

Ubiquity and quantitative significance of bacterioplankton lineages inhabiting the oxygenated hypolimnion of deep freshwater lakes

Yusuke Okazaki^{1#}, Shohei Fujinaga¹, Atsushi Tanaka², Ayato Kohzu², Hideo Oyagi³, and Shin-ichi Nakano¹

¹Center for Ecological Research, Kyoto University, 2-509-3 Hirano, Otsu, Shiga 520-2113, Japan

²National Institute for Environmental Studies, 16-2 Onogawa, Tsukuba, Ibaraki 305-8506, Japan

³College of Humanities and Sciences, Nihon University, 3-25-40 Sakurajosui Setagaya-City, Tokyo 156-8550, Japan

[#]Corresponding author: Center for Ecological Research, Kyoto University, 2-509-3 Hirano, Otsu, Shiga 520-2113, Japan, E-mail: yoka@ecology.kyoto-u.ac.jp, Phone: +81-77-549-8200, Fax: +81-77-549-8201

ABSTRACT

Freshwater bacterioplankton in the oxygenated hypolimnion are reportedly dominated by specific members that are distinct from those in the epilimnion. However, no consensus exists regarding the ubiquity and abundance of these bacterioplankton, which is necessary to evaluate their ecological importance. The present study investigated the bacterioplankton community in the oxygenated hypolimnia of 10 deep freshwater lakes. Despite the broad geochemical characteristics of the lakes, 16S rRNA gene sequencing demonstrated that many predominant lineages in the hypolimnion were shared by several lakes and consisted of members occurring in the entire water layer and members specific to the hypolimnion. Catalyzed reporter deposition fluorescence *in situ* hybridization (CARD-FISH) revealed that representative hypolimnion-specific lineages, CL500–11 (*Chloroflexi*), CL500–3, CL500–37, CL500–15 (*Planctomycetes*), and the MGI group (*Thaumarchaeota*), together accounted for 1.5–32.9% of all bacterioplankton in the hypolimnion of the lakes. Furthermore, an analysis of micro-diversification based on single-nucleotide variation in the partial 16S rRNA gene sequence (oligotyping) suggested the presence of hypolimnion-specific ecotypes among the lineages occurring in the entire water layer (e.g., *acI* and *Limnohabitans*). Collectively, these results demonstrate the uniqueness, ubiquity, and quantitative significance of bacterioplankton in the oxygenated hypolimnion, motivating future studies to focus on their eco-physiological characteristics.

INTRODUCTION

The hypolimnion is the dark cold water layer that lies below the thermocline in a thermally stratified lake. In a deep holomictic oligo-mesotrophic lake, this layer remains oxygenated throughout the year, as heterotrophic oxygen demand does not exceed the stock of hypolimnetic oxygen. In such a lake, accumulation and remineralization of organic matter in the hypolimnion are important biogeochemical processes (Wetzel, 2001). Recent studies have suggested that dissolved organic matter (DOM) in the oxygenated hypolimnion is enriched by the slowly consumed semi-labile fraction (Maki *et al.*, 2010) and can be transformed into a more recalcitrant form by microbes (Thottathil *et al.*, 2013; Hayakawa *et al.*, 2016), as shown by the microbial carbon pump theory proposed for the ocean (Ogawa *et al.*, 2001; Yamashita and Tanoue, 2008; Jiao and Zheng, 2011; Hansell, 2013). Other studies have demonstrated that nitrification (Small *et al.*, 2013), dark carbon fixation (Callieri *et al.*, 2014), and methane oxidation (Murase and Sugimoto, 2005; Bornemann *et al.*, 2016) are also present in the oxygenated hypolimnion. The bacterioplankton inhabiting this realm are responsible for these important biogeochemical processes; thus, their eco-physiological characteristics should be studied.

Bacterioplankton affiliated with the phyla *Actinobacteria*, *Proteobacteria*, and *Bacteroidetes* are globally predominant in freshwater systems (Zwart *et al.*, 2002; Newton *et al.*, 2011). However, these data are based on surface water studies, and members of other phyla could dominate the oxygenated hypolimnion. Among them, the *Chloroflexi* CL500–11 clade (Urbach *et al.*, 2001, 2007; Okazaki *et al.*, 2013; Deneff *et al.*, 2016) and the *Thaumarchaeota* Marine Group I (MGI) group (Urbach *et al.*, 2001, 2007; Auguet *et al.*, 2012; Vissers *et al.*, 2013; Coci *et al.*, 2015; Mukherjee *et al.*, 2016) are the most investigated. The relatively large cell size (1–2 μm) and high abundance (>15% of all prokaryotes) of CL500–11 suggest their quantitative importance in the oxygenated hypolimnion. MGI are ammonia-oxidizing archaea, and members in the oxygenated hypolimnion are affiliated with either *Nitrosopumilus* or *Candidatus Nitrosoarchaeum* (Coci *et al.*, 2015). Other nitrifiers such as *Nitrospira* and *Nitrospira* have also been found in the water column of the oxygenated hypolimnion (Mukherjee *et al.*, 2016; Okazaki and Nakano, 2016; Fujimoto *et al.*, 2016). Moreover, high-throughput sequencing of the 16S rRNA partial amplicon has highlighted members of *Planctomycetes* (e.g., CL500–3, CL500–15, and CL500–37) as representative lineages in the oxygenated hypolimnion (Rozmarynowycz, 2014; Okazaki and Nakano, 2016). These inhabitants are potentially important components of the microbial food web and biogeochemical cycling in the deep pelagic freshwater ecosystem. However, it remains unknown how ubiquitously and abundantly they are distributed in the oxygenated hypolimnion. Due to a lack of quantitative data, their ecological importance remains poorly understood.

The present study investigated the bacterioplankton community in the oxygenated hypolimnia of 10 deep freshwater lakes with a variety of geochemical characteristics. Community composition was investigated by 16S rRNA amplicon sequencing, and several representative members

were microscopically characterized and quantified by CARD-FISH. This approach allowed us to create a general overview of the bacterioplankton community inhabiting the oxygenated hypolimnion of deep freshwater lakes and identify abundant and ubiquitous lineages. Moreover, analyses of their habitat preference and micro-diversification (oligotyping) facilitated hypotheses about their eco-physiological characteristics and potential diversified ecotypes.

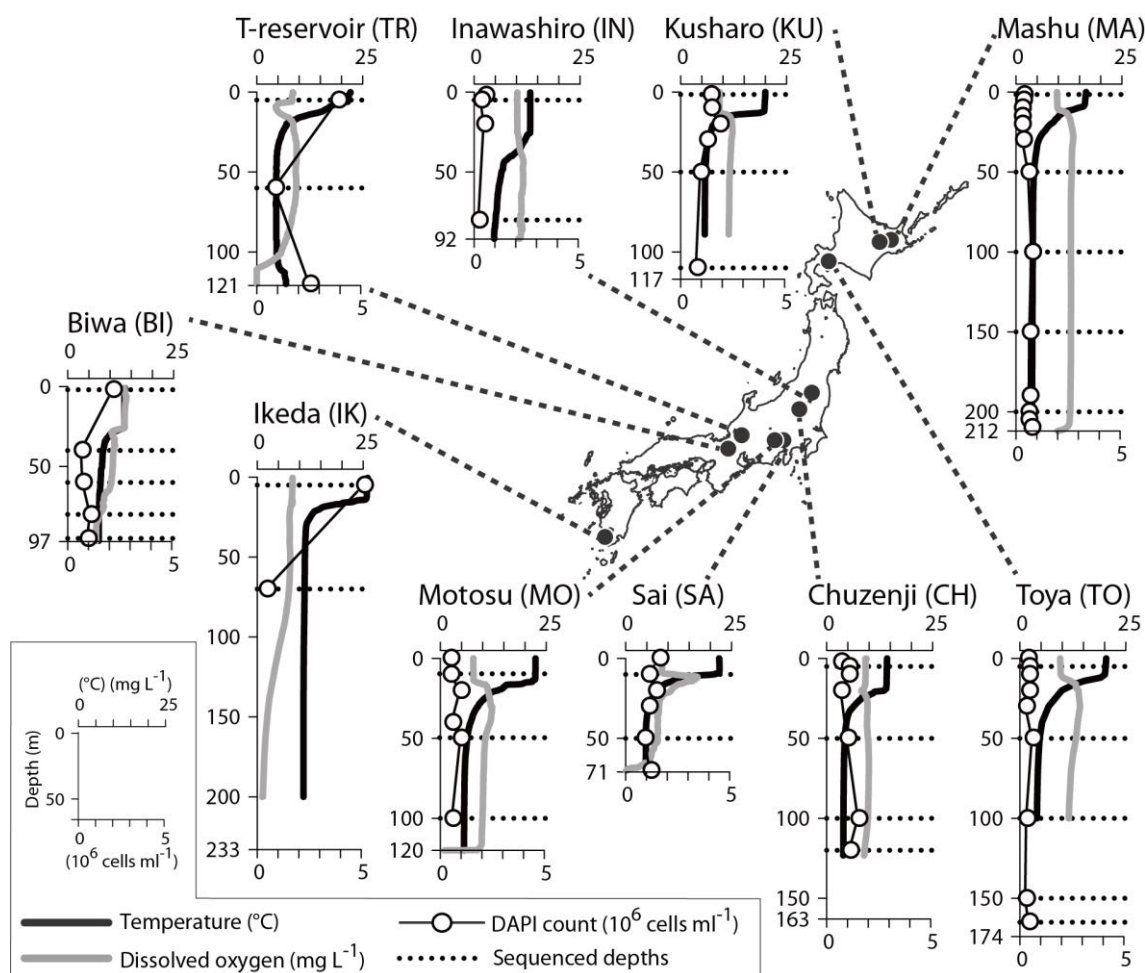


Fig. 1. Locations and vertical profiles of the sampling sites.

