

Design and proof-of-concept for targeted phage-based COVID-19 vaccination strategies with a streamlined cold-free supply chain

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Abstract

Development of effective vaccines against Coronavirus Disease 2019 (COVID-19) is a global imperative. Rapid immunization of the world human population against a widespread, continually evolving, and highly pathogenic virus is an unprecedented challenge, and many different vaccine approaches are being pursued to meet this task. Engineered filamentous bacteriophage (phage) have unique potential in vaccine development due to their inherent immunogenicity, genetic plasticity, stability, cost-effectiveness for large-scale production, and proven safety profile in humans. Herein we report the design, development, and initial evaluation of targeted phage-based vaccination approaches against Severe Acute Respiratory Syndrome Coronavirus-2 (SARS-CoV-2) by using dual ligand peptide-targeted phage and adeno-associated virus/phage (AAVP) particles. Towards a unique phage- and AAVP-based dual-display candidate approach, we first performed structure-guided antigen design to select six solvent-exposed epitopes of the SARS-CoV-2 spike (S) protein for display on the recombinant major capsid coat protein pVIII. Targeted phage particles carrying one of these epitopes induced a strong and specific humoral response. In an initial experimental approach, when these targeted phage particles were further genetically engineered to simultaneously display a ligand peptide (CAKSMGDIVC) on the minor capsid protein pIII, which enables receptor-mediated transport of phage particles from the lung epithelium into the systemic circulation (termed “dual-display”), they enhanced a systemic and specific spike (S) protein-specific antibody response upon aerosolization into the lungs of mice. In a second line of investigation, we engineered targeted AAVP particles to deliver the entire S protein gene under the control of a constitutive cytomegalovirus (CMV) promoter, which induced tissue-specific transgene expression stimulating a systemic S protein-specific antibody response. As proof-of-concept preclinical experiments, we show that targeted phage- and AAVP-based particles serve as robust yet versatile enabling platforms for ligand-directed immunization and promptly yield COVID-19 vaccine prototypes for further translational development.

Significance

The ongoing COVID-19 global pandemic has accounted for over 2.5 million deaths and an unprecedented impact on the health of mankind worldwide. Over the past several months, while a few COVID-19 vaccines have received Emergency Use Authorization and are currently being administered to the entire human population, the demand for prompt global immunization has created enormous logistical challenges—including but not limited to supply, access, and distribution—that justify and reinforce the research for additional strategic alternatives. Phage are viruses that only infect bacteria and have been safely administered to humans as antibiotics for decades. As experimental proof-of-concept, we demonstrated that aerosol pulmonary vaccination with lung-targeted phage particles that display short epitopes of the S protein on the capsid as well as preclinical vaccination with targeted AAVP particles carrying the S protein gene elicit a systemic and specific immune response against SARS-CoV-2 in immunocompetent mice. Given that targeted phage- and AAVP-based viral particles are sturdy yet simple to genetically engineer, cost-effective for rapid large-scale production in clinical grade, and relatively stable at room temperature, such unique attributes might perhaps become additional tools towards COVID-19 vaccine design and development for immediate and future unmet needs.

Introduction

Since the beginning of 2021, the World Health Organization has estimated that over 2.5 million deaths in 192 countries/territories have been caused by complications of coronavirus disease 2019 (COVID-19). This unprecedented pandemic has prompted a worldwide collaborative effort to develop antiviral therapies and vaccines to control global spread. Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is the third zoonotic coronavirus to infect humans in less than 20 years (1, 2). Previous coronavirus epidemics, such as SARS-CoV and Middle East Respiratory Syndrome (MERS), foreshadowed the risk of emergent outbreaks and the imminent need for novel and versatile technologies for rapid manufacturing and large-scale distribution of vaccines and therapies as an emergency countermeasure.

SARS-CoV-2 is a single-stranded enveloped RNA virus with four main structural proteins. The spike (S) protein mediates both host cell recognition and membrane fusion and is pivotal for viral entry. The S protein, comprised of the S1 and S2 subunits, is displayed as a trimer on the surface of the viral particle (3). Within the S1 subunit, the receptor-binding domain (RBD) adopts an open conformation that can interact with the angiotensin-converting enzyme 2 (ACE2) receptor on the host cell membrane. Upon binding, the S1 subunit is cleaved and subsequent conformational changes in the S2 subunit trigger the formation of a six-helical bundle comprised of heptapeptide repeat sequence 1 (HR1) and heptapeptide repeat sequence 2 (HR2), followed by the insertion of the fusion peptide (FP) into the host cell membrane. Given the importance of the S protein for the entry of SARS-CoV-2 in the host cells, it has served as the main target for vaccines and therapeutic antibodies. Thus, the identification and understanding of structurally-defined S protein epitopes with potential neutralizing capabilities are crucial in the design of effective and robust vaccines and antibody cocktail therapies (4, 5).

Current vaccine platforms against COVID-19 are classified into several broad categories that include nucleic acid-based vaccines (mRNA and DNA), viral vector vaccines (e.g., adenovirus), inactivated or live attenuated viral vaccines, or recombinant protein or peptide vaccines. Some have been granted Emergency Use Authorization by various regulatory agencies, including two mRNA vaccines (Pfizer-BioNtech and Moderna), three non-replicating adenovirus vaccines (Oxford/AstraZeneca, Sputnik V, and Johnson & Johnson), and an inactivated virus vaccine (CoronaVac), while many others are pending approval or undergoing clinical trials. In addition to such extraordinary global efforts, further exploration and development of vaccines that are amenable to temperature fluctuations, rapid large-scale production and distribution, and conferral of long-term immunological protection in the face of existing and emerging variant strains remain an unmet need (6, 7).

Phage particles have been used in clinical settings for nearly a century, and represent an inexpensive, and versatile tool for large-scale immunization. Phage are viruses that naturally infect bacteria; during the pre-antibiotic era, humans received phage to neutralize systemic bacterial infections without significant adverse effects (8, 9). Recently, engineered phage particles have been leveraged in different applications, particularly in vaccine development because they are: (i) easy to genetically engineer and produce in large scale in *Escherichia coli*; (ii) strong immunogens capable of stimulating both cellular and humoral immunity (10-12); (iii) highly stable under harsh environmental conditions (pH and temperature). Such features facilitate administration, storage, and transport (10); (iv) importantly, they are considered safe for administration to humans (9, 13-15). We and others have shown that the addition of peptide targeting sequences to the minor coat protein pIII and immunogenic peptide sequences on the recombinant pVIII coat protein (rpVIII) confer tropism of phage to specific eukaryotic receptors in normal or diseased tissue or to organelles (16-21). Pertinent to vaccine development, we recently demonstrated that aerosolized phage particles targeting the lung and subsequently undergoing receptor-mediated transport to the systemic circulation are safe and elicit specific and sustained local and systemic immune responses in mice and non-human primates (22). Further genome engineering was leveraged to create a hybrid adeno-associated virus/phage vector (20), which has been extensively validated for ligand-directed gene delivery and represents a viable alternative for nucleotide-based vaccines (mRNA or DNA). Several favorable features were built into the vector design, including: (i) adjuvant properties of phage particles, (ii) ligand-directed system for receptor-mediated internalization, (iii) well-characterized fate of the genome (concatemerization and integration of intact genomes), and (iv) ability to avoid neutralization after an immune response against phage, as proven in repeat-dose studies using pet dogs (23) and several animal models (20, 23-28).

We, therefore, explored the inherent biological and genetic properties of phage particles to produce novel vaccine candidates against COVID-19. First, guided by structure-based antigen design, we selected S protein epitopes that were genetically incorporated into the recombinant major capsid coat protein pVIII (rpVIII) of the f88-4 phage genome. The peptide sequence CAKSMGDIVC, which was recently described for its unique ability to target lung epithelium cells and induce the transport of phage particles into the systemic circulation (22), was then cloned into phage minor coat protein pIII of the fUSE55 phage genome, creating a dual-display system for efficient delivery and immunization against the SARS-CoV-2 S protein. This approach allows us to use the lung as the main route of vaccination, which has remarkable advantages over conventional routes. Aerosol administration is needle-free and requires minimal use of specialized medical staff; it is not subject to first-pass metabolism and is considered the most effective route for inducing local

immune protection against airborne pathogens (29-32). In a second approach, we used the AAVP vector (20), which contains a eukaryotic expression cassette flanked by AAV inverted terminal repeat (ITR) sequences within the phage genome, for the delivery and transduction of the full-length S protein gene (AAVP S) in host cells. This approach leverages the same rationale that serves as the basis for the activity of existing vaccines (mRNA or adenovirus) that trigger a specific humoral response against the full-length S protein. Integration of large DNA antigen sequences flanking the AAV transgene of the AAVP confers stability to the construct upon transduction and enhanced expression efficiency. Both phage- and AAVP-based vaccines elicited systemic S protein-specific humoral response in mice with no evidence of adverse effects, indicating that both technologies hold promise in vaccine development.

Results

Development of novel phage- and AAVP-based vaccine platforms against SARS-CoV-2

We pursued two different strategies for immunization: (i) phage-based vaccine candidates displaying various S protein epitopes, and (ii) an AAVP-based vaccine candidate that utilizes the entire SARS-CoV-2 S protein (Fig. 1). In both approaches, we incorporated a ligand peptide along with the viral antigens in the phage or AAVP to target specific cell surface receptors and facilitate the immune response.

