would be  
362 expected in their absence (Fig. 4C). In addition, reciprocal IIGEs may cause covariance in  
363 cross-species selection to contribute to the coevolutionary covariance, causing an even more  
364 complex relationship (Fig. 4D).

365         The last four terms in equation (10) show that in the presence of IIGEs, covariance  
366 between within-species and cross-species selection may contribute to the coevolutionary  
367 covariance (Fig. 4E-F). Because IIGEs inherently tie together the evolutionary responses of  
368 the two species through their effects on the coevolutionary covariance, this can occur both  
369 when there is covariance in gradients across species (terms 6 and 7) and when there is  
370 covariance in gradients in the same species (terms 8 and 9). The total effect of IIGEs may be

371 quite complex when cross-species selection varies, with subtle changes in  $\phi$  resulting in  
372 dramatic changes in the expected among population correlation (Fig. 4D, E-F).

373 An equation incorporating both IIGEs and  $G_{xy}$  quickly becomes unwieldy, but we  
374 present a compact form containing 10 (co)variance terms as equation (A2), which illustrates  
375 that when present, non-random assortment and IIGEs together interact in complex ways to  
376 influence coevolution.

377

### 378 **Incorporating specific fitness models**

379 Coevolutionary models often posit complex relationships between interacting phenotypes and  
380 fitness (Nuismer 2017). Although the selection model we present here is linear, more  
381 complex relationships can be incorporated by translating specific fitness functions to  
382 selection gradients. The adaptive landscape represents the theoretical relationship between a  
383 population's mean fitness and its phenotypic mean (Arnold et al. 2001). Selection gradients  
384 represent the partial slope of the adaptive landscape with respect to a given phenotype, and  
385 thus the multivariate selection gradient may be calculated using a vector of partial derivatives  
386 if the adaptive landscape can be written as a differentiable function (Lande 1979, Lande and  
387 Arnold 1983). In many cases, the multivariate selection gradient may be calculated using  
388 partial derivatives of the individual fitness function as well (Lande and Arnold 1983, Abrams  
389 et al. 1993, McGlothlin et al. 2021), evaluated over the phenotypic distribution (Phillips and  
390 Arnold 1989). Once selection gradients have been calculated for a given model, they may be  
391 substituted into equations (6–7) to explore the effects of a given fitness function on selection  
392 response and the coevolutionary covariance (see also Brodie and Ridenhour 2003). This

393 exercise also allows us to explore the effects of adding IIGEs to an existing coevolutionary  
394 model.

395 First, we consider a “phenotypic difference” model of ecological trait interaction  
396 (Nuismer et al. 2007 , Nuismer 2017), where absolute fitness is a function of the difference  
397 between interacting traits,  $W_x \propto \exp(z_x - z_y)$  and  $W_y \propto \exp(z_y - z_x)$ . Using the  
398 logarithm of the fitness function to calculate relative fitness (Lande and Arnold 1983),

399

$$\begin{aligned}w_x &= \alpha_x + b_x(z_x - z_y) + \varepsilon_x \\w_y &= \alpha_y + b_y(z_y - z_x) + \varepsilon_y.\end{aligned}\tag{12}$$

400

401 where  $b_x$  and  $b_y$  are constants that may vary across populations. Differentiating, the selection  
402 gradients are then  $\beta_{xx} = -\beta_{xy} = b_x$  and  $\beta_{yy} = -\beta_{yx} = b_y$ . Substituting these selection  
403 gradients into equation (7) is trivial. However, is worth noting that under this fitness model,  
404 the coevolutionary covariance simplifies to a function of just three (co)variance components,

405

$$\begin{aligned}\text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y) &= \mathcal{U}[G_{xx}G_{yy}(1 - \phi_{xy} - \phi_{yx} + 2\phi_{xy}\phi_{yx})\text{Cov}(b_x, b_y) + \\&G_{xx}^2\phi_{yx}(1 - 2\phi_{yx})\text{Var}(b_x) + G_{yy}^2\phi_{xy}(1 - 2\phi_{xy})\text{Var}(b_y) + \mathcal{V}],\end{aligned}\tag{13}$$

406

407 which corresponds to a relationship with  $\phi$  as in Figs. 4F, S1F, and S2F. This analysis  
408 illustrates that reciprocal IIGEs have greatest impact on mediating coevolutionary outcomes  
409 in a trait-matching models when IIGEs are similar in both sign and magnitude. Biologically,

410 such a situation corresponds to a scenario where, for example, trait expression is reciprocally  
411 escalated in response to heterospecific partners.

412 Another important class of fitness models to consider is the case of nonlinear fitness  
413 interactions. Nonlinearity is fundamental to many coevolutionary models that invoke  
414 epistatic interactions across species' genomes, such as the trait matching model for  
415 quantitative traits, or the single-locus matching-allele and gene-for-gene models of host-  
416 parasite coevolution (Dybdahl et al. 2014). For example, in the traditional model of trait  
417 matching, absolute fitness is assumed to be  $W_x \propto \exp(-(z_x - z_y)^2)$  and so relative fitness  
418 can be described by

419

$$\begin{aligned}w_x &= \alpha_x - b_{xy}(z_x - z_y)^2 + \varepsilon_x \\w_y &= \alpha_y - b_{yx}(z_y - z_x)^2 + \varepsilon_y.\end{aligned}\tag{14}$$

420

421 In this model the sign of the  $b$  terms correspond to different types of biological interaction;  
422 for example, in a mutualism, both species may take positive values of  $b$  and thus have a  
423 fitness peak when their trait value matches with their heterospecific partner. Taking partial  
424 derivatives over the phenotypic distribution yields the selection gradients

425

$$\begin{aligned}\beta_{xx} &= -\beta_{xy} = -2b_{xy}(\bar{z}_x - \bar{z}_y) \\ \beta_{yy} &= -\beta_{yx} = -2b_{yx}(\bar{z}_y - \bar{z}_x)\end{aligned}\tag{15}$$

426

427 In the simplest case of no IIGEs,  $G_{xy} = 0$ , and no variation in  $b$ , these gradients yield

428

$$\text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y) = -4G_{xx}G_{yy}b_{xy}b_{yx}\text{Var}(\bar{z}_x - \bar{z}_y).\tag{16}$$

429

430 Thus, within the limits of genetic variation and ignoring stochastic forces (as we have done),  
 431 evolutionary response will be perfectly correlated in the phenotypic matching model. When  
 432  $b_{xy}$  and  $b_{yx}$  share the same sign, this covariance in evolutionary response will be negative  
 433 because each species will be evolving towards (or away) from each other in trait space. This  
 434 covariance in evolutionary response occurs even without variation in  $b$  or covariance in trait  
 435 means. These results are broadly consistent with other analyses of the trait matching model,  
 436 which indicate that this model can generate relatively strong covariance in phenotypic means  
 437 across populations (Nuismer et al. 2010). Adding indirect genetic effects and assuming  
 438  $G_{xy} = 0$  and noting that  $\text{Var}(\bar{z}_x - \bar{z}_y) = \text{Var}(\bar{z}_y - \bar{z}_x)$ ,

