

1 Relationship between stemness and transcriptionally-inferred PI3K 2 activity in human breast cancer

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44 **ABSTRACT**

45 The development of reliable, prognostically informative molecular tests to direct targeted cancer therapy is a major
46 challenge. In 2019, the PI3K α inhibitor alpelisib was approved for the treatment of advanced breast cancer, in
47 combination with the oestrogen receptor degrader fulvestrant, with some evidence for improved therapeutic
48 response in patients classified as having tumours which were positive for mutation in *PIK3CA*, the gene encoding
49 PI3K α . Using human pluripotent stem cells, we recently demonstrated that the *PIK3CA*^{H1047R} oncogenic hotspot
50 variant shows marked *PIK3CA* allele dose-dependent activation of PI3K signalling and induction of self-sustained
51 stemness. Together with recent discoveries of multi-copy and double-*cis* *PIK3CA* mutations in human cancers, this
52 calls for a re-evaluation of the *PIK3CA* genotype-phenotype relationship and the current use of binary stratification
53 by *PIK3CA* mutation status. Using computational analyses, we thus investigated the relationship between *PIK3CA*
54 mutational status, PI3K activity/signalling strength and stemness. Stemness and PI3K activity scores were calculated
55 using open-source methods and well-established transcriptional signatures. We report that a high PI3K pathway
56 activity score, but not the presence of *PIK3CA* mutation *per se*, predicts increased breast cancer dedifferentiation
57 and higher stemness, correlating with reduced overall survival. Our data (1) corroborate reports that the presence of
58 a *PIK3CA* mutation *per se* does not predict high PI3K pathway activation or poor prognosis; (2) suggest that
59 stratification of breast cancer for PI3K-based therapy might benefit from the use of a PI3K pathway activity score
60 rather than binary *PIK3CA* mutation status alone; (3) suggest that combination of PI3K pathway inhibitors with
61 differentiation-promoting treatments warrants evaluation in aggressive breast cancers with high PI3K activity and
62 stemness scores.

63

64 **INTRODUCTION**

65
66 Activating mutations in *PIK3CA* are among the most common somatic point mutations in cancer, together with
67 inactivation or loss of the tumour suppressor *PTEN*, a negative regulator of PI3K [1–3]. However, experimental
68 evidence suggests that heterozygous expression of a strongly activating *PIK3CA* mutation is not sufficient on its own
69 to transform cells in vitro or to induce tumourigenesis in vivo (reviewed in Ref. [4]). This is further supported by
70 observations in people with disorders in the *PIK3CA*-related overgrowth spectrum (PROS) which often carry the
71 same *PIK3CA* mutations as found in cancer but feature benign tissue overgrowth without excess risk of adult
72 malignancy [4].

73 We and others have recently shown that many *PIK3CA*-associated cancers harbour multiple independent
74 mutations activating the PI3K pathway, including multiple *PIK3CA* mutations in cis or trans [3,5–8]. We further
75 reported that human induced pluripotent stem cells (iPSCs) with two endogenous alleles of the strongly activating
76 cancer “hotspot” mutation *PIK3CA^{H1047R}* exhibit pronounced phenotypic differences to isogenic cells heterozygous
77 for the same *PIK3CA* variant [6]. These differences include partial loss of epithelial morphology, widespread
78 transcriptional reprogramming and self-sustained stemness in vitro and in vivo [6]. Collectively, these genetic and
79 cellular observations suggested that major cellular programmes implicated in cancer maintenance and progression
80 may be exquisitely sensitive to the strength of the pathological PI3K signal.

81 As well as being a model for normal early development, PSCs share key characteristics with cancer cells,
82 including developmental plasticity, the capacity for indefinite self-renewal, rapid proliferation and high glycolytic flux
83 [9]. Indeed, oncogenesis commonly features aberrant reactivation of primitive embryonic and tissue repair pathways
84 [10–12], while accumulating evidence suggests that tumour heterogeneity and therapeutic resistance is determined
85 in part by cancer stem cells [13]. Seminal genome-wide studies have demonstrated a striking enrichment for
86 embryonic gene signatures in human cancers, indicative of tumour dedifferentiation and aggressive disease [12,14].
87 *PIK3CA^{H1047R}* (over)expression or overexpression of wild-type *PIK3CA* have previously been linked to
88 dedifferentiation and stemness in mouse cancer models [15–21], particularly of the breast, yet gene dose-dependent
89 regulation by PI3K signalling has not been addressed. Importantly, no systematic profiling has previously
90 investigated whether the link between PI3K and stemness extends to human cancer; and if it does exist, whether it
91 may be used to guide patient stratification both for prognostic and therapeutic purposes.

