

# 1 BakRep – A searchable large-scale 2 web repository for bacterial genomes, 3 characterizations and metadata

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10 classification, multilocus sequence typing

## 12 Abstract

13 Bacteria are fascinating research objects in many disciplines for countless reasons, and whole-genome  
14 sequencing has become the paramount methodology to advance our microbiological understanding. Meanwhile,  
15 access to cost-effective sequencing platforms has accelerated bacterial whole-genome sequencing to  
16 unprecedented levels introducing new challenges in terms of data accessibility, computational demands,  
17 heterogeneity of analysis workflows, and thus, ultimately its scientific usability. To that end, *Blackwell et al.*  
18 released a uniformly processed set of 661,405 bacterial genome assemblies obtained from the European  
19 Nucleotide Archive as of November 2018. Building on these accomplishments, we conducted further genome-  
20 based analyses like taxonomic classification, MLST subtyping and annotation of all genomes. Here we present  
21 BakRep, a searchable large-scale web repository of these genomes enriched with consistent genome  
22 characterizations and original metadata. The platform provides a flexible search engine combining taxonomic,  
23 genomic and metadata information, as well as interactive elements to visualize genomic features. Furthermore,  
24 all results can be downloaded for offline analyses via an accompanying command line tool. The web repository  
25 is accessible via <https://bakrep.computational.bio>.

## 28 Introduction

29 Bacteria represent a significant portion of Earth's biodiversity, showcasing an astounding variety of habitats. For  
30 the past three decades, bacterial whole-genome sequencing (WGS) has provided deep insights into the vast  
31 diversity of populations and ecosystems' complexity, just as into the organization and plasticity of single  
32 genomes - both fundamental for our perception of microbial life. In particular, WGS of bacterial pathogens has  
33 tremendously propelled our understanding of drug resistances, virulence factors, and host interactions and has  
34 become invaluable for medical microbiology. But simultaneously, the exploration and analysis of less-studied  
35 species continuously expands our knowledge of the broad and hard-to-comprehend diversity within the bacterial  
36 domain of life. However, the rapid and accelerating generation of WGS data demands substantial storage and  
37 analysis capacities. To securely store the raw DNA sequencing data, public databases like the Sequence Read  
38 Archive (SRA), the DNA Data Bank of Japan (DDBJ), or the European Nucleotide Archive (ENA) are primarily  
39 considered [1]. Consequently, these data repositories are in a constant state of growth. For example, at the time  
40 of writing, more than 4.6 billion sequences are stored in the ENA  
41 (<https://www.ebi.ac.uk/ena/browser/about/statistics>), and the latest GenBank release (v257.0) contains 2.6 billion  
42 WGS records (<https://ncbiinsights.ncbi.nlm.nih.gov/2023/08/21/genbank-release-257/>). Along with these rapidly  
43 growing data collections, several challenges arise with regard to the **FAIR** principles [2]. **Findability**: to conduct  
44 comparative analyses targeting particular sublineages or MLST types, sequenced samples often need to be  
45 processed prior to genome-based screening and filtering steps. **Accessibility**: the sheer amount of raw data  
46 needs to be handled and properly processed for analysis which poses a serious barrier for many researchers  
47 lacking necessary IT infrastructure and bioinformatics skills. **Interoperability**: common data formats,  
48 vocabularies and ontologies are crucial to facilitate data integration across different platforms. **Reproducibility**:  
49 large parts of this data are processed over and over again introducing adverse variability regarding used  
50 analysis tools, parameters and databases. Furthermore, user-provided metadata may be prone to inaccuracies  
51 and incompleteness complicating reproducibility and subsequent processing [3], [4]. In conclusion, this situation  
52 leads to inflated bioinformatic workloads, increasing analysis costs regarding computational resources and  
53 valuable staff time. The analyses of genomes of varying quality, being assembled and annotated using different  
54 algorithms and thus ultimately putting the usability of this valuable data at stake [5], [6]. In contrast to the large  
55 raw data repositories, dedicated initiatives, e.g., Enterobase [7] conduct consistent data processing procedures  
56 comprising targeted and streamlined genome characterizations. However, these platforms typically focus on  
57 distinct taxa and thus, are of limited general usability. An essential step addressing these challenges was made  
58 by a previous study by *Blackwell et al.* following a uniform approach to assemble and characterize all bacterial  
59 paired-end WGS datasets retrieved from the ENA as of November 2018 [8]. As a result, 661,405 consistently  
60 assembled genomes were made publicly available facilitating the broader access and utilization of this data for  
61 the research community. This study accomplished the systematic and standardized processing of this massive  
62 dataset, and thus fostered the usability of this genomic data. However, access to these genomes remains  
63 limited, since all genome Fasta files are provided as one comprehensive 751 GB single-file archive, thus posing  
64 a significant barrier in terms of findability and accessibility for further analyses. Even though assembled  
65 genomes are pre-indexed using various search algorithms, it remains challenging for users without sufficient

66 bioinformatics knowledge or command-line skills to find and extract genomes of interest. Hence, to fully exploit  
67 the huge potential of this highly valuable dataset, researchers would benefit from a user-friendly platform  
68 providing streamlined access to this huge amount of data via flexible search capabilities integrating the various  
69 information layers, like genome characterizations, taxonomic classifications and subtypings, annotated genomic  
70 features, and last but not least metadata. Building on these uniformly assembled bacterial genomes, here, we  
71 present BakRep, a large-scale comprehensive web repository specifically addressing these challenges. All  
72 661,405 genomes were consistently quality controlled, taxonomically classified, multilocus sequence typed, and  
73 annotated. In line with the FAIR principles, all information is findable and accessible via an interactive website  
74 providing researchers with a versatile search engine integrating genomic and taxonomic information, annotated  
75 features, and original metadata. Batch downloads of search results can be conducted via an accompanying  
76 command line tool. BakRep is publicly available at <https://bakrep.computational.bio>

## 77 **Methods / Implementation**

### 78 **Raw data processing**

79 We retrieved 661,405 assemblies and associated metadata published by *Blackwell et al.* from  
80 <http://ftp.ebi.ac.uk/pub/databases/ENA2018-bacteria-661k/>. For taxonomic classification, the GTDB-Tk (v2.2.6)  
81 classify workflow [9] based on the Genome Taxonomy Database (GTDB) release R207 [10] was used, with the  
82 '--mash\_db' argument set for enabling ANI screening. Contamination and completeness of the assemblies were  
83 estimated with CheckM2 (v1.0.1) [11]. Basic statistics of the raw assemblies were collected using assembly-scan  
84 (<https://github.com/rpetit3/assembly-scan>). Determination of multilocus sequence types (MLST) was conducted  
85 using mlst (v2.23.0) (<https://github.com/tseemann/mlst>) utilizing the PubMLST database. Furthermore,  
86 assemblies were annotated with Bakta (v1.7.0) using the 'full' database version to use all features and the  
87 '--keep-contig-headers' flag to preserve the original contig headers of the raw assemblies [12]. Results were  
88 stored as JSON files via custom Python scripts. All analyses were implemented as part of a Nextflow [13]  
89 workflow executed in the de.NBI consortiums' cloud computing infrastructure ([https://github.com/ag-](https://github.com/ag-computational-bio/bakrep)  
90 [computational-bio/bakrep](https://github.com/ag-computational-bio/bakrep)). The metacoder package (v0.3.6) was used for graphic summaries of the taxonomic  
91 abundances [14].

