

Acute and sub-chronic toxicity of 6PPD-quinone to early-life stage lake trout (*Salvelinus namaycush*)

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Abstract

N-(1,3-Dimethylbutyl)-N'-phenyl-p-phenylenediamine-quinone (6PPD-q) is a rubber-tire derivative which leaches into surface waters from roadway runoff, from tire particles and has been identified as a possible driver of urban runoff mortality syndrome in coho salmon. Sensitivity to this toxicant is highly variable across fish species and life stages. With environmental concentrations meeting or exceeding toxicity thresholds in sensitive fishes, the potential for ecologically relevant effects is significant. There is currently no data regarding the sensitivity of lake trout (*Salvelinus namaycush*) to 6PPD-q. As early-life stages of fishes are typically more sensitive than adults, the goal of these studies was to evaluate the acute and sub-

32 chronic toxicity of 6PPD-q to early-life stage lake trout. Alevins exposed from hatch until 45
33 days post hatch (dph) to time-weighted average 6PPD-q concentrations ranging from 0.22-13.5
34 µg/L exhibited a 45-day median lethal dose (LC₅₀) of 0.39 µg/L. Deformities throughout growth
35 were observed, with a unique pooling of blood observed in the caudal fin and eye. A subsequent
36 acute study with exogenously feeding lake trout fry determined a 96-hr LC₅₀ of 0.50 µg/L. From
37 these studies we can conclude that lake trout alevins and exogenously feeding fry are sensitive to
38 6PPD-q, which underscores the relevance of this chemical to inland freshwater ecosystems.

39

40 **Keywords:** Rubber tire particles; urban runoff; fish; cross-species sensitivity; N-(1,3-
41 Dimethylbutyl)-N'-phenyl-p-phenylenediamine-quinone

42

43 INTRODUCTION

44

45 The emerging contaminant of concern, 6PPD-q (N-(1,3-Dimethylbutyl)-N'-phenyl-p-
46 phenylenediamine-quinone), has become a recent focus of ecological concern, given its acute
47 toxicity to several salmonid species at concentrations lower than what is found in urban
48 stormwater runoff and receiving streams. In fact, it was recently discovered that 6PPD-q was
49 responsible for pronounced pre-spawn mortality coho salmon (*Oncorhynchus kisutch*) within the
50 Pacific Northwest of the United States have faced for decades, a trend linked to both roadway
51 density and precipitation events ^{1,2}. This phenomenon was termed Urban Runoff Mortality
52 Syndrome (URMS) ³. A follow up study by Tian et al. ⁴ reported a 24-hour median lethal
53 concentration (LC₅₀) of 95 ng/L for 6PPD-q in coho salmon. 6PPD-q is a product of the
54 environmental oxidation of N-(1,3-Dimethylbutyl)-N'-phenyl-p-phenylenediamine (6PPD), an
55 antioxidant used in rubber, notably in tire tread. Consequently, 6PPD-q can enter waterways
56 during precipitation events as tire wear particles (TWP) from roads are washed into aquatic
57 environments. Studies in a number of urban settings have revealed environmental concentrations
58 of 6PPD-q ranging from 0.08 to 2.85 µg/L ⁵⁻⁷. These concentrations surpass the LC₅₀ for coho
59 salmon and several other sensitive species studied so far, including rainbow trout (*Oncorhynchus*
60 *mykiss*) (72-hour LC₅₀ 1.00 µg/L), brook trout (*Salvelinus fontinalis*) (24-hour LC₅₀ 0.59 µg/L) ⁸,
61 and white spotted char (*Salvelinus leucomaenis pluvius*) (24-hour LC₅₀ 0.51 µg/L) ⁹.
62 Consequently, there exists a significant acute risk to wild populations of certain salmonid

63 species; however, few of the globally known 66 distinct species of salmonids have been tested to
64 date limiting current risk assessment efforts.

