

1 The vesicular trafficking system component MIN7 is required for 2 minimizing *Fusarium graminearum* infection

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11 **Short title:** New role for MIN7 in minimizing fungal infection.

12 13 **One sentence summary:**

14 Disruption of an ARF-GEF protein encoding gene *AtMin7* in *Arabidopsis thaliana* and silencing of
15 the orthologous gene in wheat result in hyper susceptibility to the fungal pathogen *Fusarium*
16 *graminearum*.

17 18 **List of author contributions:**

19 K.K. and K.E.H-K. conceived the original idea and formulated a research plan; A.K.M.W. and V.P.
20 designed and performed most of the experiments and analyzed the data; M.G.M. provided technical
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32

33 ABSTRACT

34 Plants have developed intricate defense mechanisms, referred to as innate immunity, to defend
 35 themselves against a wide range of pathogens. Plants often respond rapidly to pathogen attack by the
 36 synthesis and delivery of various antimicrobial compounds, proteins and small RNA in membrane
 37 vesicles to the primary infection sites. Much of the evidence regarding the importance of vesicular
 38 trafficking in plant-pathogen interactions comes from the studies involving model plants whereas this
 39 process is relatively understudied in crop plants. Here we assessed whether the vesicular trafficking
 40 system components previously implicated in immunity in *Arabidopsis thaliana* play a role in the
 41 interaction with *Fusarium graminearum*, a fungal pathogen notoriously famous for its ability to cause
 42 Fusarium head blight (FHB) disease in wheat. Among the analyzed vesicular trafficking mutants, two
 43 independent T-DNA insertion mutants in the *AtMin7* gene displayed a markedly enhanced
 44 susceptibility to *F. graminearum*. Earlier studies identified this gene, encoding an ARF-GEF protein,
 45 as a target for the HopM1 effector of the bacterial pathogen *Pseudomonas syringae* pv. *tomato*, which
 46 destabilizes AtMIN7 leading to its degradation and weakening host defenses. To test whether this key
 47 vesicular trafficking component may also contribute to defense in crop plants, we identified the
 48 candidate *TaMin7* genes in wheat and knocked-down their expression through Virus induced gene
 49 silencing. Wheat plants in which *TaMIN7* were silenced displayed significantly more FHB disease.
 50 This suggests that disruption of MIN7 function in both model and crop plants compromises the
 51 trafficking of innate immunity signals or products resulting in hyper-susceptibility to various
 52 pathogens.

54 INTRODUCTION

55 Being sessile in nature, plants are frequently exposed to various environmental stresses including
 56 pathogens and yet more often than not plants appear healthy or show only weak or mild disease
 57 symptoms. To maintain this healthy status, plants have evolved an elaborate and tightly regulated
 58 innate immune system that allows them to fend off pathogens or restrict pathogens invasion or slow
 59 down/minimize disease progression (Jones and Dangl, 2006).

60 Each plant cell is surrounded by the plasma membrane (PM) and a cell wall and contain a variety of
 61 membrane-enclosed organelles. Transport of various cargo molecules across different membranes and
 62 sorting these to the correct cellular compartments, i.e. the PM, cell wall and the extracellular space
 63 (apoplast) is a fundamental process, central for multiple plant cell functions. Regulation of multiple
 64 cellular responses by the membrane trafficking network during plant-microbe interactions is required
 65 to facilitate a coordinated defense response at sites of pathogen attack. These involve rapid
 66 remodeling the composition of the PM including accumulation, activation and re-localization of cell
 67 surface immune receptors, reinforcement of the cell wall, secretion of antimicrobial proteins,

phenolics and reactive oxygen species. Whereas other molecules are trafficked from the vacuole and other cellular compartments to the apoplast and to the cell wall to orchestrate cytoplasm communication during immune signal transduction. This has been well documented, with *A. thaliana* and its vast available genetic resources being particularly well exploited for functional studies of vesicular trafficking components and pathways (Ben Khaled et al., 2015; Gu et al., 2017; Yun and Kwon, 2017; Ekanayake et al., 2019).

The vesicular trafficking system comprises two main pathways, secretory and endocytic (Figure 1a), with both implicated in effective immunity against pathogens. Thus, for example, a range of defense-related proteins, antimicrobial metabolites and compounds strengthening plant cell wall such as callose are secreted to the sites of pathogen invasion. Concomitantly cell-surface immune receptors are subjected to endocytosis which is necessary for initiation of signal transduction and regulation of receptor activity (e.g. through recycling or degradation in the vacuole). Secretory pathways transport newly synthesized proteins and other macromolecules, such as phospholipids, polysaccharides, and small RNA (collectively referred to as ‘cargo’) from the endoplasmic reticulum (ER) via the Golgi apparatus to the plasma membrane (PM) or the extracellular space. In the endocytic pathway, membranous vesicles internalized in the PM undergo homotypic fusion to form early/sorting endosomes. Internalized cargo could then be sorted and recycled back to the PM through the recycling endosome, sent to the *trans*-Golgi network (TGN) via retrograde trafficking mechanisms, or trafficked through the late endosome/multivesicular body (MVB) to the vacuole (Figure 1a).

Vesicular trafficking requires the budding, transport and fusion of vesicles from the donor to the specific target membrane. Each of these processes is regulated by different components of the vesicular trafficking system. For instance, the bud formation requires small GTPases of ARF (ADP-ribosylation factor) or SAR1 (secretion-associated Ras-related protein) type as well as adaptor proteins that recognize and recruit specific cargo receptors. Phosphorylation and activation of ARF/SAR1 are regulated by guanine nucleotide exchange factor (GEF) proteins which stimulate the release of GDP to allow binding of GTP. The latter triggers a conformational change in ARF/SAR1 that allows their stable association with the membrane surface and recruitment of specific coat proteins (COP) initiating the budding process. Once the rounded vesicle shape is formed, the large Dynamin-related multidomain GTPases catalyze the membrane scission generating a transport vesicle. The coat components are rapidly lost shortly after the vesicle buds off (Figure 1b).

Fusion of vesicles to the correct target membranes is regulated by the small Rab GTPase proteins (Figure 1c). Different Rab proteins are associated with one or more membrane-enclosed organelles of the secretory pathway. Once in the GTP bound state, the Rab GTPase proteins bind to specific tethering factors in the target membrane to establish the first connection between the membranes that are going to fuse. Next, the N-ethylmaleimide-sensitive factor adaptor protein receptor (SNARE)-domain containing proteins on both the vesicle and the targeting membrane dock to mediate the fusion

of the two membranes. SNAREs are transmembrane proteins that exist in complementary sets. Those located in the vesicle are called v-SNAREs or R-SNAREs, and those located in the target membrane are known as t-SNAREs or Q-SNAREs. A trans-SNARE complex formed following binding of v-SNARE/R-SNARE to t-SNARE/Q-SNARE located on separate membranes catalyzes the membrane fusion (Collins et al., 2003; Yun and Kwon, 2017).

