

Why are viral genomes so fragile? The bottleneck hypothesis

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If they undergo new mutations at each replication cycle, why are RNA viral genomes so fragile, with most mutations being either strongly deleterious or lethal? Here we provide theoretical evidence for the hypothesis that genetic fragility evolves as a consequence of the pervasive population bottlenecks experienced by viral populations at various stages of their life cycles. Modelling within-host viral populations as multi-type branching processes, we show that mutational fragility lowers the rate at which Muller's ratchet clicks and increases the survival probability through multiple bottlenecks. In the context of a susceptible-exposed-infectious-recovered epidemiological model, we find that the attack rate of fragile viral strains can exceed that of more robust strains, particularly at low infectivities and high mutation rates. Our findings highlight the importance of demographic events such as transmission bottlenecks in shaping the genetic architecture of viral pathogens.

RNA viruses | fragility | population bottlenecks | evolutionary epidemiology | virus evolution

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Introduction

From tobacco mosaic virus to poliovirus and SARS-CoV-2, some of the most consequential plant, animal and human pathogens are RNA viruses. In spite of their extremely small genomes, these organisms find adaptive solutions to environmental challenges such as hosts' immune response, pervasive differences in susceptible cell types, switches in host and vector species, and antiviral drugs (1–3). The remarkable evolvability of RNA viruses has been linked to their error-prone replication, short generation times, and large population sizes (4). But high mutation rates are a double-edged sword: while replication errors provide the fuel necessary for rapid adaptation, they also increase the genetic load on viral populations, which in turn imposes a limit to genome size (5, 6). Moreover, evidence gathered from diverse viral systems (7–11) shows that high mutation rates coupled with strong population bottlenecks (e.g. associated with airborne or fomite transmission events) turn on Muller's ratchet (12), resulting in the loss of fit genotypes (13, 14). As fitness declines, populations risk experiencing a mutational meltdown, with low fitness genotypes unable to restore large population sizes and deleterious mutations accumulating at an ever increasing rate (15, 16). How do RNA viruses manage to persist in the face of these challenges?

RNA viruses may have evolved specific mechanisms to maintain genome integrity in the face of high mutation rates (17–20). Proposed mechanisms include complementation at high multiplicity of infection during transmission *e.g.*, by physically aggregating viral particles (21, 22); the use of stamping machine, rather than geometric, replication mechanisms (23); the segmenting of viral genomes with segment reassortments during mixed infections or increased recombination rates, two simple forms of sex that reduce mutational load (24, 25); or the co-opting of cellular chaperones (*e.g.*, heat-shock proteins) to assist at different stages of the replication cycle (26). It is possible that all of these mechanisms (and others yet to be discovered) play a role in mitigating the damage done by mutations. Indeed, general arguments suggest that neutral evolution tends to reinforce mutational robustness by moving viral populations away from the edges of neutral networks (27–29).

Somewhat paradoxically, the opposite strategy of *maximizing* mutational damage may also be a key component of the evolutionary response to low-fidelity replication. Both empirical and theoretical arguments support this hypothesis. Firstly, viral RNA genomes generally contain overlapping reading frames, encode for polyproteins that need to be precisely processed post-translationally, and express multifunctional proteins involved in different processes along the infection cycle; these structural properties predict large deleterious effects of most mutations (5, 6), as is indeed observed (30). Secondly, analyses of Muller's ratchet show that the risk of mutational meltdown is highest when deleterious effects are moderate (16). This is because weak deleterious mutations negatively impact population fitness without being strongly selected against (Fig. 1). This observation has led several authors to the conclusion that genetic fragility is in fact selected for in the high mutation rate regime of evolution (31–33).

In this paper we show that the genetic fragility of RNA viruses may be explained by the population bottlenecks they experience during their cycle. Our contribution is twofold. First, we model population bottlenecks (and the genetic drift they induce, including Muller's ratchet) explicitly using multi-type branching processes (34, 35); such bottlenecks are not easily modelled within more common approaches based on weak-mutation strong-selection limit (32) or quasi-species theory (31, 33). Using general results in branching process theory, we derive expressions for the survival probability of a

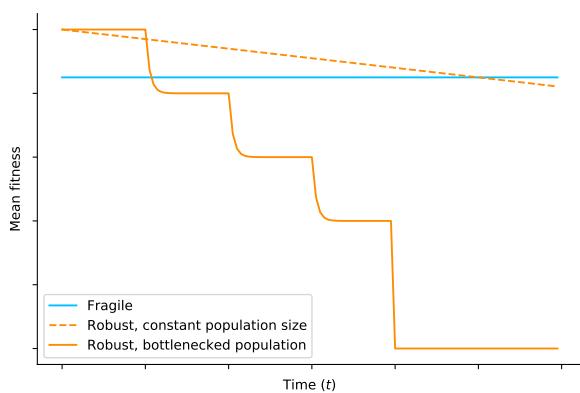


Fig. 1. Schematic of fitness dynamics. Robust genomes (orange) initially have a lower mutational load, and therefore higher population fitness, than fragile genomes (blue). But they also fix deleterious mutations more frequently, leading to a gradual decline in population fitness (dashed line); this effect is more pronounced when bottlenecks weaken the strength of selection and accelerate the clicking of Muller's ratchet (continuous line).

population through repeated bottlenecks as a function of the deleterious effect of mutations, confirming that fragility can be advantageous in the long run. Second, we consider the epidemiology of genetic fragility using a simple agent-based compartmental model. Evolutionary epidemiology (36) is an emerging field focusing on the interactions between evolutionary and epidemiological dynamics that seeks to explain the evolution of virulence and other properties of pathogens. Here we ask under what epidemiological conditions a fragile strain can have a higher attack rate than a robust one. We find that two parameters determine whether or not this is possible: the mutation rate u and the infection rate β (both relative to the recovery rate), with high u and low β both favouring fragile viruses.

Our model builds upon the standard model of Muller's ratchet (13), under which all mutations have the same deleterious effect (in particular, none is lethal), and do not interact epistatically. Positive epistasis among deleterious mutations has in fact been shown to be a pervasive phenomenon in compacted RNA genomes (37). Furthermore, the fraction of mutations that are lethal is usually large for RNA viruses (38). We rationalize these simplifications by noting that relaxing them would further increase the long-term advantage of fragile genomes, reinforcing the argument for the bottleneck hypothesis.

Results

Muller's ratchet in expanding viral populations. Consider a small viral population with absolute fitness $w_0 > 1$ and genomic mutation rate u . Assume that all mutations are deleterious with the same effect s_d , such that an individual carrying i mutations—an “ i -mutant”—has fitness $w_i \equiv w_0(1 - s_d)^i$. The dynamics of such a population falls under two broad alternatives: either it goes extinct through demographic fluctuations, or it grows to unbounded sizes with an asymptotic mean fitness $1 < \bar{w}_\infty \leq w_0$; the latter outcome is only possible if $w_0 e^{-u} > 1$. This is illustrated in Fig. 2 for a low value of s_d (a “robust” type) and a high value

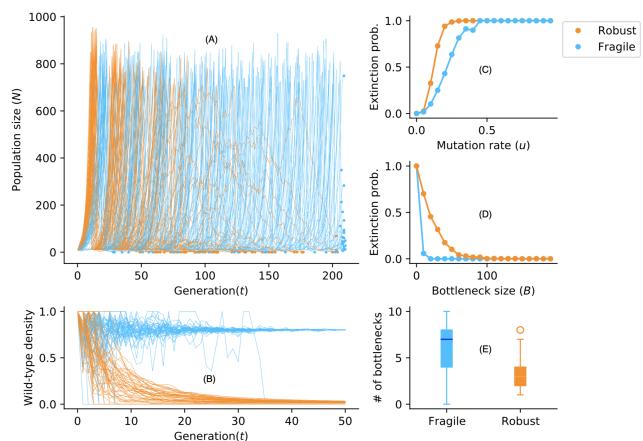


Fig. 2. Viral populations through multiple bottlenecks. For an initial population size $N = 10$ with fitness $w_0 = 1.5$ and a mutation rate $u = 0.2$, viral particles reproduce, mutate, and die out. Once the populations reach the carrying capacity $C = 800$, a sub-population of size $B = 10$ is sampled and the branching process is restarted. (A) Populations of robust viruses grow faster but go extinct more often after multiple bottlenecks. (B) The wild-type (individual with fitness w_0) density in the populations over time. (C) Effect of mutation rate on the extinction probability for a fixed bottleneck size $B = 10$. (D) Effect of B for a fixed mutation rate ($u = 0.2$) on the extinction probability. (E) Number of bottlenecks before extinction.

of s_d (a “fragile” type).

