

1 **Transparency in butterflies and moths: structural diversity, optical properties and**  
2 **ecological relevance**

3

4 **Authors :**

5 D. Gomez<sup>1</sup>, C. Pinna<sup>2</sup>, J. Pairraire<sup>3</sup>, M. Arias<sup>1,2</sup>, J. Barbut<sup>2</sup>, A. Pomerantz<sup>4,5</sup>, C. Noûs<sup>6</sup>, W. Daney de  
6 Marcillac<sup>3</sup>, S. Berthier<sup>3</sup>, N. Patel<sup>4</sup>, C. Andraud<sup>7</sup>, M. Elias<sup>2</sup>

7

8 **Corresponding author:** [doris.gomez@cefe.cnrs.fr](mailto:doris.gomez@cefe.cnrs.fr)

9

10 **Affiliations:**

11 <sup>1</sup> CEFÉ, CNRS, University of Montpellier, University of Paul Valéry Montpellier 3, EPHE, IRD,  
12 Montpellier, France

13 <sup>2</sup> ISYEB, UMR 7205, CNRS, MNHN, Sorbonne University, EPHE, France

14 <sup>3</sup> INSP, Sorbonne University, CNRS, Paris, France

15 <sup>4</sup> Marine Biological Laboratory, Woods Hole, Massachusetts, USA

16 <sup>5</sup> Department Integrative Biology, University of California Berkeley, Berkeley, USA

17 <sup>6</sup> Cogitamus Laboratory, France

18 <sup>7</sup> CRC, MNHN, Paris, France

19

20

21 **ABSTRACT:**

22 In water, transparency seems an ideal concealment strategy, as testified by the variety of transparent  
23 aquatic organisms. By contrast, transparency is nearly absent on land, with the exception of insect  
24 wings, and knowledge is scarce about its functions and evolution, with fragmentary studies and no  
25 comparative perspective. Lepidoptera (butterflies and moths) represent an outstanding group to  
26 investigate transparency on land, as species typically harbour opaque wings covered with coloured

27 scales, a key multifunctional innovation. Yet, many Lepidoptera species have evolved partially or  
28 fully transparent wings. At the interface between physics and biology, the present study investigates  
29 transparency in 123 Lepidopteran species (from 31 families) for its structural basis, optical  
30 properties and biological relevance in relation to thermoregulation and vision. Our results establish  
31 that transparency has likely evolved multiple times independently. Efficiency at transmitting light  
32 is largely determined by clearwing microstructure (scale shape, insertion, colouration, dimensions  
33 and density) and macrostructure (clearwing area, species size or wing area). Microstructural traits –  
34 density, dimensions – are tightly linked in their evolution, with different constraints according to  
35 scale shape, insertion, and colouration. Transparency appears highly relevant for vision, especially  
36 for camouflage, with size-dependent and activity-rhythm dependent variations. Links between  
37 transparency and latitude are consistent with an ecological relevance of transparency in  
38 thermoregulation, and not so for protection against UV radiation. Altogether, our results shed new  
39 light on the physical and ecological processes driving the evolution of transparency on land and  
40 underline that transparency is a more complex than previously thought colouration strategy.

41

42 **KEYWORDS:** transparency, Lepidoptera, microstructure, structural strategy, vision,  
43 thermoregulation, UV protection

44 **INTRODUCTION**

45

46 Following the invisibility myth, transparency seems an ideal camouflage strategy: being 'hidden in  
47 plain sight', invisible to go undetected by predators works whatever the background, from all  
48 viewpoints and irrespective of behaviour (Cuthill 2019). The 'success story' of transparency in  
49 water as a protection against predators (especially in pelagic habitats where there is nowhere to  
50 hide) is attested by its broad phylogenetic distribution since transparency spans 7 phyla including  
51 Arthropoda, Mollusca, Annelida, Chordata and Cnidaria (Johnsen 2001). By contrast, transparency  
52 is nearly absent on land, and almost confined to insect wings. This contrast can be explained by  
53 physical factors: compared to water, larger refractive index mismatch between air and biological  
54 tissues produces higher light reflection that ruins invisibility (Johnsen 2001). In addition, greater  
55 ultraviolet (UV) radiation on land imposes greater UV protection often through light absorption  
56 by pigments.

57 Research in transparent aquatic organisms has shown a role for concealment from visually-  
58 hunting predators, which have developed special sensorial abilities that break this camouflage (e.g.  
59 Tuthill and Johnsen 2006). As underlined by Johnsen (2014), many questions are left unanswered  
60 about transparency, like the structural bases of transparency (Bagge 2019), the functional roles of  
61 transparency in vital functions like thermoregulation and potential trade-offs with optics, and the  
62 selective pressures driving its evolution and its design. Comparative studies at broad interspecific  
63 level are absent but crucial to better understand the diversity and evolution of structures underlying  
64 transparency, the adaptive functions and evolution of this fascinating trait. The lack of studies is  
65 even more crucial for transparency on land where knowledge is scarce, with fragmentary  
66 monographic studies by physicists using bioinspired approaches, based on transparent wing  
67 antireflective, hydrophobic and antifouling properties (e.g. Deparis et al. 2014; Liu et al. 2016;  
68 Elbourne et al. 2017).

69 Within insects, Lepidoptera represent an outstanding group to explore these questions.  
70 Insect wings are made of chitin. While most insects harbour transparent wings, Lepidoptera are  
71 typically characterized by wings covered with scales. Scales are chitin extensions that are often long  
72 and large and that often contain a pigment or structures that interact with light, thereby producing  
73 opaque colour patterns (e.g. Stavenga et al. 2014). Wings covered by scales represent an  
74 evolutionary innovation involved in many functions such as antipredator defences (camouflage,  
75 deflection, aposematism... e.g. Stevens et al. 2008), communication (Kemp 2007), thermoregulation  
76 (Miaoulis and Heilman 1998; Berthier 2005; Krishna et al. 2020), or water repellency (Wagner et al.  
77 1996; Wanasekara and Chalivendra 2011). In this opaque world, many species from different  
78 families have evolved partially or totally transparent wings. The handful of existing studies in  
79 butterflies in physics or biology suggests that an important structural diversity may underlie the  
80 occurrence of transparency in many Lepidopteran lineages. The wing membrane may be nude or  
81 covered with scales, which can be of various morphologies, insertion angle on the wing membrane,  
82 and colouration (Yoshida et al. 1997; Hernandez-Chavarria et al. 2004; Berthier 2007; Goodwyn et  
83 al. 2009; Wanasekara and Chalivendra 2011; Stavenga et al. 2012; Siddique et al. 2015).

84 Lepidoptera therefore represent an evolutionary laboratory where we can examine the  
85 phylogenetic extent of transparency; the diversity and evolution of optical properties and of the  
86 underlying structures; the existence, if any, of structural constraints on transparency; and the  
87 ecological relevance of transparency in Lepidoptera. More specifically, we can ask whether, as  
88 suggested from theoretical and empirical studies on aquatic organisms, there are several structural  
89 and optical routes to transparency (Johnsen 2001) and whether there exist some structural  
90 constraints at the macroscopic and microscopic scale.

91 We can question the ecological relevance of transparency – as an optical property – for  
92 vision and camouflage. The prominent role of transparency in camouflage shown so far (Johnsen  
93 2014; McClure et al. 2019; Arias, Mappes, et al. 2019; Arias, Elias, et al. 2019) suggests that visually-  
94 hunting predators may be important selective pressures on the evolution of transparency on land,

95 too. The issue of minimizing light reflection may be even more important for diurnal species active  
96 during daytime and exposed to sun beams in many directions, than for nocturnal species less mobile  
97 during daytime.

98 We can also question the ecological relevance of transparency for thermoregulation. The  
99 thermal melanism hypothesis states that individuals from colder places, as those in higher latitudes  
100 or altitudes, can gain extra thermal benefit from being more strongly pigmented as radiation  
101 absorption helps thermoregulation in these ectotherms (Bogert 1949). This hypothesis has received  
102 support from comparative analyses at large taxonomical and geographical scales (e.g. Zeuss et al.  
103 2014; Xing et al. 2018; Stelbrink et al. 2019) as well as from analyses at species level (e.g. Colias  
104 pierids in Ellers and Boggs 2004) showing that individuals gain thermal benefits from melanisation  
105 of their proximal wing close to the body. Recent large-scale comparative analyses in opaque  
106 butterflies have shown that body and proximal wing colouration correlates to climate in the near-  
107 infrared [700-1100] nm range but not so below 700 nm where vision occurs (Munro et al. 2019).  
108 Moreover, nocturnal species harbour darker colouration than diurnal species, especially at low  
109 elevations (Xing et al. 2018), a result that is coherent with a role of coloration in thermoregulation.

110 We can finally question the ecological relevance of transparency for UV protection.  
111 Exposure to highly energetic and penetrating UVB [280-315] nm and UVA [315-400] nm radiation  
112 has detrimental effects on physiology, fecundity and survival in terrestrial living organisms (e.g. in  
113 insects Zhang et al. 2011). Levels of UV radiation are higher at low latitudes than at higher latitudes  
114 (Beckmann et al. 2014). Hence, absorption of UV radiation should be more important at low  
115 latitudes.

116 Using a large dataset comprising 123 clearwing Lepidoptera species, we examine the  
117 phylogenetic distribution of transparency, the influence of wing macrostructure (wing size, area,  
118 clearwing area, and proportion of clearwing area) and microstructure (presence of scale, type,  
119 insertion and colouration) on optical properties. We also assess to which extent structural features  
120 are conserved across the phylogeny, and whether some of these features present correlated

121 evolution, which can help us reveal evolutionary constraints. We then examine the potential  
122 relevance of transparency not only as a physical property, but most importantly in biology, in  
123 relation to camouflage and thermoregulation. More specifically, we test whether, if transparency is  
124 involved in camouflage, diurnal species, more exposed to visual predators, transmit more light  
125 through their wings than nocturnal species. We finally test whether, if transparency is involved in  
126 thermoregulation or in UV protection. In optics theory, the light received by an object can be either  
127 transmitted, reflected or absorbed. Variations in light transmission can indicate variations in  
128 absorption if reflection levels are maintained at similar levels. We posit that species living at  
129 increasing distance from equator should transmit less light through their wings (at least in the near-  
130 infrared range) if transparency plays a role in thermoregulation, but more light (clearwings should  
131 transmit more or at least proportionally more in the ultraviolet range) if transparency plays a role  
132 in UV protection.

133 The present study thus addresses for the first time the links between structure and optics  
134 at such a broad phylogenetic scale to understand the ‘small success story of transparency on land’,  
135 its evolution and putative functions in relation to camouflage and thermoregulation.

136

137

## 138 **METHODS**

### 139 **Specimens and ecological data**

140 We looked for clearwing species in the Lepidoptera collection of the French Museum of Natural  
141 History, based on our own experience, on the literature, on the knowledge of museum curators  
142 and researchers, on species names (*vitrata*, *fenestrata*, *hyalina*, *diaphanis*...), and on a systematic search  
143 for small families. We found clearwing species (transparent or translucent but called transparent in  
144 the literature) in 31 out of the 124 existing Lepidoptera families (Supplementary Table S1) and  
145 gathered a total of 123 species. We took 1 specimen per species (see Figure 1 for some examples).  
146 Those specimens were often unique or precious, which prevented us from conducting destructive

147 measurements. There were 77 specimens for which labels specified exact collect location that could  
148 be tracked down to GPS coordinates. We obtained data on diurnality / nocturnality for 114 species:  
149 59 were diurnal, 51 were nocturnal and 4 were active at day and night.

150

151 **Structure measurements**

152 Museum specimens were photographed using a D800E Nikon camera equipped with a 60mm lens,  
153 placed on a stand with an annular light. Photos were then analysed using ImageJ (Schneider et al.  
154 2012) to extract descriptors of wing macrostructure: wing length (mm), wing surface (mm<sup>2</sup>), and  
155 clearwing area (the surface of transparent area in mm<sup>2</sup>), for the forewing and hindwing separately.  
156 We defined and computed proportion of clearwing area as the ratio clearwing area/wing surface,  
157 i.e. the proportion of the total wing area occupied by transparency.

158 Museum specimens were also photographed with a powerful binocular (Zeiss Stereo  
159 Discovery V20) and with a photonic digital microscope (Keyence VHX-5000) to get close images  
160 of the dorsal side of transparent and opaque zones of each wing. These images were analysed using  
161 the built-in measurement software to describe wing microstructure. Because of the diversity in scale  
162 shapes, here we define the word *phanera* as the generic term encompassing morphological types of  
163 scales. Coining this term allows us to distinguish scale as the epicene word that encompasses a large  
164 diversity of morphological types (phanera here) and scale as a subcategory of those types (scale  
165 here). We measured phanera density, length (μm), width (μm) and we computed phanera surface  
166 as the product of length by width, and phanera coverage as the product of surface by density.

167 As shown from examples in Figure 1, the transparent zone showed diversity in  
168 morphological type (hereafter referred to as type), insertion, and coloration:

169 - in phanera morphological type (hereafter referred to as type): no phanera (N), bifid  
170 or monofid hair-like scales (H), scales (any other shape than hair-like scales) (S),  
171 scales and hair-like scales (HS);

172 - in phanera insertion on the membrane: erected (E) or flat (F) phanera, or unknown  
173 insertion (U, when phanera were absent);  
174 - in phanera colouration: coloured (C) or transparent (T, when phanera were partly  
175 or totally transparent), or unknown coloration (U, when phanera were absent).

176 We define as *structural strategy* the combination of the following traits: type of phanera, insertion and  
177 colouration. For instance, SEC is a structural strategy with erected coloured scales. For clarity  
178 reasons, the no phanera strategy (that should be called NUU) was hereafter referred to as the N  
179 strategy.

180 For most analyses (except for sample size counting and mean-pairwise distance analyses,  
181 see below), we assimilated the combined scales and hair-like scales (HS) to scales (S) of their  
182 corresponding insertion and colouration, for several reasons: (i) the exploration of structural  
183 changes in phanera length and width could only address one type of phanera, not two, (ii) strategies  
184 involving the combination of scales and hair-like scales were rare and effects could not be easily  
185 tracked. (iii) In HSF strategies (HSFC and HSFT), both scales and hair-like scales were in similar  
186 density and in HSE strategies (HSEC and HSET), scales were in greater density than hair-like  
187 scales, which suggested that scales played a similar role or a greater role than hair-like scales for  
188 aspects linked to phanera density.

189

## 190 **Optical measurements**

191 We measured specular transmittance from 300 to 1100 nm, using a deuterium-halogen lamp  
192 (Avalight DHS, Avantes). Wing samples were illuminated using an optic fibre (FC-UV200-2-1.5 x  
193 100, Avantes). We collected the transmitted light using a similar optic fibre connected to the  
194 spectrometer (Starline Avaspec-2048 L, Avantes). Fibres were aligned 5mm apart and the wing  
195 sample was placed perpendicular between them at equal distance, guaranteeing that we measured  
196 specular and not diffuse transmittance. The incident light beam made a 1mm diameter spot. Spectra

197 were taken relative to a dark (light off) and to a white reference (no sample between the fibres)  
198 measurement. For each species and wing, we took five measurements.

199 We analysed spectral shape using Avicol v6 (Gomez 2011) and the ‘pavo’ R package (Maia  
200 et al. 2019) to extract physically and biologically relevant parameters. We computed the mean  
201 transmittance over [300-700] nm, which described the level of transparency. In addition, we  
202 computed the proportion of UV transmittance as the ratio (total transmittance over [300-400]  
203 nm/total transmittance over [300-700] nm), i.e. the proportion of the total amount of transmitted  
204 light that occurred in the ultraviolet range. Wings and wing phanera are made of chitin; chitin  
205 absorption, negligible above 500 nm increases as wavelength decreases, especially in the ultraviolet  
206 range (Azofeifa et al. 2012; Stavenga et al. 2014). Given that absorption + reflection + transmission  
207 = 1, an increase in absorption causes a loss in transmission if reflection is maintained at similar  
208 levels. Separating this wavelength range allows us to assess the potential role of transparency as a  
209 parasol against UV radiation.

210 Following the method implemented by Munro et al. (2019) in their recent study on  
211 thermoregulation in opaque butterflies, we extracted the mean transmittance values separately for  
212 the ultraviolet range [300-400] nm, the human-visible range [400-700] nm and the near infrared  
213 range [700-1100] nm. Separating these ranges allows us to disentangle the near-infrared range where  
214 only thermoregulation can act as a selective pressure from shorter wavelengths where vision can  
215 also operate and drive colour design.

216 We also analysed spectra in vision modelling using Vorobyev and Osorio’s discriminability  
217 model (Vorobyev and Osorio 1998). Given that we were mainly interested in testing whether  
218 optical transparency was transferred into biologically meaningful transparency, we took the blue tit  
219 UVS vision as an example of insectivorous bird vision. We used the spectral data from the blue tit  
220 (*Cyanistes caeruleus*) and relative cone densities of 1:1.9:2.7:2.7 for UVS:S:M:L (Hart et al. 2000), we  
221 assumed a Weber fraction of 0.1 for chromatic vision (Maier and Bowmaker 1993; Lind et al. 2014)  
222 and 0.2 for achromatic vision (average of the two species studied in Lind et al. 2013). We assumed

223 that the incident light went through the wing to the eye of the bird, which was viewing the butterfly  
224 against an average green vegetation forest background, both illuminated by a large gap light (light  
225 spectra and vegetation taken from Gomez and Théry 2007). We obtained colour and brightness  
226 contrast values.

227

## 228 **Phylogeny reconstruction**

229 We built a phylogeny comprising 183 species representing the 31 families included in our dataset,  
230 as follows. First, for each of the 123 clearwing species in our dataset, we searched for DNA  
231 sequences in GenBank and BOLD (Ratnasingham and Hebert 2007), and if none was available, we  
232 took a species from the same genus, tribe, subfamily or family as a substitute (Supplementary File  
233 1 for the list of tree species and surrogate clearwing species). Second, we incorporated 60 additional  
234 species from families where species sampling was low to consolidate tree topology. We used DNA  
235 sequences for the mitochondrial CO1 and CO2 genes, and for the nuclear CAD, EF1, GADPH,  
236 IDH, MDH, RpS5, and WG genes (supplementary Table S1). We aligned the sequences with  
237 CodonCodeAligner (version 4.2.7, CodonCode Corporation, <http://www.codoncode.com/>) and  
238 concatenated them with PhyUtility (version 2.2, Smith and Dunn 2008). The dataset was then  
239 partitioned by gene and codon positions and the best models of substitution were selected over all  
240 models implemented in BEAST 1.8.3 (Drummond and Rambaut 2007), using the ‘greedy’  
241 algorithm and linked rates implemented in Partition Finder 2.1.1 (Lanfear et al. 2017, best scheme  
242 in Supplementary File 1). We constrained the topology of all families to follow Figs12 from Regier  
243 et al. (2013), and we used the following secondary calibrations from Regier et al. (2013): node  
244 joining Bombycoidea and Lasiocampoidea at 84.05 My [74.15;94.4]), Noctuidea ancestor at 77.6  
245 My [66.97;88.57]), node joining Gelechioidea to Bombycoidea at 105.23 My [93.77; 117.3]),  
246 Papilioidea ancestor at 98.34 My [86.85;110.33]); node joining Sesioidea to Coccoidea at 145.03  
247 My [93.4;118.32]), and tree root at 145.03 My [128.96;161.64]) on Cipres Science Gateway (Miller  
248 et al. 2010). We constrained monophyly from genus to family level. Four independent analyses

249 were run for 10 million generations, with one Monte Carlo Markov Chain each and a sampling  
250 frequency of one out of 10,000 generations. We examined the trace of each run, and defined a  
251 burn-in period for each run independently, using Tracer 1.6 (<http://beast.bio.ed.ac.uk/tracer>). We  
252 retained only the 2 runs that had a stable trace and combined the trees using LogCombiner 1.8.4  
253 (<http://beast.bio.ed.ac.uk>). We then computed the maximum clade credibility (MCC) tree with  
254 median node ages using TreeAnnotator 1.8.4 (Drummond et al. 2012). Additional species were  
255 then pruned from the tree and we used the resulting MCC tree in subsequent comparative analyses.

