

1 Iliski, a software for robust calculation of transfer functions

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11 1. Abstract

12 Understanding the relationships between biological events is paramount to unravel
13 pathophysiological mechanisms. These relationships can be modeled with Transfer Functions
14 (TFs), with no need of *a priori* hypotheses as to the shape of the transfer function. Here we
15 present Iliski, a software dedicated to TFs computation between two signals. It includes
16 different pre-treatment routines and TF computation processes: deconvolution, deterministic
17 and non-deterministic optimization algorithms that are adapted to disparate datasets. We
18 apply Iliski to data on neurovascular coupling, an ensemble of biological events that link
19 neuronal activity to local changes of blood flow, highlighting the software benefits and caveats
20 in the computation and evaluation of TFs. We also propose a workflow that will help users to
21 choose the best computation according to the dataset. Iliski is available under the open-source
22 license CC BY 4.0 on GitLab (https://gitlab.com/AliK_A/iliski) and can be used on the most
23 common operating systems, either within the MATLAB environment, or as a standalone
24 application.

26 2. Introduction

27 Modelling and understanding of the relationship between complex and intermingled biological
28 signals is often difficult, particularly when the drivers of the signals are unknown. The problem
29 of the relationship between two time series can be address using deconvolution, which
30 provides Transfer Functions (TFs) representative of the system processing on the input signal
31 to generate the output signal¹. Extracting the transfer function of a neuron or neurons is widely
32 used in neurobiological studies²⁻⁹, and is widely used to solve general problems in signal
33 analysis, such as predicting the output of complex electrical circuits¹⁰ or other industrial
34 systems for which a proper model is very complex due to multiple processes working in
35 parallel¹¹. In brain imaging based on blood flow dynamics, transfer functions are classically
36 used to lump the multitude of cellular and molecular processes linking neural activation to
37 changes in blood flow. This coupling between neural activity and hemodynamics is known as
38 neurovascular coupling (NVC)¹². While there are many successful phenomenological models of
39 NVC^{3,13-17}, most physiology-based models of neurovascular coupling¹⁸⁻²¹ focus on a single
40 mechanism. As NVC is mediated through multiple processes, a more integrated approach is
41 necessary. NVC has often been assessed with deconvolution^{8,22}, either in the frequency domain
42 or with matrix-based approaches, like Toeplitz matrices²³. While these approaches allow the
43 unbiased extraction of the TF, these deconvolution methods suffer from sensitivity to noise,
44 affecting the quality of the computed TFs. Reducing the noise (or bandwidth) of the signals
45 improves the estimate of the TF. Alternatively, one can opt for optimization of known functions
46 or a kernel of functions^{7,9}. The first option may lead to information loss, e.g. in cases where the
47 noise is not well characterized. Sophisticated smoothing methods partially prevent this loss,
48 like Savitzky-Golay filter, or noise modelling as proposed by Seghouane and colleagues²⁴. The
49 second option relies on parametric functions to find the TF best linking the input to the output
50 signals. The transfer function for neural activity to hemodynamic signals has been canonically

51 modeled using a gamma-distribution function^{3,14-16}. While making assumptions as to the shape
52 of the TF has some drawbacks, it is robust in the face of noise and generates parametric
53 representations of intrinsically smooth TFs. These approaches still can suffer from
54 under/overfitting and the search for the minimum of the cost function for ill-posed problems
55 may represent a challenging exercise. A valuable help comes from non-deterministic
56 optimizations like simulated annealing or genetic algorithms, which despite their
57 computational expense have potential advantages in extracting TFs from time series.

58 Recently, our group has been extensively involved in TF computation of neurovascular
59 coupling in a study based on multi-modal recordings, namely two-photon microscopy and
60 ultra-fast functional ultrasound²⁵. For the required task, we comprehensively tested many
61 deconvolution and optimization algorithms to choose the best-suited approach. We noticed
62 that there is no standard software package providing all these different TF extraction tools, nor
63 a program where all these approaches are available in a comfortable signal pre-treatment and
64 I/O workflow. Here, we present Iliski ([ilīski], meaning “relationship” in Turkish), a software
65 which contains all the functionalities that we previously used (Aydin et al.) and which, being
66 open source, can be further improved by the users.

67

68 **3. Design and Implementation**

69 Iliski can compute TFs between an input and an output time series, regardless of their nature.
70 The originality of Iliski resides in its multiple options to process and analyze input signals. Iliski
71 provides users with efficient pre-treatment and several deconvolution or optimization
72 algorithms, through a clear graphic interface. It is meant to be easy-to-use for anyone, even with
73 basic digital signal processing skills.

74 Iliski can be used either as a suite of functions or through a Graphical User Interface (**Fig 1A**).
75 Functions are grouped according to the analysis workflow to keep the interface simple. **Fig 1B**
76 shows the general purpose of Iliski.

77

78 **3.1. Data loading and pre-treatment**

79 We propose two input files format: either plain text files or HDF5 data, the latter being an open-
80 source file format with advanced database features. As experimental acquisitions are prone to
81 multiple component noise, we provided, as an option to the analysis workflow, smoothing
82 (Savitzky-Golay method) and median filter functions, to exclude outliers. The input and output
83 signals are interpolated to a chosen time interval (Δt). Both signals can be cut between two
84 given time points to study continuous recordings while computing TFs on chunks of signal (**Fig**
85 **1C**).

