

In vitro and computational analysis of the putative furin cleavage site (RRARS) in the divergent spike protein of the rodent coronavirus AcCoV-JC34 (sub-genus luchacovirus)

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Abstract

The *Coronaviridae* is a highly diverse virus family, with reservoir hosts in a variety of wildlife species that encompass bats, birds and small mammals, including rodents. Within the taxonomic group alphacoronavirus, certain sub-genera (including the luchacoviruses) have phylogenetically distinct spike proteins, which remain essentially uncharacterized. Using in vitro and computational techniques, we analyzed the spike protein of the rodent coronavirus AcCoV-JC34 from the sub-genus luchacovirus, previously identified in *Apodemus chevrieri* (Chevrier's field mouse). We show that AcCoV-JC34—unlike the other luchacoviruses—has a putative furin cleavage site (FCS) within its spike S1 domain, close to the S1/S2 interface. The pattern of basic amino acids within the AcCoV-JC34 FCS (-RR-R-) is identical to that found in “pre-variant” SARS-CoV-2—which is in itself atypical for an FCS, and suboptimal for furin cleavage. Our analysis shows that, while containing an -RR-R- motif, the AcCoV-JC34 spike “FCS” is not cleaved by furin (unlike for SARS-CoV-2), suggesting the possible presence of a progenitor sequence for viral emergence from a distinct wildlife host.

Introduction

The animal reservoirs for pandemic potential viruses (including coronaviruses) are focused on the breadth of bat species (order Chiroptera) that exist around the world [1-3]. However certain coronaviruses, notably the sub-genus embecovirus (genus betacoronavirus) currently have no bat-origin examples and have a putative reservoir in animal species within the order Rodentia, which is the most diverse mammalian order on the planet and is well-documented as an important reservoir host for human diseases [4, 5].

While rodents are generally appreciated as an important reservoir for RNA viruses, surveillance and detection of coronaviruses is currently relatively limited. Following the initial discovery of what is now the prototype luchacovirus (Lucheng Rn rat coronavirus, or LRVN), along with two *Betacoronavirus* species [6], a study from Ge *et al.* examined 177 intestinal samples from three species of rodents in Yunnan Province, China and detected both alphacoronaviruses and betacoronaviruses in three animal species (*Apodemus chevrieri*, *Eothenomys fidelis* and *Apodemis ilex*) [7]. Their study reported the full-length genome of a coronavirus (AcCoV-JC34) from *A. chevrieri* (Chevrier's field mouse) that was designated an alphacoronavirus (sub-genus luchacovirus) based on its genome structure and multiple sequence alignments, which included analysis of the whole genome and the ORF1a/b genes. However, Ge *et al.* noted that both AcCoV-JC34 and LRVN may represent a novel alphacoronavirus species. In particular, they noted that the luchacovirus S gene formed a distinct genetic lineage with low sequence identity (<25%) compared to other well characterized coronaviruses. Ge *et al.* also noted that AcCoV-JC34 S contained two predicted

proteolytic cleavage sites, one at residue 508 at the S1/S2 interface, and the other at residue 674 (the fusion peptide-proximal S2' position).

More recently, a more comprehensive sampling of rodents and other small mammals has identified a diverse range of coronaviruses in such animal reservoirs [8]. To determine the evolutionary history of rodent alphacoronaviruses in more detail, Tsolteridis *et al.* also reported sequence data from viruses sampled from European rodents, to define a single common ancestor for all rodent alphacoronaviruses with a shared recombinant betacoronavirus spike gene—also shared with batCoV HKU2, swine acute diarrhea syndrome (SADS) coronavirus and two shrew coronaviruses [9]. According to Tsolteridis *et al.*, the luchacoviruses (including AcCoV-JC34) comprised a distinct lineage within the “recombinant” viruses. In summary, it can be argued that coronaviruses of small mammals, including rodents, are still poorly understood.

We have previously reported that rodent coronavirus AcCoV-JC34 has a weakly predicted furin cleavage site (FCS) in its spike protein [10]. Here, we further analyze the AcCoV-JC34 spike and its “FCS” along with the other luchacoviruses, taking an *in vitro* and computational perspective.

Results

Phylogenetic analysis of luchacoviruses

To understand the relationship of ACoV-JC34 and the other known luchacoviruses, we first constructed a phylogenetic tree of these viruses in comparison to representatives of the diverse coronavirus family, based on spike protein sequences (Figure 1). In agreement with Ge *et al.*,

luchacoviruses formed a monophyletic group with 100% bootstrap support, indicating a common ancestor origin outside of the established alphacoronavirus branch. Luchacoviruses clustered with rhinacoviruses, which include swine acute diarrhea syndrome coronavirus, *Rhinolophus* bat coronavirus HKU2, and porcine enteric alpha coronavirus (Figure 1).

Geographical distribution of sampled luchacoviruses

The geographical location, dates and rodent species sampled for the currently identified luchacoviruses are summarized in Figure 2 and Table 1. The luchacoviruses sampled to date are from a range of rodent hosts and are from the United Kingdom and several provinces in China (Figure 2), indicating a widespread distribution. Despite being sampled in these distinct locations, as mentioned above, luchacoviruses form a monophyletic group suggesting they have been associated with rodents for an extended period of time.

Multiple sequence alignment and structural analysis of AcCoV-JC34 spike

A multiple sequence alignment of spike proteins was performed on AcCoV-JC34 spike in comparison to the prototype luchacovirus Lucheng Rn rat CoV (LRNV), as well as SARS-CoV-2, SARS-CoV, HCoV-HKU1, HCoV-OC43 and MERS-CoV. This alignment revealed that the -RR-R- motif present in AcCoV-JC34 does not align precisely with the S1/S2 motif of most coronavirus spikes (Figure 3). However, it aligned with a potential secondary MERS-CoV furin cleavage site (RSTRS).

To investigate the structural location of AcCoV-JC34 furin cleavage site, the AcCoV-JC34 spike protein structure was structurally modeled (Figure 4). We used SADS-CoV spike for our modeling due to its available structure in the RCSB protein data bank and relatively high

identity with JC34 (41.5%). In our JC34 model, the potential furin cleavage site (-RR-R-) is located in an exposed loop of the protein which is predicted to increases its accessibility to proteases. However, the potential AcCoV-JC34 furin cleavage site was within a loop upstream of the typical S1/S2 furin cleavage site found in other CoVs (see Figure 3). In SARS-CoV-2, this upstream region aligned with a DQLTP sequence upstream of the expected S1/S2 cleavage site.

Bioinformatic and biochemical analysis of potential AcCoV-JC34 spike cleavage site

To determine whether furin processes the -RR-R- motif in AcCoV-JC34, we first utilized the PiTou and ProP furin cleavage prediction tools (Figure 5). A positive score for Pitou or a score above 0.5 for ProP indicates the likelihood of furin cleavage. AcCoV-JC34 has a weakly predicted furin cleavage site based on the PiTou score (see also [10]). Although bioinformatic tools are useful for prediction, these may not represent biologically relevant cleavage events, which need to be addressed experimentally.

To directly test whether furin cleaves this site *in vitro*, we performed peptide cleavage assays using furin, along with trypsin as a control. The peptide sequences used were TFMTKARARTTF (Lucheng Rn rat CoV, LRNV), TFSRRARARTL (AcCoV-JC34), and TNSPRRARSVA(SARS-CoV-2). Trypsin cleaved all three peptides with varying efficiency. Furin, as expected from previous studies, cleaved the SARS-CoV-2 peptide; however, it did not cleave the LRNV or JC34 peptides (Figure 6). These data indicate that although AcCoV-JC34 has a minimal furin cleavage sequence (R-X-X-R) it is not able to be cleaved by furin when tested experimentally.