65
66 The broad range of sensitivity among fishes to this compound is noteworthy. To date, the only
67 fish species found to be sensitive are in the Salmonidae family. However, while some salmonid
68 species, such as white-spotted char and brook trout, exhibit sensitivity at very low concentrations
69 of 6PPD-q in the sub- $\mu\text{g}/\text{L}$ range, other closely related species such as Arctic char (*Salvelinus*
70 *alpinus*) and southern Asian dolly varden (*Salvelinus curilus*) show no acute adverse effects even
71 at very high non-environmentally relevant concentrations ^{8,9}. At the same time, experimental data
72 on acute lethality following exposure to 6PPD-q is lacking for most salmonid (and non-
73 salmonid) species, and presently, there are no reliable predictors that allow risk assessors to
74 identify species that are vulnerable to the exposure with this chemical. In addition, little is known
75 in regard to early-life stage or chronic exposure of salmonid species to 6PPD-q ^{10,11}. Given that
76 embryonic and larval life stages of fishes often tend to be more sensitive to contaminants than
77 adults ¹², there is the potential that these life stages might be more susceptible to the lethal or
78 sub-lethal effects compared to their adult conspecifics. Given that 6PPD-q can leach from TWPs,
79 the risk exists that salmonid alevins, which are closely associated with gravel and sediment, may
80 be of particular risk to exposure to 6PPD-q leaching from settled TWP. In addition, while
81 aqueous exposure to 6PPD-q through urban stormwater runoff normally occurs in pulses ⁷, one
82 might speculate that exposure of salmonids alevins through TWPs in sediments could occur over
83 prolonged periods of time.

84
85 Lake trout (*Salvelinus namaycush*) are a freshwater species which is prevalent across North
86 America, and typically found in cooler, deeper waters ¹³. Lake trout are long-lived, with lifespans
87 surpassing 20 years ¹³. Despite their longevity, the status of lake trout populations in the
88 Laurentian Great Lakes and across Ontario are of concern, with declining numbers and
89 distribution of the species ¹⁴. While this decline is largely attributed to the adverse effects of
90 climate change and habitat loss ¹⁴, there is the potential that lake trout populations could be
91 subject to 6PPD-q via urban stormwater runoff. The Canadian Great Lakes basins, where lake
92 trout reside, are subject to as high 2.23 billion m^3 of total runoff from the Greater Toronto Area
93 annually. While 6PPD-q specifically has not been studied in the Great Lakes, the Don River,

94 which feeds into Lake Ontario, has been documented to contain up to 2.85 µg/L of 6PPD-q
95 (Johannessen et al., 2021). If lake trout are sensitive to 6PPD-q exposure, particularly during
96 their early-life stages, it could represent an additional threat to population recovery and
97 maintenance efforts in these ecosystems. Therefore, lake trout are a crucial native species of
98 concern in Canada and the United States.

99

100 In this study, we aimed to determine the sub-chronic and acute sensitivity of lake trout alevins
101 and exogenously feeding fry, respectively, to exposure with 6PPD-q. In addition, this study
102 observed and described a number of sub-lethal effects of 6PPD-q exposure, which can provide
103 important insights into the mechanisms through which 6PPD-q exerts its high level of toxicity.
104 Together, this study provides important new information for environmental risk assessors and
105 regulators to understand the impacts of 6PPD-q on an important fish species with ecological,
106 commercial, and cultural relevance in North America.

107

108 MATERIALS AND METHODS

109 *Chemical Source*

110 6PPD-q was sourced from Toronto Research Chemicals, Cat# P348790, Lot # 6-ABK-122-2;
111 97% purity. Powder stock was made up in dimethyl sulfoxide (DMSO), at 10,000x the exposure
112 concentrations, resulting in a 0.01% DMSO (v/v) concentration for all treatments.
113 Deuterium-labeled 6PPD-q d5 (Cat# P348691) was also obtained from Toronto Research
114 Chemicals, and the neat compound was dissolved in LC-MS-grade methanol at 1 mg/L.
115 Exposure solutions were freshly prepared daily.

116

117 *Lake trout source and housing*

118 Lake trout eggs were sourced from fish collected from Whiteswan Lake (SK, Canada) in
119 conjunction with the Fisheries Department of the Government of Saskatchewan, Canada. Eggs
120 from ten different females were each fertilized using milt from two different males, resulting in
121 20 different genetic lines. Embryos were maintained in the dark at the University of
122 Saskatchewan Aquatic Toxicology Research Facility (ATRF) at $10 \pm 0.5^{\circ}\text{C}$ in a flow-through
123 heath tray system until they reached the eyed stage, checked twice daily for temperature and
124 mortalities, which were removed, as well as water quality (DO, ammonia, NH_3 , NH_4 , hardness)

125 (La Motte). Prior to hatching, embryos were transferred to larger glass aquaria for monitoring
126 and randomized exposure tank allocation. All experiments involving animals were reviewed and
127 approved by the University of Saskatchewan Animal Research Ethics Board (Animal Use
128 Protocol 2022-0002). Lake trout embryos were transported to the University of Saskatchewan
129 under the Government of Saskatchewan Fish Transport/Stocking permit FTP02-2022.

