

# RESEARCH

# Chromosome-scale assemblies of *Acanthamoeba castellanii* genomes provide insights into *Legionella pneumophila* infection-related chromatin re-organization

Cyril Matthey-Doret<sup>1,2†</sup>, Morgan J. Colp<sup>3†</sup>, Pedro Escoll<sup>4</sup>, Agnès Thierry<sup>1</sup>, Bruce Curtis<sup>3</sup>, Matt Sarrasin<sup>5</sup>, Michael W. Gray<sup>3</sup>, B. Franz Lang<sup>5</sup>, John M. Archibald<sup>3\*</sup>, Carmen Buchrieser<sup>4\*</sup> and Romain Koszul<sup>1\*</sup>

\*Correspondence:

[john.archibald@dal.ca](mailto:john.archibald@dal.ca);

[cbuch@pasteur.fr](mailto:cbuch@pasteur.fr);

[romain.koszul@pasteur.fr](mailto:romain.koszul@pasteur.fr)

<sup>3</sup>Department of Biochemistry and Molecular Biology and Institute for Comparative Genomics, Dalhousie University, Sir Charles Tupper Medical Building, 5850 College Street, B3H 4R2 Halifax, Nova Scotia, Canada

<sup>4</sup>Institut Pasteur, Université de Paris, CNRS UMR3525, Unité Biologie des Bactéries Intracellulaires, F-75015, Paris, France

<sup>1</sup>Institut Pasteur, Université de Paris, CNRS UMR3525, Unité Régulation Spatiale des Génomes, F-75015, Paris, France

Full list of author information is available at the end of the article

†Equal contributor

## Abstract

The unicellular amoeba *Acanthamoeba castellanii* is ubiquitous in aquatic environments, where it preys on bacteria. The organism also hosts bacterial endosymbionts, some of which are parasitic, including human pathogens such as *Chlamydia* and *Legionella* spp. Here we report complete, high quality genome sequences for two extensively studied *A. castellanii* strains, Neff and C3. Combining long- and short-read data with Hi-C, we generated near chromosome-level assemblies for both strains with 90% of the genome contained in 29 scaffolds for the Neff strain and 31 for the C3 strain. Comparative genomics revealed strain-specific functional enrichment, most notably genes related to signal transduction in the C3 strain, and to viral replication in Neff. Furthermore, we characterized the spatial organization of the *A. castellanii* genome and showed that it is reorganized during infection by *Legionella pneumophila*. Infection-dependent chromatin loops were found to be enriched in genes for signal transduction and phosphorylation processes. In genomic regions where chromatin organization changed during *Legionella* infection, we found functional enrichment for genes associated with metabolism, organelle assembly, and cytoskeleton organization, suggesting that changes in chromosomal folding are associated with host cell biology during infection.

**Keywords:** amoeba; genome organization; evolution; assembly; Hi-C

## Introduction

The first amoebae were isolated in 1913 [1], and the genus *Acanthamoeba* was established in 1931 by Volkonsky [2]. It comprises different species of free living, aerobic, unicellular protozoa, present throughout the world in soil and nearly all aquatic environments [3]. The life cycle of *Acanthamoeba* includes a dormant cyst with minimal metabolic activities under harsh conditions and a motile trophozoite that can feed on small organisms and reproduce by binary fission in optimal conditions [4]. *Acanthamoeba* is perhaps most widely known from its role as a human pathogen, acting to cause the vision-threatening eye infection *Acanthamoeba* keratitis, but it can also cause serious infections of the lungs, sinuses, skin, and a central nervous system disease called granulomatous amoebic encephalitis [5]. The species *Acanthamoeba castellanii* was first isolated in 1930 by Castellani as a contaminant of a yeast culture [6].

In their natural environment, *Acanthamoeba* spp. are characterized by the ability to change their shape through pseudopode formation and are considered professional phagocytes as they feed on bacteria, but may also phagocytose yeasts and algae. However, some bacteria are resistant to degradation and live as endosymbionts in these protozoa, and others even use the amoeba as a replication niche. Thus *Acanthamoeba* are also reservoirs of microorganisms and viruses, including human pathogens, which have adapted to survive inside these cells and resist digestion, persist or even replicate as intracellular parasites. At least 15 different bacterial species, two archaea and several eukaryotes and viruses have been shown to interact with *Acanthamoeba* in the environment and may even co-exist at the same time within the same host cell [7].

Although it was observed early on that bacteria could resist digestion of free-living amoebae [8], it was not until the discovery that *Legionella pneumophila* replicated in amoebae that researchers began studying the bacterium-amoeba relationship in depth [9]. *L. pneumophila* is the agent responsible for Legionnaires' disease, a severe pneumonia that can be fatal if not treated promptly. In addition, many species of amoebae have the ability to form highly resistant cysts in hostile environments, providing shelter for their intracellular parasites [10]. Indeed, it is thought that *L. pneumophila* may survive water disinfection treatments and contaminate water distribution systems by encystation [11, 12, 13]. From these contaminated water sources, *L. pneumophila* can reach the human lungs via aerosols contaminated with the bacteria and replicate within the alveolar macrophages that are, like amoebae, phagocytic cells.

*L. pneumophila* has the ability to escape the lysosomal degradation pathway of both *A. castellanii* and human alveolar macrophages through the formation of a protective vacuole (the Legionella-containing vacuole or LCV) where it multiplies to high numbers. Once the host cell has been fully exploited and nutrients become limited, *L. pneumophila* exits the host and infects a new cell [14].

To establish the LCV and replicate, *L. pneumophila* secretes over 300 effector proteins into the host cytoplasm via a type four secretion system (T4SS) called Dot/Icm [15], thereby manipulating host pathways and redirecting nutrients to the LCV [16, 17]. In the early stages of infection, many of these proteins target the host secretory pathway, including several small GTPases, to recruit endoplasmic reticulum-derived vesicles to the LCV [18]. During the intracellular cycle, a wider range of processes, including membrane trafficking, cytoskeleton dynamics, and signal transduction pathways, are targeted by these effectors [19, 20]. *L. pneumophila* also directly alters the genome of its host by modifying epigenetic marks of the host genome in human macrophages and *A. castellanii*. It secretes an effector named RomA with histone methyltransferase activity that is targeted to the nucleus. RomA carries out genome-wide trimethylation of K14 of histone H3 [21], leading to transcriptional changes that modulate the host response in favor of bacterial survival [21]. Concomitantly, *L. pneumophila* infection leads to genome-wide changes in gene expression [22]. In many eukaryotes, gene regulation is intertwined within the three-dimensional organization of chromosomes. The functional interplay between gene regulation and higher-order chromatin elements such as loops, self-interacting domains and active/inactive compartments is actively being studied [23, 24]. Therefore, the infection of *A. castellanii* by *L. pneumophila* provides

an amenable model with which to investigate how an intracellular bacterial infection may affect the regulation of chromosome folding, and its consequences, in a eukaryotic host.

The investigation of genome organization and regulation of *A. castellanii* in response to infection requires a highly contiguous genome assembly. The reference genome sequence for *A. castellanii*, NEFF-v1 [25], is based on the Neff strain, isolated from soil in California in 1957 [26]. This assembly is widely used by different laboratories studying *A. castellanii*, but is fragmented into 384 scaffolds comprising 3192 contigs, which makes chromosome-level analyses difficult, if not impossible, and basic features of the *A. castellanii* genome, such as the number of chromosomes and ploidy, remain undetermined. In addition, many teams investigating bacteria-amoeba interactions use the "C3" strain (ATCC 50739), isolated from a drinking water reservoir in Europe in 1994 and identified as a mouse pathogen [27]. However, genomic information is scarce for this strain and little is known about its similarity to the Neff strain. Notably, these two *A. castellanii* strains have been cultivated for several decades and were isolated from different ecological niches, but the extent of conservation between their genomes is unknown. It is difficult to investigate the factors that determine the susceptibility of different *A. castellanii* strains to the pathogen without proper genomic resources. These resources would also be required to apply genome-wide omics approaches.

The goal of this work was to study how the *A. castellanii* C3 strain responds to *L. pneumophila* infection through the lens of the three-dimensional organisation of its genome. This analysis required the generation of a high quality reference genome sequence of the C3 strain, as well as a new and improved assembly of the Neff reference genome. Illumina, Nanopore long read, and Hi-C data were used to generate near chromosome-level assemblies of the genomes of both strains. Surprisingly, the new Neff and C3 assemblies have a (gap-excluded) sequence divergence of 6.7%. We find evidence for strain-specific enrichment of a handful of functions, including ones related to signal transduction in C3, and one relating to viral replication and virion assembly in Neff. Using the C3 assembly, RNA-seq and Hi-C, we were able to analyze the genome folding and expression changes of *A. castellanii* in response to the infection by *L. pneumophila*. We found infection-dependent chromatin loops to be enriched in genes involved in signal transduction and phosphorylation.

## Results

The *A. castellanii* Neff and C3 genome assemblies are highly contiguous and complete. We used a combination of Illumina short reads, Oxford Nanopore long reads and Hi-C to assemble each genome to chromosome scale, with 90% of the Neff genome contained within 28 scaffolds. This is in contrast to a previous estimate of approximately 20 chromosomes inferred using pulsed-field gel electrophoresis [28]. For both the Neff and C3 strains, we first generated a raw *de-novo* assembly using Oxford Nanopore long reads. To account for the error prone nature of long reads, we polished the first draft assemblies with paired-end shotgun Illumina sequences using HyPo [29]. The polished assemblies were then scaffolded with long range Hi-C contacts using our probabilistic program instaGRAAL, which exploits a Markov Chain Monte Carlo algorithm to swap DNA segments until the most likely scaffolds are

achieved [30]. Following the post-scaffolding polishing step of the program (see [30]), the final genome assemblies displayed better contiguity (Table 1), completion, and mapping statistics than the previous versions, with the cumulative scaffold lengths quickly reaching a plateau (Fig. 1a). The assemblies of both strains are also slightly longer, with a smaller number of contigs than the original Neff assembly (NEFF-v1) (Fig. 1b). The BUSCO-completeness scores for both assemblies are also improved, with 90.6% (Neff) and 91.8% (C3) complete eukaryotic universal single copy orthologs, compared to 77.6% for NEFF-v1. We also noted an increased proportion of properly paired shotgun reads from 71% for NEFF-v1 to 84% for our new Neff assembly, suggesting a reduced number of short mis-assemblies. Hi-C contact maps present a convenient readout to explore large mis-assemblies in genome sequences [31]. While this allowed us to manually address major unambiguous mis-assemblies, a number of visible mis-assemblies remain in complex regions such as repeated sequences near telomeres and ribosomal DNAs (rDNAs). These mis-assemblies could not be resolved with the data generated herein. In the C3 assembly, there are also a few (at least 5) interchromosomal mis-assemblies which appear to be heterozygous and cannot be resolved without a phased genome. We also found shotgun coverage to be highly heterogeneous between scaffolds, which is suggestive of aneuploidy (Fig. S1).

#### A. *castellanii* strains Neff and C3 have partly non-overlapping gene complements

The generation of chromosome-scale genome assemblies for two different *A. castellanii* strains afforded us the first opportunity to compare and contrast their coding capacities. We used both Broccoli [32] and OrthoFinder [33] for inference of orthologous groups. A summary of the inferred orthogroups shared by, and specific to, the Neff and C3 strains of *A. castellanii* is presented in Figure 2, with orthogroup numbers from both orthologous clustering tools included. This figure only compares Neff against C3, irrespective of orthogroup presence or absence in outgroup taxa. In this analysis, each strain-specific gene that was not assigned to an orthogroup by either program was still considered to be a single strain-specific orthogroup in order to account for the presence of genes without any orthologs across the five species. Broccoli predicted more orthogroups overall and more strain-specific genes than OrthoFinder, but predicted fewer shared orthogroups. Despite these differences, the overall trend is similar for the two outputs. The number of orthogroups shared by the two strains is roughly an order of magnitude greater than the number specific to either strain, while the C3 strain has a greater number of strain-specific orthogroups than the Neff strain as predicted by both programs.

To investigate how similar the *A. castellanii* gene complement was to other members of Amoebozoa, *A. castellanii* orthogroups were evaluated for their presence in three outgroup species. Both Broccoli and OrthoFinder outputs were analyzed in this fashion. According to Broccoli, 43.5% of orthogroups shared by the two *A. castellanii* strains were not present in the other three amoebae, while OrthoFinder gave a figure of 48.4%. In the Neff strain, 49.1% of all orthogroups, shared or strain-specific, were not found in the three outgroup amoebae according to Broccoli, compared to 51.0% as predicted by OrthoFinder. In the C3 strain, the Broccoli results indicate that 52.4% of all orthogroups are not present in the outgroup amoebae,



while 52.8% were not found in the outgroup by OrthoFinder. This is in contrast with *A. castellanii* strain C3 sharing an estimated 82.5% (Broccoli) to 89.4% (OrthoFinder) of its orthogroups with the Neff strain, and the Neff strain sharing an estimated 88.9% (Broccoli) to 93.6% (OrthoFinder) of its orthogroups with the C3 strain.

#### *A. castellanii* accessory genes show strain-specific functional enrichment

In an attempt to gain insight into the functional significance of strain-specific genes in the C3 and Neff genomes, the top 30 most significantly enriched terms were identified by topGO and plotted in order of decreasing p-value for each strain/ontology combination (Supplementary Figures S8-S13). Notably, among C3-specific genes, only two terms were found to be statistically significantly enriched for each of the three ontologies at a 95% confidence level. Among Neff-specific genes, only one term was significantly enriched in each of the ‘cellular component’ and ‘molecular function’ ontologies, while three were significantly enriched in the ‘biological process’ ontology.

In C3, enriched molecular functions were ‘GTP binding’ ( $p = 5e-5$ ) and ‘protein serine/threonine phosphatase activity’ ( $p = 0.037$ ), enriched biological processes were ‘small GTPase mediated signal transduction’ ( $p = 8.5e-5$ ) and ‘ubiquitin-dependent protein catabolic processes’ ( $p = 0.029$ ), and enriched cellular components were ‘RNA polymerase II core complex’ ( $p = 0.026$ ) and ‘the Golgi membrane’ ( $p = 0.036$ ). In Neff, the enriched molecular function was ‘DNA helicase activity’ ( $p = 0.0071$ ), enriched biological processes were ‘telomere maintenance’ ( $p = 0.0027$ ), ‘protein homooligomerization’ ( $p = 0.0135$ ), and ‘DNA replication’ ( $p = 0.0403$ ), and the enriched cellular component was ‘virion parts’ ( $p = 0.012$ ). When searched against the nr database with BLASTp [34], the Neff genes found to be responsible for both DNA helicase activity enrichment and telomere maintenance enrichment had their best BLAST hits to PIF1 5'-to-3' DNA helicases, those responsible for protein homooligomerization enrichment had their best BLAST hits to K<sup>+</sup> channel tetramerization domains, and the gene annotated as being a virion part had its best BLAST hits to major capsid protein from various nucleocytoplasmic large DNA viruses (NCLDV).

