

Inheritance of somatic mutations can affect fitness in monkeyflowers

Matthew A. Streisfeld^{1*}, Jessie C. Crown¹, Jack J. McLean¹, Aidan W. Short¹, and Mitchell B. Cruzan²

¹ Institute of Ecology and Evolution, University of Oregon, Eugene, OR 97403 USA

² Department of Biology, Portland State University, Portland, OR, 97207 USA

*Author for correspondence (mstreis@uoregon.edu)

Keywords: Acquired mutations, autogamy, cell lineage selection, *Mimulus aurantiacus*, geitonogamy, natural selection

Abstract

Plants possess the unique ability to transmit mutations to progeny that arise both through meiotic and mitotic (somatic) cell divisions. This is because the same meristem cells responsible for vegetative growth also generate gametes for sexual reproduction. Despite the potential for somatic mutations to be an additional source of genetic variation for adaptation, their role in plant evolution remains largely unexplored. We performed multiple experiments in the bush monkeyflower (*Mimulus aurantiacus*) to determine the fitness effects of somatic mutations inherited across generations. We tracked somatic mutations transmitted to progeny by generating self-pollinations within a flower (autogamy) or between stems of the same plant (geitonogamy). Autogamy and geitonogamy lead to different segregation patterns of somatic mutations among stems, making it possible to compare average fitness due to somatic variants. We found increased fecundity following autogamy, as well as significant impacts on drought tolerance, survival, and biomass. The variance in fitness was also greater following autogamy, consistent with the effects of somatic mutations impacting fitness. Effect sizes were small, but predictable, given that *M. aurantiacus* is a long-lived, drought-adapted shrub. These results reveal the importance of inherited somatic mutations as a source of genetic variation that can be relevant for plant adaptation.

Introduction

1 Mutations are the ultimate source of genetic variation. While this is a well-known saying
2 in genetics, only mutations that are transmitted to subsequent generations will be relevant for
3 evolution. Mutations are generated through both mitosis and meiosis, but among most animals,
4 only mutations that arise in the germline can be transmitted to progeny. This is because the
5 germline is determined early in development and is separated from the somatic cell lineages that
6 form the rest of the organism's body. Therefore, the set of somatic mutations that form during
7 mitotic division outside of the germline are typically not heritable.

8 By contrast, plants undergo indeterminate growth, where shoot and root systems
9 continually elongate and develop throughout a significant portion of their life-cycle (Antolin &
10 Strobeck. 1985, D'Amato 1996). Growth of the shoot system in plants occurs at shoot apical
11 meristems (SAMs), which contain a population of undifferentiated cells known as the central
12 zone. In vascular plants, these cells differentiate into leaf and stem tissue necessary for growth
13 and development, and they eventually produce the gametes required for sexual reproduction.
14 This reservoir of pluripotent cells is continually replenished through mitotic division
15 (Kwiatkowska 2008), but as the shoot elongates, somatic mutations may occur due to DNA
16 replication errors. These somatic mutations can accumulate as the stem elongates, resulting in
17 distal areas of the shoot system possessing more somatic variants than their basal counterparts
18 (Schultz & Scofield 2009). In angiosperms, the gametes are not produced until later in
19 development when the SAM is first converted to a floral meristem and then to a flower,
20 indicating that somatic mutations may be transmitted to offspring. This leads to the possibility
21 that somatic mutations are an important source of genetic variation that can impact evolutionary
22 processes.

23 Despite the potential for the inheritance of somatic variants that accumulated during
24 vegetative growth, the role and relevance of somatic mutations within plants remains unsettled.
25 Since plants possess the ability to pass on both meiotic and somatic mutations to progeny, one
26 might expect the mutation rate per generation among plants would be noticeably higher than
27 animals. However, mutation rates per generation appear to be similar between plants and animals
28 (Gaut et al. 2011). Multiple explanations have been offered to explain this discrepancy.

29 For example, germline segregation in plants may occur earlier in development than
30 previously appreciated, with primordial germ cells physically separated from future somatic cells
31 within the meristem (Lanfear et al. 2018). This explanation asserts that somatic mutations arising
32 during vegetative growth are only rarely inherited by progeny, since future germ cells would
33 only be found in isolated cell lineages (Cruzan 2018). These isolated populations of germ cells
34 could potentially have a slower rate of division than their somatic counterparts, and as a result,
35 they would have a significantly lower mutation rate over time (Lanfear et al. 2018). Due to its
36 slow cell division rate relative to the peripheral zone and rib meristem, the central zone of the
37 angiosperm SAM is a candidate location for this proposed population of segregated germ cells
38 (Cutter 1965). However, additional observations in angiosperm models have revealed that
39 mitotic activity spikes both within the central zone and rib meristem during the transition from
40 vegetative to reproductive tissue, suggesting that multiple zones contribute to the formation of
41 the gametes (Kwiatkowska 2008). More recently, computational models based on quantitative
42 cell lineage data from *Arabidopsis thaliana* and tomato (*Solanum lycopersicum*) were used to
43 replicate patterns of cell division in SAMs and axillary meristems (Burian et al. 2016). These
44 models suggested that cells were not constantly replaced within the central zone of the SAM, and

45 instead persisted throughout vegetative growth. Burian et al. (2016) claimed these findings
46 indicated that plants possess mechanisms to prevent the fixation and eventual accumulation of
47 deleterious genetic load. They further asserted that plants possess germlines analogous to those
48 found within animals.

49 An alternate explanation posits that cell lineages containing deleterious somatic
50 mutations are removed from the population of meristem cells due to natural selection (Cruzan
51 2018). This has been referred to as cell lineage selection (CLS; Fagerstrom et al. 1998; Otto and
52 Hastings 1998; Monro and Poore 2009). Since the size of the central zone is fixed and is
53 constantly replenished through mitotic division, cell lineages that express deleterious mutations
54 may replicate more slowly and therefore will be replaced by cell lineages with accelerated
55 division (Pineda-Krch & Lehtila 2002). Models of stochastic growth have indicated that
56 relatively minor differences in cell replication rates during development can result in significant
57 differences in the proportion of mutant cells found within adults (Otto & Orive 1995; Pineda-
58 Krch & Lehtila 2002). These models are supported by Yu et al. (2020), who identified thousands
59 of single nucleotide polymorphisms among ramets (individual stems) of common eelgrass
60 (*Zostera marina*) that were impacted by natural selection. Furthermore, Cruzan et al. (2022)
61 observed that seep monkeyflower (*Mimulus guttatus*) exhibited extraordinary variation in fitness
62 due to the accumulation of somatic mutations during stem growth, which in some cases led to
63 higher fitness from potentially beneficial somatic mutations being inherited by progeny. This
64 increased fitness may be a result of the novel environments that the plants were grown in (i.e.,
65 salt stress), as somatic mutations that were transmitted to offspring would have a high probability
66 of being beneficial (Fisher 1930). These results suggest that somatic mutations can play a non-
67 negligible—and possibly beneficial—role in plant fitness, challenging earlier studies on the
68 topic, which have claimed that beneficial mutations should be exceedingly rare (Charlesworth &
69 Willis 2009).