In the capsid engineering system, we genetically engineered phage to display immunologically relevant S protein epitopes (see below) on the highly exposed pVIII protein of the phage capsid using the f88-4 vector (Fig. 1, Step 1) (18, 19). To enable tissue-specific targeting of these phage particles, we also subcloned the coding sequence of the novel CAKSMGDIVC peptide ligand into the pIII gene of the fUSE55 vector, yielding a dual-display phage (Fig. 1, Step 2). The CAKSMGDIVC ligand binds to $\alpha 3\beta 1$ integrins and mediates the transport of phage particles across the lung epithelium to the systemic circulation where they elicit strong and sustained pulmonary and systemic humoral responses against antigens displayed on the phage capsid (22). As a control, we used the untargeted parental phage particles (insertless phage) that display the native pVIII and pIII proteins.

For our second strategy, based on gene delivery, we inserted an expression cassette containing the full-length S protein transgene and the human CMV promoter in *cis* conformation within the 5' and 3' ITRs in the AAVP genome for gene delivery and transduction in host cells (Fig. 1, Step 2). As a control, we used the targeted AAVP empty vector (termed "AAVP S-null"). Targeting was established by display of another integrin-binding peptide, ACDCRGDCFCG (RGD4C) on the pIII protein, which has high affinity for α_v integrins (17, 33), which are highly expressed in leukocytes trafficking to draining lymph

nodes and areas of inflammation (34). The RGD motif (arginine-glycine-aspartate) facilitates particle uptake by dendritic cells and enhances the immunogenicity of peptide antigens, DNA vaccines, and adenovirus vectors (35-37).

Both approaches allow the rapid “swapping” of targeting motifs, epitopes, and/or gene coding sequences, thus providing rapid and powerful design flexibility to produce a variety of vaccines and overcome potential limitations in protein conformation of structure-based epitopes. The dual-display phage particles, the RGD4C-AAVP S particles, and corresponding controls were tested *in vivo* in mice to assess different routes of administration, and to evaluate the induced antigen-specific humoral response by ELISA (Fig. 1, Step 3). The overall vaccination schedule included at least two administered doses of 10^9 transducing units (TU) of phage or AAVP particles with an interval of 1–2 weeks.

Identification and selection of epitopes for dual-display phage-based vaccine

To identify relevant epitopes for phage capsid manipulation, *in silico* analysis of the experimentally-determined viral S protein structure of the Wuhan-Hu-1 strain (GenBank Accession number: NC_045512.2) was performed. We prioritized solvent-exposed amino acid stretches with flanking cysteine residues and cyclic conformation, because these amino acid sequences are more likely to recapitulate the composition of endogenous epitopes and thereby increase the likelihood of antigen recognition and processing by the host immune system. Other epitopes were also considered following structure-guided predictions, even in the absence of flanking cysteine residues. Also, given that phage particles are produced in prokaryotic host cells, we prioritized epitopes lacking sites expected to undergo post-translational modification.

We selected six S protein epitopes, which are accessible in both the closed and open states of the S protein. At least five of these epitopes have since been shown to be fully or partially immunogenic (Fig. S1 and Table S1) (38-45). The six epitopes range in length from 9 to 26 amino acids (aa). Four occur within the S1 subunit and two are found in the S2 subunit (Fig. 2A). Three of the S1 epitopes are located in the receptor-binding domain (RBD): epitope 1 (aa 336–361), epitope 2 (aa 379–391), and epitope 3 (aa 480–488). The remaining epitope derived from the S1 subunit, epitope 4 (aa 662–671), is located near the cleavage site between the S1 and S2 subunits. Epitopes within the S2 subunit, epitope 5 (aa 738–760) and epitope 6 (aa 1032–1043), are located near FP (aa 788–806) and HR1 (aa 912–984), respectively. Most of the selected epitopes are cyclic due to the presence of flanking cysteine residues, except epitope 2 (aa 379–391), which adopts a loop-like conformation despite the absence of disulfide bridges (Fig. 2B).

Many studies have demonstrated that the S protein is highly glycosylated, and some glycosylation sites have been reported to alter the infectivity of variants and facilitate evasion

of the host immune response (4, 7). Considering that we selected epitopes based on conformation, we identified that epitope 1 contains a glycosylation site on residue N343. Notably, this site seems to be important in viral infectivity in which the glycosylation deletion N343Q drastically reduces the D614G variant infectivity (7). In our system, however, we expect that the lack of N-glycosylation will not produce a significant structural divergence in the epitope conformation when displayed on the phage capsid, as similarly observed with other SARS-CoV-2 strains (46). Because the glycosylation site of epitope 1 is located in its N-terminus and unlikely to interrupt antibody recognition of the remaining structure, we sought to investigate the efficacy of this additional epitope.

Epitope conformational analysis by molecular dynamics

To examine whether the predicted conformation of each epitope on display on the phage capsid would recapitulate its features on the S protein, we performed *in silico* conformational analysis using microsecond-scale all-atom explicit-solvent simulations of each epitope. We hypothesized that epitope conformation is a pre-requisite for successful induction of the immune response against the native S protein. Notably, we found that epitope 4 has the lowest spatial root-mean-square deviation (RMSD) amongst the four S1 subunit epitopes. While its RMSD fluctuates between 2 Å and 4 Å (Fig. 3A), epitope 4 has the most frequent low values (RMSD~2Å), which may explain its near-native conformation when compared to the structure of the S protein (Fig. 3B) (47). As for the other selected epitopes of the S1 subunit (epitopes 1, 2, and 3) and the S2 subunit (epitopes 5 and 6), RMSD values are substantially higher (4–10 Å) (Fig. 3C, D), after an initial relaxation of each system (~100 nanoseconds). These differential RMSD trends suggest that epitope 4 is most likely to resemble the native conformation and generate IgG antibodies with high cross-reactivity with the S protein. If true, epitope 4 represents a promising candidate for display on phage capsid that would elicit an immune response towards an exposed region of the S protein. It would also demonstrate that molecular dynamics simulations can serve as a useful tool for prioritizing epitopes with higher likelihood of recapitulating native conformation when selecting epitope candidates for use in phage-based vaccines.

Characterization of structurally-defined S epitopes for immunogenicity in mice

To evaluate the immunological potential of each of S epitope and select promising candidate(s) for phage-based vaccine development, we tested their ability to induce an immune response in mice. Six f88-4 phage were produced, each exhibiting one of the six S protein epitopes fused into the rpVIII protein on ~300 copies per phage particle (termed “single-display phage”) (Fig. S2).

Immunogenicity of the epitopes expressed on the rpVIII protein was assessed in mouse serum (Swiss Webster or BALB/c) obtained after the first dose (prime) and the second dose (boost) administered subcutaneously and compared against the control insertless phage. Antigen-specific IgG titers were quantified by ELISA using an immobilized recombinant S protein (aa 16–1213) for capture. Epitope 4 from the S1 subunit induced high levels of S protein-specific IgG antibodies, and booster immunizations further increased antibody levels (Fig. 4A). None of the other five phage constructs generated S protein-specific IgG antibodies above the background levels observed for mice immunized with the control insertless phage. These results indicate that epitope 4 is the most immunogenic among the selected epitopes, suggesting that epitope display of the native conformation supports development of specific immune response as predicted by the *in silico* analysis.

Given the well-documented inherent immunogenicity of native filamentous phage, we also investigated the levels of phage-specific IgG antibodies in the sera of these mice. Notably, all single-display phage constructs generated high titers of phage-specific IgG antibodies, which were markedly increased after the second dose independent of the presence or absence of the S protein epitopes displayed on the rpVIII protein (Fig. 4B). It is remarkable that generation of phage-specific IgG antibodies does not appear to compromise the S protein-specific humoral response. A clear distinction was observed between epitope 4 and the other phage particles, including the control insertless phage. Epitope 4, therefore, was selected as the lead candidate for testing a dual-display phage-based vaccine bearing a targeting moiety.

Dual-display phage construct for pulmonary vaccination

To generate a dual-display phage-based vaccine, we optimized a simple two-step cloning strategy that allows rapid exchange of epitopes and/or targeting peptide ligands in the phage genome. This methodology has the potential to rapidly mitigate the shortcomings in the neutralization due to possible mutations and/or direct the phage to target cells or tissue to improve the immune response. Pulmonary vaccination has been shown to be the most efficient route to generate mucosal and systemic immunity against airborne pathogens (22, 29-31) and significantly increase immunological protection of non-human primates infected with SARS-CoV-2 (30, 32). Therefore, we prioritized the peptide CAKSMGDIVC for lung targeting and aerosol delivery (22) and generated dual-display phage particles that simultaneously express epitope 4 on rpVIII protein (~300 copies) and the peptide CAKSMGDIVC on pIII protein (3-5 copies).

To determine the immunogenic properties of the epitope 4/CAKSMGDIVC dual-display phage particles, groups of five-week-old BALB/c female mice were dosed intratracheally. Cohorts of mice (n=10 per group) received two doses of 10^9 TU of the

epitope 4/CAKSMGDIVC dual-display phage, or the epitope 4 single-display phage, or the control insertless phage in 3-week intervals. The presence and levels of S protein-specific IgG antibodies were evaluated in serum samples collected weekly by ELISA. Titers of S protein-specific IgG antibodies were higher in mice immunized with the epitope 4/CAKSMGDIVC dual-display phage particles compared to the control insertless phage, especially after three weeks of the first dose, and with a substantial increase after the second dose (weeks 4 and 5) (Fig. 4C). Epitope 4 single-display phage particles also induced systemic S protein-specific IgG antibodies but do so at levels lower than the dual-display phage particles. These results indicate that the addition of the CAKSMGDIVC peptide ligand mediates the transport of the dual-display phage from the pulmonary tree to the systemic circulation, thereby increasing the immunogenicity.