86

87 **3.2. TF computation options**

88 Two main types of TF computation are proposed: deconvolution or function optimization. The
89 former is straightforward, either Toeplitz or Fourier deconvolution, and does not require any
90 specific settings. The latter is the optimization of a parametric function, which requires further
91 settings depending on the chosen algorithm. Beside the proposed fitting functions, the users
92 can input their own function in the graphical interface or add it to the default ones by modifying
93 a text file (the procedure is described in the Iliski Manual). Optimization of parameters can be
94 done with various Matlab algorithms, each coming with pros and cons (see Results section) (**Fig**
95 **1C, middle**).

96

97 **3.3. Evaluation of the TF accuracy**

98 A TF is evaluated comparing its prediction – the convolution of the input signal and the TF - to
99 the expected output. Two metrics are used in Iliski: the Pearson coefficient (*corrcoef* function,
100 Matlab, **Fig 1C, right**) and the residual sum of squares. The former was chosen to have a metric
101 solely focusing on the dynamic, allowing for inter-subject comparisons, while the latter
102 evaluates the overall fit, considering the amplitude of the prediction. The cost function of all the
103 optimization algorithms tested in this article is the residual sum of squares (hereinafter
104 referred to as "residuals").

105

106 **3.4. Post-computation**

107 The results structure is arranged to be as informative as possible while avoiding useless
108 repetition of data. Iliski allows for loading previously computed results structures to check
109 them again. After TF computation, results structure can be saved either as XLS file, readable by
110 any Excel-like software, or as a MAT-file (MATLAB formatted binary file format), but it is also
111 available in Matlab workspace to be exported in various data formats by the user.

112 **3.7. Implementation**

113 Iliski is accessible both as a GUI and as a set of functions to be used in scripts. It has been
114 developed using Matlab R2018a, with the following dependencies: Optimization Toolbox,
115 Signal Processing Toolbox and Global Optimization Toolbox.

116 Common user errors are thoroughly prevented by various messages and fail safes. In parallel,
117 all errors are treated and saved in a LOG file, to allow for efficient bug-fixing by any developer.
118 We purposely kept just a few parameters to modify through the GUI, with the goal of providing
119 an easy-to-use tool to people not used to these functions. In most cases, Matlab default

120 parameters of each deconvolution/optimization function worked well with our data, and we
121 believe that it can be extended to many biological datasets. However, a user skilled with Matlab
122 and optimization algorithms can easily modify the parameters of each function used.

123 **3.8. Animal Research**

124 This study uses already published data of animal experimentation (Aydin et al.). All animal
125 care and experimentations were performed in accordance with the INSERM Animal Care and
126 Use Committee guidelines (protocol numbers CEEA34.SC.122.12 and CEEA34.SC.123.12).

127 **4. Results**

128 Here we present the use of Iliski to find the best mathematical representation of neurovascular
129 coupling, an ensemble of cellular mechanisms that links brain activation to local increases of
130 blood flow. Neural activity is reported by GCaMP6f²⁶, a calcium-sensitive protein expressed in
131 specific neurons. Blood flow is quantified by measuring red blood cells velocity changes in
132 capillaries²⁷.

133 Several deconvolution and function optimization algorithms are provided. Choosing the
134 algorithm(s) and settings to compute a TF that gives faithful and robust predictions is not
135 always a straightforward task. It must be done according to the data features. Here we use some
136 data from our published study on neurovascular coupling²⁵ to point out how TFs change with
137 different algorithms and settings, and we show the critical points in the usage of non-
138 deterministic methods. Finally, we propose a step-by-step guide to optimize the best TF on
139 practical situations.

140 **4.1. Choosing the best TF computation approach**

141 **Figure 2** shows TF computation with different settings over the same couple of signals:
142 neuronal (Ca^{2+}) and vascular (red blood cells velocity) activations recorded in a mouse upon

143 odor application. Our example data display unavoidable and complex noise coming from many
144 sources: the biological system, the optical setup, the electronics, etc. Deconvolution with
145 Fourier or Toeplitz approaches predicts the vascular responses very well for a given data set.
146 However, the high-frequency noise is amplified by deconvolution²⁴ and transmitted to the TF,
147 the predictions are not robust across data sets and the actual dynamics of neurovascular
148 coupling is completely hidden in the TF noise (**Fig 2**). In this example, we show what we regard
149 as a typical case of overfitting. The TF is capturing the high frequency noise of the system
150 because it does not have any previous expectations for the shape of the relationship between
151 the input and the output signals. This contrasts to the optimization of a parametric function
152 approach which, although it imposes constraints on the shape of the TF, gives meaningful
153 neurovascular relationship and does not need noise clearing. In blood flow-based
154 neuroimaging, the standard function used to represent neurovascular coupling is composed of
155 one or two Γ functions, depending on the nature of the signals, i.e purely vascular or based on
156 oxygen level²⁸.

157 Below is the one Γ -driven function we used with our data.

$$158 \quad TF(t) = H(t - p_3) \times p_4 \times \frac{(t - p_3)^{p_1-1} \times p_2^{p_1} \times e^{-p_2 \times (t - p_3)}}{\Gamma(p_1)}$$

159 Where p_1, \dots, p_4 are the parameters to optimize and H is the Heaviside function to include a time-
160 shift parameter (p_3) which, in some cases, significantly improved the prediction and is a known
161 biological phenomenon to consider²⁹. Its four parameters are not all independent from one
162 another, e.g. p_1 , p_2 and p_4 all impact the TF amplitude. This inter-dependency between the
163 parameters brings an ill-posed optimization problem with multiple local minima of the cost
164 function, the sum of the residual squares, in the 4D space of the parameters.

165 A derivative-free optimization method (provided by Matlab and used previously⁶) is provided
166 by the *fminsearch* function in Matlab. This approach on our data produced a TF with more than

167 one underivable point that is not representative of the smooth dynamic of neurovascular
168 coupling.

