

1 **16S rRNA Gene Amplicon Profiling of the New Zealand parasitic blowfly** 2 ***Calliphora vicina*.**

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6 **ABSTRACT**

7 Here, we present a 16S rRNA gene amplicon sequence data set and profiles demonstrating the bacterial diversity
8 of larval and adult *Calliphora vicina*, collected from Ashhurst, New Zealand (May 2020). The three dominant
9 genera among adult male and female *C. vicina* were *Serratia* and *Morganella* (phylum *Proteobacteria*) and
10 *Carnobacterium* (phylum *Firmicutes*), while the larvae were also dominated by the genera *Lactobacillus* (phylum
11 *Firmicutes*).

12 **ANNOUNCEMENT**

13 Ectoparasitic flies (blowflies) are a significant animal welfare and production issue for farmers worldwide (1).
14 Control of blowflies is problematic because the flies are unpredictable and highly mobile, and strike (or myiasis)
15 is difficult to see initially but has an immediate impact on animal production and welfare. Currently control relies
16 heavily on the prophylactic application of long-acting chemicals to all sheep, but this approach is increasingly
17 under threat due to development of resistance to current treatments (2, 3). *Calliphora vicina* NZ_CalVic_NP (4,
18 5) was selected for microbiome assessment as a representative of a New Zealand field strain of *C. vicina*. In this
19 study, we have investigated the larval, adult male and adult female bacterial microbial profiles of *C. vicina* to gain
20 a better understanding of the microbial communities of blowflies targeting the development of new interventions
21 such as probiotics, bioactive compounds, vaccines or insecticides.

The *C. vicina* specimen larvae were collected from a farm site in Ashhurst area in New Zealand (40°18' S, 175°45' E). Lab reared blowflies were maintained on beef liver as protein source and a 10% sugar solution, with the procedures for blowfly propagation and sample preparation were based done according to Dear J.P. (1985). To remove surface adherent bacteria from lab reared *C. vicina*, pools of larvae, entire adult male and adult female were separated and washed twice in sterile phosphate-buffered saline (PBS, pH 7.4), snap frozen in liquid nitrogen, and transferred to -80 °C storage prior to DNA extraction. Genomic DNA for metagenomic 16S rRNA gene amplicon sequencing of the V3-V4 hypervariable region was isolated from *C. vicina* pooled samples of 100 larvae as well as 10 entire adult males and females per replicate ($n=5$ for each). High molecular weight genomic DNA was prepared using a modified phenol:chloroform protocol recently applied to difficult samples such as parasitic roundworms (7, 8), fastidious anaerobic rumen bacteria (9-11) and spore-forming psychrotolerant *Clostridium* isolated from spoiled meat (12, 13). A DNA library was prepared using the Illumina 16S V3-V4 rRNA library preparation method (Illumina, Inc., San Diego, CA) according to the manufacturer's instructions (20), and sequenced on the Illumina MiSeq platform with the 2× 250 bp paired-end (PE) reagent kit v2 producing a total of 3,017,007 PE raw reads.

The processing of the amplicon reads followed a modified form of the pipeline described in (21). The reads produced by the sequencing instrument were paired using the program FLASH2 (22). Paired reads were then quality trimmed using Trimmomatic 0.38 (23). The trimmed reads were reformatted as fasta, and the read headers were modified to include the sample name. All reads were compiled in a single file, and the Mothur (24) program suite was used to remove reads with homopolymers longer than 10 nt and to collapse the reads into unique representatives. The collapsed reads were clustered using the program Swarm (25). The clustered reads were filtered based on their abundance, keeping representatives that were a) present in one sample with a relative abundance >0.1%, b) present in >2% of the samples with a relative abundance >0.01% or c) present in 5% of the samples at any abundance level. The selected representatives were annotated using Qiime (26) with the Silva database v138 (27). The annotated tables were then used for downstream statistical analysis. The predominant

phyla in all samples were Proteobacteria (Fig. 1) and at the genus level, *Serratia*, *Morganella* and *Carnobacterium*, while the larvae were also dominated by *Lactobacillus* (phylum Firmicutes).

The metagenomic 16S rRNA gene amplicon sequencing of *C. vicina* field strain NZ_CalVic_NP reported here is a valuable resource for future studies investigating the bacterial genetic mechanisms associated with flystrike. Management of flystrike in a world increasingly demanding fewer inputs of synthetic chemicals to food producing animals will be challenging. Equally, this research is important owing to the diminished efficacy demonstrated by current blowfly treatments due to the emergence of resistance in blowflies against many classes of insecticides.

Data availability. The 16S rRNA gene amplicon sequence data have been deposited in the GenBank Sequence Read Archive (SRA) under the BioProject accession number [PRJNA667961](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA667961).