#### The Neff strain has a divergent mannose binding protein

One particular gene of interest encodes a mannose binding protein, which is known to be used as a receptor for cell entry by *Legionella* in some *A. castellanii* strains [35]. The MEEI 0184 strain of *A. castellanii*, an isolate from a human corneal infection, was used as a reference sequence, because it is the only strain in which the mannose binding protein is biochemically characterized [36, 37]. The orthologs from C3, Neff, and *Acanthamoeba polyphaga* were retrieved, and all four sequences were aligned (Figure S14). The percent identity of each sequence to the reference was calculated over the sites in the alignment where the *A. polyphaga* sequence was not missing (Table 2). The C3 homolog was found to be 99.5% identical to the MEEI 0184 homolog, whereas the Neff and *A. polyphaga* proteins were more divergent, sharing 91.6% and 97.2% identity to MEEI 0184, respectively. Despite being of the same species as the reference, the Neff strain homolog was found to

be much more divergent than the *A. polyphaga* sequence is from the other two *A. castellanii* strains. Interestingly, we observed that *L. pneumophila* replicates worse in the Neff strain than the C3 strain in culture. This phenotype may result from impaired receptor-mediated entry by *Legionella* into Neff cells due to differences in the receptor encoding gene.

### Spatial organisation of the *A. castellanii* genome

To our knowledge, no Hi-C contact maps have been generated from species of Amoebozoa. Therefore, the Hi-C reads we used to generate the chromosome-scale scaffolding of two *A. castellanii* genomes also offer the opportunity to reveal the average genome folding in a species of this clade. Hi-C reads were realigned along the new assemblies of both the C3 and Neff strains to generate genome-wide contact maps. Visualising the Hi-C contact maps of both genomes shows that *A. castellanii* chromosomes are well resolved in our assemblies (Fig. 3). In Neff, the highest intensity contacts are concentrated on the main diagonal, suggesting an absence of large-scale mis-assemblies. On the other hand, the C3 assembly retains a few mis-assembled blocks, mostly in the rDNA region where tandem repeats could not be resolved correctly with the data available to us. However, for both strains the genome-wide contact maps reveal a grid-like pattern, with contact enrichment between chromosome extremities resulting in discrete dots. These contacts can be interpreted as a clustering of the telomeres, or subtelomeres, of the different chromosomes (Fig. 3a). Based on the presence of these inter-telomeric contacts patterns, Hi-C contact maps suggest the presence of at least 35 chromosomes in both strains, ranging from roughly 100 kbp to 2.5 Mbp in length (Fig. S15). Additionally we found 100 copies of 5S rDNA dispersed across most chromosomes for both strains, and 18S/28S rDNA genes show increased contacts with subtelomeres (Fig. 3a).

In addition to large, interchromosomal subtelomeric contacts, we also explored the existence of intrachromosomal chromatin 3D structures in the contact maps using ChromSight, a program that detects patterns reflecting chromatin structures on Hi-C contact maps [38]. For both strains, ChromSight identified arrays of chromatin loops along chromosomes, as well as boundaries separating chromatin domains (Fig. 3b). Most chromatin loops are regularly spaced, with a typical size of 20 kbp (Fig. 3c). The chromatin domains correspond to discrete squares along the diagonal (Fig. S3a). We overlapped all predicted genes in the C3 genome with the domain borders detected from Hi-C data and measured their base expression using RNA-seq we generated from that strain (see Methods). We selected the closest gene to each domain border and found that the genes overlapping domain boundaries are overall more highly expressed than those that do not (Fig. S2c). In addition, the analysis showed that gene expression is negatively correlated with the distance to the closest domain border (Fig. S2d). We performed the same comparison using chromatin loop anchors instead of domain borders. To a lesser extent, genes overlapping chromatin loops are also associated with higher expression (Fig. S2a), although it is not correlated with the distance from the closest loop (Fig. S2b). Altogether, these results suggest that the chromatin structures observed in *cis* are both associated with gene expression, although the association between gene expression and chromatin loop anchors is likely due to their co-localization with domain

borders (Fig. S2e). Some microorganisms (e.g. budding yeasts and euryarchaeotes) organize their chromosomes into micro-domains that correspond to expressed genes [39, 40]. Our findings in *A. castellanii* bear an interesting similarity to this type of organization.

#### *L. pneumophila* infection induces chromatin loop changes enriched in infection-related functions

The generation of near-complete assemblies allowed us to tackle the question of whether *L. pneumophila* infection impacts the 3D folding and transcription of the *A. castellanii* C3 strain genome. We harvested cultured *A. castellanii* cells before and 5 hours following infection by *L. pneumophila* strain Paris [41] (Methods). The cells were processed using Hi-C and RNA-seq (Methods), and the resulting reads aligned against the reference genome to assess changes in the genome structure and the host transcription program, respectively. RNA-seq was performed in triplicate, and Hi-C in duplicate (Methods). To measure changes in *trans*-chromosomal contacts, we merged the contact maps from our replicates and applied the serpentine adaptive binning method to improve the signal-to-noise ratio [42]. We then computed average interactions between each pair of chromosomes before and after infection. For each pair of chromosomes, we then used the log ratio of infected over uninfected average contacts. Following infection a global decrease in *trans*-subtelomeric contacts was observed, suggesting a slight de-clustering of chromosome ends (Fig. 4b). In addition, the scaffold bearing 18S and 28S rDNA (scaffold\_29), as well as two other small scaffolds (35 and 36) displayed weaker interactions with other scaffolds during infection (Fig. 4a).

We then assessed whether the behavior of *cis* contacts changes during infection. First, we computed the average contact frequencies according to genomic distance  $p(s)$  (Methods), which is a convenient way to unveil variations in the compaction state of chromatin [43]. The  $p(s)$  curves show a global increase in long range contacts following infection (Fig. S4b). The strengths of chromatin loops and domain borders before and 5h after infection were quantified using Chromosight [38]. However, no significant average increase or decrease in the intensity of these structures (Fig. S4a) was identified when computed over the whole genome. To focus on infection-dependent chromatin structures, we filtered the detected patterns to retain those showing the top 20% strongest change in Chromosight score during infection (either appearing or disappearing). We performed a GO term enrichment analysis for genes associated with infection-dependent chromatin loops (Methods). A significant enrichment for Rho GTPase and phosphorelay signal transduction, protein catabolism and GPI biosynthesis was found (Fig. S6a). The strongest loop changes were associated with genes encoding Rho GTPase, GOLD and SET domains as well as genes for proteins containing leucine-rich repeats and ankyrin repeats (Fig. S7).

We followed the same procedure for domain borders and found that genes associated with infection-dependent domain borders were significantly enriched in 'amino acid transport', 'cyclic nucleotide biosynthetic process', 'protein modification' and 'deubiquitination' (Fig. S6b). Our results suggest that domain borders are generally associated with highly transcribed metabolic genes, consistent with previous findings showing that such borders are associated with high transcription [44].

By analyzing the *A. castellanii* RNA-seq data after infection with *L. pneumophila*, we revealed that the expression of genes was globally impacted at 5h post infection compared to uninfected cells (Fig. S5a). This is consistent with recent results showing that transcription is globally disrupted in *A. castellanii* Neff following infection by *L. pneumophila* [22]. To investigate the relationship between this change in gene expression and chromatin structure, we assigned the closest domain border to each gene and compared their expression and border score changes during infection. For the majority of genes, we found border intensity not to be correlated with gene expression changes (Fig. S5b). Only genes undergoing extreme expression changes during infection corresponded to changes in associated borders (Fig. S5c). This raises the possibility that insulation domains in *A. castellanii* chromosomes do not dictate gene expression programs as they do in mammals.

Recently, Li et al. [22] investigated gene expression changes at 3, 8, 16 and 24h after infection of *A. castellanii* Neff by *L. pneumophila*. To further validate our finding that chromatin domains are not units of regulation in *A. castellanii*, we used these expression results and migrated the gene annotations to our C3 assembly using liftoff [45]. This allowed us to compute co-expression between gene pairs during infection (i.e., expression correlation). We found that gene pairs within the same chromatin domain did not have a higher co-expression than gene pairs from different domains at similar genomic distances (Fig. S3d).

## Discussion

### Chromosome-level assembly uncovers *A. castellanii* genome organization

Generation, analysis and comparison of the genome sequences of two *A. castellanii* strains revealed heterogeneous coverage across scaffolds, which is consistent with previous findings that *A. castellanii* has a high but variable ploidy of approximately 25n [46]. Previous estimates of the *A. castellanii* Neff karyotype using pulsed-field gel electrophoresis estimated 17 to 20 unique chromosomes ranging from 250 kbp to just over 2 Mbp [28], while our estimate suggests at least 35 unique chromosomes with a similar size range of 100 kbp to 2.5 Mbp. The discrepancy between the number of bands in the electrophoretic karyotype and our estimate may result from chromosomes of similar size co-migrating on the gel, which we were able to resolve using sequence- and contact-based information.

Considering features of the nuclear biology of *A. castellanii*, such as suspected amitosis [47] and probable aneuploidy, our finding that 5S ribosomal DNA is dispersed across all chromosomes may serve to ensure a consistent copy number of 5S rDNA in daughter cells.

It was previously estimated that *A. castellanii* has 24 copies of rDNA genes per haploid genome [48]. Our data show that both strains contain 4 times as many copies as originally thought. The decrease in interchromosomal contacts with rDNA-containing scaffolds during infection may reflect an alteration in the nucleolus structure, probably caused by a global increase in translational activity. This would be consistent with the global transcription shift observed in RNA-seq under infection conditions.

At a first glance, the contact maps show a clustering of subtelomeric regions, but do not display a Rabl conformation, where centromeres cluster to the spindle-pole

body [49]. However, the precise positions of centromeres would be needed to verify that they do not co-localize with subtelomeric regions.

### Changes in chromatin structure likely reflect transcriptional changes

Infection of *A. castellanii* with *L. pneumophila* induced significant changes in chromatin loops and borders. Our analyses showed an enrichment in several interesting GO terms at the sites of these infection-induced changes, many of them consistent with known biological processes induced by *L. pneumophila* in amoebae and macrophages. Several enriched terms are related to cell cycle regulation, including mitotic cell cycle, cell cycle processes and cell cycle checkpoints (Fig. S6), which is consistent with recent results showing that *L. pneumophila* prevents proliferation of its natural host *A. castellanii* [50, 22]. *L. pneumophila*-induced alterations of the host cell cycle may serve to avoid cell cycle phases that restrict bacterial replication [51], or to prevent amoebal proliferation, which has been proposed to increase the feeding efficiency of individual amoebae [52].

Several other GO terms that we found to be enriched at infection-dependent loops or borders are related to host cell organelles, such as organelle assembly, microtubule cytoskeleton organization, protein localization to endoplasmic reticulum, mitochondrion organization, electron transport chain, or mitochondrial respiratory chain complexes (Fig. S6). This is interesting given that it is well known that during infection, *L. pneumophila* hijacks host organelles such as the cytoskeleton, the endoplasmic reticulum, and mitochondria in both amoebae and macrophages [53, 54, 55]. Indeed, mitochondrial respiration and electron transport chain complexes were recently shown to be altered in macrophages during *L. pneumophila* infection [54, 56].

Sites of infection-dependent chromatin reorganization also show enrichment in functions related to changes in the general metabolism of the host, such as biosynthetic and catabolic processes, including nucleotide and nucleoside synthesis, lipid metabolism, or transport of amino acids and metal ions. To replicate intracellularly, *L. pneumophila* acquires all its nutrients from the cytoplasm of the host cell. Therefore, it is thought that bacteria-induced modulation of the host metabolism is key to establishing a successful infection [57]. In summary, many of the GO terms associated with changes in chromatin loops and borders during infection align with the known biology of *Legionella* infection, suggesting a link between chromatin organization and many of the observed changes in host cells during infection.

It was previously shown that *L. pneumophila* infection halts host cell division and is associated with a decrease of mRNA of the *A. castellanii* CDC2b gene, a putative regulator of the *A. castellanii* cell cycle [50]. The large scale 3D changes we observed in chromatin compaction (Fig. S4b) and interchromosomal contacts (Fig. 4) are reminiscent of cell cycle changes in yeast and could suggest that the bacterium stops the host's cell cycle at a specific checkpoint.

We identified an array of regularly spaced chromatin loops in *A. castellanii* chromosomes of approximately 20 kbp in size. This is consistent with size range of chromatin loops observed in *S. cerevisiae* during the G2/M stage [58]. This similarity in terms of regularity and size suggests that chromatin loops in *A. castellanii* may serve a similar purpose of chromosome compaction for cell division as in yeast. Our finding that DNA loop anchors and domain borders overlap highly expressed

genes is also concordant with observations made in yeast and other species that domain borders are preferentially located at highly expressed genes [38, 59], and could result from their role in blocking the processing SMC complexes [60], potentially to avoid interferences between cohesin activity and transcription.

Unlike previously shown in *Drosophila* [61], we did not find an increase in co-expression of genes sharing the same contact domain in *A. castellanii*. This suggests chromatin domains may be caused by highly transcribed genes, and do not act as units of regulation.

#### *A. castellanii* accessory genes may permit environmental adaptation

Despite the substantial number of genes predicted to be strain-specific in *A. castellanii*, few functions were found to be significantly enriched in either the Neff or C3 strain set of strain-specific genes. Of these, the most biologically interesting is the enrichment of both ‘small GTPase mediated signal transduction’ and ‘GTP binding’ genes in C3. Nearly all of the genes annotated as being involved in ‘small GTPase mediated signal transduction’ biological processes are also annotated as having ‘GTP binding’ molecular functions, which is not surprising – GTP binding is an integral part of GTPase functionality. The enrichment of these two GO terms, as well as protein serine/threonine phosphatase activity enrichment, suggests that the C3 strain may have expanded its capacity for environmental sensing and associated cellular responses by expanding gene families involved in signal transduction. Given the extensive gene repertoire in *A. castellanii* dedicated to cell signalling, environmental sensing, and the cellular response [25], which is thought to help the amoeba navigate diverse habitats and identify varied prey, it seems likely that alterations of this gene repertoire in C3 may have permitted further environmental adaptations.

Another enrichment of note is that of ‘viriion parts’ in the Neff strain of *A. castellanii*. This enrichment comprises a single gene with a best BLAST hit to major capsid proteins in various NCLDV, including a very strong hit to *Mollivirus sibericum*. Many NCLDV, including *Mollivirus*, are known viruses of *Acanthamoeba* spp. [62]. Although no phylogenetic analyses were performed to investigate the origin of this major capsid protein gene in the Neff genome, it seems plausible that it was acquired by lateral gene transfer during an NCLDV infection, perhaps by *Mollivirus* or some closely related virus.

The remaining enriched functions have no obvious biological significance. They could well be non-adaptive, having been generated through gene duplication, differential loss in the other surveyed amoebae, or lateral gene transfer, without conferring any notable selective advantage. An improved understanding of *Acanthamoeba* cell and molecular biology is needed to make sense of the gene enrichment data presented herein.