439

$$\begin{aligned}
 \text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y) = & \mathcal{U}[-4b_{xy}b_{yx}G_{xx}G_{yy}\text{Var}(\bar{z}_x - \bar{z}_y) + \\
 & 4b_{xy}^2G_{xx}^2\phi_{yx}\text{Var}(\bar{z}_x - \bar{z}_y) + 4b_{yx}^2G_{yy}^2\phi_{xy}\text{Var}(\bar{z}_x - \bar{z}_y) \\
 & - 8b_{xy}b_{yx}G_{xx}G_{yy}\phi_{xy}\phi_{yx}\text{Var}(\bar{z}_x - \bar{z}_y) + \\
 & 4b_{xy}b_{yx}G_{xx}G_{yy}\phi_{xy}\text{Var}(\bar{z}_x - \bar{z}_y) + 4b_{yx}b_{xy}G_{xx}G_{yy}\phi_{yx}\text{Var}(\bar{z}_x - \bar{z}_y) \\
 & - 8b_{xy}^2G_{xx}^2\phi_{yx}^2\text{Var}(\bar{z}_x - \bar{z}_y) - 8b_{yx}^2G_{yy}^2\phi_{xy}^2\text{Var}(\bar{z}_x - \bar{z}_y) + \mathcal{V}]
 \end{aligned} \tag{17}$$

440 This expression again shows the covariance in evolutionary response occurs even in the  
 441 absence of (co)variance in  $b$ , and the addition of IIGEs alter the magnitude of the covariance  
 442 substantially. Analysis of more complex cases of the trait matching model, for example  
 443 where selection varies across space and/or across terms in the expanded polynomial  
 444  $(z_x - z_y)^2$ , would be straightforward in this framework.

445 As a final example of how this approach can be used to understand simple variations  
 446 on classic coevolutionary models, we focus on a multiplicative model of trait interaction,

447 where fitness depends on the interaction between traits of the two species. Consider a fitness  
448 model where relative fitness depends solely on the product of the two phenotypes:

449

$$\begin{aligned}w_x &= \alpha_x + b_{xy}z_xz_y + \varepsilon_x \\w_y &= \alpha_y + b_{yx}z_yz_x + \varepsilon_y.\end{aligned}\tag{18}$$

450

451 This model is conceptually and mathematically similar to the previous trait matching model,  
452 but importantly lacks stabilizing selection terms (e.g.,  $b_{xy}z_x^2$ ) present in the trait matching  
453 model. Because of this, directional natural selection depends only on the means of the other  
454 species:

455

$$\begin{aligned}\beta_{xx} &= b_{xy}\bar{z}_y \\ \beta_{xy} &= b_{xy}\bar{z}_x \\ \beta_{yy} &= b_{yx}\bar{z}_x \\ \beta_{yx} &= b_{yx}\bar{z}_y\end{aligned}\tag{19}$$

456

457 This relationship may cause selection to (co)vary across populations even when  $b_{xy}$  and  $b_{yx}$   
458 are homogeneous. In the absence of IIGEs, this simplest case would lead to a coevolutionary  
459 covariance defined by

460

$$\text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y) = G_{xx}G_{yy}b_{xy}b_{yx}\text{Cov}(\bar{z}_x, \bar{z}_y).\tag{20}$$

461 Thus, any initial covariance in the population means leads to a covariance in the response to  
 462 selection across species, lending a runaway aspect to the coevolutionary covariance. Adding  
 463 IIGEs, this becomes

464

$$\begin{aligned}
 \text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y) = & \mathcal{U}[G_{xx}G_{yy}b_{xy}b_{yx}\text{Cov}(\bar{z}_x, \bar{z}_y) \\
 & + G_{xx}^2\phi_{yx}b_{xy}^2\text{Var}(\bar{z}_y) + G_{yy}^2\phi_{xy}b_{yx}^2\text{Var}(\bar{z}_x) + \\
 & 2G_{xx}G_{yy}\phi_{xy}\phi_{yx}b_{xy}b_{yx}\text{Cov}(\bar{z}_x, \bar{z}_y) \tag{21} \\
 & + G_{xx}G_{yy}\phi_{xy}b_{xy}b_{yx}\text{Var}(\bar{z}_y) + G_{xx}G_{yy}\phi_{yx}b_{yx}b_{xy}\text{Var}(\bar{z}_x) + \\
 & 2G_{xx}^2\phi_{yx}^2b_{xy}^2\text{Cov}(\bar{z}_x, \bar{z}_y) + 2G_{yy}^2\phi_{xy}^2b_{yx}^2\text{Cov}(\bar{z}_x, \bar{z}_y) + \mathcal{V}].
 \end{aligned}$$

465

466 This equation again shows that when IIGEs are present, any variance across populations in  
 467 the trait mean of either species leads to a cross-species covariance in the response to selection  
 468 (terms 2, 3, 5, and 6). Cross species covariance in selection response is also mediated further  
 469 by covariance in trait means in this model (terms 1, 4, 7, and 8) as well as higher order  
 470 products of IIGEs captured in  $\mathcal{V}$  (equation 11). These effects may lead trait means to become  
 471 correlated across populations in future generations. This analysis, particularly contrasting  
 472 equations 16 vs. 20 and 17 vs. 21, also reveals that coevolutionary dynamics can be  
 473 substantially different even across models that share a common polynomial relative fitness  
 474 function.

475

## 476 Discussion

477 Our model adapts the theory of trait-based intraspecific social evolution to the phenotypic  
 478 interface between two coevolving species. We show that two forms of interspecific  
 479 interaction, interspecific indirect genetic effects (IIGEs) and cross-species selection  
 480 (analogous to within-species social selection), both contribute to correlated evolution

481 between interacting species. Our analysis shows that reciprocal IIGEs modulate selection  
482 response, suggesting that IIGEs may play a major role in generating and mediating patterns  
483 of correlated evolution between species. Further, we show that constant (across space) IIGEs  
484 can generate a coevolutionary covariance even in the absence of covariance in selection, or  
485 even the absence of genetic variance in one species. When selection does covary between  
486 species across populations, reciprocal IIGEs will promote changes in the magnitude of  
487 coevolution and even reversals in the expected among-population covariance. IIGEs also  
488 allow cross-species selection, which we model as the effect of the traits of one species on the  
489 fitness of another, to influence evolutionary response. Such a response may also be mediated  
490 by cross-species genetic assortment between interacting individuals. When IIGEs and cross-  
491 species selection act together, effects on the coevolutionary covariance can be complex, with  
492 dramatic changes in the expected sign and magnitude of correlated evolution occurring with  
493 subtle changes in these parameters of interspecific social interaction. Our results indicate that  
494 whenever coevolving species socially interact to modify expression of one another's  
495 phenotypes, these interspecific social interactions are key to understanding coevolution.

