

1 **Wanted not, wasted not: Searching for non-target taxa in environmental DNA**

2 **metabarcoding by-catch**

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21 **Abstract:**

22 Metabarcoding of environmental DNA is based on primers that are specific to the target taxa
23 (e.g. bacteria, zooplankton, fishes). However, due to the nature of the commonly used protocols,
24 regardless of the chosen primers, several sequences of non-target species will inevitably be
25 generated, but are usually discarded in commonly used bioinformatics pipelines. These non-
26 target sequences might contain important biological information about the presence of other
27 species in the studied habitats and its potential for ecological studies is still poorly understood.
28 Here, we analyzed the presence of mammal and bird species in aquatic environmental samples
29 that were originally amplified targeting teleost fish species. After all cleaning and checking
30 steps, we kept 21 amplicon sequence variants (ASVs) belonging to mammals and ten to birds.
31 Most ASVs were taxonomic assigned to farm/domestic animals, such as cats, cows, and ducks.
32 Yet, we were able to identify a native semi-aquatic mammal, the capybara, in the samples. Four
33 native bird species and a non-native potentially invasive bird (*Corvus* sp.) were also detected.
34 Although the data derived from these samples for mammals and birds are of limited use for
35 diversity analyses, our results demonstrate the potential of aquatic samples to characterize non-
36 aquatic birds and highlight the presence of a potentially invasive species that had not been
37 recorded before in the region.

38 **Key-words:** Amplicon sequence variants, Birds, Fishes, High throughput sequencing, Mammals,
39 Neotropics, Vertebrata.

40

41 **1. Introduction:**

42 In recent decades, environmental DNA (eDNA) metabarcoding revolutionized our ability
43 to efficiently sample and monitor a wide range of taxa (Seymour et al., 2020; Yang et al., 2021).
44 Although eDNA metabarcoding is a cost-effective technique when compared with traditional
45 surveys (Shokralla et al., 2015) or other molecular techniques such as shotgun sequencing (Stat
46 et al., 2017), the cost of amplification and sequencing can pose an important limitation for
47 countries in the Global South. In this context, optimizing the results obtained from eDNA
48 metabarcoding sequencing is highly desirable.

49 A crucial step of eDNA metabarcoding studies is the choice of genetic marker and
50 primers. The chosen genetic marker should be variable enough to distinguish between target
51 species, whereas the used primers should be specific enough to avoid amplifying non-target taxa
52 (Collins et al., 2019; Leese et al., 2020). However, as any given environmental sample contains a
53 myriad of DNA from entire communities, amplification of non-target taxa is inevitable. Usually,
54 the bioinformatics of taxonomic assignment discards sequences that do not belong to the target
55 taxa (Andújar et al., 2018; Burgess, 2001). Yet, these non-target sequences can provide valuable
56 information about important organisms present in the sample, such as threatened or invasive
57 species, or even seasonal variation in the composition of non-target communities (Mariani et al.,
58 2021).

59 Aquatic environments have been extensively studied for a wide range of taxa, from
60 viruses to eukaryotic metazoans (Alberti et al., 2017), including many vertebrate taxa (Stat et al.,
61 2017). The most abundant and diverse vertebrates on Earth—the teleost fishes (Osteichthyes)
62 that dominate the aquatic realm — have been widely studied through eDNA metabarcoding in
63 marine and freshwater systems (McElroy et al., 2020), both locally (Sales et al., 2021) and

64 globally (Miya et al., 2020). However, aquatic samples are also able to successfully record other
65 vertebrate classes, such as mammals (Harper et al., 2019; José et al., 2021; Sales et al., 2020).
66 Although most studies that detected mammals and birds used more general primers for
67 vertebrates (Andruszkiewicz et al., 2017; Closek et al., 2019), a recent study identified such
68 organisms from primers originally designed to amplify teleost fishes (Mariani et al., 2021).

69 Here we explore, for the first time in the Neotropical region, the potential of
70 metabarcoding-derived sequences to identify non-target vertebrates from aquatic environmental
71 samples. We used data from fish monitoring of the Itaipu dam and associated fish pass system in
72 the Paraná River, in South Brazil (Dal Pont et al., 2021). We also discuss some potential caveats
73 and limitations of this approach.

