

*Systematic Biology* (2024), **0**, 0, pp. 1–83  
doi:10.1093/????/output

# Performance and Robustness of Parameter Estimation from Phylogenetic Trees Using Neural Networks

TIANJIAN QIN<sup>1</sup>, KOEN J. VAN BENTHEM<sup>1</sup>, LUIS VALENTE<sup>1,2,\*</sup>, AND RAMPAL S. ETIENNE<sup>1,\*</sup>

<sup>1</sup> GRONINGEN INSTITUTE FOR EVOLUTIONARY LIFE SCIENCES, UNIVERSITY OF GRONINGEN,  
NIJENBORGH 7, GRONINGEN, 9747 AG, THE NETHERLANDS

<sup>2</sup> NATURALIS BIODIVERSITY CENTER, DARWINWEG 2, LEIDEN, 2333 CR, THE NETHERLANDS

CORRESPONDING AUTHOR: TIANJIAN QIN, E-MAIL: T.QIN@RUG.NL

\*JOINT SENIOR AUTHORS

## ABSTRACT

1 Species diversification is characterized by speciation and extinction, the rates of which can,  
2 under some assumptions, be estimated from time-calibrated phylogenies. However,  
3 maximum likelihood estimation methods (MLE) for inferring rates are limited to simpler  
4 models and can show bias, particularly in small phylogenies. Likelihood-free methods to  
5 estimate parameters of diversification models using deep learning have started to emerge,  
6 but how robust neural network methods are at handling the intricate nature of  
7 phylogenetic data remains an open question. Here we present a new ensemble neural  
8 network approach to estimate diversification parameters from phylogenetic trees that  
9 leverages different classes of neural networks (dense neural network, graph neural network,  
10 and long short-term memory recurrent network) and simultaneously learns from graph  
11 representations of phylogenies, their branching times and their summary statistics. Our  
12 best-performing ensemble neural network (which corrects graph neural network result  
13 using a recurrent neural network) can compute estimates faster than MLE and is less  
14 affected by tree size. Our analysis suggests that the primary limitation to accurate  
15 parameter estimation is the amount of information contained within a phylogeny, as

16 indicated by its size and the strength of effects shaping it. In cases where MLE is  
17 unavailable, our neural network method provides a promising alternative for estimating  
18 phylogenetic tree parameters. If there are detectable phylogenetic signals present, our  
19 approach delivers results that are comparable to MLE but without inherent biases.

20 *Key words:* graph neural network, recurrent neural network, machine learning, regression

21

## INTRODUCTION

22 Identifying the underlying mechanisms shaping biodiversity is an important goal in  
23 the fields of evolutionary biology and ecology. Species diversification processes can often be  
24 characterized by speciation and extinction rates, which can be estimated from  
25 time-calibrated phylogenies (Nee et al., 1997) as long as the assumed model structure of  
26 diversification resembles the true underlying data generation process (Louca and Pennell,  
27 2021). Time-calibrated phylogenies contain branching times and topological relationships  
28 between species and offer a complementary source of information to the often incomplete  
29 fossil record (Kidwell and Flessa, 1996). The increasing availability of reconstructed  
30 phylogenies has empowered many studies seeking explanations for the underlying diversity  
31 patterns using modelling approaches (Etienne et al., 2016; Morlon, 2014; Wagner, 2000).

32 One type of models – birth-death models – are often used to estimate speciation,  
33 extinction and diversification rates from reconstructed phylogenetic trees (Hey, 1992; Nee,  
34 2001; Nee et al., 1994, 1997).

35 Likelihood-based approaches, such as maximum likelihood estimation (MLE) and  
36 Bayesian inference, can be used to infer not only speciation and extinction rates, but also  
37 possibly existing evolutionary and ecological signals, such as diversity-dependence or  
38 trait-dependence of rates from branching times and other information sources (Alexander  
39 et al., 2016; Etienne et al., 2014; Foote, 1997; Valente et al., 2015). However, MLE  
40 approaches are only mathematically tractable for simple diversification models (Janzen

et al., 2015; Lambert et al., 2023). In addition, MLE tends to be biased when estimating diversification parameters from phylogenetic trees. The degree of bias depends on the size of the phylogenetic tree (the number of tips): as tree size increases, the estimates become asymptotically more unbiased (Etienne et al., 2016). MLE may also perform worse on complex models with a high number of parameters (Ward, 2008).

An alternative to these likelihood-based approaches for parameter estimation is Approximate Bayesian Computation (ABC), which approximates the posterior distribution of parameters without requiring explicit calculation of a likelihood function. ABC is often seen as a good substitute to MLE when a likelihood function of a model is not available, as long as simulations of the model are fast and tractable (Beaumont, 2010; Beaumont et al., 2002; Janzen et al., 2015). However, studies using ABC for parameter estimation in phylogenetics remain scarce (Bokma, 2010; Kutsukake and Innan, 2013; Rabosky, 2009; Xie et al., 2023). This is partly due to the fact that it is often difficult to identify adequate summary statistics in ABC, which makes the application and development of this potentially powerful approach challenging.

A promising class of tools that may help overcome the limitations of likelihood-based methods and ABC are machine learning approaches, such as neural networks. Neural networks are comprised of layers of nodes, or "neurons", which process input data and learn to recognize patterns between input and output data from training data (Charu C, 2018). Classic feed-forward neural networks have achieved good results in tasks such as image recognition and natural language processing (Zhu et al., 2018).

Another class of neural networks, graph neural networks, are designed specifically for graph-structured data, such as social networks, molecular structures, and ecological interaction networks. They can capture the dependencies and relationships inherent in data types that can be naturally represented as graphs (Kipf and Welling, 2016) and have shown strong performance in various tasks involving graph representation learning (Li et al., 2020; Rampáek et al., 2022; Ying et al., 2018). Phylogenetic trees can also be viewed

68 as graphs, suggesting that graph neural networks have potential applicability in  
69 phylogenetics. Recurrent neural networks, another type of neural network, are designed to  
70 handle sequential data, such as time series, by maintaining a memory of previous inputs  
71 (Sak et al., 2014; Salehinejad et al., 2017). Recurrent neural networks can process inputs of  
72 varying lengths and capture time-dependent features, making them particularly well-suited  
73 for tasks where the order of data points is crucial, such as learning parameters from  
74 branching times when viewed as time-series data.

75 Owing to the rapid development of both hardware capability and deep learning  
76 algorithms, applications of neural networks in phylogenetic analyses have started to emerge  
77 (e.g. Moi and Dessimoz (2022), Reiman et al. (2020), Voznica et al. (2022), Lambert et al.  
78 (2023), and Lajaaiti et al. (2023)). For instance, phylogenetic deep learning approaches  
79 have been shown to provide reliable estimates of parameters in epidemiological,  
80 birth-death, and trait-dependent speciation models (Lajaaiti et al., 2023; Lambert et al.,  
81 2023; Voznica et al., 2022). Despite their potential, employing neural networks for  
82 estimating parameters based on the whole phylogenetic tree, especially those associated  
83 with diversification, poses significant challenges and requires further systematic research  
84 regarding their performance, accuracy and robustness. Specifically, feed-forward linear  
85 neural networks usually require a large amount of data to be able to generalize well on the  
86 patterns within the data (Zhu et al., 2018); producing graph representations for  
87 graph-level learning can be challenging given the need to aggregate information across  
88 diverse graph sizes and topologies (Ying et al., 2018); the capability of the recurrent neural  
89 networks to predict parameters from whole sequences is often challenging (Sak et al.,  
90 2014). Hence, how robust neural network methods are at handling the intricate nature of  
91 phylogenetic data remains an open question.

92 In this study, we explore the capabilities of neural networks in research on species  
93 diversification using phylogenies. We first develop various neural network architectures and  
94 protocols for transforming phylogenetic trees and branching times into formats compatible

95 with popular neural network frameworks. We then investigate predictive performance on  
96 simulated data for ensemble learning strategies, which combine different neural network  
97 classes to maximize data utilization and enhance performance. We also assess the  
98 determinants of estimation accuracy and robustness for both neural network and MLE  
99 methods under various diversification scenarios. Finally, we implement our trained neural  
100 networks on empirical phylogenetic datasets and compare their estimations to those of  
101 MLE.

102 Our analyses encompass three different diversification scenarios for which  
103 likelihood-based inference approaches already exist: a constant-rate birth-death (BD)  
104 scenario, with constant speciation and extinction rates over time (Stadler, 2011); a  
105 diversity-dependent diversification (DDD) scenario, where the number of species in a clade  
106 negatively affects the speciation rate (Etienne et al., 2012); and a protracted birth-death  
107 (PBD) scenario, where speciation takes time and does not always proceed to completion  
108 (Rosindell et al., 2010). Applying our new methodology to phylogenetic trees simulated  
109 under a broad range of the parameter space, our findings indicate that neural network  
110 approaches are as effective, if not more so, than MLE in recovering parameters from  
111 phylogenetic data simulated under these stochastic processes. Trained neural networks can  
112 be conveniently applied to empirical trees for parameter estimation. To facilitate this, we  
113 present a new R package, "EvoNN," capable of performing such analyses based on  
114 phylogenetic trees (empirical or simulated) supplied by the user (Qin, 2024).

115 **MATERIALS AND METHODS**

116 *Software Environment and Computational Budget*

117 We used a hybrid programming environment with PyTorch 1.12.1 (Imambi et al.,  
118 2021), PyTorch Geometric 2.3.1 (Fey and Lenssen, 2019), Python 3.7.1 (Python, 2021),  
119 CUDA 12.2.2 (Luebke, 2008) and R 4.2.1 (R Core Team, 2013). The procedures of  
120 simulation, data transformation, and maximum likelihood estimation were handled

121 through parallel CPU computations on the Hábrók high-performance computing cluster of  
122 the University of Groningen. The total computational budget for these processes was  
123 approximately 3000 hours (used CPU time). Our neural networks were trained, optimized  
124 and evaluated on the NVIDIA A100 and V100 tensor core GPUs of the Hábrók cluster.  
125 The estimated computational budget was 1500 hours (used GPU time, excluding CPU  
126 time for dataset loading and saving). We implemented a user-friendly tool to estimate  
127 parameters from phylogenetic trees using the neural network approach developed in this  
128 study in the new R package "evoNN".

129 *Simulation Approaches*

130 To train the neural networks, we simulated phylogenetic trees using different  
131 functions from different R packages. For each simulated dataset we kept trees with only  
132 extant lineages, mimicking reconstructed phylogenies. The settings for the parameters used  
133 to simulate the trees were selected to limit the maximum total number of nodes (including  
134 root, internal and tip nodes, here and after, we always refer to the total number of nodes)  
135 for the trees in each dataset. After simulation, we further filtered out all trees containing  
136 more than 3000 nodes to avoid the creation of excessively large matrices that could deplete  
137 the available memory space allocated to the GPUs during the GNN training process. Such  
138 trees are uncommon under the settings we used – typically fewer than 5 trees with more  
139 than 3000 nodes (~1500 tips) are present within each set of phylogenies we acquired from  
140 simulation. We also filtered out all trees containing less than 5 nodes (3 tips) to ensure  
141 successful data transformation and summary statistic computation. Small trees inherently  
142 carry limited informational content. The exclusion of these trees is unlikely to impact  
143 performance of the neural networks on the remaining trees (typically fewer than 100 trees  
144 with less than 5 nodes were present for each parameter setting).

145 To consider different diversification processes, we simulated 100,000 random  
146 birth-death trees (BD phylogenies), 100,000 diversity-dependent trees (DDD phylogenies)

and 100,000 protracted birth-death trees (PBD phylogenies). The amount of simulated data is bounded by the resource and time limits of the computing cluster. All trees have an identical crown age of 10 time units ( $t = 10$ ) to reduce the dimension of data complexity. This age was chosen arbitrarily, as we can always rescale the trees in time. For simulating BD trees, we used the "rlineage" function from R package "ape" (Paradis and Schliep, 2019) to generate complete trees and then pruned all the extinct lineages; for DDD trees, we used the "dd\_sim" function from R package "DDD" (Etienne et al., 2012); for PBD trees we used the "pbd\_sim" function from our R package "eveGNN" (a codebase of phylogeny simulation, data transformation, neural network training and MLE computation for our study), which is similar to the function with the same name in the original R package "PBD" (Etienne and Rosindell, 2012), but only outputs necessary data for our study.

In our simulation approach we randomly sampled the (log) parameters required for each scenario (BD, DDD and PBD) from uniform distributions. The upper bound for the extinction rates were proportionally dependent on the drawn speciation rate to avoid cases where extinction rates could be larger than speciation rates, because in such cases the whole tree likely goes extinct. Furthermore, to prevent a huge number of evolutionary events that would deplete available computational time and memory, we also imposed an overall cap of 1.5 on the extinction rates. See Table 1 for the detailed parameter distribution settings used in the simulations.

### 166 *Data Preparation*

167 We employed three different basic neural network architectures: a dense neural  
168 network (DNN), a graph neural network (GNN), and a long short-term memory (LSTM)  
169 recurrent network, as illustrated in Figure 1 (see Appendix C for a detailed description).  
170 Each of these architectures was refined through validation and required different input  
171 data. For the DNN, the input data consisted of a total of 54 summary statistics  
172 (Appendix N) for each simulated tree. In the GNN, the full phylogeny was interpreted as a

Table 1. Parameter settings for the simulated tree datasets. The type column specifies which function is used to generate the trees. The columns specify the crown age (age), the number of trees in the data set ( $N$ ), the lower ( $a$ ) and the upper ( $b$ ) bounds of the parameters for the tree simulations, all the parameters being sampled from  $U(a, b)$ , except for  $\lambda_1$  of the protracted birth-death scenario.  $\lambda_1$  is computed as  $\lambda_1 = 10^i$  where  $i$  is sampled from  $U(-3, 1)$ .  $U$  denotes uniform distribution. Sub-table A shows the parameter distributions of the constant-rate birth-death model and the diversity-dependent-diversification model,  $\lambda$ : intrinsic speciation rate/birth rate;  $\mu$ : intrinsic extinction rate/death rate;  $K$ : carrying capacity. Sub-table B shows the parameter distributions of the protracted birth-death model,  $\lambda_1$ : speciation-initiation rate of good species;  $\lambda_2$ : speciation-completion rate;  $\lambda_3$ : speciation-initiation rate of incipient species;  $\mu_1$ : extinction rate of good species;  $\mu_2$ : extinction rate of incipient species. \*In diversity-dependent-diversification simulations, the maximum extinction rate is capped at 1.5 if  $0.9\lambda > 1.5$ .

**A:** Parameter settings for BD and DDD trees

Type	Age	N	$\lambda_0$		$\mu_0$		K	
			a	b	a	b	a	b
BD	10	100,000	0.1	0.8	0.0	$0.9\lambda_0$	-	-
DDD	10	100,000	0.1	4.0	0.0	$0.9\lambda_0^*$	10	1000

**B:** Parameter settings for PBD trees

Type	Age	N	$\lambda_1$		$\log_{10}(\lambda_2)$		$\lambda_3$		$\mu_1$		$\mu_2$	
			a	b	a	b	a	b	a	b	a	b
PBD	10	100,000	0.1	1.0	-3	1	0.1	1.0	0.0	$0.8\lambda_1$	0.0	$0.8\lambda_3$

graph and could in that form be used as input data (as illustrated in Figure 2). In the LSTM, we treated branching times of the phylogenies as sequential or time-series data (Sak et al., 2014). Given its recurrent architecture, LSTM is adept at sequence prediction tasks, making it particularly suitable for estimating tree parameters from entire sequences of branching times.

Therefore, our data compound comprises three major components: the phylogenetic trees, their corresponding summary statistics, and their branching times, to maximize the use of available data. The functions needed for the data transformations are either available in PyTorch or implemented in our package eveGNN and described in more detail in Appendix A.

183 *Ensemble Learning Strategies*

184 To leverage all available data and improve prediction accuracy, we combined GNN,  
185 DNN, and LSTM using bagging, stacking, and boosting, which are typical ensemble  
186 learning strategies (Graczyk et al., 2010). With bagging, we trained GNN, DNN and  
187 LSTM independently on the same dataset, translated their original outputs to parameter  
188 predictions (we will use "readout" hereafter to refer to this translation) and then aggregated  
189 the predictions. We used four aggregation methods: mean, median, max and min.

190 With stacking, we use GNN, DNN and LSTM in the same architecture but without  
191 their own readout layers. Instead, we combined the features learned from DNN, LSTM and  
192 GNN and fed them to a meta-learner comprising linear neural network layers that learns  
193 the best readout parameter predictions from these combined features. GNN, DNN, LSTM  
194 and the meta-learner were trained simultaneously.

195 With boosting, the neural networks were trained sequentially. Boosting strategies  
196 offered various pathways for enhancing model performance. We started with a GNN to  
197 make initial predictions and explored the effectiveness of both DNN and LSTM for  
198 correcting residuals, either individually or in sequence. We used "Boost SS" to refer to  
199 correcting GNN's residuals by DNN (from summary statistics) "Boost BT" to refer to  
200 correcting GNN's residuals by LSTM (from branching times); "Boost SS+BT" to refer to  
201 correcting GNN's residuals by DNN and then correcting DNN's residuals by  
202 LSTM; "Boost BT" to refer to correcting GNN's residuals by LSTM (from branching  
203 times); "Boost BT+SS" to refer to correcting GNN's residuals by LSTM and then  
204 correcting LSTM's residuals by DNN.

205 See Figure 3 for a simplified illustration of the ensemble learning strategies.

206 *Training Neural Networks*

207 Prior to training, each dataset of 100,000 trees was randomly shuffled and  
208 subsequently divided into two segments. The first segment, consisting of 90% of the

209 dataset, was allocated for training purposes, while the remaining 10% was used as the  
210 validation dataset for monitoring and fine-tuning the neural network performance. The  
211 training session is carried out by epochs, each consisting of three major steps: first,  
212 performing forward pass on the training dataset; second, assessing the prediction accuracy;  
213 and lastly, performing back-propagation (adjusting the weights of the neuron connections  
214 to improve the neural network performance). Back-propagation, requires quantifying the  
215 error between the neural networks predictions and the actual ground truth values. We  
216 quantified the 'total loss' as the sum of the residual error and other terms for facilitating  
217 neural network training. We represented total loss using a loss function which sums up all  
218 the loss terms (see Appendix B for more detail).

219 We use the AdamW (Adaptive Moment Estimation with decoupled weight decay)  
220 optimizer (Loshchilov and Hutter, 2017) to iteratively update the neural networks'  
221 parameters to minimize the loss function. We used default AdamW argument settings.  
222 During training, we adopted mini-batches of size 64 (data of 64 simulated trees per  
223 mini-batch) to reduce GPU memory usage. The total number of epochs was manually  
224 optimized to avoid underfitting and overfitting. This was done by comparing the loss  
225 metrics for the training dataset to those of the validation datasets at every epoch.  
226 Overfitting is indicated by a training loss that continues to decrease while the validation  
227 loss starts to increase, whereas underfitting is suggested by both training and validation  
228 losses being high and decreasing at a similar rate. Analyzing these loss trends over time  
229 can help to optimize hyper-parameters ("settings" that might alter neural network behavior  
230 or impact performance).

231 Under the BD scenario, the neural networks were trained to predict two  
232 parameters: birth rate ( $\lambda$ ) and death rate ( $\mu$ ). Under the DDD scenario, the neural  
233 networks were trained to predict three parameters: speciation rate ( $\lambda$ ), extinction rate ( $\mu$ )  
234 and carrying capacity ( $K$ ). Under the PBD scenarios, the neural networks were trained to  
235 predict five parameters: speciation rate of the good species ( $\lambda_1$ ), speciation completion rate

<sup>236</sup>  $(\lambda_2)$ , speciation rate of the incipient species  $(\lambda_3)$ , extinction rate of the good species  $(\mu_1)$   
<sup>237</sup> and extinction rate of the incipient species  $(\mu_2)$ .

<sup>238</sup> *Baseline Benchmark*

<sup>239</sup> Maximum likelihood estimation (MLE) approaches have been developed for the  
<sup>240</sup> BD, DDD and PBD scenarios (Etienne et al., 2012, 2014; Etienne and Rosindell, 2012).  
<sup>241</sup> Per scenario, we simulated additional testing datasets each comprising 10,000 phylogenies  
<sup>242</sup> using the same parameter spaces as the training datasets. For these testing datasets, we  
<sup>243</sup> adopted the MLE approaches to estimate the parameters of each phylogeny from their  
<sup>244</sup> branching times under different scenarios. For the BD trees, we estimated their birth and  
<sup>245</sup> death rates. For the DDD trees, we estimated their speciation rate, extinction rate and  
<sup>246</sup> carrying capacity. There is a limitation in the MLE approach for PBD trees because to  
<sup>247</sup> allow for a computation of the likelihood, the speciation initiation rates of good species  
<sup>248</sup> and incipient species need to be equal (Etienne et al., 2014). We therefore only estimated  
<sup>249</sup> four initial parameters: speciation initiation rate (for both good and incipient species,  
<sup>250</sup> assuming they are the same), speciation-completion rate, extinction rate of good species  
<sup>251</sup> and extinction rate of incipient species, although in our simulation we have five  
<sup>252</sup> independently sampled parameters. The MLE results are used as a baseline benchmark to  
<sup>253</sup> evaluate the performance of the neural networks on tree parameter estimation.

<sup>254</sup> We estimated two types of benchmarks using the MLE approaches: one with ground  
<sup>255</sup> truth parameter values set as the starting point of the MLE searching process, the other  
<sup>256</sup> with a starting point randomly sampled from the parameter space of the simulation. We  
<sup>257</sup> consider the first type of benchmarks as a best-case MLE performance (as in real  
<sup>258</sup> applications ground truth parameters are not known) and the other type as a typical-case  
<sup>259</sup> MLE performance, which mimics the pragmatic approach if true parameter values are not  
<sup>260</sup> known. Note that in practice it is possible to achieve better performance than the  
<sup>261</sup> typical-case, e.g. by optimizing from several starting points.

262 We also explored the effectiveness of different optimization approaches for MLE on  
263 the DDD phylogenies. We used the "Simplex" (Lagarias et al., 1998) optimizer to compute  
264 the baseline benchmarks for all the analyses. See Appendix E for reasons and for a detailed  
265 comparison between the optimizers.

266

### *Performance Analysis*

267 From the same testing datasets used for baseline benchmark computation, we  
268 analysed the patterns of residuals (differences between ground truth and predicted values,  
269 which can be viewed as the goodness of fit) by visually examining their relation to true  
270 values and the total node counts of the phylogenetic trees, which include root, internal and  
271 tip nodes. Considering the complex nature of residual patterns, which may vary according  
272 to specific characteristics of the simulation processes (for instance, carrying capacity effects  
273 in DDD and protracted speciation in PBD), as well as the performance and robustness of  
274 the estimation methods, we calculated error metrics locally for three different phylogeny  
275 size ranges, as a global metric could be misleading.

276 The main case study we decided to focus on is based on simulated trees under the  
277 DDD scenario, because it involves more evolutionary mechanisms than the simple  
278 birth-death scenario while containing fewer parameters than the protracted birth-death  
279 scenario. This simplifies our analyses on the neural network performance while maintaining  
280 enough complexity to challenge the capability of our proposed methods. From this case  
281 study we identified and selected the most effective MLE optimization algorithm, neural  
282 network architecture and ensemble strategy, which we then applied to BD and PBD  
283 scenarios. We therefore only analysed the best-performing neural network methods against  
284 the typical and best MLE cases on BD and PBD. Additionally, for the PBD scenario, we  
285 computed a composite parameter called the mean duration of speciation from the  
286 speciation completion rate, the speciation rate of incipient species and the extinction rate  
287 of incipient species (Etienne et al., 2014), because MLE can arguably better estimate the

288 mean duration of speciation than the original parameters.

289 *Robustness Analysis*

290 We assessed the robustness of the estimation results of both neural networks and  
291 MLE by measuring the consistency with which these approaches produce similar estimates  
292 for phylogenies generated under identical parameter settings. In the previous simulations,  
293 each parameter combination was sampled and used only once, whereas in the robustness  
294 analysis we repeatedly use identical parameter settings to generate sets of phylogenies  
295 (bootstrapping) under the DDD scenario. Even when the same parameters are used, the  
296 resulting phylogenies can vary substantially in size, topology and structure due to  
297 stochasticity. Such an evaluation helps assess the neural networks' ability to abstract the  
298 underlying parameter influences from the phylogenetic data, regardless of heterogeneity.  
299 For each parameter combination, 1000 trees were generated randomly. We used a total of  
300 80 sets of parameter combinations, thus 80,000 phylogenies in total. Specifically, we used  
301 all combinations of speciation rates  $\lambda = 1.0, 1.5, 2.0, 2.5, 3.0$ , extinction rates  
302  $\mu = 0.2, 0.4, 0.6, 0.8$  and carrying capacities  $K = 200, 400, 600, 800$ .

303 MLE is computationally more expensive than predicting from already trained  
304 neural networks, and computational time rapidly increases with the size of the phylogenies.  
305 We thus performed MLE on only 2000 simulated phylogenies. To ensure fair visual and  
306 numerical comparisons when plotting the results of these analyses, extreme MLE estimates  
307 were not shown in the figures (they exceeded the fixed range of the y-axis) and excluded  
308 from the computation of the mean absolute errors of the MLE estimates. Neural network  
309 results were randomly sub-sampled to match the MLE data count, maintaining equivalent  
310 visual density and facilitating a more accurate performance comparison between  
311 approaches. For the neural networks, the mean absolute errors were computed on the  
312 complete dataset without sub-sampling and exclusion. In the figure, on average (we  
313 simulated the testing datasets many times throughout the study), out of 2000 samples,

314 5-150 samples per MLE figure panel, and 1-3 per neural network figure panel fell beyond  
315 the axis range. For the most underperforming method (Boost BT+SS) 600-1500 samples  
316 fell beyond the axis range.

317 We did not analyze the robustness of BD and PBD scenarios, because BD is a  
318 special case of DDD (if we set carrying capacity to an infinite value) and PBD related  
319 parameters can hardly be estimated accurately using MLE methods (Etienne et al., 2014).

320 The complete code base for this study, including simulations, data processing,  
321 neural network training, evaluation, and both data analysis and visualization tools, is  
322 available in the GitHub repository eveGNN (Qin, 2023).

323 *Empirical Tree Estimation*

324 We deployed pre-trained neural networks to estimate phylogenetic parameters from  
325 a dataset of 199 empirical phylogenetic trees curated by Condamine et al. (2019), with a  
326 tip count ranging from 20 to 1500. To align with the training conditions of our neural  
327 networks, which were trained on simulated phylogenies spanning exactly 10 time units  
328 (Myr), we rescaled the crown ages of all empirical trees to this duration. The parameter  
329 estimations we present are therefore rescaled. All the selected empirical trees are  
330 reconstructed phylogenies and fully bifurcated (each root or internal node has exactly two  
331 descendants). If an empirical tree fails an ultrametric (all tip-ends are aligned at the  
332 present) test due to branch length precision issue, we forced all its tips to end exactly at  
333 the present by extending the shorter tips to align with the longest one. See Appendix M  
334 for meta information of the empirical trees.

