

IP3 receptor depletion in a spontaneous canine model of Charcot-Marie-Tooth disease 1J with amelogenesis imperfecta

Marjo K. Hytönen^{1,2,3,#}, Julius Rönkkö^{4,#}, Sruthi Hundi^{1,2,3}, Tarja S. Jokinen⁵, Emilia Suonto^{1,2,3}, Eeva Teräväinen⁶, Jonas Donner⁷, Rita La Rovere⁸, Geert Bultynck⁸, Emil Ylikallio^{4,9}, Henna Tyynismaa^{2,4} and Hannes Lohi^{1,2,3*}

¹Department of Veterinary Biosciences, University of Helsinki, Helsinki, Finland

²Department of Medical and Clinical Genetics, University of Helsinki, Helsinki, Finland

³Folkhälsan Research Center, Helsinki, Finland

⁴Stem Cells and Metabolism Research Program, Faculty of Medicine, University of Helsinki, Helsinki, Finland

⁵Department of Equine and Small Animal Medicine, University of Helsinki, Helsinki, Finland

⁶Animal Clinic Tassuasema, Kauniainen, Finland

⁷Wisdom Panel, Mars Petcare Science and Diagnostics, Helsinki, Finland

⁸Laboratory of Molecular and Cellular Signaling, Department of Cellular and Molecular Medicine & Leuven Kanker Instituut, Campus Gasthuisberg O/N-I bus 802, KU Leuven, Leuven, Belgium.

⁹Clinical Neurosciences, Neurology, University of Helsinki and Helsinki University Hospital,

[#]M.K.H. and J.R. contributed equally to this paper

*Corresponding author: Hannes Lohi, PhD, Professor, Department of Veterinary Biosciences, Department of Medical and Clinical Genetics, Folkhälsan Research Center, P.O. Box 63, 00014 University of Helsinki, Finland. email: hannes.lohi@helsinki.fi, tel. +358 2941 25085

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Abstract

Inositol 1,4,5-trisphosphate receptors (IP₃R) mediate Ca²⁺ release from intracellular stores, contributing to complex regulation of numerous physiological responses. The involvement of the three IP₃R genes (*ITPR1*, *ITPR2* and *ITPR3*) in inherited human diseases has started to shed light on the essential roles of each receptor in different human tissues and cell types. Variants in the *ITPR3* gene, which encodes IP₃R3, have recently been found to cause demyelinating sensorimotor Charcot-Marie-Tooth neuropathy type 1J (CMT1J). In addition to peripheral neuropathy, immunodeficiency and tooth abnormalities are occasionally present. Here, we report the identification of a homozygous nonsense variant in the *ITPR3* gene in Lancashire Heeler dogs, presenting with a severe developmental enamel defect and reduced nerve conduction velocity. We studied the primary skin fibroblasts of the affected dogs and observed that the nonsense variant in *ITPR3* led to a complete absence of full-length IP₃R3 protein. Unexpectedly, the protein levels of IP₃R1 and IP₃R2 were also markedly decreased, suggesting co-regulation. Functional Ca²⁺ measurements revealed reduced IP₃R-mediated Ca²⁺ flux upon stimulation of G-protein-coupled-receptors in the affected dog fibroblasts. We were able to rescue the IP₃R1 and IP₃R2 depletion by proteasome inhibition but not the IP₃R3 loss, which was facilitated by nonsense-mediated mRNA decay. These findings highlight the first spontaneous mammalian phenotype caused by a nonsense variant in *ITPR3*, leading to the loss of IP₃R3. The human and canine IP₃R3 proteins are highly similar, and our study suggests that the tissue involvement resulting from the receptor's dysfunction is also conserved. In summary, IP₃R3 is critical for enamel formation and peripheral nerve maintenance.

Author summary

We investigated pet dogs, Lancashire Heelers, with impairments in tooth development and in the nerves that regulate limb muscles. Through genetic studies of the dog pedigree, we found that the phenotypes were caused by a recessively inherited mutation in the *ITPR3* gene, which encodes one of three IP₃ receptors (IP₃R) isoforms (IP₃R3 isoform) that are needed for intracellular Ca²⁺ signaling. Mutated IP₃R3 has been recently linked to a human inherited neuropathy called Charcot-Marie-Tooth disease type 1J, which impairs peripheral nerve function and is accompanied by immunodeficiency and abnormal teeth in some individuals. We showed that in the skin cells of the affected dogs, the full-length IP₃R3 protein was completely absent, and also the protein levels of the other two IP₃R isoforms (IP₃R1 and IP₃R2) were severely lowered. This led to impaired agonist-induced Ca²⁺ release and signaling. Our results demonstrate the high conservation between human and canine IP₃ receptors and their significance for different tissue systems. The genetic studies now highlight that IP₃R3 is vital for peripheral nerve function and enamel development.

Keywords

enamel, amelogenesis imperfecta, hypomineralization, canine, ameloblast, Ca²⁺ signaling, inositol 1,4,5-trisphosphate receptor, IP₃R1, IP₃R2, IP₃R3, ITPR3, neuropathy, CMT, Charcot-Marie-Tooth, CMT1J

Introduction

Inositol 1,4,5-trisphosphate receptors (IP₃Rs) are intracellular Ca²⁺ channels located in the endoplasmic reticulum (ER). Through Ca²⁺ signals induced by IP₃, they are involved in a wide range of physiological processes, including differentiation, metabolism, and apoptosis (1, 2). The IP₃Rs are homo- or heterotetramers composed of three homologs IP₃R1, IP₃R2, and IP₃R3 (encoded by *ITPR1*, *ITPR2* and *ITPR3* respectively), each with different tissue distribution and properties. Studies on IP₃R knockout animal models and patients with pathogenic variants in IP₃Rs have elucidated the

importance of distinct isoforms in various tissues (3). Despite their importance, the molecular mechanisms by which IP₃Rs regulate Ca²⁺ signaling and contribute to various cellular processes are still not fully understood. Nevertheless, the discoveries of disease-associated genetic variants, in addition to genetically modified animal and cell models, support the crucial role of IP₃Rs (3-9).

The genes encoding IP₃Rs have been linked to various inherited human diseases. Variants in *ITPR1* cause dominantly inherited spinocerebellar ataxia types 15 and 29, as well as Gillespie syndrome, which exhibits both dominant and recessive inheritance patterns (10-13). A homozygous missense variant in *ITPR2* has been reported to cause isolated anhidrosis (14). *Itpr1* knockout mice either died *in utero* or had severe ataxia and epilepsy after birth (15), whereas *Itpr2* knockouts had reduced sweat secretion (14) and *Itpr3* knockouts had abnormal taste perception (16).

We described dominantly inherited or *de novo* *ITPR3* variants in individuals with progressive distal muscle weakness, foot deformities and reduced nerve conduction velocity (NCV), a sign of peripheral nerve demyelination (17). Previously, a few other cases of neuropathy associated with *ITPR3* variants of unknown significance had been reported (18, 19). These findings led to a classification of Charcot-Marie-Tooth disease 1J (CMT1J), a new subtype of hereditary neuropathy caused by *ITPR3* variants (Table 1). CMT is a heterogeneous group of inherited peripheral neuropathies affecting the peripheral sensory and motor nerves. The two most common types of CMT are demyelinating CMT1, characterized by reduced NCV (< 38 m/s), and axonal CMT2 (NCV >38 m/s). Recently, two individuals with immunodeficiency were found to carry compound heterozygous variants in *ITPR3* (20). One of those patients had peripheral neuropathy, abnormal tooth eruption and mineralization and thin hair since birth. Furthermore, two unrelated individuals with immunodeficiency from low B and T cells, deciduous teeth, anhidrosis, abnormal gait, bilateral vocal fold paralysis and sensorimotor neuropathy of the lower extremities have been reported (21). Thus, the recent findings suggest that *ITPR3*-related phenotypes extend beyond peripheral nerves.

Table 1. Identified human disease-causing *ITPR3* variants and patient phenotypes

#	Variant	Genotype	Allele frequency	NCV reduction	Neuropathy	Immunological symptoms	Tooth abnormalities	Reference
1	V615M	HET	Not found	+	+	-	-	(17)
2	V615M	HET	Not found	+	+	-	-	(17)
3	V615M	HET	Not found	NA	+	-	-	(17)
4	V615M	HET	Not found	+	+	-	-	(17)
5	R2524C	HET	Not found	+	+	-	-	(17)
6	R2524C	HET	Not found	+	+	+	+	(21)
7	R2524C	HET	Not found	+	+	+	+	(21)
8	R1850Q/ R2524C	Comp. HET	8.178E-02/ Not found	NA	+	+	+	(20)
9	F1628L/ R1850Q	Comp. HET	2.05E-06/ 8.178E-02	NA	-	+	-	(20)
10	M1064V	HET	2.06E-06	NA	+	NA	NA	(19)
11	T1424M	HET	7.14E-07	+	+	NA	NA	(18)
12	T1424M	HET	7.14E-07	+	+	NA	NA	(18)
13	T1424M	HET	7.14E-07	+	+	NA	NA	(18)
14	T1424M	HET	7.14E-07	+	+	-	-	(22)
15	T1424M	HET	7.14E-07	+	-	-	-	(22)
16	P187S	HET	4.96E-05	+	+	-	-	(23)
17	P187S	HET	4.96E-05	+	+	-	-	(23)
18	P187S	HET	4.96E-05	NA	+	-	-	(23)

The allele frequencies are from gnomAD v4.1.0 (24).

In this study, we discovered a homozygous canine *ITPR3* nonsense variant leading to amelogenesis imperfecta and peripheral neuropathy in Lancashire Heeler dogs. This is the first known naturally

occurring nonsense variant of *ITPR3* linked to disease, which led to a complete loss of full-length IP₃R3 protein. Unexpectedly, we found concurrent depletion of IP₃R1 and IP₃R2 proteins, which could be rescued by proteasome inhibition. These data point to interdependency in the turnover of the three IP₃R subunits. Our results suggest a critical role for adequate IP₃R-dependent Ca²⁺ signaling in enamel development and peripheral nerve function.

Results

Enamel defect and peripheral nerve dysfunction in Lancashire Heeler dogs

The affected Lancashire Heeler dogs (Fig 1A) were originally identified by abnormal enamel in the permanent teeth, including severe yellow to brown discoloration, enamel hypoplasia and abrasion leading to exposure of dentin (Fig 1B, C). The enamel defects were already apparent upon tooth eruption. The affected dogs displayed normal breed-typical moving patterns and activity levels. The owners of the affected dogs did not report untypical clumsiness or leg movement in their respective dogs, except for one case at nine years of age. In the general clinical examination, two out of three dogs had deformity detected in their front limbs, i.e. outward rotation of front limbs from the elbow joint. Neurological examination was normal in all affected dogs. No remarkable abnormalities were detected in the complete blood count or serum biochemistry of the examined dogs. Abnormal electromyographic findings consistent with demyelinating neuropathy were detected in all three examined dogs and consisted of fibrillation potentials and positive sharp waves occurring predominantly in distal appendicular limb muscles (i.e. in muscles distal to elbow and knee joints) (Supplementary table 1). Fibrillation potentials were also detected in paraspinal musculature. Motor nerve conduction velocities (MNCVs) were below published canine reference values in both the peroneal and ulnar nerves of the two examined dogs (25). Decreased compound muscle action potential (CMAP) amplitudes and increased CMAP duration (i.e., temporal dispersion) at all stimulation points were observed in these two dogs. One of the examined dogs had normal MNCVs.

Next generation sequencing revealed a stop-gain variant in *ITPR3*

The pedigree constructed around the affected dogs suggested an autosomal recessive disease (Fig 1E). We carried out a genome-wide genotyping on a cohort of four cases and six controls to identify the runs of homozygosity shared by cases. Homozygosity mapping revealed only one case-specific region of allelic homozygosity (ROH) on chromosome 12 (g.156,577-5,127,877) of size 4,97 Mb (Fig 1D). We performed whole-exome sequencing (WES) on two affected dogs and whole-genome sequencing (WGS) on one affected dog to identify the disease-causing variant. Filtering the SNVs and indels according to a recessive model against variant data of 689 control dogs from other breeds resulted in 4 case-specific homozygous variants (Supplementary table 2). Three of the variants located within the ROH on chromosome 12, however, only one had a predicted effect on the protein: one nucleotide substitution in exon 37 of *ITPR3* (g.12:3214076C>T; XM_038553756.1:5002C>T). The variant substitutes glutamine to a translation termination codon (p.Q1668X) (XP_038409684), which is predicted to result in a significantly truncated protein (amino acids 1668-2690 missing) if translated. The resulting protein would lack part of the coupling region, the transmembrane region and the C-terminal tail, thus lacking the complete C-terminal channel-pore region (26).

We further analyzed the WGS data of the affected dog to exclude any additional non-coding variants, structural variants (SVs), and mobile element insertions (MEIs) using 388 control dogs from other breeds. After filtering the homozygous case-specific variants, 3919 SNVs/indels, 14 MEIs, but no SVs remained (Supplementary Tables 3 and 4). None of the MEIs located within the identified ROH. Of the SNVs and indels, 17 variants were within the mapped ROH, one being the above-mentioned stop-gain variant in *ITPR3*. The remaining variants were non-coding, except for a deletion in ncRNA LOC119874143 annotated by NCBI. However, Ensembl annotated this variant as intronic in *HLA-DRB1*. We investigated the *ITPR3* variant frequency in an additional cohort of 1413 dog genomes sequenced as part of the Dog10K project. All dogs were wild-type, except for one Lancashire Heeler,

which was heterozygous for the allele. Based on gene function, predicted pathogenicity and the breed-specific distribution, the nonsense variant in *ITPR3* was prioritized and selected for further validation.

We genotyped the *ITPR3* variant using standard PCR in a cohort of 266 Lancashire Heeler samples, including 10 affected dogs, which confirmed the variant and the segregation of the allele (Fig 1 E, F). The variant was observed as homozygous in all 10 affected dogs and also in one sample from the biobank. According to our records, the additional homozygous dog is deceased and we were unsuccessful in contacting the owner. Therefore, its health status remains unknown. The remaining samples were either heterozygous (n=67) or wild type (n=188). The allele frequency was 16.6% in the study cohort. An additional cohort of over 2.6 million dogs submitted for commercial genetic testing was screened for the *ITPR3* variant to explore the distribution across different breeds. We found only 16 heterozygous dogs, 13 of which had undergone DNA-based breed ancestry testing. Ten of these heterozygotes were purebred Lancashire Heelers, two were mixed breed dogs with detected genomic ancestry from the Lancashire Heeler (30% and 2% contributions, respectively), and one was a dog of highly mixed ancestry.

The IP₃R1, IP₃R2, and IP₃R3 are highly similar proteins between humans and canine, with 96%-98% of the amino acids being identical between the species (Fig 2A) (Supplementary Fig 1). The premature stop codon caused by the variant resides in a conserved site in the IP₃R3 regulatory/coupling region upstream of the channel region containing the channel pore (Fig 2B). The possible truncated protein would be non-functional without transmembrane alpha helices or the channel pore (Fig 2C).

Complete loss of IP₃R3 and reduced IP₃R1 and IP₃R2 levels in fibroblasts of affected dogs

To investigate if the p.Q1668X variant leads to truncation of IP₃R3, we obtained primary skin fibroblasts from the affected Lancashire Heelers and control dogs and performed a Western blot with

an antibody directed against the N-terminal (22-230 amino acid residue) region of IP₃R3 (Fig 2D, E). The affected dog fibroblasts showed a complete lack of full-length IP₃R3 proteins. In addition, no N-terminal fragment was visible around 170-180 kDa, the theoretical molecular weight of the truncated protein fragment (Supplementary figure 2), suggesting that the nonsense-mutant *ITPR3* mRNA is degraded by nonsense-mediated decay. Indeed, RT-qPCR analysis showed a reduction of *ITPR3* mRNA level (reduced to 35% of control level) in the affected dog fibroblasts (Fig 2F). Surprisingly, we also found decreased protein levels of IP₃R1 and IP₃R2 in the affected dog fibroblasts (79% and 71% reduction, respectively). Also, the mRNA level of *ITPR2* was decreased, whereas the level of *ITPR1* mRNA was increased in the affected dog fibroblasts.

These results were consistent with partial nonsense-mediated mRNA decay caused by the p.Q1668X variant in *ITPR3*, and if any truncated protein was produced, it was not stable. Elevation of *ITPR1* mRNA may represent a compensatory upregulation of the alternative IP₃R, which ultimately appeared futile as the protein levels of both IP₃R1 and IP₃R2 were decreased in the affected Lancashire Heeler dog fibroblasts. The latter result was unexpected and prompted further investigation of IP₃R turnover.

Proteasome inhibitor MG-132 rescues the IP₃R1 and IP₃R2-protein levels

To assess the mechanism of IP₃R depletion, we treated affected Lancashire Heeler fibroblasts with proteasome inhibitor MG-132 for 12 and 24 hours. After the MG-132 treatment, we analyzed the protein levels of IP₃R1-3 using Western blotting with IP₃R-isoform-specific antibodies (Fig 3A). The analysis revealed significant changes in the IP₃R1 and IP₃R2 levels compared to non-treated cells (Fig 3B). The MG-132 treatment rescued the depletion of IP₃R1 and IP₃R2 to levels comparable to those in the control cell lines, while the treatment had no consistent effect on the protein levels in the control cells. As anticipated, the levels of the full-length IP₃R3 protein were not affected by MG-132. Hence, the depletion of IP₃R1 and IP₃R2 proteins appears to be caused by proteasomal degradation, as MG-132 restored their protein levels.