### 92 **Implementation of the web repository**

93 The BakRep web repository is implemented as an HTTP-based API, based on Vert.x, offering public endpoints  
94 for search and data access [15]. Elasticsearch is utilized for implementing the search functionality [16]. All  
95 genomic data is stored as compressed plain text files in a S3-compatible storage. We provide a publicly  
96 available website that retrieves data via the API and visualizes it. The website's graphical user interface is  
97 implemented as a single-page application in Vue.js 3 (<https://vuejs.org>). The services are deployed on a scalable  
98 Kubernetes cluster, which is currently hosted and run within the cloud computing infrastructure of the de.NBI  
99 consortium. An additional command line tool for automated large-scale downloads was implemented in Python  
100 (<https://github.com/ag-computational-bio/bakrep-cli>).

# 101 Results

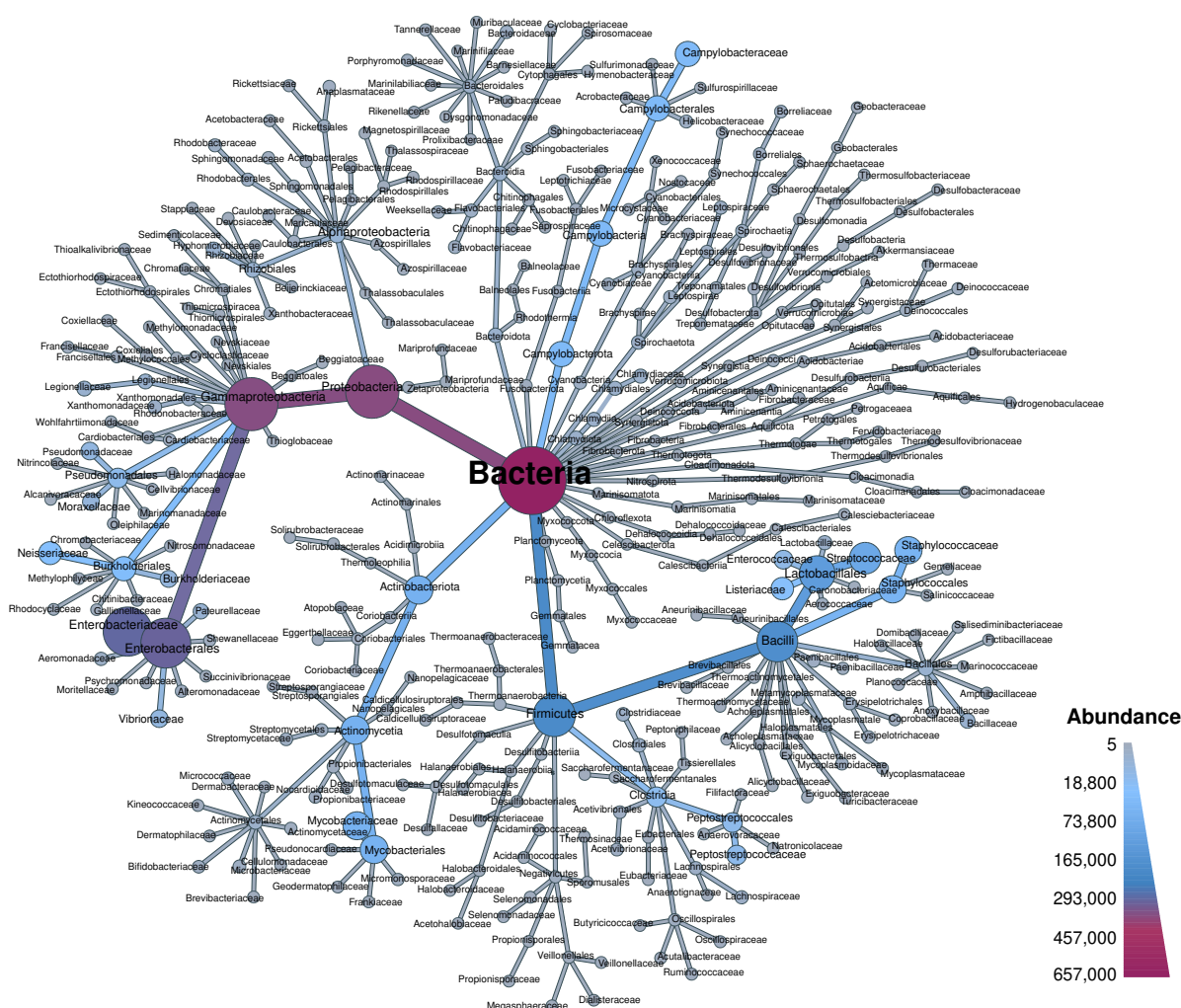
## 102 Expansion of consistent genome analyses

103 In this study, we built up on the 661,405 bacterial genome assemblies provided by *Blackwell et al.* who uniformly  
 104 assembled WGS raw data retrieved from the ENA archive as a November 2018 snapshot. We aimed to expand  
 105 the range of consistent per-genome characterizations and to provide these results as accessible and user-  
 106 friendly as possible. In this regard, all 661,405 assembled genomes (751 GB in total) were quality-checked and  
 107 basic assembly statistics were calculated. We then taxonomically classified all genomes using the robust GTDB  
 108 taxonomy, and where applicable, we further sequence-typed all genomes for which a species-specific  
 109 multilocus-sequence typing schema existed. Last but not least, we performed a robust annotation of all genomes  
 110 using Bakta taking advantage of its taxonomically untargeted full database version. From these 661,405 input  
 111 assemblies, 648,567 were successfully characterized. To streamline the technical accessibility of all results,  
 112 output files of all analysis tools were parsed, normalized, and serialized in JSON format, generating a total of  
 113 3,891,402 unique files. In addition, annotation results are also available in GenBank format, as well as nucleotide  
 114 and amino acid Fasta files for all annotated coding sequences. A total of 6.15 TB of genomic information was  
 115 generated and stored in a cloud-based S3 storage which is publicly available via an interactive web repository at  
 116 <https://bakrep.computational.bio>.

## 117 Diversity and bias across the various taxonomic ranks

118 Given the vast size of public databases, they naturally encompass a variety of species. Nevertheless, certain  
 119 species receive more frequent attention due to their clinical relevance, ease of cultivation, or long-standing  
 120 usage as model organisms. This inherent bias contributes to taxonomic imbalances in such data repositories.  
 121 *Blackwell et al.* comprehensively demonstrated an intrinsic taxonomic bias at both the genus and species level  
 122 [8]. However, we would like to address one more aspect: to what extent is there either bias or diversity at higher  
 123 taxonomic ranks? Therefore, we comprehensively explored the distribution across all taxonomic ranks, utilizing a  
 124 robust, purely genome-based, and thus objective taxonomic classification. We used GTDB-Tk, a widely utilized  
 125 tool in the community, that delineates prokaryotic taxa based on systematic criteria and phylogenetic  
 126 relationships using domain-specific marker genes in combination with mutual ANI-based genome distances. At  
 127 the species level, and in line with former results, our analysis revealed that the 24 most prevalent species  
 128 constitute 90 % of all genomes. The most abundant species were: *Salmonella enterica* (27.10 %), *Escherichia*  
 129 *coli* (13.52 %), *Streptococcus pneumoniae* (7.80 %), *Mycobacterium tuberculosis* (7.43 %), and *Staphylococcus*  
 130 *aureus* (7.28 %). At the genus level, the most prevalent genera were: *Salmonella* (27.99 %), *Escherichia*  
 131 (13.82 %), *Streptococcus* (12.89 %), *Mycobacterium* (8.6 %), and *Staphylococcus* (7.92 %). However, despite  
 132 these over-represented species and genera, the genomes contained in this repository exhibit a notable degree  
 133 of diversity at higher taxonomic ranks, comprising 66 distinct phyla divided into 132 classes, 345 orders, 722  
 134 families, 2,466 genera, and 8,207 species. In comparison, the genome sequence-based GTDB database counts  
 135 175 phyla, divided into 538 classes, 1,840 orders, 4,870 families, 23,112 genera, and 107,235 species  
 136 (<https://gtdb.ecogenomic.org/>) and the literature-based Bacterial Diversity Metadatabase (BacDive) lists 42 phyla