496 IIGEs represent a scenario where phenotypes in individuals of one species influence  
497 trait expression of individuals of another species. Thus, IIGEs are a specific type of  
498 environmental effect (Moore et al. 1997, Drown and Wade 2014) where the environment is  
499 the phenotypic value of the interspecific individual(s) with which an organism interacts. To  
500 our knowledge, this type of environmental effect on between-species coevolution has been  
501 considered in only two other theoretical studies (Scheiner et al. 2015, O'Brien et al. 2021; but  
502 see Shuster et al. 2006, Witham et al. 2020 for a variance-partitioning approach). Scheiner et  
503 al. (2015) consider a special case of our model, where evolvable IIGEs are present in only  
504 one of the two interacting species. In this non-reciprocal model, they show a much more  
505 limited role for IIGEs in coevolution. Our results are broadly consistent with this conclusion,

506 in that IIGEs in only a single species do not generate the reciprocal effects that lead to  
507 massive inflation of evolutionary response. However, IIGEs in only a single species (e.g.,  
508  $\Phi_{xy} = \mathbf{0}$ ,  $\Phi_{yx} \neq \mathbf{0}$ ) still play a role in mediating response to interspecific social selection  
509 whenever interspecific social selection is a function of individual trait values (as opposed to  
510 the population mean, as modeled by Scheiner et al. 2015). We also note that our fully  
511 multivariate model accommodates the possibility that reciprocal IIGEs act across different  
512 types or numbers of traits in the two interacting species. More recently, O'Brien et al. (2021;  
513 see also Queller 2014) developed a model of coevolution between host plant and microbial  
514 symbionts. Their parameterization differed from ours in that they consider evolution of a  
515 single joint trait governed by genetic variation in host and symbiont, and so is most directly  
516 applicable to plant-microbe systems or other intimate interactions. Nonetheless, their model  
517 shows an important role for reciprocal fitness feedbacks, consistent with the conclusions of  
518 our trait-based model.

519         Within-species models of interacting phenotypes clarify the line between genetic and  
520 environmental effects, furthering an understanding of how genotypes expressed in an  
521 individual that act as environments for other individuals can influence phenotypic change  
522 (Wolf et al. 1998, Wolf 2003). That is, IIGEs are genetically-based environments that  
523 influence phenotypic expression during interactions. Conceptually, this relationship is similar  
524 to genetically based plasticity. Extending these types of models to the case of interspecific  
525 interaction carries similar challenges and benefits. We have referred to reciprocal change in  
526 phenotypic means between interacting species as “coevolution,” even when these effects are  
527 mediated by IIGEs. This is a broad use of the term coevolution, as IIGEs are an  
528 environmental effect that can themselves evolve, again similar in concept to phenotypic  
529 plasticity. However, a critical difference is that changes in phenotypic response mediated by  
530 IIGEs represent changes driven by evolution of an interacting species (made clear in equation

531 4). Our point is not to broaden the definition of coevolution, but rather to highlight that  
532 IIGEs can have substantial impact on patterns of phenotypic divergence in coevolving  
533 species. For example, our model highlights that genetic divergence across populations of a  
534 single species is sufficient to generate tightly-coupled patterns of correlated change in an  
535 interacting species, a result that suggests the challenges of interpreting correlated phenotypes  
536 as evidence of genetic response to reciprocal selection may be even greater than already  
537 appreciated (e.g. Nuismer et al. 2010, Gomulkiewicz et al 2007, Janzen 1980).

538 Interspecific indirect genetic effects, or at least the potential for a prevalence of such  
539 effects, appear to be commonplace in many biological systems. In Table 1, we provide in a  
540 breakdown of types of biological interaction in which there is a large literature suggesting  
541 importance of IIGE-like phenomena. These types of effects on trait expression across  
542 species, widely appreciated in their own specific contexts (Weis and Abrahamson 1986,  
543 Peacor and Werner 2001, Werner and Peacor 2003, Chen 2008, Thomas et al. 2012, O'Brien  
544 et al. 2021), have taken on a variety of different forms. We suggest that these disparate  
545 biological phenomena may nonetheless share a commonality—reciprocal effects on trait  
546 expression across interspecific partners—that we have shown can affect the coevolutionary  
547 process in dramatic, and in some cases predictable, ways.

548 Cross-species selection features prominently in verbal descriptions of the  
549 coevolutionary process (Thompson 1982), and we show that such selection is especially  
550 important in the presence of interspecific indirect genetic effects. When individual trait  
551 values of one species affect individual fitness of another, focal species, this cross-species  
552 selection can manifest evolutionary change in the focal species when there is phenotypic  
553 assortment between interspecific interactants. This assortment, analogous to that required for  
554 evolutionary response to social selection within species (Wolf et al. 1999, McGlothlin et al.  
555 2010, Brodie et al. submitted), can be generated directly by a non-random genetic assortment,

556 or via IIGEs. Examples of processes that could generate direct genetic assortment between  
557 interacting individuals of two different species include shared genetic structure, to the extent  
558 that such structure manifests assortment of breeding values for the relevant traits. Such  
559 shared genetic structure could arise through shared features of the environment that limit  
560 gene flow and panmictic mating in both species, or alternatively, through variation in habitat  
561 preference across individuals of both species. Direct genetic assortment could also arise  
562 through behavioral preference for certain trait values in heterospecific partners. Such  
563 preferences may be especially common in predator-prey interactions, where, for example,  
564 predator body size may be expected to coevolve with behavioral preference for prey size  
565 (Troost et al. 2008). Currently, it is unclear how common this type of cross-species genetic  
566 assortment may be, although in part this likely reflects a lack of studies that have attempted to  
567 measure assortment between breeding values of individuals of separate species. Moreover,  
568 when it does occur, such assortment is likely to be transient because it does not rely on  
569 transmission of pleiotropic alleles that may stabilize within-species genetic correlations over  
570 multiple generations. The substantial evidence for IIGEs, but limited evidence of  $G_{xy}$   
571 suggests that IIGEs may play a prominent role in mediating any realized response to cross-  
572 species selection.

573         Our results also indicate that nonlinear effects on cross-species selection can  
574 contribute to coevolution even in the absence of genetic assortment or IIGEs. This form of  
575 selection corresponds to an interaction between focal and interspecific-partner trait values for  
576 focal individual fitness. The effect of this form of cross-species selection on evolutionary  
577 response in a population depends on the mean genotype of the other species, and thus  
578 represents a diffuse effect of population mean phenotype of the coevolving species. Such  
579 interspecific interactions are potentially less intimate, for example diffuse predator chemical  
580 cues in aquatic environments, than the individual level interactions (e.g., of host and parasite)

581 required to generate response from linear cross-species selection. Across populations,  
582 nonlinear cross-species selection contributes to coevolution via covariance in mean genetic  
583 values and/or linear selection between the species. This result is consistent with past models  
584 of coevolution, verbal and mathematical, that indicate trait interactions for fitness are a key  
585 feature of coevolution (Thompson 1982, 1994, 2005, Nuismer 2017), and in our model, such  
586 interactions lead to a dependence between selection in one species and the mean trait value of  
587 another. By defining these interaction terms in the framework of social evolution, our model  
588 adds to past work by indicating that reciprocal IIGEs can substantially increase the degree to  
589 which trait interactions for fitness contribute to reciprocal evolutionary change.

