Microbiota analysis of rural and urban surface waters and sediments in Bangladesh identifies human waste as driver of antibiotic resistance

Ross Stuart McInnes[a], Md. Hassan uz-Zaman[b], Imam Taskin Alam[b], Siu Fung Stanley Ho[a], Robert A. Moran[a], John D. Clemens[b], Md. Sirajul Islam[b, #], Willem van Schaik[a, #].

[a] Institute of Microbiology and Infection, University of Birmingham, Birmingham, B15 2TT, United Kingdom.
[b] The Laboratory of Food Safety and One Health, Laboratory Sciences and Services Division, International Centre for Diarrhoeal Disease Research Bangladesh, Dhaka 1212, Bangladesh.

[#] Corresponding authors: sislam@icddrb.org; W.vanSchaik@bham.ac.uk

Running title: surface water microbiota in Bangladesh

Keywords: Resistome, Metagenome, Antibiotic Resistance, Plasmids, Aquaculture, Bangladesh

Word count abstract: 250

Word count text: 4,552

Abstract

In many low- and middle-income countries antibiotic resistant bacteria spread in the environment due to inadequate treatment of wastewater and the poorly regulated use of antibiotics in agri- and aquaculture. Here we characterised the abundance and diversity of antibiotic-resistant bacteria and antibiotic resistance genes in surface waters and sediments in Bangladesh through quantitative culture of Extended-Spectrum Beta-Lactamase (ESBL)-producing coliforms and shotgun metagenomics. Samples were collected from highly urbanised settings (n = 7), from rural ponds with a history of aquaculture-related antibiotic use (n = 11) and from rural ponds with no history of antibiotic use (n = 6). ESBL-producing coliforms were found to be more prevalent in urban samples than in rural samples. Shotgun sequencing showed that sediment samples were dominated by the phylum Proteobacteria (on average 73.8% of assigned reads), while in the water samples Cyanobacteria (on average 60.9% of assigned reads) were the predominant phylum. Antibiotic resistance genes were detected in all samples, but their abundance varied 1,525-fold between sites, with the highest levels of antibiotic resistance genes being present in urban surface water samples. We identified an IncQ1 sulphonamide resistance plasmid ancestral to the widely studied RSF1010 in one of the urban water samples. The abundance of antibiotic resistance genes was significantly correlated (R^2 = 0.73; P = 8.9 x 10^{-15}) with the abundance of bacteria originating from the human gut, which suggests that the release of untreated sewage is a driver for the spread of environmental antibiotic resistance genes in Bangladesh, particularly in highly urbanised settings.

Importance

Low- and middle-income countries (LMICs) have higher burdens of multidrug-resistant infections than high-income countries and there is thus an urgent need to elucidate the drivers of the spread of antibiotic-resistant bacteria in LMICs. Here we study the diversity and abundance of antibiotic resistance genes in surface water and sediments from rural and urban settings in Bangladesh. We found that urban surface waters are particularly rich in antibiotic resistance genes, with a higher number of them associated with plasmids indicating that they are more likely to spread horizontally. The abundance of antibiotic resistance genes was strongly correlated with the abundance of bacteria that originate from the human gut, suggesting that uncontrolled release of human waste is a major driver for the spread of antibiotic resistance in the urban environment. Improvements in sanitation in LMICs may thus be a key intervention to reduce the dissemination of antibiotic resistant bacteria.

Introduction

The prevalence of antibiotic-resistant bacteria causing infections is increasing globally, but the clinical issues, including significant morbidity and mortality, posed by these bacteria are particularly alarming in low- and middle-income countries (LMICs) (1–4). Proposed drivers for the high burden of drug-resistant infections in LMICs include the unregulated sales of antibiotics and their misuse in clinical medicine, agriculture and aquaculture, an inadequate sewerage infrastructure, poor governance and low investments in health care (5, 6).

One of the challenges of studying AMR is to disentangle the spread of resistant bacteria and antibiotic resistance genes between humans, animals and the wider environment (7). For this reason, AMR is increasingly being studied from a collaborative and cross-disciplinary perspective that has been termed 'One Health' (8). The One Health concept for studying the spread of AMR is particularly relevant for LMICs due to the crucially important role of agriculture and aquaculture in the livelihoods of billions of people in many of these countries, especially the poorest ones (9). Asia is home to an estimated 74% of the world's 570 million farms (10) and, in 2016, 89% of the global aquaculture production was estimated to originate from this continent (11). However, there are still major knowledge gaps on the spread of AMR in Asia from a One Health perspective.

Bangladesh is an LMIC in South Asia, where antibiotic-resistant infections are common among both hospitalised patients and the non-hospitalised population (12). The country has a number of unique characteristics that may contribute to the rapid spread of AMR. The capital city of Bangladesh, Dhaka, has a population of around 16 million people, with a population density that ranks among the highest of any megacity. Less than 20% of the households in Dhaka are connected to sewerage infrastructure (13), facilitating the spread of antibiotic-resistant bacteria via the environment. While a prescription is legally required to purchase antibiotics in Bangladesh, they can be readily acquired from many of the 200,000 drug stores across Bangladesh (14). In rural Bangladesh, aquaculture is widespread with more than 2 million tonnes of freshwater fish produced

in 2017 from inland freshwater fisheries (15). A recent survey revealed that antibiotics are widely used in Bangladeshi aquaculture for disease prevention and growth promotion. The most prominent classes of antibiotics employed are the tetracyclines, but other antibiotic classes including β-lactams and sulphonamides are also used (16). The use of antibiotics in Bangladesh is regulated in line with the European Union standards for antibiotic use in aquaculture, but Bangladesh has been found to be in breach of these regulations several times (17). The causes of antibiotics overuse in aquaculture are multifactorial: pharmaceutical companies provide food which is premixed with antibiotics without the farmers' knowledge, farmers administer antibiotics too often because they do not understand the instructions, and prophylactic use of antibiotics may be used to reduce the chance of damaging losses in production caused by disease (18). The combination of a densely populated country, intensive antibiotic usage in aquaculture and the potential for the dissemination of antibiotic-resistant bacteria through surface water thus provides a unique opportunity to study the spread of AMR from a One Health perspective.

In this manuscript, we use a combination of quantitative bacterial culture and metagenomic shotgun sequencing methods to disentangle pathways that contribute to the dissemination of antibiotic resistance. Specifically, we describe the abundance and diversity of microorganisms and antibiotic resistance genes in surface water in rural and urban settings in Bangladesh.

Results

Sample collection across urban and rural sites in Bangladesh

Freshwater surface water and sediment samples were collected from 24 sites across three districts in Bangladesh (Mymensingh, Shariatpur and Dhaka; Figure 1). These sites spanned both rural and urban areas with different population densities. Among rural sites, ponds used for aquaculture with a history of antibiotic use (n = 11) and ponds with no history of antibiotic use (n = 6) were sampled. Further information on sampling locations and protocols is provided in the Materials and Methods section and in Table S1. We used culture-dependent and culture-independent methods to study the abundance of antibiotic resistance genes and the diversity of microbiotas across the different sites.

ESBL-producing coliforms were more prevalent in urban samples than in rural samples

We quantitatively determined the burden of Extended Spectrum Beta-lactamase (ESBL) producing coliforms in the water and sediment samples from the different sampling locations and found that ESBL-producing coliforms were detected in significantly more urban samples (12/14) than rural samples (15/34) (Fisher exact test; P = 0.01). However, in samples that contained detectable levels of ESBL-producing coliforms there was no statistically significant difference in the viable counts of urban or rural samples (Figure 2).

Microbiotas of surface water and sediments are distinct with higher levels of human gut bacteria in urban samples.

Shotgun metagenomic sequencing was used to study the diversity and composition of the microbial communities in the different samples. An important determinant shaping the communities was the sample type, with distinct (PERMANOVA; $P < 0.001$) clustering of sediment and water samples (Figure 3A). The sediment samples were dominated by the phylum Proteobacteria (73.8%; standard deviation (SD) 27.1) while in the water samples Cyanobacteria (60.9%; SD 29.6) was the dominant phylum (Figure 3B). However, considerable variation in the composition of the microbial

communities was observed as in five of the nine sediment samples collected in Mymensingh, the abundance of Euryarchaeota was greater than 50%, while in five Dhaka water samples Proteobacteria were present at levels greater than 45%. Water sample WAM6 had very high levels (>60%) of bacteriophage DNA. The sediment samples were dominated by typical soil bacteria such as *Pseudomonas*, *Azoarcus* and *Anaeromyxobacter* while the water samples were dominated by cyanobacteria such as *Cyanobium*, *Microcystis* and other typical aquatic bacterial species from the phyla Proteobacteria and Actinobacteria. Three bacteriophages (*Mycobacterium* phage rizal, *Microcystis aeruginosa* phage Ma LMM01 and an Epsilon15-like virus) were also identified at different sampling sites. It was apparent that many of the Dhaka water samples contained bacteria which are typically found within the gastrointestinal tract, including *Escherichia coli, Streptococcus infantarius, Bifidobacterium adolescentis* and *Prevotella copri*. Through microbial source tracking analysis of our shotgun sequencing data using the FEAST(19), we found that the urban water samples had a significantly greater (Kruskal-Wallis; $P < 0.01$) contribution from gut bacteria compared to the rural samples without previous antibiotic use (Figure 3C).