335 We used two distinct neural networks, each pre-trained on simulated trees from one  
336 of two evolutionary scenarios (BD or DDD) to estimate parameters from the empirical  
337 trees. For the BD scenario, we estimated the parameters  $\lambda$  (speciation rate) and  $\mu$   
338 (extinction rate); for the DDD scenario, we estimated  $\lambda$ ,  $\mu$ , and  $K$  (carrying capacity). We  
339 did not estimate parameters for the PBD scenarios because neither neural networks, nor

340 MLE approaches could recover the parameters accurately from the simulated phylogenies.  
341 In addition to our neural network estimates, we used MLE methods for parameter  
342 estimation to provide a comparative assessment of the results. The MLE methods were set  
343 to use default starting points of likelihood optimization, as we do not know the true  
344 parameters of the empirical phylogenies.

345 We used the same bootstrapping method described before to quantify the  
346 uncertainties of both MLE and neural network estimates from empirical data. The process  
347 involves three main steps: first, estimating parameters from empirical phylogenies using  
348 MLE and pre-trained neural networks; second, simulating a set of phylogenies under a  
349 specified diversification scenario (such as BD, DDD, or PBD) using the MLE and neural  
350 network estimates; and third, re-estimating parameters from the simulated phylogenies  
351 using MLE and neural networks. The estimates derived from the bootstrapped phylogenies  
352 form a distribution.

353 We applied this uncertainty computation to a selected set of empirical phylogenies  
354 from the Condamine dataset (Condamine et al., 2019) under the DDD scenario (see  
355 Appendix F for details). The criteria for selection were phylogenies with more than 300  
356 and less than 1000 nodes, and maximum likelihood estimates (MLE) of  $K$  (carrying  
357 capacity) being less than 1000. The distributions of MLE and neural network estimates  
358 from the bootstrapped phylogenies was compared to the original MLE and neural network  
359 estimates from empirical phylogenies. For each set of parameters estimated from empirical  
360 phylogenies, we bootstrapped 1000 simulated phylogenies.

361 Our R package "EvoNN" (Qin, 2024) provides functions to perform the uncertainty  
362 (bootstrap) analyses.

363

## RESULTS

364

### *Performance Analysis*

365

We evaluated the performances of various neural networks, both individually and in combination through ensemble strategies, in predicting parameters from simulated DDD phylogenies. These predictions were benchmarked against best-case and typical-case MLE results using the Simplex optimizer.

369

Among all the methods, we consider boosting GNN with LSTM as the most robust method based on the goodness of fit (see Figure 4, Figure 5, Figure 6 and Figure 7), the mean absolute errors (see Appendix G, Figure 14) and robustness (see rows named Boost BT in Figure 8 and Figure 15). Both neural networks and MLE approaches generally struggle with small phylogenies (see Figure 4, Figure 5 and Figure 6 for larger errors represented by the yellow data points, see also Appendix G, Figure 14). Performance improves significantly on medium and large phylogenies for both neural network and MLE approaches.

377

The MLE implementation sometimes fails to find an optimal solution. In our visualizations, failed MLE estimations are indicated by small squares spreading along the x-axis to avoid misinterpretation. MLE tended to give small or near-zero estimates, particularly on the extinction rate and the carrying capacity. This phenomenon is more prominent when starting optimization from a random point. For all figures showing the MLE error, the ideal situation is that all the data points lie near the horizontal black two-dash reference lines (at which the error is 0) and do not spread along or near the purple dotted reference lines (which suggests near-zero MLE estimations). See the last two panels of Figure 4, Figure 5 and Figure 6 for details.

386

Neural networks often return values closer to the parameter space's mid-points (indicated by red dashed lines), a result of making "safer" predictions that minimize loss compared to random guesses. Consequently, neural networks usually overestimate at low true values and underestimate at high true values (see Figure 4, Figure 5 and Figure 6).

390 These errors are mitigated or partially corrected when the neural networks are trained in  
391 tandem through boosting strategies, e.g. boosting GNN results with DNN or LSTM or  
392 both (see the panels of Boost SS, Boost BT and Boost SS+BT in Figure 4, Figure 5 and  
393 Figure 6). This happens particularly for large phylogenies (the blue data points in  
394 Appendix G, Figure 14) when the underlying true carrying capacity ( $K$ ) is large, or for  
395 small phylogenies (the yellow data points) when the underlying true speciation rate ( $\lambda$ ) is  
396 small.

397 However, boosting strategies can introduce their own challenges. When boosting  
398 GNN results first with LSTM and then with DNN, the DNN failed to identify a general  
399 pattern of errors from LSTM results. This led to overfitting on the training dataset at the  
400 second epoch of the training session (the total loss in the validation dataset started to  
401 increase and became much larger than the total loss in the training dataset), which, in  
402 turn, resulted in poor performance on the testing dataset (see the panels named Boost  
403 BT+SS in Figure 4, Figure 5, Figure 6 and Appendix G, Figure 14).

404 Upon further analysis of the residuals, we observed that inaccuracies in the  
405 predictions were largely influenced by the size of the phylogeny (Figure 4, Figure 5 and  
406 Figure 6). For neural network approaches, the prediction errors for speciation rate,  
407 extinction rate, and carrying capacity tended to increase as the size of the phylogeny  
408 decreased, especially in phylogenies with fewer than 200 nodes. Systematic error was also  
409 identified in the estimation of carrying capacity: neural networks generally overestimated  
410 this parameter in smaller phylogenies and underestimated it in larger ones. Boosting  
411 strategies were effective in mitigating or partially correcting systematic errors, and  
412 enhancing prediction accuracy, particularly for carrying capacity (see the rows of Boost SS,  
413 Boost BT and Boost SS+BT in Appendix G, Figure 14).

414 We calculated the strength of the carrying capacity effect using the formula  
415  $1/K' = (\lambda - \mu)/K$ , where  $\lambda$  represents the true speciation rate,  $\mu$  the true extinction rate,  
416  $K$  the true carrying capacity, and  $K'$  the diversity at which speciation becomes zero for

417 linear negative diversity-dependence (Etienne et al., 2012). A larger  $(\lambda - \mu)/K$  value  
418 corresponds to smaller  $K'$  and therefore a stronger carrying capacity effect. Phylogenies  
419 exhibiting a stronger carrying capacity effect typically have more accurate estimates.  
420 However, in the case of smaller phylogenies, neural networks tended to underestimate  
421 speciation and extinction rates while overestimating carrying capacity when the carrying  
422 capacity effect is weak, and the reverse is observed when the effect is strong. In contrast,  
423 MLE tends to overestimate speciation and extinction rates while underestimating carrying  
424 capacity under conditions of weak carrying capacity effect, with the reverse occurring  
425 under strong effects, except for the carrying capacity which is always underestimated (see  
426 Appendix G, Figure 14). Neural network methods tend to underestimate the carrying  
427 capacity effect. This phenomenon can be mitigated by the boosting strategies, especially  
428 the Boost BT method, which achieved similar performance to the best case MLE estimates  
429 (see Figure 7).

430 Unlike GNN and LSTM, DNN cannot by itself reliably recover speciation and  
431 extinction rates from the summary statistics of the phylogenies, with its predictions mostly  
432 clustering around the mid-points of the parameter space (around the red dashed lines in  
433 the DNN panels in Figure 4, Figure 5 and Figure 6). The overall accuracy of the carrying  
434 capacities recovered by the DNN is also inferior compared to the other approaches (see the  
435 row named DNN in Appendix G, Figure 14).

436 Among all ensemble learning strategies, boosting consistently outperformed both  
437 bagging and stacking in enhancing prediction accuracy compared to using neural networks  
438 independently, as can be seen, for instance, by the lower mean absolute prediction errors in  
439 Appendix G, Figure 14. Boosting strategies also exhibited better performance in recovering  
440 the true values of the carrying capacity effect (see Figure 7). The most effective neural  
441 network approaches overall matched or even surpassed the results of MLE while exhibiting  
442 no bias, even on smaller phylogenies. Overall, sequential boosting of GNN results first with  
443 DNN and then with LSTM (Boost SS+BT) led to best performance in terms of prediction

accuracy, except for estimating carrying capacity on large phylogenies with around 2000 nodes (see the row named Boost SS+BT in Appendix G, Figure 14 this strategy led to overestimation on very large phylogenies). However, Boost SS+BT led to more overestimation on the true values of the carrying capacity effect, as compared to the strategy boosting the GNN results with only LSTM (Boost BT, see Figure 7).

#### 449 *Robustness Analysis*

450 As a proxy for robustness of each method, we used the mean absolute errors of the  
451 parameters estimated from sets of phylogenies simulated under identical true parameters.  
452 Our analysis indicates that the robustness of the methods against phylogenetic  
453 heterogeneity (e.g., phylogenies of very different sizes, topologies and other characteristics)  
454 depends on the values of the underlying true parameters. We observed that the strength of  
455 the carrying capacity effect critically influences robustness. Generally, a weaker carrying  
456 capacity effect (associated to a smaller value of  $(\lambda - \mu)/K$ ) tends to diminish the  
457 robustness of both MLE and neural network methods across all parameters: speciation  
458 rate, extinction rate, and carrying capacity (as can be seen in Figure 8 and Appendix G,  
459 Figure 16 and Figure 15, by observing the increase of error along with the darkening  
460 background colors from light pink to dark blue).

461 When the carrying capacity effect is weak, neural network methods typically exhibit  
462 greater robustness in estimating speciation and extinction rates compared to the best-case  
463 MLE results (see Figure 8 and Appendix G, Figure 16). When the carrying capacity effect  
464 is exceptionally strong, the best-case MLE results can outperform neural networks  
465 particularly when estimating carrying capacity. Typical-case MLE results consistently  
466 show less robustness compared to all neural network methods.

467 A higher extinction rate generally decreases the robustness of all methods in  
468 estimating any parameter. A higher speciation rate enhances the robustness of carrying  
469 capacity estimates across all methods, although its impact on the robustness of speciation

20

TIANJIAN QIN, KOEN J. VAN BENTHEM, LUIS VALENTE, RAMPAL S. ETIENNE

470 and extinction rate estimates is not consistent. A higher carrying capacity generally  
471 decreases the robustness of all methods in estimating carrying capacity.

472 Note that MLE typical-case results often contained more extreme estimations than  
473 best-case results, consequently, the exclusion of extreme values could lead to a wrong  
474 impression in the figures that when the carrying capacity effect is weak, the typical-case  
475 MLE is more robust than the best-case MLE. This is particularly prominent for the  
476 speciation rate. The exclusion of these extreme values is crucial, however, as they are rare  
477 and their magnitude can obstruct meaningful interpretation and comparison.

478 We find that DNN alone (estimating parameters from summary statistics) shows  
479 the worst robustness among all the methods and LSTM alone (estimating parameters from  
480 branching times) shows the greatest robustness overall. Among all the estimation methods,  
481 the MLE best-case achieved the greatest possible robustness in estimating the extinction  
482 rate and the carrying capacity while GNN alone (estimating parameters from phylogenies)  
483 achieved the greatest possible robustness in estimating the speciation rate. Among the  
484 neural network methods, GNN alone achieved the greatest possible robustness in  
485 estimating the speciation rate and the carrying capacity while Boost BT (boosting GNN  
486 estimates with LSTM) achieved the greatest possible robustness in estimating the  
487 extinction rate. See Figure 8 and Appendix G, Figure 15 and Figure 16 for details.

488

### *Empirical Data*

489 MLE estimates of carrying capacity are typically lower than those of the neural  
490 networks, especially in smaller phylogenies (Figure 9). However, as the size of the  
491 phylogenies increases, MLE estimates tend to converge towards those produced by neural  
492 networks. Similarly, MLE estimates of net diversification rate (computed as  $\lambda - \mu$ ) also  
493 align more closely with neural network estimates in larger phylogenies Figure 9.

494 MLE generally provides a broader range of estimates on all the parameters except  
495 for carrying capacity on small phylogenies. Neural networks provide a broader range of

496 carrying capacity estimates on small phylogenies and less frequently produce zero or  
497 near-zero estimates for extinction rates which we often observe for MLE. We also observed  
498 that MLE sometimes produces extreme values (ranging from 10,000 to infinity) for  
499 carrying capacity on empirical trees, see Figure 9 for the comparison between MLE and  
500 neural networks on empirical tree parameter estimation.

501 Generally, neural network estimates of all the parameters under the DDD scenario  
502 are close to the center (mean) of the distribution generated by the bootstrapping method.  
503 See Figure 13 in Appendix F for details.

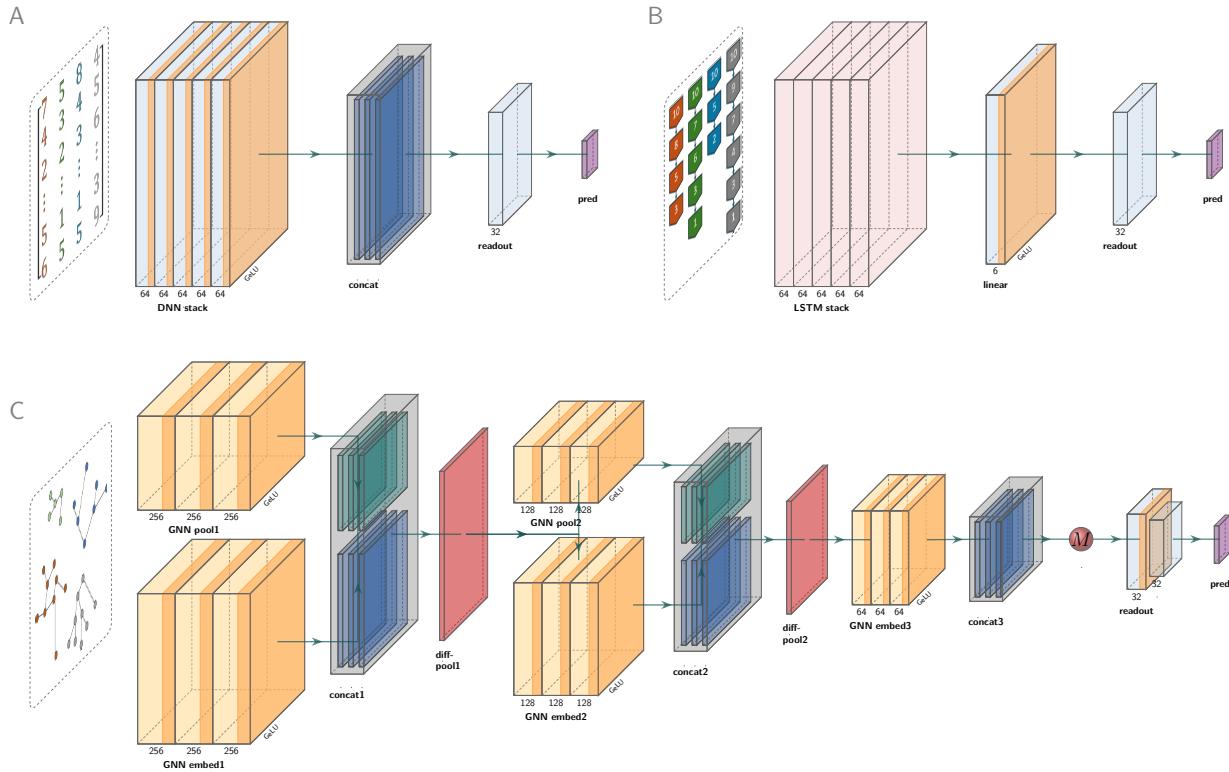


Fig. 1. Illustration of the neural network architectures. From left to right, for each neural network, the inputs are filtered through the layers, and the network ultimately outputs the final predictions of the parameters through the readout layers. A: dense neural network (DNN), whose input data are summary statistics. The major component of the DNN is a stack comprising five linear layers ("DNN stack"), each followed by a Batch Normalization for 1D Inputs operator (BatchNorm1D, not shown in figure) and Gaussian Error Linear Units (GELU, the orange band within the boxes). Learned features from all the linear layers within the stacks are collected and concatenated ("concat"). A single linear readout layer ("readout") outputs  $n$  predicted parameters ("pred"). B: long short-term memory recurrent neural network (LSTM), whose input data are the branching times. The major component of the LSTM is a stack of five LSTM recurrent neural network layers ("LSTM stack"). Learned features are processed by a linear layer accompanied by a GELU ("linear"), then passed to a single linear readout layer ("readout") that outputs  $n$  predicted parameters ("pred"). C: graph neural network (GNN), whose input data is a graph representation of the phylogeny. GNN is assembled from five modules. Each module comprises the same number of GraphSAGE (sample-and-aggregate graph convolutional neural network) operators. Each operator is accompanied by a BatchNorm1d (not shown in the figure) operator and then a GELU activation function (illustrated by the orange bands within the yellow boxes). Learned features from all the GraphSAGE operators within a module are collected and concatenated. The differentiable pooling (DiffPool) technique is adopted to perform graph coarsening. In the first coarsening operation, the graph data inputs are passed to two GNN modules ("GNN pool1" and "GNN embed1"). The pooling group reduces the graph size, while the embedding group captures the node features. The filtered data from each GraphSAGE operator are concatenated ("concat1") and then passed to a DiffPool layer ("diff-pool1"), which finalizes the first coarsening operation. The second coarsening operation is applied in the same way as the first (as represented by "GNN pool1", "GNN embed2", "concat2"), and the outputs from the second DiffPool layer ("diff-pool2") are passed to the final (fifth) GNN module ("GNN embed3"). After the final GNN module, the outputs are concatenated ("concat3") and transformed by a global mean pooling operation (red ball "M") to create a final graph representation. This graph representation is passed to a readout layer group ("readout" as represented by light blue boxes) consisting of two linear layers to perform graph-level regression which ultimately outputs a vector of  $n$  predicted parameters ("pred" as represented by a purple box). Only the first linear layer is followed by GELU (see the orange band of the first linear layer). See Appendix C for the detailed description and technical details.

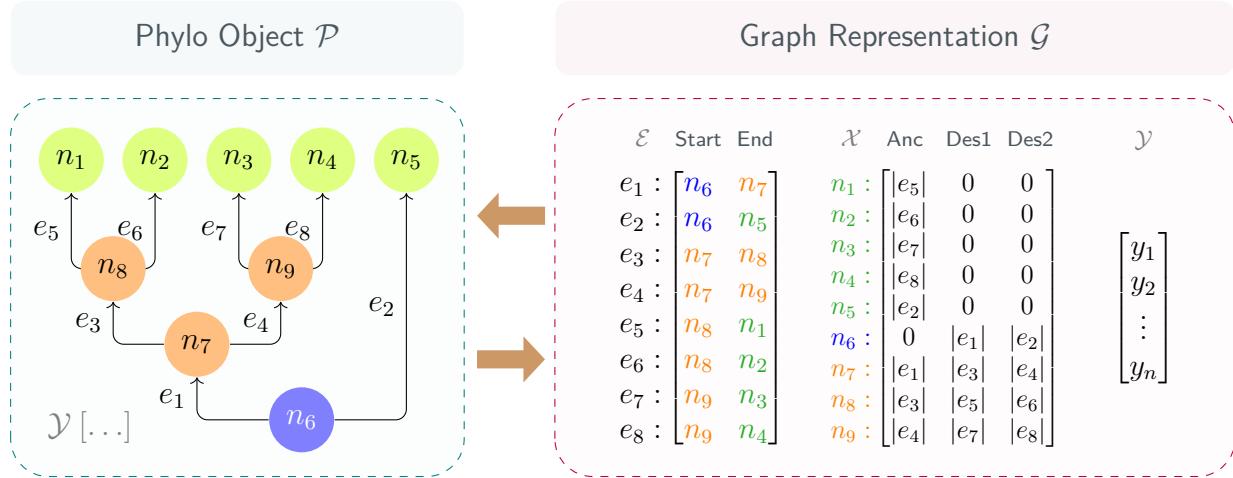
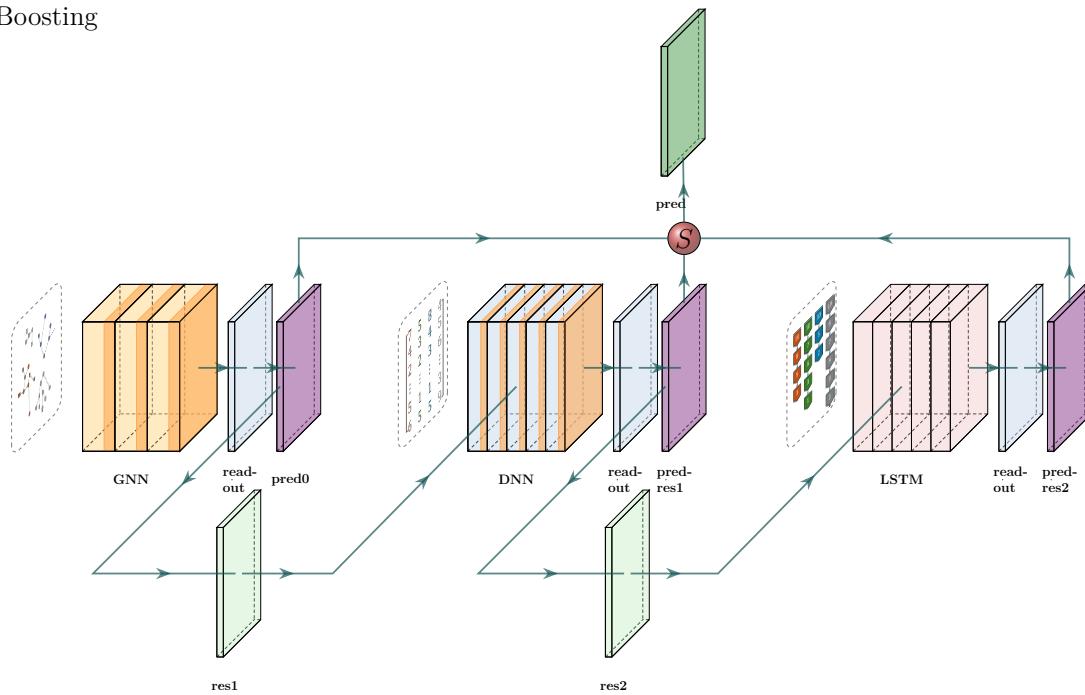
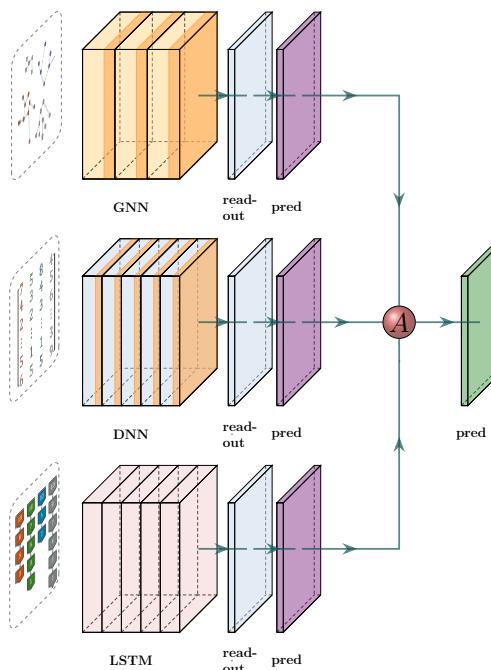


Fig. 2. Illustration of data transformation between a "phylo" object and its graph representation. The left panel shows a visualization of a "phylo" object. The blue circle represents the root node, orange circles represent the internal nodes and green circles represent the tip nodes. Arrows represent directed edges between each pair of the nodes. The right panel shows the transformed graph data structure. The adjacency list is denoted as  $\mathcal{E}$ . Each row of the adjacency list represents one edge, the first column represents the starting node and the second column represents the end node. Note that the adjacency list is transposed (in the example into  $\mathcal{E}^{2 \times 8}$ ) after converting to a tensor. The node feature matrix is denoted as  $\mathcal{X}$ . Each row of the node feature matrix represents the features contained in one node, the first column represents the distance from the node to its direct ancestor node, the second and the third columns represent the distances from the node to its two descendants. In the node feature matrix, the distances from a node to non-existing nodes (e.g. the tip nodes have no descendants, and the root node has no ancestor) are represented by zeros. The node and edge labels before the colons (including the colons) are placed here for visual assistance. After transformation, we use graph-level attributes  $\mathcal{Y}$  to store the parameters used to generate the "phylo" object. The node labels are given by  $n_1, n_2, n_3, \dots, n_9$ , the edge labels are given by  $e_1, e_2, e_3, \dots, e_8$ , the edge lengths are given by  $|e_1|, |e_2|, |e_3|, \dots, |e_8|$ . The generating parameters are given by a vector  $[y_1, y_2, \dots, y_n]$  where  $n$  is the number of parameters

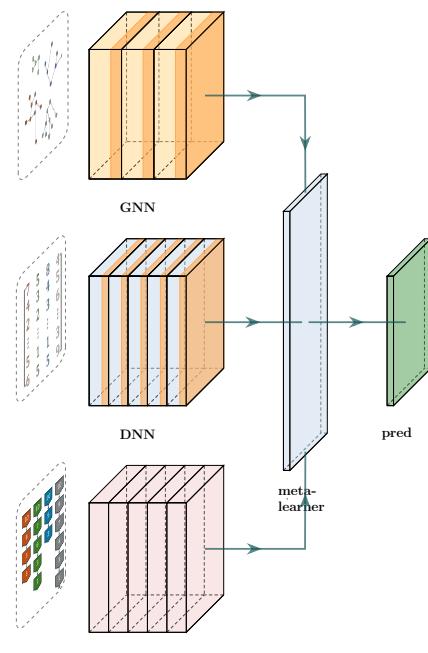
Boosting



Bagging



Stacking



(Caption on next page.)

Fig. 3. (*Figure on previous page.*) Illustration of the ensemble learning strategies to combine the graph neural network (GNN), the dense neural network (DNN) and the long short-term memory recurrent neural network (LSTM). The neural networks are largely simplified. With boosting, GNN, DNN and LSTM were trained sequentially to iteratively correct residuals. For example, DNN is trained to predict the residuals of GNN predictions. Subsequently, LSTM is trained to predict the residuals of the residuals after DNN corrected the GNN predictions. The final prediction comes from the initial prediction by GNN minus two learned residual terms by DNN and LSTM. With bagging, we trained GNN, DNN and LSTM independently, translated their original outputs to parameter predictions and then aggregated the predictions. With stacking, we trained GNN, DNN and LSTM simultaneously but without readout. We directly concatenated the outputs from GNN, DNN and LSTM and then used a meta-learner to make predictions from the outputs. With bagging, we trained GNN, DNN and LSTM independently ("GNN", "DNN" and "LSTM" blocks of boxes), translated their outputs to parameter predictions through their own readout layers (three "readout" boxes next to the neural networks and three "pred" boxes next to the readout layers) and then aggregated the predictions (red ball "A"). With stacking, we trained GNN, DNN and LSTM simultaneously ("GNN", "DNN" and "LSTM" blocks of boxes) but without their own readout layers. We combined the features from DNN, LSTM and GNN and fed to a meta-learner ("meta-learner") comprising linear neural network layers to output parameter predictions. With boosting, there can be different pathways. In our illustration, GNN, DNN and LSTM were trained sequentially to iteratively correct residuals. First, the GNN is trained from the graphs to make the initial predictions (see "GNN", "readout" and then "pred0") and from predicted and ground truth values of the parameters we computed the residuals ("res1") second, the DNN is trained to predict these residuals from the summary statistics (see "DNN", "readout" and then "pred-res1"), learning to correct the GNN's errors; lastly, the LSTM is trained to predict the residuals of the residuals (see "LSTM", "readout" and then "pred-res2"), which is the initial predictions minus the predicted residuals by the DNN, from branching times, to further improve the predictive accuracy. Finally, we subtracted the two residual terms from the initial predictions (red ball "S") to make the corrected predictions. See Appendix D for a detailed explanation.