MATERIALS AND METHODS

Field sampling

Water samples were collected at pelagic sites in 10 deep freshwater lakes in Japan from August to December in 2015, including Lake Mashu, Kusharo, Toya, Inawashiro, Chuzenji, Sai, Motosu, Biwa, Ikeda, and T-Reservoir (hereafter, MA, KU, TO, IN, CH, SA, MO, BI, IK, and TR, respectively) (Fig. 1). The profiles of the sampling locations are summarized in Table 1. In all, 3 to 13 depths were sampled in each lake, and the temperature and dissolved oxygen vertical profiles were

measured using a CTD probe *in situ*. These lakes have permanently oxygenated hypolimnia and were thermally stratified when the sampling was carried out (Fig. 1). Total prokaryotic abundance was determined by microscopic enumeration of DAPI-stained cells (Porter and Feig, 1980).

	Max. Depth (m)	Depth of sampling point (m)	Surface area (km ²)	Water Volume (10 ⁶ m ³)	Surface Altitude (m)	Lake Origin	Water Retention Time (year)	Trophic Status	pH*	Total Nitrogen (mg/L)*	Total Phosphorus (mg/L)*	Sampling Location	Sampling date
Mashu (MA)	211.4	211	19.2	2640	351	Caldera	200	Oligotrophic	7.0–8.0	0.06–0.08	<0.003–0.004	43°35'16.0"N, 144°31'50.0"E	2015.8.26–29
Kusharo (KU)	117.5	117	79.6	2261	121	Caldera	12	Oligotrophic	7.0–7.5	0.07–0.24	<0.003–0.004	43°35'26.0"N, 144°19'48.0"E	2015.8.25
Toya (TO)	179.7	174	70.7	8272	84	Caldera	9	Mesotrophic	7.1–7.8	0.24–0.38	0.003–0.01	42°36'15.1"N, 140°49'07.7"E	2015.9.24
Inawashiro (IN)	93.5	92	103.3	5320	514	Tectonic	5.4	Oligotrophic	6.6–7.0	0.17–0.34	<0.003–0.004	37°28'27.6"N, 140°05'40.5"E	2015.10.30
Chuzenji (CH)	163	163	11.8	1116	1269	Dammed	5.9	Oligotrophic	7.7–8.8	0.07–0.27	<0.003–0.009	36°43'55.7"N, 139°28'25.9"E	2015.10.21
Sai (SA)	71.7	71	2.1	81	900	Dammed	1.6	Oligotrophic	7.4–8.8	0.06–0.32	0.003–0.008	35°29'49.5"N, 138°41'09.5"E	2015.9.8
Motosu (MO)	121.6	120	4.7	319	900	Dammed	6.5	Oligotrophic	6.8–7.9	0.15–0.25	<0.003–0.006	35°28'06.5"N, 138°34'57.8"E	2015.9.7
T-Reservoir (TR)	121	121	13	660	390	Artificial Reservoir	0.27	Mesotrophic	7.2–9.2	0.18–0.46	<0.003–0.008	35°40'26.0"N, 136°29'57.0"E	2015.9.16
Biwa (BI)	103.8	97	670.3	27616	85	Tectonic	5.5	Mesotrophic	7.7–8.4	0.15–0.32	0.005–0.009	35°23'21.0"N, 136°07'51.0"E	2015.12.9
Ikeda (IK)	233	233	10.9	1368	66	Caldera	33.5	Mesotrophic	7.1–8.6	0.14–0.32	<0.003–0.006	31°14'14.0"N, 130°33'52.0"E	2015.9.17

Table 1. Profiles of the sites studied. Data were collected from Mori and Sato (2015) and the Japanese Ministry of the Environment public water database. *Annual range recorded in the epilimnion.

Partial 16S rRNA gene amplicon sequencing

In total, 33 DNA samples were collected (Fig. 1). Immediately after sampling, 70–300 mL the water collected was filtered through a 0.2 µm polycarbonate filter (47 mm diameter; Whatman, Maidstone, UK). The filter was stored at –20°C until DNA was extracted using the PowerSoil DNA Isolation Kit (MoBio Laboratories, Carlsbad, CA, USA). The V4 and V5 regions of the 16S rRNA gene were amplified using the modified 530F and 907R primers (Nunoura *et al.*, 2009) and then an eight-base-pair DNA tag (for post-sequencing sample identification) and 454 adaptors were conjugated by second PCR, as described previously (Okazaki and Nakano, 2016). The PCR products from the samples were pooled in equimolar quantities and sequenced in the 1/8 regions of a sequencing reaction on the Roche 454 GS-FLX Titanium sequencer (Roche Science, Mannheim, Germany). The nucleotide sequence data are available in the Sequence Read Archive database under accession numbers DRX062810–DRX062842 (BioProject: PRJDB5151).

Analysis of sequencing reads

The sequence data were processed using the UPARSE pipeline (Edgar, 2013) following the author's instructions (http://www.drive5.com/usearch/manual/upp_454.html). We used a fastq_maxee value of 1.0, a truncated length of 350 bp, and an OTU creation identity threshold of 97%. Thereafter, respective OTUs were taxonomically assigned with the SINA 1.2.11 (Pruessner *et al.*, 2012) online tool (<https://www.arb-silva.de/aligner/>) referring to the 123 Ref database (Quast *et al.*, 2013) and using the default parameter settings. Subsequently, non-prokaryotic OTUs (i.e., chloroplast, eukaryote, and

unclassified domains) were removed. The resulting 96 149 reads were used for subsequent analyses.

Before calculating community diversity, coverage-based rarefaction (Chao and Jost, 2012) was applied. Reads were randomly discarded from each sample until coverage was <96.59% (i.e., slope of the rarefaction curve was >0.0341), which was the minimum value recorded among samples. Subsequently, alpha (Shannon diversity) and beta diversities (non-metric multidimensional scaling [NMDS] based on Bray–Curtis dissimilarity) were calculated. These analyses were performed using the *vegan* 2.4–0 package (Oksanen *et al.*, 2016) in R 3.3.1 software (<http://www.R-project.org/>).

The OTUs were classified following the nomenclature proposed by Newton *et al.* (2011). Using the “--search” option in the SINA 1.2.11 (Pruesse *et al.*, 2012) stand-alone tool, the representative sequences of the respective OTUs (generated by the UPARSE pipeline) were searched and classified against the original ARB (Ludwig *et al.*, 2004) database provided by Newton *et al.* (2011). Dominant OTUs that failed to be classified by this procedure were named manually. If closely related (>99% identity) sequences with the prefixes “CL” (Urbach *et al.*, 2001) or “LiUU-” (Eiler and Bertilsson, 2004) were in the public sequence database, their names were preferentially used. The *Planctomycetes* phylogenetic clades were newly defined for the collective descriptions (Fig. S1). In other cases, the OTU was shown by its taxonomic affiliation (e.g., genus) based on the SILVA nomenclature.

The sequences were aligned using the SINA 1.2.11 (Pruesse *et al.*, 2012) stand-alone tool against the SILVA 123 Ref NR 99 database to construct a phylogenetic tree (Quast *et al.*, 2013). Maximum Likelihood trees were calculated by the RAxML 8.2.4 software (Stamatakis, 2014) with the GTR substitution model and the GAMMA rate heterogeneity model. The trees were drawn in MEGA 7 software (Kumar *et al.*, 2016).

Oligotyping

Intra-OTU micro-diversity of representative lineages was analyzed by oligotyping, which facilitated detection of single nucleotide variation by excluding the effects of sequencing errors based on the Shannon entropy values (Eren *et al.*, 2013). Following the author’s instructions (<http://merenlab.org/software/oligotyping/>), the quality filtered FASTA file generated by the UPARSE pipeline was split at individual OTUs and aligned against the Greengenes (DeSantis *et al.*, 2006) alignment database using the SINA 1.2.11 (Pruesse *et al.*, 2012) stand-alone tool. Uninformative columns were removed by subsequently applying the “o-trim-uninformative-columns-from-alignment” and “o-smart-trim” scripts. Several rounds of oligotyping were repeated by manually choosing the most informative (i.e., the highest entropy) column until all oligotypes with > 100 reads exceeded the purity score of 0.90. The minimum substantive abundance parameter (option-M) was always set to 10.

CARD-FISH

CARD-FISH was performed based on Pernthaler *et al.* (2002) with some modifications as described previously (Okazaki *et al.*, 2013). Specifically, the hybridization, amplification, and washing

steps were carried out at 46°C. AlexaFluor 488 (Life Technologies, Carlsbad, CA, USA) was used as the tyramide-conjugated fluorescent dye. The oligonucleotide probes were designed previously for CL500–11 (Okazaki *et al.*, 2013) and MGI (Coci *et al.*, 2015), and newly designed for the other targets (Table 2). The probes were constructed using the “Design Probes” function in ARB 6.0.3 software (Ludwig *et al.*, 2004) against the SILVA 123 Ref NR 99 database (Quast *et al.*, 2013). Specificity of the probes was confirmed by an NCBI BLAST online search and the Test Probe 3.0 tool against the SILVA 123 Parc database (<https://www.arb-silva.de/search/testprobe/>). The probes designed in this study targeted the V4 or V5 regions of the 16S rRNA gene, which were read by amplicon sequencing. We confirmed that the probes perfectly and exclusively matched all sequencing reads affiliated with the target lineage. To enhance the fluorescent signal, oligonucleotide helpers (Fuchs *et al.*, 2000) were used for several probes (Table 2). The helpers were designed to target the adjacent or opposite loci of the probe target site to loosen the secondary rRNA structure (Fuchs *et al.*, 2000) by confirming that all probe-targets in the database were not mismatched with their corresponding helpers. The hybridization buffer contained 0.5 µg mL⁻¹ probe and 0.1 µg mL⁻¹ of each helper. The formamide concentration in the buffer was determined for each probe (Table 2) by testing a series of concentrations to obtain the best stringency (i.e., highest concentration without signal loss). The stringency test was performed in samples with a high read proportion of the target determined by amplicon sequencing. The hybridized filters were counterstained with DAPI and enumerated under an epifluorescence microscope. At least 300 DAPI-positive cells and the corresponding FISH-positive cells were enumerated three times per sample (the same filter piece). A negative NON338 probe control (Wallner *et al.*, 1993) confirmed that no false-positive cells were present.