590         A key feature of our model is the development of a formal expression for the expected  
591 covariance in evolutionary response,  $\text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y)$ . This coevolutionary covariance is  
592 expected to be a key contributor to generating among-population covariation in species mean  
593 trait values, a major focus in many empirical (Thompson 1994, 1995, Zangerl and  
594 Berenbaum 2003, Toju and Sota 2005, Hanifin et al. 2008, Hague et al. 2020) and theoretical  
595 (Nuismer et al. 2010, Nuismer and Week 2019, Week and Nuismer 2019) studies of  
596 coevolution. Importantly, similar to existing within-species models of among-population  
597 quantitative genetic variation (Zeng 1988, Chenoweth et al. 2010), defining this  
598 coevolutionary covariance illustrates how selection, IIGEs, and genetics may contribute to  
599 patterns of trait variation across populations.

600         Our model subsumes mechanistic detail into broad statistical descriptions of species  
601 interactions and thus provides a general description of how IIGEs and cross-species selection,  
602 when present, contribute to reciprocal evolutionary change and correlated evolution across  
603 populations. In contrast to our approach, some models of coevolution have focused instead  
604 on specific ecological mechanisms that generate trait-fitness relationships between interacting  
605 species (reviewed in Nuismer 2017). By highlighting the key parameters that contribute to

606 coevolution—covariance in natural selection, covariance in cross-species selection, and  
607 IIGEs—our model indicates various pathways through which specific ecological mechanisms  
608 may affect coevolution. Our framework can be tailored to specific scenarios by substituting  
609 different fitness models into the general equations we present here.

610 Our model generates quantitative predictions for the shape of coevolution that are  
611 directly testable with empirical data because it focuses on estimable statistical effects of  
612 underlying ecological mechanisms rather than the mechanisms themselves, which are often  
613 unknown (Wade and Kalisz 1990). For example, using an empirical estimate of  $\Phi$  (which  
614 could be measured using methods analogous to those used to measure within-species indirect  
615 genetic effects; Bleakley and Brodie 2009, McGlothlin and Brodie 2009), one could use  
616 matrix comparison of covariances among population means and the covariance terms  
617 presented here to quantitatively test the contribution of IIGEs to among-population  
618 covariance in selection response between two interacting species (e.g., see Chenoweth et al.  
619 2010 for a within-species test of the predictions of Zeng's 1988 model). More generally,  
620 Week and Nuismer (2019; see also Nuismer and Week 2019) have shown how datasets of  
621 among-population variation in trait means can be used to test for conformation to  
622 expectations from coevolutionary models. Concomitantly, our models show how  
623 environmental effects can be partitioned into terms describing genotypes of other species in  
624 the ecological community, which could be useful in understanding when and why  
625 evolutionary response fails to conform to predictions arising from the standard breeder's  
626 equation.

627 Social interactions between individuals of the same species play a central role in the  
628 evolutionary process. Within a single lineage, indirect genetic effects and social selection  
629 fundamentally change selection response, the expression of genetic variance, and together  
630 determine the course of social evolution (Moore et al. 1997, Wolf et al. 1998, Wolf et al.

631 1999, McGlothlin et al. 2010). We have shown that these effects of interactions among  
632 individuals may transcend species boundaries and profoundly impact the dynamics of  
633 coevolution between interacting lineages.

## 634 Literature Cited

- 635 Abrahamson, W. G., and A. E. Weis. 1997. Evolutionary Ecology Across Three Trophic  
636 Levels: Goldenrods, Gallmakers, and Natural Enemies. Princeton University Press  
637 Princeton.
- 638 Abrams, P. A., Y. Harada, and H. Matsuda. 1993. On the relationship between quantitative  
639 genetic and ESS models. *Evolution* 47:982-985.
- 640 Agrawal, A. A. 2001. Phenotypic plasticity in the interactions and evolution of species.  
641 *Science* 294:321-326.
- 642 Arnold, S. J., M. E. Pfrender, and A. G. Jones. 2001. The adaptive landscape as a conceptual  
643 bridge between micro- and macroevolution. *Genetica* 112/113:9-32.
- 644 Arnqvist, G., and L. Rowe. 2002. Antagonistic coevolution between the sexes in a group of  
645 insects. *Nature* 415:787-789.
- 646 Barribeau, S. M. 2014. Gene expression differences underlying genotype-by-genotype  
647 specificity in a host-parasite system. *Proceedings of the National Academy of Sciences*  
648 111:3496-3501.
- 649 Bijma, P., and M. J. Wade. 2008. The joint effects of kin, multilevel selection and indirect  
650 genetic effects on response to genetic selection. *Journal of Evolutionary Biology*  
651 21:1175-1188.
- 652 Bleakley, B. H., and E. D. Brodie III. 2009. Indirect genetic effects influence antipredator  
653 behavior in guppies: Estimates of the coefficient of interaction  $\psi$  and the inheritance  
654 of reciprocity. *Evolution* 63:1796-1806.
- 655 Boersma, M., P. Spaak, and L. De Meester. 1998. Predator-mediated plasticity in  
656 morphology, life history, and behavior of *Daphnia*: the uncoupling of responses.  
657 *American Naturalist* 152:237-248.
- 658 Borell, E. M., A. Foggo, and R. A. Coleman. 2004. Induced resistance in intertidal  
659 macroalgae modifies feeding behavior of herbivorous snails. *Oecologia* 140:328-334.
- 660 Brodie, E. D., III, and E. D. Brodie, Jr. 1999. Predator-prey arms races. *BioScience* 49:557-  
661 568.
- 662 Brodie, E. D. Jr., B. J. Ridenhour, and E. D. Brodie III. 2002. The evolutionary response of  
663 predators to dangerous prey: hotspots and coldspots in the geographic mosaic of  
664 coevolution between garters snakes and newts *Evolution* 56:2067-2082.
- 665 Brodie, E. D., III and B. J. Ridenhour 2003 Reciprocal selection at the phenotypic interface  
666 of coevolution. *Integrative and Comparative Biology* 43:408-418
- 667 Brodie, E.D., III, P. A. Cook, R. A. Costello, V. A. Formica. Submitted. Phenotypic  
668 assortment changes the landscape of selection. *Journal of Heredity*.
- 669 Chen, M.-S. 2008. Inducible direct plant defense against insect herbivores: A review. *Insect*  
670 *Science* 15:101-114.
- 671 Chenoweth, S. F., H. D. Rundle, and M. W. Blows. 2010. The contribution of selection and  
672 genetic constraints to phenotypic divergence. *American Naturalist* 175:186-196.
- 673 Damian, R. T. 1997. Parasite immune evasion and exploitation: reflections and projections.  
674 *Parasitology* 115:S169-175.
- 675 Dawkins, R. 1982. *The Extended Phenotype*. Oxford Univ. Press, Oxford.
- 676 Drown, D. M., and M. J. Wade. 2014. Runaway coevolution: adaptation to heritable and  
677 nonheritable environments. *Evolution* 68:3039-3046.
- 678 Dybdahl, M. F., C. E. Jenkins, and S. L. Nuismer. 2014. Identifying the molecular basis of  
679 host-parasite coevolution: merging models and mechanisms. *American Naturalist*  
680 184:1-13.
- 681 Ehrlich, P. R., and P. H. Raven. 1964. Butterflies and plants: a study in coevolution.  
682 *Evolution* 18:586-608.