Discussion

The “furin cleavage site” or FCS of SARS-CoV-2 has been at the center of the many discussions on the origin of the COVID-19 pandemic; see [11] for a recent summary. Despite being interpreted as “highly unusual”, an FCS is—to the contrary—very common among the *Coronaviridae* [12], with sarbecoviruses and most alphacoronaviruses being the exception rather than the rule in lacking this important regulatory sequence. In fact, many zoonotic coronaviruses and those in reservoir hosts appear to contain sequences and structural loops at the S1/S2 interface that are sub-threshold for furin-mediated cleavage [13-16] and may be “poised” for spillover events. Examples include “pre-variant” SARS-CoV-2, as well as the sarbecoviruses RmYN02, RacCS203, BANAL-20-116, BANAL-20-246 that have potential phylogenetic homology to the SARS-CoV-2 FCS [17]—and may include the luchacovirus AcCoV-JC34 analyzed here. It is noteworthy that AcCoV-JC34 is the only luchacovirus containing this -R-RR- motif.

While containing an -RR-R- motif, as found in SARS-CoV-2, the data presented here show that this AcCoV-JC34 sequence is not cleaved by furin. The reasons for this are currently unclear. One possibility is that the upstream proline found in SARS-CoV-2, as well as in other spike cleavage site sequences, may promote cleavage by creating a structural turn beneficial for furin activity. It is also possible that the additional downstream arginine residue in AcCoV-JC34 spike may be inhibitory for the tight active site binding pocket present in furin [18]. Alternatively, the structural loop present in AcCoV-JC34 spike may be cleaved by other proprotein convertases of the furin family that have less stringent cleavage requirements, or

by trypsin-like enzymes or cathepsins. Notably, the -RR-R- motif is rare in furin substrates, and only other known example of this sequence motif in FurinDB (a database of furin substrates) is found in proaerolysin, a bacterial toxin [19].

One notable aspect of the -RR-R- motif in AcCoV-JC34 is that it does not align precisely with the S1/S2 motif of most coronavirus spikes (see Figure 3) and is a structurally exposed location above the typical S1/S2 loop (see Figure 4). Analysis of the MERS-CoV spike also shows an additional putative FCS in the MERS-CoV spike (SRSTRS); while this contains a minimal furin motif this sequence shows low scores for furin cleavage with both Pitou and ProP, and FRET-based peptides were not cleaved by furin in biochemical cleavage assays—in contrast to the PRSVRS motif at the expected S1/S2 junction (J. K. Millet, unpublished results). Nevertheless, it is possible that, as with AcCoV-Jc34, this “secondary” MERS-CoV sequence comprises a “blocked” FCS due to flanking hydrophobic and charges residues in the downstream C-terminal positions (i.e., SRSTRSMLKRRDS). This putative secondary cleavage site also lacks an upstream proline/proline-rich region, as with many other S1/S2 regions that are known to be cleaved by furin.

For SARS-CoV-2, it is clear that selection is occurring to up-regulate the spike FCS, as seen with several of the highly transmissible variants that have emerged [20-24]. The FCS can also be readily down regulated upon Vero cell adaptation; for examples see refs [25, 26]. Likewise, some coronaviruses in animal reservoirs may be “poised” for proteolytic cleavage-activation at S1/S2, with selection occurring along with modifications to their receptor binding domain. One interesting example of this may exemplified by the MERS-like bat-CoVs HKU-4 and

HKU-5, with HKU-4 binding human DPP4, but having no identifiable FCS, and with HKU-5 not able to bind hDPP4 and having a robust FCS [27].

Our studies highlight the possible presence of a distinct proteolytic cleavage loop in the coronavirus spike protein and the specific features of the luchacovirus spike—which along with that found in the rhinacoviruses (e.g., SADS-CoV) appears to represent an evolutionary disparate spike protein with apparent similarities to a betacoronavirus spike protein (see Figure 1), despite the taxonomic designation of these viruses as alphacoronaviruses.

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Methods

Furin prediction calculations. Prop: CoV sequences were analyzed using the ProP 1.0 Server hosted at: cbs.dtu.dk/services/ProP/. PiTou: CoV sequences were analyzed using the PiTou V3 software hosted at: <http://www.nuolan.net/reference.html>.

Amino acid alignments and phylogenetic trees

Multiple sequence alignment was performed on coronavirus spike protein using Geneious Prime ® (v.2019.2.3. Biomatters Ltd.). A maximum likelihood phylogenetic tree was constructed using MegaX(, 100 boot strap replicates based on the spike protein. Amino acid sequences of S were obtained from NCBI GenBank. Accession numbers are: AcCoV-Jc34 (YP_009380521), Asian leopard cat CoV (EF584908.1), Bat_Hp/Zhejiang2013(YP_009072440), Bat-Rm/Yunnan/YN02/2019 (QPD89843.1), Bat-SL-CoV_ZC45 (AVP78031.1), BCoV (P15777), Bottlenose dolphin CoV-HKU22 (AHB63508), BtRf-AlphaCoV/YN2012 (YP_009200735), CCoV (AY436637.1), ECoV-NC99 (AAQ67205.1), FCoV-Black (EU186072.1), Ferret-CoV (NC_030292.1), FIPV 79-1146 (DQ010921.1), HCoV-229E(NC_002645.1), HCoV-HKU1(NC_006577), HCoV-NL63(NC_005831.2), HCoVOC43(NC_006213.1), HeCoV(MK679660.1), HKU4(YP_001039953), HKU5(YP_001039962), HKU23(QEY10673), HKU24(QOE77327), IBV(NC_001451.1), Longquan Rl rat CoV (QOE77336.1), Lucheng Rn rat CoV(QOE77268.1), MERS-CoV(AFS88936.1), MHV-1 (ACN89742), PDCoV (MN942260.1), PEDV(NC_003436.1), PHEV(QTF73995.1), Porcine enteric alphacoronavirus GDS04 (ASK51717.1), Rabbit CoV-HKU14(AFE48827), RaTG13(QHR63300), Rhinolophus bat coronavirus HKU2 (YP_001552236.1), Rhinolophus bat CoV-BTKY72 (APO40579.1), Rhinolophus bat CoV-HKU32 (QCX35178), Rhinolophus bat CoV-HKU2 (YP_001552236.1), Rousett bat CoV-229E related(QHA24665), Rousettus bat CoV-GCCDC1(QKF94914), RtClan-CoV/GZ2015 (), RtMurf-CoV-1/JL 2014 (ATP66738), RtRl-CoV/F J2015 (KY370050), SARS-CoV (AAT74874.1), SARS-CoV-2 Wuhan-Hu-1 (YP_009724390.1), Sc-

BatCoV-512 (ABG47078), Swine acute diarrhea syndrome coronavirus(AVM41569.1), Swine acute diarrhea syndrome related coronavirus (AVM80500.1), TGEV(P07946), Turkey-CoV (QRR19172), UkMa1(QBG64648), UKRn3(QBG64657).

Spike structural modelling

Pairwise amino acid alignment between AcCoV-Jc34 (YP_009380521) and SADS-CoV (AVM80500) was performed using Geneious Prime ® (v.2019.2.3. Biomatters Ltd.). S protein models were built based on the SADS-CoV structure obtained from RCSB (PDB: 6M39), using UCSF Chimera (v.1.14, University of California) through the modeler homology tool of the Modeller extension (v.9.23, University of California).

