

Direct Genome-Scale Screening of *Gluconobacter oxydans* B58 for Rare Earth Element Bioleaching

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Abstract

The transition to a sustainable energy economy will require an enormous increase in the supply of rare earth elements (REE). Bioleaching offers a promising alternative to conventional hydrometallurgical methods for REE extraction from low-grade ores. However, exploiting this potential remains challenging due to large gaps in our understanding of the genetics involved, and inadequate biological tools to address them. We generated a highly non-redundant whole genome knockout collection for the bioleaching microbe *Gluconobacter oxydans* B58, reducing redundancy by 85% compared to the previous best collection. This new collection was directly screened for bioleaching neodymium from a synthetic monazite powder, identifying 89 genes important for bioleaching, 68 of which have not previously been associated with this mechanism. We conducted bench-scale experiments to validate the extraction efficiency of promising strains: 8 demonstrated significant increases in bioleaching by up to 111% (*G. oxydans* δ GO_1598, a disruption of the gene encoding the orotate phosphoribosyltransferase enzyme PyrE), and one strain significantly reduced it by 97% (δ GO_1096, a disruption of the gene encoding the GTP-binding protein TypA). Notable changes in biolixiviant pH were only observed for 3 strains, suggesting an important role for non-acid mechanisms in bioleaching. These findings provide valuable insights into further enhancing REE-bioleaching by *G. oxydans* through targeted genetic engineering.

Introduction

Rare Earth Elements (REE; including the lanthanides ($Z = 57$ to 71), scandium, and yttrium) are crucial ingredients in present-day sustainable energy technologies including wind turbines^{1,2} and electric vehicles³, and in future ones like high-temperature superconductors⁴⁻⁷. As a result of this, the demand for REE is expected to grow by between 3- and 7-fold by 2040⁸. However, existing hydrometallurgical methods for REE mining pose considerable environmental risks⁹⁻¹¹.

The growing demand for an environmentally-friendly supply chain for REE has fueled interest in biotechnological processes for REE mining and recycling^{9,12-20}. Bioleaching solubilizes metals from minerals and end-of-life feedstocks with a microbially-secreted mineral-dissolving cocktail called a biolixiviant^{13,14,16,21-23}. In recent years, the search for more efficient bioleaching strategies has gained significant interest²³⁻²⁵.

The Generally Regarded As Safe (GRAS) acidophilic bacterium *Gluconobacter oxydans*²⁶⁻²⁹ has played a significant role in bioleaching research over the past decade^{14,21,30-33}. This microbe offers a unique capability for secretion of organic acids^{21,29,34} and creation of a highly acidic biolixiviant (primarily containing gluconic acid¹⁴) which is particularly effective for recovering REE from sources including minerals like allanite¹⁵; industrial residues like phosphogypsum³⁰, red mud³⁵, and blast furnace slag³⁶; and recycled materials like fluid cracking catalyst (FCC)²¹, nickel-metal-hydride (NiMH) batteries³⁷, and retorted phosphor powder (RPP)^{16,21,35}.

We have already improved bioleaching of REE by *G. oxydans* by first identifying genes involved in acid production with high-throughput screening¹⁶, and then engineering their regulation¹⁵. We increased REE-bioleaching from the REE-containing mineral allanite by up to 73% by up-regulating the membrane bound dehydrogenase, *mgdh*, and knocking out the phosphate signaling and transport system *pst*¹⁵.

Despite progress in enhancing REE-bioleaching with *G. oxydans*, further genetic improvements may be necessary to allow it to leapfrog traditional thermochemical extraction processes^{33,38}. One way to further improve it is to target the regulation of additional genes implicated in bioleaching by acid production screens¹⁶. However, it is proposed that metal solubilization in bioleaching happens through three processes: acidolysis via acid production, complexolysis by creating complexes between metal ions and organic compounds, and redoxolysis, which involves electron transfer and oxidative reactions³³. *G. oxydans*' biolixiviant appears to be more effective than pure gluconic acid controls at the same pH²¹. This could suggest that the biolixiviant contains additional components that amplify its effectiveness^{21,22,30,32,39}.

We hypothesize that further characterizing the mechanism of bioleaching and then engineering the composition of the biolixiviant is the route to future significant increases in bioleaching efficiency. But the acid production screens that we have used to date may not be sensitive to complexolysis and redoxolysis. Thus, to fully characterize the biolixiviant, it is necessary to identify genes directly linked to bioleaching, rather than just acid production alone. In this study, we conducted a genome-scale screen of a new quality-controlled *G. oxydans* B58 whole genome knockout collection with a high-throughput assay designed to directly identify genes involved in REE-bioleaching.

Results

***G. oxydans*' Quality-Controlled Collection Is 85% Smaller Than Previous Collection**

To facilitate direct high-throughput screening of mineral-dissolution, we built a highly non-redundant Quality-Controlled (QC) whole genome knockout collection for *G. oxydans* B58. The QC collection was built by removing redundancy from our earlier condensed collection (CC) of *G. oxydans* transposon disruption mutants¹⁶. Throughout this article, we refer to transposon disruption mutants with a ‘δ’ symbol (as opposed to a ‘Δ’ symbol for clean deletion mutants; e.g., δGO_1096).

In total, the QC collection contains a single carefully chosen transposon disruption mutant for 2,733 non-essential genes in the *G. oxydans* B58 genome arrayed onto 57 checker-boarded 96-well plates (**Supplementary Data S1**). The QC collection is 94% smaller than our *G. oxydans* saturating-coverage transposon mutant collection used to source all mutants in our earlier work (containing 49,256 mutants)¹⁶, and 85% smaller than our previous best curated collection (17,706 mutants)¹⁶. An overview of QC collection construction is shown in **Figure 1**, and collection statistics are shown in **Table 1**. Full details of QC collection construction can be found in **Materials and Methods**.

Genome-Scale Screen of QC Collection Identifies 68 Genes Not Previously Implicated in REE-Bioleaching

We performed a genome-scale screen of the QC collection to identify disruption mutants with differential REE-bioleaching capabilities. We developed a rapid microplate-based colorimetric assay with the REE-chelating dye Arsenazo-III (As-III) to screen for REE extraction^{18,40,41} from a synthetic monazite powder (**Figures 2A and 2B**) (monazite is the mineral host for REE at the Mountain Pass Mine in California, one of the largest REE mines in the world¹⁰). Unlike natural monazite (which contains multiple light lanthanides), this synthetic mineral contains just neodymium⁴², allowing greater consistency in the determination of extraction efficiency. Two synthetic monazite powder batches were synthesized (Mon1 and Mon2) as described by Balta *et al.*⁴², presenting minor differences in Nd-phosphate composition (**Supplementary Data S2**). Mon1 was composed of monazite-Nd (NdPO₄), rhabdophane-Nd (NdPO₄·2H₂O), and neodymium oxyphosphate (Nd₃PO₇). In contrast, Mon2 consisted solely of monazite-Nd and rhabdophane-Nd.

Mon1 demonstrated greater dissolution than Mon2 during bioleaching. This discrepancy is presumed to be due to the elevated concentration of neodymium oxides in Mon1 (**Supplementary Data S2**), along with the higher thermal expansion and oxidation state of the Nd₃PO₇^{43,44}. We utilized both Mon1 and Mon2 in our screening to leverage their distinct characteristics. Mon1 enhanced the detection sensitivity for potential hits in the initial preliminary survey, thereby expanding the selection pool for subsequent validation. Mon2, being less reactive, allowed for the isolation of the most significant outliers in the initial survey, offering a closer analog to bioleaching with natural monazite.

Our preliminary survey (**Supplementary Data S3**) identified 89 gene disruptions that produced significant changes in Nd extraction from the Mon1 synthetic monazite (**Figure 2C**). These hits included disruption of 21 genes previously linked to acidification¹⁶, and 68 genes that had not previously been linked to bioleaching (**Supplementary Data S4**).

A follow-up assay of these 89 mutants found 6 mutants that produced statistically significant changes to Nd-extraction from the Mon2 synthetic monazite (**Supplementary Data S5, Table 2**). We added an additional 6 mutants for follow up study that were prominent in the initial Mon1 Nd-extraction survey (**Table 2**).

Metal Binding and DNA Repair Gene Ontologies are Highly Enriched in Bioleaching Hits

The 68 genes not previously associated with REE-bioleaching that were identified through the initial Mon1 REE-extraction screen were analyzed for gene ontology (GO) enrichment with topGO⁴⁵. A total of four GO terms were found to be significantly enriched ($p < 0.05$ by Fisher's Exact Test), and to be associated with more than one gene (**Supplementary Data S6**). These four enriched GO terms belong to the biological process (BP) and molecular function (MF) groups (**Supplementary Data S6**). No significant terms belonging to the cellular component group were found.

The two most statistically-significantly enriched GO terms are metal ion binding and nucleotide-excision repair (**Figure 2D, Supplementary Data S6**). We speculate that disruption of these genes affects bioleaching indirectly by modifying the expression of metabolic pathways that produce metal solubilizing compounds. For example, metal ion binding proteins are often associated with signaling, sensing, and regulation⁴⁶⁻⁴⁹, suggesting that their disruption could alter *G. oxydans*' bioleaching capabilities. Similarly, disruption of the nucleotide-repair system could impair the ability of *G. oxydans* to respond to damage generated by oxidative stress resulting from initial biolixiviant production^{50,51} and thus alter further biolixiviant production.

