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1 ***Title***

2 Renewed diversification following Miocene landscape turnover in a Neotropical butterfly radiation

3

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23 ***Abstract***

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26 The Neotropical region has experienced a dynamic landscape evolution throughout the Miocene, with
27 the large wetland Pebas occupying western Amazonia until 11-8 my ago and continuous uplift of the
28 Andes mountains along the western edge of South America. Although the complex dynamics between
29 the Andes and Amazonia may have strongly affected the trajectory of Neotropical biodiversity, there
30 is little evidence for such an influence from time-calibrated phylogenies of groups that diversified
31 during this period. Here, we generate one of the most comprehensive time-calibrated molecular

32 phylogenies of a group of Neotropical insects: the butterfly tribe Ithomiini. Our tree includes 340
33 species (87% of extant species), spanning 26 million years of diversification in the Neotropics. We
34 investigate temporal and spatial patterns of diversification, focusing on the influence of Miocene
35 landscape transformations on the dynamics of speciation, extinction and biotic interchanges at the
36 Amazonia/Andes interface. We find that Ithomiini likely began diversifying at the interface between
37 the Andes and the Amazonia around 26.4 my ago. Five subtribes with a very low extant diversity
38 started diversifying early in western Amazonia, but a rapid decrease in diversification rate due to
39 increased extinction rate between 20 and 10 my ago suggests a negative impact of the Pebas wetland
40 system on these early lineages. By contrast, the clade containing the five most species-rich subtribes
41 (85% of extant species) was characterized by a high, positive net diversification rate. This clade
42 diversified exclusively in the Central Andes from 20 to 10 my ago. After the demise of the Pebas
43 system (11-8 my ago), we found a sudden increase of interchanges with the Northern Andes and
44 Amazonia, followed by local diversification, which led to a substantial renewal of diversification. In
45 general, ecological turnovers throughout the Miocene strongly determined the dynamics of speciation,
46 and extinction and interchanges, and appear as a key driving force shaping the region's current
47 extraordinary biodiversity.

48

49 **Keywords**

50 Neotropics, phylogeny, diversification, biogeography, Andes, Pebas system, Western-Andean Portal,
51 Ithomiini, butterflies

52 INTRODUCTION

53 There has been a long fascination among biologists for the Neotropics and the origin of its intriguingly
54 high biodiversity. The timing of Neotropical diversification, and therefore its major driving processes,
55 is still controversial despite the large amount of publications that have addressed the question (e.g.,^{1, 2,}
56 ³).

57 Despite uncertainty about the precise timing and magnitude of surface uplift, the formation of the
58 Andean cordilleras during the Cenozoic greatly shaped Neotropical landscapes and affected
59 diversification in the Neotropics. As the Andes arose, they brought about new biotic and abiotic
60 conditions along their slopes, modified the climate of the Neotropical region and deeply affected the
61 formation of the Amazonian basin by bringing large amounts of sediments and modifying water
62 drainage ¹. There is increasing evidence that the Andes influenced the diversification of Neotropical
63 lineages, primarily by increasing speciation rate, perhaps most spectacularly in the high altitude
64 páramo habitat (e.g., ⁴). In parallel, the western part of the Amazon basin, which is connected to the
65 Andes, has experienced major turnovers of ecological conditions. During the Oligocene, western
66 Amazonia was occupied by a fluvial system flowing northward (paleo-Orinoco basin), which
67 transformed ~23 million years (my) ago into an aquatic system of shallow lakes and swamps
68 episodically invaded by marine conditions, known as the Pebas system ^{1, 5, 6, 7, 8}. The Pebas was
69 connected northward with the Caribbean Sea and likely also with the Pacific Ocean through the
70 Western-Andean portal (“WAP”, ⁵), a low-altitude gap that separated the Central Andes and the
71 Northern Andes until 13-11 my ago. During the late Miocene and during the Andean uplift, the
72 accumulation of sediments combined with a sea level decrease initiated the eastward drainage of the
73 Pebas, and by 10-8 my ago the region had changed into a fluvial system, which then turned into the
74 modern configuration of the Amazon. More recently, climatic fluctuations during the Peistocene (2.5-
75 0 my ago) may have led to episodic dryness affecting Amazonian forest habitats ⁹. The extent of the
76 influence of Pleistocene events and their effects on Neotropical diversification, and even the
77 importance of dryness episodes, are controversial (e.g., ^{2, 10, 11, 12}).

78 In this study we purposely focus our attention mostly on the Miocene and Pliocene and how the
79 interaction between the rise of the Andes and coincident large landscape modifications in western
80 Amazonia have determined diversification and dispersal over 30 million years. The Pebas ecosystem
81 covered up 1.1 million km² at its maximum ⁷ and was probably not suitable for a terrestrial fauna.
82 Therefore, between 23 and 10 my ago, diversification of terrestrial lineages may have been impeded in
83 western Amazonia or restricted to its edges ⁷. By contrast, the uplift of the Central and the Northern
84 Andes, also occurring throughout the Miocene and the Pliocene, and the ecological gradients present
85 along this mountain chain probably constituted an important driver of diversification. In the last 10 to
86 8 my, the retreat of the Pebas may have provided opportunities for terrestrial lineages to radiate in

87 western Amazonia. The Pebas may also have constrained dispersal, acting as a barrier between the
88 Andes, the Guiana shield and western Amazonia. Thus, determining whether rates of interchanges
89 have been constant throughout time since the origin of Ithomiini, or instead have increased after the
90 Pebas's retreat, would test the potential importance of this ecosystem in building the modern pattern of
91 diversity⁶.

92 Paleontological studies have shown that the Pebas greatly contributed to the diversification of aquatic
93 fauna such as molluscs¹³, ostracods¹⁴ and crocodilians¹⁵. However, the fossil record also suggests a
94 negative effect of the Pebas system on terrestrial fauna^{16, 17}. The hypothesis that the Pebas has shaped
95 patterns of terrestrial diversification and dispersal in western Amazonia has grown over the years (e.g.,
96 5, 6, 7, 16, 18) but support from molecular phylogenies mostly stems from the observation that western
97 Amazonian clades have diversified during the last 10-8 my and not before (19⁶ and reference therein).
98 Yet, there is very little information on what happened before, when the Pebas was occupying western
99 Amazonia, particularly on whether the presence of Pebas constrained diversification and interchange
100 patterns in this region. A thorough assessment of the role of the Pebas ecosystem on diversification
101 and dispersal requires phylogenies of large Neotropical clades that originated before the formation of
102 the Pebas, i.e. clades older than 23 my old. Phylogenies of Neotropical clades meeting these
103 conditions are surprisingly rare. In insects – which are among the most diverse terrestrial organisms -
104 attempts to build phylogenies of Neotropical groups to test different drivers of Neotropical
105 diversification have either suffered from a small size or a low sampling fraction (e.g. 19, 20, 21, 22, 23), and
106 therefore from low statistical power and reliability.

107 Butterflies are among the best candidates for addressing the evolution of the Neotropical biota at such
108 time scales. They are probably the best-known insects and Neotropical butterfly lineages have
109 benefited from substantial phylogenetic research compared to other insects (e. g., 19, 20, 21, 24, 25, 26, 27, 28, 29,
110 30, 31, 32, 33, 34, 35). Among the most emblematic Neotropical butterflies is the tribe Ithomiini
111 (Nymphalidae: Danainae, 393 species), also referred to as the clearwing butterflies because of the
112 transparent wings of the majority of species. Ithomiini are forest-dwellers distributed throughout the
113 Neotropics, from sea level up into montane cloud forests (to 3000 m), where their larvae feed on
114 plants of the families Solanaceae, Gesneriaceae and Apocynaceae³⁶. Species richness is primarily
115 concentrated in the Andes, where about half of the species occur (mostly on the eastern slopes) and in
116 western Amazonia. Ithomiini are chemically defended and they engage in Müllerian mimicry,
117 whereby co-occurring species exhibit convergent wing colour patterns that advertise their toxicity to
118 predators³⁷. Ithomiini butterflies represent a keystone group in Neotropical forests by numerically
119 dominating mimetic butterfly communities and sharing wing colour patterns with a large number of
120 other palatable and unpalatable Lepidoptera, such as the iconic *Heliconius* butterflies³⁸. For this
121 reason, Ithomiini were used by both Bates³⁹ and Müller³⁷ in their original descriptions of deceptive
122 (Batesian) and mutualistic (Müllerian) mimicry, respectively.

123 The diversity and the intriguing ecology of Ithomiini has generated a great interest and a broad and
124 diverse literature on topics including life history ^{40, 41, 42, 43}, chemical ecology ^{44, 45, 46}, systematics ^{19, 21,}
125 ^{26, 32, 34, 36, 47, 48}, cytogenetics ⁴⁹, community ecology ^{50, 51, 52, 53, 54, 55}, wing colour pattern evolution ⁵⁶, and
126 biogeography ^{19, 21, 26, 27}. In this study, we build on existing molecular data and provide a large amount
127 of novel DNA sequences for ithomiine species to generate the first species-level phylogeny of the
128 entire tribe, providing a large and densely sampled (340 species included out of 393 currently
129 recognized) phylogenetic dataset for a Neotropical insect clade that underwent diversification during
130 the last ~30 million years ^{27, 57}. This phylogenetic framework provides an ideal opportunity for
131 investigating Neotropical diversification over a large evolutionary time-scale. Ithomiini originated
132 before the Pebas system, thus offering the opportunity to investigate diversification before, during and
133 after the environmental changes during the Miocene with a high statistical power. Importantly,
134 contrary to many other large radiations of similar age, Ithomiini are endemic to the Neotropics. Their
135 diversification therefore occurred without interaction with other biogeographic regions such as the
136 Nearctic.

137 Here, we investigated the dynamics of speciation, extinction and dispersal rates in Ithomiini through
138 space and time, using a combination of time- and trait- dependent models of diversification and
139 historical biogeography. We focused on the interaction between the turnover of ecological conditions
140 in western Amazonia and the Andean uplift during the Miocene, and we investigated whether
141 geological and ecological events in both regions affected synergistically the diversification of
142 Ithomiini. More specifically, an important role for Andean uplift and the Pebas would be supported if:
143 (1) *During the Pebas period*: (a) Andean diversification largely exceeds Amazonian diversification,
144 due to an increased diversification in the Andes driven by the evolving ecological gradient and uplift
145 dynamics and/or a reduced diversification rate in Amazonia accompanying the loss of terrestrial
146 habitats; (b) interchanges with western Amazonia are reduced; (c) interchanges between the Central
147 and the Northern Andes are reduced, because of the existence of the WAP. (2) *During the retreat of*
148 *the Pebas*: interchanges with western Amazonia and between the Central and the Northern Andes
149 largely increase, as a result of new terrestrial habitats and the disappearance of the WAP, respectively.
150 (3) *After the Pebas period*: Diversification rates in Amazonia globally increase and biotic interchanges
151 are not constrained anymore. After the Pebas retreat, decrease of speciation rates through time suggest
152 post-Pebas radiations in Amazonia, while increase of speciation rates through time may suggest a role
153 of climatic fluctuations during the last 2.5 my.