Fluorogenic peptide cleavage assays

Fluorogenic peptide cleavage assays were performed as described previously [14]. Each reaction was performed in a 100 μ L volume consisting of buffer, protease, and AcCoV-Jc34 (TFSRRARARTL) or Lucheng Rn rat CoV (TFMTKARARTTF) or SARS-CoV-2 S1/S2 WT (TNSPRRARSVA) fluorogenic peptide in an opaque 96-well plate. For trypsin catalyzed reactions, 0.8 nM/well TPCK trypsin was diluted in PBS buffer. For furin catalyzed reactions, 1 U/well recombinant furin was diluted in buffer consisting of 20 mM HEPES, 0.2 mM CaCl₂, and 0.2 mM β -mercaptoethanol, at pH 7.0. Fluorescence emission was measured once per minute for 60 minutes using a SpectraMax fluorometer (Molecular Devices) at 30 °C with an excitation wavelength of 330 nm and an emission wavelength of 390 nm. V_{max} was calculated by fitting the linear rise in fluorescence to the equation of a line.

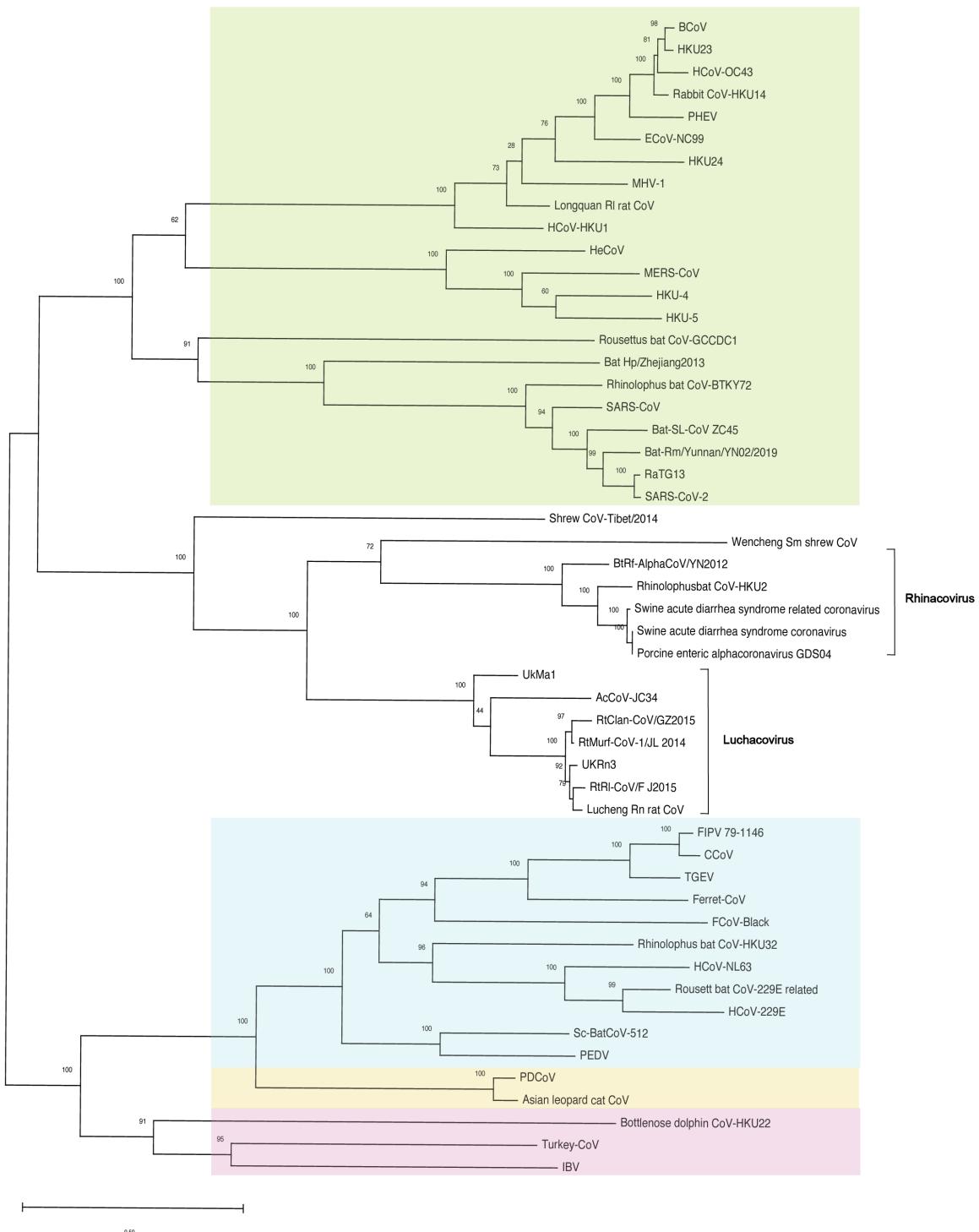


Figure 1. Phylogenetic tree of spike protein sequences.

The maximum likelihood phylogenetic tree was constructed using MegaX, 100 bootstraps, from a multiple sequence alignment of the spike sequences. Betacoronavirus spikes are shaded green, alphacoronavirus spikes are shaded blue, deltacoronavirus spikes are shaded yellow and gammacoronavirus spikes are shaded pink

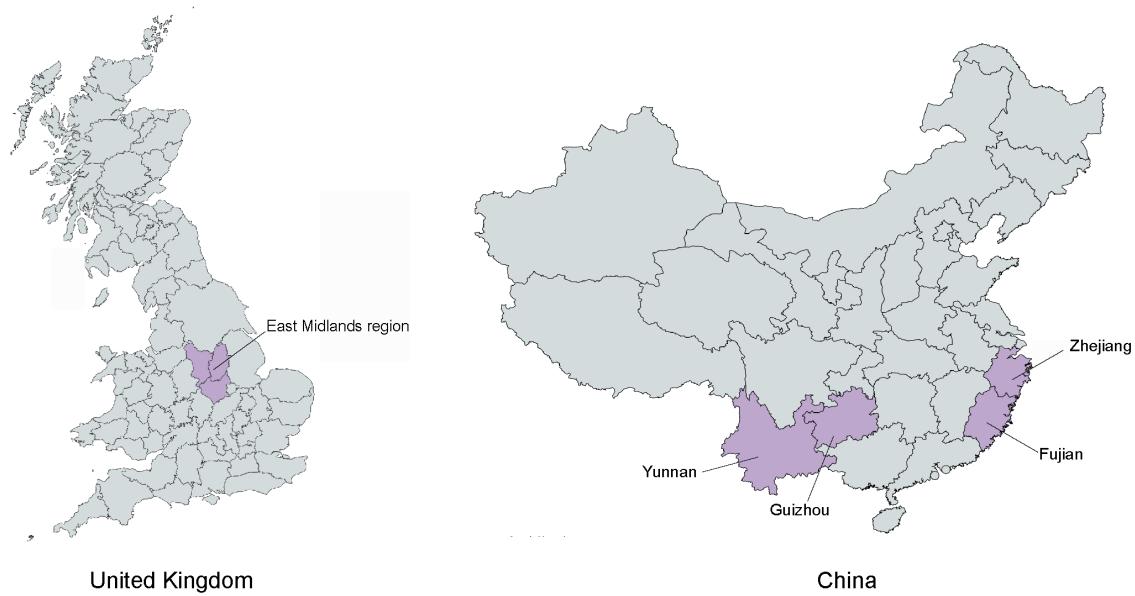
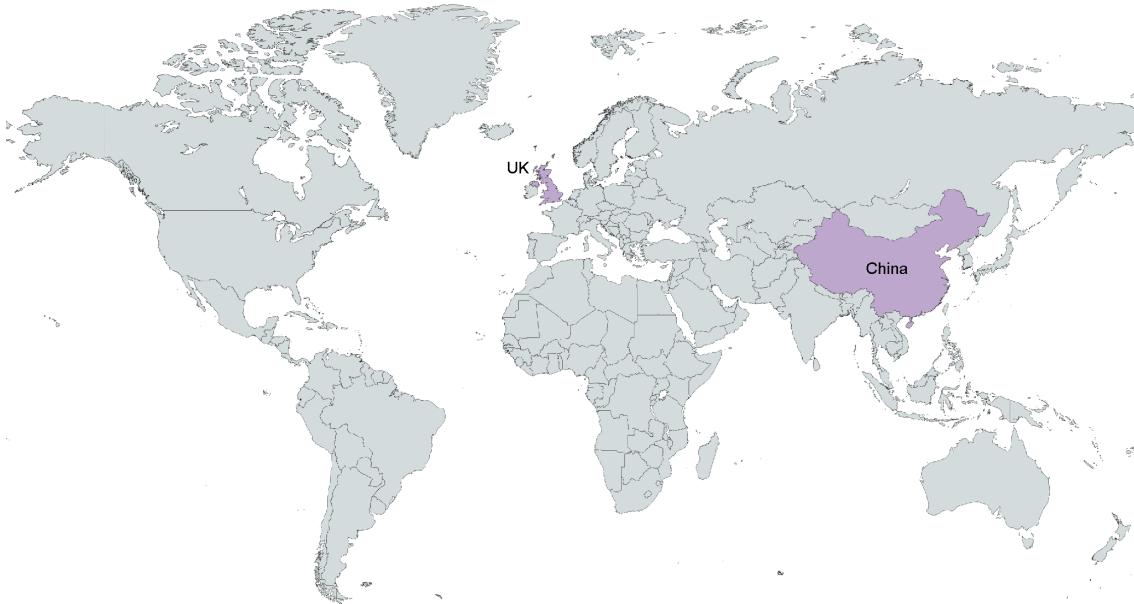