74

75 **2. Material and Methods:**

76 Our sampling design is described in Dal Pont et al. (2021). In brief, six sites were
77 sampled in the Piracema fish pass, including a site on the reservoir dam, four in the fish pass and
78 one in the Paraná River in 2019 and three sites that were re-sampled in 2020. The sites were
79 sampled in sextuplicate, each including one liter of water filtered using nitrocellulose membranes
80 (0.45- μ m pore). Filters were kept in 100% ethanol under refrigerated conditions. Total DNA
81 from samples and three negative controls were extracted using magnetic beads. We amplified the
82 12S rRNA gene using the MiFish primers designed by Miya et al. (2015) to yield 163–185 bp
83 fragments targeting teleost fish. The samples and the three negative controls were sequenced
84 with Illumina MiSeq (Illumina, USA). The raw sequences are deposited in GenBank under
85 Bioproject PRJNA750895 (biosamples SAMN20500524 – SAMN20500577).

86 To determine amplicon sequence variants (ASVs), we first removed primers with the
87 Cutadapt package (Martin, 2011) in Python v.3.3 (Van Rossum and Drake, 2009), and then used
88 the DADA2 package (Callahan et al., 2016) in R v. 4.0.2 (R Core Team, 2021) to quality filter
89 reads, merge sequences, remove chimeras, and to infer ASVs. ASVs present with a proportion >
90 0.01% of reads across all three negative controls were discarded.

91 For taxonomic inference, we build a reference dataset of 12S mitochondrial DNA
92 sequences for fish, mammal, and bird taxa that have been historically recorded in the Itaipu area
93 using the available data in GenBank (Benson et al., 2018). A total of 75 bird, 126 fish, and 78
94 mammal species had sequences available and were used. For fishes, we added an in-house
95 database which included sequences for 42 additional species (Dal Pont et al., 2021). Finally, we
96 blasted the obtained ASVs sequences with our reference database to verify the taxonomic
97 composition using the “Blastn” function of the program Blast+ (Camacho et al., 2009) for the 10
98 best hits and an e-value of < 0.001. We kept ASVs that matched a species from our reference at
99 minimum level of 75% similarity. Inconsistent results were checked manually. ASVs blasted >
100 98% similarity was considered the matched species. All other ASVs were blasted in GenBank as
101 an additional check and then replaced if there was a match with a superior e-value. ASVs with
102 similarity between 96 to 98% were considered in the same genus, 90 to 96 the same family. For
103 similarities between 75 to 90 the ASVs were just considered for the class. We used the
104 metagMisc v. 0.0.4 (Mikryukov, 2019), phyloseq v. 1.36.0 (McMurdie and Holmes, 2013), and
105 tidyverse v. 1.3.0 (Wickham, 2017) packages for data curation and ggplot2 v. 3.3.2 (Wickham,
106 2016), plotly v. 4.10.0 (Sievert, 2020), and patchwork v.1.1.1 (Pedersen, 2019), for data
107 visualization in R v. 4.1.1 (R Core Team, 2021). The script is available as Appendix 1.

109 **3. Results:**

110 We obtained a total of 17,616,032 reads of raw sequence. After all cleaning steps, we
111 retained a total of 2,280,447 reads belonging to 7,096 ASVs. After we removed ASVs with a
112 proportion of $> 0.1\%$ of reads present in the sum of three negative controls and taxonomically
113 assigned the ASVs, we kept 994,251 (44% of total) sequences belonging to 220 ASVs with at
114 least 75% similarity of one species in our reference database. As expected, most ASVs (189
115 ASVs in a total of 966,610 reads) belong to fishes, followed by mammals (26,127 reads
116 belonging to 21 ASVs), and only 10 ASVs (1,514 reads) assigned as birds (Fig. 1).

117 From the 21 ASVs assigned as mammal, 12 ASVs were assigned at species level (five
118 species including dog, cat, mouse, and cow), one at genus level, seven at family level, and one
119 only as a mammal. From the ten ASVs assigned as birds, seven were assigned at species level
120 (six species, including a *Corvus sp.*), one at genus level, and two at family level (Table S1). Most
121 non-fish ASV were recorded in 2019, with higher abundance in the Parana River and the higher
122 number of ASVs in the Brasilia stream locality (Fig. 2).