70 To shed additional light on the evolutionary consequences of somatic mutations, we
71 performed multiple experiments to determine the fitness effects of inherited somatic mutations in
72 the bush monkeyflower (*Mimulus aurantiacus* Curtis; Phrymaceae). *M. aurantiacus* is a woody,
73 perennial subshrub that is found throughout semi-arid regions of southwestern North America
74 (McMinn 1951). To track the fitness effects of somatic mutations that accumulate within a single
75 generation, we take advantage of the fact that these shrubs have separate stems. Each stem can
76 thus contain distinct germ cell lineages that are derived from the same zygote. As a consequence,
77 each stem can potentially contain different sets of somatic mutations that have accumulated
78 during growth.

79 By making crosses either within the same flower (autogamy) or between flowers on
80 separate stems of the same plant (inter-stem geitonogamy—hereafter, just geitonogamy), we can
81 produce progeny segregating for somatic mutations that vary among stems. Critically, these
82 crosses are both self-fertilizations, which leads to high homozygosity of meiotic mutants.
83 However, the offspring of each cross type will differ in the complement of somatic mutations
84 that they inherit. For a diploid plant, we can assume that somatic mutations ($a \rightarrow a'$) will be in
85 the heterozygous state when they first appear. For progeny generated via autogamy, a somatic
86 mutation will segregate as 25% homozygous ($a'a'$), 50% heterozygous (aa'), and 25% the
87 original (wildtype) homozygote (aa). By contrast, progeny from geitonogamous crosses will
88 segregate for somatic mutations that are different in each stem, such that 50% of offspring will
89 be carrying mutations in the heterozygous state and none of the progeny will be homozygous for
90 mutations that arose in a single stem. Thus, the average fitness effects of somatic mutations can

91 be evaluated by comparing the difference in fitness of progeny generated by autogamous and
92 geitonogamous crosses (Bobiwash et al. 2013; Schultz and Scofield 2009).

93 As noted above, prior studies in the herbaceous perennial *M. guttatus* demonstrated
94 substantial fitness consequences of somatic mutations when grown under salt stress (Cruzan et al
95 2022). By investigating the fitness effects of somatic mutations in a large and long-lived, woody
96 shrub (*M. aurantiacus*), we are able to compare results between two closely related plant species
97 that differ in important life history characteristics. Moreover, rather than testing progeny in a
98 novel environment, we challenged progeny under drought conditions – a stress that *M.*
99 *aurantiacus* routinely encounters in its native habitat (Sobel et al 2019). We followed fitness
100 among these two sets of progeny across multiple stages in the life cycle, including fecundity,
101 germination, early seedling growth rates, survival under terminal drought conditions, and total
102 biomass. Under a model where CLS sieves out deleterious somatic mutations while retaining
103 beneficial ones, we expect to find significant differences in fitness between progeny generated
104 from autogamous and geitonogamous pollination (Cruzan et al 2022). This difference in fitness
105 would be attributable to the accumulation of somatic mutations in vegetative tissue that were
106 subsequently transmitted to progeny. In addition, due to the different patterns of segregation of
107 somatic mutations between cross types, we expect progeny from autogamous pollinations to
108 display increased variation in fitness compared to progeny from geitonogamous crosses (Cruzan
109 et al. 2022). Findings from this study contribute to our understanding of the relevance of somatic
110 mutations in plant evolution.

111

112

113

Materials and Methods

114

115 *Experimental setup* - To estimate the fitness effects of somatic mutations, we made
116 autogamous and geitonogamous crosses in 26 *M. aurantiacus* genets that had been growing in an
117 open plot in Eugene, Oregon for four years. These genets were initially created through the
118 crossbreeding of red- and yellow-flowered ecotypes of *M. aurantiacus* ssp. *puniceus* (Sobel &
119 Streisfeld 2015; Chase et al. 2017). Using saturating pollen loads from a single flower at the end
120 of each stem, we made four crosses: one autogamous pollination on that same flower and three
121 geitonogamous pollinations to flowers on different stems of the same genet. Hereafter, we refer
122 to the offspring from a set of autogamous and geitonogamous crosses made from a single pollen
123 donor as a “unit.” Because somatic mutations can arise uniquely in any stem, we created multiple
124 units from different stems on the same genet (mean: 1.8 per genet; range 1 - 4). Specifically,
125 between 1 and 22 July 2021, we made 170 crosses, of which 163 developed into fruits. This
126 included 42 individual units that successfully produced a fruit from the autogamous cross and at
127 least two of the geitonogamous crosses. These were used in subsequent analyses.

128 We note that this approach is an improvement over the method used in Cruzan et al.
129 (2022), where pollen from two stems was reciprocally crossed to create autogamous and
130 geitonogamous pollinations. In that case, distinct somatic mutations in each stem could not be
131 controlled for, which may have impacted estimates of fitness. By using pollen from a single
132 flower to produce multiple geitonogamous crosses on different stems, we were better able to
133 control for different mutations among stems.

134

135 *Fecundity* – Fruits were collected when they turned brown and stored at room
136 temperature for two months to allow them to mature fully. Each mature fruit was weighed to the

137 nearest 0.1 mg. Seeds were carefully separated from their capsule, and all seeds from each fruit
138 were weighed. Seeds were then photographed using a Sony Alpha 6000 digital camera and
139 counted using ImageJ software.

140

141 *Seed germination and growth rate* - From four units (two units each from genets A and
142 C), we performed a germination experiment to determine if the time to germinate differed
143 between pollination treatments. For each of the four units, we filled two 96-cell plug trays with
144 moist potting soil and randomly sowed 192 seeds derived from autogamy and 192 seeds from
145 geitonogamy (64 seeds from each of the three geitonogamous crosses) across the cells (two seeds
146 of the same cross type per cell). Trays were placed in a grow room equipped with fluorescent
147 lights and maintained at 22C with a 16-hour photoperiod. Trays were bottom-watered and
148 overhead misted as needed. Seedling emergence was recorded at the same time each day for 16
149 days after the first seedling emerged. Each day, seedlings were digitally photographed from
150 above with a ruler in the frame, and we estimated total leaf area using Adobe Photoshop. To
151 estimate early seedling growth rates, we subtracted the total leaf area on the first day a seedling
152 emerged from the total leaf area on the final day of the experiment and divided this by the
153 number of days since the seedling emerged.

154

155 *Drought Sensitivity* – In the Cruzan et al (2022) study, the fitness of *M. guttatus* offspring
156 was measured in a novel greenhouse environment. However, the ecotypes of *M. aurantiacus* ssp.
157 *puniceus* are drought tolerant shrubs that have adapted to endure seasonal droughts in southern
158 California (Sobel et al. 2019). Because of these seasonal droughts, drought sensitivity likely
159 serves as a principal agent of selection for these ecotypes in the wild. Therefore, to determine if
160 somatic mutations can impact the fitness of offspring under drought conditions, we employed a
161 terminal drought experiment (as in Sobel et al 2019).

