

## **The Role of Temperature on the Development of Circadian Rhythms in Honey Bee Workers**

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# **Abstract:**

Circadian rhythms in honey bees are involved in various processes that impact colony survival. For example, young nurses take care of the brood constantly throughout the day and lack circadian rhythms, while foragers use the circadian clock to remember and predict food availability in subsequent days. Previous studies suggested that development of circadian rhythms both in the field and the laboratory began around 7-9 days of age. However, not much is understood about the postembryonic development of circadian rhythms in honey bees. In the current study, we examine the effects of socially regulated colony temperature on the ontogeny of circadian rhythms of young workers under controlled laboratory conditions. We hypothesized that temperature plays a key role in the development of circadian rhythmicity in young workers. Our results show that young workers kept at 35°C develop circadian rhythmicity faster and in greater proportion than bees kept at 25°C. In addition, we examine if the effect of colony temperature during the first 48 hours after emergence is enough to observe effects on the rate and proportion of development of circadian rhythmicity. We observed that twice as many individuals that were exposed to 35°C during the first 48 hours develop circadian rhythms compared to individuals kept at 25°C. In addition, we observed differences in the average endogenous period length consistent with temperature compensation of the circadian rhythms between the 25°C and 35°C cohorts. We also observed differences in the degree of period length variation between the 25°C and 35°C cohorts, which combined with the proportion of arrhythmic individuals and survival data suggest that development of circadian rhythms is incomplete in individuals exposed to 25°C adult emergence. This study shows that temperature, which is socially regulated inside the hive, is a key factor that influences the ontogeny of circadian rhythmicity of workers.

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## Introduction:

The circadian clock of honey bees is important in complex physiological processes, such as spatiotemporal learning, time perception and sun-compass navigation (Goodwin and Lewis, 1987; Moore et al., 1998; Van Nest and Moore, 2012; von Frisch, 1967; Wagner et al., 2013). However, when it comes to development of circadian rhythms in honey bee workers, scientists are just beginning to scratch the surface of what is thought to be a highly complex mechanism of regulation, with components at the environmental, social, hormonal and genetic levels (Eban-Rothschild et al., 2012; Moore, 2001; Moore et al., 1998; Shemesh et al., 2007). In this manuscript, we study the role of environmental temperature on the ontogeny of circadian rhythms of young honey bee workers.

The development of honey bee circadian rhythms is of particular interest because similar to human infants, young honey bees present postembryonic development of circadian rhythms before they forage (Eban-Rothschild et al., 2012; Moore et al., 1998). Furthermore, in the colony, it is thought that workers will remain arrhythmic performing in-hive tasks and will develop circadian rhythmicity just prior to the onset of foraging behavior, suggesting that ontogeny of circadian rhythms is intertwined with age-related division of labor in the colony. Studies examining the timing of in-hive tasks such as brood care found that individual 'nurses' performed this task around the clock, which is thought to benefit the developing brood (Moore et al., 1998).

In isolation, during the first days of their adult life young bees lack behavioral, metabolic or daily oscillations in circadian gene expression in the brain, that are associated with circadian rhythmicity. Under these constant conditions (DD, ~60%RH, 26-30°C), researchers have reported that ontogeny of circadian rhythmicity occurs at around 7-10 days of age in 50% of the sampled subjects (Moore, 2001; Toma et al., 2000). Furthermore, under these experimental conditions by 16 days of age around 25% of the bees were still arrhythmic.

Since ontogeny of circadian rhythms is thought to be regulated by age-related division of labor, researchers have manipulated neuroendocrine signals known to accelerate onset of foraging (such as juvenile hormone, octopamine and cGMP dependent protein kinase), hypothesizing a similar effect on circadian rhythms without success in individually isolated bees (Ben-Shahar, 2003; Bloch et al., 2002; Meshi and Bloch, 2007). A recent study examined whether the colony environment or other social cues may elicit strong circadian rhythms in young workers (Eban-Rothschild et al., 2012). Their findings reveal that experiencing the colony environment, either in a mesh cage or interacting with other bees for 48 hours after adult emergence, resulted in strong circadian rhythms when bees were brought to the laboratory. The authors of this work postulate that social cues, the colony microenvironment or a combination of both plays a role in the ontogeny of circadian rhythms of young workers. Taken together, these studies suggest the existence of a cue, which can be social or environmental, that elicits the development of circadian rhythmicity.

Honey bee colonies are able to efficiently regulate the colony microenvironment (Jones et al., 2004, 2007; Kronenberg and Heller, 1982; Seeley, 1974; Simpson, 1961). Studies have shown that bees regulate CO<sub>2</sub> levels, humidity and temperature inside the colony (Ohashi et al., 2008). In response to an increase in CO<sub>2</sub> levels inside the colony honey bee workers begin fanning until CO<sub>2</sub> levels diminish (Seeley, 1974). While the ability of honey bees to control temperature has been the main interest of researchers, humidity inside the nest is also regulated by workers (Human et al., 2006). Studies have shown that colonies with a naturally mated queen, are able to regulate temperature better than colonies that originate from a single drone artificially inseminated queen (Jones et al., 2004). This temperature control is especially important, since deviations of more than 1.5°C from 35°C at the core of the hive during larval and pupal development can have lasting changes in the adult honey bee (Winston, 1987).

Environmental temperature is also important for locomotor activity rhythms. Studies examining the endogenous rhythms of the Japanese honey bee *Apis cerana* show that environmental temperature has a direct effect on the endogenous period length of foragers (Fuchikawa and Shimizu, 2007). Recent work in our laboratory using the gentle Africanized honey bee (*gAHB*) also shows that environmental temperature affects the endogenous period length in honey bee foragers (Giannoni-Guzmán et al., 2014). However, the effect of temperature in the development of circadian rhythms in honey bee workers has yet to be explored.

In the current study we examined the effects of environmental temperature on the development of circadian rhythms in young workers. We hypothesized that temperature at the center of the colony is important for the development of circadian rhythms in young honey bee workers. In order to test this hypothesis, we isolated 1-day-old workers in locomotor activity monitors either at 25°C or 35°C. We examined the endogenous period length of rhythmic individuals in each group, the variation in period length and the mortality between the groups. Lastly, given the previous body of work that indicates that the first 48 hours after emergence are important for the development of circadian rhythms, we examined the effect of colony temperature during these 48 hours by placing individuals at 35°C and then changing the temperature to 25°C. Our results highlight the importance of socially regulated temperature of the hive in the ontogeny of circadian rhythms in honey bee workers.