Figure 2. Geographical distribution of currently identified luchacoviruses.

Luchacoviruses have been identified from surveillance studies in United Kingdom (East Midlands region) and China (Yunnan, Zhejiang, Fujian, Jilin, and Guizhou provinces).

Luchacovirus	Rodent species	Location	Time period
			2011.8
AcCoV-JC34	<i>Apodemus chevrieri</i>	China (Yuanna)	2011-2013
Lucheng Rn rat CoV	<i>Rattus norvegicus</i>	China (Lucheng)	2015.5
RtRl-CoV/FJ2015	<i>Rattus losea</i>	China (Fujian)	2014.8
RtMruf-CoV-1/JL2014	<i>Myodes rufocaninus</i>	China (Jilin)	2015.12
RtClan-CoV/GZ2015	<i>Eothenomys melanogaster</i>	China (Guizhou)	2008-2015
UKRn3	<i>Rattus norvegicus</i>	United Kingdom	2008-2015
UKMa1	<i>Microtus agrestis</i>	United Kingdom	

Table 1. Rodent luchacoviruses identified from surveillance studies.

Figure 3

AcCoV-JC34	-----MALIFVLMILITLYRCPFVLCNFQVCTDQLRQQ-----	EVYL-PNAVS	41
Lucheng Rn rat CoV	-----MYFFLLLLFVSADAAIQTCPAPGNVNLDISK-----	LYYG-TQASI	40
SARS-CoV-2	-----MFVFLVLLPLVSSQCVCNLTRTQ-LPPAYTN-----	SFTRGVYYP-DKVFR	44
SARS-CoV	-----MAIFLLFLTTSQSDLDRCTTFDDVQAPNYTQ-----	HTSSMRGVYYP-DEIFR	48
HKU 1	-----MLLIIIFILPTTLAVIGDFNCTNFAIDLNTTV-----	PRISEYVVDSYGLGTYYILDVY	57
OC43	-----MFLILLISLPTAFAVIGDNLCTLDPRLKGSFNN-RDTGPPSISIDTVDTVNLGLGTYVLDVY	63	
MERS-CoV	MIHSVFLLMFLPTESYVDVGPDSVKSCIEVDIQQTFFDKTWPR-----PIDVSKADGIYQPQGRTYS		65
AcCoV-JC34	NATYIATA--VFPT-----RHWHCGSSSNSTPGDNKFNGI-----	GVFVHRFNHPNW-----	86
Lucheng Rn rat CoV	NATFQVVQ--VLPQ-----VPWKCNSYSNG--PSNKFNGI-----	GVFVDSLASAQHS-----	83
SARS-CoV-2	SSVLH5TQDLFLPF---FSNVTFHAIHSGTNGTKRFDNP---VLPFNDGVYFASTEKSNI-----	100	
SARS-CoV	SDTLYLTQDLFLPF---YSNVTGFHTINHT---FDNP---VIPFKDGIVFAATEKSNV-----	97	
HKU 1	NTTILFTG--YFPK---SGANFRDLSLKGTTYLSTLWYQKPF---LSDFNNNGIFSRSVKNTKLYVNKTLY	118	
OC43	NTTLFLNG--YYPT---SGSTYRNMALKGTDLLSTLWFKPPF---LSDFINGIFAKVKNTKVFKDGVMY	124	
MERS-CoV	NITITYQG--LFPYQGDHGDMYVYSAHATGTTPKLFLVANYSQDVQKFANGFVVRIGAAANSTGTVII		132
AcCoV-JC34	-----WHFAAYPAAPTNKT-----WILFWWHR-----ATQAS----TVFQV	118	
Lucheng Rn rat CoV	-----WHLFVYPSMPTNKT-----WILSWADT-----HTSFEHGSVISYVQI	120	
SARS-CoV-2)	-----IRGWIFGTTLDSKT-----QSLLIVNN-----ATNVV--IKVCEFQF	135	
SARS-CoV	-----VRGWVFGSTMNNKS-----QSVIIINN-----STNVV--IRACNFEL	132	
HKU 1	SE-----FSTIVIGSVFINNS-----YTIVVQPH-----NGVLE--ITACQYTM	155	
OC43	SE-----FPAITIGSTFVNTS-----YSVVVQPRTIINSTQDGVNKLQGLLE--VSVCQYNM	173	
MERS-CoV	SPSTSATIRKIYPAFMLGSSVGNFSDGKMGFFFNHTLVL	186	
AcCoV-JC34	CKYQQPAVDVLTSGFR--CE---APTNPPCSA--IVNMAI-----ECLINSTFRPTTFSF-	166	
Lucheng Rn rat CoV	CKYPSNVVTIDNGNG---CHTNMAGPGATTCDV--ILSSPL-----ECVLNRTYSQQYAGV-	172	
SARS-CoV-2	CNDPFLGVYYHKNNKSWMESEFRVYS-SANNCTF-EYVSQPFLMDLEGKQGNFKNLREFVFKNIDG---	199	