The p.Q1668X variant leads to a reduction in IP₃R-mediated cytosolic Ca²⁺ release

To assess the effect of the *ITPR3* nonsense variant on Ca²⁺ signaling, we utilized functional Ca²⁺ imaging on the affected Lancashire Heeler dog fibroblasts and control cell lines. We analyzed the dog fibroblasts with ratiometric Fura-2 AM Ca²⁺ indicator using a fluorescent microscope. The measurements were performed in the absence of extracellular Ca²⁺ to ensure that Ca²⁺ signals originated from intracellular stores. We measured the cytosolic [Ca²⁺] rises in living cells exposed to the G-protein-coupled-receptor (GPCR) agonist ATP, while the total intracellular releasable Ca²⁺ was estimated using the Ca²⁺ ionophore ionomycin.

First, we elicited the IP₃ production of the cells by 80 μM ATP and measured the cells' capacity to generate cytosolic Ca²⁺ signals upon production of IP₃. The affected Lancashire Heeler fibroblasts had significantly reduced ATP-evoked Ca²⁺ responses compared to the control cells, as both the maximum amplitude and area-under-the-curve analysis displayed similar decreases in the affected fibroblasts (Fig 4A). We then assessed the total intracellular Ca²⁺ storage with 2 μM Ca²⁺ ionophore ionomycin. There were no differences between the affected and the control fibroblasts, indicating comparable intracellular Ca²⁺ loading between the cell lines (Fig 4B).

Next, we analyzed the dog fibroblasts with Ca²⁺ indicator Cal-520 AM using the drug screening system FDSS/μCell. The Lancashire Heeler fibroblasts had a significantly reduced response to 100 μM ATP, similarly as observed with ratio metric measurements (Supplementary figure 3A). We then assessed the total ER Ca²⁺ storages with 2 μM irreversible SERCA inhibitor thapsigargin and the total intracellular Ca²⁺ stores with 2.5 μM ionomycin. These results indicate that the Ca²⁺ loading of intracellular Ca²⁺ stores and ER was significantly different between the affected and control fibroblast cells (Supplementary figure 3B & C).

Altogether, these results indicate that the p.Q1668X variant caused a loss of IP₃R3 and affected the IP₃R1- and IP₃R2-protein levels through proteasomal degradation, resulting in deranged Ca²⁺ signaling.

DISCUSSION

We report the first spontaneous large animal model of human CMT1J with a nonsense mutation in *ITPR3*, leading to a complete loss of full-length IP₃R3 proteins in Lancashire Heeler dogs. The results provide additional evidence for the critical role of IP₃R3 in the development and function of the peripheral nerves and teeth.

IP₃Rs are major regulators of intracellular calcium signaling, however, the functional importance of each of the three receptors in different tissues and cell types is not fully established. Earlier studies of knockout mice have yielded important insights into the role of the different IP₃R isoforms in physiology, but whether such findings translate to humans has not been known. Genetic studies identifying pathogenic variants in the IP₃R genes have now elucidated that each human IP₃R has unique tissue-specific roles that the other two receptors cannot compensate for. IP₃R1 has a significant role in the central nervous system, leading to ataxia phenotypes when mutated (3), whereas IP₃R2 is critical in sweat glands, causative of anhidrosis and heat intolerance (14). IP₃R3, on the other hand, has recently been linked to peripheral neuropathy, with some patients having additional immunodeficiency and tooth abnormalities (17, 20, 21).

Here, we describe the first spontaneous model of human CMT1J with significant clinical similarities. The affected dogs showed no systematic clinical evidence of motor or sensory impairment, but the electrodiagnostic examinations revealed neurogenic changes in muscle and decreased NCV. These findings were consistent with subclinical peripheral neuropathy. The human disease CMT1J, caused by pathogenic variants in *ITPR3*, is typically a slowly progressive sensorimotor neuropathy (17-19). Electrodiagnostic studies in the human patients have shown decreased NCV, consistent with demyelination, which has also been confirmed in sural nerve biopsy (17). The demyelinating CMT type suggests the involvement of Schwann cells, which could indicate that the role of IP₃R3 in peripheral nerve maintenance is through a defect in the myelinating cells. It is noteworthy that the severity of CMT1J varies between patients. Analogously to the dogs described here, one individual

249 in her 50s who carried the *ITPR3* p.T1424M variant had no signs of neuropathy but widespread
250 slowing of NCV, confirming the presence of subclinical neuropathy (22).

251 The affected Lancashire Heelers had features of amelogenesis imperfecta, including significant
252 discoloration of teeth, enamel hypoplasia and abrasion leading to dentin exposure. Similarly, signs of
253 tooth abnormalities have been reported in some human patients with the *ITPR3* p.R2524C variant
254 (20, 21). Ca^{2+} is a critical component in enamel mineralization, as it is essential for the formation and
255 growth of hydroxyapatite crystals, which provide the structural integrity and hardness of enamel in
256 teeth (27, 28). Ca^{2+} signaling pathways are involved in the differentiation and maturation of
257 ameloblasts, the cells responsible for enamel formation (29, 30). IP_3Rs are key Ca^{2+} regulators in
258 ameloblasts, functioning as ER's main Ca^{2+} release channel (31).

259 Other important Ca^{2+} handling proteins involved in ameloblast Ca^{2+} signaling are ORAI1 and STIM1,
260 responsible for Ca^{2+} uptake and refilling ER Ca^{2+} storages via store operated calcium entry (SOCE)
261 (32). Loss-of-function variants in *STIM1* and *ORAI1* cause calcium release-activated channelopathies
262 (CRAC) with ectodermal dysplasia, leading to enamel defects and anhidrosis, immunodeficiency,
263 and muscular hypotonia with muscle weakness (33). The complex interplay between IP_3Rs , ORAI1,
264 and STIM1 in amelogenesis and dental enamel development explains the similarities between their
265 genetic defects. However, our results of the *ITPR3* nonsense mutation in Lancashire Heeler dogs,
266 leading to the absence of $\text{IP}_3\text{R3}$, suggest that it is the most important IP_3R for enamel formation,
267 which is also supported by the lack of tooth phenotypes in human *ITPR1* or *ITPR2* diseases.

268 Unlikely in the studied dogs, CMT1J is predominantly an autosomal dominant disease in humans,
269 but a few patients with compound heterozygous *ITPR3* mutations have been reported (20). The human
270 p.V615M and p.T1424M variants seem to lead to a purely neuropathic phenotype. The differences in
271 patient phenotypes might stem from the severity of each variant in the channel function, as suggested
272 by the results of a study where the mutant proteins were expressed in cells lacking all wild-type IP_3Rs

(34). The above-mentioned neuropathy-causing mutants formed abnormal but partially functional IP₃R channels. In contrast, the p.R2524C variant locating in the channel pore led to a complete absence of channel function upon stimulation (34). We showed that the *ITPR3* nonsense variant caused a loss of IP₃R3 in dog fibroblasts. It may thus be functionally similar to the dead-pore human p.R2524C variant, although the cells harboring p.R2524C can still form IP₃R tetramers and localize into the ER membrane. Tetramers containing the dead-pore variant may have lost Ca²⁺ conduction but could produce additional harmful signaling effects, which are not seen in dogs. The mutation type may also contribute to the lack of immunological phenotype in the affected dogs, which has been reported in a few human patients (20, 21). Together, these data suggest a genotype-phenotype relationship for *ITPR3* variants in humans and dogs, where the mildest dominantly inherited variants may lead to neuropathy while more severe variants produce multisystem involvement.

We unexpectedly observed that besides the loss of IP₃R3 in the affected dogs' fibroblasts, the protein levels of IP₃R1 and IP₃R2 were also severely reduced. The contribution of these additional deficiencies in IP₃Rs to the dog phenotype is not known. Furthermore, we only had access to skin biopsies of the pet dogs and cannot rule out if IP₃R1 and IP₃R2 were less or more affected in other cell types of the affected dogs. The loss of IP₃R3 was most likely mediated at least partially by nonsense-mediated mRNA decay, and if any truncated protein was produced, it could not be detected by Western blotting. What could explain the reduction in IP₃R1 and IP₃R2 levels when the production of IP₃R3 was impaired? IP₃Rs are known to have a short half-life once activated, which is achieved through their rapid degradation by the ER-associated degradation (ERAD) pathway, which is ubiquitin-proteasome dependent (35-37). This mechanism may shield cells from the deleterious effects of overactivation of Ca²⁺ signaling pathways. One possibility is that the truncated IP₃R3 may be a misfolded ER-protein that constitutively activates the ERAD pathway, and the overactivation of the system accelerated the degradation of IP₃R1 and IP₃R2. The observed rescue by the proteasomal inhibitor could support this hypothesis. Finally, the reduction in ATP-evoked Ca²⁺ responses was

moderate in the skin fibroblasts of the affected dogs, suggesting that the residual IP₃Rs were functional on a level that was likely sufficient in most tissues.

In conclusion, we have described the first nonsense mutation in *ITPR3*, causing subclinical neuropathy and abnormal enamel development in Lancashire Heeler dogs. This study extends the phenotypic range of defective IP₃R3 function due to disease-linked variants, bridging findings emerging from recent genetic studies in human patients. Our results point to the role of IP₃R3 in enamel formation and link *ITPR3* variants to CRAC channelopathies.

METHODS

Experimental model and subject details

Cell lines and culture conditions

All dogs in this study were privately owned pet dogs. The dog owners gave written informed consent to perform the experiments. The skin biopsies from two affected and four unaffected dogs were collected for fibroblast culture. Skin biopsies were performed under general anesthesia with a disposable sterilized biopsy dermal anchor puncher. The fibroblasts were cultured in Dulbecco's Modified Eagle Medium (DMEM), supplemented with 10% FBS (Life Technologies), 1% penicillin/streptomycin (Life Technologies), 1% L-glutamine (Life Technologies) and 0.2% uridine (Sigma). Cells were incubated at 37 °C in 5 % CO₂.

Method details

Study cohort

EDTA-blood samples were collected from 266 Lancashire Heeler dogs, including 10 affected dogs, their relatives, and other dogs in the canine biobank at the University of Helsinki. The samples were

stored at -20°C until genomic DNA was extracted using the semi-automated Chemagen extraction robot (PerkinElmer Chemagen Technologie GmbH). DNA concentration was determined by either DeNovix DS-11 Spectrophotometer (DeNovix Inc., Wilmington, Delaware, USA) or Qubit 3.0 Fluorometer (Thermo Fisher Scientific Inc.). All sampled animals were privately owned pet dogs enrolled in the study with the owner's informed consent. Sample collection was ethically approved by the Animal Ethics Committee of State Provincial Office of Southern Finland (ESAVI/343/04.10.07/2016 and ESAVI/25696/2020). The pedigree was compiled using GenoPro genealogy software.

Clinical examination

Three dogs underwent a physical and neurologic examination and a bloodwork assessment, including complete blood count and serum biochemistry. The neurological examination included an evaluation of the dogs' mental status, posture, gait, an assessment of postural reactions, spinal reflexes, panniculus reflex, perineal and bulbourethral reflex, and an evaluation of cranial nerves.

The dogs were anesthetized for electrodiagnostic examination. We performed all electrophysiological examinations with a Cadwell electrodiagnostic machine (Cadwell Industries, Inc., Kennewick, WA). EMG was performed bilaterally in the proximal and distal muscles of the pelvic and thoracic limbs and the paraspinal muscles. A disposable concentric needle electrode was used for EMG analysis and a subdermal needle electrode placed subcutaneously on the animal's flank served as the ground. Abnormalities detected included abnormal insertional and spontaneous activity, such as fibrillation potentials and positive sharp waves.

MNCV of ulnar and peroneal nerves were measured unilaterally performed with two stimulating monopolar needle electrodes. Subdermal needle electrodes served as a recording electrode, a

reference electrode, and a ground electrode. The rectal temperature of the dog was measured during the electrodiagnostic testing.

Homozygosity mapping

We genotyped four affected and six unaffected dogs using Illumina's CanineHD Whole-Genome Genotyping BeadChip containing 173,662 markers (Lincoln, NE, USA). We conducted the pre-analytical QC using PLINK (version 1.96b6.20) (38) and included pruning for a sample call rate of > 95 %, marker call rate of > 95 %, and Hardy-Weinberg equilibrium p-value < 1×10^{-8} . Pruning for minor allele frequency or LD was not performed as recommended by Meyermans et al. (39). After QC, all four cases and six controls, as well as 165,826 markers, remained for analysis.

Detection of runs of homozygosity (ROH) was performed with PLINK 1.9 (38). We optimized the population-dependent parameters using simulated data, and then detected ROH shared by the affected dogs. The minimum marker size for ROH (--homozyg-snp) was calculated based on the formula described by Purfield et al. (40) with $\alpha = 0.05$, $N_s = 149084$, $N_i = 6$ and mean SNP heterozygosity = 0.27. The parameters --homozyg-window-snp, --homozyg-window-missing, --homozyg-window-het, --homozyg-window-threshold and --homozyg-kb were set to a fixed value depending on the analysis (Supplementary table 5).

To establish suitable values for ROH density and maximum gap, genotypes for a fully homozygous individual were simulated based on the population map file and analyzed for maximal genome coverage as described by Meyermans et al. (39). To calculate maximum genome coverage for the simulated genome, --homozyg-gap was set to 2000 kb and --homozyg-density to 200 kb/snp; genome coverage was determined as the total length of the resulting ROH. The simulated genome was then analyzed by varying --homozyg-density from 10 to 125 kb/snp in increments of 5 kb and --homozyg-gap from 20 to 1000 kb in increments of 20 kb (Supplementary table 5).

Based on the simulation, density was set to 25 kb/snp and maximum gap to 200 kb, as genome coverage reached 100 % for --homozyg-density at 30 and increased only negligibly for --homozyg-gap after 200. The detection of ROH was performed with --homozyg-group using these parameters. The ROH included in the further analyses were required to be allelically shared by all four affected dogs and either absent or allelically different in the control dogs.

Whole-genome and exome sequencing

We performed exome sequencing on two samples, where the exome library was prepared with 140702_canFam3_exomeplus_BB_EZ_HX1 kit. The prepared library, with a total capture size of exome of 152 Mb obtained from the Roche NimbleGen SeqCap EZ target enrichment design (41) was sequenced on Illumina NextSeq500 platform at Biomedicum Functional Genomics Unit. The exome sequence fastq, with a read length of 300 bp, was aligned to GSD1.0 (42) with Y chromosome appended from ROS_Cfam_1.0 using bwa-mem and had 49X and 52X coverage. Coverage was calculated by the percentage of mapped reads on the target captured region. Duplicates were marked using Mark duplicates feature in GATK and Indel realignment was performed on the aligned bam files, followed by base quality score recalibration and variant calling in gVCF mode using GATK Haplotype caller (43). Whole-genome sequencing (WGS) was conducted on another affected dog on Illumina HiSeq2000 high-throughput sequencing platform with a read length of 300 bp and an average coverage of 19X. Post quality control using fastqc, the reads from the WGS sample were processed using SpeedSeq open-source software with bwa (v0.7.15) for alignment. Similar to the exome processing, duplicates were marked, followed by indel realignment and variant calling in gVCF mode. The gVCFs were combined using combineGVCFs and jointgenotyped using GenotypeGVCFs in GATK version 4.2. The combined VCFs files were annotated for genomic and functional annotations using annovar (44) with Ensembl release100 and NCBI *Canis lupus familiaris* Annotation Release 106. For the WGS sample, mobile element insertions were detected

using Mobile Element Locator Tool (MELT) (45). Manta 1.6 (46) was used to identify Structural Variants (SVs), which included insertions, deletions, inversions, and duplications. The identified SVs were genotyped using GraphTyper2.

Variant analysis

The detected variants were imported into an in-house server with an adapted version of webGQT for canine variants for inheritance model-based variant filtering to identify the casual variant (47). We performed the analysis of the variant data separately for SNVs and indels and, SVs and MEIs. We filtered SNV and indel variant data of three cases (two dogs with WES and one with WGS data) against 689 control genomes assuming an autosomal recessive inheritance. Specifically, the affected dogs were assumed to share the variants in a homozygous state, while no heterozygous or homozygous calls were allowed per alternative allele in the controls. Whole-exome sequencing and WGS data from 689 dogs from different breeds and 4 wolves with 25X average depth (17-59X) were utilized for filtering the SNVs and indels, while 388 of these samples were available for MEI and SV analysis. Additionally, we used 1413 control genomes that had been sequenced (20X average depth) as a part of the Dog 10K genomes project (48). The variant data of all control genomes were aligned and called as described above.

NCBI transcript XM_038553756.1 and protein XP_038409684.1 were used to count the nucleotide and amino acid positions for *ITPR3*, respectively. The protein alignment was performed with the Clustal Omega algorithm (<https://www.ebi.ac.uk/Tools/msa/clustalo>).

Variant validation

Genotyping of individual dogs was performed with PCR, followed by Sanger sequencing. The DNA template was amplified with Taq polymerase (Biotools B&M Labs, S.A.) The primers used are listed in Supplementary Table 6. The products were treated with exonuclease I (New England Biolabs) and

411 rapid alkaline phosphatase (Roche Diagnostics) and then sequenced using the PCR primers on an
412 ABI 3730 capillary sequencer (Life Technologies). The sequence data were analyzed using Unipro
413 UGENE v1.32.0 (49-51)

414 A sample of 2,651,699 pure-bred and mixed breed dogs, collected via owner submission for
415 commercial testing, was screened using the Wisdom Panel™ (Wisdom Panel, WA, USA) genetic
416 testing platform, including breed detection assessment.