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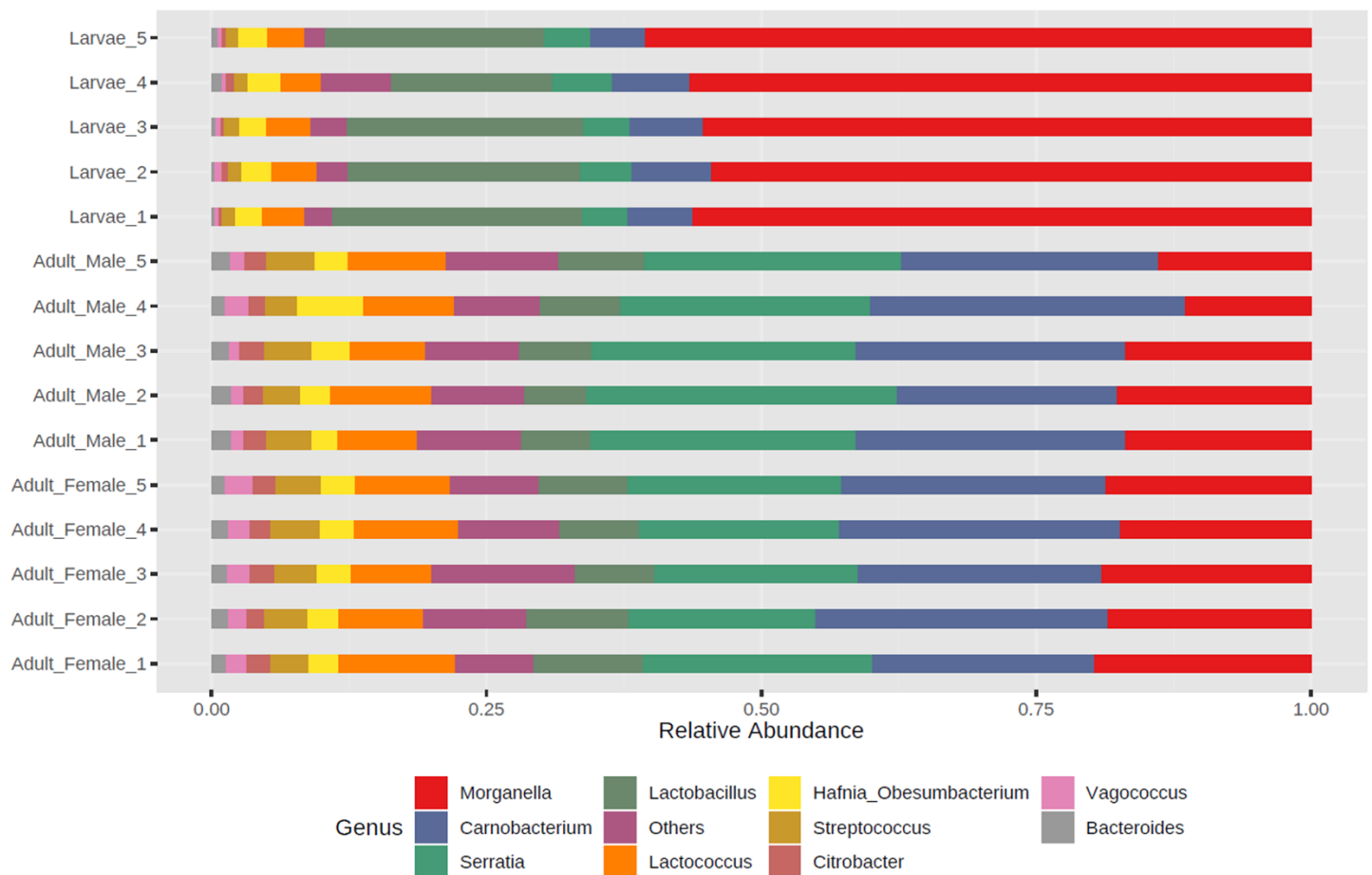


FIG 1. The taxonomic composition of the dominant bacteria of New Zealand *C. vicina*. Relative abundance of the dominant bacterial genera obtained from 16S rRNA sequencing of *C. vicina* field strain NZ_CalVic_NP larvae, adult males and female samples. Genera with a relative abundance of less than 1% and unassigned amplicon sequence variants were grouped together as Others.