169 Another common option is provided by Quasi-Newton optimization algorithms: for this
170 approach too, we tested an unconstrained built-in method (*fminunc* function, Matlab). This
171 prediction is, overall, as good as with *fminsearch* (Fig 2, Pearson coefficients, *fminunc* vs.
172 *fminsearch*: 0.95 vs. 0.96), but the onset phase is not properly fit. Moreover, although not
173 evident from the plot, the optimized time shift was negative (-120 ms), implying that the onset
174 of the vascular response precedes the neuronal activation.

175 All the optimization methods tested above are deterministic, meaning that repeating them with
176 the same initial parameters will bring the same result. The pitfall of these methods when
177 applied to ill-posed problem is that optimization process will get attracted to the nearest local
178 minimum, regardless of the many other deeper minima, which may be far away in the
179 parameters space. In other words, deterministic algorithms are sensitive to the initial
180 parameters set before starting the optimization.

181 Non-deterministic algorithms exist to overcome the local minimum issue, adding some level of
182 randomness in the optimization process, and for this purpose Iliski uses the Simulated
183 Annealing algorithm. Each optimization run can yield a different result, reaching possibly a
184 different cost function's minimum each time. We define as 'run' a single application of the
185 optimization with a given set of initial values, and 'iteration' the ensemble of runs sharing the
186 same initial values. By running the algorithm multiple times, one can choose the result with the
187 lowest residual, while avoiding TFs which shape are biologically not acceptable. In Aydin et al.
188 (2020), we described a workflow of runs and iterations to get to biologically consistent TFs (see
189 Supplementary Figure 1). To speed up computation, we imposed bonds over the parameters.
190 Note that such bonds can be set through the Iliski GUI for any constrainable algorithm.

191 Using our data, Simulated Annealing gave a smooth TF and a prediction as good as *fminsearch*
192 for the onset phase of the vascular response. The data shown in **Fig 2** is representative of the
193 rest of the data. In fact, optimization of TFs using neural and vascular recordings from other
194 mice, tested with the same odor stimulation, produced similar residual values of the cost
195 function across the 3 optimization algorithms presented above (1-way ANOVA, $F(2, 17) = 0.035$,
196 $p = 0.97$, **Fig 3A**). However, as in the example of **Fig 2**, deterministic algorithms are prone to
197 biologically inconsistent TFs (**Fig 3B**). The non-deterministic, Simulated Annealing algorithm
198 with subsequent iterations method allows to efficiently exclude these TFs and obtain the best
199 trade-off between prediction performance and biological consistency at the cost of a longer
200 computation time. Direct deconvolution is a good option when the goal is the prediction quality
201 within the training database. Deterministic optimization algorithms are fast but yield to TFs
202 that may have biologically inconsistent dynamics. Note that for all the computations we used a
203 short Δt (50 ms) for interpolation to preserve most of the information.

204

205 **4.2. Evaluating the number of runs in a non-deterministic case**

206 As already mentioned, the Simulated Annealing algorithm requires several runs and iterations
207 to obtain a good TF. In our experience, starting the optimization with a 'bad' TF - whose shape
208 is different from what is expected for the processed dataset - helps to collect more local minima
209 in a pool of optimization runs. For example, in our previous study²⁵, we proposed iterations of
210 50 runs and started with the initial values of the standard TF (one Γ HRF) which, peaking at 5
211 seconds, turned to be much slower than any of the optimized TFs. The sequence of 50-runs
212 iterations stopped when, within an iteration, no clear improvement was found in the optimized
213 TF²⁵. On average, 2 iterations were sufficient to get a stable TF with Pearson coefficient above
214 0.9. Here, we investigated if a higher number of runs is beneficial to the detection of the
215 minimum of the cost function and if it prevents the need for further iterations. We compared

216 50 and 200 runs with single and double iterations, in cascade (**Fig 4A**). In a mouse dataset, we
217 observed a non-significant trend towards more scattered TFs shapes for computation using 50
218 runs versus 200 runs (1-way ANOVA, $F(3, 16) = 2.086$, $p = 0.14$, **Fig 4B**). Similarly, the quality
219 of the TFs did not significantly improve with increasing runs (1-way ANOVA, $F(3, 16) = 2.299$,
220 $p = 0.12$). As a result, TFs with fast dynamics (peaking within 1 and 2 sec), was a common
221 feature independently of the adopted protocol (**Fig 4C**). In a dataset from another mouse (**Fig**
222 **4D,E**), TFs with sparse time to peak values after 200 runs improved after a second iteration,
223 with the same number of runs (2.3 ± 0.3 s VS 1.5 ± 0.1 s (mean \pm SEM), two-tailed T-test,
224 unpaired, $p = 0.02 < 0.05$). Note that this compression of TF dynamics was not accompanied by
225 a significant improvement of the TF quality (residuals: 10.1 ± 2.1 vs. 6.9 ± 0.8 (mean \pm SEM) for
226 200 and $200 + 200$ runs respectively, two-tailed T-test, unpaired, $p = 0.19$). To conclude,
227 depending on the input/output signals, non-deterministic algorithms can produce TFs with
228 different dynamics but close performances in the prediction.

229 **4.2. Guide to choose the algorithm best fitting your needs**

230 We provide a decision diagram to choose the best approach to compute a TF based on the
231 features of the user's dataset (**Fig 5**). Nonetheless, we believe it is always a good choice to test
232 different approaches before making the final choice.