417 **Western blot**

418 Skin fibroblasts were cultured until they reached 70-80% confluency and lysed after collection with
419 RIPA buffer (Cell Signaling #9806). Aliquot containing 20 µg of total protein was boiled at 95 °C,
420 separated in 4-20% Criterion™TGX™ gels (Bio-Rad), and transferred to 0.2 µm nitrocellulose (Bio-
421 Rad) using Trans-Blot Turbo (Bio-Rad). The membranes were blocked in 10% milk in 0.1% TBS-T.
422 Antibodies used were: IP₃R1^(CT-1), IP₃R2^(NT-2) (gift from Dr. David Yule, University of Rochester,
423 NY), IP₃R3 (BD-Transduction #610312) and Vinculin (Sigma #V9264-25UL). Secondary antibodies
424 were: Anti-rabbit and anti-mouse (Jackson ImmunoResearch#111-035-144 and #115-035-146). The
425 blots were imaged with WesternBright™ ECL-spray (Thermo Scientific) and Molecular Imager
426 ChemiDoc XRS+ (Bio-Rad).

427 **Real-time quantitative PCR**

428 The RNA was extracted from fibroblast by NucleoSpin® RNA extraction kit (Macherey-Nagel
429 #740955) and reverse transcribed by Maxima first strand cDNA synthesis kit (Thermo Fischer) using
430 Bio-Rad CFX Maestro 1.1 software (Bio-Rad). The primers used are listed in Supplementary table 6.

431 **Functional Ca²⁺ imaging**

432 We investigated IP₃R-mediated Ca²⁺ signaling using non-ratiometric manual Ca²⁺ assay with Ca²⁺
433 dye Cal-520 AM and ratiometric automated Ca²⁺ assay with Ca²⁺ dye FURA-2 AM. The non-

434 ratiometric Ca^{2+} signaling dynamics of the affected Lancashire Heeler fibroblast cells and the control
 435 cells were analyzed using the FDSS/ μ Cell functional drug screening system (Hamamatsu). The
 436 fibroblasts were seeded in 96-well plates (Greiner Bio-one #655090) and analyzed using Ca^{2+}
 437 indicator dye Cal-520 AM (Aat Bioquest). The cells were loaded with 1 μM Cal-520 and incubated
 438 at 37 °C for 30 min in modified Krebs solution (150 NaCl, 5.9 KCl, 1.2 MgCl_2 , 11.6 HEPES (pH
 439 7.3), 1.5 CaCl_2 , in mM). After incubation, the cells were washed once with the buffer and incubated
 440 for 30 min at 37 °C to allow dye de-esterification. After the second incubation, the cells were washed
 441 once with the buffer and imaged at 37 °C with Krebs solution with 0 mM Ca^{2+} . The 3 mM EGTA
 442 was added to the cells after 30 seconds, and after an additional 60 seconds, the cells' Ca^{2+} response
 443 was induced by either 100 μM ATP, 2 μM thapsigargin or 2.5 μM ionomycin, and imaged for 540
 444 seconds. In experiments with ATP and thapsigargin, the cells were additionally exposed to ionomycin
 445 after 390 seconds. The Ca^{2+} signaling experiments were quantified with GraphPad Prism 9 software.
 446 The fluorescent signal was then normalized to the baseline values, assessed by the fluorescent signal
 447 60 sec prior to the stimuli.

448 For the ratiometric Ca^{2+} assays, the dog fibroblast cells were seeded in MatTek glass bottom dish
 449 (MatTek #P35G-1.5-14-C) and imaged with Zeiss Axio Observer Z1 inverted phase contrast
 450 fluorescence microscope. The cells were loaded with 1 μM Fura-2 AM (ThermoFisher) in a fibroblast
 451 culture medium at 37 °C for 30 min. After loading, the cells were washed twice with Krebs solution
 452 and let de-esterificate for 15 min before imaging at 37 °C. The cells were washed twice before
 453 imaging with Ca^{2+} -free Krebs solution containing 3 mM EGTA and stimulated with either 80 μM
 454 ATP or 2 μM ionomycin after 30 seconds of imaging. The cells were masked, and the mean pixel
 455 intensity was measured for each frame using FIJI ImageJ software. The baseline values were selected
 456 in the first 30 seconds, and the relative intensities were calculated. The results were analyzed using
 457 Clampfit software (Axon instruments). The AUC of the peak and the peak amplitude were measured
 458 and the cells with no response (<10% increase to baseline) were discarded.

Proteasome inhibition with MG-132

The fibroblast cells were cultured as normal with 10 μ M MG-132 proteasome inhibitor added to the cell media. For the Western blot analysis, the cells were incubated with MG-132 for 12 hours or 24 hours and collected.

Quantification and statistical analysis

The statistical analysis was performed with GraphPad Prism 9 (GraphPad Software). The statistical significances between the affected and control cell lines were analyzed with an unpaired two-tailed t-test. The number of replicates is indicated in the figure legends. Significance is defined as * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ and **** $p < 0.0001$ in all graphs.

ACKNOWLEDGEMENTS

Sini Karjalainen, Jana Pennonen and Miia Nissilä (at the University of Helsinki), Anja Florizoone, and Tomas Luyten (at KU Leuven) are acknowledged for technical assistance. We acknowledge the Institute for Molecular Medicine Finland core facility (FIMM) at the University of Helsinki for sequencing services and the IT Center for Science Ltd. (CSC, Finland) for providing a high-performance computing infrastructure.

Figure legends

Figure 1. Identification of *ITPR3* p.Q1668X variant in Lancashire Heeler dogs

(A) Photograph of Lancashire Heeler dogs, the subject of this study. The image displays the distinct appearance and breed-specific traits of Lancashire Heelers.

(B & C) Close-up image of an affected Lancashire Heeler dog aged 2 years and 10 months illustrating the typical tooth abnormalities observed in the affected dogs. The typical enamel defects, such as poor enamel formation, discoloration and abrasion of the enamel, are evident.

(D) Homozygosity mapping of four affected dogs and six controls revealed a case-specific region of allelic homozygosity (ROH) on chromosome 12 (g.156,577-5,127,877) of size 4,97 Mb

(E) The pedigree compiled around the affected Lancashire Heelers indicates the *ITPR3* variant segregating as a recessive disease. The pedigree includes 10 affected dogs with homozygous (shapes filled with black) *ITPR3* variant, 17 unaffected heterozygous dogs (black and white shapes) and 8 unaffected canines with wildtype *ITPR3* (shapes with black outline). The genotype and phenotype of the rest of the dogs have not been determined (shapes with grey outline).

(F) Sanger sequencing results confirmed the segregation of the identified *ITPR3* p.Q1668X variant with the phenotype.

Figure 2. The affected dogs have a complete loss of IP₃R3 and reduced IP₃R1 and IP₃R2 protein levels

(A) Schematic representation of the secondary structure of IP₃R3 protein. The functional domains are annotated, highlighting the key functional domains of the protein. These domains include N-terminal suppressor domain, IP₃-binding ligand binding domain, regulatory domain, and C-terminal domain. Known human CMT1J variants are depicted at their respective locations in IP₃R3 in black, and the dog variant in red. The length of the potential truncated protein is depicted in green and non-translated region in grey.

(B) The position of the p.Q1668X variant in IP₃R3 3D structure. The canine p.Q1668X corresponds to the human p.Q1649. The variant site and five adjacent amino acids are highlighted in red. Two IP₃R3 subunits are illustrated. The potential truncated protein is depicted in green and the non-translated region in grey. IP₃ in the binding site is depicted in blue. Human IP₃R3 3D model: (7T3T) (52).

505 (C) The canine p.Q1668X variant site and amino acid conservation.

506 (D) Representative Western blots showing the reduced IP₃R1 and IP₃R2 protein levels and loss of

507 IP₃R3 in the affected dogs.

508 (E) Quantification of the Western blot analysis of the IP₃R1-3 protein levels. Mean ± SEM, n= 4-6

509 replicates.

510 (F) Quantification of RT-qPCR analysis of the *ITPR1*, *ITPR2* and *ITPR3* mRNA expression levels.

511 Mean ± SEM, n = 3 replicates.

512 Statistics are analyzed by Student's t-test (*p < 0.05, **p < 0.01, ***p < 0.001).

513 **Figure 3. Proteasome inhibitor MG-132 rescues the IP₃R1 and IP₃R2 protein levels**

514 (A) Representative Western blots of the IP₃R1-3 proteins after treating the cells with the proteasome

515 inhibitor MG-132 for 12 and 24 hours.

516 (B) Quantification of the Western blot analysis of the IP₃R1-3 expression levels after MG-132

517 treatments. n = 3-6 replicates.

518 Data are presented as mean ± SEM. Statistics are analyzed by one-way ANOVA with Dunnett's

519 multiple comparison test (*p < 0.05, **p < 0.01, ***p < 0.001).

520 **Figure 4. The Q1668X variant leads to reduction in IP₃R-mediated Ca²⁺ release**

521 The Ca²⁺ flux measurements were performed with ratiometric fluorescent Ca²⁺ indicator FURA-2

522 AM. The Ca²⁺ flux analysis was performed with one homozygous Lancashire Heeler dog skin

523 fibroblast line and one control dog cell line.

524 (A) Fluorescent response curves of the affected and control cell lines after stimulation with GPCR

525 agonist ATP. Average fluorescent response curves of the affected and control cell lines after

526 stimulation with 80 μM ATP. The affected fibroblast cells had significantly reduced peak amplitude

527 and AUC (n = 36 - 47 cells, 3 independent experiments).

(B) Fluorescent response curves of the affected and control cell lines after stimulation 2.5 μ M ionomycin. There were no significant differences in the peak amplitude or AUC between the affected and control cells (n = 73 – 118 cells, 3 independent experiments). Data are presented as mean $\pm$ SEM. Statistics are analyzed by Student's t-test (*p < 0.05, **p < 0.01, ***p < 0.001).

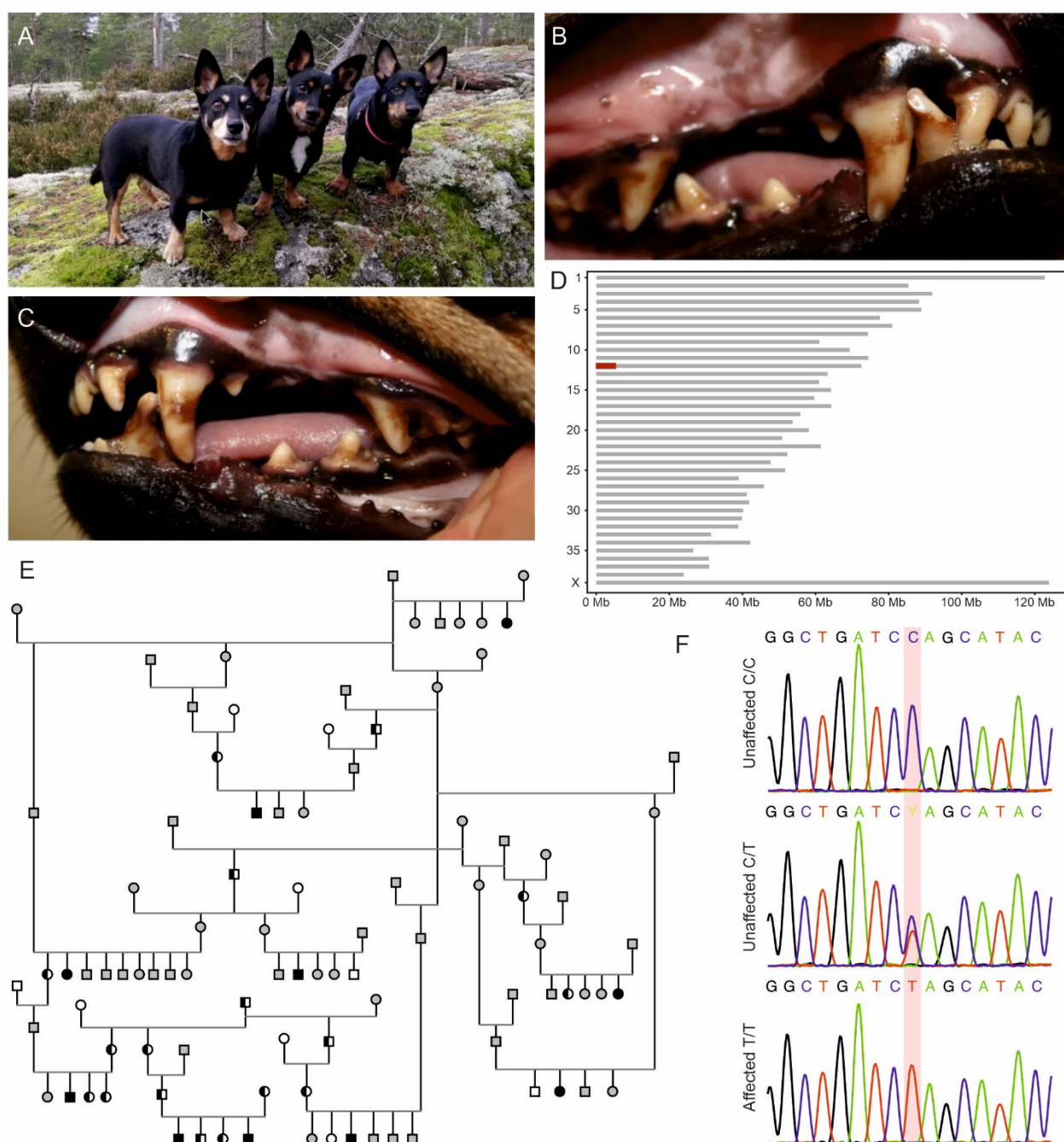


Figure 1.

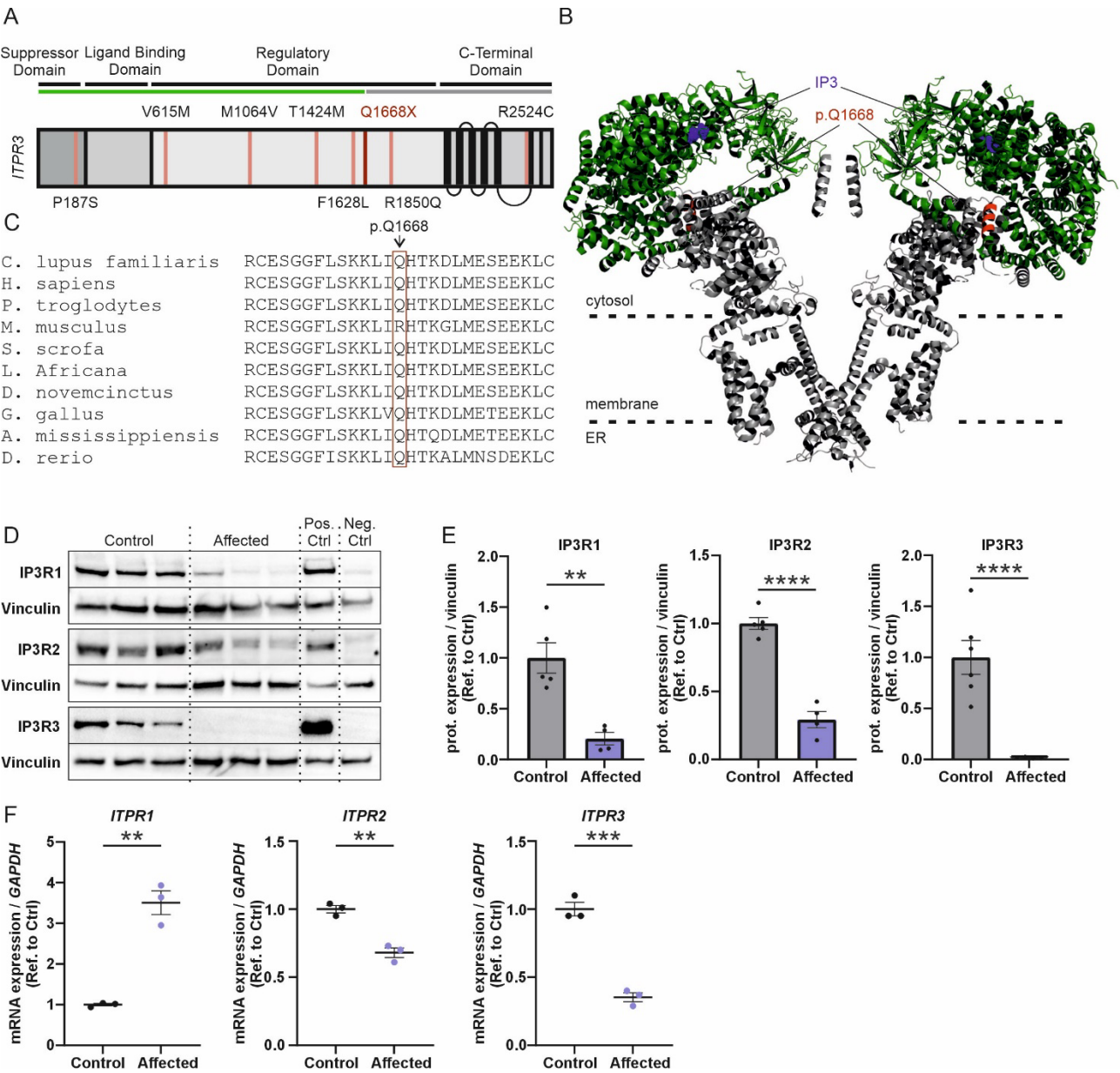


Figure 2.