Gene Disruptions of Interest Demonstrate Significant Changes in Bioleaching

The 12 gene disruption mutants that were highlighted by the Mon1 and Mon2 high-throughput assays (**Table 2**) were validated with bench-scale experiments where bioleaching was measured by Inductively Coupled Plasma - Mass Spectrometry (ICP-MS) (**Figure 3**). When compared to the proxy wild-type strain (pWT) (See "**Materials and Methods**"), 9 of the 12 disruption mutants generated significant changes in bioleaching ($p < 0.05$). Eight disruption mutants produced greater (with increases of between 56 and 111%) Nd-bioleaching than pWT, while only one mutant (δGO_1096) produced a reduction (-97%) in bioleaching (**Figure 3**). When compared to the wild-type strain (WT), all 12 disruption mutants tested produced significantly different extractions ($p < 0.01$) (**Supplementary Figure S1**), showing increases from 75 to 231%, and a 95% reduction by δGO_1096 .

Even though the increases in Nd-bioleaching created by 9 of the newly identified gene disruptions are large, they are notably smaller than those produced by our current two best-performing strains (*G. oxydans* $\Delta pstS$ and *G. oxydans* P₁₁₂:*mgdh*, $\Delta pstS$ ¹⁵; demonstrating 322% and 1,186% increase over pWT (**Figure 3**), and 564% and 1,922% over WT (**Supplementary Figure S1**).

Many Gene Disruptions of Interest Do Not Affect Biolixiviant pH

The pH of the biolixivants produced by the 12 gene disruption mutants selected was measured (**Figure 4**). Unsurprisingly, given our focus on identifying mutants absent in acid-production screens, yet still noteworthy, 9 of the 12 mutants did not exhibit significant changes in biolixiviant pH when compared to a proxy wild-type strain of *G. oxydans*. Only the δGO_102 , δGO_1096 , and δGO_1109 mutants produced

statistically-significant changes in biolixiviant pH (**Figure 4**). δGO_102 and δGO_1109 produced very modest reductions in biolixiviant pH of 0.06 and 0.07 units respectively, which are only barely above the precision-threshold of a pH meter. δGO_1096 produced a large increase in biolixiviant pH of 0.55 units.

Discussion

The generation of the QC collection facilitates the implementation of cost-effective and elaborate screening methods^{52,53}. The curated collection encompasses disruption mutants for 2,733 genes, one single representative for almost every non-essential gene in *G. oxydans*' genome¹⁶. The reduction in redundancy is crucial to improve the utility of a knockout collection, improving its value as a practical tool for the exploration of gene function^{53,54}. This streamlined collection was essential for conducting a technically challenging genome-wide screen for REE-bioleaching, leading to the identification of previously uncharacterized aspects of this mechanism. To our knowledge, this study represents the first direct whole genome screening for bioleaching of any mined metal.

In total, we identified 89 gene disruptions involved in Nd-bioleaching from synthetic monazite. Notably, 21 genes implicated in acidification, previously reported¹⁶, were among the hits found through this screen, corroborating their direct involvement in REE-bioleaching. This outcome verifies the reliability of our As-III screen for Nd-bioleaching but also underscores some challenges related to reduced sensitivity. However, we also identified an additional 68 gene disruptions that were tested in our earlier assays but were not implicated in bioleaching. Out of 12 disruption mutants selected for further study, 9 did not produce any statistically-significant change in pH (**Figure 4**). This indicates that the new screen is more comprehensive than pH-indicator screens for acid production. Furthermore, these results suggest that additional components might be present during bioleaching by *G. oxydans*^{13,30}, potentially increasing solubilization.

Notable and significant changes in bioleaching were validated by ICP-MS through bench-scale experiments (**Figures 3** and **4**). Out of the 12 mutants selected for verification, only one (δGO_1096) reduced extraction, suggesting that this gene normally facilitates bioleaching. In contrast, 11 of the mutants appear to regulate bioleaching, and their disruption removes the inhibitory controls on it.

Three results (δGO_1096 , δGO_1991 , and δGO_1968) imply that changes in stress response pathways can both improve or impair the bioleaching mechanism. This notion has been discussed by Sousa *et al.* after observing increased resistance to acetic acid at specific levels of oxidative stress in *Saccharomyces cerevisiae*⁵⁵.

Disruption of the gene encoding Formamidopyrimidine-DNA Glycosylase Fpg (*GO_1968*; belonging to both the metal ion binding and nucleotide excision repair gene ontologies terms) increased bioleaching. Fpg is a glycosylase involved in the repair of DNA damage caused by oxidative stress⁵⁶, and its disruption might lead to a heightened cellular response. For instance, it has been shown that certain levels of mistranslation can improve *Escherichia coli*'s tolerance to oxidative stress⁵¹. The metabolic response to accumulated DNA damage could involve transcription errors and shifts in secondary metabolite production. Such shifts might influence the types or quantities of solubilizing compounds produced by *G. oxydans*. Moreover, the gene coding for the Efflux ABC Transporter for Glutathione CydC (*GO_1991*) plays a role

in redox balance within the periplasm during oxidative stress conditions⁵⁷, and its disruption also increases bioleaching.

Notably, only a single gene disruption led to a decrease in bioleaching efficiency (δGO_1096). GO_1096 codes for the GTP-binding protein TypA which has been linked to stress response⁵⁸. This suggests that its disruption might hamper *G. oxydans*' ability to adapt to the stress generated during bioleaching, thereby diminishing efficiency. δGO_1096 was the only mutant that increased biolixiviant pH (out of only 3 that changed the pH at all). This result suggests TypA may facilitate acid-mediated bioleaching as well as non-acid bioleaching. We suspect that this gene was not found in our earlier acid-production screen¹⁶ because we swapped the GO_1096 disruption mutant used in our original curated collection¹⁶ for a new choice that was more likely to disrupt gene function. Interestingly, δGO_1096 's increase in biolixiviant pH is modest compared to those produced by mutants identified in our previous screen that produced similar reductions in REE-extraction. For example, disruption of the *tldD* gene, necessary for the synthesis of the PQQ molecule (a cofactor for the membrane bound glucose dehydrogenase), produced a 92% reduction in REE-extraction (admittedly from a different substrate) but raised biolixiviant pH by ≈ 0.7 units¹⁶.

Furthermore, we observed that the disruption of three genes involved in gene expression increased bioleaching: GO_1162 encoding the phosphate regulon regulatory protein PhoB, GO_1109 encoding a LysR substrate binding domain, and GO_1816 encoding the transcriptional regulator YafY.

PhoB is involved in the regulation of inorganic phosphate (Pi) homeostasis⁵⁹; it can positively or negatively regulate phosphate uptake in the presence of excess or limited Pi⁶⁰. Disrupting this transcriptional activator causes cells to lose the ability to modulate the phosphate regulon, which proves to be advantageous during bioleaching in phosphate-rich environments.

The LysR substrate binding domain binds specific ligands for signal recognition, and as a result, triggers the activity of transcriptional regulators⁶¹. Its disruption may influence gene repression or activation, altering the metabolic flux in a manner favorable for bioleaching. The regulatory protein YafY presents HTH (helix-turn-helix) and WYL (tryptophan, W; tyrosine, Y; and leucine, L) domains, which are known for their role in gene expression in response to DNA damage⁶². Disruption of the hypothetical protein GO_102 produced the second largest increase in Nd-bioleaching (Figure 3). *blastp* analysis⁶³ revealed 100% sequence alignment between GO_102 and an HTH-domain containing protein, suggesting it might also be involved in gene expression. These disruptions may lead to significant changes in gene expression levels, which could explain the increase in extraction.

Both δGO_102 and δGO_1109 disruption mutants decreased biolixiviant pH but were not identified by our earlier acid production screen. The pH changes caused by these mutants are modest compared with prominent mutants from our earlier acid production screen (which decreased pH by 0.1 to 0.2 units)¹⁶. We suspect that these pH changes were too small to be detected by our earlier screen. This leads us to presume that these mutants may be simultaneously altering acid and non-acid mechanisms of bioleaching.

We speculate that disruption of the hypothetical protein GO_2946 might enhance bioleaching by increasing the permeability of the outer membrane of *G. oxydans*. *blastp* analysis⁶³ showed homology of GO_2946 to a glycosyltransferase family 2 (GT2) protein, whose function involves the transfer of sugar moieties among

intermediates during the biosynthesis of polysaccharides⁶⁴. In a previous study, a GT2 protein was found to repress oxidative fermentation in *Gluconacetobacter intermedius*; and when disrupted, final yields of gluconic acid and acetic acid increased compared to the parental strain⁶⁴. Changes in the polysaccharide composition of the cell surface might increase the permeability of the membrane, facilitating the diffusion of substrate, therefore enhancing transport of the biolixiviant out of the cell⁶⁴.

The disruption of a TonB-dependent receptor gene (*GO_19*) increased bioleaching. This type of system is often associated with the uptake of iron bound to siderophores⁶⁵. Under iron starvation conditions, bacterial cells tend to up-regulate secretory mechanisms in an attempt to manage iron deficiency^{66,67}. Disruption of this gene might alter iron sensing and transport, prompting the cell to increase biolixiviant production.

