

Zika virus-induced TNF- α signaling dysregulates expression of neurologic genes associated with psychiatric disorders

Po-Lun Kung¹, Tsui-Wen Chou¹, Marissa Lindman¹, Nydia P. Chang¹, Benjamin D. Buckley¹, Colm Atkins¹, and Brian P. Daniels¹⁺

¹Department of Cell Biology and Neuroscience, Rutgers University, Piscataway, NJ, USA

+Correspondence:

Brian Daniels
604 Allison Road
Room B314
Piscataway, NJ 08854
(848) 445-2709
b.daniels@rutgers.edu

Abstract

Zika virus (ZIKV) is an emerging flavivirus of global concern. ZIKV infection of the central nervous system has been linked to a variety of clinical syndromes, including microcephaly in fetuses and rare but serious neurologic disease in adults. However, the potential for ZIKV to influence brain physiology and host behavior following recovery from apparently mild or subclinical infection is less well understood. Furthermore, though deficits in cognitive function are well-documented following recovery from neuroinvasive viral infection, the potential impact of ZIKV on other host behavioral domains has not been thoroughly explored. In our study, we performed transcriptomic profiling of primary neuron cultures following ZIKV infection, which revealed altered expression of key genes associated with major psychiatric disorders, such as bipolar disorder and schizophrenia. Gene ontology enrichment analysis also revealed significant changes in gene expression associated with fundamental neurobiological processes, including neuronal development, neurotransmission, and others. These alterations to neurologic gene expression were also observed in the brain *in vivo* using an immunocompetent mouse model of ZIKV infection. Mechanistic studies identified TNF- α signaling via TNFR1 as a major regulatory mechanism controlling ZIKV-induced changes to neurologic gene expression. Our studies reveal that cell-intrinsic innate immune responses to ZIKV infection profoundly shape neuronal transcriptional profiles, highlighting the need to further explore associations between ZIKV infection and disordered host behavioral states.

Introduction

Zika virus (ZIKV) is a mosquito-borne pathogen of global concern (1). Like many other members of the genus *Flaviviridae*, ZIKV is both neuroinvasive and neurotropic (2). Infection of the central nervous system has been linked to diverse clinical syndromes, including severe congenital neurodevelopmental abnormalities in infants following vertical infection *in utero* (3-5). Severe neurologic disease is less frequent in adults, though cases of encephalitis, myelitis, and, more commonly, peripheral neuropathy have been reported (2, 6). While research on ZIKV pathogenesis to date has heavily focused on severe neurologic disease, it remains unclear whether ZIKV accesses the central nervous system during mild and/or subclinical infection, though data from animal models suggest this is probable (7-9). Even in the case of established neuroinvasive infection, the long-term neurologic consequences that follow viral clearance and recovery remain poorly understood. However, recent evidence suggests that a range of potential neurologic sequalae may occur in the postinfectious brain, including changes to host behavior (10-13).

In particular, recovery from neuroinvasive infection by flaviviruses, including ZIKV, has been associated with neurocognitive deficits (14-17). These effects have been attributed in part to the activities of immune cells, including both T cells and microglia, which act in concert to aberrantly prune neuronal synapses following flavivirus recovery (18-20). Flavivirus infection has also been shown to alter neurodevelopmental programs (21-24), including adult neurogenesis (25-27), a feature of flavivirus infection that contributes to altered learning and memory following recovery in rodent models (28). Cognitive decline, including persistent memory loss, is also a common occurrence in human patients recovering from flavivirus encephalitis (29-31).

Despite these insights into the cognitive consequences of flavivirus infection, the potential impact of these viruses on other behavioral domains remains relatively unexplored. The multifaceted impact of neurotropic flaviviruses on a diverse array of neurologic functions suggests that such infections may also promote or exacerbate neuropsychiatric conditions, including mood and psychotic disorders. Indeed, depression is another common behavioral symptom reported in patients recovering from flavivirus infection (32-34). Case reports have also documented the appearance of psychotic symptoms, including hallucinations, in adult patients infected with ZIKV (35, 36). However, the cellular and molecular mechanisms that underlie these effects remain unknown. In particular, the potential for ZIKV infection to impact host behavior due to cell intrinsic effects on neuronal gene expression has been relatively unexplored.

In this study, we examined how ZIKV infection in neurons impacted the expression of key neurologic genes that promote homeostatic neural function, as well as genes associated with disordered behavioral states. Transcriptomic profiling of primary cortical neurons following ZIKV infection revealed altered expression of a large number of genes associated with psychiatric disorders, including autism, depression, and schizophrenia. Moreover, unbiased gene ontology enrichment analysis revealed that ZIKV infection disproportionately impacted expression of genes associated with neurotransmission and neurodevelopment. These patterns of altered gene expression were also observed *in vivo*, using an established model of CNS ZIKV infection in immunocompetent mice. Observed changes in gene expression were due, at least in part, to innate cytokine signaling via tumor necrosis factor receptor-1 (TNFR1). Our data describe a mechanism linking the cell intrinsic innate immune response to ZIKV with dysregulation of a diverse array of neurologic gene pathways, opening new avenues of inquiry into the effect of flavivirus infection on host behavior.

Materials and Methods

Viruses

ZIKV strain MR766 was originally provided by Dr. Andrew Oberst, University of Washington. Viral stocks were generated by infecting Vero cells (MOI 0.01) and harvesting supernatants at 72hpi. Viral titers of stocks were determined via plaque assay on Vero cells (ATCC, #CCL-81). Cells were maintained in DMEM (Corning #10-013-CV) supplemented with 10% Heat Inactivated FBS (Gemini Biosciences #100-106), 1% Penicillin-Streptomycin-Glutamine (Gemini Biosciences #400-110), 1% Amphotericin B (Gemini Biosciences #400-104), 1% Non Essential Amino Acids (Cytiva #SH30238.01), and 1% HEPES (Cytiva SH30237.01). Plaque assay basal media was 10X EMEM (Lonza # 12-684F) adjusted to 1X and supplemented with 2% Heat Inactivated FBS (Gemini Biosciences #100-106), 1% Penicillin-Streptomycin-Glutamine (Gemini Biosciences #400-110), 1% Amphotericin B (Gemini Biosciences #400-104), 1% Non Essential Amino Acids (Cytiva #SH30238.01), and 1% HEPES (Cytiva SH30237.01), 0.75% Sodium Bicarbonate (VWR #BDH9280) and 0.5% Methyl Cellulose (VWR #K390). Plaque assays were developed 4dpi by removal of overlay media and staining/fixation using 10% neutral buffered formalin (VWR #89370) and 0.25% crystal violet (VWR #0528).

Cell culture experiments

Primary cerebral cortical neurons were generated using E15 embryos as described (37). Cells were maintained on cell culture treated multiwell dishes supplemented by coating with 20 μ g/mL Poly-L-Lysine (Sigma-Aldrich, #9155). Neurobasal Plus + B-27 supplement was used for all experiments (Thermo-Fisher Scientific, #A3582901). All primary mouse cells were generated using pooled tissues derived from both male and female animals. For ZIKV infection experiments, primary neuron cultures were infected at an MOI of 0.1.

Quantitative real-time PCR

Total RNA from cultured cells was isolated with Qiagen RNeasy mini extraction kit (Qiagen, #74106) following the manufacturer's protocol. RNA concentration was measured with a Quick Drop device (Molecular Devices). cDNA was subsequently synthesized with qScript cDNA Synthesis Kit (Quantabio, #95048). qPCR was performed with SYBR Green Master Mix (Applied Biosystems, #A25742) using a QuantStudio5 instrument (Applied Biosystems). Cycle threshold (CT) values for analyzed genes were normalized to CT values of the housekeeping gene 18S ($CT_{Target} - CT_{18S} = \Delta CT$). Data were further normalized to baseline control values ($\Delta CT_{experimental} - \Delta CT_{control} = \Delta\Delta CT$). Primers were designed using Primer3 (<https://bioinfo.ut.ee/primer3/>) against murine genomic sequences. A list of primer sequences used in the study appear in **Supplemental Table 1**.

Murine model of ZIKV infection

C57BL/6J mice were bred in-house for all experiments. All animals were housed under pathogen-free conditions in the animal facilities in Nelson Biological Laboratories at Rutgers University. Both male and female mice were inoculated intracranially (10 μ l) with 10^4 PFU of ZIKV, as described previously (8). ZIKV strain MR-766 was used in all experiments.

Tissue preparation

All tissues harvested from mice were extracted following extensive cardiac perfusion with 30 mL of sterile PBS. Extracted tissues were weighed and homogenized using 1.0 mm diameter zirconia/silica beads (Biospec Products, #11079110z) in sterile PBS for ELISA (VWR #L0119) or TRI Reagent (Zymo, #R2050-1) for gene expression analysis. Homogenization was performed in an Omni BeadRupter Elite for 2 sequential cycles of 20 seconds at a speed of 4 m/s. Total RNA was extracted using Zymo Direct-zol RNA Miniprep kit, as per manufacturer instructions (Zymo, #R2051).

ELISA

A TNF- α sandwich ELISA kit (EBioscience, #MTA00B) was used for detection of cytokine levels in cell culture supernatants and brain tissue homogenates. Colorimetric reading of ELISA plates was performed with a microplate reader and Gen5 software (BioTek Instruments, Inc.).

Neutralizing-antibody studies

Neutralizing-antibody studies were performed after 30 minutes of pretreatment with purified anti-mouse TNFR1 (Invitrogen, # 16-1202-85) and anti-mouse IFNAR1 (Leinco, # I-400) antibodies. IgG isotype antibodies (eBioscience, # eBio299Arm; Leinco, # I-443) were used as controls.