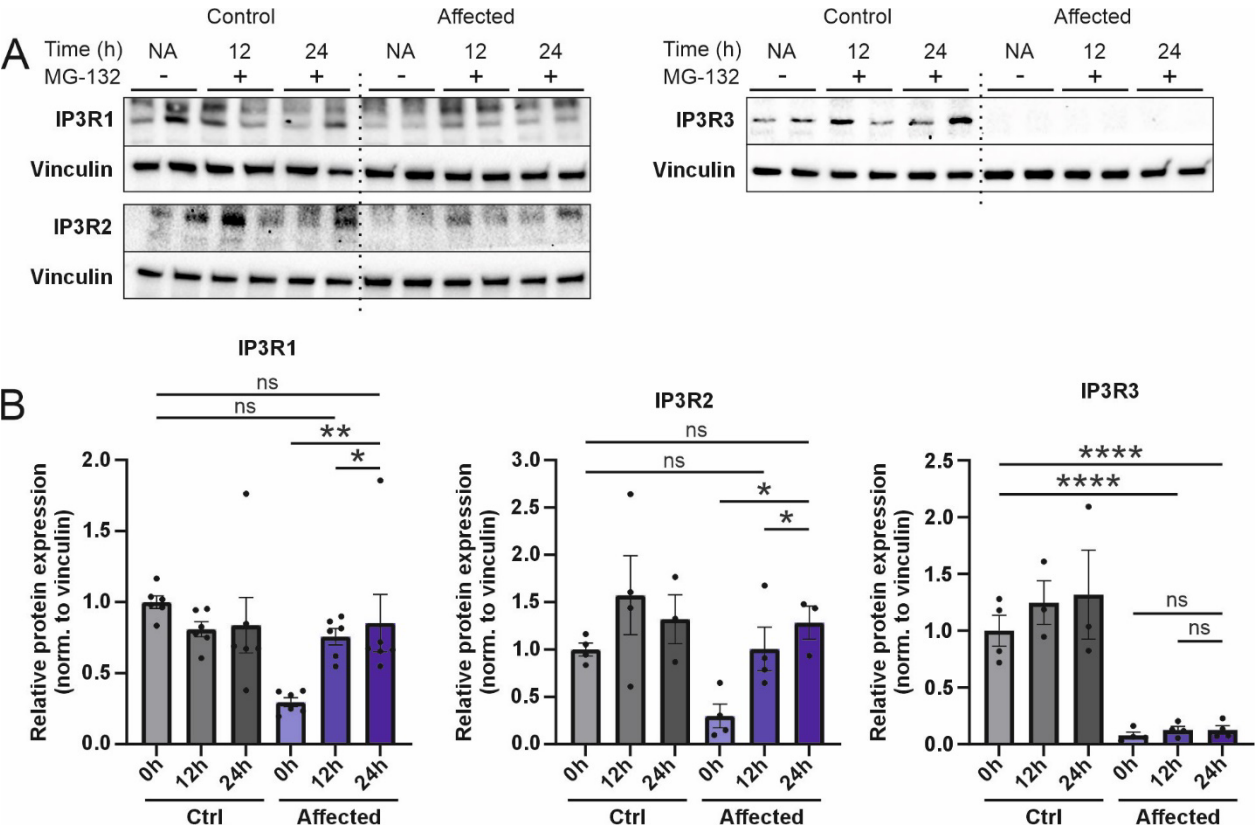


Figure 3.

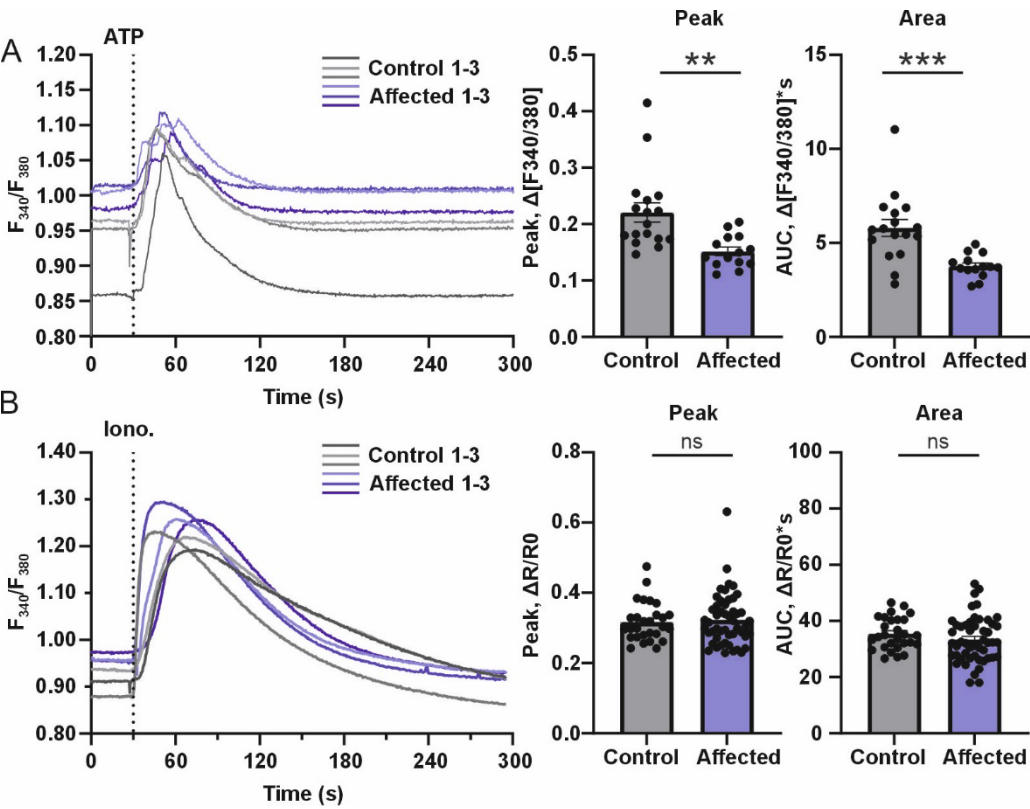


Figure 4.

Supplementary material

Supplementary figure 1. Alignment of the dog IP3R1-3 protein sequences to human, mouse and rat IP₃R1-3 sequences.

Supplementary figure 2. A Western blot of IP3R3 protein.

Supplementary figure 3. Ca²⁺ flux measurements with drug screening system FDSS/μCell

Supplementary table 1. Electromyographic findings of the affected dogs.

Supplementary table 2. Private variants (SNVs/indels) of three affected dogs after filtering the data against 689 control genomes.

Supplementary table 3. Private variants (SNVs/indels) of one affected dog genome after filtering it against 689 control genomes.

Supplementary table 4. Private mobile element insertions (MEI) of one affected dog genome after filtering it against 388 control genomes.

Supplementary table 5. Values used for parameter optimization and detection of shared ROH in PLINK.

Supplementary table 6. Primers used in the study.

Supplementary figure 1. Alignment of the dog IP₃R1-3 protein sequences to human, mouse and rat IP₃R1-3 sequences.

Nonsense variant in ITPR3 leads to depletion of IP₃ receptors in dogs with developmental dental defect and decreased nerve conduction velocity

Marjo K. Hytönen[#], Julius Rönkkö[#], Sruthi Hundi, Tarja S. Jokinen, Emilia Suonto, Eeva Teräväinen, Jonas Donner, Rita La Rovere, Geert Bultynck, Emil Ylikallio, Henna Tyynismaa and Hannes Lohi*

*Corresponding author: Hannes Lohi (hannes.lohi@helsinki.fi), Department of Veterinary Biosciences, Department of Medical and Clinical Genetics, Folkhälsan Research Center, University of Helsinki, Finland.

Percent identity matrices (Clustal 2.1).

ITPR1	Canine	Human	Mouse	Rat
Canine	100	98.73	98.69	98.62
Human	98.73	100	98.62	98.51
Mouse	98.69	98.62	100	99.6
Rat	98.62	98.51	99.6	100

ITPR2	Canine	Human	Mouse	Rat
Canine	100	97.93	95.48	95.3
Human	97.93	100	95.82	95.34
Mouse	95.48	95.82	100	98.41
Rat	95.3	95.34	98.41	100

ITPR3	Canine	Human	Mouse	Rat
Canine	100	96.41	94.27	94.23
Human	96.41	100	95.32	95.24
Mouse	94.27	95.32	100	98.73
Rat	94.23	95.24	98.73	100

Alignment of the dog ITPR1 protein sequence (A0A8I3NKB2) to human (Q14643), mouse (P11881) and rat (P29994) ITPR1 with the Clustal Omega algorithm (CLUSTAL O(1.2.4)).

sp P11881 ITPR1_MOUSE	MSDKMSSFLHIGDICSLEYAEGSTNGFISTLGLVDDRCVVQPEAGDLNPPKKFRDCLFKL	60
sp P29994 ITPR1_RAT	MSDKMSSFLHIGDICSLEYAEGSTNGFISTLGLVDDRCVVQPEAGDLNPPKKFRDCLFKL	60
sp Q14643 ITPR1_HUMAN	MSDKMSSFLHIGDICSLEYAEGSTNGFISTLGLVDDRCVVQPEAGDLNPPKKFRDCLFKL	60
tr A0A8I3NKB2 A0A8I3NKB2_CANLF	MSDKMSSFLHIGDICSLEYAEGSTNGFISTLGLVDDRCVVQPEAGDLNPPKKFRDCLFKL	60

sp P11881 ITPR1_MOUSE	CPMNRYSAQKQFWKAAKPGANSTTDAVLLNKLHHAADLEKKQNETENRKLGLTVIQYGNV	120
sp P29994 ITPR1_RAT	CPMNRYSAQKQFWKAAKPGANSTTDAVLLNKLHHAADLEKKQNETENRKLGLTVIQYGNV	120
sp Q14643 ITPR1_HUMAN	CPMNRYSAQKQFWKAAKPGANSTTDAVLLNKLHHAADLEKKQNETENRKLGLTVIQYGNV	120
tr A0A8I3NKB2 A0A8I3NKB2_CANLF	CPMNRYSAQKQFWKAAKPGANSTTDAVLLNKLHHAADLEKKQNETENRKLGLTVIQYGNV	120

sp P11881 ITPR1_MOUSE	IQLLHLKSNKYLTVNKRLPALLEKNAMRVTLDEAGNEGSWFYIQPFYKLRSIGDSVVIGD	180
sp P29994 ITPR1_RAT	IQLLHLKSNKYLTVNKRLPALLEKNAMRVTLDEAGNEGSWFYIQPFYKLRSIGDSVVIGD	180
sp Q14643 ITPR1_HUMAN	IQLLHLKSNKYLTVNKRLPALLEKNAMRVTLDEAGNEGSWFYIQPFYKLRSIGDSVVIGD	180
tr A0A8I3NKB2 A0A8I3NKB2_CANLF	IQLLHLKSNKYLTVNKRLPALLEKNAMRVTLDEAGNEGSWFYIQPFYKLRSIGDSVVIGD	180

sp P11881 ITPR1_MOUSE	KVVLNPNVAGQPLHASSHQVLNDNPGCNEVNSVNCNTSWKIVLFMKWSDNKDDILKGGDVV	240
sp P29994 ITPR1_RAT	KVVLNPNVAGQPLHASSHQVLNDNPGCNEVNSVNCNTSWKIVLFMKWSDNKDDILKGGDVV	240
sp Q14643 ITPR1_HUMAN	KVVLNPNVAGQPLHASSHQVLNDNPGCNEVNSVNCNTSWKIVLFMKWSDNKDDILKGGDVV	240
tr A0A8I3NKB2 A0A8I3NKB2_CANLF	KVVLNPNVAGQPLHASSHQVLNDNPGCNEVNSVNCNTSWKIVLFMKWSDNKDDILKGGDVV	240

sp P11881 ITPR1_MOUSE	RLFHAEQEKFLTCDEHRKKQHVFLRTTGRQSATSATSSKALWEVEVVQHDPCRGAGYWN	300
sp P29994 ITPR1_RAT	RLFHAEQEKFLTCDEHRKKQHVFLRTTGRQSATSATSSKALWEVEVVQHDPCRGAGYWN	300
sp Q14643 ITPR1_HUMAN	RLFHAEQEKFLTCDEHRKKQHVFLRTTGRQSATSATSSKALWEVEVVQHDPCRGAGYWN	300
tr A0A8I3NKB2 A0A8I3NKB2_CANLF	RLFHAEQEKFLTCDEHRKKQHVFLRTTGRQSATSATSSKALWEVEVVQHDPCRGAGYWN	300

sp P11881 ITPR1_MOUSE	SLFRFKHLATGHYLAEEVDPDFEEECLEFQPSVDPDQDASRSRLRNAQEKMVYSLVSVPE	360
sp P29994 ITPR1_RAT	SLFRFKHLATGHYLAEEVDPDFEEECLEFQPSVDPDQDASRSRLRNAQEKMVYSLVSVPE	360
sp Q14643 ITPR1_HUMAN	SLFRFKHLATGHYLAEEVDPDFEEECLEFQPSVDPDQDASRSRLRNAQEKMVYSLVSVPE	360
tr A0A8I3NKB2 A0A8I3NKB2_CANLF	SLFRFKHLATGHYLAEEVDPDFEEECLEFQPSVDPDQDASRSRLRNAQEKMVYSLVSVPE	360

sp P11881 ITPR1_MOUSE	GNDISSIFELDPPTTLRGGDSLVPFRNSYVRLRHLCNTNTWVHSTNIPIDKEEEKPVMLKIGT	420
sp P29994 ITPR1_RAT	GNDISSIFELDPPTTLRGGDSLVPFRNSYVRLRHLCNTNTWVHSTNIPIDKEEEKPVMLKIGT	420
sp Q14643 ITPR1_HUMAN	GNDISSIFELDPPTTLRGGDSLVPFRNSYVRLRHLCNTNTWVHSTNIPIDKEEEKPVMLKIGT	420
tr A0A8I3NKB2 A0A8I3NKB2_CANLF	GNDISSIFELDPPTTLRGGDSLVPFRNSYVRLRHLCNTNTWVHSTNIPIDKEEEKPVMLKIGT	420

sp P11881 ITPR1_MOUSE	SPLKEDKEAFAIVPVSPAIEVRDLDFANDASKVLGSIAGKLEKGTITQNERRSVTKLLEDL	480
sp P29994 ITPR1_RAT	SPLKEDKEAFAIVPVSPAIEVRDLDFANDASKVLGSIAGKLEKGTITQNERRSVTKLLEDL	480
sp Q14643 ITPR1_HUMAN	SPVKEDKEAFAIVPVSPAIEVRDLDFANDASKVLGSIAGKLEKGTITQNERRSVTKLLEDL	480
tr A0A8I3NKB2 A0A8I3NKB2_CANLF	SPVKEDKEAFAIVPVSPAIEVRDLDFANDASKVLGSIAGKLEKGTITQNERRSVTKLLEDL	480
;***		
sp P11881 ITPR1_MOUSE	VYFVTGGTNSGQDVLEVVFVSKPNRERQKLMREQNILKQIFKLLQAPFTDCGDGPMRLLEE	540
sp P29994 ITPR1_RAT	VYFVTGGTNSGQDVLEVVFVSKPNRERQKLMREQNILKQIFKLLQAPFTDCGDGPMRLLEE	540
sp Q14643 ITPR1_HUMAN	VYFVTGGTNSGQDVLEVVFVSKPNRERQKLMREQNILKQIFKLLQAPFTDCGDGPMRLLEE	540
tr A0A8I3NKB2 A0A8I3NKB2_CANLF	VYFVTGGTNSGQDVLEVVFVSKPNRERQKLMREQNILKQIFKLLQAPFTDCGDGPMRLLEE	540

sp P11881 ITPR1_MOUSE	LGDRHAPFRHICRLCYRVLRRHSQQDYRKNOEYIAKQFGFMQKQIGYDVLAEDTITALLH	600
sp P29994 ITPR1_RAT	LGDRHAPFRHICRLCYRVLRRHSQQDYRKNOEYIAKQFGFMQKQIGYDVLAEDTITALLH	600
sp Q14643 ITPR1_HUMAN	LGDRHAPFRHICRLCYRVLRRHSQQDYRKNOEYIAKQFGFMQKQIGYDVLAEDTITALLH	600
tr A0A8I3NKB2 A0A8I3NKB2_CANLF	LGDRHAPFRHICRLCYRVLRRHSQQDYRKNOEYIAKQFGFMQKQIGYDVLAEDTITALLH	600

sp P11881 ITPR1_MOUSE	NNRKILLEKHITAAEIDTFVSLVRKNRPRFLDYLDLCSVMNKSIPVTQELICKAVLNPT	660
sp P29994 ITPR1_RAT	NNRKILLEKHITAAEIDTFVSLVRKNRPRFLDYLDLCSVMNKSIPVTQELICKAVLNPT	660
sp Q14643 ITPR1_HUMAN	NNRKILLEKHITAAEIDTFVSLVRKNRPRFLDYLDLCSVMNKSIPVTQELICKAVLNPT	660
tr A0A8I3NKB2 A0A8I3NKB2_CANLF	NNRKILLEKHITAAEIDTFVSLVRKNRPRFLDYLDLCSVMNKSIPVTQELICKAVLNPT	660

sp P11881 ITPR1_MOUSE	NADILIIETKLVLRSFEFEGVS-TGENALEAGEDEEEVWLFWRDSNKEIRSKSVRELAQDA	719
sp P29994 ITPR1_RAT	NADILIIETKLVLRSFEFEGVS-TGENALEAGEDEEEVWLFWRDSNKEIRSKSVRELAQDA	719
sp Q14643 ITPR1_HUMAN	NADILIIETKLVLRSFEFEGVSSTGENALEAGEDEEEVWLFWRDSNKEIRSKSVRELAQDA	720
tr A0A8I3NKB2 A0A8I3NKB2_CANLF	NADILIIETKLVLRSFEFEGVS-TGENALEAGEDEEEVWLFWRDSNKEIRSKSVRELAQDA	719

