

1 **Polymorphism of fecundity genes (*BMP15* and *GDF9*) and their association  
2 with litter size in Bangladeshi prolific Black Bengal goat**

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18 **Abstract**

19 The primary objective of this study was to identify polymorphisms in two major fecundity genes  
20 (*BMP15* and *GDF9*) and their association with litter size in Black Bengal goat, a prolific goat breed  
21 in Bangladesh. Total 40 blood samples were collected from Black Bengal does with twining  
22 records in the first three parities. All sampled animals were genotyped for fragments of exon 2 of

23 *BMP15* gene and exon 1 and 2 of *GDF9* gene using DNA sequencing. The results of DNA  
24 sequence analysis revealed six polymorphic loci (g.735G>A, g.743C>A, g.754G>T, g.781C>A,  
25 g.808C>G and g.1061C>T) in *BMP15* gene and three (g.118C>T, g.302\_303insT and  
26 g.1173\_1174insA) in *GDF9* gene. Association analysis for polymorphic loci showed litter size in  
27 Black Bengal goat significantly varied between genotypes at g.735G>A and g.781C>A loci of  
28 *BMP15* gene. Further studies with a high number of genetically unrelated animals for assessing the  
29 association of these loci and others in the fecundity genes with litter size may be useful.

30 Keywords: Black Bengal goat; fecundity genes, polymorphisms, litter size

31 **Introduction**

32 Prolificacy (increase in litter size) is an essential economic trait in farm animals. This trait abides  
33 vital attention to the animal breeders since a small increase in offspring size can yield substantial  
34 gains in profit. Prolificacy in domestic species genetically influenced by multiple genes called  
35 fecundity genes.<sup>1</sup> Studies on the genetics of prolificacy in goat detected several candidate  
36 fecundity genes.<sup>2</sup> Bone Morphogenetic Protein 15 (*BMP15*), and Growth Differentiation Factor 9  
37 (*GDF9*) are two of them.<sup>2</sup> Several studies reported the correlation between prolificacy and  
38 genetic polymorphisms in *BMP15* and *GDF9* genes in different goat breeds around the globe.<sup>2-11</sup>

39 In Bangladesh, with a census size of 25.7 million goat stands for the third largest  
40 livestock species. Most of the goats in Bangladesh are Black Bengal goat (>90%), reared mainly  
41 by landless, small-scale farmers.<sup>12</sup> Black Bengal goat is a dwarf goat breed and is well-known  
42 for its high prolificacy, meat quality and skin quality.<sup>13</sup> The average litter size of this breed  
43 ranges from 1.93 to 2.33.<sup>12,14,15</sup> This breed also renders a significant source of red meat (25%)  
44 production in Bangladesh<sup>16</sup>. Considering its importance, increasing the numbers of black Bengal

45 goat has invariably been the central breeding goal for selective goat breeding program in  
46 Bangladesh.

47 The use of molecular markers in animal breeding has added benefits over conventional  
48 breeding techniques. The advancement in molecular genetics has led to the detection of DNA  
49 markers with considerable effects on the traits of economic importance. To the best of our  
50 knowledge, no screening attempted to identify polymorphisms in fecundity genes in goats of  
51 Bangladesh, using DNA based technologies. Therefore, this study aimed at the detection of  
52 genetic variants in fecundity genes (*BMP15* and *GDF9*) and their relationship with litter size in  
53 the only recognized native goat breed in Bangladesh, Black Bengal.

54

## 55 **Materials and Methods**

### 56 ***Sample acquisition***

57 In the present study, a total of forty (40) prolific Black Bengal does (with twining in the first  
58 three parities) were selected from two Upazilas (a sub-district unit) from Chattogram district,  
59 Bangladesh. We selected unrelated animals at random from their breeding tracts by picking up  
60 only two samples per smallholding and only five herds per village. Demographic data for all  
61 selected animal were collected using a predefined questionnaire.

### 62 ***Blood sample collection and DNA extraction***

63 In the present study, a total of forty (40) prolific Black Bengal does (with twining in the first  
64 three parities) were selected from two Upazilas (a sub-district unit) from Chattogram district,  
65 Bangladesh. We selected unrelated animals at random from their breeding tracts by picking up

66 only two samples per smallholding and only five herds per village. Demographic data for all  
67 selected animal were collected using a predefined questionnaire. Blood samples were collected  
68 aseptically from the jugular vein in a vacutainer tube containing ethylene diamine tetraacetic acid  
69 (EDTA) as the anticoagulant. All samples were delivered to the Poultry Research and Training  
70 Center (PRTC) laboratory at Chattogram Veterinary and Animal Sciences University using an  
71 icebox. Genomic DNA was extracted from the blood samples using GeneJET Genomic DNA  
72 Purification kit (Thermo Scientific, Waltham, Massachusetts, USA) according to the  
73 manufacturer's instructions. The quality of isolated DNA was investigated using agarose gel  
74 electrophoresis (0.8%).