SARS-CoV	CDNPFFAVSKPMGTQ---THTMIFD-NAFNCTF-EYISDAFSLDVSEKSGNFKHLREFVFKNKDGF	194	
HKU 1	CEYPHTICKSK-GSSR---NESWHFDKSEPLCLF-----KKNFTYNVSTDWL-	198	
OC43	CEYPHTICHPNLGNHF---KELWHYDTGVVSCLY-----KRNFTYDVNATYL-	217	
MERS-CoV	CE-PRSGNHCAGNSY---TSFATYHTPATDCSDGNYNRNASLNSFKEYFNLRNCFMYTYNITEDEIL	251	
AcCoV-JC34	SYVTWYNSEINAVIH-----GQRFSF--VYEGFLWSNASAFCYDA-----NGCI	208	
Lucheng Rn rat CoV	SYITWYNDHIIASIQ-----GEVFTF-DIGEVLQWSNFSAFCGTG-----NKCG	215	
SARS-CoV-2	-YFKIYSKHTPINLV---RDLPGQFSALEPLVDPPIGINITRFQTLALHRSYLTGDSSSGWTAGAAAY	265	
SARS-CoV	-YV--YKGYQPIDVV--RDLPSGFNTLKPIFKLPLGINITNFRALTAFSPAQDT-----WGTSAAY	252	
HKU 1	-YFHFYQERGTFYAY--YADSGMPTTFLF-SLYLGTLLSHYYVLPHTCNAISSNTDN---E---TLQY	256	
OC43	-YFHFYQEGGTFYAY--FTDTGFVTKFLF-NVYLGMLASHYYMPLTC--IRRPKDG---F---SLEY	273	
MERS-CoV	EWFGITQTAQGVHLFSSRYVDLYGGNMFQATLPVYDTIKYYSIIPHSIRSISQDRKA---W---A-AF	313	
AcCoV-JC34	FNIPNTTSDWLVSTDSSGAINNFVDCGFYESQLKCKNLVFELEPAVYHGAALPIE--SSVYYVANDLA	275	
Lucheng Rn rat CoV	FSYATTLSSEWLVRTDNDGTVIDYVICDTDFESQLKCKNMVFELTPAVYSGSAVELQ--SAIYYVSNELP	282	
SARS-CoV-2	YVGYLQPRTFLLKYNENGTTDAVDCALDPLSETKCTLKSFTVEKGIVYQTSNFRVQPTESIV-RFPNIT	333	
SARS-CoV	FVGYLKPTTFMLKYDENGTTDAVDCSQNPLAELKCSVKSFEIDKGIYQTSNFRVPSGDVV-RFPNIT	320	
HKU 1	WVTPLSKRQYLLKFDNRGVITNAVDCSSFFFSEIQCKTKSLLPNTGVYDLSGFTVKPVATVHRRIPDLP	325	
OC43	WVTPLTPRQYLLAFNQDGIIFNAVDCMSDFMSEIKCTQSIAPPTGVYELNGYTVQPVADVRRKPDL	342	
MERS-CoV	YVYKLQPLTFLLDFSDVGYIRRAIDCGFNDLSQLHCSYESFDVESGVYSVSSFEAKPSGSVVEQAEGVE	382	
AcCoV-JC34	D-CAFS--FADIFSDGTGNYGLRRHVFTNCWNYTSW-----FLCDDGLACIIFNAIFSEVV--	330	
Lucheng Rn rat CoV	D-CDFS--FADMFMGDTGNFEGLRRHVFSNCWNYTAW-----FACADDYSCIIFNAIFAEVR--	337	
SARS-CoV-2	NLCPFGEVFNATRFASVYAWN--RKRISSNCVADYSVLYNSASFSTFKCYGVSPTKLNDLCFTNVYADS	399	
SARS-CoV	NLCPFGEVFNATKFPSVYAWE--RKKIISNCVADYSVLYNSTFFSTFKCYGVSATKLNDLCFSNVYADS	386	
HKU 1	D-CDIDKWLNDFNVPSPLNWE--RKIFSCNCNFLSTLLRLVHTDSFSCNNFDESKIYGSCFKSIVLDK	390	
OC43	N-CNIEAWLNDKSVPSPLNWE--RKTFSNCNFNMSLMSFIQADSFTCNNIDAAKIYGMCFSSITIDK	407	
MERS-CoV	--CDFSPLLSGTP--PQVYNFK--RLVFTNCNYNLTKLLSLFSVNDFTCSQISPAAIASNCYSSLILDY	445	
AcCoV-JC34	-----YNLTQPDGLINPFLRCNGLVDYTIVKGCS-AGYVLRYQLFDSG-----	372	
Lucheng Rn rat CoV	-----YKLSQPDGLVNPFIKCNGLDLYSITKGCS-SGFVLRYQLYANG-----	379	
SARS-CoV-2	FVIRGDEVRIQIAPGQTGKIADYNYKLPPDDFTGCVIAWNSNNLDSKVGG--NYNYLYRLFRKS-----	459	
SARS-CoV	FVVKGDDVRQIAPGQTGVIADYNYKLPPDDFTGCVLAWNTRNIDATSTG--NYNYKYRLKHG-----	446	
HKU 1	FAIPNSRRSDLQLGSSGFLQSSNYKIDTSSSCQLYSLPAINVTINNNPSSWNRYYGFFNNF-----	454	
OC43	FAIPNRRKVDLQLGNGLQSSNYRIDTTATSCQLYYNLPAANVSVSRFPNPTWNKRGFIEDSVFVPQ	476	
MERS-CoV	FSYPLSMKSDLSVSSAGPISQFNYKQSFSNPTCLILATVPHNLTTITK-----PLKYSYINKC-----	503	