Disruption of genes involved in nucleotide metabolism increased bioleaching (*GO_1598*, *GO_1113*, and *GO_2555*). The largest increase in bioleaching was produced by δ *GO_1598*. *GO_1598* codes for the orotate phosphoribosyltransferase enzyme PyrE which catalyzes the production of orotidine monophosphate (OMP), using phosphoribosyl diphosphate (PRPP) and orotate⁶⁸. One way that the disruption of *GO_1598* could increase bioleaching is by increasing availability of orotate, which is known to chelate metals⁶⁸, and in particular lanthanides⁶⁹. *GO_1113* codes for XdhC, an accessory protein involved in purine metabolism that binds to Molybdenum Cofactor (MoCo) for maturation of xanthine dehydrogenase⁷⁰. We speculate that its disruption could create an accumulation of MoCo, and accelerate other enzymatic reactions used in the production of the biolixiviant. *GO_2555* codes a hypothetical protein that demonstrated partial homology⁶³ (E-value = 1×10^{-125}) to the L-dopachrome tautomerase yellow-f2 protein (EC 5.3.2.3) involved in tyrosine metabolism that interacts with quinone and flavonoids to produce melanin⁷¹⁻⁷³. The enhanced bioleaching observed from this disruption may be attributed to the accumulation of intermediates, which can be utilized in other metabolic pathways associated with bioleaching.

Our results propose that the choice of bioleaching substrate can dramatically amplify (or perhaps negate) the effectiveness of genetic engineering. In our earlier work on bioleaching from retorted phosphor powder with *G. oxydans*¹⁶, the largest genetically-caused increase in bioleaching was 18% (due to disruption of *pstC*). Likewise, an engineered double mutant of *G. oxydans* (with clean deletion of *pstS*, and up-regulation of *mgdh* with the P₁₁₂ promoter) improved bioleaching from the mineral allanite by up to 73%¹⁵. In this work on bioleaching monazite, single gene disruptions can improve bioleaching by up to 111%, while the engineered double mutant improves bioleaching by 1,186% over proxy wild-type. It is easy to infer why the effectiveness of the *G. oxydans* biolixiviant could vary from substrate to substrate, as different minerals will have different chemistries that will be affected more or less by the bioleaching mechanisms. However, it is both more challenging and intriguing to explain why the same genetic interventions would produce different relative changes in bioleaching performance.

It remains unclear whether modification of the genes discussed in this article could lead to transformative advancements in bioleaching by *G. oxydans*. It is worth noting that when we first observed the effects generated by transposon disrupted genes, they were very modest compared with their subsequent genetic modifications. For example, disruption of *pstC* only increased bioleaching from retorted phosphor powder by 18% (and disruption of *pstS* did not significantly improve it at all)¹⁶, but clean deletion of *pstS* improved bioleaching from allanite by 30%¹⁵. Remarkably, combination of this knockout and up-regulation of *mgdh*

increased bioleaching from allanite by 73%¹⁵. This suggests (and it is emphasized that further testing is necessary) that multiple modifications inspired by the results presented here could have very big impacts on bioleaching performance from some substrates.

Conclusions

Our work pivots away from the well-characterized acidolysis mechanism to examine contributions to bioleaching via alternative routes. Our findings introduce additional genes that notably influence bioleaching yet remained undetected in acidification screens (**Figures 2** and **3**). Most genes that we identified do not significantly affect biolixiviant pH (**Figure 4**).

These results, along with earlier observations by Reed *et al.*²¹, strongly suggest that the bioleaching mechanism of *G. oxydans* is not purely acid-based. Our data indicates that disrupting genes involved in nucleotide synthesis, transcriptional regulation, DNA repair, nutrient uptake, and oxidative stress response could significantly impact bioleaching. But there is no clear evidence in this list that implies a single gene coding for a redox molecule (*e.g.*, a flavin) or chelator (*e.g.*, a siderophore) amplifies the effectiveness of bioleaching.

All of the gene disruptions we have identified appear to be involved in regulatory functions. This suggests that if there are genes involved in complexolysis or redoxolysis within the *G. oxydans* genome, they could be multiply-redundant, and that knocking out a single gene contributes only marginally to changes in bioleaching. Alternatively, these genes may possess some degree of essentiality, rendering their identification challenging. This highlights the need for broader and higher-sensitivity screening approaches that can capture a more diverse array of bioleaching-related genetic activities, potentially leading to further enhanced methods for harnessing and optimizing the bioleaching mechanism.

End Notes

Data Availability

The datasets generated and analyzed during the current study are available from the corresponding author (B.B.).

Code Availability

Code used in the generation of figures and analysis of data for this article is available from the corresponding author (B.B.).

Materials and Correspondence

Correspondence and material requests should be addressed to B.B. Individual strains (up to ≈ 10 at a time) are available at no charge for academic researchers. We are happy to supply a duplicate of the entire *G. oxydans* knockout collection to academic researchers but will require reimbursement for materials, supplies, and labor costs. Commercial researchers should contact Cornell Technology Licensing for licensing details.

Acknowledgments

This work was supported by Cornell University startup funds, an Academic Venture Fund award from the Atkinson Center for Sustainability at Cornell University, a Career Award at the Scientific Interface from the Burroughs Wellcome Fund to B.B., ARPA-E award DE-AR0001341 to B.B., M.H., E.G., and M.W., and a gift from Mary Fernando Conrad and Tony Conrad to B.B. S. Marecos. was supported by a Link Foundation Graduate Fellowship. A.S. was supported by a Cornell Energy Systems Institute Postdoctoral Fellowship. S.A.M. was supported by a Cornell Presidential Life Sciences Graduate Fellowship.

Contributions

Conceptualization, S. Marecos. and B.B.; Methodology, S. Marecos. and B.B.; Investigation, S. Marecos., B.P., S.A.M., and A.S.; Writing—Original draft, S. Marecos. and B.B.; Writing—Review and editing, S. Marecos., B.B., J.B.B., M.W., E.G., S.A.M., and A.S.; Funding acquisition, S. Marecos., S.A.M., A.S., M.W., M.H., E.G., and B.B.; Resources, M.C.R., E.G., J.B.B., and B.B.; Supervision, M.W., M.H., E.G., M.C.R., and B.B.; Data curation, S. Marecos. and B.B.; Visualization, S. Marecos. and B.B.; Formal analysis, S. Marecos.

Competing Interests

The authors are pursuing patent protection for engineered organisms using knowledge gathered in this work (US provisional patent application number 63/653,203). A.S. and S.A.M. are co-founders of, and B.B. is a contributor, and uncompensated advisor to, REEgen, Inc., which is developing genetically engineered microbes for REE bio-mining. The remaining authors declare no competing interests.

Figures for:

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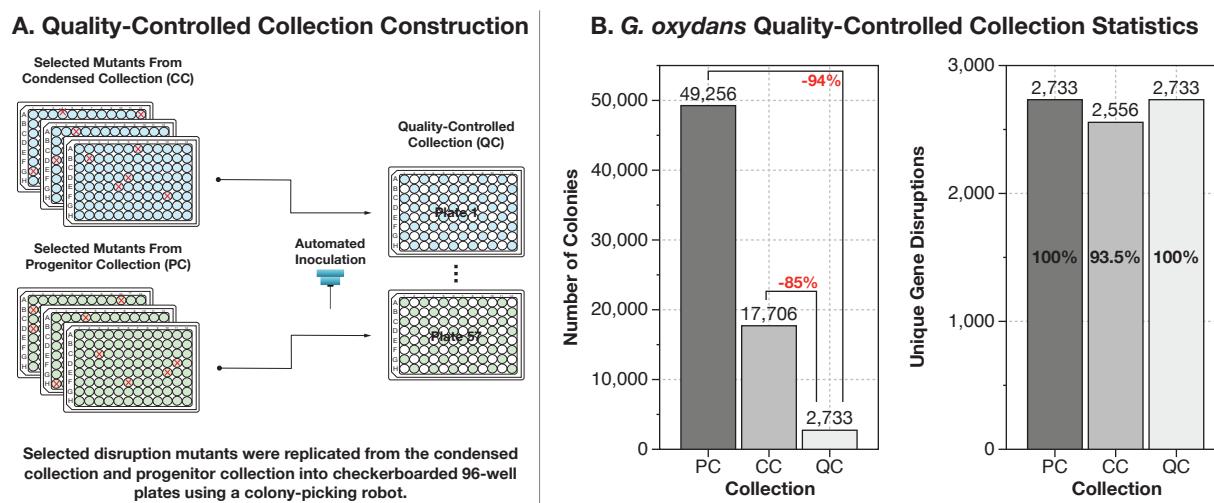


Figure 1. Construction of a quality-controlled (QC) collection containing a single disruption mutant for 2,733 genes in *Gluconobacter oxydans*' genome. (A) The QC collection was built by choosing the best available mutants from an earlier condensed collection (CC) of *G. oxydans* mutants¹⁶, and by supplementation from the originating saturating-coverage transposon collection (Progenitor Collection or PC)¹⁶. Mutants were transferred from the earlier collections by fluid transfer into fresh 96-well plates with a colony picking robot. (B) The QC collection contains 2,733 unique mutants distributed across 57 checkerboarded 96-well plates. This collection represents a 94% reduction in size from the PC, and an 85% reduction in size from the CC, indicated in red. It is worth noting, that the CC collection contained disruption mutants with confirmed identity for 2,556 genes (93.5% of the 2,733 genes disrupted in the PC), whereas the QC collection aims to include a disruption mutant for 100% of genes disrupted in the PC.