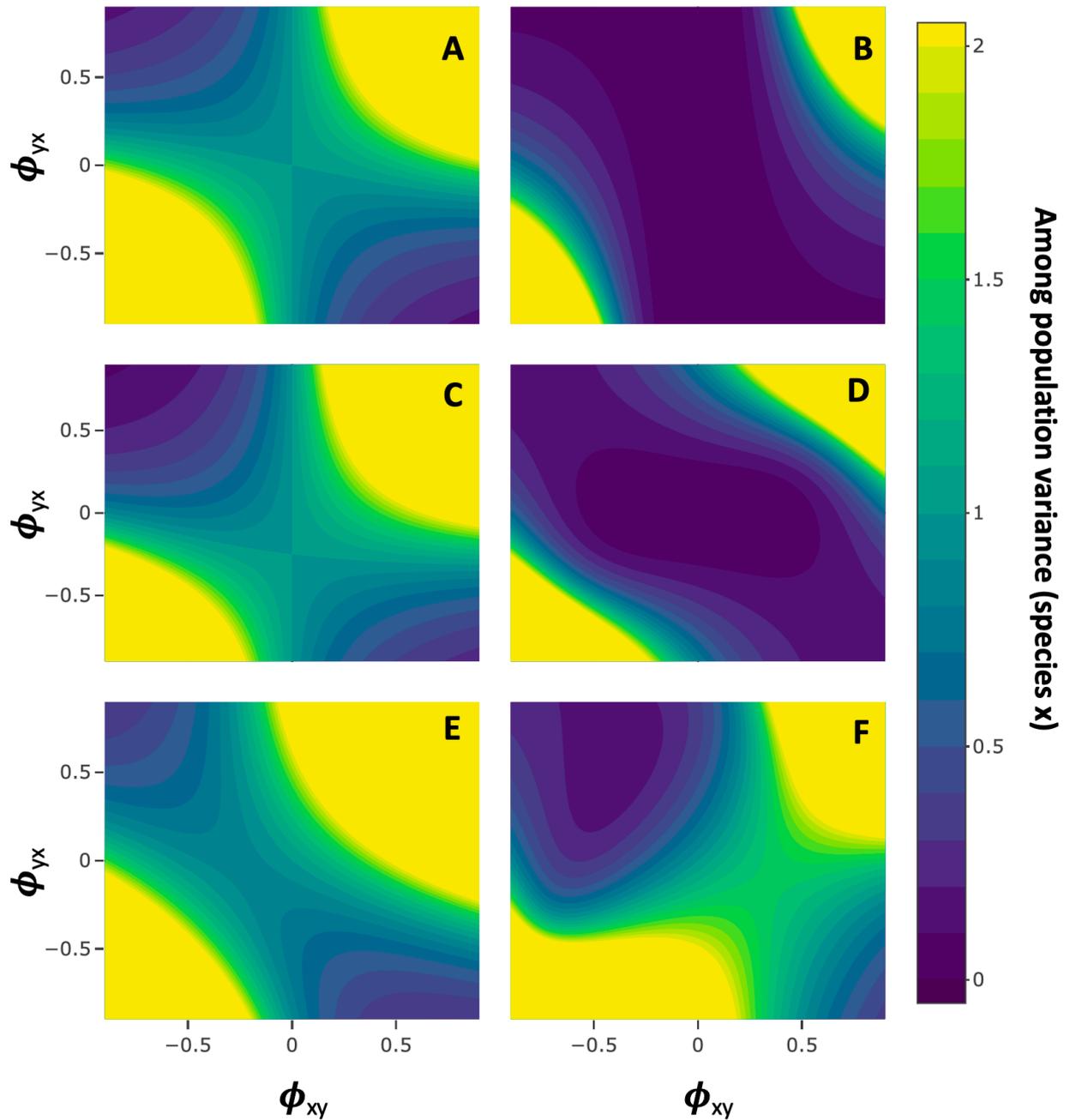
- 683 Fisher, D.N. and A. McAdam. 2019. Indirect genetic effects clarify how traits can evolve  
684 even when fitness does not. *Evolution Letters* 3:4-14.
- 685 Foster, K. R., J. Schluter, K. Z. Coyte, and S. Rakoff-Nahoum. 2017. The evolution of the  
686 host microbiome as an ecosystem on a leash. *Nature* 548:43-51.
- 687 Friesen, M. L., S. S. Porter, S. C. Stark, E. J. von Wettberg, J. L. Sachs, and E. Martinez-  
688 Romero. 2011. Microbially mediated plant functional traits. *Annual Review of*  
689 *Ecology and Systematics* 42:23-46.
- 690 Fuess, L., J. N. Weber, S. den Haan, N. C. Steinel, K. C. Shim, and D. I. Bolnick. 2020. A  
691 test of the Baldwin effect: Differences in both constitutive expression and inducible  
692 responses to parasites underlie variation in host response to a parasite. *bioRxiv*  
693 <https://doi.org/10.1101/2020.07.29.216531>.
- 694 Gianinazzi-Pearson, V., N. Séjalon-Delmas, A. Genre, S. Jeandroz, and P. Bonfante. 2007.  
695 Plants and arbuscular mycorrhizal fungi: cues and communication in the early steps of  
696 symbiotic interactions. *Advances in Botanical Research* 46:181-219.
- 697 Gomulkiewicz, R., D.M. Drown, M.F. Dybdahl, W. Godsoe, S.L. Nuismer, K.M. Pepin, B.J.  
698 Ridenhour, C.I. Smith, and J.B. Yoder. 2007. Dos and don'ts of testing the geographic  
699 mosaic theory of coevolution. *Heredity* 98:249-258.
- 700 Hague, M. T. J., A. N. Stokes, C. R. Feldman, E. D. Brodie Jr., and E. D. Brodie III. 2020.  
701 The geographic mosaic of arms race coevolution is closely matched to prey  
702 population structure. *Evolution Letters* 164:1567-16.
- 703 Hanifin, C. T., E. D. Brodie Jr., and E. D. Brodie III. 2008. Phenotypic mismatches reveal  
704 escape from arms-race coevolution. *PLoS Biology* 6:e60.
- 705 Janzen, D. H. 1980. When is it coevolution? *Evolution* 34:611-612.
- 706 Johnson, S. D., and B. Anderson. 2010. Coevolution between food-rewarding flowers and  
707 their pollinators. *Evolution: Education and Outreach* 3:32-39.
- 708 Kirkpatrick, M. and R. Lande 1989. The evolution of maternal characters. *Evolution* 43:485-  
709 503.
- 710 Korgaonkar, A., C. Han, A. L. Lemire, I. Siwanowicz, D. Bennouna, R. E. Kopec, P.  
711 Andolfatto, S. Shigenobu, and D.L. Stern. 2021. A novel family of secreted insect  
712 proteins linked to plant gall development. *Current Biology* 31:1-14
- 713 Lande, R. 1979. Quantitative genetic analysis of multivariate evolution, applied to brain:  
714 body size allometry. *Evolution* 33:402-416.
- 715 Lande, R., and S. J. Arnold. 1983. The measurement of selection on correlated characters.  
716 *Evolution* 37:1210-1226.
- 717 McAdam, A.G., S. Boutin, D. Réale, and D. Berteaux. 2002. Maternal effects and the  
718 potential for evolution in a natural population of animals. *Evolution* 56: 846-851.
- 719 McGlothlin, J. W., and E. D. Brodie III. 2009. How to measure indirect genetic effects: the  
720 congruence of trait-based and variance-partitioning approaches. *Evolution* 63:1785-  
721 1795.
- 722 McGlothlin, J. W., E. Akçay, E. D. Brodie III, A. J. Moore, and J. Van Cleve. 2021. A  
723 synthesis of game theory and quantitative genetic models of social evolution. *bioRxiv*  
724 <https://doi.org/10.1101/2021.03.27.437341>
- 725 McGlothlin, J. W., A. J. Moore, J. B. Wolf, and E. D. Brodie III. 2010. Interacting  
726 phenotypes and the evolutionary process. III. Social evolution. *Evolution* 64:2558-  
727 2574.
- 728 Moore, A. J., E. D. Brodie III, and J. B. Wolf. 1997. Interacting phenotypes and the  
729 evolutionary process: I. Direct and indirect genetic effects of social interactions.  
730 *Evolution* 51:1352-1362.
- 731 Mousseau, T.A. and C.W. Fox. 1998. The adaptive significance of maternal effects. *Trends in*  
732 *Ecology and Evolution* 13:403-407