AcCoV-JC34	-QFNPDAYTPDYM--EC-----	386
Lucheng Rn rat CoV	-SFDVNSYTPDYM--EC-----	393
SARS-CoV-2	--NLKPFERDISTEIYQAGST--PC-----	480
SARS-CoV	--KLRPFERDISNVPFSPDGK--PC-----	467
HKU 1	--LSSHSVYVSYRCFSVNNTFCPCAKPSFASSCKSHKPPSASCPIGTYRSCESTTVDLHTDRCRS	519
OC43	PTGVFTNHSVVYAQHCFKAPKNFCPC-----SSCPGKNNNGIGTCPAGTNSLCDNLCTLD-----	531
MERS-CoV	-SRLLSDDRTEVPQLVNANQYS--PC-----	528
AcCoV-JC34	--FGYFQI-----DRSYIVYNAKFIT-----	405
Lucheng Rn rat CoV	--FGYFAL-----YNGYVIYNAKFVS-----	412
SARS-CoV-2	--NGVEGFNCYF-----PLQS-----YGFQPTN-----GVGYQPYRVRVVL-----	514
SARS-CoV	--TPPALNCYW-----PLND-----YGFYTT-----GIGYQPYRVRVVL-----	500
HKU 1	CLPDPIATYDPRSCSQKKSLSVGVGEHCAGFGVDEEKCGVLGDSYNSCLCSTDALFLGWSYDTCVSNNRC	588
OC43	--PITLKAPDTYKCPQSKSLVGIGEHCSGLAVKSDYCG-----NNSCTCQPQAFLGWSADSCLQGDKC	592
MERS-CoV	IVPSTVWEDGDDYRKQLSPLEG-----GGWLVAS-----GSTVAMTEQLQMG-----	570
AcCoV-JC34	--EGLTCVVLQPVPEPE-----LGVCKQYTIDGVTQGILHTS NAN-ITTFHNLLY--YG-----	454
Lucheng Rn rat CoV	--KGLTCVVQPVPEPE-----LDVCKSYTIDGVTQGILRNTTHQ-IDSFHNI LY--YG-----	461
SARS-CoV-2	--FELLHAPATVCGPKKSTNLVKNK-----CVNFNFNGLTGTGVLTESNKK-FLPFQQFGRDIAD	571
SARS-CoV	--FELLNAPATVCGPKLSTDLIKNQ-----CVNFNFNGLTGTGVLT PSSKR-FQPFQQFGRDVSD	557
HKU 1	NIFSNFILNGINSGTTCSNDLQLQPNTEV--FTDVCVDYDLYGITQGIFKEVSAVYYNSWQNLLYDSNG	655
OC43	NIFANFILHDVNNGLTCSTDLKANTEI--ELGVCVNYDLYGISGQGIFVEVNATYYNSWQNLLYDSNG	659
MERS-CoV	--FGITVQYGTDTNSVCPKLEFANDTKIASQLGNCVEYSLYGVSGRGVFQNCTAV-GVRQQRFVYDAYQ	636
	(JC34 -RR-R- motif)	
AcCoV-JC34	DMVSH--VRIKGIVYSVEPCNSFYYSV---LKTRSEVGYLYSGANCNSSDVV--FS RRA RARTLTD-----	515
Lucheng Rn rat CoV	DMVSY--VRIRGVVYAVESCNRFYYSV---FKTLSAIGYLYSGATCDSTDVTT--FMT KAA KATTFVDS	522
SARS-CoV-2	TTDAVRDPQTLEI-LDITPCSFGGVSVITPGNTSQNVAVLYQDVNCTEVPAI--HADQLTPW RVY S	637
SARS-CoV	FTDSVRDPKTSEI-LDISPCSFGGVSVITPGTNASSEAVVLYQDVNCTDVSTAI--HADQLTPA W RIYS	623
HKU 1	NIIGFKDFVTNKT-YNIFPCYAGRVAASF--HQNASSLALLYRNLKCSYVLNNI-----SLTTQPYFDS	716
OC43	NLYGFRDYITNRT-FMIHSCYSGRVSAAY--HANSSEPALLFRNIKCNVFNNS--LTRQLQPINYSFD	723
MERS-CoV	NLVGY--YSDDGNEYCLRACVSPVSVIY--DKETKTHATLFGSVACEHISSTMSQYS RST RSML KRRD	701
	S1 / S2	
AcCoV-JC34	S-----LGCLVDVQLTNNN-YTTCSYPIGNGLCADVNVTG-----MPVVGNIYI-----QPH-----	561
Lucheng Rn rat CoV	S-----LGCFIDVSVTDGN-YTDCLNPIGNGFCDVNV-----QPVVGNIFI-----QTH-----	568
SARS-CoV-2	TGSNVFQTQAGCLIGAEHVNN--SYECDIPIGAGICASYQTQT-NSP RRA SVASQSIAYTMSLGAD-----	703
SARS-CoV	TGNNVFQTQAGCLIGAEHVDT--SYECDIPIGAGICASYHTV-----RSTSQKSIVAYTMSLGAD-----	685
HKU 1	Y-----LGCVFNADNLTDYVSSCALRMGSGFCVDNSPSSSSSS RRKRR SISASYRFVTF---EPF-----	774
OC43	S-----YLGCVVNAYNSTAISVQTCDLTVGSGYCVDFKNR-----R SRA ITTGYRFTNF---EPF-----	777
MERS-CoV	STYGPLQTPVGCVLGNVNSLF-VEDCKLPLGQSLCALPDTSTLTP RSV RSVPGEMLASIAFNHPI-----	768
AcCoV-JC34	DTDYARPILSPQ---IVSLPLDHINVKEQFVQTSAPKFDVDCERYICDVSQCKELLAKYGGYCPKI	626
Lucheng Rn rat CoV	DTDYARPILTAQ---QIELPIDHYVSVKEQFIQTSTPKFDVDCERYICDVSQDCRELLVKYGGYCSKI	633
SARS-CoV-2	SVAYSNN-----IAIPTNFTISVTEILPVSMKTSVDCTMYICGDS TECSNLLLQYGSFCTQL	763
SARS-CoV	SIAYSNN-----IAIPTNFSISITTEVMPVSMAKTSVDCNMYICGDS TECANLLLQYGSFCTQL	745
HKU 1	NVSFVNDSIESVGGLYEIKIPTNFTIVQGEEFIQTNSPKVTDCLSLFVCSNYAACHDLLSEYGTFCDNI	843
OC43	TVNSVNDSLEPVVGGLYEIQIPSEFTIGNMEEFIQTNSPKVTDCAAFVCGDYAACKLQLVEYGSFCDNI	846
MERS-CoV	QVDQLNNS-----YFKLSIPTNFSFGVTQEYIQTTLQKTVDCQKQYVCNGFQKCEQLLREYGQFCSKI	831
	S2' (FP)	
AcCoV-JC34	VSDIKSSSLQLDYQVQGIYKTLNVDVKVPDV-DFG-AFNF-----SMFESEP NGR SFIEDIL-----	681
Lucheng Rn rat CoV	LADIKSSSIQLDYQILGLYKTLAVDFKVPDI-DFG-DFNF-----SMYMS EA NG R SFIEDLL-----	688
SARS-CoV-2	NRALTGIAVEQDKNTQEVFAQVKQIYKTPPIKDFG-GFNF-----SQILPDPSKPS R SFIEDLL-----	822
SARS-CoV	NRALSGIAAEQDRNTREVFAQVKQMYKTPTLKYFG-GFNF-----SQILPDPLKPT R SFIEDLL-----	804
HKU 1	NSILDEVNGLLDTTQLHVADTLMQGVTLSNLNTNLHFDVDNINFKSLVGCLGPHCGS-SS R SFFEDLL-----	911
OC43	NAILTEVNELLDTTQLQVANSLMNGVTLSTKLKDGVNFNVDDINFS P VLGCLGSECSKASS R SAIEDLL-----	915
MERS-CoV	NQALHGANLRQDDSVRNLFASVKSSQSSPIIPGFGGDFNL-----TLLEPVSI T GSRS A SAIEDLL-----	894

fusion peptide (FP)