#### Substitutions in the Neff mannose binding protein may inhibit *Legionella* entry

Alignment of the three *A. castellanii* mannose binding proteins (MBPs) and the *A. polyphaga* homolog may help explain the difference in susceptibility to *Legionella* infection between the Neff and C3 strains. The C3 strain mannose binding protein is highly similar to its counterpart in strain MEEI 0184, which was first to



be biochemically characterized. The Neff strain MBP, however, is markedly more divergent than even the *A. polyphaga* MBP, which is not known to participate in *Acanthamoeba-Legionella* interactions [63]. These results are consistent with the hypothesis that the Neff strain of *A. castellanii* is not a very good host for infection by *Legionella* due to an accumulation of amino acid substitutions in its mannose binding protein, substitutions that may prevent *Legionella* from binding to this protein during cell entry. Whether or not *A. castellanii* uses its MBP for feeding or recognition of potential pathogens like *Legionella* is at present unclear, but it is worth noting that the Neff strain has been in axenic culture since 1957, so it may be that relaxed selective pressure on this protein, combined with repeated population bottlenecks during culture maintenance, has allowed for mutations in the Neff strain MBP gene to accumulate. At the present time, without available genome data for strains more closely related to the Neff strain, it cannot be determined whether these mutations arose in nature or in culture. However, given that the divergence of the *A. polyphaga* ortholog to the MEEI 0184 strain is much less than that of the Neff strain, despite all four strains having similar lifestyles in nature, evolution of the Neff strain since being deposited in the culture collection seems likely.

## Methods

### Strains and growth conditions

*A. castellanii* strains Neff and C3 were grown on amoeba culture medium (2% Bacto Tryptone, 0.1% sodium citrate, 0.1% yeast extract), supplemented with 0.1 M glucose, 0.1 mM CaCl<sub>2</sub>, 2.5 mM KH<sub>2</sub>PO<sub>4</sub>, 4 mM MgSO<sub>4</sub>, 2.5 mM Na<sub>2</sub>HPO<sub>4</sub>, 0.05 mM Fe<sub>4</sub>O<sub>21</sub>P<sub>6</sub> at 20°C. *L. pneumophila* strain Paris was grown for 3 days on N-(2-acetamido)-2-amino-ethanesulfonic acid (ACES)-buffered charcoal-yeast (BCYE) extract agar, at 37 °C.

### Infection timecourse

Infection of *A. castellanii* C3 with *L. pneumophila* was performed using MOI 10 over 5h in infection medium (0.5% sodium citrate supplemented with 0.1 mM CaCl<sub>2</sub>, 2.5 mM KH<sub>2</sub>PO<sub>4</sub>, 4 mM MgSO<sub>4</sub>, 2.5 mM Na<sub>2</sub>HPO<sub>4</sub>, 0.05 mM Fe<sub>4</sub>O<sub>21</sub>P<sub>6</sub> at 20°C. At 5h post-infection, amoebae were collected in a 15 mL tube, pelleted by centrifugation at 300 *g* for 10 minutes and washed twice in PBS, then crosslinked in 3% formaldehyde during 20 minutes at room temperature (RT) with gentle shaking. 2.5 M glycine was then added to reach a final concentration of 0.125 M over 20 minutes, centrifuged, washed, and pellets were stored at -80 °C until DNA extraction.

### Library preparations

#### *Hi-C*

Cell pellets were suspended in 1.2ml H<sub>2</sub>O and transferred to CK14 Precellys tubes. Cells were broken with Precellys (6 cycles: 30 sec ON / 30 sec OFF) at 7500 RPM and transferred into a tube. All Hi-C libraries for *A. castellanii* strains C3 and Neff were prepared using the Arima kit and protocol with only the *DpnII* restriction enzyme. Libraries were sequenced to produce 35 bp paired-end reads on an Illumina NextSeq machine.



HyPo v1.0.1. Contigs from the polished assembly bearing more than 60% of their sequence or 51% identity to the mitochondrial sequence from the NEFF\_v1 assembly were separated from the rest of the assembly to prevent inclusion of mitochondrial contigs into the nuclear genome during scaffolding. Polished nuclear contigs were scaffolded with Hi-C reads using *instagraal* v0.1.2 with default parameters. *Instagraal-polish* was then used to fix potential errors introduced by the scaffolding procedure. Mitotic contigs were then added at the end of the scaffolded assembly and the final assembly was polished with the Illumina shotgun library data using two rounds of *pilon* polishing. The resulting assembly was edited manually to remove spurious insertion of mitochondrial contigs in the scaffold and other contaminants. The final assembly was polished again using *pilon* with *Rcorrector*-corrected reads [64]. *Minimap2* v2.17 [65] was used for all long reads alignments, and *bowtie2* v2.3.4.1 for short reads alignments.

### Genome annotation

The structural genome annotation pipeline employed here was implemented similarly as described in [66]. Briefly, RNA-Seq reads were mapped to the genome assembly using *STAR* v2.7.3a [67], followed by both *de novo* and genome-guided transcriptome assembly by *Trinity* v2.12.0 [68]. Both runs of *Trinity* were performed with *jaccard* clipping to mitigate artificial transcript fusions. The resulting transcriptome assemblies were combined and aligned to the genome assembly using *PASA* v2.4.1 [69]. Protein sequences were aligned to the genome using *Spaln* v2.4.2 [70] to recover the most information from sequence similarity. The *ab initio* predictors employed were *Augustus* v3.3.2 [71], *Snap* [72], *Genemark* v4.33 [73], and *CodingQuarry* v2.0 [74]. Finally, the *PASA* assembly, *Spaln* alignments, as well as *Augustus*, *Snap* and *Codingquarry* gene models, were combined into a single consensus with *Evidencemodeler* v1.1.1 [75].

Functional annotations were added using *funannotate* v1.5.3. [76] Repeated sequences were masked using *repeatmasker*. Predicted proteins were fed to *Interproscan* v5.22 [77], *Phobius* v1.7.1 [78] and *Eggnog-mapper* v2.0.0 [79] were used to generate functional annotations. Ribosomal RNA genes were annotated separately using *RNAmmer* v1.2 [80] with *HMMER* 2.3.2.

As described in the [Availability of data and materials](#) section, the *funannotate*-based script "*func.annot\_from\_gene\_models.sh*" used to add functional annotations to existing gene models is provided in the Zenodo record and on the associated github repository.

### Analysis of sequence divergence

To compute the proportion of substituted positions in aligned segments between the C3 and Neff strains, the two genomes were aligned using *minimap2* with the *map-ont* preset and *-c* flag. The gap-excluded sequence divergence (mismatches / (matches + mismatches)) was then computed in each primary alignment and the average of divergences (weighted by segment lengths) was computed. This is implemented in the script "*04\_compute\_seq\_divergence.py*" available in the genome analysis repository listed in [Availability of data and materials](#)

### Orthogroup inference

Orthogroups were inferred using the predicted proteomes of both the Neff and C3 strains, with *Dictyostelium discoideum*, *Physarum polycephalum*, and *Vermamoeba vermiformis* as outgroups to improve the accuracy of orthogroup inference. The outgroup predicted proteomes were retrieved from PhyloFisher [81]. Both Broccoli [32] and OrthoFinder [33] were run with default settings for orthogroup inference.

### Gene content comparison of Neff and C3 strains

Custom Python scripts were used to retrieve genes unique to each *A. castellanii* strain, as well as orthogroups that were shared between the two strains. Genes were only determined to be strain-specific or shared if both Broccoli and OrthoFinder assigned them as such; genes were excluded from the analysis if both tools did not agree. For both strains, functional assignments for each gene ID were extracted from funannotate output and tabulated. The tabulated assignments and strain-specific gene IDs were fed into the R package topGO [82] to analyze GO term enrichment in the strain-specific genes. Fisher's exact test with the weight algorithm was implemented in topGO for the Neff- and C3-specific genes for each of the three ontologies (biological process, cellular component, and molecular function). When building the GOdata objects for these three ontologies, nodeSize was set to 10 for both the biological process and molecular function ontologies, and 5 for the cellular component ontology due to the lower number of GO terms in this ontology.

### Mannose Binding Protein Comparison

Mannose binding protein (MBP) amino acid sequences from three strains of *A. castellanii* (Neff, C3, and MEEI 0184) and one strain of *Acanthamoeba polyphaga* were retrieved, aligned using MAFFT-linsi (v7.475) [83], and visualized in Jalview (v2.11.1.3) [84]. The MEEI 0184 strain sequence was retrieved from NCBI (Accession: AAT37865.1), and the Neff and C3 sequences were retrieved from the predicted proteomes generated in this study with the MEEI 0184 sequence as a BLASTp [34] query. The *A. polyphaga* genome does not have a publicly available predicted proteome, so its MBP protein sequence was manually extracted from several contigs in the genome sequence (NCBI accession: GCA\_000826345.1) using tBLASTn with the MEEI 0184 sequence as a query (the sequence encoding the first 8 amino acids of the protein could not be found in the genome due to a truncated contig).

### Hi-C analyses

Reads were aligned with bowtie2 v2.4.1, and Hi-C matrices were generated using hicstuff v3.0.1 [85]. For all comparative analyses, matrices were downsampled to the same number of contacts using cooltools (<https://www.github.com/mirnylab/cooltools>) and balancing normalization was performed using the ICE algorithm [86]. Loops and domain borders were detected using Chromosight v1.6.1 [38] using the merged replicates at a resolution of 2 kbp. We measured the intensity changes in Chromosight scores during infection using pareidolia (v0.6.1) [87] on 3 pseudo replicates generated by sampling the merged contact maps, as described in [88]. This was done to account for contact coverage heterogeneity across replicates. The 20% threshold used to select differential patterns amounts to 1.2% false detections for loops and 2.3% for borders when comparing pseudo-replicates from the same condition.

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## Availability of data and materials

Sequencing datasets have been deposited in SRA under bioprojects PRJNA599339 and PRJNA487265. All processed data, as well as the assemblies and annotations used in this work are available on zenodo record <https://zenodo.org/record/5507417>. Strains supporting the findings of this study are available from the corresponding authors.

The analyses are packaged into the following snakemake pipelines available on github. Hybrid genome assembly: [https://github.com/cmdoret/Acastellanii\\_hybrid\\_assembly](https://github.com/cmdoret/Acastellanii_hybrid_assembly), functional annotation of *A. castellanii*: [https://github.com/cmdoret/Acastellanii\\_genome\\_annotation](https://github.com/cmdoret/Acastellanii_genome_annotation), analyses of genomic features in *A. castellanii*: [https://github.com/cmdoret/Acastellanii\\_genome\\_analysis](https://github.com/cmdoret/Acastellanii_genome_analysis), changes during infection by Legionella: [https://github.com/cmdoret/Acastellanii\\_legionella\\_infection](https://github.com/cmdoret/Acastellanii_legionella_infection). The dataset(s) supporting the conclusions of this article are available in the Zenodo repository <https://zenodo.org/record/5507417>.

## Competing interests

The authors declare that they have no competing interests.

## Consent for publication

All authors gave their consent for publication of this manuscript.

## Authors' contributions

C.M.D and M.J.C performed analyses. P.E. Performed infection experiments and DNA extractions. M.J.C performed DNA extraction. M.J.C and C.M.D did the Nanopore sequencing. A.T. constructed Hi-C and shotgun libraries. M.J.C., C.M.D., B.C., M.S., M.W.G. and B.F.L. contributed to the curation and improvement of the genome assembly. All authors contributed to writing the manuscript.

## Author details

<sup>1</sup>Institut Pasteur, Université de Paris, CNRS UMR3525, Unité Régulation Spatiale des Génomes, F-75015, Paris, France. <sup>2</sup>Collège Doctoral, Sorbonne Université, F-75005, Paris, France. <sup>3</sup>Department of Biochemistry and Molecular Biology and Institute for Comparative Genomics, Dalhousie University, Sir Charles Tupper Medical Building, 5850 College Street, B3H 4R2 Halifax, Nova Scotia, Canada. <sup>4</sup>Institut Pasteur, Université de Paris, CNRS UMR3525, Unité Biologie des Bactéries Intracellulaires, F-75015, Paris, France. <sup>5</sup>Robert Cedergren Centre for Bioinformatics and Genomics, Département de Biochimie, Université de Montréal, Montréal, QC, Canada.

## References

1. Puschikaw, B.M.: Über die Verbreitung der Süßwasser-Protozoen durch die Luft. Arch Protistent. (23), 323–362 (1913). Accessed 2021-09-07
2. Volkonsky, M.: Hartmannella castellanii Douglas et classification des Hartmannelles. Archives de zoologie expérimentale et générale de Paris **72**(3), 317–339 (1931)
3. Rodríguez-Zaragoza, S.: Ecology of Free-Living Amoeboeae. Critical Reviews in Microbiology **20**(3), 225–241 (1994). doi:[10.3109/10408419409114556](https://doi.org/10.3109/10408419409114556). Publisher: Taylor & Francis .eprint: <https://doi.org/10.3109/10408419409114556>. Accessed 2021-04-26
4. Siddiqui, R., Khan, N.A.: Biology and pathogenesis of Acanthamoeba. Parasites & Vectors **5**(1), 6 (2012). doi:[10.1186/1756-3305-5-6](https://doi.org/10.1186/1756-3305-5-6). Accessed 2021-03-25
5. Visvesvara, G.S., Moura, H., Schuster, F.L.: Pathogenic and opportunistic free-living amoebae: Acanthamoeba spp., Balamuthia mandrillaris, Naegleria fowleri, and Sappinia diploidea. FEMS Immunology & Medical Microbiology **50**(1), 1–26 (2007). doi:[10.1111/j.1574-695X.2007.00232.x](https://doi.org/10.1111/j.1574-695X.2007.00232.x). .eprint: <https://onlinelibrary.wiley.com/doi/pdf/10.1111/j.1574-695X.2007.00232.x>. Accessed 2021-09-07
6. Castellani, A.: An amoeba found in culture of yeast: preliminary note. J Trop Med Hyg **33**, 160 (1930)
7. Samba-Louaka, A., Delafont, V., Rodier, M.-H., Cateau, E., Héchar, Y.: Free-living amoebae and squatters in the wild: ecological and molecular features. FEMS Microbiology Reviews **43**(4), 415–434 (2019). doi:[10.1093/femsre/fuz011](https://doi.org/10.1093/femsre/fuz011). Accessed 2021-04-26
8. Drozanski, W.: Fatal bacterial infection in soil amoebae. Acta Microbiologica Polonica (1952) **5**(3-4), 315–317 (1956)
9. Rowbotham, T.J.: Preliminary report on the pathogenicity of Legionella pneumophila for freshwater and soil amoebae. Journal of Clinical Pathology **33**(12), 1179–1183 (1980). doi:[10.1136/jcp.33.12.1179](https://doi.org/10.1136/jcp.33.12.1179). Accessed 2021-03-25