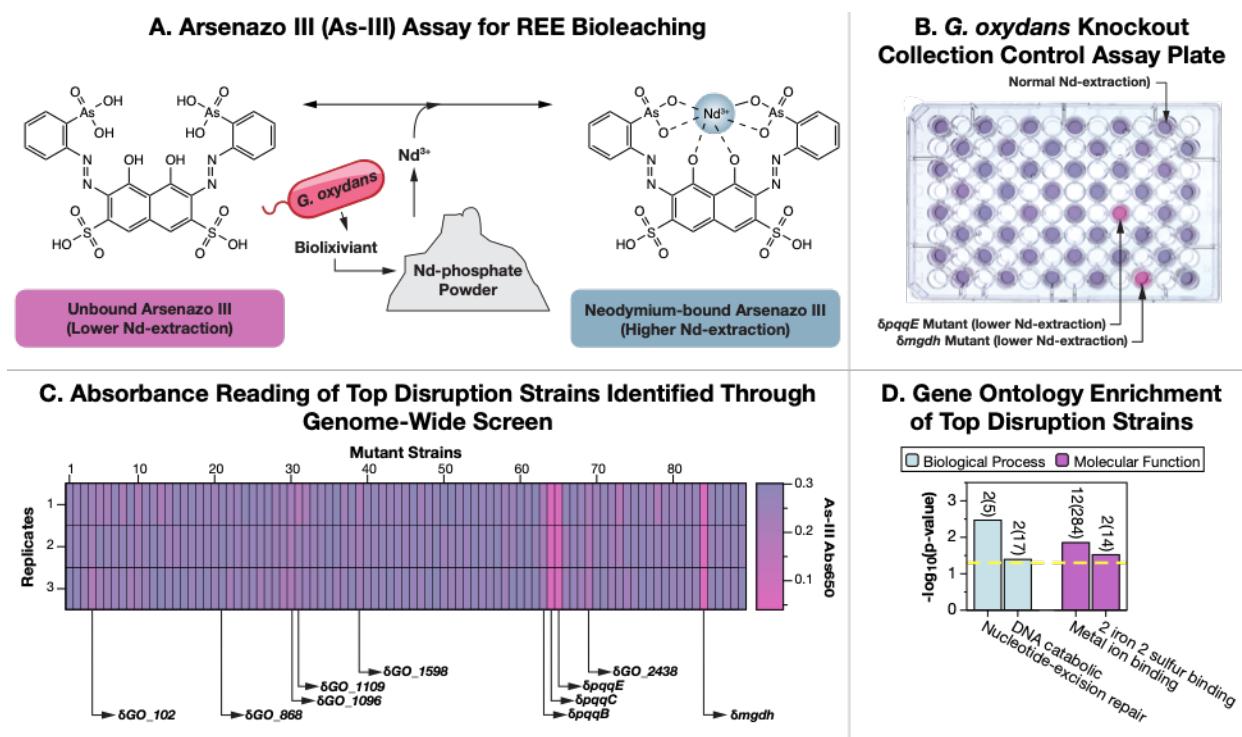


Figure 2. Direct bioleaching screening of the quality-controlled whole genome knockout collection of *Gluconobacter oxydans* identifies 68 gene disruptions not previously associated with bioleaching. (A) We developed a rapid 96-well plate based high-throughput screen for REE-bioleaching that uses a colorimetric assay with the Arsenazo III (As-III) REE-chelating dye in presence of synthetic monazite. This assay directly detects REE release, facilitating the identification of disruptions with differential bioleaching capabilities. (B) Color differences in each well identify which genes control bioleaching. Gene knockouts that reduce bioleaching leave the As-III dye unbound and pink, while knockouts that increase bioleaching turn the dye blue. (C) The genome-scale screening of the QC collection identified 89 hits including 68 gene disruptions not previously associated with the bioleaching capabilities of *G. oxydans*. (D) Gene Ontology (GO) Enrichment was performed with a Fisher's Exact Test ($p < 0.05$, yellow dashed line) to analyze the hits identified. Significantly enriched GO terms with at least 2 representatives are displayed, and numbers above the bars represent the significant hits out of the total annotated genes associated with that gene ontology in the genome of *G. oxydans* (the full list of GO terms can be found in **Supplementary Data S5**). Only terms belonging to the Biological Process and Metabolic Function GO groups were significantly enriched.

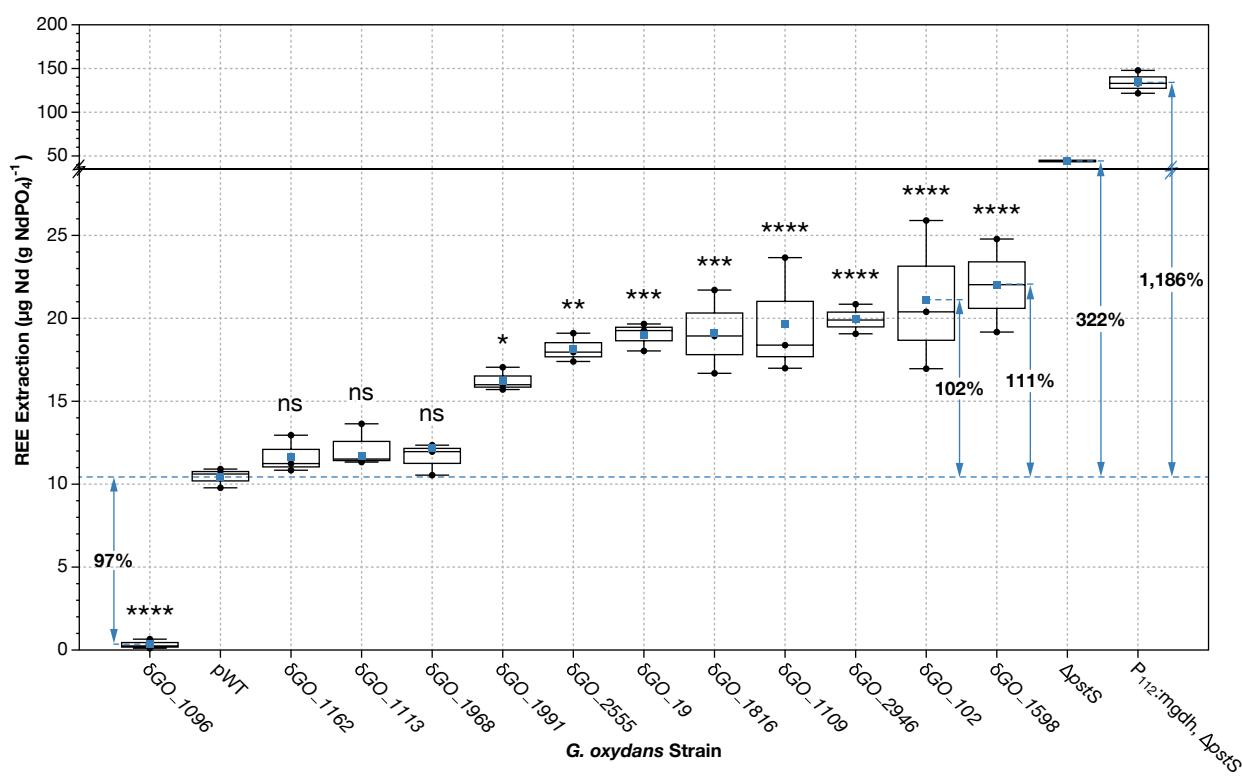


Figure 3: Identified *Gluconobacter oxydans* gene disruptions raised REE-bioleaching by up to 111% or lowered it by 97%. Direct measurement of bioleaching extraction was performed on disruption mutants of interest following bioleaching experiments with synthetic monazite, using ICP-MS analysis. REE extractions were analyzed with pairwise comparisons among disruption mutants ($n = 3$) and pWT ($n = 3$). Levels of extraction significantly different from pWT are labeled with asterisks ($*p < 0.05$; $**p < 0.01$; $***p < 0.001$, $****p < 0.0001$) or “ns” if $p > 0.05$, denoting statistical significance after Bonferroni-corrected t -tests ($N = 12$). Blue squares indicate the mean extraction for each mutant, the center line denotes the median, boxes show the upper and lower quartiles, and whiskers extend to the range of data points within 1.5 times the interquartile range. Strains $\delta\text{GO-1162}$, $\delta\text{GO-1113}$, and $\delta\text{GO-1968}$ did not show statistically-significant changes in extraction compared to pWT. Only one disruption mutant, $\delta\text{GO-1096}$, decreased bioleaching, demonstrating a 97% reduction in extraction. Current best-performing engineered strains, *G. oxydans* ΔpstS and *G. oxydans* $\text{P}_{112:\text{mgdh}}, \Delta\text{pstS}$, were included to contrast the extractions of the tested disruption mutants. Significant changes in extraction efficiency compared against the WT strain can be found in **Supplementary Figure S1**.