AcCoV-JC34	FDKIVTTGPGFYQDYYDCK--KMNLEDITCKQY	YNGIMVIIPPVMDTLITFWSSA-VAGSMTAGLFGG	746							
Lucheng Rn rat CoV	FDKIVTTGPGFYQDYYDCK--KMNLDLCKQY	YNGIMVIIPPVMDTLITFWSSA-VAGSMTAGLFGG	753							
SARS-CoV-2	FNKVTLADAGFIQKYGDCL--GDIARDLICAQK	FNGLTVLPLLLTDEMIAQYTSALLAGTITSGWTFG	889							
SARS-CoV	FNKVTLADAGFMQKYGEC--GDIARDLICAQK	FNGLTVLPLLLTDDMIAAYTAALVSGTATAGWTFG	871							
HKU 1	FDKVKLSDVGFVEAYNNCT--GGSEIRDLLCVQS	FNGIKVLPPILSESQISQGYTTAATVAAMFPPWS--	976							
OC43	FDKVKLSDVGFVEAYNNCT--GGAEIRDLLCVQS	YKGKVLPPLLSENQISQGYTLAATSASLFPWP--	980							
MERS-CoV	FDKVTIADPGYMQGYDDCMQQGPASARDLICAQY	VAGYKVLPLMDVNMEAAYTSSLGSIAGVGTAG	963							
AcCoV-JC34	--QAGMVSWSIAVAGRLNALGVMQDALVNDVNKL	ANGFNNLTQYADGFKTTSQALSAIQSVNNNAQQ	813							
Lucheng Rn rat CoV	--QAGMVSWTVALAGRLNALGVMQDALVEDVNKL	ANGFNNLTQYSDGFKTTSQALSTIQAVVNNNAQQ	820							
SARS-CoV-2	AGAALQIPFAMQMAYRFNGIGVTQNVLYENQKLI	ANQFNSAIGKIQDSSLTASALGKLQDVVNQNAQA	958							
SARS-CoV	AGAALQIPFAMQMAYRFNGIGVTQNVLYENQKQI	ANQFNKAISQIQLTSTALGKLQDVVNQNAQA	940							
HKU 1	--AAAGIPFSLNQYRINGLGVTVMDVLNKQKLI	ATAFNNALLSIQNGFSATNSALAKIQSVVNSNAQA	1043							
OC43	--AAAGVPFYLNQYRINGLGVTVMDVLSQNQKLI	ANAFNNALHAIQQGFDATNSALVKIQAVVNANAEA	1047							
MERS-CoV	LSSFAAIPFAQSIFYRLNGVGITQQVLSENQKLI	ANKFNQALGAMQTGTTTNEAFQKVQDAVNNNAQA	1032							
AcCoV-JC34	ISQLVQGLSENFGAISNNFLVIAERLERLEAQ	MQMDRLINGRMNILQNFVTNYKLSISELKSQQILAQS	882							
Lucheng Rn rat CoV	VSQLVQGLSENFGAISNNFLAERLERIEAAMQ	MDRLINGRMNILQNFVTNYKLSISELKSQQALAQS	889							
SARS-CoV-2	LNTLVQKLSSNFGAISSSVLNDILSRDKVEAEV	QIDRLITGRLQLQTYVTQQLIRAAEIRASANLAAT	1027							
SARS-CoV	LNTLVQKLSSNFGAISSSVLNDLSSRLDKVEAEV	QIDRLITGRLQLQTYVTQQLIRAAEIRASANLAAT	1009							
HKU 1	LNSLLQQLFNKFGAISSSLQEILSRDLAEEAQV	QIDRLINGRLTALNAYVSQQLSDISLVKFGAALAME	1112							
OC43	LNNLLQQLSNRFGAISASLQEILSRDLAEEAQI	DRLINGRLTALNAYVSQQLSDSTLVKFSAAQAME	1116							
MERS-CoV	LSKLASELSNTFGAISASIGDIIQRLDVLEQD	QIDRLINGRLTTLNAFVAQQLVRSESAALSAQLAKD	1101							
AcCoV-JC34	LVNECVYAQSNRNGFCGDLHLFLS	MQRAPDGIMFFHYTLVPPNNTIDVETTPGLCLGN--VCIAPRDG	949							
Lucheng Rn rat CoV	LTNECVYAQSSRNFGDGLHLFLS	MQRAPDGIMFFHYTLKPNNTIIVETTPGLCLSD--VCIAPKDG	956							
SARS-CoV-2	KMSECVLGQS	KRVDFCGKGYHLMSPQSAPHGVVFLHVTYVPAQEKNFTTAPAI	1093							
SARS-CoV	KMSECVLGQS	KRVDFCGKGYHLMSPQAAPHGVVFLHVTYVPSQERNFTTAPAI	1075							
HKU 1	KVNECVKSQS	PRINFCGNGNHILSLVQ	NAPYGLLFMHFSYKPI	KFSFTVLPGLCISGD--VGIAPKQG	1179					
OC43	KVNECVKSQS	SSRINFCGNGNHII	SLVQ	NAPYGLYFIHFNYVPTKYVTA	KVSPGLCIAGN--RGIAPKSG	1183				
MERS-CoV	KVNECVKAQSKRSG	FCGQGTHIVSFVNAPNGLYFMHV	GYYPSNHIEVVSAYGLCDAANPTNCIAPVNG	1170						
AcCoV-JC34	LFVKTNIRSDV--WHFTTRNL	YNPQAITVNNSVIVNG-GVNFTSLNQTIEGIEPPTIPS--FDEEFEDL	1013							
Lucheng Rn rat CoV	LFVRLSTAR	TD-WHFTTRNRYSP	EPIVTNNLTISG-GVNFTV	VNSTIDGIEPPANPS--FDEEFAEL	1021					
SARS-CoV-2	VFVSGNTH----	-WFVTQRNFYEP	QIITTNTFVSGNC	VDVIGIVNNTVYDPLQPELDS--FKEELDKY	1155					
SARS-CoV	VVFNGNTS----	-WFITQRNF	SPQIITTNTFVSGNC	VDVIGIINNTVYDPLQPELDS--FKEELDKY	1137					
HKU 1	YFIKHNHD----	-WMFTGSS	YYPEPISDKN	VFMNTCSVNFTKAPLVLYNHSPV	PKLSD--FESELSHW	1241				
OC43	YFVN	VNNT----WMTGSG	YYPEPITENN	VVVMS	TCAVNYTKAPYVMLNTSIPNLPD--FKEELDQW	1245				
MERS-CoV	YFIKTN	NTNTRIVDEWSY	TGSSFYA	PEPITS	LNTKYVAP-QVTYQNI	STNLPPPLGNSTGIDFQDELDEF	1238			
AcCoV-JC34	YKNITTLEQLK-NIT-FDPE	LLNLTYYIDR	DLDELSTNVSQLHVD	ISEFNKYVQYIKWPWYVWLAIFLV	1080					
Lucheng Rn rat CoV	YKNVTLEQLK-NIS-FDPE	MLNLTYYIDR	DELATNVSQLHVD	SEFNKFVQYIKWPWYVWLAIFLV	1088					
SARS-CoV-2	FKNHTSP	VDVLG-DISGINAS	VVNIQKEIDRL	NEVAKNLNESLIDLQELGKYEQYIKWPWYI	WLGFIAG	1223				
SARS-CoV	FKNHTSP	VDVLG-DISGINAS	VVNIQKEIDRL	NEVAKNLNESLIDLQELGKYEQYIKWPWYV	WLGFIAG	1205				
HKU 1	FKNQTSI	APNLTLNHTINATFL	DLYYEMNL	IQESIKLNNSYINLK	DIGTYEMYVKWPWYVWLLISFS	1310				
OC43	FKNQTSV	AAPDL--SLDY	INVFLDLQ	VMEMNRLQEA	IKVLNHSYINLK	DIGTYEYVV	KWPWYVWLLICLA	1312		
MERS-CoV	FKNV	TSIPNFG-SLTQ	INTTLLDTY	EMLSLQ	QVVKALNESYIDL	KELGNY	TYYNKWPWYI	WLGFIAG	1306	
AcCoV-JC34	LVLFSF	MLLWCCC	CATGCCGCC	MLGSACNGC	CTKPQ--TVEFEK	VHVQ--	1126			
Lucheng Rn rat CoV	LVLFSF	MLWCCC	CATGCCGCC	MLGSACNGC	CTKPQ--PIEFEK	VHVQ*-	1135			
SARS-CoV-2	LIAIVM	VMTIMLCC	MTSCS	CLKG-C	CSCGSCCKF	DEDDSEPV	LKGVKLHYT	1273		
SARS-CoV	LIAIVM	VMTILLCC	MTSCS	CLKG-A	CSCGSCCKF	DEDDSEPV	LKGVKLHYT	1255		
HKU 1	FIIFL	VLLFFF	ICCTGCGS	ACF--	SKCHNCC	DEYGGHDF	VIKTS	HDD--	1356	
OC43	GVAM	VLLFFF	ICCTGCGT	SCF--	KKCG	GCCDDY	TGQ	ELVIKTS	HDD--	1358
MERS-CoV	LVAL	ALCVFF	ILCCTGCGT	NMGK-L	KCN	RCCD	RYEEY	-DLEPH	KVHVH--	1353