## Materials and Methods:

### Honey bees Colonies and collections:

Colonies used in our experiments had mated queens that were laying eggs of gentle Africanized honey bees (Gallindo-Cardona et al., 2013). These colonies were located at the University of Puerto Rico (UPR) Gurabo Experimental Station in Gurabo, Puerto Rico. For all experiments, brood frames were collected, workers were removed and then the frame was stored in an incubator overnight (~35°C). The following morning, bees that emerged from the frames were collected and placed inside individual tubes for locomotor activity monitoring. The first colony of experiment 1 was examined on November 29, 2012 (colony 1), while the second colony was assayed beginning January 12, 2013 (colony 2). A total of 320 bees were used in this experiment, 256 for colony 1 and 64 for colony 2. Experiment 2 examined the effect of temperature during the first 48 hours after eclosion on the development of circadian rhythms, fixed began on February 26, 2016.

### Experiment 1: Development of Circadian rhythms at 25°C vs. 35°C

Locomotor activity measurements were carried out using two environmental chambers (Percival, I-30BLL) set up under constant darkness, relative humidity of 80%±5% and temperature of 25±0.5°C or 35±0.5°C and maintained constant throughout the experiments. Locomotor activity was recorded using monitors and software from Trikinetics (Waltham, MA, USA) as previously described (Giannoni-Guzmán et al., 2014). Briefly, 1-day-old workers were collected from the brood frame and placed inside individual tubes within the activity monitoring system. Food in the form of honey candy (mixed sugar and honey) and water were provided “*ad-libitum*” and changed as needed. Circadian rhythmicity was determined using 4 consecutive days of data (days 6-10), using autocorrelation analysis for 1-minute bins (Levine et al., 2002). All bees were approximately the same age for periods where rhythmicity was analyzed.

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## 126 **Experiment 2: Development of circadian rhythms after 48 hours at 35°C**

127 As in experiment 1, we carried out locomotor activity measurements using two  
128 environmental chambers. In one of these the temperature during the first 48 hours was set at  
129 35°C and afterwards lowered to 25°C for the remainder of the experiment. The other incubator  
130 was kept at 25°C throughout the experiment. Food and water were provided *ad libitum* and  
131 changed as needed.

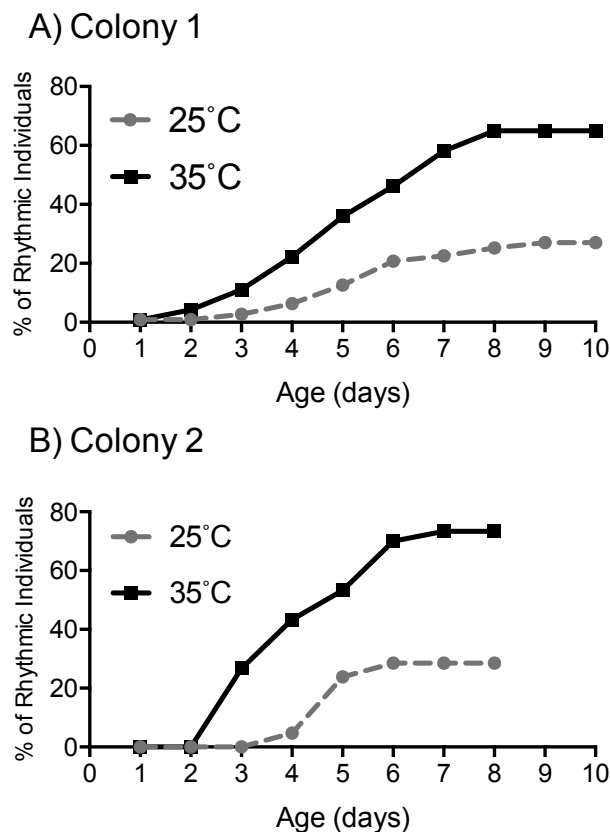
## 132 **Data analysis:**

133 All data sets were tested for normality via a Goodness of Fit test and appropriate  
134 nonparametric statistics were used where needed. The locomotor activity of each individual  
135 was processed using freely available MatLab® toolboxes developed in Jeffrey Hall's  
136 laboratory (Levine et al., 2002). Visual examination of locomotor activity for each individual in  
137 the form of actograms was utilized to determine the age at onset of circadian rhythms.  
138 Repeated measures MANOVA were utilized to determine if there were significant differences  
139 between the onset of rhythmicity between each of the experimental groups. Autocorrelation  
140 plots were utilized to confirm rhythmicity and calculate period length for each bee. Period  
141 length analysis was calculated for days. To examine differences in average period length  
142 between cohorts a two-way ANOVA was performed. To determine differences in the degree  
143 of period length variation the Levine's test for equality of variance was performed.

144 To determine if environment temperature influences survival in our experiments, we  
145 performed survival analysis via the Gehan-Breslow-Wilcoxon test. Furthermore, Proportional  
146 Hazards analysis was performed to determine if differences in mortality were the result of  
147 independent factors or a combination of different factors. All statistical analyses were  
148 performed using the JMP™ software package from SAS (SAS Institute Inc., 2009); graphs

149 and figures were created in MATLAB (MathWorks, Inc., Natick, MA, USA) and GraphPad  
150 Prism 6.00 (GraphPad Software, La Jolla, CA, USA).

# 151 Results:



**Figure 1. Rate and proportion of young workers developing circadian rhythms is greater at 35°C than at 25°C.** Cumulative distribution of rhythmic young workers at 25°C and 35°C in constant darkness for two colonies. At 35°C the rate of development and the proportion of 1-day-old bees developing strong circadian rhythms were higher than at 25°C. Repeated measures MANOVA for each of the colonies samples yielded significant differences between the 25°C and 35°C conditions for both colonies sampled **A)** Colony 1 ( $F=3.94$ ,  $df=9$ ,  $p<<0.001$ ). **B)** Colony 2 ( $F=3.29$ ,  $df=7$ ,  $p<0.01$ ).