256

## 257 **Statistical analyses**

258 All analyses were conducted using the R environment (R Development Core Team 2013).

259

260 Repeatability analysis: we assessed the repeatability of colour and structural parameters measured,  
261 using the 'rptR' package (Stoffel et al. 2017). For transparency measurements, we took the 5  
262 measurements as repetitions of the same species and wing. For wing length, we measured twice  
263 each wing and thus obtained 2 repetitions of the same wing for each species. Regarding the  
264 repeatability of the measurements of phanera density, length, and width, we measured a small  
265 number of phanera of the same type, zone, and wing, and tested whether within-group variability  
266 was lower than between-group variability. All measurements were found highly repeatable (see  
267 sample sizes and results in Table S2).

268

269 Phylogenetic signal: We implemented two complementary approaches. First, we estimated the  
270 amount of phylogenetic signal in each structural and optical variable. For continuous variables, we  
271 used both Pagel's lambda (Pagel 1999) and Blomberg's K (Blomberg et al. 2003) implemented in  
272 the 'phytools' R package (Revell 2012). For binary variables, we used Fritz and Purvis' D (Fritz and  
273 Purvis 2010) implemented in the 'caper' R package (Orme et al. 2018). Second, we assessed to

274 which extent structural features and structural strategies were conserved on the phylogeny, in other  
275 words we estimated the degree of phenotypic clustering for structures.

276 We calculated the mean pairwise phylogenetic distances (MPD) for each categorical  
277 structural parameter (Webb et al. 2002), using the ‘picante’ R package (Kembel et al. 2010). MPD  
278 measures the average phylogenetic distance that separates two species sharing a specific trait state.  
279 We computed MPD mean pairwise distance (Webb 2000) for each wing separately, or for both  
280 wings (in that case we considered all the species that presented the trait on at least one wing). For  
281 a specific trait, we first computed the observed MPD, and then simulated MPD distribution by  
282 randomly shuffling trait values on the phylogeny. We then determined whether the observed value  
283 was below the 5% lower quantile of the distribution of simulated MPD values, in which case we  
284 concluded that the trait was found in species separated by fewer nodes than expected by chance.

285

286 Correlated evolution between structural traits To assess whether phanera presence, type, insertion  
287 and colour evolved in a correlated fashion both within and across transparent and opaque zones,  
288 we computed Pagel’s discrete model for those binary traits and compared the likelihood values  
289 from the dependent and independent models, using the FitPagel function, with ARD model and  
290 (x,y) structure, from ‘phytools’ (Revell 2012).

291

292 Structural constraints. We aimed to investigate the variations of the proportion of clearwing area  
293 and the structural effort in producing transparency in clearwing Lepidoptera. For this purpose, we  
294 conducted (i) mixed models using the ‘nlme’ R package (Pinheiro et al. 2020) and (ii) Bayesian  
295 phylogenetic mixed models with Markov Chain Monte Carlo analyses using the ‘mulTree’ R  
296 package (Guillerme and Healy 2019). Both approaches are suited to repeated observations but,  
297 unlike the former, the latter controls for phylogenetic relatedness. Using both allowed us to assess  
298 the influence, if any, of phylogeny on the observed relationships. (i) In the mixed model approach,  
299 we selected the best model based on the factors supposed to play a role and based on AICc

300 minimization. (ii) In the Bayesian phylogenetic approach, we used the model formulated in the  
301 classic approach; uninformative priors were used, with variance and belief parameter set to 1 and  
302 0.002 respectively for both random effect and residual variances (Hadfield 2010). We took species  
303 as random effects. Models were run using two chains of 500,000 iterations with a burn-in of 10,000  
304 and thinning interval of 300. Fixed effects were considered statistically significant when the  
305 probabilities in the 95% credible intervals did not include zero.

306 To investigate the variations of the proportion of clearwing area, we took phanera length,  
307 insertion and colouration, as well as wing length, clearwing area and wing as factors, with  
308 meaningful interactions. To explore the structural effort in producing transparency, we computed  
309 the change in the microstructural trait (phanera density, length, width, surface or coverage) as the  
310 difference (trait in the transparent zone – trait in the opaque zone) of the same wing and species.  
311 Values departing more from zero indicate greater changes, towards a decrease (<0) or an increase  
312 (>0) in the structural parameter in the transparent zone relative to the opaque zone. We considered  
313 that structural effort increased as values departed more from zero, whatever the direction.

314 For each structural trait, we took the 2 structural effort values per species (one per wing)  
315 obtained from image analysis as observations and species as random effect. We took the structural  
316 effort in a specific trait as the dependent variable. We included as fixed effects phanera surface,  
317 density, category, insertion and colour in the transparent zone, the surface of phanera in the opaque  
318 zone, as well as wing length, clearwing area, and proportion of clearwing area, and relevant  
319 interactions between these factors.

320

321 Structure-optics relationships. We aimed to identify which structural parameters influenced optical  
322 properties and to test whether the relationships between structural parameters may explain the  
323 diversity in optical properties in clearwing Lepidoptera. For this purpose, we took the 10 spectral  
324 measurements per species as observations and species and wing within species as random effects.  
325 We took the mean transmittance over [300-700] nm or the proportion of UV transmittance as the

326 dependent variable. We included as fixed effects phanera surface, density, category, insertion and  
327 colour, wing length, clearwing area, and proportion of clearwing area and relevant interactions  
328 between these factors.

329

330 Ecological relevance. We conducted both mixed models and Bayesian phylogenetic mixed models  
331 to explore the ecological relevance of transparency for vision and thermoregulation. We took the  
332 10 spectral measurements per species as observations and species and wing within species as  
333 random effects.

334 First, we examine whether optical properties translated into perceptual transparency. We  
335 took the physical property (mean transmittance over [300-700] nm or proportion of UV  
336 transmittance) as the dependent variable. We included as fixed effects variables computed from  
337 vision modelling (brightness contrast and colour contrast) but not the interaction between these  
338 factors as we had no expectation.

339 Second, we explored the link between transparency and nocturnality. We took mean  
340 transmittance over [300-700] nm as the dependent variable. We included as fixed effects wing, wing  
341 length, and the proportion of clearwing area as potential fixed factors.

342 Third, we explored the link between transparency and habitat latitude. Munro et al. (2019)  
343 have shown that body and proximal wing colouration vary with latitude in butterflies, more strongly  
344 so in smaller species, and more strongly so in the near-infrared range. We took the mean  
345 transmittance over specific ranges of wavelengths (UV, human-visible, and near-infrared) as the  
346 dependent variable and we included latitude (absolute latitudinal distance to the equator),  
347 nocturnality, wing length and their interaction as fixed factors. We included nocturnality as this  
348 may play a role as suggested in opaque butterflies (Xing et al. 2018)

349 For the latter two hypotheses, we had no specific expectation concerning the proportion  
350 of UV transmittance; we thus did not test that variable.

351

352 **RESULTS**

353 **Diversity of microstructures**

354 Structural investigation showed a high diversity of structural strategies (gathering phanera type,  
355 insertion and colouration), as shown by a few examples (Figure 1 and associated spectra in Figure  
356 2). Transparency could be achieved by the means of a nude wing membrane (N), of hair-like scales  
357 (H), of scales (S), or of scales and hair-like scales in combination (HS). When present, phanera  
358 could be flat (F) or erected (E) on the wing membrane and coloured (C) or transparent (T) (Figure  
359 3). Rather counterintuitively, scales (S) are by far the most common structural type to achieve  
360 transparency (70/123 species, 27/31 families), followed by the absence of phanera (N, 32/123  
361 species, 12/31 families) and hair-like scales alone (H, 27/123 species and 9/31 families, Table S3,  
362 Figure S1). Rarer strategies involved either transparent erected scales (SET, 9/123 species, 6/31  
363 families), the combination of scales and hair-like scales (HS, 12/123 species, 7/31 families), with  
364 the combination of erected hair-like scales and scales being the rarest (HSE, 5/123 species, 3/31  
365 families). There was no species with transparent hair-like scales alone, be they erected or flat.  
366 Transparent hair-like scales when existing were always associated to scales (HS).

367

368 **Evolution of microstructural features**

369 Phylogenetic signal. We examined to which extent structural traits were influenced by common  
370 ancestry in their evolution. More specifically, wing macrostructure (i. e., wing dimension and  
371 clearwing area) and microstructure (i. e., phanera dimensions and density) – showed significant  
372 phylogenetic signal in both clearwing and opaque zones, for both forewing and hindwing (except  
373 in one case, Table S4). By contrast, the colorimetric variables generally showed no phylogenetic  
374 signal, except for mean transmittance and brightness contrast on the hindwing (Table S4).  
375 Concerning binary structural variables, both wings showed the same evolutionary patterns: phanera  
376 presence, type, insertion and colouration in the transparent zone showed a non-random evolution  
377 conformed to a Brownian motion process (except for insertion that was different from a Brownian

378 motion process, Table S5) while in the opaque zone, interspecific variations in phanera type and  
379 insertion evolved randomly (i. e., independently of the phylogeny), probably because there was little  
380 variation on these traits (Table S5). Several structural parameters showed significant phylogenetic  
381 clustering (i. e., species that shared these structural parameters were separated by fewer nodes than  
382 expected by chance). Specifically, in the transparent zone significant phylogenetic clustering was  
383 found on the forewing for the presence of phanera, the presence of hair-like scales alone or  
384 including the mixed category (HS, scales combined with hair-like scales), for the erected insertion,  
385 and for the absence of transparency in phanera, and on the hindwing for the absence of phanera  
386 and the presence of transparency in phanera (Table S6). In the opaque zone, there was significant  
387 clustering for the presence of scales alone (S) or in combination with hair-like scales (HS) for the  
388 forewing, and the presence of combined hair-like scales and scales (HS) for the hindwing.

389 Considering structural strategies (i. e., the combination of given type, insertion and  
390 colouration), only 2/ 11 structural strategies appeared phylogenetically clustered: erected coloured  
391 hair-like scales (HEC), erected coloured scales mixed with hair-like scales (HSEC), both strategies  
392 clustered when considering wings separately or together, and the nude membrane (N) only for  
393 hindwing (Table S6).

394 Overall, structural features appeared phylogenetically conserved while structural strategies  
395 were more labile, with a few showing a significant phylogenetic clustering. Coloration showed  
396 hardly any sign of phylogenetic signal

397

398 Correlated evolution between structural traits: Structural strategies were correlated between wings.  
399 In other words, knowing the structural strategy on one wing was a good predictor of what the  
400 structural strategy would be in the other wing. In the transparent zone, phanera presence, type (H  
401 or S), insertion, and colour were correlated between wings and in the opaque zone, phanera type  
402 (H or S), and insertion were correlated between wings (Figure S2, Table S7). We found similar  
403 correlations in both wings: the phanera type (H or S) of the opaque zone was correlated to the

404 phanera type of the transparent zone. Likewise, the phanera insertion (flat or erected) of the opaque  
405 zone was also correlated to the phanera insertion of the transparent zone. While within the opaque  
406 zone, phanera type (H or S) and insertion were not correlated, it was the case in the transparent  
407 zone. In addition, phanera insertion and colouration were correlated but only for the hindwing  
408 (Figure S2, Table S7).

409

#### 410 **Structural constraints and effort in transparency**

411 Analyses reveal some relationships between structural features that suggest the existence of  
412 evolutionary constraints. Large-sized species had only a small wing area concerned by transparency,  
413 contrary to small-sized species for which the wing area concerned by transparency could span the  
414 entire range of values for the proportion of clearwing area (Figure 6B, C, Table 2). The decrease  
415 of proportion of clearwing area with wing length was stronger for the forewing than the hindwing,  
416 although both wings had similar proportion of clearwing area in average (Table 2). Finally,  
417 transparency concerned a greater proportion of wing area when it implied coloured phanera rather  
418 than transparent phanera (Figure 6D, Table 2) and the difference in proportion of clearwing area  
419 between the wings was stronger when transparency was achieved with coloured phanera than with  
420 a nude membrane.

421 By comparing phanera structures in opaque and transparent zones, we assessed the  
422 structural effort, assuming it increased as changes between both zones got more important,  
423 whatever the direction of these changes. The parameters that conditioned the changes in the  
424 transparent zone were the surface of phanera in the opaque zone and the density of phanera in the  
425 transparent zone (factors significant in all analyses in Table 3). The greater the surface of phanera  
426 in the opaque zone, the lower the structural changes in phanera length, width, surface and coverage  
427 in the transparent zone compared to the opaque zone. The greater the density of phanera in the  
428 transparent zone, the lower the structural changes in phanera length, width, surface and coverage  
429 in the transparent zone compared to the opaque zone.

430        Regarding density, transparency generally led to a reduction in density (negative intercept),  
431    a reduction that was lower for scales and the combination hair-like scales and scales than for hair-  
432    like scales alone or a nude membrane. The reduction in density decreased as clearwing area  
433    increased (Figure 7A, B, Table 3). In other words, opaque and transparent zones could highly differ  
434    (or not) in density for small-sized transparent areas, but were similar for large-sized transparent  
435    areas (Figure 7A).

436        Regarding phanera length, there was an increase in length when pooling all species together  
437    (intercept marginally positive when controlling for phylogeny). Phanera length was maximally  
438    reduced for the nude membrane, and more reduced when phanera were erected than flat, which  
439    were increased in length (Figure 7C, Table 3). Regarding phanera width, there was a global  
440    reduction when pooling all species together (negative intercept). Phanera width was maximally  
441    reduced for the nude membrane, less in hair-like scales, and even less in scales. Phanera width was  
442    even increased in transparent scales compared to coloured scales.

443        Regarding phanera surface, there was no global change when pooling all species together  
444    (intercept non-different from zero). Phanera surface was maximally reduced for the nude  
445    membrane, less in hair-like scales, and even less in scales. Large-sized species entailed lower changes  
446    in phanera surface between transparent and opaque zones. Strategies that involved transparent  
447    scales (alone or in combination with transparent hair-like scales) showed an increase in the surface  
448    of the scales, and an increase in the coverage of phanera in the transparent zone compared to the  
449    opaque zone. Notice that given our retaining only scale dimensions and densities when scales were  
450    in combination with hair-like scales, results concerning scales effectively concern that type of  
451    phanera in structural categories that include scales alone or scales in combination with hair-like  
452    scales.

453        Overall, except for transparent scales, transparency entailed a reduction in density and in  
454    phanera dimensions and coverage for all structural strategies, reaching a maximum of such  
455    reduction with nude membranes. Transparent scales followed the reverse trend, with increase in

456 width, surface and coverage on the membrane compared to the opaque zone of the same wing and  
457 species.

458

#### 459 **Impact of structure on optical transparency**

460 Microstructural and macrostructural features of the transparent zone explained the variations in  
461 optical transparency, as estimated by mean transmittance over [300-700] nm (Figure 4, 5A, C, and  
462 Table 1) and proportion of UV transmittance (Figure 4, 5 B, D, and Table 1). These parameters  
463 showed generally similar variations with microstructure: they were higher for lower phanera surface  
464 and density, for a nude membrane than for a membrane covered with phanera, for erected phanera  
465 than flat phanera, for transparent phanera compared to coloured phanera (Table 1). Mean  
466 transmittance was also higher for no phanera or hair-like scales compared to scales alone or in  
467 combination with hair-like scales, while UV transmittance did not significantly vary between  
468 phanera types but only in contrast with nude membrane. Some relationships were only significant  
469 when controlling for phylogeny, a fact that may be explained by a high optical variation between  
470 closely-related species with similar microstructural features. The large variation of transmittance  
471 within each structural category shown in Figure 4 suggests that many key features contribute to  
472 building the optical signal. At a macroscopic level, mean transmittance and the proportion of UV  
473 transmittance were higher when transparency occupied a higher proportion of wing area (Figure  
474 6A, Table 1). In addition, mean transmittance was also higher for the forewing compared to the  
475 hindwing (Table 1), while the proportion of clearwing area did not significantly vary between wings  
476 (Table 2). Overall, transparency depended on both wing macrostructure and wing microstructure.  
477 It increased as it occupied a larger proportion of wing area and as membrane coverage decreased.

478

#### 479 **Ecological relevance of transparency for vision, thermoregulation and UV protection**

480 Relevance for vision. Mean transmittance over [300-700] nm was higher on the forewing than the  
481 hindwing; it was positively correlated to proportion of UV transmittance (Figure 2A) and this

482 relationship did not differ between wings (Table 4). However, a Pearson's correlation between  
483 mean transmittance over [300-700] nm and proportion of UV transmittance yielded a moderate  
484 coefficient ( $r=0.47$ ,  $t=8.18$ ,  $p<0.001$ ), suggesting that species can play on these aspects  
485 independently, to some extent.

486 In terms of contrast, optical transparency, defined by mean transmittance or by proportion  
487 of UV transmittance, had an impact on the visual impression given to bird predators: a greater  
488 transparency yielded a reduced chromatic and achromatic contrast between the butterfly and its  
489 background, as expected (Table 4). The significant positive interaction term between brightness  
490 and colour contrast indicated that the reduction in visual contrast decreased as transparency  
491 increased (Figure 8). Such reduction of visual contrast was stronger for the hindwing than for the  
492 forewing.

493 Mean transmittance was related to species activity rhythm, whatever the model approach  
494 taken. Mean transmittance was lower in nocturnal species than in diurnal species (Table 5), but  
495 without significant difference between nocturnal and diurnal species in the proportion of clearwing  
496 area (results not shown), supporting the hypothesis that diurnal species being more active in  
497 daylight and at risk of showing parasitic reflections would be selected by visually-hunting predators  
498 for a higher transparency than nocturnal species. Transmittance was generally higher in the  
499 forewing than the hindwing. Nocturnal species had similar transmittance on both wings and  
500 showed little variation in transmittance whatever species wing length or the proportion of wing  
501 surface occupied by clearwing zones (Figure 9). Conversely, diurnal species had lower values of  
502 transmittance on the hindwing compared to the forewing, and showed steeper variations in  
503 transmittance with increasing wing length (or decreasing wing proportion of clearwing area,  
504 Figure 9).

505

506 Relevance for UV protection and thermoregulation. Mean transmittance in the UV range decreased  
507 with increasing latitude (a relationship that disappeared when controlling for phylogeny), and the

508 proportion of UV transmittance showed no significant variation with latitude (Figure 10A, C, Table  
509 6). Patterns did not significantly differ between nocturnal and diurnal species (factor not retained  
510 in the best model). These two results were in complete contradiction with the hypothesis that  
511 transparency offered UV protection, as an increase in latitude was expected for both variables.  
512 Conversely, we found that mean transmittance decreased with increasing latitude (Table 6), a  
513 pattern that supported the hypothesis of transparency playing a role in thermoregulation. These  
514 variations were independent of species size (factor not retained in the analyses). We found these  
515 results for the near infrared range [700-1100] nm and the human-visible range [400-700] nm but  
516 only for models not controlled for phylogeny for the UV range [300-400] nm (Figure 10B, D, Table  
517 6). As shown by the tighter correlation in Figure S3B compared to Figure S3A, transmittance in  
518 the human-visible wavelength range better predicted transmittance in the near infrared than in the  
519 UV range (856 AIC counts below). Finally, all transmittance spectra show a decrease in  
520 transmittance in the UV range, resulting in a proportion of UV transmittance lower than 0.25  
521 (Figure 2). This could be interpreted as a possible UV protection.

522 Overall, optical transparency translates into perceptual achromatic and chromatic  
523 transparency, it is higher in diurnal species, and higher in species living further away from the  
524 equator, supporting its ecological relevance in the context of vision and thermoregulation but not  
525 so in the context of protection against UV radiation.

526

527

## 528 **DISCUSSION**

### 529 **Diverse structural strategies produce transparency**

530 We record the presence of species with at least partially transparent wings in at least 31 families,  
531 representing a quarter of the extant butterfly families, which collectively harbour a striking majority  
532 of opaque species. Transparency has evolved multiple times independently and may present  
533 evolutionary benefits. With these multiple evolutionary gains comes a massive diversity of

534 structural strategies (combination of phanera type, insertion, and colouration), expanding the range  
535 of strategies reported in the literature in the scarce studies conducted so far in Lepidoptera.