10. Kilvington, S., Price, J.: Survival of *Legionella pneumophila* within cysts of *Acanthamoeba polyphaga* following chlorine exposure. *Journal of Applied Bacteriology* **68**(5), 519–525 (1990). doi:[10.1111/j.1365-2672.1990.tb02904.x](https://doi.org/10.1111/j.1365-2672.1990.tb02904.x). Accessed 2021-03-25
11. Pagnier, I., Raoult, D., La Scola, B.: Isolation and identification of amoeba-resisting bacteria from water in human environment by using an *Acanthamoeba polyphaga* co-culture procedure. *Environmental Microbiology* **10**(5), 1135–1144 (2008). doi:[10.1111/j.1462-2920.2007.01530.x](https://doi.org/10.1111/j.1462-2920.2007.01530.x). Accessed 2021-03-25
12. Lasheras, A., Boulestreau, H., Rogues, A.-M., Ohayon-Courtes, C., Labadie, J.-C., Gachie, J.-P.: Influence of amoebae and physical and chemical characteristics of water on presence and proliferation of *Legionella* species in hospital water systems. *American Journal of Infection Control* **34**(8), 520–525 (2006). doi:[10.1016/j.ajic.2006.03.007](https://doi.org/10.1016/j.ajic.2006.03.007). Accessed 2021-03-25
13. Ikeda, M., Yabuuchi, E.: Ecological Studies of *Legionella* Species: I. Viable Counts of *Legionella pneumophila* in Cooling Tower Water. *Microbiology and Immunology* **30**(5), 413–423 (1986). doi:[10.1111/j.1348-0421.1986.tb02967.x](https://doi.org/10.1111/j.1348-0421.1986.tb02967.x). Accessed 2021-03-25
14. Mondino, S., Schmidt, S., Rolando, M., Escoll, P., Gomez-Valero, L., Buchrieser, C.: Legionnaires' Disease: State of the Art Knowledge of Pathogenesis Mechanisms of *Legionella*. *Annual Review of Pathology: Mechanisms of Disease* **15**(1), 439–466 (2020). doi:[10.1146/annurev-pathmechdis-012419-032742](https://doi.org/10.1146/annurev-pathmechdis-012419-032742). Publisher: Annual Reviews. Accessed 2021-04-26
15. Kubori, T., Nagai, H.: The Type IVB secretion system: an enigmatic chimera. *Current Opinion in Microbiology* **29**, 22–29 (2016). doi:[10.1016/j.mib.2015.10.001](https://doi.org/10.1016/j.mib.2015.10.001)
16. Isberg, R.R., O'Connor, T.J., Heidtman, M.: The *Legionella pneumophila* replication vacuole: making a cosy niche inside host cells. *Nature Reviews Microbiology* **7**(1), 13–24 (2009). doi:[10.1038/nrmicro1967](https://doi.org/10.1038/nrmicro1967). Accessed 2021-03-25
17. Ensminger, A.W.: *Legionella pneumophila*, armed to the hilt: justifying the largest arsenal of effectors in the bacterial world. *Current Opinion in Microbiology* **29**, 74–80 (2016). doi:[10.1016/j.mib.2015.11.002](https://doi.org/10.1016/j.mib.2015.11.002)
18. Swart, A.L., Gomez-Valero, L., Buchrieser, C., Hilbi, H.: Evolution and function of bacterial RCC1 repeat effectors. *Cellular Microbiology* **22**(10), 13246 (2020). doi:[10.1111/cmi.13246](https://doi.org/10.1111/cmi.13246)
19. Hubber, A., Roy, C.R.: Modulation of host cell function by *Legionella pneumophila* type IV effectors. *Annual Review of Cell and Developmental Biology* **26**, 261–283 (2010). doi:[10.1146/annurev-cellbio-100109-104034](https://doi.org/10.1146/annurev-cellbio-100109-104034)
20. Qiu, J., Luo, Z.-Q.: *Legionella* and *Coxiella* effectors: strength in diversity and activity. *Nature Reviews Microbiology* **15**(10), 591–605 (2017). doi:[10.1038/nrmicro.2017.67](https://doi.org/10.1038/nrmicro.2017.67)
21. Rolando, M., Sanulli, S., Rusnick, C., Gomez-Valero, L., Bertholet, C., Sahr, T., Margueron, R., Buchrieser, C.: *Legionella pneumophila* Effector RomA Uniquely Modifies Host Chromatin to Repress Gene Expression and Promote Intracellular Bacterial Replication. *Cell Host & Microbe* **13**(4), 395–405 (2013). doi:[10.1016/j.chom.2013.03.004](https://doi.org/10.1016/j.chom.2013.03.004). Accessed 2021-03-25
22. Li, P., Vassiliadis, D., Ong, S.Y., Bennett-Wood, V., Sugimoto, C., Yamagishi, J., Hartland, E.L., Pasricha, S.: *Legionella pneumophila* Infection Rewires the *Acanthamoeba castellanii* Transcriptome, Highlighting a Class of Sirtuin Genes. *Frontiers in Cellular and Infection Microbiology* **10**, 428 (2020). doi:[10.3389/fcimb.2020.00428](https://doi.org/10.3389/fcimb.2020.00428). Accessed 2021-03-25
23. Rennie, S., Dalby, M., van Duin, L., Andersson, R.: Transcriptional decomposition reveals active chromatin architectures and cell specific regulatory interactions. *Nature Communications* **9**(1), 487 (2018). doi:[10.1038/s41467-017-02798-1](https://doi.org/10.1038/s41467-017-02798-1). Accessed 2021-03-25
24. Nora, E.P., Lajoie, B.R., Schulz, E.G., Giorgetti, L., Okamoto, I., Servant, N., Piolot, T., van Berkum, N.L., Meisig, J., Sedat, J., Gribnau, J., Barillot, E., Blüthgen, N., Dekker, J., Heard, E.: Spatial partitioning of the regulatory landscape of the X-inactivation centre. *Nature* **485**(7398), 381–385 (2012). doi:[10.1038/nature11049](https://doi.org/10.1038/nature11049). Accessed 2021-03-25
25. Clarke, M., Lohan, A.J., Liu, B., Lagkourdos, I., Roy, S., Zafar, N., Bertelli, C., Schilde, C., Kianianmomeni, A., Bürglin, T.R., Frech, C., Turcotte, B., Kopec, K.O., Synnott, J.M., Choo, C., Paponov, I., Finkler, A., Heng Tan, C., Hutchins, A.P., Weinmeier, T., Rattei, T., Chu, J.S., Gimenez, G., Irimia, M., Rigden, D.J., Fitzpatrick, D.A., Lorenzo-Morales, J., Bateman, A., Chiu, C.-H., Tang, P., Hegemann, P., Fromm, H., Raoult, D., Greub, G., Miranda-Saavedra, D., Chen, N., Nash, P., Ginger, M.L., Horn, M., Schaap, P., Caler, L., Loftus, B.J.: Genome of *Acanthamoeba castellanii* highlights extensive lateral gene transfer and early evolution of tyrosine kinase signaling. *Genome Biology* **14**(2), 11 (2013). doi:[10.1186/gb-2013-14-2-r11](https://doi.org/10.1186/gb-2013-14-2-r11). Accessed 2021-03-25
26. Neff, R.J.: Purification, Axenic Cultivation, and Description of a Soil Amoeba, *Acanthamoeba* sp. *The Journal of Protozoology* **4**(3), 176–182 (1957). doi:[10.1111/j.1550-7408.1957.tb02505.x](https://doi.org/10.1111/j.1550-7408.1957.tb02505.x). Accessed 2021-03-25
27. Michel, R., Hauröder, B.: Isolation of an *Acanthamoeba* Strain with Intracellular *Burkholderia pickettii* Infection. *Zentralblatt für Bakteriologie* **285**(4), 541–557 (1997). doi:[10.1016/S0934-8840\(97\)80116-8](https://doi.org/10.1016/S0934-8840(97)80116-8). Accessed 2021-03-25
28. Rimm, D.L., Pollard, T.D., Hieter, P.: Resolution of *Acanthamoeba castellanii* chromosomes by pulsed field gel electrophoresis and construction of the initial linkage map. *Chromosoma* **97**(3), 219–223 (1988). doi:[10.1007/BF00292964](https://doi.org/10.1007/BF00292964). Accessed 2021-03-25
29. Kundu, R., Casey, J., Sung, W.-K.: HyPo: Super Fast & Accurate Polisher for Long Read Genome Assemblies. preprint, *Bioinformatics* (December 2019). doi:[10.1101/2019.12.19.882506](https://doi.org/10.1101/2019.12.19.882506). <https://doi.org/10.1101/2019.12.19.882506> Accessed 2021-03-25
30. Baudry, L., Guiglielmoni, N., Marie-Nelly, H., Cormier, A., Marbouty, M., Avia, K., Mie, Y.L., Godfroy, O., Sterck, L., Cock, J.M., Zimmer, C., Coelho, S.M., Koszul, R.: instaGRAAL: chromosome-level quality scaffolding of genomes using a proximity ligation-based scaffold. *Genome Biology* **21**(1), 148 (2020). doi:[10.1186/s13059-020-02041-z](https://doi.org/10.1186/s13059-020-02041-z). Accessed 2021-04-27
31. Marie-Nelly, H., Marbouty, M., Cournac, A., Flot, J.-F., Liti, G., Parodi, D.P., Syan, S., Guillén, N., Margeot, A., Zimmer, C., Koszul, R.: High-quality genome (re)assembly using chromosomal contact data. *Nature Communications* **5**(1), 5695 (2014). doi:[10.1038/ncomms5695](https://doi.org/10.1038/ncomms5695). Accessed 2021-03-25
32. Derelle, R., Philippe, H., Colbourne, J.K.: Broccoli: Combining Phylogenetic and Network Analyses for

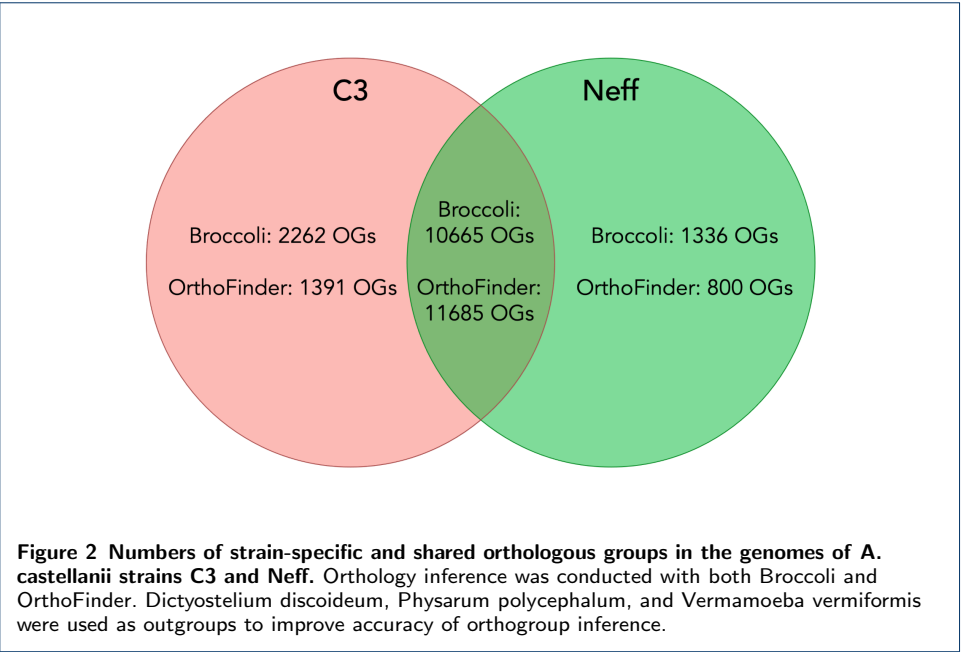
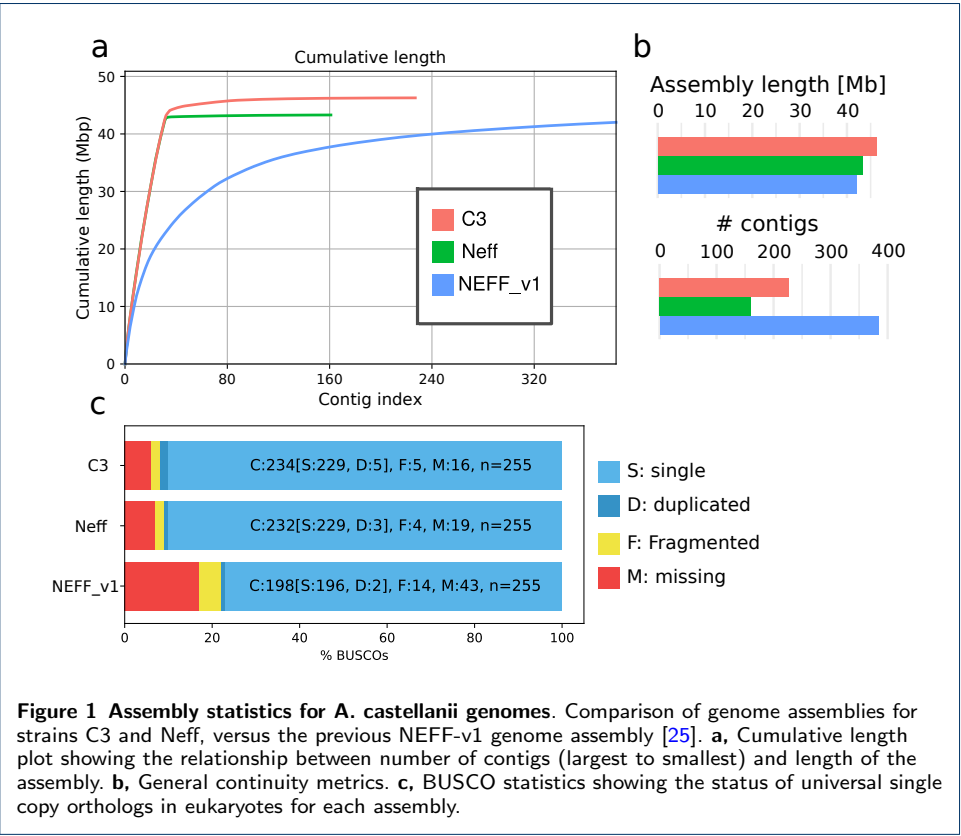