154

155 RESULTS

156 Time-calibrated phylogeny

157 *Tree topology and time-calibration*

158 We generated a time-calibrated phylogeny that comprised 340 out of 393 Ithomiini species
159 (Supporting Information S1-S2-S3), based on three mitochondrial and six nuclear gene fragments. The
160 tree topology was obtained under maximum likelihood inference, and branch lengths were estimated
161 by Bayesian inference, using six secondary calibration points from Wahlberg *et al.* (57). The tree
162 topology was generally well supported, including deep nodes (Supporting Information S4). We found
163 a crown age of Ithomiini of 26.4 (CI=22.75-30.99) my ago (Figure 1, Supporting Information S5) and
164 a divergence time from its sister clade Tellervini of 42.1 (CI=39.50-48.44) my ago. All subtribes (10
165 in total) diverged in the first 10 million years, in the following order (Figure 1): (1) Melinaeina (26.4,
166 CI=22.75-30.99 my ago), (2) Mechanitina (24.2, CI=21.00-28.59 my ago), (3) the clade consisting of
167 Tithoreina and the Methonina (23.6, CI=20.40-27.71 my ago), (4) Athesitina (22.1, CI=19.09-26.25
168 my ago) and (5) a large clade that comprises the five most species-rich subtribes (Ithomiina,
169 Napeogenina, Oleriina, Dircennina and Godyridina), hereafter called the “core-group”. The
170 relationships between tribes were similar to those recovered in a recent higher-level phylogeny of
171 Ithomiini based on a combination of 3 gene regions and morphological characters 34, except that
172 Brower *et al.* (34) recovered Mechanitina as sister to Tithoreina+Methonina. Lineage ages in our
173 phylogeny were generally younger than those inferred in Wahlberg *et al.* (57), but older than those
174 inferred in Garzon-Orduña *et al.* (58) (see De-Silva *et al.* 48 for further discussion of such differences).

175 **Diversification rates**

176 *Time-dependent diversification*

177 We combined three methods to assess patterns of diversification through time and across clades. We
178 first used MEDUSA 59, which automatically detects shifts of diversification processes across a
179 phylogeny. The analysis detected two significant shifts from the background process of diversification
180 (Figure 1, Supporting Information S6). One was at the root of the core-group accounting for ~85% of
181 present-day diversity of Ithomiini. The second shift was at the root of a subgroup within the genus
182 *Melinaea*, which appears to have diversified rapidly during the last one million years (hereafter
183 “*Melinaea*-group”).

184 We then used the method developed by Morlon *et al.* 60 to fit time-dependent models of speciation and
185 extinction on the different partitions based on the MEDUSA results. This method does not detect shifts
186 automatically, but it allows both speciation and extinction rates to vary through time and across
187 lineages. The results confirmed that the partitioned models (either or both of the two shifts detected by
188 MEDUSA) had a significantly better fit and that the two-shift model was significantly better than the
189 one-shift models (Table 1). For the core-group, no model of time-dependent diversification had a
190 significantly better fit than the null model of constant speciation rate without extinction (0.255 lineage-

191 my^{-1} , Figure 2, Table 1, Supporting Information S6). Under the null model, the core-group diversity
192 increased exponentially during the last 20 million years and reached its current diversity (334 extant
193 species). For the *Melinaea*-group, the best fitting model was an exponentially decreasing speciation
194 rate without extinction, with a very high initial speciation rate (7.62 lineage $^{-1}\text{my}^{-1}$ at the root, 0.342
195 lineage $^{-1}\text{my}^{-1}$ at present, Figure 2, Table 1, Supporting Information S6). The *Melinaea*-group radiated
196 into eight species during a time lapse of only one million years. On the remaining background
197 lineages, the best fitting model involved a time-dependency of both speciation and extinction rates.
198 The resulting diversification rate was high during the initial stages of diversification (0.75 at the root),
199 but decreased rapidly and became negative around 19 my ago. The background diversification rate
200 then started a slow recovery, but remained negative (-0.038 lineage $^{-1}\text{my}^{-1}$ at present, Figure 2, Table 1,
201 Supporting Information S6). Consequently the background tree diversity reconstructed from this
202 model shows a pattern of diversity that increased during the first 10 my up to ~60 species before
203 slowly declining toward its current diversity (51 species) during the last 15 my. This signal of high
204 relative extinction rate was not affected by initial parameters for maximum likelihood search or by
205 diversification rate heterogeneity potentially remaining from the background (see discussion in
206 Supporting Information S7). Clades within the background tree showing positive net diversification
207 rates during the last 5 my supported the recovery trend described above (Supporting Information S7).

208 We also used TreePar⁶¹ to fit models of constant diversification rate in 5 my time-bins for each
209 partition identified by MEDUSA, as a second and independent assessment of diversification through
210 time. TreePar accommodates constant birth-death models within time-intervals and allows those birth-
211 death rates to vary between time intervals. The results were congruent with those of the time-
212 dependent models of diversification obtained with Morlon *et al.*⁽⁶⁰⁾'s method (Figure 2, Supporting
213 Information S7). The diversification rates estimated for the core-group within 5 my time-bins
214 remained relatively constant through time and the turnover rate was close to 0, supporting a null or
215 extremely low relative extinction. For the background tree, we found that diversification rate was
216 highest between 25 and 20 my ago, declining toward 0 during the last 15 my, in agreement with the
217 results obtained with the method of Morlon *et al.*⁽⁶⁰⁾. Turnover largely increased and reached a
218 maximum during the last 15 million years, supporting a very high relative extinction rate, similarly to
219 our results of the time-dependent diversification models of Morlon *et al.*⁽⁶⁰⁾'s method.

220 We additionally performed an analysis with BAMM v.2.5.0⁶², which also detects shifts in
221 diversification dynamics using a Bayesian framework. The results supported the presence of a shift at
222 the root of the core-group with an increasing diversification rate compared to the lineages diverging
223 earlier. More details about this analysis can be found in Supporting Information S8.

224

225 *Diversification in the Andes*

226 We compared the pattern of diversification in the Andes and in the rest of the Neotropics and we
227 assessed the rate of interchanges between these regions using character state-dependent diversification
228 models (ClaSSE, ⁶³). Such models estimate character state-dependent rates of speciation, extinction
229 and cladogenetic state transitions (i. e., occurring at nodes). Species were classified into two
230 biogeographical states, Andean and non-Andean. We compared 10 models to test whether rates of
231 speciation, extinction or transitions were different or not between regions. For both the whole
232 Ithomiini and the core-group we failed to identify a single model that fitted the data significantly better
233 than other models (Table 2). For the whole Ithomiini, the model with the lowest AIC score had a
234 higher speciation rate in the Andes ($\lambda_{222}=0.230$) than in non-Andean regions ($\lambda_{111}=0.118$) and those
235 speciation rates were higher than the colonization rates ($\lambda_{112}=\lambda_{212}=0.079$). The second best model had,
236 again, a higher speciation rate in the Andes ($\lambda_{222}=0.231$) than in non-Andean regions ($\lambda_{111}=0.108$) and
237 the colonization rate out of the Andes was higher ($\lambda_{212}=0.095$) than into the Andes ($\lambda_{212}=0.047$). In the
238 core-group, there were four models within an AIC difference of 2. All models had extremely low
239 extinction rates. Only one of them inferred different speciation rates and three of them inferred a
240 higher colonization rate into the Andes than out of the Andes (Table 2). Therefore, there is no support
241 for a general pattern of increasing diversification in the Andes across the entire core-group (~85% of
242 the extant diversity). This interpretation was confirmed by additional analyses in which we tested the
243 presence of a hidden character using the model HiSSE ⁶⁴. The results supported the conclusion that the
244 Andes are not directly associated with higher diversification rates (see Supporting Information S9),
245 and that diversification was also fast in some non-Andean clades and slow in some Andean lineages.

246 Ancestral state inference is not implemented in ClaSSE. Instead, we used the BiSSE ⁶⁵ model, in
247 which transitions occur only along branches. To infer ancestral states on the whole tree, we fitted the
248 BiSSE models corresponding to the best-fitting ClaSSE models (i.e. model1: different speciation rates,
249 model2: different speciation and transition rates). In both cases the most likely state of the root was
250 non-Andean but with high uncertainty (probability of 0.508 and 0.543 respectively) and there was
251 uncertainty at the nodes leading to the core-group (Figure 1, Supporting Information S10). The most
252 recent common ancestor (hereafter MRCA) of the core-group was inferred as most likely Andean
253 (model1: probability of 0.558, model2: probability of 0.625). In both models the MRCA of all
254 background subtribes were inferred to be non-Andean with a strong support (except for Athesitina,
255 Figure 1, Supporting Information S10). In the core-group, the best model (with different speciation
256 rates) inferred an Andean origin for subtribes Napeogenina, Dircennina and Godyridina, whereas the
257 second best model (different speciation and transition rates) inferred an Andean origin for all five
258 subtribes (Supporting Information S10).

259 *Diversification in Amazonia*

260 We also investigated the pattern of Amazonian diversification during the post-Pebas period (8-0 my
261 ago). We fitted a model of time-dependent speciation rate using Morlon et al.'s method ⁶⁰ (no
262 extinction, based on our previous results) to assess whether speciation rates decreased through time
263 (supporting radiations accompanying the post-Pebas recolonizations) or increased through time
264 (supporting a recent diversification potentially caused by Pleistocene climatic fluctuations). We
265 identified four Amazonian diversification events – clades whose nodes were inferred to be almost all
266 Amazonian (see historical biogeography results below) – in the core-group and three in the
267 background subtribes. All core-group clades showed decreasing speciation rates through time,
268 suggesting an early diversification, perhaps following the appearance of new forest habitats
269 accompanying the Pebas retreat. Among the three clades from the background lineages only the genus
270 *Methona* followed a trend of decreasing speciation rate. The other two clades, *Mechanitis* + *Forbestra*
271 and *Melinaea* (the whole genus) supported an increasing speciation rate through time, which is
272 consistent with a potential effect of Pleistocene climatic fluctuations in driving diversification.

273

274 **Historical biogeography**

275 We divided the Neotropical region into 9 areas (Supporting Information S11) and we assigned each
276 ithomiine species to these areas according to their current distribution. We performed historical
277 biogeographic reconstruction using BioGeoBEARS ⁶⁶, under two models (DEC and DIVALIKE),
278 using a three-step procedure outlined below. We first performed a “null” model with uniform dispersal
279 multipliers. Based on the results of the null model, we computed rates of dispersal between specific
280 regions for all 1-my intervals. We then implemented those rates in a time-stratified model.

281 *Biogeographic null model*

282 We started with a “null” biogeographic model, which restricted the area adjacency but set all dispersal
283 probabilities to 1, and we compared the models DEC and DIVALIKE. The null DEC model had a
284 better fit than the null DIVALIKE model (likelihoods: DECnull: -1335.802, DIVALIKEnull: -
285 1347.869), hence we used the DEC model in all subsequent analyses.