536 The most common structural strategy involves flat scales (SF, 49 species, 22 families), either  
537 coloured and in lower densities or transparent and packed in high densities. Flat coloured scales  
538 (SFC) have been previously recorded in the nymphalid *Parantica sita* and the papilionid *Parnassius*  
539 *glacialis* (Goodwyn et al. 2009). Conversely, flat transparent scales (SFT) have not been recorded in  
540 any clearwing Lepidoptera species so far, although they have been recorded in the opaque coloured  
541 papilionid *Graphium sarpedon* (Berthier 2007; Stavenga et al. 2012). A nude membrane (N) is the  
542 second most widespread structural strategy (32 species, 12 families). This structural strategy  
543 previously recorded in the sphingid *Cephonodes hylas* (Yoshida et al. 1997) can be split into two  
544 subcategories: a fully nude membrane or a nude membrane with the presence of phanera sockets  
545 (in 7/25 species for the forewing, 5/28 species for the hindwing) that can be the remnants of fully  
546 developed scales shedding at first flight, as in the sphingid *Hemaris* sp. or that could result from the  
547 interrupted development of scales at the socket stage. Whether the latter process exists remains to  
548 be explored. Hair-like scales alone (H) are moderately represented (27 species, 9 families), and have  
549 been previously reported in the saturniid *Rothschildia lebeau* (Hernandez-Chavarria et al. 2004) or the  
550 nymphalid *Cithaerias menander* (Berthier 2007). Hair-like scales are never transparent when alone (no  
551 strategy HFT or HET) and when transparent, hair-like scales are always found in association to  
552 transparent scales. Given the relative abundance of hair-like scales in our dataset (~1/5 species and  
553 ~1/3 families), some structural or functional constraints likely limit the benefit of having  
554 transparent hair-like scales alone. Finally, combined hair-like scales and scales (HS) represents the  
555 rarest strategies in our dataset (12 species, 7 families), with erected coloured hair-like scales and  
556 scales (HSEC) showing a significant phylogenetic clustering. Another rare structural strategy  
557 involves transparent erected scales (SET, 9 species, 6 families) which shows no phylogenetic  
558 clustering, indicating that it evolved independently in distinct lineages. This structural strategy has  
559 never been recorded so far in the literature.

560 **Investment in transparency differ between structural strategies**

561 Structural strategies imply joint changes in multiple structural aspects, as shown by analyses on  
562 structural effort in transparency estimated by changes between transparent and opaque zones of  
563 the same species and wing. The general and probably intuitive picture that transparency is a  
564 reduction (here shown by general reduction in density, phanera width, membrane coverage when  
565 pooling all species together) does not hold when considering structural strategies separately. Some  
566 play on the ‘reduction’ side while others play on the opposite side of an ‘increase’.

567 At one extreme, reduction is maximal for nude membrane (absent phanera, hence no  
568 length, width, density, surface and coverage) especially when socket remnants are absent on the  
569 wing membrane (~23% of nude membrane cases) or if socket remnants reflect an interrupted  
570 development at socket stage, an hypothesis which remains to be tested. In case socket remnants  
571 result from scale full development and shedding, structural effort in transparent and opaque zones  
572 may not differ substantially. Hair-like scales alone show a decrease in density and width, an increase  
573 in length (especially for flat hair-like scales), a reduction in surface and coverage. Coloured scales  
574 showed a roughly similar pattern: a decrease in density and width an increase in length for flat scales  
575 but a decrease in length for erected scales, an increase in surface but a decrease in coverage.

576 At the other extreme, transparent scales (alone or in combination with hair-like scales) show  
577 changes towards an increase. Erected transparent scales are shorter and wider while flat transparent  
578 scales are longer and wider than their opaque counterparts. Transparent scales increase in surface  
579 and they are more densely packed, especially when flat on the wing membrane, resulting in a higher  
580 membrane coverage (density x surface). The fact that scale colouration (here present or absent)  
581 relates to scale morphology has been previously documented in the literature (e.g. Janssen et al.  
582 2001; Matsuoka and Monteiro 2018). Opaque scales of different colours have different shape and  
583 ultrastructure (lamina, crossribs, trabeculae, ridges), an aspect that remains to be investigated in  
584 future studies on transparency.

585 If we use phanera coverage to estimate the investment in chitin, transparency seems

586 maximally economical for a nude membrane without sockets while it seems costlier than the  
587 opaque zone for transparent scales. But even in the latter case, the lower investment in pigment  
588 for these transparent scales may compensate the higher investment in chitin. Still, our results  
589 suggest that transparency may be costlier than classically viewed and call for further quantification  
590 of physiological costs of transparency.

591 Although few structural strategies are phylogenetically clustered (MPD analyses), most  
592 strategies seem to have evolved independently in Lepidopteran lineages, with convergent evolution  
593 of structural parameters. Why are transparent scales often packed in higher densities and  
594 broadened, especially when erected? We can invoke two non-mutually exclusive reasons. First,  
595 melanisation may enhance mechanical resistance, as suggested in birds (Butler and Johnson 2004)  
596 and in Lepidoptera (only mentioned as pers. comm. in Brakefield 1987). In insects, cuticle  
597 melanisation and cuticle hardening (sclerotization) occur with little time lag during development  
598 and share very similar pathways with common precursors and some sclerotization components  
599 provide different coloration in insect cuticle, from colourless to yellow and brown (Sugumaran  
600 2009). Knockout mutations in genes that function in the melanin pathway affect both scale  
601 coloration and scale ultrastructure (Matsuoka and Monteiro 2018). Shortening melanin-deprived  
602 transparent scales may be a way to reinforce their mechanical resistance; it would be interesting to  
603 explore whether these shape changes come with ultrastructural changes that reinforce scale  
604 mechanical resistance. This may also explain why transparent hair-like scales are absent from our  
605 dataset, as long and thin melanin-deprived hair-like scales may be too fragile. Second, shortening  
606 erected scales and enlarging them (see the hesperid *Oxynetra semihyalina* in Figure 1) may be  
607 beneficial for water repellency and mechanical resistance. Erected phanera (HE hair-like scale  
608 alone, SE scales alone or HSE the combination of scales and hair-like scales) provide hierarchically  
609 structured geometry with asperities and air pockets between them. Such a multiscale roughness is  
610 crucial to ensure superhydrophobicity, as shown both in theoretical studies (Nosonovsky and  
611 Bhushan 2007) and in natural systems, like the cuticle of water striders (Goodwyn et al. 2008).

612 **Structural strategies differ in their optical efficiency**

613 Optical properties of transparency evolved more rapidly than structures, as shown by the absence  
614 of phylogenetic signal in most colour descriptors. Structural strategies are correlated with their  
615 efficiency at transmitting light. The nude membrane category is the most efficient while scales alone  
616 or in combination with hair-like scales are the least efficient. Erected are more efficient than flat  
617 phanera, transparent are more efficient than coloured phanera. Yet, the large variation of  
618 transmittance within structural strategies indicates that other parameters are crucial: phanera  
619 density, dimension and surface are key features. The lower the density, phanera surface and  
620 coverage on the wing membrane, the higher the transmittance. Additional parameters not  
621 measured here – wing membrane and phanera pigmentation, or nanostructures – likely play a role.  
622 For instance, wing nanostructures like nanopillars found in the glasswing nymphalid *Greta oto*  
623 (Binetti et al. 2009; Siddique et al. 2015) create by their very shape a progressive air : chitin gradient  
624 from the air interface towards the chitinous membrane which facilitates light transmission and  
625 reduces light reflection, acting as effective antireflective devices. Both the density and shape of the  
626 nanorelief are crucial to determine the amount of reduction of light reflection.

627

628 **Ecological relevance of transparency for vision, differences between small-sized and large-  
629 sized species**

630 Variations in transparency have a visual impact on the contrast offered by butterflies with a  
631 background. While translation of mean transmittance into brightness contrast is intuitive,  
632 translation of mean transmittance into colour contrast is more surprising; it underlines that  
633 transmittance spectra are far from being flat spectra. Chromatic contribution can come from  
634 structures in phanera or in the wing membrane (e.g. iridescence produced by the wing membrane  
635 in Quiroz et al. 2019) or from pigments in phanera (scales and/or hair-like scales, S, H or HS,  
636 which concerns 53 species in our dataset) or in the wing membrane. Our results show that, although  
637 increasing transmittance yields reduced visual contrast, the gain gets lower as transmittance

638 increases, which may explain why the maximal value we found in our measurements was 90.25%.

639 Even if transmittance can reach up to 95% in the [400-700] nm range, it is lower in the UV range

640 due to pigment absorption. Melanin has stronger absorptance at short than at long wavelengths

641 (Wolbarsht et al. 1981) and even a weak pigmentation produces a loss of transmittance in the UV

642 range first. Transmittance spectra reveal that transparency is fundamentally not achromatic. This

643 ‘imperfection’ may result from a weaker selection of vision in this range, either because visual

644 systems get less performing as wavelength decreases in the UV, and/or because short wavelengths

645 are cut out by vegetation (Endler 1993), thereby attenuating the importance of this range for forest-

646 dwelling species. Transparency can be efficient enough without further increase of achromaticity

647 in the UV range. Moreover, other constraints such as the need of communication and

648 thermoregulation (see below) may offset the visual gain, and species may be able to ‘play’ on the

649 UV range rather independently from longer wavelengths, as suggested by the loose relationship

650 between transmittance and proportion of UV transmittance that we revealed.

651 Following on the idea that ambient light influences perception of transparency, we found

652 that nocturnal species displayed lower mean transmittance than diurnal species. Yet, nocturnal and

653 diurnal species did not differ in their proportion of clearwing area. Nocturnal species resting on

654 vegetation or mineral support are protected by their immobility from incidentally exposing parasitic

655 reflections, which diurnal species experience when moving during the day. At night, even if

656 nocturnal species were hunted by visually-guided predators, dim light conditions would hamper

657 detection and relax selection towards high transparency levels. Transparency being similar in

658 forewing and hindwing in nocturnal species is coherent with an overall signal if both wings are

659 exposed to predators. By contrast, diurnal species show a higher transparency on the forewing

660 compared to the hindwing. Such a discrepancy between colour traits in forewing and hindwing has

661 been shown for instance in the genus *Bicyclus* (Nymphalidae, ~80 species) where forewing eyespots

662 evolve more rapidly and are more subject to sexual dimorphism than hindwing eyespots (Oliver et

663 al. 2009). Such coloration strategies likely relate to butterfly potential ability to hide or not

664 hindwings when at rest, and to butterfly shape, forewings being generally larger thus more visible  
665 than hindwings to potential predators or partners.

666 Interestingly, our analyses reveal size-dependent strategies that can be interpreted in the  
667 context of a role of transparency in vision. Small and large-sized butterfly species seem to differ  
668 optically, for two combined reasons: (i) in small-sized species, transparency spans a wide range of  
669 proportions of wing surface from spots to entire wing, but in large-sized species, transparency is  
670 always restricted to small proportion of wing surface, and (ii) the efficiency of transparency at  
671 transmitting light positively correlates with proportion of clearwing area. Hence, small and poorly  
672 transmitting spots can be present in wings of all species, such as the large saturniid *Rothschildia lebeau*  
673 (Hernandez-Chavarria et al. 2004) and the small thyridid *Dysodia speculifera*. Conversely, only small  
674 species can have almost entirely transparent wings with a high efficiency at transmitting light, as  
675 the psychiid *Chaliooides fereritrea*, the wings of which are extremely difficult to detect.

676 Why so? Small species are often nearly undetectable given their small size when they have  
677 wings with high proportion of clearwing area. Conversely, large species may not benefit from  
678 having high proportion of clearwing area, for several reasons.

679 First, optical benefits may be offset by costs entailed by transparency for other functions.  
680 For instance, efficiency at repelling water is crucial for butterflies. Among the butterfly species  
681 investigated so far (Wagner et al. 1996; Zheng et al. 2007; Goodwyn et al. 2009; Wanasekara and  
682 Chalivendra 2011; Fang et al. 2015), the largest but poorly transmitting species like the nymphalid  
683 *Parantica sita* and the papilionid *Parnassius glacialis* show moderate to high hydrophobicity (Goodwyn  
684 et al. 2009; Fang et al. 2015). Conversely, the species with the highest proportion of clearwing area  
685 and covered with hair-like scales, the nymphalid *Greta oto*, has one of the lowest hydrophobicity  
686 values found (Wanasekara and Chalivendra 2011). These potential trade-offs between optics and  
687 hydrophobicity remain to be studied. Yet, we can hypothesise that keeping clearwing area to a  
688 reduced proportion of surface wing (lower proportion of clearwing area) may bring substantial  
689 visual benefits while being beneficial for repelling water.

690        Second, optical benefits of being transparent may be limited by butterfly size in large  
691        species. In erebid moths, Kang et al (2017) have found cryptic colouration in both wings of small  
692        species and wing-dependent colouration in large species, with cryptic colouration on the forewing  
693        and hidden contrasting colour signals on the hindwing, probably involved in startle displays. They  
694        posit two hypotheses to explain this size-dependent switch of anti-predator defence: crypsis fails  
695        as body size increases and secondary defence like startle displays are more effective in large prey,  
696        hypotheses that they confirm experimentally. If such an anti-predator switch applies for  
697        transparency, crypsis by the means of transparency could potentially fail in large species for two  
698        reasons: a conspicuous large opaque body, and a wing transparency too limited in efficiency by  
699        wing membrane thickness. More studies are needed to explore this size-dependent switch of  
700        transparency wing pattern and its structural determinants.

701        In large species, what would then be the potential benefit of keeping small clear spots that  
702        sometimes account for only a few percent of wing total area? We can emit several hypotheses: (i)  
703        clear spots can function as surface disruption patterns that create false margins away from the  
704        animal outline. When combined with a cryptic opaque colouration, this can result in internal edges  
705        being more salient than the true outline form (Stevens, Winney, et al. 2009). As suggested by  
706        Costello et al. (2020), these false internal edges can create false holes, especially when transparent  
707        area edges are enhanced, as this creates false depth planes that foster incorrect visual categorization  
708        as depth cues. (ii) Clear-spotted cryptically coloured butterflies may be protected by masquerade,  
709        predators taking them for non-edible objects of their environment, like damaged or rotten leaves,  
710        an hypothesis formulated by Janzen (1984) but never tested. Such an hypothesis would function  
711        of course if birds are not attracted to damaged leaves as a cue for the presence of insects, as  
712        suggested by Costello et al. (2020). (iii) When transmission is poor, small spots could resemble  
713        white-pigmented spots, which could be involved in communication. More generally, small clear  
714        spots may function as eyespots, which are efficient at limiting predator attack and deviate them  
715        from vital parts (Stevens et al. 2008; Stevens, Cantor, et al. 2009) and used in mate choice

716 (Robertson and Monteiro 2005) in opaque species. Experimental studies are needed to clarify this  
717 point.

718

## 719 **Ecological relevance of transparency for thermoregulation**

720 The general patterns of variations of transmittance with latitudinal distance to equator support the  
721 idea that transparency is involved in thermoregulation. While transmittance shows its largest  
722 variation in the tropics, it decreases as latitude (in its absolute value) increases, with an average loss  
723 of 4 to 5% transmittance per  $10^{\circ}$  increase in latitude. Such variations primarily concern the near-  
724 infrared [700-1100] nm range and the human-visible [400-700] nm range. For the UV range, the  
725 relationship found in classic mixed models vanishes when controlling for phylogeny, which  
726 suggests it is likely due more to phylogenetic common ancestry than to adaptive convergence in  
727 response to a common environmental pressure. Given that transmission + reflection + absorption  
728 = 1, it is reasonable to think that lower transmittance values likely translate, at least in part, into  
729 higher absorption levels, if reflection values are roughly maintained at the same levels. In this  
730 context, we can interpret the decrease in transmittance with increasing latitude as an increase in  
731 absorption with increasing latitude. In this context, our results are in agreement with the thermal  
732 melanism hypothesis found in previous studies (Zeuss et al. 2014; Heidrich et al. 2018; Xing et al.  
733 2018; Stelbrink et al. 2019). Contrary to Xing et al. (2018), we did not find any differences in the  
734 latitude-dependent variations in transparency between nocturnal and diurnal species. Contrary to  
735 Munro et al. (2019) study in Australian opaque butterflies, where wing reflectance (transmission =  
736 0 in opaque patches) decreases with increasing latitude, and more in the near infrared range than  
737 at shorter wavelengths, we find similar results in the human-visible [400-700] nm and the near-  
738 infrared [700-1100] nm range. Why so? Why would these two ranges be so tightly correlated? Is it  
739 part of the intrinsic nature of transparency? Indeed, chitin and melanin refraction indexes and  
740 absorption coefficients vary less in wavelengths at long than at short wavelengths (Azofeifa et al.  
741 2012; Stavenga et al. 2014), ensuring more similar properties at longer wavelengths. To this optical

742 explanation of physical properties of materials, we can add a biological one. Increasing ambient  
743 light intensity levels determine the maximal sighting distance at which a transparent object can be  
744 detected (Johnsen and Widder 1998; Ruxton et al. 2004). A poorly transmitting target seen in  
745 dimmer light can be as poorly detectable as a highly transmitting target seen in brighter light. Light  
746 intensities are higher in the tropics than in the temperate zone (Gueymard and Ruiz-Arias 2016)  
747 and visual systems may therefore select for a higher transmittance in the tropics. As a consequence,  
748 thermoregulation (in the infrared range) and vision (at shorter wavelengths) may act concurrently  
749 and contribute to creating an autocorrelation. More studies are needed to decipher the relative  
750 contribution of both selective pressures on variations of transparency at large geographical scale.

751 Contrary to Munro et al. (2019) who have found that climate correlates more to body and  
752 proximal wing colouration (the body parts that are most relevant for thermoregulation given that  
753 wing poorly transmits heat) than distal wing colouration, we find a correlation between  
754 transmittance and latitude for varied locations across the wing. We measured 5 points per wing at  
755 varied locations within the transparent zone and the transparent zone was not always at the same  
756 distance from the insect body. Whether there exist optical variations within transparent zones in  
757 relation to the distance to the insect body requires further investigation. Contrary to Munro et al.'s  
758 (2019) study, we did not find that latitudinal variations were more pronounced in smaller species.  
759 Overall, our results that the optical properties of transparency change with latitude suggests that,  
760 even if the thermal gain may appear really small given that it concerns transparency (where  
761 absorption is minimal), benefits accumulated over a life-time can be substantial (Stuart-Fox et al.  
762 2017).

763

#### 764 **Ecological relevance of transparency for UV protection?**

765 Given higher UV radiation levels at lower latitudes (Beckmann et al. 2014), transparent patches  
766 should absorb more UV radiation at lower latitudes if transparency helps protecting species from  
767 these harmful radiation. Given that transmission + reflection + absorption = 1, and if reflection

768 levels are roughly similar between species, we expect that clearwing species living at lower latitudes  
769 should transmit less in the UV range, in absolute (transmittance over [300-400] nm) or in  
770 proportion (that the UV range should contribute less to the overall transmittance). Our results that  
771 transmittance in the UV range transmit more in the UV range at lower latitudes and the absence  
772 of relationship between the proportion of UV transmittance and latitude are in contradiction with  
773 the hypothesis that wing melanisation may help protection against UV radiation and pathogens  
774 (True 2003) which are more important in warmer and more humid regions, hence in lower latitudes.  
775 Likewise, agreement with thermoregulation but not with UV protection has been found in the  
776 coloration of bird eggs, for which thermoregulation is crucial to ensure embryo development  
777 (Wisocki et al. 2020). Still, the fact that UV transmittance is always lower than transmittance at  
778 longer wavelengths shows an absorption in the UV range, as shown by chitin and melanin  
779 absorption curves (Wolbarsht et al. 1981; Azofeifa et al. 2012; Stavenga et al. 2014).

780

## 781 Conclusion

782 Transparency has evolved multiple times independently in an insect order characterised by wing  
783 opacity. These multiple gains have led to a large diversity of structural strategies to achieve  
784 transparency. Optical transparency is determined by both macrostructure ((length, area, proportion  
785 of clearwing area) and microstructure (phanera dimensions, type, insertion, colouration, and  
786 density). Microstructural traits are tightly linked in their evolution leading to differential investment  
787 in chitin and pigments between structural strategies. Physical transparency translates into visually  
788 effective concealment with interesting size-dependent and rhythm-dependent differences likely  
789 selected for camouflage and communication. The links between transparency and latitude are  
790 consistent with thermal benefits, and much less with UV protection. These results often echo the  
791 results that have been found in opaque Lepidoptera, showing that transparency is more complex  
792 than just enhancing concealment and is likely a multifunctional compromise. Experimental studies

793 are needed to shed light on some hypotheses emitted to explain the patterns of transparency with  
794 species ecology.