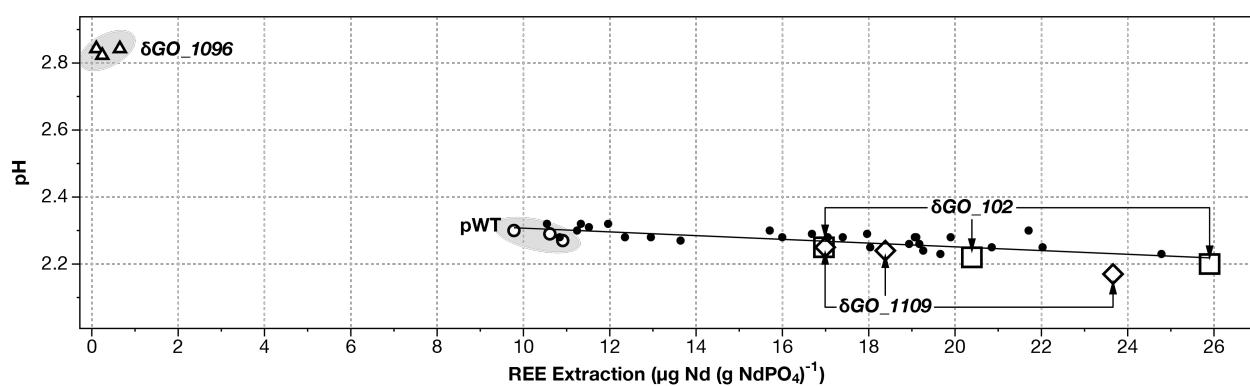


Figure 4: Only three of the identified *Gluconobacter oxydans* gene disruptions produce statistically-significant changes in biolixiviant pH. REE extraction and biolixiviant pH were plotted for each strain selected for direct bioleaching measurement. Strain replicates that significantly changed the biolixiviant pH compared to proxy wild-type are indicated as follows, pWT “○”, δGO_102 “□”, δGO_1109 “◇”, and δGO_1096 “Δ”. Other gene disruptions with data points falling within the model are represented as “●”, including data points for δGO_19, δGO_1113, δGO_1162, δGO_1598, δGO_1816, δGO_1968, δGO_1991, δGO_2555, and δGO_2946.

Collection	Number of Mutants	Distance from Translation Start				
		< 50bp	≥ 50bp	≤ 0.5	> 0.5	≥ 50 bp AND ≤ 0.5
Progenitor Collection (PC)	285	21	264	172	113	153
Condensed Collection (CC)	2,448	240	2,208	2,037	411	1,797
Total (QC Collection)	2,733	261	2,472	2,209	524	1,950

Table 1. Overview of *Gluconobacter oxydans* B58 quality-controlled (QC) whole genome knockout collection statistics. This table summarizes the transposon location characteristics of the mutants derived from the progenitor collection (PC) and condensed collection (CC) used to generate the QC collection. Mutants were categorized based on the distance of the disruption from the translation start of the gene: less than 50 base pairs, at least 50 base pairs, within 0.5 fractional distance of the gene, beyond 0.5 fractional distance, and disruptions located at least 50 base pairs downstream and within 0.5 fractional distance, which were considered the optimal locations for disruption of gene function.

Gene Identifier	Feature Disrupted	Extraction Against pWT (%)
<i>GO_1096</i>	GTP-binding protein TypA/BipA	-97
<i>GO_1598</i>	Orotate phosphoribosyltransferase (EC 2.4.2.10)	+111
<i>GO_102</i>	hypothetical protein	+102
<i>GO_1109</i>	Transcriptional regulator, LysR family	+89
<i>GO_1113</i>	Xanthine and CO dehydrogenases maturation factor, XdhC/CoxF family	+12
<i>GO_2555</i>	hypothetical protein	+74
<i>GO_2946</i>	hypothetical protein	+91
<i>GO_1162</i>	Phosphate regulon transcriptional regulatory protein PhoB (SphR)	+11
<i>GO_19</i>	TonB-dependent receptor	+82
<i>GO_1968</i>	Formamidopyrimidine-DNA glycosylase (EC 3.2.2.23)	+17
<i>GO_1991</i>	Efflux ABC transporter for glutathione/L-cysteine, CydC subunit	+56
<i>GO_1816</i>	Transcriptional regulator, YafY family	+83

Table 2. Selected gene disruptions for direct measurement of bioleaching. This table lists the selected gene disruption mutants for analysis, specifying their gene identifiers and disrupted features. It also presents the percentage change in extraction efficiency compared to pWT, where positive values indicate enhanced bioleaching, and negative values denote a reduction.

Material and Methods for:

Direct Genome-Scale Screening of *Gluconobacter oxydans* B58 for Rare Earth Element Bioleaching

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Materials and Methods

Curation of the Quality-Controlled Whole Genome Knockout Collection

The QC collection was built by removing redundancy from an earlier condensed collection (CC) of *G. oxydans* transposon mutants and supplementing with additional mutants from a saturating-coverage progenitor collection (PC) from the same work¹⁶. The progenitor collection of transposon mutants was catalogued using the Knockout Sudoku probabilistic inference combinatorial pooling method^{52,74}.

The best-choice disruption mutant for each gene in the QC collection was selected by considering the location of the transposon disruption in the gene, and the reliability of the location assignment of the mutant. We followed the generally-accepted consensus that the best location for a transposon to disrupt gene function is between 50 bp from the start of the coding region and the middle of the gene (a fractional distance of ≤ 0.5 into the gene). Furthermore, we preferred to select mutants which location was directly inferred by the Knockout Sudoku algorithm (also known as unambiguous location inference^{52,74}), rather than relying upon probabilistic location inference (also known as ambiguous location inference^{52,74}). If given the choice between a mutant with non-ideal locations and unambiguous location inference, and a mutant in the same gene with ideal location and ambiguous location inference we chose the former. Mutant genomic location statistics are shown in **Table 1**. A catalog of the QC collection is included in **Supplementary Data S1**.

Mutants were transferred from the earlier collections by fluid transfer with a colony picking robot (CP7200, Norgren Systems, Ronceverte, West Virginia, USA) into fresh 96-well polypropylene storage plates containing yeast peptone mannitol (YPM) media (5 g L⁻¹ yeast extract (catalog number C7341, Hardy Diagnostics, Santa Maria, California, USA); 3 g L⁻¹ peptone (catalog number 211677, BD, Franklin Lakes, New Jersey, USA); 25 g L⁻¹ mannitol (catalog number BDH9248, VWR Chemicals, Radnor, Pennsylvania, USA)) and 100 μ g mL⁻¹ kanamycin. Plates were incubated at 30 °C and 800 rpm in a Multitron shaker (Infors Inc., Laurel, Maryland, USA) for 3 nights to achieve saturation. Following incubation, stocks were prepared by adding glycerol (catalog number G5516, Sigma-Aldrich, St. Louis, Missouri, USA) to 20% (v/v) to each well for long term storage at -80 °C.

Genome-Scale REE-Bioleaching Screen

REE concentrations were measured using the REE-chelating Dye Arsenazo-III (As-III; catalog number 110107, Sigma-Aldrich, St. Louis, Missouri, USA). The As-III solution was prepared following the method described by Hogendoorn *et al.*⁴⁰.

The high-throughput screen consisted of a preliminary survey carried out in duplicate followed by a validation screen performed in triplicate. It spanned several weeks and was partitioned into batches of 10-15 plates. Plates containing mutants to be analyzed were replicated from the QC collection using a 96-pin cryo-replicator press (catalog number CR1100, EnzyScreen, Heemstede, Netherlands) into 96-well polypropylene round-bottom plates filled with 150 μ L YPM supplemented with 100 μ g mL⁻¹ kanamycin.

Replicated plates were incubated for two nights at 30 °C, 80% humidity, and 800 rpm in a Multitron shaker (Infors Inc., Laurel, Maryland, USA). Then, cultures were back-diluted either in duplicate or triplicate, by inoculating 5 μ L of bacterial culture in 120 μ L of the same medium using the PLATEMASTER pipetting

system (catalog number F110761, Gilson Inc., Middleton, Wisconsin, USA). After two nights of incubation, the optical density was measured using a plate reader (catalog number 18531, Biotek, Charlotte, Vermont, USA), and the biolixiviant production was set up by measuring the remaining volume in each well and adding glucose (catalog number 0188, VWR Chemicals, Radnor, Pennsylvania, USA) to 20% w/v. Plates were incubated for 24 hours at 30 °C, 80% humidity, and 800 rpm. For all the aforementioned steps, 96-well plates were covered with a breathable rayon film (catalog number 60941-086, VWR Chemicals, Radnor, Pennsylvania, USA).

For bioleaching, 150 µL of biolixiviant was transferred from each well into plates containing synthetic monazite powder covered with an adhesive polypropylene sealing Film (PCR-TS, Axygen Scientific, Union City, CA) and were incubated for 24 hours at 30 °C and 800 rpm. Final plates containing the leachate were centrifuged at 3,214 × g for 12 minutes to obtain the supernatant to be used for analysis.