137 divided into 106 classes, 255 orders, 648 families, 3,801 genera and 21,203 species  
138 (<https://bacdiv.dsmz.de/dashboard>). Thus, this repository covers 37 % and 157 % of phyla, 24 % and 124 % of  
139 classes, 18 % and 135 % of orders, 14 % and 111 % of families, 10 % and 64 % of genera and 7 % and 38 % of  
140 species available in the genome-based GTDB and described in the literature-based BacDive databases,  
141 respectively. To illustrate both the diversity and bias of this repository, a taxonomic tree weighted by aggregated  
142 genome counts along all ranks was created (Fig. 1). For better visualization, taxa were clipped and aggregated  
143 at the family level. A more detailed version including all ranks is available in the supplemental data (Suppl. Fig  
144 1). Notably, 1,634 assemblies (0.25 %) could not be assigned to any species epithet, of which 122 (0.02 %)   
145 could not be assigned to a genus. A closer examination of the unclassified genomes revealed that those lacking  
146 a genus assignment exhibit an average estimated completeness of only 46.50 %. Genomes lacking a species  
147 epithet classification exhibited a higher average completeness of 67.02 %, albeit with increased variability  
148 (Suppl. Fig 2).



150 **Figure 1:** Overview of the taxonomic composition at the family level. Nodes and branches are colored and sized by  
151 aggregated genome counts at each taxonomic rank. The figure was created using the Metacoder package.

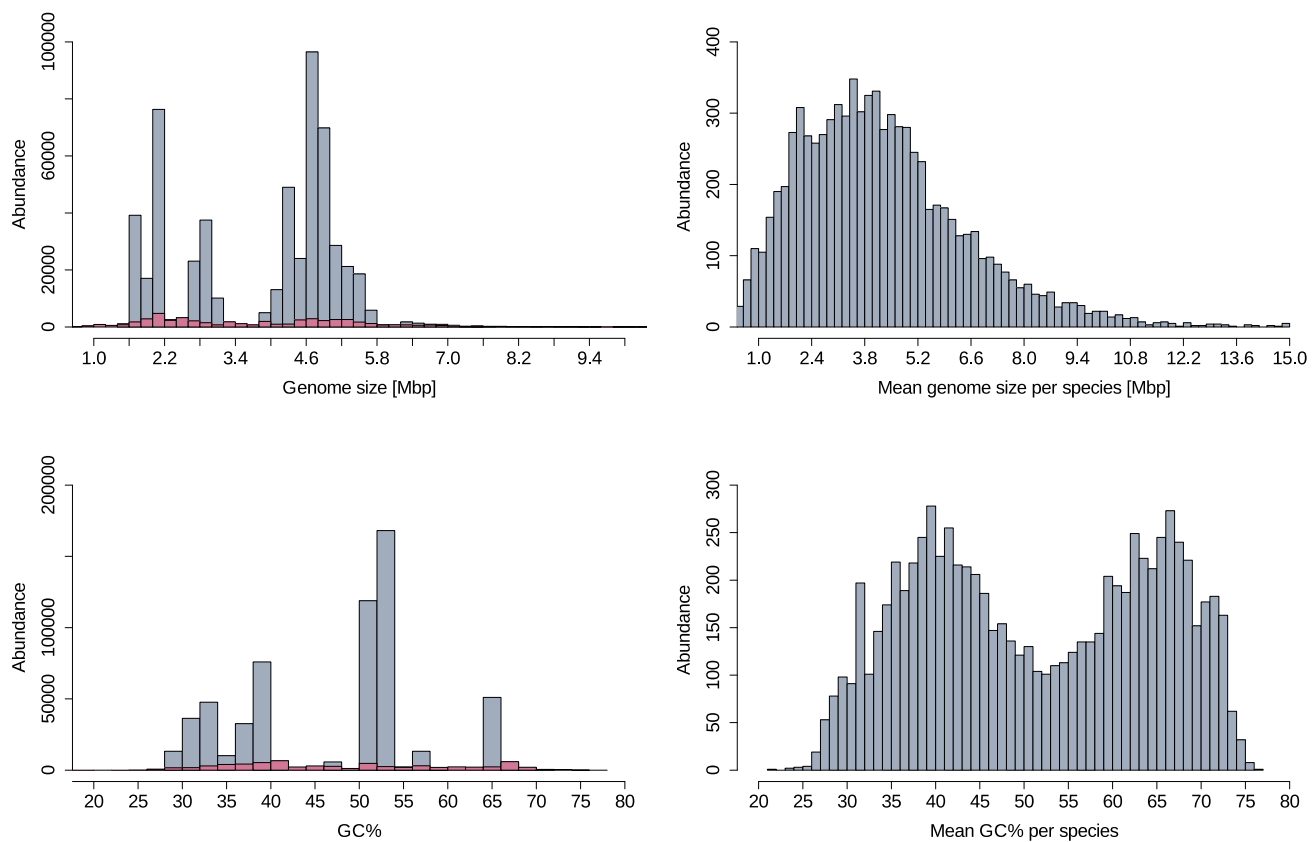


152 Various comparative analyses begin with the selection of suitable genomes from public repositories. Here, the  
 153 reliability of pre-assigned taxa is of utmost importance, immediately impacting the outcome of comparative  
 154 studies. Unfortunately, user-provided taxonomic information stored as metadata in raw sequencing data archives  
 155 is known to be error-prone and often does not correspond to genome-based taxonomic classifications. To  
 156 address this issue, we compared the scientific taxonomic names associated with the raw data in the ENA with  
 157 the genome-based taxonomic classifications conducted with GTDB-Tk. For 45,275 (6.98 %) genomes, we  
 158 observed discrepancies at the species epithet. Variations at the genus level occurred in 25,913 (4.0 %)   
 159 genomes. In 21,349 (3.29 %) cases, both the genus and species epithets differed. On further review, a  
 160 substantial portion of these discrepancies (54.96 %) is attributed to the genus *Shigella*, which was consistently  
 161 classified as *Escherichia*. Frequent inconsistencies were also evident for *Mycobacteroides abscessus*,  
 162 designated as *Mycobacterium abscessus* in 2,675 cases (10.32 %), and *Burkholderia pseudomallei* classified as  
 163 *Burkholderia mallei* in 1,763 cases (6.80 %). Among the 2,774 (10.71 %) species discrepancies within the  
 164 *Salmonella* genus, variations arose from assigning distinct subspecies, designated as full species names by  
 165 GTDB-Tk. Considering these examples, 7,106 (33.28 %) cases remained for which neither the genus nor the  
 166 species epithets matched (Suppl. Tab. 1).