Together, our findings demonstrate that epitope 4 induces a robust S protein-specific humoral response when displayed on the rpVIII protein as a single display entity (single-display phage particles) and, when combined with the lung transport peptide CAKSMGDIVC, immunogenicity is enhanced. These results provide encouraging evidence that epitope 4/CAKSMGDIVC dual-display phage particles are suitable candidates for pulmonary vaccination against SARS-CoV-2 and pave the way for further validation of phage-based cocktails as vaccines that cover a broad spectrum of antigenic sites that elicit the production of antibodies with well-established and/or new neutralizing capabilities.

A novel AAVP-based vaccine for efficient gene delivery and humoral response against the viral S protein

As a parallel approach to the dual-display phage design, we adapted a long-standing and well validated targeted AAVP gene therapy platform (20) to deliver the S protein gene (Wuhan-Hu-1 strain, GenBank Accession Number: NC_045512.2) (termed “AAVP S”). AAVP S displays the ACDCRGDCFCG (RGD4C) targeting peptide ligand on the pIII protein, allowing it to bind α_v integrins. These integrins are known to regulate trafficking of lymphocytes and antigen-presenting cells (i.e., dendritic cells) into secondary lymphoid organs (37). For the control, we generated a RGD4C-AAVP empty vector that is identical to RGD4C-AAVP S but does not contain the S protein gene (RGD4C-AAVP *S-null*) (Fig. 5A, B).

To assess the immunogenicity of RGD4C-AAVP S, we first immunized five-week-old female outbred Swiss Webster mice. Cohorts of mice (n=5) received 10^9 TU of targeted RGD4C-AAVP S *via* intraperitoneal (group 1), intravenous (group 2), intratracheal (group 3), or subcutaneous (group 4) routes. The S protein-specific IgG antibody response was evaluated 14 days later by ELISA (Fig. 5C). Baseline sera were used as controls. Administration of RGD4C-AAVP S particles elicited serum IgG responses against the S

protein in all experimental groups, whereas mice immunized *via* subcutaneous injection had higher serum IgG titers than the other groups. Thus, we elected to administer RGD4C-AAVP S subcutaneously for subsequent *in vivo* assays.

We tested our immunization regimen in five-week-old female inbred BALB/c mice. Cohorts of mice (n=12 per group) received weekly subcutaneous doses of 10^9 TU of RGD4C-AAVP S or the control, RGD4C-AAVP S-*null*. Higher titers of S protein-specific IgG antibodies were observed in mice vaccinated with RGD4C-AAVP S relative to the baseline sera and RGD4C-AAVP S-*null*, particularly five weeks after the first dose, confirming that RGD4C-AAVP S is a suitable vector for transgene delivery and elicits a systemic humoral immune response (Fig. 5D).

To gain insight into the S protein transgene expression mediated by RGD4C-AAVP S, we investigated the fate of the phage genome, namely, the sites in which transduced cells were detected in mouse tissues, including main regional lymph nodes (axillary, inguinal, mesenteric, and mediastinal) 4 weeks after the first dose. Transgene expression at varying levels was detected mainly in the draining lymph nodes. Skeletal muscle and spleen were used as control organs and show only background levels of transgene expression (Fig. 5E). As expected, transcripts for the S protein were not detected in mice immunized with RGD4C-AAVP S-*null*. These results confirm that gene delivery by AAVP and the expression of the S protein in the draining lymph nodes trigger a systemic S protein-specific humoral response. Moreover, the data recapitulate the well-established attribute of AAVP particles in preventing off-target effects, even upon clearance *via* the reticulum-endothelial system (RES), sparing non-targeted or distal tissues, while a strong promoter drives the expression of the transgene in the transduced cells (20, 23-28). This finding is particularly important for evaluating potential adverse effects in novel, candidate AAVP-based vaccines, since off-site transduction-associated toxicities have been reported in toxicological studies of adenovirus vaccines (48, 49). Therefore, the extensive body of data generated to date with AAVP in cancer gene therapies (20, 23-28) can help accelerate future clinical development of AAVP vaccine candidates, potentially in a highly cost-effective and efficient manner.

Finally, we examined the antibody response against phage in mice vaccinated with RGD4C-AAVP S or RGD4C-AAVP S-*null* phage. We observed a strong and sustained phage-specific IgG antibody response upon administration of RGD4C-AAVP S by all routes of administration (Fig. 5F) and the response increased after boost injections in both RGD4C-AAVP S (Fig. 5G) and RGD4C-AAVP S-*null* (Fig. 5G), indicating that AAVP are strong immunogens and likely serve as adjuvants for AAVP-based vaccines. Together, these results indicate that targeted AAVP S is an efficient tool for delivery and expression of viral proteins and can induce an immune response against the S protein.

Discussion

In this study we designed, generated, and evaluated the translational potential of phage- and AAVP-based vaccine candidates using capsid epitope display and gene delivery as strategies for immunization against SARS-CoV-2. We demonstrated that both systems can be successfully used to induce an antigen-specific humoral response against the S protein and therefore represent valid candidate approaches for vaccine development. Notably, as part of ongoing work, we have optimized Good Manufacturing Practice (GMP) generation, production, and purification of engineered phage particles so that manufacturing could be accomplished on an industrial scale for rapid commercialization.

One of the main challenges associated with vaccines deployed under Emergency Use Authorization (EUA) is to predict the potency and duration of the protective immune response against host cell-engaging epitopes on the S protein, particularly in the face of new genetic variants. In principle, focusing on structural antigen mapping and immunodominant B- and T-cell epitopes that trigger immune responses associated with both potent neutralizing activity and lower transmissibility would lead to long-term protection. As such, many studies have aimed to predict an/or map epitopes from B- and T-cells derived from the SARS-CoV-2 S protein and other structural proteins (50). In this study, we selected six exposed regions of the S protein with specific structural constraints for display on the phage capsid and to increase the likelihood of antigen recognition and processing by the host immune system. We found that epitope 4 (aa 662–671) triggered a strong and specific systemic humoral response against the S protein, presumably by recapitulating the near-native conformation of the epitope when expressed on the rpVIII protein as predicted by molecular dynamics analyses. Thus, the combinatorial approach of selecting regions of an antigen based on conformational constraints and evaluating their structural dynamics *in silico* can be used to identify epitopes likely to replicate the natural immune response to an infection. Our findings suggest that antigen-engineering strategies, such as the vaccine candidate for the Zika virus (51), have the potential to generate vaccines with high efficacy in inducing a broadly functional repertoire of neutralizing antibodies and cell-mediated immune responses.

To support the translational applications of phage-based vaccination, we designed a protocol for immunization in mice as a proof of principle towards pulmonary vaccination against SARS-CoV-2. The design of dual-display phage particles relies on a clever, yet simple bioengineering exercise: the simultaneous display of both epitope 4 on the pVIII protein and the CAKSMGDIVC targeting ligand on the pIII protein. Because CAKSMGDIVC mediates selective targeting and transport of phage particles to the systemic circulation (22), we showed that an aerosol strategy of immunization may confer biologically significant advantages over conventional routes of immunization. First, unlike subcutaneous or

intramuscular injections, inhalation is needle-free and minimally invasive, eliminating the requirement for specialized medical staff, and leading to the potential for self-administration for isolated and/or vulnerable populations sheltering in place, particularly the elderly and the immunocompromised. Moreover, rapid access to the upper and lower respiratory tract leads to a reduction in viral shedding, which results in lower transmission (52-54).

Antigen exposure to the lung surface, which is lined by the highly vascularized pulmonary epithelium, is a unique feature that is known to induce a local immune response that decreases infection and transmission of airborne pathogens. Emerging studies of intranasal or intratracheal immunizations have already proven successful protection against SARS-CoV-2 in mice and non-human primates (32, 54).

We also uncovered the potential value of using AAV-Phage particles to deliver the S protein gene as an additional alternative SARS-CoV-2 vaccine candidate. In the last decade, AAVP technology has proven to be a modular platform that can be appropriately tailored to image and treat a variety of human solid tumors in mouse models and spontaneous tumors in pet dogs (20, 23-28). These attributes make AAVP a unique platform for gene delivery. Indeed, we showed that administration of RGD4C-AAVP S particles elicits an antibody response in mice against the encoded S protein. Because our prototype AAVP S vaccine is targeted with the integrin-binding peptide (RGD4C), which has a high affinity binding for α_v integrins, this approach may facilitate targeting of inflammatory cells trafficking to the lymph nodes where gene expression and antigen presentation occur. The identification of a RGD motif within the RBD domain of the S protein suggests that integrins may act as co-receptors or an alternate path for coronavirus entry (55). Therefore, it is plausible that different functional ligands for tissue-specific transgene expression within lymph nodes (12), lymphatic vessels (56), or lung epithelial cells (e.g., CAKSMGDIVC) which has proven to be highly efficient in inducing local and systemic immune response upon pulmonary delivery (22), may enhance the efficacy and broad administration of AAVP-based vaccines.

In conclusion, here we bring forward and validate the first proof-of-concept for the design, structure-function relationship, and translation of both phage- and AAVP-based vaccines against SARS-CoV-2. A targeted pulmonary vaccination strategy with a simplified supply chain may also be applicable for potential utility against other viruses.

Materials and Methods

Animals. Four-to-six-week-old Swiss Webster and BALB/c mice were purchased from the Jackson Laboratory (Sacramento, CA) and were housed in specific pathogen and opportunist free (SOPF) rooms with controlled temperature ($20 \pm 2^\circ\text{C}$), humidity ($50 \pm 10\%$), light cycle (light, 7:00-19:00; dark, 19:00 – 7:00), and access to food and water *ad libitum* at the research animal facilities of the Rutgers Cancer Institute of New Jersey. Littermates were randomly assigned to experimental groups. The Institutional Animal Care and Use Committee (IACUC) from the Rutgers Cancer Institute of New Jersey approved all animal experiments.