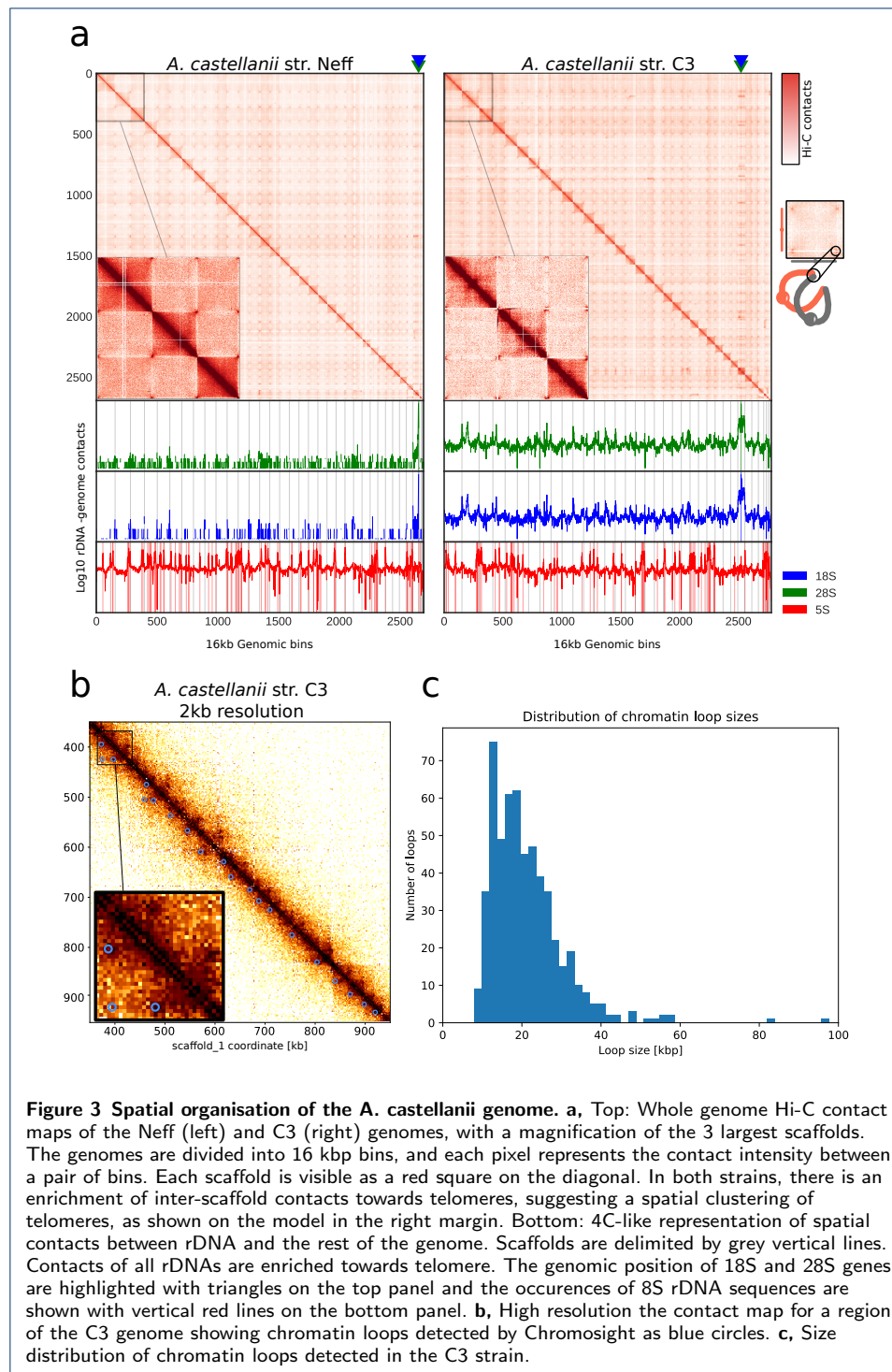
- Orthology Assignment. *Molecular Biology and Evolution* **37**(11), 3389–3396 (2020). doi:[10.1093/molbev/msaa159](https://doi.org/10.1093/molbev/msaa159). Accessed 2021-10-06
33. Emms, D.M., Kelly, S.: OrthoFinder: phylogenetic orthology inference for comparative genomics. *Genome Biology* **20**(1), 1–14 (2019). doi:[10.1186/s13059-019-1832-y](https://doi.org/10.1186/s13059-019-1832-y). Number: 1 Publisher: BioMed Central. Accessed 2021-10-06
34. Altschul, S.F., Gish, W., Miller, W., Myers, E.W., Lipman, D.J.: Basic local alignment search tool. *Journal of Molecular Biology* **215**(3), 403–410 (1990). doi:[10.1016/S0022-2836\(05\)80360-2](https://doi.org/10.1016/S0022-2836(05)80360-2). Accessed 2021-10-06
35. Declerck, P., Behets, J., De Keersmaecker, B., Ollevier, F.: Receptor-mediated uptake of *Legionella pneumophila* by *Acanthamoeba castellanii* and *Naegleria lovaniensis*. *Journal of Applied Microbiology* **103**(6), 2697–2703 (2007). doi:[10.1111/j.1365-2672.2007.03530.x](https://doi.org/10.1111/j.1365-2672.2007.03530.x). Preprint: <https://onlinelibrary.wiley.com/doi/pdf/10.1111/j.1365-2672.2007.03530.x>. Accessed 2021-10-04
36. Garate, M., Cao, Z., Bateman, E., Panjwani, N.: Cloning and Characterization of a Novel Mannose-binding Protein of *Acanthamoeba*\*. *Journal of Biological Chemistry* **279**(28), 29849–29856 (2004). doi:[10.1074/jbc.M402334200](https://doi.org/10.1074/jbc.M402334200). Publisher: Elsevier. Accessed 2021-10-04
37. Garate, M., Cubillos, I., Marchant, J., Panjwani, N.: Biochemical Characterization and Functional Studies of *Acanthamoeba* Mannose-Binding Protein. *Infection and Immunity* **73**(9), 5775–5781 (2005). doi:[10.1128/IAI.73.9.5775-5781.2005](https://doi.org/10.1128/IAI.73.9.5775-5781.2005). Publisher: American Society for Microbiology. Accessed 2021-10-04
38. Matthey-Doret, C., Baudry, L., Breuer, A., Montagne, R., Guiguelmon, N., Scolari, V., Jean, E., Campeas, A., Chanut, P.H., Oriol, E., Méot, A., Politis, L., Vigouroux, A., Moreau, P., Koszul, R., Cournac, A.: Computer vision for pattern detection in chromosome contact maps. *Nature Communications* **11**(1), 5795 (2020). doi:[10.1038/s41467-020-19562-7](https://doi.org/10.1038/s41467-020-19562-7). Number: 1 Publisher: Nature Publishing Group. Accessed 2021-05-07
39. Hsieh, T.-H.S., Weiner, A., Lajoie, B., Dekker, J., Friedman, N., Rando, O.J.: Mapping Nucleosome Resolution Chromosome Folding in Yeast by Micro-C. *Cell* **162**(1), 108–119 (2015). doi:[10.1016/j.cell.2015.05.048](https://doi.org/10.1016/j.cell.2015.05.048). Accessed 2021-08-23
40. Cockram, C., Thierry, A., Gorlas, A., Lestini, R., Koszul, R.: Euryarchaeal genomes are folded into SMC-dependent loops and domains, but lack transcription-mediated compartmentalization. *Molecular Cell* **81**(3), 459–472 (2021). doi:[10.1016/j.molcel.2020.12.013](https://doi.org/10.1016/j.molcel.2020.12.013). Accessed 2021-08-23
41. Cazalet, C., Rusniok, C., Brüggemann, H., Zidane, N., Magnier, A., Ma, L., Tichit, M., Jarraud, S., Bouchier, C., Vandenesch, F., Kunst, F., Etienne, J., Glaser, P., Buchrieser, C.: Evidence in the *Legionella pneumophila* genome for exploitation of host cell functions and high genome plasticity. *Nature Genetics* **36**(11), 1165–1173 (2004). doi:[10.1038/ng1447](https://doi.org/10.1038/ng1447). Accessed 2021-03-25
42. Baudry, L., Millot, G.A., Thierry, A., Koszul, R., Scolari, V.F.: Serpentine: a flexible 2D binning method for differential Hi-C analysis. *Bioinformatics* **36**(12), 3645–3651 (2020). doi:[10.1093/bioinformatics/btaa249](https://doi.org/10.1093/bioinformatics/btaa249). Accessed 2021-08-24
43. Barbieri, M., Chotalia, M., Fraser, J., Lavitas, L.-M., Dostie, J., Pombo, A., Nicodemi, M.: Complexity of chromatin folding is captured by the strings and binders switch model. *Proceedings of the National Academy of Sciences* **109**(40), 16173–16178 (2012). doi:[10.1073/pnas.1204799109](https://doi.org/10.1073/pnas.1204799109). Publisher: National Academy of Sciences Section: Biological Sciences. Accessed 2021-08-24
44. Le, T.B.K., Imakaev, M.V., Mirny, L.A., Laub, M.T.: High-Resolution Mapping of the Spatial Organization of a Bacterial Chromosome. *Science* **342**(6159), 731–734 (2013). doi:[10.1126/science.1242059](https://doi.org/10.1126/science.1242059). Publisher: American Association for the Advancement of Science Section: Report. Accessed 2021-08-10
45. Shumate, A., Salzberg, S.L.: Liftoff: accurate mapping of gene annotations. *Bioinformatics* **37**(12), 1639–1643 (2021). doi:[10.1093/bioinformatics/btaa1016](https://doi.org/10.1093/bioinformatics/btaa1016). Accessed 2021-08-24
46. Byers, T.J.: Molecular Biology of DNA in *Acanthamoeba*, *Amoeba*, *Entamoeba*, and *Naegleria*. In: *International Review of Cytology* vol. 99, pp. 311–341. Elsevier, ??? (1986). doi:[10.1016/S0074-7696\(08\)61430-8](https://doi.org/10.1016/S0074-7696(08)61430-8). <https://linkinghub.elsevier.com/retrieve/pii/S0074769608614308> Accessed 2021-03-25
47. Gicquaud, C., Tremblay, N.: Observations with Hoechst Staining of Amitosis in *Acanthamoeba castellanii*. *The Journal of Protozoology* **38**(3), 221–224 (1991). doi:[10.1111/j.1550-7408.1991.tb04432.x](https://doi.org/10.1111/j.1550-7408.1991.tb04432.x). Accessed 2021-03-25
48. Yang, Q., Zwick, M.G., Paule, M.R.: Sequence organization of the *Acanthamoeba* rRNA intergenic spacer: identification of transcriptional enhancers. *Nucleic Acids Research* **22**(22), 4798–4805 (1994). doi:[10.1093/nar/22.22.4798](https://doi.org/10.1093/nar/22.22.4798). Accessed 2021-03-25
49. Rabl, C.: Über zelltheilung. *Morphol. Jahrbuch* (10), 214–330 (1885)
50. Mengue, L., Régnacq, M., Aucher, W., Portier, E., Héchard, Y., Samba-Louaka, A.: *Legionella pneumophila* prevents proliferation of its natural host *Acanthamoeba castellanii*. *Scientific Reports* **6**(1), 36448 (2016). doi:[10.1038/srep36448](https://doi.org/10.1038/srep36448). Accessed 2021-03-25
51. de Jesús-Díaz, D.A., Murphy, C., Sol, A., Dorer, M., Isberg, R.R.: Host Cell S Phase Restricts *Legionella pneumophila* Intracellular Replication by Destabilizing the Membrane-Bound Replication Compartment. *mBio* **8**(4), 02345–16 (2017). doi:[10.1128/mBio.02345-16](https://doi.org/10.1128/mBio.02345-16). Publisher: American Society for Microbiology. Accessed 2021-09-07
52. Quinet, T., Samba-Louaka, A., Héchard, Y., Van Doninck, K., Van der Henst, C.: Delayed cytokinesis generates multinuclearity and potential advantages in the amoeba *Acanthamoeba castellanii* Neff strain. *Scientific Reports* **10**(1), 12109 (2020). doi:[10.1038/s41598-020-68694-9](https://doi.org/10.1038/s41598-020-68694-9). Bandiera\_abtest: a Cc.license.type: cc.by Cg.type: Nature Research Journals Number: 1 Primary\_atype: Research Publisher: Nature Publishing Group Subject.term: Evolutionary ecology;Microbial ecology;Parasitology Subject.term.id: evolutionary-ecology;microbial-ecology;parasitology. Accessed 2021-09-07
53. Rothmeier, E., Pfaffinger, G., Hoffmann, C., Harrison, C.F., Grabmayr, H., Repnik, U., Hannemann, M., Wölke, S., Bausch, A., Griffiths, G., Müller-Taubenberger, A., Itzen, A., Hilbi, H.: Activation of Ran GTPase by a *Legionella* Effector Promotes Microtubule Polymerization, Pathogen Vacuole Motility and Infection. *PLOS Pathogens* **9**(9), 1003598 (2013). doi:[10.1371/journal.ppat.1003598](https://doi.org/10.1371/journal.ppat.1003598). Publisher: Public Library of Science. Accessed 2021-09-07
54. Escoll, P., Song, O.-R., Viana, F., Steiner, B., Lagache, T., Olivo-Marin, J.-C., Impens, F., Brodin, P., Hilbi,