The urban sediment samples were significantly more diverse than both the rural samples with and without previous antibiotic use (Browne-Forsythe and Welch; $P < 0.05$). There was no significant difference in diversity between either of the rural sediment sample types (Figure 3D). On the other hand, the rural water samples without previous antibiotic use were significantly more diverse than the rural samples with previous antibiotic use (Browne-Forsythe and Welch; $P < 0.005$) but there was no significant difference between the urban water samples and either of the rural sample types.

Urban samples carry the highest antibiotic resistance gene loads.

A total of 114 different antibiotic resistance genes (ARGs) that confer resistance to 16 antibiotic classes were identified in the 48 samples from sediment and surface water. The urban samples had the greatest number of ARGs (n = 99) followed by the rural samples with previous antibiotic use (n

= 49), while the rural samples with no previous antibiotic use had the fewest resistance genes (n = 36) (Figure 4). There was a large overlap between the ARGs present in the different sample types with the urban and rural + antibiotic samples sharing the greatest number of resistance genes (n = 24). There were 17 ARGs shared between all three samples types including five different beta-lactamase genes belonging to the *bla*$_{OXA}$ and *bla*$_{RSA}$ families.

The abundance of antibiotic resistance genes varied 1,525-fold between sites, with sample SAM6 (rural sediment sample with previous antibiotic exposure collected in Shariatpur) having the lowest abundance (0.078 Reads per Kilobase per Million reads [RPKM]) and sample WD7 (surface water sample collected in Dhaka) having the highest ARG abundance (120.45 RPKM). Of the paired sediment and water samples, the ARG abundance was on average 3 times greater in the water samples than the sediment samples (Wilcoxon; $P < 0.0001$). The urban sediment samples collected from around the city of Dhaka were found to have a significantly (Kruskal-Wallis; $P < 0.05$) greater total ARG abundance (median RPKM = 4.01, interquartile range (IQR) = 0.95 – 12.79) than the rural samples with prior antibiotic use (median RPKM = 0.60, IQR = 0.20 – 1.27) (Figure 5A). However, the urban sediment samples were not significantly different to the rural samples without antibiotic use (median RPKM = 0.72, IQR = 0.64 – 1.36). There was also no statistically significant difference (Kruskal-Wallis; $P > 0.99$) between ARG abundance in rural sediment with prior antibiotic use versus sediment from rural sites in which antibiotics had not been used. ARG levels in the water samples reflected that of the sediment samples, with the total ARG abundance in urban samples (median RPKM = 37.08, IQR = 5.71 – 97.74) being significantly higher (Kruskal-Wallis; $P < 0.05$) than the rural samples with previous antibiotic use (median RPKM = 4.30, IQR = 2.39 – 7.60) but not significantly different to the rural samples with no previous antibiotic use (median RPKM = 5.09, IQR = 1.80 – 11.68). As with the sediment samples there was no significant difference found between either of the rural sample types (Kruskal-Wallis, $P > 0.99$).

The individual antibiotic resistance genes were collated into 16 classes that cover resistance to specific antibiotics and a separate class for genes conferring antibiotic efflux mechanisms (Figure 5B). Efflux genes were present in 47 of 48 samples making it the most widespread ARG class. Other abundant antibiotic resistance classes were resistance to sulphonamides, macrolides and aminoglycosides. Urban water samples WD2, WD6, WD7 and WD1 and an urban sediment sample SD7 clustered together, with high levels of resistance genes from these classes.

Abundance of human gut bacteria predicts levels of antibiotic resistance genes.

There was a statistically significant correlation (R^2 = 0.73; P = 8.9 x 10^{-15}) between the aggregated abundance of ARGs and the levels of human gut bacteria across our study (Figure 6A). We also determined whether the levels of ESBL-producing coliforms are correlated with the total abundance of ARGs and observed a relatively weak but statistically significant correlation (R^2 = 0.38; P = 1.8 x 10^{-6}) (Figure 6B).

Urban sites were enriched in plasmids carrying antibiotic resistance genes.

As antibiotic resistance genes were particularly abundant in water samples, we performed a metagenomic assembly of the short-read data from the surface water samples to recover complete plasmid sequences and study their potential association with antibiotic resistance. The metagenomic assemblies were queried against the PlasmidFinder database (20) to identify contigs which contained plasmid replication (*rep*) genes. Eleven contigs in our dataset contained *rep* genes (Table S2). Seven Gram-negative replicons were found which were related to representatives of the P and Q incompatibility groups or to small theta- or rolling circle-replicating plasmids. A single Gram-positive replicon, repUS43, was identified in sample WD1. Two plasmid contigs, k141_206349 (2113 bp) and k141_304072 (8535 bp), could be circularised (Figure S1). The latter plasmid, which we named pWD1, contained the sulphonamide resistance gene *sul2* adjacent to a complete copy of the mobile element CR2 (blue box in Figure 7), an IncQ1 replicon, three mobilisation genes

(*mobABC*) and an origin-of-transfer (*oriT*). pWD1 was found to have 99.97% identity over 81% of its sequence to the canonical broad-host range mobilisable plasmid RSF1010 (21). Alignment and annotation of these two plasmids revealed that they were identical apart from in the region immediately downstream of *sul2*. In RSF1010 the insertion of the streptomycin resistance genes *strA-strB,* is associated with truncation of CR2 and the *rcr2* gene (Figure 7). While the RSF1010 configuration is common, the *sul2*-CR2 configuration in pWD1 was not found in any other IncQ1 plasmids in GenBank (searched December 9, 2020).

As metagenomic assemblies are often fragmented and plasmid replication genes may not be on the same contigs as ARGs that are carried on another region of the plasmid, we employed PlasFlow (22) to classify contigs in our metagenomic assembly as either chromosomal or plasmid. We identified a total of 93 plasmid contigs containing ARGs. The urban sediment samples contained significantly more plasmid contigs with ARGs than either of the rural sample types (Kruskal-Wallis; $P < 0.001$) whereas the urban water samples had significantly more ARG bearing plasmid contigs than the rural samples with no previous antibiotic use (Kruskal-Wallis*;* $P < 0.05$) (Figure 8). There was no significant difference in the number of ARG-containing plasmid contigs between rural samples with and without prior antibiotic use. Of the 93 contigs identified which contained ARGs, 78 contigs contained only one resistance gene with the remaining 15 contigs containing two or more ARGs (Table S3). All of the contigs that contained multiple resistance genes were found in urban samples and were closely related to known proteobacterial plasmids.

Discussion

In this study we used quantitative culture and metagenomic techniques to understand the community composition and the level of antibiotic resistance genes in rural and urban surface water sites across Bangladesh. Selective plating showed that ESBL-producing coliforms were more prevalent in urban surface water compared to rural settings, consistent with reports of antibiotic resistant faecal coliforms in rivers across Asia (23, 24). However, the predictive value of the abundance of ESBL-producing coliforms for the total abundance of antibiotic resistance genes was found to be limited, suggesting that ESBL-producing coliforms are not necessarily a valid proxy to determine AMR load in environmental ecosystems.

In addition to quantitative culture of ESBL-producing coliforms, a metagenomic shotgun sequencing approach was used to characterise the microbiota of each sample and quantify the abundance of antibiotic resistance genes in water and sediment samples. We found that the water and sediment samples grouped together by their type (water or sediment) rather than the location they were collected from. Sediment samples were dominated by bacteria belonging to the genera *Pseudomonas*, *Azoarcus* and *Hydrogenophilacea* which is in line with other studies which have shown that sediment is dominated by the phylum Proteobacteria (25). Water samples were dominated by the cyanobacteria *Cyanobium* and *Microcystis* that cause harmful blooms in aquaculture ponds (26). *Microcystis* produces potent toxins which can kill fish but are also harmful to humans (27). The two river water samples and a public pond water sample collected in Dhaka clustered away from the other water samples and were defined by an increased abundance of bacteria associated with the human intestinal tract. The presence of increased amounts of the faecal indicator bacteria *E. coli* suggests that human waste is contaminating urban surface water (28).

Several different types of antibiotic were used in the rural aquaculture ponds which we surveyed (Table S1). The antibiotics were either mixed with feed or added directly to the ponds for the treatment of disease. Fluoroquinolone antibiotics such as ciprofloxacin and levofloxacin were the most widely used antibiotics in the rural aquaculture ponds, however high levels of fluoroquinolone resistance were not observed in the rural sites with prior antibiotic use. Resistance to fluoroquinolone drugs is mainly mediated by chromosomal mutations in the *parC* and *gyrA* genes, so the absence of dedicated resistance genes in these ecosystems may be unsurprising (29). However, we note that the multidrug efflux pump genes *mexV*, *mexF*, *adeI* and *adeH* were exclusively found in the rural sites with prior antibiotic use and these efflux systems are capable of exporting fluoroquinolones from the cell (30–33). In addition, other multidrug efflux pump genes capable of exporting fluoroquinolones such as *evgA* and *qacH* were found in these sites and in urban samples (34, 35). The macrolide drug erythromycin was another antibiotic which was widely used in aquaculture ponds that were sampled in this study. However, levels of macrolide resistance genes were low in the rural aquaculture ponds but extremely high in a subset of the urban samples. Notably, the erythromycin resistance gene *msrA* (36) was only present in the aquaculture ponds with prior antibiotic use. This gene was previously found in the intestinal contents of farmed rainbow trout and may thus be more commonly associated with aquaculture (37). Perhaps surprisingly, we did not observe a difference in the total load of antibiotic resistance genes in rural ponds with and without a history of antibiotic use. It may be possible that the widespread use of poultry manure as fish feed in both types of ponds (38–40) has introduced antibiotic resistant bacteria and/or antibiotics and could thus have minimised differences. Further research is needed to quantify the impact of these practices on the selection for antibiotic resistance in aquaculture ponds.