123

124 **4. Discussion:**

125 Our results demonstrate that environmental DNA metabarcoding by-catch is a viable source
126 of genetic information for non-target species. Indeed, this is the second study to explore this
127 possibility (Mariani et al., 2021), and the first in the Neotropics. Interestingly, contrary to that
128 study, our data recorded mostly farm/urban animals, such as cat, dog, rat, cow, and duck. It is
129 interesting to note that these records were detected in samples from 2019, whereas in 2020
130 almost no sequences of non-target organisms were found. In 2020 there was an extreme drought

131 in southeastern Brazil (de Oliveira Bueno et al., 2020), which potentially impacted fish
132 assemblages (Dal Pont et al., 2021) and probably other organisms present in the region as well.
133 Another relevant result is the lower number ASVs recorded in the reservoir site, probably
134 associated with the high water volume near the Itaipu Hydroelectric Power Plant dam and, then
135 low eDNA density.

136 A unique native non-farm / urban mammal species recorded was the capybara *Hydrochoerus*
137 *hydrochaeris*. Capybara is the largest living rodent of the world (Nowak and Walker, 1999) with
138 a semi-aquatic habit (Corriale and Herrera, 2014) and occurring throughout most of South
139 America (Moreira et al., 2013), being common in the region (Corriale and Herrera, 2014; Dias et
140 al., 2020). Other mammals were only assigned at the family level, including other rodents
141 (Cricetidae and Hydrocharidae), one bat (Phyllostomidae), and one carnivore (Procyonidae), that
142 is a New World family (Duszynski et al., 2018). Although two species of Procyonidae, *Nasua*
143 *nasua* and *Procyon cancrivorous*, are common in the region (Brocardo et al., 2019), these
144 species were present in our reference database matching with a low similarity. These sequences
145 may have low similarity due to the lack of publicly available representative sequences of the
146 species which them belong, highlighting the need of further studies in the Neotropical region.

147 For birds, beyond the very common non-native species house sparrow (*Passer*
148 *domesticus*) and duck (*Cairina moschata*), we recorded four additional species. From these, three
149 are native and common species, demonstrating the potential of eDNA of aquatic samples to
150 record non-aquatic bird species. One species we found was a crow (*Corvus* sp.) that is non-native
151 of South America (Burton and Burton, 2002). The corvid species occurring in the region is
152 *Cyanocharax chrysops* (<http://www.ultimaarcadenoe.com.br/aves-do-pq-nacional-do-iguacu/>),
153 and although no sequence for this species is available other species of the genera had it and

154 match with very low similarity, being the *Corvus* sp. the most probable species. To the best of
155 our knowledge, only one species of crow is known to occur in Brazil, *Corvus albus*, which was
156 reported for the first time in 2004 (Silva e Silva and Olmos, 2007), followed by additional
157 records (Adelino et al., 2017; Lima and Kamada, 2009). This species is considered a native
158 invader in its home range that has benefited from human infrastructure (Cunningham et al.,
159 2016) and is a potentially invasive species in Brazil (Adelino et al., 2017). It is alarming, since
160 invasive species are detrimental to both biodiversity, ecosystem process, human welfare, and
161 economy (Blackburn et al., 2014). In particular, *C. albus* has established populations outside
162 their native range in several countries, where they are responsible for ecological impacts (Ryall,
163 1992), economic loss (Kamel, 2014), and human health problems (Yap and Sodhi, 2004).
164 Although the record derived from metabarcoding data targeting fishes should be considered
165 carefully, it indicates a possible occurrence of a potentially invasive species that deserves to be
166 further investigated.

167

168 **5. Conclusions:**

169 Metabarcoding studies generate hundreds to thousands of non-target sequences. Although
170 this data is severely biased, it can contain important biological information. It is important to
171 note that it is unlikely that environmental DNA metabarcoding by-catch will provide sufficient
172 information for comprehensive surveys and diversity estimates of non-target species. However,
173 we envision two particularly useful applications of this approach. First, it might provide valuable
174 information on population fluctuations of the most common species that live close or associated
175 to bodies of water, such as the capybara. Second, it might show general trends in the anthropic
176 influence in the region, as evidence of native species might be slowly replaced by domestic ones.

177 In addition, we provide evidence for the potential presence on an invasive species that may be
178 controlled before it becomes a major problem. Also, we showed that aquatic samples are suitable
179 to detect Neotropical bird species. The use of primers specific for birds in aquatic samples can
180 optimize sampling in highly diverse and remote areas.