- 733 Nuismer, S. L. 2017. Introduction to Coevolutionary Theory. W.H. Freeman and Company  
734 New York
- 735 Nuismer, S. L., B. J. Ridenhour, and B. P. Oswald. 2007. Antagonistic coevolution mediated  
736 by phenotypic differences between quantitative traits. *Evolution* 61:1823-1834.
- 737 Nuismer, S.L., R. Gomulkiewicz, and B.J. Ridenhour. 2010. When is correlation coevolution.  
738 *The American Naturalist* 175:525-537.
- 739 Nuismer, S.L., and B. Week. 2019. Approximate Bayesian estimation of coevolutionary arms  
740 races. *PLoS Computational Biology* 15:e1006988.
- 741 Ohgushi, T. 2005. Indirect interaction webs: Herbivore-induced effects through trait change  
742 in plants *Annual Review of Ecology, Evolution, and Systematics* 36:81-105.
- 743 Oladiran, A., and M. Belosevic. 2012. Immune evasion strategies of trypanosomes: a review.  
744 *Journal of Parasitology* 98:284-292.
- 745 O'Brien, A. M., C. N. Jack, M. L. Friesen, and M. E. Frederickson. 2021. Whose trait is it  
746 anyways? Coevolution of joint phenotypes and genetic architecture in mutualisms.  
747 *Proceedings of the Royal Society of London Series B.* 288:20202483.
- 748 Peacor, S. D., and E. E. Werner. 2001. The contribution of trait-mediated indirect effects to  
749 the net effects of a predator. *Proceedings of the National Academy of Sciences*  
750 98:3904-3908.
- 751 Pellmyr, O. 2003. Yuccas, yucca moths, and coevolution: A review. *Annals of the Missouri*  
752 *Botanical Garden* 90:35-55.
- 753 Phillips, P.C., and S.J. Arnold. 1989. Visualizing multivariate selection. *Evolution* 43:1209-  
754 1222.
- 755 Preisser, E. L., D. I. Bolnick, and M. F. Benard. 2005. Scared to death? The effects of  
756 intimidation and consumption in predator-prey interactions. *Ecology* 86:501-509.
- 757 Price, G. R. 1970. Selection and covariance. *Nature* 227:520-521.
- 758 Price, G. R. 1972. Extension of covariance mathematics. *Annals of Human Genetics* 35:485-  
759 490.
- 760 Queller, D. C. 2014. Joint phenotypes, evolutionary conflict and the fundamental theorem of  
761 natural selection. *Philosophical Transactions of the Royal Society B.* 369:20130423.
- 762 Ridenhour, B. J. 2005. Identification of selective sources: Partitioning selection based on  
763 interactions. *The American Naturalist* 166:12-25.
- 764 Robertson, A. 1966. A mathematical model of the culling process in dairy cattle. *Animal*  
765 *Production* 8:95-108.
- 766 Scharsack, J. P., A. Gossens, F. Franke, and J. Kurtz. 2013. Excretory products of the  
767 cestode, *Schistocephalus solidus*, modulate in vitro responses of leukocytes from its  
768 specific host, the three-spined stickleback (*Gasterosteus aculeatus*). *Fish and*  
769 *Shellfish Immunology* 35:1779-1787.
- 770 Scharsack, J. P., M. Kalbe, R. Derner, J. Kurtz, and M. Milinski. 2004. Modulation of  
771 granulocyte responses in three-spined sticklebacks *Gasterosteus aculeatus* infected with  
772 the tapeworm *Schistocephalus solidus*. *Diseases of Aquatic Organisms* 59:141-150.
- 773 Scharsack, J. P., K. Koch, and K. Hammerschmidt. 2007. Who is in control of the stickleback  
774 immune system: interactions between *Schistocephalus solidus* and its specific  
775 vertebrate host. *Proceedings of the Royal Society of London Series B.* 274:3151-  
776 3158.
- 777 Scheiner, S. M., R. Gomulkiewicz, and R. D. Holt. 2015. The genetics of phenotypic  
778 plasticity. XIV. Coevolution. *American Naturalist* 185:594-609.
- 779 Schmid-Hempel, P. 2008. Parasite immune evasion: a momentous molecular war. *Trends in*  
780 *Ecology and Evolution* 23:318-326.