## 167 Distribution of genome-based key metrics

168 In the NGS era, a multitude of sequencing platforms, as well as constantly evolving bioinformatics methods and  
 169 implementations, contribute to a variety of assembly approaches. To quickly assess biological key features and  
 170 the technical quality of assembled genomes, several metrics have evolved as gold standard indicators. For  
 171 instance, the mere size of a genome alone can provide important information, for instance regarding its  
 172 completeness. Also, the GC content is widely used as a rough proxy for the nucleotide composition of a genome,  
 173 that is typically found in a narrow range specific to a particular bacterial species. We used the available  
 174 information stored in BakRep to get an overview of the distribution of some of the most important and widely  
 175 used metrics for this repository. To better understand the extent of variation within the bacterial diversity, we  
 176 summarized the overall distribution of genome size and GC content. The total genome size ranges from a  
 177 minimum of 100,943 base pairs (bp) to a maximum of 20,285,777 bp with a mean value of 3,901,303 bp and a  
 178 median of 4,379,349 bp. To account for the observed taxonomic biases, we excluded the 24 most abundant  
 179 species accounting for 90 % of all genomes. For this taxonomically clipped set of genomes, the maximum and  
 180 minimum genome sizes remain unchanged, while the mean genome size increased to 3,962,704 bp, and the  
 181 median decreased to 3,853,294 bp. To further mitigate the influence of over-represented regions in the genome  
 182 size distribution, primarily attributed to the *Enterobacteriaceae* in the range of 4.5 - 5.5 Mbp, the  
 183 *Mycobacteriaceae* in the range of 1.5 - 2.0 Mbp, and the *Vibrionaceae* and *Neisseriaceae* in the range of  
 184 2.5 - 3.5 Mbp, we calculated the mean genome size per species. In contrast, this reveals a notably  
 185 homogeneous distribution with a peak at approximately 3.8 Mbp, followed by a rapid decline extending to a  
 186 maximum of 15 Mbp (Fig. 2). The GC content of all genomes ranges from a minimum of 23.6 % to a maximum of  
 187 76.5 % with a mean value of 47.2 % and a median of 50.7 %. Distinct peaks are observed at approximately  
 188 40 %, within the 50 - 55 % range, and at 65 %, mostly attributed to the 24 overrepresented species. The GC

173 content was likewise normalized based on species, resulting in a bimodal distribution that peaks at 40 % and  
174 between 60 - 70 % (Fig. 2).



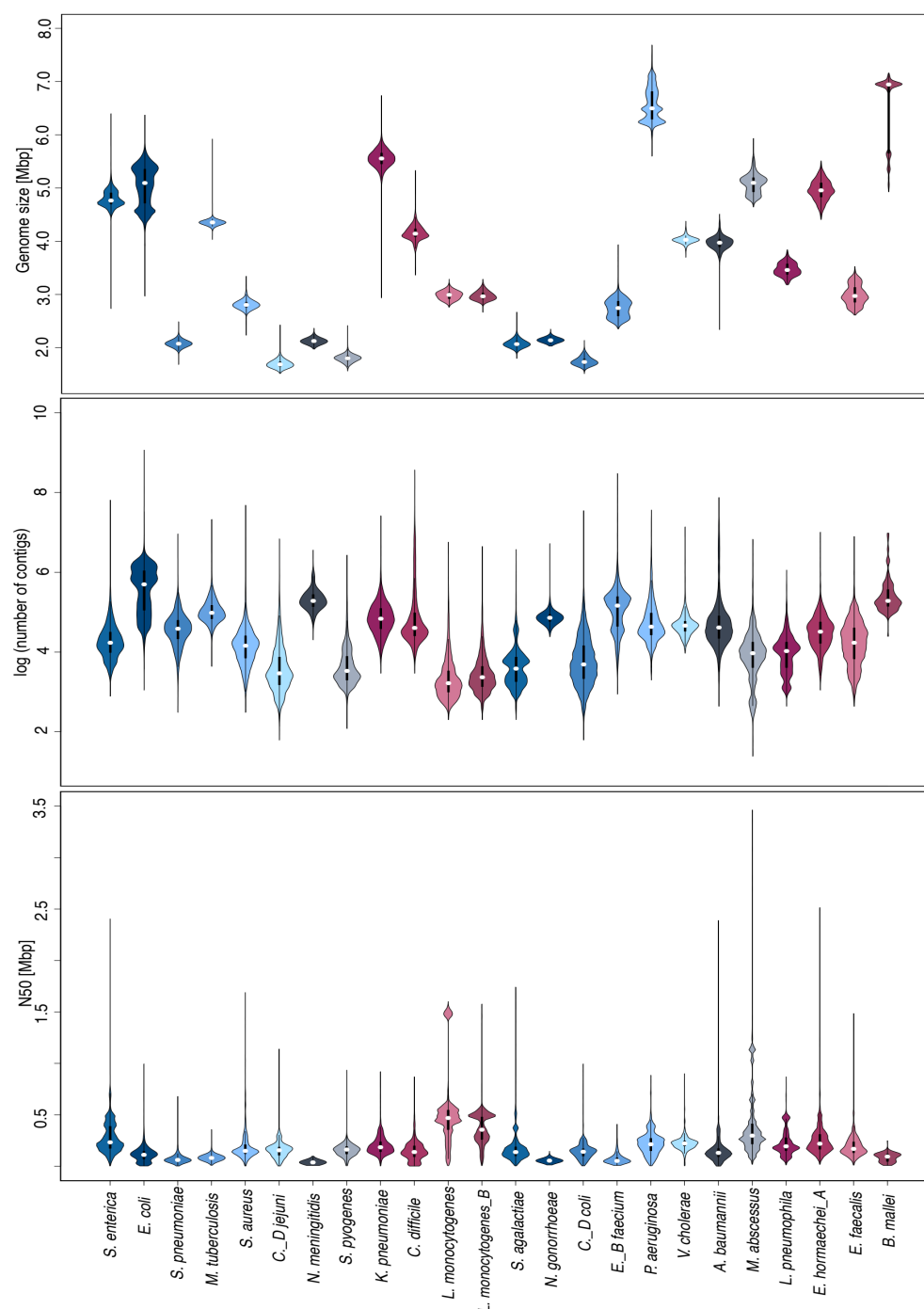
175 **Figure 2:** Distribution of genomic metrics in the repository. The genome size (top left) and the GC content (bottom left) are  
176 displayed for all genomes. In comparison, the mean genome size (top right) and the mean GC content (bottom right) per  
177 species are shown. The magenta-highlighted region illustrates the distribution excluding the 24 most abundant species.  
178

179 In addition to genome size and GC content, further metrics evolved to quickly assess the technical quality of a  
180 sequenced and assembled genome, e.g. the number of contigs and the well-known N50 metric. However, actual  
181 values for these metrics can vary widely not only between sequencing platforms and assembly approaches, but  
182 also between species due to biological factors, like the existence and abundance of sequence repeats and  
183 mobile elements. Furthermore, due to the lack of common guidelines, it is often far from obvious which actual  
184 values are acceptable for a given metric. Hence, we leveraged the robust taxonomic classifications and vast size  
185 of this repository to aid with the provision of potential guidelines for acceptable value ranges of these key metrics  
186 per species. Hence, we examined the distributions of the aforementioned key metrics for each of the most  
187 prevalent species, including many of significant medical relevance. As anticipated, we observed substantially  
188 varying value ranges for these metrics across species (Fig. 4). Additionally, the distribution ranges within  
189 individual species also showed considerable variability. For instance, for *Klebsiella pneumoniae*, we observed  
190 notable downward deviations, with some isolates exhibiting a minimum genome size ranging from 2.8 to  
191 4.0 Mbp, while the mean is 5.5 Mbp. However, upon closer observation, most of these outliers were identified as

192 several isolates from the same study that utilized transposon-directed insertion-site sequencing, suggesting that  
 193 these samples were not whole-genome sequenced. Discrepancies also exist for *Burkholderia mallei* with  
 194 likewise noticeable downward deviations for which no clear explanation could be found within the metadata.  
 195 Despite some outliers, core ranges of these key metric values might help to establish guidelines for quality  
 196 assessment.