Much of the evidence regarding the role for vesicular trafficking in plant immunity comes from the studies involving model plants *Arabidopsis thaliana* and *Nicotiana benthamiana* and to a lesser extent crops such as barley (*Hordeum vulgare*) and a small number of their respective biotrophic or hemi-biotrophic bacterial (*Pseudomonas syringae*), oomycete (*Phytophthora infestans*) or the powdery mildew fungal pathogens (*Golovinomyces cichoracearum*, *Blumeria graminis*, and *Erysiphe pisi*) (Kwon et al., 2008a; Kwon et al., 2008b; Dodds and Rathjen, 2010; Bozkurt et al., 2011; Nomura et al., 2011; Ellinger et al., 2013). However, little is known about the role of plant vesicular trafficking in the interactions involving other pathogens and those which infect non-leaf tissue.

Fusarium Head Blight (FHB) disease caused by the ascomycete fungus *Fusarium graminearum* and related *Fusarium* species causes substantial yield losses and reduced grain quality and safety in a number of major cereal crops, such as wheat, barley, maize and oat, worldwide (Bottalico and Perrone, 2002). Moreover, under laboratory conditions *F. graminearum* is able to infect floral tissue of intact *A. thaliana* plants as well as detached leaves (Urban et al., 2003; Chen et al., 2006; Cuzick et al., 2008; Koch et al., 2013). For this plant species a large mutant collection is readily available.

The aim if this study was to assess whether knock-out mutations in the individual components of the vesicular trafficking system in *A. thaliana* previously implicated in plant immunity had any impact on the interaction with *F. graminearum*. Screening of the assembled mutants collection using a detached-leaf bioassay identified two independent T-DNA insertion mutants in the *AtMin7* gene, which encodes an ADP ribosylation factor (ARF) guanine nucleotide exchange factor (GEF) protein, that displayed striking hyper-susceptibility to *F. graminearum* strain PH-1 infection compared to the parental wild-type *A. thaliana* ecotype Col-0. Utilizing a recently released high-quality fully annotated wheat genome reference sequence assembly (International Wheat Genome Sequencing Consortium (IWGSC), 2018) and well established bioinformatics tools enabling identification of putative gene orthologs from different plant species (Adamski et al., 2019) we identified the three homoeologous *TaMin7* genes in hexaploid wheat (*Triticum aestivum*). Knock-down of these genes using Virus-induced gene silencing (VIGS) (Lee et al., 2012) significantly promoted FHB disease formation in this crop species.

RESULTS

Assembling a collection of *A. thaliana* mutants with defects in membrane trafficking

To investigate whether vesicular trafficking plays a role in a compatible interaction (i.e. disease) between an ascomycete fungus *F. graminearum* and its laboratory host *A. thaliana*, we assembled a collection of twelve mutants containing T-DNA insertions in nine immunity associated genes regulating different vesicular trafficking pathways (**Figure 1a**, and **Table 1**). Homozygous mutants were obtained from the Nottingham Arabidopsis Stock Centre (NASC, UK), and each mutant was verified by PCR amplification using gene-specific and T-DNA-specific primers as described in **Supplemental Methods** and **Supplemental Table S1**. No obvious developmental or growth defects were observed in any of the untreated mutant plants when compared with the corresponding wild type *A. thaliana* ecotype Col-0 plants grown under standard controlled environment conditions (data not shown).

AtMIN7, an ARF-GEF protein is required to minimize *F. graminearum* infections

To gain insight into whether mutations in any of the selected vesicular trafficking gene increase or decrease susceptibility to the virulent *F. graminearum* strain PH-1, young 5-weeks-old *A. thaliana* plants were point inoculated with *F. graminearum* conidial suspension supplemented with the mycotoxin deoxynivalenol (DON) in a detached leaf bioassay (**Chen et al., 2006**). At 6 days post inoculation (dpi), the inoculated leaves were photographed, and disease levels were quantified by measuring the proportion of lesioned/necrotic area compared to the total leaf area by analyzing the images using the LemnaGrid software module (LemnaTec GmbH, Aachen, Germany).

In the first two experiments, each involving a partially overlapping subset of the mutants, *F. graminearum* inoculated leaves of the two independent T-DNA insertion mutants in the *AtMin7* gene developed extensive necrotic lesions covering up to 75-100% of the total leaf area and showed almost complete loss of green photosynthetic tissue, while most of the remaining mutants displayed much milder disease symptoms with smaller lesions, the rest of the leaf remained unaffected and green (**Figure 2, Supplemental Figure S1**, and data not shown). Experiments involving nine out of twelve mutants, including *atmin7-1* and *atmin7-2*, and the wild-type *A. thaliana* Col-0 were replicated three times with similar results.

AtMin7 is a large gene of 5,857-nt containing 33 exons and is located on chromosome 3. The two loss of function mutants, *atmin7-1* and *atmin7-2*, studied here carry T-DNA insertions in the exon 1 and exon 18, respectively (**Nomura et al., 2006**). *AtMin7* is a member of small family comprising 8 genes encoding ARF-GEF proteins (**Supplemental Figure S2**), which play important roles in the budding of transport vesicles from the membranes (**Steinmann et al., 1999; Mossessova et al., 2003**). This vesicular trafficking component has been shown to contribute to resistance to the bacterial pathogen

Pseudomonas syringae pv. *tomato* (*Pst*), possibly through regulating the trafficking of immunity-associated cargo molecules (Nomura et al., 2006; Nomura et al., 2011).

Identification of candidate wheat *TaMin7* genes

To identify homologs of the *A. thaliana AtMin7* gene in wheat, a natural, crop host of *F. graminearum*, we used the BioMart data mining tool available through Ensembl Plants (Smedley et al., 2015). A total of 26 gene sequences were identified in the reference hexaploid wheat (*Triticum aestivum*) genome (AABBDD) by searching for genes encoding proteins containing the catalytic SEC7 domain (PF12783) characteristic of ARF-GEF proteins. We then aligned the proteins coded by the identified wheat genes with all eight members of the *A. thaliana* ARF-GEF family proteins and used the resulting multiple alignment for phylogenetic analysis. The constructed maximum likelihood phylogenetic tree revealed the three closely sequence related wheat proteins formed a distinct clade with AtMIN7, suggesting these sequences represent the wheat A-, B- and D-genome encoded orthologues of AtMIN7 (Figure 3).