75 ***Polymerase chain reaction (PCR) amplification***

76 All selected goats were genotyped for segments of *BMP15* (Gene ID: 100861233) and *GDF9*  
77 (Gene ID: 100860859), respectively, associated with fecundity in sheep and goat<sup>3,17-19</sup>. To  
78 amplify exon 1 of *GDF9* gene and, exon 2 of *BMP15* gene, we used two sets primers reported in  
79 the literature (Table 1).

80 Polymerase chain reaction (PCR) was performed in a final reaction volume of 25 µL on a  
81 Thermo-cycler (2720 Thermal cycler, Applied Biosystems, USA). The PCR reaction was run  
82 under the following thermal condition: initial denaturation for 1 min at 94 °C; 30 cycles of  
83 denaturation at 94 °C for 45 s, annealing at 60°C for 45 s, extension step at 72 °C for 45 s with a  
84 final extension at 72 °C for 5 min. The amplified PCR product was electrophoresed by running  
85 10 µl of through a 2% agarose gel stained with ethidium bromide (0.5µg/ml) (Sigma Aldrich,  
86 USA). The specific sizes of the fragments were distinguished by using 1kb plus DNA ladder  
87 (O, GeneRuler, 1 kb Plus, Thermo Scientific Fermentas) in the gel and visualized by a UV  
88 transilluminator gel-documentation system (BDA digital, Biometra GmbH, Germany).

89 ***Sequencing and analysis***

90 Amplified PCR products were bidirectionally sequenced in Macrogen Co., Korea. Nucleotide  
91 sequence data were edited and analysed by MEGA version 10.0.5.<sup>20</sup> The detected  
92 polymorphisms (SNPs) were compared to the *Capra hircus* nucleotide database in NCBI da-  
93 tabase using BLAST.<sup>21</sup> The sequences were submitted with the accession number of MN401415  
94 for *BMP15* and MN401414 for *GDF9* gene, respectively.

95 ***Phylogenetic analysis***

96 We performed a phylogenetic analysis of exon 2 of *BMP15* gene and exon 1 of *GDF9* gene with  
97 accessible published sequences of the same regions of genes in different goat breeds to identify  
98 the genetic diversity of Black Bengal goat. CLUSTALW<sup>22</sup> was used for multiple alignment and  
99 iTOL<sup>23</sup> to visualize the phylogenetic trees.

100 ***Statistical analysis***

101 We used SHEsis online platform (<http://analysis.bio-x.cn>)<sup>24</sup> to calculate genotype frequencies,  
102 allele frequencies and  $\chi^2$  values for Hardy–Weinberg equilibrium (HWE) test. The deviation  
103 from HWE for each polymorphism was tested using the Hardy–Weinberg  
104 law.<sup>25,26</sup> Heterozygosity (He), polymorphism information content (PIC) and effective allele  
105 number (Ne) for each polymorphism were estimated employing an online computing software  
106 (<http://www.msrgcall.com/Gdical.aspx>). For PIC, following classifications was used i) low  
107 polymorphism if PIC value  $<0.25$ , ii) moderate polymorphism if PIC value  $\geq 0.25$  to  $\leq 0.50$  and  
108 iii) high polymorphism if PIC 0.50.

109 A generalized linear model was used to analyse the association of polymorphisms in  
110 *BMP15* and *GDF9* gene with litter size by applying the least-squares method in SPSS 25  
111 statistical software, SPSS Inc., Chicago, IL, USA.

112 
$$Y_i = \mu + P_i + e_{ij}$$

113 where,  $Y_i$  is the phenotypic value of litter size;  $\mu$ , is the overall population mean;  $P_i$ , is  
114 the genotype and  $e_{ij}$ , is the random error.

115 **Results**

116 ***Sequence analysis of the BMP15 gene exon 2***

117 In a comparison of caprine *BMP15* gene sequence, we identified six single nucleotide  
118 polymorphisms (SNPs): g.735G>A, g.743C>A, g.754G>T, g.781C>A, g.808C>G (Fig.1) and  
119 g.1061C>T in the exon 2 of *BMP15* gene (Fig.1 and Supplementary Fig. S1). Five of these  
120 polymorphisms leading to amino substitutions (Table 2). The predicted possible effect of  
121 identified polymorphism on the structure and function of the functional protein revealed two  
122 mutations: g.754G>T and g.1061C>T have a significant impact on the coding BMP15 peptide  
123 (Fig. 2) indicating a substantial phenotypic effect. The g.781C>A mutation deduced to cause a  
124 moderate change while remaining two mutations did not show significant effects on the  
125 functional BMP15 protein.

126 ***Sequence analysis of the GDF9 gene exons 1 and 2***

127 The sequence of the exon 1 of *GDF9* gene showed a SNP g.118C>T and an insertion mutation  
128 g.302\_303insT. The g.302\_303insT mutation causes a frameshift in the reading frame of the  
129 *GDF9* gene which leads to a premature stop codon at third codon downstream of the mutated

130 codon (Supplementary Fig. S2). We identified a one bp insertion in between nucleotide 1173 and  
131 1174 in the exon 2 of *GDF9* gene, which deduced a premature stop codon at 16<sup>th</sup> codon  
132 downstream of the mutated codon (Supplementary Fig. S3).