Key to the fate of the population is the onset of Muller's ratchet, *i.e.* the extinction of fit genotypes through genetic drift in small populations. The ratchet mostly clicks during the early states of the expansion, when the population size is smallest and extinction is likely. But it may also start clicking later through some rare fluctuation; if this click is followed by another click, and then another, the population can start shrinking again into mutational meltdown. If the ratchet clicks too many times, namely more than $K = \max\{k : w_k e^{-u} > 1\}$ times, extinction is unavoidable: any mutant carrying more than K mutations has absolute fitness smaller than one, and therefore generates a subcritical branching process.

Robust and fragile populations experience Muller's ratchet differently. For a robust genotype, mutations have a small deleterious effect, and are therefore under weak negative selection. This makes a click of the ratchet quite likely. On the other hand, each click comes at a low fitness cost, and so the population can withstand a relatively large number of clicks. Fragile populations, by contrast, are under strong negative selection against deleterious mutations, hence clicks are rarer; when they do occur, though, extinction becomes almost certain. Which fares better?

Using branching process theory we compute analytically the probability p_k that the fittest surviving individual carries exactly k mutations, *i.e.* that Muller's ratchet click k times (Eq. (1) in Materials and Methods). From this, we find that both the survival probability $p_{\text{surv}} = \sum_{k=0}^K p_k$ and the expected asymptotic population mean fitness $\mathbb{E}(\bar{w}_\infty)$ (Eq. (2)) are decreasing functions of s_d , consistent with the idea that, when all mutations are deleterious, mutational robustness is an evolutionary advantage. But there is a caveat: because Muller's ratchet clicks less frequently when genomes are fragile, the populations that do emerge from the expansion

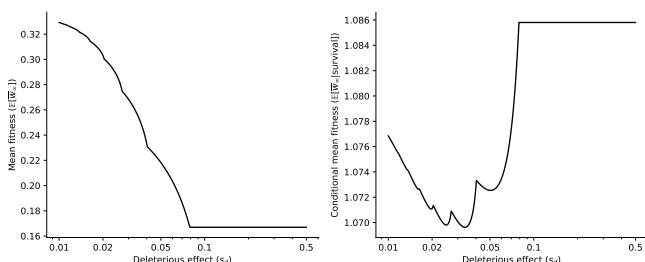


Fig. 3. Analytical results for expected asymptotic population mean fitness, both unconditional (left) and conditional on survival (right) as a function of the deleterious effect of mutations s_d , for a population with $w_0 = 1.2$ and $u = 0.1$. While a more robust strain has a lower mutational load and therefore a higher population mean fitness (left), it is also more vulnerable to Muller's ratchet, implying that surviving populations tend to have lower mean fitness (right).

tend to be mutations-free. As a result, the asymptotic mean population fitness *conditional on survival* $\mathbb{E}(\bar{w}_\infty | \text{survival})$ turns out to be non-monotonic in s_d , and in fact to be maximized for large deleterious effects (Fig. 3).

Survival of viral population through multiple bottlenecks. The adaptive value of virus' mutational fragility becomes apparent when we consider a succession of bottleneck-expansion cycles (Fig. 2), corresponding to viral transmission followed by within-host replication. This process can be modeled as a Markov chain on the space of post-bottleneck populations. Let B denote the size of the population after the transmission bottleneck. (In some cases, B can be a small as 1 (39, 40)).

We model virus transmission as sampling without replacement from a surviving population as described above, resulting in a new founding population with composition $\mathbf{n} = (n_0, \dots, n_K)$, where n_i is the number of i -mutants and we omit the subcritical mutants whose lineage will go extinct with probability one (hence $\sum_{k=0}^K n_k \leq B$). After within-host expansion and transmission sampling, this population will give rise to a new founding population with composition $\mathbf{m} = (m_0, \dots, m_K)$ with a probability $P(\mathbf{n} \rightarrow \mathbf{m})$ given explicitly in Eq. (15) in Materials and Methods. From this Markov chain, we can compute the probability that a population can survive any given number of bottlenecks (Eq. (4)). Fig. 4 shows the survival probability after up to five bottlenecks of size $B = 5$ as a function of the deleterious effect s_d . Although non-monotonic, this probability is maximized at large s_d when the number of bottlenecks increases, *i.e.* extinction becomes less likely for more fragile genomes. The results in the previous paragraph explain why: fragile populations are protected against Muller's ratchet, hence each new infection starts from fit founders. By contrast, robust genomes accumulate deleterious mutations; after several transmission bottlenecks, the founder particles tend to have low fitness and become increasingly unlikely to give rise to surviving lineages. It is as if robustness promoted mutational meltdown on a longer time scale—a meta-population meltdown.

Epidemiology of fragility. How would these effects play out in an epidemic outbreak? Would the lower propensity

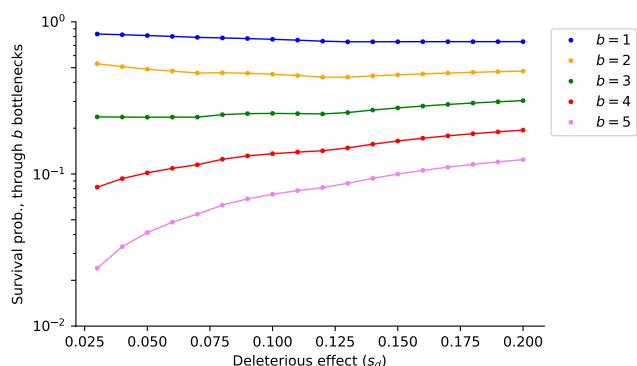


Fig. 4. Analytical results for the survival probability vs. mutational fragility for a population with $w_0 = 1.2$ and $u = 0.05$ undergoing b bottlenecks of size $B = 5$, computed using the bottleneck-to-bottleneck Markov chain $P(\mathbf{n} \rightarrow \mathbf{m})$. The higher the number of bottlenecks a populations has to survive, the more fragility is selected for.

of fragile genomes to suffer meltdown make up for their lower initial population fitness? To investigate this question we consider a Susceptible-Exposed-Infectious-Recovered (SEIR) model defined as follows. When a susceptible S meets an infectious individual I , a sample of the latter's viral population with size B is transmitted and S becomes exposed E . After this event, the within-host viral population carried by E can either (i) go extinct, in which case E returns to the susceptible compartment ($E \rightarrow S$) or (ii) grow exponentially during an incubation period τ until it reaches a critical threshold C which makes the host infectious ($E \rightarrow I$). Which is more likely depends on the viral genetic parameters (w_0, s_d, u) and, as shown in the previous section, on the numbers of supercritical viral particles transmitted to the new host, described by some vector \mathbf{n} . The incubation time is random as well, with a probability distribution depending on \mathbf{n} and on the number of clicks of Muller's ratchet during the expansion, see Eq. (5). (We show in *SI text* that this distribution can be approximated by a Gamma distribution depending on these parameters.) The host population is assumed to be well-mixed (no spatial structure): at each time step and for each pair (S, I) , transmission occurs with a probability β . Recovery in turn takes place at a rate γ .

Fig. 5 presents the results of simulations where a susceptible population of size $N = 5000$ is seeded with 100 infectious individuals, half of which carry a robust viral strain ($s_d = 0.05$) and the other half a fragile one ($s_d = 0.9$). Three regimes emerge depending on the transmission rate β and the mutation rate u (at fixed bottleneck size $B = 5$). When infectivity is high and the population quickly becomes completely infected, the robust strain fares better due to its higher initial fitness and faster growth rate. When infectivity is low, however, fragile strains prove to have a much higher attack rate. (We define the attack rate of a strain as the fraction of the host population which is exposed, infectious or recovered with that strain.). As noted in Fig. 5A, the robust strains are more virulent in the early stages of the epidemic. But as time goes on, the robust genomes loose fitness and increase their post-bottleneck extinction probabilities. That is also the reason why there is a small decrease in the total number of