197 The demonstrated varying ranges in genome sizes in the preceding section is an outcome of different habitats  
 198 and evolutionary mechanisms constantly introducing and removing genes. Due to the intricate and diverse set of  
 199 ecosystems, bacterial genomes exhibit significant variability in size and complexity, encompassing a fluid  
 200 continuum between compact genomes and those with larger and more elaborate structures. As a rule of thumb,  
 201 it is accepted as common knowledge that bacterial gene lengths average approximately 1 kbp per gene. To  
 202 assess this assumption, we juxtaposed the mean genome sizes with the mean number of genes per species. A  
 203 regression analysis revealed a slope of approximately 915 genes per 1 Mbp, resulting in a mean gene length of  
 204 1,093 bp, roughly validating but specifying this assumption with a deviation of 9.3 % and a determination  
 205 coefficient ( $R^2$ ) of 0.98 confirming the postulation of a linear relationship between genome size and the number  
 206 of coding genes. Besides, non-coding RNA features also play pivotal roles in bacterial genomes and cellular  
 207 processes, like for example, non-coding RNAs (ncRNAs), recognized for their regulatory functions, as well as  
 208 transfer and ribosomal RNAs (tRNAs/rRNAs) as essential components of the protein synthesis machinery.  
 209 Hence, we likewise compared numbers of annotated non-coding RNA features to the mean genome size per  
 210 species. Here, tRNAs showed a linear correlation however with significant variability ( $R^2=0.56$ ). In contrast, for  
 211 ncRNAs ( $R^2=0.35$ ), and rRNAs ( $R^2=0.15$ ), no clear linear trend could be observed (Suppl. Fig. 3).





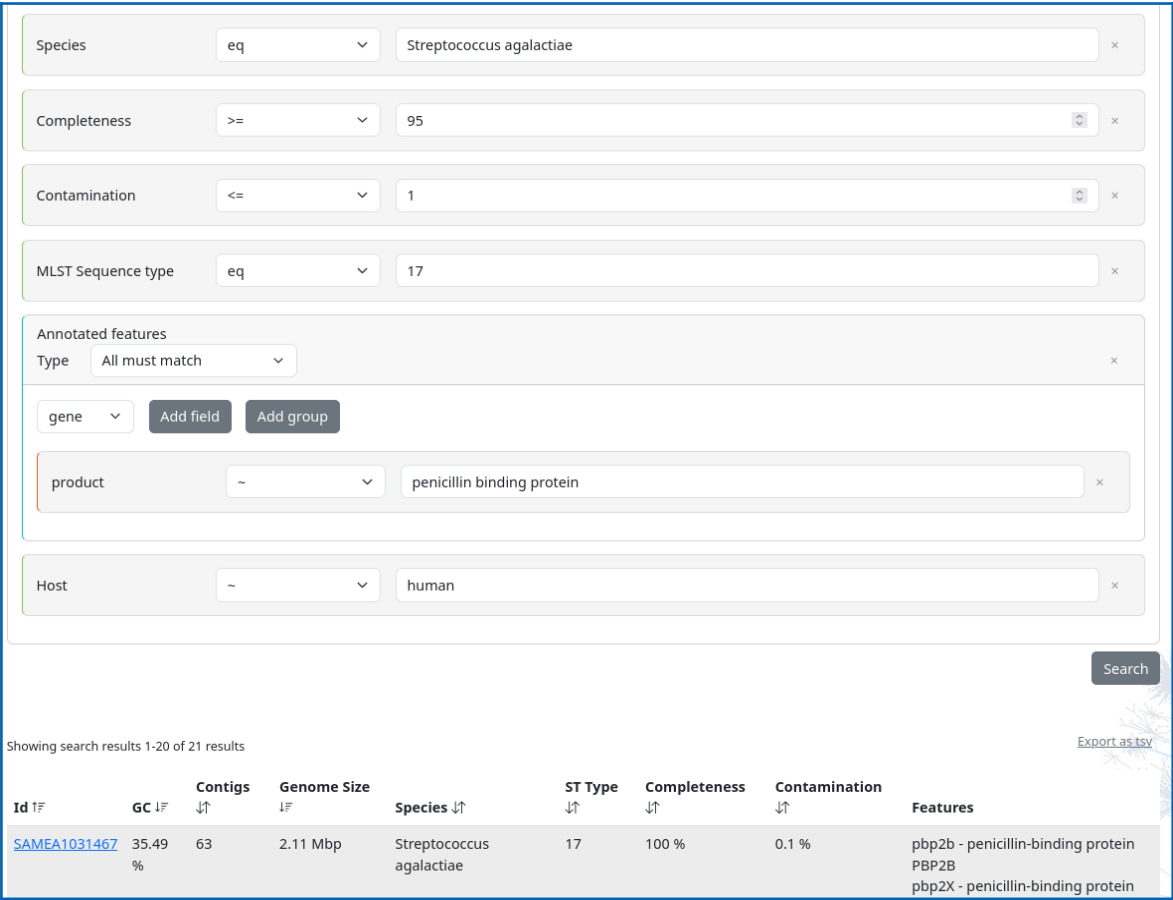
213 **Figure 4:** Distribution of genome assembly metrics for the 24 most abundant species. The genome size (top), the number of  
214 contigs (middle), and the N50 values (bottom) are displayed. White points indicate medians; bold black bars represent  
215 interquartile ranges; thin black lines represent outliers. Genomes were filtered for 95 % completeness and less than 1 %  
216 contamination.