Curation of psychiatric disorder-associated gene list

Genes associated with autism spectrum disorder (ASD) for our bioinformatics study were identified using the Sfari Gene database (<https://gene.sfari.org>) (38). The Sfari database includes a ranked list of genes with known associations to ASD. We included genes within the top 3 levels of evidential strength of association (syndromic, category 1, and category 2). All genes within this curation have at least two reported de novo likely-gene-disrupting mutations. We were unable to identify similar database resources for other psychiatric disorders. We thus assembled gene lists for additional disorders by consultation of recent and/or highly cited literature in these areas, including metaanalyses and systematic reviews. More information about our gene list can be found in **Supplemental Table 2**.

Bioinformatics and statistical analysis

Secondary analysis of our previously published microarray dataset (accession # GSE122121) was performed in GEO2R and the GO Enrichment Analysis tool (geneontology.org). Biological pathways were defined using the PANTHER (Protein Analysis Through Evolutionary Relationships) classification system. Corrected p values (false discovery rate) were determined using the Benjamini & Hochberg procedure. For molecular biology assays, two-way analysis of variance (ANOVA) with Sidak's correction for multiple comparisons was performed using GraphPad Prism Software v8 (GraphPad Software, San Diego, CA). $P < 0.05$ was considered statistically significant. Data points in all experiments represent biological replicates unless otherwise noted.

Results

We first assessed whether ZIKV infection in neurons resulted in altered expression of genes associated with abnormal and/or disordered behavioral states. To do so, we first generated a list of 676 genes that have been linked in previous studies to psychiatric disorders, including autism, attention deficit hyperactivity disorder (ADHD), bipolar disorder, major depressive disorder, and schizophrenia (39-86) (**Supplemental Table 2**). This list of genes includes a combination of known risk genes as well as genes associated with behavioral abnormalities in each of the above human disorders and related animal models. While we stress that the list is not designed to be comprehensive or definitive, it serves as a starting point for probing the behavioral consequences of neuronal ZIKV infection. We assessed the impact of ZIKV infection on the expression of these genes using a dataset previously published by our group and others in which primary cortical neurons derived from C57BL/6J mice were infected with 0.01 MOI ZIKV-MR766 (8). Gene expression in this study was profiled via microarray analysis at 24h following infection.

Cross-comparison of the differentially expressed genes (DEGs) from the microarray analysis with our curated gene list revealed that 181 out of 676 (26.8%) genes exhibited significant differential expression following ZIKV infection (**Figure 1A**). Of these significant DEGs, 85 (46.96%) were

downregulated by ZIKV infection, while 96 (53.04%) were upregulated by ZIKV infection. The 13 genes with the lowest p value in this analysis included *Nr4a2*, *Itgb3*, and *Slc7a5*, which were downregulated by ZIKV infection, along with *Nlgn1*, *Myt1l*, *Dpp6*, *Thsd7a*, *Hepcam*, *Syn3*, *Cacna1e*, *Met*, *Pyhin1*, and *Tor3a*, which were upregulated by ZIKV infection (**Figure 1B**). The functions of these genes are summarized in **Figure 1C**, and generally include synaptic function, ion channel physiology, and neurodevelopmental processes. None of the specific psychiatric disorder gene lists were significantly overrepresented among the list of significant DEGs (**Figure 1D**). Together, these data suggest that ZIKV infection in neurons dysregulates expression of a broad set of genes associated with abnormal or pathologic behavioral states.

To better understand the consequences of ZIKV infection on neurologic gene expression, we next took an unbiased approach by performing gene ontology (GO) enrichment analysis on the DEGs derived from our microarray dataset. Similar to our results using the curated psychiatric disorder-associated gene list, this unbiased analysis revealed significant enrichment of several GO terms related to neurotransmission, neuronal stress responses, and neurodevelopment, a subset of which are highlighted in **Figure 2A-B**. Notably, regulatory pathways influencing ion transport and ion homeostasis were particularly enriched in our dataset. We next questioned whether there were clear patterns in the direction of differential expression among the significantly enriched GO terms. Heatmaps depicting the expression of genes associated with several representative GO terms are shown in **Figure 2C-E**, each of which reveal a mixed set of both up- and down-regulated genes. These findings suggest that the impact of ZIKV infection on the transcriptional state of neurons likely involves complex alterations to a variety of fundamental neurobiological processes.

While our microarray data revealed profound alterations to neurologic gene expression in primary neuronal culture following ZIKV infection, we next wanted to assess whether similar changes to gene expression occur *in vivo*. To do so, we inoculated male and female wildtype (C57BL/6J) mice intracranially with 10^4 pfu ZIKV-MR766. We performed these studies in separate cohorts of adolescent (3 week old) and adult (8 week old) animals to account for potential differences in the expression of neurologic genes across development (**Figure 3A**). On days 2 and 4 following infection, we harvested brains and used qRT-PCR to assess the expression of a panel of genes derived from the top DEGs identified in our microarray analysis. This candidate gene panel included genes from both our curated psychiatric disorder-associated gene list, as well as DEGs from the highly enriched GO terms identified in Figure 2.

These experiments revealed that a majority of genes in our panel did exhibit differential expression at the whole-brain level following intracranial ZIKV infection *in vivo*. However, the magnitude and direction of differential expression exhibited complex patterns that differed across time post infection and between 3 week old and 8 week old animals. In particular, 3 week old animals exhibited a mix of significantly downregulated and upregulated expression of selected neurologic genes 2 days post infection (dpi), with a marked shift to primarily upregulated expression at 4dpi (**Figure 3B-C**). In contrast, 8 week old animals exhibited an essentially inverse pattern, with nearly uniform upregulation of significant DEGs at 2dpi, but a mixture of down- and up-regulated DEGs at 4dpi (**Figure 3D-E**). These patterns of differential expression were evident across GO terms, including neurogenesis (**Figure 3F**), neuropeptide signaling pathway (**Figure 3G**), and regulation of behavior (**Figure 3H**), as well as genes taken from our curated list of psychiatric disorder-associated genes (**Figure 3I**). Together, these data confirm that gene expression associated with disease-relevant neurologic pathways is significantly dysregulated following ZIKV infection of the brain *in vivo*, but the mechanisms that control these transcriptional responses are under complex regulation by factors that vary with host age.

We next questioned whether innate immune activation following ZIKV infection in neurons may be linked to observed changes in neurologic gene expression. To answer this, we returned to our gene ontology enrichment analysis to identify innate immune signaling pathways that were most significantly impacted by neuronal ZIKV infection. Well-known antiviral cytokine responses were among the most enriched pathways in this analysis, particularly those related to innate cytokines, including type I

interferon (IFN), interleukin (IL)-1 and IL-6, and tumor necrosis factor (TNF)- α (**Figure 4A-B**). Pathways related to signaling by the inflammatory transcription factor nuclear factor kappa B (NF- κ B) were also particularly enriched within the list of significant DEGs. We next confirmed that each of these major cytokine responses was induced in the brains of 8 week old mice following intracranial infection with ZIKV. Expression analysis via qRT-PCR showed that ZIKV infection induced significant upregulation of each of the cytokines analyzed, including *Ifna6*, *Ifnb*, *Ifng*, *Il1b*, *Il6*, and *Tnfa* (**Figure 4C-H**). Together, these data confirm that wildtype neurons mount a robust innate immune cytokine response to ZIKV infection, including several cytokines previously established to influence brain function and behavior.

To examine whether neuronal cytokines may be implicated in ZIKV-induced changes in neurologic gene expression, we generated primary cultures of cortical neurons and examined expression of targets in our candidate gene panel following treatment with exogenous cytokines, including IFN- β , IL-6, and TNF- α . We then compared the direction of differential expression for each gene to that induced by ZIKV to see which, if any, cytokine most closely phenocopied the pattern of gene expression induced by ZIKV infection. While IFN- β and IL-6 did significantly alter the expression of some target genes, significant DEGs following these treatments did not closely follow the pattern of downregulation (**Figure 5A**) or upregulation (**Figure 5B**) induced by ZIKV infection. In contrast, TNF- α treatment induced a strikingly similar pattern of differential expression to that induced by ZIKV infection. Moreover, while IFN- β and IL-6 only induced significant changes in expression of a handful of neurologic genes in our analysis (**Figure 5C-D**), TNF- α significantly altered 17 out of 18 genes in our panel (**Figure 5E**), and of these, all but 2 matched the pattern of up- or downregulated expression observed in ZIKV-infected neuronal cultures. These data identified TNF- α signaling as a promising candidate mechanism for the altered neurologic gene expression observed in the setting of neuronal ZIKV infection.

While we previously confirmed that *Tnfa* was induced at the transcriptional level in the brain *in vivo* following ZIKV infection, we next wanted to confirm that a robust TNF- α -dependent signature could indeed be observed following infection. We thus performed enzyme-linked immunosorbent assay (ELISA) to confirm that TNF- α was upregulated at the protein level in both supernatants of primary neuronal cultures at 24h following infection (**Figure 5F**) and whole brain homogenates derived from 8 week old animals on days 2 and 4 following intracranial ZIKV inoculation (**Figure 5G**). We also confirmed upregulation of known TNF- α transcriptional targets, including *Cd69*, *Ccl2*, *Ccl5*, and *Stat1*, in the brains of infected 8 week old mice (**Figure 5H-K**). In contrast, transcript expression of the TNF- α receptors TNFR1 (*Tnfrsf1a*) and TNFR2 (*Tnfrsf1b*) were not altered in the brain following infection (**Figure 5L-M**), suggesting that enhanced TNF- α signaling in this setting is mediated primarily through induction of cytokine expression. Together, these data confirm that TNF- α signaling is active in both cultured neurons and the brain *in vivo* following ZIKV infection.