92 Recently, the PI3K α -specific inhibitor alpelisib (Piqray/NVP-BYL719; Novartis) received FDA approval for the
93 treatment of advanced hormone-receptor (HR)-positive, HER2-negative breast cancers, following a randomised
94 phase III trial evaluating alpelisib with fulvestrant *versus* fulvestrant alone [22]. The trial concluded that a clinically-
95 relevant benefit of the combination therapy was more likely in patients with *PIK3CA*-mutant tumours [22]. The FDA-
96 approval of alpelisib was accompanied by approval of the companion diagnostic therascreen® *PIK3CA* test
97 (QIAGEN) which detects 11 *PIK3CA* “hotspot” mutations. Despite these advances, a substantial proportion of
98 patients with *PIK3CA*-mutant tumours failed to improve on the combination therapy [22], highlighting the need for
99 further refinements of current patient stratification strategies.

100 Mutation-centric approaches for stratification of patients with cancer are relatively easy to implement, using well-
101 established technical protocols and analytical pipelines. Such “hard-wired” genomic information nevertheless has
102 limited predictive value for cellular behaviour, which is governed by additional layers of biological complexity and
103 buffering, including transcriptional and translational regulation. Gene expression signatures are used to classify
104 functionally distinct cell types, and “functional genomic tests” are also gaining traction in clinical oncology [23]. This is
105 exemplified by the FDA-approved MammaPrint® 70-gene signature test which is used to aid treatment decisions in
106 early-stage breast cancers. Moreover, computational initiatives like the Broad Institute’s Molecular Signatures
107 Database Hallmark Gene Set Collection (mSigDb) have provided the community with “refined” gene sets that can
108 be used to evaluate the activity of major biological pathways based on transcriptional data [24].

109 In this study, we computed PI3K “activity” scores for breast cancers based on well-established and publicly-
110 available transcriptional signatures. The scores represent inferred PI3K activity, equivalent to a ‘record’ of past PI3K
111 pathway activation. A similar approach was used to compute transcriptional stemness scores. We next used these
112 scores to evaluate the relationship between PI3K signalling strength, stemness and dedifferentiation in two large
113 breast cancer cohorts (METABRIC and TCGA breast carcinoma). We demonstrate that a high PI3K pathway
114 activity score correlates strongly with cancer dedifferentiation/grade, stemness gene expression and reduced
115 survival. In line with previous studies [25,26], binary classification of samples according to *PIK3CA* genotype resulted
116 in a negative relationship between *PIK3CA* mutant status, PI3K signalling strength and stemness. Further
117 stratification that also took into account *PIK3CA* mutant allele dosage revealed the expected positive correlation with
118 PI3K signalling strength and stemness. Nevertheless, given the multitude of known genetic and non-genetic causes
119 of PI3K hyperactivation in cancer, including complex signalling feedback loops, we conclude that *PIK3CA* mutational
120 status alone is not sufficient to predict PI3K pathway activation and/or stemness. The implications of our findings in
121 the context of cancer therapy are discussed.

123 **RESULTS**

124 **Transcriptional PI3K pathway activity in breast cancer is associated with increased stemness and tumour**
125 **dedifferentiation**

126
127 In order to determine whether dose-dependent PI3K activation – irrespective of its genetic basis – was linked to
128 stemness in human cancer, we implemented openly available tools to calculate phenotypic scores for PI3K signalling
129 strength and stemness (see Supplementary Data for annotated source code to reproduce all of the following steps).

130 For the PI3K activity score, we used Gene Set Variation Analysis (GSVA) [27] and the
131 "HALLMARK_PI3K_AKT_MTOR_SIGNALING" gene set from the Broad Institute's Molecular Signature Database
132 (MSigDB). This gene set consists of 105 genes upregulated upon PI3K pathway activation across multiple studies
133 [24] (Supplementary Table 1). To compute a stemness score, we used the PLURINET gene signature (n = 299
134 genes), developed based on machine learning methods to facilitate robust classification of human pluripotent stem
135 cells [28] (Supplementary Table 2). Of note, only 4 genes were shared between the PI3K activity and stemness
136 gene lists, thus precluding a confounding effect on the relationship between stemness and PI3K activity scores
137 reported below.