REFERENCES

1. **Githeko AK, Lindsay SW, Confalonieri UE, Patz JA.** 2000. Climate change and vector-borne diseases: a regional analysis. *Bulletin of the World Health Organization* **78**:1136-1147.
2. **Hall M, Wall R.** 1995. Myiasis of humans and domestic animals. *Advances in parasitology* **35**:257-334.
3. **Fischer O, Matlova L, Dvorska L, Svastova P, Bartl J, Weston R, Pavlik I.** 2004. Blowflies *Calliphora vicina* and *Lucilia sericata* as passive vectors of *Mycobacterium avium* subsp. *avium*, *M. a. paratuberculosis* and *M. a. hominissuis*. *Medical and veterinary entomology* **18**:116-122.
4. **Palevich N, Carvalho L, Maclean P.** 2020. Characterization of the complete mitochondrial genome of the New Zealand parasitic blowfly *Calliphora vicina* (Insecta: Diptera: Calliphoridae). *bioRxiv* doi:10.1101/2020.11.05.370361:2020.2011.2005.370361.
5. **Palevich N, Carvalho, Luis, Maclean, Paul.** 2020. The Complete Mitochondrial Genome of the New Zealand Parasitic Blowfly *Lucilia sericata* (Insecta: Diptera: Calliphoridae). Preprints **2020100601**.
6. **Dear JP.** 1986. Calliphoridae (Insecta: Diptera). *Fauna of New Zealand*, **8**: 88.
7. **Palevich N, Maclean PH, Baten A, Scott RW, Leathwick DM.** 2019. The Genome Sequence of the Anthelmintic-Susceptible New Zealand *Haemonchus contortus*. *Genome biology and evolution* **11**:1965-1970.
8. **Palevich N, Maclean PH, Choi Y-J, Mitreva M.** 2020. Characterization of the Complete Mitochondrial Genomes of Two Sibling Species of Parasitic Roundworms, *Haemonchus contortus* and *Teladorsagia circumcincta*. *Frontiers in Genetics* **11**.
9. **Palevich N, Kelly WJ, Ganesh S, Rakonjac J, Attwood GT.** 2019. *Butyrivibrio hungatei* MB2003 Competes Effectively for Soluble Sugars Released by *Butyrivibrio proteoclasticus* B316^T during Growth on Xylan or Pectin. *Applied and Environmental Microbiology* **85**:e02056-02018.
10. **Palevich N, Kelly WJ, Leahy SC, Denman S, Altermann E, Rakonjac J, Attwood GT.** 2019. Comparative genomics of rumen *Butyrivibrio* spp. uncovers a continuum of polysaccharide-degrading capabilities. *Applied and environmental microbiology* **86**.
11. **Palevich N, Maclean PH, Kelly WJ, Leahy SC, Rakonjac J, Attwood GT.** 2020. Complete Genome Sequence of the Polysaccharide-Degrading Rumen Bacterium *Pseudobutyrvibrio xylanivorans* MA3014 Reveals an Incomplete Glycolytic Pathway. *Genome Biology and Evolution* **12**:1566-1572.
12. **Palevich N, Palevich FP, Maclean PH, Altermann E, Gardner A, Burgess S, Mills J, Brightwell G.** 2021. Comparative genomics of *Clostridium* species associated with vacuum-packed meat spoilage. *Food Microbiology* **95**:103687.
13. **Palevich N, Palevich FP, Maclean PH, Jauregui R, Altermann E, Mills J, Brightwell G.** 2019. Draft genome sequence of *Clostridium estertheticum* subsp. *laramiense* DSM 14864^T, isolated from spoiled uncooked beef. *Microbiology resource announcements* **8**.
14. **Kozich JJ, Westcott SL, Baxter NT, Highlander SK, Schloss PD.** 2013. Development of a dual-index sequencing strategy and curation pipeline for analyzing amplicon sequence data on the MiSeq Illumina sequencing platform. *Applied and environmental microbiology* **79**:5112-5120.
15. **Camarinha-Silva A, Jáuregui R, Pieper DH, Wos-Oxley ML.** 2012. The temporal dynamics of bacterial communities across human anterior nares. *Environmental microbiology reports* **4**:126-132.
16. **Magoč T, Salzberg SL.** 2011. FLASH: fast length adjustment of short reads to improve genome assemblies. *Bioinformatics* **27**:2957-2963.
17. **Bolger AM, Lohse M, Usadel B.** 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* **30**:2114-2120.
18. **Schloss PD, Westcott SL, Ryabin T, Hall JR, Hartmann M, Hollister EB, Lesniewski RA, Oakley BB, Parks DH, Robinson CJ.** 2009. Introducing mothur: open-source, platform-independent, community-supported software for describing and comparing microbial communities. *Applied and environmental microbiology* **75**:7537-7541.

- 112 19. **Mahé F, Rognes T, Quince C, de Vargas C, Dunthorn M.** 2014. Swarm: robust and fast clustering method
113 for amplicon-based studies. *PeerJ* **2**:e593.
- 114 20. **Caporaso JG, Kuczynski J, Stombaugh J, Bittinger K, Bushman FD, Costello EK, Fierer N, Pena AG,**
115 **Goodrich JK, Gordon JL.** 2010. QIIME allows analysis of high-throughput community sequencing data.
116 *Nature methods* **7**:335-336.
- 117 21. **Quast C, Pruesse E, Yilmaz P, Gerken J, Schweer T, Yarza P, Peplies J, Glöckner FO.** 2012. The SILVA
118 ribosomal RNA gene database project: improved data processing and web-based tools. *Nucleic acids research*
119 **41**:D590-D596.