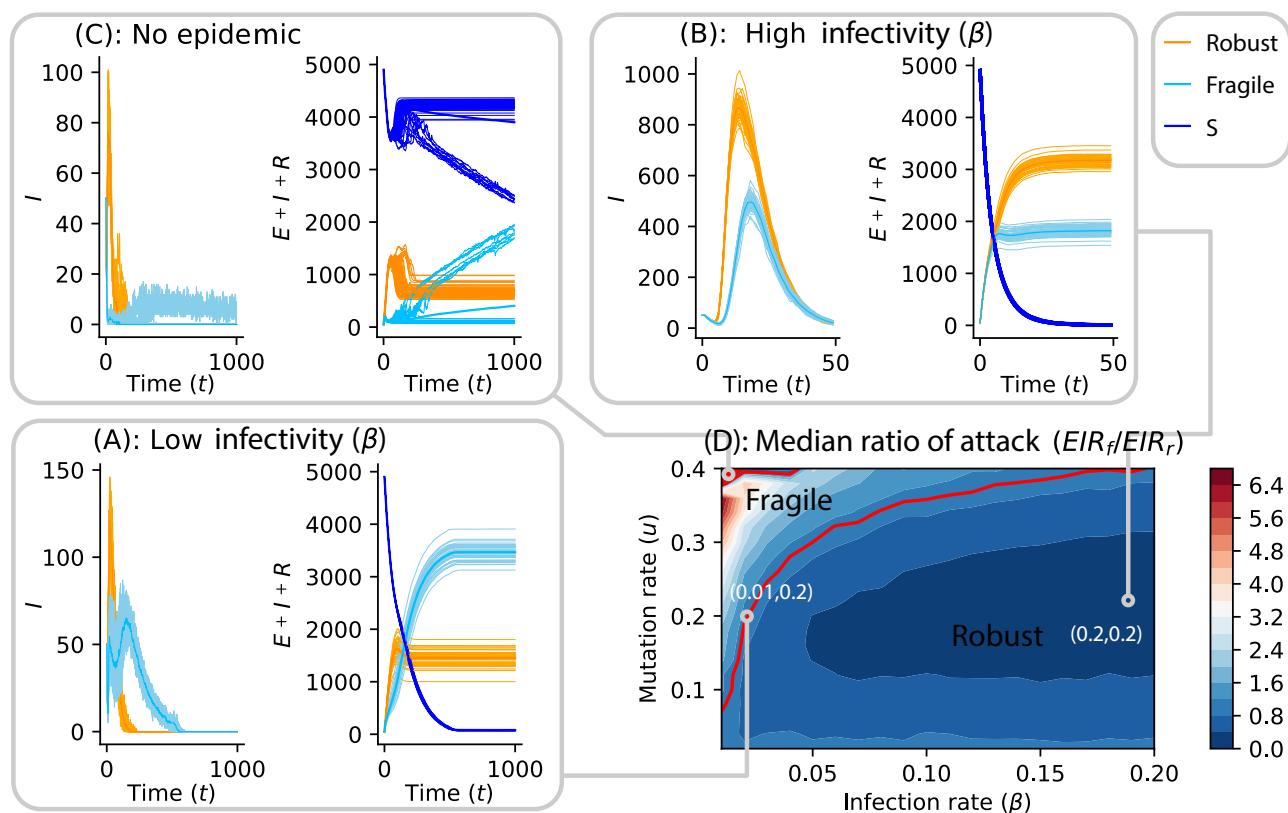


Fig. 5. Agent-based simulation of the SEIR model. (A) At low infection rate ($\beta = 0.01$) and high mutation rate $u = 0.2$, the robust genomes are more infectious in the beginning of the epidemic than the fragile ones, but as time goes on, it turns out that the fragile genomes become more virulent. (B) Using the same mutation rate and the highest infectivity gives the robust genome a chance to become the most infectious because it reproduces faster, and its population undergoes fewer bottlenecks. (C) At low infection rate and very high mutation rate ($u = 0.4$), the disease does not spread in the population, due to the high extinction probability of the viral populations. (D) Median ratio of attack rate (which is the total number of fragile infections divided by the robust ones) across 50 runs for each (β, u) parameters.

robust genome infections in $(E + R + I)$ -populations, as exposed hosts return to the susceptible compartment (Fig. 5A). At a late stage ($t > 200$), mutationally robust viruses are too degraded to make their host infectious. Finally, very high mutation rates and low infectivities do not allow for the epidemic to spread, as all viral populations (robust and fragile) quickly go extinct. These findings are summarized in Fig. 5D in terms of the median ratio of attack rates across 50 runs for each parameter set.

Underlying this contrast is the distribution of incubation times, which differs for the robust and fragile strains (Fig. 6A and Fig. 6B). The same observation can be drawn by looking at the mean basic reproduction number of both robust and fragile strains at low infectivity (and respectively at high infectivity) (see Fig. 6C and Fig. 6D). Over time, the basic reproduction number R_t of fragile strains stays constant while the robust one decreases.

Discussion

There is a growing appreciation for the fact that population bottlenecks are not just a fundamental aspect of the life cycle of viruses, but that they also play a key role in their evolution (41, 42). The effect of bottlenecks is not always detrimental: bottlenecks can effectively remove cheaters, *e.g.* defective interfering viruses (43), or enhance the effectiveness of selection if beneficial alleles act in *trans* (44). In addition,

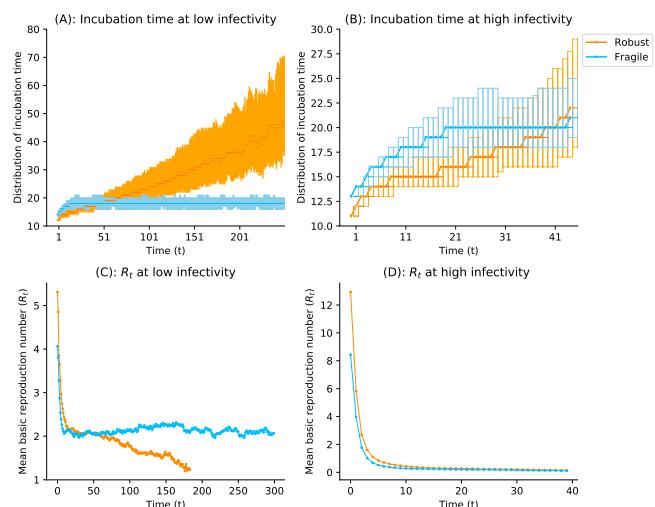


Fig. 6. Incubation time distributions and mean basic reproduction number at low infectivity ($\beta = 0.01$) (respectively at high infectivity ($\beta = 0.2$)), with viral parameters ($w_0 = 1.5$, $u = 0.1$) and bottleneck size $B = 5$. (A) The epidemic lasts longer and the robust viral populations lose fitness. The later transmitted viral particles take a longer time to grow up to C . In contrast, the incubation times of the fragile strains stay on average constant. (B) On short-time epidemic, the mean incubation time of the robust strain stays smaller than the fragile ones. (C) The epidemic lasts longer, wherein the beginning, the reproduction number of the robust strain is higher than the fragile one but later on, when the robust strain's R_t decreases, the fragile one stays constant. (D) The virus takes over the population in the early stage of the epidemic when the robust strains are the most virulent.

genetic bottlenecks can facilitate traveling across the characteristically rugged fitness landscapes of RNA viruses (37), where it is easy for viruses to become trapped at suboptimal fitness peaks (45, 46); by relaxing the intensity of selection, bottlenecks enable the exploration of new regions of the landscape.

In this paper we have explored another aspect of highly mutable populations subjected to periodic bottlenecks: they experience a strong evolutionary pressure towards genetic fragility. Earlier work has established that intermediate deleterious effects s_d maximize the strength and speed of Muller's ratchet (16, 31); similar results have been reported more recently in terms of “ratchet robustness” (47) or “drift robustness” (32). We find this U-shaped pattern in the context of expanding populations as well, *e.g.* for the asymptotic population mean fitness given survival (Fig. 3); the unconditional population mean fitness, by contrast, always decreases with fragility, which is consistent with another analysis of the advantage of mutational robustness (33). By modelling changes in population sizes with branching processes, we allowed a more complete picture to emerge. In this picture, the evolution of high neutrality (27, 28) is not incompatible with selection for maximally deleterious mutations (31) and anti-redundancy (33). An agent-based SEIR epidemiological model further reveals the determinants of genetic fragility, highlighting the importance of epidemic transmission parameters.

The evolution of viruses is often described in terms of fitness landscapes and their topographies. Our findings highlight the limitation of this picture: the motion of evolving population in genotype space depends on the structure of the genotype-phenotype-fitness mapping, but also on mutation rates and life cycle parameters such as the frequency and stringency of population bottlenecks. As a result, the evolution of mutational robustness—or mutational fragility—cannot be construed solely as the search for an optimal region in the fitness landscape, be it the highest peak or the flattest plateau. Our multi-type branching process approach is suggestive that a composite definition of fitness might be more predictive of evolutionary success in the present context, namely, a definition that takes account of both offspring number (Malthusian fitness) and long-term survival probability. The augmented evolutionary relevance of this definition to the present context is manifest in the comparison between the left and right panels of Fig. 3.

To be sure, our model relies on simplifying assumptions, mainly pertaining to the nature of underlying mutational landscapes and to epidemiological details. The assumption that all mutations having the same deleterious effect is a common one that simplifies the mathematics. Fitness effects of deleterious mutations are more realistically modeled as a random variable with a continuous, heavy-tailed distribution (*e.g.* the Gamma and Weibull distributions have been previously used to satisfactorily fit experimental data (48, 49)) or a U-shaped distribution to incorporate lethal mutations. As already noted, we make another common assumption that does not necessarily hold for real viral genomes, namely, the inde-

pendence of mutational effects. Evidence pervasively suggests that positive epistasis is the norm for compacted viral RNA genomes (37), including many instances of compensatory mutations. Indeed, it was shown long ago that if deleterious alleles interact synergistically, they are more efficiently removed from the population and thereby slow the advance of Muller's ratchet (50). Finally, our evolutionary epidemiology approach ignores co- or super-infection cases wherein multiple viral strains infect the same host. On the one hand, multiple infections allow competition among strains to occur at the level of individual lineages. On the other hand, multiple infections allow the sharing of gene products among different genotypes within a cell, thus compensating for deleterious effects and eventually contributing to the accumulation of more mildly deleterious mutations in the population (51). In its current form, our model may well serve as the null against which to test the effect of all these important factors in the fate of evolving viral populations.