## 217 **Interactive and command line access via a searchable web repository**

218 A major part of research would be constrained or rendered infeasible without accessible data. While a high level  
 219 of standardization is crucial, it is equally essential to consider the ease of data findability and accessibility. For  
 220 example, in outbreak analyses, for which the presence of specific antibiotic resistance genes is pivotal, it is  
 221 essential to systematically search for genomes of a particular species characterized by distinct features such as  
 222 multilocus sequence-type or virulence factors. To ensure the accessibility of our results, all data was stored in a  
 223 public S3 bucket. To furthermore ensure the findability of genomes of interest, we developed and provide an  
 224 interactive web page that is publicly available at <https://bakrep.computational.bio>. It offers diverse search and  
 225 filter options, allowing and streamlining the compilation of customized cohorts. To obtain an initial  
 226 comprehensive overview, all available genomes can be browsed by GC content, number of contigs, genome  
 227 size, as well as estimated completeness and contamination levels. To conduct comprehensive and detailed  
 228 large-scale searches, BakRep offers an advanced search engine that enables robust scalable queries flexibly  
 229 combining various information like genome size, GC content, number of contigs, sequence type, different  
 230 taxonomic ranks and annotated gene symbols or protein product descriptions. Furthermore, and in addition to  
 231 genome analysis-based information, users also have access to quality-controlled metadata associated with each  
 232 dataset upon initial raw data submission to the ENA. So, the repository supports the filtering of genomes based  
 233 on various metadata, including isolation source and time, associated host species, and projekt affiliation,  
 234 enabling targeted searches by criteria such as country of origin, isolation period, or host organism. A more  
 235 detailed list including all possible search tags is available in the supplemental data (Suppl. Tab. 2). To name an  
 236 example, in one of our ongoing research projects, we utilized this search engine to identify all *Streptococcus*  
 237 *agalactiae* genomes that met specific quality criteria, were isolated from humans, belonged to sequence type 17,  
 238 and contained the penicillin-binding proteins *pbp1a*, *pbp1b*, *pbp2a* or *pbp2X* (Fig. 5). A summary of the particular  
 239 search results can be exported in TSV format. All individual genomes are displayed in human-readable formats  
 240 such as a summary table, a feature table, and an igv.js-based genome browser [17], and provide cross-links to  
 241 databases such as the GTDB, RefSeq [18] or UniProt [19]. Each analysis result can be accessed and  
 242 downloaded per genome via the website. To facilitate extensive analyses with the download of larger genome  
 243 cohorts, we offer access to the download backend through a dedicated command line tool accessible via  
 244 <https://github.com/ag-computational-bio/bakrep-cli>.

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248 **Figure 5:** Overview of the search function of the BakRep web repository. Advanced queries for genomes with specific  
249 characteristics such as species, completeness, contamination levels, sequence type, annotated features, or host species are  
250 possible. The search outcomes are presented in a concise summary table. Details for each dataset are provided on a  
251 separate page.

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## 265 Discussion

266 Given the current capabilities for rapid and cost-efficient sequencing approaches, vast amounts of bacterial  
 267 sequence data are generated daily. Submitting genomic data to public repositories has become a pivotal and  
 268 obligatory procedure to underpin research findings and guarantee unrestricted and open access to this valuable  
 269 data. Such databases consequently contain a wealth of bacterial WGS data, representing genetic reservoirs with  
 270 vast potential for various applications. Nevertheless, publicly accessible genomic data often exhibit  
 271 inconsistencies or insufficient processing, hindering accessibility for researchers. Challenges arise from diverse  
 272 assembly methods and variations in quality control, potentially introducing batch artifacts in large-scale analyses.  
 273 A critical stride in tackling these challenges was taken by *Blackwell et al.*, through the uniform processing of all  
 274 WGS data found in the ENA as of November 2018, which yielded 661,405 standardized assemblies. BakRep  
 275 follows up on this endeavor by adding results of additional analyses, like assembly metrics, taxonomic  
 276 classifications, MLST subtyping, genome annotations, and metadata of original submissions. Finally, it facilitates  
 277 streamlined access to this broad amount of information via an interactive website providing powerful search  
 278 capabilities.

279 The sequencing of bacterial genomes has become routine, significantly reshaping our understanding of the  
 280 bacterial world with information gleaned from tens of thousands of genomes. Nevertheless, this quantity exhibits  
 281 a notable skew toward specific phyla housing for example particular model organisms [20]. This taxonomic bias  
 282 of just a few species making up the majority of genomic data may be due to various factors, such as the over-  
 283 representation of well-researched and easily cultivable species, leading to gaps in the representation of the  
 284 lesser researched or uncultivable microbial diversity. Furthermore, many sequencing projects focus on certain  
 285 pathogens or organisms with global significance. For example, the GenomeTrakr network represents the  
 286 inaugural distributed collaboration of laboratories employing WGS for pathogen identification [21], or the “10,000  
 287 *Salmonella* genomes project”, which sequenced more than 10,000 *Salmonella* isolates [22]. This shows the  
 288 impact of funding and scientific emphasis on the diversity of sequences. In contrast to the approach employed by  
 289 *Blackwell et al.*, our study presents a more intricate portrayal of the taxonomic distribution through systematic  
 290 species assignment utilizing the GTDB. They already acknowledged that certain aspects of sequence diversity  
 291 within the assemblies might have been overlooked due to constraints inherent in the Kraken 2 database, which  
 292 they used for taxonomic assignment and abundance estimation [3]. In contrast, the taxonomic classification in  
 293 this study was conducted using the GTDB, employing a normalized genome-based classification derived from  
 294 phylogenetic trees. These trees were constructed using a concatenated protein phylogeny, serving as the  
 295 foundation for bacterial taxonomy. This approach conservatively eliminates polyphyletic groups and normalizes  
 296 taxonomic ranks based on the relative evolutionary divergence [23]. However, the GTDB currently enumerates  
 297 175 phyla, divided into 538 classes, 1,840 orders, 4,870 families, 23,112 genera, and 107,235 species. This  
 298 indicates that our dataset covers 37% of those phyla, 24% of classes, 18% of orders, 14% of families, 10% of  
 299 genera, and 7% of species, highlighting its limited scope and underscoring that it encompasses only a fraction of  
 300 the extensive bacterial diversity. There is a need to shift emphasis from a strong focus on known pathogens in

301 sequencing projects, towards underrepresented and unknown species. This approach is crucial for a more  
302 comprehensive understanding of the patterns within bacterial diversity.

303 Examining several assembly metrics provides valuable insights into bacterial genomes, aiding in understanding  
304 their genetic diversity, evolutionary relationships, functional roles, and taxonomic classification. The bias of  
305 overrepresented species in the repository is also evident regarding these metrics. Nevertheless, the absence of  
306 prominently discernible gaps in the distribution of the mean genome size per species instills confidence that this  
307 snapshot may nevertheless encapsulate a substantial portion of bacterial diversity. However, while bacteria can  
308 attain genome sizes of up to 16 Mbp [24], the upper ranges are only poorly represented here. A recent study  
309 mentions a connection between the distribution of genome sizes and ecosystem type or associations with hosts,  
310 and it also discusses the ongoing challenge of precisely defining the distribution of genome size beyond the  
311 confines of laboratory settings [25]. Another study postulated an indirect mechanism of natural selection whereby  
312 ancient adaptations have induced alterations in the bacterial genome, contributing to a bimodal distribution  
313 pattern of genomic GC, which we also observed here [25]. While the genome size of a species may show some  
314 variability, caution should be exercised when encountering pronounced outliers. Substantial deviations from the  
315 mean literature value may indicate potential issues with quality, possible contamination, or the sequencing of  
316 partial segments rather than the entire genome, as exemplified in *Klebsiella pneumoniae*. Knowledge gained  
317 from a comprehensive and standardized analysis of numerous bacterial genomes has the potential to contribute  
318 valuable insights, aiding in the formulation of robust guidelines specific to certain species. Empirical values  
319 derived from diverse biological samples might offer more reliable guidelines than solely relying on literature  
320 values established over the years only using a few type strains or reference genomes. By encompassing a  
321 broader array of datasets, our repository helps to generate such guidelines. This extensive collection allows for  
322 more robust analysis and comparisons. Consequently, researchers can develop more nuanced and reliable  
323 guidelines that better reflect the complexity of bacterial genomes.