130

131 *Sub-chronic exposure*

132 Tanks (2.5 L) were individually aerated and randomized on shelves and allowed to equilibrate
133 with exposure water for 24 hours prior to exposure. Upon hatch, 15 lake trout alevins were
134 placed randomly in each tank and were exposed to nominal concentrations of 6PPD-q of 0.625,
135 1.25, 2.5, 5, 10, and 20 $\mu\text{g/L}$, as well as a solvent control (0.01% DMSO) with five replicate
136 tanks per treatment. Tanks were maintained under semi-static conditions, with a 70% water
137 change daily in an environmental chamber at a temperature of $10\pm0.5^\circ\text{C}$. Exposure water was
138 sampled at a time zero and time 24h (following a water change and before the subsequent water
139 change), over four separate time points throughout the exposure period, and frozen at -20°C
140 until analysis. Water quality measurements were taken weekly. As alevins approached swim-up,
141 they were fed *Artemia nauplii* once per day, then increased to twice per day after one week.
142 Upon the completion of the exposure, fry were euthanized in 150 mg/L buffered tricaine
143 mesylate (MS-222), weighed, and measured for total and standard length.

144

145 *Juvenile Acute Exposure*

146 Following the sub-chronic study, lake trout from the same genetic pool were exposed to 6PPD-q
147 for 96 hours at an age of eight weeks post-hatch. Based on results from the sub-chronic study,
148 fish were exposed to nominal 6PPD-q concentrations of 0.1, 0.3, 0.9, and 2.7 $\mu\text{g/L}$. Tanks were
149 again maintained in an environmental chamber at a temperature of $10\pm0.5^\circ\text{C}$. under semi-static
150 conditions, with a daily water change of 70%. Fish were not fed over the exposure period.
151 Following initiation of the exposure, fry were monitored for behavioral changes and mortality
152 hourly for the first seven hours, and twice daily for the remaining 96 hours. Exposure water was
153 again sampled at a time zero and time 24h, over three time points throughout the exposure
154 period, and frozen at -20°C . Upon the completion of the exposure, surviving fry were

155 euthanized in 150 mg/L of buffered MS-222, weighed, and measured for total and standard
156 length.

157

158 *Chemical Analysis*

159 Water samples, consisting of 950 μ L exposure water, and 50 μ L of deuterium-labeled internal
160 standard solution (1mg/L) were frozen at -20 °C in amber autosampler vials. Analytical
161 confirmation of exposure concentrations was achieved using ultra-high-performance liquid
162 chromatography in tandem with high-resolution mass spectrometry following the methods
163 described by ⁵. Calibration standard concentrations were within 15% of nominal, and target
164 quantification of 6PPD-q was done using TraceFinder 4.1. Concentrations were reported as time-
165 weighted average (TWA) across all replicates in a treatment.

166

167 *Data Analysis and Statistics*

168 Chronic percent mortality was calculated using the Kaplan-Meier function $S_{t+1} = S_t * ((N_{t+1} -$
169 $D_{t+1}) / N_{t+1})$, and LC50s were interpolated using the [agonist] vs. response – variable slope (four
170 parameters) model via Prism 10.1.2 for Windows (GraphPad Software, Boston, MA).

171

172 **RESULTS**

173 *Exposure Concentration Validation*

174 LC-MS/MS quantification resulted in TWAs of 0.22, 0.58, 1.3, 3.4, 7.6, and 13.5 μ g/L 6PPD-q
175 for the sub-chronic exposure. For the 96-hour juvenile exposure, TWA concentrations of 0.05
176 μ g/L in the lowest treatment, as levels were below the limit of detection, this represents half the
177 limit of detection, and 0.16, 0.55, and 2.1 μ g/L in the three highest exposure groups.
178 Concentrations in the lowest treatment were below the limit of detection, and were reported as
179 half the limit of detection (0.05 μ g/L). Water quality parameters were ammonia 0.03 ± 0.18 ppm,
180 nitrite 0.47 ± 0.58 ppm, nitrate 0.73 ± 0.23 ppm, total hardness 92.67 ± 9.27 , pH 8.23 ± 0.20 , and DO
181 $93 \pm 4.65\%$.