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187 **Declaration of Interest Statement:**

188 The authors declare no conflict of interest.

189

190 **Author Contribution Statement:**

191 M.P. study conceptualization, G.D.P, A.H, N.C., O.S.M.N., A.O., MP. experimental design.
192 A.O.A, P.V.S., and E.B. molecular analysis. C.D.R. bioinformatics and statistical analysis. A.O.
193 an M.P. fund-raising. C.D.R. wrote the first version with the collaboration of all authors. All
194 authors read and approved the final version.

195

196 **References**

197 Adelino, J.R.P., dos Anjos, L., Lima, M.R., 2017. Invasive potential of the pied crow (*Corvus*
198 *albus*) in eastern Brazil: best to eradicate before it spreads. *Perspect. Ecol. Conserv.* 15,
199 227–233.

200 Alberti, A., Poulain, J., Engelen, S., Labadie, K., Romac, S., Ferrera, I., Albini, G., Aury, J.-M.,
201 Belser, C., Bertrand, A., 2017. Viral to metazoan marine plankton nucleotide sequences
202 from the Tara Oceans expedition. *Sci. data* 4, 1–20.

203 Andruszkiewicz, E.A., Starks, H.A., Chavez, F.P., Sassoubre, L.M., Block, B.A., Boehm, A.B.,
204 2017. Biomonitoring of marine vertebrates in Monterey Bay using eDNA metabarcoding.
205 *PLoS One* 12, e0176343.

206 Andújar, C., Arribas, P., Gray, C., Bruce, C., Woodward, G., Yu, D.W., Vogler, A.P., 2018.
207 Metabarcoding of freshwater invertebrates to detect the effects of a pesticide spill. *Mol.*
208 *Ecol.* 27, 146–166.

209 Benson, D.A., Cavanaugh, M., Clark, K., Karsch-Mizrachi, I., Ostell, J., Pruitt, K.D., Sayers,
210 E.W., 2018. GenBank. *Nucleic Acids Res.* 46, D41–D47.

211 Blackburn, T.M., Essl, F., Evans, T., Hulme, P.E., Jeschke, J.M., Kühn, I., Kumschick, S.,
212 Marková, Z., Mrugała, A., Nentwig, W., 2014. A unified classification of alien species
213 based on the magnitude of their environmental impacts. *PLoS Biol.* 12, e1001850.

214 Brocardo, C.R., da Silva, M.X., Ferracioli, P., Cândido-Jr, J.F., Bianconi, G.V., Moraes, M.F.D.,
215 Galetti, M., Passamani, M., Policena, A., dos Reis, N.R., 2019. Mamíferos do Parque
216 Nacional do Iguaçu. *Oecologia Aust.* 23.

217 Burgess, R., 2001. An improved protocol for separating meiofauna from sediments using
218 colloidal silica sols. *Mar. Ecol. Prog. Ser.* 214, 161–165.

219 Burton, M., Burton, R., 2002. *International Wildlife Encyclopedia: Index Volume*. Marshall
220 Cavendish.

221 Callahan, B.J., McMurdie, P.J., Rosen, M.J., Han, A.W., Johnson, A.J.A., Holmes, S.P., 2016.
222 DADA2: high-resolution sample inference from Illumina amplicon data. *Nat. Methods* 13,
223 581–583.

224 Camacho, C., Coulouris, G., Avagyan, V., Ma, N., Papadopoulos, J., Bealer, K., Madden, T.L.,
225 2009. BLAST+: architecture and applications. *BMC Bioinformatics* 10, 421.
226 <https://doi.org/10.1186/1471-2105-10-421>

227 Closek, C.J., Santora, J.A., Starks, H.A., Schroeder, I.D., Andruszkiewicz, E.A., Sakuma, K.M.,
228 Bograd, S.J., Hazen, E.L., Field, J.C., Boehm, A.B., 2019. Marine vertebrate biodiversity
229 and distribution within the central California Current using environmental DNA (eDNA)
230 metabarcoding and ecosystem surveys. *Front. Mar. Sci.* 6, 732.

231 Collins, R.A., Bakker, J., Wangensteen, O.S., Soto, A.Z., Corrigan, L., Sims, D.W., Genner,
232 M.J., Mariani, S., 2019. Non- \square specific amplification compromises environmental DNA
233 metabarcoding with COI. *Methods Ecol. Evol.* 10, 1985–2001.