138 We next used breast cancer transcriptomic data to assess correlations among PI3K scores, stemness scores
139 and clinical characteristics/outcomes. We used the METABRIC breast cancer dataset [29] due to its large sample
140 size and high-quality information on cancer grade, a surrogate measure of dedifferentiation and stemness. The PI3K
141 activity score in METABRIC breast tumours correlated significantly with the stemness score (**Fig. 1A**; Spearman's
142 Rho = 0.49, p < 2.2e-16) as well as tumour grade and thus dedifferentiation (**Fig. 1B**).

143 Upon stratification of METABRIC breast cancers into those with "high" and "low" PI3K pathway activity, we
144 found that around 90% of ER-negative tumours but only up to 40% of ER-positive tumours had "high" PI3K activity
145 score (**Fig. 2A**). PI3K activity and stemness scores were highest in the more aggressive PAM50 breast cancer
146 subtypes (**Fig. 2B**), including Basal, HER2 and Luminal B [30]. This contrasts with the known enrichment of *PIK3CA*
147 mutations in the ER-positive tumours, in particular the Luminal A subtype (**Fig. S1A**) (demonstrated previously
148 [26,31]). Several of these findings, including the strong relationship between inferred PI3K activity and stemness
149 scores, were reproduced using available TCGA breast cancer transcriptomic data (**Fig. S1B, S1C**).

150
151 **A "high" PI3K activity score, but not mutant *PIK3CA* status, predicts reduced survival in breast cancer**

152 Across all breast cancers, a "high" PI3K activity score was associated with reduced patient survival, with the
153 median survival probability reduced by over 3 years in patients with a high PI3K pathway activity score (**Fig. 3A**).
154 However, no such correlation was seen when patients were grouped by binary *PIK3CA* mutation status (**Fig. 1E**).
155 The prognostic value of the PI3K score remained apparent when tested in ER-positive tumours only (**Fig. S1B**),
156 suggesting it was not confounded by the higher PI3K activity score in the more aggressive ER-negative breast
157 cancer subtypes (**Fig. 2**).

158
159 **Stratification of breast cancers according to mutant *PIK3CA* allele dosage results in an unexpected**
160 **biphasic relationship with PI3K activity and stemness scores**

161 Next, using TCGA data and our previous allele copy number data for TCGA tumours [6], we tested whether
162 breast cancers with multiple copies of *PIK3CA* "hotspot" mutations exhibit higher PI3K pathway activity and
163 stemness scores than tumours with a single copy, as predicted by our iPSC findings [6,32]. In the presence of a
164 single oncogenic *PIK3CA* missense variant, we found a paradoxical reduction in transcriptional PI3K pathway activity
165 score, a relationship that was also reflected in lower stemness scores when only one copy of oncogenic *PIK3CA*
166 was present (**Fig. 4A**). In contrast, in breast cancers with multiple oncogenic *PIK3CA* copies, both PI3K and
167 stemness scores increased (**Fig. 4A**). This relationship was lost upon simple binary classification based on *PIK3CA*
168 genotypes (i.e. wild-type vs mutant) (**Fig. S1E**). Nearly identical patterns were observed in the METABRIC cohort
169 when we used publicly available allele copy number data from cBioPortal (**Fig. 4B**).

170 Taken together, these multiple analyses in large human breast cancer datasets provide strong evidence for an
171 association between strong PI3K pathway activation, breast cancer stemness and clinical outcome (visually
172 summarised in **Fig. 4C**), contrasting with the lack of prognostic value found on binary stratification based on *PIK3CA*
173 mutation status. Our findings raise the possibility that stratification of cancers in general according to transcriptional
174 indices of PI3K pathway activity may be of greater prognostic value and more useful in precision therapy than simple
175 stratification by *PIK3CA* mutation status.

176
177 **DISCUSSION**

178 *PIK3CA*^{H1047R} is the most common activating *PIK3CA* mutation in human cancers and in PROS, a group of largely
179 benign overgrowth disorders [4]. We recently found that *PIK3CA*-associated cancers often harbour multiple mutated
180 *PIK3CA* copies, and demonstrated that homozygosity but not heterozygosity for *PIK3CA*^{H1047R} leads to self-

181 sustained stemness in iPSCs [6]. Here, we use computational analyses of large human breast cancer datasets to
182 demonstrate a strong, positive relationship between the transcriptionally-inferred PI3K pathway activity, stemness
183 gene expression and tumour grade. Importantly, we show that stratification of breast tumours according to single vs
184 multiple copies of "hotspot" *PIK3CA* mutations results in subgroups with "low" and "high" PI3K activity scores,
185 respectively. This is not observed upon binary classification into *PIK3CA* mutant and *PIK3CA* wild-type cancers, due
186 to a paradoxical decrease in transcriptional PI3K pathway activity conferred by single *PIK3CA* mutations. This agrees
187 with the negative relationship between *PIK3CA* mutant status and indices of transcriptomic PI3K pathway activity
188 previously reported [25,26].