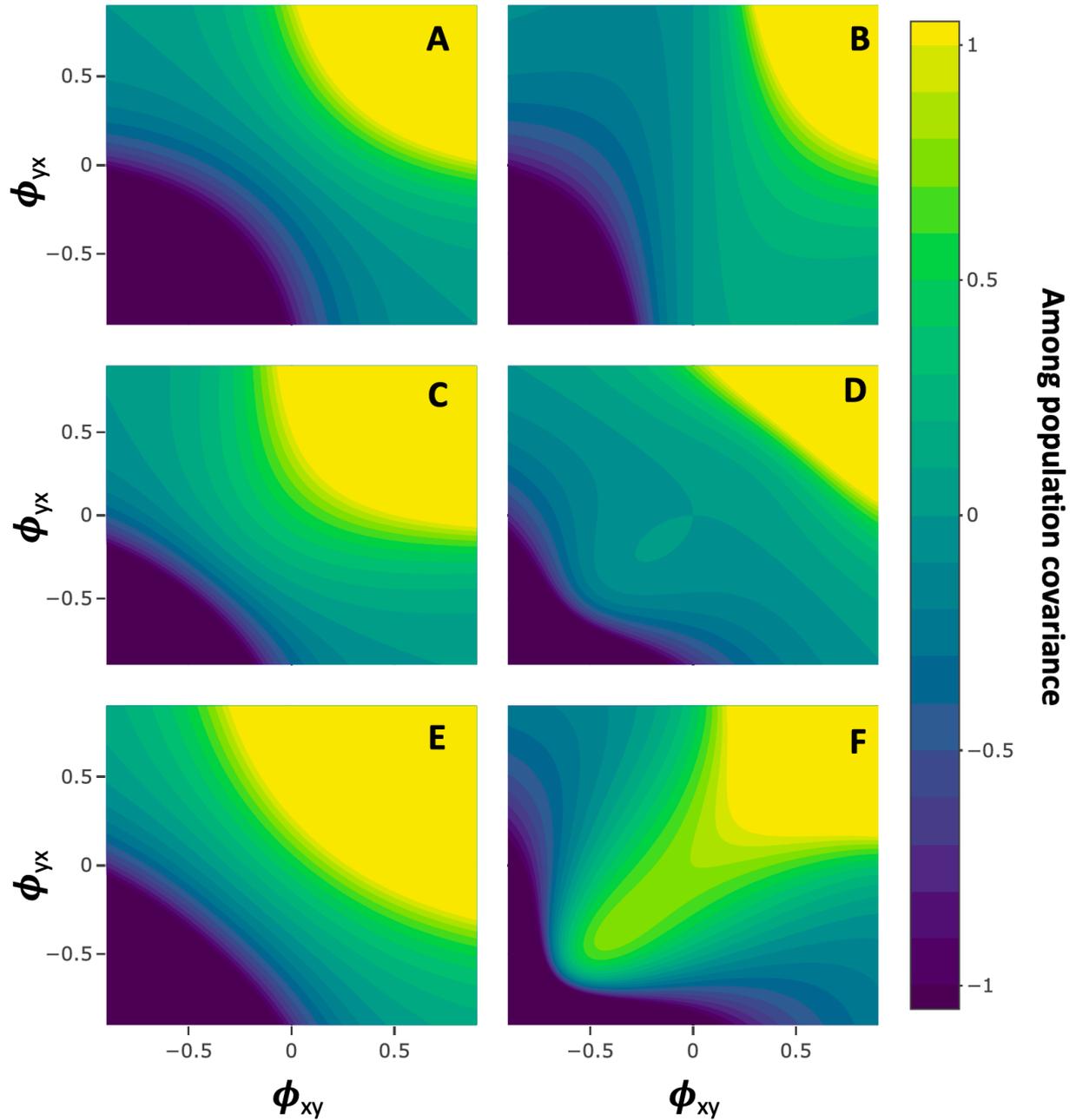
- 781 Shuster, S. M., E. V. Lonsdorf, G. M. Wimp, J. K. Bailey, and T. G. Whitham. 2006.  
782 Community heritability measures the evolutionary consequences of indirect genetic  
783 effects on community structure. *Evolution* 60:991-1003.
- 784 Siepielski, A. M., K. M. Gotanda, M. B. Morrissey, S. E. Diamond, J. D. DiBattista, and S.  
785 M. Carlson. 2013. The spatial patterns of directional phenotypic selection. *Ecology*  
786 *Letters* 16:1382-1392.
- 787 Stappenbeck, T. S., and H. W. Virgin. 2016. Accounting for reciprocal host-microbiome  
788 interactions in experimental science. *Nature* 534:191-199.
- 789 Stearns, S. C. 2012. Evolutionary routes leading to host manipulation by parasites: Afterward  
790 *in* F. Thomas, J. Brodeur, and D. P. Hughes, editors. *Host manipulation by parasites*.  
791 Oxford University Press, Oxford.
- 792 Tams, V., J. H. Nickel, A. Ehring, and M. Cordellier. 2019. Insights into the genetic basis of  
793 predator-induced response in *Daphnia* - a comparative transcriptomic approach.  
794 bioRxiv. <https://doi.org/10.1101/503904>
- 795 Thomas, F., J. Brodeur, and D. P. Hughes. 2012. *Host manipulation by parasites* Oxford  
796 University Press, Oxford.
- 797 Thompson, J. N. 1982. *Interaction and Coevolution*. University of Chicago Press, Chicago.
- 798 Thompson, J. N. 1994. *The Coevolutionary Process*. University of Chicago Press, Chicago.
- 799 Thompson, J. N. 2005. *The Geographic Mosaic of Coevolution*. University of Chicago Press,  
800 Chicago.
- 801 Toju, H., and T. Sota. 2005. Imbalance of predator and prey armament: geographic clines in  
802 phenotypic interface and natural selection. *American Naturalist* 167:105–117
- 803 Troost, T. A., B. W. Kooi, and U. Dieckmann. 2008. Joint evolution of predator body size  
804 and prey size preference. *Evolutionary Ecology* 22:771-799.
- 805 Urban, M.C. 2011. The evolution of species interactions across natural landscapes. *Ecology*  
806 *Letters* 14:723-732.
- 807 Wade, M. J., and Kalisz. 1990. The causes of natural selection. *Evolution* 44:1947-1955.
- 808 Walsh, M. R., and D. M. Post. 2011. The impact of intraspecific variation in a fish predator on  
809 the evolution of phenotypic plasticity and investment in sex in *Daphnia ambigua*.  
810 *Journal of Evolutionary Biology* 25:80-89.
- 811 Weber, A., and S. Declerck. 1997. Phenotypic plasticity of *Daphnia* life history traits in  
812 response to predator kairomones: genetic variability and evolutionary potential  
813 *Hydrobiologia* 360:89-99.
- 814 Weber, J. N., N. C. Steinel, K. C. Shim, and D. I. Bolnick. 2017. Recent evolution of extreme  
815 cestode growth suppression by a vertebrate host. *Proceedings of the National*  
816 *Academy of Sciences* 114:6575-6580.
- 817 Week, B. and S.L. Nuismer. 2019. The measurement of coevolution in the wild. *Ecology*  
818 *Letters* 22:717-725
- 819 Weis, A. E., and W. G. Abrahamson. 1986. Evolution of host-plant manipulations by gall  
820 makers: ecological and genetic factors in the Solidago-Eurosta system. *American*  
821 *Naturalist* 127:681-695.
- 822 Weis, A.E., W.G. Abrahamson, and M.C. Anderson. 1992. Variable selection on Eurosta's  
823 gall size, I: The extent and nature of variation in phenotypic selection. *Evolution*  
824 46:1674-1697.
- 825 Werner, E. E., and S. D. Peacor. 2003. A review of trait-mediated indirect interactions in  
826 ecological communities. *Ecology* 84:1083-1100.
- 827 West-Eberhard, M. J. 1979. Sexual selection, social competition, and evolution. *Proceedings*  
828 *of the American Philosophical Society* 123:222-234.
- 829 West-Eberhard, M. J. 1983. Sexual selection, social competition, and speciation. *Quarterly*  
830 *Review of Biology* 58:155-183.