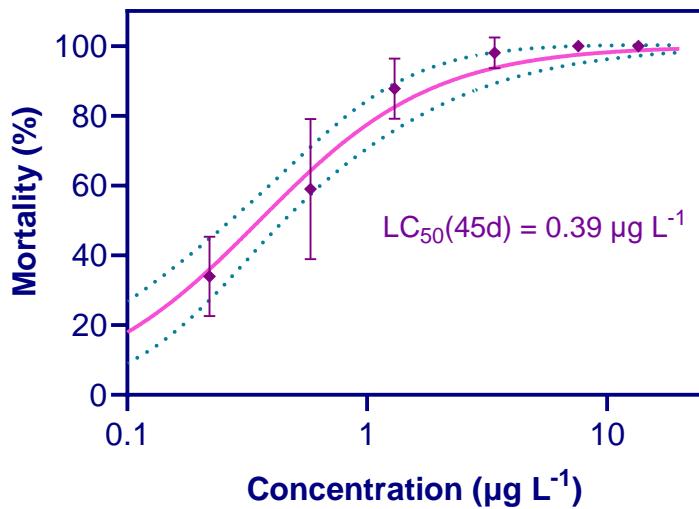
182

183 *Sub-chronic toxicity*

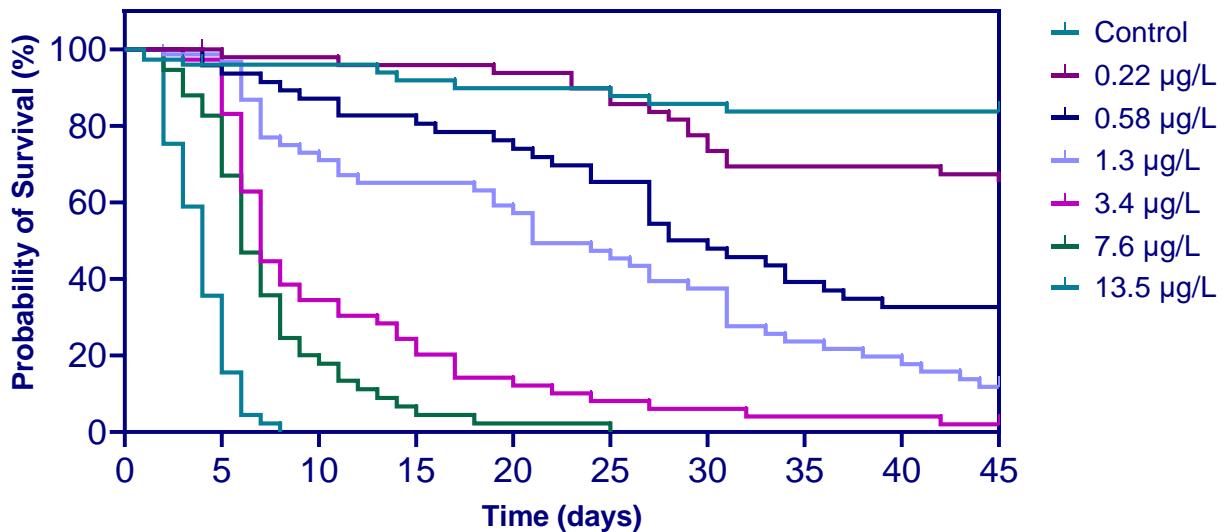
184 Lake trout were sensitive to exposure with 6PPD-q, resulting in significant mortality and
185 deformities (Figures 1-3). Significant mortality was observed four days post-initiation of

186 exposure at hatch and continued throughout the exposure period with a 45-day LC₅₀ of 0.39 µg/L
187 [95% Confidence Interval (CI) of 0.26 to 0.41 µg/L] (Figure 2). No behavioral changes were
188 noted. Several types of deformities were observed, with yolk sac edema, spinal curvature, and
189 pooling of blood in the tail and eye being the most prominent (Figure 3). These deformities were
190 noted within 72-hours of commencing exposure, and continued to occur until day 28, after which
191 no new deformities were observed. The most common deformity was yolk sac edema, which
192 occurred in all but one treatment. After 45 days of exposure, there was no significant difference
193 in total length or weight among treatments in those fish remaining.