233

234 **5. Discussion**

235 Iliski provides a user friendly, interactive and rich in option software for quickly testing
236 different methods and settings to compute TFs between biological events. In addition to its
237 standard integrated functions, it also allows for user-defined functions of any number of
238 parameters. Using data from the NVC field, we demonstrate how critical is the choice of the
239 method for computing TFs and the caveats of some parameters such as the number of iterations

240 necessary to non-deterministic algorithms. Note that we did not report the influence of
241 smoothing, interpolation, fitting and cost functions choice which are also known to affect the
242 final result. The use of multimodal datasets, i.e. neuronal calcium signal, measurements of
243 vascular responses at both the microscopic and mesoscopic scales enabled us to demonstrate
244 that NVC is represented by a similar TF which is much faster than the classical HRF, a finding
245 which is getting accepted in the field of brain imaging based on blood flow^{5,25,30,31}.

246 **6. Availability and Future Directions**

247 Iliski is open-source and freely available under the Creative Commons Attribution 4.0
248 International (CC BY 4.0) license. Iliski is maintained on GitLab, enabling user-friendly bug
249 report and community work to make the tool fit the users' need. It can be found here:
250 https://gitlab.com/AliK_A/iliski/.

251 In the neurovascular imaging field, computing the hemodynamic response function is
252 paramount to interpreting vascular activation in terms of neural activation. In any other field,
253 computing TFs may be of help to go deeper in the interpretation of the results. For these
254 reasons, we think it is extremely helpful to have a data analysis tool which lets fast testing of
255 different algorithms with a user-friendly interface.

256 **7. Supporting information**

- 257 • GitLab repository: https://gitlab.com/AliK_A/iliski/
- 258 • Example data: <http://doi.org/10.5281/zenodo.3773863>
- 259 • User Manual: https://gitlab.com/AliK_A/iliski/-/raw/master/Iliski_User_Manual.pdf?inline=false

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274 9. Figures

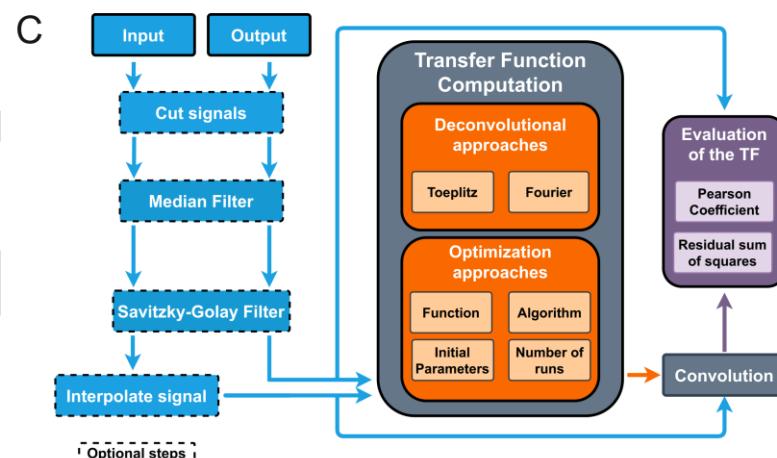
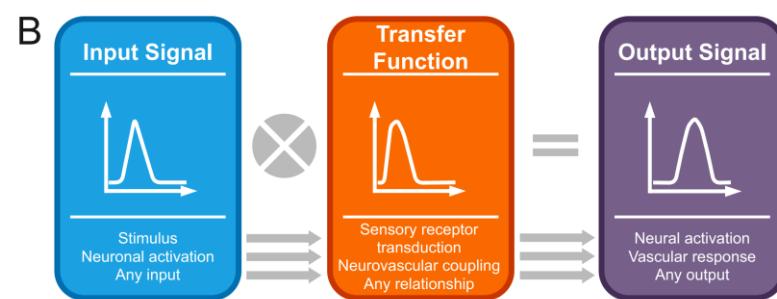
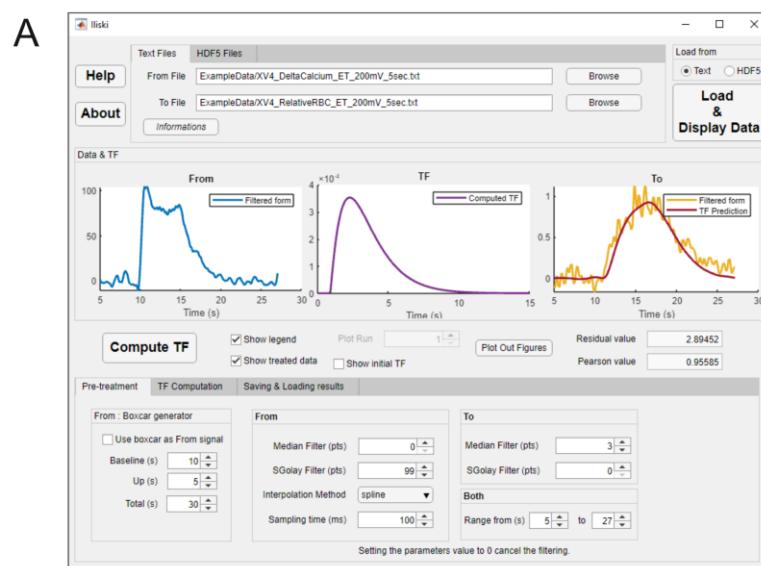


Fig 1. Overview of Iliski.

(A) Iliski has a clear interface with tabs bringing through the analysis steps. (B) The usage of Iliski are many; although it has been conceived for biological data, there is no limitation to load any discretized signal, serving as a tool for fast testing different approaches for TFs computation. (C) Iliski workflow is modular so that signal pre-processing is optional and functions to compute TFs can be modified by the user preserving the I/O modules.

