

Title: Structural insights into hormone recognition by the human glucose-dependent insulinotropic polypeptide receptor

Authors

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Abstract

Glucose-dependent insulinotropic polypeptide (GIP) is a peptide hormone that exerts crucial metabolic functions by binding and activating its cognate receptor, GIPR. As an important therapeutic target, GIPR has been subjected to intensive structural studies without success. Here, we report the cryo-EM structure of the human GIPR in complex with GIP and a G_s heterotrimer at a global resolution of 2.9 Å. GIP adopts a single straight helix with its N terminus dipped into the receptor transmembrane domain (TMD), while the C-terminus is closely associated with the extracellular domain and extracellular loop 1. GIPR employs conserved residues in the lower half of the TMD pocket to recognize the common segments shared by GIP homologous peptides, while uses non-conserved residues in the upper half of the TMD pocket to interact with residues specific for GIP. These results provide a structural framework of hormone recognition and GIPR activation.

MAIN TEXT

Introduction

Glucose-dependent insulinotropic polypeptide (GIP) is a 42-amino acid peptide hormone that plays crucial role in glucose regulation and fatty acid metabolism. In response to food intake, GIP is secreted by intestinal K cells to enhance insulin secretion and peripheral fatty acid uptake (1), as well as a number of neuronal effects (2). The pleiotropic functions of GIP is mediated by its cognate receptor (GIPR), a member of class B1 G protein-coupled receptors (GPCRs) that also include glucagon receptor (GCGR) and glucagon-like peptide-1 (GLP-1R). GIPR, together with GCGR and GLP-1R, form the central endocrine network in regulating insulin sensitivity and energy homeostasis, and they are validated drug targets (3-5). Intensive efforts were made in drug discovery targeting these receptors (6). A number of GLP-1R selective ligands have been developed successfully to treat type 2 diabetes and obesity. Encouragingly, peptide ligands that bind both

GIPR and GLP-1R show better clinical efficacy than the GLP-1R agonist alone. As such, GIPR has emerged as a hot target pursued by pharmaceutical research community.

GIPR contains a large extracellular domain (ECD) and a 7-transmembrane domain (TMD). Both are involved in ligand recognition and receptor activation (7-9). Cryo-electron microscopy (cryo-EM) structures of GCGR and GLP-1R, as well as several other class B1 GPCRs have been solved, providing a general mechanism of two-domain model for peptide recognition and receptor activation. However, GIP displays an exquisite sequence specificity towards GIPR as it does not bind to other class B1 GPCRs. However, the efforts to understand the ligand selectivity by GIPR have been hampered by technical difficulties in expression and stabilization of the liganded GIPR complexes for structural studies. We have overcome such challenges and determined a high-resolution (2.9 Å) structure of the human GIPR in complex with the stimulatory G protein (G_s) using single-particle cryo-EM approach in conjunction with NanoBiT strategy (10). Together with functional studies, our results demonstrate several unique structural features that distinguish GIPR from other members of the glucagon subfamily of class B1 GPCRs and provide an important template for rational design of GIPR agonists for therapeutic development.

Results

Structure determination

To prepare a high quality human GIPR- G_s complex, we overcame several technical obstacles to enhance the expression level and protein stability by adding a double tag of maltose binding protein at the C terminus and a BRIL fusion protein at the N terminus (Fig. S1A), as well as employing the NanoBiT tethering strategy (10-12) (Fig. S1A, B). To solve the GIP₁₋₄₂-GIPR- G_s structure, we further introduced one mutation (T345F) to stabilize the assembly of complex (Fig. S1C, D). This mutation does not affect the ligand binding or potency of GIP₁₋₄₂ in cAMP accumulation assay (Fig. S1G, H). Large-scale purification was followed and the GIP₁₋₄₂-GIPR- G_s complexes were collected by size-exclusion chromatography (SEC) for cryo-EM studies (Fig. S1E, F). The activity of the modified GIPR construct was confirmed by cAMP accumulation assay showing a similar response as that of the wild-type (WT; Fig. S1G).

The GIP₁₋₄₂-GIPR- G_s complexes were imaged using a Titan Krios equipped with a Gatan K3 Summit direct electron detector (Fig. S2). 2D classification showed a clear secondary structure feature and random distribution of the particles. Different directions of the particles enabled a high-resolution cryo-EM map reconstruction (Fig. S2B). A total of 295,021 particles were selected after 3D refinement and polishing, leading to an overall resolution of 2.9 Å (Fig. S2C, D).

Overall structure

Apart from the α -helical domain (AHD) of $G\alpha_s$ which is flexible in most cryo-EM GPCR-G protein complex structures, the bound GIP₁₋₄₂, GIPR and G_s were well defined in the EM density maps (Fig. 1, Fig. S3). Except for the ECD, side-chains of the majority of amino acid residues are well resolved in all protein components. The final model contains 30 GIP₁₋₄₂ residues, the $G\alpha\beta\gamma$ subunits of G_s , and the GIPR residues from Q30^{ECD} to S415^{8.66b} (class B GPCR numbering in superscript) (13), with six amino acid residues missing at helix 8. Owing to the high resolution map, notable conformation difference from GCGR (14) or GLP-1R (15) was observed in the ECL1.

Similar to other class B1 GPCR- G_s complexes, the TM6 of GIPR shows a sharp kink in the middle and TM7 displays an outward movement. Like PTH1R- G_s and CRF1R- G_s cryo-EM structures (16, 17), the TMD of GIPR is surrounded by annular detergent micelle, with a diameter of 12 nm thereby mimicking the lipid bilayer morphology (Fig. 1). In addition, we also observed several cholesterol molecules in the cryo-EM map.

97 Ligand recognition

98 In the complex, GIP adopts a single continuous helix that penetrates into the TMD core through its
 99 N-terminal half (residues 1 to 15), while the C-terminal half (residues 16 to 30) is recognized by
 100 the ECD and ECL1 (Fig. 2A-C). Y1^P (P indicates that the residue belongs to the peptide ligand) of
 101 GIP points to TMs 2-3, forms hydrogen bonds with R190^{2.67b} and Q224^{3.37b}, and makes hydrophobic
 102 contacts with V227^{3.40b} and W296^{5.36b}. This observation received support of the mutagenesis study,
 103 where mutant W296A decreased GIP potency by 50-fold (Fig. 2D), and mutants R190A and Q224A
 104 diminished the potency of GIP by 71- and 5-fold, respectively, as reported in a previous report (18).
 105 N-terminal truncation of either Y1^P or both Y1^P and A2^P led to reduced low efficacy or loss of
 106 activity (19, 20), highlighting a crucial role of Y1^P. E3^P, D9^P and D15^P are three negatively charged
 107 residues in the N-terminal half of GIP and form salt bridges with R183^{2.60b}, R370^{7.35b} and R289^{ECL2},
 108 respectively. Removal of these salt bridges by alanine substitution at either R183^{2.60b} (18) or
 109 R370^{7.35b} (Fig. 2D) greatly reduced GIP potency (by 76- and 55-fold, respectively), whereas the
 110 effect on mutant R289A was mild (6-fold, Fig. 2D). Polar interactions also occurred between S8^P
 111 and N290^{ECL2} as well as Y10^P and Q138^{1.40b}. The GIP-TMD interface was further stabilized by a
 112 complementary nonpolar network involving TM1 (L134^{1.36b}, L137^{1.39b} and Y141^{1.43b}) and TM7
 113 (L374^{7.39b} and I378^{7.43b}) via A2^P, F6^P and Y10^P of GIP (Fig. 2C), in line with decreased ligand
 114 potencies observed in Y141A (by 103-fold), L374A (by 41-fold) and I378A (by 8-fold) mutants
 115 (Fig. 2D).