286 In both models, the ancestral area of the Ithomiini MRCA was unclear (Central Andes + Western
287 Amazonia for the highest probability). The areas where the two first divergences occurred, which led
288 to the Melinaeina and the Mechanitina subtribes, were also unclear (Figure1, Supporting Information
289 S12). The ancestor of the remaining ithomiine lineages was recovered to be only occupying the
290 Central Andes. Following this node (23.6 my ago) all the divergences occurred in the Central Andes
291 until 9.4 my ago, when the first colonization event out of the Central Andes occurred (MRCA of
292 *Oleria*, which dispersed into Western Amazonia). Hence, our null biogeographic reconstruction found
293 that all subtribes except Melinaeina and Mechanitina originated and started diversifying in the Central

294 Andes. Interchanges between regions appeared to have increased during the last 10 my. However, all
295 node reconstructions at the basal nodes of the background lineages were highly uncertain.

296 *Biogeographic diversification of the core-group*

297 Using this null model, we investigated more specifically the biogeographic pattern of the core-group
298 by computing rates of dispersal among different regions. We applied the null model to 100 trees
299 randomly sampled from the BEAST posterior distribution and extracted the state with the highest
300 probability at each node. Then, for each 1-my interval, we computed the number of specific transitions
301 divided by the number of lineages existing during this interval and fitted a spline line on the
302 distribution of points.

303 As observed in the ancestral state reconstruction on the maximum clade credibility tree (MCC), no
304 dispersal event occurred during the initial Central-Andean phase of diversification in the core-group
305 (Figure 3). Between 13-8 my ago a major peak of interchanges between the Andes and Amazonia
306 occurred, followed by a second peak between 4-0 my ago (Figure 3). The first peak was almost
307 entirely driven by colonization from the Andes toward the Amazonia, whereas the second peak
308 involved many reverse colonizations toward the Andes. We also recorded a large peak of
309 colonizations from the Central Andes toward the Northern Andes between 13-8 my ago, also followed
310 by a second peak 4-0 my ago (Figure 3, Supporting Information S11-S12). Colonization of Central
311 America may have started 8 my ago, but interchanges largely increased during the last 4 my
312 (Supporting Information S11). Colonizations of the Atlantic Forest also started early (around 13 my
313 ago), but the rate of interchanges between the Atlantic Forest and the remaining Neotropical regions
314 remained relatively constant during the last 10 my (Supporting Information S11-S12).

315 We also used the biogeographic reconstruction to estimate local diversification, namely the cumulative
316 number of divergences inferred to have occurred exclusively in a given region. As described above,
317 until ~10 my ago speciation events occurring in the Central Andes fully account for the core-group
318 diversification (no dispersal events). During the last ~10 my, we observed a dampening of speciation
319 events in the Central Andes (Figure 3). At the same time, following the peaks of dispersal identified
320 above, Northern Andean and Amazonian lineages started diversifying, although the latter diversified at
321 a slower pace than the former. This reflects the large number of dispersal events into the Northern
322 Andes that were followed by important local diversification, for example in the genera *Hypomenitis*
323 (17/20 species in the phylogeny occur in the Northern Andes) and *Pteronymia* (30/45 species in the
324 phylogeny occur in the Northern Andes), or in subclades of the genera *Oleria* or *Napeogenes*. We also
325 identified some important transitions to lowland Amazonia, for example at the origin of the
326 *Brevioleria*-clade, during early divergence in the genus *Oleria*, or in the genus *Hypothyris* (see results
327 on *Diversification in Amazonia*).

328 *Time-stratified biogeographic model*

329 We used the results highlighted above to refine the biogeographic model by incorporating the
330 variations of dispersal rates identified into a model accounting for time-stratified dispersal multipliers.
331 This time-stratified model designed from rates of colonization computed above led to a significant
332 improvement of the model (likelihoods: DECnull: -1335.802, DECstrat: -1321.805). Both ancestral
333 state reconstructions were very congruent but the time-stratified model increased the resolution of
334 several nodes throughout the tree (Supporting Information S11-S12). We identified one major
335 difference in the ancestral states. From the null model the ancestral state of the subtribe Melinaeina,
336 the first lineage to diverge, was highly unclear and the first nodes within Melinaeina were identified as
337 Central Andean, although this was not strongly supported. Likewise, in the null model, Mechanitina,
338 the second lineage to diverge, was inferred to have diversified in the Atlantic Forest but this was
339 poorly supported (Supporting Information S11-S12). The time-stratified model greatly increased the
340 resolution of all these deep nodes, inferring that both Mechanitina and Melinaeina likely initially
341 occupied Western Amazonia. For Melinaeina and Mechanitina this result was in agreement with the
342 BiSSE ancestral state reconstruction. The ancestral state reconstruction inferred that the Ithomiini
343 occupied the Andes from the root, but this was very weakly supported. By contrast, all
344 reconstructions, either BioGeoBEARS or BiSSE, inferred an Andean origin for the core-group.

345

346 **DISCUSSION**

347 We generated one of the largest species-level phylogenies to date for a tropical insect group, the
348 emblematic Neotropical butterfly tribe Ithomiini. With 340 out of 393 species included and a crown
349 age of 26.4 my, this phylogeny offers a unique opportunity to investigate the dynamics of
350 diversification of an insect group throughout the Neotropical region during the major geological and
351 ecological events that have occurred since the Miocene. We discuss our findings below and we
352 propose that the dynamics of multiple landscape transformations during the Miocene, and more
353 specifically the interactions between the Andes and the Pebas system, have strongly affected the
354 dynamics of speciation, extinction and biotic interchanges of Ithomiini butterflies in the Neotropical
355 region.

356 *Early diversification at the interface of the Pebas and Central Andes: has the Pebas driven extinction?*

357 The Ithomiini probably originated along the early Andean foothills at the transition with western
358 Amazonia. The onset of the uplift of the eastern cordillera of the Central Andes during late Oligocene
359 coincides with the origin of Ithomiini⁶⁷ and our reconstruction of the ancestral biogeographic area for
360 the MRCA of the tribe was unable to distinguish between Central Andes or western Amazonia. The
361 Pebas ecosystem replaced the previous western Amazonian terrestrial ecosystem from 23 to 10 my

362 ago. Wesselingh *et al.* (7) described the Pebas as an ecosystem “which was permanently aquatic with
363 minor swamps and fluvial influence, and was connected to marine environments”, and may have
364 reached a maximum size of 1.1 million km². The presence of fossil marine fishes ⁶⁸ and molluscs ¹³
365 testifies to the presence of saline waters. More recently, Boonstra *et al.* ⁶⁹ found evidence from
366 foraminifera and dinoflagellate cysts that marine incursions reached 2000 km inland from the
367 Caribbean sea during the early to middle Miocene during periods of high sea levels. The extent and
368 duration of these marine influences is controversial (see ⁷⁰ and references therein). Yet, it is undeniable
369 that the Pebas system was not suitable for terrestrial fauna and flora, and therefore was likely to affect
370 diversification and dispersal of the terrestrial fauna, including early Ithomiini lineages.

371 The timing of diversification of background lineages reveals a fast early diversification, perhaps
372 following the colonization of South America during the pre-Pebas period – the sister clade of
373 Ithomiini, Tellervini, is found in Australia and Papua New Guinea. Diversification was perhaps partly
374 facilitated by an early shift to a new and diverse hostplant family, the Solanaceae, which is particularly
375 diverse in the Neotropics ³⁶, but the possible effects of new hostplants on early diversification will be
376 difficult to distinguish from the effects of newly available landscapes. Yet, diversification rate rapidly
377 decreased through time, driven by an increasing relative extinction rate and at a time corresponding to
378 the replacement of the terrestrial habitats by the Pebas ecosystem, i.e., ca. 23 my ago. Although the
379 ancestral area of Ithomiini is ambiguous, the two first diverging Ithomiini lineages (Melinaeina and
380 Mechanitina) were clearly endemic to western Amazonia (time-stratified model in BioGeoBEARS and
381 BiSSE reconstruction) and therefore were likely to be affected by the dramatic landscape
382 modifications of the Miocene. However, there are uncertainties surrounding the other deep nodes and
383 also the time when first colonization of the Andes occurred. Two scenarios can be envisioned: (1) The
384 remaining lineages (Tithoreina, Methonina, Athesitina and the core-group) became endemic to the
385 Central Andes (supported by the BioGeoBEARS reconstruction) and we do not know what has driven
386 the shift of diversification at the root of the core-group; and (2) All background lineages were
387 ancestrally western Amazonian (supported by BiSSE ancestral state reconstruction and the sister-clade
388 Tellervini being a group restricted to lowlands) and central Andean endemicity occurred at the root of
389 the core-group only, but extinction events in the background lineages (potentially higher in western
390 Amazonian lineages) may have falsified the BioGeoBEARS ancestral state reconstruction. Indeed,
391 asymmetrical extinction across different geographical regions, as suspected here, may lead to
392 inaccurate inferences of past geographic ranges ^{71, 72}. In our case, if western Amazonian lineages were
393 more prone to extinction than Andean lineages due to the presence of the Pebas, ancestral
394 reconstruction of the distribution areas based on a phylogeny of extant taxa, i. e. those that survived
395 extinction, will be biased towards Andean regions. Such a scenario, where background lineages were
396 ancestrally western Amazonian, would explain the common pattern of high relative extinction rate in
397 the background lineages and a shift of diversification process at the root of the core-group.

398 The idea that the Pebas may have driven extinction is well supported by a recent evaluation of an
399 Amazonian fossil record, which pointed at a major decline of diversity in western Amazonia during
400 the early and middle Miocene ¹⁶. This study concludes that mammalian diversity dropped from 11
401 orders, 29 families and 38 species during late Oligocene down to 1 order, 2 families and 2 species
402 during middle Miocene (see also ¹⁷). These results and the pattern of extinction we found in (at least
403 some) early Amazonian Ithomiini, which occurred during the Pebas period, strongly suggest that the
404 late Oligocene fauna occupying western Amazonia suffered from extinction during the Pebas period.
405 The progressive recovery of these background lineages toward the present, including positive
406 diversification rates in some recent lineages, also concurs with the idea that the retreat of the Pebas
407 released the constraints on diversification during the last 10 my (Supporting Information S7).

408 Parallel to the events occurring in western Amazonia during the Pebas period, the core-group MRCA
409 (19.1-22.1 my old) occupied the Central Andes. This event was of major importance in shaping the
410 diversification of Ithomiini since it is the origin of 85% of the current Ithomiini diversity. Firstly, from
411 this event until ~10 my ago, all core-group lineages exclusively diversified in the Central Andes,
412 meaning that from 19.1-22.1 to ~10 my ago not a single dispersal event occurred out of the Central
413 Andes. Secondly, the core-group corresponds to a shift of diversification dynamics, characterized by a
414 low (or zero) extinction rate and a constant speciation rate, which greatly contrasts with the slow and
415 even negative diversification dynamics of the background lineages during the same period.
416 Consequently, the Central Andes hosted most of the diversification during the first half of Ithomiini
417 history. A two-fold higher Andean diversification rate was found across the whole Ithomiini, which
418 may be mainly the consequence of the diversification rate shift found at the root of the core-group. By
419 contrast, when considering only the core-group, Andean and non-Andean lineages had similar
420 diversification rates. The lack of support for a general increase in diversification rate in the Andes
421 within the core-group is also supported by analyses performed independently on different core-group
422 subtribes. For example, in both Oleriina ⁷³ and Godyridina ¹⁹, radiations occurred in both Andean and
423 Amazonian genera.