162 Using the seedlings from the germination experiment, we randomly selected 48 plants
163 from autogamous crosses and 48 seedlings from geitonogamous crosses (16 per cross) to
164 transplant into individual cone-shaped pots (21 cm deep) filled with potting soil, which were
165 placed into random positions within a separate 98-cell rack for each unit. Racks were placed in
166 the University of Oregon greenhouse and bottom watered as needed for two weeks to allow
167 seedlings to recover and to establish their roots in the deep cones. After this, no water was added.
168 On each subsequent day, a single researcher categorically scored plant health using a scale
169 between 0 and 4 (as described in Sobel et al. 2019). A score of 0 indicated no sign of drought
170 stress. A score of 1 indicated initial signs of drought stress, including the adaxial side of the
171 leaves curling under. A score of 2 indicated the first sign of true wilting. A score of 3 indicated
172 systemic and severe wilting. A score of 4 indicated death of the plant. The experiment ended
173 once all plants were assigned a score of 4. Plants were measured at the same time each day
174 throughout the experiment, and the identity of the pollination treatment was kept blind to the
175 evaluator until the end of the experiment. At the end of the experiment, the above ground plant
176 material was harvested, dried, and weighed to provide a final estimate of biomass at the time the
177 plant died. To test the effects of somatic mutations across a broader set of stems, we repeated the
178 drought experiment using six additional units (one unit from each of six additional genets; a total
179 of 960 seedlings measured among the 10 units), but we did not collect germination, growth rate,
180 or biomass data from these plants.

181 To provide an estimate of drought tolerance from these time-series data, we fit a three-
182 parameter logistic curve to the drought scores estimated in each plant on each day of the

183 experiment. Then, we estimated the parameter ‘ b ,’ which occurs at the time (in days) when the
184 drought score reaches 50% of its maximum. This corresponds to the rate at which each plant
185 begins showing obvious signs of drought stress, such that a larger value of ‘ b ’ indicates a more
186 drought tolerant plant. This was repeated separately for each plant within each of the 10 units
187 used in the drought experiments. We also estimated the time (in days) for plants to reach a
188 drought score of 4 (i.e., the survival time). Prior to analysis, we removed 11 plants that died too
189 quickly to obtain accurate parameter estimates.

190
191 *Data Analysis* – Our primary goal was to determine if there were fitness differences
192 between offspring generated from autogamy and geitonogamy. To begin, we averaged the seed
193 counts and seed and fruit weights from the multiple geitonogamous crosses per unit and used
194 separate paired t-tests to determine if fruit weight, seed weight, and seed count differed
195 significantly between autogamous and geitonogamous pollinations. We then standardized the
196 individual fitness components from each unit to a mean of zero and standard deviation of one.
197 We performed separate MANOVAs for each unit to test if the five fitness components estimated
198 on each seedling differed between pollination treatments. Statistical significance was tested using
199 Pillai’s trace, and effect size was calculated using the partial eta-squared method (Cohen 1988).
200 Individual linear models were then performed with each fitness component as the response
201 variable and cross type as the predictor variable in each of the four units to determine which
202 aspects of fitness differed between pollination type. Finally, we estimated the coefficient of
203 variation between autogamous and geitonogamous treatments for each fitness component across
204 the four units to determine if the variance in fitness was higher in progeny from autogamous
205 crosses, as predicted under a model of cell lineage selection. All analyses were performed in R.
206

207 Results

208

209 *Fecundity* – We identified significant effects of cross type on fecundity. Specifically, among the
210 42 units from 26 genets generated in this experiment, autogamous pollination consistently
211 resulted in more seeds than geitonogamous pollination (paired t-test, $p = 0.014$; Fig 1). Indeed, in
212 29 of the 42 units (69%), the total seed count per fruit was higher from autogamous crosses. This
213 pattern was similar for fruit weight and total seed weight as well (both $p = 0.004$), which were
214 each strongly correlated with seed count (seed count vs seed weight: $r = 0.87$; seed count vs fruit
215 weight: $r = 0.58$).
216
217