Probe	Sequence	Length (base)	Formamide	Target	Reference
CLGNS_584	GCCGACTTGCCCAACCTC	18	20%	CL500-11	Okazaki et al. (2013)
CLGNS_567h	CTACACGCCCTTTACGCC	18			
CL500-3_829	CGTTCAACCCCTCGACC	18	20%	CL500-3	This study
CL500-3_847h	ATCAGTTTTCTACGGCCGAGAAGG	25			
CL500-3_804h	AAGTGCGCATCGTTTACGGCGTGGA	25			
CL500-37_658	CGTTCCACCCGCCTCTACCTG	21	30%	CL500-37	This study
CL500-15_826	ACAGGGAGACCCCAAACCTAG	20	30%	CL500-15	This study
CL500-15_846h	TCACTTTTGTTCGCCTGGGATCCC	25			
CL500-15_801h	TGCCCATCGTTTACAGCTAGGACTA	25			
MGI_535	TCCTGACCACTTGAGGTGCTGG	22	25%	MGI	Coci et al. (2015)

Table 2. Catalyzed reporter deposition fluorescence *in situ* hybridization (CARD-FISH) probes, helpers, and formamide concentrations (at 46°C) used in this study.

RESULTS

16S rRNA partial amplicon sequencing generated 666 OTUs from the 96 149 reads originating from the 33 samples collected from the water columns of the 10 lakes. The complete dataset generated by the pipeline is available in Supplementary Information.

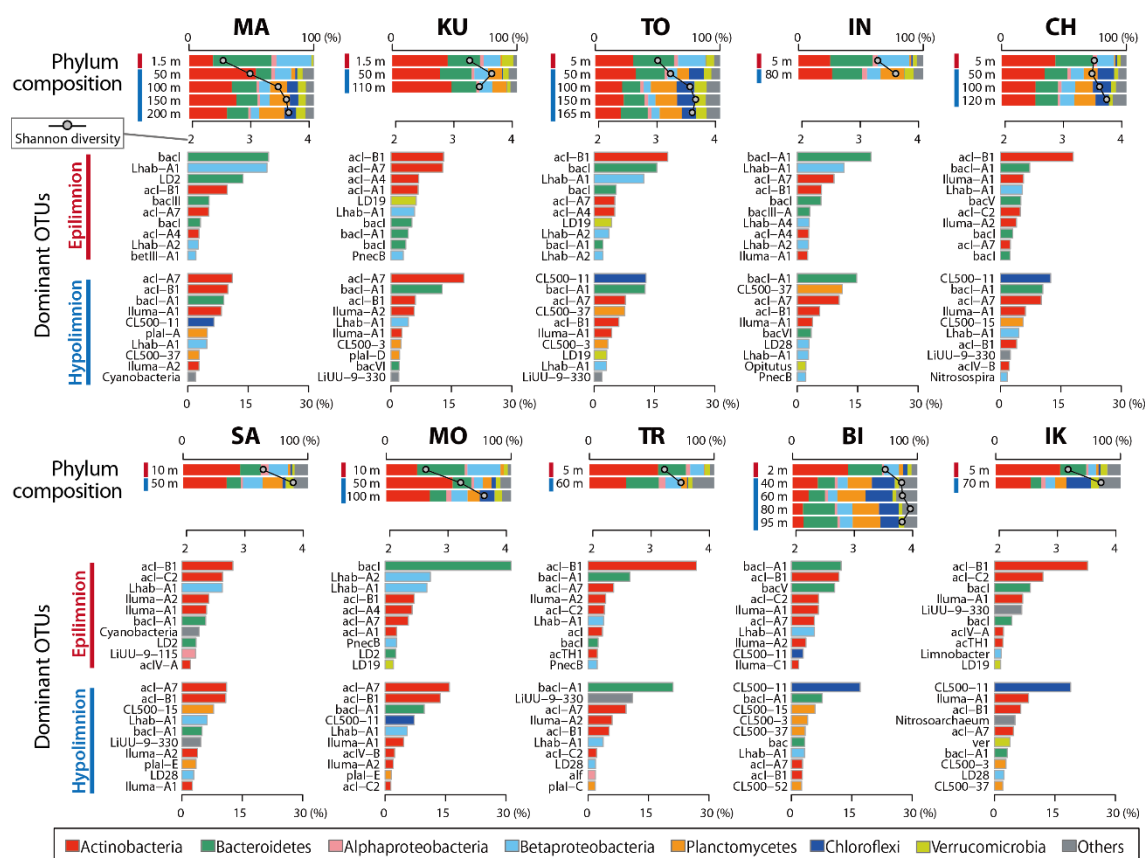


Fig. 2. Composition of the 16S amplicon reads. The top panel (band graphs) displays phylum-resolved community composition at each depth for each lake, with an overwritten line graph indicating the alpha (Shannon) diversity. The two lower panels show the 10 dominant operational taxonomic units (OTUs) in the epilimnion and hypolimnion, composed of averaged data for each layer (the depths averaged are illustrated by red and blue lines in the top panel). Bar colors indicate phyla to which individual OTUs were assigned.

Phyla *Actinobacteria*, *Bacteroidetes*, and class *Betaproteobacteria* generally dominated throughout the water column, whereas *Chloroflexi* and *Planctomycetes* were also predominant phyla in the hypolimnion (Fig. 2). The epilimnetic and hypolimnetic communities were compared at the OTU level by averaging the samples from each layer (Fig. 2). The dominant members in the epilimnion were generally shared between the lakes (e.g., acI-B1, acI-A7, Lhab-A1, and bacI-A1). In addition to the lineages common to the epilimnion, *Chloroflexi* and *Planctomycetes* were also ranked as dominant OTUs in the hypolimnion (Fig. 2). CL500-11 alone accounted for most of the *Chloroflexi* reads, whereas *Planctomycetes* consisted of diverse OTUs (e.g., CL500-3, CL500-15, CL500-37, and plaI-A) (Fig. 2), which were affiliated with three classes in the phylum (Fig. S1). The alpha (Shannon) diversity was higher in the hypolimnion than that in the epilimnion of all 10 lakes (Fig. 2). The beta diversity analysis (NMDS) clearly separated the hypolimnetic communities from the epilimnetic communities, but no clear separation between the lakes was observed (Fig. 3).

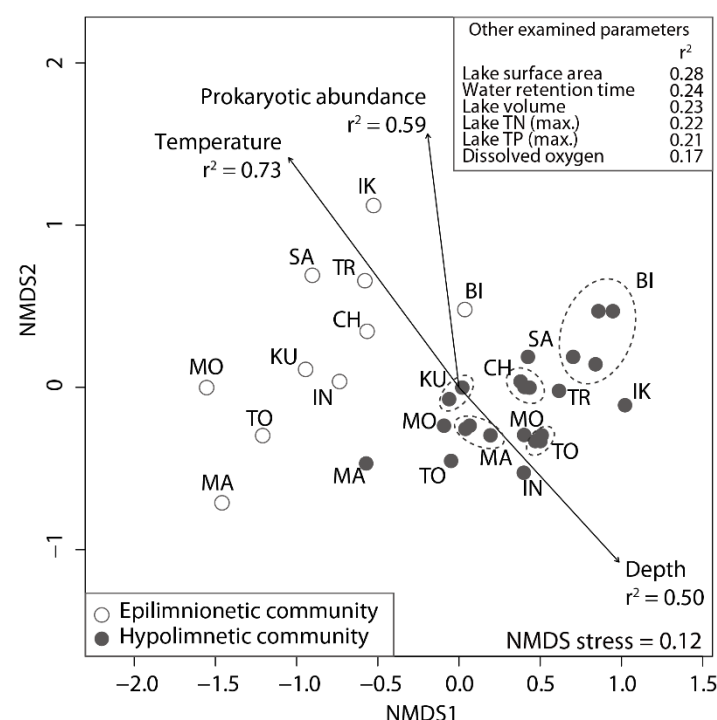


Fig. 3. Non-metric multidimensional scaling (NMDS) ordination of all 33 bacterioplankton communities. The arrows are fitted vectors for the environmental variables calculated using the envfit function. The direction of the arrow indicates the direction at which the gradient of the environmental variable was maximum. The length of the arrow is proportional to the squared correlation coefficient (r^2). Only significant variables ($p < 0.01$; based on 999 permutations) were visualized. The other variables examined are shown in the top right box with the r^2 values.

Vertical preference of the bacterioplankton in each OTU was determined based on the number of lakes where it was abundant ($>1\%$ of all amplicon reads) in each water layer (Fig. 4). Seven OTUs were defined as “whole-layer generalists,” which were abundant in either the epilimnion or hypolimnion of more than eight lakes (Fig. 4). Twenty-two OTUs were defined as “hypolimnion specialists,” which were abundant in the hypolimnion of multiple lakes but not abundant in the epilimnion of almost all (nine or ten) lakes. Conversely, 19 OTUs were identified as “epilimnion specialists” (Fig. 4). Several bacterioplankton in the hypolimnion showed a habitat preference between the lakes, but only few environmental parameters significantly explained the patterns (Fig. 5). The oligotyping analysis revealed that the whole-layer generalists exhibited more intra-OTU diversity than that of the hypolimnion specialists (Fig. 5). The composition of the oligotypes in each sample is shown in Supplementary Figure S2.

The newly constructed FISH probes targeted a monophyletic clade of the target lineages (Fig. S3). Enumeration of the positive cells revealed that CL500–11 accounted for $>10\%$ of all prokaryotic cells in four lakes (maximum was 25.9% at 60 m in BI). CL500–3, CL500–37, and MG1 respectively accounted for $>3\%$ in several lakes. CL500–15 were less abundant but still detectable, with a maximum percentage of 1.6% (Fig. 6). The cells detected in each target shared identical morphology between the lakes: CL500–11 were curved rods 1–2 μm long; CL500–3, CL500–37, and

CL500–15 were cocci approximately 1 μm diameter; and MG1 were rods around 1 μm long (Fig. 6). CL500–15 cells were often found to be particle-associated (51% of the cells detected at 50 m in SA), whereas the other lineages were mostly planktonic. The correlation between relative abundance determined by amplicon sequencing and CARD-FISH was significant for all targets, but the CARD-FISH estimates tended to be lower than those of amplicon sequencing (Fig. S4).