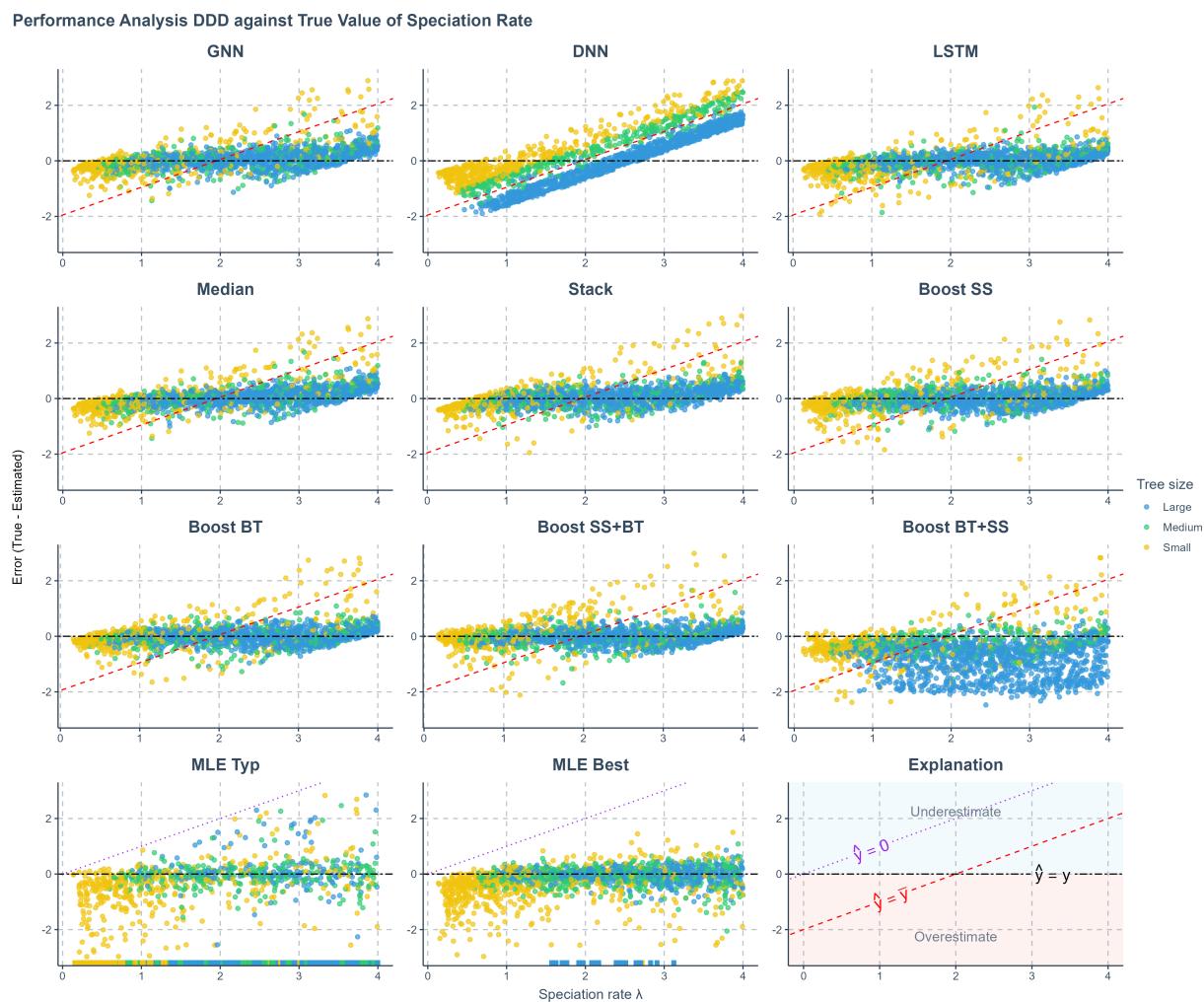


Fig. 4. Prediction error of various methods applied to phylogenies simulated under a diversity-dependent diversification scenario, against true values of the speciation rate. The errors shown (y-axis) are the differences between the true parameters (x-axis) used to simulate the phylogenies and the values predicted or estimated by each method. Each panel represents an estimation method. Phylogenies are categorized based on their size: yellow for small phylogenies with fewer than 200 nodes (including root, internal, and tip nodes), green for medium-sized phylogenies with 200 to 500 nodes, and blue for large phylogenies with more than 500 nodes, refer to Appendix G, Figure 17 for how the tree sizes are distributed. GNN: Predictions obtained by the graph neural network using the phylogenies. DNN: Predictions by the dense neural network using summary statistics. LSTM: Predictions by the long short-term memory recurrent neural network using branching times. Median: Bagging strategy that takes the median value of the predictions from GNN, DNN, and LSTM. Stack: Stacking strategy that utilizes a meta-learner to integrate results from GNN, DNN, and LSTM. Boost SS: Boosting strategy that corrects GNN results using DNN. Boost BT: Boosting strategy that corrects GNN results using LSTM. Boost SS+BT: Sequential correction of GNN errors first using DNN, followed by LSTM. Boost BT+SS: Sequential correction of GNN errors first using LSTM, followed by DNN. MLE Typ: Maximum Likelihood Estimation results using a random starting point within the parameter space of the training dataset for each parameter's optimization. MLE Best: MLE results using the true parameter values as the starting points for optimization. Red dashed lines in panels representing neural network results indicate the mid-points of the parameter spaces ( $\hat{y} = \bar{y}$  where  $\hat{y}$  denotes an estimated parameter and  $\bar{y}$  denotes the mid-point of the parameter space). Data points close to purple dotted lines ( $\hat{y} = 0$ ) in MLE result panels indicate near-zero estimates. Black two-dash lines indicate accurate estimates ( $\hat{y} = y$  where  $y$  denotes the true parameter value). In the MLE result panels, small squares spreading along the x-axis signify optimization failures. Due to significantly lower accuracy, other aggregation methods from the bagging strategy are not displayed on the plot.  $\lambda$ : Speciation rate.

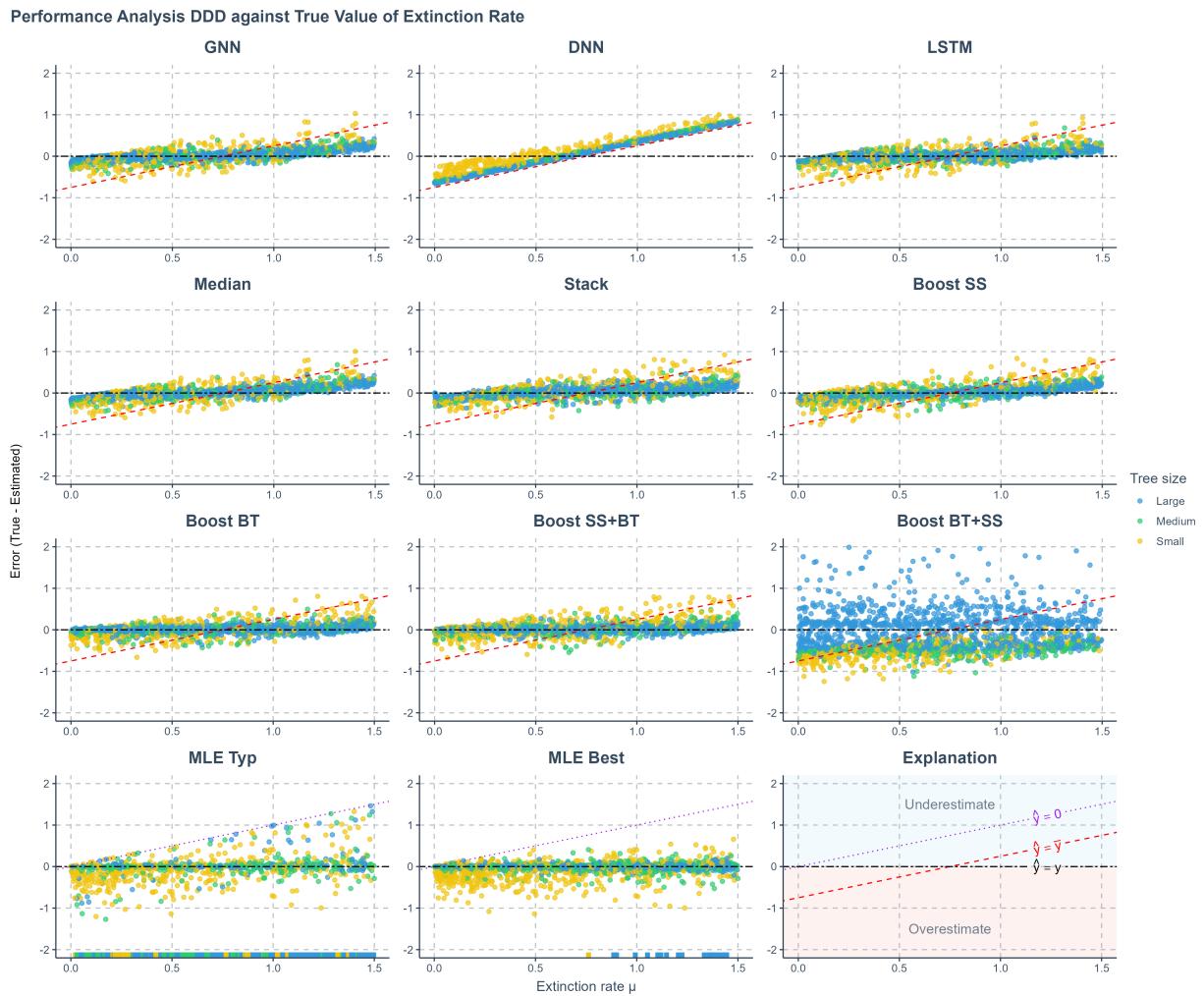


Fig. 5. Prediction error of various methods applied to phylogenies simulated under a diversity-dependent diversification scenario, against true values of the extinction rate. The errors shown (y-axis) are the differences between the true parameters (x-axis) used to simulate the phylogenies and the values predicted or estimated by each method. Each panel represents an estimation method. Phylogenies are categorized based on their size: yellow for small phylogenies with fewer than 200 nodes (including root, internal, and tip nodes), green for medium-sized phylogenies with 200 to 500 nodes, and blue for large phylogenies with more than 500 nodes. GNN: Predictions obtained by the graph neural network using the phylogenies. DNN: Predictions by the dense neural network using summary statistics. LSTM: Predictions by the long short-term memory recurrent neural network using branching times. Median: Bagging strategy that takes the median value of the predictions from GNN, DNN, and LSTM. Stack: Stacking strategy that utilizes a meta-learner to integrate results from GNN, DNN, and LSTM. Boost SS: Boosting strategy that corrects GNN results using DNN. Boost BT: Boosting strategy that corrects GNN results using LSTM. Boost SS+BT: Sequential correction of GNN errors first using DNN, followed by LSTM. Boost BT+SS: Sequential correction of GNN errors first using LSTM, followed by DNN. MLE Typ: Maximum Likelihood Estimation results using a random starting point within the parameter space of the training dataset for each parameter's optimization. MLE Best: MLE results using the true parameter values as the starting points for optimization. Red dashed lines in panels representing neural network results indicate the mid-points of the parameter spaces ( $\hat{y} = \bar{y}$  where  $\hat{y}$  denotes an estimated parameter and  $\bar{y}$  denotes the mid-point of the parameter space). Data points close to purple dotted lines ( $\hat{y} = 0$ ) in MLE result panels indicate near-zero estimates. Black two-dash lines indicate accurate estimates ( $\hat{y} = y$  where  $y$  denotes the true parameter value). In the MLE result panels, small squares spreading along the x-axis signify optimization failures. Due to significantly lower accuracy, other aggregation methods from the bagging strategy are not displayed on the plot.  $\mu$ : extinction rate.

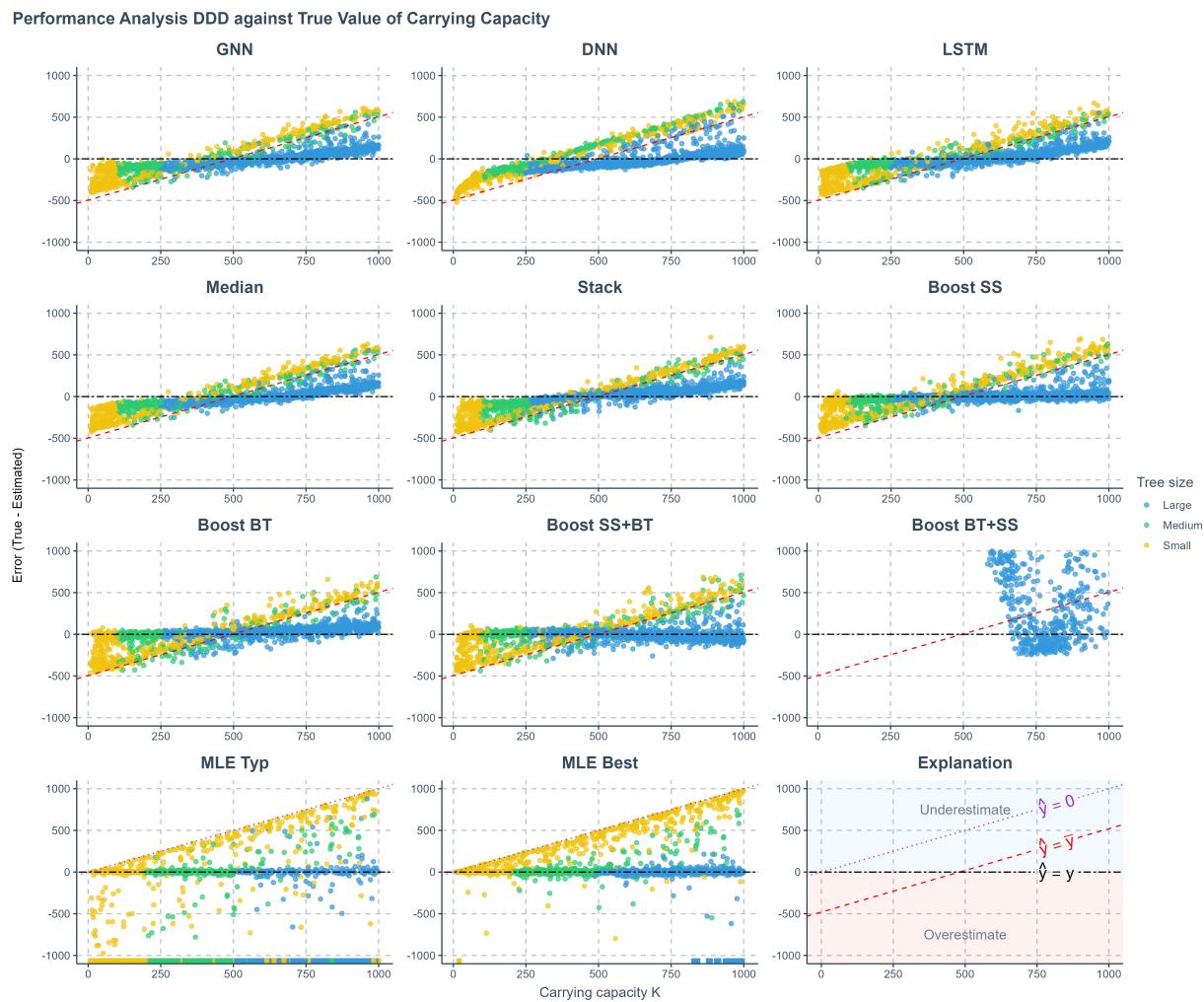


Fig. 6. Prediction error of various methods applied to phylogenies simulated under a diversity-dependent diversification scenario, against true values of the carrying capacity. The errors shown (y-axis) are the differences between the true parameters (x-axis) used to simulate the phylogenies and the values predicted or estimated by each method. Each panel represents an estimation method. Phylogenies are categorized based on their size: yellow for small phylogenies with fewer than 200 nodes (including root, internal, and tip nodes), green for medium-sized phylogenies with 200 to 500 nodes, and blue for large phylogenies with more than 500 nodes. GNN: Predictions obtained by the graph neural network using the phylogenies. DNN: Predictions by the dense neural network using summary statistics. LSTM: Predictions by the long short-term memory recurrent neural network using branching times. Median: Bagging strategy that takes the median value of the predictions from GNN, DNN, and LSTM. Stack: Stacking strategy that utilizes a meta-learner to integrate results from GNN, DNN, and LSTM. Boost SS: Boosting strategy that corrects GNN results using DNN. Boost BT: Boosting strategy that corrects GNN results using LSTM. Boost SS+BT: Sequential correction of GNN errors first using DNN, followed by LSTM. Boost BT+SS: Sequential correction of GNN errors first using LSTM, followed by DNN. MLE Typ: Maximum Likelihood Estimation results using a random starting point within the parameter space of the training dataset for each parameter's optimization. MLE Best: MLE results using the true parameter values as the starting points for optimization. Red dashed lines in panels representing neural network results indicate the mid-points of the parameter spaces ( $\hat{y} = \bar{y}$  where  $\hat{y}$  denotes an estimated parameter and  $\bar{y}$  denotes the mid-point of the parameter space). Data points close to purple dotted lines ( $\hat{y} = 0$ ) in MLE result panels indicate near-zero estimates. Black two-dash lines indicate accurate estimates ( $\hat{y} = y$  where  $y$  denotes the true parameter value). In the MLE result panels, small squares spreading along the x-axis signify optimization failures. Due to significantly lower accuracy, other aggregation methods from the bagging strategy are not displayed on the plot.  $K$ : carrying capacity.

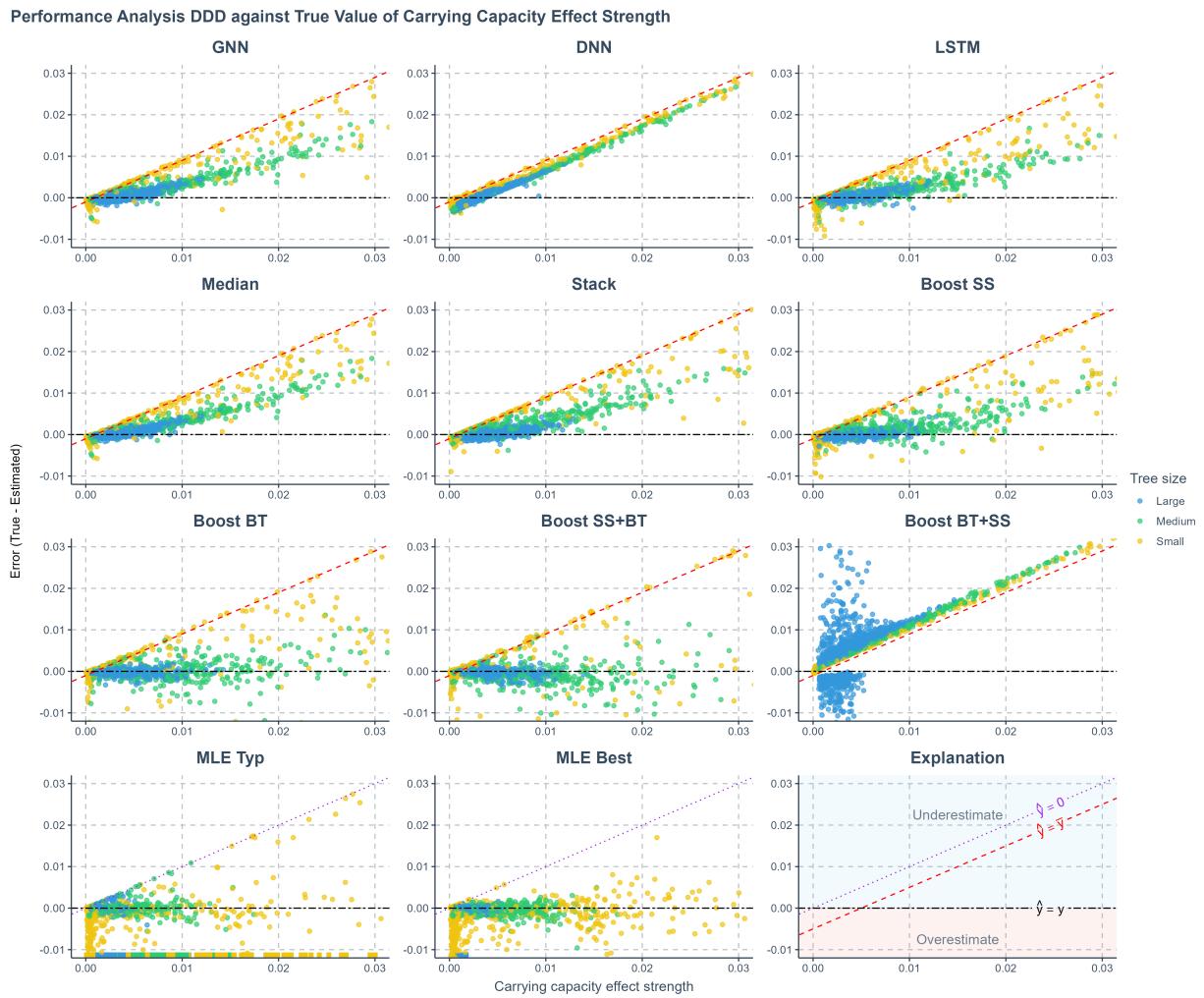
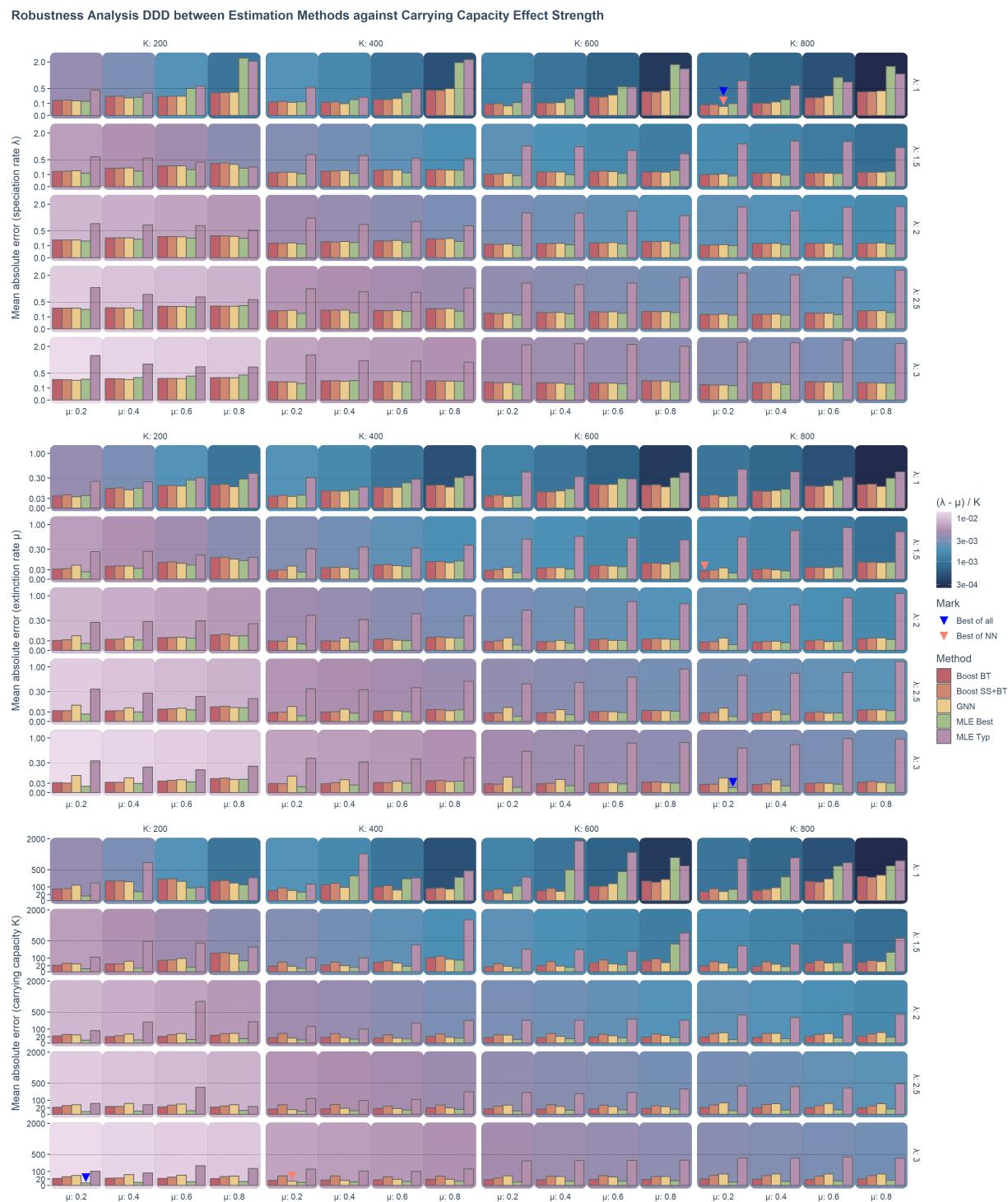


Fig. 7. Prediction error of estimated carrying capacity effect computed from estimated values of speciation rate, extinction rate and carrying capacity using various methods applied to phylogenies simulated under a diversity-dependent diversification scenario, plotted against the true carrying capacity effect computed from true parameters. The errors shown (y-axis) are the differences between the values of true carrying capacity effect (x-axis) used to simulate the phylogenies and the values predicted or estimated by each method. Each panel represents an estimation method. Phylogenies are categorized based on their size: yellow for small phylogenies with fewer than 200 nodes (including root, internal, and tip nodes), green for medium-sized phylogenies with 200 to 500 nodes, and blue for large phylogenies with more than 500 nodes. GNN: Predictions obtained by the graph neural network using the phylogenies. DNN: Predictions by the dense neural network using summary statistics. LSTM: Predictions by the long short-term memory recurrent neural network using branching times. Stack: Stacking strategy that utilizes a meta-learner to integrate results from GNN, DNN, and LSTM. Boost SS: Boosting strategy that corrects GNN results using DNN. Boost BT: Boosting strategy that corrects GNN results using LSTM. Boost SS+BT: Sequential correction of GNN errors first using DNN, followed by LSTM. MLE Typ: Maximum Likelihood Estimation results using random starting points for parameter optimization. MLE Best: MLE results using the true parameter values as the starting points for optimization. Red dashed lines in panels representing neural network results indicate the mid-points of the parameter spaces ( $\hat{y} = \bar{y}$  where  $\hat{y}$  denotes an estimated parameter and  $\bar{y}$  denotes the mid-point of the parameter space). Data points close to purple dotted lines ( $\hat{y} = 0$ ) in MLE result panels indicate near-zero estimates. Black two-dash lines indicate accurate estimates ( $\hat{y} = y$  where  $y$  denotes the true parameter value). In the MLE result panels, small squares spreading along the x-axis signify optimization failures. The title of the figure shows how the carrying capacity effect is computed,  $\lambda$ : Speciation rate.  $\mu$ : extinction rate.  $K$ : carrying capacity.