Our data suggest that antibiotic use in Bangladeshi aquaculture does not have a significant effect on the aggregated abundance of all antibiotic resistance genes in this ecosystem in comparison to urban surface water sites. Antibiotic resistance was the highest in urban areas which suggests that human

276 factors contribute to the accumulation of antibiotic resistant bacteria in the environment. This was
277 further corroborated by the correlation between the abundance of bacteria originating from the
278 human gut and antibiotic resistance gene abundance observed in our study. The rivers and lakes of
279 Dhaka are surrounded by areas with high population densities with 13.7% of households reporting
280 that human waste is untreated and released directly into lakes, ponds or rivers (41). Our study thus
281 extends on previous observations that link the introduction of human sewage into river and lake
282 systems to high levels of antibiotic resistance genes (42).
283
284 By creating a metagenomic assembly of our short-read sequencing data we were able to identify
285 contigs which contained plasmid replication initiation genes, contigs which could be circularised
286 into complete plasmids and contigs which were predicted to be plasmids by PlasFlow and contained
287 antibiotic resistance genes. We found that IncP, IncQ and various small plasmid types were most
288 common. All of these plasmid types can replicate in a number of species belonging to the
289 Enterobacteriaceae (43). Two complete plasmid sequences were recovered from the metagenomic
290 assemblies of samples WCM1 and WD1. The small rolling-circle plasmid pWCM1 is closely
291 related to plasmids such as pNMEC-O75D and p124_D that have been found in human and
292 environmental *E. coli* isolates (44). The IncQ1 plasmid pWD1 is closely related and clearly
293 ancestral to the well-characterised RSF1010. Although RSF1010 has been circulating globally since
294 at least the 1960s, the structures of ancestral IncQ1 plasmids that only contain *sul2* have been
295 predicted (45) but never found. The discovery of pWD1 in an urban water sample from 2018 is
296 therefore surprising and demonstrates that this ancestral plasmid lineage has persisted stably for
297 over 50 years. Due to the difficulties of assembling complete plasmid sequences from short-read
298 metagenomic datasets, we were only able to circularise two plasmid sequences. For this reason, we
299 also used additional methods to reconstruct plasmids revealing that urban samples had a higher
300 number of plasmids carrying antibiotic resistance genes. This suggests that particularly in urban

water bodies there exists an increased potential of horizontal gene transfer of mobile genetic elements carrying antibiotic resistance genes.

The microbiotas of surface water and sediment samples across Bangladesh are diverse, but antibiotic resistance genes are highly abundant in urban samples and are more commonly associated with plasmids in this setting. While the abundance of antibiotic resistance genes was considerably lower in rural than in urban settings, we nonetheless observed evidence for the selection for fluoroquinolone resistance mechanisms in ponds used for fish farming. Policies to minimise the use of antibiotics in aquaculture should thus remain a priority to reduce selection for antibiotic resistance. The presence of human gut bacteria was associated with high levels of antibiotic resistance genes, suggesting that contamination by human waste is an important driver for the presence of antibiotic resistance genes in surface water. Interventions aimed at improving access to clean water, sanitation and sewerage infrastructure may thus be important to reduce the risk of AMR dissemination in Bangladesh and other low- and middle-income countries.

Materials and Methods

Site selection

Paired surface water and sediment samples were collected in Bangladesh from 24 freshwater sites across three districts (Mymensingh, Shariatpur and Dhaka; Figure 1) in May and June of 2018. These sites spanned both rural and urban areas with different population densities. Samples were collected from 11 aquaculture ponds in the rural areas of two districts (Mymensingh and Shariatpur) with high commercial aquaculture activity. These ponds all had a history of antibiotic use within the past three months of collection. Six ponds with no history of antibiotic use were also sampled from these rural areas. In Mymensingh, 3 ponds used for domestic purposes were selected, while in Shariatpur, these were aquaculture ponds with no prior antibiotic use, which were used for culturing fingerlings. Antibiotic use information for the ponds was collected from local dealers who were responsible for supplying fish feed for these ponds. In addition to rural surface water sites, 7 water bodies (rivers, lakes and public ponds) were sampled in Dhaka. The public ponds were heavily used for domestic purposes and, while some had history of casual (non-commercial) fish cultivation, none of them had any prior antibiotic use.

Sample collection

Samples were named using the following scheme; water (W) or sediment (S) followed by aquaculture (A) or control (C; ponds without antibiotic use). Sample sites were designated using (M) Mymensingh, (S) Shariatpur or (D) Dhaka and a number was included to differentiate samples. Further metadata on the samples, including temperature, pH and dissolved oxygen levels are provided in Table S1. Water samples were collected by submerging a sterile 500 ml Nalgene plastic bottle approximately 15 cm below the water's surface. Bottles were capped before being removed from the water. The water samples were filtered through a 0.22 µm Sterivex-GP filter (Millipore) until water would no longer be passed through the filter. The filter units were then capped and

342 stored in a cool box and transported to the laboratory within 12 hours of sampling. In addition to the
343 water samples, approximately 10 g of sediment was taken from either the bed of the pond or from
344 the bank 30 – 50 cm below the surface of the water. The sediment samples were stored in sterile 50
345 ml Falcon tubes and were transported with the water samples.

346

347 **Selective culturing for coliforms in surface water and sediment samples**

348 Water and sediment samples were screened for the presence of Extended-Spectrum Beta-Lactamase
349 (ESBL) producing coliforms by quantitative plating on Brilliance ESBL agar (Oxoid). Water and
350 sediment samples were spread onto the plates and incubated for 48 hours at 37°C. In accordance
351 with the manufacturer's instructions blue, pink and green colonies were designated as coliforms and
352 counted.

353

354 **DNA extraction and Illumina sequencing**

355 DNA was extracted from the Sterivex filters and sediment samples using the DNeasy PowerWater
356 Kit (Qiagen) and the DNeasy PowerSoil kit (Qiagen), respectively, in accordance with the
357 manufacturer's instructions. DNA concentrations were quantified using the Qubit dsDNA HS assay
358 kit (Thermo Fisher) with all samples yielding more than 0.2 ng/µl. Negative control runs were
359 performed for both kits by isolating DNA from sterile, distilled water: these yielded no detectable
360 DNA. Metagenomic DNA libraries were prepared using the Nextera XT Library Prep Kit
361 (Illumina). The libraries were pooled and sequenced on the HiSeq 2500 sequencing platform
362 (Illumina) using a 150 bp paired-end protocol. Paired reads were adapter trimmed and both
363 duplicates and reads less than 50 bp were removed using Trimmomatic 0.30 with Q15 as the
364 sliding-window quality cut-off (46). The short-read sequencing data for this project has been
365 deposited at the European Nucleotide Archive under accession number PRJEB39306.

366

Taxonomic Profiling

To perform taxonomic profiling, the paired-end sequencing reads were mapped against clade specific markers using the MetaPhlAn2 package v.2.7.7 (47). The MetaPhlAn2 package was run with default parameters. The utility script merge_metaphlan_tables.py was used to merge all of the output files into a single tab delimited file.

Source-sink analysis

Raw sequence reads from projects PRJNA254927, PRJEB7626 and PRJEB6092, which had previously been used as sources for source-sink analysis (48), were downloaded from the European Nucleotide Archive (ENA). These sequences represented freshwater, soil and gut metagenomes respectively. Adapters were removed from the sequence reads using fastp (49). Taxonomic counts were created for these metagenomic sequences and the 48 samples in this study by kraken2 v.2.0.9 (50) and Bracken v.2.6.0 (51) using a database containing bacterial, archaeal, viral and fungal sequences. A metadata table was created which described the environment that the sample was from and designated it as either a source or a sink. The taxonomic count table and the metadata table were used an input to the R package FEAST v.0.1.0 (19) which determined the proportion that each source contributed to each sink.

Resistome profiling

Antibiotic resistance genes were identified using the ShortBRED package v.0.9.5 (52). The CARD database (53) (downloaded 1st July 2019) and the UniRef90 database (downloaded 4 July 2019) were used by ShortBRED-Identify to construct a marker database which the metagenomic reads could be mapped against. ShortBRED-Quantify.py was then used to map these paired-end reads against the database. The relative abundance in Reads Per Kilobase per Million reads (RPKM) was generated for each resistance gene family in the database. The RPKMs were summed for antibiotic

resistance genes belonging to the same class and visualised with the pheatmap package (https://cran.r-project.org/web/packages/pheatmap/pheatmap.pdf) in R (54).