234 Corriale, M.J., Herrera, E.A., 2014. Patterns of habitat use and selection by the capybara
235 (*Hydrochoerus hydrochaeris*): a landscape-scale analysis. *Ecol. Res.* 29, 191–201.

236 Cunningham, S.J., Madden, C.F., Barnard, P., Amar, A., 2016. Electric crows: powerlines,
237 climate change and the emergence of a native invader. *Divers. Distrib.* 22, 17–29.

238 Dal Pont, G., Ritter, C.D., Agostinis, A.O., Stika, P.V., Horodesky, A., Cozer, N., Balsanelli, E.,
239 Netto, O.S.M., Henn, C., Ostrensky, A., 2021. Monitoring fish communities through DNA
240 metabarcoding in the fish pass system of the second largest hydropower plant in the world.
241 bioRxiv.

242 de Oliveira Bueno, E., Alves, G.J., Mello, C.R., 2020. Hydroelectricity water footprint in Parana
243 hydrograph region, Brazil. *Renew. Energy* 162, 596–612.

244 Dias, T.C., Stabach, J.A., Huang, Q., Labruna, M.B., Leimgruber, P., Ferraz, K.M., Lopes, B.,
245 Luz, H.R., Costa, F.B., Benatti, H.R., 2020. Habitat selection in natural and human-
246 modified landscapes by capybaras (*Hydrochoerus hydrochaeris*), an important host for
247 *Amblyomma sculptum* ticks. *PLoS One* 15, e0229277.

248 Duszynski, D.W., Kvicerova, J., Seville, R.S., 2018. The biology and identification of the
249 Coccidia (Apicomplexa) of carnivores of the world. Academic Press.

250 Harper, L.R., Handley, L.L., Carpenter, A.I., Ghazali, M., Di Muri, C., Macgregor, C.J., Logan,
251 T.W., Law, A., Breithaupt, T., Read, D.S., 2019. Environmental DNA (eDNA)
252 metabarcoding of pond water as a tool to survey conservation and management priority
253 mammals. *Biol. Conserv.* 238, 108225.

254 José, L., Yagui, H., Tejeda, V., Bonifaz, E., Bellemain, E., Valentini, A., Tobler, M.W.,
255 SÁNCHEZ-VENDIZ, P., Lyet, A., 2021. Environmental DNA metabarcoding as a useful
256 tool for evaluating terrestrial mammal diversity in tropical forests. *Ecol. Appl.* 31, e02335.

257 Kamel, A.M., 2014. Potential impacts of invasive house crows (*Corvus splendens*) bird species
258 in Ismailia Governorate, Egypt; ecology, control and risk management. *J. Life Sci. Technol*
259 2, 86–89.

260 Leese, F., Sander, M., Buchner, D., Elbrecht, V., Haase, P., Zizka, V.M.A., 2020. Improved
261 freshwater macroinvertebrate detection from eDNA through minimized non-target
262 amplification. *bioRxiv*.

263 Lima, B., Kamada, B., 2009. Registros de corvo-bicolor *Corvus albus* (Passeriformes: Corvidae)
264 em território brasileiro. *Atualidades Ornitológicas* 150, 10–11.

265 Mariani, S., Harper, L.R., Collins, R.A., Baillie, C., Wangensteen, O.S., McDevitt, A.D.,
266 Heddell-Cowie, M., Genner, M.J., 2021. Estuarine molecular bycatch as a landscape-wide
267 biomonitoring tool. *Biol. Conserv.* 261, 109287.
268 <https://doi.org/10.1016/J.BIOCON.2021.109287>

269 Martin, M., 2011. Cutadapt removes adapter sequences from high-throughput sequencing reads.
270 *EMBnet. J.* 17, 10–12.

271 McElroy, M.E., Dressler, T.L., Titcomb, G.C., Wilson, E.A., Deiner, K., Dudley, T.L., Eliason,
272 E.J., Evans, N.T., Gaines, S.D., Lafferty, K.D., 2020. Calibrating environmental DNA
273 metabarcoding to conventional surveys for measuring fish species richness. *Front. Ecol.*
274 *Evol.* 8, 276.

275 McMurdie, P.J., Holmes, S., 2013. phyloseq: an R package for reproducible interactive analysis
276 and graphics of microbiome census data. *PLoS One* 8, e61217.