133 ***Genetic parameters for the detected polymorphisms in BMP15 and GDF9 gene***

134 The genotypic and allelic frequencies, PIC, HE, Ne and  $\chi^2$  values for detected polymorphisms in  
135 two fecundity genes are presented in Table 3. The g.735G>A of *BMP15* showed all the three  
136 possible genotypic combinations while remaining SNPs recorded with two genotypes. All 40  
137 tested individuals had only homozygous mutant genotypes for g.1061C>T of *BMP15* and  
138 g.118C>T of *GDF9* gene. All tested goats expressed heterozygous genotype at g.808C>G and  
139 homozygous mutant genotype at g.1061C>T of *BMP15* gene. Genotype frequencies for most of  
140 the detected polymorphisms were significantly different from the expectations of Hardy-  
141 Weinberg equilibrium in Black goat population in this study ( $P < 0.01$ ), except g.735G>A and  
142 g.754G>T in *BMP15* gene. Three polymorphisms in *BMP15* genes (g.743C>A, g.781C>A and  
143 g.808C>G) and two in *GDF9* gene (g.302\_303insT and g.1173\_1174insA) were found to  
144 moderate polymorphic according to the classification of PIC. The remainder of the detected  
145 polymorphisms was found to be low polymorphic.

146 ***Phylogenetic analysis***

147 Results of phylogenetic analysis show that the sequences of the exon 2 of *BMP15* gene in this  
148 study clustered in a shared cluster with previously published sequences for Black Bengal goat,  
149 however, diverges from that of other goat breeds (Fig. 4). On the other hand, sequences of the  
150 exon 1 of *GDF9* gene did not form a breed-specific cluster (Fig. 5).

151 **Association between SNPs of *BMP15* and *GDF9* and litter size trait**

152 The litter size of AA genotype individuals at G735A locus of *BMP15* was significantly higher  
153 than that of GG genotype ( $p<.05$ ), however, there is no significant difference between  
154 individuals with GA and GG genotypes at this locus. Individuals with heterozygous CA  
155 genotype at C781A locus of *BMP15* had a significantly higher ( $p<.01$ ) litter size than those of  
156 CC genotype. Individuals with different genotypes at other loci in both *BMP15* and *GDF9* gene  
157 did not show any significant difference for litter size. In this study, the mean litter size was  
158 highest ( $3.04\pm0.45$ ) in individuals with heterozygous CA genotype at C781A locus of *BMP15*  
159 (Table 4).

160 **Discussion**

161 We herein reported nine polymorphic sites in two well-studied fecundity genes namely *BMP15*  
162 and *GDF9* in Black Bengal goat, the only prolific goat breed in Bangladesh. The genetic  
163 parameters of the detected loci and their association with the litter size in the Black Bengal goat  
164 were also analysed. The g.735G>A locus of *BMP15* recorded with three possible combinations  
165 of genotypes while the remaining loci expressed either two or one genotypes in the study  
166 population. The frequency distribution of genotype in a population is the simplest way to  
167 describe mendelian variation<sup>27</sup>, which might not be the fact in this study since we sequenced only  
168 40 goats for each locus. However, other factors such as such matting pattern, random genetic  
169 drift, individual survival, reproductive success, and migration also generate genetic variation in a  
170 population.<sup>28</sup>

171 In this study, four SNPs in *BMP15* gene viz. g.743C>A, 754G>T, g.781C>A and  
172 g.1061C>T were found to be novel based on an extensive literature search. Remaining two SNPs  
173 identified by us in *BMP15* gene viz g.735G>A and g.808C>G have also been reported in Black

174 Bengal goats in the neighbouring country Indian.<sup>2,4,18</sup> None of the polymorphisms in *GDF9* gene  
175 detected by us was reported so far in goat. However, several studies reported polymorphisms in  
176 the flanking sequences of *GDF9* gene.<sup>2,25,29</sup>

177 The diversity of the impact on molecular function determines the effect of a mutation.

178 Mutational effects can be neutral, harmful or beneficial depending on their context or location<sup>30</sup>.

179 In our study, two of the identified polymorphisms in *BMP15* gene predicted to have a significant  
180 effect on resulting protein sequence, hence might contribute to phenotypic change. An  
181 advantageous phenotypic effect of two insertions detected by us in *GDF9* gene might be  
182 excluded since frameshifting generally assume to cause a loss of function resulting from  
183 degrading mutant mRNA by nonsense-mediated or non-stop-mediated mRNA decay<sup>31-33</sup>.

184 However, any harmful or neutral effect<sup>34</sup> of these frameshift mutations on GDF9 protein function  
185 cannot be ruled out.