For the preliminary survey, bioleaching was performed in presence of synthetic monazite mineral Mon1 (**Supplementary Data S2**) at 0.33% pulp density to facilitate an initial identification of disruption strains with differential bioleaching capabilities. For the screen with Mon2 (**Supplementary Data S2**), the pulp density was 1%. After centrifugation, a 1:10 dilution of the supernatant from each well was transferred to polystyrene flat-bottom 96-well plates filled with 100 µL of 60 µM AS-III Solution. For validation with Mon2, 100 µL of the supernatant was used instead. Analysis plates were shaken for 1 minute, then the absorbance at 650 nm was measured with the plate reader.

REE-Bioleaching Screen Analysis

Several factors were considered to confirm the quality of the data collected, namely cross contamination of wells, systematic differences among wells caused by their position in the plate, and additional errors generated during manipulation.

To address these challenges, the raw data collected through our preliminary high-throughput survey with Mon1 was analyzed with a correcting software⁷⁵ to detect and remove systematic errors. This software allowed us to normalize our data based on Z-score transformation within plates⁷⁶. Wells with growth OD600 < 0.2 were not considered to avoid the potential offsets during the normalization process.

An initial hit identification was carried out by comparing the normalized absorbance values of the disruption mutants to the mean of the plate they were analyzed within. In a conservative manner, disruption mutants presenting changes of 1.5 standard deviation away from the plate mean were selected for further screening.

For our screen with Mon2, selected disruption mutants' wells were picked to inoculate new 96-well plates filled with media and supplemented with antibiotic. To guarantee reliable data collection we again arrayed the mutants into a checkerboard pattern; included each mutant in triplicate; avoided perimeter wells to reduced media evaporation; and included a proxy wild-type strain within each plate.

Raw absorbance data was used for statistical analysis, which included pairwise comparisons conducted utilizing the emmeans package in R, while applying a false discovery rate (FDR) correction for *p*-value adjustment for multiple comparisons⁷⁷. Disruption mutants that demonstrated significant differences in REE extraction were considered promising.

Gene Ontology Enrichment Analysis

A gene ontology (GO) enrichment analysis was carried out as described by Schmitz *et al.*¹⁶. Enrichment was performed with the BioConductor topGO package^{45,78}, employing the TopGO Fisher test with the default weight algorithm, applying a significance threshold of 0.05.

Direct REE-Bioleaching Measurements

Selected disruption mutants that included five significant hits identified through the REE-bioleaching screen and other seven additional disruption mutants were used for further validation through bench-scale bioleaching experiments⁷⁹ with Mon2. Additional disruption mutants were picked based on the features disrupted that represented a trait of interest, these included specific enzymes or disruptions with interesting behavior throughout the screen.

A total of 12 disruption mutant strains were plated on YPM and kanamycin agar (05039, VWR Chemicals, Radnor, PA) petri dishes for colony isolation. Three biological replicates for each disruption mutant were used to inoculate 3 mL cultures, as well as for wild-type controls in addition to no-bacteria control. After incubation for two nights at 30 °C and 200 rpm in a shaking incubator (29314, Infors Inc., Laurel, MD), cultures were normalized to OD590 0.01 in 50 mL flasks with a final volume of 5 mL and grown for another two nights. Following incubation, the cultures were diluted in half glucose 40% (w/v) to reach a final concentration in solution of 20%. Flasks were incubated at 30 °C and 200 rpm for 48 hours to allow biolixiviant production.

Bioleaching experiments were set up to digest Mon2 at 1% pulp density, for 24 hours at 30 °C and 200 rpm. Once obtained, the leachate was filtered through a 0.45 µm AcroPrep Advance 96-well filter plate (8029, Pall Corporation, Show Low, AZ, USA) by centrifugation at 1,500 × g for 5 minutes.

Each sample underwent a 20-fold dilution using 2% trace metal grade nitric acid (JT9368, J.T. Baker, Radnor, Pennsylvania, USA). Neodymium concentration in each sample was measured using an Agilent 7800 ICP-MS. Calibration and verification utilized a REE mix standard (67349, Sigma-Aldrich, St. Louis, Missouri, USA) and a rhodium internal standard (catalog number 04736, Sigma-Aldrich, *m/Z* = 103). Rigorous quality checks included intermittent assessment of standards, blanks, and replicate samples⁸⁰. Data interpretation was carried out using MassHunter software, version 4.5.

A proxy wild-type strain (pWT) was selected for comparison to account for changes in the wild-type background introduced by the presence of the antibiotic cassette in the genome. The selected pWT contains a transposon insertion in an intergenic region (at base pair 1,262,471; between genes *GO_2147* and *GO_2148*) and does not demonstrate any notable phenotypic changes. This strain can be located in plate 130, well A2 in the progenitor collection.

For each disruption mutant, bioleaching measurements ($n = 3$) were evaluated against those of the wild-type and proxy wild-type strains ($n = 3$). This was done performing pairwise comparisons with Bonferroni corrected *t*-tests using the emmeans package in R. The total REE extracted was deemed significantly different if $p < 0.05/N$ ($N = 12$).

Limitations of the Study

During high-throughput screening, growth hindrance due to strain disruption poses considerable challenges. For example, despite the *pstS* gene disruption being flagged as a key player in REE-bioleaching during preliminary testing, its pronounced effect on microbial growth required its exclusion from final analyses because it did not meet the established growth threshold for inclusion.

Future studies could address these growth-related issues by incorporating an up-regulation collection into the screening process. This approach would not only enhance the insights gained from knockout collections but also allow for the potential identification of essential genes as significant contributors to bioleaching. Furthermore, confirmation of gene involvement of hits identified in this study will require further experiments, including gene deletion and complementation, to fully evaluate gene function and interaction.

While our study adds to the growing body of work on microbial metal recovery, the application of engineered microbes in bioleaching is still emerging. Successes have been noted with engineered strains in controlled settings such as stirred-tank reactors. Nonetheless, advancing this technology will require careful consideration of containment strategies and regulatory adjustments to appropriately mitigate potential risks.

Statistics and Reproducibility

In our genome-scale screen, statistical robustness was ensured through multiple replicates and a rigorous validation approach. Our screen included a preliminary survey carried out in duplicate, followed by a validation screen performed in triplicate. This multistep approach allowed for the establishment of robustness in our findings. To control for potential biases, we considered factors such as cross-contamination of wells, position-related systematic noise, and errors during manipulation.

The data from our preliminary survey with Mon1 was corrected using specialized software to remove systematic errors⁷⁵, and Z-score transformation was applied within plates for normalization. Wells that showed growth OD600 less than 0.2 were excluded to avoid potential offsets during this normalization process. Disruption mutants exhibiting changes of 1.5 standard deviations from the plate mean were selected for further screening.

In the subsequent validation screen with Mon2, disruption mutants were subjected to a more stringent validation process. This included a balanced distribution of mutants across the plates, triplicate data collection, and within-plate comparison strains. Statistical significance for the REE extraction capabilities of each mutant was determined using pairwise comparisons with the emmeans package in R and applying a false discovery rate (FDR) correction for *p*-value adjustment in multiple comparisons.

Bench-scale bioleaching experiments further confirmed the reproducibility of our findings, with each mutant strain's performance benchmarked against that of the wild-type and the proxy wild-type strains. Neodymium extraction levels measured by ICP-MS were compared pairwise, with significance established at *p* < 0.05 after Bonferroni correction (*N* = 12). These stringent statistical analyses ensured that the results presented are both robust and reproducible, providing a reliable foundation for future explorations into the genetic determinants of REE-bioleaching.