To more carefully assess whether TNF- α signaling was required for ZIKV-mediated alterations to neurologic gene expression, we cultured primary cortical neurons and pretreated with neutralizing antibodies against cytokine receptors for 2h prior to infection. After 24h, we then performed qRT-PCR analysis of major DEGs from our microarray analysis to assess the impact of cytokine signaling on ZIKV-induced gene expression. These experiments revealed that blockade of type I IFN signaling via neutralization of the IFN α/β receptor (IFNAR) had no impact on ZIKV-induced changes in expression of the neurologic genes we analyzed (**Figure 5N**), findings which mirrored our previous result showing that exogenous IFN- β treatment did not phenocopy ZIKV-induced patterns of expression in our target gene list (**Figure 5A-B**). In contrast, blockade of TNFR1 rescued ZIKV-induced changes in each of the 6 genes we analyzed, including *Chac1*, *Nr4a2*, *Lhx6*, *Dpp6*, *Cacna1e*, and *Tor3a* (**Figure 5O**). Taken together, these data suggest that the induction of TNF- α following neuronal ZIKV infection is a major regulatory mechanism that alters expression of genes relevant to neuronal function.

Discussion

Emerging flaviviruses represent a significant and growing challenge to global public health. While most famously associated with rare but severe clinical manifestations, including encephalitis, congenital abnormalities, etc., the consequences of apparently mild and/or asymptomatic infection by neuroinvasive flaviviruses remain poorly understood (2, 87). The observation of behavioral sequelae following recovery from severe flavivirus infections raises the possibility that subclinical neuroinvasive infection may also impact brain function in ways that promote or exacerbate psychiatric disorders. This idea is supported by some case reports (15, 35, 88-90), though, to our knowledge, this hypothesis has not been rigorously tested in the clinical literature. The prevalence of psychiatric sequelae following ZIKV infection, in particular, may be hard to discern due to the relatively low neurovirulence of ZIKV compared to other flaviviruses, resulting in symptoms that may not be severe enough to warrant clinical attention and the documentation of infection status. Our study highlights the need for increased attention to behavioral symptoms in patients who are seropositive for ZIKV and other neuroinvasive flaviviruses, as well as further mechanistic investigation into the cellular and molecular impacts of flavivirus infection on brain physiology and function.

In our study, we show that neurons mount a robust innate cytokine response to ZIKV infection, including a number of cytokines with previously established effects on behavior. A large body of evidence has established that neuroinflammation and inflammatory cytokine signaling is associated with psychiatric disorders, including major depressive disorder (91-94) and schizophrenia (95-97). TNF- α is a major pleiotropic cytokine induced strongly in the CNS by ZIKV and other flaviviruses (98-101). Notably, recent work has described complex neuromodulatory effects of TNF- α signaling, including direct effects on glutamatergic neurotransmission (102-104), neuronal differentiation (105-107), and other fundamental neurologic processes (108-110). In our study, ZIKV-mediated changes to neurologic gene expression greatly overlapped those induced by TNF- α , and gene expression changes induced by ZIKV could be rescued in part by blockade of TNFR1 signaling. These data identify TNF- α as a candidate for further mechanistic investigation of the potential impacts of flavivirus infection on neuronal function.

To date, the most well-described behavioral outcomes of neuroinvasive flavivirus infection in animal models are changes to learning and memory (18, 19, 27, 111-113). While it is clear that a variety of pathogenic processes related both to viral infection and neuroinflammation can impact cognition, comparatively less attention has been devoted to how flavivirus infection impacts other behavioral domains, including mood, affect, and emotional regulation. This discrepancy is likely due, in part, to technical limitations, including difficulty modeling these behavioral domains in rodents and containment issues related to using ABSL2 and ABSL3 pathogens within behavioral laboratories. Nevertheless, our data identify a need for more robust assessment of behavioral changes in models of flavivirus infection, particularly measures of anxiety, fear/avoidance, and other paradigms with relevance to human psychiatric disorders.

Finally, our data add to a growing body of evidence suggesting that the impact of flavivirus infection varies across the lifespan. In our study, intracranial ZIKV infection resulted in very different impacts on neurologic gene expression in adolescent compared to adult animals, suggesting that factors such as developmental states and immune system maturation may significantly influence the neurologic outcomes of flavivirus infection. Recent work has shown that neuroimmune responses to other flaviviruses are impacted by aging (114, 115), and thus differential engagement of cytokine signaling, adaptive immune priming, and blood-brain barrier function may all be relevant variables in determining how flavivirus infection might impact behavior differentially across life stages. Moreover, while the potential for ZIKV to induce severe congenital abnormalities following vertical transmission *in utero* has now been well established, it remains less clear what the impact of ZIKV infection is on apparently developmentally normal fetuses, including those who are exposed to ZIKV late in gestation, when rates of microcephaly and severe birth defects are exceedingly rare (113, 116, 117). Further work will be

needed to assess whether ZIKV infection in this context may result in changes to neurodevelopment and brain function that impact behavior postnatally and beyond.

Declarations

Ethics Approval and Consent to Participate

All animal experiments were performed with approval of the Rutgers University Institutional Animal Care and Use Committee (IACUC).

Consent for Publication

Not applicable

Availability of Data and Materials

All data are available upon reasonable request to the corresponding author. Microarray data used in this study are deposited in NCBI's Gene Expression Omnibus and can be accessed under accession number GSE122121.

Competing Interests

The authors declare they have no competing interests.

Funding

This work was supported by NIH Grant R21 MH125034 and startup funds from Rutgers University (to BPD). PLK was supported in part by a Division of Life Sciences Summer Undergraduate Research Fellowship from Rutgers University.

Contributions

Conceptualization: PLK and BPD; Investigation and analysis: PLK, TWC, ML, NPC, BDB, CA, and BPD.; Writing- original draft: PLK and BPD; Writing- review and editing: PLK, TWC, NPC, CA, and BPD; Supervision: CA and BPD; Funding acquisition: BPD.

Acknowledgements

Not applicable.

Figures

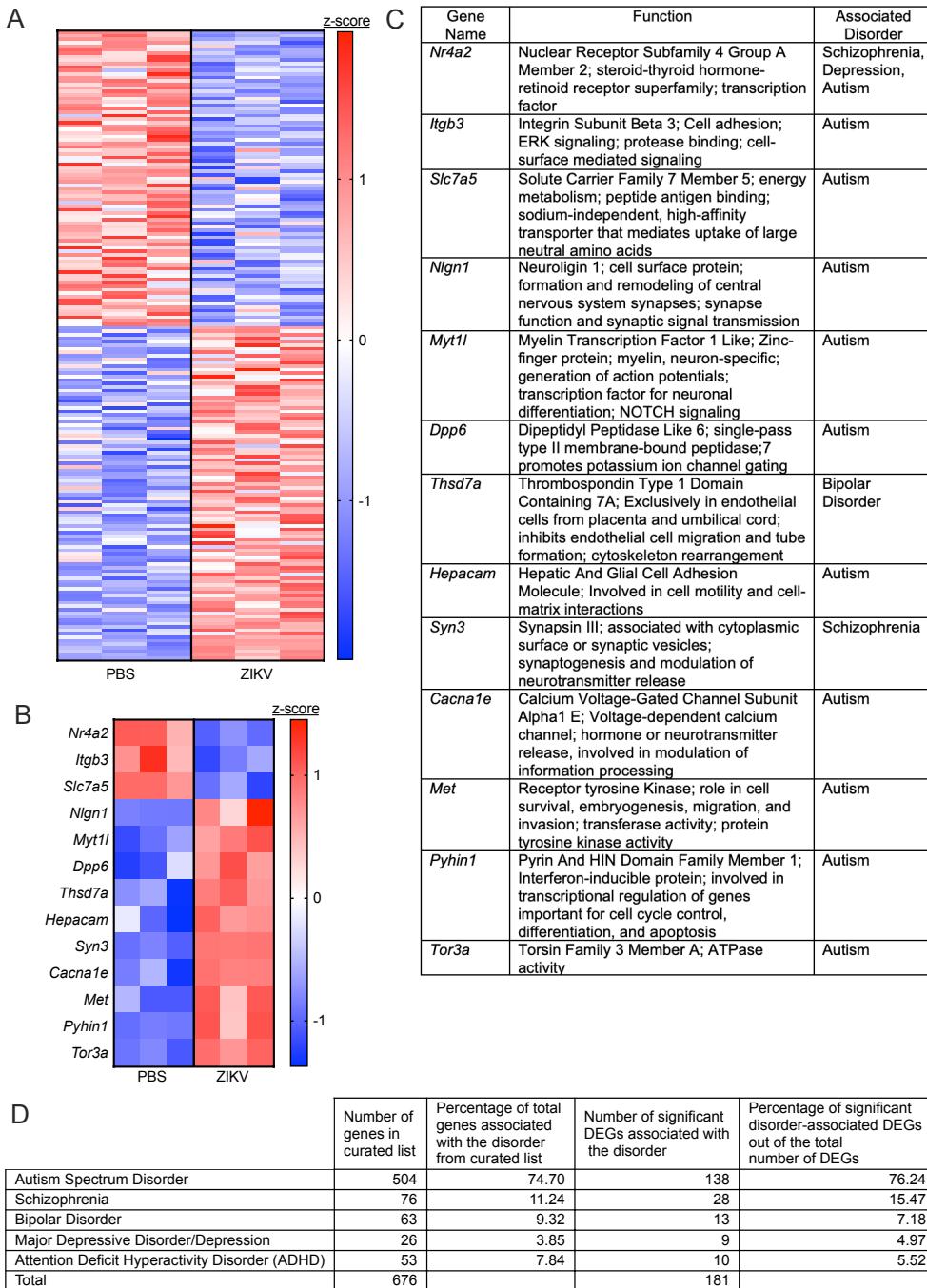
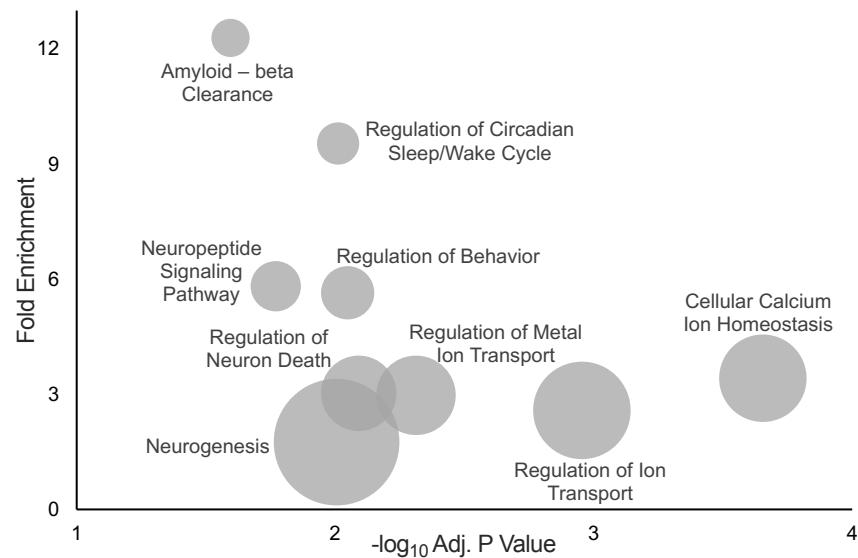