186 Phylogenetic analysis of *BMP15* gene revealed Black Bengal goat including the present  
187 study assorted in a common cluster which differs from that of other goat breeds. The *GDF9* gene  
188 sequences show more genetic diversity in different goat breeds. A similar trend of genetic  
189 divergence in the coding region of *BMP15* and *GDF9* gene has reported by Xue-qin et al.<sup>35</sup>

190 Results of association study showed that goat with AA genotype at G735A of *BMP15*  
191 had significantly higher litter size than that of GG genotypic individuals ( $p = <0.05$ ). The effect  
192 of G735A in *BMP15* gene on litter size also reported in Indian Black Bengal goat<sup>2,4</sup>. Besides, the  
193 phenotypic effects of this synonymous mutation could not be ruled out since there are many  
194 mechanisms exist by which a synonymous mutation can affect a phenotype.<sup>36</sup> In this study,  
195 Black Bengal goat with genotype CA at C781A of *BMP15* gene had significantly higher litter  
196 size than those with CC at the same locus. Ahlawat et al<sup>2,4</sup> and Feng et al<sup>37</sup> have reported a

197 significant effect of allelic variants in *BMP15* gene on the litter size in different goat breeds  
198 including Black Bengal.

199 Variations of a quantitative trait are controlled by several genes, genetic variants and their  
200 interactions. Hence, detection of polymorphisms that are underlying the differences in a  
201 quantitative trait, for instance, litter size remains as a challenge in modern genetics. *BMP15*,  
202 *GDF9* and *BMPR1B* are three well-documented candidate genes for litter size in sheep and goat.  
203 Till date, no association of *BMPR1B* gene with litter size in goat has been established. However,  
204 researchers have explored the association between litter size in goat and polymorphisms in  
205 *BMP15* and *GDF9* genes. These two genes are part of the ovary-derived transforming growth  
206 factor  $\beta$  (TGF  $\beta$ ) that have an integral role as growth factors and receptors in follicular  
207 development in the ovaries.<sup>38</sup> Both bone morphogenetic proteins (BMPs) and growth  
208 differentiation factors (GDFs) have critical role in follicle growth and cell-survival signalling  
209 hence causal mechanism underlying the high prolificacy or fertility in female animal.<sup>39</sup>  
210 Considering the biological importance of *BMP15* and *GDF9* genes, this study investigated  
211 polymorphisms in these genes and their association with the litter size in Bangladeshi prolific  
212 Black Bengal goat.

## 213 Conclusion

214 This is the first study exploring polymorphisms in *BMP15* and *GDF9* genes and investigating  
215 their association with litter size in Bangladeshi goat. The findings of the study reveal that  
216 different genotypes at two loci in *BMP15* gene had significant ( $p \leq 0.05$ ) effect on litter size in  
217 the prolific Black Bengal breed of Bangladesh. Hence, there is a need for further research with a  
218 substantially large number of animals across a wide range of geographically divergent  
219 populations of this breed. Our results enrich the repository of molecular markers database of

220 caprine fecundity genes which pave the way for association studies with fecundity trait, hence  
221 contribute to molecular breeding in goat.

222 **Data availability**

223 The assembled and annotated sequences for *BMP15* and *GDF9* genes were deposited to  
224 GenBank database under accession numbers MN401414 and MN401415, respectively.

225 **Conflict of Interest**

226 The authors declare that they have no competing financial interests.

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230 for marker-assisted selection in Black Bengal goat’.

231 **Author contributions**

232 A. Das conceived this project. M. Shaha performed sample collection. M. Shaha and A. Dutta  
233 extracted genomic DNA and performed PCR. A. Das and M. Das Gupta performed data curation  
234 and formal data analysis. A. Das and O. F. Miazi involved in funding acquisition, project  
235 administration and supervision. A. Das drafted the original manuscript. All authors read and  
236 approved the final manuscript.

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**Table 1.** List of primer used to amplify specific segment of *GDF9* and *BMP15* gene.

Gene	Region	Oligonucleotide sequence (5'-3')	Amplicon size (bp)
<i>BMP15</i>	Exon2	F: CACTGTCTTCTTGTACTGTATTCATGAC	141
		R: GATGCAATACTGCCTGCTTG	
	Exon1	F: TCCCTAAAGGCCTGAAAGAGT	575
		R: GCTGAAGGCAAGGAATAGAAC	
<i>GDF9</i>	Exon1	F: GAAGACTGGTATGGGGAAATG	462
		R: CCAATCTGCTCCTACACACCT	
	Exon2	F: CCACACAAATACAACCCCTCGATAC	183
		R: AGGCTCGATGCCAAACACT	

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**Table 2.** Identified polymorphisms in *GDF9* and *BMP15* genes in Bangladeshi Black Bengal goat breed.