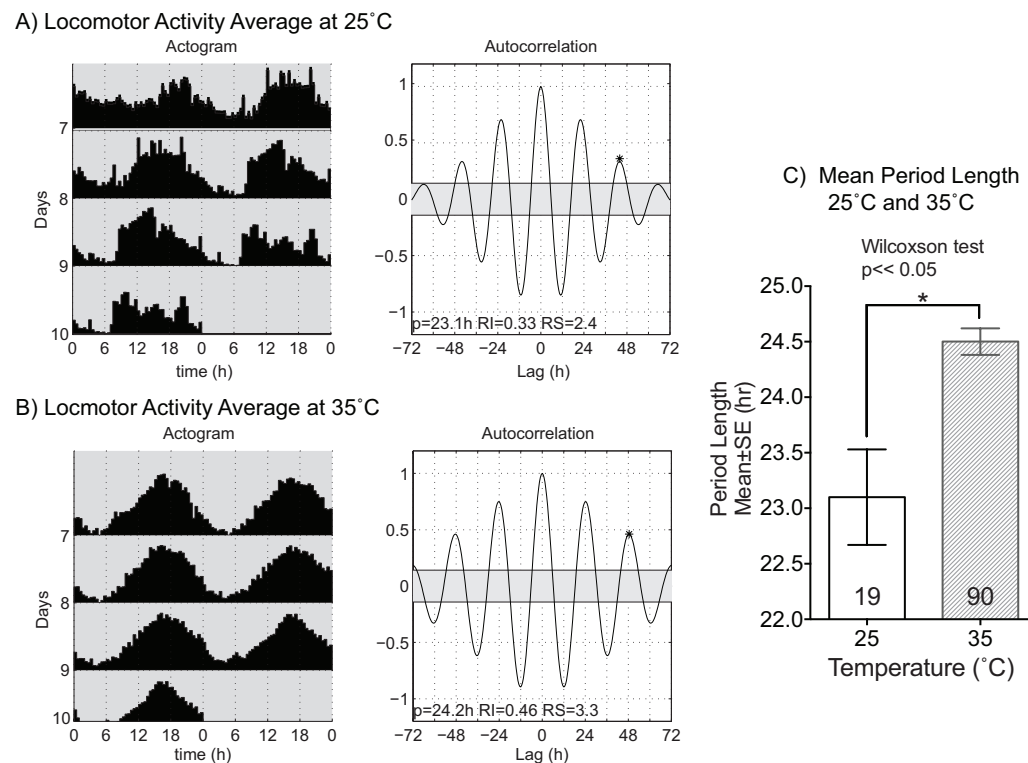
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170 Consistent with the hypothesis that brood nest temperature is important for the  
171 ontogeny of circadian rhythms, our results show that young workers kept at 35°C developed  
172 circadian rhythms as early as 2 days of age compared to young workers kept at 25°C, which  
173 began developing rhythms between 4-5 days of age (Figure 1). In addition, at 35°C between  
174 60-80% of workers developed circadian rhythms, while at 25°C less than 30% of the bees  
175 developed rhythmicity (Repeated measures MANOVA, colony 1:  $F=3.94$ ,  $df=9$ ,  $p<<0.001$ ;  
176 colony 2:  $F=3.29$ ,  $df=7$ ,  $p<0.01$ ) (Figure 1). This result indicates that colony temperature plays  
177 a key role in the development of circadian rhythmicity. Further examination of locomotor





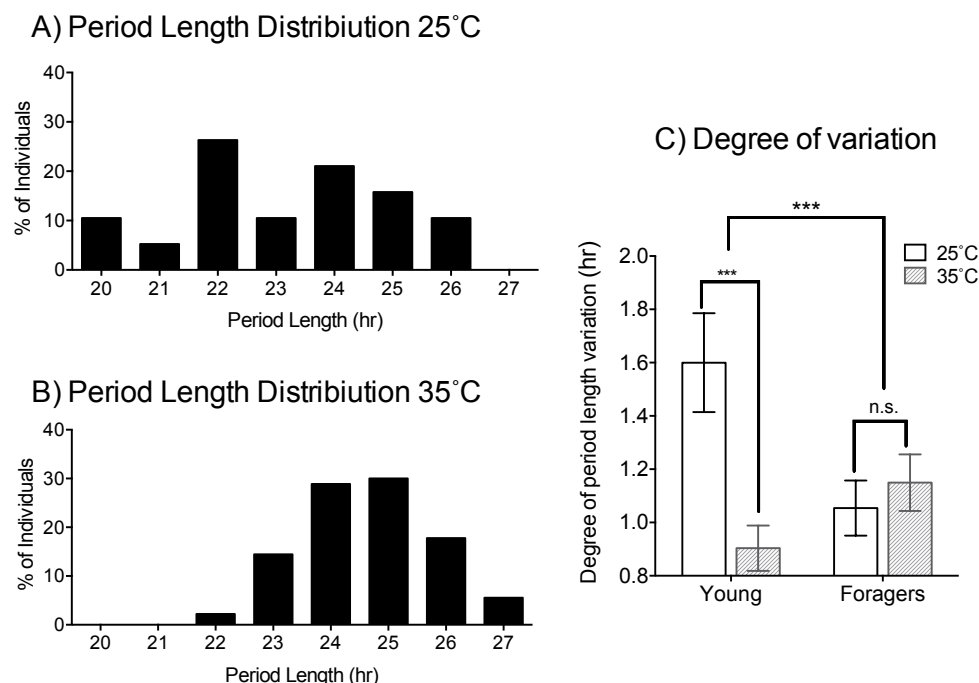
Recent work on different species of honey bees has shown that environmental temperature affects endogenous period length of foragers (Fuchikawa and Isamu Shimizu, 2007; Giannoni-Guzmán et al., 2014). We hypothesized that rhythmic young workers would present endogenous rhythms closer to 24 hours when assayed at 35°C than those assayed at 25°C. To test our hypothesis, we compared the endogenous periods of days 6-10 for rhythmic bees kept at 25°C or 35°C. The resulting analysis revealed that bees kept at 25°C have an average endogenous period length of 23.10hr, compared to that of bees kept at 35°C, whose average period was 24.5hr (Figure 3). This finding is consistent with previous work testing the endogenous period length in foragers (Giannoni-Guzmán et al., 2014; Moore and Rankin, 1985; Spangler, 1972; Toma et al., 2000).



**Figure 3. Average endogenous period length of young bees at 35°C is closer to 24 hours.** Double-plotted actograms and autocorrelations of the average locomotor profile rhythmic bees at **A)** 25°C and **B)** 35°C for days 6-10. **C)** The mean period length at 35°C (24.5±0.13h SEM) was closer to 24 hours and significantly different from that measured in the 25°C cohort (23.10±0.29h SEM) (ANOVA  $F=18.59$ ,  $df=1$ ,  $p<<0.01$ ).