Figure 1: ZIKV infection in neurons dysregulates expression of a broad set of genes associated with abnormal or pathologic behavioral states. A) Heatmap depicting relative expression values of 181 candidate genes associated with psychiatric disorders. Values are derived from microarray analysis of primary cortical neurons 24h following ZIKV-MR766 infection (MOI 0.1) or PBS control treatment. **B-C)** Expression values (B) of the top 13 significant differentially expressed genes (DEGs) in our microarray analysis (identified by lowest p values). Table in (C) lists known functions and associated disorders for these genes. **D)** Descriptive statistics for the curated psychiatric disorder-associated gene list and the significant DEGs observed for each disorder. Data in (A) and (B) represent normalized and z-transformed expression values. Data in (A) include all genes with a False Discovery Rate (FDR) <0.1.

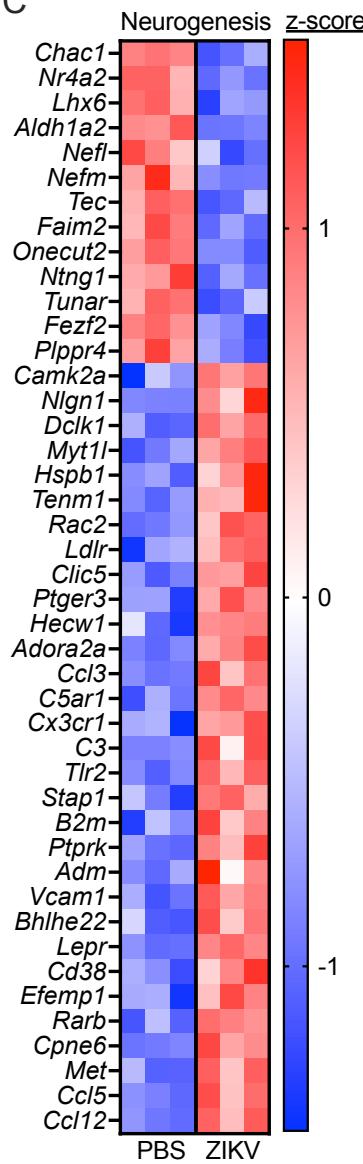
A

| | Number of Genes | Fold Enrichment | FD Rate (adj. P-value) |
|--|-----------------|-----------------|------------------------|
| Regulation of Ion Transport | 27 | 2.60 | 0.001110 |
| Cellular Calcium Ion Homeostasis | 22 | 3.43 | 0.000222 |
| Regulation of Metal Ion Transport | 18 | 2.99 | 0.004870 |
| Regulation of Neuron Death | 16 | 3.04 | 0.008140 |
| Regulation of Behavior | 8 | 5.65 | 0.008930 |
| Regulation of Circadian Sleep/Wake Cycle | 5 | 9.55 | 0.009770 |
| Neurogenesis | 45 | 1.77 | 0.009880 |
| Amyloid – beta Clearance | 4 | 12.29 | 0.025400 |
| Neuropeptide Signaling Pathway | 7 | 5.82 | 0.017000 |

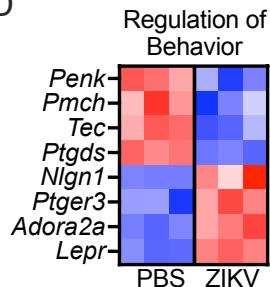
B



C



D



E

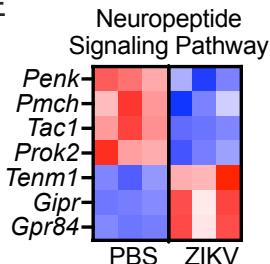


Figure 2: ZIKV infection impacts transcriptional pathways associated with fundamental neurobiological processes. A-B) Selected overrepresented GO terms obtained from GO enrichment analysis of DEGs resulting ZIKV-MR766 infection in primary cortical neurons. Tabular results (A) are graphically represented in a bubble plot (B) to demonstrate associations between fold enrichment, FDR, and number of associated genes for each GO term. **C-E)** Heatmaps depicting expression values for genes associated with neurogenesis (C), regulation of behavior (D), and neuropeptide signaling pathway (E). Data in (C), (D) and (E) represent normalized and z-transformed expression values.

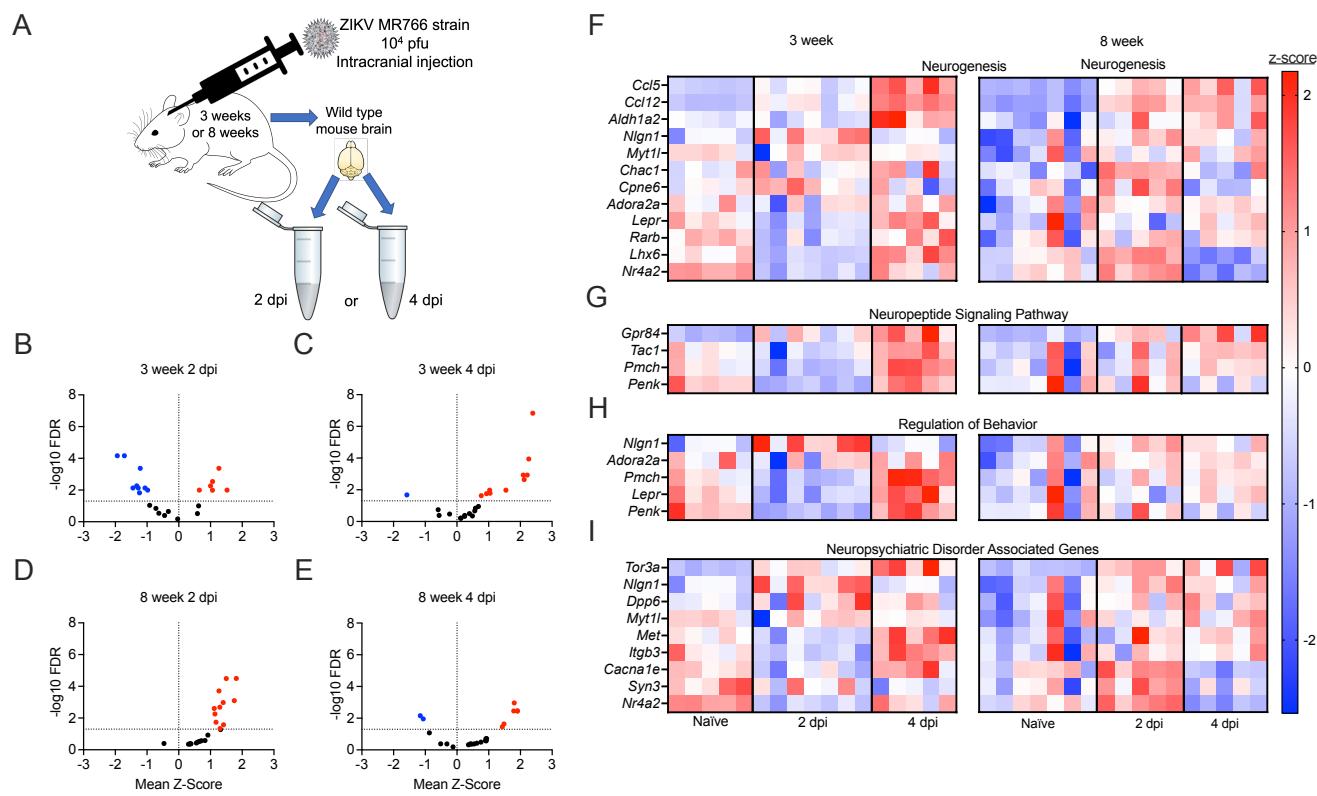
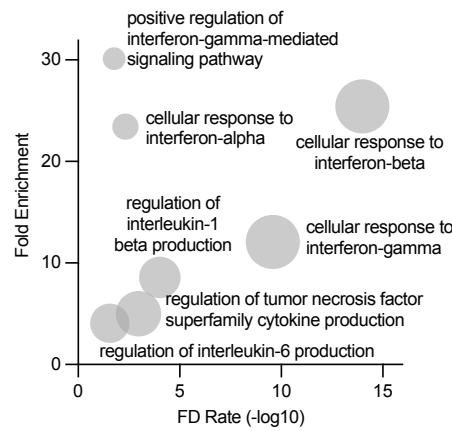


Figure 3: Gene expression associated with disease-relevant neurologic pathways is significantly dysregulated following ZIKV infection of the brain *in vivo*. **A)** Schematic of experimental design of *in vivo* murine model. C57/BL/6J WT mice were infected with 10⁴ pfu ZIKV-MR766 via intracranial inoculation at 3 and 8 weeks of age. Whole brain homogenates were collected at 2 or 4 days post infection (dpi). **B-E)** Expression profiles of 23 candidate genes assessed by qRT-PCR analysis are described and separated into 3-week mice at 2dpi (**B**), 3-week mice at 4dpi (**C**), 8-week mice at 2dpi (**D**), and 8-week mice at 4dpi (**E**). Significant differences ($p < 0.05$) are noted in red (upregulated DEGs) or blue (downregulated DEGs). **F-I)** Heatmaps showing individual expression values per mouse for candidate genes from microarray and GO enrichment analyses in 3 week old adolescent and 8 week old adult mouse brains following ZIKV-MR766 infection. Relative gene expression values are reported for naïve, 2 dpi, or 4 dpi groups. Data in (**F**), (**G**), (**H**) and (**I**) represent normalized and z-transformed values of qRT-PCR expression data. n= 5-7 mice/group.