424 *Dispersal out of the Central Andes at the demise of the Pebas*

425 Gentry (⁷⁴) pointed at a dichotomy observed in the geographic distribution of Neotropical plant
426 diversity, showing that groups could be divided into Andean-centred *versus* Amazonian-centred
427 patterns. Clades tend to be species-rich in one of these centres and relatively species-poor in the other.
428 Antonelli & Sanmartín (⁶) coined this observation the “Gentry-pattern”. They also suggested that in
429 the absence of a barrier between the Andes and the Amazon basin we should observe continuous
430 interchanges between these regions. Antonelli & Sanmartín (⁶) proposed that the Pebas could be this
431 “missing long-lasting barrier needed for creating the disjunction between Andean-centred and
432 Amazonian-centred groups”. Therefore, in addition to the constraints on diversification discussed

433 above, we predicted that the Pebas ecosystem should have influenced interchanges toward or across
434 western Amazonia.

435 Our results conform surprisingly well to the scenario proposed by Antonelli & Sanmartín (6). Ithomiini
436 are Andean-centered with more than a half of their current diversity occurring in the Andes (see also
437 75). Here we show that interchanges have been virtually absent during the Pebas period, with a period
438 as long as 9-12 my without interchanges. However, rates of interchanges from the Central Andes
439 toward the Northern Andes and Amazonia suddenly peaked ~10 my ago (between 13-8 my ago) and
440 more recently (4-0 my ago). The Western Andean Portal (WAP) is a low-altitude gap that separated
441 the Central Andes and the Northern Andes until 13-11 my ago, and which may have connected the
442 Pebas system and the Pacific Ocean (6). The closure of the WAP may have allowed multiple
443 colonizations of the Northern Andes facilitated by the presence of connecting higher altitude habitats.
444 In parallel, between 10-8 my ago the Pebas system was drained eastward, leading to the formation of
445 the present-day configuration of Amazonian drainage basin. It was accompanied by the expansion of
446 terrestrial forest habitats in western Amazonia. This corresponds precisely to the timing at which core-
447 group lineages colonized western Amazonia and then diversified.

448 *Diversification across the whole Neotropics following the demise of the Pebas*

449 We found a strong dampening of local speciation in the Central Andes during the last 10 my.
450 However, colonizations following the retreat of the Pebas system were followed by large local bursts
451 of diversification within the Northern Andes and Amazonia. As an illustration, from our
452 biogeographic reconstruction, 69 divergence events occurred strictly in the Central Andes in the core-
453 group from 20 my ago until present-day. However, multiple independent dispersal events followed by
454 local diversification lead to the exact same number of divergences occurring strictly in the Northern
455 Andes during the last 9 my only. The genera *Hypomenitis* and *Pteronymia*, for example, diversified
456 extensively within the Northern Andes with 23 and 53 valid species respectively. We also identified
457 four Amazonian radiations in the core-group and three in the background lineages. Lineages that
458 dispersed into the Northern Andes and Amazonia after the demise of the Pebas system probably
459 benefited from a large range of free ecological niches, including a diversity of host-plants that had
460 already diversified or that radiated concomitantly. Two of the background Amazonian radiations, the
461 genus *Melinaea* and the clade *Mechanitis* + *Forbestra*, showed increasing speciation rate toward the
462 present, due in the former case to the shift detected by MEDUSA (*Melinaea*-group). The recent and
463 dramatic increase in diversification rate of the *Melinaea*-group, which produced at least eight species
464 and 50 subspecies ⁴³ in just 1 my, is particularly intriguing. This radiation may be interpreted as
465 support for an effect of recent climatic fluctuations during the Pleistocene on the diversification of this
466 group (as well as the *Mechanitis* + *Forbestra* clade), although ecological drivers of speciation
467 classically invoked in mimetic butterfly diversification, such as colour pattern and hostplant shifts,

468 cannot be ruled out ^{42, 43}. Five other Amazonian radiations showed diversification rates decreasing
469 through time, meaning that diversification was highest just after the retreat of the Pebas. Recent
470 radiations in western Amazonia that post-date the Pebas period have been repeatedly reported. For
471 example, in the genus *Astrocaryum* (Arecaceae), the upper Amazonian clade started diversifying only
472 ~6 my ago ⁷⁶. In *Taygetis* butterflies, Amazonian lineages show rapid diversification during the last 7-
473 8 my ³⁵. Such convergent timing of diversification in western Amazonia strongly supports the scenario
474 of a post-Pebas recovery of terrestrial habitats, which triggered dispersal followed by local
475 diversification.

476

477 *Conclusion*

478 Our research shows that the timing of diversification and biogeographic interchanges in Ithomiini
479 butterflies are tightly associated with the turnover of ecological conditions that occurred during the
480 Miocene. Our findings suggest that the ecological turnover that first accompanied the expansion of the
481 Pebas system has led to a decline of diversification, potentially driven by increasing extinction, in
482 early lineages adapted to the ecological conditions that existed during the Oligocene in western
483 Amazonia. Such a decline of diversity has also been documented in the fossil record ¹⁶, which calls for
484 further investigations on the role of the Pebas in driving extinction during the Miocene. By contrast,
485 lineages that colonized the Central Andes 20 my ago rapidly diversified. However, during the entire
486 existence of the Pebas, these lineages remained trapped in the Central Andes (at least 9-12 my without
487 dispersal events out of the Andes). The closure of the West Andean Portal, connecting the Central and
488 North Andes, and the associated demise of the Pebas (10-8 my ago), apparently released these long-
489 lasting barriers, allowing interchanges with the Northern Andes and Amazonia and opening new
490 opportunities for diversification. As a result of these multiple events, major differences appear
491 between the different faunas. Central Andean lineages started diversifying early (at least 19 my ago),
492 allowing species to accumulate over a long period of time, but diversification slowed down during the
493 last 10 my. In contrast, the Northern Andean fauna is recent (13-11 my old at most), driven by
494 multiple colonization events sometimes followed by important bursts of diversification. In parallel,
495 some Amazonian lineages may be old (late Oligocene), but modern diversity almost entirely arose
496 during the last 8-10 my, after the demise of the Pebas ecosystem. Taken together, all this information
497 points to a robust scenario for Neotropical diversification, which highlights the role of Miocene
498 ecosystem turnover in determining the timing of interchanges, speciation and extinction in the world's
499 most biologically diverse region.

500

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514

515 **Author contributions:**

516 NC and ME conceived the study, with contribution from KRW, GL and AVLF. All co-authors
517 provided specimens and sequences. NC, ME, FPP, CFA, DLDS performed the labwork. NC
518 performed the analyses. NC wrote the paper with major contributions from ME, and contributions
519 from all co-authors.

520

521 **MATERIAL AND METHODS**

522 **Time-calibrated phylogeny**

523 *Molecular data*

524 We compiled sequences of 1460 Ithomiini individuals (Supporting Information S1) that included
525 sequences newly generated for this study and previously published sequences^{19, 21, 26, 27, 32, 47, 52, 77}. We
526 used a concatenation of nine gene fragments, a mitochondrial fragment spanning genes COI-tRNA-
527 COII, and fragments of nuclear genes EF1 α , Tektin, CAD, RPS2, MDH, GAPDH, representing a total
528 of 7083 bp⁷⁸. Primers and PCR conditions followed³² and⁷⁸. We obtained at least one gene fragment
529 for 340 species out of 393 currently known in the group, which represents 87% of the known species
530 richness of the tribe. For each species we produced the consensus sequence of all sequences belonging
531 to individuals of that species to obtain the longest sequence possible. We added 41 outgroups, which
532 spanned all Danainae genera as well as representatives of the main Nymphalidae clades. In total, seven
533 concatenated genes from 381 taxa were used to generate the time-calibrated phylogeny of the
534 Ithomiini.

535 *Tree topology and time calibrations*

536 First, we generated a phylogeny under maximum likelihood inference (ML), using IQ-Tree software
537 as implemented in the W-ID-TREE server^{79, 80} in order to obtain a tree topology (Supporting
538 Information S2). Then, using BEAST v1.8.2⁸¹, we time calibrated this tree by enforcing the ML
539 topology and preventing BEAST from searching for a new topology in the xml file and following the
540 calibration procedure described below.

541 Gene partitioning and substitution models were estimated using PartitionFinder v.1.1⁸². We divided
542 all gene fragments into codon positions and allowed all partitions to be tested. Only substitution
543 models available in BEAST were tested. The models of linked partitions had a better fit than unlinked
544 partition schemes, hence the former was used. The best linked partition scheme contained 13 partitions
545 (Supporting Information S3).

546 Branch lengths were estimated using BEAST v.1.8.2⁸¹ with a birth-death tree prior and a lognormal
547 relaxed molecular clock for each gene partition. In order to time-calibrate the tree we also used a
548 combination of secondary calibrations and host-plant calibrations (Supporting Information S4). Four
549 secondary calibration points were retrieved from Wahlberg *et al.* (⁵⁷)'s phylogeny of Nymphalidae
550 genera and were placed only outside of Ithomiini. We used uniform distribution priors, corresponding
551 to the 95% HSPD inferred by Wahlberg *et al.* (⁵⁷). Host-plant calibrations were used as constraints
552 only within the Ithomiini. Almost all Ithomiini species feed on Solanaceae with a relatively high
553 specificity. A phylogeny of the Solanaceae was published by Särkinen *et al.* (⁸³), and recalibrated by
554 De-Silva *et al.* (⁴⁸). We identified six relevant hostplant clades (Supporting Information S4), which
555 were used as maximum age constraints. Priors for host-plant calibration followed a uniform
556 distribution. The minimum value of the uniform was 0 (present). The maximum value was the upper
557 boundary of the 95% HSPD of the stem age of the host-plant subclade on which the calibrated
558 Ithomiini lineage feeds. To get a starting tree suitable for time-calibration priors, the ML tree was
559 ultrametrised and rescaled using PATHD8⁸⁴ and Mesquite v.3.2⁸⁵.

560 We performed two independent runs of BEAST v.1.8.2⁸¹ on the CIPRES server⁸⁶ of 87 million
561 generations, sampling every 10000. Using Tracer v.1.6 we checked that the two runs had converged
562 and the parameter's ESS values. Both runs were combined using Logcombiner v.1.8.3⁸¹, applying a
563 15% burn-in for each run. Finally using TreeAnnotator v.1.8.3⁸¹ we extracted the Maximum Clade
564 Credibility tree with median branch lengths and we computed the posterior probabilities of each node
565 (Supporting Information S5). The outgroups were pruned and the remaining tree was used for the
566 following analyses (hereafter, MCC tree).