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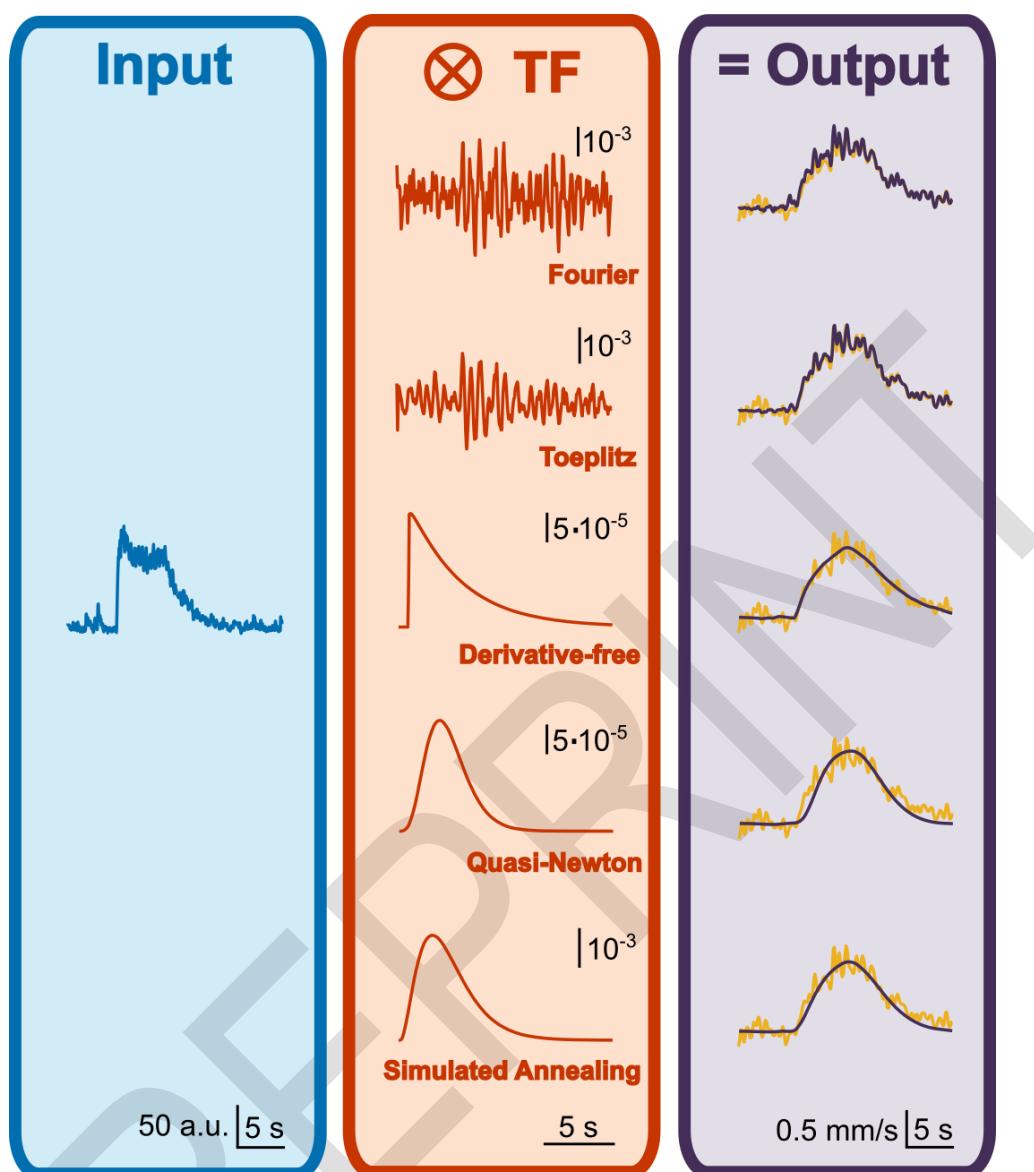


Fig 2. Comparison of deconvolution and optimization algorithms on a batch of data.

Odor stimulation elicited a neuronal response in the Olfactory Bulb of a mouse, reported by a calcium-dependent fluorescent signal (in blue, left panel), providing the input of TF computation. Output is given by the vascular response, measured as the change in speed of red blood cells flowing inside a capillary proximal to the recorded neuronal activation (in yellow, right panel). Both experimental data have been resampled at 50ms and used to compute a set of TFs (in orange) either with direct deconvolution approaches (Fourier or Toeplitz methods, middle-upper panel TFs) or with $1-\Gamma$ function optimization performed by 3 different algorithms (middle-lower panel TFs). Complex TFs bring accurate prediction but amplify the noise of the data used to deconvolve them, with a consequent loss of robustness on other datasets. Smoother TFs are less accurate on the training dataset, but much robust when applied to test datasets.