(Caption on next page.)

Fig. 8. (Figure on previous page.) The robustness (mean absolute error) of neural network and maximum likelihood estimation was assessed for 80 sets of phylogenies, each containing 1000 trees randomly simulated under a diversity-dependent diversification scenario, employing identical parameter settings but varying in size, topology, and structure. The robustness results of the speciation rate, the extinction rate and the carrying capacity are shown from top to bottom. For each panel group associated to a parameter, each panel contains the robustness of different estimation methods (the MLE and neural networks) under a combination of parameters indicated by the facet strip labels. Each facet column represents the robustness under a specific carrying capacity ( $K$ ) setting used in the simulation of the phylogenies. Each facet row represents a specific speciation rate ( $\lambda$ ). Each group of the bars represents a specific extinction rate ( $\mu$ ) as shown by the x-axis. The background color of a panel represents the carrying capacity effect strength (calculated as  $(\lambda - \mu)/K$  and visualized in "log10" scale), from bottom-left to top-right, the carrying capacity effect strength decreases. The color of a bar represents the associated estimation method. Boosting BT: Graph neural network with long short-term memory recurrent neural network correcting its residuals using branching times. Boosting SS + BT: Graph neural network with dense neural network and long short-term memory recurrent neural network correcting residuals sequentially using summary statistics and branching times. GNN: Graph neural network. MLE Best: Maximum likelihood estimation using true parameters as the starting points. MLE Typ: Maximum likelihood estimation using a random value as the starting point of optimization for each parameter. X-axis: Represents extinction rate ( $\mu$ ) settings. Y-axis: Represents the mean absolute error in a square-root transformed scale. Some bars are marked; for each parameter, the blue triangle represents the greatest possible robustness achieved among all the estimation methods, the red triangle represents the greatest possible robustness achieved among the neural network methods.

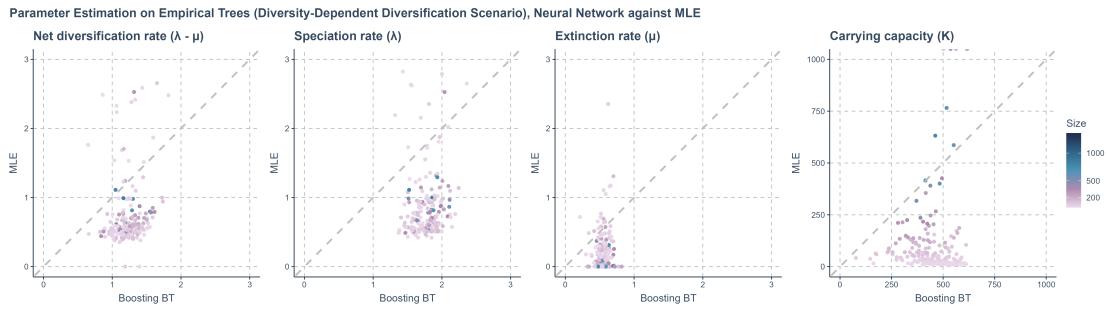


Fig. 9. Comparison of the estimations of Maximum Likelihood Estimation (MLE) and neural network methods (specifically, Boosting BT, which refers to using graph neural network to make first predictions and then using a long short-term memory recurrent neural network to correct for residuals) on empirical trees under a diversity-dependent diversification scenario. Each panel, arranged from left to right, focuses on a specific parameter being estimated. X-axis: Represents the estimated values of the neural network. Y-axis: Represents estimated the values from MLE. A gray dashed line is included in each panel to indicate where the estimations from the neural network and MLE are exactly the same. The color of the points varies from purple to blue, with the gradient representing the size of the phylogenies measured by the total number of nodes (including root, internal, and tip nodes).

504

### Other Scenarios

505

*Birth-Death Scenario* Neural network methods outperformed MLE in accuracy,

506

particularly on smaller phylogenies, under the BD scenario in the simulated dataset (see Appendix H, Figure 19 and Figure 18). Both MLE and neural network methods give less accurate estimates on small phylogenies; this is more prominent for the MLE estimates.

509

On empirical phylogenies, similar to the DDD scenario, neural network methods

510

seldom produce zero-estimation of extinction rate, unlike MLE, which often gives zero or near-zero estimation on extinction rate. Neural networks tend to give estimates within the parameter space of the training dataset. They predict conservative speciation and

511

extinction rates yet are highly consistent with MLE estimation on the net diversification rate, defined as the difference between speciation and extinction rates ( $\lambda - \mu$ ). The

512

consistency of prediction increases on larger empirical phylogenies. See Figure 10 for details.

513

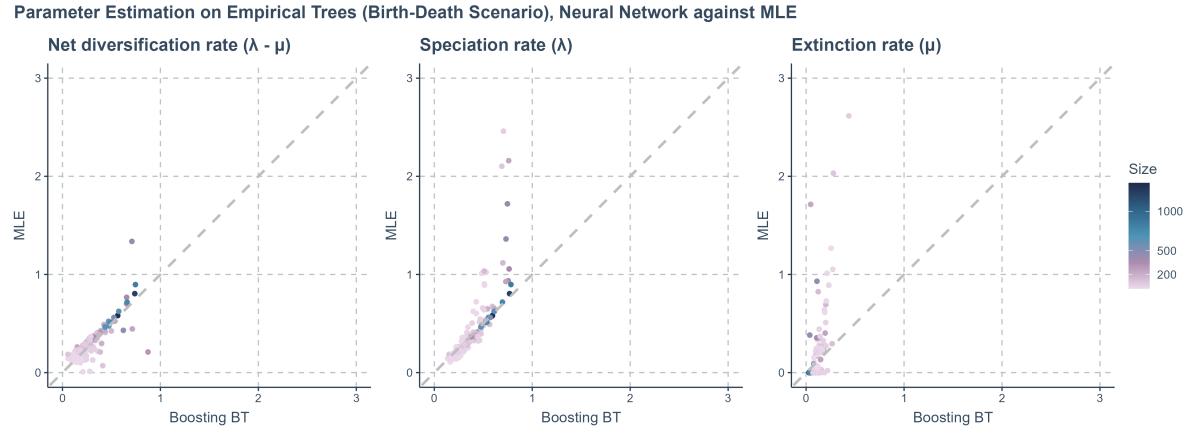


Fig. 10. Comparing the estimations of Maximum Likelihood Estimation (MLE) and neural network methods (specifically, Boosting BT, which refers to using graph neural network to make first predictions and then using long short-term memory recurrent neural network to correct for residuals) on empirical trees under a birth-death scenario. Each panel, arranged from left to right, focuses on a specific parameter being estimated. X-axis: Represents the estimated values of the neural network. Y-axis: Represents estimated the values from MLE. A gray dashed line is included in each panel to indicate where the estimations from the neural network and MLE are exactly the same. The color of the points varies from purple to blue, with the gradient representing the size of the phylogenies measured by the total number of nodes (including root, internal, and tip nodes).

517        *Protracted Birth-Death Scenario*    Both maximum likelihood estimation (MLE) and  
 518        neural network methods did not perform well on estimating parameters under the PBD  
 519        scenario; MLE estimates were generally less accurate, but neural networks also failed to  
 520        predict the parameters as all the parameter estimates are close to the mid-points of  
 521        corresponding parameter spaces (Appendix I, Figure 20,). However, there are exceptions:  
 522        neural networks seem to perform better on the speciation rate of the incipient species ( $\lambda_3$ )  
 523        and on the mean duration of speciation ( $\tau$ ) when the true value is between 0 and 2.

524        MLE estimates become significantly inaccurate as phylogenies become increasingly  
 525        small (Appendix I, Figure 21); it is also noticeable that MLE estimates of the speciation  
 526        completion rate ( $\lambda_2$ ) are very inaccurate, especially when the phylogenies are large. A  
 527        general pattern is that both MLE and neural network methods achieve more accurate  
 528        estimates on phylogenies with higher true values of the mean duration of speciation  
 529        (Appendix I, Figure 22).

530

## DISCUSSION

531 We have developed an ensemble learning based neural network approach that  
532 matches and sometimes outperforms the accuracy and robustness of maximum likelihood  
533 estimation (MLE) for estimating phylogenetic tree parameters. Our approach leverages  
534 different classes of neural networks by learning from the phylogenies, their branching times  
535 and their summary statistics simultaneously.

536 When trained, our neural networks can compute estimates faster than MLE on  
537 larger phylogenies as computation time is less affected by increases in phylogeny size. We  
538 considered boosting strategies most effective in eliminating systematic prediction errors in  
539 neural network estimates. Among them, Boost BT (which corrects GNN results using  
540 LSTM) achieved overall best performance which is comparable to, or even surpassing, the  
541 best case MLE, in terms of accuracy and robustness. We observed that generally the  
542 performance of the typical MLE was second-worst (Boost BT+SS was worst).  
543 Interestingly, some phylogenies, such as small trees and those shaped by relatively weak  
544 effects, pose significant challenges to both MLE and neural network methods.

545 Previous neural network methods applied to phylogenies have experimented with  
546 various architectures such as convolutional neural networks (CNN), GNN, and LSTM  
547 (Lajaaiti et al., 2023; Lambert et al., 2023; Voznica et al., 2022). The deep learning  
548 architectures we employed differ from those used in prior studies, making direct  
549 comparisons challenging. Additionally, while previous research focused on birth-death  
550 (Lambert et al., 2023) and trait-state-dependent models (Lajaaiti et al., 2023), our  
551 approach is novel in its application to models such as diversity-dependent diversification  
552 (DDD) and protracted birth-death (PBD) from a neural network perspective. Despite  
553 these differences, our findings align with recent studies in underscoring the potential of  
554 neural networks to infer diversification processes, offering a viable alternative to  
555 mathematically complex methods.

556 *Rethinking Neural Networks*

557        Although performance was equal or better than MLE, the neural network approach  
558    is not without its shortcomings. The neural networks often defaulted to predicting values  
559    close to the mid-point of the true parameter space of the training dataset, indicating that  
560    they struggle to extract meaningful features from the dataset. This conservative prediction  
561    strategy minimizes overall error compared to random guessing. Examples can be found in  
562    the GNN predictions of carrying capacity from simulated DDD trees, the DNN estimates  
563    of the speciation and extinction rates (Figure 4, Figure 5 and Figure 6, but see  
564    Appendix L for a detailed investigation of possible under-performance of DNN on the  
565    summary statistics) and most neural network predictions of PBD related parameters  
566    (Appendix I, Figure 20), especially for smaller phylogenies.

567        This behavior, while effective in reducing apparent error metrics, can skew our  
568    understanding of a neural networks performance particularly when the focal parameter  
569    space is relatively narrow. Neural networks may consistently show smaller overall error  
570    compared to MLE, because the latter has no prior knowledge of the limits of the  
571    parameter space, which would lead to a false impression of better accuracy of the neural  
572    networks. We therefore recommend performing case-specific residual analyses on the neural  
573    network predictions and the MLE estimates, which are often overlooked or over-simplified.

574        We propose two strategies to minimize the influence of possible under-representing  
575    training dataset. The first strategy is to estimate parameters using MLE (if possible) and  
576    then training neural networks with a dataset generated by true parameter values that  
577    cover the MLE estimates, if the estimates seem reasonable. When MLE does not exist, or  
578    the estimates seem unrealistic, we can use the second strategy, that is, to train the neural  
579    networks and predict parameters on a relatively narrow training dataset, then retrain on a  
580    broader dataset generated from a parameter space with different means. We can examine if  
581    our predictions are subject to the range of the training dataset by observing whether the  
582    prediction changes. Generally, we recommend to train the neural networks with as large a

36

TIANJIAN QIN, KOEN J. VAN BENTHEM, LUIS VALENTE, RAMPAL S. ETIENNE

583 dataset (sample size) and as broad a parameter space as possible.

584 Improving neural network predictions that are close to the mean is unlikely to be  
585 achieved by increasing the amount of training data: we did not observe major performance  
586 improvement when changing the size of the datasets (from 1,000 to 100,000 phylogenies  
587 per dataset). Instead, one might consider increasing the complexity of the network  
588 architecture, such as increasing their depth or adapting the scale of the hidden nodes  
589 (Zhang et al., 2021), but note that this may require larger data.

590 Although potentially beneficial, increasing the depth can also harm predictive  
591 power. In particular for GNNs, increasing their depth may lead to "over-smoothing" and  
592 "over-squashing". Over-smoothing causes node features to become increasingly similar as  
593 more layers are added (Li et al., 2018), leading to a loss of distinct node embeddings across  
594 different clusters. Over-squashing involves the compression of expansive node information  
595 through bottleneck edges into a fixed-size vector, which is problematic in graphs with large  
596 diameters and long-range dependencies (Alon and Yahav, 2021), e.g. phylogenies. Both  
597 issues degrade node representations and distort information flow, making deeper GNNs  
598 potentially less effective than shallower ones (Dwivedi et al., 2022). Moreover,  
599 over-smoothing and over-squashing are intrinsically linked, creating a trade-off that cannot  
600 be easily resolved (Giraldo et al., 2023).

601 In our analyses, we observed that increasing the number of GraphSAGE layers  
602 beyond three in the differentiable pooling architecture destabilized the training process  
603 and reduced the accuracy of estimates on validation datasets, introducing more outliers.  
604 We therefore opted to maintain two layers throughout our study. We explored newer  
605 algorithms designed to mitigate deep GNN issues (Chen et al., 2020; Gravina et al., 2022;  
606 Li et al., 2018), but found that these deeper architectures performed worse than our  
607 differentiable pooling approach with fewer layers. For DNN and LSTM, we also  
608 experimented with more complex architectures, different activation functions and various  
609 hyper-parameter optimizations but failed to achieve better performance.

610 *Fundamental Problems with Phylogenies*

611 The lack of improvement when changing the amount of training data or the  
612 network architecture suggests that the real challenges of estimating parameters might not  
613 lie in the architecture of the networks, but might instead be attributed to underlying weak  
614 or absent phylogenetic signals. Whenever this is the case, we expect similarities in  
615 inaccuracies of both MLE and neural network approaches. This occurs, for example, for  
616 the carrying capacity when it is high and thus has a weak effect (measured by  $(\lambda - \mu)/K$ ).  
617 Here, the phylogeny is typically not near the carrying capacity, allowing the number of  
618 species to grow (almost) unbounded. This may result in carrying capacity estimates that  
619 are arbitrarily high, especially in the MLE methods. The PBD scenario is known to  
620 present difficulties in recovering parameters reliably with MLE (Etienne et al., 2014) and  
621 we find similar poor performance with neural networks. A second case where accurate  
622 parameter estimation is complicated occurs when extinction processes erase critical  
623 information (Louca and Pennell, 2021), as observed in the decline of estimation robustness  
624 associated with increasing extinction rate.

625 More generally, small phylogenies tend to contain less information than large ones.  
626 In our results we see that estimation accuracy and robustness decline with decreasing size  
627 of the phylogenies. This trend is observed across both MLE and neural network methods.  
628 In the BD and PBD scenarios, where datasets have greater variability in phylogeny sizes,  
629 poor estimations for small trees could be explained by both low information content, or  
630 under-representation of such trees. To account for the potential effects of under- and  
631 over-representation of phylogenies of different sizes in our datasets, we conducted a  
632 supplementary study to explore whether the patterns we observed persist in a dataset with  
633 a re-balanced distribution of phylogeny sizes (see Appendix J, Figure 24). This shows the  
634 same patterns and they are therefore unlikely to be a result of under- or  
635 over-representation of different phylogeny sizes, and instead reflect low information content  
636 of small trees (compare Figure 18 and Figure 24).

637 Low information content for specific parameters is expected to be more likely as the  
638 complexity of the underlying diversification models increases (see the results of BD in  
639 Appendix H, the results of DDD and the results of PBD in Appendix I, from BD to DDD  
640 to PBD, the complexity increases). This difficulty may stem from the increasingly complex  
641 and substantial signals that the phylogenies are required to convey, which may not be fully  
642 captured in the stochastically generated data. When applying these methods to empirical  
643 phylogenies, there is a noticeable decline in the agreement between MLE and neural  
644 network estimates from the BD scenario to the DDD scenario.

645 *Confronting the Empirical Phylogenies*

646 The processes of evolution within natural systems are often unknown. Determining  
647 the "true parameters" of an empirical phylogeny is challenging, even when they meet  
648 theoretical assumptions, making it difficult to evaluate which tool provides more accurate  
649 estimates. Therefore, choosing the right tool is crucial.

650 With neural networks, it is possible that the true parameter value is not part of the  
651 assumed parameter range for simulating the training data. In such cases, neural network  
652 accuracy decreases notably, as shown in a supplementary study (explained in  
653 Appendix K). We also noticed that when comparing the estimates of MLE and neural  
654 network methods on the empirical phylogenies (see Figure 9, Figure 10), MLE estimates  
655 spread wider than the neural networks (e.g. our BD training dataset comprises phylogenies  
656 simulated using speciation rate between 0 and 0.8 and extinction rate between 0 and 0.72,  
657 our neural networks never predict speciation rate larger than 0.8 or extinction rate larger  
658 than 0.72, see Figure 10, similar results can be found under the DDD scenario in Figure 9).  
659 Expanding the training dataset's parameter space can resolve the generalization issue (we  
660 expanded our training datasets several times in the experiments), but this approach  
661 requires significantly more computational resources for both simulation and training of the  
662 neural networks.

663 Our supplementary study (explained in Appendix K) also reveals that neural  
664 networks tend to provide more accurate estimates of speciation and extinction rates from  
665 complete phylogenies than from extant ones under the BD scenario. This increase in  
666 accuracy was not observed in the DDD scenario. While complete phylogenies offer a  
667 broader picture and more contextual information, obtaining them is challenging because it  
668 is nearly impossible to account for all extinct species.

669 Our analyses indicate that GNN is more robust but more prone to systematic errors  
670 (GNN achieved the greatest possible robustness in estimating the speciation rate and  
671 carrying capacity among neural network methods). We show that using GNN as a base  
672 and other neural networks like LSTM to enhance GNN might effectively combine the  
673 advantages of different methods and information sources, thus strengthening overall  
674 generalization ability. Our boosting methods (e.g. Boost BT) perform the best in this  
675 context.

676 In conclusion, when applied with caution, neural network methods can be applied  
677 to other diversification scenarios where MLE is absent or non-tractable, as our  
678 best-performing neural network method showed comparable or even better performance to  
679 the best-case MLE. Our neural networks particularly perform better than MLE in terms of  
680 accuracy and robustness on small phylogenies and can be significantly faster when  
681 estimating very large phylogenies. Thus, if properly trained, neural network methods may  
682 substitute for or at least cross-reference with MLE estimates where they exist.

683

## ACKNOWLEDGEMENTS

684 We thank the Center for Information Technology of the University of Groningen for  
685 their support and for providing access to the Hábrók high performance computing cluster.

40

REFERENCES

686 FUNDING

687 Tianjian Qin was funded by a joint scholarship program of China Scholarship  
688 Council and University of Groningen. Luis Valente was funded by a NWO VIDI grant.

689 REFERENCES

690 Alexander HK, Lambert A, Stadler T. 2016. Quantifying Age-dependent Extinction from  
691 Species Phylogenies. *Systematic Biology*. 65:35–50.

692 Alon U, Yahav E. 2021. On the Bottleneck of Graph Neural Networks and its Practical  
693 Implications. *ArXiv:2006.05205*.

694 Ardia D, Boudt K, Carl P, Mullen K, Peterson BG. 2010. Differential Evolution with  
695 DEoptim: An Application to Non-Convex Portfolio Optimization. *SSRN:1584905*.

696 Beaumont MA. 2010. Approximate Bayesian Computation in Evolution and Ecology.  
697 Annual Review of Ecology, Evolution, and Systematics. 41:379–406.

698 Beaumont MA, Zhang W, Balding DJ. 2002. Approximate Bayesian Computation in  
699 Population Genetics. *Genetics*. 162:2025–2035.

700 Bokma F. 2010. Time, Species, and Separating Their Effects on Trait Variance in Clades.  
701 *Systematic Biology*. 59:602–607.

702 Charu C A. 2018. Neural networks and deep learning: a textbook. Springer.

703 Chen M, Wei Z, Huang Z, Ding B, Li Y. 2020. Simple and Deep Graph Convolutional  
704 Networks. *ArXiv:2007.02133v1*.

705 Condamine FL, Rolland J, Morlon H. 2019. Assessing the Causes of Diversification  
706 Slowdowns: Temperature-Dependent and Diversity-Dependent Models Receive  
707 Equivalent Support. *Ecology Letters*. 22:1900–1912.

708 Dwivedi VP, Rampáek L, Galkin M, Parviz A, Wolf G, Luu AT, Beaini D. 2022. Long  
709 range graph benchmark. *Advances in Neural Information Processing Systems*.  
710 35:22326–22340.

711 Etienne RS, Haegeman B, Stadler T, Aze T, Pearson PN, Purvis A, Phillimore AB. 2012.  
712 Diversity-Dependence Brings Molecular Phylogenies Closer to Agreement with the Fossil  
713 Record. *Proceedings of the Royal Society B: Biological Sciences*. 279:1300–1309.

714 Etienne RS, Morlon H, Lambert A. 2014. Estimating the Duration of Speciation from  
715 Phylogenies. *Evolution*. 68:2430–2440.

716 Etienne RS, Pigot AL, Phillimore AB. 2016. How Reliably Can We Infer  
717 Diversity-Dependent Diversification from Phylogenies? *Methods in Ecology and*  
718 *Evolution*. 7:1092–1099.

719 Etienne RS, Rosindell J. 2012. Prolonging the Past Counteracts the Pull of the Present:  
720 Protracted Speciation Can Explain Observed Slowdowns in Diversification. *Systematic*  
721 *Biology*. 61:204.

722 Fey M, Lenssen JE. 2019. Fast Graph Representation Learning with PyTorch Geometric.

723 Foote M. 1997. Estimating Taxonomic Durations and Preservation Probability.  
724 *Paleobiology*. 23:278–300.

725 Giraldo JH, Skianis K, Bouwmans T, Malliaros FD. 2023. On the Trade-off between  
726 Over-smoothing and Over-squashing in Deep Graph Neural Networks. In: *Proceedings*  
727 of the 32nd ACM International Conference on Information and Knowledge Management.  
728 Birmingham United Kingdom: ACM. p. 566–576. doi:10.1145/3583780.3614997.

729 Graczyk M, Lasota T, Trawiski B, Trawiski K. 2010. Comparison of Bagging, Boosting  
730 and Stacking Ensembles Applied to Real Estate Appraisal. In: D Hutchison, T Kanade,  
731 J Kittler, JM Kleinberg, F Mattern, JC Mitchell, M Naor, O Nierstrasz,  
732 C Pandu Rangan, B Steffen, M Sudan, D Terzopoulos, D Tygar, MY Vardi, G Weikum,

733 NT Nguyen, MT Le, J witek, editors. Intelligent Information and Database Systems.  
734 volume 5991. Berlin, Heidelberg: Springer Berlin Heidelberg. p. 340–350.  
735 doi:10.1007/978-3-642-12101-2\_35. Series Title: Lecture Notes in Computer Science.

736 Gravina A, Bacciu D, Gallicchio C. 2022. Anti-Symmetric DGN: a Stable Architecture for  
737 Deep Graph Networks. The Eleventh International Conference on Learning  
738 Representations.

739 Hamilton WL, Ying R, Leskovec J. 2017. Inductive Representation Learning on Large  
740 Graphs. ArXiv:1706.02216v4.

741 Hendrycks D, Gimpel K. 2016. Gaussian Error Linear Units (GELUs).  
742 ArXiv:1606.08415v5.

743 Hey J. 1992. Using Phylogenetic Trees to Study Speciation and Extinction. Evolution.  
744 46:627–640.

745 Huber PJ. 1992. Robust Estimation of a Location Parameter. In: Breakthroughs in  
746 Statistics. New York, NY: Springer New York. p. 492–518.

747 Imambi S, Prakash KB, Kanagachidambaresan GR. 2021. PyTorch. In: KB Prakash,  
748 GR Kanagachidambaresan, editors. Programming with TensorFlow. Cham: Springer  
749 International Publishing. p. 87–104. doi:10.1007/978-3-030-57077-4\_10. Series Title:  
750 EAI/Springer Innovations in Communication and Computing.

751 Ioffe S, Szegedy C. 2015. Batch Normalization: Accelerating Deep Network Training by  
752 Reducing Internal Covariate Shift. ArXiv:1502.03167v3.

753 Janzen T, Höhna S, Etienne RS. 2015. Approximate Bayesian Computation of  
754 diversification rates from molecular phylogenies: introducing a new efficient summary  
755 statistic, the nLTT. Methods in Ecology and Evolution. 6:566–575.

756 Kidwell SM, Flessa KW. 1996. The Quality of the Fossil Record: Populations, Species, and  
757 Communities. Annual Review of Earth and Planetary Sciences. 24:433–464.

758 Kipf TN, Welling M. 2016. Semi-Supervised Classification with Graph Convolutional  
759 Networks. ArXiv:1609.02907v4.

760 Kutsukake N, Innan H. 2013. Simulation-Based Likelihood Approach for Evolutionary  
761 Models of Phenotypic Traits on Phylogeny. Evolution. 67:355–367.

762 Lagarias JC, Reeds JA, Wright MH, Wright PE. 1998. Convergence Properties of the  
763 NelderMead Simplex Method in Low Dimensions. SIAM Journal on Optimization.  
764 9:112–147. \_eprint: <https://doi.org/10.1137/S1052623496303470>.

765 Lajaaiti I, Lambert S, Voznica J, Morlon H, Hartig F. 2023. A Comparison of Deep  
766 Learning Architectures for Inferring Parameters of Diversification Models from Extant  
767 Phylogenies. doi:10.1101/2023.03.03.530992. Pages: 2023.03.03.530992 Section: New  
768 Results.

769 Lambert S, Voznica J, Morlon H. 2023. Deep Learning from Phylogenies for Diversification  
770 Analyses. Systematic Biology. 72:1262–1279.

771 Li G, Xiong C, Thabet A, Ghanem B. 2020. DeeperGCN: All You Need to Train Deeper  
772 GCNs. ArXiv:2006.07739v1.

773 Li Q, Han Z, Wu XM. 2018. Deeper Insights into Graph Convolutional Networks for  
774 Semi-Supervised Learning. In: Proceedings of the AAAI conference on artificial  
775 intelligence. volume 32. Issue: 1.

776 Loshchilov I, Hutter F. 2017. Decoupled Weight Decay Regularization.  
777 ArXiv:1711.05101v3.

778 Louca S, Pennell MW. 2021. Why extinction estimates from extant phylogenies are so  
779 often zero. Current Biology. 31:3168–3173. e4. Type: Journal Article.

780 Luebke D. 2008. CUDA: Scalable parallel programming for high-performance scientific  
781 computing. In: 2008 5th IEEE international symposium on biomedical imaging: from  
782 nano to macro. IEEE. p. 836–838.