References

- 1 Dent, P. C. Rare earth elements and permanent magnets (invited). *Journal of Applied Physics* **111** (2012). <https://doi.org/10.1063/1.3676616>
- 2 Bauer, D. *et al.* U.S. Department of Energy Critical Materials Strategy. Medium: ED (United States, 2010).
- 3 Humphries, M. Rare Earth Elements: The Global Supply Chain. Report No. <https://digital.library.unt.edu/ark:/67531/metadc272008/>, (Washington D.C., 2013).
- 4 Schubert, E. F. & Kim, J. K. Solid-State Light Sources Getting Smart. *Science* **308**, 1274-1278 (2005). <https://doi.org/doi:10.1126/science.1108712>
- 5 Peng, F. *et al.* Hydrogen Clathrate Structures in Rare Earth Hydrides at High Pressures: Possible Route to Room-Temperature Superconductivity. *Physical Review Letters* **119**, 107001 (2017). <https://doi.org/10.1103/PhysRevLett.119.107001>
- 6 Ivanovskii, A. L. New high-temperature superconductors based on rare-earth and transition metal oxyarsenides and related phases: synthesis, properties and simulations. *Physics-Uspekhi* **51**, 1229 (2008). <https://doi.org/10.1070/PU2008v05n12ABEH006703>
- 7 Zurek, E. & Bi, T. High-temperature superconductivity in alkaline and rare earth polyhydrides at high pressure: A theoretical perspective. *The Journal of Chemical Physics* **150** (2019). <https://doi.org/10.1063/1.5079225>
- 8 IEA (2021). (International Energy Agency, Paris).
- 9 Castro, L., Blázquez, M. L., González, F. & Muñoz, J. Biohydrometallurgy for Rare Earth Elements Recovery from Industrial Wastes. *Molecules* **26** (2021). <https://doi.org/10.3390/molecules26206200>
- 10 Bauer, D. *et al.* (ed U.S. Department of Energy) (2011).
- 11 Lucas, J., Lucas, P., Le Mercier, T., Rollat, A. & Davenport, W. in *Rare Earths* (eds Jacques Lucas *et al.*) 47-67 (Elsevier, 2015).
- 12 Rasoulnia, P., Barthen, R. & Lakaniemi, A.-M. A critical review of bioleaching of rare earth elements: The mechanisms and effect of process parameters. *Critical Reviews in Environmental Science and Technology* **51**, 378-427 (2021). <https://doi.org/10.1080/10643389.2020.1727718>
- 13 Brisson, V. L., Zhuang, W.-Q. & Alvarez-Cohen, L. Bioleaching of rare earth elements from monazite sand. *Biotechnology and Bioengineering* **113**, 339-348 (2016). <https://doi.org/https://doi.org/10.1002/bit.25823>
- 14 Thompson, V. S. *et al.* Techno-economic and Life Cycle Analysis for Bioleaching Rare-Earth Elements from Waste Materials. *ACS Sustainable Chemistry & Engineering* **6**, 1602-1609 (2018). <https://doi.org/10.1021/acssuschemeng.7b02771>
- 15 Schmitz, A. M. *et al.* High Efficiency Rare Earth Element Biomining with Systems Biology Guided Engineering of *Gluconobacter oxydans*. *bioRxiv*, 2023.2002.2009.527855 (2023). <https://doi.org/10.1101/2023.02.09.527855>
- 16 Schmitz, A. M. *et al.* Generation of a *Gluconobacter oxydans* knockout collection for improved extraction of rare earth elements. *Nature Communications* **12**, 6693 (2021). <https://doi.org/10.1038/s41467-021-27047-4>
- 17 Bonificio, W. D. & Clarke, D. R. Rare-Earth Separation Using Bacteria. *Environmental Science & Technology Letters* **3**, 180-184 (2016). <https://doi.org/10.1021/acs.estlett.6b00064>
- 18 Medin, S. *et al.* Genomic characterization of rare earth binding by *Shewanella oneidensis*. *Scientific Reports* **13**, 15975 (2023). <https://doi.org/10.1038/s41598-023-42742-6>
- 19 Mattocks, J. A. & Cotruvo, J. A. Biological, biomolecular, and bio-inspired strategies for detection, extraction, and separations of lanthanides and actinides. *Chemical Society Reviews* **49**, 8315-8334 (2020). <https://doi.org/10.1039/D0CS00653J>
- 20 Skovran, E. & Martinez-Gomez, N. C. Just add lanthanides. *Science* **348**, 862-863 (2015). <https://doi.org/doi:10.1126/science.aaa9091>

21 Reed, D. W., Fujita, Y., Daubaras, D. L., Jiao, Y. & Thompson, V. S. Bioleaching of rare earth elements from waste phosphors and cracking catalysts. *Hydrometallurgy* **166**, 34-40 (2016). <https://doi.org/https://doi.org/10.1016/j.hydromet.2016.08.006>

22 Gumulya, Y. et al. In a quest for engineering acidophiles for biomining applications: challenges and opportunities. *Genes* **9**, 116 (2018).

23 Vera, M., Schippers, A., Hedrich, S. & Sand, W. Progress in bioleaching: fundamentals and mechanisms of microbial metal sulfide oxidation – part A. *Applied Microbiology and Biotechnology* **106**, 6933-6952 (2022). <https://doi.org/10.1007/s00253-022-12168-7>

24 Naseri, T., Beiki, V., Mousavi, S. M. & Farnaud, S. A comprehensive review of bioleaching optimization by statistical approaches: recycling mechanisms, factors affecting, challenges, and sustainability. *RSC Advances* **13**, 23570-23589 (2023). <https://doi.org/10.1039/D3RA03498D>

25 Shen, L., Zhou, H., Qiu, G. & Zhao, H. A review of bioleaching combined with low-cost fermentation and metabolic engineering technology to leach rare earth elements. *Journal of Environmental Chemical Engineering* **12**, 112117 (2024). <https://doi.org/https://doi.org/10.1016/j.jece.2024.112117>

26 De Muynck, C. et al. The Genus *Gluconobacter Oxydans*: Comprehensive Overview of Biochemistry and Biotechnological Applications. *Critical Reviews in Biotechnology* **27**, 147-171 (2007). <https://doi.org/10.1080/07388550701503584>

27 Kazunobu Matsushita, H. T., Naoto Tonouchi, Akiko Okamoto-Kainuma. *Acetic Acid Bacteria*. 350 (Springer Tokyo, 2016).

28 Macauley, S., McNeil, B. & Harvey, L. M. The Genus *Gluconobacter* and Its Applications in Biotechnology. *Critical Reviews in Biotechnology* **21**, 1-25 (2001). <https://doi.org/10.1080/20013891081665>

29 Ripoll, M., Lerma-Escalera, J. A., Morones-Ramírez, J. R., Rios-Solis, L. & Betancor, L. New perspectives into *Gluconobacter*-catalysed biotransformations. *Biotechnology Advances* **65**, 108127 (2023). <https://doi.org/https://doi.org/10.1016/j.biotechadv.2023.108127>

30 Antonick, P. J. et al. Bio- and mineral acid leaching of rare earth elements from synthetic phosphogypsum. *The Journal of Chemical Thermodynamics* **132**, 491-496 (2019). <https://doi.org/https://doi.org/10.1016/j.jct.2018.12.034>

31 Jin, H. et al. Sustainable Bioleaching of Rare Earth Elements from Industrial Waste Materials Using Agricultural Wastes. *ACS Sustainable Chemistry & Engineering* **7**, 15311-15319 (2019). <https://doi.org/10.1021/acssuschemeng.9b02584>

32 Owusu-Fordjour, E. Y. & Yang, X. Bioleaching of rare earth elements challenges and opportunities: A critical review. *Journal of Environmental Chemical Engineering* **11**, 110413 (2023). <https://doi.org/https://doi.org/10.1016/j.jece.2023.110413>

33 Vo, P. H. N. et al. Biomining for sustainable recovery of rare earth elements from mining waste: A comprehensive review. *Science of The Total Environment* **908**, 168210 (2024). <https://doi.org/https://doi.org/10.1016/j.scitotenv.2023.168210>

34 Bösecker, K. Bioleaching: metal solubilization by microorganisms. *FEMS Microbiology Reviews* **20**, 591-604 (1997). <https://doi.org/10.1111/j.1574-6976.1997.tb00340.x>

35 Abhilash, Hedrich, S. & Schippers, A. Distribution of scandium in red mud and extraction using *Gluconobacter oxydans*. *Hydrometallurgy* **202**, 105621 (2021). <https://doi.org/https://doi.org/10.1016/j.hydromet.2021.105621>

36 Abhilash et al. Extraction of REEs from Blast Furnace Slag by *Gluconobacter oxydans*. *Minerals* **12**, 701 (2022).

37 Rasoulnia, P., Barthen, R., Puhakka, J. A. & Lakaniemi, A.-M. Leaching of rare earth elements and base metals from spent NiMH batteries using gluconate and its potential bio-oxidation products. *Journal of Hazardous Materials* **414**, 125564 (2021). <https://doi.org/https://doi.org/10.1016/j.jhazmat.2021.125564>

38 Banerjee, I., Burrell, B., Reed, C., West, A. C. & Banta, S. Metals and minerals as a biotechnology feedstock: engineering biomining microbiology for bioenergy applications. *Current Opinion in Biotechnology* **45**, 144-155 (2017). <https://doi.org/https://doi.org/10.1016/j.copbio.2017.03.009>

39 Corbett, M. K., Eksteen, J. J., Niu, X.-Z., Croue, J.-P. & Watkin, E. L. J. Interactions of phosphate solubilising microorganisms with natural rare-earth phosphate minerals: a study utilizing Western Australian monazite. *Bioprocess and Biosystems Engineering* **40**, 929-942 (2017). <https://doi.org/10.1007/s00449-017-1757-3>

40 Hogendoorn, C. et al. Facile Arsenazo III-Based Assay for Monitoring Rare Earth Element Depletion from Cultivation Media for Methanotrophic and Methylotrophic Bacteria. *Applied and Environmental Microbiology* **84**, e02887-02817 (2018). <https://doi.org/doi:10.1128/AEM.02887-17>

41 Rohwer, H., Collier, N. & Hosten, E. Spectrophotometric study of arsenazo III and its interactions with lanthanides. *Analytica Chimica Acta* **314**, 219-223 (1995). [https://doi.org/https://doi.org/10.1016/0003-2670\(95\)00279-9](https://doi.org/https://doi.org/10.1016/0003-2670(95)00279-9)

42 Balta, J. B., Holycross, M.E., Barstow, B., Gazel, E. Co-generation of NaREE(MoO₄)₂ and REPO₄ in multiple habits by solid-flux crystal growth. *Minerals Engineering* (2024).