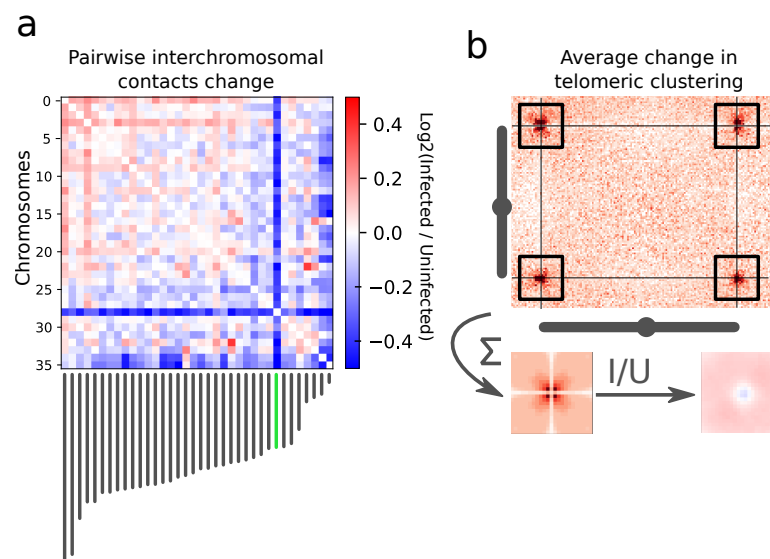
- H., Buchrieser, C.: Legionella pneumophila Modulates Mitochondrial Dynamics to Trigger Metabolic Repurposing of Infected Macrophages. *Cell Host & Microbe* **22**(3), 302–3167 (2017). doi:[10.1016/j.chom.2017.07.020](https://doi.org/10.1016/j.chom.2017.07.020). Accessed 2021-03-25
55. Escoll, P., Mondino, S., Rolando, M., Buchrieser, C.: Targeting of host organelles by pathogenic bacteria: a sophisticated subversion strategy. *Nature Reviews Microbiology* **14**(1), 5–19 (2016). doi:[10.1038/nrmicro.2015.1](https://doi.org/10.1038/nrmicro.2015.1). Bandiera\_abtest: a Cg\_type: Nature Research Journals Number: 1 Primary\_atype: Reviews Publisher: Nature Publishing Group Subject\_term: Bacterial host response;Bacterial pathogenesis;Bacterial secretion;Pathogens Subject\_term\_id: bacterial-host-response;bacterial-pathogenesis;bacterial-secretion;pathogens. Accessed 2021-09-07
56. Escoll, P., Platon, L., Dramé, M., Sahr, T., Schmidt, S., Rusniok, C., Buchrieser, C.: Reverting the mode of action of the mitochondrial FOF1-ATPase by Legionella pneumophila preserves its replication niche. Technical report (May 2021). doi:[10.1101/2021.05.12.443790](https://doi.org/10.1101/2021.05.12.443790). Company: Cold Spring Harbor Laboratory Distributor: Cold Spring Harbor Laboratory Label: Cold Spring Harbor Laboratory Section: New Results Type: article. <https://www.biorxiv.org/content/10.1101/2021.05.12.443790v1> Accessed 2021-09-07
57. Escoll, P., Buchrieser, C.: Metabolic reprogramming: an innate cellular defence mechanism against intracellular bacteria? *Current Opinion in Immunology* **60**, 117–123 (2019). doi:[10.1016/j.coi.2019.05.009](https://doi.org/10.1016/j.coi.2019.05.009). Accessed 2021-09-07
58. Garcia-Luis, J., Lazar-Stefanita, L., Gutierrez-Escribano, P., Thierry, A., Cournac, A., García, A., González, S., Sánchez, M., Jarmuz, A., Montoya, A., Dore, M., Kramer, H., Karimi, M.M., Antequera, F., Koszul, R., Aragon, L.: FACT mediates cohesin function on chromatin. *Nature structural & molecular biology* **26**(10), 970–979 (2019). doi:[10.1038/s41594-019-0307-x](https://doi.org/10.1038/s41594-019-0307-x). Accessed 2021-08-23
59. Marbouty, M., Le Gall, A., Cattoni, D., Cournac, A., Koh, A., Fiche, J.-B., Mozziconacci, J., Murray, H., Koszul, R., Nollmann, M.: Condensin- and Replication-Mediated Bacterial Chromosome Folding and Origin Condensation Revealed by Hi-C and Super-resolution Imaging. *Molecular Cell* **59**(4), 588–602 (2015). doi:[10.1016/j.molcel.2015.07.020](https://doi.org/10.1016/j.molcel.2015.07.020). Accessed 2021-03-25
60. Anchimiuk, A., Lioy, V.S., Bock, F.P., Minnen, A., Boccard, F., Gruber, S.: A low Smc flux avoids collisions and facilitates chromosome organization in *Bacillus subtilis*. *eLife* **10**, 65467 (2021). doi:[10.7554/eLife.65467](https://doi.org/10.7554/eLife.65467). Publisher: eLife Sciences Publications, Ltd. Accessed 2021-08-23
61. Ramírez, F., Bhardwaj, V., Arrigoni, L., Lam, K.C., Grüning, B.A., Villaveces, J., Habermann, B., Akhtar, A., Manke, T.: High-resolution TADs reveal DNA sequences underlying genome organization in flies. *Nature Communications* **9**(1), 189 (2018). doi:[10.1038/s41467-017-02525-w](https://doi.org/10.1038/s41467-017-02525-w). Number: 1 Publisher: Nature Publishing Group. Accessed 2021-04-28
62. Legendre, M., Lartigue, A., Bertaux, L., Jeudy, S., Bartoli, J., Lescot, M., Alempic, J.-M., Ramus, C., Bruley, C., Labadie, K., Shmakova, L., Rivkina, E., Couté, Y., Abergel, C., Claverie, J.-M.: In-depth study of *Mollivirus sibericum*, a new 30,000-y-old giant virus infecting *Acanthamoeba*. *Proceedings of the National Academy of Sciences* **112**(38), 5327–5335 (2015). doi:[10.1073/pnas.1510795112](https://doi.org/10.1073/pnas.1510795112). Accessed 2021-03-25
63. Harb, O.S., Venkataraman, C., Haack, B.J., Gao, L.-Y., Kwaik, Y.A.: Heterogeneity in the Attachment and Uptake Mechanisms of the Legionnaires' Disease Bacterium, *Legionella pneumophila*, by Protozoan Hosts. *Applied and Environmental Microbiology* **64**(1), 126–132 (1998). doi:[10.1128/AEM.64.1.126-132.1998](https://doi.org/10.1128/AEM.64.1.126-132.1998). Publisher: American Society for Microbiology. Accessed 2021-10-07
64. Song, L., Florea, L.: Rcorrector: efficient and accurate error correction for Illumina RNA-seq reads. *GigaScience* **4**(1), 1–8 (2015). doi:[10.1186/s13742-015-0089-y](https://doi.org/10.1186/s13742-015-0089-y). Number: 1 Publisher: BioMed Central. Accessed 2021-08-05
65. Li, H.: Minimap2: pairwise alignment for nucleotide sequences. *Bioinformatics (Oxford, England)* **34**(18), 3094–3100 (2018). doi:[10.1093/bioinformatics/bty191](https://doi.org/10.1093/bioinformatics/bty191)
66. Gray, M.W., Burger, G., Derelle, R., Klimeš, V., Leger, M.M., Sarrasin, M., Vlcek, C., Roger, A.J., Elias, M., Lang, B.F.: The draft nuclear genome sequence and predicted mitochondrial proteome of *Andalucia godoyi*, a protist with the most gene-rich and bacteria-like mitochondrial genome. *BMC biology* **18**(1), 22 (2020). doi:[10.1186/s12915-020-0741-6](https://doi.org/10.1186/s12915-020-0741-6)
67. Dobin, A., Davis, C.A., Schlesinger, F., Drenkow, J., Zaleski, C., Jha, S., Batut, P., Chaisson, M., Gingeras, T.R.: STAR: ultrafast universal RNA-seq aligner. *Bioinformatics (Oxford, England)* **29**(1), 15–21 (2013). doi:[10.1093/bioinformatics/bts635](https://doi.org/10.1093/bioinformatics/bts635)
68. Grabherr, M.G., Haas, B.J., Yassour, M., Levin, J.Z., Thompson, D.A., Amit, I., Adiconis, X., Fan, L., Raychowdhury, R., Zeng, Q., Chen, Z., Mauceli, E., Hacohen, N., Gnirke, A., Rhind, N., di Palma, F., Birren, B.W., Nusbaum, C., Lindblad-Toh, K., Friedman, N., Regev, A.: Full-length transcriptome assembly from RNA-Seq data without a reference genome. *Nature Biotechnology* **29**(7), 644–652 (2011). doi:[10.1038/nbt.1883](https://doi.org/10.1038/nbt.1883). Bandiera\_abtest: a Cg\_type: Nature Research Journals Number: 7 Primary\_atype: Research Publisher: Nature Publishing Group Subject\_term: Software;Transcriptomics Subject\_term\_id: software;transcriptomics. Accessed 2021-10-06
69. Haas, B.J., Delcher, A.L., Mount, S.M., Wortman, J.R., Smith Jr, R.K., Hannick, L.I., Maiti, R., Ronning, C.M., Rusch, D.B., Town, C.D., Salzberg, S.L., White, O.: Improving the Arabidopsis genome annotation using maximal transcript alignment assemblies. *Nucleic Acids Research* **31**(19), 5654–5666 (2003). doi:[10.1093/nar/gkg770](https://doi.org/10.1093/nar/gkg770). Accessed 2021-10-06
70. Gotoh, O.: A space-efficient and accurate method for mapping and aligning cDNA sequences onto genomic sequence. *Nucleic Acids Research* **36**(8), 2630–2638 (2008). doi:[10.1093/nar/gkn105](https://doi.org/10.1093/nar/gkn105). Accessed 2021-10-06
71. Stanke, M., Schöffmann, O., Morgenstern, B., Waack, S.: Gene prediction in eukaryotes with a generalized hidden Markov model that uses hints from external sources. *BMC Bioinformatics* **7**(1), 1–11 (2006). doi:[10.1186/1471-2105-7-62](https://doi.org/10.1186/1471-2105-7-62). Number: 1 Publisher: BioMed Central. Accessed 2021-10-06
72. Korf, I.: Gene finding in novel genomes. *BMC Bioinformatics* **5**(1), 1–9 (2004). doi:[10.1186/1471-2105-5-59](https://doi.org/10.1186/1471-2105-5-59). Number: 1 Publisher: BioMed Central. Accessed 2021-10-06
73. Lomsadze, A., Burns, P.D., Borodovsky, M.: Integration of mapped RNA-Seq reads into automatic training of eukaryotic gene finding algorithm. *Nucleic Acids Research* **42**(15), 119 (2014). doi:[10.1093/nar/gku557](https://doi.org/10.1093/nar/gku557). Accessed 2021-10-06

74. Testa, A.C., Hane, J.K., Ellwood, S.R., Oliver, R.P.: CodingQuarry: highly accurate hidden Markov model gene prediction in fungal genomes using RNA-seq transcripts. *BMC Genomics* **16**(1), 1–12 (2015). doi:[10.1186/s12864-015-1344-4](https://doi.org/10.1186/s12864-015-1344-4). Number: 1 Publisher: BioMed Central. Accessed 2021-10-06
75. Haas, B.J., Salzberg, S.L., Zhu, W., Pertea, M., Allen, J.E., Orvis, J., White, O., Buell, C.R., Wortman, J.R.: Automated eukaryotic gene structure annotation using EVIDENCEModeler and the Program to Assemble Spliced Alignments. *Genome Biology* **9**(1), 1–22 (2008). doi:[10.1186/gb-2008-9-1-r7](https://doi.org/10.1186/gb-2008-9-1-r7). Number: 1 Publisher: BioMed Central. Accessed 2021-10-06
76. Palmer, J., Stajich, J.: nextgenusfs/funcannotate: funcannotate v1.5.3. Zenodo (2019). doi:[10.5281/zenodo.2604804](https://doi.org/10.5281/zenodo.2604804). <https://zenodo.org/record/2604804> Accessed 2021-10-06
77. Quevillon, E., Silventoinen, V., Pillai, S., Harte, N., Mulder, N., Apweiler, R., Lopez, R.: InterProScan: protein domains identifier. *Nucleic Acids Research* **33**(Web Server issue), 116–120 (2005). doi:[10.1093/nar/gki442](https://doi.org/10.1093/nar/gki442)
78. Käll, L., Krogh, A., Sonnhammer, E.L.L.: Advantages of combined transmembrane topology and signal peptide prediction—the Phobius web server. *Nucleic Acids Research* **35**(Web Server issue), 429–432 (2007). doi:[10.1093/nar/gkm256](https://doi.org/10.1093/nar/gkm256)
79. Cantalapiedra, C.P., Hernández-Plaza, A., Letunic, I., Bork, P., Huerta-Cepas, J.: eggNOG-mapper v2: Functional Annotation, Orthology Assignments, and Domain Prediction at the Metagenomic Scale. *Molecular Biology and Evolution*, 293 (2021). doi:[10.1093/molbev/msab293](https://doi.org/10.1093/molbev/msab293)
80. Lagesen, K., Hallin, P., Rødland, E.A., Staerfeldt, H.-H., Rognes, T., Ussery, D.W.: RNAmmer: consistent and rapid annotation of ribosomal RNA genes. *Nucleic Acids Research* **35**(9), 3100–3108 (2007). doi:[10.1093/nar/gkm160](https://doi.org/10.1093/nar/gkm160)
81. Tice, A.K., Žihala, D., Pánek, T., Jones, R.E., Salomaki, E.D., Nenarokov, S., Burki, F., Eliáš, M., Eme, L., Roger, A.J., Rokas, A., Shen, X.-X., Strasser, J.F.H., Kolisko, M., Brown, M.W.: PhyloFisher: A phylogenomic package for resolving eukaryotic relationships. *PLOS Biology* **19**(8), 3001365 (2021). doi:[10.1371/journal.pbio.3001365](https://doi.org/10.1371/journal.pbio.3001365). Publisher: Public Library of Science. Accessed 2021-10-06
82. Alexa, A., Rahnenfuhrer, J.: topGO: Enrichment Analysis for Gene Ontology. Bioconductor version: Release (3.13) (2021). doi:[10.18129/B9.bioc.topGO](https://doi.org/10.18129/B9.bioc.topGO). <https://bioconductor.org/packages/topGO/> Accessed 2021-10-06
83. Katoh, K., Standley, D.M.: MAFFT Multiple Sequence Alignment Software Version 7: Improvements in Performance and Usability. *Molecular Biology and Evolution* **30**(4), 772–780 (2013). doi:[10.1093/molbev/mst010](https://doi.org/10.1093/molbev/mst010). Accessed 2021-10-06
84. Waterhouse, A.M., Procter, J.B., Martin, D.M.A., Clamp, M., Barton, G.J.: Jalview Version 2—a multiple sequence alignment editor and analysis workbench. *Bioinformatics* **25**(9), 1189–1191 (2009). doi:[10.1093/bioinformatics/btp033](https://doi.org/10.1093/bioinformatics/btp033). Accessed 2021-10-06
85. Matthey-Doret, C., baudry, axelcournac, Amaury, Remi-Montagne, Guiguelmoni, N., Foutel-Rodier, T., Scolari, V.F.: koszullab/hicstuff: Standardized help messages. Zenodo (2021). doi:[10.5281/zenodo.4722873](https://doi.org/10.5281/zenodo.4722873). <https://zenodo.org/record/4722873> Accessed 2021-10-06
86. Imakaev, M., Fudenberg, G., McCord, R.P., Naumova, N., Goloborodko, A., Lajoie, B.R., Dekker, J., Mirny, L.A.: Iterative correction of Hi-C data reveals hallmarks of chromosome organization. *Nature Methods* **9**(10), 999–1003 (2012). doi:[10.1038/nmeth.2148](https://doi.org/10.1038/nmeth.2148). Number: 10 Publisher: Nature Publishing Group. Accessed 2021-04-27
87. Matthey-Doret, C.: koszullab/pareidolia: v0.6.1 (2021). doi:[10.5281/zenodo.5062485](https://doi.org/10.5281/zenodo.5062485). <https://zenodo.org/record/5062485> Accessed 2021-07-02
88. Yang, T., Zhang, F., Yardimci, G.G., Song, F., Hardison, R.C., Stafford, W., Yue, F., Li, Q.: HiCRep: assessing the reproducibility of Hi-C data using a stratum-adjusted correlation coefficient, 37

Figures







**Figure 4** Changes in trans-chromosomal contacts between *A. castellanii* chromosomes following *L. pneumophila* infection. **a**, Average contact change during infection between each pair of chromosomes. Chromosome lengths are shown below the interaction matrix, with the chromosome bearing 18S and 28S rDNA highlighted in green **b**, Representative inter-telomeric contacts between a pair of chromosomes (Neff scaffolds 3 and 11). The average inter-telomeric contact profile generated from all pairs of chromosomes is shown as a pileup. The Log ratio between the infected (I) and uninfected (U) profiles is shown as a ratio (right).