783 Moi D, Dessimoz C. 2022. Reconstructing Protein Interactions across Time Using  
784 Phylogeny-Aware Graph Neural Networks. doi:10.1101/2022.07.21.501014. Pages:  
785 2022.07.21.501014 Section: New Results.

786 Morgan SL, Deming SN. 1974. Simplex Optimization of Analytical Chemical Methods.  
787 Analytical Chemistry. 46:1170–1181.

788 Morlon H. 2014. Phylogenetic Approaches for Studying Diversification. Ecology Letters.  
789 17:508–525.

790 Nee S. 2001. Inferring Speciation Rates from Phylogenies. Evolution. 55:661–668.

791 Nee S, Holmes EC, May RM, Harvey PH. 1994. Extinction Rates Can Be Estimated from  
792 Molecular Phylogenies. Philosophical Transactions of the Royal Society of London.  
793 Series B: Biological Sciences. 344:77–82.

794 Nee S, May RM, Harvey PH. 1997. The reconstructed evolutionary process. Philosophical  
795 Transactions of the Royal Society of London. Series B: Biological Sciences. 344:305–311.  
796 Publisher: Royal Society.

797 Paradis E, Schliep K. 2019. ape 5.0: an Environment for Modern Phylogenetics and  
798 Evolutionary Analyses in R. Bioinformatics. 35:526–528.

799 Python W. 2021. Python. Python releases for windows. 24. Publisher: Citeseer.

800 Qin T. 2023. eveGNN - Codebase for Phylogenetic Tree Parameter Estimation with Neural  
801 Networks. <Https://github.com/EvoLandEco/eveGNN>.

802 Qin T. 2024. EvoNN - Neural Networks for Evolution.  
803 <Https://github.com/EvoLandEco/EvoNN>.

804 R Core Team R. 2013. R: A language and environment for statistical computing.  
805 Publisher: Vienna, Austria.

REFERENCES

45

806 Rabosky DL. 2009. Heritability of Extinction Rates Links Diversification Patterns in  
807 Molecular Phylogenies and Fossils. *Systematic Biology*. 58:629–640.

808 Rampáek L, Galkin M, Dwivedi VP, Luu AT, Wolf G, Beaini D. 2022. Recipe for a  
809 General, Powerful, Scalable Graph Transformer. *ArXiv*:2205.12454v4.

810 Reiman D, Metwally AA, Sun J, Dai Y. 2020. PopPhy-CNN: A Phylogenetic Tree  
811 Embedded Architecture for Convolutional Neural Networks to Predict Host Phenotype  
812 From Metagenomic Data. *IEEE Journal of Biomedical and Health Informatics*.  
813 24:2993–3001. Conference Name: *IEEE Journal of Biomedical and Health Informatics*.

814 Rosindell J, Cornell SJ, Hubbell SP, Etienne RS. 2010. Protracted speciation revitalizes  
815 the neutral theory of biodiversity. *Ecology Letters*. 13:716–727. *eprint*:  
816 <https://onlinelibrary.wiley.com/doi/pdf/10.1111/j.1461-0248.2010.01463.x>.

817 Rowan TH. 1990. Functional Stability Analysis of Numerical Algorithms. The University  
818 of Texas at Austin.

819 Sak H, Senior A, Beaufays F. 2014. Long Short-Term Memory Based Recurrent Neural  
820 Network Architectures for Large Vocabulary Speech Recognition. *ArXiv*:1402.1128v1.

821 Salehinejad H, Sankar S, Barfett J, Colak E, Valaee S. 2017. Recent Advances in  
822 Recurrent Neural Networks. *ArXiv*:1801.01078v3.

823 Stadler T. 2011. Simulating Trees with a Fixed Number of Extant Species. *Systematic  
824 Biology*. 60:676–684.

825 Valente LM, Phillimore AB, Etienne RS. 2015. Equilibrium and Non-Equilibrium  
826 Dynamics Simultaneously Operate in the Galápagos Islands. *Ecology Letters*.  
827 18:844–852.

828 Voznica J, Zhukova A, Boskova V, Saulnier E, Lemoine F, Moslonka-Lefebvre M, Gascuel  
829 O. 2022. Deep learning from phylogenies to uncover the epidemiological dynamics of  
830 outbreaks. *Nature Communications*. 13:3896. Publisher: Nature Publishing Group.

831 Wagner PJ. 2000. Phylogenetic Analyses and the Fossil Record: Tests and Inferences,  
832 Hypotheses and Models. *Paleobiology*. 26:341–371.

833 Ward EJ. 2008. A Review and Comparison of Four Commonly Used Bayesian and  
834 Maximum Likelihood Model Selection Tools. *Ecological Modelling*. 211:1–10.

835 Xie S, Valente L, Etienne RS. 2023. Can We Ignore Trait-Dependent Colonization and  
836 Diversification in Island Biogeography? *Evolution*. 77:670–681.

837 Ying Z, You J, Morris C, Ren X, Hamilton W, Leskovec J. 2018. Hierarchical Graph  
838 Representation Learning with Differentiable Pooling. In: *Advances in Neural*  
839 *Information Processing Systems*. volume 31. Curran Associates, Inc.

840 Zhang W, Sheng Z, Jiang Y, Xia Y, Gao J, Yang Z, Cui B. 2021. Evaluating Deep Graph  
841 Neural Networks. *ArXiv*:2108.00955v1.

842 Zhu H, Akroud M, Zheng B, Pelegris A, Jayarajan A, Phanishayee A, Schroeder B,  
843 Pekhimenko G. 2018. Benchmarking and Analyzing Deep Neural Network Training. In:  
844 *2018 IEEE International Symposium on Workload Characterization (IISWC)*. p. 88–100.  
845 doi:10.1109/IISWC.2018.8573476.

846

## APPENDIX

847

### A. DATA TRANSFORMATION PROTOCOL

848

#### *Protocol Transforming Phylogenies for Graph Neural Network*

849

Phylogenetic trees are usually stored in the "phylo" data format in R. This data format is not directly compatible with GNN implementations. To facilitate graph convolutional operations, we transformed phylogeny from a "phylo" object into three major components: adjacency list, node feature matrix and graph attributes (see Figure 2). The adjacency list contains information on the connectivity between nodes and tips, the node feature matrix contains distances between nodes and tips, and the graph-level attributes include the true initial values to generate the phylogenies. These components are stored in separate tensors. In machine learning, a tensor is a mathematical object that generalizes scalars, vectors, and matrices to higher dimensions, allowing complex operations to be performed efficiently on multi-dimensional arrays.

859

*Adjacency List* In the context of a phylogenetic tree, tip nodes usually represent taxonomic units such as species, while root nodes and internal nodes represent the points where two taxonomic units depart from each other. An edge in a phylogenetic tree represents the connection between two nodes, and as such describes the evolutionary relatedness between taxa. Each root node, internal node and tip node in an R "phylo" object is indexed sequentially, each edge is also sequentially indexed independently of node indices. The sub-list "edge" of a "phylo" object contains the adjacency list of a phylogenetic tree which describes the relationships between nodes. Each row of the adjacency list represents an edge, the first column contains the index (or numbering) of the ancestor node, and the second column contains the index of the descendant node.

869

870

871

This data structure effectively captures the tree's branching pattern, showing how each taxon (or node) is connected to others. The adjacency list in "phylo" object uses a "1-based" indexing in R, we therefore element-wise deduct 1 from the list to convert it into

872 "0-based" indexing which is compatible with the python environment.

873 We output the converted adjacency list within the "phylo" object as the adjacency  
874 list  $\mathcal{E}$  of the graph representation, in PyTorch Geometric, which is conventionally named as  
875 "data.edge\_index". We store  $\mathcal{E}$  as a "torch.long" long integer type tensor and transpose it  
876 such that it has shape  $[2, \text{num\_edges}]$ , where "num\_edges" is the number of edges in the  
877 "phylo" object. This tensor has two dimensions. This way, the connections between nodes  
878 in the transformed graph are all single-directional, from the ancestor nodes to their  
879 descendants (if any). Training the GNN with graphs of non-directed edges gives no  
880 performance advantage, according to our tests in phylogenetic tree parameter estimation  
881 tasks. Single-directional data structure can save GPU memory and reduce the computation  
882 complexity.

883 *Node Feature Matrix* In a "phylo" object in R, the "edge.length" sub-list defines the  
884 lengths of the edges in the phylogenetic tree. In a phylogenetic context, these lengths often  
885 correspond to evolutionary distances, time, or genetic change. "edge.length" is a numeric  
886 vector where each element corresponds to the length of the edge as defined in the  
887 adjacency list. The order of lengths in the "edge.length" vector aligns with the order of  
888 edges in the adjacency list.

889 For each tree, we aggregate information contained in "edge.length" to a node feature  
890 matrix. Each row of the matrix represents features contained in a node. The first column  
891 contains the edge length from a node to its direct ancestor node, the second and the third  
892 columns contain the edge lengths from a node to its two daughter nodes. We pad the row  
893 of the root node with an 0 in the first column as it has no ancestor. We also pad the rows of  
894 the tip nodes with two 0s in the second and the third columns as they have no descendants.  
895 The row order of feature matrix aligns with the order of edges in the adjacency list.

896 We output the node feature matrix of each tree as the node feature matrix  $\mathcal{X}$  of the  
897 graph representation, in PyTorch Geometric, this is conventionally named as "data.x". We  
898 store  $\mathcal{X}$  as a "torch.float" floating point type tensor, it has shape

899 [num\_nodes, num\_node\_features], where "num\_nodes" is the number of nodes  
900 (including tip nodes) in the "phylo" object and "num\_node\_features" in our case is 3, i.e.  
901 the phylogenetic distances from a node to its ancestor (if any) and two descendants (if  
902 any). This tensor has two dimensions. We do not store the phylogenetic distance  
903 information in edge features because GCN operators will eventually pass and aggregate the  
904 edge features into each of the node. Our data structure is simpler and so is the GNN  
905 architecture.

906 *Graph-Level Attributes as Training Targets* We store all the parameters used to  
907 simulate a tree (ground truth values) in the graph-level attributes  $\mathcal{Y}$ . These can have  
908 arbitrary length, which should be consistent with the number of the parameters to be  
909 estimated (the three diversification scenarios, BD, DDD and PBD, have different number  
910 of parameters). We store graph-level attributes as a "torch.float" floating point type tensor  
911 with length of the number of parameters we want to predict for each type of the  
912 phylogenetic tree. In PyTorch Geometric, graph-level attributes can be named as "data.y".  
913 The graph-level attributes are used as training targets to compute loss (see Appendix B for  
914 the definition of loss).

915 *Protocol Transforming Summary Statistics for Dense Neural Network*

916 The summary statistics of a phylogeny are represented by a 1D vector, so the  
917 protocol for DNN is straightforward: we convert the vector into a tensor containing  
918 floating type data, with the shape [num\_stats], where "num\_stats" denotes the total  
919 number of statistics. This tensor has only one dimension. This conversion guarantees that  
920 each tensor is associated with its respective tree, with all contained statistics maintaining  
921 their original order. Within the PyTorch Geometric framework, these statistics are  
922 encapsulated as "data.stats" for each tree. When using DNN alone to estimate parameters  
923 from the summary statistics, the ground truth values of the parameters of the trees are

924 stored in the same way as the graph-level attributes, as model training targets. When  
925 using DNN with other neural networks (e.g. in stacking and boosting strategies), they  
926 share the same ground truth values which are the graph-level attributes.

927 *Protocol Transforming Branching Times for Recurrent Neural Network*

928 To address the varying lengths in branching times across different phylogenetic  
929 trees, we standardize these sequences by padding them to match the length of the longest  
930 branching time sequence. This is achieved by appending zeros to the shorter sequences  
931 until they match the predefined maximum length. The padded sequences are stored in  
932 tensors containing floating type data. As the original branching times do not contain zero  
933 values, this padding strategy allows us to distinguish between original data and padding.  
934 Consequently, we can pass masks of the sequences to the LSTM, which indicates the  
935 positions of the paddings, making LSTM concentrate only on the informative portions of  
936 the sequences, thereby optimizing its performance. When using LSTM alone to estimate  
937 parameters from the branching times, the ground truth values of the parameters of the  
938 trees are stored in the same way as the graph-level attributes, as model training targets.  
939 When using LSTM with other neural networks (e.g. in stacking and boosting strategies),  
940 they share the same ground truth values which are the graph-level attributes.

941

## B. TOTAL LOSS

942 Total loss comprises three key components: Huber loss, link prediction loss and  
943 entropy of regularization. Huber loss was used for optimizing regression accuracy while the  
944 remaining components focused on alleviating a possible issue where GNN can be hard to  
945 train, if incorporating the differentiable pooling method (Ying et al., 2018).

946 The Huber loss (Huber, 1992) for vectors  $y$  and  $\hat{y}$ , each with  $n$  elements, computed  
947 as the average loss across all elements, is given by:

$$L_\delta(\mathbf{y}, \hat{\mathbf{y}}) = \frac{1}{n} \sum_{i=1}^n \begin{cases} \frac{1}{2}(y_i - \hat{y}_i)^2 & \text{for } |y_i - \hat{y}_i| \leq \delta, \\ \delta(|y_i - \hat{y}_i| - \frac{1}{2}\delta) & \text{otherwise,} \end{cases} \quad (\text{B.1})$$

948 where  $\mathbf{y}$  is the true value vector comprising the ground truth parameters used for  
949 simulating a phylogenetic tree,  $\hat{\mathbf{y}}$  is the predicted value vector comprising the parameter  
950 predictions,  $y_i$  and  $\hat{y}_i$  are the  $i$ -th elements of  $\mathbf{y}$  and  $\hat{\mathbf{y}}$  respectively,  $n$  is the number of  
951 elements in the vectors  $\mathbf{y}$  and  $\hat{\mathbf{y}}$  and  $\delta$  is the threshold parameter that defines the  
952 transition from squared to linear loss (here loss refers to the difference between ground  
953 truth and predicted values). In our research, we set  $\delta = 0.8$  for all the training sessions,  
954 making the neural networks more sensitive to smaller errors and more robust to outliers .

955 The total loss  $L$  is given by

$$L = L_\delta(\mathbf{y}, \hat{\mathbf{y}}) + L_{\text{LP}} + L_{\text{E}}, \quad (\text{B.2})$$

956 where  $L_{\text{LP}}$  is the link prediction loss and  $L_{\text{E}}$  is the entropy of regularization, see  
957 Ying et al. (2018) for their definitions.

958 C. NEURAL NETWORK ARCHITECTURE

959 For the graph neural network, we used GraphSAGE (Hamilton et al., 2017), a  
960 sample-and-aggregate graph convolutional neural network, to capture a graph-level  
961 representation. GraphSAGE has achieved strong performance of learning from large  
962 graphs. We use graph neural network (GNN) to refer to the graph neural network  
963 approach which incorporates GraphSAGE.

964 GNN is mainly assembled from five GNN modules (see Figure 1-C for five blocks of  
965 boxes in yellow and orange colors). Each module comprises the same number of  
966 GraphSAGE operators (Hamilton et al., 2017), where the number of layers (GraphSAGE  
967 operators, as illustrated by the number of combined boxes within each GNN modules in  
968 Figure 1-C)  $N_L = 1, 2, \dots, 6$ . Each GNN operator is accompanied by a Batch  
969 Normalization for 1D Inputs (BatchNorm1d, not shown in Figure 1) operator (Ioffe and  
970 Szegedy, 2015) and then a Gaussian Error Linear Units (GELU, as illustrated by the  
971 orange bands within the yellow boxes in Figure 1-C) activation function (Hendrycks and  
972 Gimpel, 2016). The GraphSAGE operators facilitate the convolution operation over  
973 graphs, capturing both local node features and their neighborhood information. The  
974 BatchNorm1d operator is commonly employed in neural networks to stabilize and  
975 accelerate the training process. The GELU activation layer is used for introducing  
976 non-linearity into the data. Learned features from all the GraphSAGE operators within a  
977 module are collected and concatenated. Larger  $N_L$  will result in the GNN modules to  
978 aggregate information into each node from its more distantly connected neighbors.  
979 According to our experiments, the optimal case is  $N_L = 2$ , all figures and results relating  
980 to GNN were reported on the optimal case.

981 The graph-learning process also involves graph coarsening operations. We  
982 incorporated the differentiable pooling (DiffPool hereafter) technique to better learn  
983 hierarchical representations of the graphs. DiffPool can aggregate graph nodes into clusters  
984 after each operation. It facilitates graph coarsening and captures intricate hierarchical

985 structure, which makes it particularly suitable for graph-level tasks (Ying et al., 2018). In  
986 the first coarsening operation, the graph data inputs are passed to two GNN modules  
987 (pooling and embedding, see Figure 1-C for the blocks marked as "GNN pool1" and "GNN  
988 embed1"). The pooling group reduces the graph size, while the embedding group captures  
989 the node features. The filtered data from each GraphSAGE operator are concatenated (see  
990 Figure 1-C for the blocks of boxes marked as "concat1") then passed to a DiffPool layer  
991 (see Figure 1-C for the red box marked as "diff-pool1"), which finalizes the first coarsening  
992 operation. The second coarsening operation is applied in the same way as the first (as  
993 represented by "GNN pool1", "GNN embed2", "concat2" in Figure 1-C), and the outputs  
994 from the second DiffPool layer ("diff-pool2" in Figure 1-C) are passed to the final (fifth)  
995 GNN module ("GNN embed3" in Figure 1-C). The nodes in a graph are dynamically  
996 clustered and reduced after each coarsening operation. The coarsening ratio at each  
997 operation is determined by a pre-set DiffPool pooling ratio. Let  $N_{\text{coarsened}}$  represent the  
998 number of nodes in the coarsened graph and  $N_{\text{original}}$  the number of nodes in the original  
999 graph. The DiffPool pooling ratio  $\rho_{\text{pool}}$  is given by  $\rho_{\text{pool}} = \frac{N_{\text{coarsened}}}{N_{\text{original}}}$ . Throughout the study,  
1000 we used a manually optimized value  $\rho_{\text{pool}} = 0.25$ . This is a manually optimized  
1001 hyper-parameter.

1002 After the final GNN module, the outputs are concatenated ("concat3" in  
1003 Figure 1-C) and transformed by a global mean pooling operation (red ball "M" in  
1004 Figure 1-C) to create a final graph representation. This graph representation is passed to a  
1005 readout layer group ("readout" as represented by light blue boxes in Figure 1-C) consisting  
1006 of two linear layers to perform graph-level regression which ultimately outputs a vector of  
1007  $n$  predicted parameters ("pred" as represented by a purple box in Figure 1-C). Only the  
1008 first linear layer is followed by GELU (see the orange band of the first linear layer). All the  
1009 linear layers incorporate dropout operations with a pre-set dropout ratio to prevent  
1010 over-fitting and to utilize as many neuron connections as possible. Let  $\rho_{\text{dropout}}$  represent  
1011 the probability  $p$  of disabling a connection between an input node and a hidden node of a

1012 linear layer in each epoch. The dropout ratio  $\rho_{\text{dropout}}$  is simply given by  $\rho_{\text{dropout}} = p$ .  
1013 Throughout the study, we used a commonly picked value  $\rho_{\text{dropout}} = 0.5$ . This is a manually  
1014 optimized hyper-parameter.

1015 DNN's major component is a stack comprises 5 linear layers ("DNN stack" in  
1016 Figure 1-A), each followed by a BatchNorm1D (not shown in figure) and a GELU (the  
1017 orange band within the boxes). All the linear layers within the stack incorporate dropout  
1018 operations with  $\rho_{\text{dropout}} = 0.5$ . Learned features from all the linear layers within the stacks  
1019 are collected and concatenated ("concat" in Figure 1-A). A single linear readout layer  
1020 ("readout" in Figure 1-A) outputs  $n$  predicted parameters ("pred" in Figure 1-A).  
1021 According to our experiments, stacking more linear layers gives no substantial  
1022 improvement to the performance.

1023 LSTM's major component is a stack of 5 LSTM recurrent neural network layers  
1024 ("LSTM stack" in Figure 1-B). The final hidden state from the last recurrent neural  
1025 network layer is processed by a linear layer with  $\rho_{\text{dropout}} = 0.5$  accompanied by a GELU  
1026 ("linear" in Figure 1-B), then passed to a single linear readout layer ("readout" in  
1027 Figure 1-B) that outputs  $n$  predicted parameters ("pred" in Figure 1-B). According to our  
1028 experiments, stacking more recurrent neural network layers provides no substantial  
1029 improvement to the performance.

1030 The hyper-parameters not mentioned are set by their default values. The  
1031 dimensions of the boxes do not map to any hyper-parameter settings, they are set for the  
1032 best visual effect. The values below the boxes indicate their respective number of hidden  
1033 neurons, their input and output neurons are not shown in the figure, they can be found in  
1034 the configuration files in our GitHub repository "eveGNN" (Qin, 2023).

1035

## D. ENSEMBLE LEARNING

1036 With bagging, we trained GNN, DNN and LSTM independently ("GNN", "DNN"  
1037 and "LSTM" blocks of boxes in Figure 3-Bagging), translated their outputs to parameter  
1038 predictions through their own readout layers (three "readout" boxes next to the neural  
1039 networks and three "pred" boxes next to the readout layers in Figure 3-Bagging) and then  
1040 aggregated the predictions (red ball "A" in Figure 3-Bagging). We experimented with four  
1041 aggregation methods: taking the mean, median, max and min values among the three  
1042 predictions. We also recorded the individual predictions without aggregation.

1043 With stacking, we trained GNN, DNN and LSTM simultaneously ("GNN", "DNN"  
1044 and "LSTM" blocks of boxes in Figure 3-Stacking) but without their own readout layers.  
1045 We combined the features from DNN, the LSTM's final hidden state, and GNN's graph  
1046 representation and fed to a meta-learner ("meta-learner" in Figure 3-Stacking) comprising  
1047 linear neural network layers that learns to best readout parameter predictions from these  
1048 combined outputs.

1049 With boosting, there can be different pathways. In our illustration, GNN, DNN and  
1050 LSTM were trained sequentially to iteratively correct residuals. For example, firstly, the  
1051 GNN ("GNN" in Figure 3-Boosting) is trained from the graphs to make the initial  
1052 predictions ("readout" and then "pred0" in Figure 3-Boosting) and from predicted and  
1053 ground truth values of the parameters we computed the residuals ("res1" in  
1054 Figure 3-Boosting)secondly, the DNN ("DNN" in Figure 3-Boosting) is trained to predict  
1055 these residuals from the summary statistics ("readout" and then "pred-res1" in  
1056 Figure 3-Boosting), learning to correct the GNN's errorslastly, the LSTM ("LSTM" in  
1057 Figure 3-Boosting) is trained to predict the residuals of the residuals ("readout" and then  
1058 "pred-res2" in Figure 3-Boosting), which is the initial predictions minus the predicted  
1059 residuals by the DNN, from branching times, to further improve the predictive accuracy.  
1060 Finally, we subtracted the two residual terms from the initial predictions (red ball "S" in  
1061 Figure 3-Boosting) to make the corrected predictions.

1062        E. COMPARISON BETWEEN MLE OPTIMIZERS

1063        On the phylogenies from the diversity-dependent diversification (DDD) dataset, we  
1064        compared between three approaches: "Simplex", "Subplex" and "DEoptim". Simplex is a  
1065        derivative-free optimization method that uses a simplex of solutions to iteratively explore  
1066        and adjust within the parameter space, suitable for non-smooth objective functions but  
1067        potentially slow for high-dimensional problems (Morgan and Deming, 1974). Subplex is an  
1068        enhancement of the Simplex method, Subplex breaks high-dimensional optimization into  
1069        smaller subproblems, each optimized using Simplex techniques, providing improved  
1070        efficiency and effectiveness in complex parameter landscapes (Rowan, 1990). DEoptim  
1071        (Differential Evolution) is a more recent population-based algorithm that applies  
1072        evolutionary strategies such as mutation, crossover, and selection to efficiently navigate  
1073        and optimize multimodal and complex objective functions (Ardia et al., 2010).

1074        All three MLE methods encountered consistent optimization challenges, likely due  
1075        to numerical issues related to machine precision limits or unexpected negative values during  
1076        matrix operations. From a random sample of 2000 DDD phylogenies, the completion rates  
1077        for each method were as follows: Simplex achieved 1966 completions from true parameter  
1078        starts and 1910 from random starts; Subplex completed 1681 from true starts and 1612  
1079        from random starts; DEoptim finished 1122 from true starts and 999 from random starts.  
1080        It is more difficult to estimate parameters from random starts, comparing to true starts.

1081        For all the three optimization approaches, -1 will be returned as a parameter  
1082        estimation if the likelihood becomes too small in the searching process. This means that  
1083        the algorithm cannot find optima given the initial starting point of the parameters. It is  
1084        highly possible that the unfinished estimations consisted of inaccurate or even -1 values.  
1085        The comparison between MLE optimizers can be skewed due to less completion rate of the  
1086        Subplex and DEoptim results.

1087        In instances where optimization starting points were randomly set, a significant  
1088        number of outcomes were trapped at local optima, failing to achieve global optima and

1089 often leading to inaccurate parameter estimates. This issue was less prevalent when  
1090 starting points were the true parameters. For visual reference, see Figure 11 and Figure 12.  
1091 Notably, in DEoptim’s best-case scenarios, estimation accuracy deteriorated significantly  
1092 on larger phylogenies, as shown in the last row of Figure 12.

1093 In the best cases, all the MLE optimizers are more likely to give accurate  
1094 estimations on larger phylogenies while in the typical cases, larger phylogenies become a  
1095 burden. Nevertheless, the MLE optimizers generally perform better on larger trees. All the  
1096 MLE results showed similar trends of bias. We calculated the strength of the carrying  
1097 capacity effect using the formula  $(\lambda - \mu)/K$ , where  $\lambda$  represents the true speciation rate,  $\mu$   
1098 the true extinction rate, and  $K$  the true carrying capacity. Phylogenies exhibiting a  
1099 stronger carrying capacity effect typically link to more accurate estimates.

1100 In the best-case scenarios, all MLE methods tended to yield more accurate  
1101 estimates on larger phylogenies, while in typical cases, larger phylogenies posed challenges.  
1102 However, all MLE methods generally performed better with larger trees, and all displayed  
1103 similar trends of bias. We calculated the strength of the carrying capacity effect with the  
1104 formula  $(\lambda - \mu)/K$ , where  $\lambda$  is the true speciation rate,  $\mu$  is the true extinction rate, and  
1105  $K$  is the true carrying capacity.

1106 Subplex was the fastest among the tested algorithms, Simplex and DEoptim were  
1107 slower. Simplex, although slower, completed the most computations and did not show a  
1108 definitive performance disadvantage compared to Subplex or DEoptim. For this reason, in  
1109 all comparisons between MLE and neural network methods, we consistently used results  
1110 from the Simplex optimizer due to data coverage and reliability.