43 Brierley, C. L. Microbiological Mining. *Scientific American* **247**, 44-53 (1982).

44 Charkin, D. O. et al. Structural, thermal, and IR studies of β -[Nd₂O₂](CrO₄), an unexpected analog of a slag phase [Ba₂F₂](S₆+O₃S₂-). *Zeitschrift für Kristallographie - Crystalline Materials* **234**, 1-8 (2019). <https://doi.org/doi:10.1515/zkri-2018-2065>

45 Alexa, A. & Rahnenfuhrer, J. Gene set enrichment analysis with topGO. (2007).

46 Capdevila, Daiana A., Edmonds, Katherine A. & Giedroc, David P. Metallochaperones and metalloregulation in bacteria. *Essays in Biochemistry* **61**, 177-200 (2017). <https://doi.org/10.1042/ebc20160076>

47 Chunqiang Zhang, F. Z., Ping Zhou, Caiguo Zhang. Functional role of metalloproteins in genome stability. *Front. Biol.* **11**, 119-131 (2016). <https://doi.org/10.1007/s11515-016-1392-4>

48 Nakayama, H. in *Handbook of Metal Biotechnology* (ed Mitsuo Yamashita Michihiko Ike, Satoshi Soda) Ch. 17, 32 (Jenny Stanford Publishing, 2012).

49 Urbina, J. et al. A new approach to biomining: bioengineering surfaces for metal recovery from aqueous solutions. *Scientific Reports* **9**, 16422 (2019).

50 Carvajal-Garcia, J., Samadpour, A. N., Hernandez Viera, A. J. & Merrikh, H. Oxidative stress drives mutagenesis through transcription-coupled repair in bacteria. *Proceedings of the National Academy of Sciences* **120**, e2300761120 (2023). <https://doi.org/doi:10.1073/pnas.2300761120>

51 Fan, Y. et al. Protein mistranslation protects bacteria against oxidative stress. *Nucleic Acids Research* **43**, 1740-1748 (2015). <https://doi.org/10.1093/nar/gku1404>

52 Baym, M., Shaket, L., Anzai, I. A., Adesina, O. & Barstow, B. Rapid construction of a whole-genome transposon insertion collection for *Shewanella oneidensis* by Knockout Sudoku. *Nat Commun* **7**, 13270 (2016). <https://doi.org/10.1038/ncomms13270>

53 Giaever, G. & Nislow, C. The Yeast Deletion Collection: A Decade of Functional Genomics. *Genetics* **197**, 451-465 (2014). <https://doi.org/10.1534/genetics.114.161620>

54 Gallagher, L. A. et al. Sequence-defined transposon mutant library of *Burkholderia thailandensis*. *mBio* **4**, e00604-00613 (2013). <https://doi.org/10.1128/mBio.00604-13>

55 Sousa, M. et al. Genome-wide identification of genes involved in the positive and negative regulation of acetic acid-induced programmed cell death in *Saccharomyces cerevisiae*. *BMC Genomics* **14**, 838 (2013). <https://doi.org/10.1186/1471-2164-14-838>

56 Landová, B. & Šilhán, J. Conformational changes of DNA repair glycosylase MutM triggered by DNA binding. *FEBS Letters* **594**, 3032-3044 (2020). <https://doi.org/https://doi.org/10.1002/1873-3468.13876>

57 Pittman, M. S., Robinson, H. C. & Poole, R. K. A Bacterial Glutathione Transporter (*Escherichia coli* CydDC) Exports Reductant to the Periplasm*. *Journal of Biological Chemistry* **280**, 32254-32261 (2005). <https://doi.org/https://doi.org/10.1074/jbc.M503075200>

58 Kumar, V. et al. Structure of BipA in GTP form bound to the ratcheted ribosome. *Proceedings of the National Academy of Sciences* **112**, 10944-10949 (2015). <https://doi.org/10.1073/pnas.1513216112>

59 Jha, V., Dafale, N. A. & Purohit, H. J. Regulatory rewiring through global gene regulations by PhoB and alarmone (p)ppGpp under various stress conditions. *Microbiological Research* **227**, 126309 (2019). <https://doi.org/https://doi.org/10.1016/j.micres.2019.126309>

60 Yamada, M., Makino, K., Shinagawa, H. & Nakata, A. Regulation of the phosphate regulon of Escherichia coli Properties of phoR deletion mutants and subcellular localization of PhoR protein. *Molecular and General Genetics MGG* **220**, 366-372 (1990). <https://doi.org/10.1007/BF00391740>

61 Matilla, M. A., Velando, F., Martín-Mora, D., Monteagudo-Cascales, E. & Krell, T. A catalogue of signal molecules that interact with sensor kinases, chemoreceptors and transcriptional regulators. *FEMS Microbiology Reviews* **46** (2021). <https://doi.org/10.1093/femsre/fuab043>

62 Keller, L. M. L., Flattich, K. & Weber-Ban, E. Novel WYL domain-containing transcriptional activator acts in response to genotoxic stress in rapidly growing mycobacteria. *Communications Biology* **6**, 1222 (2023). <https://doi.org/10.1038/s42003-023-05592-6>

63 Camacho, C. et al. BLAST+: architecture and applications. *BMC Bioinformatics* **10**, 421 (2009). <https://doi.org/10.1186/1471-2105-10-421>

64 Iida, A., Ohnishi, Y. & Horinouchi, S. Identification and characterization of target genes of the GinL/GinR quorum-sensing system in *Gluconacetobacter intermedius*. *Microbiology* **155**, 3021-3032 (2009). <https://doi.org/https://doi.org/10.1099/mic.0.028613-0>

65 Delepelaire, P. Bacterial ABC transporters of iron containing compounds. *Research in Microbiology* **170**, 345-357 (2019). <https://doi.org/https://doi.org/10.1016/j.resmic.2019.10.008>

66 Folsom, J. P., Parker, A. E. & Carlson, R. P. Physiological and Proteomic Analysis of *Escherichia coli* Iron-Limited Chemostat Growth. *Journal of Bacteriology* **196**, 2748-2761 (2014). <https://doi.org/doi:10.1128/jb.01606-14>

67 Sasnow, S. S., Wei, H. & Aristilde, L. Bypasses in intracellular glucose metabolism in iron-limited *Pseudomonas putida*. *MicrobiologyOpen* **5**, 3-20 (2016). <https://doi.org/https://doi.org/10.1002/mbo3.287>

68 Silva, R., Aguiar, T. Q. & Domingues, L. Orotic acid production from crude glycerol by engineered *Ashbya gossypii*. *Bioresource Technology Reports* **17**, 100992 (2022). <https://doi.org/https://doi.org/10.1016/j.biteb.2022.100992>

69 Kostova, I., Mojžiš, J. & Chiş, V. Theoretical and Experimental Vibrational Characterization of Biologically Active Nd(III) Complex. *Molecules* **26**, 2726 (2021).

70 Neumann, M., Schulte, M., Jünemann, N., Stöcklein, W. & Leimkühler, S. Rhodobacter capsulatus XdhC Is Involved in Molybdenum Cofactor Binding and Insertion into Xanthine Dehydrogenase*. *Journal of Biological Chemistry* **281**, 15701-15708 (2006). <https://doi.org/https://doi.org/10.1074/jbc.M601617200>

71 Choi, K.-Y. Bioprocess of Microbial Melanin Production and Isolation. *Frontiers in Bioengineering and Biotechnology* **9** (2021). <https://doi.org/10.3389/fbioe.2021.765110>

72 El-Naggar, N. E.-A. & Saber, W. I. A. Natural Melanin: Current Trends, and Future Approaches, with Especial Reference to Microbial Source. *Polymers* **14**, 1339 (2022).

73 Solano, F. On the Metal Cofactor in the Tyrosinase Family. *International Journal of Molecular Sciences* **19** (2018).

74 Anzai, I. A., Shaket, L., Adesina, O., Baym, M. & Barstow, B. Rapid curation of gene disruption collections using Knockout Sudoku. *Nature Protocols* **12**, 2110-2137 (2017). <https://doi.org/10.1038/nprot.2017.073>

75 Makarenkov, V. et al. HTS-Corrector: software for the statistical analysis and correction of experimental high-throughput screening data. *Bioinformatics* **22**, 1408-1409 (2006). <https://doi.org/10.1093/bioinformatics/btl126>

76 Kevorkov, D. & Makarenkov, V. Statistical Analysis of Systematic Errors in High-Throughput Screening. *Journal of Biomolecular Screening* **10**, 557-567 (2005). <https://doi.org/10.1177/1087057105276989>

77 Benjamini, Y. & Hochberg, Y. Controlling the false discovery rate: a practical and powerful approach to multiple testing. *Journal of the Royal statistical society: series B (Methodological)* **57**, 289-300 (1995).

78 topGO: Enrichment Analysis for Gene Ontology. R package version 2.52.0. (2023).

79 Silberbach, M., Maier, B., Zimmermann, M. & Büchs, J. Glucose oxidation by *Gluconobacter oxydans*: characterization in shaking-flasks, scale-up and optimization of the pH profile. *Applied Microbiology and Biotechnology* **62**, 92-98 (2003). <https://doi.org/10.1007/s00253-003-1222-x>

80 Al-Hakkani, M. F. Guideline of inductively coupled plasma mass spectrometry “ICP-MS”: fundamentals, practices, determination of the limits, quality control, and method validation parameters. *SN Applied Sciences* **1**, 791 (2019). <https://doi.org/10.1007/s42452-019-0825-5>