Structural Analysis of S Protein for Epitope Selection. The structure of SARS-CoV-2 S protein (PDB IDs: 6VXX, 6VYB) (4) was analyzed using UCSF Chimera software (57) for selection of epitopes to display on rpVIII protein. Though epitope 3 (aa 480–488) was not resolved in these early structures, the flanking cysteine residues of this region were predicted to form a disulfide bridge (58). This has been confirmed experimentally with since-determined structures (e.g., PDB IDs: 6ZP0, 6ZGG) (59, 60).

Molecular Dynamics Simulations. All-atom explicit-solvent simulations of the epitope sequences were performed with the GROMACS 2020 software package (61, 62). The initial configuration for each epitope was taken from a cryo-EM structure of the full-length S protein (PDB ID: 6XR8) (63). Each epitope was solvated using the TIP3P water model (64), where the box size was defined to have a 10 Å buffer between the edge of the box and the epitope. Depending on the charge of each molecule, neutralization with either Cl^- or Na^+ ions was applied. The AMBER99SB-ILDN protein force field (65) was used for all simulations. After steepest-descent energy minimization, each system was equilibrated at 270 K using the NVT ensemble for 5 nanoseconds (ns), followed by the NPT ensemble for 5 ns, while position restraints were imposed on all heavy atoms (1000 kJ/nm^2). Restraints were then removed and steepest-descent minimization was performed, followed by NVT and NPT equilibration simulations (5 ns each, at 270 K). All production runs were performed in the NPT ensemble using the Nose-Hoover thermostat (61, 66) at 310 K and the Parrinello-Rahman barostat (67) set at 1 bar. Each production simulation was performed for a minimum of 1 microsecond (ms) (21 ms, aggregate simulated time). To ensure reproducibility of the results, a second set of simulations were equilibrated using identical protocols, except that the temperature during equilibration was 310 K. The overall dynamics were not sensitive to the equilibration temperature.

Generation of S Protein Epitopes Single-Display Phage Particles. To generate single-display phage constructs, we used the fd-tet-derived vector f88-4 containing a recombinant gene VIII (GenBank Accession Number: AF218363.1). Single colonies were selected on Luria-Bertani (LB) agar plates with tetracycline (40 µg/mL) and cultured overnight (O.N.). Each plasmid DNA was isolated by plasmid purification (Qiagen). Next, annealed oligonucleotides encoding for each of the six selected epitopes: epitope 1: fwd 5'AGCTTTGCCTGTCCGTTCCGGCGAAGTGTTCAACGCGACCCGCTTCGCGAGCGTGTAT GCGTGGAACCGCAAACGCATCAGCAACTGTCCTGCA 3', rev 5' GGACAGTTGCTGATGCGTTTTCGGTTCCACGCATACGCTCGCGAAGCGGGTCGCG TTGAACACTTCGCCGAACGGACAGGCAA 3'; epitope 2: fwd 5' AGCTTTGCCTGTTATGGCGTGAGCCCGACCAAACCTGAACGATCTGTGTCCTGCA 3', rev 5' GGACACAGATCGTTTCAGTTTGGTCGGGCTCACGCCATAACAGGCAA 3'; epitope 3: 5' AGCTTTGCCTGTAACGGCGTGGAAGGCTTCAACTGTCCTGCA 3', rev 5' GGACAGTTGAAGCCTTCCACGCCGTTACAGGCAA 3'; epitope 4: fwd 5' AGCTTTGCCTGTGATATCCCGATCGGCGCGGGCATCTGTCCTGCA 3', rev 5' GGACAGATGCCCGCGCCGATCGGGATATCACAGGCAA 3'; epitope 5: fwd 5' AGCTTTGCCTGTACCATGTATATCTGTGGCGATAGCACCGAATGTAG 3', rev 5' CAACCTGCTGCTGCAGTATGGCAGCTTCTGTCCTGCA 3'; epitope 6: fwd 5' GGACAGAAGATCGCCACAGATATACATGGTACAGGCAA 3', rev 5' AGCTTTGCCTGTGTGCTGGGCCAGAGCAAACGCGTGATTCTGTCC 3' were mixed at equimolar ratio and annealed using a thermocycler (93°C for 3 min, 80°C for 20 min, 75°C for 20 min, 70°C for 20 min, 65°C for 20 min, 40°C for 60 min). The f88-4 plasmid was digested with HindIII and PstI restriction endonucleases and ligated with the annealed double stranded oligonucleotides as described (18, 19). Each ligation product was electroporated into electrocompetent DH5α *E.coli*. Sequence-verified individual clones were used to infect K91kan *E.coli*. Phage particles were cultured in LB media containing 1 mM IPTG, tetracycline (40 µg/mL), and kanamycin (100 µg/mL), and were purified by polyethylene glycol (PEG)-NaCl precipitation method (17). Titration of single-display phage particles was carried out by infection of host bacterial cells K91kan *E. coli* for colony counting and represented as transducing units (TU/µL).

Generation of Dual-Display Phage Particles. To produce phage particles simultaneously displaying SARS-CoV-2 S protein epitope 4 (aa 662–671) on rpVIII protein and the lung transport peptide CAKSMGDIVC on pIII protein, we fused the single-display phage constructs (described above) and the fUSE55 genome to create a chimeric vector. The f88-4 vector-derived DNA fragment containing the rpVIII gene was inserted into the fUSE55 phage vector by double digestion of both vectors with XbaI and BamHI restriction enzymes at 37

°C, 4 h. After incubation, DNA fragments were loaded onto an agarose gel (0.8%, wt/vol). Under an ultraviolet transilluminator, the 3,925 bp fUSE55 DNA fragment and the 5,402-bp f88-4 DNA fragment containing epitope 4 in-frame with the rpVIII gene were excised. Fifty nanograms (ng) of the fUSE55 DNA fragments were ligated to 68.8 ng of the f88-4 DNA fragment with T4 DNA ligase (1U) in a final volume of 20 µL, O.N. at 16 °C for 16 h. An aliquot of the ligation reaction was transformed into electrocompetent DH5α *E.coli* and plated onto LB agar plates containing 40 µg/mL of tetracycline. Positive clones were verified by Sanger sequencing analysis and the plasmid containing the chimeric vector was purified with a QIAprep Spin Miniprep kit (Qiagen). The chimeric vector containing the f88-4 epitope 4 (aa 662–671) and fUSE55 was then digested with SfiI at 50°C for 4 h. The digested chimera was ligated to annealed oligonucleotides encoding the CAKSMGDIVC peptide into the fUSE55 pIII gene to generate the dual-display phage vector. The titration of dual-display phage particles was carried out by infection of host bacterial cells K91kan *E.coli*.

Genetic Engineering and Production of RGD4C-AAVP S and RGD4C-AAVP S-null

Particles. The 3.821 kb SARS-CoV-2 S protein coding sequence (Genbank Accession number NC_045512.2) was synthesized at GeneWiz (South Plainfield, NJ) with modifications to simplify subcloning into the RGD4C-AAVP-TNF genome. The single EcoRI restriction site at 1371 bp was deleted by replacing the thymidine nucleotide at position 1374 to cytosine in the wobble codon position without changing the translated asparagine residue at position 437. The 69-nucleotide sequence of the human interferon leader sequence and the 19-nucleotide sequence of the poly A region in RGD4C-AAVP-TNF (Genbank Accession number URN.LOCAL. P-7734T13) were added to the 5' or 3' ends, respectively, to the modified synthetic SARS-CoV-2 S gene to produce a 3.909 kb modified SARS-CoV-2 S gene that was subcloned into the EcoRI and Sall restriction sites of pUC57/Amp^R at GeneWiz. The first EcoRI restriction site at 829 bp within the AgeI and KsaI restriction sites in RGD4C-AAVP-TNF was deleted in two steps to mutate a thymidine to cytosine nucleotide at position 833 without altering the translated amino acid using the Q5 site-directed mutagenesis kit (New England Biolabs, Ipswich, MA) by following the manufacturer's protocol. Positive clones were verified by EcoRI restriction mapping, confirmed with overlapping sense and antisense primers by Sanger sequencing (SeqStudio, Thermo Fisher Scientific) using the BigDye Terminator v.3.1 Cycle Sequencing and XTerminator Purification kits (Applied Biosystems, Thermo Fisher Scientific) and analyzed using SnapGene software (GSL Biotech, San Diego, CA). Positive RGD4C-AAVP-TNFΔEcoRI829/MC1061 F⁻ colonies containing a unique EcoRI site at 10,191 kb were identified by EcoRI restriction mapping of dsDNA purified from 50 mL overnight animal-free