218

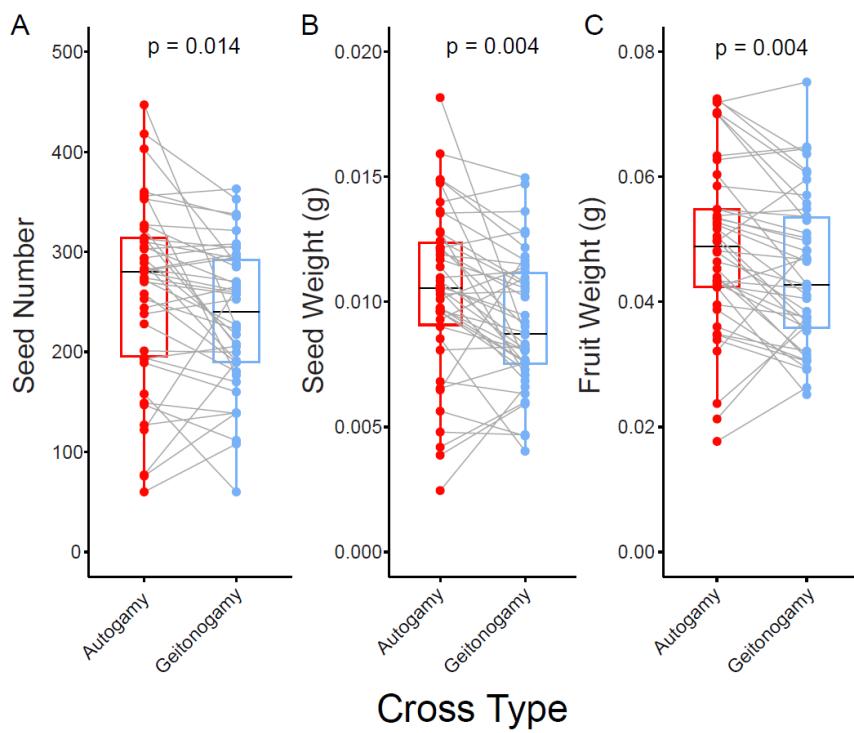


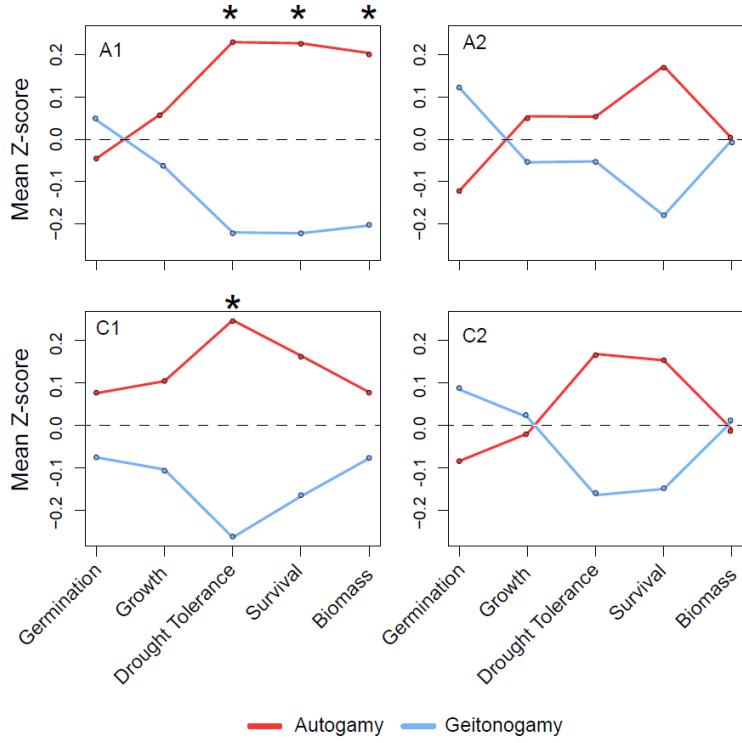
Figure 1. Fecundity is higher following autogamous pollination compared to geitonogamous pollination. In each panel, gray lines connect the fecundity estimates from autogamous and geitonogamous pollinations from each of the 42 units in the experiment. Box plots show the median (in black), the bottom and top of the boxes correspond to the first and third quartile, respectively, and whiskers represent 1.5 times the interquartile range. P-values above each plot were estimated using paired t-tests. A) Seed number per fruit, B) total seed weight, C) total fruit and seed weight. Values for geitonogamous crosses were averaged from the two or three crosses made within each unit.

219

220 *Patterns of selection in offspring* - We measured variation in five aspects of fitness among the
221 offspring of autogamous and geitonogamous pollinations. These components of selection acted
222 at different stages of the plant life cycle, beginning with germination, and continuing through
223 early seedling growth rates, drought tolerance, survival, and total biomass. Using MANOVA, we
224 found an overall significant difference in fitness for progeny derived from autogamous and
225 geitonogamous crosses in two of the four units (A1 and C2; Table S1). In both cases, the partial
226 eta-squared value > 0.14 , indicating a moderate to large effect of cross type on the multivariate
227 fitness estimates (Cohen 1988). By contrast, the other two units (A2 and C1), which were
228 derived from different stems of these same two genets, showed no difference between pollination
229 types ($P > 0.155$). These results demonstrate variation in fitness among stems of the same genet,
230 likely due to different complements of somatic mutations having accumulated in each stem.

231 In the offspring of unit A1, there were significant differences in drought tolerance,
232 survival, and biomass between pollination treatments (Table S2), with the mean fitness being
233 higher in autogamous crosses for drought tolerance and survival and lower for biomass (Fig 2,

234 Fig S1). Differences in drought tolerance were negatively correlated with both early seedling
235 growth rates and biomass (Fig S2, S3), consistent with previous findings in this species that
236 revealed smaller plants were better able to withstand drought conditions (Sobel et al 2019).
237 Interestingly, even though we found an overall significant effect of cross type on multivariate



238
239

Figure 2. Mean fitness varies between cross types across the four experimental units. The five fitness components are listed at the bottom, in order of their occurrence across the life-cycle. The data are presented as mean Z-scores for each fitness component broken down by cross type, such that the overall mean is 0 and the standard deviation is 1. The gray dashed line is at zero, corresponding to the expected mean values across both pollination treatments. Values above and below zero correspond to the number of standard deviations above and below the mean, respectively. Individual boxplots of each fitness component are presented in Fig. S1. To aid in visual presentation, individual Z-scores for growth rate and biomass were inverted by multiplying values by -1, and the mean was estimated. Asterisks indicate statistically significant differences between pollination treatments ($p < 0.05$).

240 fitness in unit C2, none of the five selection components were individually significant between
241 pollination types (Table S2). This implies that despite an effect of cross type when all
242 components are tested together, that effect is not driven strongly by any one measure of fitness.
243 By contrast, in unit C1, drought tolerance was significantly higher in offspring derived from
244 autogamous crosses (Table S2), even though the overall MANOVA was not significant (Table
245 S1).

246 Across all estimated fitness components, we did not find an effect of cross type on the
247 timing of germination or early seedling growth rates. However, we did observe differences in

249 drought tolerance in two of the four units. Therefore, we tested whether this pattern was
250 consistent among a larger set of units from six additional genets. In one of the six units, there
251 was a significant difference in drought tolerance between cross types, with plants derived from
252 geitonogamous pollination having a slightly higher mean value of drought tolerance (Table S3).
253 There were no differences between cross types in the other five units.

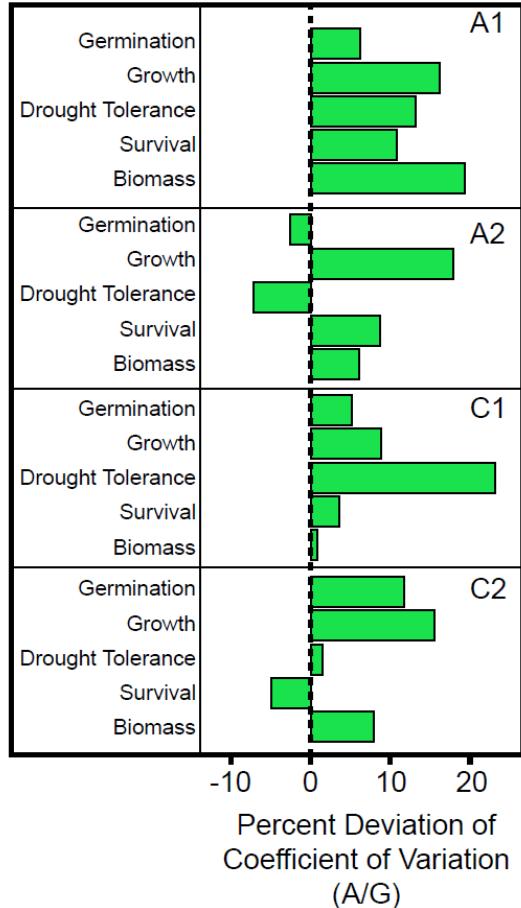


Figure 3. The variance in fitness in offspring is higher following autogamy compared to geitonogamy. Shown is the percent deviation in the coefficient of variation between autogamous and geitonogamous pollination treatments for the different fitness estimates across all four units. A value of zero would indicate that the coefficient of variation for a particular fitness estimate is the same between autogamy and geitonogamy, but positive values reveal higher variation in fitness among offspring derived from autogamy compared to geitonogamy.