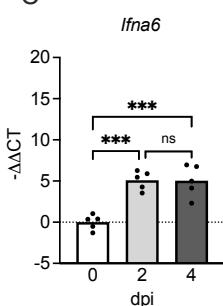
A

| Gene Ontology Term | Number of Associated Genes | Fold Enrichment | FD Rate (adj. P-value) |
|---|----------------------------|-----------------|------------------------|
| cellular response to interferon-beta | 17 | 25.41 | 1.03e-014 |
| positive regulation of cytokine production | 33 | 5.17 | 2.84e-011 |
| cellular response to interferon-gamma | 17 | 12.06 | 2.64e-010 |
| positive regulation of NF-kappaB transcription factor activity | 14 | 6.69 | 1.23e-005 |
| regulation of interleukin-1 beta production | 10 | 8.57 | 9.69e-005 |
| regulation of interleukin-1 production | 10 | 7.02 | 4.23e-004 |
| regulation of I-kappaB kinase/NF-kappaB signaling | 13 | 5.05 | 4.62e-004 |
| regulation of tumor necrosis factor superfamily cytokine production | 12 | 4.99 | 1.08e-003 |
| regulation of toll-like receptor signaling pathway | 8 | 8.39 | 1.09e-003 |
| chemokine-mediated signaling pathway | 7 | 9.11 | 2.16e-003 |
| cellular response to interferon-alpha | 4 | 23.42 | 4.80e-003 |
| positive regulation of toll-like receptor signaling pathway | 5 | 12.11 | 7.52e-003 |
| cellular response to tumor necrosis factor | 9 | 5.14 | 7.53e-003 |
| cellular response to interleukin-1 | 7 | 6.47 | 1.09e-002 |
| positive regulation of interferon-gamma-mediated signaling pathway | 3 | 30.11 | 1.72e-002 |
| regulation of interleukin-6 production | 9 | 4.05 | 2.80e-002 |

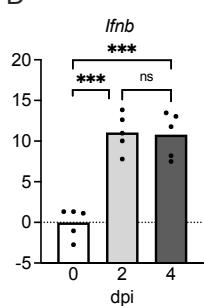
B



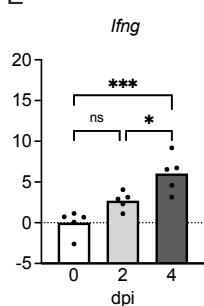
C



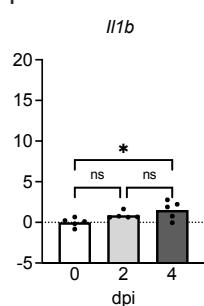
D



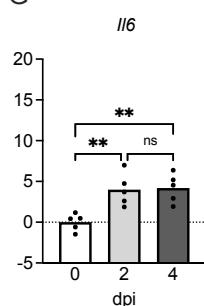
E



F



G



H

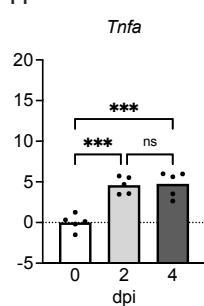


Figure 4: Neurons mount a robust innate immune cytokine response to ZIKV infection. A-B)

Results of GO enrichment analysis of microarray data derived from primary cortical neurons following ZIKV-MR766 infection (MOI 0.1) compared to PBS-treated controls after 24 hours. Selected GO terms focus on cytokine activation or inflammatory transcription factor responses. Tabular results (A) are graphically represented in a bubble plot (B) to demonstrate associations between fold enrichment, FDR, and number of associated genes for each GO term. C-H) qRT-PCR analysis was performed measuring cytokine genes *Ifna6* (C), *Ifnb* (D), *Ifng* (E), *Il1b* (F), *Il6* (G), and *Tnfa* (H) at indicated time points in 8 week old adult mouse brains following intracranial ZIKV-MR766 infection. n=5. ns not significant, * p < 0.05, ** p < 0.01, *** p < 0.001. Bars represent group means.

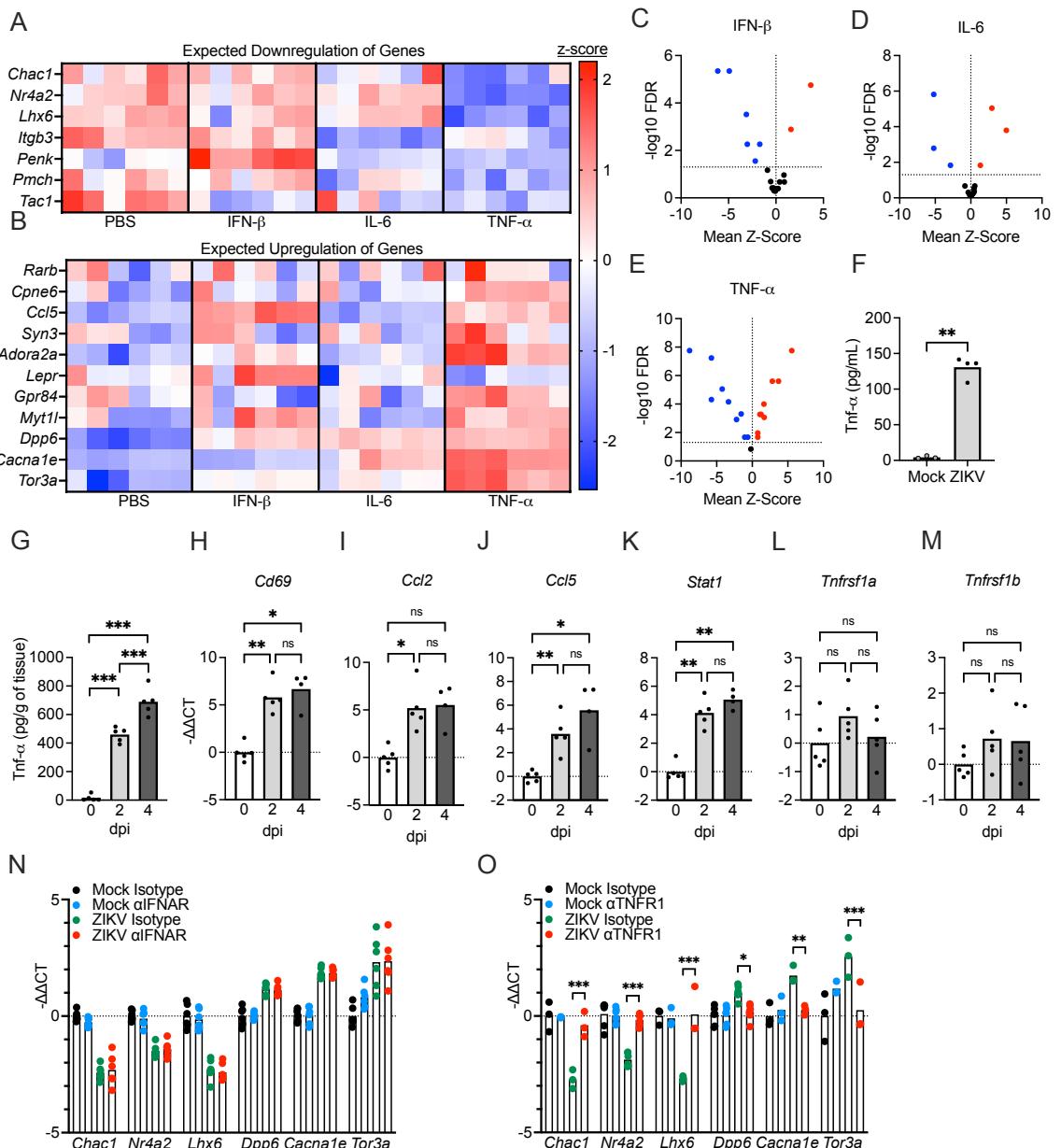


Figure 5: Induction of TNF- α following neuronal ZIKV infection is a major regulatory mechanism that alters expression of genes relevant to neuronal function. A-B) Expression values of candidate genes previously shown to be either downregulated (A) or upregulated (B) by ZIKV infection were measured via qRT-PCR in neuronal cultures treated for 24h with exogenous cytokines: IFN- β , IL-6, or TNF- α . n=6. **C-E)** Volcano plots depicting mean z-score and -log10FDR of representative DEGs in neurons treated with IFN- β (C), IL-6 (D), or TNF- α (E). Significant differences ($p < 0.05$) are noted in red (upregulated DEGs) or blue (downregulated DEGs). n=6. **F-G)** Concentrations of TNF- α in supernatants of *in vitro* neuronal cell cultures infected with ZIKV for 24h (n=4) (F) and brains harvested following *in vivo* intracranial infection (n=5) (G). Cytokine concentrations were quantified via ELISA assay. **H-M)** qRT-PCR analysis was performed for known TNF- α transcriptional target genes, Cd69 (H), Ccl2 (I), Ccl5 (J), Stat1 (K), and TNF- α receptor genes, Tnfrsf1a (L) and Tnfrsf1b (M), in 8 week old adult mouse brains following ZIKV-MR766 infection at 2 or 4 dpi. n=5. **N-O)** qRT-PCR analysis of representative DEGs associated with neurological functions (Chac1, Nr4a2, Lhx6, Dpp6, Cacna1e, and Tor3a) following pretreatment with neutralizing antibodies against IFN α / β receptor (IFNAR) (N) or TNFR1 (O) and subsequent 24h infection with ZIKV-MR766. n=3-6 biological replicates per group. Data in (A) and (B) represent normalized and z-transformed values of qRT-PCR expression data. ns not significant, * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$. Bars represent group means.