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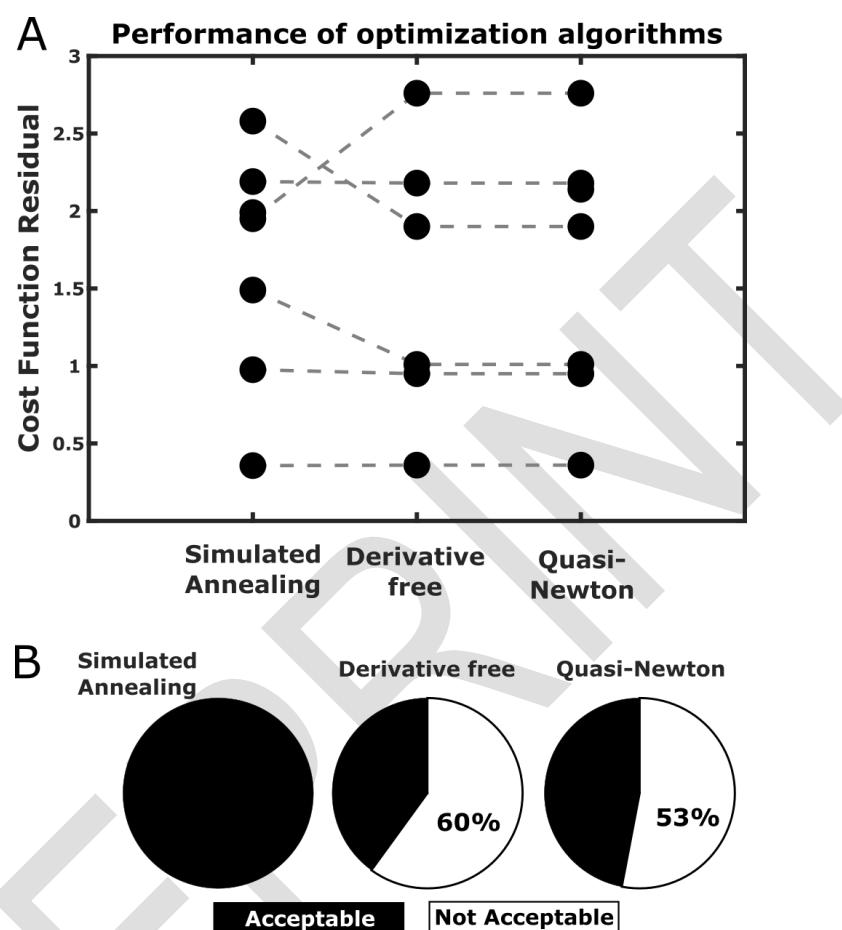


Fig 3. Prediction performance of different optimization algorithms.

(A) 3 algorithms were compared in terms of the residuals of the cost function of the optimized TF on 7 mice datasets (Derivative free algorithm failed in optimizing a TF in a mouse). No significant difference was found across the 3 methods. (B) However, simulated annealing was the only approach to provide TFs consistent with the nature of biological data (TF with no more than 1 non-derivable point), while both the other deterministic methods run into inconsistent TFs in roughly 60% of the cases.

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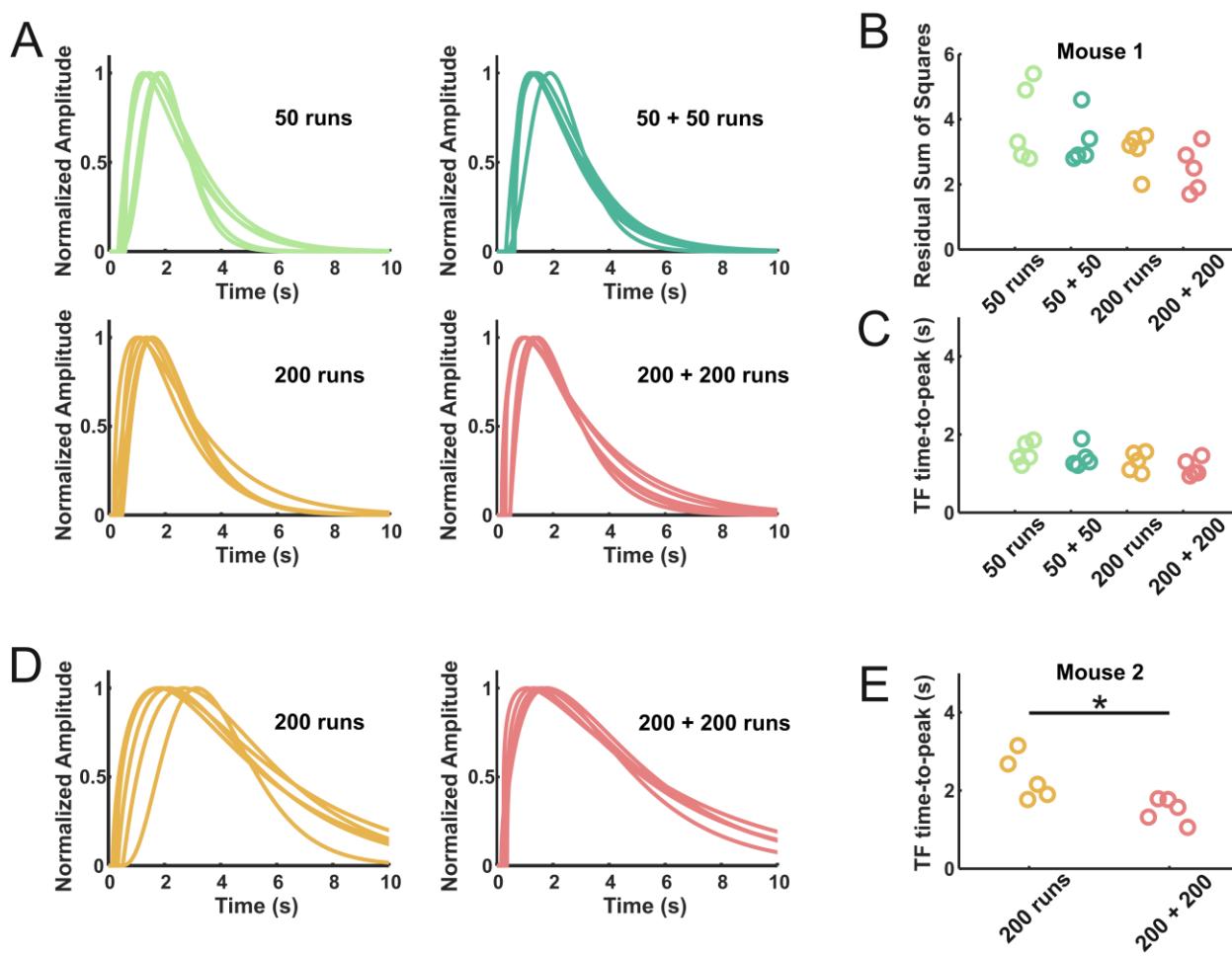


Fig 4. Influence of the number of runs and iterations on the TF shape and quality.