254
255
256 In addition to differences in fitness, we also predicted that selection occurring in progeny
257 following the transmission of somatic mutations that accumulated in stems would result in a
258 higher variance in fitness in the offspring from autogamous pollinations compared to
259 geitonogamous pollinations. If somatic mutations affect offspring fitness, variation in fitness
260 should be greater for progeny groups from autogamy than from geitonogamy, as long as
261 mutations are not completely dominant. This is because somatic mutations will segregate as
262 homozygotes and heterozygotes in autogamous progeny but will remain heterozygous in the
263 progeny of geitonogamous crosses. To investigate this, we compared the coefficient of variation
264 for each of the five fitness components between autogamous and geitonogamous treatments. We
265 find that the coefficient of variation is higher in offspring from autogamy in 17 of the 20 cases,
266 with a deviation that averages 10.9% higher following autogamy than geitonogamy. In
267 particular, in unit C1, we see that the variation in drought tolerance is 23.2% higher in the
268 autogamy treatment compared to geitonogamy. Similarly, we see higher variance among
269 autogamous offspring for drought tolerance, survival, and biomass in unit A1 (range 10.8 – 13.9
270 %), the three components that were significantly different between pollination treatments. The
271 excess variance in the autogamy treatment compared to geitonogamy is highly significant across

272 all units combined (binomial probability, $p = 0.001$), implying that there is overall higher
273 variance in fitness following autogamy, consistent with the effects of somatic mutations
274 accumulating within stems and affecting fitness as they segregate in offspring.

275

276 Discussion

277

278 In this study, we demonstrated that the accumulation of somatic mutations in vegetative tissue
279 can impact the fitness of plants in the following generation. In addition, rather than somatic
280 mutations being uniformly deleterious, we show that they can occasionally have a net beneficial
281 effect, resulting in an increase of average fitness. This finding is consistent with expectations
282 from models of cell lineage selection (Fagerstrom et al. 1998; Otto and Hastings 1998; Monro
283 and Poore 2009), which argue that cell lineages with faster growth can displace slower ones
284 (Poethig 1987; Klekowski 2003). If these differences in division rates are determined by somatic
285 mutations, we would expect CLS to contribute to the purging of mutational load (Pineda-Krch
286 and Fagerstrom 1999; Monro and Poore 2009). Similarly, we expect mutations enhancing growth
287 to be retained. Therefore, cell lineage selection during vegetative growth has the potential to
288 modify the distribution of fitness effects of accumulated mutations by filtering expressed
289 deleterious mutations and allowing the transmission of beneficial variants. Specifically, we
290 found evidence that supports the accumulation and transmission of somatic mutations, which can
291 lead to higher fecundity, and in some cases, increased tolerance under drought conditions. These
292 results provide evidence for the potential importance of somatic mutations for plant evolution.

293 In spite of slow division rates and possibly enhanced DNA repair capacity (Yadav et al.
294 2009; Heyman et al. 2013), plant meristem cells are expected to accumulate substantial levels of
295 mutational load during stem elongation. The effects of this deleterious variation often are
296 apparent as reduced fecundity (or increased embryo abortion) following autogamous compared
297 to geitonogamous pollinations, which has been referred to as autogamy depression (Schultz and
298 Scofield 2009). Autogamy depression for seed and fruit abortion has been observed in several
299 species (reviewed in Bobiwash et al. 2013), including *M. guttatus* (Cruzan et al. 2022), and it is
300 expected to be stronger in longer lived plants, as longer lifespan should correspond to more
301 mitotic cell divisions and thus a greater opportunity for somatic mutation accumulation (Schultz
302 and Scofield 2009, Ally et al. 2010, Barrett 2015). Although *M. aurantiacus* is a long-lived
303 perennial shrub, we did not find evidence for autogamy depression. By contrast, we found an
304 overall average increase in fecundity following autogamy. Given that both cross types are self-
305 fertilizations, these differences cannot be attributable to variation in the strength of inbreeding
306 depression between treatments. Rather, the absence of autogamy depression in this system could
307 be due to the transmission of beneficial somatic variants whose fitness effects outweigh those of
308 deleterious mutations, resulting in a net increase in fecundity. Because deleterious mutations can
309 be filtered out due to CLS prior to fertilization, the presumed larger number of mitotic divisions
310 in these plants may actually result in a shift in the distribution of fitness effects that is skewed
311 toward the transmission of more beneficial mutations rather than deleterious ones. Although
312 these findings conflict with trends seen in other species that show autogamy depression is
313 common (Klekowski 1998, Bobiwash et al 2013), they are consistent with the pattern of an
314 unexpectedly high transmission of beneficial mutations in mutation accumulation studies in
315 *Arabidopsis thaliana* (Shaw et al. 2002; Rutter et al. 2010; Rutter et al. 2012; Rutter et al. 2018).
316 Moreover, it is important to note that the study of the fitness consequences of somatic mutations

317 is in its infancy. Therefore, further investigation on the consistency of these patterns among
318 closely related plants with different life history strategies is needed.

319 In addition to fecundity, we also measured variation in five aspects of fitness among the
320 offspring of autogamous and geitonogamous pollinations. These components of selection acted
321 at different stages of the plant life cycle, beginning with germination, and continued through
322 early seedling growth rates, drought tolerance, survival, and total biomass. While we did not find
323 significant differences in fitness between pollination treatments for germination or early seedling
324 growth rates in any of the units, we did find evidence for increased tolerance to drought, higher
325 survival, and lower total biomass in offspring derived from autogamy. We also found higher
326 variance in fitness among offspring derived from autogamy, which is in line with our results
327 demonstrating an increase in fitness in seedlings following autogamous pollination (Cruzan et al
328 2022). In one case (unit E3), we also found significantly higher fitness in seedlings following
329 geitonogamy, which suggests that the transmission of deleterious somatic mutations may have
330 occurred in the autogamous lines. Regardless, these results are consistent with the hypothesis
331 that somatic variants that accumulated during vegetative growth can be transmitted to offspring
332 where they can occasionally impact fitness. Observed increases in fitness after autogamy suggest
333 a potential role for somatic variation in local adaptation.

334 Despite detecting significant differences for fitness components in some of the units, the
335 individual effect sizes are rather small. This result is not unexpected for at least two reasons.
336 First, offspring were grown in an environment that closely matches their native habitat. The
337 populations of *M. aurantiacus* used in this experiment occur in chaparral communities of
338 southern California, which is dominated by hot, dry summers and cool, moist winters (Beeks
339 1962). Seedling recruitment tends to be very low due to the rapid drying of the soil after
340 seedlings emerge. Thus, the terminal drought experiment we conducted closely mimics the
341 conditions of natural seedlings (Sobel et al 2019). As a result, we would expect plants to already
342 be near their adaptive peaks for drought tolerance, suggesting that most new mutations would not
343 greatly improve fitness (Orr 2005). By contrast, previous work in the herbaceous *M. guttatus*
344 revealed that somatic mutations accumulating during vegetative growth had large, beneficial
345 effects on offspring fitness in five of the 14 stems tested (Cruzan et al 2022). In this case, the
346 progeny were grown in a novel environment (hydroponic salt-stress), implying that there was a
347 broader spectrum of mutations that could have phenotypic effects capable of moving the
348 population closer to its optimum. Second, our analyses focused on testing for average differences
349 in fitness between pollination treatments. Following autogamous pollination, only 25% of
350 offspring on average are expected to be homozygous for a somatic mutation that arose in that
351 stem. Therefore, provided that new mutations are not completely dominant, most somatic
352 variants will fail to be expressed in offspring, resulting in few plants that show differences in
353 fitness between cross types. As a consequence, our findings are consistent with a prediction of
354 small differences in average fitness between pollination treatments.