References

1. Pierson TC, Diamond MS. The continued threat of emerging flaviviruses. *Nat Microbiol*. 2020;5(6):796-812.
2. Munoz LS, Parra B, Pardo CA, Neuroviruses Emerging in the Americas S. Neurological Implications of Zika Virus Infection in Adults. *J Infect Dis*. 2017;216(suppl_10):S897-S905.
3. Freitas DA, Souza-Santos R, Carvalho LMA, Barros WB, Neves LM, Brasil P, et al. Congenital Zika syndrome: A systematic review. *PLoS One*. 2020;15(12):e0242367.
4. Marques VM, Santos CS, Santiago IG, Marques SM, Nunes Brasil MDG, Lima TT, et al. Neurological Complications of Congenital Zika Virus Infection. *Pediatr Neurol*. 2019;91:3-10.
5. Coyne CB, Lazear HM. Zika virus - reigniting the TORCH. *Nat Rev Microbiol*. 2016;14(11):707-15.
6. Schwartzmann PV, Ramalho LN, Neder L, Vilar FC, Ayub-Ferreira SM, Romeiro MF, et al. Zika Virus Meningoencephalitis in an Immunocompromised Patient. *Mayo Clin Proc*. 2017;92(3):460-6.
7. Gorman MJ, Caine EA, Zaitsev K, Begley MC, Weger-Lucarelli J, Uccellini MB, et al. An Immunocompetent Mouse Model of Zika Virus Infection. *Cell Host Microbe*. 2018;23(5):672-85 e6.
8. Daniels BP, Kofman SB, Smith JR, Norris GT, Snyder AG, Kolb JP, et al. The Nucleotide Sensor ZBP1 and Kinase RIPK3 Induce the Enzyme IRG1 to Promote an Antiviral Metabolic State in Neurons. *Immunity*. 2019;50(1):64-76 e4.
9. Berry N, Ferguson D, Ham C, Hall J, Jenkins A, Giles E, et al. High susceptibility, viral dynamics and persistence of South American Zika virus in New World monkey species. *Sci Rep*. 2019;9(1):14495.
10. Raper J, Kovacs-Balint Z, Mavigner M, Gumber S, Burke MW, Habib J, et al. Long-term alterations in brain and behavior after postnatal Zika virus infection in infant macaques. *Nat Commun*. 2020;11(1):2534.
11. Mavigner M, Raper J, Kovacs-Balint Z, Gumber S, O'Neal JT, Bhaumik SK, et al. Postnatal Zika virus infection is associated with persistent abnormalities in brain structure, function, and behavior in infant macaques. *Sci Transl Med*. 2018;10(435).
12. Nem de Oliveira Souza I, Frost PS, Franca JV, Nascimento-Viana JB, Neris RLS, Freitas L, et al. Acute and chronic neurological consequences of early-life Zika virus infection in mice. *Sci Transl Med*. 2018;10(444).
13. Adams Waldorf KM, Olson EM, Nelson BR, Little ME, Rajagopal L. The Aftermath of Zika: Need for Long-Term Monitoring of Exposed Children. *Trends Microbiol*. 2018;26(9):729-32.
14. Patel H, Sander B, Nelder MP. Long-term sequelae of West Nile virus-related illness: a systematic review. *Lancet Infect Dis*. 2015;15(8):951-9.
15. Zucker J, Neu N, Chiriboga CA, Hinton VJ, Leonardo M, Sheikh A, et al. Zika Virus-Associated Cognitive Impairment in Adolescent, 2016. *Emerg Infect Dis*. 2017;23(6):1047-8.
16. Ripamonti E, Gaffuri M, Molteni F. Cognitive, neuropsychiatric, and motor profile in post tick-borne flaviviral encephalomyelitis. *Neurol Sci*. 2020;41(12):3759-60.
17. Belaunzaran-Zamudio PF, Ortega-Villa AM, Mirenza-Alvarado AJ, Guerra-De-Blas PDC, Aguilar-Navarro SG, Sepulveda-Delgado J, et al. Comparison of the Impact of Zika and Dengue Virus Infection, and Other Acute Illnesses of Unidentified Origin on Cognitive Functions in a Prospective Cohort in Chiapas Mexico. *Front Neurol*. 2021;12:631801.
18. Vasek MJ, Garber C, Dorsey D, Durrant DM, Bollman B, Soung A, et al. A complement-microglial axis drives synapse loss during virus-induced memory impairment. *Nature*. 2016;534(7608):538-43.

19. Garber C, Soung A, Vollmer LL, Kanmogne M, Last A, Brown J, et al. T cells promote microglia-mediated synaptic elimination and cognitive dysfunction during recovery from neuropathogenic flaviviruses. *Nat Neurosci*. 2019;22(8):1276-88.
20. Figueiredo CP, Barros-Aragao FGQ, Neris RLS, Frost PS, Soares C, Souza INO, et al. Zika virus replicates in adult human brain tissue and impairs synapses and memory in mice. *Nat Commun*. 2019;10(1):3890.
21. Bayless NL, Greenberg RS, Swigut T, Wysocka J, Blish CA. Zika Virus Infection Induces Cranial Neural Crest Cells to Produce Cytokines at Levels Detrimental for Neurogenesis. *Cell Host Microbe*. 2016;20(4):423-8.
22. Rosa-Fernandes L, Cugola FR, Russo FB, Kawahara R, de Melo Freire CC, Leite PEC, et al. Zika Virus Impairs Neurogenesis and Synaptogenesis Pathways in Human Neural Stem Cells and Neurons. *Front Cell Neurosci*. 2019;13:64.
23. Brault JB, Khou C, Basset J, Coquand L, Fraisier V, Frenkiel MP, et al. Comparative Analysis Between Flaviviruses Reveals Specific Neural Stem Cell Tropism for Zika Virus in the Mouse Developing Neocortex. *EBioMedicine*. 2016;10:71-6.
24. Yoon KJ, Song G, Qian X, Pan J, Xu D, Rho HS, et al. Zika-Virus-Encoded NS2A Disrupts Mammalian Cortical Neurogenesis by Degrading Adherens Junction Proteins. *Cell Stem Cell*. 2017;21(3):349-58 e6.
25. Li H, Saucedo-Cuevas L, Regla-Nava JA, Chai G, Sheets N, Tang W, et al. Zika Virus Infects Neural Progenitors in the Adult Mouse Brain and Alters Proliferation. *Cell Stem Cell*. 2016;19(5):593-8.
26. Tang H, Hammack C, Ogden SC, Wen Z, Qian X, Li Y, et al. Zika Virus Infects Human Cortical Neural Progenitors and Attenuates Their Growth. *Cell Stem Cell*. 2016;18(5):587-90.
27. Soung AL, Dave VA, Garber C, Tycksen ED, Vollmer LL, Klein RS. IL-1 reprogramming of adult neural stem cells limits neurocognitive recovery after viral encephalitis by maintaining a proinflammatory state. *Brain Behav Immun*. 2021.
28. Garber C, Vasek MJ, Vollmer LL, Sun T, Jiang X, Klein RS. Astrocytes decrease adult neurogenesis during virus-induced memory dysfunction via IL-1. *Nat Immunol*. 2018;19(2):151-61.
29. Murray KO, Garcia MN, Rahbar MH, Martinez D, Khuwaja SA, Arafat RR, et al. Survival analysis, long-term outcomes, and percentage of recovery up to 8 years post-infection among the Houston West Nile virus cohort. *PLoS One*. 2014;9(7):e102953.
30. Mehta R, Soares CN, Medialdea-Carrera R, Ellul M, da Silva MTT, Rosala-Hallas A, et al. The spectrum of neurological disease associated with Zika and chikungunya viruses in adults in Rio de Janeiro, Brazil: A case series. *PLoS Negl Trop Dis*. 2018;12(2):e0006212.
31. Carson PJ, Konewko P, Wold KS, Mariani P, Goli S, Bergloff P, et al. Long-term clinical and neuropsychological outcomes of West Nile virus infection. *Clin Infect Dis*. 2006;43(6):723-30.
32. Murray KO, Resnick M, Miller V. Depression after infection with West Nile virus. *Emerg Infect Dis*. 2007;13(3):479-81.
33. Nolan MS, Hause AM, Murray KO. Findings of long-term depression up to 8 years post infection from West Nile virus. *J Clin Psychol*. 2012;68(7):801-8.
34. John CC, Carabin H, Montano SM, Bangirana P, Zunt JR, Peterson PK. Global research priorities for infections that affect the nervous system. *Nature*. 2015;527(7578):S178-86.
35. Correa-Oliveira GE, do Amaral JL, da Fonseca BAL, Del-Ben CM. Zika virus infection followed by a first episode of psychosis: another flavivirus leading to pure psychiatric symptomatology. *Braz J Psychiatry*. 2017;39(4):381-2.
36. Carteaux G, Maquart M, Bedet A, Contou D, Brugieres P, Fourati S, et al. Zika Virus Associated with Meningoencephalitis. *N Engl J Med*. 2016;374(16):1595-6.