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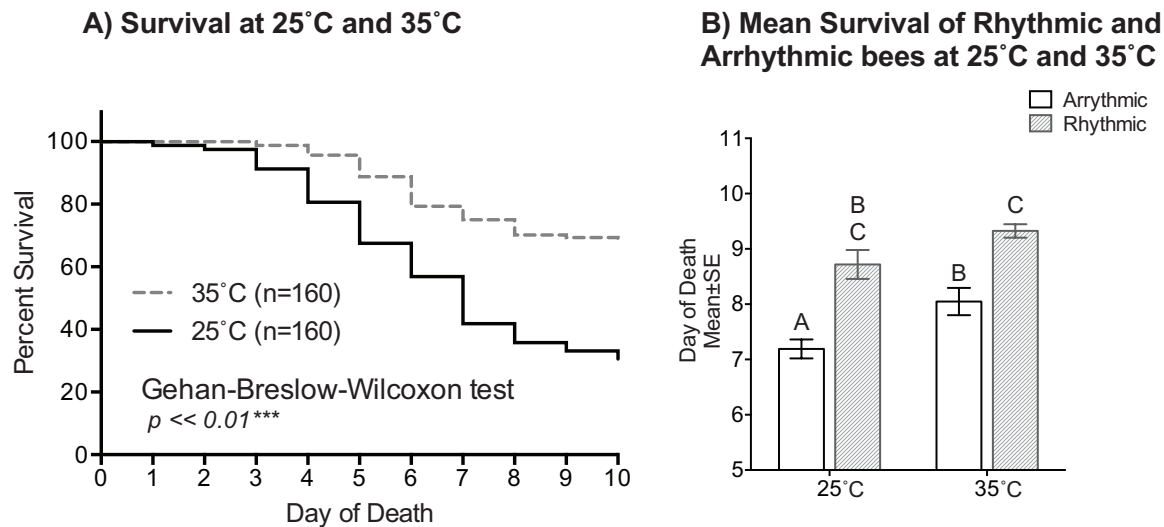
206 Interestingly, we observed that the period length standard error of the 25°C group was  
 207 higher than that of the 35°C group. By observing the distributions of period length for each of  
 208 the group it was evident that the 25°C group presented a larger degree of variation than the  
 209 35°C group (Figure 4A, B). To quantify this variation, we performed Levene's test of equality  
 210 of variance, which confirmed that period length in the 25°C cohort varies significantly more  
 211 than that of the 35°C cohort ( $F=17.9$ ,  $df=1$ ,  $p<<0.01$ ) (Figure 4C). Interestingly, this result does  
 212 not translate to foragers, where the degree of variation in endogenous period length was not  
 213 significantly different between foragers at 25°C or 35°C conditions (Levene's test,  $F=0.35$   
 214  $df=1$ ,  $p=0.56$ ) (Figure 4C). Multiple comparisons between young workers and foragers at 25°C  
 215 and 35°C, revealed that the degree of variation of foragers was similar to that of young workers  
 216 at 35°C and significantly different from that of young bees at 25°C (Figure 4C). These results  
 217 suggest that colony temperature after adult emergence plays an important role in the  
 218 development of circadian circuitry in the honey bee system.



**Figure 4. Individual variation of endogenous period length is greater at 25°C than at 35°C**

Frequency distributions of endogenous period length of days 6-10 from rhythmic young workers at **A)** 25°C and **B)** 35°C. **C)** Bar graph comparing the degree of period length variation as calculated by Levene's test of equality of variance for young workers and foragers at 25°C (white bars) and 35°C (gray shaded bars). Significant differences were observed between the young worker cohorts ( $F=17.9$ ,  $df=1$ ,  $p<<0.01$ ), while differences comparison within foragers was not significant ( $F=0.35$   $df=1$ ,  $p=0.56$ ). Multiple comparisons test revealed significant differences ( $p<0.05$ ) between young workers at 25°C and foragers at either 25°C or 35°C.

During the data analysis of the experiments, another difference that was noticed between the 25°C and 35°C cohorts was their mortality. When we compared the mortality of each group we observed that by day 10 only ~30% of bees in the 25°C cohort survived (Figure 5A). Significantly, this was less than half of the mortality observed in the 35°C cohort, where more than ~65% of the bees were still alive (Gehan-Breslow-Wilcoxon test,  $p<<0.01$ ). This result is somewhat surprising since our experiments with foragers under the same experimental setup did not reveal significant differences in mortality (unpublished results). Furthermore, by separating each cohort by individuals who developed or did not develop circadian rhythms, we observed a relationship between arrhythmicity and mortality in both groups (Figure 5B). Nonparametric Kruskal-Wallis rank sums test revealed significant differences between arrhythmic and rhythmic individuals at 25° and at 35°C ( $F=78.13$ ,  $df=3$ ,  $p<<0.01$ ). Post hoc analysis using Wilcoxon each pair test uncovered significant differences between 3 of the 4 groups tested, the exception being the comparison of rhythmic individuals at 25°C and arrhythmic individuals at 35°C. In order to ascertain potential factors playing a role in the mortality of honey bee workers, we used a proportional hazards model analysis.

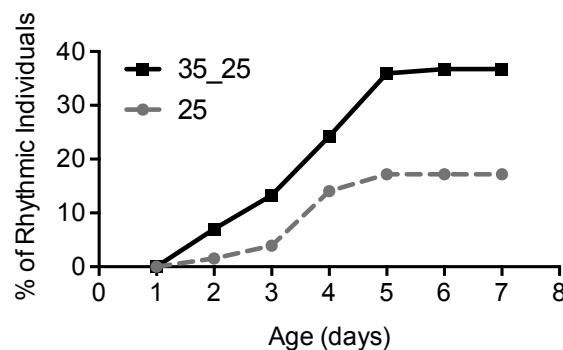


**Figure 5. Mortality of isolated young workers greatly increases at 25°C and in arrhythmic individuals.** A) Survival plot of 1-day-old honey bee cohorts at 25°C (solid line) and at 35°C (intermittent line). Both visual and statistical comparison of the cohorts revealed that survival of individuals was higher in the 35°C cohort (Gehan-Breslow-Wilcoxon,  $n=320$ ,  $p < 0.01$ ). B) Bar graph of mean survival and standard error of arrhythmic and rhythmic individuals separated by experimental cohort (25°C or 35°C). Proportional Hazards model revealed that temperature and rhythmicity have independent effects on mortality in young workers (Temperature:  $X^2=12.35$ ,  $df=1$ ,  $p < 0.001$ ; Rhythm:  $X^2=15.64$ ,  $df=1$ ,  $p < 0.001$ ; Temperature\*Rhythm:  $X^2=0.055$ ,  $df=1$ ,  $p=0.8142$ ). Wilcoxon each Pair test revealed significant differences ( $p < 0.05$ ) between paired comparisons represented by different letters.

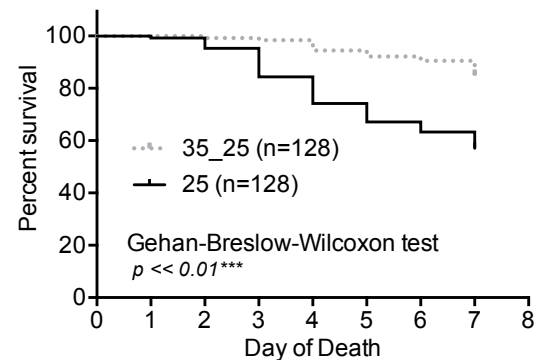
In this analysis environmental temperature, rhythmicity (whether the individual developed rhythms or was arrhythmic throughout the experiment) and the interaction of these factors were tested as the variables causing the observed mortality. The resulting analysis revealed that environmental temperature and rhythmicity, independently, have a significant effect on the mortality of young workers in our assay, while their interaction was not significant (Temperature:  $X^2=12.35$ ,  $df=1$ ,  $p < 0.001$ ; Rhythm:  $X^2=15.64$ ,  $df=1$ ,  $p < 0.001$ ; Temperature\*Rhythm:  $X^2=0.055$ ,  $df=1$ ,  $p=0.8142$ ). The combined results suggest that in our experiments mortality is caused by the environmental temperature and the inability to develop a circadian rhythm independently and not their combination.