795

796

797 **ACKNOWLEDGEMENTS**

798 This work was funded by Clearwing ANR project (ANR-16-CE02-0012), HFSP project on  
799 transparency (RGP0014/2016) and a France-Berkeley fund grant (FBF #2015---58). We warmly  
800 thank Jacques Pierre and Rodolphe Rougerie for help with species choice, identification, and data  
801 on species ecology, Edgar Attivissimo for contributing to Keyence imaging, and Thibaud Decaëns,  
802 Daniel Herbin, and Claude Tautel for species selection and identification.

803

804

805 **REFERENCES**

806 Arias M, Elias M, Andraud C, Berthier S, Gomez D. 2019. Transparency improves concealment in  
807 cryptically coloured moths. *J Evol Biol.*:jeb.13560. doi:10.1111/jeb.13560.

808 Arias M, Mappes J, Desbois C, Gordon S, McClure M, Elias M, Nokelainen O, Gomez D. 2019.  
809 Transparency reduces predator detection in chemically protected clearwing butterflies. *Funct Ecol.*  
810 doi:10.1111/1365-2435.13315. <https://www.biorxiv.org/content/10.1101/410241v1>.

811 Azofeifa DE, Arguedas HJ, Vargas WE. 2012. Optical properties of chitin and chitosan  
812 biopolymers with application to structural color analysis. *Opt Mater.* 35(2):175–183.  
813 doi:10.1016/j.optmat.2012.07.024.

814 Bagge LE. 2019. Not as clear as it may appear: challenges associated with transparent camouflage  
815 in the ocean. *Integr Comp Biol.* 59(6):1653–1663. doi:10.1093/icb/icz066.

816 Beckmann M, Václavík T, Manceur AM, Šprtová L, Wehrden H von, Welk E, Cord AF. 2014.  
817 glUV: a global UV-B radiation data set for macroecological studies. *Methods Ecol Evol.* 5(4):372–  
818 383. doi:10.1111/2041-210X.12168.

819 Berthier S. 2005. Thermoregulation and spectral selectivity of the tropical butterfly *Prepona meander*.  
820 a remarkable example of temperature auto-regulation. *Appl Phys -Mater Sci Process.* 80(7):1397–  
821 1400. doi:10.1007/s00339-004-3185-x.

822 Berthier S. 2007. Iridescent colors of insects. Berlin, Germany: Springer.

823 Binetti VR, Schiffman JD, Leaffer OD, Spanier JE, Schauer CL. 2009. The natural transparency  
824 and piezoelectric response of the *Greta oto* butterfly wing. *Integr Biol.* 1(4):324–329.  
825 doi:10.1039/b820205b.

826 Blomberg SP, Garland T, Ives AR. 2003. Testing for phylogenetic signal in comparative data:  
827 behavioral traits are more labile. *Evolution.* 57(4):717–745. doi:10.1111/j.0014-  
828 3820.2003.tb00285.x.

829 Bogert CM. 1949. Thermoregulation in reptiles, a factor in evolution. *Evolution.* 3(3):195–211.  
830 doi:10.1111/j.1558-5646.1949.tb00021.x.

831 Brakefield PM. 1987. Industrial melanism: do we have the answers? *Trends Ecol Evol.* 2(5):117–  
832 122.

833 Butler M, Johnson AS. 2004. Are melanized feather barbs stronger? *J Exp Biol.* 207(2):285–293.  
834 doi:10.1242/jeb.00746.

835 Costello LM, Scott-Samuel NE, Kjernsmo K, Cuthill IC. 2020. False holes as camouflage. *Proc R  
836 Soc B Biol Sci.* 287(1922):20200126. doi:10.1098/rspb.2020.0126.

837 Cuthill IC. 2019. Camouflage. *J Zool.* 308(2):75–92. doi:10.1111/jzo.12682.

838 Deparis O, Mouchet S, Dellieu L, Colomer J-F, Sarrazin M. 2014. Nanostructured surfaces:  
839 bioinspiration for transparency, coloration and wettability. *Mater Today Proc.* 1:122–129.  
840 doi:10.1016/j.matpr.2014.09.008.

841 Drummond AJ, Rambaut A. 2007. BEAST: Bayesian evolutionary analysis by sampling trees. *BMC  
842 Evol Biol.* 7(1):214. doi:10.1186/1471-2148-7-214.

843 Drummond AJ, Suchard MA, Xie D, Rambaut A. 2012. Bayesian Phylogenetics with BEAUti and  
844 the BEAST 1.7. *Mol Biol Evol.* 29(8):1969–1973. doi:10.1093/molbev/mss075.

845 Elbourne A, Crawford RJ, Ivanova EP. 2017. Nano-structured antimicrobial surfaces: from nature  
846 to synthetic analogues. *J Colloid Interface Sci.* 508:603–616. doi:10.1016/j.jcis.2017.07.021.

847 Ellers J, Boggs CL. 2004. Functional ecological implications of intraspecific differences in wing  
848 melanization in *Colias* butterflies. *Biol J Linn Soc.* 82(1):79–87.

849 Endler JA. 1993. The color of light in forests and its implications. *Ecol Monogr.* 63(1):1–27.

850 Fang Y, Sun G, Bi YH, Zhi H. 2015. Multiple-dimensional micro/nano structural models for  
851 hydrophobicity of butterfly wing surfaces and coupling mechanism. *Sci Bull.* 60(2):256–263.  
852 doi:10.1007/s11434-014-0653-3.

853 Fritz SA, Purvis A. 2010. Selectivity in Mammalian Extinction Risk and Threat Types: a New  
854 Measure of Phylogenetic Signal Strength in Binary Traits. *Conserv Biol.* 24(4):1042–1051.  
855 doi:10.1111/j.1523-1739.2010.01455.x.

856 Gomez D. 2011. AVICOL v6. a program to analyse spectrometric data. Free program available  
857 from the author upon request at [dodogomez@yahoo.fr](mailto:dodogomez@yahoo.fr) or by download from  
858 <http://sites.google.com/site/avicolprogram/>.

859 Gomez D, Théry M. 2007. Simultaneous crypsis and conspicuousness in color patterns:  
860 comparative analysis of a Neotropical rainforest bird community. *Am Nat.* 169:S42–S61.

861 Goodwyn PP, De Souza E, Fujisaki K, Gorb S. 2008. Moulding technique demonstrates the  
862 contribution of surface geometry to the super-hydrophobic properties of the surface of a water  
863 strider. *Acta Biomater.* 4(3):766–770. doi:10.1016/j.actbio.2008.01.002.

864 Goodwyn PP, Maezono Y, Hosoda N, Fujisaki K. 2009. Waterproof and translucent wings at the  
865 same time: problems and solutions in butterflies. *Naturwissenschaften.* 96(7):781–787.  
866 doi:10.1007/s00114-009-0531-z.

867 Gueymard CA, Ruiz-Arias JA. 2016. Extensive worldwide validation and climate sensitivity analysis  
868 of direct irradiance predictions from 1-min global irradiance. *Sol Energy.* 128:1–30.  
869 doi:10.1016/j.solener.2015.10.010.

870 Guillerme T, Healy K. 2019. *mulTree*: performs MCMCglmm on multiple phylogenetic trees. R  
871 package version 1.3.6.

872 Hadfield JD. 2010. MCMC methods for multi-response generalized linear mixed models: The  
873 MCMCglmm R Package. *J Stat Softw.* 33(2). doi:10.18637/jss.v033.i02.  
874 <http://www.jstatsoft.org/v33/i02/>.

875 Hart NS, Partridge JC, Cuthill IC, Bennett ATD. 2000. Visual pigments, oil droplets, ocular media  
876 and cone photoreceptor distribution in two species of passerine bird: the blue tit (*Parus caeruleus* L.)  
877 and the blackbird (*Turdus merula* L.). *J Comp Physiol Ser A.* 186(4):375–387.

878 Heidrich L, Friess N, Fiedler K, Braendle M, Hausmann A, Brandl R, Zeuss D. 2018. The dark  
879 side of Lepidoptera: colour lightness of geometrid moths decreases with increasing latitude. *Glob  
880 Ecol Biogeogr.* 27(4):407–416. doi:10.1111/geb.12703.

881 Hernandez-Chavarria F, Hernandez A, Sittenfeld A. 2004. The ‘windows’ scales, and bristles of the  
882 tropical moth *Rothschildia lebeau* (Lepidoptera : Saturniidae). *Rev Biol Trop.* 52(4):919–926.

883 Janssen JM, Monteiro A, Brakefield PM. 2001. Correlations between scale structure and  
884 pigmentation in butterfly wings. *Evol Dev.* 3(6):415–423. doi:10.1046/j.1525-142X.2001.01046.x.

885 Janzen DH. 1984. Weather-Related Color Polymorphism of *Rothschildia lebeau* (Saturniidae). *Bull  
886 Entomol Soc Am.* 30(2):16–21. doi:10.1093/besa/30.2.16.

887 Johnsen S. 2001. Hidden in plain sight: The ecology and physiology of organismal transparency.  
888 *Biol Bull.* 201(3):301–318. doi:10.2307/1543609.

889 Johnsen S. 2014. Hide and seek in the open sea: pelagic camouflage and visual countermeasures.  
890 *Annu Rev Mar Sci Vol 6.* 6:369–392. doi:10.1146/annurev-marine-010213-135018.

891 Johnsen S, Widder EA. 1998. Transparency and visibility of gelatinous zooplankton from the  
892 Northwestern Atlantic and Gulf of Mexico. *Biol Bull.* 195(3):337–348. doi:10.2307/1543145.

893 Kang C, Zahiri R, Sherratt TN. 2017. Body size affects the evolution of hidden colour signals in  
894 moths. *Proc R Soc B Biol Sci.* 284(1861):20171287. doi:10.1098/rspb.2017.1287.

895 Kembel SW, Cowan PD, Helmus MR, Cornwell WK, Morlon H, Ackerly DD, Blomberg SP, Webb  
896 CO. 2010. Picante: R tools for integrating phylogenies and ecology. *Bioinformatics*. 26(11):1463–  
897 1464. doi:10.1093/bioinformatics/btq166.

898 Kemp DJ. 2007. Female butterflies prefer males bearing bright iridescent ornamentation. *Proc R  
899 Soc B-Biol Sci.* 274(1613):1043–1047. doi:10.1098/rspb.2006.0043.

900 Krishna A, Nie X, Warren AD, Llorente-Bousquets JE, Briscoe AD, Lee J. 2020. Infrared optical  
901 and thermal properties of microstructures in butterfly wings. *Proc Natl Acad Sci.* 117(3):1566–  
902 1572. doi:10.1073/pnas.1906356117.

903 Lanfear R, Frandsen PB, Wright AM, Senfeld T, Calcott B. 2017. PartitionFinder 2: new methods  
904 for selectingpartitioned models of evolution for molecular and morphologicalphylogenetic  
905 analyses. *Mol Biol Evol.* 34(3):772–773. doi:10.1093/molbev/msw260.

906 Lind O, Karlsson S, Kelber A. 2013. Brightness discrimination in budgerigars (*Melopsittacus  
907 undulatus*). Dyer AG, editor. *PLoS ONE*. 8(1):e54650. doi:10.1371/journal.pone.0054650.

908 Lind O, Mitkus M, Olsson P, Kelber A. 2014. Ultraviolet vision in birds: the importance of  
909 transparent eye media. *Proc R Soc B-Biol Sci.* 281(1774). doi:10.1098/rspb.2013.2209.

910 Liu Y, Song Y, Niu S, Zhang Y, Han Z, Ren L. 2016. Integrated super-hydrophobic and  
911 antireflective PDMS bio-templated from nano-conical structures of cicada wings. *RSC Adv.*  
912 6(110):108974–108980. doi:10.1039/C6RA23811D.

913 Maia R, Gruson H, Endler JA, White TE. 2019. pavo 2: New tools for the spectral and spatial  
914 analysis of colour in r. *Methods Ecol Evol.* 10(7):1097–1107. doi:10.1111/2041-210X.13174.

915 Maier EJ, Bowmaker JK. 1993. Colour vision in the passeriform bird, *Leiothrix lutea*: correlation of  
916 visual pigment absorbance and oil droplet transmission with spectral sensitivity. *J Comp Physiol  
917 Ser A.* 172:295–301. doi:10.1007/BF00216611.

918 Matsuoka Y, Monteiro A. 2018. Melanin Pathway Genes Regulate Color and Morphology of  
919 Butterfly Wing Scales. *Cell Rep.* 24(1):56–65. doi:10.1016/j.celrep.2018.05.092.

920 McClure M, Clerc C, Desbois C, Meichanetzoglou A, Cau M, Bastin-Hélène L, Bacigalupo J,  
921 Houssin C, Pinna C, Nay B, et al. 2019. Why has transparency evolved in aposematic butterflies?  
922 Insights from the largest radiation of aposematic butterflies, the Ithomiini. *Proc R Soc B Biol Sci.*  
923 286(1901):20182769. doi:10.1098/rspb.2018.2769.

924 Miaoulis IN, Heilman BD. 1998. Butterfly thin films serve as solar collectors. *Ann Entomol Soc  
925 Am.* 91(1):122–127. doi:10.1093/aes/91.1.122.

926 Miller MA, Pfeiffer W, Schwartz T. 2010. Creating the CIPRES Science Gateway for inference of  
927 large phylogenetic trees. In: 2010 Gateway Computing Environments Workshop (GCE). p. 1–8.

928 Munro JT, Medina I, Walker K, Moussalli A, Kearney MR, Dyer AG, Garcia J, Rankin KJ, Stuart-  
929 Fox D. 2019. Climate is a strong predictor of near-infrared reflectance but a poor predictor of  
930 colour in butterflies. *Proc R Soc B Biol Sci.* 286(1898):20190234. doi:10.1098/rspb.2019.0234.

931 Nosonovsky M, Bhushan B. 2007. Hierarchical roughness makes superhydrophobic states stable.  
932 *Microelectron Eng.* 84(3):382–386. doi:10.1016/j.mee.2006.10.054.

933 Oliver JC, Robertson KA, Monteiro A. 2009. Accommodating natural and sexual selection in  
934 butterfly wing pattern evolution. Proc R Soc Lond Ser B-Biol Sci. 276(1666):2369–2375.  
935 doi:10.1098/rspb.2009.0182.

936 Orme CDL, Freckleton RP, Thomas GH, Petzoldt T, Fritz SA, Isaac NJB, Pearse W. 2018. Caper:  
937 comparative analyses of phylo-genetics and evolution in R. R package version 1.0.1.  
938 <http://CRAN.R-project.org/package=caper>.

939 Pagel M. 1999. Inferring the historical patterns of biological evolution. Nature. 401(6756):877–884.  
940 doi:10.1038/44766.

941 Pinheiro J, Bates D, DebRoy D, Sarkar DR. 2020. nlme: linear and nonlinear mixed effects models.  
942 R package version 3.1-145. <https://CRAN.R-project.org/package=nlme>.

943 Quiroz HP, Barrera-Patiño CP, Rey-González RR, Dussan A. 2019. Optical properties of *Greta oto*  
944 butterfly wings: relation of iridescence with photonic properties. J Nanosci Nanotechnol.  
945 19(5):2833–2838. doi:10.1166/jnn.2019.16028.

946 R Development Core Team. 2013. R: a language and environment for statistical computing. R  
947 Foundation for Statistical Computing. <http://www.R-project.org>.

948 Ratnasingham S, Hebert PDN. 2007. BOLD: the barcode of life data system  
949 (<http://www.barcodinglife.org>). Mol Ecol Notes. 7(3):355–364. doi:10.1111/j.1471-  
950 8286.2007.01678.x.

951 Regier JC, Mitter C, Zwick A, Bazinet AL, Cummings MP, Kawahara AY, Sohn J-C, Zwickl DJ,  
952 Cho S, Davis DR, et al. 2013. A large-scale, higher-level, molecular phylogenetic study of the insect  
953 order Lepidoptera (moths and butterflies). PLoS One. 8(3). doi:10.1371/journal.pone.0058568.

954 Revell LJ. 2012. phytools: an R package for phylogenetic comparative biology (and other things).  
955 Methods Ecol Evol. 3(2):217–223. doi:10.1111/j.2041-210X.2011.00169.x.

956 Robertson KA, Monteiro A. 2005. Female *Bicyclus anynana* butterflies choose males on the basis of  
957 their dorsal UV-reflective eyespot pupils. Proc R Soc Lond Ser B-Biol Sci. 272(1572):1541–1546.  
958 doi:10.1098/rspb.2005.3142.

959 Ruxton GD, Sherratt TN, Speed MP. 2004. Avoiding attack - the evolutionary ecology of crypsis,  
960 warning signals, and mimicry. New York: Oxford University Press.

961 Schneider CA, Rasband WS, Eliceiri KW. 2012. NIH Image to ImageJ: 25 years of image analysis.  
962 Nat Methods. 9(7):671–675. doi:10.1038/nmeth.2089.

963 Siddique RH, Gomard G, Holscher H. 2015. The role of random nanostructures for the  
964 omnidirectional anti-reflection properties of the glasswing butterfly. Nat Commun. 6.  
965 doi:10.1038/ncomms7909.

966 Smith SA, Dunn CW. 2008. Phyutility: a phyloinformatics tool for trees, alignments and molecular  
967 data. Bioinformatics. 24(5):715–716. doi:10.1093/bioinformatics/btm619.

968 Stavenga DG, Leertouwer HL, Wilts BD. 2014. The colouration toolkit of the pipevine swallowtail  
969 butterfly, *Battus philenor*: thin films, papiliochromes, and melanin. J Comp Physiol A. 200(6):547–  
970 561. doi:10.1007/s00359-014-0901-7.

971 Stavenga DG, Matsushita A, Arikawa K, Leertouwer HL, Wilts BD. 2012. Glass scales on the wing  
972 of the swordtail butterfly *Graphium sarpedon* act as thin film polarizing reflectors. *J Exp Biol.*  
973 215(4):657–662. doi:10.1242/jeb.066902.

974 Stelbrink P, Pinkert S, Brunzel S, Kerr J, Wheat CW, Brandl R, Zeuss D. 2019. Colour lightness of  
975 butterfly assemblages across North America and Europe. *Sci Rep.* 9:1760. doi:10.1038/s41598-  
976 018-36761-x.

977 Stevens M, Cantor A, Graham J, I.S. W. 2009. The function of animal ‘eyespots’: conspicuousness  
978 but not eye mimicry is key. *Curr Zool.* 55(5):319–326.

979 Stevens M, Stubbins CL, Hardman CJ. 2008. The anti-predator function of ‘eyespots’ on  
980 camouflaged and conspicuous prey. *Behav Ecol Sociobiol.* 62(11):1787–1793. doi:10.1007/s00265-  
981 008-0607-3.

982 Stevens M, Winney IS, Cantor A, Graham J. 2009. Outline and surface disruption in animal  
983 camouflage. *Proc R Soc B Biol Sci.* 276(1657):781–786. doi:10.1098/rspb.2008.1450.

984 Stoffel MA, Nakagawa S, Schielzeth H. 2017. rptR: repeatability estimation and variance  
985 decomposition by generalized linear mixed-effects models. *Methods Ecol Evol.* 8(11):1639–1644.  
986 doi:10.1111/2041-210X.12797.

987 Stuart-Fox D, Newton E, Clusella-Trullas S. 2017. Thermal consequences of colour and near-  
988 infrared reflectance. *Philos Trans R Soc B Biol Sci.* 372(1724):20160345.  
989 doi:10.1098/rstb.2016.0345.

990 Sugumaran M. 2009. Complexities of cuticular pigmentation in insects. *Pigment Cell Melanoma  
991 Res.* 22(5):523–525. doi:10.1111/j.1755-148X.2009.00608.x.

992 True JR. 2003. Insect melanism: the molecules matter. *Trends Ecol Evol.* 18(12):640–647.  
993 doi:10.1016/j.tree.2003.09.006.

994 Tuthill JC, Johnsen S. 2006. Polarization sensitivity in the red swamp crayfish *Procambarus clarkii*  
995 enhances the detection of moving transparent objects. *J Exp Biol.* 209(9):1612–1616.  
996 doi:10.1242/jeb.02196.