Reconstruction of plasmids from metagenomic datasets

Metagenomic sequencing reads were assembled using the MEGAHIT v.1.1.3 assembler using default parameters (55). Contigs produced by MEGAHIT were then classified as plasmid or chromosomal by trained neural networks in the PlasFlow v1.1 program (22). Contigs designated to be of plasmid origin were queried against the CARD database by ABRicate v.0.9.8 (https://github.com/tseemann/abricate) to identify the presence of antibiotic resistance genes. Resistance genes were identified which had at least 95% identity and 50% coverage compared to the CARD database. Plasmid contigs were similarly queried against the PlasmidFinder database (20) to identify replication genes. Plasmids were circularised by comparing 300 bp from either end of putative plasmid-containing contigs using BLASTn (56). When ends were found to overlap, one copy of the overlapping sequence was removed to generate a complete, circularised plasmid sequence.

Statistical analyses

The Shannon Diversity Index of the samples was calculated in R v.3.4.3 using the diversity function of the vegan package v.2.5-7 (57). Non-metric multidimensional scaling (NMDS) was also performed in R using the metaNMDS function of the vegan package. Permutational multivariate analysis of variance (PERMANOVA) was performed on a Bray-Curtis distance matrix of species abundance in R using the adonis function of the vegan package. Correlation between total ARG abundance and human gut bacterial contribution was calculated using the lm function in base R. Additional tests for determining statistical significance were performed as described in the text, implemented in GraphPad Prism v.8.3.1.

Acknowledgments

This study was funded by the Royal Society through a Challenge Grant (CHG\R1\170015) and a Wolfson Research Merit Award (WM160092) to W.v.S. Metagenome sequencing was provided by MicrobesNG (http://www.microbesng.uk), which is supported by the BBSRC (grant number BB/L024209/1). R.S.M is funded by the Wellcome Trust Antimicrobials and Antimicrobial Resistance Doctoral Training Programme (215154/Z/18/Z).

Author Contributions

W.V.S. and M.S.I. conceived this study. R.S.M., M.H.Z, I.T.A. and W.V.S. collected and processed the environmental samples with support from J.D.C. R.S.M., S.H. and R.A.M. analysed the data. R.S.M. and W.V.S. wrote the manuscript with input from all authors.

Data Availability

Raw sequencing data have been submitted to the European Nucleotide Archive with accession number PRJEB39306.

Figure legends

Figure 1. Map of Bangladesh showing the districts that the samples were collected from and the population of each district (obtained through https://data.humdata.org/dataset/bangladesh-administrative-level-0-3-population-statistics). Green circles represent sampling locations.

Figure 2. The abundance of ESBL producing coliforms, in $\log_{10}$ (cfu/ml), isolated from sediment and surface water in urban sites and rural settings with antibiotic use (+Abx) and without antibiotic use (-Abx) across Bangladesh. The horizontal dashed line represents the detection limit of 20 cfu/ml. Samples with ESBL-producing coliforms below the detection limit were plotted at $\log_{10}$ (cfu/ml) = 1.

Figure 3. **A**. Non-metric multidimensional scaling (NMDS) analysis of a Bray Curtis distance matrix of species abundance. Stress 0.15. Ellipses represent standard deviation. **B.** Relative abundance (%) of Phyla across the 48 samples from sediment and surface water. **C.** Source-sink analysis, percentage contribution of human gut bacteria to the bacterial composition of the water and sediments samples. Kruskal-Wallis. $^{**}P < 0.01$. **D.** Shannon diversity values of species present in sediment and water samples from across Bangladesh. Brown-Forsythe ANOVA. $^{*}P < 0.05$ $^{**}P < 0.005$.

Figure 4. Distribution of antibiotic resistance genes across urban, rural without prior antibiotic use and rural with prior antibiotic use sample types. Circles are proportional to the number of antibiotic resistance genes present within each sample type.

Figure 5. **A.** Abundance in RPKM of Antibiotic Resistance Genes (ARGs) in each sample (sediments and surface water; urban, rural with antibiotic use and rural without antibiotic use).

Kruskal-Wallis test * $P < 0.05$. **B.** Heatmap representing the summed abundance (log_{10} transformed RPKM) of antibiotic resistance gene classes present in water and sediment samples from surface water sites across Bangladesh.

Figure 6. **A.** Correlation between the total antibiotic resistance gene (ARG) abundance (RPKM) and the percentage of bacteria contributed from the human gut within each sample. $R^2 = 0.73$. $P = 8.9 \times 10^{-15}$. **B.** Correlation between the total ARG abundance (RPKM) and the number of ESBL producing coliforms (cfu/ml) in each sample. $R^2 = 0.38$. $P = 1.8 \times 10^{-6}$. The grey area represents the 95% confidence interval.

Figure 7. Comparison of plasmids pWD1 and RSF1010. Plasmid sequence is shown as a black line with the positions of genes indicated by labelled arrows below and the location of *oriT* shown above. The mobile element CR2 is shown as a thicker blue box. The light-blue shading highlights the region that differs between the plasmids and includes the *strAB* genes in RSF1010. Drawn to scale from GenBank accessions MW363525 and M28829 for pWD1 and RSF1010, respectively.

Figure 8. The number of contigs identified as plasmid by PlasFlow that carry an antibiotic resistance gene from Bangladesh surface water sites in sediment and rural, with and without antibiotic use (Abx), sediment and surface water samples. Kruskal-Wallis test: * $P < 0.05$ and *** $P < 0.001$.

References

1. Founou RC, Founou LL, Essack SY. 2017. Clinical and economic impact of antibiotic resistance in developing countries: A systematic review and meta-analysis. PLOS ONE 12:e0189621.

2. Gandra S, Tseng KK, Arora A, Bhowmik B, Robinson ML, Panigrahi B, Laxminarayan R, Klein EY. 2019. The mortality burden of multidrug-resistant pathogens in India: a retrospective, observational study. Clin Infect Dis 69:563–570.

3. Laxminarayan R, Duse A, Wattal C, Zaidi AKM, Wertheim HFL, Sumpradit N, Vlieghe E, Hara GL, Gould IM, Goossens H, Greko C, So AD, Bigdeli M, Tomson G, Woodhouse W, Ombaka E, Peralta AQ, Qamar FN, Mir F, Kariuki S, Bhutta ZA, Coates A, Bergstrom R, Wright GD, Brown ED, Cars O. 2013. Antibiotic resistance—the need for global solutions. Lancet Infect Dis 13:1057–1098.

4. Lim C, Takahashi E, Hongsuwan M, Wuthiekanun V, Thamlikitkul V, Hinjoy S, Day NP, Peacock SJ, Limmathurotsakul D. 2016. Epidemiology and burden of multidrug-resistant bacterial infection in a developing country. eLife 5:e18082.

5. Chokshi A, Sifri Z, Cennimo D, Horng H. 2019. Global contributors to antibiotic resistance. J Glob Infect Dis 11:36–42.

6. Collignon P, Beggs JJ, Walsh TR, Gandra S, Laxminarayan R. 2018. Anthropological and socioeconomic factors contributing to global antimicrobial resistance: a univariate and multivariable analysis. Lancet Planet Health 2:e398–e405.

7. Woolhouse M, Ward M, van Bunnik B, Farrar J. 2015. Antimicrobial resistance in humans, livestock and the wider environment. Philos Trans R Soc B Biol Sci 370:20140083.

8. McEwen SA, Collignon PJ. 2018. Antimicrobial resistance: a One Health perspective. Microbiol Spectr 6 doi:10.1128/microbiolspec.ARBA-0009-2017.

9. Robinson TP, Bu DP, Carrique-Mas J, Fèvre EM, Gilbert M, Grace D, Hay SI, Jiwakanon J, Kakkar M, Kariuki S, Laxminarayan R, Lubroth J, Magnusson U, Thi Ngoc P, Van Boeckel TP, Woolhouse MEJ. 2016. Antibiotic resistance is the quintessential One Health issue. Trans R Soc Trop Med Hyg 110:377–380.

10. Lowder SK, Skoet J, Raney T. 2016. The Number, Size, and Distribution of Farms, Smallholder Farms, and Family Farms Worldwide. World Dev 87:16–29.

11. Garlock T, Asche F, Anderson J, Bjørndal T, Kumar G, Lorenzen K, Ropicki A, Smith MD, Tveterås R. 2020. A Global Blue Revolution: Aquaculture Growth Across Regions, Species, and Countries. Rev Fish Sci Aquac 28:107–116.

12. Ahmed I, Rabbi MdB, Sultana S. 2019. Antibiotic resistance in Bangladesh: A systematic review. Int J Infect Dis 80:54–61.

13. Peal A, Evans B, Blackett I, Hawkins P, Heymans C. 2014. Fecal sludge management: a comparative analysis of 12 cities. J Water Sanit Hyg Dev 4:563–575.

14. Rousham EK, Islam MA, Nahar P, Lucas PJ, Naher N, Ahmed SM, Nizame FA, Unicomb L. 2019. Pathways of antibiotic use in Bangladesh: qualitative protocol for the PAUSE study. BMJ Open 9:e028215.

15. 2018. FAO yearbook. Fishery and Aquaculture Statistics 2016.

16. Ali H, Rico A, Murshed-e-Jahan K, Belton B. 2016. An assessment of chemical and biological product use in aquaculture in Bangladesh. Aquaculture 454:199–209.