Overall, our findings can be summarized with a classic metaphor. Robust genomes are hares: they grow fast but accumulate mutational damage through Muller's ratchet, which jeopardizes their potential for long-term survival. Fragile genomes are turtles: they grow more slowly but weather bottlenecks more reliably and have higher long-term survival probabilities. Turtles may seem weak individually, but as a group they have survived for hundreds of millions of years. Similarly, fragile viral genomes are individually vulnerable to deleterious mutations; at the meta-population level, however, they may hold the key to evolutionary resilience.

Materials and Methods

The evolution of viral populations experiencing Muller's ratchet and going through bottlenecks is modeled by a multi-type branching process, in which a “type” corresponds to the number of accumulated deleterious mutations. We refer to *SI text* for a detailed description of the model and a rigorous demonstration of the following results.

Starting with n supercritical mutants, the extinction probability $p_{\text{ext},k}(\mathbf{n})$ of all types up to type k in the population is given by the element-wise product \mathbf{q}_k^n , where \mathbf{q}_k is the smallest solution to a fixed-point equation involving the generating function of the branching process. Muller's ratchet click probability $p_k(\mathbf{n})$ that the fittest surviving individuals carry k mutations is then

$$p_k(\mathbf{n}) = p_{\text{ext},k-1}(\mathbf{n}) - p_{\text{ext},k}(\mathbf{n}) \quad (1)$$

and the population survival probability $p_{\text{surv}}(\mathbf{n}) = \sum_{k=0}^K p_k(\mathbf{n})$.

On the event that the fittest surviving individuals carry k mutations, the asymptotic mutant spectrum of the population is given by $(0, \dots, 0, f_0, f_1, \dots)$, where f_i is the Poisson frequency $f_i = e^{-u/s_d} (u/s_d)^i / i!$. The asymptotic mean population fitness is thus a random variable \bar{w}_∞ equal to $w_k e^{-u}$ with probability $p_k(\mathbf{n})$ for $0 \leq k \leq K$, and to 0 otherwise.

Therefore

$$\begin{cases} \mathbb{E}(\bar{w}_\infty) = e^{-u} \sum_{k=0}^K w_k p_k(\mathbf{n}), \\ \mathbb{E}(\bar{w}_\infty \mid \text{survival}) = e^{-u} \sum_{k=0}^K w_k p_k(\mathbf{n}) / p_{\text{surv}}(\mathbf{n}). \end{cases} \quad (2)$$

When going through a bottleneck, this population gives rise to a new founding population with composition \mathbf{m} , according to transition probability

$$P(\mathbf{n} \rightarrow \mathbf{m}) = \sum_{k=0}^K Q_k(\mathbf{m}) p_k(\mathbf{n}) + \mathbf{1}_{\mathbf{m}=\mathbf{0}} (1 - p_{\text{surv}}(\mathbf{n})), \quad (3)$$

where $Q_k(\mathbf{m})$ is the probability of getting \mathbf{m} supercritical mutants in the sample of size B . The resulting transition matrix P then provides an explicit expression of the probability for a viral population with initial state \mathbf{n} to become extinct after going through at most b bottlenecks:

$$p_{\text{bottlenecks},b}(\mathbf{n}) = \mathbf{P}^b(\mathbf{n} \rightarrow \mathbf{0}). \quad (4)$$

In our SEIR epidemiological model, the time spent in the exposed state depends on \mathbf{n} and on the number of Muller's ratchet clicks during the expansion, or equivalently on the type k of the fittest surviving individuals. We show that this incubation period $\tau_{\mathbf{n},k}$ is

$$\tau_{\mathbf{n},k} = \left\lceil \frac{\ln(C/W_{\mathbf{n},k}) - u/s_d}{\ln w_k - u} \right\rceil, \quad (5)$$

where $W_{\mathbf{n},k}$ is a positive random variable and $\lceil \cdot \rceil$ stands for the ceiling function. As exposed in *SI text*, $W_{\mathbf{n},k}$ can be expressed as the sum of a random number of independent exponential random variables with common parameter

$$1 + \frac{1}{w_k e^{-u}} W \left[-w_k e^{-u} e^{-w_k e^{-u}} \right],$$

where W stands for the principal solution of the Lambert function.

Simulations were carried out using an agent-based approach in Python. We have created a repository at github.com/strevol-mpi-mis/EvoEpi containing the code and minimum documentation.

ACKNOWLEDGEMENTS

We thank the members of the Structure of Evolution group at MPI MIS for useful discussions. Funding for this work was provided by the Alexander von Humboldt Foundation in the framework of the Sofja Kovalevskaja Award endowed by the German Federal Ministry of Education and Research to M.S. Work in València was supported by Spain Agencia Estatal de Investigación - FEDER grant PID2019-103998GB-I00 and Generalitat Valenciana grant PROMETEO2019/012 to S.F.E.