116 The C-terminal half of GIP was clasped by the GIPR ECD, closely resembling the crystal
 117 structure of GIP-GIPR ECD (PDB code: 2QKH) (21). Specifically, three adjacent amino acids
 118 (H18^P, Q19^P and Q20^P) form multiple hydrogen bonds with the side-chains of Q30 and Y36 at the
 119 N-terminal α -helix of ECD, as well as N120 and E122 at the hinge region between ECD and TM1.
 120 The hydrophobic residues of GIP (F22^P, V23^P, L26^P and L27^P) occupy a complementary binding
 121 groove of the GIPR ECD, consisting of a series of hydrophobic residues (L35, Y36, W39, M67,
 122 Y68, Y87, L88, P89 and W90). Alanine substitutions in W39, D66 and Y68 significantly reduced
 123 the potency of GIP (Fig. 2D). Notably, the cryo-EM map suggests that the ECL1 stands upwards
 124 to approach the N-terminal α -helix of ECD and forms hydrogen bonds with the side-chains of R43
 125 and Y36 (Fig. 2A, B), resulting in a close contact between TMD and ECD for GIP-bound GIPR
 126 (interface area = 571 Å²), significantly larger than that of GLP-1 bound GLP-1R (362 Å²),
 127 reinforcing the importance of ECD in GIP recognition.

128 Receptor activation

129 GIPR shares ~50% sequence similarity with GCGR, especially in the TMD region (75%), thus
 130 GCGR structures published previously provide a good template for the present study (Fig. S4) (14,
 131 22-25). It was found the TMD of activated GIPR exhibits a conformation similar to that of GCGR
 132 activated by glucagon or ZP3780 (C α RMSD = 1.2 and 0.7 Å, respectively) (14, 22) and distinct
 133 from that of GCGR bound by the negative allosteric modulator NNC0640 or partial agonist
 134 NNC1702 (C α RMSD = 4.0 and 3.9 Å, respectively) (26). Facilitated by Gly^{7.50b} located in the
 135 middle of TM7, the extracellular half of TM7 bends towards TM6 by 8.0 Å (measured by C α atom
 136 of Gly^{7.32b}) (Fig. S4). This feature and the outward movement of ECL3 expanded the ligand binding
 137 pocket. Meanwhile, the extracellular tip of TM1 was extended by one turn and moved inward by
 138 8.0 Å (measured by C α atom of the residues at 1.30b) (Fig. S4). Together with the raised ECL1,
 139 these conformational changes stabilized ligand binding.

140 In the intracellular side, the sharp kink in the middle of TM6 led to an outward movement of
 141 its intracellular portion measured by C α atom of R336^{6.35b} (18.9 Å, similar to that of other Gs-
 142 coupled class B1 receptors). This was accompanied by the movement of the intracellular tip of TM5
 143 towards TM6 by 7.6 Å (measured by C α atom of the residues at 5.67b), thereby creating an
 144 intracellular cavity for G protein coupling (Fig. S4).

145 G protein coupling

146 In our model, G_s protein is anchored by the $\alpha 5$ helix of G α_s (GaH5), thereby fitting to the
 147 cytoplasmic cavity formed by TMs 3, 5 and 6, intracellular loops (ICLs) 1-2 and H8 (Fig. 3). In
 148 general, the GIPR–G_s complex shows a similar receptor–G protein interface as other reported class
 149 B1 receptor structures such as GLP-1R (27), GLP-2R (12), GCGR (14), PTH1R (16), SCTR
 150 (secretin receptor) (28) and GHRHR (11), suggesting a common G protein signaling mechanism
 151 (Fig. 3A). The hydrophobic residues at the C-terminal of G α_s (L388^{GaH5}, Y391^{GaH5}, L393^{GaH5}
 152 and L394^{GaH5}) insert into a small hydrophobic pocket formed by Y240^{3.53b}, L241^{3.54b}, L244^{3.57b},
 153 L245^{3.58b}, I317^{5.58b}, I320^{5.60b}, L321^{5.61b} and L325^{5.65b} (Fig. 3B). The side-chain of R338^{6.37b} points
 154 to G α_s and makes one hydrogen bonds with L394^{GaH5}. Of note is the interaction between R380^{GaH5}
 155 and ICL2 resulting in five hydrogen bonds with the backbone atoms of L245^{3.58b}, V246^{3.59b},
 156 L247^{3.60b} and V248^{ICL2}, significantly more than that observed in GLP-1R, SCTR or GCGR (Fig.
 157 3C). The polar residues in ICL2 (S251^{ICL2} and E253^{ICL2}) produce two hydrogen bonds with K34
 158 and Q35 of G α_s , while H8 forms several hydrogen bonds with ICL1, then contacts with G β
 159 (E398^{8.49b}-R164^{ICL1}-D312^{G β} , E402^{8.53b}-R164^{ICL1}-D312^{G β}) (Fig. 3D). Together, these specific
 160 interactions contribute to the G_s coupling specificity of GIPR.

161 Ligand specificity

162 GIP, GLP-1 and glucagon are three important metabolic hormones exerting distinct functions in
 163 glucose homeostasis, in spite of high degrees of sequence similarity. Superimposing the TMD of
 164 GIP-bound GIPR with that of GLP-1-bound GLP-1R (27) or glucagon-bound GCGR (14) displays
 165 a similar ligand-binding pocket and the three peptides all adopt a single continuous helix, with the
 166 N-terminus penetrating to the TMD core to the same depth, while the C-terminus anchors the ECD
 167 and ECL1 in a receptor-specific manner (Fig. 4). Notably, the ECL1 of GIPR stands upwards in
 168 line with TMs 2 and 3, and moves towards the TMD core by 5~7 Å. Such a movement, together
 169 with a α -helical extension in TM1 by six residues, allows GIP to shift to TM1 by 2.7 and 3.3 Å
 170 (measured by C α atom of L27^P) relative to GLP-1 (27) and glucagon (14), respectively (Fig. S5).

171 Based on the sequence similarity, the three peptides can be divided into four segments: two
 172 common segments (residues 4-11 and 21 to 30 in GIP) and two unique segments (residues 1-3 and
 173 12-20 in GIP) (Fig. 4H). The N-terminus (residues 1-3) makes massive contacts with the conserved
 174 central polar network of class B1 GPCRs including one hydrogen bond with Q^{3.37b} stabilized by the
 175 hydrophobic residue at 3.40b; one hydrogen bond with Y^{1.47b} made by the third peptide residue (Fig.
 176 4B, H); residues 4-11 interact with salt bridges of R^{7.35b}, pi-stacking of Y^{1.43b}, hydrophobic L^{2.71b},
 177 W^{5.36b} and L^{7.39b}, as well as several hydrogen bonds in ECL2 (Fig. 4C, H); residues 12-20 are
 178 divergent and mainly interact with ECLs 1-2 and TMs 1-2 (Fig. 4D, H).

179 To accommodate varying lengths of side-chains at A^{13P}/Y/Y, I^{17P}/Q/R, Q^{19P}/A/A and Q^{20P}/K/Q,
 180 both TM1 and ECL1 adjusted their conformations to avoid clashes (Fig. 4D, Fig. S5). For example,
 181 ECL1 of GLP-1R is more distant from GLP-1 than that of GIPR from GIP, whereas repulsion of
 182 the side-chain of R^{18P} was seen between GCGR and glucagon. Therefore, receptor-specific
 183 interaction may reside in this region which precludes the binding of GLP-1 or glucagon to GIPR
 184 revealed by MD simulations (Fig. S6). As far as C-terminus is concerned, all three peptides form
 185 extensive hydrophobic contacts with the ECD, resulting from the hydrophobic composition of
 186 amino acids in both sides (Fig. 4E-H). It appears that GIPR, GLP-1R and GCGR employ conserved
 187 residues to recognize the common segments of their endogenous peptides and use non-conserved
 188 residues to make specific interaction that govern the ligand selectivity.