- 831 West-Eberhard, M. J. 1984. Sexual selection, competitive communication and species-  
832 specific signals in insects. Pages 283-342 *in* T. Lewis, editor. Insect communication.  
833 Academic Press, New York.
- 834 Whitham, T. H., G. J. Allan, H. F. Cooper, and S. M. Shuster. 2020. Intraspecific genetic  
835 variation and species interactions contribute to community evolution. *Annual Review*  
836 *of Ecology and Systematics* 51:587-612.
- 837 Wolf, J. B., E. D. Brodie III, J. M. Cheverud, A. J. Moore, and M. J. Wade. 1998.  
838 Evolutionary consequences of indirect genetic effects. *Trends in Ecology and*  
839 *Evolution* 13:64-69.
- 840 Wolf, J. B., E. D. Brodie III, and A. J. Moore. 1999. Interacting phenotypes and the  
841 evolutionary process. II. Selection resulting from social interactions. *American*  
842 *Naturalist* 153:254-266.
- 843 Wolf, J.B. 2003. Genetic architecture and evolutionary constraint when the environment  
844 contains genes. *Proceedings of the National Academy of Sciences* 100:4655-4660.
- 845 Zangerl, A. R., and M. R. Berenbaum. 2003. Phenotype matching in wild parsnip and parsnip  
846 webworms: causes and consequences. *Evolution* 57:806–815
- 847 Zeng, Z.-B. 1988. Long-term correlated response, interpopulation covariation, and  
848 interspecific allometry. *Evolution* 42 363-374.
- 849 Züst, T., S. Mou, and A.A. Agrawal. 2018. What doesn't kill you makes you stronger: The  
850 burdens and benefits of toxin sequestration in a milkweed aphid. *Functional Ecology*  
851 8:1972-1981

## Supplemental Figures



**Figure S1.** IIGEs accelerate evolutionary rate in a single species. Panels show the evolutionary rate,  $\text{Var}(\Delta\bar{z}_x)$ , of species  $x$  as a function of the indirect genetic effect parameters  $\phi$ , under the same parameter values as in Figure 3. Reciprocal IIGEs between interacting species generally accelerate evolutionary rate. Note that in the absence of any other effects, the evolutionary rate is equal to the variance in natural selection, which is unity in Panels A, C, E, and F. In panel D, evolutionary rate is driven entirely by cross-species selection and IIGEs. In panel B, where  $G_{xx}=0$ , evolutionary rate in species  $x$  is driven entirely by reciprocal IIGEs and evolutionary change in species  $y$ .



**Figure S2.** Among population covariances. Panels show the coevolutionary covariance between two interacting species as a function of the indirect genetic effect parameters  $\phi$ , from equation (7) unstandardized. Parameter values are as in Figs. 4 and S1. For all panels, genetic variances were set to unity.

852 **Appendix**

853 **A1. Phenotypic covariance between interacting species**

854 For a single trait, we can solve for the covariance between  $z_y$  and  $z_x$  to partition the  
 855 phenotypic covariance between individuals of two interacting species into terms describing  
 856 the contribution of IIGEs and terms describing non-random genetic assortment. Assuming  
 857 cross-species environmental covariance is zero,

858

$$C_{xy} = (1 - \phi_{yx}\phi_{xy})^{-2}[(1 + \phi_{yx}\phi_{xy})G_{xy} + \phi_{yx}(G_x + E_x) + \phi_{xy}(G_{yy} + E_x)]$$

859

860 where  $E_x$  and  $E_y$  represent within-species environmental variance and  $G_{xy} = G_{yx}$ . When  
 861 IIGEs are absent, non-random genetic assortment  $G_{xy}$  is the sole contributor to the  
 862 phenotypic association between individuals of coevolving species. When IIGEs are present,  
 863 they can substantially change this phenotypic association.

864

865 **A2. Covariance in selection response with nonzero IIGEs and genetic assortment**

866 We expand the covariance in evolutionary response in two species when IIGEs are present  
 867 and constant and when genetic assortment  $G_{xy} = G_{yx}$  is present and constant,

868

$$\begin{aligned} \text{Cov}(\Delta\bar{z}_x, \Delta\bar{z}_y) = & \mathcal{U}[\mathcal{A}^2\phi_{yx}\text{Var}(\beta_{xx}) + \mathcal{B}^2\phi_{yx}\text{Var}(\beta_{xy}) + \mathcal{C}^2\phi_{xy}\text{Var}(\beta_{yy}) \\ & + \mathcal{D}^2\phi_{xy}\text{Var}(\beta_{yx}) + \mathcal{A}\mathcal{B}\mathcal{E}\text{Cov}(\beta_{xx}, \beta_{xy}) + \mathcal{A}\mathcal{C}\mathcal{E}\text{Cov}(\beta_{xx}, \beta_{yy}) \\ & + \mathcal{A}\mathcal{D}\mathcal{E}\text{Cov}(\beta_{xx}, \beta_{yx}) + \mathcal{B}\mathcal{C}\mathcal{E}\text{Cov}(\beta_{xy}, \beta_{yy}) + \mathcal{B}\mathcal{D}\mathcal{E}\text{Cov}(\beta_{xy}, \beta_{yx}) \\ & + \mathcal{C}\mathcal{D}\mathcal{E}\text{Cov}(\beta_{yy}, \beta_{yx})] \end{aligned}$$

873

874 where

875

876 
$$\mathcal{U} = (1 - \phi_{xy}\phi_{yx})^{-4}$$

877 
$$\mathcal{A} = G_{xx} + G_{xy}\phi_{xy}$$

878 
$$\mathcal{B} = G_{xy} + G_{xx}\phi_{yx}$$

879 
$$\mathcal{C} = G_{yy} + G_{xy}\phi_{yx}$$

880 
$$\mathcal{D} = G_{xy} + G_{yy}\phi_{xy}$$

881 
$$\mathcal{E} = 1 + \phi_{xy}\phi_{xy}$$

882