(A) Using the Simulated Annealing algorithm, we tested 4 protocols of 50 or 200 optimization runs, either done a single time or repeated (5 TFs computed for each protocol). (B) Residuals of the cost function do not significantly differ across the protocols, although the protocols with the highest number of runs show a trend of smaller residuals. (C) Similarly, there was no significant difference for TFs time-to-peak values. (D, left) Same protocols comparison on a dataset from a different mouse revealed a sparse dynamic of optimized TFs, even if the best TFs were selected on a pool of many TFs (200 runs). (D, right) A second iteration of 200 runs gave more homogeneous TFs dynamics. (E) Quantification of the dynamic heterogeneity was made by measuring the time-to-peak which resulted in a scattered distribution for the 200 runs protocol, packed up repeating the same iteration a second time.

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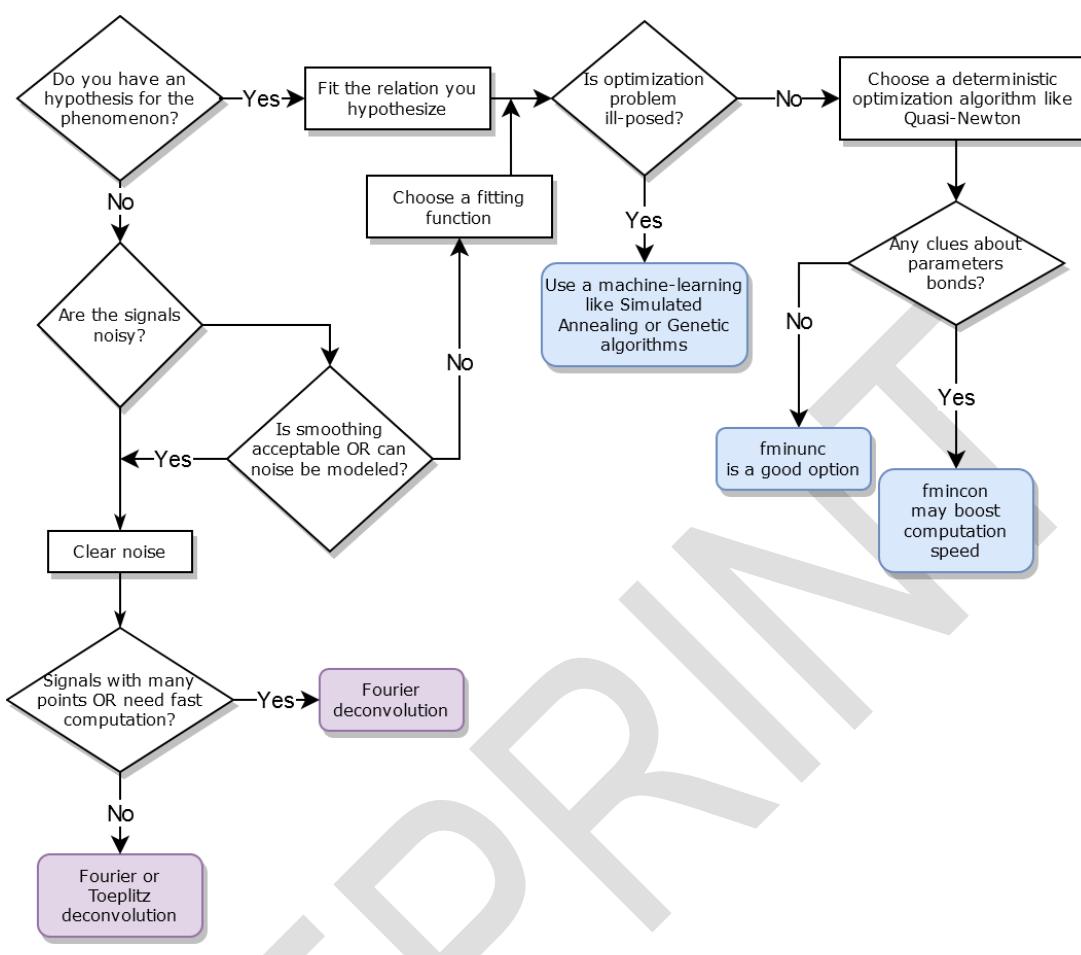


Fig 5. Decision tree to help choosing the most efficient method to compute a TF with Iliski, based on the data features.

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288 10. Bibliography

289

290 1. Dijk, G. van. *Distribution Theory Convolution, Fourier Transform, and Laplace Transform, 2013.pdf.* (2013).

291

292 2. Marmarelis, P. Z. & Naka, K.-I. White-Noise Analysis of a Neuron Chain: An Application of the 293 Wiener Theory. *Science* **175**, 1276–1278 (1972).

294 3. Silva, A. C., Koretsky, A. P. & Duyn, J. H. Functional MRI impulse response for BOLD and 295 CBV contrast in rat somatosensory cortex. *Magnet Reson Med* **57**, 1110–1118 (2007).

296 4. Theunissen, F. E. *et al.* Estimating spatio-temporal receptive fields of auditory and visual neurons 297 from their responses to natural stimuli. *Netw Comput Neural Syst* **12**, 289–316 (2009).

298 5. Lambers, H. *et al.* A cortical rat hemodynamic response function for improved detection of
299 BOLD activation under common experimental conditions. *NeuroImage* **116**446 (2019)
300 doi:10.1016/j.neuroimage.2019.116446.