190 Discussion

As one of the incretin hormones, GIP modulates glucose metabolism by stimulating the β -cells to release insulin (29). Unlike GLP-1, it does not suppress gastric emptying and appetite, while exerting opposite actions on pancreatic α -cells as well as adipocytes leading to glucagon secretion and lipogenesis (29). Coupled with reduced sensitivity in type 2 diabetic patients, development of GIPR-based therapeutics met little success (30).

Comparison of the full-length structures of six glucagon subfamily of GPCRs demonstrates that bound peptides (GLP-1, exendin-P5, glucagon, ZP3780, secretin, GHRH, GLP-2, and GIP) all adopt a single straight helix with their N-terminus inserted into the TMD core, while the C-terminal is recognized by the ECD (11, 12, 14, 15, 28). For parathyroid hormone subfamily of GPCRs, the long-acting PTH analog (LA-PTH) predominantly exhibits an extended helix with its N-terminus inserted deeply into the TMD, where the peptide C-terminus may bend occasionally (16). In the case of corticotropin-releasing factor (CRF) subfamily of GPCRs, the N-terminus (first seven residues) of urocortin 1 (UCN1) and CRF1 present an extended loop conformation, and its C-terminal residues (8–40) adopt a single extended helix (17, 31). As far as calcitonin subfamily of GPCRs is concerned, calcitonin gene-related peptide (CGRP) has an unstructured loops in both N- and C-terminal regions (32). Looking at pituitary adenylate-cyclase-activating peptide (PACAP) and vasoactive intestinal polypeptide (VIP) receptor subfamily, PACAP displays an extended α -helix, while maxadilan, a natural PAC1R agonist (61-amino acid long), forms the N- and C-terminal helices that are linked as a loop (10, 31, 33). These observations highlight diversified peptide binding modes among class B1 GPCRs (Fig. S7).

Species differences in class B1 receptor responsiveness are diversified and receptor-specific, which is tolerable for some receptors such as GLP-1R and GCGR, but leads to concerns for others like GIPR and PTH2R (34, 35). Interestingly, the sequence identities between human and mouse at both ligand and receptor levels are more conserved between GLP and GLP-1R (100% and 93%) than that between GIP and GIPR (92% and 81%)(34). Such a divergence is not caused by changes in peptide potency, but resides in the biological property of either GIP or the receptor (34, 36). However, it may affect GIP-related pharmacology markedly (34). Indeed, a previous study found that human GIP is a comparatively weak partial agonist in rodent models (34). Human (Pro3)GIP is a full agonist with identical maximum response as human GIP, whereas both rat and mouse (Pro3)GIPs are partial agonists (34, 36). Of note is that among rat, mouse and human GIPs, the only residue change (from His to Arg) occurs at the 18th position (34). From a structural biology perspective, the variation in the sequences of both GIP (H^{18P}/R/R for human, rat and mouse) and GIPR, and the consequent alterations in either peptide-binding or G protein-coupling may offer an explanation. Nonetheless, it may also complicate knowledge transfer from rodents to humans for clinical development of GIPR-based therapeutics.

The interactions between three receptors (GIPR, GLP-1R and GCGR) and their endogenous peptides transduce precise cellular signals responsible for glucose control. This is instituted by a common and closely related mechanism where the extracellular portion of the receptor mainly binds to a cognate ligand while the TMD activates a cascade of signaling events. The upper half of the TMD pocket composed of the top parts of ECL1, TM1 and TM2 interacts with unique residues in the peptide through flexible movement of ECL1 and complementary shape formation by TM1 and TM2, thereby conferring selectively and discriminating unrelated ligands. The lower half of the TMD pocket composed of TMs 3, 6 and 7 displays conserved sequences for recognition of common residues in the peptide. Its key function is to converge external signal into the cytoplasm and executes transduction with high efficiency. This mechanistic design reflects evolutionary advantages because multiple polypeptides could be accurately recognized via different sequences in the upper half of the TMD pocket.

Finally, GIPR, combined with GLP-1R and GCGR, have been intensively studied as targets of dual- or tri- agonists (37, 38). Combined activation of GLP-1R and GIPR by dual agonists would

provide synergistic and improved effects in glycemic and body weight control (39). The GLP-1R/GIPR dual-agonists LY3298176 (developed by Eli Lilly) and NN9709 (developed by Novo Nordisk/Marcadia) as well as GLP-1R/GCGR/GIPR tri-agonist HM15211 (developed by Hamni Pharmaceuticals) are undergoing phase II or III clinical trials (6). The detailed structural information on GIPR reported here will certainly be of value to better understand the mode of actions of these therapeutic peptides.

Materials and Methods

Cell culture

Spodoptera frugiperda (Sf9) (Invitrogen) and High-Five™ insect cells (ThermoFisher Scientific) were cultured in ESF 921 serum-free medium (Expression Systems) at 27 °C and 120 rpm.

Constructs

The human GIPR DNA (Genewiz) with one mutation (T345F) was cloned into a modified pFastBac vector (Invitrogen). The native signal peptide was replaced by the haemagglutinin signal peptide (HA) to enhance receptor expression. A BRIL fusion protein was added at the N terminal of the ECD with a TEV protease site and 2GSA linker between them. Forty-five amino acids (Q422-C466) were truncated at the C terminus where LgBiT was added with a 15-amino acid (15AA) polypeptide linker in between, followed by a TEV protease cleavage site and an optimized maltose binding protein- maltose binding protein tag (OMBP-MBP). A dominant-negative bovine Gαs (DNGαs)(S54N, G226A, E268A, N271K, K274D, R280K, T284D and I285T) construct was used to stabilize the complex (11, 40). SmBiT34 (peptide 86, Promega) subunit was added to the C terminus of Rat Gβ1 with a 15AA polypeptide linker between them. The modified rat Gβ1 and bovine Gγ2 were both cloned into a pFastBac vector.

Protein expression

Baculoviruses containing the above complex construct were prepared by the Bac-to-Bac system (Invitrogen). GIPR and Gs heterotrimer were co-expressed in High-Five™ cells. Briefly, insect cells were grown in ESF 921 culture medium (Expression Systems) to a density of 3.2×10^6 cells/mL, and then cells were infected with four kinds of viral preparations: BRIL-TEV-2GSA-GIPR(22-421)T345F-15AA-LgBiT-TEV-OMBP-MBP, Gαs, Gβ1-peptide 86, and Gγ2 at a ratio of 1:3:3:3. After 48 h incubation at 27 °C, the cells were collected by centrifugation and stored at -80 °C until use.

Nb35 expression and purification

Nanobody-35 (Nb35) with a 6× his tag at the C terminal was expressed in the periplasm of *E. coli* BL21 (DE3) cells. Briefly, Nb35 target gene was transformed in the bacterium and amplified in TB culture medium with 100 µg/mL ampicillin, 2 mM MgCl₂, 0.1 % (w/v) glucose at 37 °C, 180 rpm. When OD₆₀₀ reached 0.7-1.2, 1 mM IPTG was added to induce expression followed by overnight incubation at 28 °C. The cell pellet was then collected at 3000 rpm under 4 °C and stored at -80 °C. Nb35 was purified as by size-exclusion chromatography using a HiLoad 16/600 Superdex 75 column (GE Healthcare) with running buffer containing 20 mM HEPES, 100 mM NaCl, pH 7.4. Fractions of Nb35 were concentrated to ~3 mg/mL and quickly frozen in the liquid nitrogen with 10% glycerol and stored in -80 °C.