324 Genome fragmentation is a prevalent issue associated with short-read sequencing technologies. This challenge  
325 stems from the generation of shorter DNA fragments during the sequencing process, leading to genome  
326 assemblies typically consisting of an increased number of contigs. A recent article mentioned that the quality of  
327 these genome sequences may suffice for most analyses but need to be more practical for comparative genomics  
328 [26]. Given the typical size of a bacterial genome, a genome with a high number of contigs would result in  
329 smaller contig sizes. Referring to the average gene size of 1,093 bp, which we have calculated here, smaller  
330 contigs may contain at most one complete gene, with fragmented genes may frequently appear at contig  
331 boundaries. Significant variability in the number of contigs, especially with numerous upward outliers, should  
332 therefore, be approached cautiously. Due to the fact that the underlying assemblies in this study are based  
333 solely on Illumina sequencing, to improve the dataset it would be beneficial to include other sequencing  
334 techniques.

335 The presence of taxonomic misclassified species in public repositories is of significant concern to researchers,  
 336 as it can introduce inaccuracies into various analyses, thus impacting the reliability of the findings. Furthermore,  
 337 classification errors can propagate over time as incorrectly labeled genomes are used as references to identify  
 338 novel sequences. Specific errors may stem from taxonomic naming inconsistencies or the frequent  
 339 reclassification of organisms prompted by new discoveries. In our study, this applies, *e.g.*, to the discrepancies  
 340 found with the genus *Shigella*, as *Shigella* species were reclassified as later heterotypic synonyms of  
 341 *Escherichia coli* in the GTDB [27]. The variations in the nomenclature of *Burkholderia* species can be similarly  
 342 explained, given that *Burkholderia mallei* can be characterized as a recently evolved, host-adapted clonal  
 343 lineage derived from *Burkholderia pseudomallei* [28], [29]. This may also explain the observable variations in the  
 344 genome size. During host adaptation, *B. mallei* experienced considerable genome reduction [28], [29]. Given  
 345 that *B. mallei* and *B. pseudomallei* share over 99 % genetic homology, taxonomic transitions between them can  
 346 be fluid [30]. As GTDB-Tk uses an operational average nucleotide identity-based approach relying on type  
 347 strains, only a few different genes will not lead to species differentiation. Unfortunately, we were initially unaware  
 348 of the extensiveness of taxonomic discrepancies in the dataset, and thus we decided to use species information  
 349 associated as metadata for our genome annotation processes. This will certainly be addressed in future versions  
 350 by using GTDB-Tk species classifications ensuring accurate and consistent species listings down to annotation  
 351 result files.

352 Adherence to the FAIR Principles - Findability, Accessibility, Interoperability, and Reusability - is crucial for  
 353 advancing genomic research. With our public web repository we ensure that genomes are easily findable with  
 354 persistent identifiers being accessible to a wide range of users across different platforms. By providing genome  
 355 annotations in common file formats such as GenBank, we foster compatibility with various bioinformatic tools for  
 356 targeted downstream analyses, thus reducing technical barriers, increasing efficiency and supporting the  
 357 reproducibility of research results. Furthermore, we provide streamlined access to the valuable raw assemblies  
 358 of *Blackwell et al.*, ensuring that these results can be used for further studies.

## 359 Conclusion / Outlook

360 The BakRep web repository provides a consistent and comprehensive characterization of one of the largest  
 361 collections of bacterial genomes comprising assembly metrics, robust taxonomic classifications, MLST  
 362 subtypings, genome annotations, and original metadata. Its implementation and underlying cloud infrastructure  
 363 facilitate scalability and allow for swift adjustments to extended analyses and expanding datasets. Our long-term  
 364 plan includes the addition of more genomes to our repository, aiming for the continuous expansion of this  
 365 standardized dataset. We envision BakRep as a high-quality open resource for microbial researchers worldwide.

366



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377 **Author contributions**

378 OS designed and supervised the study. LJ developed the website and the servers. LF conducted data analyses,  
379 interpreted the data, and wrote the manuscript. AG supervised the study and was responsible for funding. All  
380 authors critically checked and contributed to the final version of the manuscript.

382 **Conflicts of interest**

383 The authors declare that they have no conflicts of interest.

385 **Data availability**

386 The website can be accessed via [bakrep.computational.bio](https://bakrep.computational.bio). The workflow used for data analysis is available at  
387 [github.com/ag-computational-bio/bakrep](https://github.com/ag-computational-bio/bakrep). Original data was retrieved via [ftp.ebi.ac.uk/pub/databases/ENA2018-](ftp://ftp.ebi.ac.uk/pub/databases/ENA2018-bacteria-661k)  
388 [bacteria-661k](ftp://ftp.ebi.ac.uk/pub/databases/ENA2018-bacteria-661k). The accompanying command line tool is available at  
389 <https://github.com/ag-computational-bio/bakrep-cli>.