37. Daniels BP, Snyder AG, Olsen TM, Orozco S, Oguin TH, 3rd, Tait SWG, et al. RIPK3 Restricts Viral Pathogenesis via Cell Death-Independent Neuroinflammation. *Cell*. 2017;169(2):301-13 e11.

38. Satterstrom FK, Kosmicki JA, Wang J, Breen MS, De Rubeis S, An JY, et al. Large-Scale Exome Sequencing Study Implicates Both Developmental and Functional Changes in the Neurobiology of Autism. *Cell*. 2020;180(3):568-84 e23.

39. Abrahams BS, Arking DE, Campbell DB, Mefford HC, Morrow EM, Weiss LA, et al. SFARI Gene 2.0: a community-driven knowledgebase for the autism spectrum disorders (ASDs). *Molecular autism*. 2013;4(1):36-.

40. Arcos-Burgos M, Vélez JI, Solomon BD, Muenke M. A common genetic network underlies substance use disorders and disruptive or externalizing disorders. *Human Genetics*. 2012;131(6):917-29.

41. Arias B, Fabbri C, Serretti A, Drago A, Mitjans M, Gastó C, et al. DISC1-TSNAx and DAOA genes in major depression and citalopram efficacy. *J Affect Disord*. 2014;168:91-7.

42. Balu DT, Li Y, Puhl MD, Benneyworth MA, Basu AC, Takagi S, et al. Multiple risk pathways for schizophrenia converge in serine racemase knockout mice, a mouse model of NMDA receptor hypofunction. *Proceedings of the National Academy of Sciences of the United States of America*. 2013;110(26):E2400-E9.

43. Barkus C, Sanderson DJ, Rawlins JN, Walton ME, Harrison PJ, Bannerman DM. What causes aberrant salience in schizophrenia? A role for impaired short-term habituation and the GRIA1 (GluA1) AMPA receptor subunit. *Mol Psychiatry*. 2014;19(10):1060-70.

44. Baum AE, Akula N, Cabanero M, Cardona I, Corona W, Klemens B, et al. A genome-wide association study implicates diacylglycerol kinase eta (DGKH) and several other genes in the etiology of bipolar disorder. *Mol Psychiatry*. 2008;13(2):197-207.

45. Beaulieu JM, Gainetdinov RR. The physiology, signaling, and pharmacology of dopamine receptors. *Pharmacol Rev*. 2011;63(1):182-217.

46. Bhat S, Dao DT, Terrillion CE, Arad M, Smith RJ, Soldatov NM, et al. CACNA1C (Cav1.2) in the pathophysiology of psychiatric disease. *Progress in neurobiology*. 2012;99(1):1-14.

47. Charney AW, Ruderfer DM, Stahl EA, Moran JL, Chambert K, Belliveau RA, et al. Evidence for genetic heterogeneity between clinical subtypes of bipolar disorder. *Translational Psychiatry*. 2017;7(1):e993-e.

48. Chaste P, Leboyer M. Autism risk factors: genes, environment, and gene-environment interactions. *Dialogues in clinical neuroscience*. 2012;14(3):281-92.

49. Chen DT, Jiang X, Akula N, Shugart YY, Wendland JR, Steele CJ, et al. Genome-wide association study meta-analysis of European and Asian-ancestry samples identifies three novel loci associated with bipolar disorder. *Mol Psychiatry*. 2013;18(2):195-205.

50. Chen Q, Che R, Wang X, O'Neill FA, Walsh D, Tang W, et al. Association and expression study of synapsin III and schizophrenia. *Neuroscience letters*. 2009;465(3):248-51.

51. Christensen JH, Børglum AD. Modeling the cooperativity of schizophrenia risk genes. *Nature Genetics*. 2019;51(10):1434-6.

52. Cichon S, Mühlleisen TW, Degenhardt FA, Mattheisen M, Miró X, Strohmaier J, et al. Genome-wide association study identifies genetic variation in neurocan as a susceptibility factor for bipolar disorder. *Am J Hum Genet*. 2011;88(3):372-81.

53. Demontis D, Walters RK, Martin J, Mattheisen M, Als TD, Agerbo E, et al. Discovery of the first genome-wide significant risk loci for attention deficit/hyperactivity disorder. *Nature Genetics*. 2019;51(1):63-75.

54. Escamilla MA, Zavala JM. Genetics of bipolar disorder. *Dialogues in clinical neuroscience*. 2008;10(2):141-52.

55. Gordovez FJA, McMahon FJ. The genetics of bipolar disorder. *Molecular Psychiatry*. 2020;25:544+.
56. Hawi Z, Segurado R, Conroy J, Sheehan K, Lowe N, Kirley A, et al. Preferential Transmission of Paternal Alleles at Risk Genes in Attention-Deficit/Hyperactivity Disorder. *The American Journal of Human Genetics*. 2005;77(6):958-65.
57. Hayman V, Fernandez TV. Genetic Insights Into ADHD Biology. *Frontiers in Psychiatry*. 2018;9(251).
58. Hou L, Heilbronner U, Degenhardt F, Adli M, Akiyama K, Akula N, et al. Genetic variants associated with response to lithium treatment in bipolar disorder: a genome-wide association study. *Lancet*. 2016;387(10023):1085-93.
59. Huang J, Perlis RH, Lee PH, Rush AJ, Fava M, Sachs GS, et al. Cross-disorder genomewide analysis of schizophrenia, bipolar disorder, and depression. *Am J Psychiatry*. 2010;167(10):1254-63.
60. Kandasamy R, McQuillin A, Sharp SI, Fiorentino A, Anjorin A, Blizzard RA, et al. Genetic Association, Mutation Screening, and Functional Analysis of a Kozak Sequence Variant in the Metabotropic Glutamate Receptor 3 Gene in Bipolar Disorder. *JAMA Psychiatry*. 2013;70(6):591-8.
61. Kerner B. Genetics of bipolar disorder. The application of clinical genetics. 2014;7:33-42.
62. Kerner B, Lambert CG, Muthén BO. Genome-Wide Association Study in Bipolar Patients Stratified by Co-Morbidity. *PLOS ONE*. 2011;6(12):e28477.
63. LeDoux MS. The genetics of dystonias. *Advances in genetics*. 2012;79:35-85.
64. Liu C, Kanazawa T, Tian Y, Mohamed Saini S, Mancuso S, Mostaid MS, et al. The schizophrenia genetics knowledgebase: a comprehensive update of findings from candidate gene studies. *Translational Psychiatry*. 2019;9(1):205.
65. Lohoff FW. Overview of the Genetics of Major Depressive Disorder. *Current Psychiatry Reports*. 2010;12(6):539-46.
66. Marshall CR, Noor A, Vincent JB, Lionel AC, Feuk L, Skaug J, et al. Structural variation of chromosomes in autism spectrum disorder. *Am J Hum Genet*. 2008;82(2):477-88.
67. McMahon F, Detera-Wadleigh S. Genetics of bipolar disorder. 2020. p. 735-43.
68. Mei L, Nave KA. Neuregulin-ERBB signaling in the nervous system and neuropsychiatric diseases. *Neuron*. 2014;83(1):27-49.
69. Mei L, Xiong W-C. Neuregulin 1 in neural development, synaptic plasticity and schizophrenia. *Nature reviews Neuroscience*. 2008;9(6):437-52.
70. Mill J, Xu X, Ronald A, Curran S, Price T, Knight J, et al. Quantitative trait locus analysis of candidate gene alleles associated with attention deficit hyperactivity disorder (ADHD) in five genes: DRD4, DAT1, DRD5, SNAP-25, and 5HT1B. *American Journal of Medical Genetics Part B: Neuropsychiatric Genetics*. 2005;133B(1):68-73.
71. Mühlleisen TW, Leber M, Schulze TG, Strohmaier J, Degenhardt F, Treutlein J, et al. Genome-wide association study reveals two new risk loci for bipolar disorder. *Nature Communications*. 2014;5(1):3339.
72. Neves FS, Silveira G, Romano-Silva MA, Malloy-Diniz L, Ferreira AA, De Marco L, et al. Is the 5-HTLPR polymorphism associated with bipolar disorder or with suicidal behavior of bipolar disorder patients? *Am J Med Genet B Neuropsychiatr Genet*. 2008;147b(1):114-6.
73. Ni X, Trakalo JM, Mundo E, Maciardi FM, Parikh S, Lee L, et al. Linkage disequilibrium between dopamine D1 receptor gene (DRD1) and bipolar disorder. *Biological Psychiatry*. 2002;52(12):1144-50.
74. Pagnamenta AT, Khan H, Walker S, Gerrelli D, Wing K, Bonaglia MC, et al. Rare familial 16q21 microdeletions under a linkage peak implicate cadherin 8 (CDH8) in susceptibility to autism and learning disability. *Journal of Medical Genetics*. 2011;48(1):48.

75. Palladino VS, McNeill R, Reif A, Kittel-Schneider S. Genetic risk factors and gene-environment interactions in adult and childhood attention-deficit/hyperactivity disorder. *Psychiatric Genetics*. 2019;29(3).

76. Pasquinelli AE. MicroRNAs and their targets: recognition, regulation and an emerging reciprocal relationship. *Nat Rev Genet*. 2012;13(4):271-82.

77. Poquet H, Faivre L, Chehadeh S, Morton J, McMullan D, Hamilton S. Further Evidence for Dlgap2 as Strong Autism Spectrum Disorders/Intellectual Disability Candidate Gene. *Autism-Open Access*. 2016;06.

78. Satterstrom FK, Kosmicki JA, Wang J, Breen MS, De Rubeis S, An JY, et al. Large-Scale Exome Sequencing Study Implicates Both Developmental and Functional Changes in the Neurobiology of Autism. *Cell*. 2020;180(3):568-84.e23.