Complex formation and purification

Cell pellets were lysed in a buffer consisting of 20 mM HEPES, 100 mM NaCl, pH 7.4, 10 mM MgCl₂, 1 mM MnCl₂ and 10% glycerol supplemented with protease inhibitor cocktail, EDTA-free (TragetMol). Subsequently, cell membranes were collected by ultracentrifugation at 4 °C, 90,000 g for 35 min. The membranes were resuspended with a buffer containing 20 mM HEPES, 100 mM

NaCl, pH 7.4, 10 mM MgCl₂, 1 mM MnCl₂ and 10% glycerol. The complex of GIPR-G_s was assembled by adding 15 μM GIP₁₋₄₂ (GenScript), 100 μM TCEP, 25 mU/mL Apyrase (Sigma-Aldrich), 15 μg/mL Nb35 and 100 U salt active nuclease (Sigma-Aldrich) supplemented with protease inhibitor cocktail for 1.5 h incubation at room temperature (RT). The preparation was then solubilized with 0.5% (w/v) lauryl maltose neopentylglycol (LMNG, Anatrace) and 0.1% (w/v) cholesterol hemisuccinate (CHS, Anatrace) with additional 1 μM GIP₁₋₄₂ for 3 h at 4 °C. The supernatant was isolated by centrifugation at 90,000 g for 35 min and the solubilized complex was incubated with amylose resin (NEB) for 2.5 h at 4 °C. After batch binding, the resin was collected by centrifugation at 550 g and loaded onto a gravity flow column. The resin in column was firstly washed with 5 column volumes of buffer containing 20 mM HEPES, pH 7.4, 100 mM NaCl, 10% (v/v) glycerol, 5 mM MgCl₂, 1 mM MnCl₂, 25 μM TCEP, 3 μM GIP₁₋₄₂, 0.1% (w/v) LMNG and 0.02% (w/v) CHS. Subsequently, the resin was washed with 25 column volumes of buffer containing 20 mM HEPES, pH 7.4, 100 mM NaCl, 10% (v/v) glycerol, 5 mM MgCl₂, 1 mM MnCl₂, 25 μM TCEP, 3 μM GIP₁₋₄₂, 0.03% (w/v) LMNG, 0.01% (w/v) glyco-diosgenin (GDN, Anatrace) and 0.008% (w/v) CHS. The protein was then incubated with a buffer containing 20 mM HEPES, pH 7.4, 100 mM NaCl, 10% (v/v) glycerol, 5 mM MgCl₂, 1 mM MnCl₂, 25 μM TCEP, 50 μM GIP₁₋₄₂, 10 μg/mL Nb35, 0.03% (w/v) LMNG, 0.01% (w/v) glyco-diosgenin, 0.008% (w/v) CHS and 30 μg/mL His-tagged TEV protease on the column overnight at 4 °C. The flow through was collected and concentrated to 500 μL using a 100 kDa filter (Merck Millipore). Size-exclusion chromatography was performed by loading the protein onto Superose 6 Increase 10/300GL (GE Healthcare) column with running buffer containing 20 mM HEPES, pH 7.4, 100 mM NaCl, 10 mM MgCl₂, 100 μM TCEP, 5 μM GIP₁₋₄₂, 0.00075% (w/v) LMNG, 0.00025% (w/v) glyco-diosgenin, 0.0002% (w/v) CHS and 0.00025% digitonin (Anatrace). Monomeric GIPR-G_s complexes were collected and concentrated for cryo-EM analysis.

Data acquisition and image processing

The purified GIP₁₋₄₂-GIPR-G_s-Nb35 complex at a concentration of 6-7 mg/mL was mixed with 100 μM GIP₁₋₄₂ at 4 °C and applied to glow-discharged holey carbon grids (Quantifoil R1.2/1.3, Au 300 mesh) that were subsequently vitrified by plunging into liquid ethane using a Vitrobot Mark IV (ThermoFisher Scientific). A Titan Krios equipped with a Gatan K3 Summit direct electron detector was used to acquire Cryo-EM images. The microscope was operated at 300 kV accelerating voltage, at a nominal magnification of 46685× in counting mode, corresponding to a pixel size of 1.071 Å. Totally, 8023 movies were obtained with a defocus range of -1.2 to -2.2 μm. An accumulated dose of 80 electrons per Å² was fractionated into a movie stack of 36 frames.

Dose-fractionated image stacks were subjected to beam-induced motion correction using MotionCor2.1. A sum of all frames, filtered according to the exposure dose, in each image stack was used for further processing. Contrast transfer function parameters for each micrograph were determined by Gctf v1.06. Particle selection, 2D and 3D classifications were performed on a binned dataset with a pixel size of 2.142 Å using RELION-3.0-beta2. Auto-picking yielded 4,895,399 particle projections that were subjected to reference-free 2D classification to discard false positive particles or particles categorized in poorly defined classes, producing 2,754,623 particle projections for further processing. This subset of particle projections was subjected to a round of maximum-likelihood-based three dimensional classifications with a pixel size of 2.142 Å, resulting in one well-defined subset with 1,395,031 projections. Further 3D classifications with mask on the receptor produced one good subset accounting for 565,239 particles, which were subjected to another round of 3D classifications with mask on the ECD. A selected subset containing 295,021 projections was then subjected to 3D refinement and Bayesian polishing with a pixel size of 1.071 Å. After the last round of refinement, the final map has an indicated global resolution of 2.94 Å at a Fourier shell correlation (FSC) of 0.143. Local resolution was determined using the Bsoft package with half maps as input maps.

Model building and refinement

The cryo-EM structure of GCGR-G_s-Nb35 complex (PDB code 6WPW)(14) and the crystal structure (PDB code 2QKH) (21) were used as the start for model building and refinement against the EM map. The model was docked into the EM density map using Chimera (41), followed by iterative manual adjustment and rebuilding in COOT (42). Real space refinement was performed using Phenix (43). The model statistics were validated using MolProbity (44). Structural figures were prepared in Chimera and PyMOL (<https://pymol.org/2/>). The final refinement statistics are provided in Table S1.

cAMP accumulation assay

GIP₁₋₄₂ stimulated cAMP accumulation was measured by a LANCE Ultra cAMP kit (PerkinElmer). Briefly, HEK 293T cells were cultured in DMEM (Gibco) supplemented with 10% (v/v) fetal bovine serum (FBS, Gibco) and 1% (v/v) sodium pyruvate (Gibco) at 37 °C, 5% CO₂. Cells were seeded onto 6-well cell culture plates and transiently transfected with different GIPR constructs using Lipofectamine 2000 transfection reagent (Invitrogen). All the mutant constructs were modified by single-point mutation in the setting of the WT construct (HA-Flag-3GSA-GIPR(22-466)). After 24 h culture, the transfected cells were seeded onto 384-well microtiter plates at a density of 3000 cells per well in HBSS supplemented with 5 mM HEPES, 0.1% (w/v) bovine serum albumin (BSA) and 0.5 mM 3-isobutyl-1-methylxanthine. The cells were stimulated with different concentrations of GIP₁₋₄₂ for 40 min at RT. Eu and Ulight were then diluted by cAMP detection buffer and added to the plates separately to terminate the reaction. Plates were incubated at RT for 40 min and the fluorescence intensity measured at 620 nm and 650 nm by an EnVision multilabel plate reader (PerkinElmer).