Bibliography

1. Santiago F. Elena and Rafael Sanjuán. Virus Evolution: Insights from an Experimental Approach. *Annual Review of Ecology Evolution, and Systematics*, 38(1):27–52, dec 2007. doi: 10.1146/annurev.ecolsys.38.091206.095637.
2. Edward C. Holmes. The Evolutionary Genetics of Emerging Viruses. *Annual Review of Ecology Evolution, and Systematics*, 40(1):353–372, dec 2009. doi: 10.1146/annurev.ecolsys.110308.120248.
3. Raul Andino and Esteban Domingo. Viral quasispecies. *Virology*, 479-480:46–51, may 2015. doi: 10.1016/j.virol.2015.03.022.
4. Esteban Domingo and Celia Perales. Viral quasispecies. *PLOS Genetics*, 15(10):e1008271, oct 2019. doi: 10.1371/journal.pgen.1008271.
5. Santiago F. Elena and Rafael Sanjuán. Adaptive Value of High Mutation Rates of RNA Viruses: Separating Causes from Consequences. *Journal of Virology*, 79(18):11555–11558, sep 2005. doi: 10.1128/jvi.79.18.11555-11558.2005.
6. Robert Belshaw, Andy Gardner, Andrew Rambaut, and Oliver G. Pybus. Pacing a small cage: mutation and RNA viruses. *Trends in Ecology & Evolution*, 23(4):188–193, apr 2008. doi: 10.1016/j.tree.2007.11.010.
7. Lin Chao. Fitness of RNA virus decreased by Muller's ratchet. *Nature*, 348(6300):454–455, nov 1990. doi: 10.1038/348454a0.
8. Ruben O. Donis. Muller's ratchet and flu virus. *Nature*, 353(6342):308–309, sep 1991. doi: 10.1038/353308b0.
9. E. Duarte, D. Clarke, A. Moya, E. Domingo, and J. Holland. Rapid fitness losses in mammalian RNA virus clones due to Muller's ratchet. *Proceedings of the National Academy of Sciences*, 89(13):6015–6019, jul 1992. doi: 10.1073/pnas.89.13.6015.
10. Eloisa Yuste, Sonsoles Sánchez-Palomino, Concha Casado, Esteban Domingo, and Cecilio López-Gálvez. Drastic Fitness Loss in Human Immunodeficiency Virus Type 1 upon Serial Bottleneck Events. *Journal of Virology*, 73(4):2745–2751, apr 1999. doi: 10.1128/jvi.73.4.2745-2751.1999.
11. Francisca de la Iglesia and Santiago F. Elena. Fitness Declines in Tobacco Etch Virus upon Serial Bottleneck Transfers. *Journal of Virology*, 81(10):4941–4947, mar 2007. doi: 10.1128/jvi.02528-06.
12. H.J. Muller. The relation of recombination to mutational advance. *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, 1(1):2–9, may 1964. doi: 10.1016/0027-5107(64)90047-8.
13. John Haigh. The accumulation of deleterious genes in a population—Muller's Ratchet. *Theoretical Population Biology*, 14(2):251–267, oct 1978. doi: 10.1016/0040-5809(78)90027-8.
14. Wolfgang Stephan, Lin Chao, and Joanne Guna Smale. The advance of Muller's ratchet in a haploid asexual population: approximate solutions based on diffusion theory. *Genetical Research*, 61(3):225–231, jun 1993. doi: 10.1017/s0016672300031384.
15. M. Lynch, R. Bürger, D. Butcher, and W. Gabriel. The Mutational Meltdown in Asexual Populations. *Journal of Heredity*, 84(5):339–344, sep 1993. doi: 10.1093/oxfordjournals.jhered.a111354.
16. W. Gabriel, M. Lynch, and R. Bürger. Muller's ratchet and mutational meltdowns. *Evolution*, 47(6):1744–1757, dec 1993. doi: 10.1111/j.1558-5646.1993.tb01266.x.
17. Andreas Wagner and Peter F. Stadler. Viral RNA and evolved mutational robustness. *Journal of Experimental Zoology*, 285(2):119–127, aug 1999. doi: 10.1002/(sici)1097-010x(19990115)285:2<119::aid-jez4>3.0.co;2-d.
18. Santiago F. Elena, Purificación Carrasco, José-Antonio Daròs, and Rafael Sanjuán. Mechanisms of genetic robustness in RNA viruses. *EMBO reports*, 7(2):168–173, feb 2006. doi: 10.1038/sj.embo.7400636.
19. Santiago F. Elena. RNA virus genetic robustness: possible causes and some consequences. *Current Opinion in Virology*, 2(5):525–530, oct 2012. doi: 10.1016/j.coviro.2012.06.008.
20. Adam S. Lauring, Judith Frydman, and Raul Andino. The role of mutational robustness in RNA virus evolution. *Nature Reviews Microbiology*, 11(5):327–336, mar 2013. doi: 10.1038/nrmicro3003.
21. José M. Cuevas, María Durán-Moreno, and Rafael Sanjuán. Multi-virion infectious units arise from free viral particles in an enveloped virus. *Nature Microbiology*, 2(7), may 2017. doi: 10.1038/nmicrobiol.2017.78.
22. J. Sardanyés and SF. Elena. Error threshold in RNA quasispecies models with complementation. *J Theor Biol*, 265:278–86, Aug 2010.
23. Fernanda Martínez, Josep Sardanyés, Santiago F. Elena, and José-Antonio Daròs. Dynamics of a Plant RNA Virus Intracellular Accumulation: Stamping Machine vs. Geometric Replication. *Genetics*, 188(3):637–646, apr 2011. doi: 10.1534/genetics.111.129114.
24. Rebecca Montville, Remy Froidart, Susanna K. Remold, Olivier Tenaillon, and Paul E. Turner. Evolution of Mutational Robustness in an RNA Virus. *PLoS Biology*, 3(11):e381, nov 2005. doi: 10.1371/journal.pbio.0030381.
25. S. Ojosnegros, J. García-Arriaza, C. Escarmís, SC. Manrubia, C. Perales, A. Arias, MG. Mateu, and E. Domingo. Viral genome stability can result from a trade-off between genetic content and particle stability. *PLoS Genet*, 7:e1001344, Mar 2011.
26. Ron Geller, Shuhui Taguwa, and Judith Frydman. Broad action of Hsp90 as a host chaperone required for viral replication. *Biochimica et Biophysica Acta (BBA) - Molecular Cell Research*, 1823(3):698–706, mar 2012. doi: 10.1016/j.bbamcr.2011.11.007.
27. E. van Nimwegen, J. P. Crutchfield, and M. Huynen. Neutral evolution of mutational robustness. *Proceedings of the National Academy of Sciences*, 96(17):9716–9720, aug 1999. doi: 10.1073/pnas.96.17.9716.
28. CO. Wilke, JL. Wang, C. Ofría, RE. Lenski, and C. Adami. Evolution of digital organisms at high mutation rates leads to survival of the flattest. *Nature*, 412:331–3, Jul 2001.
29. Francisco M. Codoñer, José-Antonio Daròs, Ricard V. Solé, and Santiago F. Elena. The Fittest versus the Flattest: Experimental Confirmation of the Quasispecies Effect with Sub-viral Pathogens. *PLoS Pathogens*, 2(12):e136, 2006. doi: 10.1371/journal.ppat.0020136.
30. Rafael Sanjuán. Mutational fitness effects in RNA and single-stranded DNA viruses: common patterns revealed by site-directed mutagenesis studies. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 365(1548):1975–1982, jun 2010. doi: 10.1098/rstb.2010.0063.
31. Brendan D. O'Fallon, Frederick R. Adler, and Stephen R. Proulx. Quasi-species evolution in subdivided populations favours maximally deleterious mutations. *Proceedings of the Royal Society B: Biological Sciences*, 274(1629):3159–3164, oct 2007. doi: 10.1098/rspb.2007.1228.
32. Thomas LaBar and Christoph Adami. Evolution of drift robustness in small populations. *Nature Communications*, 8(1), oct 2017. doi: 10.1038/s41467-017-01003-7.
33. D. C. Krakauer and J. B. Plotkin. Redundancy antiredundancy, and the robustness of genomes. *Proceedings of the National Academy of Sciences*, 99(3):1405–1409, jan 2002. doi: 10.1073/pnas.032668599.
34. Marek Kimmel and David E. Axelrod. *Branching Processes in Biology*. Springer New York, 2002. doi: 10.1007/b97371.
35. Patsy Haccou, Peter Jagers, and Vladimir A. Vatutin. *Branching Processes*. Cambridge

University Press, may 2005. doi: 10.1017/cbo9780511629136.

- 36. Olivier Restif. Evolutionary epidemiology 20 years on: Challenges and prospects. *Infection Genetics and Evolution*, 9(1):108–123, jan 2009. doi: 10.1016/j.meegid.2008.09.007.
- 37. SF Elena, RV Solé, and J Sardanyés. Simple genomes, complex interactions: epistasis in RNA virus. *Chaos*, 20:026106, Jun 2010.
- 38. R Sanjuán. Mutational fitness effects in RNA and single-stranded DNA viruses: common patterns revealed by site-directed mutagenesis studies. *Philos Trans R Soc Lond B Biol Sci*, 365:1975–82, Jun 2010.
- 39. Mark P. Zwart, Lia Hemerik, Jenny S. Cory, J. Arjan G.M. de Visser, Felix J.J.A. Bianchi, Monique M. Van Oers, Just M. Vlak, Rolf F. Hoekstra, and Wopke Van der Werf. An experimental test of the independent action hypothesis in virus–insect pathosystems. *Proceedings of the Royal Society B: Biological Sciences*, 276(1665):2233–2242, mar 2009. doi: 10.1098/rspb.2009.0064.
- 40. Mark P. Zwart, José-Antonio Darós, and Santiago F. Elena. One Is Enough: In Vivo Effective Population Size Is Dose-Dependent for a Plant RNA Virus. *PLoS Pathogens*, 7(7): e1002122, jul 2011. doi: 10.1371/journal.ppat.1002122.
- 41. C. T. Bergstrom, P. McElhany, and L. A. Real. Transmission bottlenecks as determinants of virulence in rapidly evolving pathogens. *Proceedings of the National Academy of Sciences*, 96(9):5095–5100, apr 1999. doi: 10.1073/pnas.96.9.5095.
- 42. Santiago F Elena, Rafael Sanjuán, Antonio V Bordería, and Paul E Turner. Transmission bottlenecks and the evolution of fitness in rapidly evolving RNA viruses. *Infection Genetics and Evolution*, 1(1):41–48, jul 2001. doi: 10.1016/s1567-1348(01)00006-5.
- 43. Mark P. Zwart and Santiago F. Elena. Matters of Size: Genetic Bottlenecks in Virus Infection and Their Potential Impact on Evolution. *Annual Review of Virology*, 2(1):161–179, nov 2015. doi: 10.1146/annurev-virology-100114-055135.
- 44. S Miyashita, K Ishibashi, H Kishino, and M Ishikawa. Viruses roll the dice: the stochastic behavior of viral genome molecules accelerates viral adaptation at the cell and tissue levels. *PLoS Biol*, 13:e1002094, Mar 2015.
- 45. CL Burch and L Chao. Evolvability of an RNA virus is determined by its mutational neighbourhood. *Nature*, 406:625–8, Aug 2000.
- 46. H Cervera, J Lalíć, and SF Elena. Efficient escape from local optima in a highly rugged fitness landscape by evolving RNA virus populations. *Proc Biol Sci*, 283, Aug 2016.
- 47. Yinghong Lan, Aaron Trout, Daniel Michael Weinreich, and Christopher Scott Wylie. Natural selection can favor ratchet robustness over mutational robustness. *BioRxiv*, mar 2017. doi: 10.1101/121087.
- 48. R. Sanjuán, A. Moya, and S. F. Elena. The contribution of epistasis to the architecture of fitness in an RNA virus. *Proceedings of the National Academy of Sciences*, 101(43): 15376–15379, oct 2004. doi: 10.1073/pnas.0404125101.
- 49. Purificación Carrasco, Francisca de la Iglesia, and Santiago F Elena. Distribution of fitness and virulence effects caused by single-nucleotide substitutions in tobacco etch virus. *Journal of virology*, 81(23):12979–12984, 2007.
- 50. Alexey S Kondrashov. Muller's ratchet under epistatic selection. *Genetics*, 136(4):1469–1473, 1994.
- 51. Nicolas Tromas, Mark P Zwart, Guillaume Lafforgue, and Santiago F Elena. Within-host spatiotemporal dynamics of plant virus infection at the cellular level. *PLoS Genet*, 10(2): e1004186, 2014.
- 52. K. B. Athreya and P. E. Ney. *Branching processes*. Dover Publications, Inc., Mineola, NY, 2004.
- 53. Sophie Hautphenne. Extinction probabilities of supercritical decomposable branching processes. *J. Appl. Probab.*, 49(3):639–651, 2012. ISSN 0021-9002. doi: 10.1239/jap/134695323.
- 54. H Kesten and B.P. Stigum. Limit theorems for decomposable multi-dimensional galton-watson processes. *Journal of Mathematical Analysis and Applications*, 17(2):309 – 338, 1967. ISSN 0022-247X. doi: [https://doi.org/10.1016/0022-247X\(67\)90155-2](https://doi.org/10.1016/0022-247X(67)90155-2).
- 55. F. Antoneli, F. Bosco, D. Castro, and L. M. Janini. *Viral evolution and adaptation as a multivariate branching process*, pages 217–243. World Scientific, 2013. doi: 10.1142/9789814520829_0013.