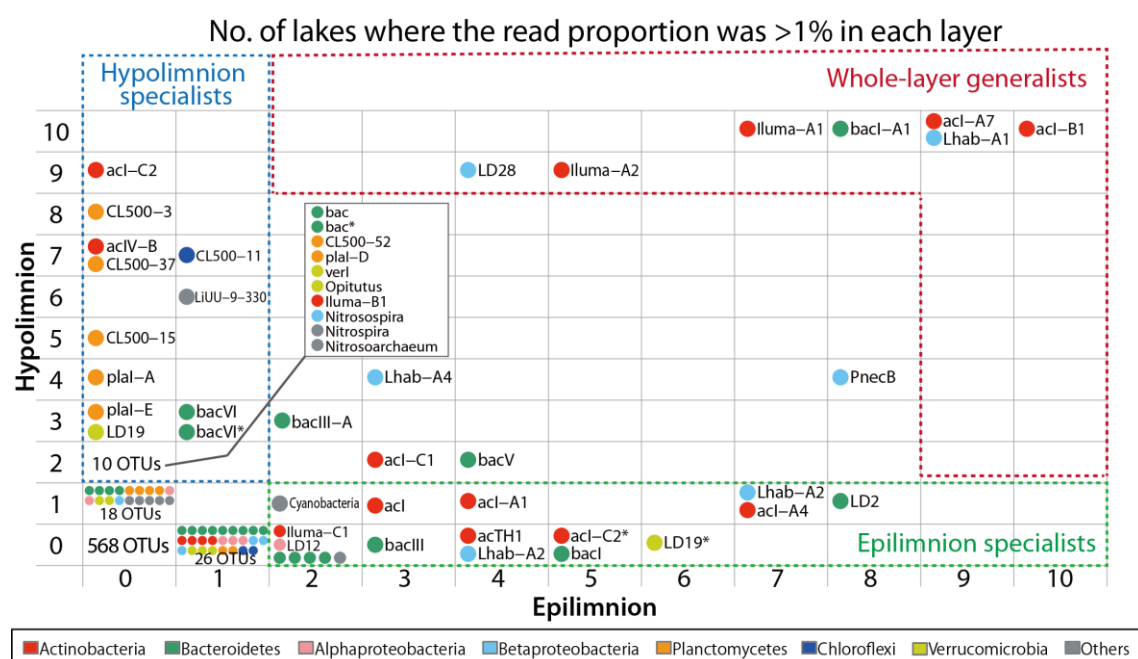


Fig. 4. Vertical preferences of individual operational taxonomic units (OTUs), mapped by the number of lakes where individual OTUs accounted for >1% of all amplicon reads in each water layer. Data for the hypolimnion were generated by averaging the data at multiple depths in the hypolimnion. Dotted boxes highlight the groups defined in this study (see main text). Point color indicates the phylum to which an individual OTU was assigned. Asterisks in the group name distinguish the different OTUs assigned to the same group.

DISCUSSION

Structure of the bacterioplankton community in the oxygenated hypolimnion

The present study investigated lakes with a variety of environmental characteristics, ranging from a cold (hypolimnetic temperature = 4°C) oligotrophic lake (MA) to a warm (11°C) mesotrophic lake (IK) (Fig. 1 and Table 1). Nevertheless, the bacterioplankton communities in the oxygenated hypolimnion were relatively similar among the lakes and separated from communities in the epilimnion (Fig. 3), indicating that the oxygenated hypolimnion is an independent niche with distinct microbial communities. The community consisted of members that were ubiquitous and predominant across the lakes and water layers (whole-layer generalists), and members that exclusively inhabited the oxygenated hypolimnion (hypolimnion specialists) (Fig. 4). The whole-layer generalists were composed of several commonly known freshwater bacterioplankton lineages (e.g., acI, Iluma-A1, Iluma-A2, Lhab-A1, LD28, and bacI-A1), whereas the hypolimnion specialists were represented by phyla that were not common to the epilimnion, including *Chloroflexi* CL500–11, members of

Planctomycetes (e.g., CL500–3, CL500–15, CL500–37, and plaI-A), and *Ca. Nitrosoarchaeum* in *Thaumarchaeota* (Figs. 2, 4). Their abundance and ubiquity were demonstrated by the CARD-FISH analysis, showing that they collectively accounted for 1.5% (TR) to 32.9% (BI) of all bacterioplankton in the hypolimnion. Our analysis also revealed that these groups were not cosmopolitan and were actually absent in some lakes (e.g., IN and KU for CL500–11, CH for CL500–3, and MO for CL500–37) (Figs. 5, 6), suggesting that they have respective habitat preferences and ecological niches.

Ubiquity, quantitative importance, and potential eco-physiology of the hypolimnion specialists

The predominance (~25% of all bacterioplankton) of planktonic CL500–11 cells in several lakes (Fig. 6) suggests that their resources are diffuse, abundant and ubiquitous. The metagenome-assembled genome and *in situ* transcriptional evidence of CL500–11 in Lake Michigan suggests their importance in peptide turnover (Denef *et al.*, 2016). Peptides in aquatic systems are mainly derived from peptidoglycans in the bacterial cell wall (McCarthy *et al.*, 2013; Nagata *et al.*, 2003) and from proteins released by other bacteria (Tanoue *et al.*, 1995) or phytoplankton (Nguyen and Harvey, 1997; Yamada *et al.*, 2012). Previous studies in BI have collectively demonstrated that N-rich (by stoichiometry) or protein-like (by fluorescence properties) semi-labile dissolved organic matter (DOM) derived from autochthonous phytoplankton production that accumulates in the hypolimnion is slowly remineralized during stratification (Kim *et al.*, 2006; Maki *et al.*, 2010; Thottathil *et al.*, 2013). Thus, it is possible that CL500–11 is scavenging protein-like debris that accumulates in the lake due to its relatively recalcitrant nature. Given that the water retention time of a system affects DOM composition (Kellerman *et al.*, 2014; Catalán *et al.*, 2016) and that autochthonous dissolved proteins can accumulate in a lake even at centennial time scales (Goldberg *et al.*, 2015), we expected that lakes with a longer water retention time would contain more bacterioplankton lineages specialized to consume relatively recalcitrant DOM. In the present study, the water retention time of the lakes ranged from 0.27 years (TR) to 200 years (MA) (Table 1). However, most of the hypolimnion specialists, including CL500–11, were not distributed in a manner that was associated with water retention time (Fig. 5). Nonetheless, it is still possible that a water retention time (<2 years) too short to accumulate semi-labile DOM would have resulted in the absence of CL500–11 in SA and TR (Fig. 6). The absence of CL500–11 in KU and IN (Fig. 6) is more challenging to explain, but may have resulted from the low pH compared to the other lakes (Table 1). As CL500–11 is a large cell (Fig. 6), protistan size-selective grazing (Pernthaler, 2005) may also be a factor controlling CL500–11 dynamics, and little information is available on the grazer communities inhabiting the deep oxygenated hypolimnion (Masquelier *et al.*, 2010; Mukherjee *et al.*, 2015). Although these assumptions are speculative, future studies should verify them given their ubiquity and quantitative importance in deep freshwater systems. Indeed, the dominance of CL500–11 has been reported in the two largest deep freshwater systems on Earth, the Laurentian Great Lakes (Rozmarynowycz, 2014; Denef *et al.*, 2016) and Lake Baikal (Kurilkina *et al.*, 2016).