277 Mikryukov, V., 2019. metagMisc: miscellaneous functions for metagenomic analysis.

278 Miya, M., Gotoh, R.O., Sado, T., 2020. MiFish metabarcoding: a high-throughput approach for
279 simultaneous detection of multiple fish species from environmental DNA and other
280 samples. *Fish. Sci.* 1–32.

281 Miya, M., Sato, Y., Fukunaga, T., Sado, T., Poulsen, J.Y., Sato, K., Minamoto, T., Yamamoto,
282 S., Yamanaka, H., Araki, H., 2015. MiFish, a set of universal PCR primers for
283 metabarcoding environmental DNA from fishes: detection of more than 230 subtropical
284 marine species. *R. Soc. open Sci.* 2, 150088.

285 Moreira, J.R., Alvarez, M.R., Tarifa, T., Pacheco, V., Taber, A., Tirira, D.G., Herrera, E.A.,
286 Ferraz, K.M.P.M.B., Aldana-Domínguez, J., Macdonald, D.W., 2013. Taxonomy, natural
287 history and distribution of the capybara, in: *Capybara*. Springer, pp. 3–37.

288 Nowak, R.M., Walker, E.P., 1999. *Walker's Mammals of the World*. JHU press.

289 Pedersen, T.L., 2019. Patchwork: The composer of plots. R Packag. version 1.

290 R Core Team, 2021. R: A Language and Environment for Statistical Computing. URL
291 <https://www.r-project.org/> [accessed 2020-03-30].

292 Ryall, C., 1992. Predation and harassment of native bird species by the Indian house crow,
293 *Corvus splendens*, in Mombasa, Kenya. *Scopus* 16, 1–8.

294 Sales, N.G., Kaizer, M. da C., Coscia, I., Perkins, J.C., Highlands, A., Boubli, J.P., Magnusson,
295 W.E., Da Silva, M.N.F., Benvenuto, C., McDevitt, A.D., 2020. Assessing the potential of
296 environmental DNA metabarcoding for monitoring Neotropical mammals: a case study in
297 the Amazon and Atlantic Forest, Brazil. *Mamm. Rev.* 50, 221–225.

298 Sales, N.G., Wangensteen, O.S., Carvalho, D.C., Deiner, K., Präbel, K., Coscia, I., McDevitt,
299 A.D., Mariani, S., 2021. Space-time dynamics in monitoring neotropical fish communities
300 using eDNA metabarcoding. *Sci. Total Environ.* 754, 142096.

301 Seymour, M., Edwards, F.K., Cosby, B.J., Kelly, M.G., de Bruyn, M., Carvalho, G.R., Creer, S.,

302 2020. Executing multi-taxa eDNA ecological assessment via traditional metrics and
303 interactive networks. *Sci. Total Environ.* 729, 138801.

304 Shokralla, S., Porter, T.M., Gibson, J.F., Dobosz, R., Janzen, D.H., Hallwachs, W., Golding,
305 G.B., Hajibabaei, M., 2015. Massively parallel multiplex DNA sequencing for specimen
306 identification using an Illumina MiSeq platform. *Sci. Rep.* 5, 1–7.

307 Sievert, C., 2020. Interactive web-based data visualization with R, plotly, and shiny. CRC Press.

308 Silva e Silva, R., Olmos, F., 2007. Adendas e registros significativos para a avifauna dos
309 manguezais de Santos e Cubatão, SP. *Rev. Bras. Ornitol.* 15, 551–560.

310 Stat, M., Huggett, M.J., Bernasconi, R., DiBattista, J.D., Berry, T.E., Newman, S.J., Harvey,
311 E.S., Bunce, M., 2017. Ecosystem biomonitoring with eDNA: metabarcoding across the tree
312 of life in a tropical marine environment. *Sci. Rep.* 7, 12240.

313 Van Rossum, G., Drake, F.L., 2009. Python 3 References Manual. Scotts Valley CA:
314 CreateSpace.

315 Wickham, H., 2017. tidyverse: Easily Install and Load “Tidyverse” Packages (Version R
316 package version 1.1. 1).

317 Wickham, H., 2016. ggplot2: elegant graphics for data analysis. Springer.

318 Yang, J., Zhang, X., Jin, X., Seymour, M., Richter, C., Logares, R., Khim, J.S., Klymus, K.,
319 2021. Recent advances in environmental DNA-based biodiversity assessment and
320 conservation. *Divers. Distrib.* 27, 1876–1879.