Whole cell binding assay

CHO-K1 cells were cultured in F12 medium with 10% FBS and seeded at a density of 30,000 cells/well in Isoplate-96 plates (PerkinElmer). The WT (HA-Flag-3GSA-GIPR(22-466)) or mutant GIPR were transiently transfected using Lipofectamine 2000 transfection reagent. The mutant construct was modified by single-point mutation in the setting of the WT construct. Twenty-four hours after transfection, cells were washed twice, and incubated with blocking buffer (F12 supplemented with 33 mM HEPES and 0.1% bovine serum albumin (BSA), pH 7.4) for 2 h at 37°C. For homogeneous binding, cells were incubated in binding buffer with a constant concentration of ¹²⁵I-GIP (40 pM, PerkinElmer) and increasing concentrations of unlabeled GIP₁₋₄₂ (3.57 pM to 1 μM) at RT for 3 h. Following incubation, cells were washed three times with ice-cold PBS and lysed by addition of 50 μL lysis buffer (PBS supplemented with 20 mM Tris-HCl, 1% Triton X-100, pH 7.4). Fifty μL of scintillation cocktail (OptiPhase SuperMix, PerkinElmer) was added and the plates were subsequently counted for radioactivity (counts per minute, CPM) in a scintillation counter (MicroBeta2 Plate Counter, PerkinElmer).

Receptor surface expression

Cell surface expression was determined by flow cytometry to the N-terminal Flag tag on the WT GIPR (HA-Flag-3GSA-GIPR(22-466)) and its mutants transiently expressed in HEK 293T cells. All the mutant constructs were modified by single-point mutation in the setting of the WT construct. Briefly, approximately 2 × 10⁵ cells were blocked with PBS containing 5% BSA (w/v) at RT for 15 min, and then incubated with 1:300 anti-Flag primary antibody (diluted with PBS containing 5% BSA, Sigma-Aldrich) at RT for 1 h. The cells were then washed three times with PBS containing 1% BSA (w/v) followed by 1 h incubation with 1:1000 anti-mouse Alexa Fluor 488 conjugated secondary antibody (diluted with PBS containing 5% BSA, Invitrogen) at RT in the dark. After washing three times, cells were re-suspended in 200 μL PBS containing 1% BSA for detection by NovoCyte (Agilent) utilizing laser excitation and emission wavelengths of 488 nm and 530 nm, respectively. For each sample, 20,000 cellular events were collected, and the total fluorescence

intensity of positive expression cell population was calculated. Data were normalized to the WT receptor.

Molecular dynamics simulations

Molecular dynamic simulations were performed by Gromacs 2018.5. The peptide–GIPR complexes were built based on the cryo-EM GIP–GIPR–G_s complex and prepared by the Protein Preparation Wizard (Schrodinger 2017-4) with the G protein and Nb35 nanobody removed. The receptor chain termini were capped with acetyl and methylamide, and the titratable residues were left in their dominant state at pH 7.0. The complexes were embedded in a bilayer composed of 200 POPC lipids and solvated with 0.15 M NaCl in explicitly TIP3P waters using CHARMM-GUI Membrane Builder (45). The CHARMM36-CAMP force field (46) was adopted for protein, peptides, lipids and salt ions. The Particle Mesh Ewald (PME) method was used to treat all electrostatic interactions beyond a cut-off of 10 Å and the bonds involving hydrogen atoms were constrained using LINCS algorithm (47). The complex system was firstly relaxed using the steepest descent energy minimization, followed by slow heating of the system to 310 K with restraints. The restraints were reduced gradually over 50 ns. Finally, restrain-free production run was carried out for each simulation, with a time step of 2 fs in the NPT ensemble at 310 K and 1 bar using the Nose-Hoover thermostat and the semi-isotropic Parrinello-Rahman barostat (48), respectively. The buried interface areas were calculated with FreeSASA (49) using the Sharke-Rupley algorithm with a probe radius of 1.2 Å. The last 700 ns trajectory of each simulation was used to root mean square fluctuation (RMSF) calculation.

Statistical analysis

All functional data were presented as means ± standard error of the mean (SEM). Statistical analysis was performed using GraphPad Prism 7 (GraphPad Software). Concentration-response curves were evaluated with a three-parameter logistic equation. The significance was determined with either two-tailed Student's *t*-test or one-way ANOVA. Significant difference is accepted at *P* < 0.05.

References

1. S. J. Kim, C. Nian, C. H. McIntosh, Activation of lipoprotein lipase by glucose-dependent insulinotropic polypeptide in adipocytes. A role for a protein kinase B, LKB1, and AMP-activated protein kinase cascade. *J Biol Chem* **282**, 8557-8567 (2007).
2. E. Faivre, C. Holscher, Neuroprotective effects of D-Ala(2)GIP on Alzheimer's disease biomarkers in an APP/PS1 mouse model. *Alzheimers Res Ther* **5**, 20 (2013).
3. M. C. Lagerstrom, H. B. Schioth, Structural diversity of G protein-coupled receptors and significance for drug discovery. *Nat Rev Drug Discov* **7**, 339-357 (2008).
4. B. Finan, C. Clemmensen, Z. Zhu, K. Stemmer, K. Gauthier, L. Muller, M. De Angelis, K. Moreth, F. Neff, D. Perez-Tilve, K. Fischer, D. Lutter, M. A. Sanchez-Garrido, P. Liu, J. Tuckermann, M. Malehmir, M. E. Healy, A. Weber, M. Heikenwalder, M. Jastroch, M. Kleinert, S. Jall, S. Brandt, F. Flamant, K. W. Schramm, H. Biebermann, Y. Doring, C. Weber, K. M. Habegger, M. Keuper, V. Gelfanov, F. Liu, J. Kohrle, J. Rozman, H. Fuchs, V. Gailus-Durner, M. Hrabe de Angelis, S. M. Hofmann, B. Yang, M. H. Tschop, R. DiMarchi, T. D. Muller, Chemical hybridization of glucagon and thyroid hormone optimizes therapeutic impact for metabolic disease. *Cell* **167**, 843-857 e814 (2016).
5. C. Longuet, E. M. Sinclair, A. Maida, L. L. Baggio, M. Maziarz, M. J. Charron, D. J. Drucker, The glucagon receptor is required for the adaptive metabolic response to fasting. *Cell Metab* **8**, 359-371 (2008).
6. D. Yang, Q. Zhou, V. Labroska, S. Qin, S. Darbalaei, Y. Wu, E. Yuliantie, L. Xie, H. Tao, J. Cheng, Q. Liu, S. Zhao, W. Shui, Y. Jiang, M. W. Wang, G protein-coupled receptors: structure- and function-based drug discovery. *Signal Transduct Target Ther* **6**, 7 (2021).
7. C. Parthier, S. Reedtz-Runge, R. Rudolph, M. T. Stubbs, Passing the baton in class B GPCRs:

peptide hormone activation via helix induction? *Trends Biochem Sci* **34**, 303-310 (2009).