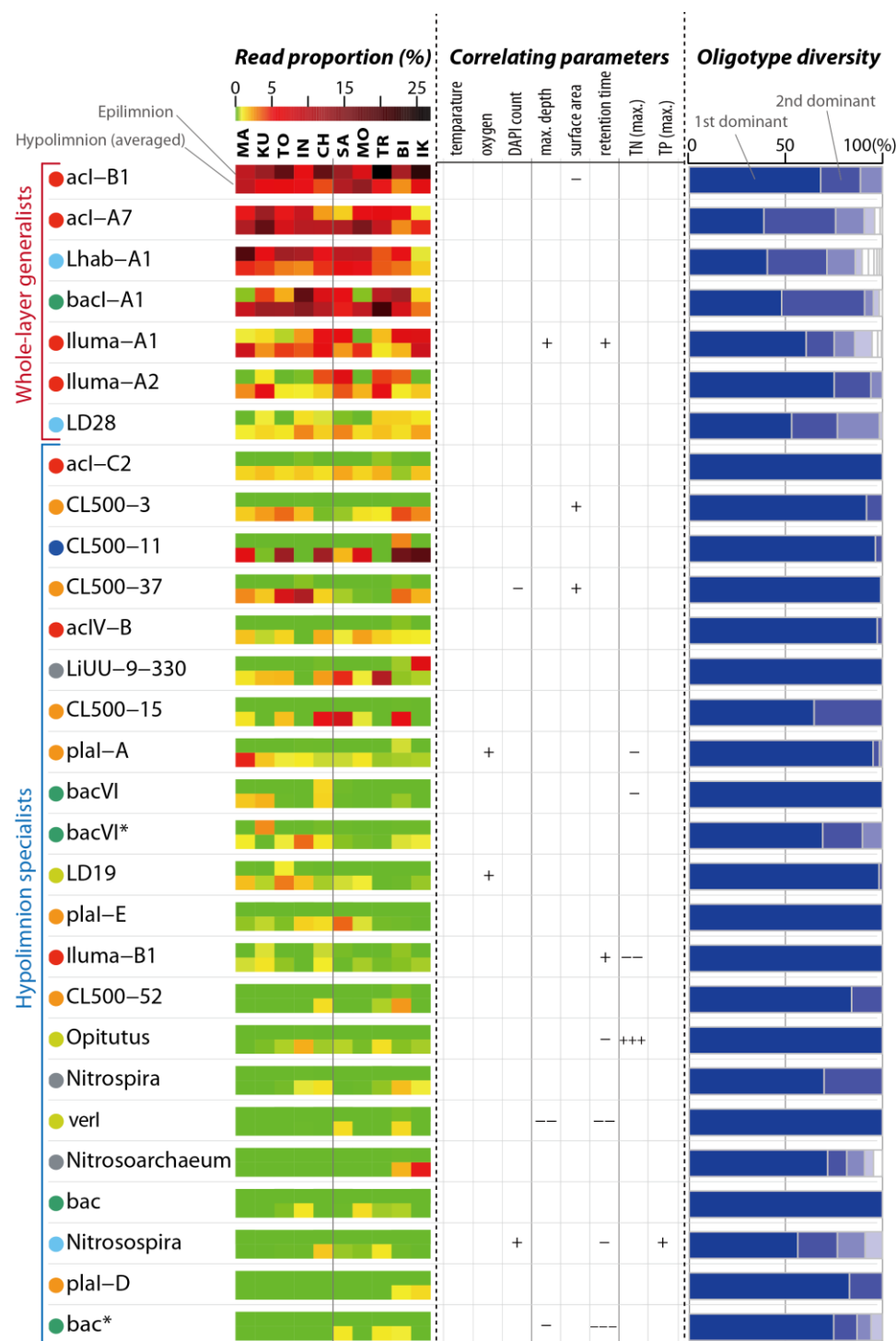


Fig. 5. Distribution patterns (left column), correlating environmental parameters (center), and oligotype diversity (right) of the hypolimnion inhabitants. The distribution pattern is illustrated by the read proportion to total amplicon reads. The hypolimnion data were generated by averaging the data at multiple depths in the hypolimnion. The correlation between read proportion and environmental parameters in the hypolimnion was evaluated by Spearman's test. For a positive correlation, +++, $p < 0.005$; ++, $p < 0.01$; +, $p < 0.05$. For a negative correlation, "-" was shown instead of "+". Maximum values recorded in TN and TP were used based on the database. The right column (band graphs) indicates composition of the oligotypes among all amplicon reads assigned to each operational taxonomic unit (OTU). Point colors indicate phyla to which individual OTUs were assigned (See Figs. 2 and 4 for legend). Asterisks in the group name distinguish the different OTUs assigned to the same group.

CL500–3 and CL500–37 were the two most abundant *Planctomycetes* in the present study (Fig. 6). Each was affiliated with their respective phylogenetic clade in the class *Phycisphaerae* (Fig. S1). Aquatic *Planctomycetes* are often associated with algal blooms (Morris *et al.*, 2006; Pizzetti *et al.*, 2011), and genomic evidence indicates their potential to aerobically consume sulfated polysaccharides derived from algae (Glöckner *et al.*, 2003; Woebken *et al.*, 2007; Erbilgin *et al.*, 2014). In a marine study, sequences closely related to CL500–3 were enriched in DNA extracted from bacterioplankton that assimilate protein secreted by phytoplankton (Orsi *et al.*, 2016). Consequently, it can be hypothesized that CL500–3 and CL500–37 consume polysaccharides or proteins derived from phytoplankton. Given that no photosynthesis occurs in the dark hypolimnion, these organisms presumably rely on substrates released from sinking phytoplankton. Indeed, substantial sinking fluxes of green algae (*Staurastrum*), a diatom (*Fragilaria*), and cyanobacteria (*Synechococcus*) into the hypolimnion have been reported in BI (Kagami *et al.*, 2006; Takasu *et al.*, 2015), where CL500–3 and CL500–37 were both abundant (Fig. 6). Their dependence on pelagic autochthonous resources was also supported by the finding that their read proportions were positively correlated with lake surface area (Fig. 5). However, they did not always co-occur, and disproportional dominance of CL500–3 was found in MO and TR, and of CL500–37 in CH and IN (Fig. 6). Amplicon sequencing data taken from the oxygenated hypolimnion in Lake Michigan (Fujimoto *et al.*, 2016) showed that only CL500–37 was abundant (Fig. S5). These observations indicate that the ecological niches of CL500–3 and CL500–37 are similar but not the same. Given that algal exudates from different phytoplankton species select different bacterial communities (Šimek *et al.*, 2011; Paver *et al.*, 2013), the difference might be attributable to differences in the sinking phytoplankton species in a lake. It should also be noted that their closely related sequences were not necessarily retrieved from the oxygenated hypolimnion and have been found in an Antarctic lake (Karlova *et al.*, 2016), arctic lake (Ntougias *et al.*, 2016), and Baltic Sea ice (Eronen-Rasimus *et al.*, 2015) (Fig. S3A). More information is needed to elucidate the eco-physiological characteristics of these widespread and abundant yet largely overlooked bacterial lineages.

The CL500–15 clade belonged to the uncultured OM190 class (Fig. S1) with only three sequences in the database: two from deep freshwater lakes (Urbach *et al.*, 2001; Pollet *et al.*, 2011) and one from deep sea sediment (Zhang *et al.*, 2013) (Fig. S3B). Another sequence was reported from the littoral water of Lake Baikal (Parfenova *et al.*, 2013). In the present study, CL500–15 was detected in half of the lakes (Fig. 4), indicating that they are one of the most common lineages in the oxygenated hypolimnion. The FISH analysis revealed a high proportion of particle-associated cells (Fig. 6) and microscopic observations revealed that the particles were not cells but rather looked like transparent exopolymer particles (TEP), which are gel-like sticky particles mainly composed of polysaccharides (Passow, 2002). Members of the OM190 class in marine systems have been preferentially detected in the particle-associated fraction (Salazar *et al.*, 2015; Bižić-Ionescu *et al.*, 2015), suggesting that their

particle-associated form is preserved across the class. Particle-associated bacteria can contribute disproportionately to total bacterial activity (Lemarchand *et al.*, 2006; Grossart *et al.*, 2007), and CL500–15 is presumably one of the major TEP decomposers that plays a substantial role in substrate remineralization in the oxygenated hypolimnion.

Other representative *Planctomycetes* were affiliated with class *Planctomycetacia* (plAI-A–F) (Fig. S1). They usually had a smaller proportion of reads than those of CL500–3, CL500–37, and CL500–15 (Fig. 2). However, plAI-A was the most represented *Planctomycetes* based on the read proportion in the oxygenated hypolimnion of MA (Fig. 2) and Lake Michigan (Fig. S5). Each of the other members of plAI (e.g., plAI-B [CL500–52], plAI-D, and plAI-E) showed their respective distribution patterns between the lakes (Fig. 5). Overall, *Planctomycetacia* (plAI group) was generally less ubiquitous and abundant, but more diverse than the other two classes (i.e., *Phycisphaerae* and OM190) inhabiting the oxygenated hypolimnion.

In the present study, the MG1 group was detected only from two lakes (BI and IK) (Fig. 5), and their maximum percentage determined by FISH was 3.8% (Fig. 6). These numbers are lower than those of a previous study that detected 8.7–19% MG1 in the oxygenated hypolimnion of all six subalpine lakes investigated (Callieri *et al.*, 2016). In the present study, MG1 was exclusively affiliated with *Ca. Nitrosoarchaeum* (Blainey *et al.*, 2011), but *Nitrosopumilus*, another predominant MG1 member in the oxygenated hypolimnion (Berdjeb *et al.*, 2013; Vissers *et al.*, 2013; Coci *et al.*, 2015), was not detected. *Ca. Nitrosoarchaeum* has been reported in the oxygenated hypolimnion of Crater Lake (Urbach *et al.*, 2001, 2007), Lake Redon (Auguet *et al.*, 2012), and Lake Superior (Mukherjee *et al.*, 2016), which are oligotrophic lakes with a hypolimnetic temperature of 4°C. The occurrence of *Ca. Nitrosoarchaeum* in BI and IK, which are mesotrophic lakes with a hypolimnetic temperature of 8–11°C (Fig. 1), revealed their broad habitat spectrum. In other lakes (e.g., CH and TR), ammonia-oxidizing bacteria, *Nitrospira*, were detected in the oxygenated hypolimnion (Fig. 5), in agreement with the previously suggested niche separation between ammonia-oxidizing archaea and bacteria (Coci *et al.*, 2015; Mukherjee *et al.*, 2016). Another nitrifier, *Nitrospira*, was also detected in the oxygenated hypolimnion (Fig. 5), in line with previous studies (Small *et al.*, 2013; Okazaki and Nakano, 2016). However, no nitrifiers were detected in the three northern lakes (MA, KU, and TY) (Fig. 5). Notably, their absence should not be concluded by the present data, as the nitrifier community can shift over seasons (Okazaki and Nakano, 2016). Nevertheless, our results indicate the potential diversity of the nitrification systems in the oxygenated hypolimnion, yet information remains scarce to conclude the cause and effects of the diversity.

The discussion above is based on the assumption that each of the bacterioplankton lineages prefer their particular suitable habitat. However, it is also possible that the occurrence of the members is controlled by occurrence of other lineages, as bacterioplankton often have streamlined genomes and are dependent on each other for lost metabolic functions (Morris *et al.*, 2012; Garcia *et al.*, 2015; Mas

et al., 2016). In the present study, many pairs of hypolimnion specialists showed correlating distribution patterns between the lakes (Fig. S6). For example, CL500-3 was positively correlated with CL500-37 and *Ca. Nitrosoarchaeum*, and negatively correlated with *Nitrosospira* (Fig. S6). Although these results do not directly support an interaction, they suggest that some hypolimnion specialists are presumably co-occurring or sharing similar ecological niches.