997 Vorobyev M, Osorio D. 1998. Receptor noise as a determinant of colour thresholds. *Proc R Soc  
998 Lond Ser B-Biol Sci.* 265(1394):351–358.

999 Wagner T, Neinhuis C, Barthlott W. 1996. Wettability and contaminability of insect wings as a  
1000 function of their surface sculptures. *Acta Zool.* 77(3):213–225. doi:10.1111/j.1463-  
1001 6395.1996.tb01265.x.

1002 Wanasekara ND, Chalivendra VB. 2011. Role of surface roughness on wettability and coefficient  
1003 of restitution in butterfly wings. *Soft Matter.* 7(2):373–379. doi:10.1039/c0sm00548g.

1004 Webb CO. 2000. Exploring the phylogenetic structure of ecological communities: An example for  
1005 rain forest trees. *Am Nat.* 156(2):145–155.

1006 Webb CO, Ackerly DD, McPeek MA, Donoghue MJ. 2002. Phylogenies and community ecology.  
1007 *Annu Rev Ecol Syst.* 33:475–505. doi:10.1146/annurev.ecolys.33.010802.150448.

1008 Wisocki PA, Kennelly P, Rivera IR, Cassey P, Burkey ML, Hanley D. 2020. The global distribution  
1009 of avian eggshell colours suggest a thermoregulatory benefit of darker pigmentation. *Nat Ecol  
1010 Evol.* 4(1):148–155. doi:10.1038/s41559-019-1003-2.

1011 Wolbarsht ML, Walsh AW, George G. 1981. Melanin, a unique biological absorber. *Appl Opt.*  
1012 20(13):2184–2186. doi:10.1364/AO.20.002184.

1013 Xing S, Bonebrake TC, Ashton LA, Kitching RL, Cao M, Sun Z, Ho JC, Nakamura A. 2018. Colors  
1014 of night: climate-morphology relationships of geometrid moths along spatial gradients in  
1015 southwestern China. *Oecologia.* 188(2):537–546. doi:10.1007/s00442-018-4219-y.

1016 Yoshida A, Motoyama M, Kosaku A, Miyamoto K. 1997. Antireflective nanoprotuberance array in  
1017 the transparent wing of a hawkmoth, *Cephonodes hylas*. *Zoolog Sci.* 14(5):737–741.  
1018 doi:10.2108/zsj.14.737.

1019 Zeuss D, Brandl R, Brändle M, Rahbek C, Brunzel S. 2014. Global warming favours light-coloured  
1020 insects in Europe. *Nat Commun.* 5:3874. doi:10.1038/ncomms4874.

1021 Zhang C-Y, Meng J-Y, Wang X-P, Zhu F, Lei C-L. 2011. Effects of UV-A exposures on longevity  
1022 and reproduction in *Helicoverpa armigera*, and on the development of its F1 generation. *Insect Sci.*  
1023 18(6):697–702. doi:10.1111/j.1744-7917.2010.01393.x.

1024 Zheng YM, Gao XF, Jiang L. 2007. Directional adhesion of superhydrophobic butterfly wings.  
1025 *Soft Matter.* 3(2):178–182. doi:10.1039/B612667G.

1026

**Table 1. Relationships between optics and structure**

Factor	Mean Transmittance over 300-700 nm			Proportion of UV transmittance		
	Mixed model		Bayesian	Mixed model		Bayesian
	Estimate ± se	t-value	Estimate [CI95%]	Estimate ± se	t-value	Estimate [CI95%]
intercept	<b>60.2 ± 2.79</b>	<b>21.6***</b>	<b>52.42 [27.42, 76.4]</b>	<b>0.16 ± 0.01</b>	<b>23.45***</b>	<b>0.16 [0.089, 0.233]</b>
(HW>FW)	<b>-1.86 ± 0.93</b>	<b>-2.01*</b>	<b>-1.56 [-2.52, -0.59]</b>	-	-	-
Phanera surface	-0.0006 ± 0.0003	-1.77~	<b>-0.0008 [-0.0012, -0.0004]</b>	<b>-1.5 10<sup>-6</sup> ± 0.6 10<sup>-6</sup></b>	<b>-2.29*</b>	<b>-1 10<sup>-6</sup> [-2 10<sup>-6</sup>, -1 10<sup>-6</sup>]</b>
Phanera density	<b>-0.02 ± 0.01</b>	<b>-3.77***</b>	-0.005 [-0.012, 0.002]	-1.6 10 <sup>-5</sup> ± 1.1 10 <sup>-5</sup>	-1.45	<b>-1.5 10<sup>-5</sup> [-2.9 10<sup>-5</sup>, -0.2 10<sup>-5</sup>]</b>
Phanera presence ((S,HS,H)>N)	<b>-8.68 ± 3.43</b>	<b>-2.53*</b>	<b>-5.8 [-10.45, -0.93]</b>	-0.009 ± 0.006	-1.38	<b>-0.013 [-0.02, -0.005]</b>
Phanera type (S,HS)>(H,N)	<b>-10 ± 3.37</b>	<b>-2.96**</b>	<b>-9.7 [-15.56, -3.46]</b>	-	-	-
Insertion (E>(F,U))	<b>10.26 ± 2.44</b>	<b>4.2***</b>	<b>10.28 [6.79, 13.73]</b>	<b>0.024 ± 0.006</b>	<b>4.17***</b>	<b>0.03 [0.023, 0.036]</b>
Proportion of clearwing area	<b>0.16 ± 0.04</b>	<b>4.63***</b>	<b>0.17 [0.13, 0.21]</b>	<b>0.0003 ± 0.00007</b>	<b>4.94***</b>	<b>0.0003 [0.0003, 0.0004]</b>
Colouration (T>(C,U))	<b>8.07 ± 3.14</b>	<b>2.57*</b>	<b>15.7 [11, 20.62]</b>	0.01 ± 0.007	1.35	<b>0.022 [0.013, 0.031]</b>
Phylogenetic variance	-	-	812.19 [607.22, 1082.99]	-	-	0.007 [0.005, 0.009]
Residual variance	-	-	62.86 [57.59, 68.61]	-	-	0.0002 [0.0002, 0.0002]

Retained classic mixed model and Bayesian phylogenetically controlled mixed model for mean transmittance over 300-700 nm and proportion of UV transmittance. For all analyses, we took all 10 measurements per species. We took species and wing within species as random factors in classic mixed models and species as random factor in Bayesian models. The symbol (-) indicated factors not retained in the retained model or of no concern (for phylogenetic and residual variances). FW=forewing, HW=Hindwing, phanera type (S=scale, HS=scale and hair-like scale, H=hair-like scale, N=none), phanera insertion on the wing membrane (U=unknown (for absent phanera), F=flat, E=erected), and phanera colouration (T=transparent, C=coloured). All factors relating to phanera concerned phanera from the transparent zone. Bold values are significant factors (\* p<0.05; \*\* p<0.01; \*\*\* p<0.001 for the mixed model and significance for 95%CI not including zero for Bayesian models).

**Table 2. Variations in the proportion of clearwing area, as explained by macrostructural and microstructural wing features.**

Factor	Proportion of clearwing area		
	Mixed model	Bayesian	
	Estimate ± se	t-value	Estimate [CI95%]
intercept	<b>57.46 ± 3.93</b>	<b>14.62***</b>	<b>61.55 [43.31, 80.41]</b>
(HW>FW)	4.26±3.29	1.29	4.45[-1.8;10.99]
Colouration (U>C)	7.9±4.62	1.71~	5.3[-3.73;14.15]
Colouration (T>C)	<b>-18.78±4.8</b>	<b>-3.91***</b>	<b>-20.1[-29.79;-10.05]</b>
Clearwing area	<b>0.19±0.02</b>	<b>11.73***</b>	<b>0.21[0.17;0.24]</b>
Wing length	<b>-1.27±0.12</b>	<b>-10.34***</b>	<b>-1.07[-1.33;-0.82]</b>
(HW>FW) x Colouration (U>C)	<b>-11.53±3.95</b>	<b>-2.92**</b>	<b>-9.46[-17.29;-1.54]</b>
(HW>FW) x Colouration (T>C)	6.06±3.48	1.74~	6.03[-0.81;12.75]
(HW>FW) x Clearwing area	<b>0.05±0.02</b>	<b>2.27*</b>	<b>0.05[0.01;0.09]</b>
Colour (U>C) x Clearwing area	0.01±0.02	0.39	0.01[-0.04;0.06]
Colour (T>C) x Clearwing area	<b>0.12±0.05</b>	<b>2.35*</b>	<b>0.11[0.02;0.2]</b>
(HW>FW) x Wing length	<b>-0.47±0.13</b>	<b>-3.52***</b>	<b>-0.41[-0.68;-0.14]</b>
Phylogenetic variance	-	-	394.72 [258.62, 566.48]
Residual variance	-	-	92.69 [70.22, 120.99]

Retained classic mixed model and Bayesian phylogenetically controlled mixed model for the proportion of surface occupied by the clearwing area. FW=forewing, HW=Hindwing, phanera colouration (T=transparent, C=coloured, U=unknown (for absent phanera)). Bold values are significant factors (\* p<0.05; \*\* p<0.01; \*\*\*\* p<0.001 for the mixed model and significance for 95%CI not including zero for Bayesian models).

**Table 3. Structural effort in transparency as change in structural traits between zones.**

Variable	Factors	Mixed model		Bayesian
		Estimate $\pm$ se	t-value	Estimate [CI95%]
Phanera density in the transparent zone – Phanera density in the opaque zone	Intercept	<b>-494.11 <math>\pm</math> 26.56</b>	<b>-18.6***</b>	<b>-471.77 [-597.7, -340.97]</b>
	OZ Phanera surface	<b>0.019 <math>\pm</math> 0.003</b>	<b>7.1***</b>	<b>0.02 [0.01, 0.02]</b>
	Phanera density	<b>0.82 <math>\pm</math> 0.06</b>	<b>14.65***</b>	<b>0.77 [0.65, 0.89]</b>
	Phanera (S,HS)>(H,N)	<b>58.46 <math>\pm</math> 25.9</b>	<b>2.26*</b>	49.73 [-5.37, 104.53]
	Clearwing area	<b>0.36 <math>\pm</math> 0.12</b>	<b>3.12**</b>	<b>0.32 [0.06, 0.59]</b>
	Phylogenetic variance	–	–	17362.31 [5613.06, 31707.7]
	Residual variance	–	–	18117.56 [13844.73, 23842.22]
	Intercept	7.77 $\pm$ 10.52	0.74	7.12 [-21.53, 33.5]
Phanera length in the transparent zone – Phanera length in the opaque zone	OZ Phanera surface	<b>-0.0048 <math>\pm</math> 0.0009</b>	<b>-5.35***</b>	<b>-0.005 [-0.007, -0.003]</b>
	Phanera density	<b>-0.05 <math>\pm</math> 0.02</b>	<b>-2.3*</b>	<b>-0.044 [-0.083, -0.003]</b>
	Insertion (F>E)	17.14 $\pm$ 9.2	1.86~	<b>19.33 [2.13, 37.49]</b>
	Insertion (U>E)	<b>-115.05 <math>\pm</math> 12.32</b>	<b>-9.34***</b>	<b>-109.68 [-135.8, -83.62]</b>
	Phylogenetic variance	–	–	381.02 [0.003, 1494.31]
	Residual variance	–	–	3053.07 [2393.11, 3769.69]
	Intercept	<b>-13.18 <math>\pm</math> 4.23</b>	<b>-3.11**</b>	-3.18 [-22.82, 17.19]
	OZ Phanera surface	<b>-0.0027 <math>\pm</math> 0.0003</b>	<b>-8.39***</b>	<b>-0.003 [-0.004, -0.002]</b>
Phanera width in the transparent zone – Phanera width in the opaque zone	Phanera density	<b>-0.02 <math>\pm</math> 0.007</b>	<b>-2.09*</b>	<b>-0.021 [-0.037, -0.005]</b>
	Colouration (T>(C,U))	<b>27.31 <math>\pm</math> 4.26</b>	<b>6.42***</b>	<b>22.64 [14.14, 31.05]</b>
	Phanera (N>H)	<b>-13.22 <math>\pm</math> 4.6</b>	<b>-2.87**</b>	<b>-20.78 [-30.63, -11.02]</b>
	Phanera ((S,HS)>H)	<b>21.46 <math>\pm</math> 4.27</b>	<b>5.02***</b>	<b>22.07 [13.54, 30.92]</b>
	Phylogenetic variance	–	–	410.31 [198.06, 678.99]
	Residual variance	–	–	194.99 [143.11, 258.68]
	Intercept	1036.99 $\pm$ 638.85	1.62	2102.84 [-437.54, 4560.3]
	OZ Phanera surface	<b>-0.66 <math>\pm</math> 0.05</b>	<b>-14.51***</b>	<b>-0.69 [-0.79, -0.59]</b>
Phanera surface in the transparent zone – Phanera surface in the opaque zone	Phanera density	<b>-3.79 <math>\pm</math> 0.95</b>	<b>-3.97***</b>	<b>-4.35 [-6.39, -2.22]</b>
	Wing Size	<b>-36.56 <math>\pm</math> 14.23</b>	<b>-2.57*</b>	<b>-35.83 [-67.77, -3.57]</b>
	Colouration (T>(C,U))	<b>2314.51 <math>\pm</math> 544.8</b>	<b>4.25***</b>	<b>2540.09 [1450.64, 3629.96]</b>
	Phanera (N>H)	<b>-2409.19 <math>\pm</math> 603.25</b>	<b>-3.99***</b>	<b>-3060.3 [-4326.08, -1791.24]</b>
	Phanera ((S,HS)>H)	<b>2847.9 <math>\pm</math> 545.65</b>	<b>5.22***</b>	<b>2495.06 [1366.67, 3644.65]</b>
	Phylogenetic variance	–	–	$6.4 \cdot 10^6$ [ $3 \cdot 10^6$ , $1.1 \cdot 10^7$ ]
	Residual variance	–	–	$3.4 \cdot 10^6$ [ $2.5 \cdot 10^6$ , $4.6 \cdot 10^6$ ]
	Intercept	<b>-245441 <math>\pm</math> 106767</b>	<b>-2.3*</b>	-25356 [-202223, 154055]
Phanera coverage in the transparent zone – Phanera coverage in the opaque zone	OZ Phanera surface	<b>-0.8 <math>\pm</math> 0.06</b>	<b>-12.76***</b>	<b>-0.77 [-0.9, -0.64]</b>
	Phanera density	<b>1707.7 <math>\pm</math> 212.4</b>	<b>8.04***</b>	<b>1959.47 [1502, 2425.54]</b>
	Colouration (T>(C,U))	<b>1043405 <math>\pm</math> 122752</b>	<b>8.5***</b>	<b>314507 [146461, 481013]</b>
	Phylogenetic variance	–	–	$2.9 \cdot 10^{11}$ [ $1.3 \cdot 10^{11}$ , $4.9 \cdot 10^{11}$ ]
	Residual variance	–	–	$3.4 \cdot 10^{11}$ [ $2.1 \cdot 10^{11}$ , $4.3 \cdot 10^{11}$ ]

Retained classic mixed model and Bayesian phylogenetically controlled mixed model. All factors relating to phanera concerned phanera from the transparent zone, except when stated otherwise. OZ=opaque zone, FW=forewing, HW=Hindwing, phanera type (H=hair-like scale, S=scale, HS=scale and hair-like scale, N=none), insertion on the wing membrane (F=flat, E=erected, U=unknown (for absent phanera)), and phanera colouration (T=transparent, C=coloured, U=unknown (for absent phanera)). Bold values are significant factors (\* p<0.05; \*\* p<0.01; \*\*\* p<0.001 for the mixed model and significance for 95%CI not including zero for Bayesian models).

**Table 4. Relationships between optical parameters.**

Variable	Factor	Mixed model		Bayesian
		Estimate ± se	t-value	Estimate [CI95%]
Mean transmittance	intercept	<b>7.05 ± 2.75</b>	<b>2.57*</b>	-1.37 [-30.57, 29.13]
	Proportion of UV transmittance (HW>FW)	<b>280.24 ± 13.27</b>	<b>21.12***</b>	<b>322.99 [297.69, 347.33]</b>
	Phylogenetic variance	<b>-1.78 ± 0.68</b>	<b>-2.61*</b>	<b>-1.72 [-2.47, -0.97]</b>
	Residual variance	-	-	1139.83 [869.43, 1492.36]
Mean transmittance	intercept	<b>84.62 ± 0.75</b>	<b>113.39***</b>	<b>83.03 [74.92, 90.82]</b>
	Colour contrast	<b>-6.12 ± 0.25</b>	<b>-24.36***</b>	<b>-6.27 [-6.74, -5.81]</b>
	Brightness contrast (HW>FW)	<b>-8.59 ± 0.15</b>	<b>-59.15***</b>	<b>-8.09 [-8.35, -7.82]</b>
	Colour contrast x Brightness contrast	<b>0.85 ± 0.04</b>	<b>23.23***</b>	<b>0.8 [0.74, 0.87]</b>
	Colour contrast x (HW>FW)	<b>-0.39 ± 0.15</b>	<b>-2.66**</b>	<b>-0.26 [-0.43, -0.09]</b>
	Brightness contrast x (HW>FW)	<b>0.88 ± 0.12</b>	<b>7.18***</b>	<b>0.56 [0.39, 0.72]</b>
	Phylogenetic variance	-	-	88.47 [66.32, 117.3]
	Residual variance	-	-	8.64 [7.93, 9.43]
Proportion of UV transmittance	intercept	<b>0.23 ± 0.002</b>	<b>110.33***</b>	<b>0.23 [0.2, 0.26]</b>
	Colour contrast	<b>-0.03 ± 0.0005</b>	<b>-59.72***</b>	<b>-0.03 [-0.03, -0.03]</b>
	Brightness contrast	<b>-0.0035 ± 0.0003</b>	<b>-13.95***</b>	<b>-0.004 [-0.005, -0.004]</b>
	Colour contrast x Brightness contrast	<b>0.0013 ± 0.0001</b>	<b>17.52***</b>	<b>0.001 [0.001, 0.001]</b>
	Phylogenetic variance	-	-	0.001 [0.001, 0.002]
	Residual variance	-	-	0.00004 [0.00004, 0.00004]

Retained classic mixed model and Bayesian phylogenetically controlled mixed model for the relationships between physically and biologically relevant descriptors of transparency. For all analyses, we took all 10 measurements per species. We took species and wing within species as random factors in classic mixed models and species as random factor in Bayesian models. FW=forewing, HW=Hindwing. Bold values are significant factors (\* p<0.05; \*\* p<0.01; \*\*\* p<0.001 for the mixed model and significance for 95%CI not including zero for Bayesian models).