Supplementary Note 1: Muller's ratchet in expanding populations

Branching process setting. The composition of the viral population is determined by the number of virions carrying 0, 1, 2, *etc.* deleterious mutations. Its evolution is modeled by a branching process (52): each viral particle in generation n randomly produces particles in generation $n+1$, independently of the rest of the population and according to some probability distribution which depends solely on the "type" of the particle. Since there are no back mutations, the resulting process $(\mathbf{X}_n)_{n \in \mathbb{N}}$ is a reducible branching process with infinitely many types, where $\mathbf{X}_n = (X_{n,i})_{i \in \mathbb{N}}$ is the population composition at time n , and $X_{n,i}$ the number of i -mutants (particles carrying i mutations) at this time. All the properties of the branching process are contained in its offspring generating functions F_0, F_1, \dots . The generating function F_i determines the distribution of the number of various mutants to be produced by an i -mutant. By definition, for $\mathbf{r} \in [0, 1]^{\mathbb{N}}$, $F_i(\mathbf{r})$ is the expected value of $r_0^{X_{1,0}} r_1^{X_{1,1}} \dots$ when the process is initiated by one i -mutant. Because there are no back mutations, it solely depends on r_{i+j} , $j \geq 0$, and we write $F_i(r_i, r_{i+1}, \dots)$.

We assume throughout this paper that the average number of $(i+j)$ -mutants produced by an i -mutant is $w_i e^{-u} u^j / j!$. This is achieved for instance if the total number of offspring of an i -mutant follows a Poisson distribution $\mathcal{P}(w_i)$, and if each of its offspring accumulates a number of additional deleterious mutations according to $\mathcal{P}(u)$. In this particular case, the i -th generating function is

$$F_i(r_i, r_{i+1}, \dots) = \sum_{k \in \mathbb{N}} e^{-w_i} \frac{w_i^k}{k!} \sum_{\substack{k_i, k_{i+1}, \dots \in \mathbb{N} \\ \sum_{j \geq 0} k_{i+j} = k}} \frac{k!}{k_i! k_{i+1}! \dots} \prod_{j \geq 0} \left(e^{-u} \frac{u^j}{j!} r_{i+j} \right)^{k_{i+j}}$$

which simplifies to

$$F_i(r_i, r_{i+1}, \dots) = \exp \left[-w_i \left(1 - e^{-u} \sum_{j \geq 0} \frac{u^j}{j!} r_{i+j} \right) \right]. \quad (6)$$

Survival probability. Let $m_i = w_i e^{-u}$ the average number of i -mutants produced by an i -mutant. It follows from a general result of branching processes theory that if $m_i \leq 1$, any population starting with one i -mutant is doomed to extinction; if $m_i > 1$, this population has positive survival probability. Let $K = \max\{k : m_k > 1\}$ the largest number of mutations which can be carried by an individual without leading to its progeny's extinction, and call *supercritical* any k -mutant with $k \leq K$. By independence of the lineages, the fate of the population $(\mathbf{X}_n)_{n \in \mathbb{N}}$ only depends on the initial numbers of supercritical mutants $\mathbf{n} = (X_{0,0}, \dots, X_{0,K})$. First, we compute for each $k \leq K$ the *partial extinction probability* $p_{\text{ext},k}(\mathbf{n})$ of all mutants up to type k , given the initial condition \mathbf{n} , namely the probability of $\{X_{n,i} \rightarrow 0, \forall i \leq k\}$. It corresponds to the extinction probability of the reducible branching process with finite set of types $\{0, 1, \dots, k\}$ and generating functions $F_i(r_i, \dots, r_k, 1, 1, \dots)$, $i \leq k$. As such (53), the partial extinction probability is given by

$$p_{\text{ext},k}(\mathbf{n}) = \prod_{i=0}^k q_{ki}^{n_i}, \quad (7)$$

where (q_{k0}, \dots, q_{kk}) is the smallest nonnegative solution of the system of equations

$$F_i(q_{k0}, \dots, q_{kk}, 1, 1, \dots) = q_{ki}, \quad 0 \leq i \leq k, \quad (8)$$

(see Eq. (24) for an explicit solution when F_i is given by Eq. (6)). The overall survival and extinction probabilities of the population are then obtained as

$$p_{\text{surv}}(\mathbf{n}) = 1 - p_{\text{ext}}(\mathbf{n}) = 1 - p_{\text{ext},K}(\mathbf{n}). \quad (9)$$

Muller's ratchet click probabilities. For a population starting with supercritical mutants \mathbf{n} and $k \leq K$, let $p_k(\mathbf{n})$ the probability that the fittest surviving individuals carry k mutations. Equivalently, it is the probability that Muller's ratchet clicks $k - k_0$ times, where $k_0 = \min\{i : n_i > 0\}$ is the fittest initial supercritical type. It corresponds to event $\{X_{n,k} \rightarrow 0, X_{n,i} \rightarrow 0, \forall i < k\}$ (all i -mutants with $i < k$ go extinct but k -mutants survive), therefore

$$\begin{cases} p_0(\mathbf{n}) = 1 - p_{\text{ext},0}(\mathbf{n}), \\ p_k(\mathbf{n}) = p_{\text{ext},k-1}(\mathbf{n}) - p_{\text{ext},k}(\mathbf{n}), \end{cases} \quad (10)$$

It comes in particular $p_{\text{surv}}(\mathbf{n}) = \sum_{k=0}^K p_k(\mathbf{n})$, consistent with the fact that any click beyond the threshold K leads to extinction.

Mutant spectra and asymptotic mean fitness. The late-time composition of the population is completely determined by the number of clicks of the ratchet. On the event Ω_k that the type of its fittest surviving individuals is k , the asymptotic behavior of the population can be described with the help of the branching process \mathbf{Z}_n with offspring generating functions $G_i = F_{i+k}$, $i \geq 0$, which only considers particles with type greater than k . To study this process we use results from the theory on decomposable branching processes (54), also used in (55) in the context of a viral population. The offspring mean matrix of the process is the upper triangular matrix $\mathbf{M} = (m_{k+i,k+j})_{i,j \geq 0}$, where m_{ij} is the average number of j -mutants born from an i -mutant in the original process \mathbf{X}_n . Its largest eigenvalue is $m_k > 1$, and $\mathbf{u} = (1, 0, \dots)$, $\mathbf{v} = ((u/s_d)^i/i!)_{i \geq 0}$ are respectively non-negative right and left eigenvectors of \mathbf{M} corresponding to m_k , satisfying $\mathbf{u} \cdot \mathbf{v}' = 1$. Assuming $\mathbf{Z}_0 = (1, 0, \dots)$, then $m_k^{-n} \mathbf{Z}_n$ almost surely converges to $W\mathbf{v}$, where W is a non-negative random variable with mean value 1, and $\{W = 0\} = \{\mathbf{Z}_{n,0} \rightarrow 0\}$. If $\mathbf{Z}_0 = (z_0, 0, \dots)$, then by independence of the lineages $m_k^{-n} \mathbf{Z}_n \rightarrow W^{*z_0} \mathbf{v}$, where W^{*z_0} is the sum of z_0 independent copies of W . The limit remains the same if $\mathbf{Z}_0 = (z_0, z_1, \dots)$, since the z_i i -lineages with $i > 0$ have growth rate $m_{k+i} < m_k$ and tend to 0 if normalized by m_k^n . It follows that for any initial value \mathbf{Z}_0 , $\mathbf{Z}_n / \sum_j Z_{n,j} \rightarrow e^{-u/s_d} \mathbf{v}$ almost surely on the event $\{\mathbf{Z}_{n,0} \rightarrow 0\}$.