LB Broth, pH 7.4 cultures containing 100 µg/mL streptomycin (VWR, Radnor, PA) and 40 µg/mL tetracycline (Gold Biotechnology, Inc., St. Louis, MO) using the PureLink HiPure Plasmid MidiPrep kit (Invitrogen, Thermo Fisher Scientific). Sequences near the AgeI and KsaI cloning sites were confirmed by Sanger sequencing in both directions as described above. The modified, synthetic SARS CoV-2 S gene was ligated into the EcoRI/SalI sites of dephosphorylated, gel-purified, RGD4C-AAVPΔEcoRI829 digested with EcoRI-HF and SalI-HF (New England Biolabs) using a 1:3 vector:insert ratio and a vector mass of 15 ng. Ligation products were transformed into animal-free, electrocompetent MC1061 F⁻ *E.coli* and plated onto animal-free LB agar containing 100 µg/mL streptomycin and 40 µg/mL tetracycline (MilliporeSigma). Positive clones were verified by restriction mapping and confirmed by Sanger sequencing of purified dsDNA as described above. The 84 bp transgene null sequence containing the upstream AAVP human interferon leader sequence ending with a stop codon was synthesized at IDT (San Diego, CA) as a G-block, amplified by PCR, digested with EcoRI-HF and SalI-HF, subcloned into dephosphorylated, gel purified RGD4C-AAVP-TNFΔEcoRI829 and transformed into electrocompetent MC1061 F⁻ to produce the RGD4C-AAVP-transgene null genome. Transformed colonies were screened by colony PCR and restriction mapping. The transgene null insert was verified in putative positive clones by Sanger sequencing in both directions as described above. A single transformed RGD4C-AAVP SARS CoV-2 S/MC1061 F⁻ or RGD4C-AAVP SARS-CoV-2 S-null/MC1061 F⁻ colony was used to inoculate 10 mL of animal-free LB Broth, pH 7.4 containing 100 µg/mL streptomycin and 40 µg/mL tetracycline and grown to mid-log phase at 37 °C and 250 rpm in the dark. One milliliter of each mid-log phase pre-culture was used to inoculate 750 mL of animal-free LB Broth, pH 7.4 containing 100 µg/mL streptomycin and 40 µg/mL tetracycline in a sterile 2 L shaker baffle flasks for phage amplification at 30 °C, 250 rpm for 20 h in the dark. Phage were precipitated twice in sterile 16.7% PEG 8000/3.3 M NaCl (15% v/v), and the final phage pellet was resuspended in 1 mL sterile phosphate-buffered saline, pH 7.4, centrifuged to remove residual bacterial debris and filtered sterilized through a 0.2 µm syringe filter. Infective phage titers [(TU)/µL] were determined by infecting K91kan *E.coli* bacteria grown in animal-free terrific broth with 1E10⁷, 1E10⁸ or 1E10⁹ diluted phage, plating the infected bacteria onto animal-free LB agar plates containing 100 µg/mL kanamycin sulfate and 40 µg/mL tetracycline and counting bacterial colonies the following day.

Immunization Studies in Mice

Swiss Webster or BALB/C mice were randomized in groups of 3 to 12 animals. Group size was calculated based on statistical considerations to yield sufficient statistical significance.

The animals were inoculated with 10^9 TU phage or AAVP constructs intraperitoneally, intravenously, intratracheally or subcutaneously. For subcutaneous injections, 10^9 TU phage or AAVP particles were administered with 100 μ L on the front and hind limbs, and behind the neck (~ 20 μ L per site). For intratracheal vaccination, 10^9 TU of single-, dual-display phage particles or negative control insertless phage particles were administered in two serial doses in 50 μ L of PBS with a MicroSprayer® Aerosolizer coupled to a high-pressure syringe (Penn-Century) and a small animal laryngoscope (Penn-Century). The devices were used to administer air-free liquid aerosol directly into the trachea of animals deeply anesthetized with 1% isoflurane (22). For the tail vein blood collections, mice were locally anesthetized with a topic solution. On day 0, blood samples were collected for the baseline, followed by consecutive blood collection every 1-2 weeks post-immunization. Endotoxin removal was performed for each phage or AAVP preparation prior to administration of each dose, regardless of the route of administration. Purified phage or AAVP containing endotoxin was mixed with 10% Triton X-114 in endotoxin-free water, incubated on ice for 10 min, warmed to 37°C degrees for 10 min followed by separation of the Triton X-114 phase by centrifugation at 14,000 rpm for 1 min. The upper aqueous phase containing phage was withdrawn into a sterile microcentrifuge tube. This process was repeated from 3-5 times followed by PEG-NaCl precipitation. Resuspended phage was sterile filtered through 0.2 μ m (Pall Corporation) syringe filters. The levels of endotoxin were measured using the Limulus Amebocyte Lysate (LAL) Kinetic- QCL kit (Lonza). Phage or AAVP preparations containing endotoxin levels < 0.05 EU/mL were used in this study.

Serological Analysis

ELISA assays were performed in 96-microwell plates coated with 150 ng/well of SARS-CoV-2 Spike (aa 16-1213) His-tagged recombinant protein (ThermoFisher) and 10^{10} phage or AAVP particles/50 μ L of PBS O.N. at 4°C (Nunc MaxiSorp flat bottom, ThermoFisher Scientific). Coated plates were blocked with PBS containing 5% low-fat milk and 1% BSA for 1 h at 37°C. Two-fold serial dilutions (starting at 1:32) or 1:50 fixed dilution of sera in blocking buffer were added to separate the wells and incubated for 1-2 h at 37°C. Following three washes with PBS and PBS containing 0.1% of Tween 20, bound antibodies were detected with an anti-mouse IgG HRP-conjugated (Jackson ImmunoResearch) at optical density (OD) at 450 nm. Commercially available polyclonal IgG anti-Spike protein antibody (Thermo Fisher, MA5-35949) or anti-fd bacteriophage antibody (Sigma Aldrich) served as positive controls.

RNA Isolation and Quantitative Real-Time PCR. Total RNA from mice tissues were obtained with the RNeasy Mini Kit (Qiagen). First-strand cDNA synthesis was carried out

with the ImProm-II Reverse Transcription System (Promega). Quantitative real-time PCR analysis was performed in a QuantStudio 5 Real-Time PCR System (Applied Biosystems). Primers and TaqMan probes were as follows: fwd 5' GCTTTTCAGCTCTGCATCGTT 3' and rev 5' GACTAGTGGCAATAAAACAAGAAAAACA 3, customized AAVP S 6FAM 5' TGGGTTCTCTTGGCATGT 3' NFQ, Mm04277571_s1 for 18S, and Mm99999915_g1 for Gapdh. The gene expression ratio was normalized to that of 18S.

Statistical Analysis

Differences between groups were tested for statistical significance with Student's *t*-test or analysis of variance (one-way or two-way ANOVA) using GraphPad Prism 8. Statistical significance was set at $P < 0.05$.

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Author contribution

D.I.S., F.H.F.T., C.M., V.J.Y., F.I.S., E.D.R., V.G.C., D.D., P.O'B., N.H., T.L.S., R.L.S., P.C.W., J.N.O., W.A., and R.P. designed the research; D.I.S., F.H.F.T., C.M., V.J.Y., F.I.S., E.D.R., V.G.C., D.D., P.O'B., N.H., T.L.S., N.B., M.L.G. and P.C.W. performed the research; E.C.L., S.K.L., S.K.B., J.N.O. contributed new reagents/analytic tools; D.I.S., F.H.F.T., C.M., V.J.Y., F.I.S., E.D.R., V.G.C., D.D., P.O'B., N.H., T.L.S., N.B., R.L.S., M.L.G., P.C.W., J.N.O., W.A. and R.P analyzed the data; D.I.S., F.H.F.T., C.M., V.J.Y., F.I.S., V.G.C., E.D.R., P.C.W, J.N.O., W.A. and R.P. wrote the initial draft of the manuscript, to which all of the authors contributed with edits. J.N.O., W.A. and R.P. supervised the overall project.

Conflict of interest statement

D.I.S., C.M., S.K.L., W.A., and R.P. are inventors on patent applications related to this technology. PhageNova Bio has licensed this intellectual property and the inventors are entitled to standard royalties. R.P., S.K.L., and W.A. are founders and equity stockholders of PhageNova Bio. S.K.L. is a board member and R.P. is Chief Scientific Officer and a paid consultant of PhageNova Bio. R.P. and W.A. are founders and shareholders of MBrace Therapeutics; R.P. serves as Chief Scientific Officer and W.A. is a board member at MBrace Therapeutics. These arrangements are managed in accordance with the established institutional conflict of interest policies of Rutgers, The State University of New Jersey.

Figure Legends

Fig 1. Schematic representation of the phage- and AAVP-based vaccine candidates.

The scheme represents the approach used for the conception, design, and application of two strategies of immunization against SARS-CoV-2 S protein using phage particles. Step 1: Structural analysis, selection of structurally-defined epitopes, and cloning steps for the generation of dual-display phage particles and AAVP encoding the full-length S protein. Step 2: Molecular engineering of single- and dual-display phage particles, and AAVP S constructs. Step 3: Functional validation and vaccination studies *in vivo* in mice.

Fig 2. Identification of structural epitopes on S protein trimer. (A) Six epitopes (orange) spanning the SARS-CoV-2 S protein were selected for display on rpVIII protein. Four epitopes are located within the S1 subunit: epitope 1 (aa 336–361), epitope 2 (aa 379–391), epitope 3 (aa 480–488), epitope 4 (aa 662–671); and two within the S2 subunit: epitope 5 (aa 738–760), and epitope 6 (aa 1032–1043). These epitopes are solvent-exposed in the surface representation of the predominantly closed-state conformation of the S protein trimer (gray, cornflower blue, and rosy brown) (PDB ID: 6ZP0) (60). Only epitope 1 (aa 336–361) contains a site for glycosylation (at N343) (purple). (B) All of the epitopes (orange) maintain a cyclic conformation in the ribbon representation of a S protein protomer (gray); disulfide bridges (yellow) are present between the flanking cysteine residues of all epitopes except on epitope 2 (aa 379–391). The open-state conformation of the S protein trimer with one RBD erect displays a change in orientation of epitopes 1, 2, and 3, though all remain solvent-exposed (PDB ID: 6ZGG) (60).

Fig 3. Explicit-solvent simulations reveal the near-native conformation of epitope 4 relative to the full-length S protein. (A) RMSD from the S protein conformation for each epitope within the S1 subunit: epitope 1 (aa 336–361), epitope 2 (aa 379–391), epitope 3 (aa 480–488) and epitope 4 (aa 662–671). Epitope 4 frequently samples low values ($\sim 2\text{\AA}$), while the other epitopes undergo substantial conformational rearrangements ($\text{RMSD} < 4\text{\AA}$). (B) Probability as a function of RMSD shows that epitope 4 is most likely to sample conformations that are similar ($\text{RMSD} \sim 2.5\text{\AA}$) to the S protein conformation. (C) RMSD as a function of time for the epitopes within the S2 subunit: epitope 5 (aa 738–760) and epitope 6 (aa 1032–1043). Both epitopes rapidly adopt and maintain large RMSD values (D).