321 Yap, C.A.M., Sodhi, N.S., 2004. Southeast Asian invasive birds: ecology, impact and
322 management. *Ornithol. Sci.* 3, 57–67.

Figures

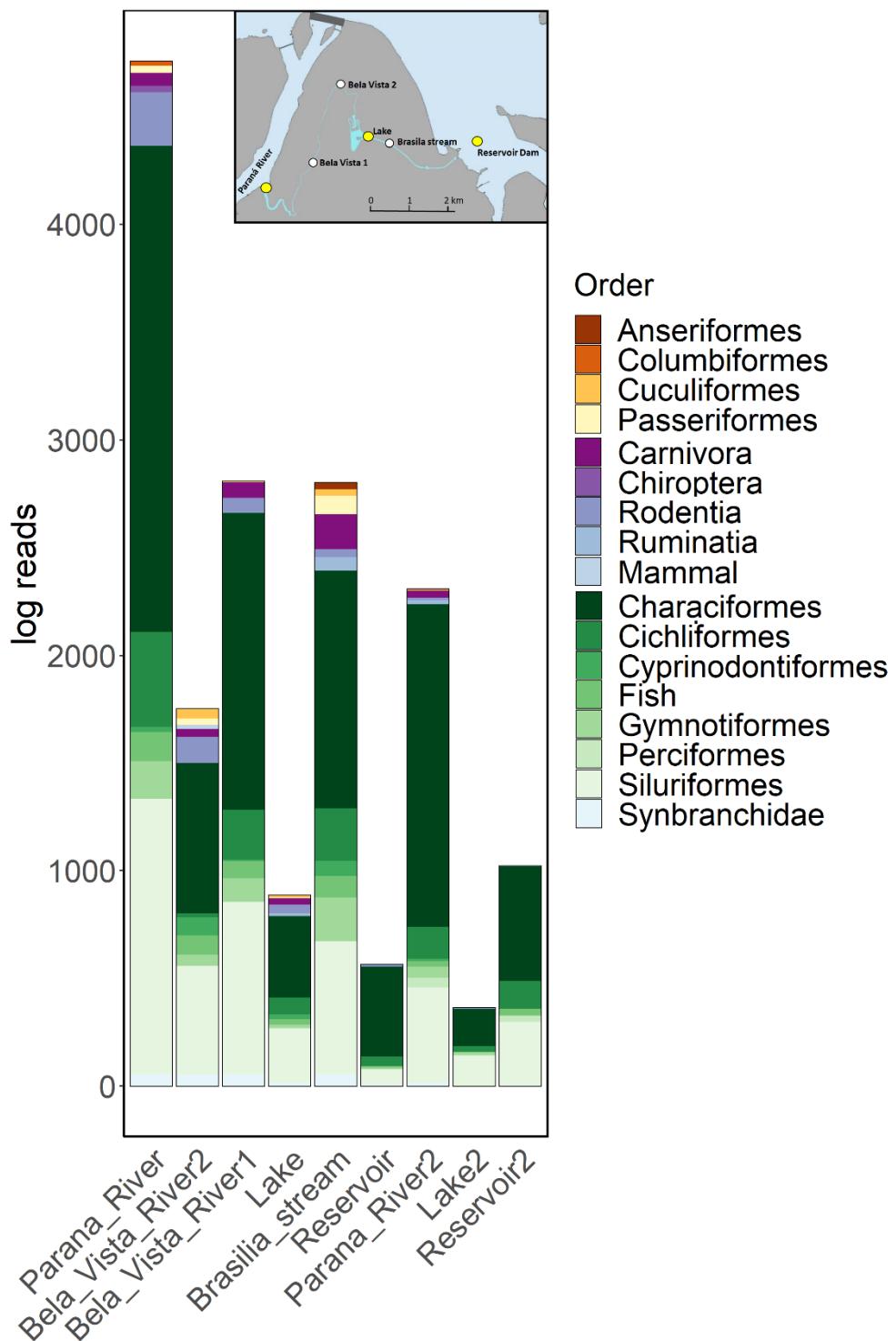


Figure 1. Taxonomic composition by order and class in Piracema fish pass. Inset panel show the point location from left to right as the bars order sampled in 2019. Point in yellow were re-sampled in 2020 (last three bars).