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## 401 References

- 402 [1] M. Blaxter *et al.*, 'Reminder to deposit DNA sequences', *Science*, vol. 352, no. 6287, pp. 780–780, May  
403 2016, doi: [10.1126/science.aaf7672](https://doi.org/10.1126/science.aaf7672).
- 404 [2] M. D. Wilkinson *et al.*, 'The FAIR Guiding Principles for scientific data management and stewardship', *Sci.*  
405 *Data*, vol. 3, no. 1, Art. no. 1, Mar. 2016, doi: [10.1038/sdata.2016.18](https://doi.org/10.1038/sdata.2016.18).
- 406 [3] H. Bagheri, A. J. Severin, and H. Rajan, 'Detecting and correcting misclassified sequences in the large-  
407 scale public databases', *Bioinformatics*, vol. 36, no. 18, pp. 4699–4705, Sep. 2020, doi:  
408 [10.1093/bioinformatics/btaa586](https://doi.org/10.1093/bioinformatics/btaa586).
- 409 [4] F. Keck, M. Couton, and F. Altermatt, 'Navigating the seven challenges of taxonomic reference databases  
410 in metabarcoding analyses', *Mol. Ecol. Resour.*, vol. 23, no. 4, pp. 742–755, 2023, doi: [10.1111/1755-](https://doi.org/10.1111/1755-0998.13746)  
411 [0998.13746](https://doi.org/10.1111/1755-0998.13746).
- 412 [5] 'Extensive Error in the Number of Genes Inferred from Draft Genome Assemblies | PLOS Computational  
413 Biology'. Accessed: May 22, 2024. [Online]. Available: [https://journals.plos.org/ploscompbiol/article?](https://journals.plos.org/ploscompbiol/article?id=10.1371/journal.pcbi.1003998)  
414 [id=10.1371/journal.pcbi.1003998](https://journals.plos.org/ploscompbiol/article?id=10.1371/journal.pcbi.1003998)
- 415 [6] S. L. Salzberg, 'Next-generation genome annotation: we still struggle to get it right', *Genome Biol.*, vol. 20,  
416 no. 1, p. 92, May 2019, doi: [10.1186/s13059-019-1715-2](https://doi.org/10.1186/s13059-019-1715-2).
- 417 [7] Z. Zhou *et al.*, 'The Enterobase user's guide, with case studies on Salmonella transmissions, Yersinia  
418 pestis phylogeny, and Escherichia core genomic diversity', *Genome Res.*, vol. 30, no. 1, pp. 138–152, Jan.  
419 2020, doi: [10.1101/gr.251678.119](https://doi.org/10.1101/gr.251678.119).
- 420 [8] G. A. Blackwell *et al.*, 'Exploring bacterial diversity via a curated and searchable snapshot of archived DNA  
421 sequences', *PLoS Biol.*, vol. 19, no. 11, p. e3001421, Nov. 2021, doi: [10.1371/journal.pbio.3001421](https://doi.org/10.1371/journal.pbio.3001421).
- 422 [9] P.-A. Chaumeil, A. J. Mussig, P. Hugenholtz, and D. H. Parks, 'GTDB-Tk v2: memory friendly classification  
423 with the genome taxonomy database', *Bioinformatics*, vol. 38, no. 23, pp. 5315–5316, Dec. 2022, doi:  
424 [10.1093/bioinformatics/btac672](https://doi.org/10.1093/bioinformatics/btac672).
- 425 [10] 'GTDB: an ongoing census of bacterial and archaeal diversity through a phylogenetically consistent, rank  
426 normalized and complete genome-based taxonomy | Nucleic Acids Research | Oxford Academic'.  
427 Accessed: Sep. 20, 2023. [Online]. Available: [https://academic.oup.com/nar/article/50/D1/D785/6370255?](https://academic.oup.com/nar/article/50/D1/D785/6370255?login=true)  
428 [login=true](https://academic.oup.com/nar/article/50/D1/D785/6370255?login=true)
- 429 [11] 'CheckM2: a rapid, scalable and accurate tool for assessing microbial genome quality using machine  
430 learning | Nature Methods'. Accessed: Sep. 20, 2023. [Online]. Available:  
431 <https://www.nature.com/articles/s41592-023-01940-w>
- 432 [12] O. Schwengers, L. Jelonek, M. A. Dieckmann, S. Beyvers, J. Blom, and A. Goesmann, 'Bakta: rapid and  
433 standardized annotation of bacterial genomes via alignment-free sequence identification', *Microb.*  
434 *Genomics*, vol. 7, no. 11, Nov. 2021, doi: [10.1099/mgen.0.000685](https://doi.org/10.1099/mgen.0.000685).
- 435 [13] P. Di Tommaso, M. Chatzou, E. W. Floden, P. P. Barja, E. Palumbo, and C. Notredame, 'Nextflow enables  
436 reproducible computational workflows', *Nat. Biotechnol.*, vol. 35, no. 4, pp. 316–319, Apr. 2017, doi:  
437 [10.1038/nbt.3820](https://doi.org/10.1038/nbt.3820).
- 438 [14] 'Metacoder: An R package for visualization and manipulation of community taxonomic diversity data |  
439 PLOS Computational Biology'. Accessed: Jan. 30, 2024. [Online]. Available:  
440 <https://journals.plos.org/ploscompbiol/article?id=10.1371/journal.pcbi.1005404>
- 441 [15] T. Parviainen, *Real-time Web Application Development using Vert.x 2.0*. Packt Publishing, 2013.
- 442 [16] C. Gormley and Z. Tong, *Elasticsearch: The Definitive Guide*, 1st ed. O'Reilly Media, Inc., 2015.
- 443 [17] J. T. Robinson, H. Thorvaldsdottir, D. Turner, and J. P. Mesirov, 'igv.js: an embeddable JavaScript  
444 implementation of the Integrative Genomics Viewer (IGV)', *Bioinformatics*, vol. 39, no. 1, p. btac830, Jan.  
445 2023, doi: [10.1093/bioinformatics/btac830](https://doi.org/10.1093/bioinformatics/btac830).
- 446 [18] N. A. O'Leary *et al.*, 'Reference sequence (RefSeq) database at NCBI: current status, taxonomic  
447 expansion, and functional annotation', *Nucleic Acids Res.*, vol. 44, no. D1, pp. D733–745, Jan. 2016, doi:  
448 [10.1093/nar/gkv1189](https://doi.org/10.1093/nar/gkv1189).
- 449 [19] The UniProt Consortium, 'UniProt: the Universal Protein Knowledgebase in 2023', *Nucleic Acids Res.*, vol.  
450 51, no. D1, pp. D523–D531, Jan. 2023, doi: [10.1093/nar/gkac1052](https://doi.org/10.1093/nar/gkac1052).
- 451 [20] M. Land *et al.*, 'Insights from 20 years of bacterial genome sequencing', *Funct. Integr. Genomics*, vol. 15,  
452 no. 2, pp. 141–161, Mar. 2015, doi: [10.1007/s10142-015-0433-4](https://doi.org/10.1007/s10142-015-0433-4).
- 453 [21] R. E. Timme, M. Sanchez Leon, and M. W. Allard, 'Utilizing the Public GenomeTrakr Database for  
454 Foodborne Pathogen Traceback', *Methods Mol. Biol. Clifton NJ*, vol. 1918, pp. 201–212, 2019, doi:  
455 [10.1007/978-1-4939-9000-9\\_17](https://doi.org/10.1007/978-1-4939-9000-9_17).
- 456 [22] M. Achtman *et al.*, 'Genomic diversity of Salmonella enterica -The UoWUCC 10K genomes project',  
457 *Wellcome Open Res.*, vol. 5, p. 223, Feb. 2021, doi: [10.12688/wellcomeopenres.16291.2](https://doi.org/10.12688/wellcomeopenres.16291.2).

- 458 [23] D. H. Parks *et al.*, 'A standardized bacterial taxonomy based on genome phylogeny substantially revises  
459 the tree of life', *Nat. Biotechnol.*, vol. 36, no. 10, Art. no. 10, Nov. 2018, doi: 10.1038/nbt.4229.
- 460 [24] 'Minicystis rosea gen. nov., sp. nov., a polyunsaturated fatty acid-rich and steroid-producing soil  
461 myxobacterium | Microbiology Society'. Accessed: Jan. 30, 2024. [Online]. Available:  
462 <https://www.microbiologyresearch.org/content/journal/ijsem/10.1099/ijs.0.068270-0>
- 463 [25] A. Rodríguez-Gijón *et al.*, 'A Genomic Perspective Across Earth's Microbiomes Reveals That Genome Size  
464 in Archaea and Bacteria Is Linked to Ecosystem Type and Trophic Strategy', *Front. Microbiol.*, vol. 12, p.  
465 761869, Jan. 2022, doi: 10.3389/fmicb.2021.761869.
- 466 [26] T. H. M. Smits, 'The importance of genome sequence quality to microbial comparative genomics', *BMC*  
467 *Genomics*, vol. 20, no. 1, p. 662, Aug. 2019, doi: 10.1186/s12864-019-6014-5.
- 468 [27] D. H. Parks, M. Chuvochina, P. R. Reeves, S. A. Beatson, and P. Hugenholtz, 'Reclassification of *Shigella*  
469 species as later heterotypic synonyms of *Escherichia coli* in the Genome Taxonomy Database'. bioRxiv, p.  
470 2021.09.22.461432, Sep. 22, 2021. doi: 10.1101/2021.09.22.461432.
- 471 [28] S. Appelt *et al.*, 'Genetic diversity and spatial distribution of *Burkholderia mallei* by core genome-based  
472 multilocus sequence typing analysis', *PLoS ONE*, vol. 17, no. 7, 2022, doi: 10.1371/journal.pone.0270499.
- 473 [29] L. Losada *et al.*, 'Continuing evolution of *Burkholderia mallei* through genome reduction and large-scale  
474 rearrangements', *Genome Biol. Evol.*, vol. 2, pp. 102–116, Jan. 2010, doi: 10.1093/gbe/evq003.
- 475 [30] C. L. Hatcher, L. A. Muruato, and A. G. Torres, 'Recent Advances in *Burkholderia mallei* and *B.*  
476 *pseudomallei* Research', *Curr. Trop. Med. Rep.*, vol. 2, no. 2, pp. 62–69, Jun. 2015, doi: 10.1007/s40475-  
477 015-0042-2.

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