189 The apparent biphasic relationship between single versus multiple copies of *PIK3CA* mutation and stemness
190 scores warrants further study, but is likely to reflect poorly understood regulatory aspects of intracellular signalling
191 networks. We therefore caution against the use of a *PIK3CA*-mutant-centric approach to predict PI3K pathway
192 activity, given that numerous alternative genetic changes – including *PIK3CA* amplification, *PTEN* and *INPP4B* loss
193 – may converge on increased PI3K pathway activation [3,26,33,34]. These will be captured by the PI3K activity
194 signature used in our study. Indeed, the aggressive basal breast cancer subtype, despite showing a relative lack of
195 *PIK3CA* mutations, exhibits some of the highest PI3K activity and stemness scores (Fig. S1A). Although their mutual
196 relationships has not been addressed previously, indices for PI3K activity and stemness have separately been
197 associated with the basal breast cancer subtype in previous studies [11,12,26,33]. Moreover PI3K signalling was
198 recently shown to promote stem cell-like traits in basal-like breast cancers [35], consistent with the notion that the
199 aggressive nature of this cancer subtype is driven by a stem cell component [36].

200 In a separate study of iPSCs with heterozygous and homozygous *PIK3CA*^{H1047R} expression, BYL719 (alpelisib;
201 Novartis) failed to reverse the increased stemness gene expression in homozygous *PIK3CA*^{H1047R} iPSCs [32]. This
202 PI3K α -selective inhibitor was recently approved for use in combination with anti-estrogen therapy in ER-positive
203 breast cancers. In a randomised phase 3 trial that compared BYL719 with fulvestrant to fulvestrant alone, increased
204 progression-free survival was seen in 26.6 % of patients with *PIK3CA*-mutant tumours vs 12.8 % of those without a
205 *PIK3CA* variant [22]. This demonstrates the utility of *PIK3CA*-centric stratification, yet a substantial proportion of
206 patients with *PIK3CA*-mutant tumours did not benefit from the BYL719 and fulvestrant combination [22]. Although
207 recent studies demonstrated that double *PIK3CA* mutations in *cis* confer greater sensitivity to PI3K α inhibition, these
208 studies focused on inhibition of proliferation/growth [7,8]. Such parameters do not strictly correlate with
209 dedifferentiation or 'stemness' which are more strongly linked with metastasis and ultimately death. This is
210 exemplified in a recent pre-clinical study of ER-negative breast cancer models in which rapalogs effectively blunted
211 primary tumour growth but failed to reduce the number of lung metastases [37]. Moreover, rapalog resistance was
212 linked to stem cell-like features in both ER-positive and ER-negative breast cancer cell lines [37].

213 Our results raise the possibility that tumours with high PI3K activity score may respond well to combined
214 inhibition of the PI3K pathway and reversal of the stemness phenotype. A recent RNA-interference screen in breast
215 cancer cell lines revealed several potential pro-differentiation agents, including the BET bromodomain inhibitor JQ1
216 [38]. The bromodomain 9 (BRD9) subunit of the SWI-SNF chromatin-remodelling complex has also been implicated
217 as a driver of the high MYC transcriptional signature in *PIK3CA*^{H1047R}/*KRAS*^{G12V} double knock-in breast epithelial
218 cells relative to single-mutant counterparts [39]. Our data in human iPSCs suggest TGF β pathway inhibition as yet
219 another strategy in the context of strong PI3K pathway activation [32]. Assessing efficacy of such treatments will,
220 however, require monitoring of phenotypes beyond bulk tumour growth, such as stemness and metastatic potential.

221 Given a previous study which found a PI3K/AKT/mTOR gene signature score to correlate negatively with pan-
222 cancer survival [25], dual PI3K-stemness score assessment may also be prognostically useful beyond breast
223 cancer. Our computational method to infer phenotypic scores is applicable to other cancer contexts given sufficient
224 transcriptomic data (N>30 tumour samples). It is, however, important to emphasise that our approach is unlikely to
225 report instantaneous PI3K pathway activity, which is better assessed, for example, by phosphoprotein-based
226 analyses [25]. By focusing on the transcriptome, we instead infer PI3K activity over time, as recorded in wider gene
227 expression changes. We also note that a limitation of the current approach is the use of bulk transcriptomic data
228 which fail to report on the heterogeneity in single-cell gene expression within a tumour sample.