Gene	Region	Mutation	Amino acid substitution	Type of mutation
<i>BMP15</i>	Exon2	g.735G>A	-	Synonymous
		g.743C>A	p.Pro248His	Non-synonymous
		g.754G>T	p. Gly252Cys	Non-synonymous
		g.781C>A	p. Pro261Thr	Non-synonymous
		g.808C>G	p.Gln270Glu	Non-synonymous
		g.1061C>T	p.Ala354Val	Non-synonymous
<i>GDF9</i>	Exon1	g.118C>T	-	Synonymous
		g.302_303insT	p.Gly101Glyfs2X	Frameshift
	Exon2	g.1173_1174insA	p.Arg392Glnfs16X	Frameshift

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342 **Table 3.** Genotypic and allelic frequencies and population parameters for mutations detected in *GDF9* and *BMP15* genes in Bangladeshi Black  
 343 Bengal Goat breed (N=40).

Gene	Mutation	Genotypic frequencies			Allelic frequencies		PIC	He	Ne	H-W ( $\chi^2$ )	test	p value
		PP	Pq	qq	P	q						
<i>BMP15</i>	g.735G>A	0.625	0.275	0.100	0.762	0.237	0.237	0.361	1.567	2.317	-	0.127
	g.743C>A	0.300	0.700	0	0.650	0.350	0.351	0.455	1.834	11.597	-	0.000
	g.754G>T	0.700	0.300	0	0.85	0.15	0.222	0.255	1.342	1.245	-	0.264
	g.781C>A	0.450	0.550	0	0.725	0.275	0.319	0.398	1.663	5.755	-	0.016
	g.808C>G	0	1.000	0	0.500	0.500	0.375	0.500	2.000	40.000	-	0.000
	g.1061C>T	0	0	1.000	0	1.000	0	0	1.000	-	-	-
<i>GDF9</i>	g.118C>T	0	0	1.000	0	1.000	0	0	1.000	-	-	-
	g.302_303insT	0.200	0.800	0	0.600	0.400	0.364	0.480	1.923	17.778	-	0.000
	g.1173_1174insA	0.300	-	0.700	0.300	0.700	0.331	0.420	1.724	39.999	-	0.000

344 PP, homozygous reference allele, Pq, heterozygous mutant allele; qq, homozygous mutant allele; PIC, polymorphism information content; He,  
 345 effective number of heterozygosities; Ne, effective number of alleles.

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**Table 4.** Genotype wise least squares mean  $\pm$  standard error of litter size in Bangladeshi Black Bengal goat.

Gene	Locus	Genotype	N	Litter size	Significance
<i>BMP15</i>	G735A	GG	25	3.00 <sup>a</sup> $\pm$ 0.33	*
		GA	11	2.52 <sup>ab</sup> $\pm$ 0.13	
		AA	4	2.27 <sup>b</sup> $\pm$ 0.20	
	C743A	CC	12	2.42 $\pm$ 0.20	NS
		CA	28	2.53 $\pm$ 0.13	
		AA	0	-	
	G754T	GG	28	2.39 $\pm$ 0.14	NS
		GT	12	2.75 $\pm$ 0.13	
		TT	0	-	
<i>GDF9</i>	C781A	CC	18	1.83 <sup>a</sup> $\pm$ 0.09	***
		CA	22	3.04 <sup>b</sup> $\pm$ 0.45	
	302_303insT	GG	8	2.47 $\pm$ 0.12	NS
		GGT	32	2.63 $\pm$ 0.24	
<i>GDF9</i>	1173_1174insA	TT	7	2.86 $\pm$ 0.69	NS
		TTA	28	2.42 $\pm$ 0.66	

353 \*, p value  $<0.05$  \*\*\*, p value  $<0.01$ ; rows with different superscripts differed significantly.

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360 **Figure Legends**

361 **Figure 1.** Sequencing chromatograms of the detected single nucleotide polymorphisms (SNPs) in exon 2 of the *BMP15* gene in Black Bengal  
362 goat breed. a. g.735G>A (heterozygous); b. g.743C>A (heterozygous); c. g.754G>T (heterozygous); d. g.781C>A (heterozygous); d. g.808C>G  
363 (heterozygous) and f. g.1061C>T (homozygous TT). Positions of the mutations are based on the full sequences of the *BMP15* gene (Gene ID:  
364 100861233).

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366 **Figure 2.** Sequencing chromatograms of the detected polymorphisms in the *GDF9* gene in Black Bengal goat breed. a. g.118C>T (homologous  
367 TT) and b. 302\_303insT in the exon 1 of *GDF9* gene; c. g.1173\_1174insA in the exon 2 of *GDF9* gene. Positions of the mutations are based on  
368 the full sequences of the *GDF9* gene (Gene ID: 100861233).

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370 **Figure 3.** Predicted effects (using Ployphen2) of five single nucleotide polymorphisms (SNPs). a, b, c, d and e represent the of effect of  
371 g.743C>A, g.754G>T, g.781C>A, g.808C>G and g.1061C>T on the functional BMP15 protein, respectively.

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373 **Figure 4.** Phylogenetic tree of *BMP15* gene.

374 **Figure 5.** Phylogenetic tree of *GDF9* gene.