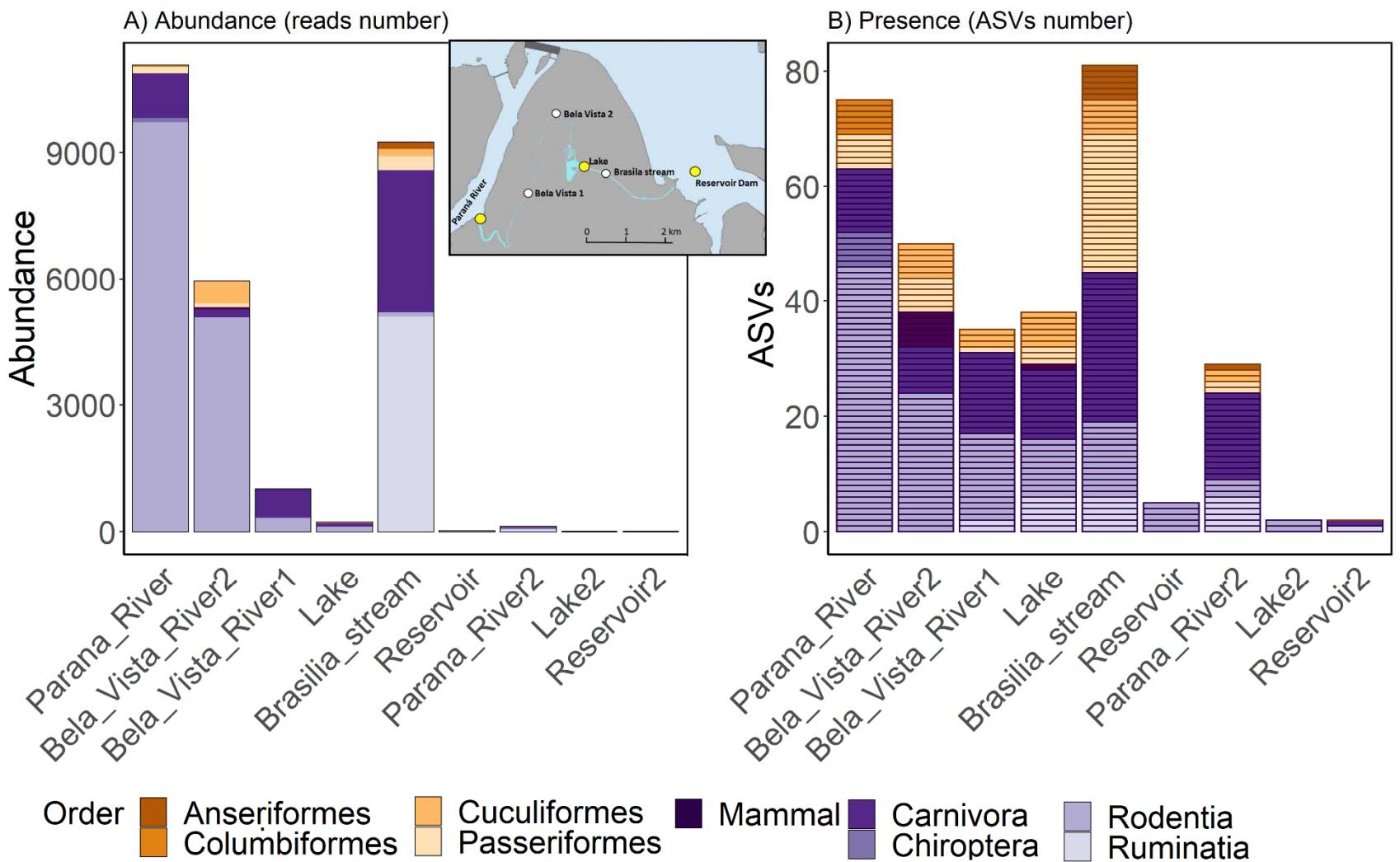


Figure 2. Taxonomic composition order abundance (A) and ASVs number (B) for birds and mammals registered in the Piracema fish pass. Inset panel show the point location from left to right as the bars order sampled in 2019. Point in yellow were re-sampled in 2020 (last three bars). One ASV in Bela Vista River 2 point assigned as mammal could not be assigned by any order by the low similarity (79.6%) and was kept as “mammal”.