355 Although the segregation of somatic mutations in offspring can obscure overall statistical
356 patterns between pollination treatments, we can still see the net fitness effects of these variants in
357 individual progeny. Specifically, we observed individual plants derived from autogamous
358 pollination that have exceptional values of fitness, especially for drought tolerance, survival, and
359 biomass. For example, in units A1 and C1 (the units that show significant differences in drought
360 tolerance between pollination treatments), we see that the plants with the highest drought
361 tolerance are derived from autogamy. These plants have drought tolerance values that are more
362 than three standard deviations above the mean (Fig S4). This trend continues with the later-

363 acting fitness components, such that these same plants also have consistently extreme values of
364 survival and biomass. We also found a strong, negative relationship between drought tolerance
365 and early seedling growth rate, such that smaller plants tended to survive longer under drought
366 conditions. These results are consistent with those of Sobel et al (2019), who also found that
367 smaller *M. aurantiacus* plants tended to better withstand desiccation. They suggested that the
368 reduced leaf area of smaller plants likely resulted in lower transpiration, leading to greater
369 drought tolerance and thus longer survival under terminal drought conditions. Thus, the
370 segregation of somatic variants can result in progeny with extreme values of fitness, providing an
371 additional source of genetic variation for adaptation. Future experiments that take advantage of
372 the power of deep sequencing can be used to identify individual somatic variants that
373 accumulated in parents, which would allow us to track the fitness consequences of these variants
374 after they are transmitted to offspring.

375 In conclusion, we find evidence for the transmission of both beneficial and deleterious
376 somatic variation in offspring, revealing that somatic variation can occasionally underlie
377 adaptation. By comparing these results with those from the closely related *M. guttatus* with
378 different life history characteristics (Cruzan et al 2022), we found similar, though more subtle,
379 fitness consequences following autogamy. Furthermore, as noted earlier, the current study
380 improved on the crossing design used by Cruzan et al (2022). In this case, we used pollen from
381 the same flower for both an autogamous pollination, as well as multiple geitonogamous crosses
382 on different stems of the same genet, which allowed us to control for somatic variation among
383 stems. Thus, the fact that our results are consistent with those from Cruzan et al (2022), despite
384 differences in crossing design, environmental conditions, and life history, reveals the potential
385 relevance of somatic mutation for plant evolution.

386 Considering that the accumulation and transmission of somatic mutations may be a
387 general feature of plant evolution can provide some insight into the success and diversification of
388 flowering plants. The evolution of apical meristems and indeterminate growth in early land
389 plants may have influenced the potential for cell lineage selection to affect the distribution of
390 mutations acquired during vegetative growth. While the primary selective advantage for
391 producing reproductive structures at the ends of growing stems may have been for improved
392 dispersal, this architecture also maximized the potential for selection among cell lineages to
393 affect the distribution of mutations passed on to offspring. Even though plants are sedentary over
394 much of their life cycle and may be subjected to substantial environmental variation within a
395 single lifespan, we show here that the accumulation of somatic variation during vegetative
396 growth has the potential to contribute significantly to plant adaptation in subsequent generations.
397 Future work in population genetics should not ignore somatic mutations as an important source
398 of genetic variation that can impact plant evolution.

399
400

401 Acknowledgments

402
403
404
405
406
407
408

We would like to thank members of the Cruzan lab for their feedback on earlier drafts of this manuscript. We also thank S. Medbury for his assistance with plant care in the University of Oregon greenhouses. This work was supported by NSF-DEB 2051242 to MAS and MBC.

References

- 409 Ally, D., K. Ritland, and S. P. Otto. 2010. Aging in a Long-Lived Clonal Tree. *PLoS Biol.* 8.
410
- 411 Antolin, M. F. & Strobeck, C. (1985). The population genetics of somatic mutation in plants.
412 *Am. Nat.* 126:52-62.
- 413 Barrett, S. C. H. 2015. Influences of clonality on plant sexual reproduction. *Proceedings of the*
414 *National Academy of Sciences* 112:8859-8866
- 415 Beeks, R. M. 1962. variation and hybridization in southern California populations of *Diplacus*
416 (Scrophulariaceae). *El Aliso* 5:83-122.
- 417
- 418 Bobiwash, K., S. T. Schultz, and D. J. Schoen. 2013. Somatic deleterious mutation rate in a
419 woody plant: estimation from phenotypic data. *Heredity* 111:338-344.
- 420
- 421 Burian, A. & Barbier de Reuille, K. (2016). Patterns of stem cell divisions contribute to plant
422 longevity. *Current Biology*. 26(11):1385-1394.
- 423
- 424 Charlesworth, D. & Willis, J. H. (2009). The genetics of inbreeding depression. *Nature Reviews*
425 *Genetics*. 10: 783-796.
- 426
- 427 Chase, M. A., S. Stankowski, and M. A. Streisfeld. 2017. Genomewide variation provides insight
428 into evolutionary relationships in a monkeyflower species complex (Mimulus sect.
429 *Diplacus*). *American Journal of Botany* 104:1510-1521.
- 430 Cohen, J (1988) Statistical power analysis for the behavioral sciences (2nd ed.). Hillsdale, NJ:
431 Erlbaum.
- 432 Cruzan, M. B. 2018. *Evolutionary Biology - A Plant Perspective*. Oxford University Press, New
433 York.
- 434
- 435 Cruzan, M. B., M. A. Streisfeld, and J. A. Schwoch. 2022. Fitness effects of somatic mutations
436 accumulating during vegetative growth. *Evol. Ecol.* 36:767-785.
- 437
- 438 Cutter, E. G. (1965). Recent experimental studies of the shoot apex and shoot morphogenesis.
439 *Botanical Review*. 31(1): 7-21.
- 440
- 441 D'Amato, F. (1996). Role of somatic mutations in the evolution of higher plants. *Caryologia*.
442 50(1).
- 443
- 444 Fagerstrom, T., D. A. Briscoe, and P. Sunnucks. 1998. Evolution of mitotic cell-lineages in
445 multicellular organisms. *Trends Ecol. Evol.* 13:117-120.
- 446 Fisher, Ronald (1930). *The Genetical Theory of Natural Selection*. Oxford, UK: Oxford
447 University Press.