8. C. M. Koth, J. M. Murray, S. Mukund, A. Madjidi, A. Minn, H. J. Clarke, T. Wong, V. Chiang, E. Luis, A. Estevez, J. Rondon, Y. Zhang, I. Hotzel, B. B. Allan, Molecular basis for negative regulation of the glucagon receptor. *Proc Natl Acad Sci U S A* **109**, 14393-14398 (2012).
9. L. Yang, D. Yang, C. de Graaf, A. Moeller, G. M. West, V. Dharmarajan, C. Wang, F. Y. Siu, G. Song, S. Reedtz-Runge, B. D. Pascal, B. Wu, C. S. Potter, H. Zhou, P. R. Griffin, B. Carragher, H. Yang, M. W. Wang, R. C. Stevens, H. Jiang, Conformational states of the full-length glucagon receptor. *Nat Commun* **6**, 7859 (2015).
10. J. Duan, D. D. Shen, X. E. Zhou, P. Bi, Q. F. Liu, Y. X. Tan, Y. W. Zhuang, H. B. Zhang, P. Y. Xu, S. J. Huang, S. S. Ma, X. H. He, K. Melcher, Y. Zhang, H. E. Xu, Y. Jiang, Cryo-EM structure of an activated VIP1 receptor-G protein complex revealed by a NanoBiT tethering strategy. *Nat Commun* **11**, 4121 (2020).
11. F. Zhou, H. Zhang, Z. Cong, L. H. Zhao, Q. Zhou, C. Mao, X. Cheng, D. D. Shen, X. Cai, C. Ma, Y. Wang, A. Dai, Y. Zhou, W. Sun, F. Zhao, S. Zhao, H. Jiang, Y. Jiang, D. Yang, H. Eric Xu, Y. Zhang, M. W. Wang, Structural basis for activation of the growth hormone-releasing hormone receptor. *Nat Commun* **11**, 5205 (2020).
12. W. Sun, L. N. Chen, Q. Zhou, L. H. Zhao, D. Yang, H. Zhang, Z. Cong, D. D. Shen, F. Zhao, F. Zhou, X. Cai, Y. Chen, Y. Zhou, S. Gadgaard, W. J. C. van der Velden, S. Zhao, Y. Jiang, M. M. Rosenkilde, H. E. Xu, Y. Zhang, M. W. Wang, A unique hormonal recognition feature of the human glucagon-like peptide-2 receptor. *Cell Res* **30**, 1098-1108 (2020).
13. D. Wootten, J. Simms, L. J. Miller, A. Christopoulos, P. M. Sexton, Polar transmembrane interactions drive formation of ligand-specific and signal pathway-biased family B G protein-coupled receptor conformations. *Proc Natl Acad Sci U S A* **110**, 5211-5216 (2013).
14. A. Qiao, S. Han, X. Li, Z. Li, P. Zhao, A. Dai, R. Chang, L. Tai, Q. Tan, X. Chu, L. Ma, T. S. Thorsen, S. Reedtz-Runge, D. Yang, M. W. Wang, P. M. Sexton, D. Wootten, F. Sun, Q. Zhao, B. Wu, Structural basis of Gs and Gi recognition by the human glucagon receptor. *Science* **367**, 1346-1352 (2020).
15. Y. Zhang, B. Sun, D. Feng, H. Hu, M. Chu, Q. Qu, J. T. Tarrasch, S. Li, T. Sun Kobilka, B. K. Kobilka, G. Skiniotis, Cryo-EM structure of the activated GLP-1 receptor in complex with a G protein. *Nature* **546**, 248-253 (2017).
16. L. H. Zhao, S. Ma, I. Sutkeviciute, D. D. Shen, X. E. Zhou, P. W. de Waal, C. Y. Li, Y. Kang, L. J. Clark, F. G. Jean-Alphonse, A. D. White, D. Yang, A. Dai, X. Cai, J. Chen, C. Li, Y. Jiang, T. Watanabe, T. J. Gardella, K. Melcher, M. W. Wang, J. P. Vilardaga, H. E. Xu, Y. Zhang, Structure and dynamics of the active human parathyroid hormone receptor-1. *Science* **364**, 148-153 (2019).
17. S. Ma, Q. Shen, L. H. Zhao, C. Mao, X. E. Zhou, D. D. Shen, P. W. de Waal, P. Bi, C. Li, Y. Jiang, M. W. Wang, P. M. Sexton, D. Wootten, K. Melcher, Y. Zhang, H. E. Xu, Molecular basis for hormone recognition and activation of corticotropin-releasing factor receptors. *Mol Cell* **77**, 669-680 e664 (2020).
18. T. Yaqub, I. G. Tikhonova, J. Lattig, R. Magnan, M. Laval, C. Escrieut, C. Boulegue, C. Hewage, D. Fourmy, Identification of determinants of glucose-dependent insulinotropic polypeptide receptor that interact with N-terminal biologically active region of the natural ligand. *Mol Pharmacol* **77**, 547-558 (2010).
19. B. D. Kerr, A. J. Flatt, P. R. Flatt, V. A. Gault, Characterization and biological actions of N-terminal truncated forms of glucose-dependent insulinotropic polypeptide. *Biochem. Biophys. Res. Commun* **404**, 870-876 (2011).
20. M. B. N. Gabe, W. J. C. van der Velden, F. X. Smit, L. S. Gasbjerg, M. M. Rosenkilde, Molecular interactions of full-length and truncated GIP peptides with the GIP receptor - A comprehensive review. *Peptides* **125**, 170224 (2020).