567

568 **Diversification rates**

569 *Time*

570 We investigated the dynamics of speciation and extinction rates through time and across the
571 phylogeny. We chose a combination of three methods to infer the dynamics of diversification:
572 MEDUSA⁵⁹ followed by time-dependent models of diversification proposed by Morlon *et al.* (60) and
573 Stadler (61, TreePar). MEDUSA is a maximum likelihood method that uses stepwise AIC to
574 automatically identify the number and the position of different diversification processes that maximize
575 the fit of the model to the tree. However, MEDUSA makes the strong assumption that rates of
576 speciation and extinction are constant through time. To relax this assumption, we combined MEDUSA
577 with two alternative methods. (1) Morlon *et al.* (60) developed a method that allows both speciation
578 and extinction rates to vary as a function of time and across lineages, and where extinction is allowed
579 to be higher than speciation, a situation leading to declining diversity. However, it does not
580 automatically detect rate shift points, which therefore have to be specified by the user. (2) TreePar⁶¹
581 accommodates models where diversification rates can vary at points in time but are constant between
582 these points. Similarly to Morlon *et al.* (60)'s method, TreePar does not automatically detect shifts
583 within a tree. We proceeded as follows: we ran MEDUSA on the whole tree (MCC tree) in order to
584 partition the tree into different diversification processes but rate estimates were ignored. Instead, we
585 used the partition inferred by MEDUSA to estimate diversification rate (speciation - extinction) and
586 turnover rate (extinction/speciation) by fitting time-dependent models of diversification using Morlon
587 *et al.* (60)'s method and TreePar⁶¹. MEDUSA detected two shifts from the background diversification
588 rates (see Results and Supporting Information S6-7): 1) one shift at the root of a large clade hereafter
589 referred to as the core-group, and 2) one shift at the root of a subclade of the genus *Melinaea* that, for
590 simplicity, is referred to as the *Melinaea*-group. We fitted time-dependent models using the method
591 provided in Morlon *et al.* (2011) on all four possible partitions: no shift, one shift (either the core-
592 group or the *Melinaea*-group) and both shifts. Sampling fraction was indicated for each partition.
593 Comparing the fit of all partitions allowed us to confirm that the two shifts identified by MEDUSA
594 were indeed significant. In each case the stem branch of the shifting clade was included in the
595 subclade, as designed in the method⁶⁰, but we excluded the stem branch of the background tree. For
596 each distinct part of the tree (background tree, *Melinaea*-group and core-group) we fitted the following
597 models: constant speciation (no extinction), time-dependent speciation (no extinction), constant
598 speciation and extinction, time-dependent speciation and constant extinction, constant speciation and
599 time-dependent extinction, time-dependent speciation and extinction. In the cases of time-dependent
600 rates we fitted an exponential dependence to time. Sampling fraction was specified for each of the
601 three partitions. All models were compared using AIC scores. The model with the lowest AIC score
602 that was significantly different from the null model of constant speciation rate was used to plot the
603 diversification rate through time. Finally we ran TreePar⁶¹ in order to obtain a second, independent,
604 estimation of diversification and turnover rates across time. TreePar uses a vector of speciation times,

605 which allowed us to run it separately on the core-group and the background tree. For the background
606 tree we added the times of divergence at which the core-group and the *Melinaea*-group diverged from
607 the background to keep track of these cladogenetic events. We split time into time bins of five million
608 years, for which diversification and turnover rates were estimated. We used these estimates to obtain a
609 second estimate of diversification rate “through time” that could be compared with Morlon *et al.* (60)’s
610 method. We did not fit TreePar on the *Melinaea*-group since it is only 1 million year old, and therefore
611 does not even span an entire time bin. We allowed diversification rate to be negative but we did not
612 allow mass-extinction events.

613 In addition to MEDUSA, Morlon *et al.* (2011) and TreePar we also performed analyses with BAMM v
614 2.5.0 (62). We ran BAMM for 30 million generations sampling every 3000 generations, using genus
615 level sampling fraction. The results of the MCMC were analyzed in the R package BAMMTOOLS (87).
616 First we checked for MCMC convergence using trace plots and ESS statistics using the R package
617 CODA, after removing 10% of the chain as burnin. Second we investigated the number and
618 configurations of diversification rate shifts (Details and results can be found in Supporting Information
619 S8).

620 *Diversification in the Andes*

621 To investigate the pattern of diversification in the Andes with respect to rest of the Neotropical
622 regions, we classified species as Andean or non-Andean, based on a combination of published (19, 21, 27,
623 48, 52, 68) and unpublished (i.e., databases generated from museum collections and our own field
624 collection) georeferenced distribution data and elevation ranges of species. In general species can be
625 unambiguously classified to either of the two categories, because Andean species are never found in
626 the lowlands, whereas species that occur in the lowlands and that sometimes also occur at the Andean
627 foothills never occur above ca. 800m, and are therefore classified as non-Andean. Species that do not
628 occur in the Andean region (e.g., Atlantic Forest) are obviously considered as non-Andean. Species
629 biogeographic distribution was used as character states. Then we used trait-dependent models of
630 diversification (19, 88) to compare the rates of speciation, of extinction and of transition between the
631 Andean area and the non-Andean regions. We used the ClasSE model (63), which accounts for up to 10
632 parameters (2 speciation rates without character state change, 4 cladogenetic transition rates, 2
633 extinction rates and 2 anagenetic transition rates). However, we constrained parameters that were not
634 biogeographically meaningful to zero (19). Those include the anagenetic transition rates, considering that
635 transition rates from one region to the other were always accompanied by a speciation event, and the
636 cladogenetic transition rates involving a transition in both descendant lineages since we considered the
637 scenario as unrealistic. We therefore ended up with at most 6 parameters. We tested all models with
638 only one parameter (speciation, extinction, transitions) free to vary as well as all models combining
639 two or more parameters free to vary. This allows comparing the relative contribution of one

640 biogeographic model to the others, as well as their combination since they are not mutually exclusive.
641 Models were compared using AIC scores. All models were fitted on the MCC tree.

642 Furthermore, we considered two important sources of potential biases. (1) The major shift of
643 diversification at the root of the core-group could affect our results of trait-dependent diversification
644 models. Thus we fitted ClaSSE on both the whole tree and the core-group alone to compare the
645 results. (2) We performed additional analyses to test the hypothesis that a hidden character not
646 considered here explained the pattern of trait-dependent diversification measured with ClaSSE. To do
647 so we fitted the model HiSSE ⁶⁴. HISSE results were fully congruent with the model ClaSSE and our
648 interpretation. The details about the models fitted, results and interpretations can be found in
649 Supporting Information S9.

650 Finally, we also conducted ancestral state reconstructions based on the models of trait-dependent
651 diversification. Since ancestral state reconstruction is not available for ClaSSE models we used the
652 BiSSE model ⁶⁵ for this purpose. BiSSE also includes 2 speciation rates and two extinction rates, but it
653 allows the transitions to occur only along branches (anagenetic transition rates). Therefore, we fitted
654 the BiSSE model corresponding to the best fitting ClaSSE model, and we used these parameters to
655 infer the ancestral states at the nodes of the phylogeny. This ancestral state reconstruction was
656 compared to that obtained using the historical biogeography analyses outlined below. All these
657 analyses were performed on both the whole phylogeny and the core-group only, to account for the
658 diversification rate shift identified by our time-dependent diversification analyses.

659 *Diversification in Amazonia*

660 We further investigated the pattern of diversification in Amazonia during the post-Pebas period. The
661 Amazonian basin appears to be the second most important place for diversification after the Andes and
662 there is a longstanding hypothesis that speciation in this region has been driven by climatic
663 fluctuations during the Quaternary ⁹. An interpretation of this scenario is that speciation rate should
664 increase during the last 2.5 million years ¹². To test this hypothesis we identified the major Amazonian
665 diversification events, i.e. clades which nodes were inferred to be almost all Amazonian from the
666 BioGeoBEARS ancestral state reconstruction. We fitted a model of time-dependent speciation rate (no
667 extinction) to see whether speciation rates increased through time (supporting a recent diversification
668 potentially caused by Pleistocene climatic fluctuations) or decreased through time (supporting
669 radiations accompanying the post-Pebas recolonizations).

670

671 **Historical biogeography**

672 We proceeded in three steps to reconstruct the historical biogeography of Ithomiini. First we
673 performed an ancestral state reconstruction using a model with refined area adjacency but uniform
674 dispersal multipliers (null-model). Second, we used the results of this model to compute rates of
675 dispersal between specific regions per million years. This allowed us to test some biogeographic
676 hypotheses but also to identify relevant time frames for which dispersal probabilities might vary.
677 Third, we implemented a time-stratified model designed from the previous information to refine our
678 biogeographic reconstruction.

679 *Biogeographic null model*

680 We inferred the historical biogeography of Ithomiini using BioGeoBEARS v.0.2.1 ⁶⁶. We divided the
681 Neotropics into nine distinct biogeographic regions (Supporting Information S11): 1) Central America,
682 2) Caribbean Islands, 3) lowlands on the western part of the Andes, including the Magdalena valley, 4)
683 Northern Andes that comprise the western and eastern Ecuadorian and Colombian cordilleras and the
684 Venezuelan cordillera, 5) Central Andes, 6) western Amazonia, 7) eastern Amazonia, 8) Guiana
685 Shield and 9) Atlantic Forest (Supporting Information S11). In this model, we constrained the
686 combinations of areas to avoid unrealistic distributions (e.g., disjunct distributions) but all dispersal
687 multipliers were set to 1 to avoid biasing the ancestral state reconstruction. We compared the models
688 DIVALIKE and DEC as implemented in BioGeoBEARS v.0.2.1 ⁶⁶ using log likelihoods. We used the
689 best fitting model (DEC) for the following analyses.

690 *Biogeographic interchanges within the coregroup*

691 One important hypothesis is that the Pebas influenced the interchanges among biogeographic regions,
692 especially toward or across western Amazonia. To test this hypothesis, we computed “rates” of
693 colonization from BioGeoBEARS ancestral-state reconstructions performed with the null-model. For
694 100 trees randomly sampled from the BEAST posterior distribution we applied the null-model with
695 BioGeoBEARS and reconstructed the ancestral states. For each tree, we extracted the state with the
696 highest probability at each node. When a descendent node had a range different from the ancestral
697 node, we considered the middle of the branch connecting these nodes as the timing of the event and
698 recoded it. Then for each million-year interval, we computed the proportion of specific state
699 transitions compared to the maximum number of lineages existing during this interval. Hence, for each
700 million-year interval we obtained a proportion of lineages that dispersed for example from the Central-
701 Andes toward the Northern-Andes or from the Andes toward the Amazonia, which we refer to as
702 “rates” of colonization. Therefore, for each million-year interval and each state transition we obtained
703 a distribution of values. We fitted a spline line on this distribution against time to assess the pattern of
704 variation of colonization rates through time. That way we computed rates of colonization between
705 Andean and non-Andean regions, between the Andes and the Amazonia, between the Central and the
706 Northern Andes, between the Atlantic forest and the other regions, and between Central America and

707 the rest of the Neotropics. These rates were computed only on the core-group, because (1) this group
708 contains 85% of Ithomiini species, (2) the diversification process is homogeneous throughout the clade
709 and (3) we found high extinction in background lineages, which may falsify the ancestral state
710 reconstruction in these lineages, which were therefore excluded. We focused more specifically on the
711 timing of transitions between the Andes and the Amazonian basin and the timing of transitions from
712 the Central Andes (ancestral Andean region of the core-group) to the Northern Andes, because a
713 strong influence of the Pebas and the WAP should have affected these rates of interchanges (see
714 Supporting Information S11 for the complete results).