229 Finally, the proposed stratification according to PI3K pathway activity does not require expensive transcriptomic
230 studies; instead, we suggest development of a simple qPCR "scorecard" that captures the hallmark PI3K gene
231 signature and key stemness signature genes in one assay (see SI Appendix for a proposed outline of such as "PI3K-
232 stemness scorecard"), akin to multigene panels such as Oncotype Dx® and MammaPrint® for use in breast cancer
233 [23]. This proposal will require evaluation of sensitivity and specificity and long-term assessment but may ultimately
234 help the clinical translation of PI3K-based therapies, an effort which has turned out to be very challenging to date.

235

236

237

238 **MATERIALS AND METHODS**

239 **METABRIC and TCGA transcriptomic data access and pre-processing**

240
241 Normalised microarray-based gene expression for METABRIC breast tumour samples were obtained from
242 Curtis et al. [29], and clinical data from Rueda et al. [40]. The relevant METABRIC mutation data were downloaded
243 from cBioPortal in January (mutation-only) and March (mutation and copy number) 2020 [41]. TCGA breast invasive
244 carcinoma (BRCA) RNAseq, mutational and clinical data were retrieved from the GDC server (using the legacy
245 database) using the TCGAbiolinks package [42]. This package was also used for subsequent quantile filtering
246 (quantile value = 0.4) of lowly-expressed gene and removal of tumour samples with low purity (cpe = 0.6). The
247 resulting raw RSEM counts were normalised with the TMM method [43] and log2-transformed using the voom()
248 function in the limma package prior to downstream use in GSVA computations. The TCGA BRCA mutation data
249 with available copy number estimates for individual mutations were obtained from Madsen et al. [6].
250

251 **Gene signature analyses of METABRIC and TCGA breast cancer cohorts**

252
253 For a detailed workflow of all computational steps, the reader is referred to the annotated RNotebook provided
254 on the accompanying OSF project page (doi:10.17605/OSF.IO/G8RF3). All computational analyses were
255 performed using the R software. Briefly, normalised microarray-based gene expression for METABRIC breast
256 tumour samples were obtained from Curtis et al. [29], and clinical data from Rueda et al. [40]. The relevant
257 METABRIC mutation data were downloaded from cBioPortal in January (mutation-only) and March (mutation and
258 copy number) 2020 [41]. TCGA breast invasive carcinoma (BRCA) RNAseq, mutational and clinical data were
259 retrieved from the GDC server (using the legacy database) using the TCGAbiolinks package [42]. This package was
260 also used for subsequent quantile filtering (quantile value = 0.4) of lowly-expressed gene and removal of tumour
261 samples with low purity (cpe = 0.6). The resulting raw RSEM counts were normalised with the TMM method [43]
262 and log2-transformed using the voom() function in the limma package prior to downstream use in GSVA
263 computations. The TCGA BRCA mutation data with available copy number estimates for individual mutations were
264 obtained from Madsen et al. [6].

265 The “HALLMARK_PI3K_AKT_MTOR_SIGNALING” (PI3K pathway activity) and “MUELLER_PLURINET”
266 (“stemness”) signatures were retrieved from The Molecular Signature Database (MSigDB). We note that the
267 “HALLMARK_PI3K_AKT_MTOR_SIGNALING” gene set also includes mTORC1-dependent gene expression
268 changes, in contrast to other studies which have sought to separate AKT- and mTORC1-driven gene expression
269 changes [44,45]. Individual signature scores were computed with the GSVA package, using the default Gaussian
270 kernel and selecting ESdiff enrichment values as output [27]. The correlation between PI3K pathway activity and
271 “stemness” scores in the two breast cancer cohorts was computed using Spearman’s rank correlation. Binary
272 classification of PI3K pathway activity scores into “low” and “high” used the 0.25 and 0.75 score quantiles,
273 respectively. The scores were next assessed in the context of available clinical and/or genetic attributes (*PIK3CA*
274 missense mutations only). *PIK3CA* “hotspot” mutations (C420R, E542K, E545K, H1047L, H1047R) were defined
275 according to their prevalence in cancer and evidence for severe functional impact in developmental overgrowth
276 disorders [4,46].