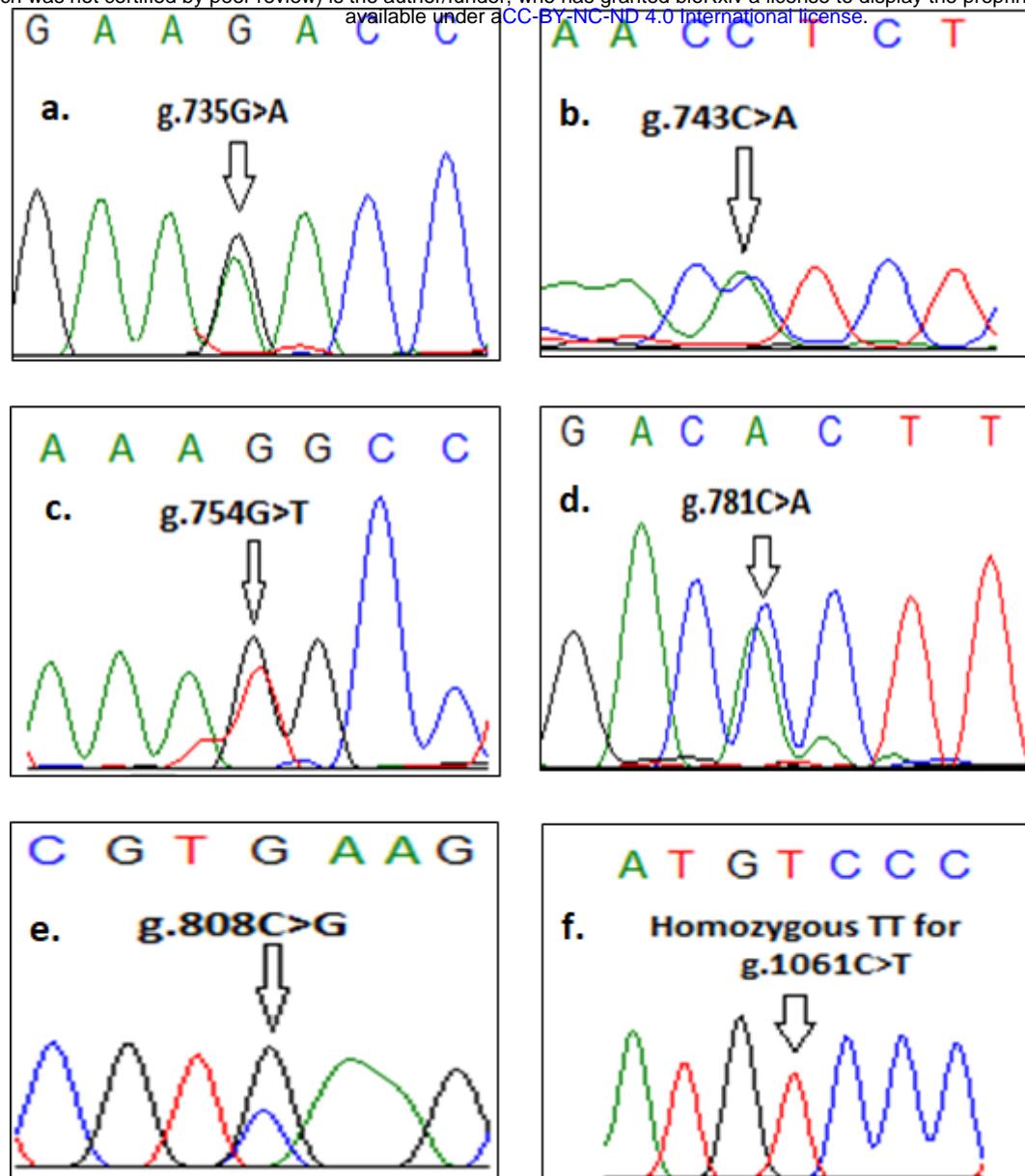
375 **Supplementary Figure S1.** Multiple alignment for g.1061C>T of *BMP15* gene (indicated by box arrow). *BMP15* gene sequences from this study  
376 aligned with BMP15 mRNA sequence from the NCBI database using MEGA version 10.0.5.<sup>20</sup>