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195
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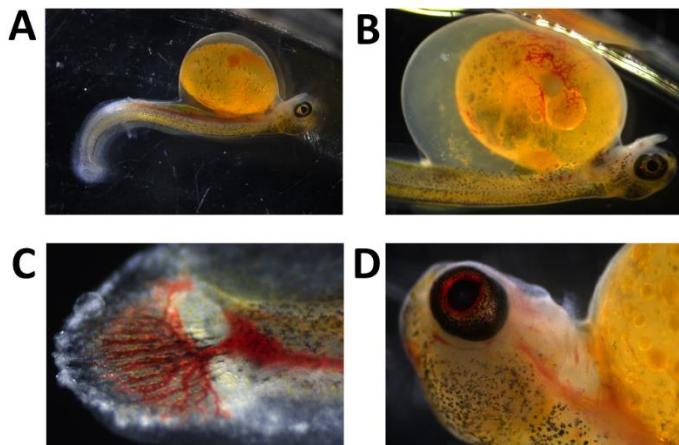


197
198 **Figure 1.** Percent mortality of early-life stage lake trout exposed to 6PPD-q for 45 days
199 beginning at hatch. The 45-day LC₅₀ was 0.39 µg/L, with a 95% CI of 0.30 to 0.48 µg/L. Points
200 represent mean of replicates for each concentration; standard deviation is represented by the bars.
201 The dotted lines indicate 95% confidence interval.



202

203 **Figure 2.** Kaplan-Meier survival analysis for early-life stag lake trout exposed to 6PPD-q for 45
204 days beginning at hatch.



205

206

207 **Figure 3.** Examples of morphological changes observed in early-life stage lake trout exposed to
208 6PPD-q for 45 days beginning at hatch. **A.** Example of spinal curvature (7.6 µg/L treatment). **B.**

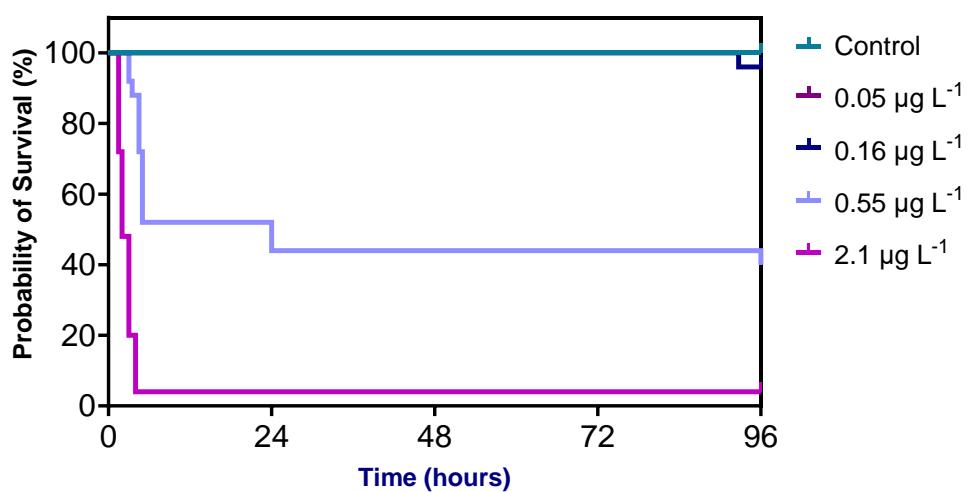
| | 13.5 µg/L | 7.6 µg/L | 3.4 µg/L | 1.3 µg/L | 0.58 µg/L | 0.22 µg/L | Solvent Control |
|------------------|-----------|----------|----------|----------|-----------|-----------|-----------------|
| Yolk sac edema | 2 | 4 | 11 | 4 | 0 | 5 | 1 |
| Blood pooling | 2 | 3 | 4 | 0 | 2 | 0 | 0 |
| Spinal curvature | 0 | 2 | 0 | 2 | 1 | 1 | 0 |

209 Example of yolk sac edema (3.4 $\mu\text{g/L}$ treatment). **C.** Example of caudal fin pathology (3.4 $\mu\text{g/L}$
210 treatment). **D.** Pooling of blood in the eye (7.6 $\mu\text{g/L}$ treatment). **E.** Total incidences of
211 morphological abnormalities over 45 days in each treatment (note: these represent total numbers
212 and were not normalized to mortalities).