Silencing the candidate *TaMin7* genes in wheat spikes enhances susceptibility to *F. graminearum*

To explore the potential function for the wheat homologs of *AtMin7* in the *F. graminearum* - wheat interaction we tested the effect of silencing the three homoeologous wheat genes, *TraesCS2A02G202900*, *TraesCS2B02G230000* and *TraesCS2D02G212800*, using Barley stripe mosaic virus-mediated Virus induced gene silencing (VIGS) on the FHB disease development (Lee et al., 2012). A 209-nt fragment highly conserved between coding sequences of the three *TaMin7* homoeologs was selected as a target for VIGS using si-Fi21 software (Lück et al., 2019). This target fragment was predicted to generate a high number of silencing-effective siRNAs ($n = 91$), and a very low likelihood of off-target silencing.

An important factor for successful application of VIGS is the ability of the virus to infect and spread without causing any deleterious effect in the host plant. Therefore, the feasibility of BSMV-VIGS approach to induce systemic silencing in the spike tissue of wheat cv. Bobwhite susceptible to *F. graminearum* PH-1 (Brown et al., 2011) was first tested by visualizing the phenotype induced by silencing the *Magnesium-chelatase subunit H gene* (*TaChlH*; *TraesCS7A02G480700*) involved in chlorophyll biosynthesis, which is often used as a visible marker of VIGS (Yuan et al., 2011). The recombinant BSMV carrying a 250-nt fragment of *TaChlH* gene in antisense orientation and BSMV:*mcs4D* (harboring a 275-nt noncoding fragment corresponding to the multiple cloning site sequence of the vector pBluescript II KS) were inoculated onto the flag leaves of wheat plants at the early boot stage. At 13 days post inoculation (dpi) the plants infected with BSMV:*asTaChlH* developed yellow-orange coloration of the lemma and palea of spikes indicative of the loss of chlorophyll and successful silencing of the *TaChlH* gene, whereas the spikes of plants infected with the control construct BSMV:*mcs4D* showed typical mosaic symptoms (Supplemental Figure S3). No

visible developmental abnormalities were observed in the wheat plants challenged with either of the two BSMV-VIGS constructs. Moreover, similar levels of FHB disease were observed on wheat plants pre-infected with BSMV and on virus-free plants challenged with *F. graminearum*, indicating that the susceptibility to the fungus was not compromised in the virus-infected wheat (Figure 4). These results suggest that BSMV-mediated VIGS can be used to silence genes in the reproductive tissue of wheat cv. Bobwhite and the approach appeared suitable for assessing the role of *TaMin7* during *F. graminearum* infection in wheat.

The ability of BSMV:*asTaMin7* to induce silencing of the corresponding endogenous genes was confirmed using quantitative reverse transcription PCR (qRT-PCR). The *TaMin7* transcripts were decreased in abundance by 77 % in the spikes of the VIGS-treated plants compared to those inoculated with BSMV:*mcs4D*, and no significant difference in *TaMin7* expression levels was observed in the mock- versus BSMV:*mcs4D*-inoculated control plants. To analyze the effect of *TaMin7* silencing on *F. graminearum* infection, spikes of VIGS-treated plants showing typical virus-induced symptoms were point inoculated with the fungal conidia suspension and observed for disease symptoms for 15 days after fungal inoculation. Reduction of *TaMin7* mRNA expression levels in spikes of the BSMV:*asTaMin7* treated plants was associated with significantly enhanced susceptibility to *F. graminearum* (Figure 4). In contrast, no effect on FHB disease development was found in mock-inoculated and BSMV:*mcs4D*-inoculated control plants (Figure 4).

DISCUSSION

Membrane trafficking plays an important role in plant-pathogen interactions and mutants with lesions in several different plant vesicular trafficking genes exhibiting compromised responses to bacterial or fungal pathogens in the model plant species *A. thaliana* have been identified in previous studies (Gu et al., 2017; Yun and Kwon, 2017; Ekanayake et al., 2019). Moreover, it appears that some pathogens evolved host cell translocated effector proteins that promote disease by interfering with plant membrane trafficking pathways (Ben Khaled et al., 2015). One of the vesicular trafficking genes whose role during plant disease has been previously well explored in the interactions between *A. thaliana* and the bacterial pathogen *Pst* is *AtMin7* (Nomura et al., 2006; Nomura et al., 2011). The *AtMIN7* protein localizes to the TGN/early endosome (EE) and is involved in endocytic recycling of PM resident proteins but it has also been hypothesized to regulate secretion (Tanaka et al., 2009; LaMontagne and Heese, 2017). Mutants that lack this protein allow increased bacterial multiplication, possibly due to miss-regulation of membrane trafficking of the plant defense-related cargo (Nomura et al., 2006; Nomura et al., 2011). *AtMIN7* has also been shown to contribute to the cytosol-initiated immune responses triggered by the *Pst* effectors such as *AvrRpt2* and *AvrPphB* (Nomura et al., 2011) and to the apoplast-initiated immune responses through an unknown mechanism by preventing apoplast water soaking and therefore presumably restricting the flow of nutrients to the bacteria (Xin et al., 2016). To achieve successful disease, *Pst* secretes a conserved effector protein *HopM1* that is translocated to the TGN/EE of its host during infection where it mediates destabilization of *AtMIN7* followed by degradation via the 26S proteasome (Nomura et al., 2011). Here we demonstrated that *AtMIN7* also contributes to defense against a fungal pathogen *F. graminearum* as the absence of this protein in *A. thaliana* resulted in a markedly enhanced disease (Figure 2). However, whereas *atmin7* mutants displayed increased susceptibility to the *Pst* Δ CEL mutant that lacks *HopM1* along with several other conserved effectors including *AvrE* and only modest increase in susceptibility to the wild type *Pst* (Nomura et al., 2011), we show that these same *A. thaliana* mutants are clearly and unmistakably hyper-susceptible to the wild type strain of *F. graminearum* (Figure 2).