79. Schmidt-Kastner R, Guloksuz S, Kietzmann T, van Os J, Rutten BPF. Analysis of GWAS-Derived Schizophrenia Genes for Links to Ischemia-Hypoxia Response of the Brain. *Frontiers in Psychiatry*. 2020;11(393).

80. Shadrina M, Bondarenko EA, Slominsky PA. Genetics Factors in Major Depression Disease. *Frontiers in psychiatry*. 2018;9:334-.

81. Stahl EA, Breen G, Forstner AJ, McQuillin A, Ripke S, Trubetskoy V, et al. Genome-wide association study identifies 30 loci associated with bipolar disorder. *Nature genetics*. 2019;51(5):793-803.

82. Thomson PA, Malavasi ELV, Grünwald E, Soares DC, Borkowska M, Millar JK. DISC1 genetics, biology and psychiatric illness. *Frontiers in biology*. 2013;8(1):1-31.

83. Verrall L, Burnet PWJ, Betts JF, Harrison PJ. The neurobiology of D-amino acid oxidase and its involvement in schizophrenia. *Molecular psychiatry*. 2010;15(2):122-37.

84. Weber H, Kittel-Schneider S, Heupel J, Weißflog L, Kent L, Freudenberg F, et al. On the role of NOS1 ex1f-VNTR in ADHD—allelic, subgroup, and meta-analysis. *American Journal of Medical Genetics Part B: Neuropsychiatric Genetics*. 2015;168(6):445-58.

85. Yoshikawa A, Kushima I, Miyashita M, Toriumi K, Suzuki K, Horiuchi Y, et al. Dysregulation of post-transcriptional modification by copy number variable microRNAs in schizophrenia with enhanced glycation stress. *Translational Psychiatry*. 2021;11(1):331.

86. Zamanian Azodi M, Rezaei-Tavirani M, Rezaei-Tavirani M. Identification of the Key Genes of Autism Spectrum Disorder Through Protein-Protein Interaction Network. *Galen Medical Journal*; Vol 8 (2019): 2019. 2019.

87. Burger-Calderon R, Bustos Carrillo F, Gresh L, Ojeda S, Sanchez N, Plazaola M, et al. Age-dependent manifestations and case definitions of paediatric Zika: a prospective cohort study. *Lancet Infect Dis*. 2020;20(3):371-80.

88. Simoes ESAC, Moreira JM, Romanelli RM, Teixeira AL. Zika virus challenges for neuropsychiatry. *Neuropsychiatr Dis Treat*. 2016;12:1747-60.

89. Tucci V, Moukaddam N, Meadows J, Shah S, Galwankar SC, Kapur GB. The Forgotten Plague: Psychiatric Manifestations of Ebola, Zika, and Emerging Infectious Diseases. *J Glob Infect Dis*. 2017;9(4):151-6.

90. Joob B, Wiwanitkit V. Zika virus infection and psychosis. *Braz J Psychiatry*. 2018;40(1):113.

91. Euteneuer F, Dannehl K, Del Rey A, Engler H, Schedlowski M, Rief W. Peripheral Immune Alterations in Major Depression: The Role of Subtypes and Pathogenetic Characteristics. *Front Psychiatry*. 2017;8:250.

92. Liu JJ, Wei YB, Strawbridge R, Bao Y, Chang S, Shi L, et al. Peripheral cytokine levels and response to antidepressant treatment in depression: a systematic review and meta-analysis. *Mol Psychiatry*. 2020;25(2):339-50.

93. Pedrotti Moreira F, Wiener CD, Jansen K, Portela LV, Lara DR, Souza LDM, et al. Childhood trauma and increased peripheral cytokines in young adults with major depressive: Population-based study. *J Neuroimmunol.* 2018;319:112-6.

94. Dowlati Y, Herrmann N, Swardfager W, Liu H, Sham L, Reim EK, et al. A meta-analysis of cytokines in major depression. *Biol Psychiatry.* 2010;67(5):446-57.

95. Fineberg AM, Ellman LM. Inflammatory cytokines and neurological and neurocognitive alterations in the course of schizophrenia. *Biol Psychiatry.* 2013;73(10):951-66.

96. Rodrigues-Amorim D, Rivera-Baltanas T, Spuch C, Caruncho HJ, Gonzalez-Fernandez A, Olivares JM, et al. Cytokines dysregulation in schizophrenia: A systematic review of psychoneuroimmune relationship. *Schizophr Res.* 2018;197:19-33.

97. Reale M, Costantini E, Greig NH. Cytokine Imbalance in Schizophrenia. From Research to Clinic: Potential Implications for Treatment. *Front Psychiatry.* 2021;12:536257.

98. Leis AA, Grill MF, Goodman BP, Sadiq SB, Sinclair DJ, Vig PJS, et al. Tumor Necrosis Factor-Alpha Signaling May Contribute to Chronic West Nile Virus Post-infectious Proinflammatory State. *Front Med (Lausanne).* 2020;7:164.

99. Daniels BP, Jujjavarapu H, Durrant DM, Williams JL, Green RR, White JP, et al. Regional astrocyte IFN signaling restricts pathogenesis during neurotropic viral infection. *J Clin Invest.* 2017;127(3):843-56.

100. Olmo IG, Carvalho TG, Costa VV, Alves-Silva J, Ferrari CZ, Izidoro-Toledo TC, et al. Zika Virus Promotes Neuronal Cell Death in a Non-Cell Autonomous Manner by Triggering the Release of Neurotoxic Factors. *Front Immunol.* 2017;8:1016.

101. Chen Z, Wang X, Ashraf U, Zheng B, Ye J, Zhou D, et al. Activation of neuronal N-methyl-D-aspartate receptor plays a pivotal role in Japanese encephalitis virus-induced neuronal cell damage. *J Neuroinflammation.* 2018;15(1):238.

102. Chen W, Sheng J, Guo J, Gao F, Zhao X, Dai J, et al. Tumor necrosis factor-alpha enhances voltage-gated Na(+) currents in primary culture of mouse cortical neurons. *J Neuroinflammation.* 2015;12:126.

103. Clark IA, Vissel B. Excess cerebral TNF causing glutamate excitotoxicity rationalizes treatment of neurodegenerative diseases and neurogenic pain by anti-TNF agents. *J Neuroinflammation.* 2016;13(1):236.

104. Pickering M, Cumiskey D, O'Connor JJ. Actions of TNF-alpha on glutamatergic synaptic transmission in the central nervous system. *Exp Physiol.* 2005;90(5):663-70.

105. Bernardino L, Agasse F, Silva B, Ferreira R, Grade S, Malva JO. Tumor necrosis factor-alpha modulates survival, proliferation, and neuronal differentiation in neonatal subventricular zone cell cultures. *Stem Cells.* 2008;26(9):2361-71.

106. Keohane A, Ryan S, Maloney E, Sullivan AM, Nolan YM. Tumour necrosis factor-alpha impairs neuronal differentiation but not proliferation of hippocampal neural precursor cells: Role of Hes1. *Mol Cell Neurosci.* 2010;43(1):127-35.

107. Borsini A, Zunszain PA, Thuret S, Pariante CM. The role of inflammatory cytokines as key modulators of neurogenesis. *Trends Neurosci.* 2015;38(3):145-57.

108. Heir R, Stellwagen D. TNF-Mediated Homeostatic Synaptic Plasticity: From in vitro to in vivo Models. *Front Cell Neurosci.* 2020;14:565841.

109. Maggio N, Vlachos A. Tumor necrosis factor (TNF) modulates synaptic plasticity in a concentration-dependent manner through intracellular calcium stores. *J Mol Med (Berl).* 2018;96(10):1039-47.

110. Pozniak PD, Darbinyan A, Khalili K. TNF-alpha/TNFR2 Regulatory Axis Stimulates EphB2-Mediated Neuroregeneration Via Activation of NF-kappaB. *J Cell Physiol.* 2016;231(6):1237-48.

111. Cornelius ADA, Hosseini S, Schreier S, Fritzsch D, Weichert L, Michaelsen-Preusse K, et al. Langat virus infection affects hippocampal neuron morphology and function in mice without disease signs. *J Neuroinflammation*. 2020;17(1):278.
112. Wu YH, Cui XY, Yang W, Fan DY, Liu D, Wang PG, et al. Zika Virus Infection in Hypothalamus Causes Hormone Deficiencies and Leads to Irreversible Growth Delay and Memory Impairment in Mice. *Cell Rep*. 2018;25(6):1537-47 e4.
113. Stanelle-Bertram S, Walendy-Gnirss K, Speiseder T, Thiele S, Asante IA, Dreier C, et al. Male offspring born to mildly ZIKV-infected mice are at risk of developing neurocognitive disorders in adulthood. *Nat Microbiol*. 2018;3(10):1161-74.
114. Funk KE, Arutyunov AD, Desai P, White JP, Soung AL, Rosen SF, et al. Decreased antiviral immune response within the central nervous system of aged mice is associated with increased lethality of West Nile virus encephalitis. *Aging Cell*. 2021;20(8):e13412.
115. Montgomery RR. Age-related alterations in immune responses to West Nile virus infection. *Clin Exp Immunol*. 2017;187(1):26-34.
116. Pecanha PM, Gomes Junior SC, Pone SM, Pone M, Vasconcelos Z, Zin A, et al. Neurodevelopment of children exposed intra-uterus by Zika virus: A case series. *PLoS One*. 2020;15(2):e0229434.
117. Vouga M, Pomar L, Panchaud A, Musso D, Baud D. A critical analysis of the neurodevelopmental and neurosensory outcomes after 2 years for children with in utero Zika virus exposure. *Nat Med*. 2019;25(11):1641-2.