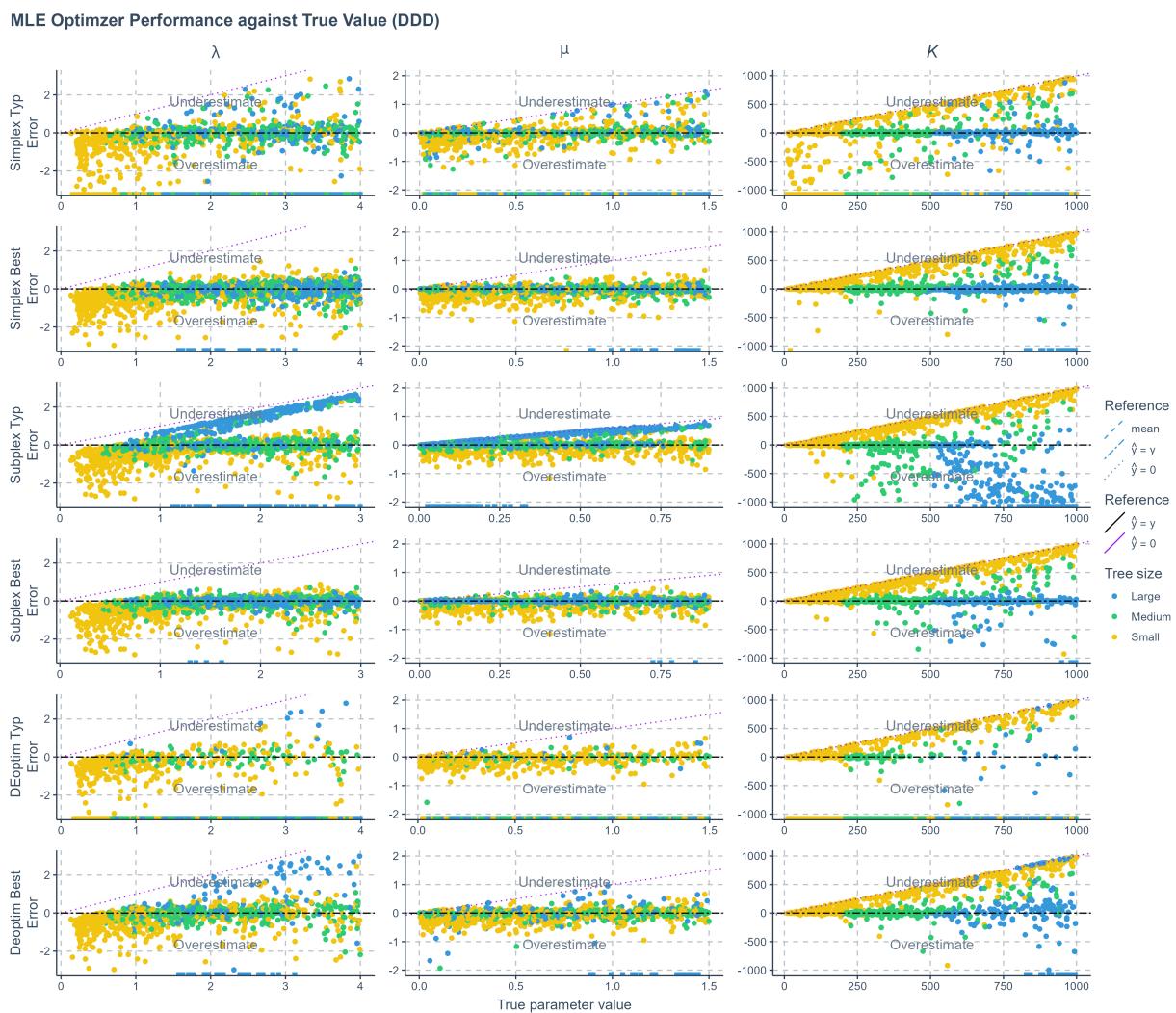


Fig. 11. Error of maximum likelihood estimation using Simplex, Subplex and DEoptim optimizers applied to phylogenies simulated under a diversity-dependent diversification scenario, against true values. For each optimizer there were two cases. The typical case (Typ) refers to using the true parameter values as the starting points for the searching process; the best case (Best) refers to using randomly sampled values from the true parameter space as the starting points. The errors shown (y-axis) are the differences between the true parameters (x-axis) used to simulate the phylogenies and the values estimated by each method. Each row represents a method, and each column corresponds to the results for one specific parameter. Phylogenies are categorized based on their size: yellow for small phylogenies with fewer than 200 nodes (including root, internal, and tip nodes), green for medium-sized phylogenies with 200 to 500 nodes, and blue for large phylogenies with more than 500 nodes. Data points close to purple dotted lines ( $\hat{y} = 0$ ) in MLE result panels indicate near-zero estimates. Black two-dash lines indicate accurate estimates ( $\hat{y} = y$  where  $y$  denotes the true parameter value). Small squares spreading along the x-axis signify optimization failures. Extremely deviating estimates are not shown in the figure.  $\lambda$ : Speciation rate.  $\mu$ : extinction rate.  $K$ : carrying capacity.

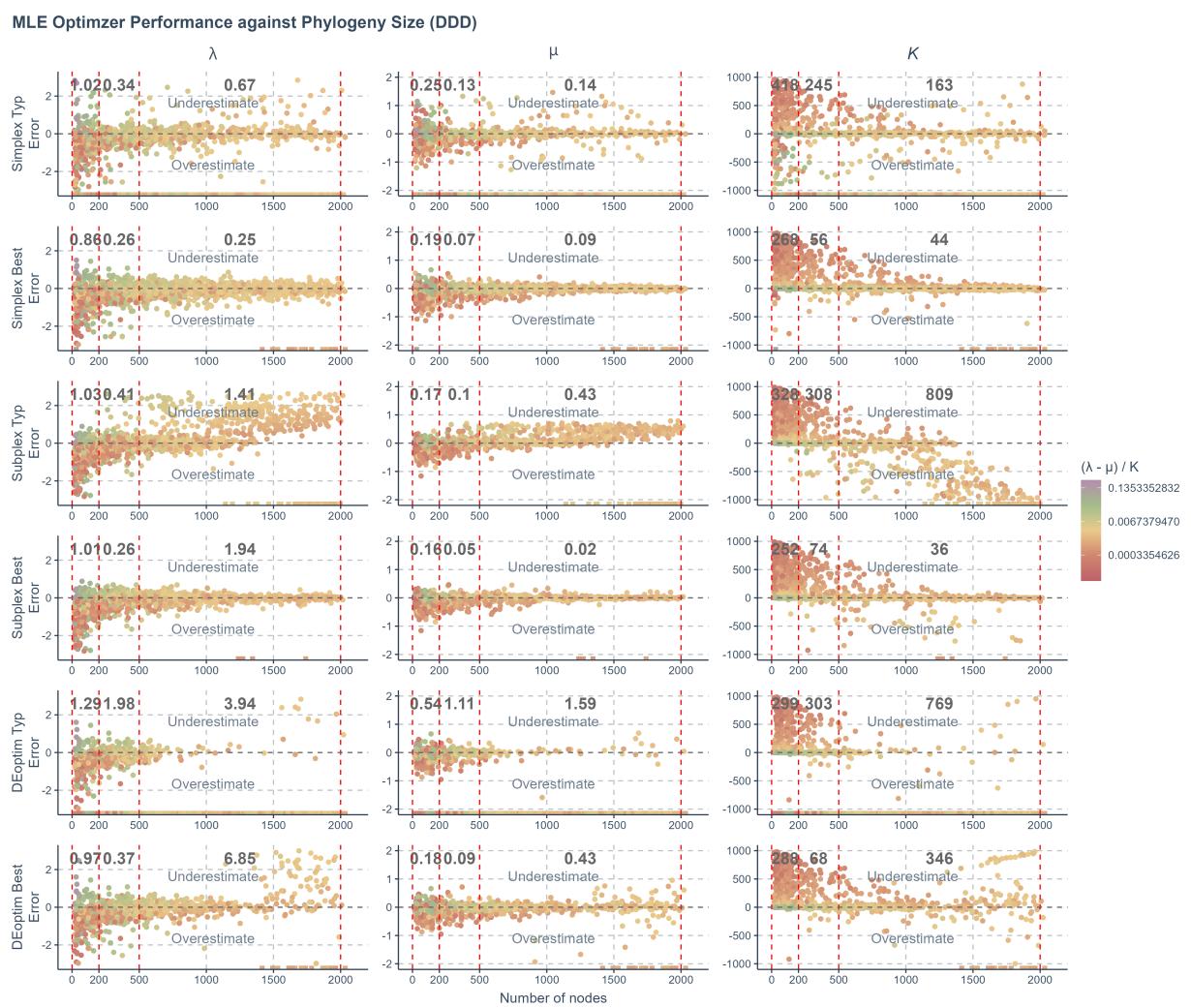


Fig. 12. Error of maximum likelihood estimation using Simplex, Subplex and DEoptim optimizers applied to phylogenies simulated under a diversity-dependent diversification scenario, against the total number of nodes (including root, internal, and tip nodes) in the phylogenies. For each optimizer there were two cases. The typical case (Typ) refers to using the true parameter values as the starting points for the searching process; the best case (Best) refers to using randomly sampled values from the true parameter space as the starting points. The errors shown (y-axis) are the differences between the true parameters used to simulate the phylogenies and the values estimated by each method. Each row represents a method, and each column corresponds to the results for one specific parameter. Phylogenies are categorized based on their size into three sectors within each panel, separated by four vertical red dashed lines. From left to right, the sectors are: small phylogenies with fewer than 200 nodes, medium-sized phylogenies with 200 to 500 nodes, and large phylogenies with more than 500 nodes. The values shown in black within each sector are the mean absolute prediction errors of all data points in the sectors. Color coding: The color of the data points illustrates the strength of the carrying capacity effect, calculated as  $(\lambda - \mu)/K$ . The color gradient transitions from red to purple, indicating increasing strength of the effect. This scale is transformed using log10 for clearer visual differentiation. Small squares spreading along the x-axis signify optimization failures. Extremely deviating estimates are not shown in the figure. X-axis: Size of the phylogenies. Y-axis: Error.  $\lambda$ : Speciation rate.  $\mu$ : extinction rate.  $K$ : carrying capacity.

1111 F. ESTIMATION UNCERTAINTY FOR EMPIRICAL TREES

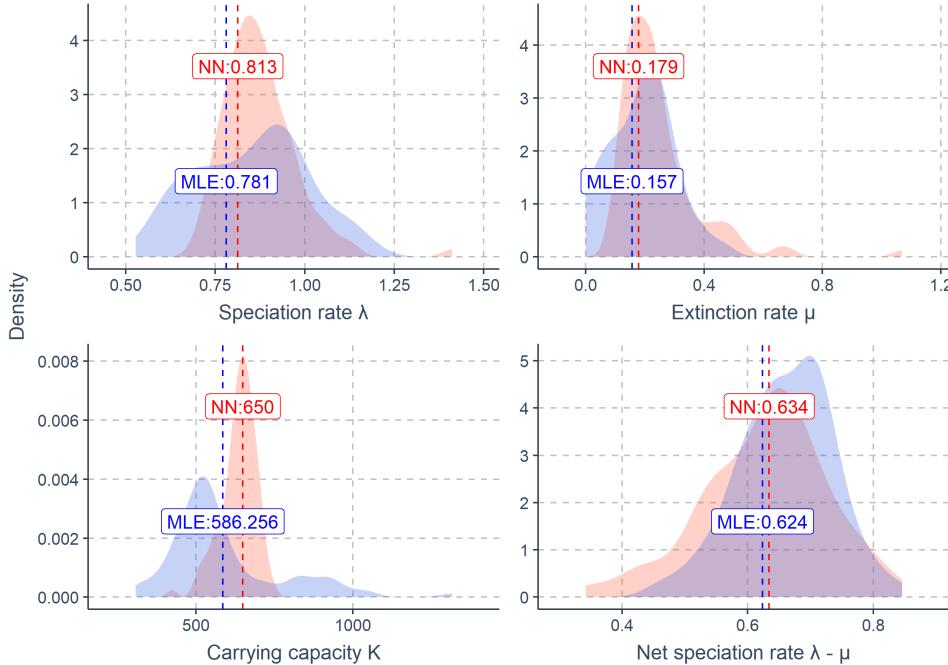


Fig. 13. The neural network estimation uncertainty for a bird phylogeny (Furnariidae). The parameters are estimated using a pre-trained neural network (Boost BT, boosting strategy that corrects GNN results using LSTM) under a diversity-dependent diversification scenario. For reference, maximum likelihood estimation (MLE) is also used to estimate the same parameters. Each panel shows one parameter's estimates using neural network and MLE methods with their uncertainties. The red dashed lines with red numbers indicate the estimates by the neural network method. The blue dashed lines with blue numbers indicate the estimates by the MLE method. Each pink area indicates the density distribution of a neural network estimate from 1000 bootstrap-simulated phylogenies, showing the uncertainty of neural network. Each blue area indicates the density distribution of an MLE estimate from the same set of simulated phylogenies, showing the uncertainty of MLE. X-axis: Parameter (Estimate) values. Y-axis: Density.  $\lambda$ : Speciation rate.  $\mu$ : Extinction rate.  $K$ : Carrying capacity.  $\lambda - \mu$ : Net speciation rate.

1112 The rest of the figures of neural network estimation uncertainty on the empirical

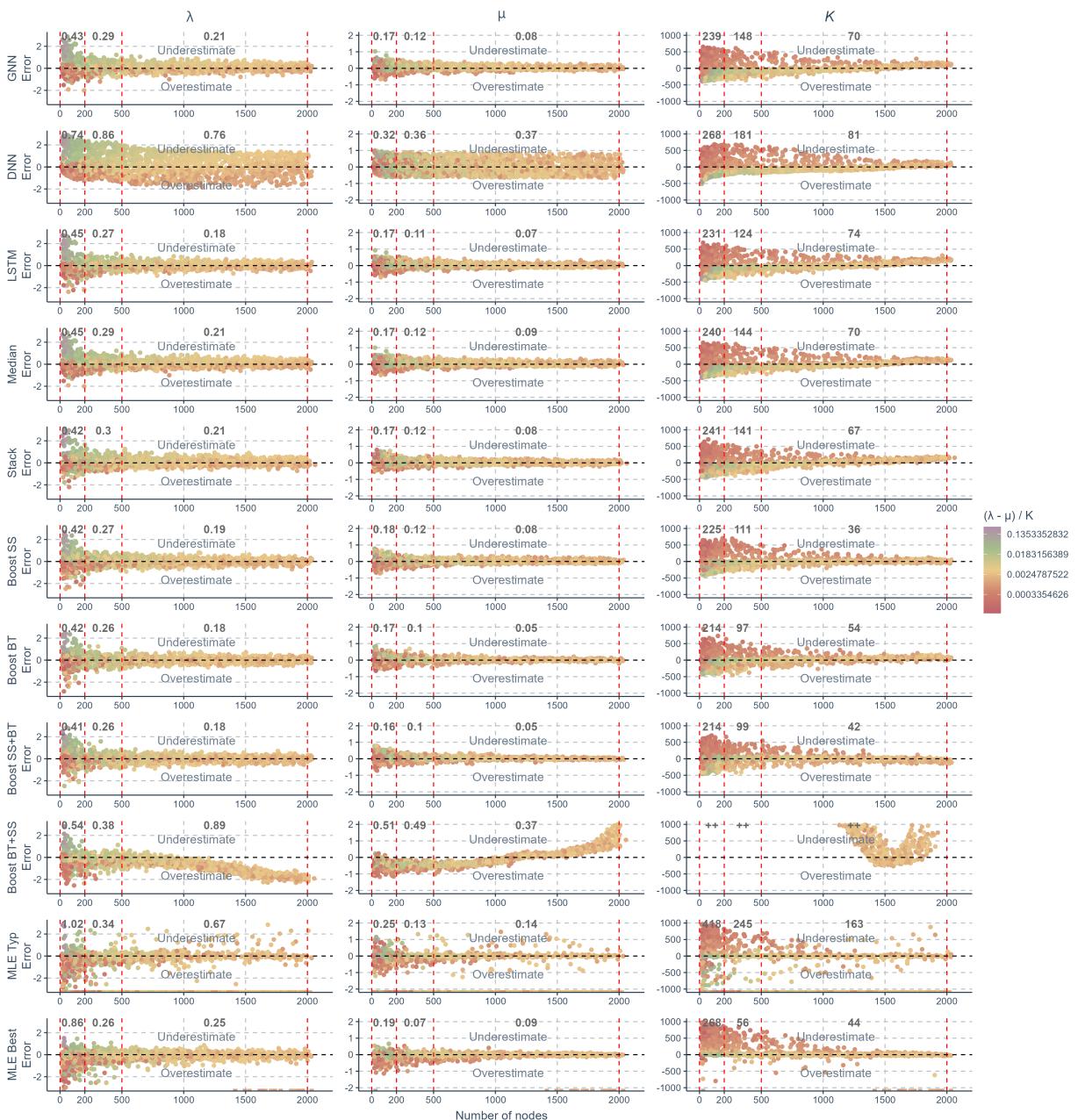
1113 phylogenies under the DDD scenario can be found at

1114 <https://github.com/EvoLandEco/eveGNN/tree/master/uncertainty>

1115 G. RESULTS UNDER THE DIVERSITY-DEPENDENT DIVERSIFICATION SCENARIO

1116

Performance Analysis DDD against Phylogeny Size



1117

1118

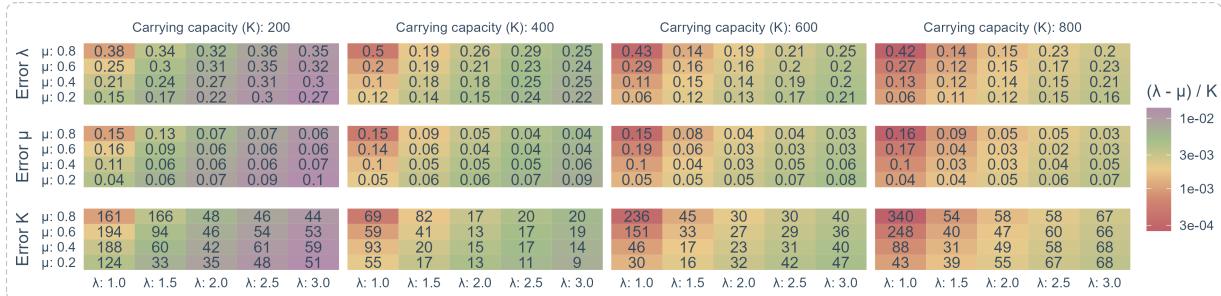
(Caption on next page.)

Fig. 14. (*Figure on previous page.*) The prediction error of various methods applied to phylogenies simulated under a diversity-dependent diversification scenario, against the total number of nodes (including root, internal, and tip nodes) in the phylogenies. The errors shown are the differences between the true parameters used to simulate the phylogenies and the values predicted or estimated by each method. Each row represents a method, and each column corresponds to the results for one specific parameter. Phylogenies are categorized based on their size into three sectors within each panel, separated by four vertical red dashed lines. From left to right, the sectors are: small phylogenies with fewer than 200 nodes, medium-sized phylogenies with 200 to 500 nodes, and large phylogenies with more than 500 nodes. The values shown in black within each sector are the mean absolute prediction errors of all data points in the sectors. Color coding: The color of the data points illustrates the strength of the carrying capacity effect, calculated as  $(\lambda - \mu)/K$ . The color gradient transitions from red to purple, indicating increasing strength of the effect. This scale is transformed using log10 for clearer visual differentiation. GNN: Predictions obtained by the graph neural network using the phylogenies transformed to graph format. DNN: Predictions by the dense neural network using summary statistics. LSTM: Predictions by the long short-term memory recurrent neural network using branching times. Median: Bagging strategy that takes the median value of the predictions from GNN, DNN, and LSTM. Stack: Stacking strategy that utilizes a meta-learner to integrate results from GNN, DNN, and LSTM. Boost SS: Boosting strategy that corrects GNN results using DNN. Boost BT: Boosting strategy that corrects GNN results using LSTM. Boost SS+BT: Sequential correction of GNN errors first using DNN, followed by LSTM. Boost BT+SS: Sequential correction of GNN errors first using LSTM, followed by DNN. MLE Typ: Maximum Likelihood Estimation results using random starting points for parameter optimization. MLE Best: MLE results using the true parameter values as the starting points for optimization. In the MLE result panels, small squares spreading along the x-axis signify optimization failures. Due to significantly lower accuracy, other aggregation methods from the bagging strategy are not displayed on the plot. X-axis: Size of the phylogenies. Y-axis: Error.  $\lambda$ : Speciation rate.  $\mu$ : Extinction rate.  $K$ : Carrying capacity.

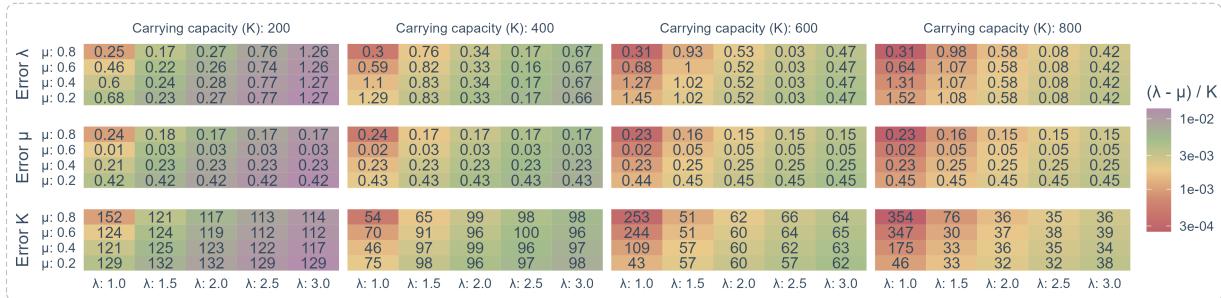
REFERENCES

63

Neural Network (GNN) Robustness



Neural Network (DNN) Robustness



Neural Network (LSTM) Robustness

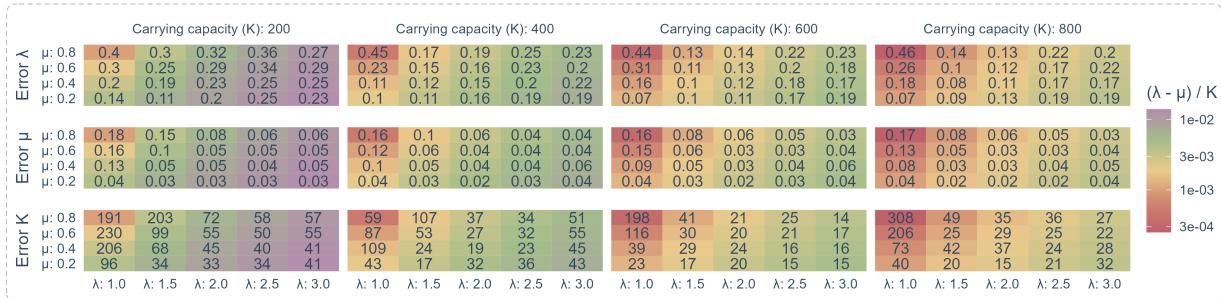
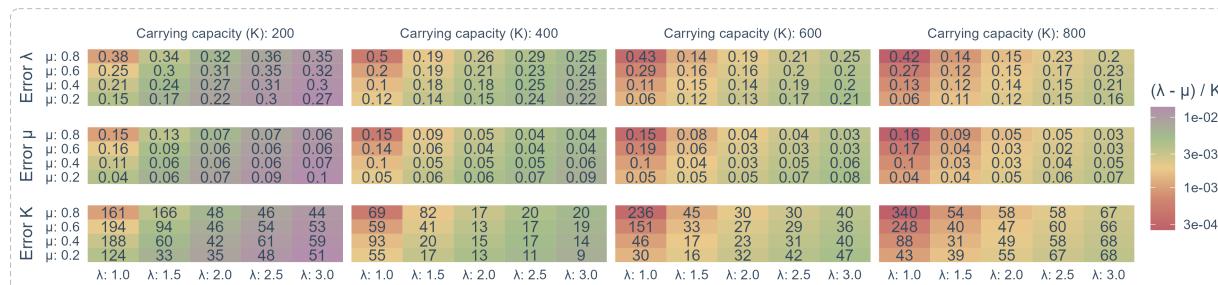
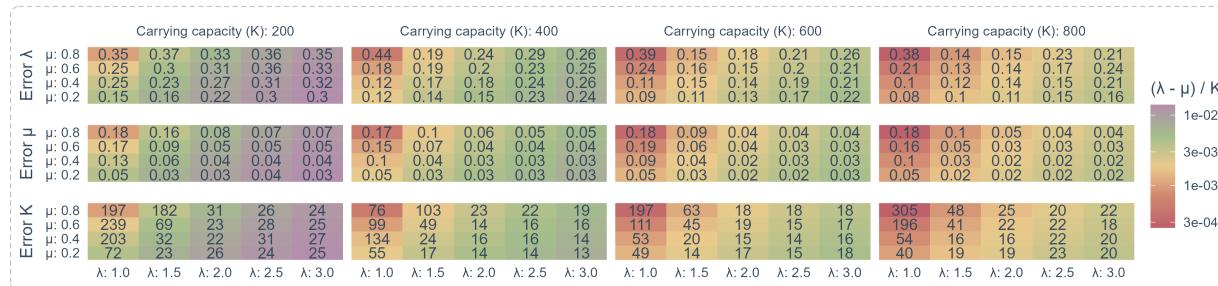


Fig. 15. Robustness comparison between graph neural network (GNN), dense neural network (DNN) and long short-term memory recurrent neural network (LSTM) when operating independently. Same structure as the previous figure.