# Tables

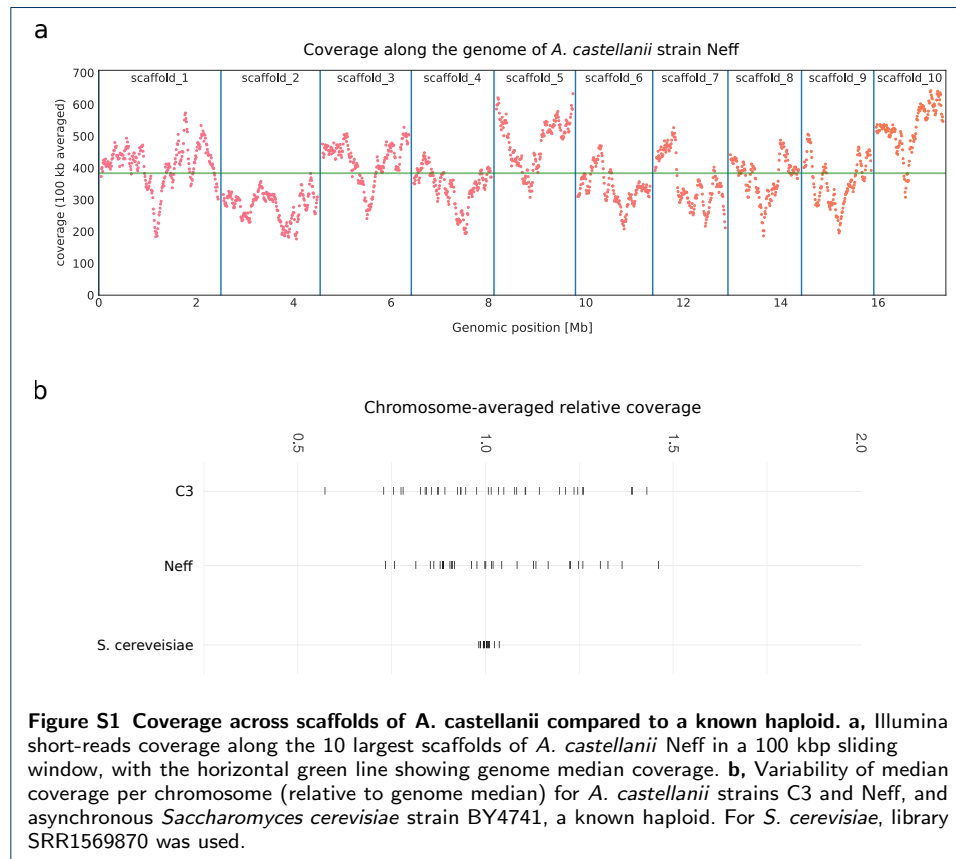
Assembly	Neff-v1	Neff	C3
Genome size (Mbp)	42.0	43.8	46.1
# scaffolds	384	111	174
# of Ns (Mbp)	2.6 (6.1%)	0 (0%)	0 (0%)
N50 (Mbp)	0.3	1.3	1.4
Largest scaffold (Mbp)	2.0	2.5	2.4
GC%	57.90	58.44	58.64
# protein coding genes	14,974	15,497	16,837

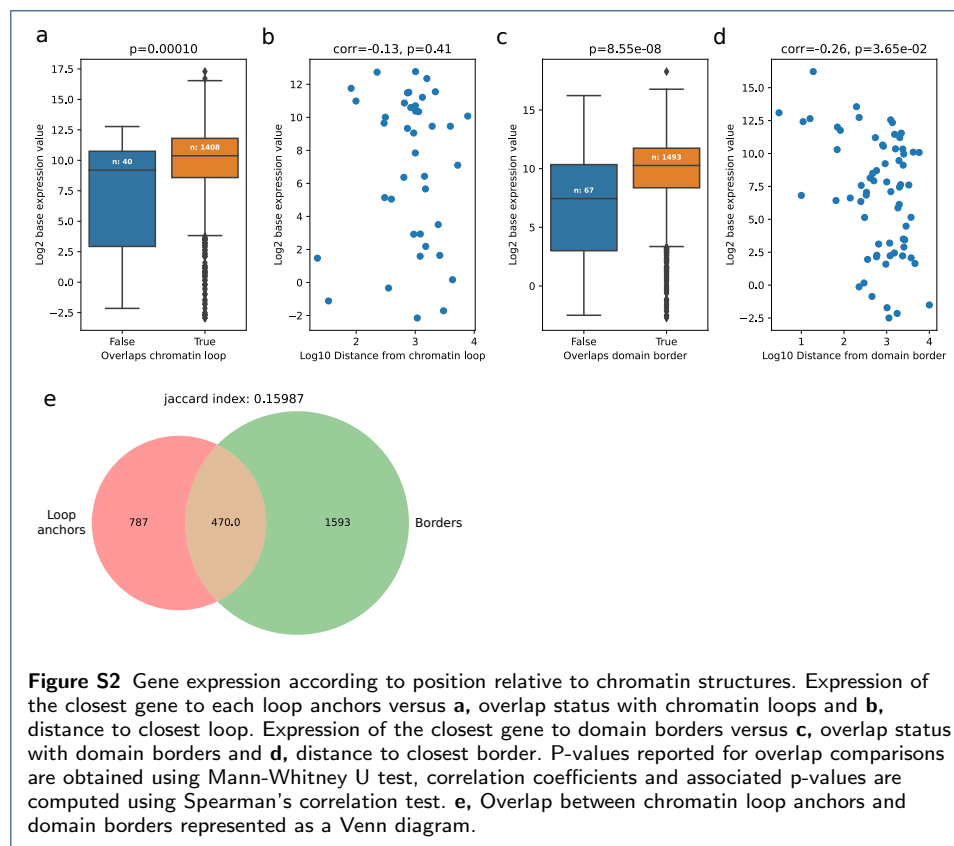
**Table 1** Genome statistics for the finished assemblies of Neff, C3 (this study) and the reference Neff-v1 genome.

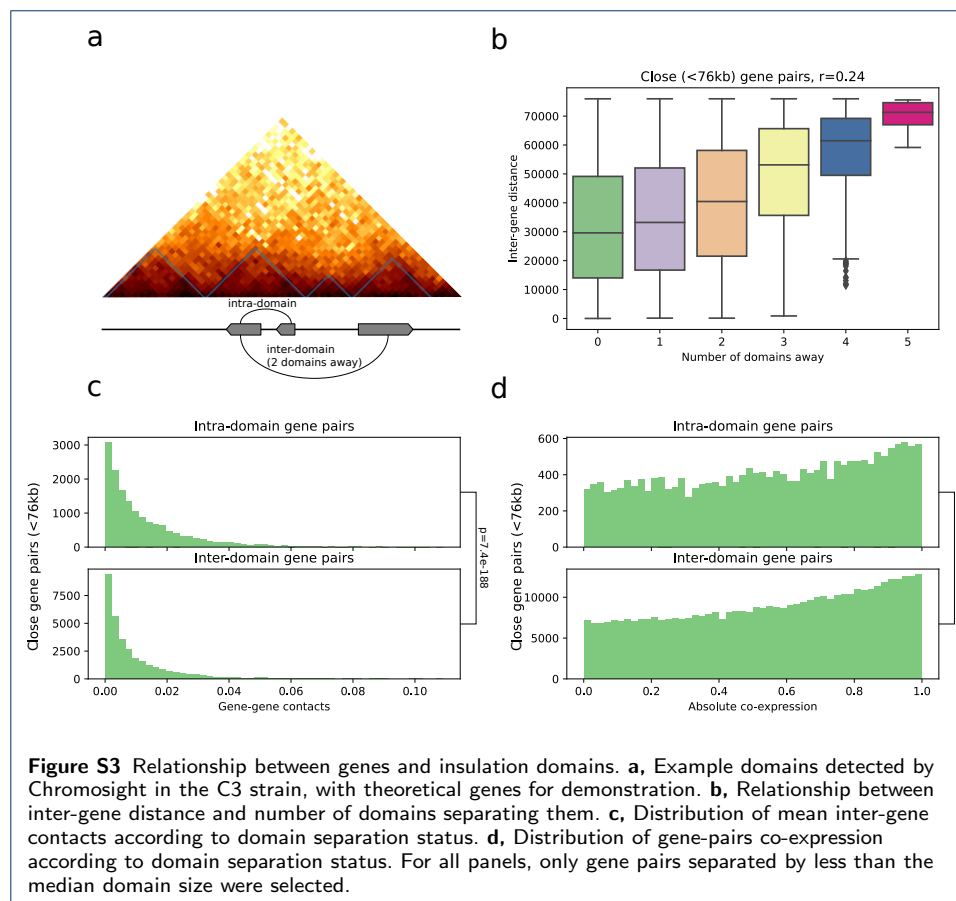
Strain	Identity	Gaps
Neff	757/826 (91.6%)	1/826 (0.12%)
C3	821/825 (99.5%)	0
<i>A. polyphaga</i>	802/825 (97.5%)	0

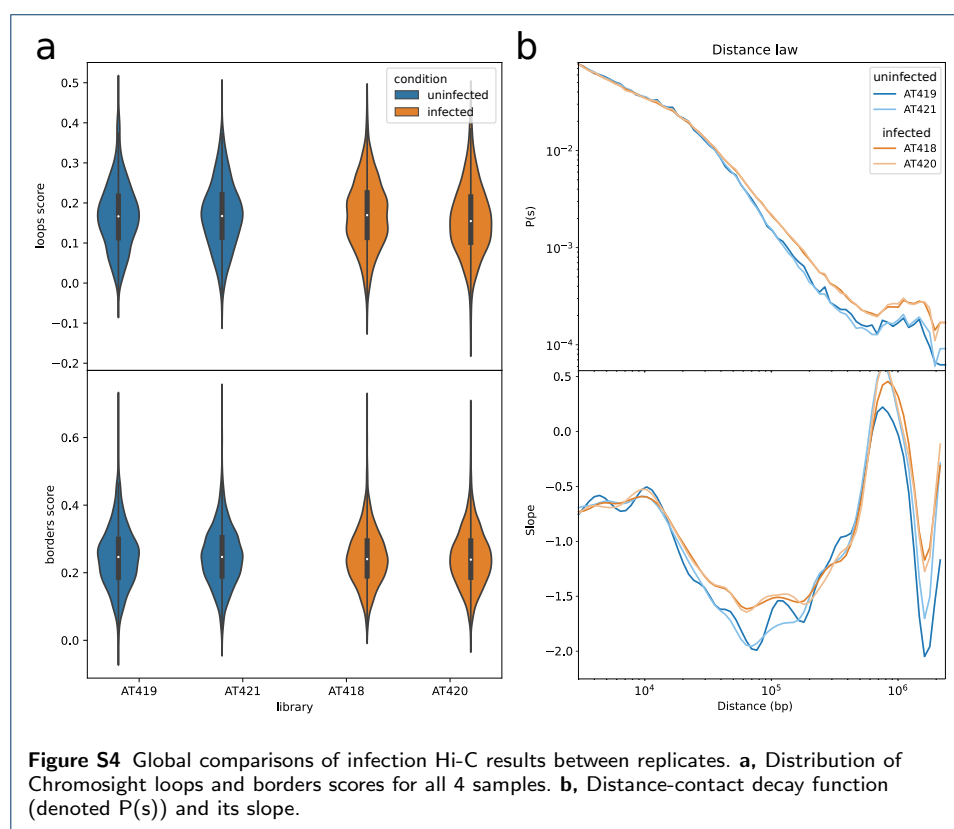
**Table 2** Identity of mannose binding proteins from *A. polyphaga* and *A. castellanii* strains Neff and C3 to their homolog in *A. castellanii* strain MEEI 0184 across 788 sites of a 834-site amino acid alignment. The first 46 sites of the alignment were excluded from the calculation because the 5' end of the gene in *A. polyphaga* was missing due to a truncated contig.