Our result that temperature positively influences the rate and proportion of individuals developing circadian rhythms combined with the findings from a recent study (Eban-Rothschild et al., 2012) that the first 48 hours in the colony influenced development of strong circadian rhythms led us to postulate the following prediction: If temperature is a key factor in the development of circadian rhythmicity during the first 48 hours after emergence in young workers, then placing 1-day old workers at 35°C for the first 48 hours after emergence and afterwards changing environmental temperature to 25°C, will result in a greater proportion of individuals developing circadian rhythms than 1 day-old workers placed at 25°C. To test this hypothesis, we placed 1-day old bees at either 35°C or 25°C group, after 48 hours, we changed the temperature to 25°C in the first group (35-25°C). Consistent with this prediction we found that exposure to 35°C during the first 48 hours after emergence plays a significant role in the development of circadian rhythms in young workers (Figure 6A). Repeated measures comparison of the cumulative distribution of rhythmic individuals for the 35-25°C group and bees continuously at the 25°C group, was significantly different ( $F=3.28$ ,  $df=6$ ,  $p<0.01$ ). In addition to the effects of temperature on the development of circadian rhythm, we also observed significant differences in the survival of individuals exposed to 35°C for the first 48 hours and those that were kept at 25°C. By day 7 less than 12 individuals had died in the 35-25°C group, while more than 50 had died in the 25°C (Gehan-Breslow-Wilcoxon,  $n=256$ ,  $p<<0.01$ ). Taken together, temperature in the colony plays a key role in the development of circadian rhythms of workers.

### A) Cumulative distribution of Rhythmic bees



### B) Survival Analysis of bees at 35°C for the first 48hrs after emergence vs bees at 25°C



**Figure 6. Temperature (35°C) during the first 48 hours after emergence is sufficient to rescue the rhythmicity and mortality effects of 25°C.** **A)** Cumulative distribution of rhythmic young workers exposed to 35°C during the first 48 hours after emergence and afterwards placed at 25°C for the remainder of the experiment (35-25°C) compared to that of bees placed at 25°C after emergence. Repeated measures MANOVA revealed significant differences between the rate and proportion of individuals developing rhythmic behavior under these conditions ( $F=3.28$ ,  $df=6$ ,  $p<0.01$ ). **B)** Survival plot of 1-day-old honey bee cohorts at 25°C (solid line) and bees exposed to 35°C for the first 48 hours after emergence (intermittent line). Individuals in the 35-25°C cohort presented significantly better survival rates than bees placed at 25°C since the beginning of the experiment (Gehan-Breslow-Wilcoxon,  $n=256$ ,  $p<0.01$ ).

### Discussion:

In the current study, we show that colony temperature plays a key role in the ontogeny of circadian rhythms of young honey bee workers. Previous studies exploring the ontogeny of circadian rhythms of young workers established that circadian rhythms both in the field and in isolation commence around 7-9 days after eclosion (Bloch et al., 2001; Moore et al., 1998; Toma et al., 2000). Experiments that followed uncovered that exposure to the colony environment during the first 48 hours after eclosion significantly impacts the development of circadian rhythms under isolation (Eban-Rothschild et al., 2012). Here we show evidence indicating that the regulation of temperature (~35°C) in the colony is a key social factor

determining the development of circadian rhythms. Placing 1-day old workers at the hive's core temperature in the laboratory results in an accelerated rate and increased proportion of individuals developing rhythmicity (Figure 1). Moreover, we show that this temperature is particularly important during the first 48 hours after eclosion and kept exposure to 35°C in this period is sufficient for early rhythm development (Figure 6). Analysis of the endogenous period length variation of rhythmic bees, suggests that temperature may play a role in the development of the neural circuitry that regulate circadian rhythms (Figures 2,3,4). Lastly, mortality differences between experimental groups were associated with development of circadian rhythms. Taken together, socially regulated temperature plays a key role in the ontogeny of circadian rhythms in honey bee workers.

The proportion and rate of honey bee workers developing circadian rhythms in the 35°C cohort is consistent with work examining the effect of colony environment on circadian rhythms, where after 48 hours of colony exposure between 60-80% of bees presented circadian rhythms (Eban-Rothschild et al., 2012). In addition, individuals exposed to 35°C for the first 48 hours after eclosion presented twice the proportion (40%) of rhythmic bees than bees that at 25°C (20%). This suggests that temperature is one of several factors that play a role in the development of circadian rhythmicity. Furthermore, our results in the 25°C groups are very similar to those of bees that only spent the the first 24 hours after eclosion inside the colony (Eban-Rothschild et al., 2012). Further studies are needed to ascertain the relative importance of temperature on the ontogeny of circadian rhythmicity compared to other colony factors. Studying the neural changes that may occur during the critical period of 24-48 hours after emergence in the honey bee nervous system, may provide clues as to the other factors that influence development of circadian rhythms.

In our data and that of previous studies examining the ontogeny of circadian rhythms in workers, we can observe that not all individuals develop circadian rhythms by the end of



the experiment (Eban-Rothschild et al., 2012; Meshi and Bloch, 2007; Toma et al., 2000). While the percent of arrhythmic individuals at 35°C is similar to that of previous studies at the end of the experiment, at 25°C the percent of arrhythmic individuals more than double of that in previous studies (Figure 1). One possible factor that is influencing this result is the time of the year the experiments were carried out, which was winter in Puerto Rico. During winter there are drastic changes in the colony demography and dynamics, such as reduction of brood and complete cease of foraging behavior (Doke et al., 2015). These changes have been mostly studied in temperate zones, where the seasons are marked by drastic changes in weather and may not be necessarily applicable to Puerto Rico. Based on our current data set, further experiments are required to accept or discard the effect of season in the ontogeny of circadian rhythmicity.