213

214 *Juvenile Acute Exposure*

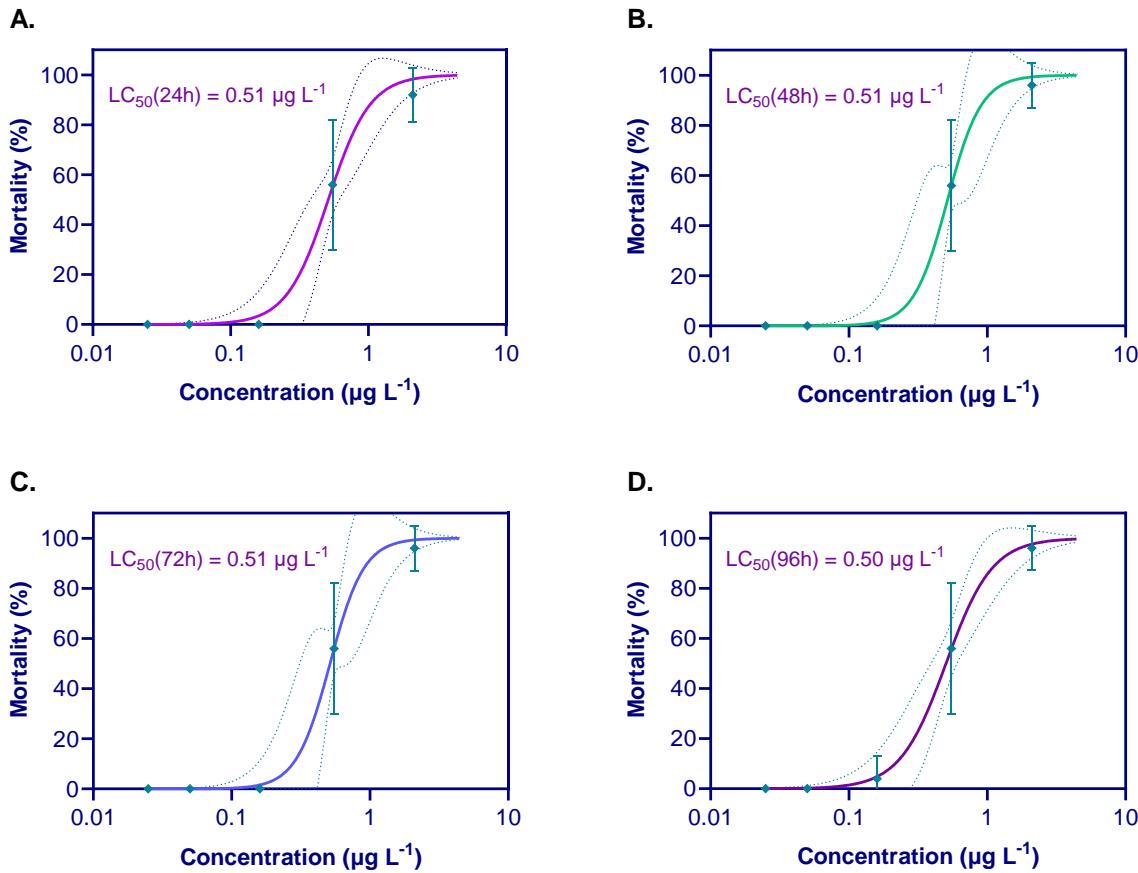
215 Exposure to 6PPD-q in the juvenile life stage resulted in a concentration-dependent increase in
216 mortality. Acute toxicity of 6PPD-q was apparent after only one hour of exposure of juvenile
217 lake trout exposed to a concentration of 2.1 $\mu\text{g/L}$ (Figure 4). The two highest treatment
218 concentrations of 2.1 and 0.55 $\mu\text{g/L}$ resulted in behavioural symptoms consistent with other
219 6PPD-q acute exposures, including loss of coordination, gasping, and surface swimming. All fish
220 which exhibited these symptoms died within six hours, and no symptomatic fish recovered. No
221 fish in the control, 0.16, or 0.05 $\mu\text{g/L}$ treatments exhibited any symptoms or
222 changes in behavior. There was no significant difference in total length or weight across treatments
223 following completion of the exposure (data not shown). The 96-hour LC₅₀ for the 8-week-old
224 juvenile lake trout was 0.50 $\mu\text{g/L}$ (95% CI of 0.42 to 0.60 $\mu\text{g/L}$) (Figure 5).



225

226 **Figure 4.** Time to survival analysis for 8-week-old lake trout exposed to 6PPD-q for 96 hours.