**Table 5. Variations of transmittance with nocturnality.**

Factor	Mean transmittance over [300-700] nm		
	Mixed model		Bayesian Estimate [CI95%]
	Estimate ± se	t-value	
intercept	<b>61.51 ± 5.8</b>	<b>10.6***</b>	<b>69.69 [39.01, 98.88]</b>
Noc>Diu	<b>-16.91 ± 8.42</b>	<b>-2.01*</b>	<b>-28.09 [-41.63, -14.11]</b>
Wing length	<b>-0.56 ± 0.14</b>	<b>-4.13***</b>	<b>-0.6 [-0.76, -0.43]</b>
HW>FW	<b>-14.97 ± 3.44</b>	<b>-4.35***</b>	<b>-15.63 [-19.17, -12]</b>
Prop clearwing area	<b>0.27 ± 0.07</b>	<b>3.95***</b>	<b>0.25 [0.18, 0.33]</b>
(Noc>Diu) x Wing length	<b>0.64 ± 0.2</b>	<b>3.17**</b>	<b>0.51 [0.18, 0.83]</b>
(Noc>Diu) x (HW>FW)	<b>18.9 ± 4.93</b>	<b>3.83***</b>	<b>16.78 [10.83, 22.8]</b>
(Noc>Diu) x Prop clearwing area	-0.09 ± 0.1	-0.93	-0.07 [-0.19, 0.04]
(HW>FW): Prop clearwing area	0.12 ± 0.07	1.68~	<b>0.13 [0.06, 0.2]</b>
(Noc>Diu) x (HW>FW) x Prop clearwing area	<b>-0.2 ± 0.09</b>	<b>-2.12*</b>	<b>-0.18 [-0.27, -0.08]</b>
Phylogenetic variance	-	-	1073.05 [810.05, 1423.09]
Residual variance	-	-	70.21 [64.14, 76.81]

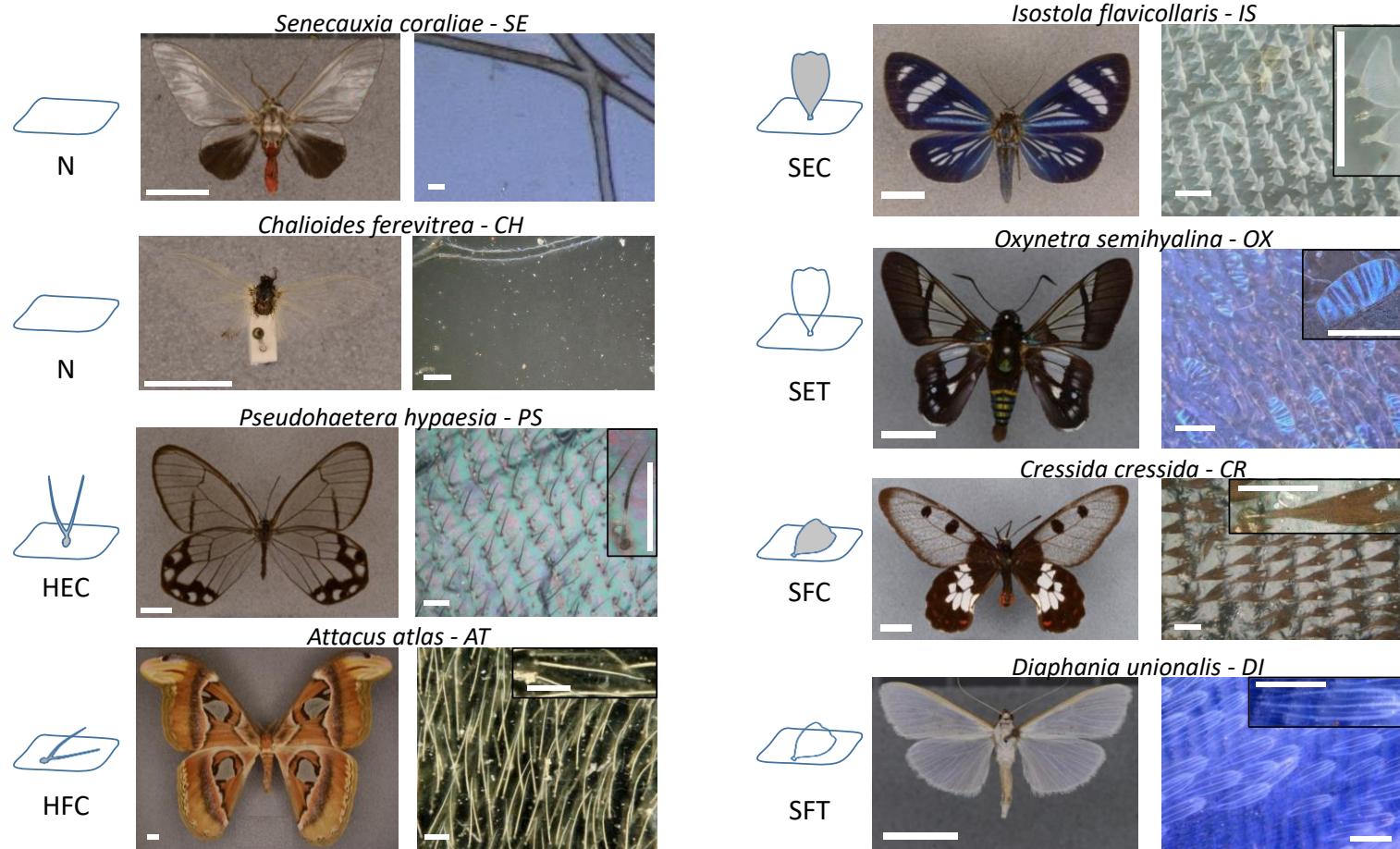
Retained classic mixed model and Bayesian phylogenetically controlled mixed model for the relationship between mean transmittance over 300-700 nm and nocturnality. For all analyses, we took all 10 measurements per species. We took species and wing within species as random factors in classic mixed models and species as random factor in Bayesian models. FW=forewing, HW=Hindwing, Noc= nocturnal, Diu=diurnal. Species active during both night and day were excluded from the model. (\* p<0.05; \*\* p<0.01; \*\*\* p<0.001 for the mixed model and significance for 95%CI not including zero for Bayesian models). Bold values are significant factors.

**Table 6. Variations of transmittance with latitude.**

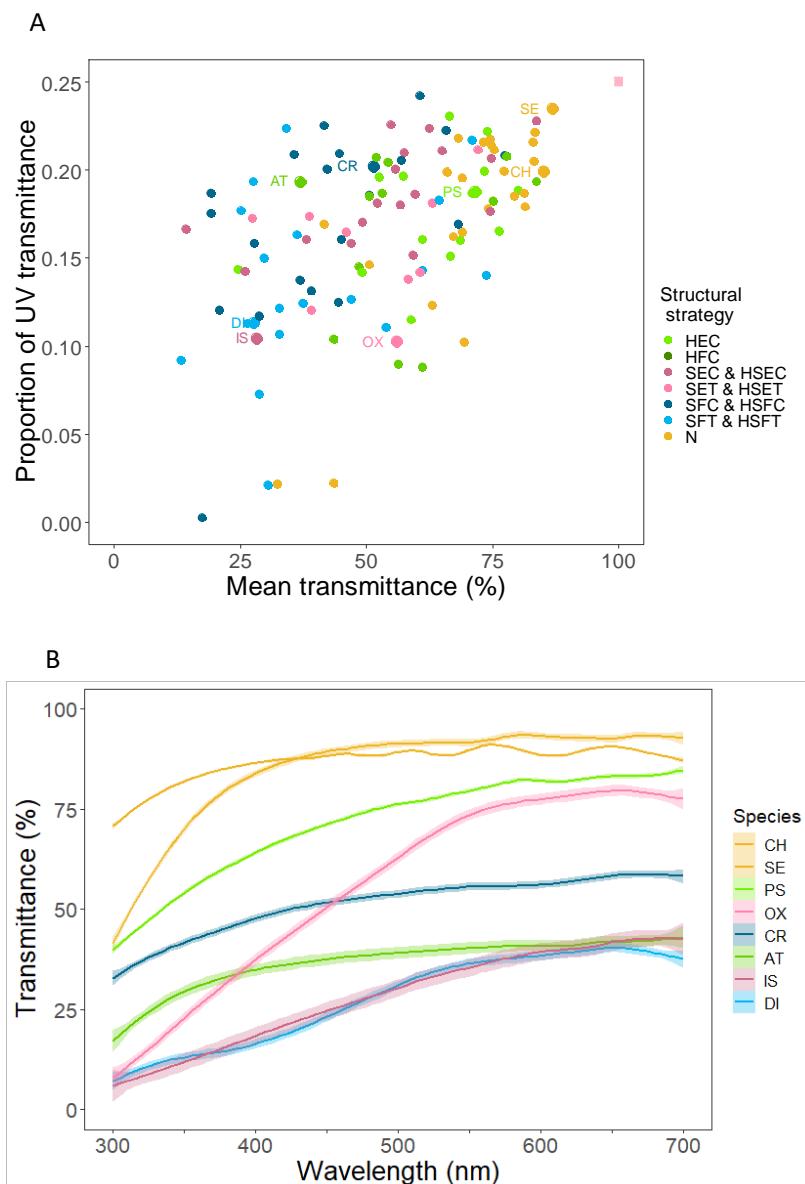
		Mixed model		Bayesian	
		Factor	Estimate ± se	t-value	Estimate [CI95%]
Proportional UV transmittance	intercept	0.17 ± 0.01	<b>23.25***</b>	<b>0.19 [0.11, 0.26]</b>	
	Abs(Latitude)	-0.0005 ± 0.0004	-1.14	-0.0008 [-0.0019, 0.0003]	
	Phylogenetic variance	-	-	0.01 [0.01, 0.01]	
	Residual variance	-	-	0.0002 [0.0002, 0.0003]	
Mean transmittance over [300-400] nm	intercept	40.41 ± 3.01	<b>13.43***</b>	<b>23.12 [0.12, 70.32]</b>	
	Abs(Latitude)	-0.4 ± 0.18	<b>-2.21*</b>	-0.28 [-0.91, 0.0001]	
	Phylogenetic variance	-	-	524.02 [0.01, 1357.47]	
	Residual variance	-	-	24.33 [0.0002, 53.07]	
Mean transmittance over [400-700] nm	intercept	62.54 ± 3.19	<b>19.59***</b>	<b>67.45 [41.05, 93.82]</b>	
	Abs(Latitude)	-0.51 ± 0.19	<b>-2.67**</b>	<b>-0.68 [-1.06, -0.31]</b>	
	Phylogenetic variance	-	-	889.34 [645.12, 1211.22]	
	Residual variance	-	-	68.87 [62.2, 76.42]	
Mean transmittance over [700-1100] nm	intercept	69.75 ± 3.05	<b>22.9***</b>	<b>73.58 [47.41, 99.32]</b>	
	Abs(Latitude)	-0.41 ± 0.18	<b>-2.25*</b>	<b>-0.57 [-0.96, -0.2]</b>	
	Phylogenetic variance	-	-	875.84 [640.03, 1195.06]	
	Residual variance	-	-	75.81 [68.15, 83.62]	

Retained classic mixed model and Bayesian phylogenetically controlled mixed model for the relationship between optics, namely proportional UV transmittance, mean transmittance in the UV, in the human visible range, and in the near infrared range, and fixed factors, namely absolute value of latitude and wing length. For all analyses, we took all 10 measurements per species. We took species and wing within species as random factors in classic mixed models and species as random factor in Bayesian models. (\* p<0.05; \*\* p<0.01; \*\*\* p<0.001 for the mixed model and significance for 95%CI not including zero for Bayesian models). Bold values are significant factors.

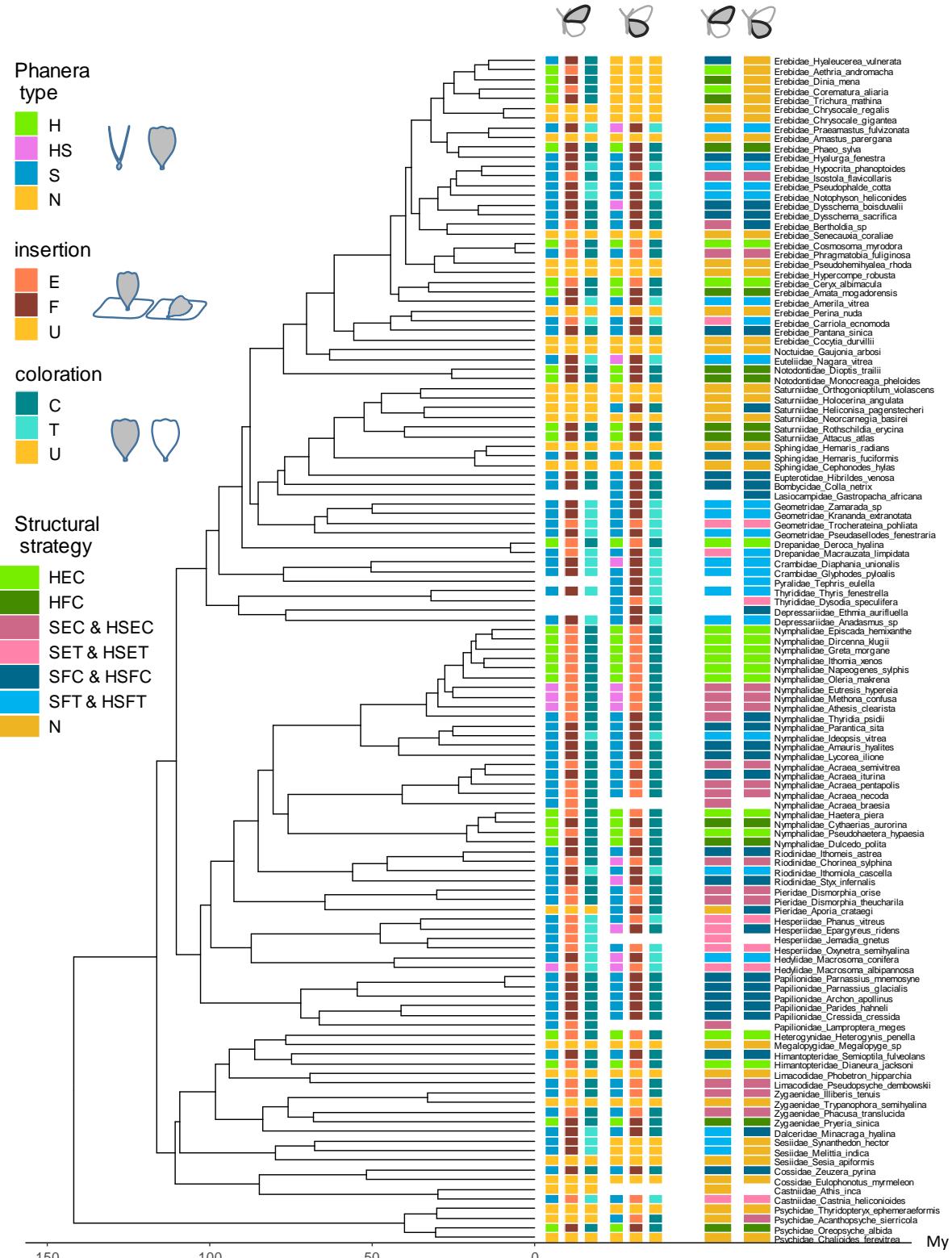
**Figure 1.** Examples of structural strategies in Lepidoptera. A structural strategy is defined as the combination of phanera type, insertion and colouration. Type is H=hair-like scales, S=scales (hair-like scales and scales were assimilated to scales), insertion is E=erected, F=flat and colouration is C=coloured, T=transparent. The N strategy has no phanera, no insertion and no colouration. Bar scales are 1cm for entire specimens (left columns), 100  $\mu$ m for microscopic imaging (right columns and phanera details). Species are the erebid *Senecauxia coraliae* (SE), the psychid *Chalioides ferevitrea* (CH), the nymphalid *Pseudohaetera hypaesia* (PS), the saturnid *Attacus atlas* (AT), the erebid *Isostola flavigollaris* (IS), the hesperid *Oxynetra semihyalina* (OX), the papilionid *Cressida cressida* (CR) and the crambid *Diaphania unionalis* (DI).



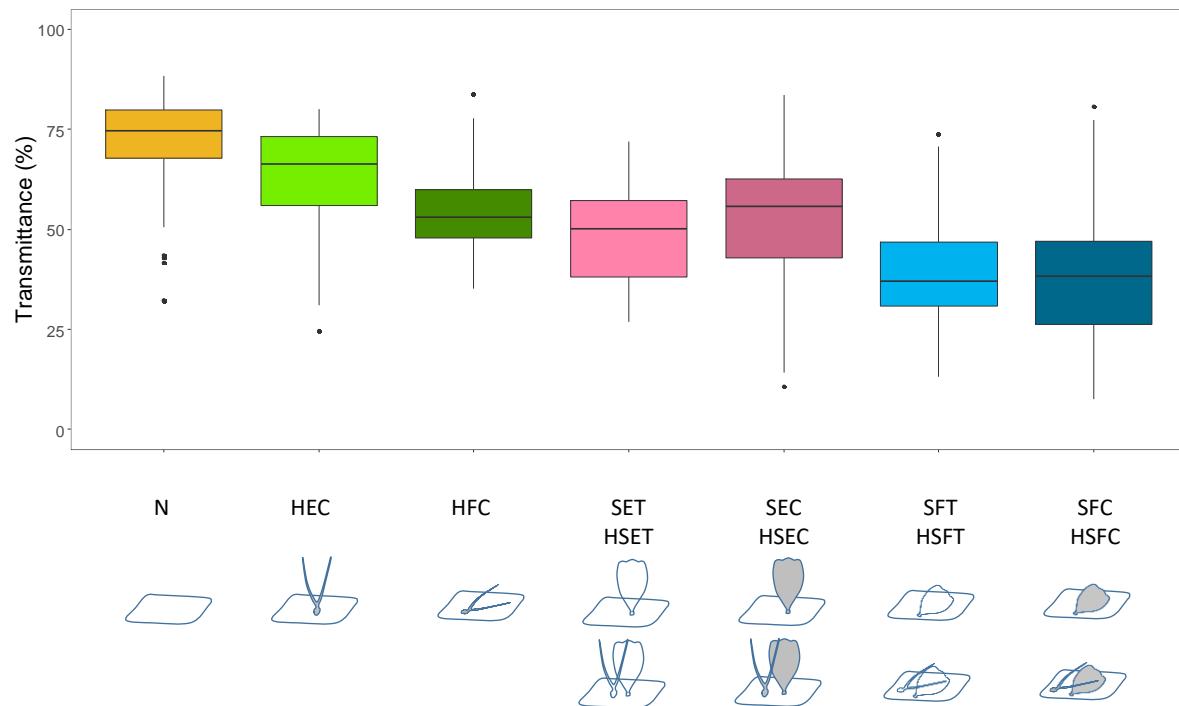
**Figure 2.** (A) Relationship between the proportion of UV transmittance and the mean transmittance over 300-700 nm. Species are represented by their forewing value. Perfect transparency (100% transmittance over 300-700 nm, resulting in a 0.25 proportion of UV transmittance, upper right corner) is represented by a pink square and species examples from Figure 1 are indicated by their two-letter code inside the plot. (B) Transmittance spectra of the 5 points of the forewing for the species listed in Figure 1, mean and standard error. Names are ordered from up to down according to decreasing transmittance values at 700nm. A structural strategy is defined as the combination of phanera type, insertion and colouration. Type is H=hair-like scales, S=scales (hair-like scales and scales were assimilated to scales), insertion is E=erected, F=flat and colouration is C=coloured, T=transparent. The N strategy has no phanera, no insertion and no colouration.



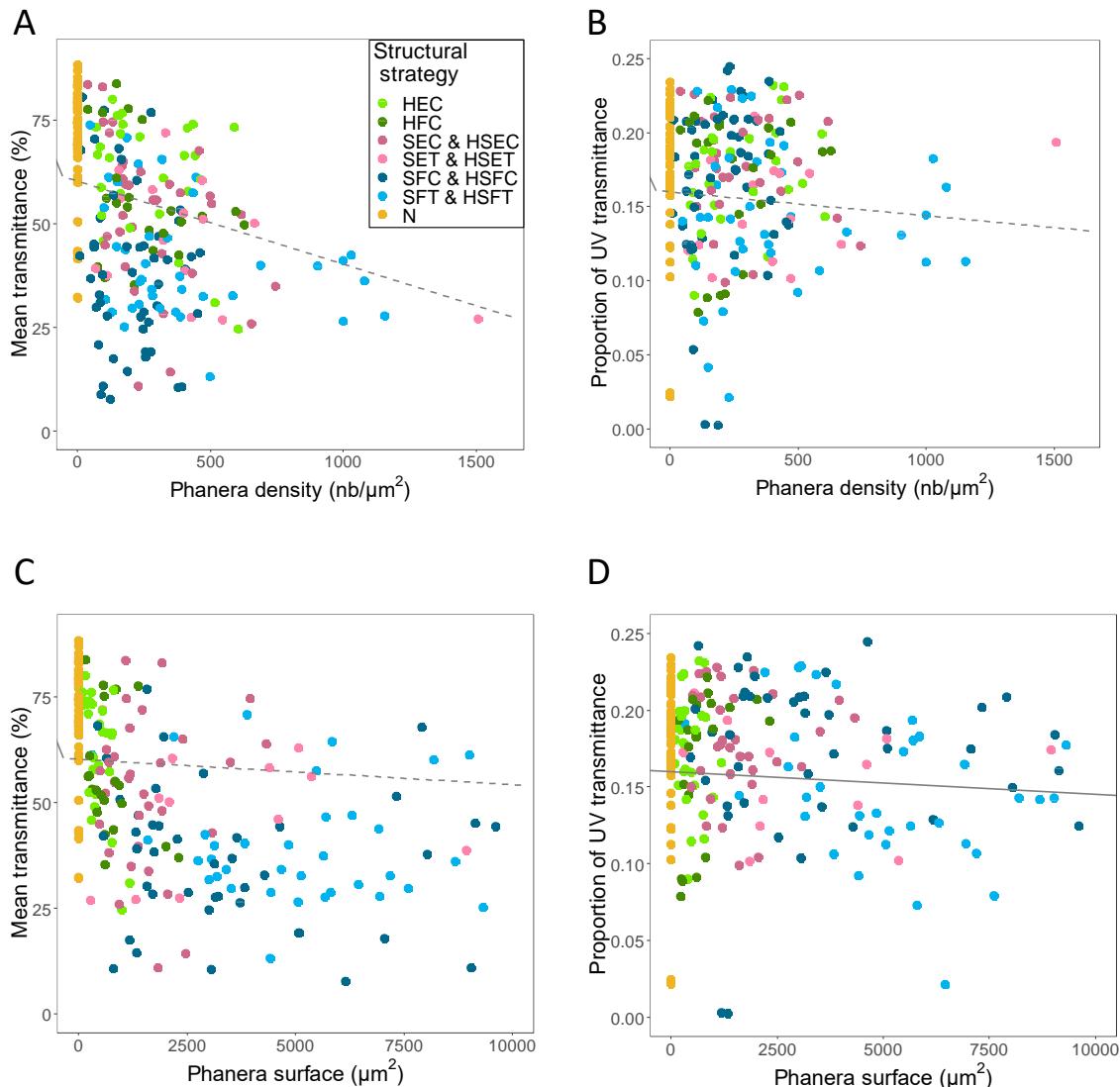
**Figure 3.** Distribution of phanera structural traits and structural strategies in the study species for the forewing (columns 1,2,3,7) and the hindwing (columns 4,5,6,8). A structural strategy is defined as the combination of phanera type, insertion and colouration. Type is H=hair-like scales, S=scales (hair-like scales were assimilated to scales), N=no phanera. Insertion is E=erected, F=flat, and U=unknown (for N strategy). Colouration is C=coloured, T=transparent, and U=unknown (for N strategy). The N strategy has no phanera, no insertion and no colouration.