17. Lulijwa R, Rupia EJ, Alfaro AC. 2020. Antibiotic use in aquaculture, policies and regulation, health and environmental risks: a review of the top 15 major producers. Rev Aquac 12:640–663.

18. Kawsar A, Alam T, Ahamed S, Mou H. 2018. Aqua drugs and antibiotics used in freshwater aquaculture of North Chittagong, Bangladesh. Int J Fish Aquat Stud 7:7.

19. Shenhav L, Thompson M, Joseph TA, Briscoe L, Furman O, Bogumil D, Mizrahi I, Pe'er I, Halperin E. 2019. FEAST: fast expectation-maximization for microbial source tracking. Nat Methods 16:627–632.

20. Carattoli A, Zankari E, García-Fernández A, Voldby Larsen M, Lund O, Villa L, Møller Aarestrup F, Hasman H. 2014. In silico detection and typing of plasmids using PlasmidFinder and plasmid multilocus sequence typing. Antimicrob Agents Chemother 58:3895–3903.

21. Scholz P, Haring V, Wittmann-Liebold B, Ashman K, Bagdasarian M, Scherzinger E. 1989. Complete nucleotide sequence and gene organization of the broad-host-range plasmid RSF1010. Gene 75:271–288.

22. Krawczyk PS, Lipinski L, Dziembowski A. 2018. PlasFlow: predicting plasmid sequences in metagenomic data using genome signatures. Nucleic Acids Res 46:e35–e35.

23. Lamba M, Gupta S, Shukla R, Graham DW, Sreekrishnan TR, Ahammad SZ. 2018. Carbapenem resistance exposures via wastewaters across New Delhi. Environ Int 119:302–308.

24. Yu Y, Wu G, Wang C, Lu N, Yuan X, Zhu X. 2019. Pollution characteristics of antibiotics and antibiotic resistance of coliform bacteria in the Yitong River, China. Environ Monit Assess 191:516.

547 25. Nho SW, Abdelhamed H, Paul D, Park S, Mauel MJ, Karsi A, Lawrence ML. 2018. Taxonomic
548 and Functional Metagenomic Profile of Sediment From a Commercial Catfish Pond in
549 Mississippi. Front Microbiol 9:2855.

550 26. Zhong F, Gao Y, Yu T, Zhang Y, Xu D, Xiao E, He F, Zhou Q, Wu Z. 2011. The management
551 of undesirable cyanobacteria blooms in channel catfish ponds using a constructed wetland:
552 Contribution to the control of off-flavor occurrences. Water Res 45:6479–6488.

553 27. Paerl HW, Tucker CS. 1995. Ecology of Blue-Green Algae in Aquaculture Ponds. J World
554 Aquac Soc 26:109–131.

555 28. Ouattara NK, de Brauwere A, Billen G, Servais P. 2013. Modelling faecal contamination in the
556 Scheldt drainage network. J Mar Syst 128:77–88.

557 29. Hooper DC, Jacoby GA. 2015. Mechanisms of drug resistance: quinolone resistance. Ann N Y
558 Acad Sci 1354:12–31.

559 30. Coyne S, Rosenfeld N, Lambert T, Courvalin P, Périchon B. 2010. Overexpression of
560 resistance-nodulation-cell division pump AdeFGH confers multidrug resistance in
561 *Acinetobacter baumannii*. Antimicrob Agents Chemother 54:4389–4393.

562 31. Damier-Piolle L, Magnet S, Brémont S, Lambert T, Courvalin P. 2008. AdeIJK, a resistance-
563 nodulation-cell division pump effluxing multiple antibiotics in *Acinetobacter baumannii*.
564 Antimicrob Agents Chemother 52:557–562.

565 32. Köhler T, Epp SF, Curty LK, Pechère JC. 1999. Characterization of MexT, the regulator of the
566 MexE-MexF-OprN multidrug efflux system of *Pseudomonas aeruginosa*. J Bacteriol
567 181:6300–6305.

568 33. Li Y, Mima T, Komori Y, Morita Y, Kuroda T, Mizushima T, Tsuchiya T. 2003. A new
569 member of the tripartite multidrug efflux pumps, MexVW-OprM, in *Pseudomonas*
570 *aeruginosa*. J Antimicrob Chemother 52:572–575.

571 34. Ceccarelli D, Salvia AM, Sami J, Cappuccinelli P, Colombo MM. 2006. New cluster of
572 plasmid-located class 1 integrons in *Vibrio cholerae* O1 and a *dfrA*15 cassette-containing
573 integron in *Vibrio parahaemolyticus* isolated in Angola. Antimicrob Agents Chemother
574 50:2493–2499.

575 35. Hirakawa H, Nishino K, Yamada J, Hirata T, Yamaguchi A. 2003. Beta-lactam resistance
576 modulated by the overexpression of response regulators of two-component signal transduction
577 systems in *Escherichia coli*. J Antimicrob Chemother 52:576–582.

578 36. Reynolds E, Ross JI, Cove JH. 2003. Msr(A) and related macrolide/streptogramin resistance
579 determinants: incomplete transporters? Int J Antimicrob Agents 22:228–236.

580 37. Muziasari WI, Pitkänen LK, Sørum H, Stedtfeld RD, Tiedje JM, Virta M. 2017. The Resistome
581 of Farmed Fish Feces Contributes to the Enrichment of Antibiotic Resistance Genes in
582 Sediments below Baltic Sea Fish Farms. Front Microbiol 7:2137.

583 38. Hoque R, Ahmed SM, Naher N, Islam MA, Rousham EK, Islam BZ, Hassan S. 2020. Tackling
584 antimicrobial resistance in Bangladesh: A scoping review of policy and practice in human,
585 animal and environment sectors. PLOS ONE 15:e0227947.

586 39. Hossen MS, Hoque Z, Nahar BS. 2015. Assessment of poultry waste management in Trishal
587 upazila, Mymensingh. 2. Res Agric Livest Fish 2:293–300.

588 40. Masud AA, Rousham EK, Islam MA, Alam M-U, Rahman M, Mamun AA, Sarker S,
589 Asaduzzaman M, Unicomb L. 2020. Drivers of Antibiotic Use in Poultry Production in
590 Bangladesh: Dependencies and Dynamics of a Patron-Client Relationship. Front Vet Sci 7.

41. Arias Granada, Yurani Haque, Sabrina Sharmin, George Joseph, Monica Yanez Pagans. 2018. Water and Sanitation in Dhaka Slums : Access, Quality, and Informality in Service Provision. Policy Research Working Paper; No. 8552. World Bank, Washington, DC.

42. Karkman A, Pärnänen K, Larsson DGJ. 2019. Fecal pollution can explain antibiotic resistance gene abundances in anthropogenically impacted environments. Nat Commun 10:80.

43. Rozwandowicz M, Brouwer MSM, Fischer J, Wagenaar JA, Gonzalez-Zorn B, Guerra B, Mevius DJ, Hordijk J. 2018. Plasmids carrying antimicrobial resistance genes in Enterobacteriaceae. J Antimicrob Chemother 73:1121–1137.

44. Nielsen DW, Ricker N, Barbieri NL, Wynn JL, Gómez-Duarte OG, Iqbal J, Nolan LK, Allen HK, Logue CM. 2018. Complete Genome Sequence of the Multidrug-Resistant Neonatal Meningitis *Escherichia coli* Serotype O75:H5:K1 Strain mcjchv-1 (NMEC-O75). Microbiol Resour Announc 7: e01043-18.

45. Yau S, Liu X, Djordjevic SP, Hall RM. 2010. RSF1010-Like Plasmids in Australian *Salmonella enterica* Serovar Typhimurium and Origin of Their *sul2-strA-strB* Antibiotic Resistance Gene Cluster. Microb Drug Resist 16:249–252.

46. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120.

47. Truong DT, Franzosa EA, Tickle TL, Scholz M, Weingart G, Pasolli E, Tett A, Huttenhower C, Segata N. 2015. MetaPhlAn2 for enhanced metagenomic taxonomic profiling. Nat Methods 12:902–903.

48. McGhee JJ, Rawson N, Bailey BA, Fernandez-Guerra A, Sisk-Hackworth L, Kelley ST. 2020. Meta-SourceTracker: application of Bayesian source tracking to shotgun metagenomics. PeerJ 8:e8783.

614 49. Chen S, Zhou Y, Chen Y, Gu J. 2018. fastp: an ultra-fast all-in-one FASTQ preprocessor.
615 Bioinformatics 34:i884–i890.

616 50. Wood DE, Lu J, Langmead B. 2019. Improved metagenomic analysis with Kraken 2. Genome
617 Biol 20:257.

618 51. Lu J, Breitwieser FP, Thielen P, Salzberg SL. 2017. Bracken: estimating species abundance in
619 metagenomics data. PeerJ Comput Sci 3:e104.

620 52. Kaminski J, Gibson MK, Franzosa EA, Segata N, Dantas G, Huttenhower C. 2015. High-
621 Specificity Targeted Functional Profiling in Microbial Communities with ShortBRED. PLOS
622 Comput Biol 11:e1004557.