- 448 Gaut, B., Yang, L., Takuno, S., Eguiarte, L. E. (2011). The patterns and causes of variation in
449 plant nucleotide substitution rates. *Annual review of ecology, evolution, and systematics*.
450 42: 245-266.
- 451
- 452 Heyman, J., T. Cools, F. Vandenbussche, K. S. Heyndrickx, J. Van Leene, I. Vercauteren, S.
453 Vanderauwera, K. Vandepoele, G. De Jaeger, D. Van Der Straeten, and L. De Veylder.
454 2013. ERF115 Controls Root Quiescent Center Cell Division and Stem Cell
455 Replenishment. *Science* 342:860-863.
- 456
- 457 Klekowski, E. J. 1988. Mutation, Developmental Selection, and Plant Evolution. Columbia
458 University Press, New York.
- 459
- 460 Klekowski, E. J. 2003. Plant clonality, mutation, diplontic selection and mutational meltdown.
461 *Biol. J. Linn. Soc.* 79:61-67.
- 462
- 463 Kwiatkowska, D. (2008). Flowering and apical meristem growth dynamics. *Journal of
464 Experimental Botany*. 59(2): 187-201.
- 465
- 466 Lanfear, R. (2018). Do plants have a segregated germline?. *PLOS Biology*. 16(5).
- 467
- 468 McMinn, H. E. 1951. Studies in the genus *Diplacus*, Scrophulariaceae. *Madrono* 11:33-128.
- 469
- 470 Monro, K., and A. G. B. Poore. 2009. The Potential for Evolutionary Responses to Cell-Lineage
471 Selection on Growth Form and Its Plasticity in a Red Seaweed. *Am. Nat.* 173:151-163.
- 472
- 473 Orr, H. A. 2005 The genetic theory of adaptation: a brief history. *Nature Reviews Genetics*.
474 6:119-127.
- 475
- 476 Otto, S. P., and I. M. Hastings. 1998. Mutation and selection within the individual. *Genetica* 102-
477 3:507-524.
- 478
- 479 Otto, S. P., Orive, M. E. (1995). Evolutionary consequences of mutation and selection within an
480 individual. *Genetics*. 141(3): 1173-1187.
- 481
- 482 Pineda-Krch, M., and T. Fagerstrom. 1999. On the potential for evolutionary change in
483 meristematic cell lineages through intraorganismal selection. *J. Evol. Biol.* 12:681-688.
- 484
- 485 Pineda-Krch, M. P. & Lehtila, K. (2002). Cell lineage dynamics in stratified shoot apical
486 meristems. *Journal of Theoretical Botany*. 219(4): 495-505.
- 487
- 488 Poethig, R. S. 1987. Clonal analysis of cell lineage patterns in plant development. *Am. J. Bot.*
489 74:581-594.
- 490
- 491 Rutter, M. T., A. Roles, J. K. Conner, R. G. Shaw, F. H. Shaw, K. Schneeberger, S. Ossowski, D.
492 Weigel, and C. B. Fenster. 2012. Fitness of *Arabidopsis thaliana* mutation accumulation
493 lines whose spontaneous mutations are known. *Evolution* 66:2335-2339.

- 494
- 495 Rutter, M. T., A. J. Roles, and C. B. Fenster. 2018. Quantifying natural seasonal variation in
496 mutation parameters with mutation accumulation lines. *Ecology and Evolution* 8:5575-
497 5585.
- 498
- 499 Rutter, M. T., F. H. Shaw, and C. B. Fenster. 2010. Spontaneous mutation parameters for
500 *Arabidopsis thaliana* measured in the wild. *Evolution* 64:1825-1835.
- 501
- 502 Schultz, S. T. & Scofield, D. G. (2009). Mutation accumulation in real branches: fitness assays
503 for genomic deleterious mutation rate and effect in large-statured plants. *The American
504 Naturalist*. 174:163-175.
- 505
- 506 Shaw, F. H., C. J. Geyer, and R. G. Shaw. 2002. A comprehensive model of mutations affecting
507 fitness and inferences for *Arabidopsis thaliana*. *Evolution* 56:453-463.
- 508
- 509 Sobel, J. M., Stankowski, S., Streisfeld, M. A. (2019). Variation in ecophysiological traits might
510 contribute to ecogeographic isolation and divergence between parapatric ecotypes of
511 *Mimulus aurantiacus*. *Journal of Evolutionary Biology*. 32: 604-618
- 512
- 513 Sobel, J. M., Streisfeld, M. A. (2015). Strong premating reproductive isolation drives incipient
514 speciation in *Mimulus aurantiacus*. *International Journal of Organic Evolution*. 69(2):
515 447-461.
- 516
- 517 Yadav, R. K., T. Girke, S. Pasala, M. T. Xie, and V. Reddy. 2009. Gene expression map of the
518 *Arabidopsis* shoot apical meristem stem cell niche. *Proceedings of the National Academy
519 of Sciences of the United States of America* 106:4941-4946.
- 520
- 521

522

Supplemental Material

523

524

525 Table S1. Results from the MANOVAs for each unit, testing the combined effects of the five
526 fitness components by pollination treatment.

527

Unit	Pillai	Eta-squared	P
A1	0.16865	0.17	0.0055
A2	0.08913	0.09	0.1522
C1	0.07724	0.08	0.2124
C2	0.1358	0.14	0.0215

528

529

530

531 Table S2. Results from linear models testing each fitness estimate against pollination treatment
532 for each of the four units. The F-value of the test, degrees of freedom (Df) and the P-value are
533 reported. P-values less than 0.05 are in bold.

534

Unit	Fitness estimate	F	Df	P
A1	Germination	0.197	1, 94	0.659
A1	Growth rate	0.409	1, 94	0.524
A1	Drought tolerance	4.937	1, 92	0.029
A1	Survival	4.975	1, 93	0.028
A1	Biomass	4.103	1, 94	0.046
A2	Germination	1.440	1, 94	0.233
A2	Growth rate	0.281	1, 94	0.597
A2	Drought tolerance	0.251	1, 89	0.617
A2	Survival	2.980	1, 92	0.088
A2	Biomass	0.002	1, 94	0.988
C1	Germination	0.548	1, 94	0.461
C1	Growth rate	1.043	1, 94	0.310
C1	Drought tolerance	6.464	1, 91	0.012
C1	Survival	2.665	1, 93	0.106
C1	Biomass	0.582	1, 94	0.447
C2	Germination	0.684	1, 94	0.410
C2	Growth rate	0.036	1, 94	0.851
C2	Drought tolerance	2.684	1, 93	0.105
C2	Survival	2.216	1, 93	0.140
C2	Biomass	0.008	1, 94	0.927