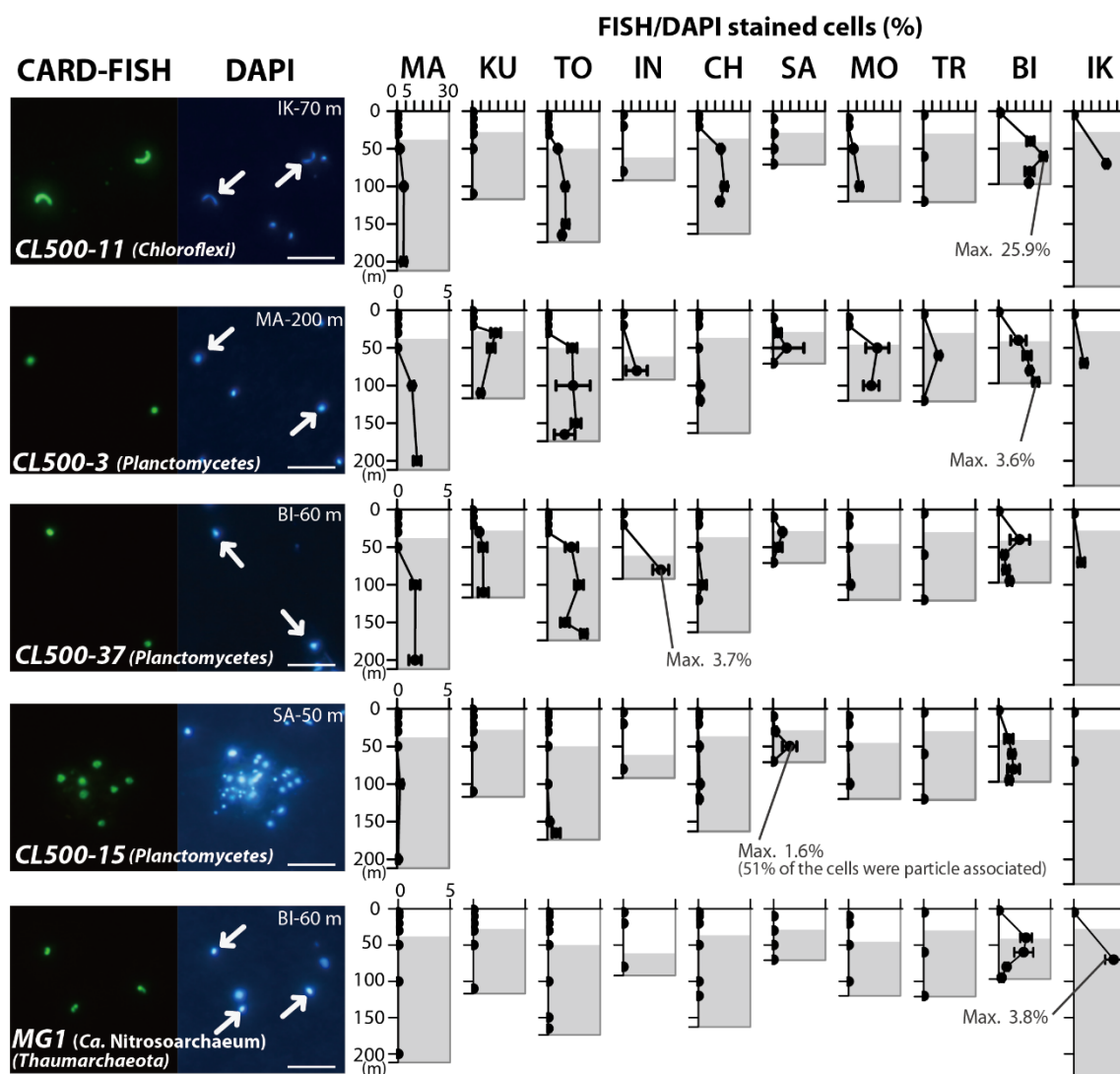


Fig. 6. Catalyzed reporter deposition fluorescence *in situ* hybridization (CARD-FISH) images and the enumeration results. Positive cells and the corresponding DAPI-stained image are shown in each micrograph. Arrows in the DAPI-image indicate cells with FISH-positive signals. Scale bar = 5 μm. The line graphs show percentages of FISH-positive cells to DAPI-positive cells, and the error bars indicating the standard deviation determined by triplicate enumeration of an identical filter. The maximum value recorded for each lineage is designated. Gray background illustrates the hypolimnion (i.e., below the thermocline).

Notable but less represented lineages

Although several lineages originally described for Crater Lake (with the prefix “CL”) (Urbach *et al.*, 2001) were identified as representative hypolimnion specialists (Figs. 4–6), several other members dominant in Crater Lake were not highly represented in our study: CL120–10 of *Verrucomicrobia* (OTU65 in the present study; see Supplementary Data), CL0–1 of *Armatimonadetes* (OTU77), and CL500–9 of *Chloroflexi* (not detected in the present study). It is plausible that further investigations in lakes on other continents or those with depths > 250 m will detect bacterioplankton not found in the present study. Methanotrophs, such as *Methylococcaceae* and *Methylocystaceae*, also accounted for a very minor fraction of all amplicon reads (see Supplementary Data). A more intriguing result is the limited distribution and low relative abundance of LD12 (Figs. 2, 4), which is one of the most dominant and ubiquitous freshwater bacterioplankton (Zwart *et al.*, 1998; Newton *et al.*, 2011; Salcher *et al.*, 2011). Notably, data produced using another sequencing platform (Miseq) from the same DNA samples showed a higher read proportion of LD12 (Fujinaga *et al.*, unpublished data), despite the fact that both analysis used PCR primers that perfectly matched the LD12 16S rRNA sequence. Although the reason behind this discrepancy is unknown, it is possible that the relative abundance of LD12 was underestimated in the present study, which, in turn, might have overestimated other lineages among the reads, as in the discrepancy with the FISH results (Fig. S4). Direct cell enumeration using FISH should be considered an accurate abundance estimate.

Potential ecotype diversification revealed by the oligotyping analysis

The oligotyping analysis revealed the intra-OTU diversification of the whole-layer generalists (Fig. 5). Some of the oligotypes were disproportionally distributed among depths or lakes (Fig. S2), suggesting that their ubiquity was achieved collectively by heterogeneous ecotypes that specialized in a respective niche. Such cryptic ecotype diversification within a ubiquitous freshwater lineage with an almost identical 16S rRNA sequence has previously been reported in *Limnohabitans* (Kasalický *et al.*, 2013; Jezbera *et al.*, 2013) and *Polynucleobacter* (Jezbera *et al.*, 2011; Hahn *et al.*, 2015, 2016), who reported diversification between habitats with different temperatures, pHs, organic and inorganic substrate availability, and geography. Recently, a horizontal oligotype profile in Lake Michigan indicated oligotype diversification within predominant bacterioplankton lineages between estuarine and pelagic sites (Newton and McLellan, 2015). The present study discovered micro-diversification between the epilimnion and hypolimnion, suggesting the presence of hypolimnion-specific ecotype within the common freshwater bacterioplankton. Further comparative studies of individual ecotypes will clarify their adaptation strategies to the respective water layers, which differ considerably in temperature, substrate availability, and grazing pressure.

In contrast to the whole-layer generalists, the low oligotype diversity of the hypolimnion specialists (Fig. 5) was intriguing, as we expected that the ecotypes would be diverse between the hypolimnia of different lakes, which are physically isolated and differ in physicochemical properties

(Table 1). The occurrence of CL500–3 in TR (Fig. 6), which is a reservoir constructed just 10 years prior to the sampling, support the idea that hypolimnion specialists migrate between lakes; thus, diversification is limited. On the other hand, diverse ecotypes are likely present at least on a continental scale, as CL500–11 from European and North American lakes have a conserved single nucleotide difference from the sequences read in the present study (Fig. S7).

Contrary to the lineages described above, acI-C2 and LD19 were separated into epilimnion- and hypolimnion-specific OTUs by the >97% OTU clustering threshold (Fig. 4), and the hypolimnetic OTUs were composed of an almost identical oligotype (Fig. 5). This indicates that the phylogenetic resolution required to resolve intra-OTU diversification within a given sequencing region depends on the lineage. As the present study used the finest phylogenetic resolution (i.e., single-nucleotide level) but analyzed only a 350 bp (V4 + V5 of the 16S rRNA gene) region, further micro-diversification studies should be carried out by increasing the number of sequenced regions.

Conclusion

This study provides the first comprehensive overview of the bacterioplankton community inhabiting the oxygenated hypolimnion in 10 deep freshwater lakes with a variety of environmental characteristics. The community was composed of members occurring in common regardless of water layer or lake (whole-layer generalist) and members that were exclusively distributed in the hypolimnion (hypolimnion specialists). CARD-FISH revealed that predominant hypolimnion specialists, represented by *Chloroflexi*, *Planctomycetes*, and *Thaumarchaeota*, together accounted for 1.5–32.9% of the total bacterioplankton in the hypolimnion of the lakes. Moreover, an oligotyping analysis revealed the potential diversification of ecotypes in the common freshwater lineages (e.g., acI and *Limnohabitans*) among the lakes and water layers. These results reveal the uniqueness, ubiquity, and quantitative importance of bacterioplankton inhabiting the oxygenated hypolimnion. Collectively, the present results provide valuable suggestions for future ecological studies on deep freshwater ecosystems and eco-physiological and phylogeographical perspectives of general microbial ecology.

CONFLICT OF INTEREST

The authors declare no conflict of interest

ACKNOWLEDGMENTS

We are grateful to Y. Goda, T Denboh, Y Ito, M Sugiyama, and Japan Water Agency for their assistance in field sampling. This work is supported by JSPS KAKENHI Grant No. 15J00971 and 15J01065, and by the Environment Research and Technology Development Fund (No. 5-1304 and 5-1607) of the Ministry of the Environment, Japan.

Supplementary Information for this paper is available on the website.