sp P11881 ITPR1_MOUSE	KEGQKEDRDVLSYYRYQLNLFARMCLDRQYLAINESGQLDVLILRCMSDENLPYDLRA	779
sp P29994 ITPR1_RAT	KEGQKEDRDVLSYYRYQLNLFARMCLDRQYLAINESGQLDVLILRCMSDENLPYDLRA	779
sp Q14643 ITPR1_HUMAN	KEGQKEDRDVLSYYRYQLNLFARMCLDRQYLAINESGQLDVLILRCMSDENLPYDLRA	780
tr A0A8I3NKB2 A0A8I3NKB2_CANLF	KEGQKEDRDVLSYYRYQLNLFARMCLDRQYLAINESGQLDVLILRCMSDENLPYDLRA	779

sp P11881 ITPR1_MOUSE	SFCRLMLHMHVDRDPQEQTVPVKYARLWSEIPSEIAIDDYSSGTSKDEIKERFAQTMEF	839
sp P29994 ITPR1_RAT	SFCRLMLHMHVDRDPQEQTVPVKYARLWSEIPSEIAIDDYSSGTSKDEIKERFAQTMEF	839
sp Q14643 ITPR1_HUMAN	SFCRLMLHMHVDRDPQEQTVPVKYARLWSEIPSEIAIDDYSSGTSKDEIKERFAQTMEF	840
tr A0A8I3NKB2 A0A8I3NKB2_CANLF	SFCRLMLHMHVDRDPQEQTVPVKYARLWSEIPSEIAIDDYSSGTSKDEIKERFAQTMEF	839

sp P11881 ITPR1_MOUSE	VEEYLRDVCQRFPPFSDKEKNKLTFEVNLARNLIYFGFYNSDLLRLTKILLAILDCVH	899
sp P29994 ITPR1_RAT	VEEYLRDVCQRFPPFSDKEKNKLTFEVNLARNLIYFGFYNSDLLRLTKILLAILDCVH	899
sp Q14643 ITPR1_HUMAN	VEEYLRDVCQRFPPFSDKEKNKLTFEVNLARNLIYFGFYNSDLLRLTKILLAILDCVH	900
tr A0A8I3NKB2 A0A8I3NKB2_CANLF	VEEYLRDVCQRFPPFSDKEKNKLTFEVNLARNLIYFGFYNSDLLRLTKILLAILDCVH	899

sp P11881 ITPR1_MOUSE	VTTIFPISKMTKGEENKG-----SNVMRSIHGVGELMTQVVLRGGGFLPMTMAAA	950
sp P29994 ITPR1_RAT	VTTIFPISKMTKGEENKG-----SNVMRSIHGVGELMTQVVLRGGGFLPMTMAAA	950
sp Q14643 ITPR1_HUMAN	VTTIFPISKMAKGEENKGNDVEKLSNVMSRSHGVGELMTQVVLRGGGFLPMTMAAA	960
tr A0A8I3NKB2 A0A8I3NKB2_CANLF	VTTIFPISKMAKGEENKGKNNVAKWRSSNVMSRSHGVGELMTQVVLRGGGFLPMTMAAA	959
*****;*****		
sp P11881 ITPR1_MOUSE	PEGNVKQAEPEKEDIMVMDTKLKIIEILQFILNVRLDYRISCLLCIFKREFDESNSQSSE	1010
sp P29994 ITPR1_RAT	PEGNVKQAEPEKEDIMVMDTKLKIIEILQFILNVRLDYRISCLLCIFKREFDESNSQSSE	1010
sp Q14643 ITPR1_HUMAN	PEGNVKQAEPEKEDIMVMDTKLKIIEILQFILNVRLDYRISCLLCIFKREFDESNSQTSE	1020
tr A0A8I3NKB2 A0A8I3NKB2_CANLF	PEGNVKQAEPEKEDIMVMDTKLKIIEILQFILNVRLDYRISCLLCIFKREFDESNSQTSE	1019
*****;*****		
sp P11881 ITPR1_MOUSE	TSSGNSSQEGPSNVPGALDFEHIIEQAEGIFGGSEENTPLDLDHGGRTFLRVLLHLTMH	1070
sp P29994 ITPR1_RAT	TSSGNSSQEGPSNVPGALDFEHIIEQAEGIFGGSEENTPLDLDHGGRTFLRVLLHLTMH	1070
sp Q14643 ITPR1_HUMAN	TSSGNSSQEGPSNVPGALDFEHIIEQAEGIFGGSEENTPLDLDHGGRTFLRVLLHLTMH	1080
tr A0A8I3NKB2 A0A8I3NKB2_CANLF	TSSGNSSQEGPSNVPGTLDFFEHIIEQAEGIFGGSEENTPLDLDHGGRTFLRVLLHLTMH	1079
*****;*****		
sp P11881 ITPR1_MOUSE	DYPPLVSGALQLLFRHFSQRQEVQLQAFKQVQLLVTSQDVVDNYKQIKQDLQQLRSIVEKSE	1130
sp P29994 ITPR1_RAT	DYPPLVSGALQLLFRHFSQRQEVQLQAFKQVQLLVTSQDVVDNYKQIKQDLQQLRSIVEKSE	1130
sp Q14643 ITPR1_HUMAN	DYPPLVSGALQLLFRHFSQRQEVQLQAFKQVQLLVTSQDVVDNYKQIKQDLQQLRSIVEKSE	1140
tr A0A8I3NKB2 A0A8I3NKB2_CANLF	DYPPLVSGALQLLFRHFSQRQEVQLQAFKQVQLLVTSQDVVDNYKQIKQDLQQLRSIVEKSE	1139

sp P11881 ITPR1_MOUSE	LWVYKGGQGPDEPMDGASGENEHKKTTEEGTSKPLKHESTSSYNRVVKEILIRLSKLCVQE	1190
sp P29994 ITPR1_RAT	LWVYKGGQGPDEPMDGASGENEHKKTTEEGTSKPLKHESTSSYNRVVKEILIRLSKLCVQE	1190
sp Q14643 ITPR1_HUMAN	LWVYKGGQGPDEPMDGASGENEHKKTTEEGNNKPKQKHESTSSYNRVVKEILIRLSKLCVQE	1200
tr A0A8I3NKB2 A0A8I3NKB2_CANLF	LWVYKGGQGPDEAMDGASGENEHKKTTEEGHNSQKHESTSSYNRVVKEILIRLSKLCVQE	1199

sp P11881 ITPR1_MOUSE	SASVRSRKQQQRLLRNMGAHAVVLELLQIPYEKAEDTKMQEIMRLAHEFLQNFCAGNQ	1250
sp P29994 ITPR1_RAT	SASVRSRKQQQRLLRNMGAHAVVLELLQIPYEKAEDTKMQEIMRLAHEFLQNFCAGNQ	1250
sp Q14643 ITPR1_HUMAN	SASVRSRKQQQRLLRNMGAHAVVLELLQIPYEKAEDTKMQEIMRLAHEFLQNFCAGNQ	1260
tr A0A8I3NKB2 A0A8I3NKB2_CANLF	SASVRSRKQQQRLLRNMGAHAVVLELLQIPYEKAEDTKMQEIMRLAHEFLQNFCAGNQ	1259

sp P11881 ITPR1_MOUSE	NQALLHKKHINFLNPGILEAVTMQHI FMNNFQLCSEINERVVQHFVHCIEHGRNVQYIK	1310
sp P29994 ITPR1_RAT	NQALLHKKHINFLNPGILEAVTMQHI FMNNFQLCSEINERVVQHFVHCIEHGRNVQYIK	1310
sp Q14643 ITPR1_HUMAN	NQALLHKKHINFLNPGILEAVTMQHI FMNNFQLCSEINERVVQHFVHCIEHGRNVQYIK	1320
tr A0A8I3NKB2 A0A8I3NKB2_CANLF	NQALLHKKHINFLNPGILEAVTMQHI FMNNFQLCSEINERVVQHFVHCIEHGRNVQYIK	1319

sp P11881 ITPR1_MOUSE	FLQITVKAEGKFIKKQDMVMAELVNSGEDVLVFNDRASFQTLIQMMSRSDRMDENSP	1370
sp P29994 ITPR1_RAT	FLQITVKAEGKFIKKQDMVMAELVNSGEDVLVFNDRASFQTLIQMMSRSDRMDENSP	1370
sp Q14643 ITPR1_HUMAN	FLQITVKAEGKFIKKQDMVMAELVNSGEDVLVFNDRASFQTLIQMMSRSDRMDENSP	1380
tr A0A8I3NKB2 A0A8I3NKB2_CANLF	FLQITVKAEGKFIKKQDMVMAELVNSGEDVLVFNDRASFQTLIQMMSRSDRMDENSP	1379

727	sp P11881 ITPR1_MOUSE	L-MYHIHLVELLAVCTEGKNVYTEIKCNSLLPLDDIVRVVTHEDCIPEVKIAYINFLNHC	1429
728	sp P29994 ITPR1_RAT	LFMYHIHLVELLAVCTEGKNVYTEIKCNSLLPLDDIVRVVTHEDCIPEVKIAYINFLNHC	1430
729	sp Q14643 ITPR1_HUMAN	L-MYHIHLVELLAVCTEGKNVYTEIKCNSLLPLDDIVRVVTHEDCIPEVKIAYINFLNHC	1439
730	tr A0A8I3NKB2 A0A8I3NKB2_CANLF	L-MYHIHLVELLAVCTEGKNVYTEIKCNSLLPLDDIVRVVTHEDCIPEVKIAYINFLNHC	1438
731		* ****	
732	sp P11881 ITPR1_MOUSE	YVDTEVEMKEIYTSNHMMKLFENFLVDICRACNNTSDRKHADSILEKYVTEIVMSIVTTF	1489
733	sp P29994 ITPR1_RAT	YVDTEVEMKEIYTSNHMMKLFENFLVDICRACNNTSDRKHADSVLEKYVTEIVMSIVTTF	1490
734	sp Q14643 ITPR1_HUMAN	YVDTEVEMKEIYTSNHMMKLFENFLVDICRACNNTSDRKHADSILEKYVTEIVMSIVTTF	1499
735	tr A0A8I3NKB2 A0A8I3NKB2_CANLF	YVDTEVEMKEIYTSNHMMKLFENFLVDICRACNNTSDRKHADSILEKYVTEIVMSIVTTF	1498
736		*****;*****	
737	sp P11881 ITPR1_MOUSE	FSSPFSQDSTTLQTRQPVFVQLLQGVFRVYHCNWLMPQSQKASVESCIRVLSDVAKSRAIA	1549
738	sp P29994 ITPR1_RAT	FSSPFSQDSTTLQTRQPVFVQLLQGVFRVYHCNWLMPQSQKASVESCIRVLSDVAKSRAIA	1550
739	sp Q14643 ITPR1_HUMAN	FSSPFSQDSTTLQTRQPVFVQLLQGVFRVYHCNWLMPQSQKASVESCIRVLSDVAKSRAIA	1559
740	tr A0A8I3NKB2 A0A8I3NKB2_CANLF	FSSPFSQDSTTLQTRQPVFVQLLQGVFRVYHCNWLMPQSQKASVESCIRVLSDVAKSRAIA	1558
741		*****	
742	sp P11881 ITPR1_MOUSE	IPVDLDSQVNNLFLKSHNIVQKTALNWRLSARNAARRDSVLAASRDYRNIIERLQDIVSA	1609
743	sp P29994 ITPR1_RAT	IPVDLDSQVNNLFLKSHNIVQKTALNWRLSARNAARRDSVLAASRDYRNIIERLQDIVSA	1610
744	sp Q14643 ITPR1_HUMAN	IPVDLDSQVNNLFLKSHNIVQKTAMNWRLSARNAARRDSVLAASRDYRNIIERLQDIVSA	1619
745	tr A0A8I3NKB2 A0A8I3NKB2_CANLF	IPVDLDSQVNNLFLKSHNIVQKTAMNWRLTARNAARRDSVLAASRDYRNIIERLQDIVSA	1618
746		*****;*****	
747	sp P11881 ITPR1_MOUSE	LEDRLRPLVQAELSVLVDVLRHPELLFPENTDARRKCESGGFICKLIKHTKQLEENEK	1669
748	sp P29994 ITPR1_RAT	LEDRLRPLVQAELSVLVDVLRHPELLFPENTDARRKCESGGFICKLIKHTKQLEENEK	1670
749	sp Q14643 ITPR1_HUMAN	LEDRLRPLVQAELSVLVDVLRHPELLFPENTDARRKCESGGFICKLIKHTKQLEENEK	1679
750	tr A0A8I3NKB2 A0A8I3NKB2_CANLF	LEDRLRPLVQAELSVLVDVLRHPELLFPENTDARRKCESGGFICKLIKHTKQLEENEK	1678
751		*****	
752	sp P11881 ITPR1_MOUSE	LCIKVLQTLREMMTKDRGYGEKQISIDSENAELPQAPAEANSTEQELEPSPLRQLEDH	1729
753	sp P29994 ITPR1_RAT	LCIKVLQTLREMMTKDRGYGEKQISIDSENAELPQAPAEANSTEQELEPSPLRQLEDH	1730
754	sp Q14643 ITPR1_HUMAN	LCIKVLQTLREMMTKDRGYGEKLIISIDELDNAELPPAPDSEENAT-EELEPSPLRQLEDH	1738
755	tr A0A8I3NKB2 A0A8I3NKB2_CANLF	LCIKVLQTLREMMTKDRGYGEKLIISIDELDNAELPPAPDSEENAT-EELEPSPLRQLEDH	1737
756		*****;*****;*****;*****;*****;*****	
757	sp P11881 ITPR1_MOUSE	KRGEALRQILVNRYGNIRPSGRRESLTSFGNGPLSPGGPSKPGGGGGPGSSSTSRGEM	1789
758	sp P29994 ITPR1_RAT	KRGEALRQILVNRYGNIRPSGRRESLTSFGNGPLSPGGPSKPGGGGGPGSSSTSRGEM	1790
759	sp Q14643 ITPR1_HUMAN	KRGEALRQILVNRYGNIRPSGRRESLTSFGNGPLSAGGPGKPGGGGGSGSSMSRGEM	1798
760	tr A0A8I3NKB2 A0A8I3NKB2_CANLF	KRGEALRQILVNRYGNIRPSGRRESLTSFGNGPLSPGGPSKPGGGGGSGSSSTSRGEM	1797
761		*****;*****;*****;*****;*****;*****	
762	sp P11881 ITPR1_MOUSE	SLAEVQCHLDKEGASNLVIDLIMNASSDRVFHESILLAIALLLEGGNTTIQHSFFCRLTED	1849
763	sp P29994 ITPR1_RAT	SLAEVQCHLDKEGASNLVIDLIMNASSDRVFHESILLAIALLLEGGNTTIQHSFFCRLTED	1850
764	sp Q14643 ITPR1_HUMAN	SLAEVQCHLDKEGASNLVIDLIMNASSDRVFHESILLAIALLLEGGNTTIQHSFFCRLTED	1858
765	tr A0A8I3NKB2 A0A8I3NKB2_CANLF	SLAEVQCHLDKEGASNLVIDLIMNASSDRVFHESILLAIALLLEGGNTTIQHSFFCRLTED	1857
766		*****	
767	sp P11881 ITPR1_MOUSE	KKSEKFFKVFYDRMKVAQQEIKATVTVNTSDLGNNKKDDDEVDRDAPSRKKAKEPTTQITE	1909
768	sp P29994 ITPR1_RAT	KKSEKFFKVFYDRMKVAQQEIKATVTVNTSDLGNNKKDDDEVDRDAPSRKKAKEPTTQITE	1910
769	sp Q14643 ITPR1_HUMAN	KKSEKFFKVFYDRMKVAQQEIKATVTVNTSDLGNNKKDDDEVDRDAPSRKKAKEPTTQITE	1918
770	tr A0A8I3NKB2 A0A8I3NKB2_CANLF	KKSEKFFKVFYDRMKVAQQEIKATVTVNTSDLGNNKKDDDEVDRDAPSRKKAKEPTTQITE	1917
771		*****;*****;*****;*****;*****;*****	
772	sp P11881 ITPR1_MOUSE	EVDRDQLEESAATRKAFTTFRREADPDDHYQSSEGTQATTDKAKDDLEMSAVITIMQPIL	1969
773	sp P29994 ITPR1_RAT	EVDRDQLEESAATRKAFTTFRREADPDDHYQSSEGTQATTDKAKDDLEMSAVITIMQPIL	1970
774	sp Q14643 ITPR1_HUMAN	EVDRDQLEESAATRKAFTTFRREADPDDHYQPEGTQATADKAKDDLEMSAVITIMQPIL	1978
775	tr A0A8I3NKB2 A0A8I3NKB2_CANLF	EARDQLEESAATRKAFTTFRREADPDDHYQSSEGAQATADKTKDEMSAVITIMQPIL	1977
776		*.*****;*****;*****;*****;*****;*****	
777	sp P11881 ITPR1_MOUSE	RFLQLLCCENHNRLQNLFRQCQNNKNTYNLVCETLQFLDCICGSTTGGLGLGLYINEKNV	2029
778	sp P29994 ITPR1_RAT	RFLQLLCCENHNRLQNLFRQCQNNKNTYNLVCETLQFLDCICGSTTGGLGLGLYINEKNV	2030
779	sp Q14643 ITPR1_HUMAN	RFLQLLCCENHNRLQNLFRQCQNNKNTYNLVCETLQFLDCICGSTTGGLGLGLYINEKNV	2038
780	tr A0A8I3NKB2 A0A8I3NKB2_CANLF	RFLQLLCCENHNRLQNLFRQCQNNKNTYNLVCETLQFLDCICGSTTGGLGLGLYINEKNV	2037
781		*****	
782	sp P11881 ITPR1_MOUSE	ALINQTLLESLEYCQGPCHENQNCIATHESENGIDIITAILINDINPLGKKRMDLVLELKN	2089
783	sp P29994 ITPR1_RAT	ALINQTLLESLEYCQGPCHENQNCIATHESENGIDIITAILINDINPLGKKRMDLVLELKN	2090
784	sp Q14643 ITPR1_HUMAN	ALINQTLLESLEYCQGPCHENQNCIATHESENGIDIITAILINDINPLGKKRMDLVLELKN	2098
785	tr A0A8I3NKB2 A0A8I3NKB2_CANLF	ALINQTLLESLEYCQGPCHENQNCIATHESENGIDIITAILINDINPLGKKRMDLVLELKN	2097
786		*****	
787	sp P11881 ITPR1_MOUSE	NASKLLLAIMESRHDSENAERILYNMRPKELVEVIKKAYMQGEVEFEDGENGEDGAASPR	2149
788	sp P29994 ITPR1_RAT	NASKLLLAIMESRHDSENAERILYNMRPKELVEVIKKAYMQGEVEFEDGENGEDGAASPR	2150
789	sp Q14643 ITPR1_HUMAN	NASKLLLAIMESRHDSENAERILYNMRPKELVEVIKKAYMQGEVEFEDGENGEDGAASPR	2158
790	tr A0A8I3NKB2 A0A8I3NKB2_CANLF	NASKLLLAIMESRHDSENAERILYNMRPKELVEVIKKAYMQGEVEFEDGENGEDGAASPR	2157
791		*****	
792	sp P11881 ITPR1_MOUSE	NVGHNIYILAHQLARHNKELQTMLKPGGQVDGDEALEFYAKHTAQIEIVRLDRTMEQIVF	2209
793	sp P29994 ITPR1_RAT	NVGHNIYILAHQLARHNKELQTMLKPGGQVDGDEALEFYAKHTAQIEIVRLDRTMEQIVF	2210
794	sp Q14643 ITPR1_HUMAN	NVGHNIYILAHQLARHNKELQSMKPGGQVDGDEALEFYAKHTAQIEIVRLDRTMEQIVF	2218
795	tr A0A8I3NKB2 A0A8I3NKB2_CANLF	NVGHNIYILAHQLARHNKELQTMLKPGGQVDGDEALEFYAKHTAQIEIVRLDRTMEQIVF	2217
796		*****;*****	
797	sp P11881 ITPR1_MOUSE	PVPSICEFLTKEKSLRIYYTTERDEQGSKINDFFLRSEDLFNEMNWQKKLRAQPVLYWCA	2269
798	sp P29994 ITPR1_RAT	PVPSICEFLTKEKSLRIYYTTERDEQGSKINDFFLRSEDLFNEMNWQKKLRAQPVLYWCA	2270
799	sp Q14643 ITPR1_HUMAN	PVPSICEFLTKEKSLRIYYTTERDEQGSKINDFFLRSEDLFNEMNWQKKLRAQPVLYWCA	2278
800	tr A0A8I3NKB2 A0A8I3NKB2_CANLF	PVPSICEFLTKEKSLRIYYTTERDEQGSKINDFFLRSEDLFNEMNWQKKLRAQPVLYWCA	2277
801		*****	
802	sp P11881 ITPR1_MOUSE	RNMSFWSSISFNLAVLMNLLVAFFYPFKGVRGGTLEPHWSGLLWTAMLISLAIVIALPKP	2329
803	sp P29994 ITPR1_RAT	RNMSFWSSISFNLAVLMNLLVAFFYPFKGVRGGTLEPHWSGLLWTAMLISLAIVIALPKP	2330
804	sp Q14643 ITPR1_HUMAN	RNMSFWSSISFNLAVLMNLLVAFFYPFKGVRGGTLEPHWSGLLWTAMLISLAIVIALPKP	2338
805	tr A0A8I3NKB2 A0A8I3NKB2_CANLF	RNMSFWSSISFNLAVLMNLLVAFFYPFKGVRGGTLEPHWSGLLWTAMLISLAIVIALPKP	2337
806		*****	
807	sp P11881 ITPR1_MOUSE	HGIRALIASTILRLIFSGLQPTLFLLGAFNVCNKIIIFLMSFVGNCGTFTRGYRAMVLVDV	2389
808	sp P29994 ITPR1_RAT	HGIRALIASTILRLIFSGLQPTLFLLGAFNVCNKIIIFLMSFVGNCGTFTRGYRAMVLVDV	2390
809	sp Q14643 ITPR1_HUMAN	HGIRALIASTILRLIFSGLQPTLFLLGAFNVCNKIIIFLMSFVGNCGTFTRGYRAMVLVDV	2398
810	tr A0A8I3NKB2 A0A8I3NKB2_CANLF	HGIRALIASTILRLIFSGLQPTLFLLGAFNVCNKIIIFLMSFVGNCGTFTRGYRAMVLVDV	2397
811		*****	