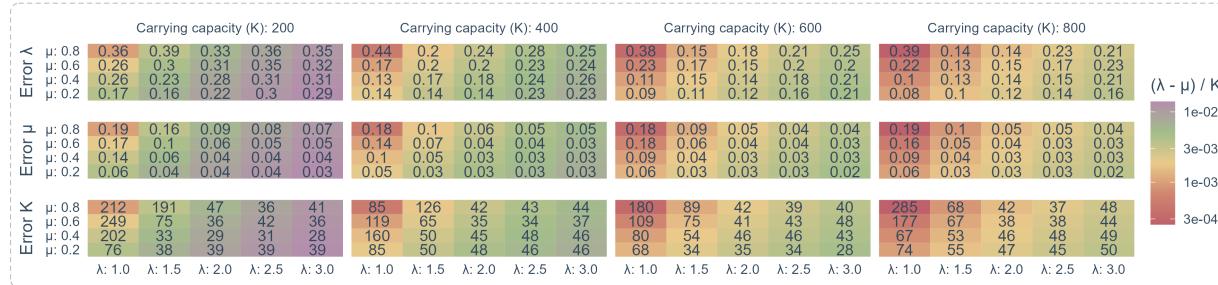
Neural Network (GNN) Robustness



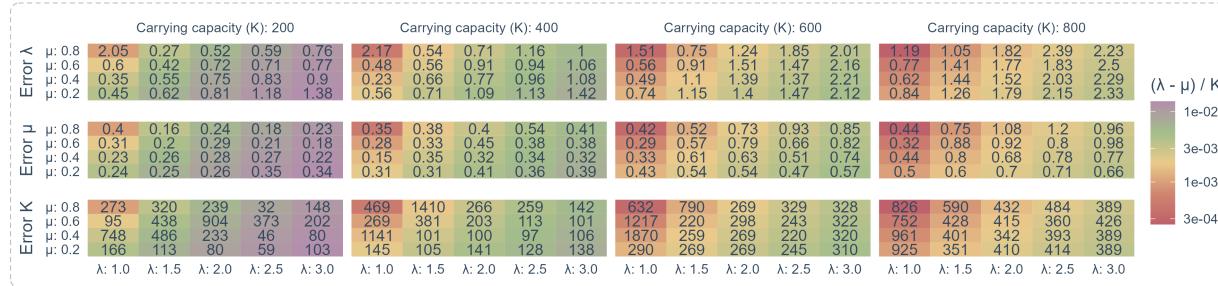
Neural Network (Boosting BT) Robustness



Neural Network (Boosting SS + BT) Robustness



Maximum Likelihood Estimation (Typical Case) Robustness



Maximum Likelihood Estimation (Best Case) Robustness

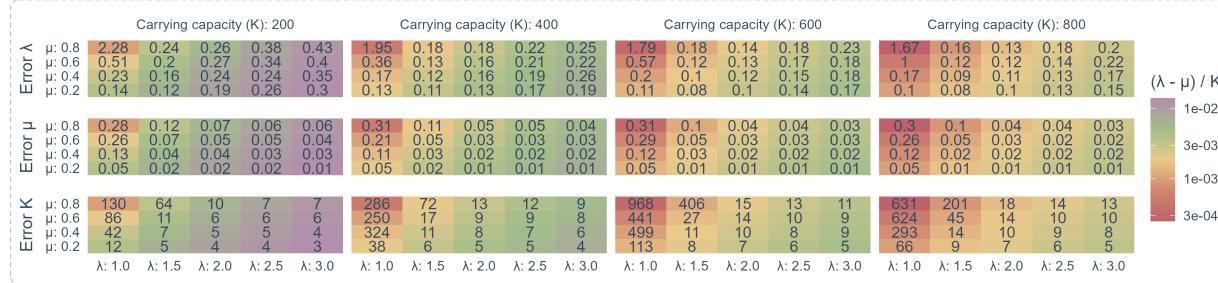


Fig. 16. (Figure on previous page.) The robustness of neural network and maximum likelihood estimation was assessed on 80 sets of phylogenies, each containing 1000 trees randomly simulated under a diversity-dependent diversification scenario, employing identical parameter settings but varied in size, topology, and structure. Each segment delineated by dashed lines corresponds to distinct methods. Each column within a segment is associated with a specific carrying capacity ( $K$ ) used in the simulation of the phylogenies. Each row within a segment details the mean absolute errors between the true and estimated values of a specific parameter, with parameter names labeled on the left side of each row. GNN: Graph neural network. Boosting BT: Graph neural network with long short-term memory recurrent neural network correcting its residuals using branching times. Boosting SS + BT: Graph neural network with dense neural network and long short-term memory recurrent neural network correcting residuals sequentially using summary statistics and branching times. Typical Case: Maximum likelihood estimation using random initial parameter as the starting point. Best Case: Maximum likelihood estimation using true parameter as the starting point. X-axis: Represents the true speciation rate ( $\lambda$ ) used to simulate phylogenies. Y-axis: Represents the true extinction rate ( $\mu$ ) used to simulate phylogenies. Cell Content: The numbers displayed within each heatmap cell indicate the mean absolute error for a parameter, given the specific  $\lambda$ ,  $\mu$  and  $K$  settings. Color Coding: The background color of each cell illustrates the strength of the carrying capacity effect, calculated as  $(\lambda - \mu)/K$ . The color gradient transitions from red to purple, indicating increasing strength of the effect. This scale is transformed using  $\log_{10}$  for clearer visual differentiation. Note that the numerical values within the cells are not mapped to the background colors. For a detailed reference to the effect strength values corresponding to the background colors, refer to the figure legends.

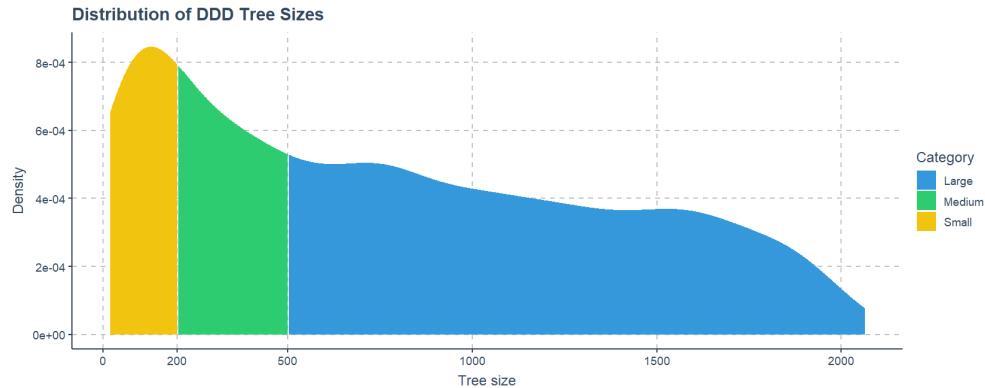


Fig. 17. The density distribution of phylogeny sizes under the diversity-dependent diversification (DDD) scenario. The colors of the areas under the density curve indicate the three categories used in our analyses. Yellow area: Small-sized phylogenies with less than 200 nodes (approx. 100 tips). Green area: Medium-sized phylogenies with more than 200 nodes and less than 500 nodes (approx. 250 tips). Blue area: Large-sized phylogenies with more than 500 nodes.

## H. RESULTS UNDER THE BIRTH-DEATH SCENARIO

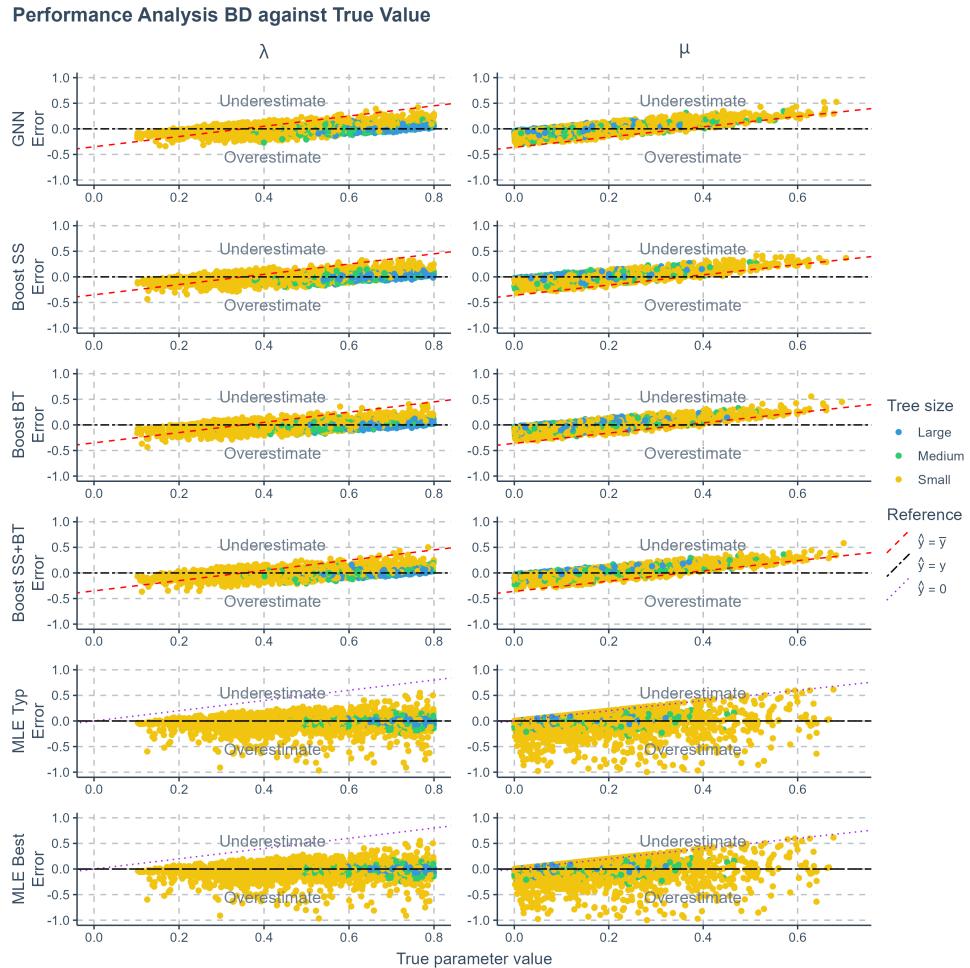


Fig. 18. The prediction error (absolute error) of various methods applied to phylogenies simulated under a birth-death scenario, against true values. The errors shown are the differences between the true parameters used to simulate the phylogenies and the values predicted or estimated by each method. Each row represents a method, and each column corresponds to the results for one specific parameter. Phylogenies are categorized based on their size: yellow for small phylogenies with fewer than 200 nodes (including root, internal, and tip nodes), green for medium-sized phylogenies with 200 to 500 nodes, and blue for large phylogenies with more than 500 nodes. GNN: Predictions obtained by the graph neural network using the phylogenies. Boost SS: Boosting strategy that corrects GNN results using DNN. Boost BT: Boosting strategy that corrects GNN results using LSTM. Boost SS+BT: Sequential correction of GNN errors first using DNN, followed by LSTM. MLE Typ: Maximum Likelihood Estimation results using random starting points for parameter optimization. MLE Best: MLE results using the true parameter values as the starting points for optimization. Red dashed lines in panels representing neural network results indicate the mid-points of the parameter spaces ( $\hat{y} = \bar{y}$  where  $\hat{y}$  denotes a estimated parameter and  $\bar{y}$  denotes the mid-point of the parameter space). Data points close to purple dotted lines ( $\hat{y} = 0$ ) in MLE result panels indicate near-zero estimates. Black two-dash lines indicate accurate estimates ( $\hat{y} = y$  where  $y$  denotes the true parameter value). In the MLE result panels, small squares spreading along the x-axis signify optimization failures. X-axis: True parameter values. Y-axis: Error, or difference between true and predicted values.  $\lambda$ : Speciation rate.  $\mu$ : Extinction rate.

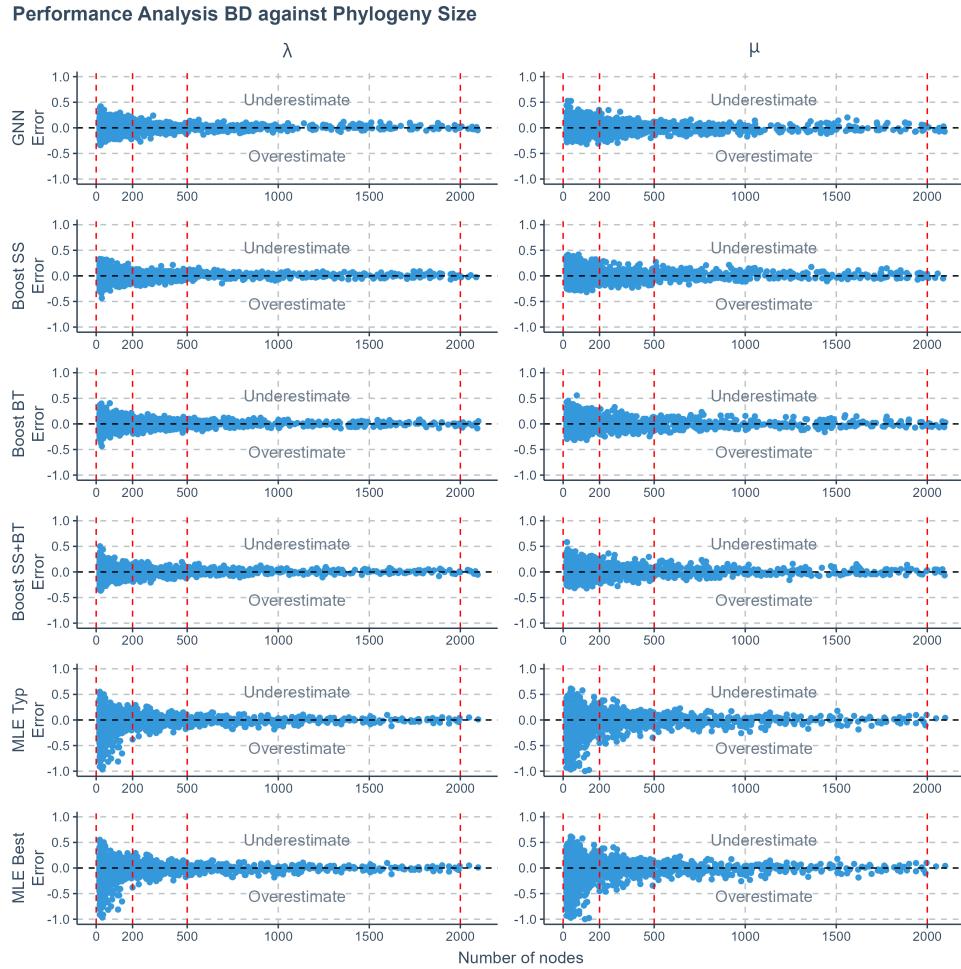


Fig. 19. The prediction error (absolute error) of various methods applied to phylogenies simulated under a birth-death scenario, against the total number of nodes in the phylogenies. The errors shown are the differences between the true parameters used to simulate the phylogenies and the values predicted or estimated by each method. Each row represents a method, and each column corresponds to the results for one specific parameter. Phylogenies are categorized based on their size into three sectors within each panel, separated by four vertical red dashed lines. From left to right, the sectors are: small phylogenies with fewer than 200 nodes (including root, internal, and tip nodes), medium-sized phylogenies with 200 to 500 nodes, and large phylogenies with more than 500 nodes. GNN: Predictions obtained by the graph neural network using the phylogenies. Boost SS: Boosting strategy that corrects GNN results using DNN. Boost BT: Boosting strategy that corrects GNN results using LSTM. Boost SS+BT: Sequential correction of GNN errors first using DNN, followed by LSTM. MLE Typ: Maximum Likelihood Estimation results using random starting points for parameter optimization. MLE Best: MLE results using the true parameter values as the starting points for optimization. X-axis: Size of the phylogenies. Y-axis: Error.  $\lambda$ : Speciation rate.  $\mu$ : Extinction rate.

## I. RESULTS UNDER THE PROTRACTED BIRTH-DEATH SCENARIO

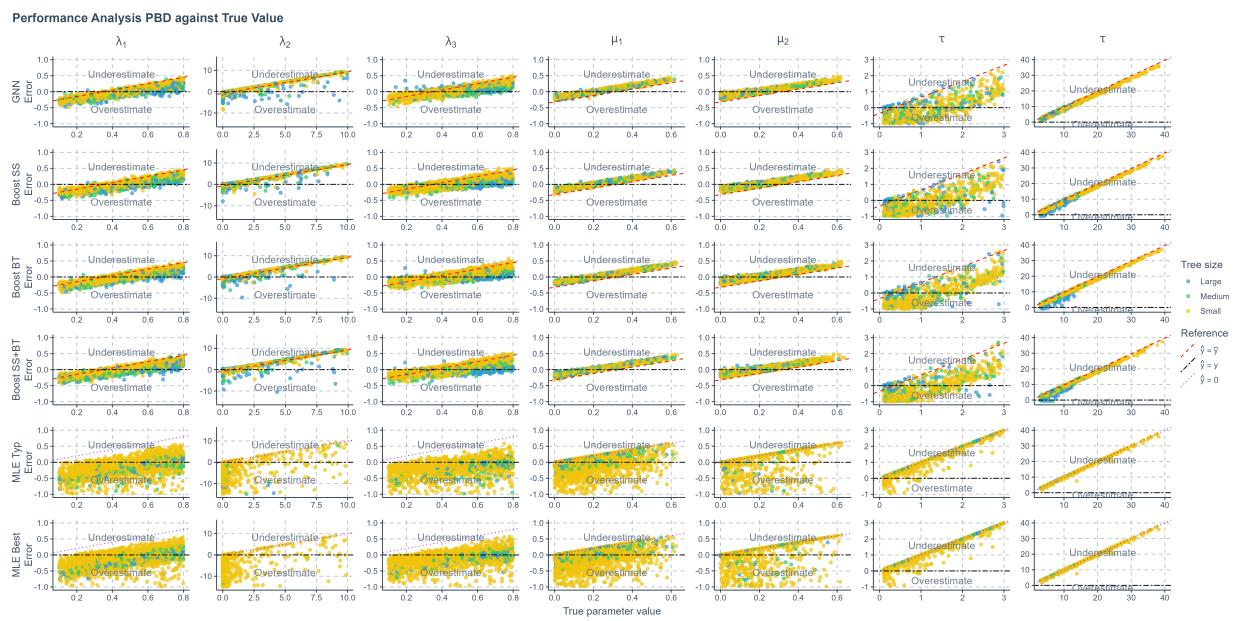


Fig. 20. The prediction error (absolute error) of various methods applied to phylogenies simulated under a protracted birth-death scenario, against true values. The errors shown are the differences between the true parameters used to simulate the phylogenies and the values predicted or estimated by each method. Each row represents a method, and each column corresponds to the results for one specific parameter. Phylogenies are categorized based on their size: yellow for small phylogenies with fewer than 200 nodes (including root, internal, and tip nodes), green for medium-sized phylogenies with 200 to 500 nodes, and blue for large phylogenies with more than 500 nodes. GNN: Predictions obtained by the graph neural network using the phylogenies. Boost SS: Boosting strategy that corrects GNN results using DNN. Boost BT: Boosting strategy that corrects GNN results using LSTM. Boost SS+BT: Sequential correction of GNN errors first using DNN, followed by LSTM. MLE Typ: Maximum Likelihood Estimation results using random starting points for parameter optimization. MLE Best: MLE results using the true parameter values as the starting points for optimization. Red dashed lines in panels representing neural network results indicate the mid-points of the parameter spaces. Purple dotted lines in MLE result panels signify where estimated values are 0. X-axis: True parameter values. Y-axis: Error, or difference between true and predicted values.  $\lambda_1$ : Speciation initiation rate of the good species.  $\lambda_2$ : Speciation completion rate.  $\lambda_3$ : Speciation initiation rate of the incipient species.  $\mu_1$ : Extinction rate of the good species.  $\mu_2$ : Extinction rate of the incipient species.  $\tau$ : Expected duration of speciation.

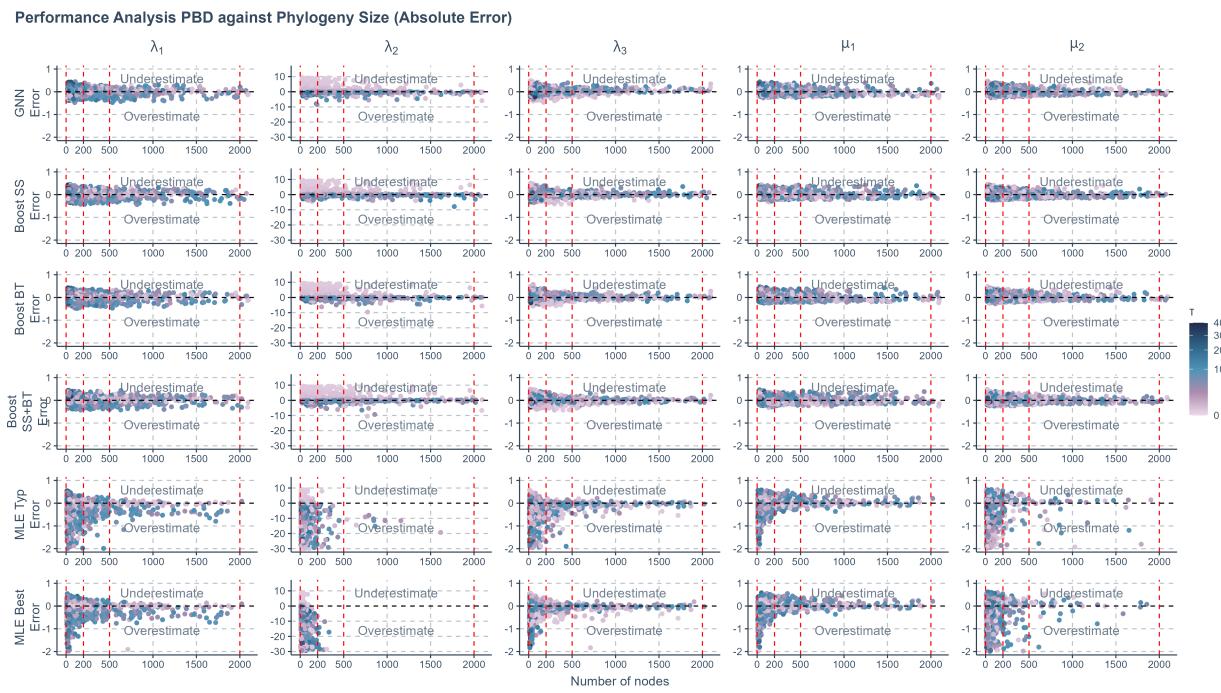


Fig. 21. The prediction error (absolute error) of various methods applied to phylogenies simulated under a protracted birth-death scenario, against the total number of nodes in the phylogenies. The errors shown are the differences between the true parameters used to simulate the phylogenies and the values predicted or estimated by each method. Each row represents a method, and each column corresponds to the results for one specific parameter. Phylogenies are categorized based on their size into three sectors within each panel, separated by four vertical red dashed lines. From left to right, the sectors are: small phylogenies with fewer than 200 nodes (including root, internal, and tip nodes), medium-sized phylogenies with 200 to 500 nodes, and large phylogenies with more than 500 nodes. Color Coding: The color of the data points illustrates the expected duration of speciation. The color gradient transitions from light purple to dark blue, indicating increasing value of the duration. This scale is transformed using square root for clearer visual differentiation. GNN: Predictions obtained by the graph neural network using the phylogenies. Boost SS: Boosting strategy that corrects GNN results using DNN. Boost BT: Boosting strategy that corrects GNN results using LSTM. Boost SS+BT: Sequential correction of GNN errors first using DNN, followed by LSTM. MLE Typ: Maximum Likelihood Estimation results using random starting points for parameter optimization. MLE Best: MLE results using the true parameter values as the starting points for optimization. X-axis: Size of the phylogenies. Y-axis: Error.  $\lambda_1$ : Speciation initiation rate of the good species.  $\lambda_2$ : Speciation completion rate.  $\lambda_3$ : Speciation initiation rate of the incipient species.  $\mu_1$ : Extinction rate of the good species.  $\mu_2$ : Extinction rate of the incipient species.  $\tau$ : Expected duration of speciation.

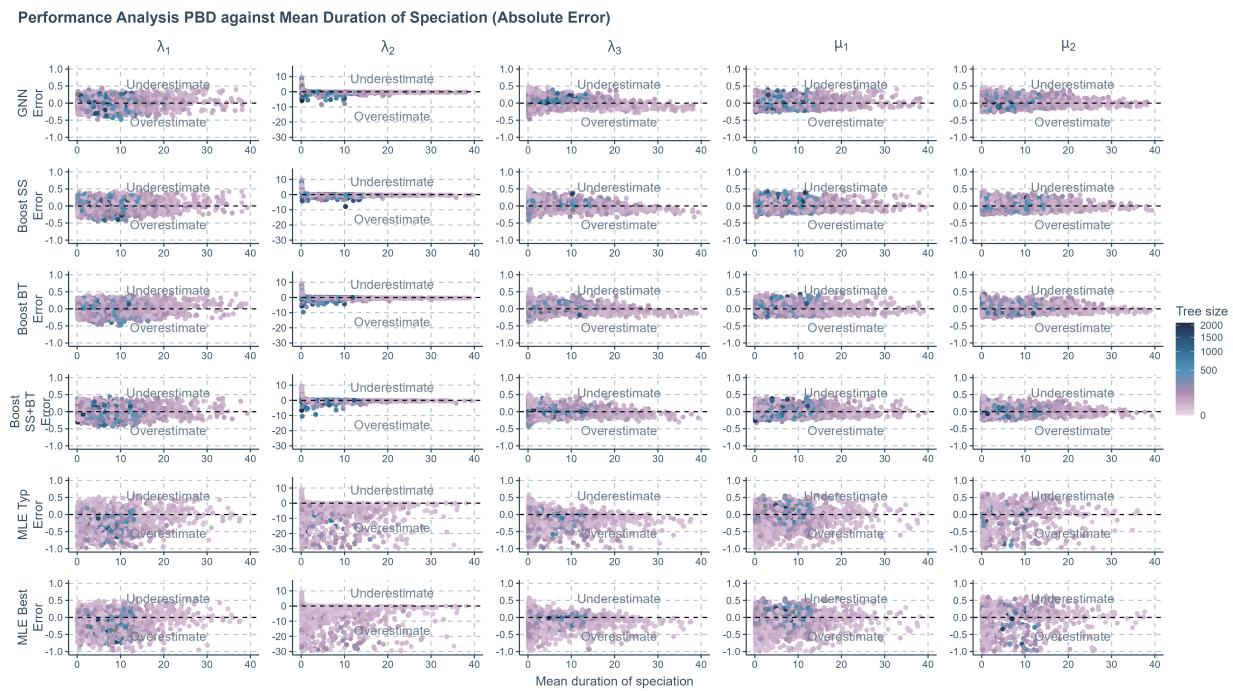


Fig. 22. The prediction error (absolute error) of various methods applied to phylogenies simulated under a protracted birth-death scenario, against the true mean duration of speciation. The errors shown are the differences between the true parameters used to simulate the phylogenies and the values predicted or estimated by each method. Each row represents a method, and each column corresponds to the results for one specific parameter. Color Coding: The color of the data points illustrates the total number of nodes of the phylogenies. The color gradient transitions from light purple to dark blue, indicating increasing value of the node number. This scale is transformed using square root for clearer visual differentiation. GNN: Predictions obtained by the graph neural network using the phylogenies. Boost SS: Boosting strategy that corrects GNN results using DNN. Boost BT: Boosting strategy that corrects GNN results using LSTM. Boost SS+BT: Sequential correction of GNN errors first using DNN, followed by LSTM. MLE Typ: Maximum Likelihood Estimation results using random starting points for parameter optimization. MLE Best: MLE results using the true parameter values as the starting points for optimization. X-axis: Size of the phylogenies. Y-axis: Error.  $\lambda_1$ : Speciation initiation rate of the good species.  $\lambda_2$ : Speciation completion rate.  $\lambda_3$ : Speciation initiation rate of the incipient species.  $\mu_1$ : Extinction rate of the good species.  $\mu_2$ : Extinction rate of the incipient species.  $\tau$ : Expected duration of speciation.