Disruption of *AtMin7* may compromise trafficking of specific molecules and cargo protein. These molecules could include plant defense-related proteins or secondary metabolites that help minimize *F. graminearum* infection. For example, callose (a (1,3)- β -glucan polymer) can act as a physical barrier reducing fungal penetration, and an increased callose deposition and the *F. graminearum* induced callose synthase activity in the infected spikelets was found to be correlated with increased disease resistance (Ribichich et al., 2000; Blümke et al., 2017). Identification of proteins and/or other molecules transported specifically by membrane vesicles regulated by *AtMIN7* is challenging. But comparison of vesicles cargo between wild-type and *atmin7* *A. thaliana* mutants during *F. graminearum* infection could provide valuable clues to the role of this gene during fungal disease establishment.

Although *A. thaliana* has proven to be a very useful model organism for unraveling the key mechanisms underlying plant-pathogen interactions, some findings cannot be translated directly to crop plants. Hence studies involving interactions between the pathogens and their natural host plants could provide more relevant information and facilitate exploration of new strategies for disease control in crops. Here we utilized a transient gene silencing approach (VIGS) to assess the role of *AtMin7* homologues in wheat, which is one of the most important staple food crops whose production is regularly threatened by fungal diseases including FHB. These results from the analysis of the wheat - *F. graminearum* pathosystem (Figure 4) are consistent with those from the study of the *A. thaliana* - *F. graminearum* pathosystem (Figure 2), and provide evidence that disruption of MIN7 function in both dicotyledonous and monocotyledonous hosts compromises the plant innate immunity resulting in more severe disease.

Whether *F. graminearum* utilizes similar mechanisms of suppressing the host immunity to promote disease as in the case of *Pst* which involve effector-mediated degradation of AtMIN7 remains to be determined. However, our preliminary data (not shown) indicates that *TaMin7* transcript levels may be reduced in the wheat spikes infected with *F. graminearum* compared to the mock-inoculated samples. If confirmed this would suggest a different mechanism operating in wheat as *Pst* infection of *A. thaliana* seem to induce AtMIN7 protein degradation while not affecting the *AtMin7* transcript levels (Nomura et al., 2006).

AtMIN7 was also shown to play a role in polar localization and dynamic repolarization of the PIN (PIN-formed) efflux carrier proteins enabling the directional transport of auxin in the tissues (Tanaka et al., 2013). Elevated levels of reactive oxygen species (ROS) induced during stress responses in *A. thaliana* affect AtMIN7-dependent PIN endocytic recycling resulting in increased accumulation of auxin in the affected tissues (Zwiewka et al., 2019). A recent study demonstrated that higher levels of auxin are accumulated during *F. graminearum* infection in a susceptible wheat cultivar Roblin compared to the moderately resistant cultivars Wuhan 1 and Nyubai, indicating that auxin may promote susceptibility to this fungal pathogen (Brauer et al., 2019). Moreover, analysis of the recently released transcriptome data indicated that wheat homologs of *AtMin7* and *PIN* genes are down-regulated during *F. graminearum* infection whereas numerous transcripts of genes belonging to auxin pathway are being up-regulated (Pan et al., 2018). The data from these previous studies together with findings from our study form a foundation to an alternative hypothesis regarding the mechanisms employed by *F. graminearum* for achieving successful infection. That is, it is conceivable that *F. graminearum* infection induces temporally coordinated waves of gene expression which regulate MIN7-dependent distribution and accumulation of auxin during infection. Further studies are necessary to confirm or refute this hypothesis.

This study also uncovered potential additional regulators of membrane trafficking contributing to host immunity such as SYP122. Significant increase ($P < 0.05$) in the size of lesions induced by *F.*

graminearum in leaves of *syp122-1* compared to those in wild type *A. thaliana* Col-0 and other mutants (apart from *atmin7-1* and *atmin7-2*) was seen in one nonreplicated experiment (**Supplemental Figure S1**). This warrants further examination in a follow-on study, as well as analysis of response to *F. graminearum* in additional vesicular trafficking gene mutants that were not investigated in the current study and including those whose association with host immunity has not yet been demonstrated. It would also be interesting to investigate contribution of unconventional secretion pathways related to pathogen attack or stress conditions (Wang et al., 2018), which involve direct fusion of the central vacuole, multivesicular bodies or EXPO (a double-membrane exocyst-positive organelle) with the PM, to plant interactions with *F. graminearum*. Particularly interesting in this respect are the exosomes, small membranous vesicles that are thought to be released to the apoplast following fusion of MVBs with the PM (Rutter and Innes, 2017). These were reported to carry various defense related proteins and antimicrobial compounds, and small regulatory RNAs and appear to be able to cross the plant cell wall and deliver this defense-related cargo to the cells of invading fungal pathogens (Rutter and Innes, 2017; Cai et al., 2018).

Elucidating the mechanisms by which membrane trafficking regulates plant immune responses and enhanced understanding of the vesicular trafficking components and pathways manipulated by microbial pathogens to promote disease will provide fundamental knowledge for the development of novel methods of disease intervention.

MATERIALS AND METHODS

Plant material and growth conditions

The *A. thaliana* mutants (**Table 1**) and the corresponding wild-type parental ecotype Col-0 used in the study were obtained from the Nottingham Arabidopsis Stock Center. The corresponding mutations were verified by PCR using the primers listed in **Supplemental Table S1** and confirmed homozygous mutants selected for further study (**Supplemental Methods**). *A. thaliana* seeds were sown in Levington F2 + S compost (Everris Ltd.) and stratified in the dark for four days at 5°C before transferring to a controlled environment growth chamber operating at 20°C/17°C during day/night and a 16-h photoperiod (light intensity of approximately 100 $\mu\text{mol m}^{-2} \text{s}^{-1}$).

The bread wheat (*Triticum aestivum*) cultivar (cv.) Bobwhite, was used in this study. The plants were grown in a controlled environment growth chamber with day/night temperatures of 23°C/20°C at around 65% relative humidity and a 16-h photoperiod with light intensity of approximately 180 $\mu\text{mol m}^{-2} \text{s}^{-1}$.

Fungal growth conditions, plants inoculation, and disease assessment

The *F. graminearum* strain PH-1 was used in this study. Routine culturing of the fungus, conidiospore induction, and preparation of conidial suspensions followed essentially the same procedures as described in [Brown et al. \(2011\)](#). Conidiospore suspensions harvested in sterile distilled water were adjusted to a concentration of 5×10^5 or 1×10^5 conidia/ml⁻¹ for inoculation of *A. thaliana* or wheat, respectively.