Fig 4. Immunogenicity of S protein epitopes on single-display phage particles. Five-week-old female Swiss Webster mice were immunized via subcutaneous injection with single-display phage constructs containing each of the six different epitopes expressed on rpVIII protein or the control insertless phage. Animals received a boost injection three weeks

after the first administration. (A) S protein-specific IgG antibodies and (B) phage-specific IgG antibodies were evaluated in sera of mice after two- and five-weeks by ELISA (n=3 mice per group). (C) Five-week-old female BALB/c mice were immunized via intratracheal administration with the epitope 4/CAKSMGDIVC dual-display phage particles, epitope 4 single-display phage particles, or the control insertless phage. Animals received a boost three weeks after the first administration. S protein-specific IgG antibodies were weekly evaluated by ELISA (n=10 mice per group). Data represent $\pm$ SEM (* $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$).

Fig 5. Immunogenicity of the RGD4C-AAVP S in mice. Schematic representation of the AAVP-based vaccine candidate. (A) The modified, synthetic SARS-CoV-2 S protein gene was excised from the pUC57 and cloned into the RGD4C-AAVP -TNFΔEcoRI829. Expression of the S protein transgene cassette is driven by the cytomegalovirus (CMV) promoter and flanked by AAV ITRs. (B) Schematic representation of the RGD4C-AAVP S and the control RGD4C-AAVP empty vector (RGD4C AAVP *S-null*). (C) S protein-specific IgG antibody response in the sera of mice immunized with RGD4C-AAVP S via different routes of administration (n=5 mice per group) by ELISA. (D) S protein-specific IgG antibodies in sera of mice weekly immunized with RGD4C-AAVP S or the control RGD4C-AAVP *S-null* (n=12 mice per group) via subcutaneous administration. Data represent $\pm$ SEM (** $P < 0.01$). (E) Tissue-specific expression of the S protein transgene in mice immunized with AAVP S five weeks after the first immunization. Data represent $\pm$ SEM (*** $P < 0.001$). (F) Phage-specific IgG antibody response in the sera of mice immunized with RGD4C-AAVP S via different routes of administration (n=5 mice per group). Phage-specific IgG antibody response in mice immunized with RGD4C-AAVP S (G) or RGD4C-AAVP *S-null* (H) at two and five weeks after the first immunization. Phage-specific IgG antibody response was evaluated by ELISA in 96-well plates coated with 10^{10} AAVP particles per well. Tet^R, tetracycline resistance gene. Amp^R, ampicillin resistance gene. Ori origin of replication.

Supplemental Figures

Fig S1. S protein epitope mapping from peer-reviewed publications. Primary sequence representation of immunogenic regions of SARS-CoV-2 S protein spanning S1 and S2 subunits identified by B-cells, T-cells, and antibody screenings on patients with COVID-19 (also listed in Table S1). The six structurally-selected epitopes displayed on the rpVIII protein are highlighted (orange).

Fig S2. The single- and dual-display phage particles cloning strategy. To generate the single-display phage particles, the f88-4 phage vector was used. This vector contains two genes encoding for the major capsid protein pVIII: the wild-type (pVIII, depicted in grey) and the recombinant (rpVIII, depicted in green). rpVIII contains a foreign DNA insert between the HindIII and PstI cloning site, which allows the cloning of annealed oligonucleotides encoding the S protein epitopes in-frame with the rpVIII gene. For the dual-display phage particles, the f88-4 vector containing the epitope 4 (CDIPGAGIC) and the fUSE55 phage vector are digested with BamHI and XbaI restriction enzymes. The digestion products are then purified and fused according to a standard ligation protocol. The result is a chimeric vector (f88-4/fUSE55). Next, oligonucleotide encoding for the CAKSMGDIVC targeting motif was cloned within the SfiI restriction site of the pIII gene (pIII, depicted in light blue), generating the dual-display phage vector, which contains epitope 4 on the rpVIII protein and the CAKSMGDIVC motif on the pIII protein. Both, single- or dual-display phage particle vectors were used to transform electrocompetent DH5α *E. coli*. Phage particles were produced in k91Kan *E. coli*. Tet^R, tetracycline resistance gene.

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Table S1: Cross-reference analysis of epitope mapping of immunogenic regions of SARS-CoV-2 S protein.

Authors	Title	Residues
Amrun <i>et al.</i>	Linear B-cell epitopes in the spike and nucleocapsid proteins as markers of SARS-CoV-2 exposure and disease severity	209–226 553–570 769–786 809–826
Farrera-Soler <i>et al.</i>	Identification of immunodominant linear epitopes from SARS-CoV-2 patient plasma	655–672 787–822 1147–1158
Li <i>et al.</i>	Linear epitopes of SARS-CoV-2 spike protein elicit neutralizing antibodies in COVID-19 patients	553–564 577–588 595–612 625–642 661–684 764–829 1148–1159
Noy-Parat <i>et al.</i>	A panel of human neutralizing mAbs targeting SARS-CoV-2 spike at multiple epitopes	369–386
Peng <i>et al.</i>	Broad and strong memory CD4+ and CD8+ T cells induced by SARS-CoV-2 in UK convalescent individuals following COVID-19	166–180 751–765 801–815 866–880
Poh <i>et al.</i>	Two linear epitopes on the SARS-CoV-2 spike protein that elicit neutralising antibodies in COVID-19 patients	553–570 809–826

Shrock <i>et al.</i>	Viral epitope profiling of COVID-19 patients reveals cross-reactivity and correlates of severity	406–417*
		414–427
		418–430*
		424–428*
		438–448*
		454–463
		459–467*
		478–488*
		504–507*
		514–518*
Zhang <i>et al.</i>	Mining of epitopes on spike protein of SARS-CoV-2 from COVID-19 patients	551–570
		766–785
		811–830
		1144–1163
		21–45
		221–245
		261–285
		330–349
		370–394
		375–394
		405–469
		450–469
		480–499
		495–521
		522–646**
		902–926

Note: All amino acid ranges aligned to Wuhan-Hu-1 strain (GenBank Accession number: NC_045512.2).

* Extrapolated from Shrock *et al.* Fig. 7 since not explicitly stated in text.

** Not included in Fig. S1.

Phage- and AAVP-based vaccine candidates

Ligand-directed dual-display phage vaccine

Ligand-directed AAVP vaccine

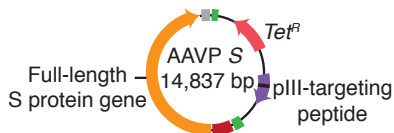
Protein structure analysis

Full-length S protein transgene cloning into the AAVP vector

Selection of S protein epitopes

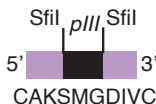
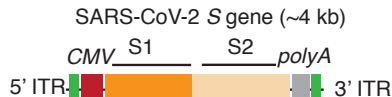
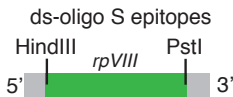
Sequencing and genomic validation

rpVIII-S epitopes



Solvent-exposed cyclic epitopes (n=6)