227



228

229 **Figure 5.** Percent mortality of 8-week-old lake trout exposed to 6PPD-q at times **A.** 24 hours **B.**
230 48 hours **C.** 72 hours **D.** 96 hours. Points represent mean of replicates for each concentration,
231 standard deviation is represented by the bars. The dotted lines indicate 95% confidence interval.
232

233 DISCUSSION AND CONCLUSIONS

234 The present study demonstrated that early-life stages of lake trout are sensitive to the exposure
235 with low, environmentally relevant concentrations of 6PPD-q, adding another species to the list
236 of salmonids susceptible to this emerging contaminant. Thus far, studies have elucidated a
237 notable range of sensitive species within the *Oncorhynchus* genus, including coho salmon and
238 rainbow trout^{8,15}, while other species in this genus such as Atlantic salmon (*Oncorhynchus keta*)
239 and westslope cutthroat trout are not^{16,17}. Interestingly, char species also exhibit either tolerance,
240 such as Arctic char⁸ and southern Asian dolly varden⁹, or high sensitivity, such as white-spotted
241 char⁹ and brook trout⁸. The acute 24-96-hr LC₅₀s of juvenile lake trout were very similar to the
242 24-hr LC₅₀s of white-spotted char (0.51 $\mu\text{g/L}$) and brook trout (0.59 $\mu\text{g/L}$)^{8,9}, suggesting that

243 these species of char may share some key similarity in terms of 6PPD-q toxicity. The wide
244 variation among species is a consistent finding in the study of 6PPD-q as an aquatic contaminant,
245 and so far, phylogeny alone does not seem to be a reliable predictor of sensitivity. Mitochondrial
246 dysfunction has been proposed as a mechanism of action for 6PPD-q toxicity ¹⁸, which has been
247 extrapolated upon ^{10,19} to include subsequent production of reactive oxygen species and changes
248 in membrane permeability as a mechanism of toxicity. Given the conservation of mitochondrial
249 and electron transport chain physiology among species, it is possible that toxicokinetic factors
250 are a key aspect of the wide-ranging sensitivity of species. The ability to clear 6PPD-q at an
251 adequate rate, or the production of a toxic metabolite(s) ^{9,20} may be the causative factor in
252 determining sensitivity of a species.

253

254 While early-life stages cannot be directly compared to adults, juvenile lake trout were among the
255 most sensitive species when comparing their acute LC₅₀ with those obtained from previously
256 reported experiments with adult or sub-adult fishes, surpassed only by juvenile and adult coho
257 salmon ^{11,15}. This high acute lethality may have significant implications for lake trout
258 populations, especially in Canada. According to the Lake Ontario Technical Committee: Lake
259 Trout Working Group, the failure of early-life survival is “likely the largest impediment to lake
260 trout restoration in contemporary Lake Ontario,” ²¹. With both early-life stage sub-chronic and
261 juvenile acute LC₅₀ falling within environmentally relevant levels, exposure to 6PPD-q may
262 further stress an already struggling population. The loss of early-life stage alevins not only
263 hampers all subsequent adult population maintenance efforts through a reduction in recruitment
264 but also severely diminishes population diversity by creating genetic bottlenecks ²²⁻²⁴.

265

266 In addition, it is possible that early-life stages of salmonids are exposed to greater concentrations
267 of 6PPD-q compared to those that are measured in the water column, or over longer periods of
268 time, given that TWP settle into sediments (Li et al., 2023). As TWP are washed into aquatic
269 environments, the 6PPD on the surface of the TWP is converted to 6PPD-q, the more stable
270 compound (Hu et al., 2023), which is still short-lived in the water column. This results in surface
271 waters experiencing intermittent pulses of 6PPD-q, correlated with precipitation (Greer et al.,
272 2023). However, the settling of TWPs in sediment may provide another, more constant and long-
273 term source of 6PPD-q. It is worth noting that early-life stage lake trout alevins inhabit this

274 gravel environment, in contrast to their adult counterparts which are predominantly found in the
275 water column in deeper parts of lakes. The newly-laid eggs incubate in the gravel for several
276 months, and after hatching, the alevins spend several weeks consuming their yolk-sac in the
277 gravel before swimming up to feed exogenously. Therefore, the period from hatching to swim-up
278 may be a critical time during which lake trout alevins are most vulnerable to the toxic effects of
279 6PPD-q.