Detached *A. thaliana* leaves were inoculated as described in [Chen et al. \(2006\)](#) with the following modification. Fully expanded rosette leaves were detached from the five-week-old plants using razor blades and placed adaxial surface facing upwards on the surface of 1% water agar in 10 × 10 cm square sterile Petri dishes, with 8 leaves per dish. Each leaf was then superficially wounded by gently puncturing over the mid rib with a glass Pasteur pipette and a 5 µL droplet of *F. graminearum* conidiospore suspension supplemented with 20 µM deoxynivalenol (DON) was deposited on the fresh wound. Mock inoculation was carried out using a 5 µL droplet of sterile distilled water supplemented with 20 µM DON. After inoculation, the plates were transferred to the controlled environment growth chamber operating at 20°C/17°C during day/night and 16-h photoperiod but kept in the dark for the first three days following which they were incubated under low light (light intensity of 40 µmol m⁻² s⁻¹) for further four days before the disease assessment took place.

Color (RGB) photographs were taken at six days after inoculation using a Nikon (D90) camera and backlighting to ensure consistent illumination. Image analysis to quantify the diseased areas was conducted using the LemnaTec LemnaGrid software module (LemnaTec GmbH, Aachen, Germany). Leaf areas were segmented using a combination of a colour-based classification and thresholding after converting the images to grayscale. Filters were applied to remove misclassified pixels and to fill in gaps. Finally, a customized colour-based classification was applied to score leaf-area pixels as belonging to diseased or healthy tissue.

Intact spikes of adult wheat cv. Bobwhite plants were point inoculated at the first signs of anther extrusion by depositing 5 µL of conidial suspension in the floral cavity between the palea and lemma of the outer two florets located in the upper one third of the spike as previously described ([Brown et al., 2011](#)). Control plants were inoculated with sterile water only. Inoculated plants were incubated in a humid chamber for 48 hr of which the first 24 h were in darkness. The inoculated plants were then kept in a controlled environment growth chamber at approximately 65% humidity, and the progression of the disease was visually monitored every 3 days and the number of bleached spikelets below the inoculated spikelet on each spike was recorded ([Urban et al., 2003](#)).

Identification of putative wheat *Min7* genes

Protein domain analysis of predicted *ARF-GEF* genes in the hexaploid wheat (*T. aestivum*) was carried out using BioMart tool in Ensembl. Initially we searched for proteins that contained the Sec7_N domain (Guanine nucleotide exchange factor in Golgi transport N-terminal domain;

PF12783). The wheat genome assembly used for this analysis was the IWGSCrefseq1 ([International Wheat Genome Sequencing Consortium \(IWGSC\), 2018](#)). Coding sequences of eight previously identified genes comprising the *ARF-GEF* gene family in *A. thaliana* ([Vernoud et al., 2003](#)) were also extracted using BioMart tool in Ensembl. Multiple protein sequences alignment was carried out in ClustalW, linked to Geneious 10. For phylogenetic reconstruction, the TVM+I+G nucleotide substitution model was selected by AIC in jModeltest 2.1.10 ([Posada, 2008; Darriba et al., 2012](#)). The Maximum Likelihood phylogeny was reconstructed using PhyML ([Guindon and Gascuel, 2003](#)), with the substitution model selected in jModeltest; starting tree with optimized topology, length and rate parameters; topology searching by the best of NNI and SPR; and 500 bootstraps.

Vector construction for VIGS and fungal inoculations of wheat plants

The total RNA extracted from healthy wheat cv. Bobwhite leaf tissue was used as a template in an RT-PCR for amplification of 209-bp *TaMin7* gene fragment for BSMV-VIGS vector construction. The primers *TaMin7*-2A-seg1-R (5'-AACCACCACCACCGTAAAAGGGTCGCCTCGTCAAT-3') and *TaMin7*-2A-seg1-F (5'-AAGGAAGTTTAATGTTGCAAGCAAAGGCCATC-3') were designed to clone the *TaMin7* gene segment in an antisense orientation into BSMV RNA γ vector pCa- γ bLIC using a ligation independent cloning ([Yuan et al., 2011](#)). VIGS vector for silencing the control, *TaChlH* gene was kindly provided by Dawei Li (China Agricultural University, Beijing, China). To prepare the virus inoculum for wheat inoculation the BSMV α , BSMV β and the recombinant BSMV γ derivatives containing the *TaMin7* and *TaChlH* inserts were transformed into the *Agrobacterium tumefaciens* strain GV3101 (pMP90). Agroinfiltration of *Nicotiana benthamiana* leaves was carried out as previously described ([Lee et al., 2015](#)). The infiltrated *N. benthamiana* leaves were harvested 5-7 days post agroinfiltration, homogenized in 10 mM Na-phosphate buffer (pH 6.8) containing 1% Celite 545 AW (Sigma-Aldrich, UK), and the sap was mechanically inoculated onto wheat leaves just prior to appearance of a flag leaf. Fungal inoculation of wheat plants was carried out using *F. graminearum* wild type strain PH-1 as described in [Brown et al. \(2011\)](#). Briefly, 5 μ L of conidial suspension was inoculated to the outer two florets between the lemma and palea in the upper part of the spike. Control plants were inoculated with sterile water only.

Quantitative real time PCR (QPCR)

Total RNA was isolated from spike tissue of wheat plants infected with BSMV:*asTaMin7*, BSMV:*mcs4D* and mock-inoculated plants prior to *F. graminearum* inoculations using the TRIzol reagent (Invitrogen, Carlsbad, CA, USA) following the manufacturer instructions. To remove any traces of gDNA contamination, RNA samples were treated with TURBO DNaseI (Invitrogen) using methods as described by the manufacturer. The first strand cDNA was synthesized from 1 μ g of total RNA in a total volume of 20 μ L using the SuperScript IV Reverse Transcriptase (Invitrogen) and oligo (dT)₁₈ primers according to manufacturer's instructions. *TaMin7*-specific primers for qPCR,

TaMIN7-Pair1F-QPCR (5'-ATCTTGCGGCAAAAACCAAGT-3') and TaMIN7-Pair1R-QPCR (5'-ACCTGCTGAGCCACATGAAA-3'), were designed using Primer3. Expression of *TaMin7* in silenced and mock-inoculated leaves was normalized to the housekeeping *Cell division control 48* gene (*CDC48*) essentially as previously described (Lee et al., 2014). A no template control was included in all qPCR experiments.

Statistical Analyses

For *A. thaliana* leaf inoculations assays, disease was quantified by expressing the diseased leaf area relative to the total leaf area. Mean disease levels for each genotype were compared using a multi-stratum analysis of variance (ANOVA). Independent lines were compared with the wild type using a Dunnett's test at the 5% ($P < 0.05$) level of significance using wild-type *A. thaliana* Col-0 as the control test. GenStat (release 20.1, 2019, VSN International Ltd.) was used for the statistical analyses.