Validation of AAVP S or AAVP S-null



Vaccination studies *in vivo*

10⁹ TU of phage or AAVP particles/dose

Multiple routes of administration

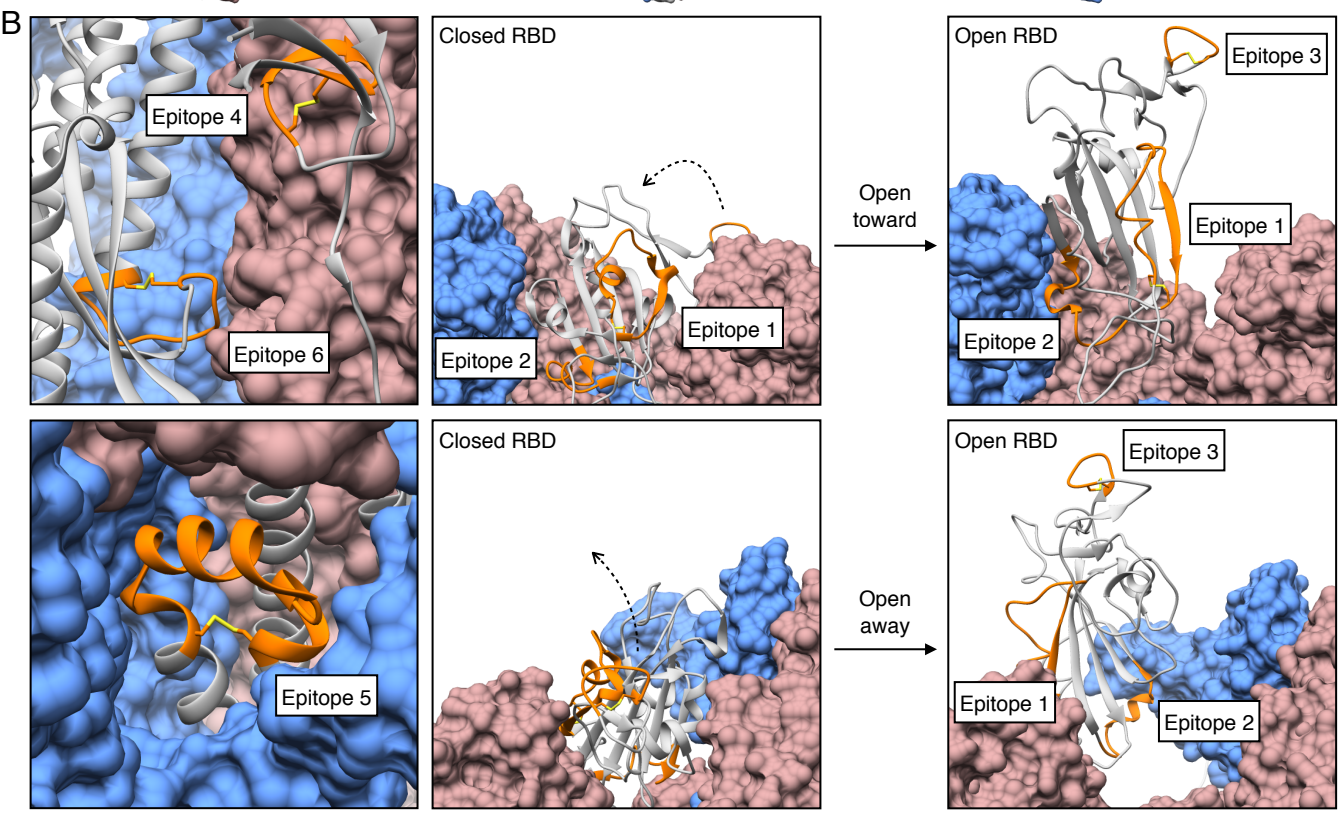
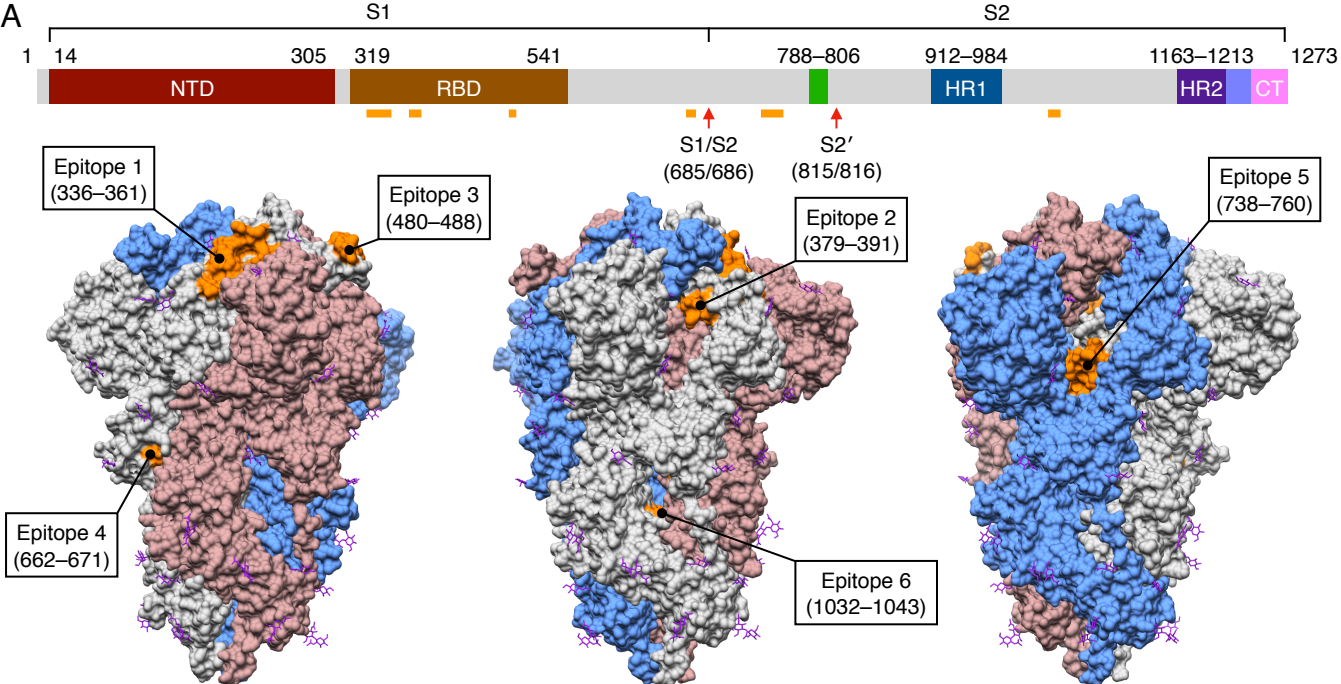


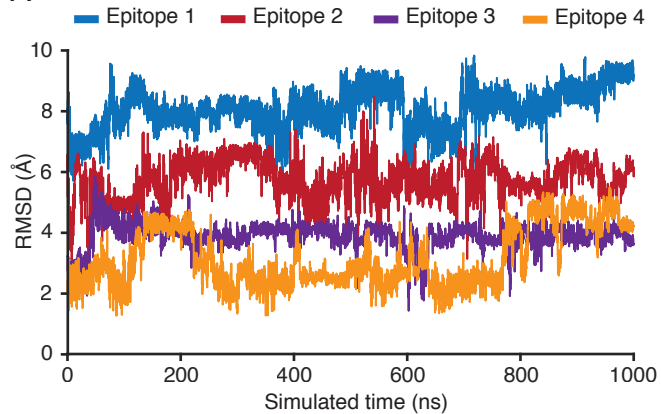
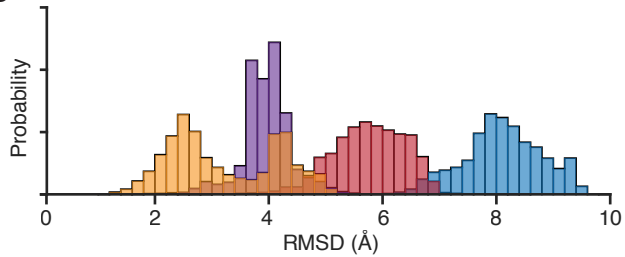
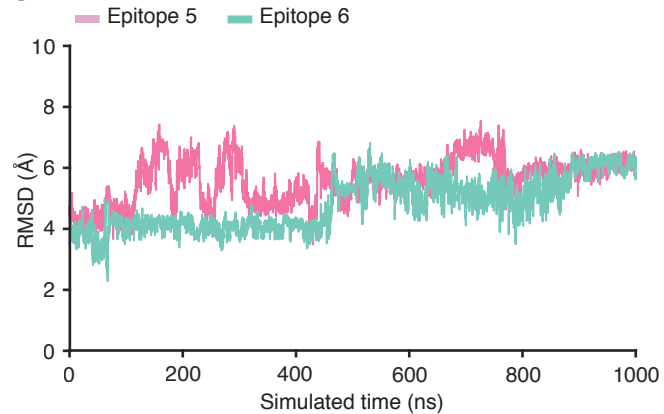
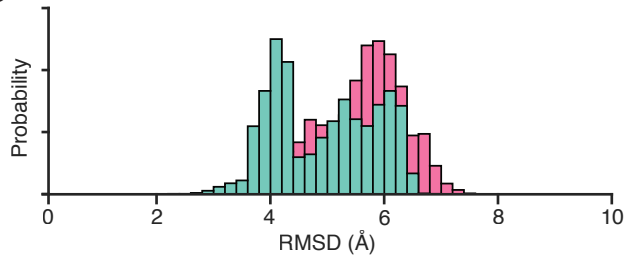
First dose