623 53. Alcock BP, Raphenya AR, Lau TTY, Tsang KK, Bouchard M, Edalatmand A, Huynh W,
624 Nguyen A-LV, Cheng AA, Liu S, Min SY, Miroshnichenko A, Tran H-K, Werfalli RE, Nasir
625 JA, Oloni M, Speicher DJ, Florescu A, Singh B, Faltyn M, Hernandez-Koutoucheva A,
626 Sharma AN, Bordeleau E, Pawlowski AC, Zubyk HL, Dooley D, Griffiths E, Maguire F,
627 Winsor GL, Beiko RG, Brinkman FSL, Hsiao WWL, Domselaar GV, McArthur AG. 2020.
628 CARD 2020: antibiotic resistome surveillance with the comprehensive antibiotic resistance
629 database. Nucleic Acids Res 48:D517–D525.

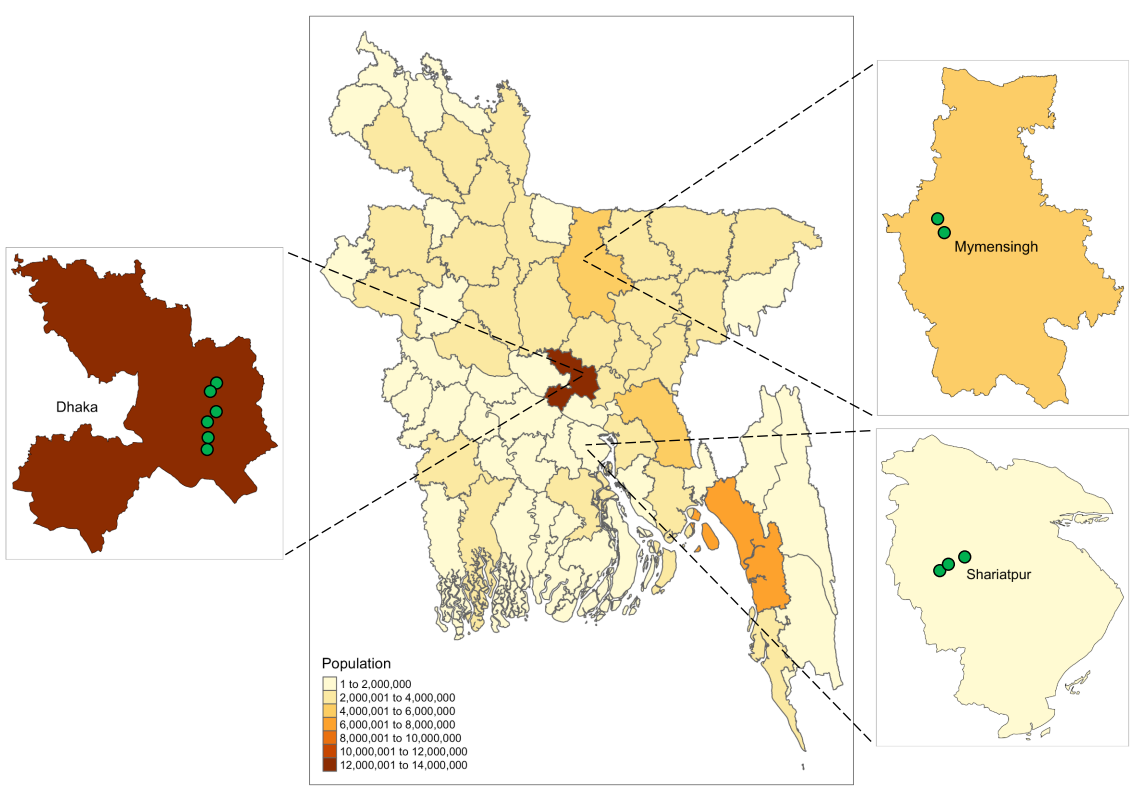
630 54. R Core Team. 2017. R: A language and environment for statistical computing. R Found Stat
631 Comput Vienna Austria.

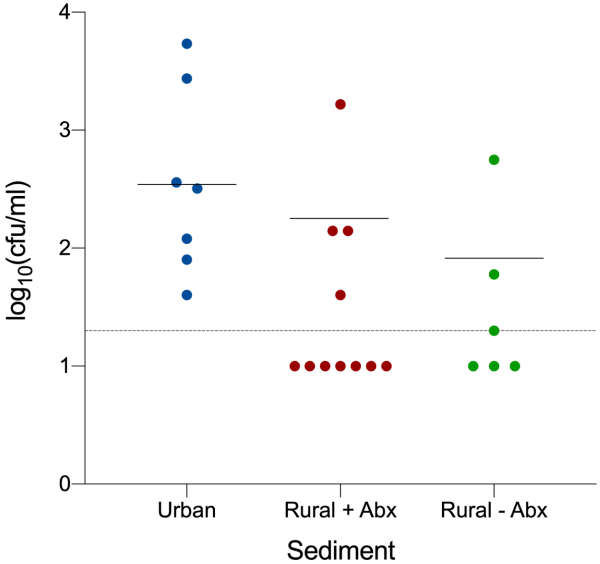
632 55. Li D, Liu C-M, Luo R, Sadakane K, Lam T-W. 2015. MEGAHIT: an ultra-fast single-node
633 solution for large and complex metagenomics assembly via succinct de Bruijn graph.
634 Bioinformatics 31:1674–1676.

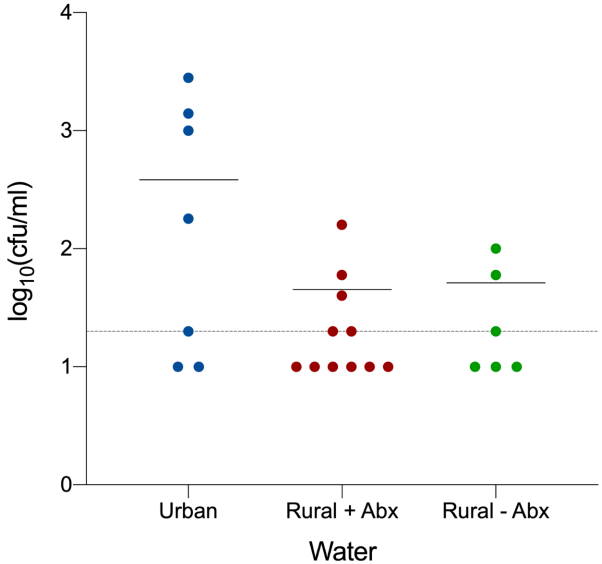
635 56. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search
636 tool. J Mol Biol 215:403–410.

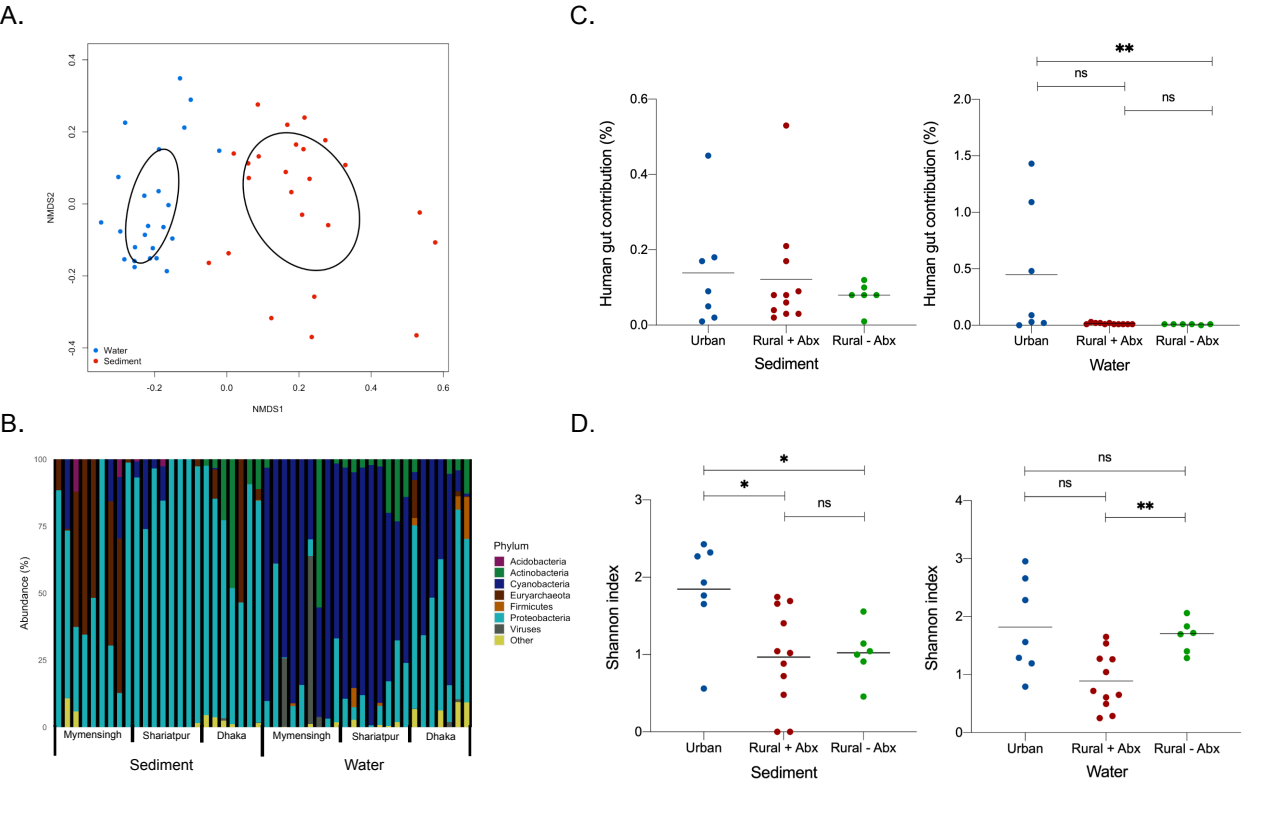
637 57. Oksanen J, Guillaume B, Friendly M, Kindt R, Legendre P, McGlinn D, Minchin PR, O'Hara
638 RB, Simpson GL, Solymos P, Henry M, Stevens H, Szoecs E, Wagner, Helene. 2019. Vegan:
639 Community Ecology Package. https://cran.r-project.org/web/packages/vegan/index.html.