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419 TABLES

420 **Table 1.** *Arabidopsis thaliana* mutants used in this study.

421

422 FIGURE LEGENDS

423 **Figure 1. Schematic diagram of vesicular trafficking pathways in *Arabidopsis thaliana*.** A, Major
424 cellular vesicular trafficking pathways: secretory (red), endocytic (green) and retrieval (blue). B,
425 Vesicles formation and budding. C, Fusion of vesicles with the correct target membranes.

426 **Figure 2. *F. graminearum* infection of different *Arabidopsis thaliana* mutants measured as the**
427 **proportion of infected leaf area compared to the total leaf area.** GLM analysis ($P < 0.01$). Data
428 representing results from three different experiments with at least 30 leaves from 12 different plants
429 from each mutant for each experiment.

430 **Figure 3. Maximum Likelihood phylogenetic tree displaying representative wheat sequence**
431 **orthologues of *Arabidopsis thaliana* ARF-GEF genes (protein coding sequences).** Node labels
432 indicate percentage bootstrap support (500 replicates). The *A. thaliana* ARF-GEF gene names are
433 shown in blue regular font with *AtMin7* accentuated in bold. Names of wheat genes most likely
434 representing orthologs of *AtMin7* are shown in red bold font, while names of all other candidate wheat
435 ARF-GEF genes are shown in regular black font.

436 **Figure 4. Effect of silencing *TaMin7* genes on the *Fusarium graminearum* infection in wheat.** A,
437 Schematic representation of the *Barley stripe mosaic virus* (BSMV)-derived vectors for silencing
438 *TaMin7* (adapted from Lee et al., 2012). cDNAs of the three BSMV genomic RNAs (α , β , and γ) each
439 of which is required for full infection are cloned into a binary vector pCaBS under control of a double
440 CaMV 35S promoter (2X35S) and flanked by a ribozyme sequence originating from TRSV satellite
441 RNA (Rz), which allows cis-cleavage of transcribed RNA at the 3' end of viral genomic RNA. The
442 RNA γ genome is modified to include a small fragment of *TaMin7* protein-coding sequence in the
443 antisense orientation immediately downstream of the BSMV *yb* cistron. Viral cistrons are shown as
444 yellow rectangles. B, images of representative wheat spikes at 15 days post inoculation with *Fusarium*
445 *graminearum* from virus-untreated (No virus) plants and those treated with the control
446 (BSMV:*mcs4D*) and the VIGS construct for silencing *TaMIN7* (BSMV:*asTaMin7*). C, *TaMin7*
447 silenced wheat plants show increased susceptibility to *F. graminearum* infection as determined by
448 counting the number of infected (bleached) spikelets per spike. Values represent mean of three
449 independent experiments. D), qRT-PCR measurement of *TaMin7* transcripts abundance in wheat
450 plants. RNA extracted prior to *F. graminearum* inoculation and the wheat *CDC48* (*Cell division*
451 *control 48*) gene used as an internal reference. A single star (*) indicates $P < 0.05$ and three stars
452 (***) indicate $P < 0.001$; n = number of plants.

453

454 SUPPLEMENTAL DATA

455 **Supplemental Methods.** PCR-based confirmation of specific mutations in the T-DNA insertion
456 mutant *Arabidopsis thaliana* plants obtained from the seed stock center.

457 **Supplemental Table S1.** Primers used in this study for genotyping *Arabidopsis thaliana* mutants.

458 **Supplemental Figure S1.** *Fusarium graminearum* infection of additional different *Arabidopsis*
459 *thaliana* mutants measured as proportion of infected leaf area compared to the total leaf area in a
460 single nonreplicated experiment. Significant differences between wild-type *A. thaliana* ecotype Col-0
461 plants and vesicle trafficking mutants indicated by letter according Tukey's test subsequent
462 to ANOVA ($P < 0.05$).

463 **Supplemental Figure S2.** Maximum Likelihood phylogenetic tree indicating the relationship among
464 *Arabidopsis thaliana* ARF-GEF genes (coding sequences only). Protein sequences were aligned using
465 the ClustalW in the Geneious 10 software. Node labels indicate percentage bootstrap support (500
466 replicates).

467 **Supplemental Figure S3.** Silencing of *TaChlH* (*Mg-chelatase subunit H*) gene in wheat spikes.
468 *TaChlH* silencing phenotype could be identified approximately two weeks after inoculation of flag
469 leaves with BSMV:*asTaChlH* whereas those inoculated with a negative control BSMV:*mcs4D*
470 construct and mock-inoculated plants showed no spike yellowing phenotype.

471

472

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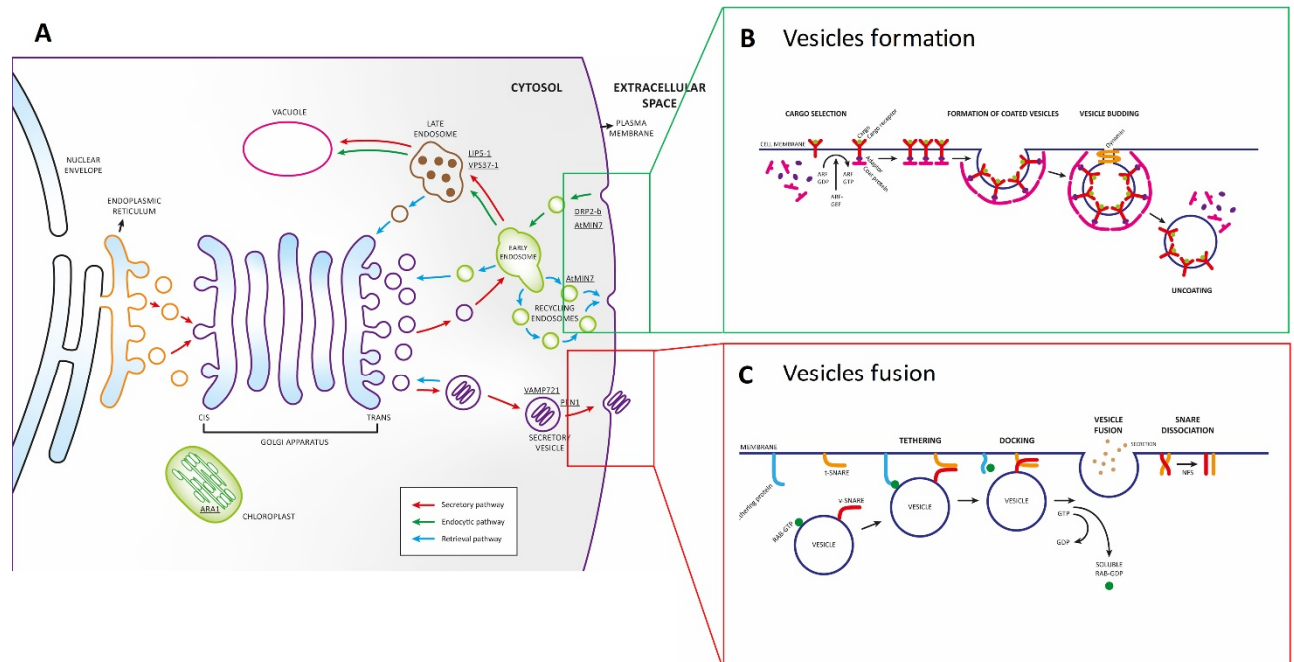