21. C. Parthier, M. Kleinschmidt, P. Neumann, R. Rudolph, S. Manhart, D. Schlenzig, J. Fanghanel, J. U. Rahfeld, H. U. Demuth, M. T. Stubbs, Crystal structure of the incretin-bound extracellular domain of a G protein-coupled receptor. *Proc Natl Acad Sci U S A* **104**, 13942-13947 (2007).
22. D. Hilger, K. K. Kumar, H. Hu, M. F. Pedersen, E. S. O'Brien, L. Giehm, C. Jennings, G. Eskici, A. Inoue, M. Lerch, J. M. Mathiesen, G. Skiniotis, B. K. Kobilka, Structural insights into differences in G protein activation by family A and family B GPCRs. *Science* **369**, (2020).
23. A. Jazayeri, A. S. Dore, D. Lamb, H. Krishnamurthy, S. M. Southall, A. H. Baig, A. Bortolato, M. Koglin, N. J. Robertson, J. C. Errey, S. P. Andrews, I. Teobald, A. J. Brown, R. M. Cooke, M. Weir, F. H. Marshall, Extra-helical binding site of a glucagon receptor antagonist. *Nature* **533**, 274-277 (2016).
24. H. Zhang, A. Qiao, L. Yang, N. Van Eps, K. S. Frederiksen, D. Yang, A. Dai, X. Cai, H. Zhang, C. Yi, C. Cao, L. He, H. Yang, J. Lau, O. P. Ernst, M. A. Hanson, R. C. Stevens, M. W. Wang, S. Reedtz-Runge, H. Jiang, Q. Zhao, B. Wu, Structure of the glucagon receptor in complex with a glucagon analogue. *Nature* **553**, 106-110 (2018).
25. R. Chang, X. Zhang, A. Qiao, A. Dai, M. J. Belousoff, Q. Tan, L. Shao, L. Zhong, G. Lin, Y. L. Liang, L. Ma, S. Han, D. Yang, R. Danev, M. W. Wang, D. Wootten, B. Wu, P. M. Sexton, Cryo-electron microscopy structure of the glucagon receptor with a dual-agonist peptide. *J Biol Chem* **295**, 9313-9325 (2020).
26. H. Zhang, A. Qiao, D. Yang, L. Yang, A. Dai, C. de Graaf, S. Reedtz-Runge, V. Dharmarajan, H. Zhang, G. W. Han, T. D. Grant, R. G. Sierra, U. Weierstall, G. Nelson, W. Liu, Y. Wu, L. Ma, X. Cai, G. Lin, X. Wu, Z. Geng, Y. Dong, G. Song, P. R. Griffin, J. Lau, V. Cherezov, H. Yang, M. A. Hanson, R. C. Stevens, Q. Zhao, H. Jiang, M. W. Wang, B. Wu, Structure of the full-length glucagon class B G-protein-coupled receptor. *Nature* **546**, 259-264 (2017).
27. X. Zhang, M. J. Belousoff, P. Zhao, A. J. Kooistra, T. T. Truong, S. Y. Ang, C. R. Underwood, T. Egebjerg, P. Senel, G. D. Stewart, Y. L. Liang, A. Glukhova, H. Venugopal, A. Christopoulos, S. G. B. Furness, L. J. Miller, S. Reedtz-Runge, C. J. Langmead, D. E. Gloriam, R. Danev, P. M. Sexton, D. Wootten, Differential GLP-1R binding and activation by peptide and non-peptide agonists. *Mol Cell* **80**, 485-500 e487 (2020).
28. M. Dong, G. Deganutti, S. J. Piper, Y. L. Liang, M. Khoshouei, M. J. Belousoff, K. G. Harikumar, C. A. Reynolds, A. Glukhova, S. G. B. Furness, A. Christopoulos, R. Danev, D. Wootten, P. M. Sexton, L. J. Miller, Structure and dynamics of the active Gs-coupled human secretin receptor. *Nat Commun* **11**, 4137 (2020).
29. Y. Seino, M. Fukushima, D. Yabe, GIP and GLP-1, the two incretin hormones: Similarities and differences. *J Diabetes Investig* **1**, 8-23 (2010).
30. T. Coskun, K. W. Sloop, C. Loghin, J. Alsina-Fernandez, S. Urva, K. B. Bokvist, X. Cui, D. A. Briere, O. Cabrera, W. C. Roell, U. Kuchibhotla, J. S. Moyers, C. T. Benson, R. E. Gimeno, D. A. D'Alessio, A. Haupt, LY3298176, a novel dual GIP and GLP-1 receptor agonist for the treatment of type 2 diabetes mellitus: From discovery to clinical proof of concept. *Mol Metab* **18**, 3-14 (2018).
31. Y. L. Liang, M. J. Belousoff, P. Zhao, C. Koole, M. M. Fletcher, T. T. Truong, V. Julita, G. Christopoulos, H. E. Xu, Y. Zhang, M. Khoshouei, A. Christopoulos, R. Danev, P. M. Sexton, D. Wootten, Toward a structural understanding of class B GPCR peptide binding and activation. *Mol Cell* **77**, 656-668 e655 (2020).
32. Y. L. Liang, M. Khoshouei, G. Deganutti, A. Glukhova, C. Koole, T. S. Peat, M. Radjainia, J. M. Plitzko, W. Baumeister, L. J. Miller, D. L. Hay, A. Christopoulos, C. A. Reynolds, D. Wootten, P. M. Sexton, Cryo-EM structure of the active, Gs-protein complexed, human CGRP receptor. *Nature* **561**, 492-497 (2018).
33. J. Wang, X. Song, D. Zhang, X. Chen, X. Li, Y. Sun, C. Li, Y. Song, Y. Ding, R. Ren, E. H.

Harrington, L. A. Hu, W. Zhong, C. Xu, X. Huang, H. W. Wang, Y. Ma, Cryo-EM structures of PAC1 receptor reveal ligand binding mechanism. *Cell Res* **30**, 436-445 (2020).

34. A. H. Sparre-Ulrich, L. S. Hansen, B. Svendsen, M. Christensen, F. K. Knop, B. Hartmann, J. J. Holst, M. M. Rosenkilde, Species-specific action of (Pro3)GIP - a full agonist at human GIP receptors, but a partial agonist and competitive antagonist at rat and mouse GIP receptors. *Br J Pharmacol* **173**, 27-38 (2016).

35. S. R. Hoare, T. I. Bonner, T. B. Usdin, Comparison of rat and human parathyroid hormone 2 (PTH2) receptor activation: PTH is a low potency partial agonist at the rat PTH2 receptor. *Endocrinology* **140**, 4419-4425 (1999).

36. C. J. Bailey, GIP analogues and the treatment of obesity-diabetes. *Peptides* **125**, 170202 (2020).

37. M. A. Skow, N. C. Bergmann, F. K. Knop, Diabetes and obesity treatment based on dual incretin receptor activation: 'twincretins'. *Diabetes Obes Metab* **18**, 847-854 (2016).

38. K. Alexiadou, O. Anyiam, T. Tan, Cracking the combination: Gut hormones for the treatment of obesity and diabetes. *J Neuroendocrinol* **31**, e12664 (2019).

39. M. Bastin, F. Andreelli, Dual GIP-GLP1-receptor agonists in the treatment of type 2 diabetes: a short review on emerging data and therapeutic potential. *Diabetes Metab Syndr Obes* **12**, 1973-1985 (2019).

40. Y. L. Liang, P. Zhao, C. Draper-Joyce, J. A. Baltos, A. Glukhova, T. T. Truong, L. T. May, A. Christopoulos, D. Wootten, P. M. Sexton, S. G. B. Furness, Dominant negative G proteins enhance formation and purification of agonist-GPCR-G protein complexes for structure determination. *ACS Pharmacol Transl Sci* **1**, 12-20 (2018).

41. E. F. Pettersen, T. D. Goddard, C. C. Huang, G. S. Couch, D. M. Greenblatt, E. C. Meng, T. E. Ferrin, UCSF Chimera--a visualization system for exploratory research and analysis. *Journal of computational chemistry* **25**, 1605-1612 (2004).

42. P. Emsley, K. Cowtan, Coot: model-building tools for molecular graphics. *Acta Crystallogr D Biol Crystallogr* **60**, 2126-2132 (2004).

43. P. D. Adams, P. V. Afonine, G. Bunkoczi, V. B. Chen, I. W. Davis, N. Echols, J. J. Headd, L. W. Hung, G. J. Kapral, R. W. Grosse-Kunstleve, A. J. McCoy, N. W. Moriarty, R. Oeffner, R. J. Read, D. C. Richardson, J. S. Richardson, T. C. Terwilliger, P. H. Zwart, PHENIX: a comprehensive Python-based system for macromolecular structure solution. *Acta Crystallogr D Biol Crystallogr* **66**, 213-221 (2010).

44. V. B. Chen, W. B. Arendall, 3rd, J. J. Headd, D. A. Keedy, R. M. Immormino, G. J. Kapral, L. W. Murray, J. S. Richardson, D. C. Richardson, MolProbity: all-atom structure validation for macromolecular crystallography. *Acta Crystallogr D Biol Crystallogr* **66**, 12-21 (2010).

45. E. L. Wu, X. Cheng, S. Jo, H. Rui, K. C. Song, E. M. Davila-Contreras, Y. Qi, J. Lee, V. Monje-Galvan, R. M. Venable, J. B. Klauda, W. Im, CHARMM-GUI Membrane Builder toward realistic biological membrane simulations. *J. Comput. Chem* **35**, 1997-2004 (2014).

46. O. Guvench, S. S. Mallajosyula, E. P. Raman, E. Hatcher, K. Vanommeslaeghe, T. J. Foster, F. W. Jamison, 2nd, A. D. Mackerell, Jr., CHARMM additive all-atom force field for carbohydrate derivatives and its utility in polysaccharide and carbohydrate-protein modeling. *J Chem Theory Comput* **7**, 3162-3180 (2011).

47. B. Hess, P-LINCS: a parallel linear constraint solver for Molecular Simulation. *J Chem Theory Comput* **4**, 116-122 (2008).

48. K. M. Aoki, F. Yonezawa, Constant-pressure molecular-dynamics simulations of the crystal-smectic transition in systems of soft parallel spherocylinders. *Phys Rev A* **46**, 6541-6549 (1992).