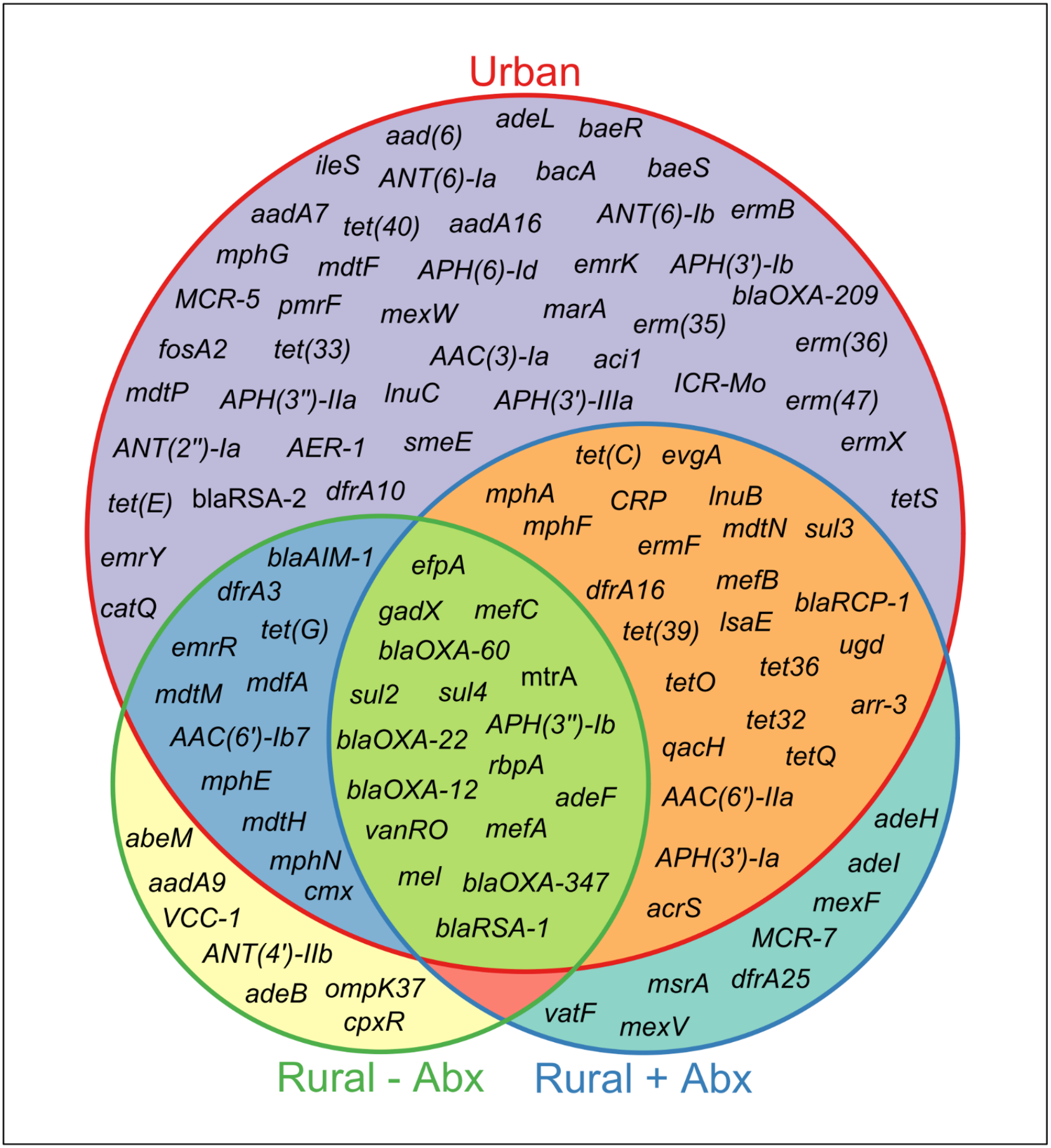
640


Dhaka
Mymensingh
Shariatpur
Population
1 to 2,000,000
2,000,001 to 4,000,000
4,000,001 to 6,000,000
6,000,001 to 8,000,000
8,000,001 to 10,000,000
10,000,001 to 12,000,000
12,000,001 to 14,000,000

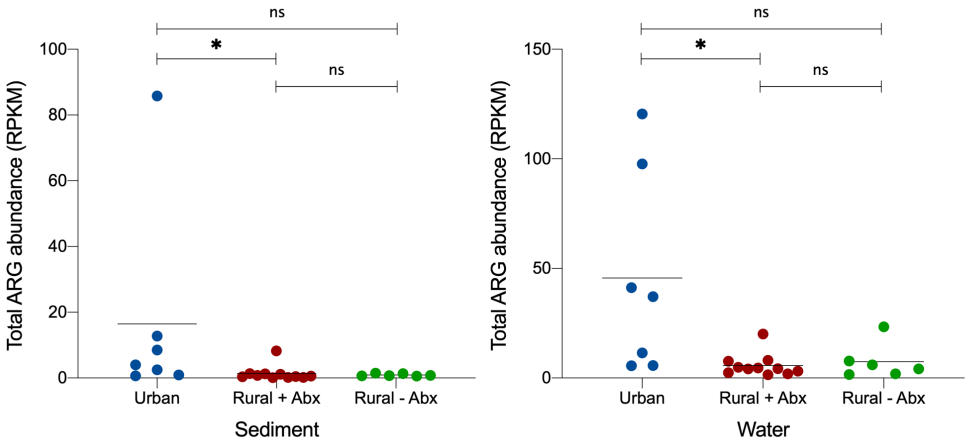

4
3
2
1
0
log10(cfu/ml)
Urban
Rural + Abx
Rural - Abx
Sediment


4
3
2
1
0
log10(cfu/ml)
Urban
Rural + Abx
Rural - Abx
Water

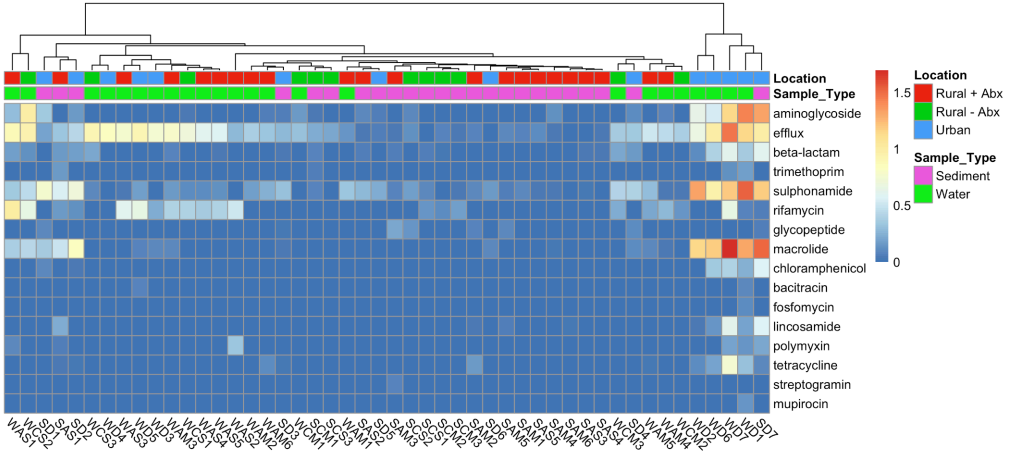

A.
NMDS2
NMDS1
Water
Sediment
B.
Abundance (%)
Mymensingh
Shariatpur
Dhaka
Sediment
Water
Phylum
Acidobacteria
Actinobacteria
Cyanobacteria
Euryarchaeota
Firmicutes
Proteobacteria
Viruses
Other
C.
Human gut contribution (%)
Urban
Rural + Abx
Rural - Abx
Sediment
Water
ns
**
D.
Shannon index
*
ns
**