715 Additionally, we recorded the divergence times of nodes inferred to be strictly in either the Central-
716 Andes, the Northern Andes or Amazonia (the region comprising the upper and lower Amazon and the
717 Guiana shield). We excluded all nodes inferred to be in more than one of these areas (for Amazonia,
718 only all combinations of the three areas above were considered) so that we obtained times of
719 divergences inferred to have occurred strictly within the region considered (which we call local
720 diversification). These divergence times were obtained from the unconstrained BioGeoBEARS
721 reconstruction (DEC) on the MCC tree and we used them to represent lineage accumulation through
722 time (exclusively due to speciation) in each region.

723 *Time-stratified biogeographic model*

724 We used our estimations of colonization rates among the major biogeographic regions to design a
725 time-stratified biogeographic model and improve the resolution of our biogeographic reconstruction
726 (Supporting Information S11). We created four time frames: i) 0-4, ii) 4-8, iii) 8-13, and iv) 13-30 my
727 ago, where dispersal multipliers between areas reflected the colonization rate variations identified
728 previously. This ancestral state reconstruction was compared to the null model as well as to the
729 ancestral state reconstruction obtained from trait-dependent diversification models (Supporting
730 Information S10).

731 Table 1. Results of time-dependent models of diversification fitted on the different partitions: 0 shift, 1
 732 shift (core-group or *Melinaea*-group), 2 shifts. For each subclade or the background tree, only the best
 733 fitting model is shown (see Supporting Information S6 for more details). BCST=constant speciation,
 734 BVAR=time-dependent speciation, DCST=constant extinction, DVAR time-dependent extinction. λ
 735 =speciation rate at present, α =coefficient of time variation of the speciation rate, μ =extinction rate at
 736 present, β =coefficient of time variation of the extinction rate.

		model	par	logL	AIC	λ	α	μ	β
0 shift	whole tree	BCST	1	-861.800	1725.613	0.227			
	background	BCSTDVAR	3	-144.886	296.294	0.311	0.397	-0.055	
1 shift	core-group	BCST	1	-703.033	1408.080	0.255			
	Joint model		4	-847.920	1704.375				
	background	BCST	1	-850.489	1702.990	0.223			
1 shift	<i>Melinaea</i> -group	BVAR	2	-2.135	8.270	0.342	3.044		
	Joint model		3	-852.624	1711.261				
	background	BVARDVAR	4	-129.863	267.727	0.151	0.159	0.190	0.147
2 shifts	<i>Melinaea</i> -group	BVAR	2	-2.135	8.270	0.342	3.044		
	core-group	BCST	1	-703.033	1408.080	0.255			
	Joint model		7	-835.032	1684.065				

737

738 Table 2. Results of ClaSSE models fitted on the whole Ithomiini tree (a.) and the core-group (b.)
 739 sorted by increasing AIC. Constraints of each model are indicated in the four first columns. 1=non-
 740 Andean, 2=Andean, λ_{111} and λ_{222} represent within-region speciation rates, λ_{112} and λ_{212} represent
 741 cladogenetic transition rates, μ = extinction rates, df = degree of freedom (number of parameters),
 742 logL=log-likelihood, AIC= Akaike information criterion score, Δ AIC = AIC difference with the best
 743 fitting model.

744 a. Whole tree

$\lambda_{111}/\lambda_{222}$	$\lambda_{112}/\lambda_{212}$	μ	df	logL	AIC	Δ AIC	λ_{111}	λ_{222}	λ_{112}	λ_{212}	μ_1	μ_2	
$\lambda_{111} \neq \lambda_{222}$	\neq	$\lambda_{112} = \lambda_{212}$	$\mu_1 = \mu_2$	4	-250.061	508.122	0	0.118	0.230	0.079	0.079	6.10E-06	6.10E-06
$\lambda_{111} \neq \lambda_{222}$	\neq	$\lambda_{112} \neq \lambda_{212}$	$\mu_1 = \mu_2$	5	-249.801	509.602	1.48	0.108	0.231	0.047	0.094	5.20E-08	5.20E-08
$\lambda_{111} \neq \lambda_{222}$	\neq	$\lambda_{112} = \lambda_{212}$	$\mu_1 \neq \mu_2$	5	-250.061	510.122	2	0.118	0.230	0.079	0.079	3.75E-06	9.11E-06
$\lambda_{111} = \lambda_{222}$	\neq	$\lambda_{112} = \lambda_{212}$	$\mu_1 = \mu_2$	3	-252.359	510.718	2.596	0.187	0.187	0.077	0.077	1.00E-09	1.00E-09
$\lambda_{111} = \lambda_{222}$	\neq	$\lambda_{112} = \lambda_{212}$	$\mu_1 \neq \mu_2$	4	-251.94	511.88	3.758	0.198	0.198	0.080	0.080	0.061	1.26E-06
$\lambda_{111} \neq \lambda_{222}$	\neq	$\lambda_{112} \neq \lambda_{212}$	$\mu_1 \neq \mu_2$	6	-249.801	511.602	3.48	0.108	0.231	0.047	0.094	9.95E-07	2.79E-07
$\lambda_{111} = \lambda_{222}$	\neq	$\lambda_{112} \neq \lambda_{212}$	$\mu_1 = \mu_2$	4	-252.335	512.67	4.548	0.189	0.189	0.066	0.081	5.26E-07	5.26E-07
$\lambda_{111} = \lambda_{222}$	\neq	$\lambda_{112} = \lambda_{212}$	$\mu_1 \neq \mu_2$	5	-251.726	513.452	5.33	0.204	0.204	0.052	0.101	0.089	9.00E-09
$\lambda_{111} = \lambda_{222}$	$=$	$\lambda_{112} = \lambda_{212}$	$\mu_1 = \mu_2$	2	-256.181	516.362	8.24	0.133	0.133	0.133	0.133	1.87E-07	1.87E-07
$\lambda_{111} = \lambda_{222}$	$=$	$\lambda_{112} = \lambda_{212}$	$\mu_1 \neq \mu_2$	3	-255.657	517.314	9.192	0.108	0.108	0.047	0.094	9.95E-07	2.79E-07

745

746 b. Core-group

$\lambda_{111}/\lambda_{222}$	$\lambda_{112}/\lambda_{212}$	μ	df	logL	AIC	ΔAIC	λ_{111}	λ_{222}	λ_{112}	λ_{212}	μ_1	μ_2
$\lambda_{111}=\lambda_{222}$	\neq	$\lambda_{112}\neq\lambda_{212}$	$\mu_1=\mu_2$	4	-169.942	347.884	0	0.156	0.156	0.068	3.48E-07	7.12E-05
$\lambda_{111}=\lambda_{222}$	\neq	$\lambda_{112}=\lambda_{212}$	$\mu_1=\mu_2$	3	-171.109	348.218	0.334	0.156	0.156	0.037	0.037	5.00E-09
$\lambda_{111}\neq\lambda_{222}$	\neq	$\lambda_{112}\neq\lambda_{212}$	$\mu_1=\mu_2$	5	-169.399	348.798	0.914	0.177	0.129	0.07	1.71E-07	2.96E-07
$\lambda_{111}=\lambda_{222}$	\neq	$\lambda_{112}\neq\lambda_{212}$	$\mu_1\neq\mu_2$	5	-169.94	349.88	1.996	0.156	0.156	0.068	1.63E-07	3.95E-06
$\lambda_{111}\neq\lambda_{222}$	\neq	$\lambda_{112}=\lambda_{212}$	$\mu_1=\mu_2$	4	-171.075	350.15	2.266	0.163	0.15	0.037	0.037	1.90E-07
$\lambda_{111}=\lambda_{222}$	\neq	$\lambda_{112}=\lambda_{212}$	$\mu_1\neq\mu_2$	4	-171.109	350.218	2.334	0.156	0.156	0.037	0.037	7.89E-06
$\lambda_{111}\neq\lambda_{222}$	\neq	$\lambda_{112}\neq\lambda_{212}$	$\mu_1\neq\mu_2$	6	-169.399	350.798	2.914	0.177	0.129	0.07	4.03E-07	1.43E-06
$\lambda_{111}\neq\lambda_{222}$	\neq	$\lambda_{112}=\lambda_{212}$	$\mu_1\neq\mu_2$	5	-171.075	352.15	4.266	0.163	0.15	0.037	0.037	4.11E-07
$\lambda_{111}=\lambda_{222}$	$=$	$\lambda_{112}=\lambda_{212}$	$\mu_1=\mu_2$	2	-179.29	362.58	14.696	0.097	0.097	0.097	0.097	4.00E-09
$\lambda_{111}=\lambda_{222}$	$=$	$\lambda_{112}=\lambda_{212}$	$\mu_1\neq\mu_2$	3	-179.291	364.582	16.698	0.177	0.177	0.07	4.00E-06	1.43E-06

747

748

749 Figure 1. Time-calibrated phylogeny of the Ithomiini tribe. Coloured branches depict the partitions
750 identified by MEDUSA and used for fitting diversification rate models. Red lineages constitute the
751 core-group, green lineages the *Melinaea*-group, blue lineages the background tree. Black and white
752 circles indicate the biogeographic ancestral states reconstructed at the basal nodes of the tree:
753 black=Central Andes, white=Western Amazonia. Question mark and arrows indicate the position of
754 two alternative scenarios for the first colonization of the Andes: BioGeoBEARS at the root of the
755 Ithomiini, BiSSE at the root of the core-group. Both methods agree for Melinaeina and Mechanitina
756 but diverge for Tithoreina, Methonina and Athesitina. The names and position of the different
757 subtribes are indicated.

758 Figure 2. Results of diversification rate estimates through time using MEDUSA's partition. A.
759 Diversification rate estimated using the method designed by Morlon *et al.* (60) for the background and
760 the core-group. Results for the *Melinaea*-group are not plotted because of its very high diversification
761 rate. B. Diversity trajectories inferred from the best fitting models of diversification obtained using
762 Morlon *et al.* (60)'s method. C. and D. Diversification rate and turnover of the background and core-
763 group respectively, estimated using 5 million-year time-bins with TreePar ⁶¹.

764 Figure 3. A. Colonization rates and lineage accumulation through time (by speciation) extracted from
765 BioGeoBEARS ancestral state reconstruction for the core-group. Colonization rates correspond to the
766 proportion of transitions compared to the number of lineages existing during each 1-million year
767 interval. The lines depicted are spline lines fitted over 100 reconstructions on the posterior
768 distribution. The blue line is the rate of total interchanges between the Andes and Amazonia, the
769 orange line the rate of colonization from the Central Andes toward the Northern Andes (see
770 Supporting Information S11 for details). Dots represent the additional contribution to lineage
771 accumulation of local diversification in three regions. Red dots represent the cumulative number of
772 divergence events through time in Central Andes; orange dots represent the cumulative number of
773 divergence events through time in Central Andes and Northern Andes; blue dots represent the
774 cumulative number of divergence events through time in Central Andes, Northern Andes and
775 Amazonia; Grey dots represent the cumulative number of divergence events through time in the entire
776 Neotropics. B. Time-dependent speciation rates estimated on seven Amazonian clades (four in the
777 core-group and three in the background lineages). All those clades originated during the last 8 my.