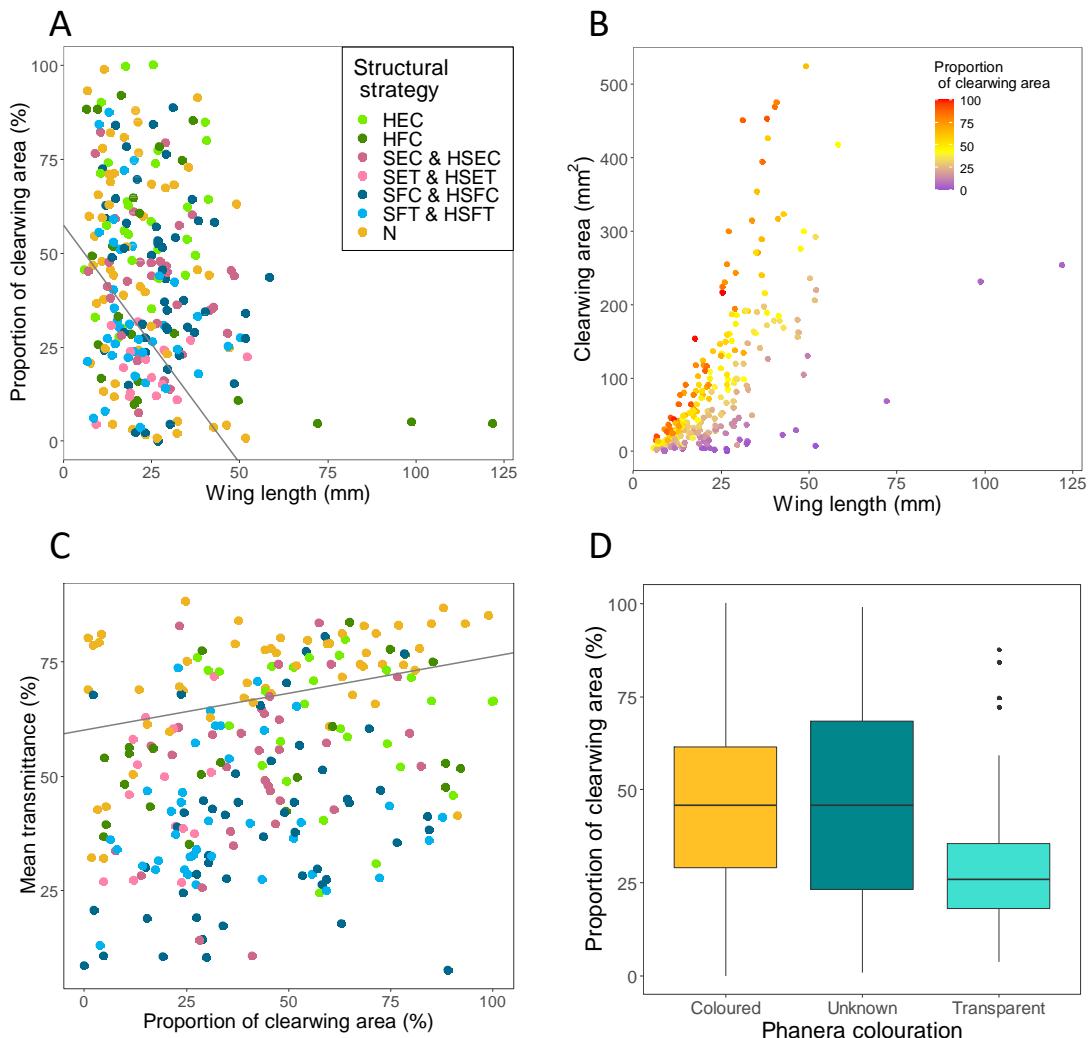
**Figure 4.** Variations of transparency (mean transmittance) with structural strategies. A structural strategy is defined as the combination of phanera type, insertion and colouration. A structural strategy is defined as the combination of phanera type, insertion and colouration. Type is H=hair-like scales, S=scales (hair-like scales and scales were assimilated to scales), insertion is E=erected, F=flat and colouration is C=coloured, T=transparent. The N strategy has no phanera, no insertion and no colouration. The categories mixing scales and hair-like scales HS were assimilated to the corresponding strategies with scales S with same insertion and colouration. Mean transmittance was computed over [300-700] nm, for the graph, we took one average per wing and species.



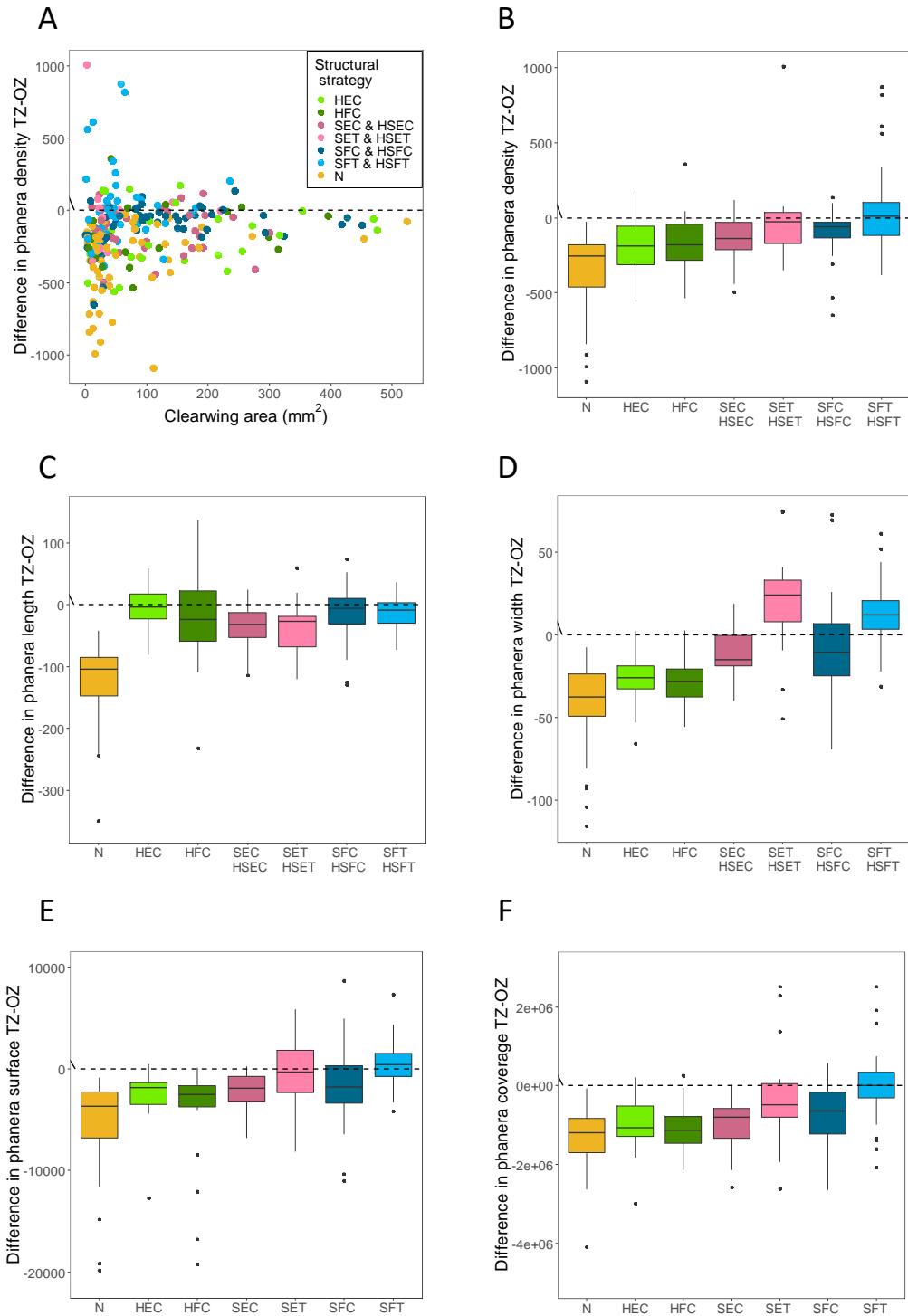
**Figure 5.** Relationship between optical transparency, estimated by mean transmittance over 300-700 nm (A, C) and proportion of UV transmittance (B,D), and phanera density (A,B) or phanera surface (C,D). Lines were drawn from the coefficients of the best classic mixed model when the factor was significant in both classic and Bayesian models (plain line) or in only one of the models (dashed line). Species are represented by two values, one for forewing and one for hindwing. In (C,D) two points were removed for clarity reasons from the graph but not from the analyses. A structural strategy is defined as the combination of phanera type, insertion and colouration. Type is H=hair-like scales, S=scales (hair-like scales and scales were assimilated to scales), insertion is E=erected, F=flat and colouration is C=coloured, T=transparent. The N strategy has no phanera, no insertion and no colouration.



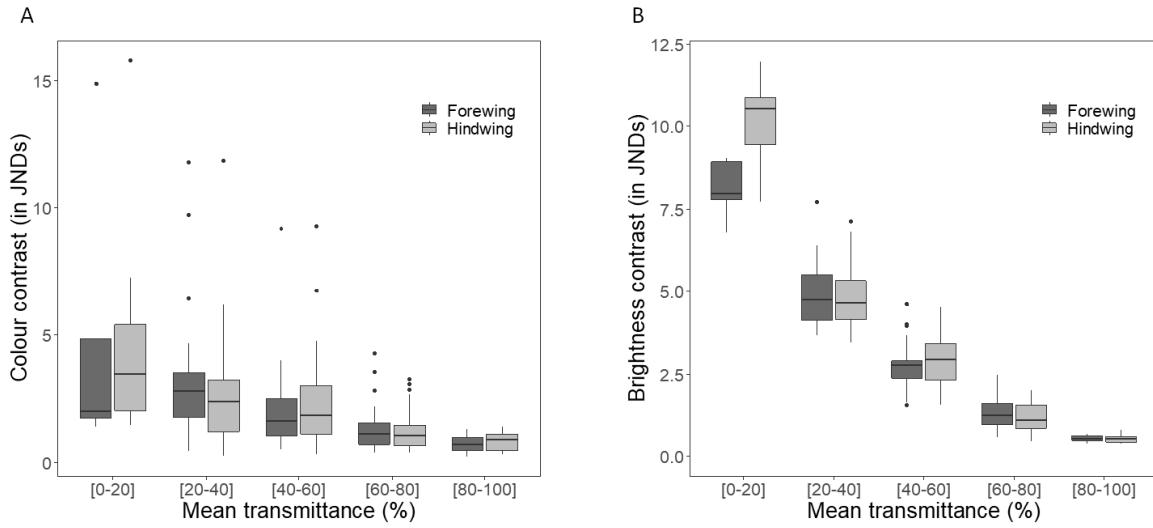
**Figure 6.** Mean transmittance over [300-700] nm in relation to the proportion of clearwing area and structural strategies (A). Proportion of clearwing area in relation to wing length and structural strategies (B), to wing total area and structural strategies (C), and to phanera colour (D). A structural strategy is defined as the combination of phanera type, insertion and colouration. Type is H=hair-like scales, S=scales (hair-like scales and scales were assimilated to scales), insertion is E=erected, F=flat and colouration is C=coloured, T=transparent, and U=unknown (for N strategy). The N strategy has no phanera, no insertion and no colouration. Structural strategies are described in plot B and the same as in Figure 6. Species are represented by two values, one for forewing and one for hindwing. In (C), two outliers for wing area were removed for clarity reasons from the plot but not from analyses.



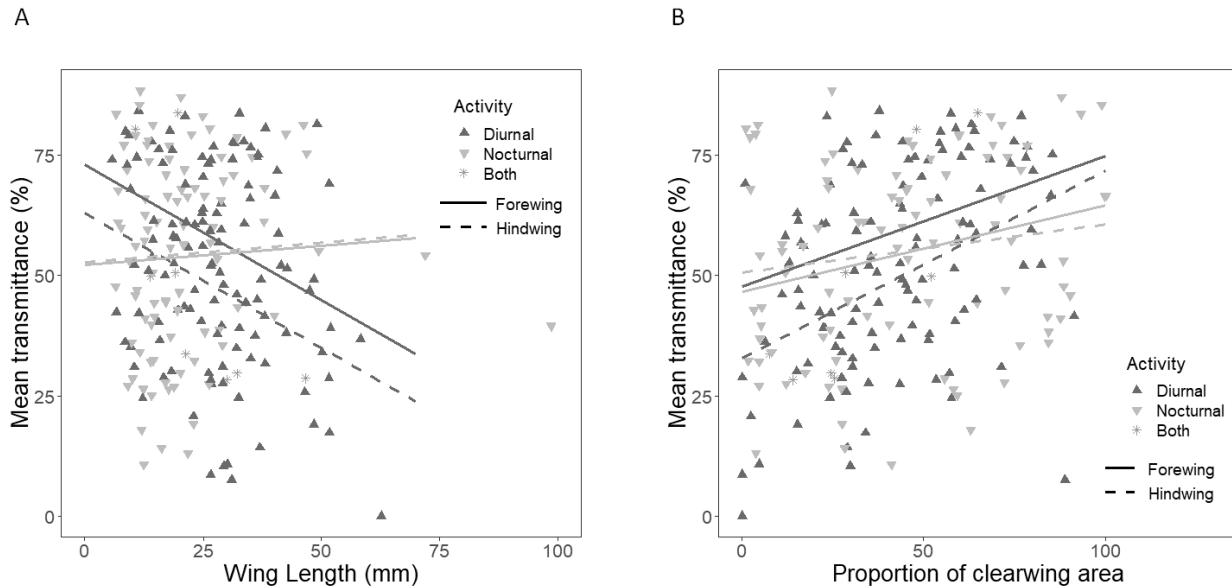
**Figure 7.** Structural effort in transparency measuring to what extent phanera structural features in the transparent zone are modified relatively to the opaque zone. Plots detail the relationships between wing clearwing area and difference in phanera density (A), between structural strategy and difference in phanera density (B), length (C), width (D), surface (E) and coverage (surface \* density) (F). A structural strategy is defined as the combination of phanera type, insertion and colouration. Type is H=hair-like scales, S=scales (hair-like scales and scales were assimilated to scales), insertion is E=erected, F=flat and colouration is C=coloured, T=transparent. The N strategy has no phanera, no insertion and no colouration. A few outliers were withdrawn from plots for clarity reasons but not from analyses.



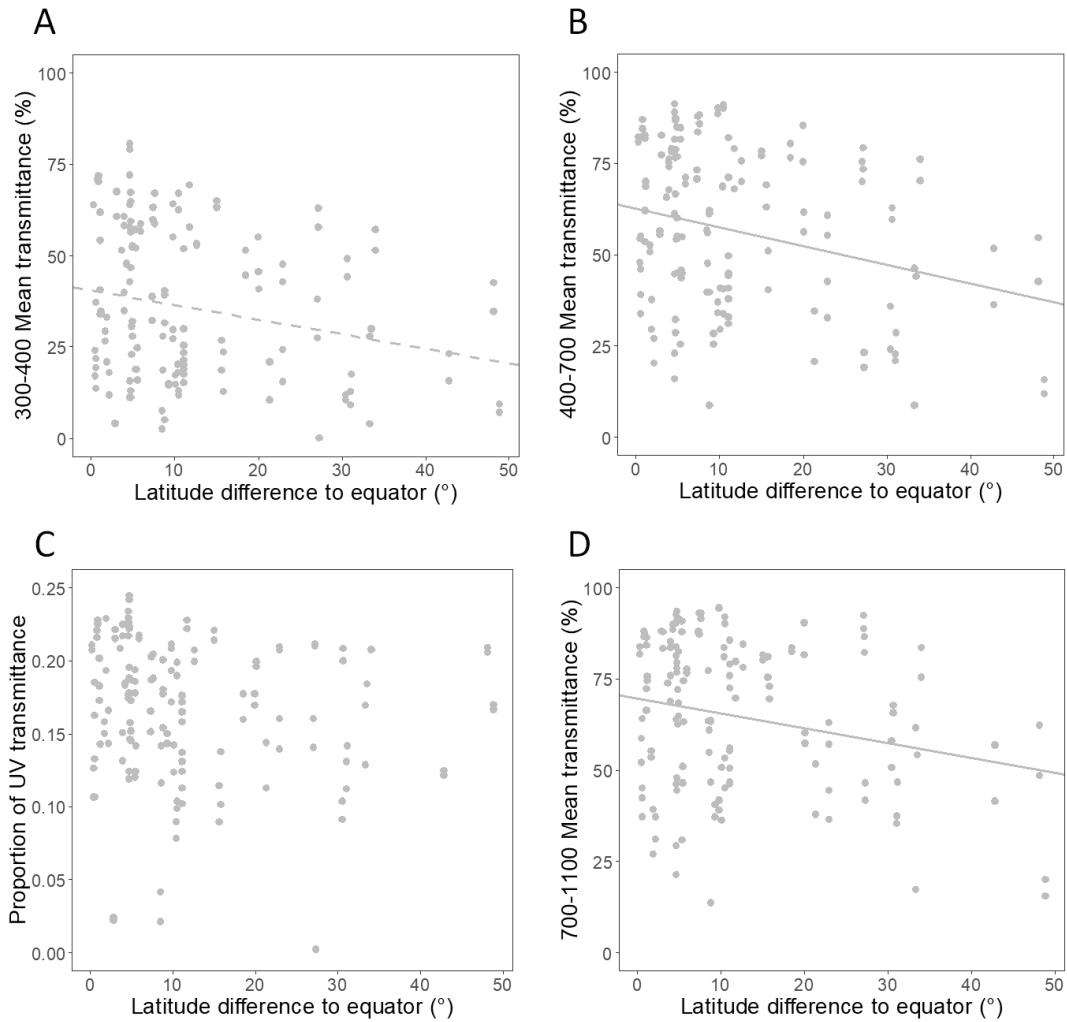
**Figure 8.** Relationships between transmittance and colour (A) or brightness (B) contrasts, for the forewing (dark grey) and the hindwing (light grey). Colour and brightness contrast values were obtained from vision modelling (see methods for details) and reflected the contrast offered by a butterfly seen against a green vegetation by a bird predator. The attenuation of the decrease with the increase in transmittance can be seen in mixed models.