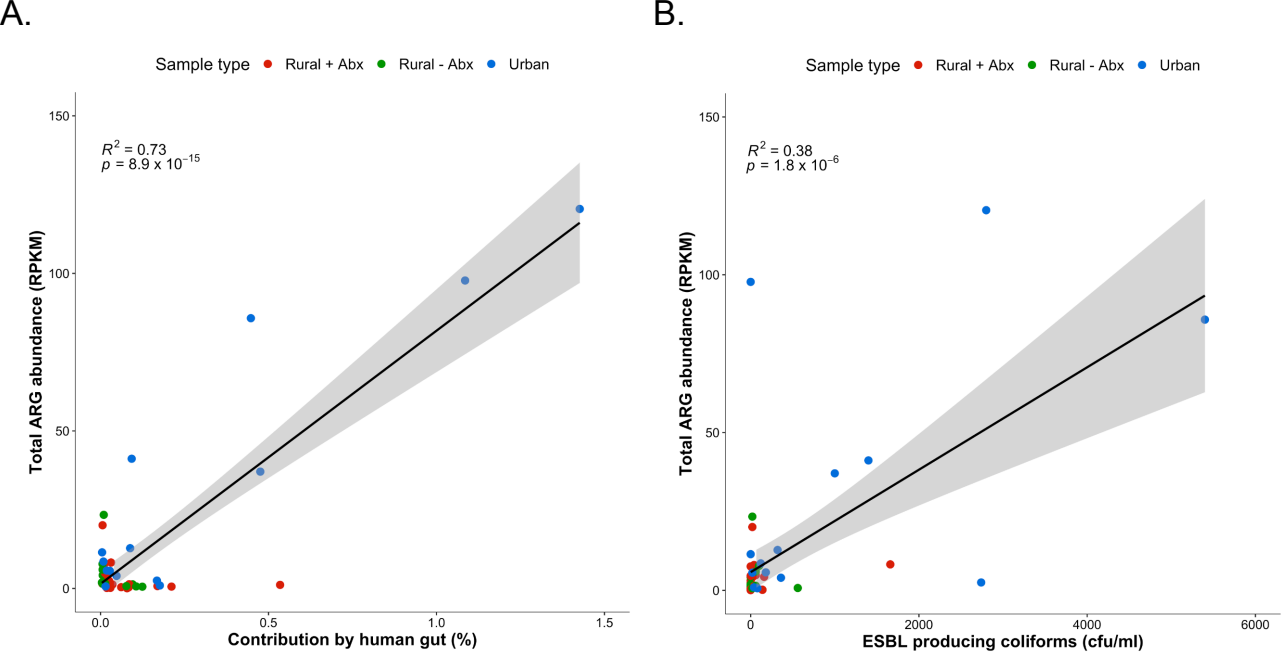

Urban
adeL
baeR
aad(6)
ileS
ANT(6)-Ia
bacA
baeS
aadA7
tet(40)
aadA16
ANT(6)-Ib
ermB
mphG
mdtF
APH(6)-Id
emrK
APH(3')-Ib
MCR-5
pmrF
mexW
marA
blaOXA-209
erm(35)
fosA2
tet(33)
AAC(3)-Ia
aci1
erm(36)
mdtP
APH(3'')-IIa
lnuC
APH(3')-IIIa
ICR-Mo
erm(47)
ANT(2'')-Ia
AER-1
smeE
ermX
tet(C)
evgA
tet(E)
blaRSA-2
dfrA10
mphA
CRP
lnuB
tetS
mphF
mdtN
sul3
ermF
emrY
blaAIM-1
efpA
dfrA3
dfrA16
mefB
catQ
gadX
mefC
blaRCP-1
tet(G)
tet(39)
lsaE
emrR
blaOXA-60
ugd
mtrA
tetO
tet36
mdtM
mdfA
sul2
sul4
arr-3
APH(3'')-Ib
tet32
AAC(6')-Ib7
blaOXA-22
qacH
tetQ
rbpA
mphE
blaOXA-12
adeF
AAC(6')-IIa
mdtH
vanRO
mefA
adeH
abeM
APH(3')-Ia
mphN
adeI
aadA9
mel
blaOXA-347
cmx
acrS
mexF
VCC-1
MCR-7
blaRSA-1
ANT(4')-IIb
msrA
dfrA25
adeB
ompK37
vatF
cpxR
mexV
Rural - Abx
Rural + Abx

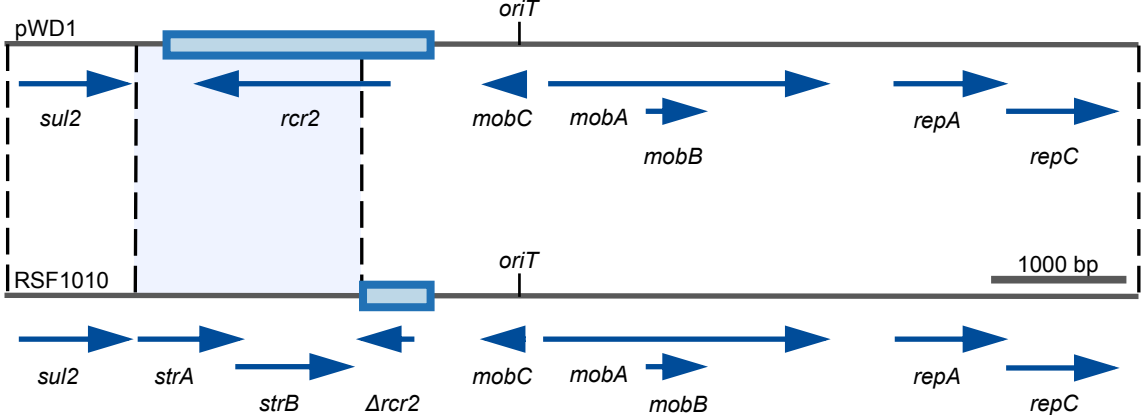
A.

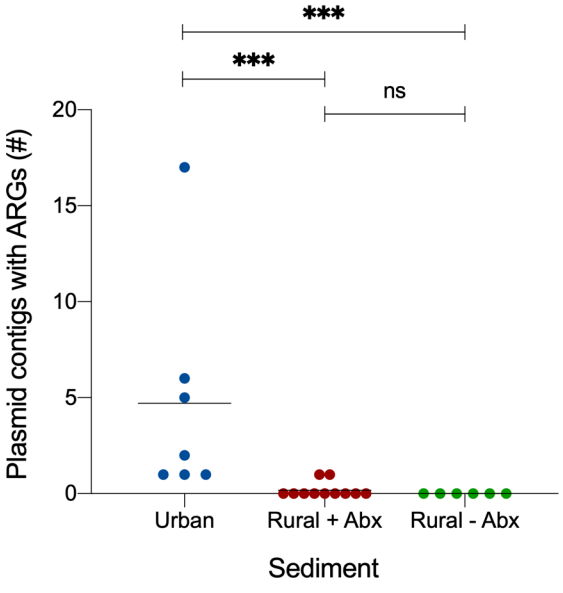

ns
*
ns
Total ARG abundance (RPKM)
100
80
60
40
20
0
Urban
Rural + Abx
Rural - Abx
Sediment
ns
*
ns
Total ARG abundance (RPKM)
150
100
50
0
Urban
Rural + Abx
Rural - Abx
Water

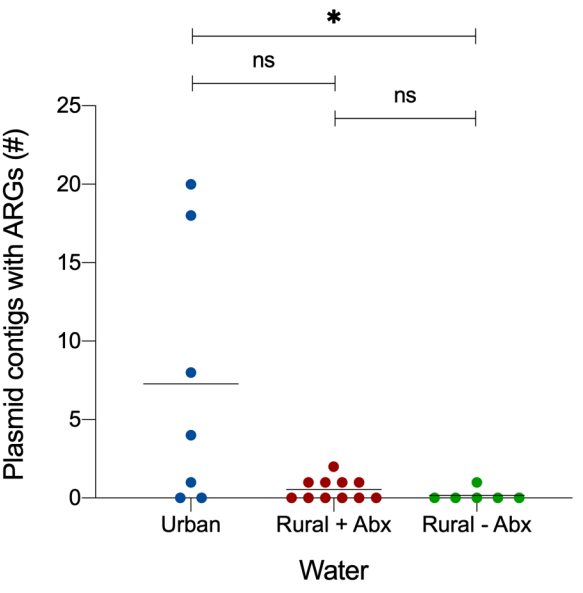
B.


Location
Sample_Type
aminoglycoside
efflux
beta-lactam
trimethoprim
sulphonamide
rifamycin
glycopeptide
macrolide
chloramphenicol
bacitracin
fosfomycin
lincosamide
polymyxin
tetracycline
streptogramin
mupirocin
1.5
1
0.5
0
Location
Rural + Abx
Rural - Abx
Urban
Sample_Type
Sediment
Water


A.
Sample type
Rural + Abx
Rural - Abx
Urban
R^2 = 0.73
p = 8.9 x 10^{-15}
Total ARG abundance (RPKM)
Contribution by human gut (%)
B.
Sample type
Rural + Abx
Rural - Abx
Urban
R^2 = 0.38
p = 1.8 x 10^{-6}
Total ARG abundance (RPKM)
ESBL producing coliforms (cfu/ml)


pWD1
oriT
sul2
rcr2
mobC
mobA
mobB
repA
repC
RSF1010
oriT
1000 bp
sul2
strA
strB
Δrcr2
mobC
mobA
mobB
repA
repC


***
***
ns
20
15
10
5
0
Plasmid contigs with ARGs (#)
Urban
Rural + Abx
Rural - Abx
Sediment


*
ns
ns
25
20
15
10
5
0
Plasmid contigs with ARGs (#)
Urban
Rural + Abx
Rural - Abx
Water