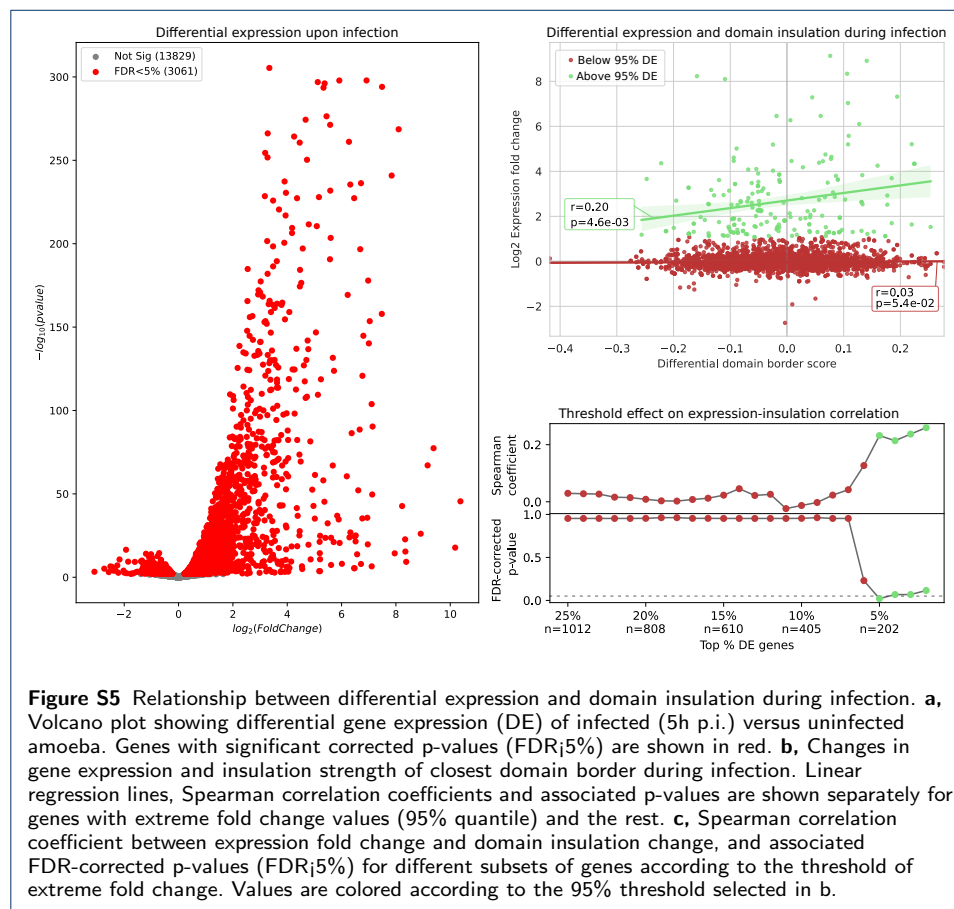
# Supplementary figures



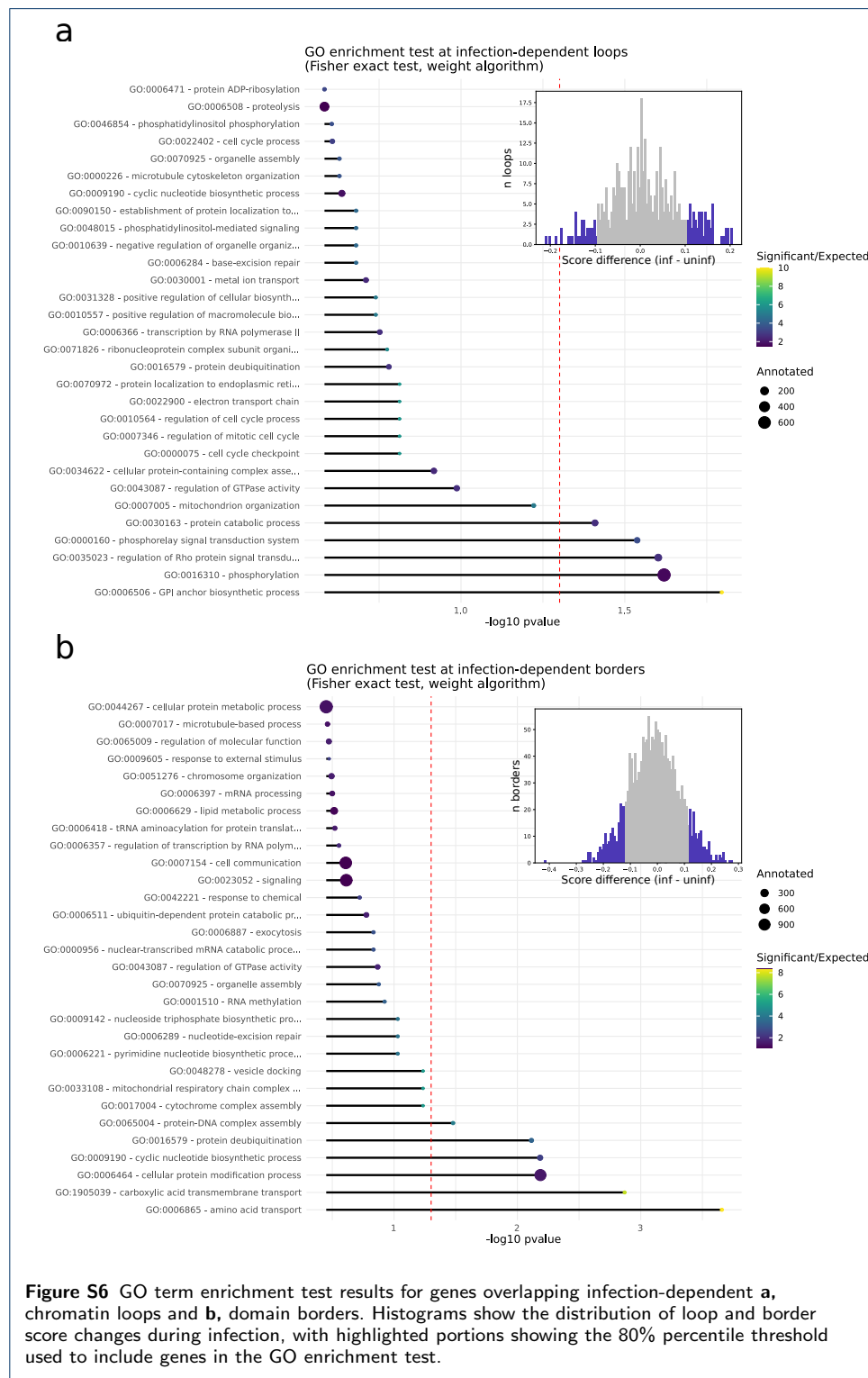


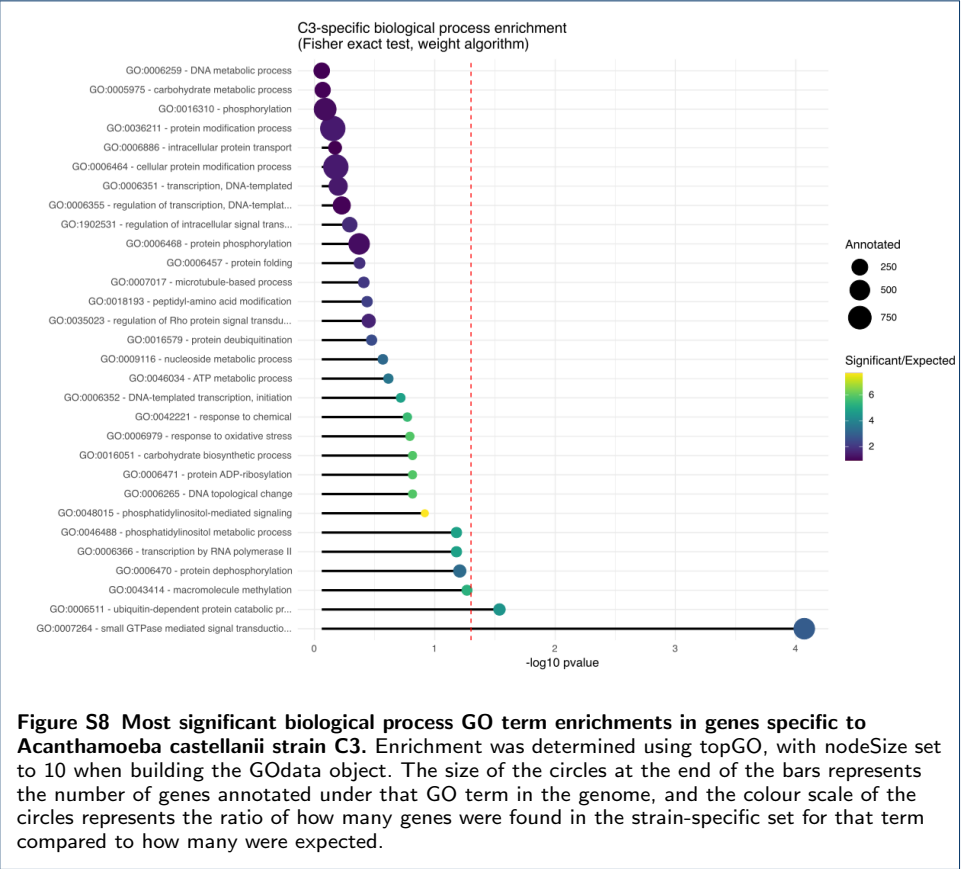
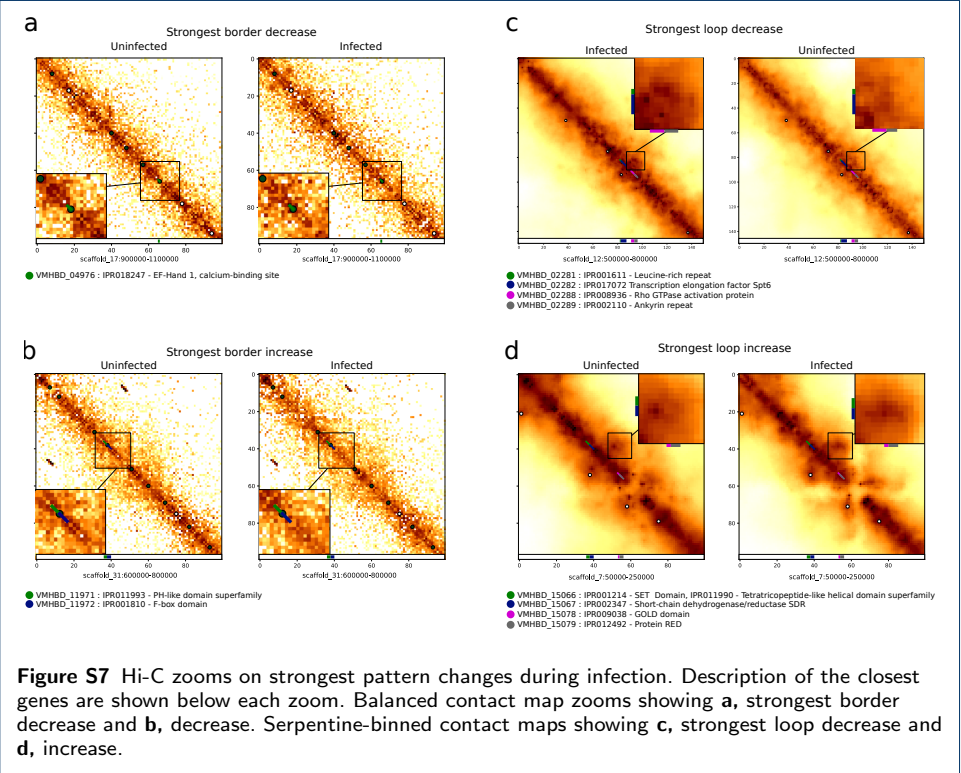


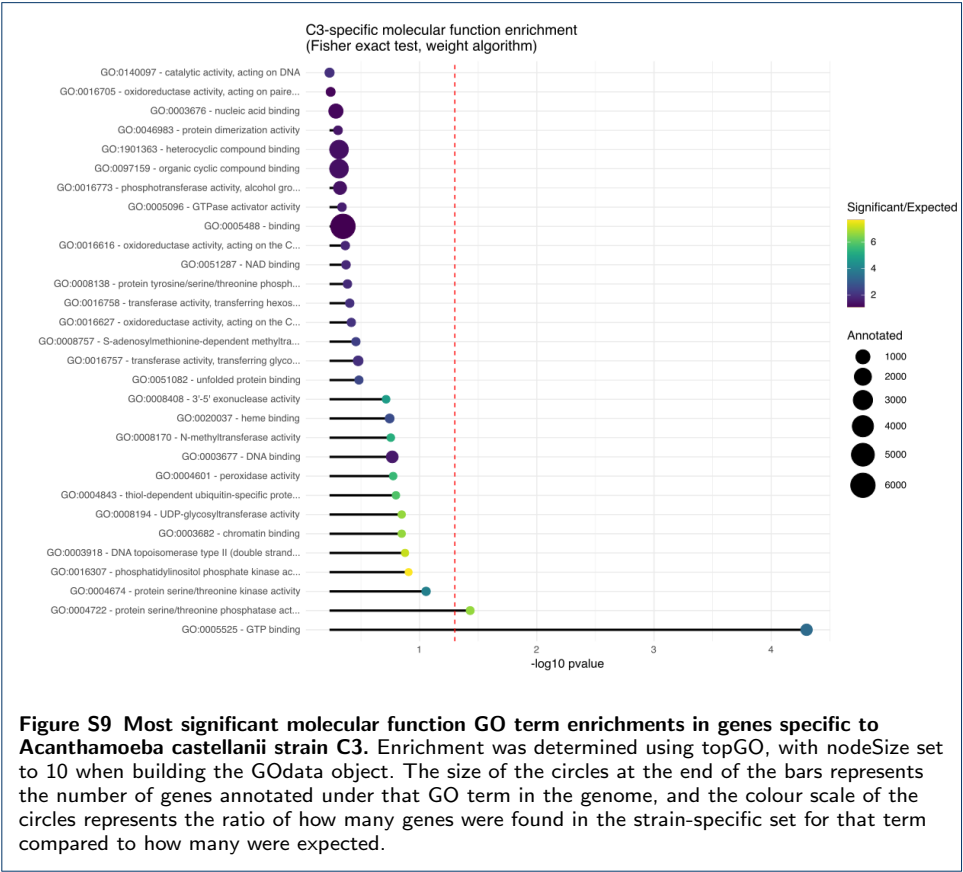


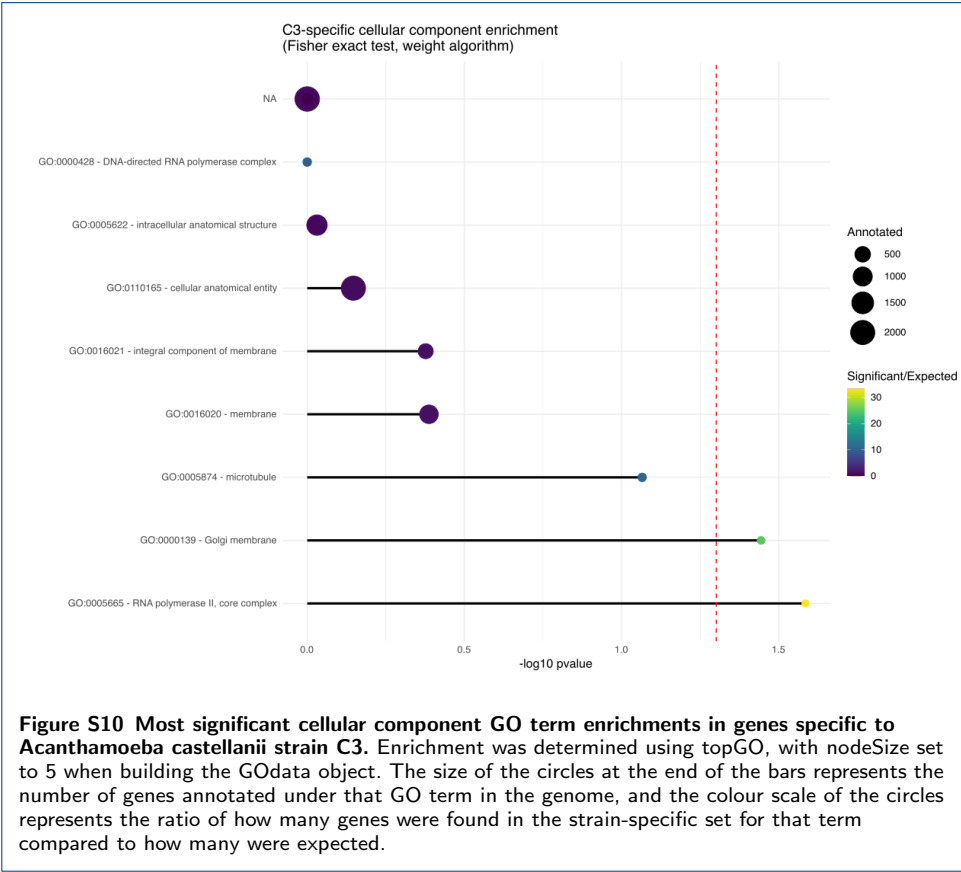


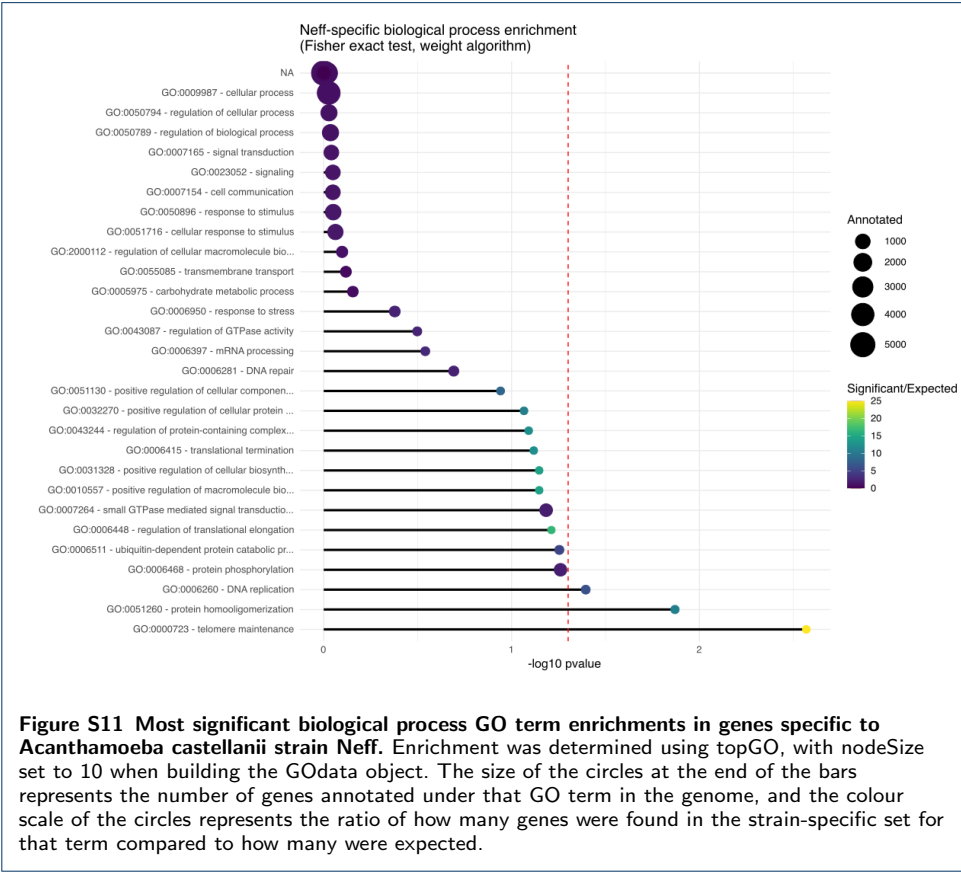












**Figure S11 Most significant biological process GO term enrichments in genes specific to *Acanthamoeba castellanii* strain Neff.** Enrichment was determined using topGO, with nodeSize set to 10 when building the GOdata object. The size of the circles at the end of the bars represents the number of genes annotated under that GO term in the genome, and the colour scale of the circles represents the ratio of how many genes were found in the strain-specific set for that term compared to how many were expected.