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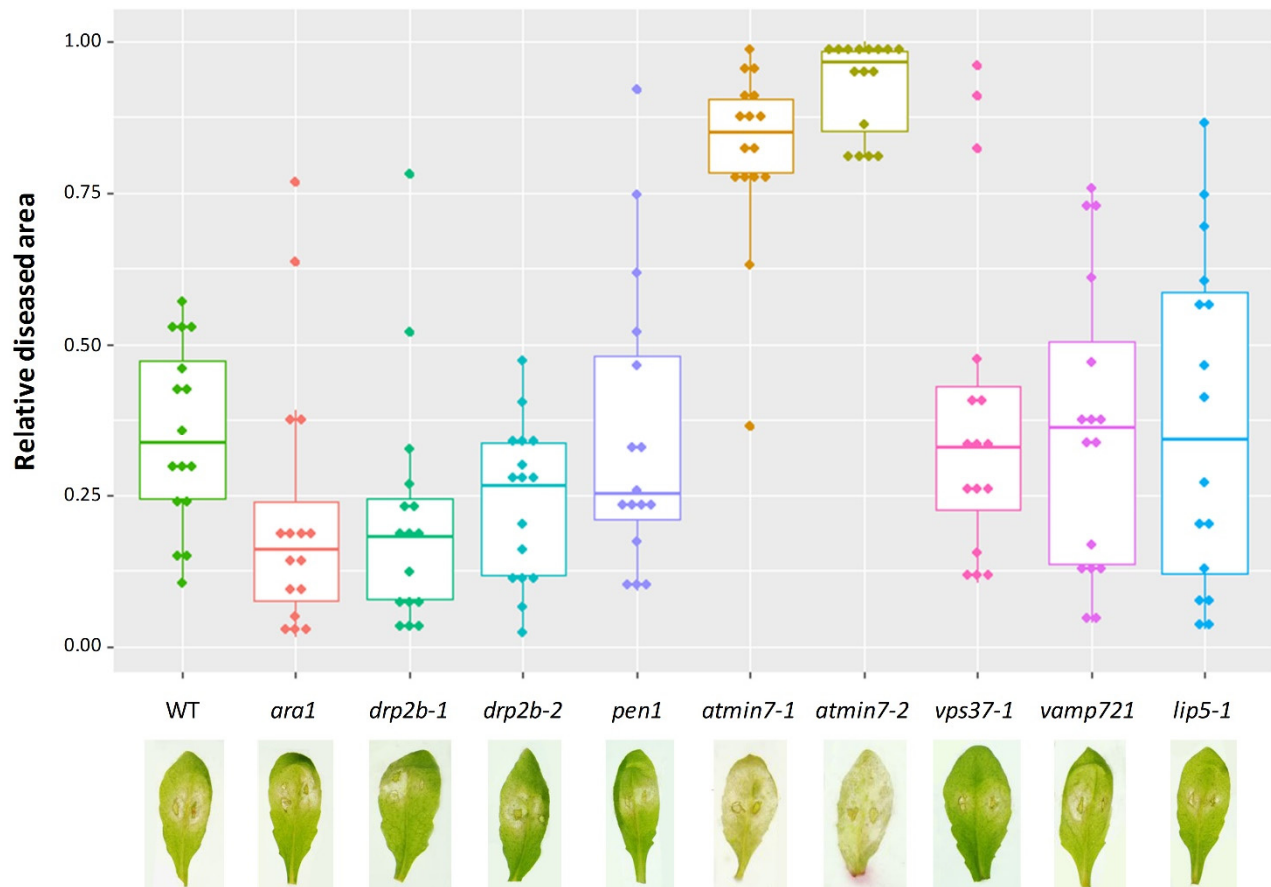


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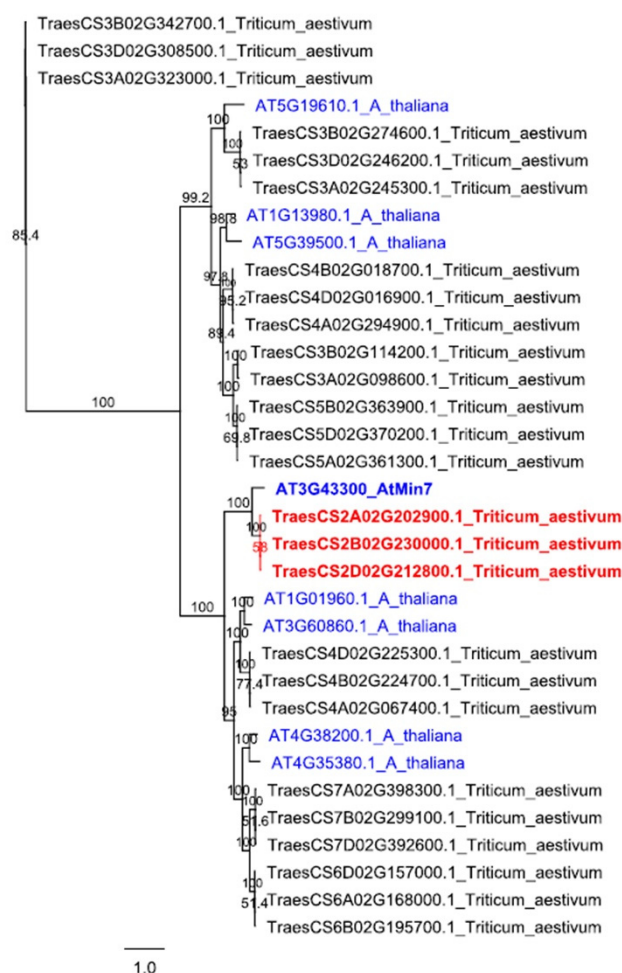