REFERENCES

- Auguet J-C, Triadó-Margarit X, Nomokonova N, Camarero L, Casamayor EO. (2012). Vertical segregation and phylogenetic characterization of ammonia-oxidizing Archaea in a deep oligotrophic lake. *ISME J* **6**: 1786–1797.
- Berdjeb L, Pollet T, Chardon C, Jacquet S. (2013). Spatio-temporal changes in the structure of archaeal communities in two deep freshwater lakes. *FEMS Microbiol Ecol* **86**: 215–230.
- Bižić-Ionescu M, Zeder M, Ionescu D, Orlic S, Fuchs BM, Grossart HP, *et al.* (2015). Comparison of bacterial communities on limnic versus coastal marine particles reveals profound differences in colonization. *Environ Microbiol* **17**: 3500–3514.
- Blainey PC, Mosier AC, Potanina A, Francis CA, Quake SR. (2011). Genome of a Low-Salinity Ammonia-Oxidizing Archaeon Determined by Single-Cell and Metagenomic Analysis. *PLoS One* **6**: e16626.
- Bornemann M, Bussmann I, Tichy L, Deutzmann J, Schink B, Pester M. (2016). Methane release from sediment seeps to the atmosphere is counteracted by highly active Methylococcaceae in the water column of deep oligotrophic Lake Constance. *FEMS Microbiol Ecol* **92**: fiw123.
- Callieri C, Coci M, Eckert EM, Salcher MM, Bertoni R. (2014). Archaea and bacteria in deep lake hypolimnion: In situ dark inorganic carbon uptake. *J Limnol* **73**: 31–38.
- Callieri C, Hernández-Avilés S, Salcher MM, Fontaneto D, Bertoni R. (2016). Distribution patterns and environmental correlates of Thaumarchaeota abundance in six deep subalpine lakes. *Aquat Sci* **78**: 215–225.
- Catalán N, Marcé R, Kothawala DN, Tranvik LJ. (2016). Organic carbon decomposition rates controlled by water retention time across inland waters. *Nat Geosci* **9**: 1–7.
- Chao A, Jost L. (2012). Coverage-based rarefaction and extrapolation: Standardizing samples by completeness rather than size. *Ecology* **93**: 2533–2547.
- Coci M, Odermatt N, Salcher MM, Pernthaler J, Corno G. (2015). Ecology and Distribution of Thaumarchaea in the Deep Hypolimnion of Lake Maggiore. *Archaea* **2015**: 590434.
- Denef VJ, Mueller RS, Chiang E, Liebig JR, Vanderploeg HA. (2016). Chloroflexi CL500-11 Populations That Predominate Deep-Lake Hypolimnion Bacterioplankton Rely on Nitrogen-Rich Dissolved Organic Matter Metabolism and C 1 Compound Oxidation. *Appl Environ Microbiol* **82**: 1423–1432.
- DeSantis TZ, Hugenholtz P, Larsen N, Rojas M, Brodie EL, Keller K, *et al.* (2006). Greengenes, a chimera-checked 16S rRNA gene database and workbench compatible with ARB. *Appl Environ Microbiol* **72**: 5069–5072.
- Edgar RC. (2013). UPARSE: highly accurate OTU sequences from microbial amplicon reads. *Nat Methods* **10**: 996–8.
- Eiler A, Bertilsson S. (2004). Composition of freshwater bacterial communities associated with

- cyanobacterial blooms in four Swedish lakes. *Environ Microbiol* **6**: 1228–1243.
- Erbilgin O, McDonald KL, Kerfeld CA. (2014). Characterization of a planctomycetal organelle: A novel bacterial microcompartment for the aerobic degradation of plant saccharides. *Appl Environ Microbiol* **80**: 2193–2205.
- Eren AM, Maignien L, Sul WJ, Murphy LG, Grim SL, Morrison HG, *et al.* (2013). Oligotyping: differentiating between closely related microbial taxa using 16S rRNA gene data. *Methods Ecol Evol* **4**: 1111–1119.
- Eronen-Rasimus E, Lyra C, Rintala JM, Jürgens K, Ikonen V, Kaartokallio H. (2015). Ice formation and growth shape bacterial community structure in Baltic Sea drift ice. *FEMS Microbiol Ecol* **91**: 1–13.
- Fuchs BM, Glöckner FO, Wulf J, Amann R. (2000). Unlabeled helper oligonucleotide increase the in situ accessibility of 16S rRNA of fluorescently labeled oligonucleotide probes. *Appl Environ Microbiol* **66**: 3603–3607.
- Fujimoto M, Cavaletto J, Liebig JR, McCarthy A, Vanderploeg HA, Denef VJ. (2016). Spatiotemporal distribution of bacterioplankton functional groups along a freshwater estuary to pelagic gradient in Lake Michigan. *J Great Lakes Res* **42**: 1036–1048.
- Garcia SL, Buck M, McMahon KD, Grossart HP, Eiler A, Warnecke F. (2015). Auxotrophy and intrapopulation complementarity in the interactome of a cultivated freshwater model community. *Mol Ecol* **24**: 4449–4459.
- Glöckner FO, Kube M, Bauer M, Teeling H, Lombardot T, Ludwig W, *et al.* (2003). Complete genome sequence of the marine planctomycete *Pirellula* sp. strain 1. *Proc Natl Acad Sci U S A* **100**: 8298–8303.
- Goldberg SJ, Ball GI, Allen BC, Schladow SG, Simpson AJ, Masoom H, *et al.* (2015). Refractory dissolved organic nitrogen accumulation in high-elevation lakes. *Nat Commun* **6**: 6347.
- Grossart HP, Tang KW, Kjørboe T, Ploug H. (2007). Comparison of cell-specific activity between free-living and attached bacteria using isolates and natural assemblages. *FEMS Microbiol Lett* **266**: 194–200.
- Hahn MW, Jezberova J, Koll U, Saueressig-Beck T, Schmidt J. (2016). Complete ecological isolation and cryptic diversity in *Polynucleobacter* bacteria not resolved by 16S rRNA gene sequences. *Isme J* **10**: 1–14.
- Hahn MW, Koll U, Jezberová J, Camacho A. (2015). Global phylogeography of pelagic *Polynucleobacter* bacteria: Restricted geographic distribution of subgroups, isolation by distance and influence of climate. *Environ Microbiol* **17**: 829–840.
- Hansell DA. (2013). Recalcitrant dissolved organic carbon fractions. *Ann Rev Mar Sci* **5**: 421–445.
- Hayakawa K, Kojima R, Wada C, Suzuki T, Sugiyama Y. (2016). Distribution and characteristics of ultraviolet absorption and fluorescence of dissolved organic matter in a large lake (Lake Biwa ,

- Japan). *J Great Lakes Res* **42**: 2–10.
- Jezbera J, Jezberová J, Brandt U, Hahn MW. (2011). Ubiquity of *Polynucleobacter necessarius* subspecies *asymbioticus* results from ecological diversification. *Environ Microbiol* **13**: 922–931.
- Jezbera J, Jezberová J, Kasalický V, Šimek K, Hahn MW. (2013). Patterns of Limnohabitans Microdiversity across a Large Set of Freshwater Habitats as Revealed by Reverse Line Blot Hybridization. *PLoS One* **8**: e58527.
- Jiao N, Zheng Q. (2011). The microbial carbon pump: From genes to ecosystems. *Appl Environ Microbiol* **77**: 7439–7444.
- Kagami M, Gurung TB, Yoshida T, Urabe J. (2006). To sink or to be lysed? Contrasting fate of two large phytoplankton species in Lake Biwa. *Limnol Oceanogr* **51**: 2775–2786.
- Karlov DS, Marie D, Sumbatyan DA, Chuvochina MS, Kulichevskaya IS, Alekhina IA, *et al.* (2016). Microbial communities within the water column of freshwater Lake Radok, East Antarctica: predominant 16S rDNA phylotypes and bacterial cultures. *Polar Biol.* e-pub ahead of print, doi: 10.1007/s00300-016-2008-9.
- Kasalický V, Jezbera J, Hahn MW, Šimek K. (2013). The Diversity of the Limnohabitans Genus, an Important Group of Freshwater Bacterioplankton, by Characterization of 35 Isolated Strains. *PLoS One* **8**: e58209.
- Kellerman AM, Dittmar T, Kothawala DN, Tranvik LJ. (2014). Chemodiversity of dissolved organic matter in lakes driven by climate and hydrology. *Nat Commun* **5**: 1–8.
- Kim C, Nishimura Y, Nagata T. (2006). Role of dissolved organic matter in hypolimnetic mineralization of carbon and nitrogen in a large, monomictic lake. *Limnol Oceanogr* **51**: 70–78.
- Kumar S, Stecher G, Tamura K. (2016). MEGA7: Molecular Evolutionary Genetics Analysis Version 7.0 for Bigger Datasets. *Mol Biol Evol* **33**: 1870–1874.
- Kurilkina MI, Zakharova YR, Galachyants YP, Petrova DP, Bukin YS, Domysheva VM, *et al.* (2016). Bacterial community composition in the water column of the deepest freshwater Lake Baikal as determined by next-generation sequencing. *FEMS Microbiol Ecol* **92**: fiw094.
- Lemarchand C, Jardillier L, Carrias JF, Richardot M, Debroas D, Sime-Ngando T, *et al.* (2006). Community composition and activity of prokaryotes associated to detrital particles in two contrasting lake ecosystems. *FEMS Microbiol Ecol* **57**: 442–451.
- Ludwig W, Strunk O, Westram R, Richter L, Meier H, Yadhukumar A, *et al.* (2004). ARB: A software environment for sequence data. *Nucleic Acids Res* **32**: 1363–1371.
- Maki K, Kim C, Yoshimizu C, Tayasu I, Miyajima T, Nagata T. (2010). Autochthonous origin of semi-labile dissolved organic carbon in a large monomictic lake (Lake Biwa): Carbon stable isotopic evidence. *Limnology* **11**: 143–153.
- Mas A, Jamshidi S, Lagadeuc Y, Eveillard D, Vandenkoornhuysen P. (2016). Beyond the Black Queen Hypothesis. *ISME J* **10**: 2085–2091.