Our results and those of other studies provide strong evidence that temperature plays a key role in the ontogeny of circadian rhythms in young workers. However, other studies in the field and laboratory provide evidence that other factors influence the development of circadian rhythmicity, such as genetic background and social environment. Monitoring the behavior of individual bees in the colony as they aged, researchers have shown that rhythmicity, measured using standing behavior as the measure of inactivity in the colony, found that bees of fast genotypes, which show accelerated behavioral development into foragers (Giray and Robinson, 1994), present circadian rhythms as early as 4-7 days of age in some cases, while the slow genotype bees did not show rhythms until the 16-19-day interval (Moore et al., 1998). This finding is consistent with the onset of foraging in slow and fast genotype groups (Giray et al., 1999). The authors of this study conclude that bees inside the hive present rhythmic activity much earlier than onset of foraging. While in the current study we did not control the genetic background of our bees, we did examine two different colonies and obtained similar results.

Since our experiments were performed in the laboratory and individuals were isolated, we cannot measure in the current data set the effects of pheromone on the development of circadian rhythms. However, studies have shown that exposure to the foragers advances development of circadian rhythmicity, while bees housed with young bees of their same age cohort develop rhythmicity later (Meshi and Bloch, 2007). Furthermore, young workers that had direct contact with the brood did not show circadian rhythms even when outside the hive and under light/dark cycles (Shemesh et al., 2007, 2010). Taken together, ontogeny of circadian rhythms in the honey bee colony context is regulated by the socially regulated factors of temperature, social interactions with brood, foragers and young workers, and potentially genetic background.

Endogenous period length of young bees at 35°C ( $24.5 \pm 0.12$  hr SE) was on average closer to the Earth's rotational period than that of individuals at 25°C ( $23.1 \pm 0.43$  hr SE) (Figure 2 and Figure 3). This result is consistent with our previous work on honey bees and work on *Apis cerana* where environmental temperature influenced endogenous period length, suggesting that the circadian clock of young workers is able to compensate for environmental temperature changes (Fuchikawa and Shimizu, 2007; Giannoni-Guzmán et al., 2014). While differences between 25°C and 35°C cohorts in average period length was consistent with that of foragers, the degree of period length variation was different between young workers at 25°C and 35°C, while the degree of variation in foragers at both 25°C and 35°C was similar to that of young workers at 35°C (Figure 4). Since foraging is the last job a worker performs before dying, these similarities in period length variation between young workers kept at 35°C and foragers at both temperatures is most likely related to foragers having spent the majority of their life inside the colony. The foragers in this study were captured at the entrance of the colony, so we can assume that they had fully developed circadian rhythms. This result suggests that bees exposed to 25°C from a young age may have differences in the

development of the circadian network or present a lack of communication between different clocks in the honey bee circadian system.

With regard to the survival rates observed in bees at 35°C and those at 25°C (Figure 5), our analysis indicates that environmental temperature and lack of rhythmicity are independently decreasing survival rates. The effect of temperature on mortality is consistent with our data that temperature is important for the development of circadian rhythmicity and that changes in temperature during development can have long lasting effects later in the honey bee's life (Becher et al., 2009; Jones et al., 2004; Tautz et al., 2003). It is possible that in addition to development of circadian rhythms, other systems are still under development and do not develop properly at 25°C causing the observed mortality.

Comparing the development of circadian rhythms of honey bee workers with that of other insects suggests that the postembryonic ontogeny in honey bees may be a product of the colony's social context. Studies examining the circadian rhythms of various insects show rhythmic activity at even pre-adult stages (Fantinou et al., 1998; Kaneko and Hall, 2000; Page and Block, 1980; Tomioka and Chiba, 1982). In the case of crickets and cockroaches, circadian rhythmicity has been documented in pre-adult nymph stages and its patterns change as individuals age (Page and Block, 1980; Tomioka and Chiba, 1982). In other insects such as egg-parasitic wasp *Telenomus busseolae*, adult emergence is timed by their entrainment of light/dark cycles, providing evidence of early development of the circadian system (Fantinou et al., 1998). In contrast to honey bee brood which is kept at almost constant conditions, in these insects the pre-adults (eggs, larvae, pupae, nymphs) are at the mercy of the external environment and having a working circadian system becomes necessary for their survival. In the case of honey bees, since conditions are constant during development, the ability to predict changes in the environment during larval and pupal development becomes less necessary, thus it is possible that honey bee circadian rhythms have evolved to developed

after adult emergence when they are needed. For example, in marsupials, such as kangaroos, where gestation is short and many developmental processes occur after birth, the front limbs are much more developed than other systems because upon birth they are required in order to climb to the maternal pouch and to the mother's nipple to feed (Wittmann, 1981, 1984). At what exact stage of development and what processes are driving the ontogeny of circadian rhythms in honey bee workers is a subject of further research.

In order to present circadian rhythms of locomotion, the connectivity between various systems is necessary. At the brain level, it is known that multiple oscillators that control the timing of locomotor activity, at different times of the day (e.g. morning and evening cells), not only need to communicate but they need to be synchronized in a specific manner (Stoleru et al., 2004, 2005). One of the possible processes that may be occurring in the first 48 hours after emergence in workers is the establishment of connections between the multiple oscillators in the honey bee brain. Another circuit that is necessary for locomotor rhythms is the connectivity between motor neurons and the different oscillators in the brain (Blanchardon et al., 2001). Motor neurons are organized forming central pattern generators that coordinate the movement of extremities independently of the brain. However, without a signal from the brain, the initiation and regulation of locomotor rhythms is not possible (Allada et al., 1998). If the formation of this connection is regulated by temperature in honey bees and is occurring in this 48-hour window after emergence, then arrhythmicity may be explained by the failure to establish this connection.

An additional process that is important in the regulation of circadian locomotion is the connections between motor neurons and muscles (i.e. Neuromuscular junction (NMJ)). This connectivity has been studied extensively in multiple insects, and the cellular and molecular processes have been well characterized in *Drosophila* (H Keshishian et al., 2003). Experiments exploring the effects of temperature on these connections show a temperature

dependent plasticity of motor nerve terminal arborization, where at higher temperatures more arborization of the nerve terminal occurs (Zhong, 2004). In honey bees, measuring circadian gene expression in the brain and muscle of young arrhythmic workers indicates that the muscle clock oscillates, while the brain's clock did not seem to oscillate (Ben Attia, 2014). Based on what is known in other models and this finding it is possible that different oscillators in the brain have not synchronized with each other and that the connection between the brain's oscillators and peripheral oscillators, at the time of collection, has not been established and requires further research.

In conclusion, this study shows for the first time the effects of colony temperature on the ontogeny of circadian rhythms, specifically during the first two days after adult emergence. Future studies will examine the weight of temperature as a factor in the development of circadian rhythms and examine the weight of other factors, such as genetic background and social cues. In addition, carefully examining the changes at the neural and gene expression levels occurring during the first 48 hours may provide insight into the mechanisms driving the ontogeny of circadian rhythms in honey bee workers, which remain to be elucidated.