778 List of supporting information:

779 S1. List of all individuals and species used in this study, and biogeographic distribution of species.

780 S2. Gene fragment partitions and substitution models associated obtained by Partition Finder v.1.1

781 S3. Node constraints used for time-calibration of the tree.

782 S4. Maximum likelihood tree obtained with IQ-tree, with bootstrap support indicated at the nodes.

783 S5. Bayesian time-calibrated tree obtained with BEAST v.1.8 with median node ages and 95%HPD
784 indicated at the nodes.

785 S6. Full results of diversification analyses.

786 S7. Testing the effect of diversification rate heterogeneity within the background on diversification
787 rate estimates

788 S8. Diversification analysis with BAMM.

789 S9. Trait-dependent diversification using HiSSE.

790 S9. Ancestral state reconstruction performed using trait-dependent diversification models

791 S10. Rates of colonization between different region, computed on the core-group and used to design a
792 time-stratified biogeographic model.

793 S11. Results of biogeographic ancestral state reconstruction obtained with BioGeoBEARS and using
794 the “null” or time-stratified model.

795

796 **REFERENCES**

- 797 1. Hoorn C, *et al.* Amazonia through time: Andean uplift, climate change, landscape evolution, 798 and biodiversity. *science* **330**, 927-931 (2010).
- 799 2. Rull V. Speciation timing and neotropical biodiversity: the Tertiary–Quaternary debate in the 800 light of molecular phylogenetic evidence. *Molecular Ecology* **17**, 2722-2729 (2008).
- 802 3. Smith BT, *et al.* The drivers of tropical speciation. *Nature* **515**, 406-409 (2014).
- 804 4. Madriñán S, Cortés AJ, Richardson JE. Páramo is the world's fastest evolving and coolest 805 biodiversity hotspot. *Frontiers in genetics* **4**, 192 (2013).
- 807 5. Antonelli A, Nylander JA, Persson C, Sanmartín I. Tracing the impact of the Andean uplift on 808 Neotropical plant evolution. *Proceedings of the National Academy of Sciences* **106**, 9749- 809 9754 (2009).
- 811 6. Antonelli A, Sanmartín I. Why are there so many plant species in the Neotropics? *Taxon* **60**, 812 403-414 (2011).
- 814 7. Wesselingh F, *et al.* Lake Pebas: a palaeoecological reconstruction of a Miocene, long-lived 815 lake complex in western Amazonia. *Cainozoic Research* **1**, 35-68 (2001).
- 817 8. Wesselingh FP, *et al.* On the origin of Amazonian landscapes and biodiversity: a synthesis. 818 *Amazonia: Landscape and Species Evolution: A look into the past*, 419-431 (2010).
- 820 9. Haffer J. Speciation in Amazonian forest birds. *Science* **165**, 131-137 (1969).
- 822 10. Colinvaux PA, De Oliveira P, Bush M. Amazonian and neotropical plant communities on 823 glacial time-scales: the failure of the aridity and refuge hypotheses. *Quaternary Science 824 Reviews* **19**, 141-169 (2000).
- 826 11. Garzón-Orduña IJ, Benetti-Longhini JE, Brower AV. Timing the diversification of the 827 Amazonian biota: butterfly divergences are consistent with Pleistocene refugia. *Journal of 828 biogeography* **41**, 1631-1638 (2014).
- 830 12. Matos-Maraví P. Investigating the timing of origin and evolutionary processes shaping 831 regional species diversity: Insights from simulated data and neotropical butterfly 832 diversification rates. *Evolution* **70**, 1638-1650 (2016).
- 834 13. Wesselingh F. Molluscs from the Miocene Pebas Formation of Peruvian and Colombian 835 Amazonia. *Scripta Geologica* **133**, (2006).

837

838 14. Muñoz-Torres F, Whatley R, Van Harten D. Miocene ostracod (Crustacea) biostratigraphy of
839 the upper Amazon Basin and evolution of the genus *Cyprideis*. *Journal of South American
840 Earth Sciences* **21**, 75-86 (2006).

841

842 15. Salas-Gismondi R, Flynn JJ, Baby P, Tejada-Lara JV, Wesselingh FP, Antoine P-O. A Miocene
843 hyperdiverse crocodylian community reveals peculiar trophic dynamics in proto-Amazonian
844 mega-wetlands. *Proceedings of the Royal Society of London B: Biological Sciences* **282**,
845 20142490 (2015).

846

847 16. Antoine P-O, *et al.* A 60-million-year Cenozoic history of western Amazonian ecosystems in
848 Contamana, eastern Peru. *Gondwana Research* **31**, 30-59 (2016).

849

850 17. Antoine P-O, Salas-Gismondi R, Pujos F, Ganerød M, Marivaux L. Western Amazonia as a
851 Hotspot of Mammalian Biodiversity Throughout the Cenozoic. *Journal of Mammalian
852 Evolution* **24**, 5-17 (2017).

853

854 18. Wesselingh F, Salo J. A Miocene perspective on the evolution of the Amazonian biota. *Scripta
855 Geologica* **133**, 439-458 (2006).

856

857 19. Chazot N, *et al.* Into the Andes: multiple independent colonizations drive montane diversity
858 in the Neotropical clearwing butterflies Godyridina. *Molecular Ecology* **25**, 5765-5784 (2016).

859

860 20. Condamine FL, Silva-Brandao KL, Kergoat GJ, Sperling FA. Biogeographic and diversification
861 patterns of Neotropical Troidini butterflies (Papilionidae) support a museum model of
862 diversity dynamics for Amazonia. *BMC evolutionary biology* **12**, 82 (2012).

863

864 21. Elias M, *et al.* Out of the Andes: patterns of diversification in clearwing butterflies. *Molecular
865 Ecology* **18**, 1716-1729 (2009).

866

867 22. Penz CM, Devries PJ, Wahlberg N. Diversification of Morpho butterflies (Lepidoptera,
868 Nymphalidae): a re-evaluation of morphological characters and new insight from DNA
869 sequence data. *Systematic Entomology* **37**, 670-685 (2012).

870

871 23. Price SL, Powell S, Kronauer DJ, Tran LA, Pierce NE, Wayne R. Renewed diversification is
872 associated with new ecological opportunity in the Neotropical turtle ants. *Journal of
873 evolutionary biology* **27**, 242-258 (2014).

874

875 24. Casner KL, Pyrcz TW. Patterns and timing of diversification in a tropical montane butterfly
876 genus, *Lymanopoda* (Nymphalidae, Satyrinae). *Ecography* **33**, 251-259 (2010).

877

878 25. Chazot N, *et al.* Morpho morphometrics: Shared ancestry and selection drive the evolution of
879 wing size and shape in Morpho butterflies. *Evolution* **70**, 181-194 (2016).

880

881 26. De-Silva DL, Day JJ, Elias M, Willmott K, Whinnett A, Mallet J. Molecular phylogenetics of the
882 neotropical butterfly subtribe Oleriina (Nymphalidae: Danainae: Ithomiini). *Molecular*
883 *Phylogenetics and Evolution* **55**, 1032-1041 (2010).

884

885 27. De-Silva DL, Elias M, Willmott K, Mallet J, Day JJ. Diversification of clearwing butterflies with
886 the rise of the Andes. *Journal of biogeography* **43**, 44-58 (2016).

887

888 28. Ebel ER, *et al.* Rapid diversification associated with ecological specialization in Neotropical
889 Adelpha butterflies. *Molecular ecology* **24**, 2392-2405 (2015).

890

891 29. Espeland M, *et al.* Ancient Neotropical origin and recent recolonisation: Phylogeny,
892 biogeography and diversification of the Riodinidae (Lepidoptera: Papilionoidea). *Molecular*
893 *phylogenetics and evolution* **93**, 296-306 (2015).

894

895 30. Kozak KM, Wahlberg N, Neild AF, Dasmahapatra KK, Mallet J, Jiggins CD. Multilocus species
896 trees show the recent adaptive radiation of the mimetic Heliconius butterflies. *Systematic*
897 *biology*, syv007 (2015).

898

899 31. Lewis DS, Sperling FA, Nakahara S, Cotton AM, Kawahara AY, Condamine FL. Role of
900 Caribbean Islands in the diversification and biogeography of Neotropical Heraclides
901 swallowtails. *Cladistics* **31**, 291-314 (2015).

902

903 32. Mallarino R, Bermingham E, Willmott KR, Whinnett A, Jiggins CD. Molecular systematics of
904 the butterfly genus Ithomia (Lepidoptera: Ithomiinae): a composite phylogenetic hypothesis
905 based on seven genes. *Molecular phylogenetics and evolution* **34**, 625-644 (2005).

906

907 33. Massardo D, Fornel R, Kronforst M, Gonçalves GL, Moreira GRP. Diversification of the
908 silverspot butterflies (Nymphalidae) in the Neotropics inferred from multi-locus DNA
909 sequences. *Molecular phylogenetics and evolution* **82**, 156-165 (2015).

910

911 34. Brower AV, Willmott KR, Silva-Brandão KL, Garzón-Orduña IJ, Freitas AV. Phylogenetic
912 relationships of ithomiine butterflies (Lepidoptera: Nymphalidae: Danainae) as implied by
913 combined morphological and molecular data. *Systematics and Biodiversity* **12**, 133-147
914 (2014).

915

916 35. Matos-Maravi PF, Pena C, Willmott KR, Freitas AV, Wahlberg N. Systematics and evolutionary
917 history of butterflies in the “Taygetis clade”(Nymphalidae: Satyrinae: Euptychiina): towards a
918 better understanding of Neotropical biogeography. *Molecular Phylogenetics and Evolution*
919 **66**, 54-68 (2013).

920

921 36. Willmott KR, Freitas AV. Higher-level phylogeny of the Ithomiinae (Lepidoptera:
922 Nymphalidae): classification, patterns of larval hostplant colonization and diversification.
923 *Cladistics* **22**, 297-368 (2006).

924
925 37. Müller F. Ituna and Thyridia: a remarkable case of mimicry in butterflies. *Trans Entomol Soc*
926 *Lond* **1879**, 20-29 (1879).

927
928 38. Brown Jr KS, Benson WW. Adaptive polymorphism associated with multiple Müllerian
929 mimicry in *Heliconius numata* (Lepid. Nymph.). *Biotropica*, 205-228 (1974).

930
931 39. Bates HW. Contributions to an insect fauna of the Amazon valley (Lepidoptera: Heliconidae).
932 *Transactions of the Linnean Society of London* **23**, 495-566 (1862).

933
934 40. Bolanos Martinez I, Zambrano Gonzalez G, Willmott K. Descripción de los estados inmaduros
935 de *Pteronymia zerlina zerlina*, *P. zerlina machay*, *P. medellina* y *P. veia* n. ssp.(Lepidoptera:
936 Nymphalidae: Ithomiini) de Colombia y del Ecuador. *Tropical Lepidoptera Research* **21**, 27-33
937 (2011).

938
939 41. Hill RI, *et al.* Ecologically relevant cryptic species in the highly polymorphic Amazonian
940 butterfly *Mechanitis mazaeus* sl (Lepidoptera: Nymphalidae; Ithomiini). *Biological Journal of*
941 *the Linnean Society* **106**, 540-560 (2012).

942
943 42. McClure M, Elias M. Unravelling the role of host plant expansion in the diversification of a
944 Neotropical butterfly genus. *BMC Evolutionary Biology* **16**, 128 (2016).