277 **Statistical analyses**

278
279 The lm() function in R was used to fit linear models to METABRIC tumour grade and gene signature score
280 data; the F statistic of each model and the accompanying R2 values are reported within the respective plots. Of note,
281 the normality assumption was violated for the stemness versus grade linear model; however, as the sample size is
282 large and as the assumption terms in the specified models are formed as a sum of several other independent
283 quantities, this violation is expected to have a minimal impact on the reported results. Tukey’s Honest Significant
284 Differences method was used to test for statistically significant (adjusted p-value < 0.05) differences in score means
285 across different tumour grades or stages. Wilcoxon pairwise-comparison with Benjamini-Hochberg correction was
286 used to assess differences in PI3K and stemness scores across tumours stratified according to *PIK3CA* copy
287 number status. The relationship between PI3K pathway activity score and survival in METABRIC was assessed
288 using a non-parametric log-rank test. Differences in score distributions across tumour subtypes and/or genotypes
289 were assessed using a Chi-squared goodness-of-fit test.
290

291 **Data and materials availability**

292
293 Raw data and bespoke RNotebooks containing guided scripts and plots are available via the Open Science
294 Framework (doi: 10.17605/OSF.IO/G8RF3). Individual scripts include information on the name and version of

295 applied R packages. Further information requests should be directed to and will be fulfilled by the corresponding
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308