- Masquelier S, Lepe C, Domaizon I, Curie M, Lepère C, Masquelier S, *et al.* (2010). Vertical structure of small eukaryotes in three lakes that differ by their trophic status: a quantitative approach. *ISME J* **4**: 1509–19.
- McCarthy MD, Hedges JJ, Benner R. (2013). Major Bacterial Contribution to Marine Dissolved Organic Nitrogen. *Physics (College Park Md)* **6**: 35.
- Mori K, Sato Y. (2015). Japanese Lakes. Asakura Publishing: Tokyo, Japan.
- Morris JJ, Lenski RE, Zinser ER. (2012). The Black Queen Hypothesis: Evolution of Dependencies through Adaptive Gene Loss. *MBio* **3**: e00036-12-e00036-12.
- Morris RM, Longnecker K, Giovannoni SJ. (2006). Pirellula and OM43 are among the dominant lineages identified in an Oregon coast diatom bloom. *Environ Microbiol* **8**: 1361–1370.
- Mukherjee I, Hodoki Y, Nakano S. (2015). Kinetoplastid flagellates overlooked by universal primers dominate in the oxygenated hypolimnion of Lake Biwa, Japan. *FEMS Microbiol Ecol* **91**: fiv083.
- Mukherjee M, Ray A, Post AF, McKay RM, Bullerjahn GS. (2016). Identification, enumeration and diversity of nitrifying planktonic archaea and bacteria in trophic end members of the Laurentian Great Lakes. *J Great Lakes Res* **42**: 39–49.
- Murase J, Sugimoto A. (2005). Inhibitory effect of light on methane oxidation in the pelagic water column of a mesotrophic lake (Lake Biwa, Japan). *Limnol Oceanogr* **50**: 1339–1343.
- Nagata T, Meon B, Kirchman DL. (2003). Microbial degradation of peptidoglycan in seawater. *Limnol Oceanogr* **48**: 745–754.
- Newton RJ, Jones SE, Eiler A, McMahon KD, Bertilsson S. (2011). A guide to the natural history of freshwater lake bacteria. *Microbiol Mol Biol Rev* **75**: 14–49.
- Newton RJ, McLellan SL. (2015). A unique assemblage of cosmopolitan freshwater bacteria and higher community diversity differentiate an urbanized estuary from oligotrophic Lake Michigan. *Front Microbiol* **6**: 1–13.
- Nguyen RT, Harvey HR. (1997). Protein and amino acid cycling during phytoplankton decomposition in oxic and anoxic waters. *Org Geochem* **27**: 115–128.
- Ntougias S, Polkowska Ż, Nikolaki S, Dionyssopoulou E, Stathopoulou P, Doudoumis V, *et al.* (2016). Bacterial Community Structures in Freshwater Polar Environments of Svalbard. *Microbes Environ*. e-pub ahead of print, doi: 10.1264/jsme2.ME16074.
- Nunoura T, Takaki Y, Kazama H, Hirai M, Ashi J, Imachi H, *et al.* (2009). Microbial Diversity in Deep-sea Methane Seep Sediments Presented by SSU rRNA Gene Tag Sequencing. *Microbes Environ* **27**: 382–390.
- Ogawa H, Amagai Y, Koike I, Kaiser K, Benner R. (2001). Production of refractory dissolved organic matter by bacteria. *Science* **292**: 917–20.
- Okazaki Y, Hodoki Y, Nakano SI. (2013). Seasonal dominance of CL500-11 bacterioplankton (phylum Chloroflexi) in the oxygenated hypolimnion of Lake Biwa, Japan. *FEMS Microbiol Ecol*

83: 82–92.

- Okazaki Y, Nakano SI. (2016). Vertical partitioning of freshwater bacterioplankton community in a deep mesotrophic lake with a fully oxygenated hypolimnion (Lake Biwa, Japan). *Environ Microbiol Rep* **8**: 780–788.
- Oksanen J, Blanchet FG, Friendly M, Kindt R, Legendre P, McGlinn D, *et al.* (2016). *vegan*: Community Ecology Package. version 2.4-0. <https://cran.r-project.org/package=vegan>.
- Orsi WD, Smith JM, Liu S, Liu Z, Sakamoto CM, Wilken S, *et al.* (2016). Diverse, uncultivated bacteria and archaea underlying the cycling of dissolved protein in the ocean. *ISME J* **10**: 2158–2173.
- Parfenova VV, Gladkikh AS, Belykh OI. (2013). Comparative analysis of biodiversity in the planktonic and biofilm bacterial communities in Lake Baikal. *Microbiology* **82**: 91–101.
- Passow U. (2002). Transparent exopolymer particles (TEP) in aquatic environments. *Prog Oceanogr* **55**: 287–333.
- Paver SF, Hayek KR, Gano KA, Fagen JR, Brown CT, Davis-Richardson AG, *et al.* (2013). Interactions between specific phytoplankton and bacteria affect lake bacterial community succession. *Environ Microbiol* **15**: 2489–2504.
- Pernthaler A, Pernthaler J, Amann R. (2002). Fluorescence In Situ Hybridization and Catalyzed Reporter Deposition for the Identification of Marine Bacteria Fluorescence In Situ Hybridization and Catalyzed Reporter Deposition for the Identification of Marine Bacteria. *Appl Environ Microbiol* **68**: 3094–3101.
- Pernthaler J. (2005). Predation on prokaryotes in the water column and its ecological implications. *Nat Rev Microbiol* **3**: 537–546.
- Pizzetti I, Fuchs BM, Gerdt G, Wichels A, Wiltshire KH, Amann R. (2011). Temporal variability of coastal Planctomycetes clades at Kabeltonne station, North Sea. *Appl Environ Microbiol* **77**: 5009–5017.
- Pollet T, Tadonl  k   RD, Humbert JF. (2011). Spatiotemporal changes in the structure and composition of a less-abundant bacterial phylum (Planctomycetes) in two perialpine lakes. *Appl Environ Microbiol* **77**: 4811–4821.
- Porter KG, Feig YS. (1980). The use of DAPI for identifying and counting aquatic microflora1. *Limnol Oceanogr* **25**: 943–948.
- Pruesse E, Peplies J, Gl  ckner FO. (2012). SINA: Accurate high-throughput multiple sequence alignment of ribosomal RNA genes. *Bioinformatics* **28**: 1823–1829.
- Quast C, Pruesse E, Yilmaz P, Gerken J, Schweer T, Yarza P, *et al.* (2013). The SILVA ribosomal RNA gene database project: Improved data processing and web-based tools. *Nucleic Acids Res* **41**: D590–6.
- Rozmarynowycz MJ. (2014). Spatio-Temporal Distribution Of Microbial Communities In The

- Laurentian Great Lakes. (Doctoral Dissertation) Bowling Green State University.
- Salazar G, Cornejo-Castillo FM, Borrull E, Díez-Vives C, Lara E, Vaqué D, *et al.* (2015). Particle-association lifestyle is a phylogenetically conserved trait in bathypelagic prokaryotes. *Mol Ecol* **24**: 5692–5706.
- Salcher MM, Pernthaler J, Posch T. (2011). Seasonal bloom dynamics and ecophysiology of the freshwater sister clade of SAR11 bacteria ‘that rule the waves’ (LD12). *ISME J* **5**: 1242–1252.
- Šimek K, Kasalický V, Zapomělová E, Horňák K. (2011). Alga-derived substrates select for distinct betaproteobacterial lineages and contribute to niche separation in Limnhabitans strains. *Appl Environ Microbiol* **77**: 7307–7315.
- Small GE, Bullerjahn GS, Sterner RW, Beall BFN, Brovold S, Finlay JC, *et al.* (2013). Rates and controls of nitrification in a large oligotrophic lake. *Limnol Oceanogr* **58**: 276–286.
- Stamatakis A. (2014). RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* **30**: 1312–1313.
- Takasu H, Ushio M, LeClair JE, Nakano SI. (2015). High contribution of Synechococcus to phytoplankton biomass in the aphotic hypolimnion in a deep freshwater lake (Lake Biwa, Japan). *Aquat Microb Ecol* **75**: 69–79.
- Tanoue E, Nishiyama S, Kamo M, Tsugita A. (1995). Bacterial membranes: possible source of a major dissolved protein in seawater. *Geochim Cosmochim Acta* **59**: 2643–2648.
- Thottathil SD, Hayakawa K, Hodoki Y, Yoshimizu C, Kobayashi Y, Nakano S. (2013). Biogeochemical control on fluorescent dissolved organic matter dynamics in a large freshwater lake (Lake Biwa, Japan). *Limnol Oceanogr* **58**: 2262–2278.
- Urbach E, Vergin KL, Larson GL, Giovannoni SJ. (2007). Bacterioplankton communities of Crater Lake, OR: Dynamic changes with euphotic zone food web structure and stable deep water populations. *Hydrobiologia* **574**: 161–177.
- Urbach E, Vergin KL, Young L, Morse A, Larson GL, Giovannoni SJ. (2001). Unusual bacterioplankton community structure in ultra-oligotrophic Crater Lake. *Limnol Oceanogr* **46**: 557–572.
- Visser EW, Blaga CI, Bodelier PLE, Muyzer G, Schleper C, Sinninghe Damsté JS, *et al.* (2013). Seasonal and vertical distribution of putative ammonia-oxidizing thaumarchaeotal communities in an oligotrophic lake. *FEMS Microbiol Ecol* **83**: 515–526.
- Wallner G, Amann R, Beisker W. (1993). Optimizing fluorescent in situ hybridization with rRNA-targeted oligonucleotide probes for flow cytometric identification of microorganisms. *Cytometry* **14**: 136–143.
- Wetzel. (2001). Limnology: lake and river ecosystems. Academic Press: San Diego, CA.
- Woebken D, Teeling H, Wecker P, Dumitriu A, Kostadinov I, DeLong EF, *et al.* (2007). Fosmids of novel marine Planctomycetes from the Namibian and Oregon coast upwelling systems and their

- cross-comparison with planctomycete genomes. *ISME J* **1**: 419–435.
- Yamada E, Hirota T, Hatori N, Kitao Y, Fuse Y, Aoki S, *et al.* (2012). Characterization of protein-like fluorophores released from lake phytoplankton on the basis of fractionation and electrophoresis. *Anal Sci* **28**: 595–600.
- Yamashita Y, Tanoue E. (2008). Production of bio-refractory fluorescent dissolved organic matter in the ocean interior. *Nat Geosci* **1**: 579–582.
- Zhang H, Wu H, Wang G, Xiang W, Yan W. (2013). Prokaryote diversity in the surface sediment of northern South China Sea. *Wei Sheng Wu Xue Bao* **53**: 915–26.
- Zwart G, Crump BC, Kamst-van Agterveld MP, Hagen F, Han SK. (2002). Typical freshwater bacteria: An analysis of available 16S rRNA gene sequences from plankton of lakes and rivers. *Aquat Microb Ecol* **28**: 141–155.
- Zwart G, Hiorns WD, Methé BA, van Agterveld MP, Huismans R, Nold SC, *et al.* (1998). Nearly identical 16S rRNA sequences recovered from lakes in North America and Europe indicate the existence of clades of globally distributed freshwater bacteria. *Syst Appl Microbiol* **21**: 546–56.