49. S. Mitternacht, FreeSASA: An open source C library for solvent accessible surface area calculations. *FI000Res* **5**, 189 (2016).

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Competing interests: Authors declare that they have no competing interests.

Data and materials availability: All data are available in the main text or the supplementary materials.

Figures

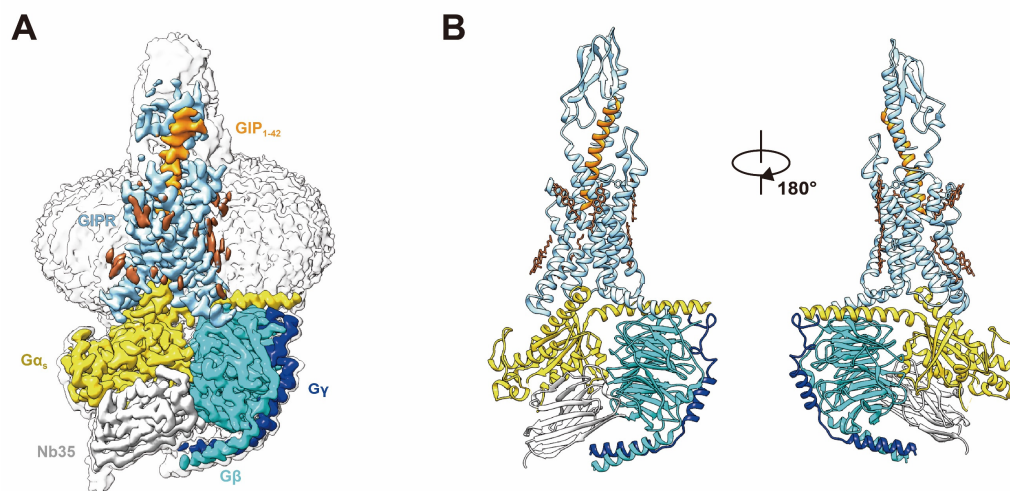
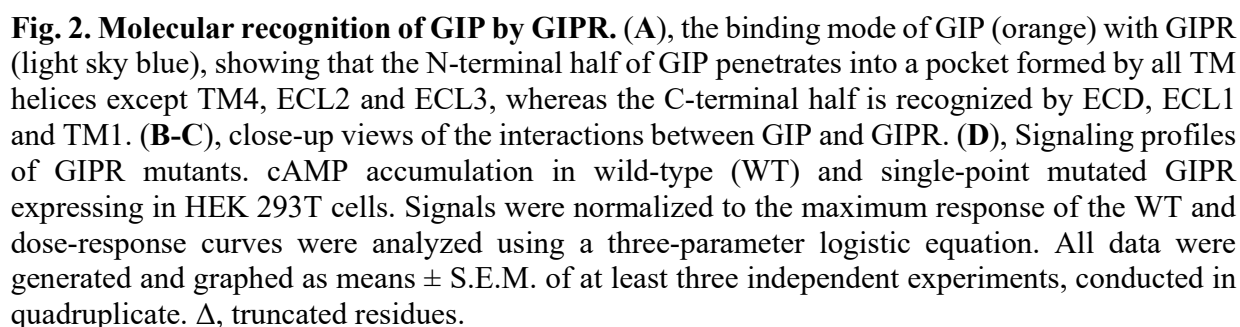


Fig. 1. Cryo-EM structure of the GIP₁₋₄₂-GIPR-G_s complex. (A), Cut-through view of the cryo-EM density map that illustrates the GIP₁₋₄₂-GIPR-G_s complex and the disc-shaped micelle. The unsharpened cryo-EM density map at the 0.07 threshold shown as light gray surface indicates a micelle diameter of 11 nm. The colored cryo-EM density map is shown at the 0.16 threshold. (B), Model of the complex as a cartoon, with GIP₁₋₄₂ as helix in orange. The receptor is shown in light sky blue, Gα_s in yellow, Gβ subunit in cyan, Gγ subunit in navy blue and Nb35 in gray.



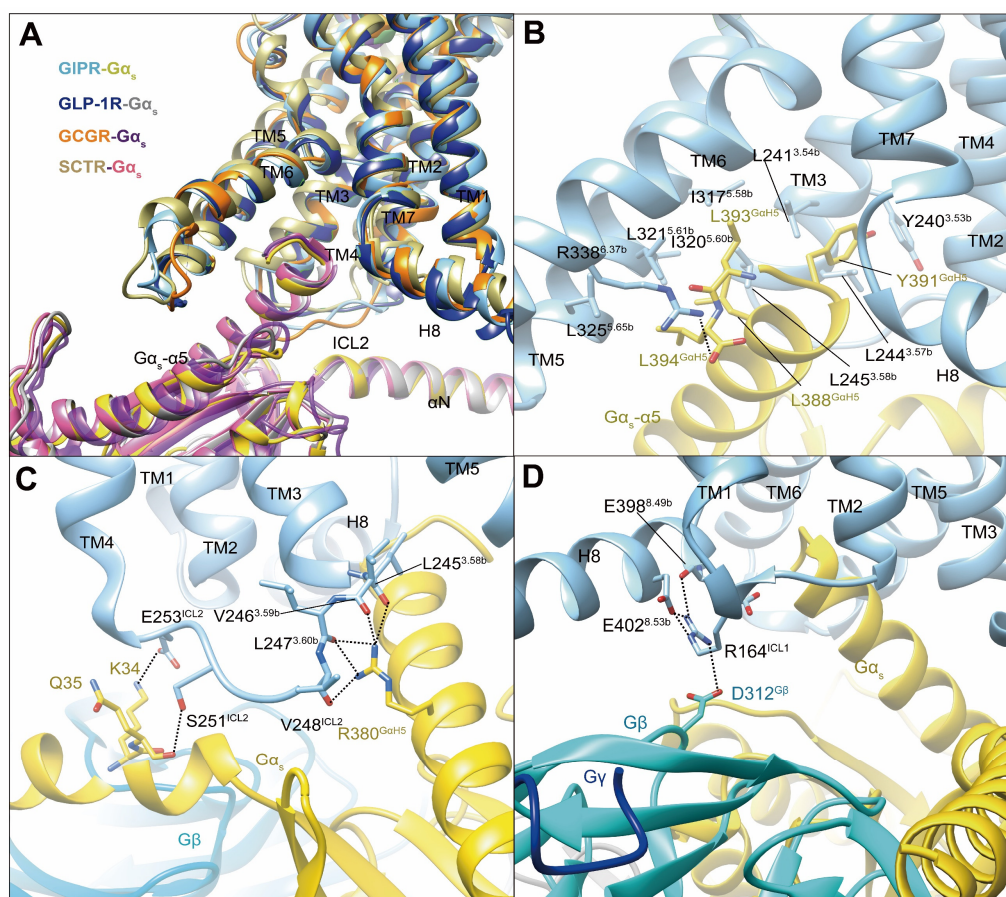


Fig. 3. G protein coupling of GIPR. (A), Comparison of G protein coupling among GIPR, GLP-1R (27), GCGR (14) and SCTR (28). The Gα_s α5-helix of the Gα_s Ras-like domain inserts into an intracellular crevice of GIPR TMD. (B), Interaction between GIPR and the C-terminus of Gα_s. (C), Polar interactions between ICL2 and Gα_s. (D), Polar interactions between H8 and ICL1 of the GIPR and Gβ. The GIP₁₋₄₂–GIPR–Gα_s structure is colored light sky blue (GIPR), gold (Gα_s) and cyan (Gβ). Residues involved in interactions are shown as sticks. Polar interactions are shown as black dashed lines.