309 **Competing interests**

310
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314

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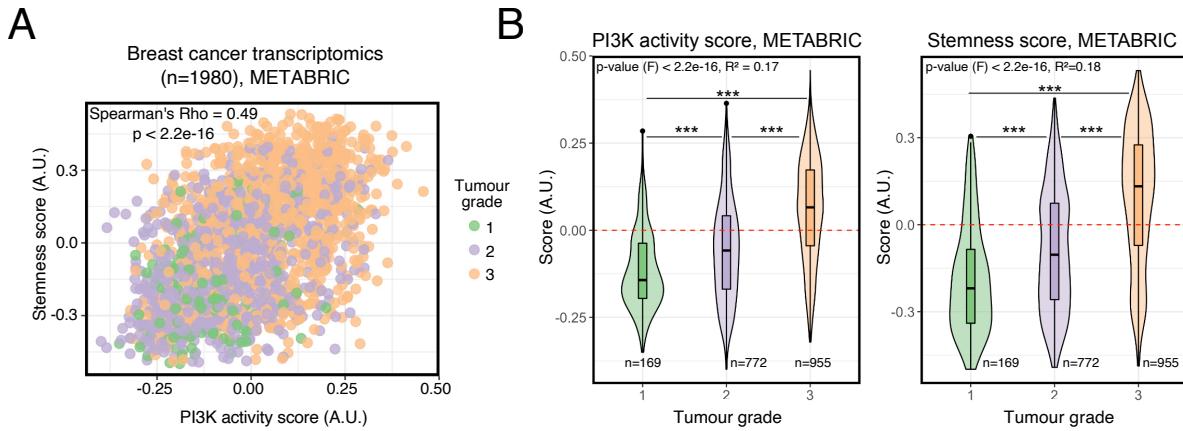
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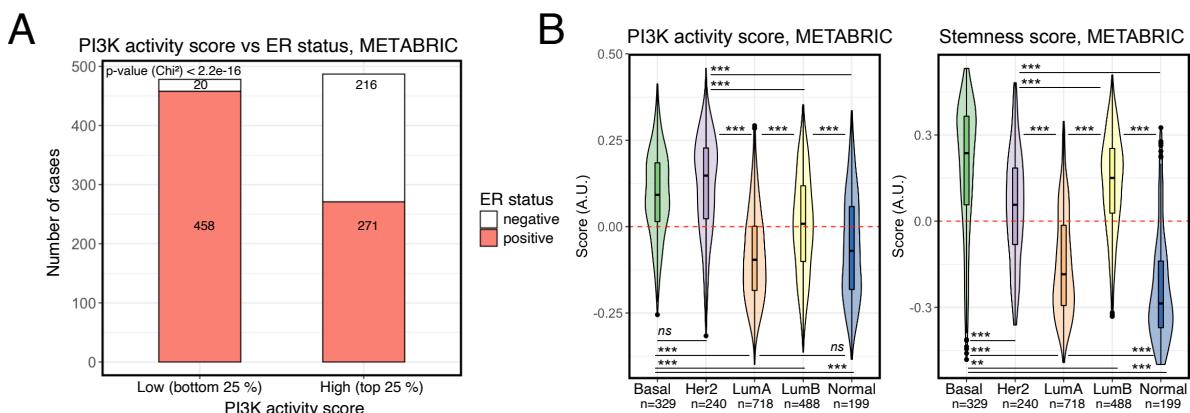
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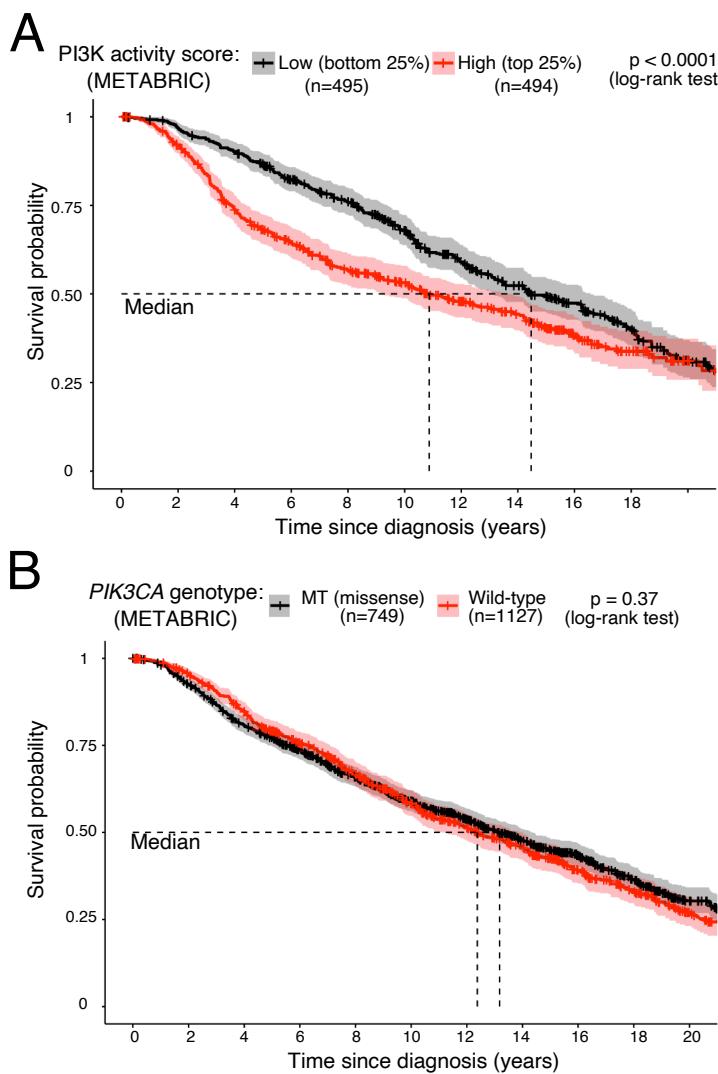
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415 **Fig. 1. Inferred PI3K pathway activation correlates with increased stemness and breast tumour**
416 **dedifferentiation.** (A) Correlation plot of PI3K activity and stemness scores, based on METABRIC breast cancer
417 transcriptomes. The scores were determined using Gene Set Variation Analysis (GSVA) with mSigDb
418 “HALLMARK_PI3K_AKT_MTOR_SIGNALING” and “MUELLER_PLURINET” gene signatures [24,27,28]. The
419 gene lists used are included in Supplementary Tables 1 and 2. (B) PI3K activity and stemness score distributions
420 across breast cancer grade (METABRIC). *** p ≤ 0.001 according to Tukey’s Honest Significant Differences
421 method. The global p-value for each linear model is indicated within each plot. See also Fig. S1A, B.
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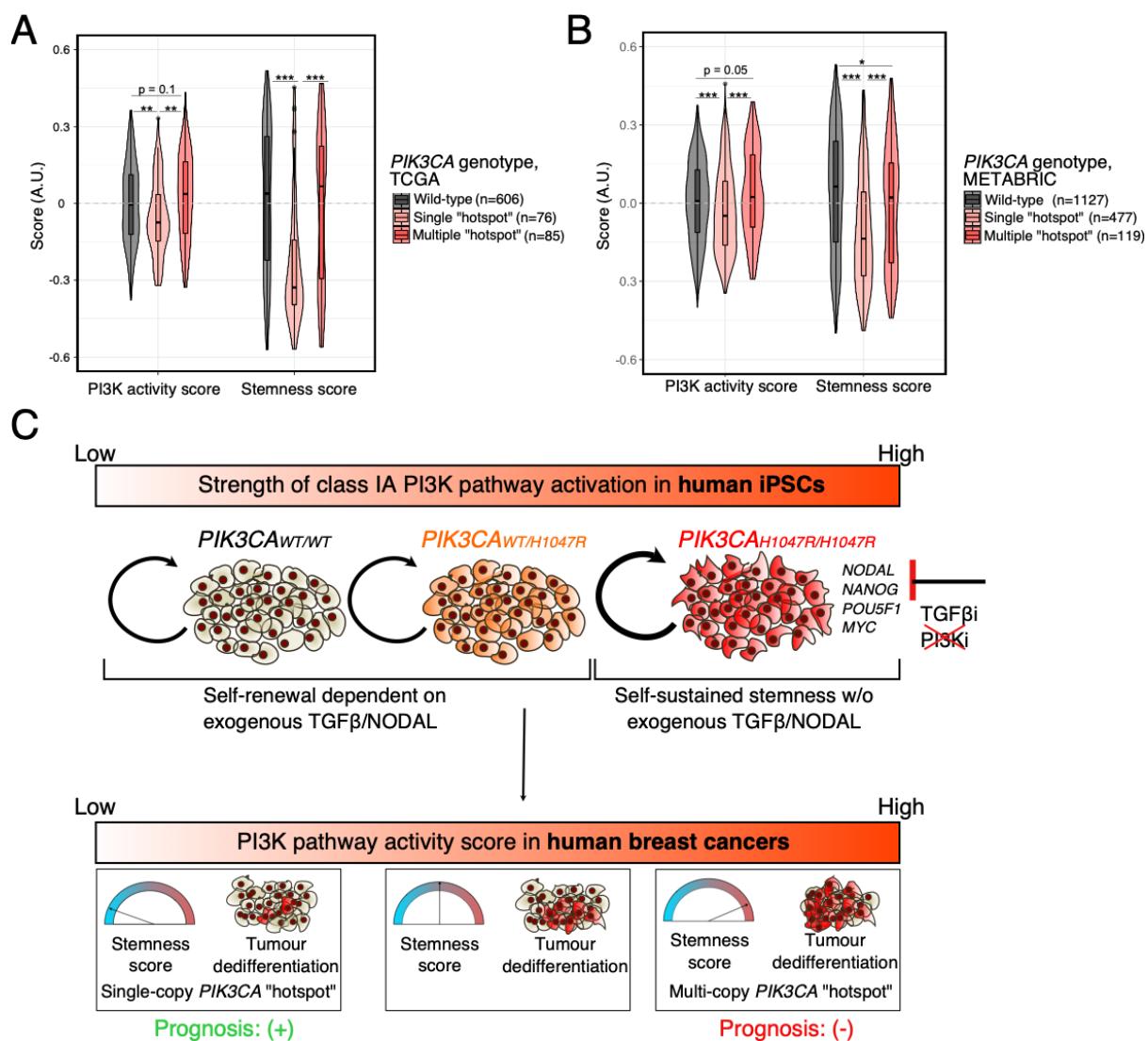