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## References:

- Allada R; NE White; W V So; JC Hall and M Rosbash (1998) A mutant *Drosophila* homolog of mammalian Clock disrupts circadian rhythms and transcription of period and timeless. *Cell* 93(5): 791–804. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/9630223>.
- Becher MA; H Scharpenberg and RF Moritz (2009) Pupal developmental temperature and behavioral specialization of honeybee workers (*Apis mellifera* L.). *Journal of comparative physiology. A, Neuroethology, sensory, neural, and behavioral physiology* 195(7): 673–679. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/19390855>.
- Ben-Shahar Y (2003) cGMP-dependent changes in phototaxis: a possible role for the foraging gene in honey bee division of labor. *Journal of Experimental Biology* 206(14): 2507–2515. Available from: <http://jeb.biologists.org/cgi/doi/10.1242/jeb.00442> (accessed 1 November 2014).
- Blanchardon E; B Grima; A Klarsfeld; E Chelot; PE Hardin; T Preat and F Rouyer (2001) Defining the role of *Drosophila* lateral neurons in the control of circadian rhythms in motor activity and eclosion by targeted genetic ablation and PERIOD protein overexpression. *European Journal of Neuroscience* 13(5): 871–888. Available from: <http://doi.wiley.com/10.1046/j.0953-816x.2000.01450.x> (accessed 23 May 2016).
- Bloch G; DP Toma and GE Robinson (2001) Behavioral Rhythmicity, Age, Division of Labor and period Expression in the Honey Bee Brain. *Journal of biological rhythms* 16(5): 444–456.
- Bloch G; JP Sullivan and GE Robinson (2002) Juvenile hormone and circadian locomotor activity in the honey bee *Apis mellifera*. *Journal of insect physiology* 48: 1123–1131.
- Doke MA; M Frazier and CM Grozinger (2015) Overwintering honey bees: biology and

484 management. *Current Opinion in Insect Science* 10: 185–193. Available from:  
 485 <http://www.sciencedirect.com/science/article/pii/S2214574515000930> (accessed 16  
 486 June 2015).

487 Eban-Rothschild A; Y Shemesh and G Bloch (2012) The colony environment, but not direct  
 488 contact with conspecifics, influences the development of circadian rhythms in honey  
 489 bees. 2012/06/02 ed. *Journal of biological rhythms* 27(3): 217–225. Available from:  
 490 <http://www.ncbi.nlm.nih.gov/pubmed/22653890>.

491 Fantinou AA; MP Alexandri and JA Tsitsipis (1998) Adult emergence rhythm of the egg-  
 492 parasitoid *Telenomus busseolae*. *Biocontrol* 43(2): 141–151.

493 Fuchikawa T and I Shimizu (2007) Circadian rhythm of locomotor activity in the Japanese  
 494 honeybee, *Apis cerana japonica*. *Physiological Entomology* 32(1): 73–80.

495 Fuchikawa T and I Shimizu (2007) Effects of temperature on circadian rhythm in the  
 496 Japanese honeybee, *Apis cerana japonica*. *Journal of insect physiology* 53(11): 1179–  
 497 1187. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/17655856> (accessed 20  
 498 April 2014).

499 Giannoni-Guzmán MA; A Avalos; J Marrero Perez; EJ Otero Loperena; M Kayım; JA  
 500 Medina; SE Massey; M Kence; A Kence; T Giray and JL Agosto-Rivera (2014)  
 501 Measuring individual locomotor rhythms in honey bees, paper wasps and other similar-  
 502 sized insects. *The Journal of experimental biology* 217(Pt 8): 1307–15. Available from:  
 503 <http://www.ncbi.nlm.nih.gov/pubmed/24436380> (accessed 30 July 2014).

504 Giray T and GE Robinson (1994) Effects of intracolony variability in behavioral development  
 505 on plasticity of division of labor in honey bee colonies. *Behavioral Ecology and*  
 506 *Sociobiology* 35(1): 13–20. Available from:  
 507 <http://link.springer.com/10.1007/BF00167054> (accessed 25 September 2014).

508 Giray T; E Guzman-Novoa; CW Aron; B Zelinsky; SE Fahrbach and GE Robinson (1999)  
509 Genetic variation in worker temporal polyethism and colony defensiveness in the honey  
510 bee, *Apis mellifera*. *Behavioral Ecology* 11(1): 44–55. Available from: <Go to  
511 ISI>://WOS:000088355900007.

512 Goodwin RM and D Lewis (1987) Honeybees use a biological clock to incorporate sun  
513 positions in their waggle dances after foraging under heavy overcast skies. *New Zealand*  
514 *Entomologist* 10: 138–140.

515 H Keshishian; K Broadie; and A Chiba and M Bate (2003) The *Drosophila* Neuromuscular  
516 Junction: A Model System for Studying Synaptic Development and Function. Annual  
517 Reviews 4139 El Camino Way, P.O. Box 10139, Palo Alto, CA 94303-0139, USA.  
518 Available from:  
519 [http://www.annualreviews.org/doi/abs/10.1146/annurev.ne.19.030196.002553?journalC](http://www.annualreviews.org/doi/abs/10.1146/annurev.ne.19.030196.002553?journalCode=neuro)  
520 [ode=neuro](http://www.annualreviews.org/doi/abs/10.1146/annurev.ne.19.030196.002553?journalCode=neuro) (accessed 23 May 2016).

521 Human H; SW Nicolson and V Dietemann (2006) Do honeybees, *Apis mellifera* scutellata,  
522 regulate humidity in their nest? *Die Naturwissenschaften* 93(8): 397–401. Available  
523 from: <http://www.ncbi.nlm.nih.gov/pubmed/16670906> (accessed 29 February 2016).

524 Jones JC; MR Myerscough; S Graham and BP Oldroyd (2004) Honey bee nest  
525 thermoregulation: diversity promotes stability. *Science* 305(5682): 402–404. Available  
526 from: <http://www.ncbi.nlm.nih.gov/pubmed/15218093>.

527 Jones JC; P Nanork and BP Oldroyd (2007) The role of genetic diversity in nest cooling in a  
528 wild honey bee, *Apis florea*. *Journal of comparative physiology. A, Neuroethology,*  
529 *sensory, neural, and behavioral physiology* 193(2): 159–65. Available from:  
530 <http://www.ncbi.nlm.nih.gov/pubmed/17013621> (accessed 27 January 2014).

531 Kaneko M and JC Hall (2000) Neuroanatomy of cells expressing clock genes in *Drosophila*:



532 Transgenic manipulation of the period and timeless genes to mark the perikarya of  
533 circadian pacemaker neurons and their projections. *Journal of Comparative Neurology*  
534 422(1): 66–94.