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sp|P11881|ITPR1_MOUSE      EFLYHLLYLLICAMGLFVHEFFYSLLLFDFLVYREETLLNVIKSVTRNGRSIIITAVLALI      2449
sp|P29994|ITPR1_RAT        EFLYHLLYLLICAMGLFVHEFFYSLLLFDFLVYREETLLNVIKSVTRNGRPIIITAAALALI      2450
sp|Q14643|ITPR1_HUMAN      EFLYHLLYLVICAMGLFVHEFFYSLLLFDFLVYREETLLNVIKSVTRNGRSIIITAVLALI      2458
tr|A0A8I3NKB2|A0A8I3NKB2_CANLF EFLYHLLYLLICAMGLFVHEFFYSLLLFDFLVYREETLLNVIKSVTRNGRSIIITAVLALI      2457
*****:*****

sp|P11881|ITPR1_MOUSE      LVYLFISVIGYLFKKDDFILEVDRLPNETAVPETGESLANDFLYSDVCRVETGENCTSPAP      2509
sp|P29994|ITPR1_RAT        LVYLFISVIGYLFKKDDFILEVDRLPNETAGPETGESLANDFLYSDVCRVETGENCTSPAP      2510
sp|Q14643|ITPR1_HUMAN      LVYLFISVIGYLFKKDDFILEVDRLPNETAVPETGESLASEFLFSDVCRVESGENCSPAP      2518
tr|A0A8I3NKB2|A0A8I3NKB2_CANLF LVYLFISVIGYLFKKDDFILEVDRLPNETALPEAGESLASEFLYSDVCRVETGENCSPAP      2517
*****:*****

sp|P11881|ITPR1_MOUSE      KEELLPAEETEQDKHEHTCETLLMCIVTVLSHGLRSGGGVGDVLRKPSKEEPLFAARVIYD      2569
sp|P29994|ITPR1_RAT        KEELLPVEETEQDKHEHTCETLLMCIVTVLSHGLRSGGGVGDVLRKPSKEEPLFAARVIYD      2570
sp|Q14643|ITPR1_HUMAN      REELVPAEETEQDKHEHTCETLLMCIVTVLSHGLRSGGGVGDVLRKPSKEEPLFAARVIYD      2578
tr|A0A8I3NKB2|A0A8I3NKB2_CANLF KEELVLAETEQDKHEHTCETLLMCIVTVLSHGLRSGGGVGDVLRKPSKEEPLFAARVIYD      2577
:***: *****

sp|P11881|ITPR1_MOUSE      LLFFFMVIIIVLNLIFGVIIIDTFADLRSEKQKKEEILKTTTCFICGLERDKFDNKTVTTEE      2629
sp|P29994|ITPR1_RAT        LLFFFMVIIIVLNLIFGVIIIDTFADLRSEKQKKEEILKTTTCFICGLERDKFDNKTVTTEE      2630
sp|Q14643|ITPR1_HUMAN      LLFFFMVIIIVLNLIFGVIIIDTFADLRSEKQKKEEILKTTTCFICGLERDKFDNKTVTTEE      2638
tr|A0A8I3NKB2|A0A8I3NKB2_CANLF LLFFFMVIIIVLNLIFGVIIIDTFADLRSEKQKKEEILKTTTCFICGLERDKFDNKTVTTEE      2637
*****

sp|P11881|ITPR1_MOUSE      HIKEEHNMWHYLCFIVLVVKDSTEYTGPESYVAEMIRERNLDWFFMRAMSLVSSDSEG      2689
sp|P29994|ITPR1_RAT        HIKEEHNMWHYLCFIVLVVKDSTEYTGPESYVAEMIRERNLDWFFMRAMSLVSSDSEG      2690
sp|Q14643|ITPR1_HUMAN      HIKEEHNMWHYLCFIVLVVKDSTEYTGPESYVAEMIKERNLDWFFMRAMSLVSSDSEG      2698
tr|A0A8I3NKB2|A0A8I3NKB2_CANLF HIKEEHNMWHYLCFIVLVVKDSTEYTGPESYVAEMIKERNLDWFFMRAMSLVSSDSEG      2697
*****:*****

sp|P11881|ITPR1_MOUSE      EQNELRNLQEKLESTMKLVNLSGQLSELKDMQTEQRKQKQRIGLLGHPPHMNVNPQQPA      2749
sp|P29994|ITPR1_RAT        EQNELRNLQEKLESTMKLVNLSGQLSELKDMQTEQRKQKQRIGLLGHPPHMNVNPQQPA      2750
sp|Q14643|ITPR1_HUMAN      EQNELRNLQEKLESTMKLVNLSGQLSELKDMQTEQRKQKQRIGLLGHPPHMNVNPQQPA      2758
tr|A0A8I3NKB2|A0A8I3NKB2_CANLF EQNELRNLQEKLESTMKLVNLSGQLSELKDMQTEQRKQKQRIGLLGHPPHMNVNPQQPA      2757
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Alignment of the dog ITPR2 protein sequence (A0A8I3PW88) to human (Q14571), mouse (Q9Z329) and rat (P29995) ITPR2 with the Clustal Omega algorithm (CLUSTAL O(1.2.4)).

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sp|Q9Z329|ITPR2_MOUSE      MSDKMSSFLYIGDIVSLYAEGSVNGFISTLGLVDDRCVVHPEAGDLANPPKKFRDCLFKV      60
sp|P29995|ITPR2_RAT        MSDKMSSFLYIGDIVSLYAEGSVNGFISTLGLVDDRCVVHPEAGDLTNPPKKFRDCLFKV      60
tr|A0A8I3PW88|A0A8I3PW88_CANLF MSDKMSSFLYIGDIVSLYAEGSVNGFISTLGLVDDRCVVHPEAGDLANPPKKFRDCLFKV      60
sp|Q14571|ITPR2_HUMAN      MTEKMSSFLYIGDIVSLYAEGSVNGFISTLGLVDDRCVVHPEAGDLANPPKKFRDCLFKV      60
*:*****

sp|Q9Z329|ITPR2_MOUSE      CPMNRYSAQKQYWKAKQAKQGNHTEAALLKKLQHAAELEQKQNESENKKLLGEIVKYSNV      120
sp|P29995|ITPR2_RAT        CPMNRYSAQKQYWKAKQAKQGNHTEAALLKKLQHAAELEQKQNESENKKLLGEIVKYSKV      120
tr|A0A8I3PW88|A0A8I3PW88_CANLF CPMNRYSAQKQYWKAKQAKQGNHTEAALLKKLQHAAELEQKQNESENKKLLGEIVKYSNV      120
sp|Q14571|ITPR2_HUMAN      CPMNRYSAQKQYWKAKQAKQGNHTEAALLKKLQHAAELEQKQNESENKKLLGEIVKYSNV      120
*****:*

sp|Q9Z329|ITPR2_MOUSE      IQLLHIKSNKYLTVNKRLPALLEKNAMRVSLDAAGNEGSWFYIHPFWKLRSSEGDNIIVGVD      180
sp|P29995|ITPR2_RAT        IQLLHIKSNKYLTVNKRLPALLEKNAMRVSLDAAGNEGSWFYIHPFWKLRSSEGDNIIVGVD      180
tr|A0A8I3PW88|A0A8I3PW88_CANLF IQLLHIKSNKYLTVNKRLPALLEKNAMRVSLDAAGNEGSWFYIHPFWKLRSSEGDNIIVGVD      180
sp|Q14571|ITPR2_HUMAN      IQLLHIKSNKYLTVNKRLPALLEKNAMRVSLDAAGNEGSWFYIHPFWKLRSSEGDNIIVGVD      180
*****

sp|Q9Z329|ITPR2_MOUSE      KVVLMFPVNAGQPLHASNVLELDNPGCKEIVNAVNCNTSWKITLFMKFSSYREDVLKGGDVV      240
sp|P29995|ITPR2_RAT        KVVLMFPVNAGQPLHASNVLELDNPGCKEIVNAVNCNTSWKITLFMKFSSYREDVLKGGDVV      240
tr|A0A8I3PW88|A0A8I3PW88_CANLF KVVLMFPVNAGQPLHASNIELLDNPGCKEIVNAVNCNTSWKITLFMKYSSYREDVLKGGDVV      240
sp|Q14571|ITPR2_HUMAN      KVVLMFPVNAGQPLHASNIELLDNPGCKEIVNAVNCNTSWKITLFMKYSSYREDVLKGGDVV      240
*****:*****

sp|Q9Z329|ITPR2_MOUSE      RLFHAEQEKFLTCDDEYKKQHIFLRTTLRQSATSATSSKALWEIEVVHHDPCRGGAGQWN      300
sp|P29995|ITPR2_RAT        RLFHAEQEKFLTCDDEYKKQHIFLRTTLRQSATSATSSKALWEIEVVHHDPCRGGAGQWN      300
tr|A0A8I3PW88|A0A8I3PW88_CANLF RLFHAEQEKFLTCDDEYKKQHIFLRTTLRQSATSATSSKALWEIEVVHHDPCRGGAGQWN      300
sp|Q14571|ITPR2_HUMAN      RLFHAEQEKFLTCDDEYKKQHIFLRTTLRQSATSATSSKALWEIEVVHHDPCRGGAGQWN      300
*****:*****

sp|Q9Z329|ITPR2_MOUSE      SLFRFKHLATGNYLAAELNPDYRDAQNEGNKVDGEIPTPKKKRQAGEKIMYTLVSVPHG      360
sp|P29995|ITPR2_RAT        SLFRFKHLATGNYLAAELNPDYRDAQNEGKTVRDGEIPTSKKKKHQAGEKIMYTLVSVPHG      360
tr|A0A8I3PW88|A0A8I3PW88_CANLF SLFRFKHLATGNYLAAELNPDYRDAQNEGNKVRDGEIPTSKKKKRQAGEKIMYTLVSVPHG      360
sp|Q14571|ITPR2_HUMAN      SLFRFKHLATGNYLAAELNPDYRDAQNEGNKVRDGVPTSSKKKKRQAGEKIMYTLVSVPHG      360
*****:*****

sp|Q9Z329|ITPR2_MOUSE      NDIASLFELDATTLQRADCLVPRNSYVRLRHLCTNTWVTSTTIPIDTEERPVMLKIGTC      420
sp|P29995|ITPR2_RAT        NDIASLFELDATTLQRADCLVPRNSYVRLRHLCTNTWVTSTTIPIDTEERPVMLKIGTC      420
tr|A0A8I3PW88|A0A8I3PW88_CANLF NDIASLFELDATTLQRADCLVPRNSYVRLRHLCTNTWVTSTTIPIDTEERPVMLKIGTC      420
sp|Q14571|ITPR2_HUMAN      NDIASLFELDATTLQRADCLVPRNSYVRLRHLCTNTWVTSTTIPIDTEERPVMLKIGTC      420
*****:*****

sp|Q9Z329|ITPR2_MOUSE      QTKEDKEAFAIVCVPLSEVRDLDFANDANKVLATTVKKLENGSITQNERRFVTKLLEDLI      480
sp|P29995|ITPR2_RAT        QTKEDKEAFAIVCVPLSEVRDLDFANDANKVLATTVKKLENGSITQNERRFVTKLLEDLI      480
tr|A0A8I3PW88|A0A8I3PW88_CANLF QTKEDKEAFAIVSVPLSEVRDLDFANDANKVLATTVKKLENGITQNERRFVTKLLEDLI      480
sp|Q14571|ITPR2_HUMAN      QTKEDKEAFAIVSVPLSEVRDLDFANDANKVLATTVKKLENGITQNERRFVTKLLEDLI      480
*****:*****

sp|Q9Z329|ITPR2_MOUSE      FFFADVNTNNGQDVLDDVITKPNRERQKLMREQNI LAQVFGILKAPFKEKAGEGSMRLLED      540
sp|P29995|ITPR2_RAT        FFFADVNTNNGQDVLDDVITKPNRERQKLMREQNI LAQVFGILKAPFKEKAGEGSMRLLED      540
tr|A0A8I3PW88|A0A8I3PW88_CANLF FFFADVNTNNGQEVLDVVITKPNRERQKLMREQNI LAQVFGILKAPFKEKAGEGSMRLLED      540
sp|Q14571|ITPR2_HUMAN      FFFADVNTNNGQEVLDVVITKPNRERQKLMREQNI LAQVFGILKAPFKEKAGEGSMRLLED      540
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sp|Q9Z329|ITPR2_MOUSE      LGDQRYAPYKYVRLCYRVLRRHSQQDYRKNOEYIAKNFCVMQSQIGYDILAEDTITALLH      600
sp|P29995|ITPR2_RAT        LGDQRYAPYKYVRLCYRVLRRHSQQDYRKNOEYIAKNFCVMQSQIGYDILAEDTITALLH      600
tr|A0A8I3PW88|A0A8I3PW88_CANLF LGDQRYAPYKYMRLCYRVLRRHSQQDYRKNOEYIAKNFCVMQSQIGYDILAEDTITALLH      600
sp|Q14571|ITPR2_HUMAN      LGDQRYAPYKYMRLCYRVLRRHSQQDYRKNOEYIAKNFCVMQSQIGYDILAEDTITALLH      600
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sp|Q9Z329|ITPR2_MOUSE      SEQNEIRNLQEKLESTMSLVKQLSGQLAELKEQMTQQRKNKQRLGFLGSNTPHVNHMPP 2700
sp|P29995|ITPR2_RAT        SEQNEIRNLQEKLESTMSLVKQLSGQLAELKEQMTQQRKNKQRLGFLGSNTPHENHMP 2700
tr|A0A8I3PW88|A0A8I3PW88_CANLF  SEQNEIRNLQEKLESTMSLVKQLSGQLAELKEQMTQQRKNKQRLGFLGSNTPHVNHMPP 2700
sp|Q14571|ITPR2_HUMAN      SEQNEIRSLQEKLESTMSLVKQLSGQLAELKEQMTQQRKNKQRLGFLGSNTPHVNHMPP 2700
*****

sp|Q9Z329|ITPR2_MOUSE      H 2701
sp|P29995|ITPR2_RAT        H 2701
tr|A0A8I3PW88|A0A8I3PW88_CANLF H 2701
sp|Q14571|ITPR2_HUMAN      H 2701
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Alignment of the dog ITPR3 protein sequence (XP_038409684.1) to human (NP_002215.2), mouse (P70227) and rat (Q63269) ITPR3 with the Clustal Omega algorithm (CLUSTAL O(1.2.4)).