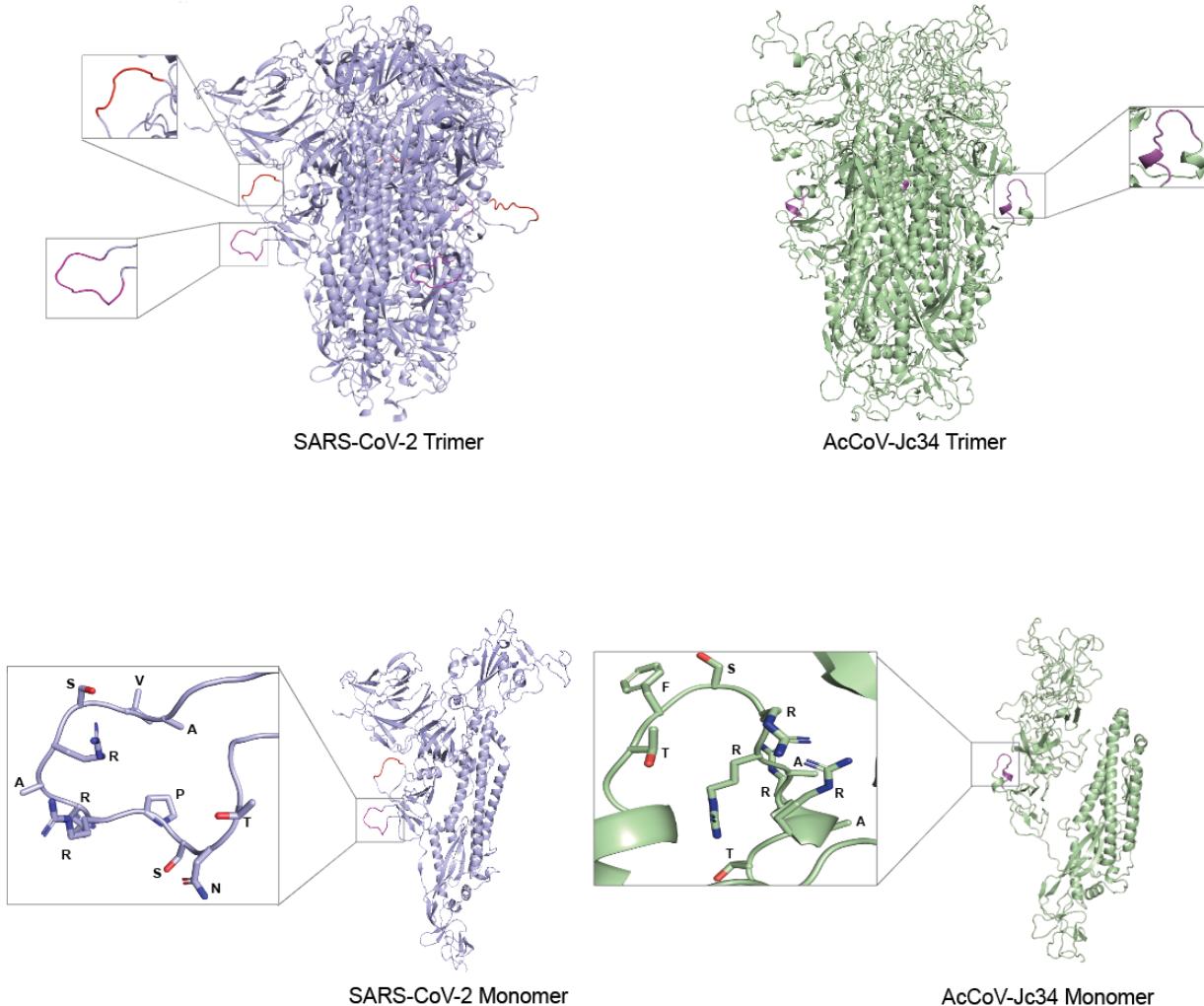


Figure 4. Predicted structure of AcCoV-JC34 spike protein.

The AcCoV-JC34 spike protein was modelled based on SADS-CoV spike. In SARS-CoV-2, the pink highlight indicates where the furin cleavage sequence (PRRAR) is located. The red highlight is the location that aligns with AcCoV-JC34 potential furin cleavage site. In the AcCoV-JC34 structural model, the pink highlight indicates location of the potential furin cleavage site (SRRAR).

Virus	Predicted S1/S2 sequence	PiTou	ProP
AcCoV-JC34	495 - CNSSDVVTFS RRAR ARTLTD - 514	+0.15	0.279
Lucheng Rn rat CoV	502 - CDSTDVTTFMT KAR ATTFVD - 521	-4.577	0.132
MERS-CoV	681 - HISSTMSQYS RSTR SMLKRR - 699	-2.662	0.497
SARS-CoV-2	672 - ASYQTQTNSP RRAR SVASQS - 691	+9.196	0.62
SARS-CoV-1	654 - AGICASYHTVSLL R STSQKS - 673	-5.167	0.123
HCoV - HKU1	747 - YNSPSSSSS RRKRR SISASY - 766	+14.634	0.88
OC43 (clinical)	750 - GYCVDYF KNRRSRR AITTGY - 769	+10.1	0.753

Figure 5. Furin cleavage analysis of CoV S1/S2 cleavage site

CoV S sequences were analyzed using the ProP 1.0 and PiTou 3.0 furin prediction algorithm, generating a score with bold numbers indicating predicted furin cleavage. (|) denotes the position of the predicted S1/S2 cleavage site. Basic resides, arginine (R) and lysine (K), are highlighted in blue.

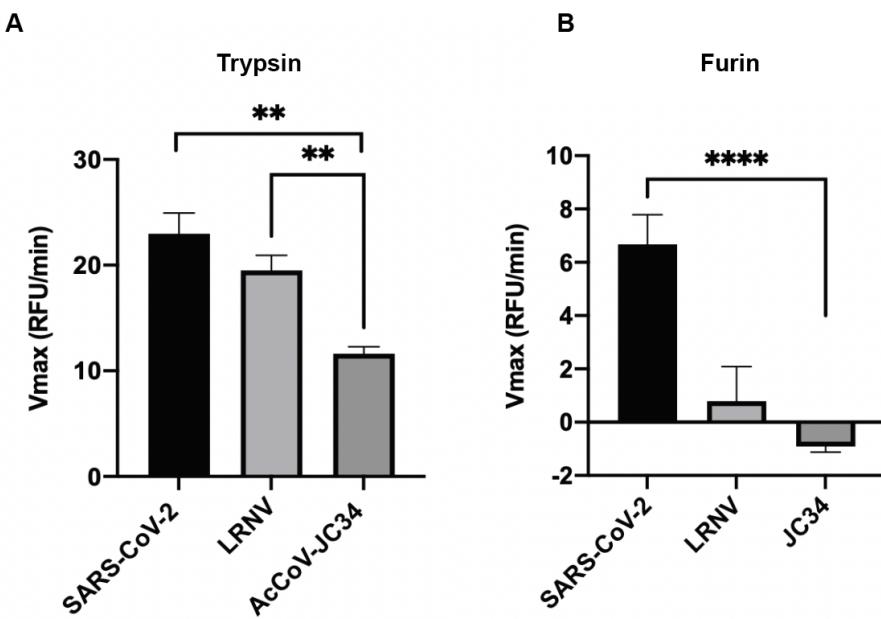


Figure 6. Fluorogenic peptide cleavage assays of the predicted AcCoV-JC34 furin cleavage site.

Peptides mimicking the S1/S2 site of the SARS-CoV-2 WT, AcCoV-JC34, and Lucheng Rn rat CoV (LRNV) were evaluated for in vitro cleavage with A) trypsin and B) furin proteases under pH 7.4 (trypsin), and 7.5 (furin) conditions. Trypsin cleaved all three peptides, while furin only cleaved SARS-CoV-2.

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