280
281 While we observed significant mortality in both the juvenile acute and sub-chronic groups, only
282 those alevins exposed from hatch, in a sub-chronic manner, exhibited significant pathologies,
283 including spinal curvature, yolk sac edema, and pooling of blood in the caudal fin and eye. This
284 may be due to both the life stage and the duration of exposure. We found that the earliest
285 pathologies occurred within 72 hours of exposure in the post-hatch alevins, but not in juveniles.
286 This suggests that early growth and development are significantly disrupted by 6PPD-q at this
287 life stage. However, we also noted new pathologies up to 28 days into the exposure, indicating
288 that chronic exposure to 6PPD-q can result in sub-lethal effects in lake trout fry. These sub-lethal
289 effects could have population-level implications for overall survival in fry that are exposed to
290 low, sub-chronic, or chronic levels of 6PPD-q. It is important to note that spinal curvature can
291 impact a fish's ability to swim effectively ²⁵, affecting their ability to find food, escape from
292 predators, and reproduce. This has broad implications for the long-term survival and
293 development of lake trout fry. While none of these pathologies have been reported in associated
294 with 6PPD-q exposure, pooling of blood in the caudal fin and eye are consistent with previous
295 studies that have shown changes in vascular and blood-brain barrier permeability in coho salmon
296 following exposure to 6PPD-q ^{10,19}. It is worth noting that none of the fish exhibiting these
297 pathologies survived the exposure, suggesting that the physiological changes which lead to the
298 development of these pathologies, might be predictive, if not causative, of mortality.

299
300 In all previous studies, acute exposure of sensitive salmonid species to 6PPD-q resulted in
301 similar symptomologies of gaping, loss of coordination, and surface swimming. We observed
302 these same changes in our juvenile acute study, but no symptoms were observed in the sub-
303 chronic exposure. In addition, we also did not observe mortality at the same rate in the newly
304 hatched larvae versus the juveniles. This may be due to the under-development of the fish's brain

305 and nervous system or possibly an initial adaptive period in which 6PPD-q may be sequestered
306 in lipid-rich tissues like the yolk sac. However, it is important to consider that deformities in the
307 caudal fin, as well as pooling of blood in the eye, have the potential to impact a fish's swimming
308 ability and vision, respectively. Nevertheless, these changes only become relevant if any fry with
309 these deformities are able to survive for a significant period of time.

310

311 *Future Outlook*

312 This research demonstrated the high sensitivity of another salmonid species to the exposure with
313 6PPD-q, deepening the question as to what physiological components are involved in toxicity.
314 Given the conservation of mitochondrial function and ROS effects in fishes, this wide range of
315 species sensitivity may indicate a toxicokinetic driver of toxicity. Proposed by Hiki et al., the
316 presence of a monohydroxylated metabolite in exposed fishes may indicate that toxicity stems
317 from the production of a toxic metabolite in those fish species which are sensitive, and not in
318 those tolerant⁹. However, given the insensitivity of RT-Hep in contrast with RT-W1 gill, it is
319 also possible that sensitivity is determined by an organism's ability to clear 6PPD-q itself. The
320 difference in sensitive and tolerant species may come down to something as simple as a change
321 in a CYP isomer. This also brings into question potential differences in sensitivity between early
322 life stages and adult, as if toxicity is driven by toxicokinetic factors, those early-life stages with
323 reduced capacity for metabolism may also be an important factor in population-level effects of
324 6PPD-q exposure.

325 Thus, further studies looking at metabolite production in tolerant and sensitive species is key to
326 determining the role of toxicokinetics in this unusual species sensitivity distribution. In addition,
327 work to determine changes in gene expression could also help point toward a greater
328 understanding of the mechanism(s) of toxicity. Differential gene expression could provide key
329 insight as to these differences in sensitivity, as well as physiological changes, particularly those
330 at a sub-lethal level. Further to this, there are a number of other salmonid species for which there
331 is no sub-chronic or early-life stage information in regards to 6PPD-q exposure. Given that early-
332 life stages fishes are, on average 60%, more sensitive to contaminants¹², the degree of risk to
333 those species where the adults are known to be sensitive may be even greater. Understanding the
334 potential risk to these populations is integral to protecting these species, and in research and

335 recommendations for mitigation efforts toward limiting 6PPD-q exposure and Urban Runoff
336 Mortality Syndrome.

337

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349

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