301 6. Sirotin, Y. B. & Das, A. Anticipatory haemodynamic signals in sensory cortex not predicted by
302 local neuronal activity. *Nature* **457**, 475 (2009).

303 7. Cardoso, M., Lima, B., Sirotin, Y. B. & Das, A. Task-related hemodynamic responses are
304 modulated by reward and task engagement. *PLOS Biology* **17**, e3000080 (2019).

305 8. Winder, A. T., Echagarruga, C., Zhang, Q. & Drew, P. J. Weak correlations between
306 hemodynamic signals and ongoing neural activity during the resting state. *Nat Neurosci* **20**, 1761–
307 1769 (2017).

308 9. Herman, M., Cardoso, M., Lima, B., Sirotin, Y. B. & Das, A. Simultaneously estimating the task-
309 related and stimulus-evoked components of hemodynamic imaging measurements. *Neurophotonics*
310 **4**, (2017).

311 10. Basso, C. P. Linear Circuit Transfer Functions. doi:10.1002/9781119236344.

312 11. Sayyafzadeh, M., Pourafshary, P., Haghghi, M. & Rashidi, F. Application of transfer functions
313 to model water injection in hydrocarbon reservoir. *J Petrol Sci Eng* **78**, 139–148 (2011).

314 12. Iadecola, C. The Neurovascular Unit Coming of Age: A Journey through Neurovascular
315 Coupling in Health and Disease. *Neuron* **96**, 17–42 (2017).

316 13. Nishimoto, S. *et al.* Reconstructing Visual Experiences from Brain Activity Evoked by Natural
317 Movies. *Curr Biol* **21**, 1641–1646 (2011).

318 14. Goense, J. B. M. & Logothetis, N. K. Neurophysiology of the BOLD fMRI Signal in Awake
319 Monkeys. *Current Biology* **18**, 631–640 (2008).

320 15. Boynton, G. M., Engel, S. A. & Heeger, D. J. Linear systems analysis of the fMRI signal.
321 *NeuroImage* **62**, 975–984 (2012).

322 16. Boynton, G. M., Engel, S. A., Glover, G. H. & Heeger, D. J. Linear Systems Analysis of
323 Functional Magnetic Resonance Imaging in Human V1. *J Neurosci* **16**, 4207–4221 (1996).

324 17. Huo, B.-X., Gao, Y.-R. & Drew, P. J. Quantitative separation of arterial and venous cerebral
325 blood volume increases during voluntary locomotion. *NeuroImage* **105**, 369–79 (2014).

326 18. Buxton, R. B., Wong, E. C. & Frank, L. R. Dynamics of blood flow and oxygenation changes
327 during brain activation: The balloon model. *Magnet Reson Med* **39**, 855–864 (1998).

328 19. Haselden, W. D., Kedarasetti, R. T. & Drew, P. J. Spatial and temporal patterns of nitric oxide
329 diffusion and degradation drive emergent cerebrovascular dynamics. *Plos Comput Biol* **16**,
330 e1008069 (2020).

331 20. Witthoft, A., Filosa, J. A. & Karniadakis, G. Potassium Buffering in the Neurovascular Unit:
332 Models and Sensitivity Analysis. *Biophysical Journal* **105**, 2046–2054 (2013).

333 21. Moshkforoush, A. *et al.* The capillary Kir channel as sensor and amplifier of neuronal signals:
334 Modeling insights on K⁺-mediated neurovascular communication. *Proc National Acad Sci* **117**,
335 16626–16637 (2020).

336 22. Huo, B.-X., Greene, S. E. & Drew, P. J. Venous cerebral blood volume increase during
337 voluntary locomotion reflects cardiovascular changes. *NeuroImage* **118**, 301312 (2015).

338 23. Hansen, P. C. Deconvolution and Regularization with Toeplitz Matrices. *Numer Algorithms* **29**,
339 323–378 (2002).

340 24. Seghouane, A.-K., Shah, A. & Ting, C.-M. fMRI hemodynamic response function estimation in
341 autoregressive noise by avoiding the drift. *Digital Signal Processing* **66**, 29–41 (2017).

342 25. Aydin, A.-K. *et al.* Transfer functions linking neural calcium to single voxel functional
343 ultrasound signal. *Nat Commun* **11**, 2954 (2020).

344 26. Chen, T.-W. *et al.* Ultrasensitive fluorescent proteins for imaging neuronal activity. *Nature* **499**,
345 295–300 (2013).

346 27. Chaigneau, E., Oheim, M., Audinat, E. & Charpak, S. Two-photon imaging of capillary blood
347 flow in olfactory bulb glomeruli. *Proceedings of the National Academy of Sciences of the United
348 States of America* **100**, 13081–6 (2003).

349 28. Friston, K. J., Josephs, O., Rees, G. & Turner, R. Nonlinear event-related responses in fMRI.
350 *Magnetic Resonance in Medicine* **39**, 41–52 (1998).

351 29. Rungta, R. L., Chaigneau, E., Osmanski, B.-F. & Charpak, S. Vascular Compartmentalization
352 of Functional Hyperemia from the Synapse to the Pia. *Neuron* **99**, 362-375.e4 (2018).

353 30. Gao, Y.-R., Greene, S. E. & Drew, P. J. Mechanical restriction of intracortical vessel dilation by
354 brain tissue sculpts the hemodynamic response. *NeuroImage* **115**, 162–176 (2015).

355 31. Drew, P. J. Vascular and neural basis of the BOLD signal. *Current Opinion in Neurobiology* **58**,
356 61–69 (2019).

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