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sp|P70227|ITPR3_MOUSE      -----MNMSSSFLHIGDIV---SLYAEGSVNGFISTLGLVDDRCVVEPAAGDLNPP 49
sp|Q63269|ITPR3_RAT        -----MNMSSSFLHIGDIV---SLYAEGSVNGFISTLGLVDDRCVVEPAAGDLNPP 49
XP_038409684.1            MRTHRNGGRLSRGAWGHLGLHTSCRWLLCLPATWGAALGTRLVDDRCVVEPAAGDLNPP 60
NP_002215.2                -----MSEMSSFLHIGDIV---SLYAEGSVNGFISTLGLVDDRCVVEPAAGDLNPP 49
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sp|P70227|ITPR3_MOUSE      KKFRDCLFKVCPMNRYSASQKQYWKAKQTKQDKEKIADVLLQKLQHAAQMEQKQNDTENK 109
sp|Q63269|ITPR3_RAT        KKFRDCLFKVCPMNRYSASQKQYWKAKQTKQDKEKIADVLLQKLQHAAQMEQKQNDTENK 109
XP_038409684.1            KKFRDCLFKVCPMNRYSASQKQYWKAKQTKQDKEKIADVLLQKLQHAAQMEQKQNDTENK 120
NP_002215.2                KKFRDCLFKVCPMNRYSASQKQYWKAKQTKQDKEKIADVLLQKLQHAAQMEQKQNDTENK 109
                             *****

sp|P70227|ITPR3_MOUSE      KVHGDVVVKYGSVIQLLHMKS NKYLT VNKRLPALLEKNAMRVTL DATGNEG SWLFIQPFWK 169
sp|Q63269|ITPR3_RAT        KVHGDVVVKYGSVIQLLHMKS NKYLT VNKRLPALLEKNAMRVTL DATGNEG SWLFIQPFWK 169
XP_038409684.1            KVHGDVVVKYGSVIQLLHMKS NKYLT VNKRLPALLEKNAMRVTL DATGNEG SWLFIQPFWK 180
NP_002215.2                KVHGDVVVKYGSVIQLLHMKS NKYLT VNKRLPALLEKNAMRVTL DATGNEG SWLFIQPFWK 169
                             *****

sp|P70227|ITPR3_MOUSE      LRSNGDNVVVGDKVILNPNVNAQPLHASNYELSDNAGCKEVNSVNCNTSWKINLFMQFRD 229
sp|Q63269|ITPR3_RAT        LRSNGDNVVVGDKVILNPNVNAQPLHASNYELSDNAGCKEVNSVNCNTSWKINLFMQFRD 229
XP_038409684.1            LRSNGDNVVVGDKVILNPNVNAQPLHASNYELSDNAGCKEVNSVNCNTSWKINLFMQFRD 240
NP_002215.2                LRSNGDNVVVGDKVILNPNVNAQPLHASNYELSDNAGCKEVNSVNCNTSWKINLFMQFRD 229
                             *****

sp|P70227|ITPR3_MOUSE      HLEEV LKGGDVVRLFHAEQEKFLTCDEYRGKLVFLRTTLRQSATSATSSNALWEVEVVH 289
sp|Q63269|ITPR3_RAT        HLEEV LKGGDVVRLFHAEQEKFLTCDEYRGKLVFLRTTLRQSATSATSSNALWEVEVVH 289
XP_038409684.1            HLEEV LKGGDVVRLFHAEQEKFLTCDEYRGKLVFLRTTLRQSATSATSSNALWEVEVVH 300
NP_002215.2                HLEEV LKGGDVVRLFHAEQEKFLTCDEYRGKLVFLRTTLRQSATSATSSNALWEVEVVH 289
                             *****

sp|P70227|ITPR3_MOUSE      HDPCRGGAGHWNGLYRFKHLATGNYLAAEENPSYKGDVSDPKAAGLGAQGRTRGRNAGEK 349
sp|Q63269|ITPR3_RAT        HDPCRGGAGHWNGLYRFKHLATGNYLAAEENPSYKGDVSDPKAAGLGAQGRTRGRNAGEK 349
XP_038409684.1            HDPCRGGAGHWNGLYRFKHLATGNYLAAEENPSYKGDVCEPKAAGMGAQGRTRGRNAGEK 360
NP_002215.2                HDPCRGGAGHWNGLYRFKHLATGNYLAAEENPSYKGDASDPKAAGMGAQGRTRGRNAGEK 349
                             *****

sp|P70227|ITPR3_MOUSE      IKYRLVAVPHGNDIASL FELDPTTLQKTD SFVPRNSYVRLRHLCTNTWIQSTNAPIDVEE 409
sp|Q63269|ITPR3_RAT        IKYRLVAVPHGNDIASL FELDPTTLQKTD SFVPRNSYVRLRHLCTNTWIQSTNAPIDVEE 409
XP_038409684.1            IKYRLVAVPHGNDIASL FELDPTTLQKTD SFVPRNSYVRLRHLCTNTWIQSTNAPIDIEE 420
NP_002215.2                IKYCLVAVPHGNDIASL FELDPTTLQKTD SFVPRNSYVRLRHLCTNTWIQSTNAPIDIEE 409
                             *** *****

sp|P70227|ITPR3_MOUSE      ERPIRLMGLTCTPTKEDKEAFAIVSVPVSEIRDLDFANDASSMLASAVEKLN EGFISQNDR 469
sp|Q63269|ITPR3_RAT        ERPIRLMGLTCTPTKEDKEAFAIVSVPVSEIRDLDFANDASSMLASAVEKLN EGFISQNDR 469
XP_038409684.1            ERPIRLMGLTCTPTKEDKEAFAIVSVPVSEIRDLDFANDASSMLASAVEKLN EGFISQNDR 480
NP_002215.2                ERPIRLMGLTCTPTKEDKEAFAIVSVPVSEIRDLDFANDASSMLASAVEKLN EGFISQNDR 469
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sp|P70227|ITPR3_MOUSE      RFVIQLLEDLVFFVSDVPNNGQNVLDIMVTKPNRERQKLMREQNILKQIFGILKAPFRDK 529
sp|Q63269|ITPR3_RAT        RFVIQLLEDLVFFVSDVPNNGQNVLDIMVTKPNRERQKLMREQNILKQIFGILKAPFRDK 529
XP_038409684.1            RFVIQLLEDLVFFVSDVPNNGQNVLDIMVTKPNRERQKLMREQNILKQIFGILKAPFRDK 540
NP_002215.2                RFVIQLLEDLVFFVSDVPNNGQNVLDIMVTKPNRERQKLMREQNILKQVFGILKAPFREK 529
                             *****

sp|P70227|ITPR3_MOUSE      GGEGPLVRL EELS DQKNAPYQMFRLCYRVLRLHSQEDYRKNQEHI AKQFGMMQSQIGYDI 589
sp|Q63269|ITPR3_RAT        GGEGPLVRL EELS DQKNAPYQMFRLCYRVLRLHSQEDYRKNQEHI AKQFGMMQSQIGYDI 589
XP_038409684.1            GGEGPLVRL EELS DQKNAPYQHMFRLCYRVLRLHSQEDYRKNQEHI AKQFGMMQSQIGYDI 600
NP_002215.2                GGEGPLVRL EELS DQKNAPYQHMFRLCYRVLRLHSQEDYRKNQEHI AKQFGMMQSQIGYDI 589
                             *****

sp|P70227|ITPR3_MOUSE      LAEDTITALLHNNRKLLEKHITKTEVETFVSLVRKNREPRFLDYSLDLCVSNRIAIPTVQ 649
sp|Q63269|ITPR3_RAT        LAEDTITALLHNNRKLLEKHITKTEVETFVSLVRKNREPRFLDYSLDLCVSNRIAIPTVQ 649
XP_038409684.1            LAEDTITALLHNNRKLLEKHITKTEVETFVSLVRKNREPRFLDYSLDLCVSNHIAIAVTQ 660
NP_002215.2                LAEDTITALLHNNRKLLEKHITKTEVETFVSLVRKNREPRFLDYSLDLCVSNHIAIAPTVQ 649
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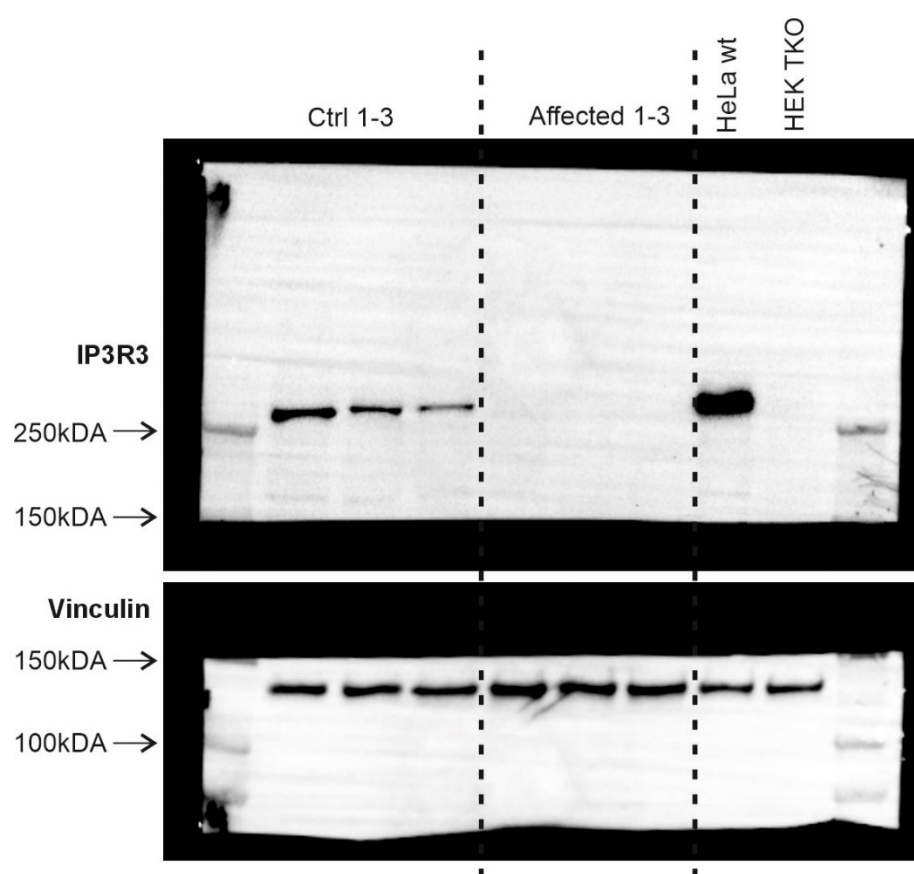
sp|P70227|ITPR3_MOUSE      ELICKCVLDPKNSDILIQTEL RPVKEMAQSHEYLSIEYSEEEVWL TWDRNNEHHEKSVR 709
sp|Q63269|ITPR3_RAT        ELICKCVLDPKNSDILIQTEL RPVKEMAQSHEYLSIEYSEEEVWL TWDRNNEHHEKSVR 709
XP_038409684.1            ELICKCVLDPKNSDILIQTEL RPVKEMAQSHEYLSIEYSEEEVWL TWDKNNEHHEKSVR 720
NP_002215.2                ELICKCVLDPKNSDILIRTEL RPVKEMAQSHEYLSIEYSEEEVWL TWTKNNEHHEKSVR 709
                             *****

sp|P70227|ITPR3_MOUSE      QLAQEARAGNAHDENVLSYRYQLKLFARMCLDRQYLAIDEISKQLGVLLFLCMADEML 769
sp|Q63269|ITPR3_RAT        QLAQEARAGNAHDENVLSYRYQLKLFARMCLDRQYLAIDEISKQLGVLLFLCMADEML 769
XP_038409684.1            QLAQEARAGNAHDENVLSYRYQLKLFARMCLDRQYLAIDEISQQLGVDLIFLCMADEML 780
NP_002215.2                QLAQEARAGNAHDENVLSYRYQLKLFARMCLDRQYLAIDEISQQLGVDLIFLCMADEML 769
                             *****

sp|P70227|ITPR3_MOUSE      PFDLRASFCHMLMLHVHVRDPQELVTPVKFARLWTEIPTAITIKDYDSNLNASRDDKKNK 829
sp|Q63269|ITPR3_RAT        PFDLRASFCHMLMLHVHVRDPQELVTPVKFARLWTEIPTAITIKDYDSNLNASRDDKKNK 829
XP_038409684.1            PFDLRASFCHMLMLHVHVRDPQELVTPVKFARLWTEIPTAITIKDYDSNLNASRDDKKNK 840
NP_002215.2                PFDLRASFCHMLMLHVHVRDPQELVTPVKFARLWTEIPTAITIKDYDSNLNASRDDKKNK 829
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1422

1423 **Supplementary figure 2. A Western blot of IP₃R3 protein.** The potential truncated IP₃R3 protein
 1424 with the nonsense mutation at p.Q1668, would correspond to a protein with 1667 amino acids if
 1425 translated. The theoretical size of the protein would be 170-185 kDA. In the Western blot, there are
 1426 no bands visible on the affected dogs' fibroblasts at the region corresponding to 150-300 kDA. Lanes:
 1427 1-3: healthy control dogs' fibroblasts, 4-6: affected dogs' fibroblasts, 7: HeLa wildtype, 8: triple-KO
 1428 (missing IP₃R1-3) HEK293.



1429

1430 **Supplementary figure 3. Ca²⁺ flux measurements with drug screening system FDSS/μCell.**

1431 The Ca²⁺ flux measurements were performed with FDSS/μCell functional drug screening system,
1432 using cell permeant Cal-520 AM fluorescent Ca²⁺ indicator.

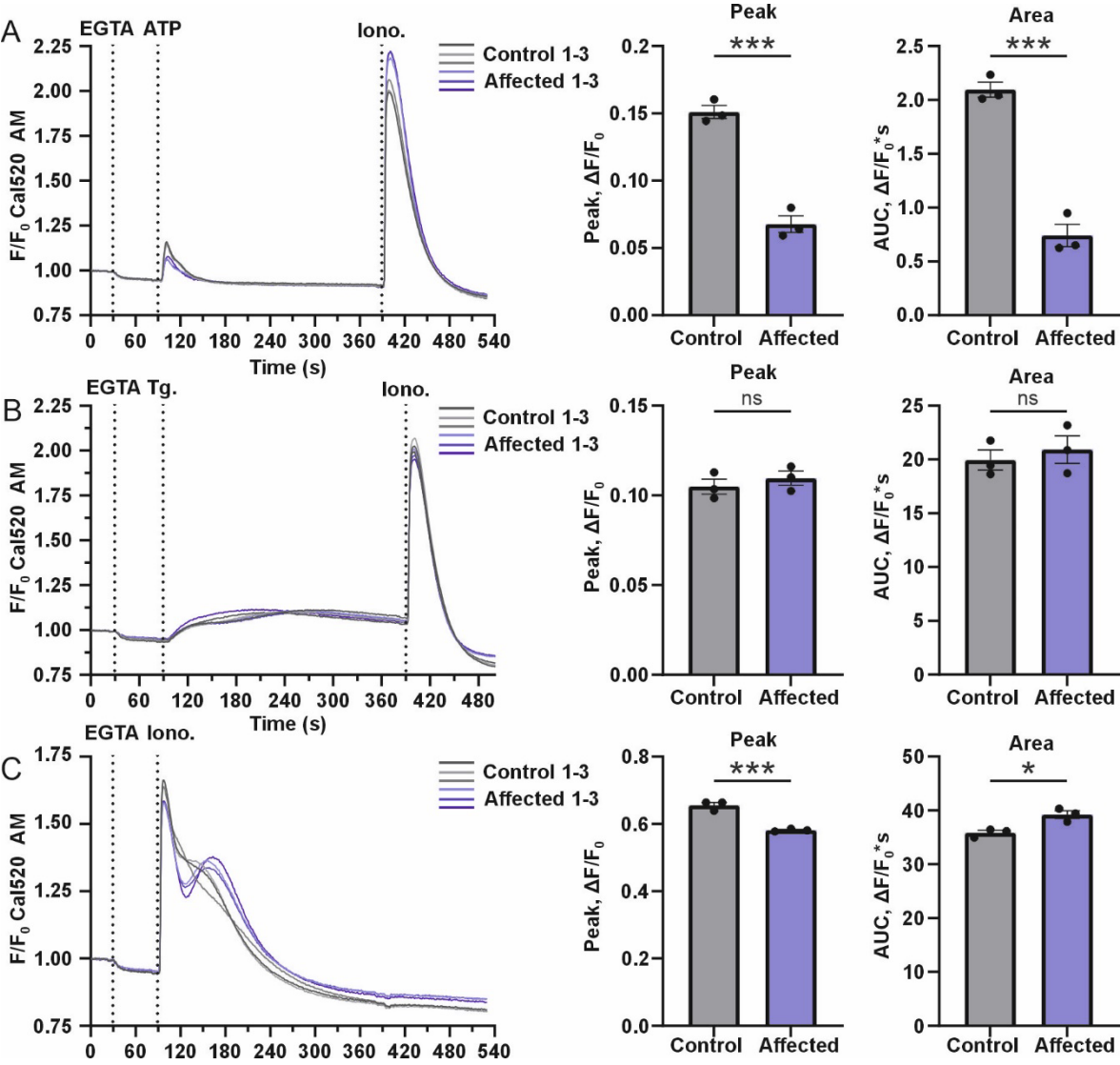
1433 **(A)** Fluorescent response curves of the affected and control cell lines after stimulation with GPCR
1434 agonist ATP. The cells were stimulated with 3 mM Ca²⁺ chelator EGTA, 100 μM ATP and 2.5 μM
1435 ionomycin. In response to GPCR agonist ATP, the affected fibroblast cells had significantly reduced
1436 peak amplitude and area under curve (AUC) (n = 3).