945
946 43. McClure M, Elias M. Ecology, life history, and genetic differentiation in Neotropical Melinaea
947 (Nymphalidae: ithomiini) butterflies from north-eastern Peru. *Zoological Journal of the*
948 *Linnean Society* **179**, 110-124 (2017).

949
950 44. Brown KS. Adult-obtained pyrrolizidine alkaloids defend ithomiine butterflies against a spider
951 predator. *Nature* **309**, 707-709 (1984).

952
953 45. Schulz S, *et al.* Semiochemicals derived from pyrrolizidine alkaloids in male ithomiine
954 butterflies (Lepidoptera: Nymphalidae: Ithomiinae). *Biochemical Systematics and Ecology* **32**,
955 699-713 (2004).

956
957 46. Trigo JR, Brown KS. Variation of pyrrolizidine alkaloids in Ithomiinae: a comparative study
958 between species feeding on Apocynaceae and Solanaceae. *Chemoecology* **1**, 22-29 (1990).

959
960 47. Brower AV, Freitas AV, Lee MM, Silva-Brandão KL, Whinnett A, Willmott KR. Phylogenetic
961 relationships among the Ithomiini (Lepidoptera: Nymphalidae) inferred from one
962 mitochondrial and two nuclear gene regions. *Systematic Entomology* **31**, 288-301 (2006).

963
964 48. De-Silva DL, *et al.* North Andean origin and diversification of the largest ithomiine butterfly
965 genus. *Scientific Reports* **7**, 45966 (2017).

966

967 49. Brown K, Von Schoultz, B., Suomalainen E. Chromosome evolution in neotropical Danainae
968 and Ithomiinae (Lepidoptera). *Hereditas* **141**, 216-236 (2004).

969
970 50. Willmott KR, Willmott JCR, Elias M, Jiggins CD. Maintaining mimicry diversity: optimal
971 warning colour patterns differ among microhabitats in Amazonian clearwing butterflies. In:
972 *Proc. R. Soc. B* (ed^eds). The Royal Society (2017).

973
974 51. Beccaloni GW. Vertical stratification of ithomiine butterfly (Nymphalidae: Ithomiinae)
975 mimicry complexes: the relationship between adult flight height and larval host-plant height.
976 *Biological journal of the Linnean Society* **62**, 313-341 (1997).

977
978 52. Chazot N, *et al.* Mutualistic mimicry and filtering by altitude shape the structure of Andean
979 butterfly communities. *The American Naturalist* **183**, 26-39 (2013).

980
981 53. DeVries P, Lande R, Murray D. Associations of co-mimetic ithomiine butterflies on small
982 spatial and temporal scales in a neotropical rainforest. *Biological Journal of the Linnean
983 Society* **67**, 73-85 (1999).

984
985 54. Elias M, Gompert Z, Jiggins C, Willmott K. Mutualistic interactions drive ecological niche
986 convergence in a diverse butterfly community. *PLoS Biol* **6**, e300 (2008).

987
988 55. Hill RI. Habitat segregation among mimetic ithomiine butterflies (Nymphalidae). *Evolutionary
989 ecology* **24**, 273-285 (2010).

990
991 56. Jiggins CD, Mallarino R, Willmott KR, Bermingham E, Funk D. The phylogenetic pattern of
992 speciation and wing pattern change in neotropical *Ithomia* butterflies (Lepidoptera:
993 Nymphalidae). *Evolution* **60**, 1454-1466 (2006).

994
995 57. Wahlberg N, *et al.* Nymphalid butterflies diversify following near demise at the
996 Cretaceous/Tertiary boundary. *Proceedings of the Royal Society of London B: Biological
997 Sciences* **276**, 4295-4302 (2009).

998
999 58. Garzón-Orduña IJ, Silva-Brandão KL, Willmott KR, Freitas AV, Brower AV. Incompatible ages
1000 for clearwing butterflies based on alternative secondary calibrations. *Systematic biology*,
1001 syv032 (2015).

1002
1003 59. Alfaro ME, *et al.* Nine exceptional radiations plus high turnover explain species diversity in
1004 jawed vertebrates. *Proceedings of the National Academy of Sciences* **106**, 13410-13414
1005 (2009).

1006
1007 60. Morlon H, Parsons TL, Plotkin JB. Reconciling molecular phylogenies with the fossil record.
1008 *Proceedings of the National Academy of Sciences* **108**, 16327-16332 (2011).

1009

1010 61. Stadler T. Mammalian phylogeny reveals recent diversification rate shifts. *Proceedings of the*
1011 *National Academy of Sciences* **108**, 6187-6192 (2011).

1012 62. Rabosky DL. Automatic detection of key innovations, rate shifts, and diversity-dependence
1013 on phylogenetic trees. *PLoS one* **9**, e89543 (2014).

1015 63. Goldberg EE, Igić B. Tempo and mode in plant breeding system evolution. *Evolution* **66**, 3701-
1016 3709 (2012).

1018 64. Beaulieu JM, O'Meara BC. Detecting hidden diversification shifts in models of trait-
1019 dependent speciation and extinction. *Systematic biology* **65**, 583-601 (2016).

1021 65. FitzJohn RG. Diversitree: comparative phylogenetic analyses of diversification in R. *Methods*
1022 *in Ecology and Evolution* **3**, 1084-1092 (2012).

1024 66. Matzke NJ. Model selection in historical biogeography reveals that founder-event speciation
1025 is a crucial process in island clades. *Systematic Biology*, syu056 (2014).

1027 67. Eude A, *et al.* Controls on timing of exhumation and deformation in the northern Peruvian
1028 eastern Andean wedge as inferred from low-temperature thermochronology and balanced
1029 cross section. *Tectonics* **34**, 715-730 (2015).

1031 68. Monsch KA. Miocene fish faunas from the northwestern Amazonia basin (Colombia, Peru,
1032 Brazil) with evidence of marine incursions. *Palaeogeography, Palaeoclimatology,*
1033 *Palaeoecology* **143**, 31-50 (1998).

1035 69. Boonstra M, Ramos M, Lammertsma E, Antoine P-O, Hoorn C. Marine connections of
1036 Amazonia: Evidence from foraminifera and dinoflagellate cysts (early to middle Miocene,
1037 Colombia/Peru). *Palaeogeography, Palaeoclimatology, Palaeoecology* **417**, 176-194 (2015).

1039 70. Lovejoy NR, Albert JS, Crampton WG. Miocene marine incursions and marine/freshwater
1040 transitions: Evidence from Neotropical fishes. *Journal of South American Earth Sciences* **21**, 5-
1041 13 (2006).

1043 71. Lieberman BS. Phylogenetic biogeography with and without the fossil record: gauging the
1044 effects of extinction and paleontological incompleteness. *Palaeogeography,*
1045 *Palaeoclimatology, Palaeoecology* **178**, 39-52 (2002).

1047 72. Sanmartín I, Meseguer AS. Extinction in phylogenetics and biogeography: from timetrees to
1048 patterns of biotic assemblage. *Frontiers in genetics* **7**, (2016).

1050 73. Chazot N, *et al.* Contrasting patterns of Andean diversification among three diverse clades of
1051 Neotropical clearwing butterflies. *Ecology & Evolution* **in press**, (2017).

1053
1054 74. Gentry AH. Neotropical floristic diversity: phytogeographical connections between Central
1055 and South America, Pleistocene climatic fluctuations, or an accident of the Andean orogeny?
1056 *Annals of the Missouri Botanical Garden* **69**, 557-593 (1982).

1057
1058 75. Chazot N, Willmott KR, Freitas AVL, de Silva DL, Pellens R, Elias M. Patterns of Species,
1059 Phylogenetic and Mimicry Diversity of Clearwing Butterflies in the Neotropics. In: *Biodiversity*
1060 *Conservation and Phylogenetic Systematics: Preserving our evolutionary heritage in an*
1061 *extinction crisis* (ed^(eds Pellens R, Grandcolas P). Springer International Publishing (2016).

1062
1063 76. Roncal J, Kahn F, Millan B, Couvreur TL, Pintaud JC. Cenozoic colonization and diversification
1064 patterns of tropical American palms: evidence from Astrocaryum (Arecaceae). *Botanical*
1065 *Journal of the Linnean Society* **171**, 120-139 (2013).

1066
1067 77. Whinnett A, Brower AV, Lee M-M, Willmott KR, Mallet J. Phylogenetic utility of Tektin, a
1068 novel region for inferring systematic relationships among Lepidoptera. *Annals of the*
1069 *Entomological Society of America* **98**, 873-886 (2005).

1070
1071 78. Wahlberg N, Wheat CW. Genomic outposts serve the phylogenomic pioneers: designing
1072 novel nuclear markers for genomic DNA extractions of Lepidoptera. *Systematic Biology* **57**,
1073 231-242 (2008).

1074
1075 79. Nguyen L-T, Schmidt HA, von Haeseler A, Minh BQ. IQ-TREE: a fast and effective stochastic
1076 algorithm for estimating maximum-likelihood phylogenies. *Molecular biology and evolution*
1077 **32**, 268-274 (2015).

1078
1079 80. Trifinopoulos J, Nguyen L-T, von Haeseler A, Minh BQ. W-IQ-TREE: a fast online phylogenetic
1080 tool for maximum likelihood analysis. *Nucleic acids research*, gkw256 (2016).

1081
1082 81. Drummond AJ, Suchard MA, Xie D, Rambaut A. Bayesian phylogenetics with BEAUti and the
1083 BEAST 1.7. *Molecular biology and evolution* **29**, 1969-1973 (2012).

1084
1085 82. Lanfear R, Calcott B, Ho SY, Guindon S. PartitionFinder: combined selection of partitioning
1086 schemes and substitution models for phylogenetic analyses. *Molecular biology and evolution*
1087 **29**, 1695-1701 (2012).

1088
1089 83. Särkinen T, Bohs L, Olmstead RG, Knapp S. A phylogenetic framework for evolutionary study
1090 of the nightshades (Solanaceae): a dated 1000-tip tree. *BMC Evolutionary Biology* **13**, 214
1091 (2013).

1092
1093 84. Britton T, Anderson CL, Jacquet D, Lundqvist S, Bremer K. Estimating divergence times in
1094 large phylogenetic trees. *Systematic biology* **56**, 741-752 (2007).

1095

1096 85. Maddison WP, Maddison DR. Mesquite: a modular system for evolutionary analysis. (2001).

1097

1098 86. Miller MA, Pfeiffer W, Schwartz T. Creating the CIPRES Science Gateway for inference of large

1099 phylogenetic trees. In: *Gateway Computing Environments Workshop (GCE), 2010* (ed^eds).

1100 Ieee (2010).

1101

1102 87. Rabosky DL, *et al.* BAMMtools: an R package for the analysis of evolutionary dynamics on

1103 phylogenetic trees. *Methods in Ecology and Evolution* **5**, 701-707 (2014).

1104

1105 88. Beckman EJ, Witt CC. Phylogeny and biogeography of the New World siskins and goldfinches:

1106 Rapid, recent diversification in the Central Andes. *Molecular phylogenetics and evolution* **87**,

1107 28-45 (2015).

1108

1109