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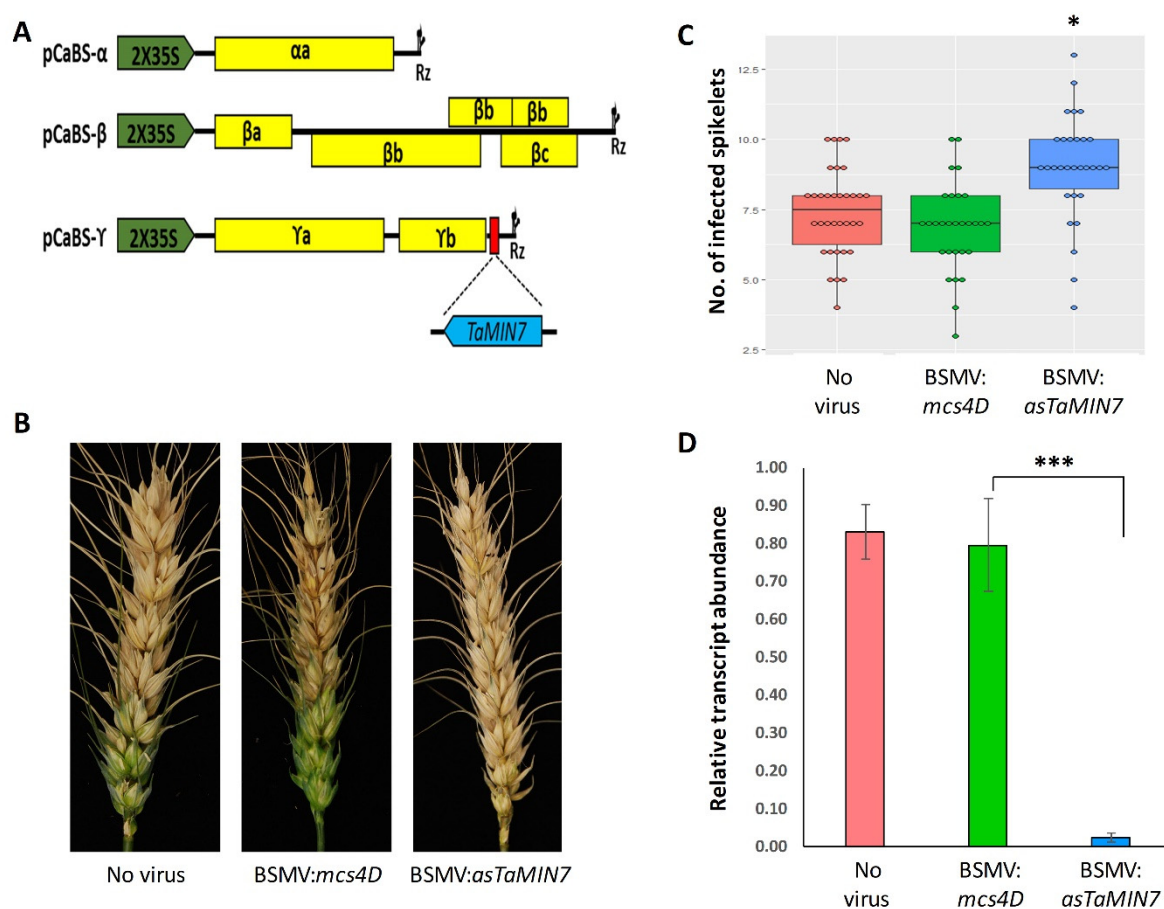


Table 1. *Arabidopsis thaliana* mutants used in this study.

Gene name	Gene ID	Gene description	Mutant name	Mutant accession No.	Reference	Evidence in Arabidopsis plants
<i>Ara1/AtRabA5E</i>	At1g05810	Arabidopsis gene homologous to the Ras oncogene 1/ Rab GTPase-like A5A	<i>ara1</i>	SM_3.15870	Karim et al., 2014	Interacts with proteins involved in multiple stress responses
<i>Drp2b</i>	At1g59610	Dynamin related protein 2B	<i>drp2b-1</i> <i>drp2b-2</i> <i>drp2b-3*</i>	SALK_003049 SALK_134887 SALK_124686	Smith et al., 2014	Functions as a novel component of flagellin-mediated defense responses against bacteria <i>P. syringae</i>
<i>Pen1/Syp121</i>	At3g11820	Penetration resistance 1/ Syntaxin of plants 121	<i>pen1</i>	SALK_004484C	Johansson et al., 2014	Absence of <i>pen1</i> is associated with increased tissue penetration by the fungal pathogen <i>Blumeria graminis</i> fsp. <i>hordei</i> (<i>Bgh</i>)
<i>Syp122</i>	At3g52400	Syntaxin of plants 122	<i>syp122-1*</i>	SALK_008617	Zhang et al., 2007	Acts together with SYP121 as a negative regulator of PCD, SA, JA and ET pathways
<i>AtMin7/Ben1/Big5</i>	At3g43300	HopM1 interactor 7/ BFA-visualized endocytic trafficking defective 1/ Brefeldin A-inhibited guanine nucleotide-exchange protein 5	<i>atmin7-1</i> <i>atmin7-2</i>	SALK_012013 SALK_013761	Nomura et al., 2006	Encodes an immunity associated ARF-GEF protein family protein targeted by HopM1, conserved bacterial <i>P. syringae</i> virulence protein
<i>Vps37</i>	At3g53120	Vacuolar protein sorting 37	<i>vps37-1</i>	SALK_042859C	Spallek et al., 2013	Co-localizes with FLS2 (FLAGELLIN SENSING 2, immune receptor) at endosomes and immunoprecipitates with this receptor upon flg22 elicitation
<i>Vps28-2</i>	At4g05000	Vacuolar protein sorting 28	<i>vps28-2*</i>	SALK_040274	Spallek et al., 2013	Critical for immunity against bacterial infection through a role in stomatal closure; regulates late FLS2 endosomal sorting

<i>Vamp721</i>	At1g04750	Vesicle-associated membrane protein 721	<i>vamp721</i>	SALK_037273C	Kim et al., 2014	Forms complex with PEN1; mediates trafficking the powdery mildew resistance protein RPW8.2 to the extrahaustorial membrane of haustorial complexes
<i>Lip5</i>	AT4G26750	LYST-interacting protein 5	<i>lip5-1</i>	SAIL_854_F08	Wang et al., 2014	Disruption of <i>LIP5</i> causes increased susceptibility to the bacterial pathogen <i>P. syringae</i>

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