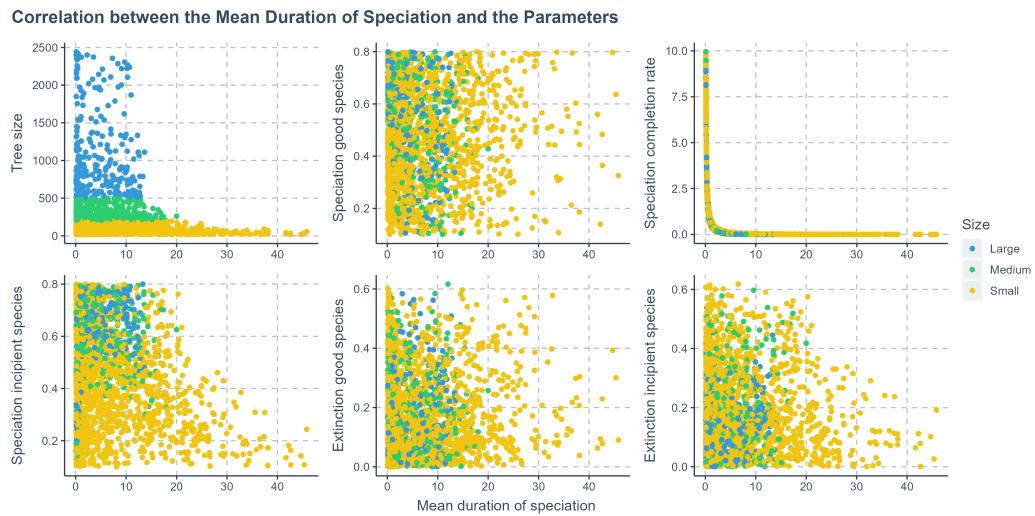


Fig. 23. The correlation between the mean duration of speciation and the true parameter values under the protracted birth-death diversification scenario. Phylogenies are categorized based on their size: yellow for small phylogenies with fewer than 200 nodes (including root, internal, and tip nodes), green for medium-sized phylogenies with 200 to 500 nodes, and blue for large phylogenies with more than 500 nodes. X-axis: Mean duration of speciation. Y-axis: Tree size.

1122 J. DATASET RE-BALANCING

1123 Birth-death processes without carrying capacity effect may have larger variance of

1124 tree size than the DDD trees. A skew in the frequency of tree size across datasets may

1125 appear that leads to a non-representative sample.

1126 To address this issue, we re-balanced the BD dataset by creating 10 bins, each

1127 designated to hold phylogenies within specific size ranges, spanning from 10 to 2000 nodes

1128 in increments of 200 nodes per bin (the first bin accepts phylogeny of sizes 10 to 200). We

1129 randomly simulated phylogenies using parameters sampled from the same space as the BD

1130 training dataset and allocated them to these bins according to their sizes, continuing this

1131 process until each bin reached its target capacity of 10,000 phylogenies. This method leads

1132 to a more equal representation of phylogenies of each size range, reducing size-based

1133 sampling bias. The filled bins were subsequently combined to form a re-balanced dataset,

1134 which in total has 100,000 phylogenies.

1135 To compare with the original BD dataset, we trained neural networks on the

1136 re-balanced dataset, and validated neural network performance on an additional testing

1137 dataset (10,000 phylogenies simulated using the same parameter space). We computed

1138 MLE estimates on 2,000 randomly sampled phylogenies from the testing dataset.

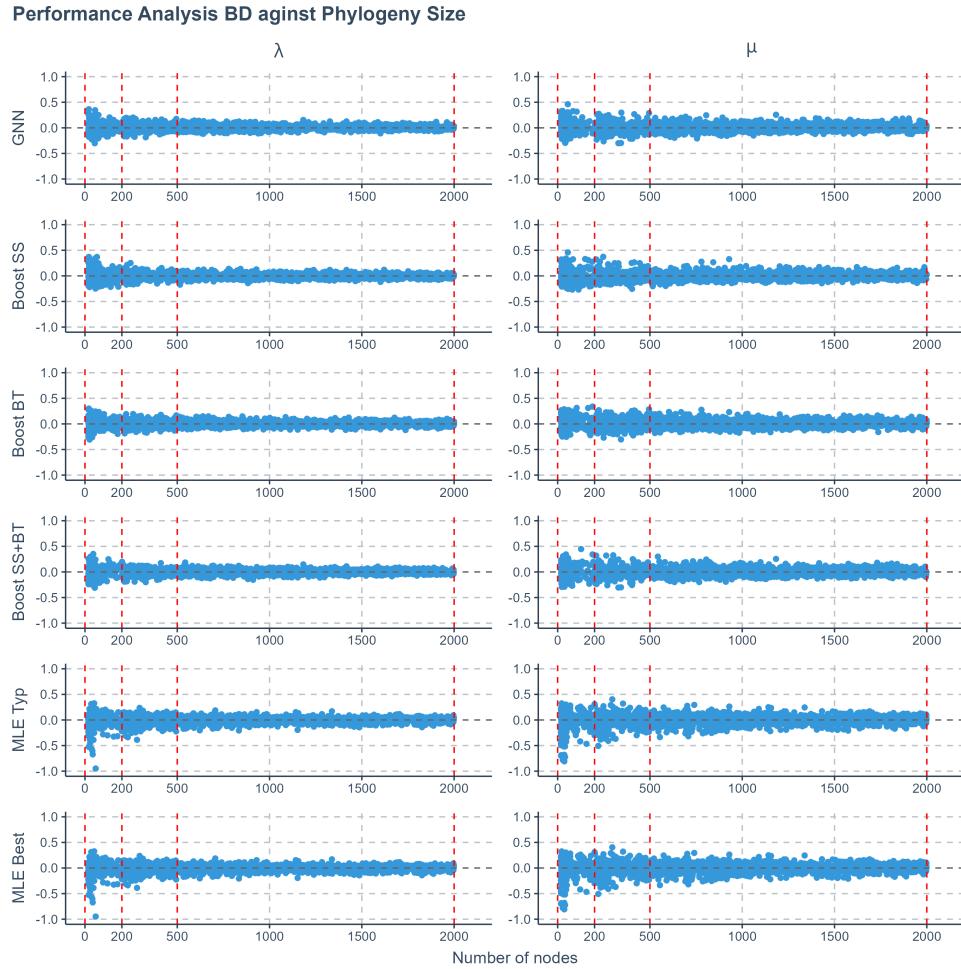


Fig. 24. The prediction error (absolute error) of various methods applied to re-balanced phylogenies simulated under a birth-death scenario, against the total number of nodes in the phylogenies. The errors shown are the differences between the true parameters used to simulate the phylogenies and the values predicted or estimated by each method. Each row represents a method, and each column corresponds to the results for one specific parameter. Phylogenies are categorized based on their size into three sectors within each panel, separated by four vertical red dashed lines. From left to right, the sectors are: small phylogenies with fewer than 200 nodes (including root, internal, and tip nodes), medium-sized phylogenies with 200 to 500 nodes, and large phylogenies with more than 500 nodes. This scale is transformed using square root for clearer visual differentiation. GNN: Predictions obtained by the graph neural network using the phylogenies. Boost SS: Boosting strategy that corrects GNN results using DNN. Boost BT: Boosting strategy that corrects GNN results using LSTM. Boost SS+BT: Sequential correction of GNN errors first using DNN, followed by LSTM. MLE Typ: Maximum Likelihood Estimation results using random starting points for parameter optimization. MLE Best: MLE results using the true parameter values as the starting points for optimization. X-axis: Size of the phylogenies. Y-axis: Error.  $\lambda$ : Speciation rate.  $\mu$ : Extinction rate.

1139 K. UNSEEN DATA AND COMPLETE PHYLOGENY

1140 We simulated additional datasets to explore the generalization ability of the neural

1141 networks when facing data with completely unseen true parameters, as well as to compare

1142 neural network performances between extant and complete phylogenies. Each simulated

1143 dataset was divided into in-sample and out-of-sample datasets. We used the in-sample

1144 datasets for training and testing and the out-of-sample datasets for evaluating the

1145 generalization ability of the trained neural networks. Validating trained neural networks on

1146 the out-of-sample datasets can provide insights into whether their performances are

1147 tailored to the peculiarities of the already seen data and whether they are robust to new,

1148 unseen phylogenies. For each tree we kept two versions: tree of all species (TAS) and tree

1149 of extant species (TES). See Table 2 for the parameter settings of the additional datasets,

1150 see Table 3 for the criteria of in-sample and out-of-sample dataset separation. To conserve

1151 GPU memory, the parameter space for additional datasets was deliberately kept smaller,

1152 given that the TAS dataset inherently contains far more information than the TES.

**A:** Parameter settings for BD and DDD trees

Type	Age	N	$\lambda_0$		$\mu_0$		K	
			a	b	a	b	a	b
BD	10	60k	0.1	0.6	0.0	$0.9\lambda_0$	-	-
DDD	10	100k	0.1	3.0	0.0	$0.9\lambda_0^*$	10	1000

**B:** Parameter settings for PBD trees

Type	Age	N	$b_1$		$\lambda_1$		$b_2$		$\mu_1$		$\mu_2$	
			a	b	a	b	a	b	a	b	a	b
PBD	10	100k	0.1	0.8	0.001	10	0.1	0.8	0.0	$0.8b_1$	0.0	$0.8b_2$

Table 2. List of simulated tree datasets. The type column specifies which function is used to generate the trees. The age column specifies the crown age of the trees. The N column specifies the number of trees in the dataset. The rest of the columns specify the lower (a) and the upper (b) bounds of the initial parameters for the tree simulations, all the parameters are sampled from  $U(a, b)$  except for  $\lambda_1$  of the protracted birth-death scenario.  $\lambda_1$  is computed as  $\lambda_1 = 10^e$  where  $e$  is sampled from  $U(-3, 1)$ .  $U$  denotes uniform distribution. List A shows the parameter distributions of the birth-death trees and the diversity-dependent-diversification trees,  $\lambda$ : intrinsic speciation rate/birth rate;  $\mu$ : intrinsic extinction rate/death rate;  $K$ : carrying capacity. List B shows the parameter distributions of the protracted birth-death trees,  $\lambda_1$ : speciation-initiation rate of good species;  $\lambda_2$ : speciation-completion rate;  $\lambda_3$ : speciation-initiation rate of incipient species;  $\mu_1$ : extinction rate of good species;  $\mu_2$ : extinction rate of incipient species. \*In diversity-dependent-diversification simulations, the maximum extinction rate is capped at 1.5 if  $0.9\lambda > 1.5$ .

Model	Parameter	Left Out	In Sample	Right Out
BD	$\lambda_0$	[0.10, 0.18)	[0.18, 0.52]	(0.52, 0.60]
BD	$\mu_0$	[0.00, 0.08)	[0.08, 0.46]	(0.46, 0.54]
DDD	$\lambda_0$	[0.00, 0.30)	[0.30, 2.70]	(2.70, 3.00]
DDD	$\mu_0$	[0.00, 0.10)	[0.10, 0.80]	(0.80, 0.90]
DDD	$K$	[10, 100)	[100, 900]	(900, 1000]
PBD	$b_1$	[0.10, 0.18)	[0.18, 0.72]	(0.72, 0.80]
PBD	$\lambda_1$	[0.001, 0.002)	[0.002, 5]	(5, 10]
PBD	$b_2$	[0.10, 0.18)	[0.18, 0.72]	(0.72, 0.80]
PBD	$\mu_1$	[0.00, 0.06)	[0.06, 0.58]	(0.58, 0.64]
PBD	$\mu_2$	[0.00, 0.06)	[0.06, 0.58]	(0.58, 0.64]

Table 3. Criteria for in-sample and out-of-sample dataset separation. Trees generated from each model are separated into left out-of-sample group, in-sample group and right out-of-sample group, based on the parameter ranges. The Model column shows the model of a parameter; the Parameter column shows the corresponding parameter; the Left Out column shows the criteria for the left out-of-sample group; the In Sample column shows the criteria for the in-sample group; the Right Out column shows the criteria of the right out-of-sample group.  $\lambda$ : intrinsic speciation rate/birth rate;  $\mu$ : intrinsic extinction rate/death rate;  $K$ : carrying capacity. List B shows the parameter distributions of the protracted birth-death trees,  $\lambda_1$ : speciation-initiation rate of good species;  $\lambda_2$ : speciation-completion rate;  $\lambda_3$ : speciation-initiation rate of incipient species;  $\mu_1$ : extinction rate of good species;  $\mu_2$ : extinction rate of incipient species.

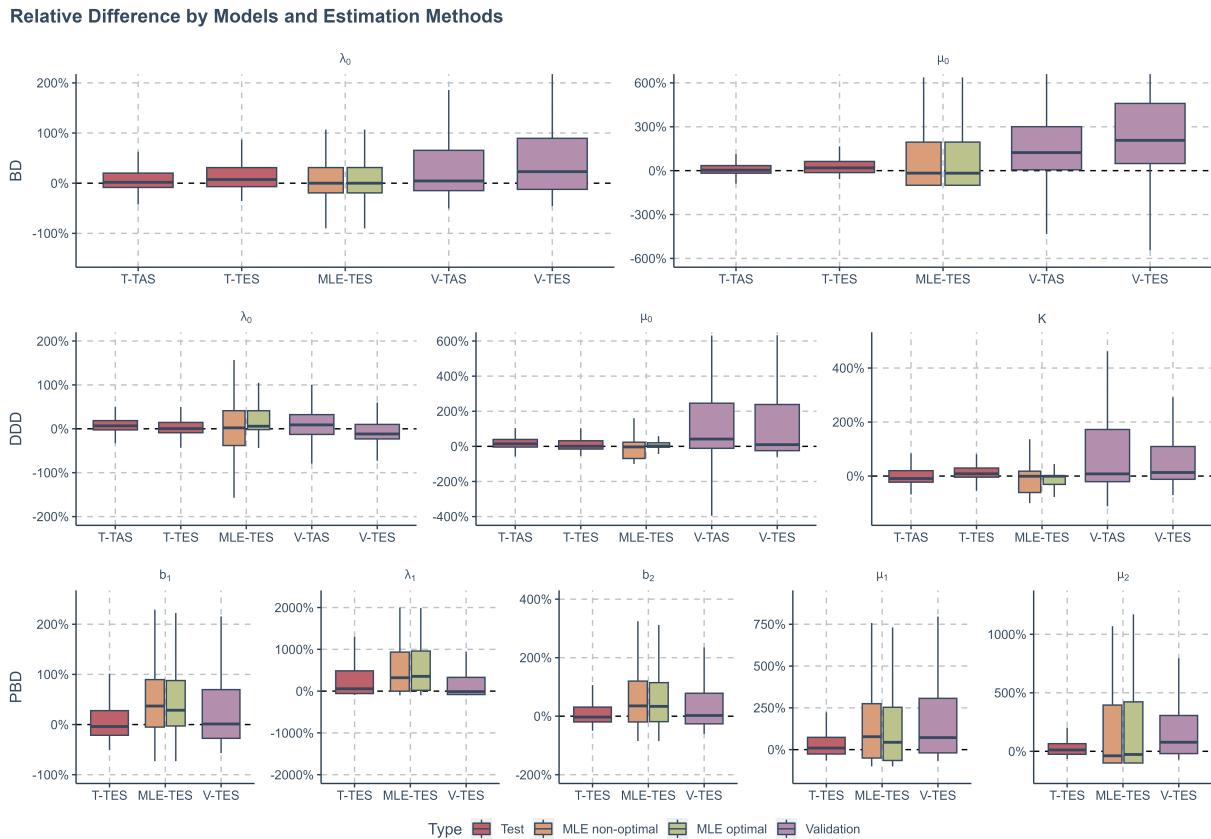


Fig. 25. Comparisons of relative differences (in percentage) between ground true and estimated parameter values. From top to bottom, the panels in each row present the relative differences of trees generated by a specific diversification process. From left to right, the panels in each column present the relative differences of a specific parameter used when simulating the trees. Within each panel, each box represents a specific method for parameter estimation on a specific data set (as described in x-axis labels). Red boxes represent parameter estimation by using only graph neural network (GNN) on the in-sample datasets (Test in figure), yellow boxes represent the non-optimal maximum likelihood estimation (MLE) method on the complete datasets (direct outputs from simulation, without any separation), green boxes represent the optimal MLE method on the complete datasets, purple boxes represent parameter estimation by GNN on the out-of-sample datasets. BD - birth-death trees; DDD - diversity-dependent-diversification trees; PBD - protracted birth-death trees.  $\lambda$  - birth rate/intrinsic speciation rate;  $\mu$  death rate/intrinsic extinction rate;  $K$  - carrying capacity;  $\lambda_1$  - speciation rate of good species;  $\lambda_2$  - speciation-completion rate;  $\lambda_3$  - speciation rate of incipient species;  $\mu_1$  - extinction rate of good species;  $\mu_2$  extinction rate of incipient species. T-TAS - GNN parameter estimation on full trees (with extinct lineages) in the in-sample data set; T-TES - GNN parameter estimation on extant trees (without extinct lineages) in the in-sample dataset; MLE-TES - MLE parameter estimation on extant trees in the complete dataset; V-TAS - GNN parameter estimation on full trees in the out-of-sample dataset; V-TES - GNN parameter estimation on extant trees in the out-of-sample dataset.

1153

## L. LEARNING FROM SUMMARY STATISTICS

1154

To analyze the relationships between summary statistics and the true parameters

1155

used in the diversity-dependent diversification simulations, we computed Pearson

1156

correlation indices for each summary statistic against the true values of the three

1157

parameters. The absolute values of the correlations are visualized as a heatmap, detailed in

1158

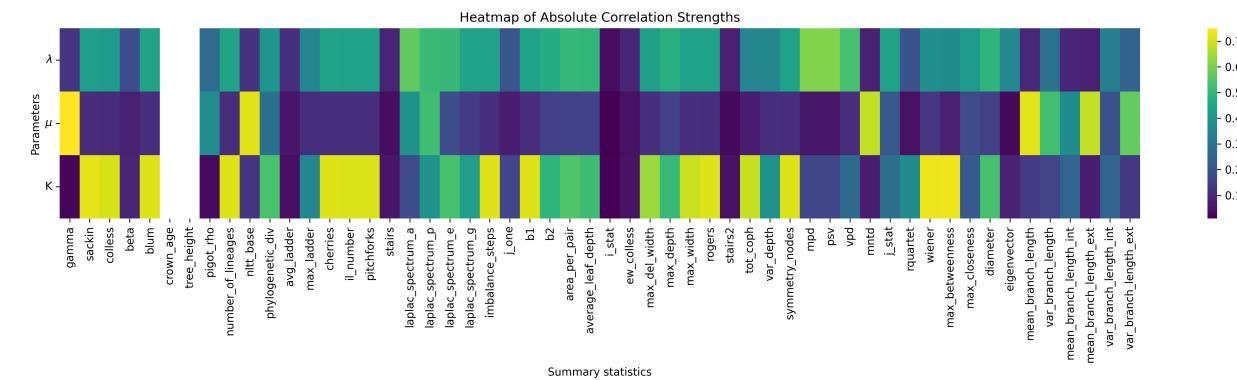


Fig. 26. Heatmap of absolute correlation strengths between true parameters and summary statistics from simulated trees under a diversity-dependent diversification scenario. Each column corresponds to a specific summary statistic, while each row corresponds to a true parameter that was used to simulate the phylogenies. The true parameters are denoted as follows:  $\lambda$  for speciation rate,  $\mu$  for extinction rate, and  $K$  for carrying capacity. The color gradient, ranging from dark purple to yellow, represents the increasing values of the absolute Pearson correlations between the summary statistics and the true parameters, See Appendix N for the details of the statistics.

1159

The heatmap analysis reveals that a large number of summary statistics showed a

1160

high correlation with carrying capacity, whereas strong correlations with speciation and

1161

extinction rates are considerably less frequent. Generally, the correlation strength between

1162

the true parameters and extinction rate is markedly lower. These findings align with the

1163

observed performance of DNN, which generally yields good estimates for carrying capacity

1164

but is much less effective in accurately predicting speciation rates and particularly poor at

1165

estimating extinction rates.

1166

We further evaluated the DNN's performance by benchmarking it against a range of

1167

non-neural-network regression techniques using the same summary statistics. These

1168 included multivariate linear regression, Ridge regression, Lasso regression, random forest  
1169 regression, and gradient boosting regression. Our findings indicated that while these  
1170 traditional regression and machine learning methods generally outperformed the DNN,  
1171 they still did not match the performance of our more complex neural networks.

1172 Traditional regression methods and other machine learning techniques often  
1173 outperform linear feed-forward neural networks in classical regression tasks. Such methods  
1174 can stabilize performance with less data compared to neural networks, which usually  
1175 require large datasets to generalize effectively. In our study, the dataset size – consisting of  
1176 100,000 entries across 54 statistics – may seem substantial, but it is still relatively modest  
1177 when tasked with regressing multiple parameters simultaneously.

1178 Despite these findings, DNNs have shown efficacy in enhancing the performance of  
1179 other neural networks, particularly through the prediction of residuals using summary  
1180 statistics. DNN might not be the optimal choice for estimating parameters from summary  
1181 statistics alone, but they can be valuable in ensemble learning strategies.

1182

M. META INFORMATION OF THE SELECTED EMPIRICAL TREES

Family	Tree	Ntip
Amphibia	Caecilidae	31
Amphibia	Hynobiidae	46
Amphibia	Salamandridae	42
Amphibia	Plethodontidae	278
Amphibia	Pipidae	23
Amphibia	Eleutherodactylidae	145
Amphibia	Ranidae	218
Bird	Tyrannidae	419
Bird	Thraupidae	370
Bird	Psittacidae	330
Bird	Trochilidae	334
Bird	Columbidae	306
Bird	Furnariidae	302
Bird	Muscicapidae	279
Bird	Accipitridae	242
Bird	Picidae	223
Bird	Thamnophilidae	219
Bird	Fringillidae	194
Bird	Strigidae	191
Bird	Turdidae	170
Bird	Meliphagidae	177
Bird	Phasianidae	176
Bird	Emberizidae	163
Bird	Anatidae	157
Bird	Cisticolidae	142
Bird	Pycnonotidae	124
Bird	Rallidae	125
Bird	Cuculidae	138
Bird	Estrildidae	140
Bird	Nectariniidae	127
Bird	Leiothrichidae	127
Bird	Corvidae	120
Bird	Zosteropidae	120
Bird	Sturnidae	109
Bird	Parulidae	109
Bird	Ploceidae	108
Bird	Icteridae	102
Bird	Apodidae	99
Bird	Laridae	99
Bird	Alaudidae	91
Bird	Monarchidae	87

REFERENCES

Bird	Scolopacidae	89
Bird	Caprimulgidae	88
Bird	Alcedinidae	91
Bird	Campephagidae	80
Bird	Procellariidae	81
Bird	Hirundinidae	83
Bird	Troglodytidae	79
Bird	Phylloscopidae	71
Bird	Ardeidae	61
Bird	Pellorneidae	66
Bird	Sylviidae	62
Bird	Cardinalidae	68
Bird	Charadriidae	64
Bird	Falconidae	64
Bird	Motacillidae	62
Bird	Acanthizidae	63
Bird	Cotingidae	65
Bird	Vireonidae	58
Bird	Acrocephalidae	52
Bird	Bucerotidae	55
Bird	Paridae	53
Bird	Pachycephalidae	50
Bird	Locustellidae	53
Bird	Rhinocryptidae	53
Bird	Timaliidae	55
Bird	Cracidae	50
Bird	Pipridae	52
Bird	Grallariidae	49
Bird	Malacoptidae	46
Bird	Passeridae	48
Bird	Rhipiduridae	42
Bird	Dicaeidae	45
Bird	Tinamidae	47
Bird	Petroicidae	44
Bird	Ramphastidae	35
Bird	Tityridae	41
Bird	Trogonidae	42
Bird	Lybiidae	41
Bird	Paradisaeidae	40
Bird	Phalacrocoracidae	33
Bird	Bucconidae	35
Bird	Threskiornithidae	34
Bird	Mimidae	34
Bird	Odontophoridae	34

REFERENCES

81

Bird	Oriolidae	30
Bird	Laniidae	29
Bird	Pittidae	31
Bird	Platysteiridae	30
Bird	Cettiidae	32
Bird	Megalaimidae	28
Bird	Maluridae	27
Bird	Sittidae	24
Bird	Meropidae	26
Bird	Dicruridae	24
Bird	Otididae	25
Bird	Alcidae	23
Bird	Hydrobatidae	22
Bird	Musophagidae	23
Bird	Megapodiidae	21
Bird	Cacatuidae	21
Bird	Diomedeidae	21
Bird	Vangidae	21
CrocoTurtle	Crocodylia	25
CrocoTurtle	Testudines	233
Mammal	Vespertilionidae	386
Mammal	Soricidae	329
Mammal	Sciuridae	276
Mammal	Pteropodidae	174
Mammal	Phyllostomidae	150
Mammal	Bovidae	138
Mammal	Cercopithecidae	127
Mammal	Molossidae	98
Mammal	Didelphidae	84
Mammal	Hipposideridae	74
Mammal	Rhinolophidae	73
Mammal	Echimyidae	69
Mammal	Dasyuridae	63
Mammal	Mustelidae	59
Mammal	Heteromyidae	58
Mammal	Leporidae	58
Mammal	Macropodidae	56
Mammal	Nesomyidae	55
Mammal	Ctenomyidae	51
Mammal	Dipodidae	51
Mammal	Emballonuridae	49
Mammal	Cebidae	48
Mammal	Cervidae	45
Mammal	Felidae	40

REFERENCES

Mammal	Talpidae	39
Mammal	Geomyidae	38
Mammal	Pitheciidae	37
Mammal	Canidae	34
Mammal	Delphinidae	34
Mammal	Viverridae	34
Mammal	Herpestidae	33
Mammal	Spalacidae	31
Mammal	Ochotonidae	28
Mammal	Gliridae	27
Mammal	Tenrecidae	25
Mammal	Atelidae	24
Mammal	Erinaceidae	22
Mammal	Phalangeridae	22
Squamate	Xantusiidae	26
Squamate	Gerrhosauridae	28
Squamate	Cordylidae	42
Squamate	Varanidae	53
Squamate	Chamaeleonidae	142
Squamate	Iguanidae	31
Squamate	Phrynosomatidae	114
Squamate	Pythonidae	26
Squamate	Viperidae	209

1183

## N. LIST OF SUMMARY STATISTICS

Summary Statistics	
Gamma	Area Per Pair (aPP)
Sackin	Average Leaf Depth (aLD)
Colless	I Statistic
Aldous' Beta Statistic	ewColless
Blum	Max Delta Width (maxDelW)
Crown Age	Maximum of Depth
Tree Height	Variance of Depth
Pigot's Rho	Maximum Width
Number of Lineages	Rogers
nLTT with Empty Tree	Total Cophenetic Distance
Phylogenetic Diversity	Symmetry Nodes
AvgLadder Index	Mean of Pairwise Distance (mpd)
Cherries	Variance of Pairwise Distance (vpd)
ILnumber	Phylogenetic Species Variability (psv)
Pitchforks	Mean Nearest Taxon Distance (mndt)
Stairs	J Statistic of Entropy
Stairs2	Rquartet Index
Laplacian Spectrum Asymmetry	Laplacian Spectrum Log Eigen
Laplacian Spectrum Peakedness	Laplacian Spectrum Eigengap
Number of Nodes	Wiener Index
B1	Max Betweenness
B2	Max Closeness
Diameter, Without Branch Lengths	Maximum Eigen Vector Value
Mean Branch Length	Variance of Branch Length
Mean External Branch Length	Variance of External Branch Length
Mean Internal Branch Length	Variance of Internal Branch Length
Number of Imbalancing Steps	J_One Statistic

Table 5. List of phylogenetic summary statistics used in neural network training