**Figure 9.** Variations of mean transmittance with wing length (A) and proportion occupied by the clearwing zone on the wing (B) in clearwing Lepidoptera showing activity during day, night or both. Lines were drawn from the coefficients of the best classic mixed model computed excluding species that were both diurnal and nocturnal, for the mean values found in the population for the factor not presented (clearwing proportion in A, wing length in B). For clarity reasons, we plotted only two points per species, forewing and hindwing mean transmittance values but models were run on all ten measurements acquired per species. Two outliers (for *Attacus atlas*) were removed from plot (A) but not from analyses.



**Figure 10.** Variation in transmittance in the ultraviolet 300-400 nm range (A), the 400-700 nm range (B), in the near infrared 700-1100 nm range (D) and variation in proportion of UV transmittance (C) with the difference in latitude to equator (in degrees) of where the specimen was collected. Lines were drawn from the coefficients of the best classic mixed model when the factor was significant in both classic and Bayesian models (plain line) or in only one of the models (dashed line). For (C) for which the null model was the best model. For clarity reasons, we plotted only two points per species, forewing and hindwing mean transmittance values but models were run on all ten measurements acquired per species.



## APPENDICES

**Table S1. Supplementary file with species names**

**Table S2. Repeatability of variables computed from colour and structure measurements.**

Variable	Nb species/measures	Level	R ( $\pm$ se)
Mean transmittance over [300-400]	123 / 1200	SpWg	0.907 ( $\pm$ 0.009) ***
Mean transmittance over [400-700]	123 / 1200	SpWg	0.903 ( $\pm$ 0.009) ***
Mean transmittance over [700-1100]	123 / 1200	SpWg	0.886 ( $\pm$ 0.011) ***
Mean transmittance	123 / 1200	SpWg	0.893 ( $\pm$ 0.01) ***
Proportion of UV transmittance	123 / 1200	SpWg	0.883 ( $\pm$ 0.011) ***
Colour contrast	123 / 1200	SpWg	0.963 ( $\pm$ 0.004) ***
Brightness contrast	123 / 1200	SpWg	0.889 ( $\pm$ 0.011) ***
Wing length	123 / 492	SpWg	0.999 ( $\pm$ <0.01) ***
Phanera density	30 / 60	SpWgZoPha	0.974 ( $\pm$ 0.011) ***
Phanera width	51 / 168	SpWgZoPha	0.874 ( $\pm$ 0.028) ***
Phanera length	51 / 168	SpWgZoPha	0.651 ( $\pm$ 0.063) ***
Phanera surface	51 / 168	SpWgZoPha	0.899 ( $\pm$ 0.021) ***

For each variable, we specify the number of species measured and the total number of measurements included in the repeatability analysis. For each analysis the level of relevance for the measurement depends on the variable. For colour and wing length, measurements relate to a specific wing (SpWg), while for phanera density and dimensions, measurements relate to a specific wing, zone, phanera (SpWgZoPha) defined by its shape, insertion and colouration. If not specified otherwise, mean transmittance was measured across the 300-700 nm range. We specify the value of repeatability R and the standard error associated *se*, as well as the significance level (\*\*\* stands for p-value <0.001).

**Table S3. Sample sizes for each structural category.**

type+insertion+colouration		type+insertion		type	
Species number	Family number	Species number	Family number	Species number	Family number
HEC	15	5	HEC	15	5
HFC	12	6	HFC	12	6
HSEC	4	2	HSE	5	3
HSET	1	1			
HSFC	3	3	HSF	7	6
HSFT	4	4			
SEC	16	8	SE	25	13
SET	9	6			
SFC	28	14	SF	49	22
SFT	22	13			
N*	32	12	N*	32	12

Type is H=hair-like scales, S=scales, HS=hair-like scales, insertion is. E=erected, F=flat and colouration is C=coloured, T=transparent. The N strategy has no phanera, no insertion and no colouration. Species can belong to 1 or 2 different strategies depending on whether they had similar or different strategies on their forewing and hindwing. We investigated a total of 123 species and 31 families. \* within the N strategy, 25 species and 10 families had a nude membrane only, while 7 species and 4 families had a nude membrane with presence of scale sockets.

**Table S4. Phylogenetic signal on continuous structural and colour variables.**

Wing zone	Variable	Forewing		Hindwing	
		K	$\lambda$	K	$\lambda$
Opaque	Phanera density	0.465***	0.611**	0.422*	0.592*
	Phanera width	0.542**	0.845***	0.607***	0.82***
	Phanera length	0.601***	0.841***	0.42ns	0.162ns
	Phanera surface	0.696***	0.952***	0.746***	0.942***
Transparent	Phanera density	0.59***	0.918***	0.656***	0.962***
	Phanera width	0.545*	0.852***	0.588**	0.899***
	Phanera length	0.745**	1.001***	0.641**	1.001***
	Phanera surface	0.657*	0.927***	0.639**	0.938***
Entire wing	Wing area	0.681**	1.044***	0.668*	1.043***
	Clearwing area	0.649***	0.717***	0.603***	0.697***
	Proportion of clearwing area	0.49***	0.633***	0.526***	0.683***
	Wing length	1.068***	1.045***	0.939***	1.032***
Transparent	Mean transmittance over [300-400]	0.326ns	0ns	0.339ns	0.287~
	Mean transmittance over [400-700]	0.325ns	0ns	0.383~	0.537**
	Mean transmittance over [700-1100]	0.316ns	0ns	0.383~	0.55**
	Mean transmittance over [300-700]	0.32ns	0ns	0.363ns	0.481**
	Proportion of UV transmittance	0.361ns	0ns	0.367ns	0 ns
	Colour contrast	0.355ns	0ns	0.357ns	0 ns
	Brightness contrast	0.277ns	0ns	0.525~	0.878***

Blomberg's K (Blomberg et al. 2003) and Pagel's  $\lambda$  (Pagel 1999) computed for continuous variables. K and  $\lambda$  were tested if significantly different from 0 (no phylogenetic signal). Symbols are: ns=non-significant,  $\sim p < 0.1$ , \*  $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ . Lambda different from 0 indicates an influence of phylogeny.

**Table S5. Phylogenetic signal on binary structural variables.**

Zone	Variable	Forewing			Hindwing		
		D	p(D≠1)	p(D≠0)	D	p(D≠1)	p(D≠0)
Transparent	Presence of phanera	0.265	**	ns	0.04	***	ns
	Phanera type	-0.093	***	ns	-0.152	***	ns
	Phanera insertion	0.55	**	*	0.591	*	*
	Phanera colouration	0.021	***	ns	0.188	***	ns
Opaque	Phanera type	0.431	ns	ns	0.538	ns	ns
	Phanera insertion	11.195	ns	**	16.074	ns	**

Fritz and Purvis' D (Fritz and Purvis 2010) was computed for the presence of phanera (wing membrane covered with phanera or nude), the phanera type (hair-like scales or scales, the category hair-like scales + scales being assigned to scales), phanera insertion on wing membrane (erected or flat) and phanera colouration (containing transparent scales or not). We tested whether D was significantly different from 0 (departing from Brownian motion) and different from 1 (departing from random process) and mentioned the associated p-value levels: ns=non-significant,  $\sim p < 0.1$ ,  $* p < 0.05$ ,  $** p < 0.01$ ,  $*** p < 0.001$ .

**Table S6. Mean-Pairwise Distance (MPD) for binary structure variables**

Zone	Variable	Forewing		Hindwing		At least on one wing	
		Species number	Mean age	Species number	Mean age	Species number	Mean age
	No phanera N	25	104.3	28	<b>94.8*</b>	32	99.5
	Phanera present	94	<b>101*</b>	91	102.6	101	102.2
	HEC	15	<b>89.5*</b>	13	<b>87.1*</b>	15	<b>89.5*</b>
	HFC	12	98	10	103.9	12	98
	SEC	15	99.6	11	107.8	16	104.9
	SET	8	95.2	5	105.6	9	97.9
	SFC	21	100	25	102.4	28	101.2
	SFT	19	99.7	15	93	22	98.9
	HSEC	3	<b>28.1*</b>	4	<b>56.5*</b>	4	<b>56.5*</b>
Transparent	HSET	1	-	1	-	1	-
	HSFC	0	-	3	106.8	3	106.8
	HSFT	0	-	4	99.3	4	99.3
	Phanera H	27	<b>96.4*</b>	23	98.9	27	<b>96.4*</b>
	Phanera S	63	102.2	56	103.8	70	103.6
	Phanera HS	4	<b>63.9*</b>	12	96.4	12	96.4
	Insertion E	42	<b>98.1*</b>	34	100.6	44	100.5
	Insertion F	52	100.6	57	100.9	62	100.6
	Colour C	66	<b>99.1*</b>	66	102.3	73	101.3
	Colour T	53	104.1	53	<b>98*</b>	60	101.3
Opaque	SEC	1	-	1	-	1	-
	SFC	116	<b>101.8*</b>	104	103.2	116	<b>101.8*</b>
	HSFC	1	-	12	<b>90.1*</b>	12	<b>90.1*</b>
	Phanera H	5	120	6	118.7	6	118.7
	Phanera S	117	<b>101.8*</b>	105	103.1	117	<b>101.8*</b>
	Phanera HS	1	-	12	<b>90.1*</b>	12	<b>90.1*</b>
	Insertion F	122	103.1	122	103.1	122	103.1

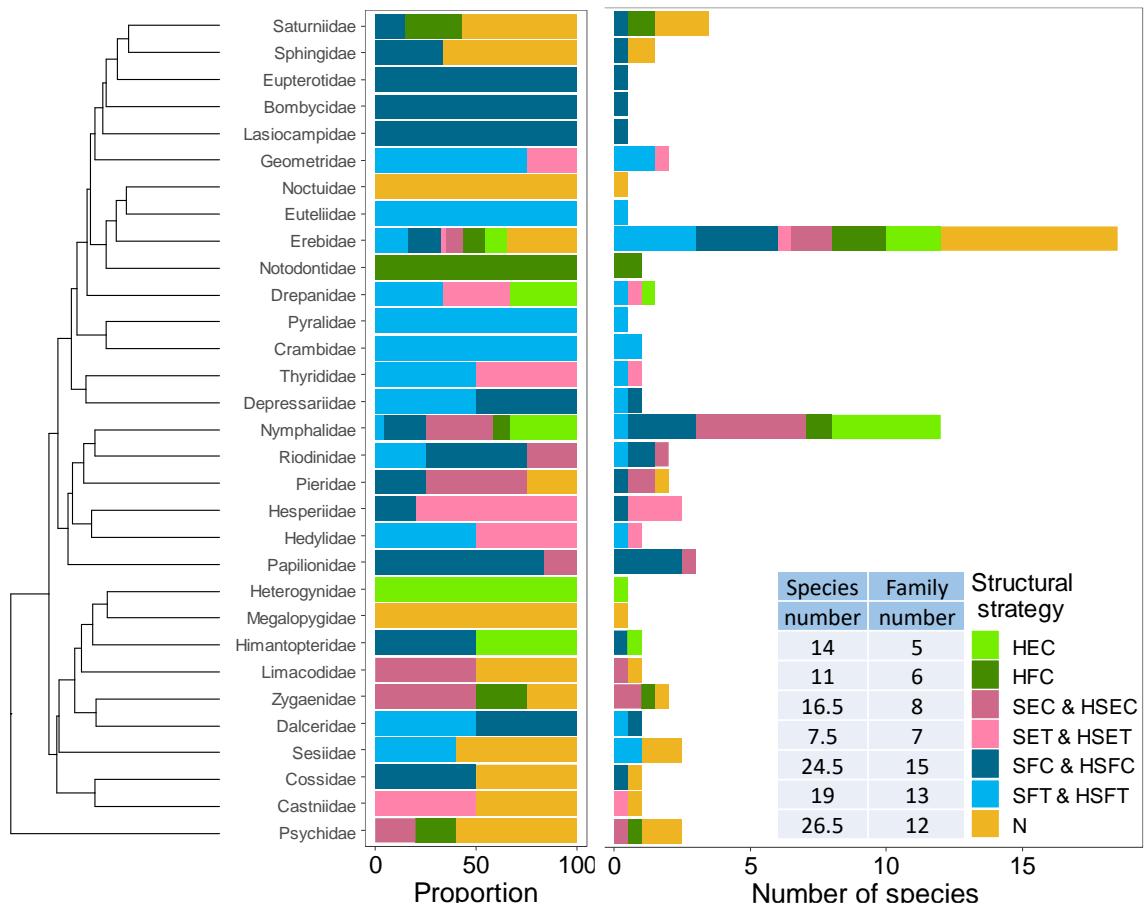
For a given variable, we considered all the species which had the value indicated for the variable in the table on the forewing, on the hindwing, or on at least one of the wings. We computed the observed MPD, the distribution of 1000 simulated values by randomly shuffling the trait on the phylogeny, and its 5% inferior quantile threshold. An observed MPD lower than the threshold indicated a significant phylogenetic clustering (indicated by a bold value and \* for p-value<0.05). Pooling both wings dilutes potential clustering compared to wings taken separately. Instead of expressing MPD values as the length of the branches separating two species, we presented mean ages as MPD/2 to express age depth. A trait shared by all species yielded an MPD-age value of 141.9 My, the root age. H=hair-like scales, S=scales, HS=hair-like scales and scales, F=flat, E=erected, T=transparent, C=coloured. Pooling both wings dilutes potential clustering if any, that is why significant values are a subset of significant values from wings taken separately. The number of species included in the analysis was 119 for the transparent zone of the forewing or the hindwing, 123 otherwise. The symbol ‘-‘ means that there was no or only one species showing that character, preventing any computation of MPD value. When there was a ‘-‘ for all the cases tested for one trait (e.g. Colour C in the opaque zone), we withdrew the trait from the table.

**Table S7. Correlation between structural traits taken as binary traits.**

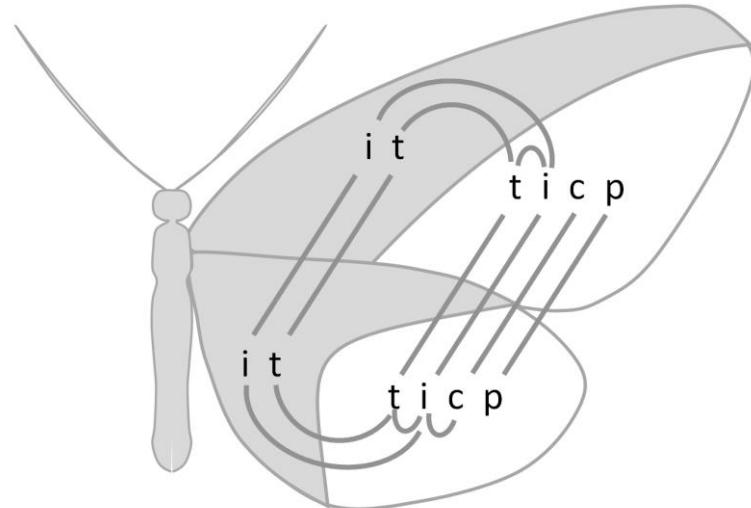
Zone	Wing	Variable	Variable	Nb	LRT (type=h)
Transparent	Forewing and Hindwing	presence		115	<b>69.73***</b>
		type <sup>†</sup>		84	<b>84.01***</b>
		insertion		84	<b>73.98***</b>
		colouration		84	<b>80.08***</b>
Opaque	Forewing and Hindwing	type <sup>†</sup>		123	<b>36.41***</b>
		insertion		123	<b>11.61*</b>
Opaque and Transparent	Forewing	type <sup>†</sup>		119	<b>26.26***</b>
		insertion		94	<b>1.8***</b>
Opaque and Transparent	Hindwing	type <sup>†</sup>		119	<b>23.85***</b>
		insertion		91	<b>2.24***</b>
Transparent	Forewing	type <sup>†</sup>	insertion	94	3.15ns
		type <sup>†</sup>	colouration	94	<b>18.42**</b>
		insertion	colouration	94	5.85ns
Transparent	Hindwing	type <sup>†</sup>	insertion	91	3.28ns
		type <sup>†</sup>	colouration	91	<b>19.07***</b>
		insertion	colouration	91	<b>9.67*</b>
Opaque	Forewing	type <sup>†</sup>	insertion	123	0.13ns
		type <sup>†</sup>	insertion	123	0.10ns

For each analysis, we indicate the number of species included in the analysis (Nb), the likelihood ratio value with the associated p-value (ns p>0.10, ~p<0.10, \* p<0.05, \*\*p<0.01, \*\*\*p<0.001) between the model with dependent traits (4 parameters estimated) and the model with independent traits (8 parameters estimated). A significant p-value indicated that the dependent model did significantly better than the independent model, indicating a correlated evolution between the traits (underlined in bold). To test the correlations involving type, insertion or colouration of phanera in the transparent zone, we excluded the species for which phanera were absent in the transparent zone. Insertion was then a binary trait (flat versus erected), as well as colouration (transparent versus coloured). <sup>†</sup>For phanera type, we considered that the combination hair-like scales and scales was assimilated to scales and we built the binary variable (hair-like scales versus (scales alone or the combination hair-like scales and scales).

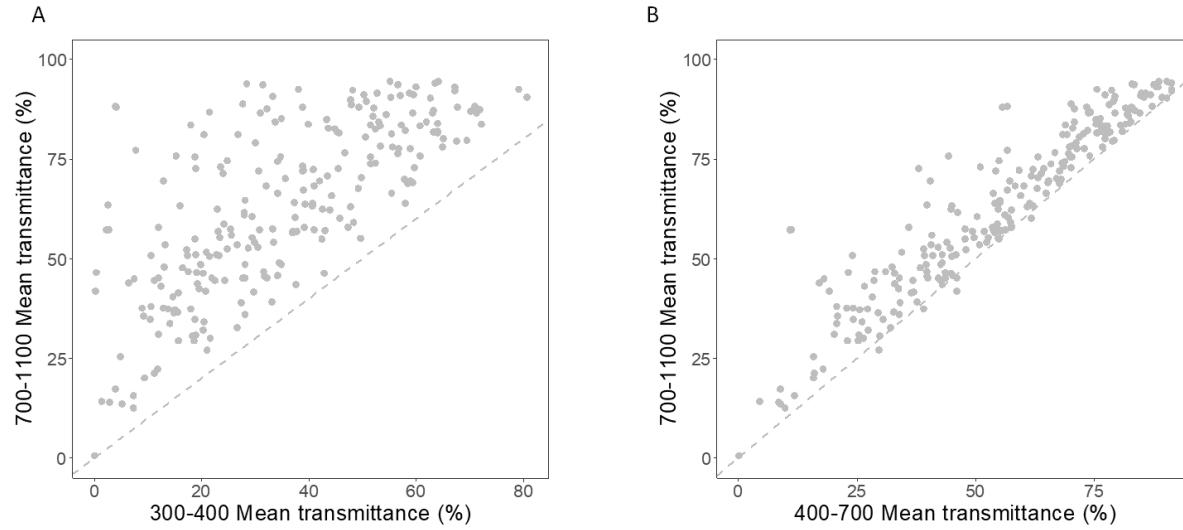
**Figure S1.** Distribution of structural strategies for transparency across the families of the study dataset, in proportion of the total number of species and wings (left), or in number of species (right), with the count of species and families showing a specific strategy. A structural strategy is defined as the combination of phanera type, insertion and colouration. Type is H=hair-like scales, S=scales (hair-like scales and scales were assimilated to scales), insertion is E=erected, F=flat and colouration is C=coloured, T=transparent. The N strategy has no phanera, no insertion and no colouration. Species that had different strategies on their forewing and hindwing were counted for 0.5 for each strategy.



**Figure S2.** Phylogeny-controlled correlations between structural binary traits as tested with Pagel's method. Phanera type was separated into two binary variables: p = presence (yes/no) and t = type (hair-like scales versus scales alone or combination of hair-like scales and scales). i=insertion (flat versus erected), and c=colouration (transparent versus coloured). Presence of a link indicates significant correlation, between wings, between the opaque zone (grey) and the transparent zone (white) of the same wing, within a zone. In the opaque zone, phanera were always present and coloured, which made impossible to test correlations involving p or c variables.



2 **Figure S3.** Correlation between mean transmittance in the near infrared 700-1100 nm range and  
3 mean transmittance in the UV range 300-400 nm (A), or the human-visible range 400-700 nm (B).  
4 Dashed lines indicate identical values.



5  
6