535 Kronenberg F and HC Heller (1982) Colonial thermoregulation in honey bees (*Apis*  
536 *mellifera*). *Journal of Comparative Physiology ? B* 148(1): 65–76. Available from:  
537 <http://link.springer.com/10.1007/BF00688889> (accessed 18 April 2014).

538 Levine JD; P Funes; HB Dowse and JC Hall (2002) Signal analysis of behavioral and  
539 molecular cycles. *BMC neuroscience* 3: 1. Available from:  
540 [http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=65508&tool=pmcentrez&ren](http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=65508&tool=pmcentrez&rendertype=abstract)  
541 [dertype=abstract](http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=65508&tool=pmcentrez&rendertype=abstract).

542 Meshi A and G Bloch (2007) Monitoring circadian rhythms of individual honey bees in a  
543 social environment reveals social influences on postembryonic ontogeny of activity  
544 rhythms. *Journal of biological rhythms* 22(4): 343–355. Available from:  
545 <http://www.ncbi.nlm.nih.gov/pubmed/17660451>.

546 Moore D (2001) Honey bee circadian clocks: behavioral control from individual workers to  
547 whole-colony rhythms. *Journal of insect physiology* 47: 843–857.

548 Moore D and MA Rankin (1985) Circadian locomotor rhythms in individual honey bees.  
549 *Physiological Entomology* 10: 191–197.

550 Moore D; JE Angel; IM Cheeseman; SE Fahrbach and GE Robinson (1998) Timekeeping in  
551 the honey bee colony: integration of circadian rhythms and division of labor. *Behav*  
552 *Ecol Sociobiol* 43: 147–160.

553 Ohashi M; H Ikeno; T Kimura; T Akamatsu; R Okada and E Ito (2008) Control of hive  
554 environment by honeybee (*Apis mellifera*) in Japan. *Proceeding(1961)*: 2008.

555 Page TL and GD Block (1980) Circadian rhythmicity in cockroaches: Effects of early post-

embryonic development and aging. *Physiological Entomology* 5(3): 271–281.

SAS Institute Inc. (2009) *JMP® 8 User Guide*. Second Edi. Cary, NC: SAS.

Seeley TD (1974) Atmospheric carbon dioxide regulation in honey-bee (*Apis mellifera*) colonies. *Journal of Insect Physiology* 20(11): 2301–2305. Available from: <http://www.sciencedirect.com/science/article/pii/0022191074900523> (accessed 17 March 2016).

Shemesh Y; M Cohen and G Bloch (2007) Natural plasticity in circadian rhythms is mediated by reorganization in the molecular clockwork in honeybees. 2007/03/16 ed. *FASEB* 21(10): 2304–2311. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/17360847>.

Shemesh Y; A Eban-Rothschild; M Cohen and G Bloch (2010) Molecular dynamics and social regulation of context-dependent plasticity in the circadian clockwork of the honey bee. 2010/09/17 ed. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 30(37): 12517–12525. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/20844146> (accessed 1 April 2014).

Simpson J (1961) Nest Climate Regulation in Honey Bee Colonie: Honey bees control their domestic environment by methods based on their habit of clustering together. *Science (New York, N.Y.)* 133(3461): 1327–33. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/17744947> (accessed 17 March 2016).

Spangler HG (1972) Daily Activity Rhythms of Individual Worker and Drone Honey Bees. *Annals of the Entomological Society of America* 65(5): 1073–1076.

Stoleru D; Y Peng; JL Agosto-Rivera and M Rosbash (2004) Coupled oscillators control morning and evening locomotor behaviour of *Drosophila*. 2004/10/16 ed. *Nature* 431(7010): 862–868. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/15483615>.

Stoleru D; Y Peng; P Nawathean and M Rosbash (2005) A resetting signal between

580 *Drosophila* pacemakers synchronizes morning and evening activity. 2005/11/11 ed.  
581 *Nature* 438(7065): 238–242. Available from:  
582 <http://www.ncbi.nlm.nih.gov/pubmed/16281038>.

583 Tautz J; S Maier; C Groh; W Rossler and A Brockmann (2003) Behavioral performance in  
584 adult honey bees is influenced by the temperature experienced during their pupal  
585 development. *Proceedings of the National Academy of Sciences of the United States of*  
586 *America* 100(12): 7343–7347. Available from:  
587 <http://www.ncbi.nlm.nih.gov/pubmed/12764227>.

588 Toma DP; G Bloch; D Moore and GE Robinson (2000) Changes in period mRNA levels in  
589 the brain and division of labor in honey bee colonies. *Proceedings of the National*  
590 *Academy of Sciences of the United States of America* 97(12): 6914–6919.

591 Tomioka K and Y Chiba (1982) Post-embryonic development of circadian rhythm in the  
592 cricket, *Gryllus bimaculatus*: A rhythm reversal. *Journal of Comparative Physiology ???*  
593 *A* 147(3): 299–304.

594 Van Nest BN and D Moore (2012) Energetically optimal foraging strategy is emergent  
595 property of time-keeping behavior in honey bees. *Behavioral Ecology* 23(3): 649–658.  
596 Available from: <http://www.beheco.oxfordjournals.org/cgi/doi/10.1093/beheco/ars010>  
597 (accessed 26 May 2014).

598 von Frisch K (1967) *The Dance Language and Orientation of Bees*. Cambridge, MA.:  
599 Harvard University Press.

600 Wagner AE; BN Van Nest; CN Hobbs and D Moore (2013) Persistence, reticence and the  
601 management of multiple time memories by forager honey bees. *The Journal of*  
602 *experimental biology* 216(Pt 7): 1131–1141. Available from:  
603 <http://www.ncbi.nlm.nih.gov/pubmed/23197093>.

604 Winston ML (1987) *The biology of the honey bee*. Cambridge, MA: Harvard University  
605 Press.

606 Wittmann KJ (1981) Comparative biology and morphology of marsupial development in  
607 Leptomysis and other Mediterranean Mysidacea (Crustacea). *Journal of experimental*  
608 *marine Biology and Ecology*, Elsevier 52(2): 243–270.

609 Wittmann KJ (1984) Ecophysiology of marsupial development and reproduction in  
610 Mysidacea (Crustacea). *Oceanogr Mar Biol Annu Rev* 22: 393–428.

611 Zhong Y (2004) Neuronal Activity and Adenylyl Cyclase in Environment-Dependent  
612 Plasticity of Axonal Outgrowth in *Drosophila*. *Journal of Neuroscience* 24(6): 1439–  
613 1445. Available from: [http://www.jneurosci.org/cgi/doi/10.1523/JNEUROSCI.0740-](http://www.jneurosci.org/cgi/doi/10.1523/JNEUROSCI.0740-02.2004)  
614 02.2004.

615

616