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425 **Fig. 2. High PI3K activity and stemness scores are enriched for in aggressive breast cancer subtypes. (A)**
426 PI3K activity score distribution in METABRIC breast tumours stratified according to ER status. **(B)** PI3K activity and
427 stemness score distributions across METABRIC breast cancers stratified according to PAM50 subtype; ** p ≤ 0.01,
428 *** p ≤ 0.001 according to Tukey's Honest Significant Differences method; ns: non-significant. See also Fig.
429 S1C.
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Fig. 3. PI3K activity score, but not *PIK3CA* genotype, is prognostic in breast cancer. (A) Kaplan-Meier plot of overall survival of patients stratified according to PI3K activity score (all METABRIC breast cancers). **(B)** Kaplan-Meier plot of overall survival of patients stratified according to the presence/absence of *PIK3CA* missense mutations (all METABRIC breast cancers). See also Fig. S1D.



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440 **Fig. 4. A biphasic relationship between mutant *PIK3CA* allele dosage and PI3K activity / stemness scores**
441 (A) PI3K activity and stemness score distributions across TCGA breast cancers following stratification according to
442 the presence or absence of single vs multiple copies of the *PIK3CA* "hotspot" alleles (C420R, E542K, E545K,
443 H1047L, H1047R); * p < 0.05, ** p < 0.01, *** p < 0.001 according to Wilcoxon pairwise-comparison with Benjamini-
444 Hochberg correction. (B) As in (A) but performed using METABRIC breast cancer data. (C) Graphical summary of
445 key conclusions. The current study builds on previous findings of increased stemness in human induced pluripotent
446 stem cells (iPSCs) with homozygous but not heterozygous *PIK3CA*^{H1047R} expression [6]. We extend these findings
447 to breast cancer and demonstrate that a high transcriptomic PI3K pathway activity score associates strongly with
448 high stemness score, tumour dedifferentiation and reduced survival.