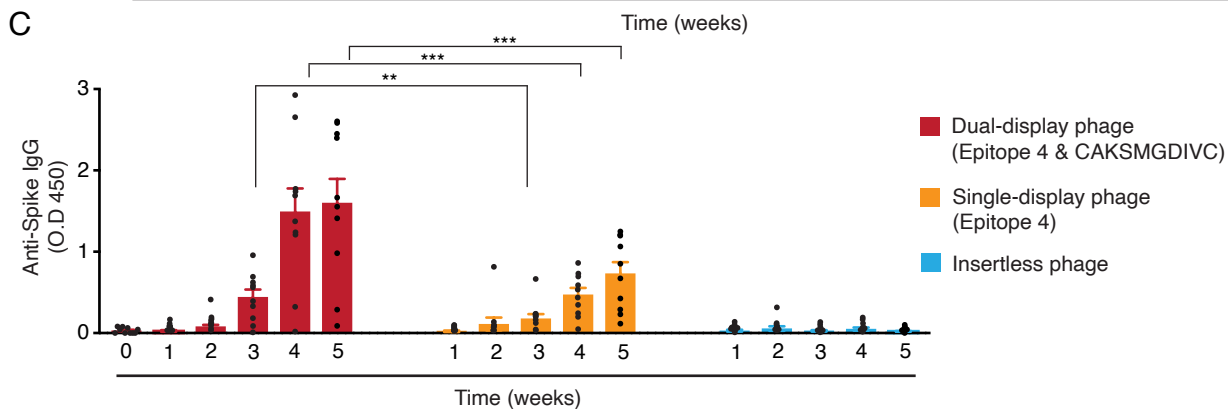
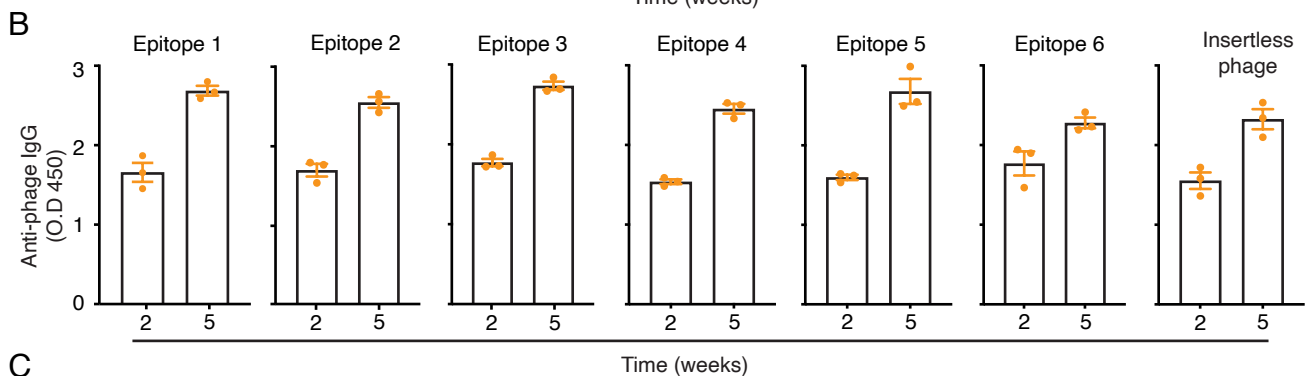
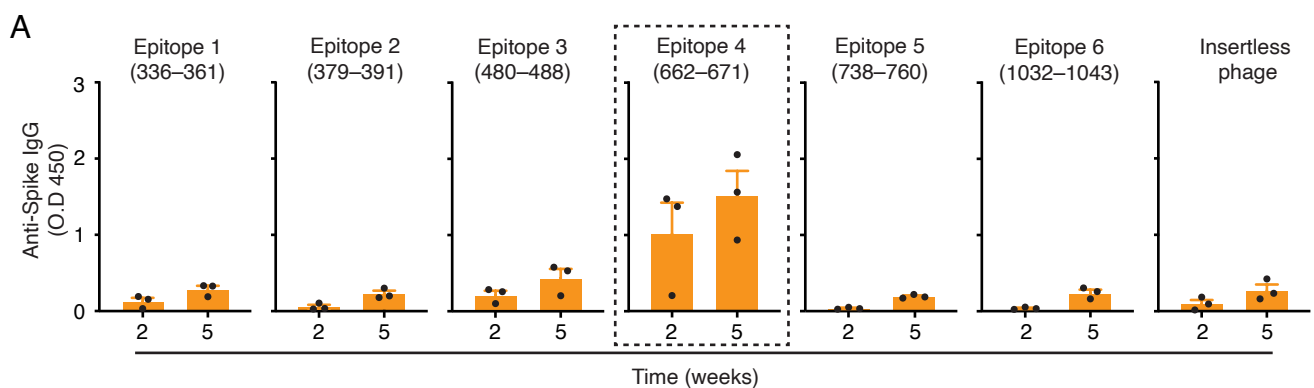
Second dose

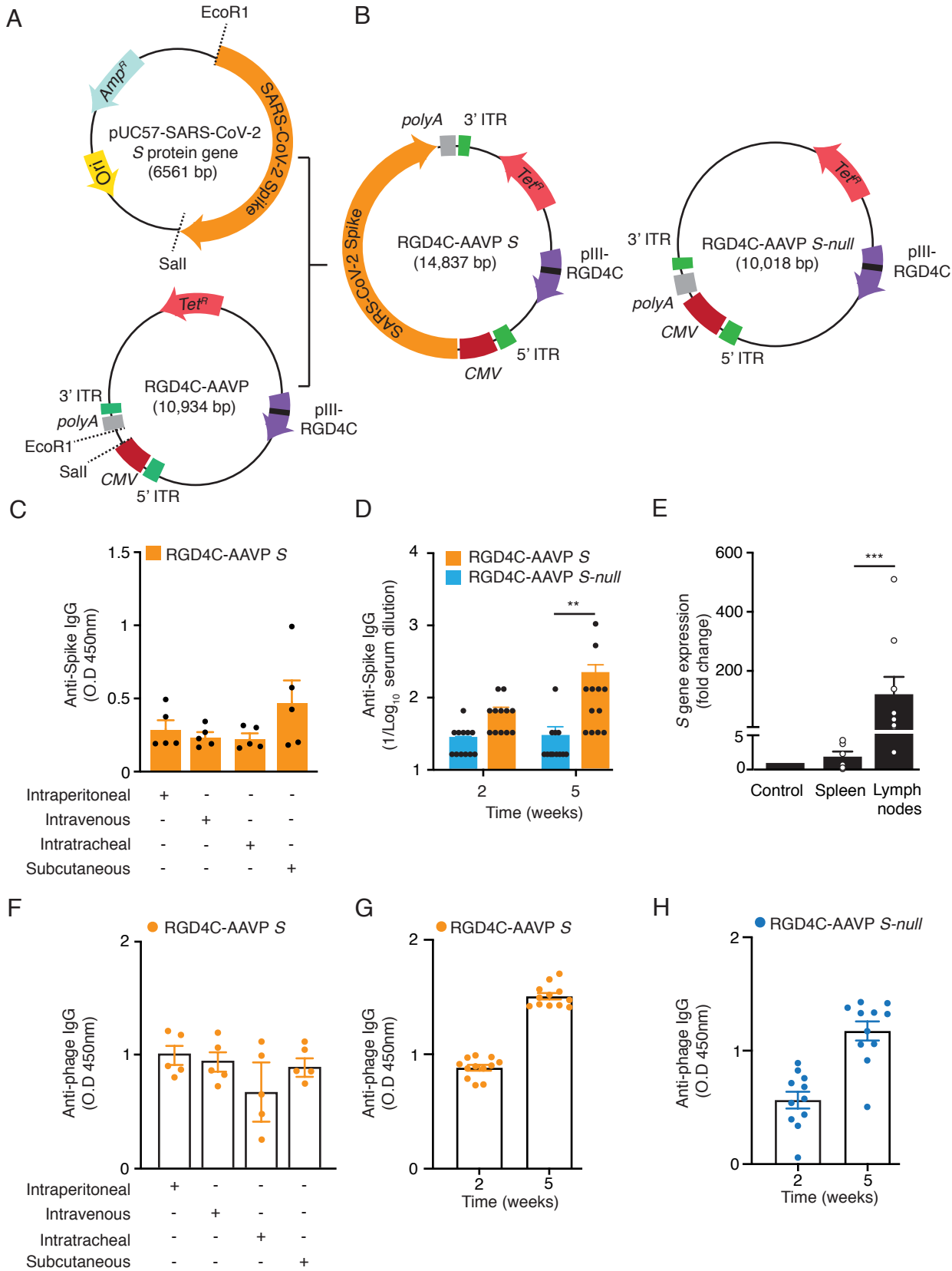
Time(weeks)

Serum collection and antibody detection



A**B****C****D**





19 TTRTQLPPAYTNSFTRGVYYPDKVFRSSV 47 164 NNCTFEYVVSQPFILMDLEGK 182
RTQLPPAYTNSFTRGVYYPDKVFRS CTFEYVVSQPFILMDLE

207 HTPINLVRDLPQGFSALEPLVDLPIGINITRFQTLALLHRSYLTPGDSSSGWTAGAAAYYVGYLQPRTFLLKYNENGTITD 287
PINLVRDLPQGFSALEPL GAAAYYVGYLQPRTFLLKYNENGTI
SALEPLVDLPIGINITRFQTLALLH

328 RFPNITNLCPFGGEVFNATRFASVYAWNRRKRISNCVADYSVLYNSASFSTFKCYGVSPTKLNDLCFTNVY 396
PNITNLCPFGGEVFNATRFAS YNSASFSTFKCYGVSPTK
NSASFSTFKCYGVSPTKLNDLCFTN
STFKCYGVSPTKLNDLCFTN

403 RGDEVQRQIAPGGQTGKIADYNYKLPDDFTGCVIAWNSNNLDSKVGGNYNYLYRLFRKSNLKPFFERDISTE 471
DEVQRQIAPGGQTGKIADYNYKLPDDFTGCVIAWNSNNLDSKVGGNYNYLYRLFRKSNLKPFFERDIS
EVQRQIAPGGQTGK KLPDD SNNLDSKVGGN NYLYRLFRKSNLKPFFERDIS
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IADYNYKLPDDFT SNLKPFFERD

476 GSTPCNGVEGFNCYFPLQSYGFQPTNGVGYPYRVVLSFELLHAPAT 523
TPCNGVEGFNC YGFQPTNGVGYPYRVVLSFELLHAP
CNGVEGFNCYFPLQSYGFQ GYQP SFELL

549 TGVLTESNKKFLPFQQFGRDIAIDTTDAVRDPQTLEILDITPCSFGGVSVITPGTNTSNQVAVLY 614
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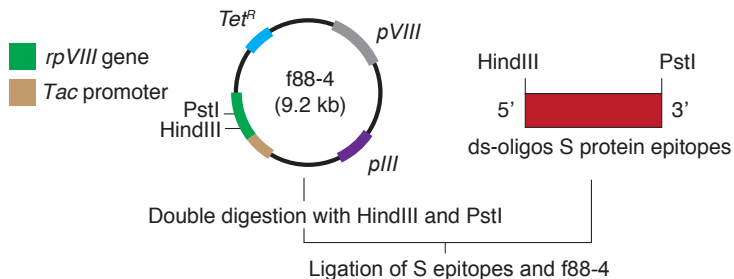
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PSKPSKRSFIEDLLFNKV
PSKPSKRSFIEDLLFNKV
KPSKRSFIEDLLFNKVTLAD

864 LLTDEMIAQYTSALLAGTITSGWTFGAGAALQIPFAMQMAYRFNGIGVTQNVLYENQKLIANQFN 928
TDEMIAQYTSALLAG MAYRFNGIGVTQNVLYENQKLIANQ

1142 QPELDSFKEELDKYFKNHTSPDVD 1165
ELDSFKEELDKYFKNHTSPD
SFKEELDKYFKN
FKEELDKYFKN

Filamentous phage display vector



Filamentous phage display vector