Let $N_n = \sum_i X_{n,i}$ be the total population size at time n and let $f_i = e^{-u/s_d} (u/s_d)^i / i!$. We deduce from what precedes that the fraction of i -mutants ($i \geq k$) in the population satisfies

$$\lim_n \frac{X_{n,i}}{N_n} = f_{i-k} \quad \text{almost surely on } \Omega_k, \quad (11)$$

while this proportion converges to 0 if $i < k$. Since $(\Omega_k)_{0 \leq k \leq K}$ forms a partition of the survival set, the previous result can be stated as follows (setting the frequency to 0 if $N_n = 0$):

$$\lim_n \frac{X_{n,i}}{N_n} = Y_i \quad \text{almost surely}, \quad (12)$$

where Y_i is a random variable equal to f_{i-k} with probability $p_k(\mathbf{n})$, $k \leq \min(i, K)$, and equal to 0 with probability $p_{\text{ext}}(\mathbf{n}) + \sum_{k=i+1}^K p_k(\mathbf{n})$. Therefore, the mean population fitness satisfies

$$\lim_n \sum_{i \geq 0} w_i \frac{X_{n,i}}{N_n} = \bar{w}_\infty \quad \text{almost surely}, \quad (13)$$

where $\bar{w}_\infty = \sum_{i \geq 0} w_i Y_i$ is a random variable equal to $w_k e^{-u}$ with probability $p_k(\mathbf{n})$, $0 \leq k \leq K$, and equal to 0 with probability $p_{\text{ext}}(\mathbf{n})$. It follows that the expected asymptotic population mean fitness is given by

$$\begin{cases} \mathbb{E}(\bar{w}_\infty) = e^{-u} \sum_{k=0}^K w_k p_k(\mathbf{n}), \\ \mathbb{E}(\bar{w}_\infty \mid \text{survival}) = e^{-u} \sum_{k=0}^K w_k p_k(\mathbf{n}) / p_{\text{surv}}(\mathbf{n}). \end{cases} \quad (14)$$

Supplementary Note 2: Survival through multiple bottlenecks

Markov chain model. In order to assess the effect of successive bottleneck events on the viral population, we introduce a stochastic model describing the evolution of the random composition of the population after each size reduction event. If the population reaches its carrying capacity C , a sample of size B is taken, resulting in a new founding population. As described earlier, the fate of this new population only depends on its initial numbers of supercritical mutants \mathbf{n} , with $|\mathbf{n}| = \sum_{k=0}^K n_k \leq B$ (the sample might include some particles which are not supercritical). We choose C large enough such that (i) the probability of not reaching C is close to the extinction probability of the population, (ii) if it reaches C , the mutant frequencies in the population follow the asymptotic random distribution Eq. (12). The evolution of the viral population going through these multiple size reductions is then entirely determined by the successive states $\mathbf{n}_0, \mathbf{n}_1, \dots$, where \mathbf{n}_b describes the supercritical fitness classes after b bottleneck events. We describe this random evolution via a Markov chain with state space $\mathcal{S} = \{\mathbf{n} \in \mathbb{N}^{K+1}, |\mathbf{n}| \leq B\}$, and transition probability from state \mathbf{m} to state \mathbf{n} :

$$P(\mathbf{m} \rightarrow \mathbf{n}) = \sum_{k=0}^K Q_k(\mathbf{n}) p_k(\mathbf{m}) + \mathbf{1}_{\mathbf{n}=\mathbf{0}} p_{\text{ext}}(\mathbf{m}), \quad (15)$$

where $Q_k(\mathbf{n})$ is the probability of obtaining \mathbf{n} supercritical mutants in the sample of size B , if the fittest surviving individuals in the population founded by \mathbf{m} are of type k (event of probability $p_k(\mathbf{m})$). It follows from Eq. (11) that on the latter event, the asymptotic proportion of non-supercritical mutants is $1 - \sum_{i=0}^{K-k} f_i$, while the asymptotic proportion of supercritical i -mutants is f_{i-k} if $i \geq k$, and 0 otherwise. The random composition of the sample of size B thus follows a multinomial distribution

with B trials and success probabilities $\{0, \dots, 0, f_0, \dots, f_{K-k}, 1 - \sum_{i=0}^{K-k} f_i\}$, the last coordinate corresponding to the non-supercritical mutants. Therefore, the probability of getting \mathbf{n} supercritical mutants is

$$Q_k(\mathbf{n}) = \frac{B! \sum_{i=0}^{k-1} n_i \left(1 - \sum_{i=0}^{K-k} f_i\right)^{B-|\mathbf{n}|} \prod_{i=0}^{K-k} f_i^{n_i+k}}{n_0! \dots n_K! (B - |\mathbf{n}|)!}. \quad (16)$$

Extinction probability under multiple bottlenecks. Let $\mathbf{P} = (P(\mathbf{m} \rightarrow \mathbf{n}))_{\mathbf{m}, \mathbf{n} \in \mathcal{S}}$ the stochastic matrix of this Markov chain, and \mathbf{P}^b its b -th power. Then the probability for a viral population with initial state \mathbf{n}_0 to become extinct after going through at most b bottlenecks is

$$p_{\text{bottlenecks}, b}(\mathbf{n}_0) = \mathbf{P}^b(\mathbf{n}_0, \mathbf{0}), \quad (17)$$

i.e. entry $(\mathbf{n}_0, \mathbf{0})$ of matrix \mathbf{P}^b . This probability can thus be computed explicitly as soon as the extinction and click probabilities p_{ext} and p_k are known, which is the case if for instance the numbers of offspring and of accumulated mutations follow Poisson distributions (see Eq. (24)).

Supplementary Note 3: Epidemiology of genetic fragility

Incubation period distribution. How long does it take for the in-host population carried by an exposed individual E to reach size C ? Assuming C is large enough, this random time can be approximated thanks to the asymptotic behavior of the population. Let \mathbf{n} the initial numbers of supercritical viral particles in the in-host population. As described earlier, if the population does not go extinct, its limit behavior depends on the type k of its fittest surviving particles. The distribution of the incubation period thus depends on \mathbf{n} and k . When a host is infected, we therefore specify these parameters \mathbf{n} and k and denote by $\tau_{\mathbf{n}, k}$, $E_{\mathbf{n}, k}$ and $I_{\mathbf{n}, k}$ the corresponding incubation period, exposed and infectious states.

More specifically, when a susceptible S meets an infectious individual $I_{\mathbf{m}, l}$, it receives \mathbf{n} supercritical viral particles with probability $Q_l(\mathbf{n})$, $\mathbf{n} \in \mathcal{S}$, given by Eq. (16). The susceptible individual then either remains in state S with probability $p_{\text{ext}}(\mathbf{n})$, or enters in exposed state $E_{\mathbf{n}, k}$ with probability $p_k(\mathbf{n})$, $0 \leq k \leq K$, given by Eq. (1). If so, it remains in $E_{\mathbf{n}, k}$ for $\tau_{\mathbf{n}, k}$ time steps Eq. (5) before entering infectious state $I_{\mathbf{n}, k}$. Let N_n the total viral population size carried by $E_{\mathbf{n}, k}$ at time n . We deduce from previously mentioned results that $m_k^{-n} N_n$ converges almost surely to $e^{u/s_d} W_{\mathbf{n}, k}$, where $W_{\mathbf{n}, k}$ is a positive random variable whose distribution is given by Eq. (20), and explicitly by Eq. (26) in the particular case of Poisson distributed numbers of offspring and mutations described by Eq. (6). For C large enough, the time needed for N_n to reach size C can consequently be approximated by

$$\tau_{\mathbf{n}, k} = \left\lceil \frac{\ln(C/W_{\mathbf{n}, k}) - u/s_d}{\ln m_k} \right\rceil. \quad (18)$$

In the general setting, the random variable $W_{\mathbf{n}, k}$ is approximately the sum of L independent copies of V , where (i) $L \geq 1$ is the random number of surviving k -lineages (i.e. a line of descent of a k -mutant with no k -mutants as ancestors), and (ii) V is the limiting distribution of $m_k^{-n} X_n$ on its survival set, if $(X_n)_n$ is a monotype branching process modeling only k -mutants. Although not all explicit, we can state the following general results on L and V .