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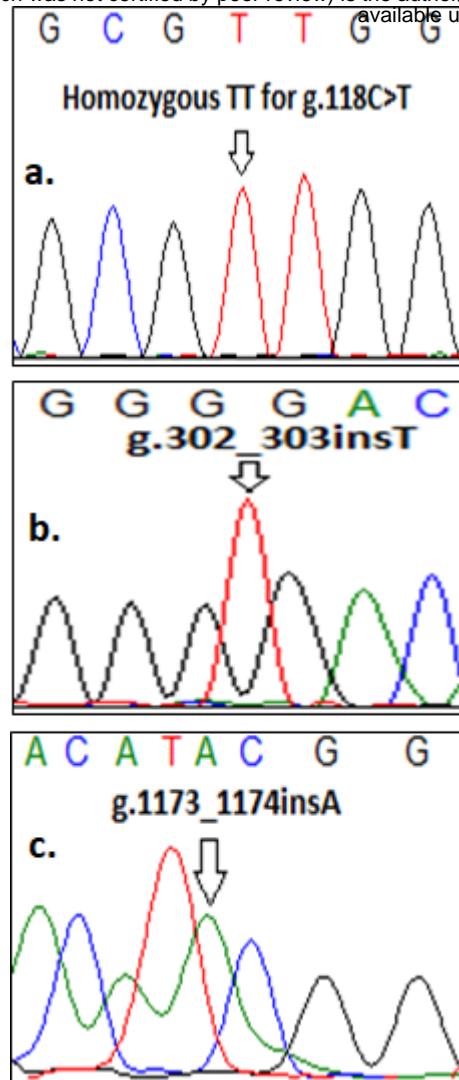
378 **Supplementary Figure S2.** Multiple alignment for 302\_303insT of *GDF9* gene (Condon changes are indicated by box arrow). *GDF9* gene  
379 sequences from this study aligned with GDF9 mRNA sequence from the NCBI database using MEGA version 10.0.5.<sup>20</sup>

380

381 **Supplementary Figure S2.** Multiple alignment for 302\_303insT of *GDF9* gene (Condon changes are indicated by box arrow). *GDF9* gene  
382 sequences from this study aligned with GDF9 mRNA sequence from the NCBI database using MEGA version 10.0.5.<sup>20</sup>



**Figure 1.** Sequencing chromatograms of the detected single nucleotide polymorphisms (SNPs) in exon 2 of the *BMP15* gene in Black Bengal goat breed. a. g.735G>A (heterozygous); b. g.743C>A (heterozygous); c. g.754G>T (heterozygous); d. g.781C>A (heterozygous); d. g.808C>G (heterozygous) and f. g.1061C>T (homozygous TT). Positions of the mutations are based on the full sequences of the *BMP15* gene (Gene ID: 100861233).



**Figure 2.** Sequencing chromatograms of the detected polymorphisms in the *GDF9* gene in Black Bengal goat breed. a. g.118C>T (homologous TT) and b. 302\_303insT in the exon 1 of *GDF9* gene; c. g.1173\_1174insA in the exon 2 of *GDF9* gene. Positions of the mutations are based on the full sequences of the *GDF9* gene (Gene ID: 100861233).

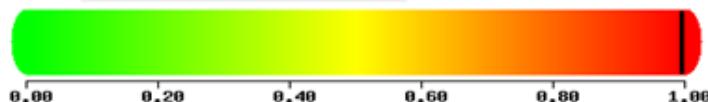
This mutation is predicted to be **BENIGN** with a score of 0.022 (sensitivity: 0.95; specificity: 0.80)

a.



This mutation is predicted to be **PROBABLY DAMAGING** with a score of 0.996 (sensitivity: 0.55; specificity: 0.98)

b.



This mutation is predicted to be **BENIGN** with a score of 0.450 (sensitivity: 0.89; specificity: 0.90)

c.



This mutation is predicted to be **BENIGN** with a score of 0.001 (sensitivity: 0.99; specificity: 0.15)

d.