1437 **(C)** Fluorescent response curves of the affected and control cell lines after stimulation with 3 mM
1438 EGTA, 2 μM thapsigargin and 2.5 μM ionomycin. The loading of ER Ca²⁺ storages were measured
1439 with SERCA inhibitor thapsigargin. There were no significant differences in the peak amplitude and
1440 AUC between the affected and control cell lines (n = 3).

1441 **(D)** Fluorescent response curves of the affected and control cell lines after stimulation with 3 mM
1442 EGTA and 2.5 μM ionomycin. The total intracellular Ca²⁺ levels were measured with Ca²⁺ ionophore
1443 ionomycin. The affected cells had reduced peak amplitude, while the AUC was elevated in
1444 comparison to control cells (n = 3).

1445 Data are presented as mean ± SEM. Statistics are analyzed by Student's t-test (*p < 0.05, **p < 0.01,
1446 ***p < 0.001).

1447



Supplementary table 1. Electromyographic findings of the affected dogs.

Lancashire Heeler	Ulnar (normal >60 m/s)	Ishiadicus/peroneal (normal >80 m/s)	Fibrillation potentials	Positive sharp waves	CMAP amplitude	CMAP duration
Affected 1 (age 10y 5m)	58 m/s	62 m/s	+	+	↓	↑
Affected 2 (6y 10m)	44 m/s	76 m/s	+	+	↓	↑
Affected 3 (6y 5m & 7y 6m)	80 m/s	95 m/s	+	+	-	-

Supplementary tables 2-4.

Supplementary tables 2-4 are in .xlsx format (Excel sheets).

1457 **Supplementary table 5. Values used for parameter optimization and detection of shared ROH**
1458 **in PLINK.**

	values by analysis type	
parameter	parameter optimization	detection of shared ROH
--homozyg-window-snp	20	20
--homozyg-window-missing	1	1
--homozyg-window-het	0	1
--homozyg-window-threshold	0.05	0.05
--homozyg-snp	70	54
--homozyg-kb	1000 kb	1000 kb
--homozyg-gap	2000 kb (when unvaried)	200 kb
	20-1000 kb (when varied)	
--homozyg-density	200 kb/snp (when unvaried)	25 kb/snp
	10-125 kb/snp (when varied)	

1459

Supplementary table 6. Primers used in the study.

<u>Oligonucleotide</u>	<u>Sequence (5'->3')</u>	<u>Application</u>	<u>Citation</u>
canine ITPR3-Fw	CCTGTCCAAGTGAGCGAGAT	PCR & Sanger sequencing	This paper
canine ITPR3-Rv	CGTACTTGGCCTTCTTGAGC	PCR & Sanger sequencing	This paper
canine ITPR1-qPCR-Fw	GTCCCAAGAAACTCCTATGTCC	qRT-PCR	This paper
canine ITPR1-qPCR-Rv	CGGGTTTCTCTTCTTCCTTGT	qRT-PCR	This paper
canine ITPR2-qPCR-Fw	GAGAAGCGAGGGTGACAATATC	qRT-PCR	This paper
canine ITPR2-qPCR-Rv	CTCTATGTTGCTGGCATGTAGT	qRT-PCR	This paper
canine ITPR3-qPCR-Fw	CCATCGTGTCTGTTCTGTATC	qRT-PCR	This paper
canine ITPR3-qPCR-Rv	TGACAAACCTGCGATCATTCT	qRT-PCR	This paper
canine GAPDH-qPCR-Fw	CTGGGGCTCACTTGAAAGG	qRT-PCR	This paper
canine GAPDH-qPCR-Rv	CAAACATGGGGGCATCAG	qRT-PCR	This paper

References

1. Prole DL, Taylor CW. Structure and Function of IP(3) Receptors. Cold Spring Harb Perspect Biol. 2019;11(4).
2. Bootman MD, Bultynck G. Fundamentals of Cellular Calcium Signaling: A Primer. Cold Spring Harb Perspect Biol. 2020;12(1).
3. Kerkhofs M, Seitaj B, Ivanova H, Monaco G, Bultynck G, Parys JB. Pathophysiological consequences of isoform-specific IP(3) receptor mutations. Biochim Biophys Acta Mol Cell Res. 2018;1865(11 Pt B):1707-17.
4. Ronkko J, Rodriguez Y, Rasila T, Torregrosa-Munumer R, Pennonen J, Kvist J, et al. Human IP(3) receptor triple knockout stem cells remain pluripotent despite altered mitochondrial metabolism. Cell Calcium. 2023;114:102782.
5. Sugawara H, Kurosaki M, Takata M, Kurosaki T. Genetic evidence for involvement of type 1, type 2 and type 3 inositol 1,4,5-trisphosphate receptors in signal transduction through the B-cell antigen receptor. EMBO J. 1997;16(11):3078-88.
6. Yue L, Wang L, Du Y, Zhang W, Hamada K, Matsumoto Y, et al. Type 3 Inositol 1,4,5-Trisphosphate Receptor is a Crucial Regulator of Calcium Dynamics Mediated by Endoplasmic Reticulum in HEK Cells. Cells. 2020;9(2).
7. Alzayady KJ, Wang L, Chandrasekhar R, Wagner LE, 2nd, Van Petegem F, Yule DI. Defining the stoichiometry of inositol 1,4,5-trisphosphate binding required to initiate Ca²⁺ release. Sci Signal. 2016;9(422):ra35.
8. Ando H, Hirose M, Mikoshiba K. Aberrant IP(3) receptor activities revealed by comprehensive analysis of pathological mutations causing spinocerebellar ataxia 29. Proc Natl Acad Sci U S A. 2018;115(48):12259-64.
9. Wang YJ, Huang J, Liu W, Kou X, Tang H, Wang H, et al. IP3R-mediated Ca²⁺ signals govern hematopoietic and cardiac divergence of Flk1⁺ cells via the calcineurin-NFATc3-Etv2 pathway. J Mol Cell Biol. 2017;9(4):274-88.
10. Gerber S, Alzayady KJ, Burglen L, Bremond-Gignac D, Marchesin V, Roche O, et al. Recessive and Dominant De Novo ITPR1 Mutations Cause Gillespie Syndrome. Am J Hum Genet. 2016;98(5):971-80.
11. Huang L, Chardon JW, Carter MT, Friend KL, Dudding TE, Schwartzentruber J, et al. Missense mutations in ITPR1 cause autosomal dominant congenital nonprogressive spinocerebellar ataxia. Orphanet J Rare Dis. 2012;7:67.
12. McEntagart M, Williamson KA, Rainger JK, Wheeler A, Seawright A, De Baere E, et al. A Restricted Repertoire of De Novo Mutations in ITPR1 Cause Gillespie Syndrome with Evidence for Dominant-Negative Effect. Am J Hum Genet. 2016;98(5):981-92.
13. van de Leemput J, Chandran J, Knight MA, Holtzclaw LA, Scholz S, Cookson MR, et al. Deletion at ITPR1 underlies ataxia in mice and spinocerebellar ataxia 15 in humans. PLoS Genet. 2007;3(6):e108.
14. Klar J, Hisatsune C, Baig SM, Tariq M, Johansson AC, Rasool M, et al. Abolished InsP3R2 function inhibits sweat secretion in both humans and mice. J Clin Invest. 2014;124(11):4773-80.
15. Matsumoto M, Nakagawa T, Inoue T, Nagata E, Tanaka K, Takano H, et al. Ataxia and epileptic seizures in mice lacking type 1 inositol 1,4,5-trisphosphate receptor. Nature. 1996;379(6561):168-71.

16. Hisatsune C, Yasumatsu K, Takahashi-Iwanaga H, Ogawa N, Kuroda Y, Yoshida R, et al. Abnormal taste perception in mice lacking the type 3 inositol 1,4,5-trisphosphate receptor. *J Biol Chem*. 2007;282(51):37225-31.
17. Ronkko J, Molchanova S, Revah-Politi A, Pereira EM, Auranen M, Toppila J, et al. Dominant mutations in ITPR3 cause Charcot-Marie-Tooth disease. *Ann Clin Transl Neurol*. 2020;7(10):1962-72.
18. Schabhuttl M, Wieland T, Senderek J, Baets J, Timmerman V, De Jonghe P, et al. Whole-exome sequencing in patients with inherited neuropathies: outcome and challenges. *J Neurol*. 2014;261(5):970-82.
19. Lassuthova P, Safka Brozkova D, Krutova M, Neupauerova J, Haberlova J, Mazanec R, et al. Improving diagnosis of inherited peripheral neuropathies through gene panel analysis. *Orphanet J Rare Dis*. 2016;11(1):118.
20. Neumann J, Van Nieuwenhove E, Terry LE, Staels F, Knebel TR, Welkenhuyzen K, et al. Disrupted Ca(2+) homeostasis and immunodeficiency in patients with functional IP(3) receptor subtype 3 defects. *Cell Mol Immunol*. 2023;20(1):11-25.
21. Wysocki C, Moses A, Zhang H, Satterthwaite A, Ottens K, Xing C, et al. A human ITPR3 variant causes a dominant negative attenuation of calcium responses with immunodeficiency and growth delay confirmed in a mouse model. *Clinical Immunology*. 2023;250:109379.
22. Record CJ, Pipis M, Skorupinska M, Blake J, Poh R, Polke JM, et al. Whole genome sequencing increases the diagnostic rate in Charcot-Marie-Tooth disease. *Brain*. 2024.
23. Panigrahi B, Agarwal A, Garg D, Vishnu VY, Faruq M, Srivastava AK. Dominant Mutation in ITPR3 Gene Causing Charcot-Marie-Tooth Disease 1J in a Family from India. *Annals of Indian Academy of Neurology*. 2024.
24. Karczewski KJ, Francioli LC, Tiao G, Cummings BB, Alfoldi J, Wang Q, et al. The mutational constraint spectrum quantified from variation in 141,456 humans. *Nature*. 2020;581(7809):434-43.
25. Walker TL, Redding RW, Braund KG. Motor nerve conduction velocity and latency in the dog. *Am J Vet Res*. 1979;40(10):1433-9.
26. Wu L, Chen J. Type 3 IP3 receptor: Its structure, functions, and related disease implications. *Channels (Austin)*. 2023;17(1):2267416.
27. Smith CE. Cellular and chemical events during enamel maturation. *Crit Rev Oral Biol Med*. 1998;9(2):128-61.
28. Hubbard MJ. Calcium transport across the dental enamel epithelium. *Crit Rev Oral Biol Med*. 2000;11(4):437-66.
29. Lacruz RS, Smith CE, Bringas P, Jr., Chen YB, Smith SM, Snead ML, et al. Identification of novel candidate genes involved in mineralization of dental enamel by genome-wide transcript profiling. *J Cell Physiol*. 2012;227(5):2264-75.
30. Lacruz RS, Habelitz S, Wright JT, Paine ML. Dental Enamel Formation and Implications for Oral Health and Disease. *Physiol Rev*. 2017;97(3):939-93.
31. Nurbaeva MK, Eckstein M, Concepcion AR, Smith CE, Srikanth S, Paine ML, et al. Dental enamel cells express functional SOCE channels. *Sci Rep*. 2015;5:15803.
32. Liou J, Fivaz M, Inoue T, Meyer T. Live-cell imaging reveals sequential oligomerization and local plasma membrane targeting of stromal interaction molecule 1 after Ca2+ store depletion. *Proc Natl Acad Sci U S A*. 2007;104(22):9301-6.
33. Silva-Rojas R, Laporte J, Bohm J. STIM1/ORAI1 Loss-of-Function and Gain-of-Function Mutations Inversely Impact on SOCE and Calcium Homeostasis and Cause Multi-Systemic Mirror Diseases. *Front Physiol*. 2020;11:604941.
34. Terry LE, Arige V, Neumann J, Wahl AM, Knebel TR, Chaffer JW, et al. Missense mutations in inositol 1,4,5-trisphosphate receptor type 3 result in leaky Ca(2+) channels and activation of store-operated Ca(2+) entry. *iScience*. 2022;25(12):105523.

35. Lu JP, Wang Y, Sliter DA, Pearce MM, Wojcikiewicz RJ. RNF170 protein, an endoplasmic reticulum membrane ubiquitin ligase, mediates inositol 1,4,5-trisphosphate receptor ubiquitination and degradation. *J Biol Chem*. 2011;286(27):24426-33.
36. Pearce MM, Wormer DB, Wilkens S, Wojcikiewicz RJ. An endoplasmic reticulum (ER) membrane complex composed of SPFH1 and SPFH2 mediates the ER-associated degradation of inositol 1,4,5-trisphosphate receptors. *J Biol Chem*. 2009;284(16):10433-45.
37. Wojcikiewicz RJ, Pearce MM, Sliter DA, Wang Y. When worlds collide: IP(3) receptors and the ERAD pathway. *Cell Calcium*. 2009;46(3):147-53.
38. Chang CC, Chow CC, Tellier LC, Vattikuti S, Purcell SM, Lee JJ. Second-generation PLINK: rising to the challenge of larger and richer datasets. *Gigascience*. 2015;4:7.
39. Meyermans R, Gorssen W, Buys N, Janssens S. How to study runs of homozygosity using PLINK? A guide for analyzing medium density SNP data in livestock and pet species. *BMC Genomics*. 2020;21(1):94.
40. Purfield DC, Berry DP, McParland S, Bradley DG. Runs of homozygosity and population history in cattle. *BMC Genet*. 2012;13:70.
41. Broeckx BJ, Hitte C, Coopman F, Verhoeven GE, De Keulenaer S, De Meester E, et al. Improved canine exome designs, featuring ncRNAs and increased coverage of protein coding genes. *Sci Rep*. 2015;5:12810.
42. Wang C, Wallerman O, Arendt ML, Sundstrom E, Karlsson A, Nordin J, et al. A novel canine reference genome resolves genomic architecture and uncovers transcript complexity. *Commun Biol*. 2021;4(1):185.
43. Poplin R, Ruano-Rubio V, DePristo MA, Fennell TJ, Carneiro MO, Van der Auwera GA, et al. Scaling accurate genetic variant discovery to tens of thousands of samples. *bioRxiv*. 2018.
44. Wang K, Li M, Hakonarson H. ANNOVAR: functional annotation of genetic variants from high-throughput sequencing data. *Nucleic Acids Res*. 2010;38(16):e164.
45. Gardner EJ, Lam VK, Harris DN, Chuang NT, Scott EC, Pittard WS, et al. The Mobile Element Locator Tool (MELT): population-scale mobile element discovery and biology. *Genome Res*. 2017;27(11):1916-29.
46. Chen X, Schulz-Trieglaff O, Shaw R, Barnes B, Schlesinger F, Kallberg M, et al. Manta: rapid detection of structural variants and indels for germline and cancer sequencing applications. *Bioinformatics*. 2016;32(8):1220-2.
47. Arumilli M, Layer RM, Hytonen MK, Lohi H. webGQT: A Shiny Server for Genotype Query Tools for Model-Based Variant Filtering. *Front Genet*. 2020;11:152.
48. Ostrander EA, Wang GD, Larson G, vonHoldt BM, Davis BW, Jagannathan V, et al. Dog10K: an international sequencing effort to advance studies of canine domestication, phenotypes and health. *Natl Sci Rev*. 2019;6(4):810-24.
49. Rose R, Golosova O, Sukhomlinov D, Tiunov A, Prosperi M. Flexible design of multiple metagenomics classification pipelines with UGENE. *Bioinformatics*. 2019;35(11):1963-5.
50. Golosova O, Henderson R, Vaskin Y, Gabrielian A, Grekhov G, Nagarajan V, et al. Unipro UGENE NGS pipelines and components for variant calling, RNA-seq and ChIP-seq data analyses. *PeerJ*. 2014;2:e644.
51. Okonechnikov K, Golosova O, Fursov M, team U. Unipro UGENE: a unified bioinformatics toolkit. *Bioinformatics*. 2012;28(8):1166-7.
52. Schmitz EA, Takahashi H, Karakas E. Structural basis for activation and gating of IP(3) receptors. *Nat Commun*. 2022;13(1):1408.