(i) Let k_0 the fittest initial type in \mathbf{n} . If Muller's ratchet does not click, i.e. if $k = k_0$, then the number L of surviving k -lineages is at least 1 and at most n_k . Since the survival probability of a k -lineage is $1 - q_{kk}$, L follows a binomial distribution with parameters n_k and $1 - q_{kk}$, conditioned on being positive. If $k > k_0$ however, there might be more than n_k surviving k -lineages (namely also those stemming from the $\sum_{i=k_0}^{k-1} n_i$ initial mutants). We therefore approximate the maximum number of surviving k -lineages by the average number of k -mutants born from \mathbf{n} after one time-step, conditioned on the event that the surviving individuals are of type k (which we denote Ω_k). This average is equal to $\sum_{i=k_0}^k n_i \tilde{m}_{ik}$, where \tilde{m}_{ik} is the mean number of k -mutants born from one i -mutant, conditionally on Ω_k . In the following computation, the subscript i indicates that the process starts with one i -mutant, and \mathbf{q}_k stands for the infinite vector $(q_{k0}, \dots, q_{kk}, 1, \dots)$ defined in Eq. (9). It comes from Eq. (7) and Eq. (1) that $\mathbb{P}_i(\Omega_k) = q_{k-1,i} - q_{ki}$, and that for any infinite vector \mathbf{x} , using the notation $\mathbf{x}^y = \prod_{i \in \mathbb{N}} x_i^{y_i}$, $\mathbb{P}_{\mathbf{x}}(\Omega_k) = \mathbf{q}_{k-1}^{\mathbf{x}} - \mathbf{q}_k^{\mathbf{x}}$. With this notation, $F_i(\mathbf{r}) = \mathbb{E}_i(\mathbf{r}^{\mathbf{X}_1})$ and $r_k \frac{\partial F_i}{\partial r_k}(\mathbf{r}) = \mathbb{E}_i(X_{1k} \mathbf{r}^{\mathbf{X}_1})$. As a result, the mean \tilde{m}_{ik} is obtained as

$$\begin{aligned} \tilde{m}_{ik} &= \mathbb{E}_i(X_{1k} \mid \Omega_k) = \frac{\mathbb{E}_i(X_{1k} \mathbb{P}_{\mathbf{X}_1}(\Omega_k))}{\mathbb{P}_i(\Omega_k)} \\ &= \frac{\mathbb{E}_i(X_{1k} \mathbf{q}_{k-1}^{\mathbf{X}_1}) - \mathbb{E}_i(X_{1k} \mathbf{q}_k^{\mathbf{X}_1})}{q_{k-1,i} - q_{ki}} \\ &= \frac{\frac{\partial F_i}{\partial r_k}(\mathbf{q}_{k-1}) - q_{kk} \frac{\partial F_i}{\partial r_k}(\mathbf{q}_k)}{q_{k-1,i} - q_{ki}}. \end{aligned} \quad (19)$$

(ii) Let $(X_n)_n$ a branching process with $X_0 = 1$ and generating function $F(r) := F_k(r, 1, 1, \dots)$, $r \in [0, 1]$. It is therefore supercritical with mean offspring number $m_k > 1$. From the theory on monotype branching processes (52) we know that its extinction probability is the smallest nonnegative solution of $F(r) = r$, which is by definition q_{kk} given by Eq. (9). Moreover, $m_k^{-n} X_n \rightarrow W$ almost surely, where W is a nonnegative random variable whose Laplace transform $\varphi(s) = \mathbb{E}(e^{-sW})$ is the unique solution of $\varphi(m_k s) = F(\varphi(s))$. Then the desired random variable V is distributed as W conditioned on $W > 0$. Since $\mathbb{P}(W = 0) = q_{kk}$, the Laplace transform of V is $\phi(s) = (\varphi(s) - q_{kk}) / (1 - q_{kk})$.

To summarize, we assume that the random variable $W_{n,k}$ appearing in Eq. (5) is given by

$$W_{n,k} = \sum_{i=1}^L V^{(i)}, \quad (20)$$

where L is a random integer such that

$$\mathbb{P}(L = l) = \binom{\tilde{n}_k}{l} \frac{(1 - q_{kk})^l q_{kk}^{\tilde{n}_k - l}}{1 - q_{kk}^{\tilde{n}_k}}, \quad 1 \leq l \leq \tilde{n}_k, \quad (21)$$

with

$$\tilde{n}_k = \begin{cases} n_k, & k = k_0, \\ \left\lceil \sum_{i=k_0}^k n_i \tilde{m}_{ik} \right\rceil, & k > k_0, \end{cases} \quad (22)$$

and where the $V^{(i)}$ are independent copies of the random variable V with Laplace transform

$$\phi(s) = \frac{\varphi(s) - q_{kk}}{1 - q_{kk}}, \quad (23)$$

φ being the unique Laplace transform solution of $\varphi(m_k s) = F_k(\varphi(s), 1, 1, \dots)$

Supplementary Note 4: Explicit formulas

Poisson distributed number of offspring. In the particular case of a Poisson distributed number of offspring and mutations (with generating functions Eq. (6)), an exact computation of (q_{k0}, \dots, q_{kk}) can be found, leading to explicit expressions of the extinction and Muller's ratchet click probabilities Eq. (9)-Eq. (1) and of the expected asymptotic population mean fitness Eq. (2). Indeed, system Eq. (8) becomes

$$\exp \left[-m_i \left(\sum_{j=i}^k \frac{u^{j-i}}{(j-i)!} (1 - q_{kj}) \right) \right] = q_{ki}, \quad 0 \leq i \leq k,$$

leading to

$$\begin{cases} q_{kk} = -\frac{1}{m_k} W[-m_k e^{-m_k}], \\ q_{ki} = -\frac{1}{m_i} W \left[-m_i e^{-m_i \left(1 + \sum_{j=i+1}^k u^{j-i} (1 - q_{kj}) / (j-i)! \right)} \right], \quad 0 \leq i \leq k-1, \end{cases} \quad (24)$$

where $W[\cdot]$ stands for the principal solution of the Lambert W function.

In this particular case, we can also compute Eq. (19) and Eq. (23), and therefore provide the explicit distribution Eq. (20) of $W_{n,k}$ involved in the incubation period of the exposed state Eq. (5). Indeed, with F_i given by Eq. (6), we have $\frac{\partial F_i}{\partial r_k}(\mathbf{r}) = m_{ik} F_i(\mathbf{r})$, where $m_{ik} = w_i e^{-u} u^{k-i} / (k-i)!$. In addition, since \mathbf{q}_k satisfies by definition $F_i(\mathbf{q}_k) = q_{ki}$, Eq. (19) becomes

$$\tilde{m}_{ik} = m_{ik} \frac{q_{k-1,i} - q_{kk} q_{ki}}{q_{k-1,i} - q_{ki}}. \quad (25)$$

Finally, we show that, approximately, the random variable V involved in Eq. (20) follows an exponential distribution with parameter $1 - q_{kk}$. Indeed, the latter means that $\phi(s) = \frac{1 - q_{kk}}{1 - q_{kk} + s}$, and thus $\varphi(s) = 1 - \frac{(1 - q_{kk})s}{1 - q_{kk} + s}$. We have on the one hand

$$\varphi(m_k s) = 1 - m_k \frac{(1 - q_{kk})s}{1 - q_{kk} + s},$$

while on the other hand

$$F_k(\varphi(s), 1, \dots) = \exp \left[-m_k \frac{(1 - q_{kk})s}{1 - q_{kk} + s} \right],$$

hence for m_k close to 1, $\varphi(m_k s) \approx F_k(\varphi(s), 1, \dots)$, leading to our rough approximation. As the sum of independent exponentially distributed variables, $W_{\mathbf{n},k}$ thus follows a Gamma distribution with shape and rate parameters

$$W_{\mathbf{n},k} \sim \text{Gamma}(L, 1 - q_{kk}) \quad (26)$$

where L is given by Eq. (21) with

$$\tilde{n}_k = \begin{cases} n_k, & k = k_0, \\ \left[\sum_{i=k_0}^k n_i m_{ik} \frac{q_{k-1,i} - q_{kk} q_{ki}}{q_{k-1,i} - q_{ki}} \right], & k > k_0. \end{cases} \quad (27)$$

Binary reproduction. Other reproduction mechanisms can be considered, with the same key assumption that the average number of $(i+j)$ -mutants produced by an i -mutant is $w_i e^{-u} u^j / j!$. For example, we can assume a binary reproduction (although less relevant for viral populations) where an i -mutant either dies, or produces two offspring with probability $w_i/2$, each of them accumulating a random number of additional mutations following $\mathcal{P}(u)$. In this case, the generating function is quadratic

$$F_i(r_i, r_{i+1}, \dots) = 1 - \frac{w_i}{2} + \frac{w_i}{2} \left(\sum_{j \in \mathbb{N}} e^{-u} \frac{u^j}{j!} r_{i+j} \right)^2,$$

and system Eq. (8) comes down to

$$1 - \frac{w_i}{2} + \frac{w_i}{2} \left(1 - \sum_{j=i}^k e^{-u} \frac{u^{j-i}}{(j-i)!} (1 - q_{kj}) \right)^2 = q_{ki}, \quad 0 \leq i \leq k,$$

which can be solved explicitly as well.