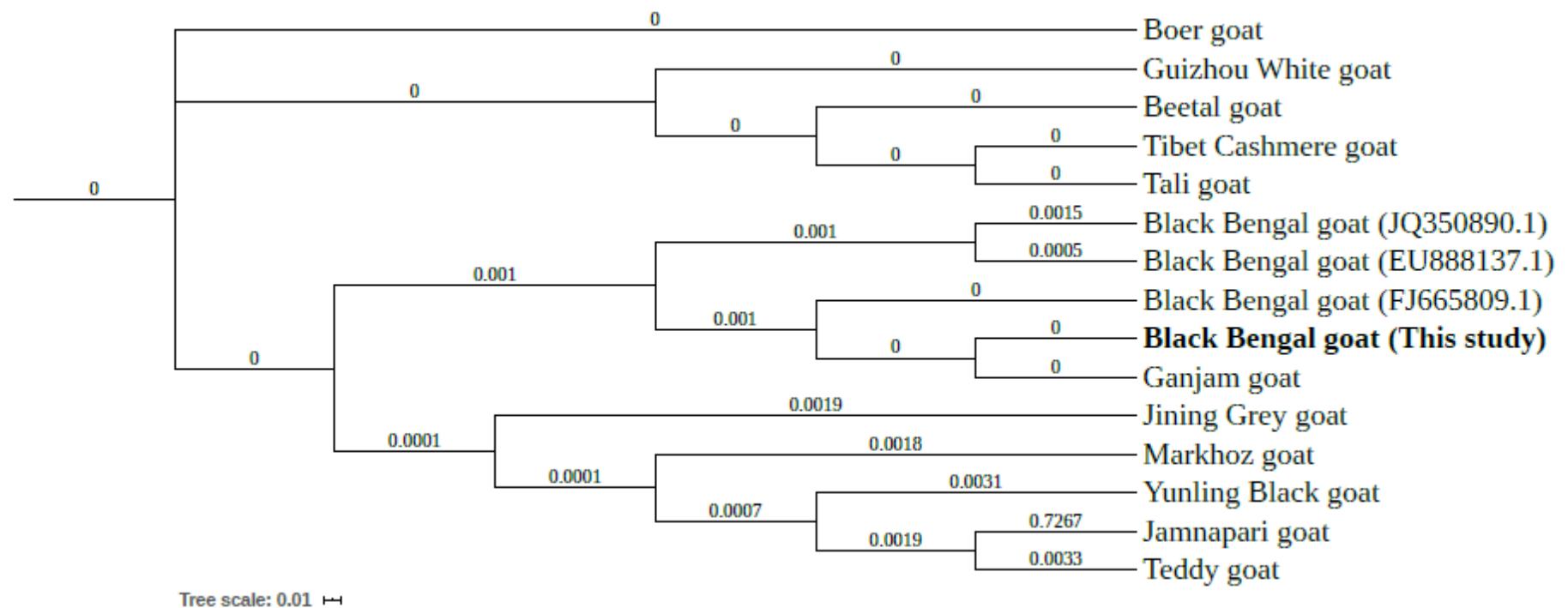


This mutation is predicted to be **PROBABLY DAMAGING** with a score of 0.999 (sensitivity: 0.14; specificity: 0.99)

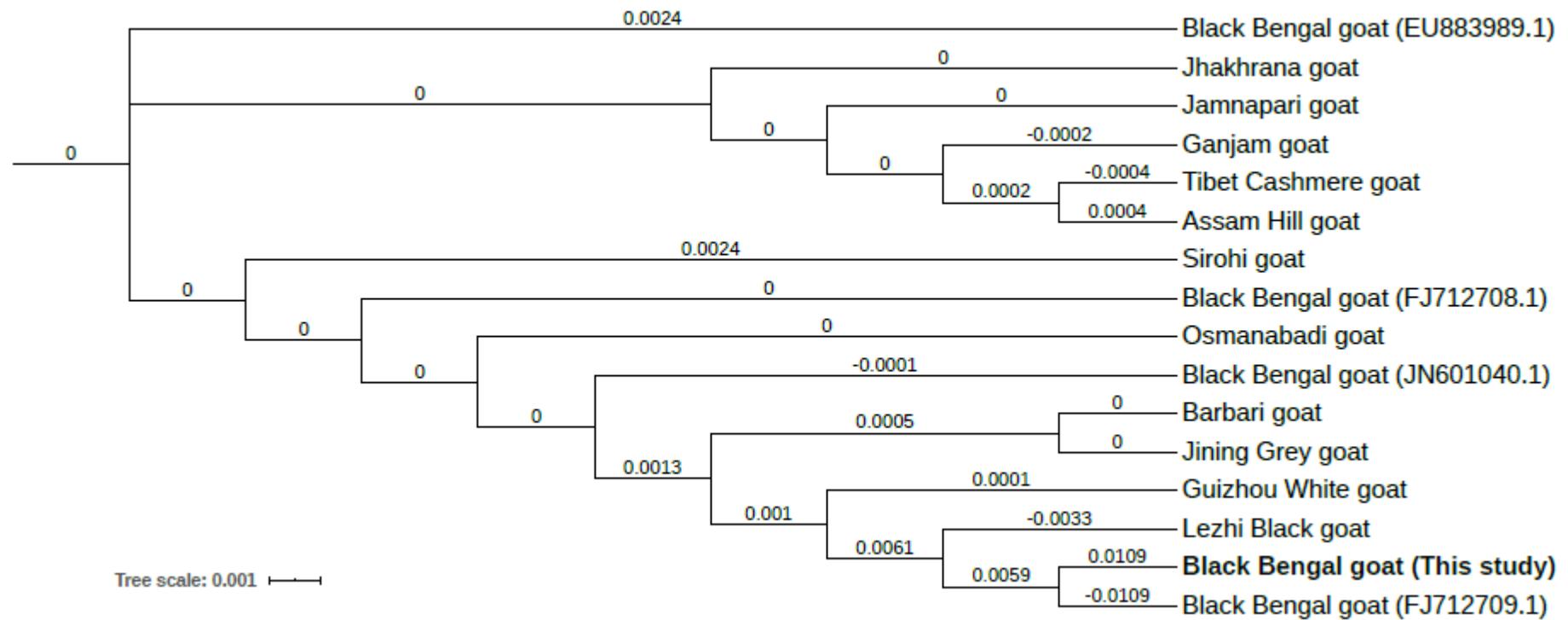
e.



**Figure 3.** Predicted effects (using Ployphen2) of five single nucleotide polymorphisms (SNPs). a, b, c, d and e represent the effect of g.743C>A, g.754G>T, g.781C>A, g.808C>G and g.1061C>T on the functional BMP15 protein, respectively.



**Figure 4.** Phylogenetic tree of *BMP15* gene.



**Figure 5.** Phylogenetic tree of *GDF9* gene.