535

536

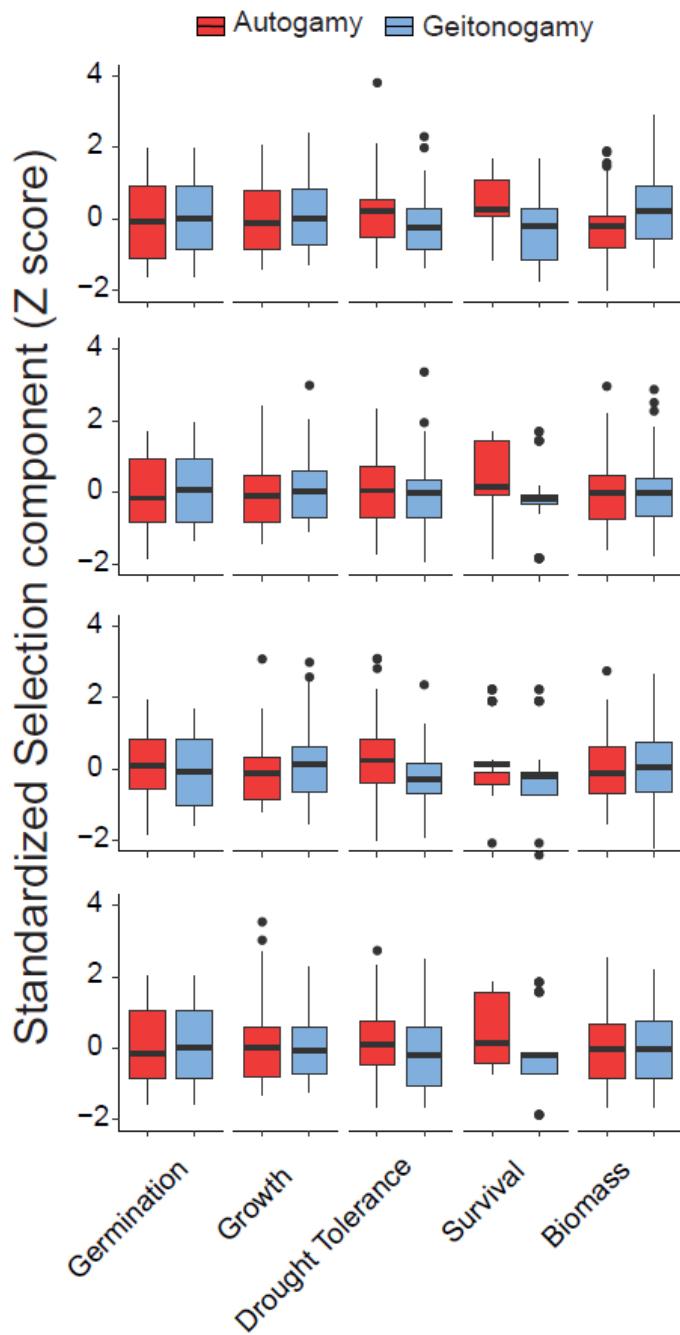
537 Table S3. Results from linear models testing the effects of pollination treatment on drought
538 tolerance in six additional units. The F-value of the test, degrees of freedom (Df) and the P-value
539 are reported. P-values less than 0.05 are in bold.

540

Unit	F	Df	P
E3	6.312	1, 94	0.0137
F1	1.01	1, 94	0.3175
H1	0.295	1, 94	0.5883
K1	0.112	1, 94	0.7382
O1	0.081	1, 94	0.7765
R2	0.396	1, 94	0.5308

541

542



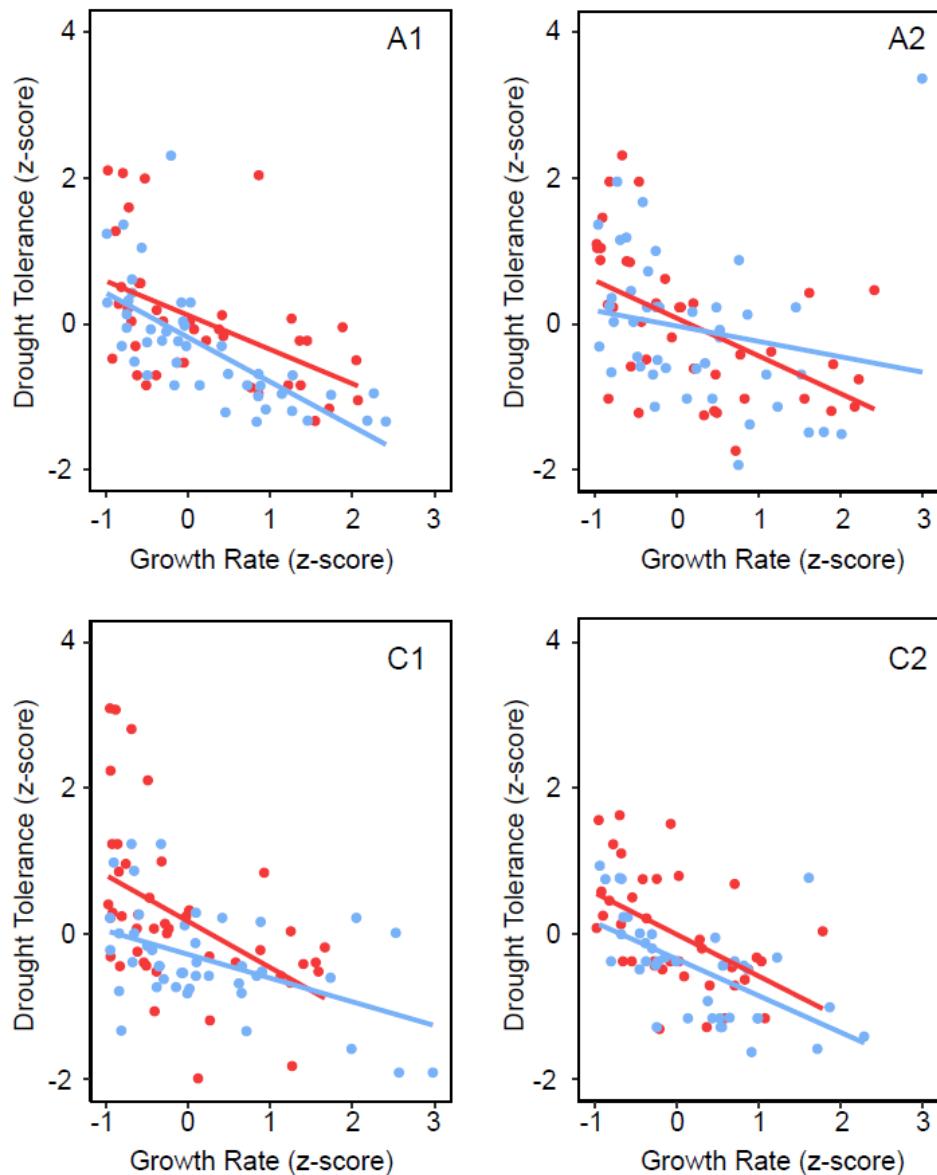
543

544

545 Figure S1. Boxplots of the fitness estimates for each fitness component across the four units
546 following autogamy (red) and geitonogamy (blue). Fitness values are standardized to z-scores,
547 with a mean of zero and standard deviation of 1. The black horizontal line corresponds to the
548 median, box heights indicate the lower and upper quartile, and whiskers correspond to 1.5 times
549 the interquartile range. From top to bottom, plots are for units A1, A2, C1, C2.

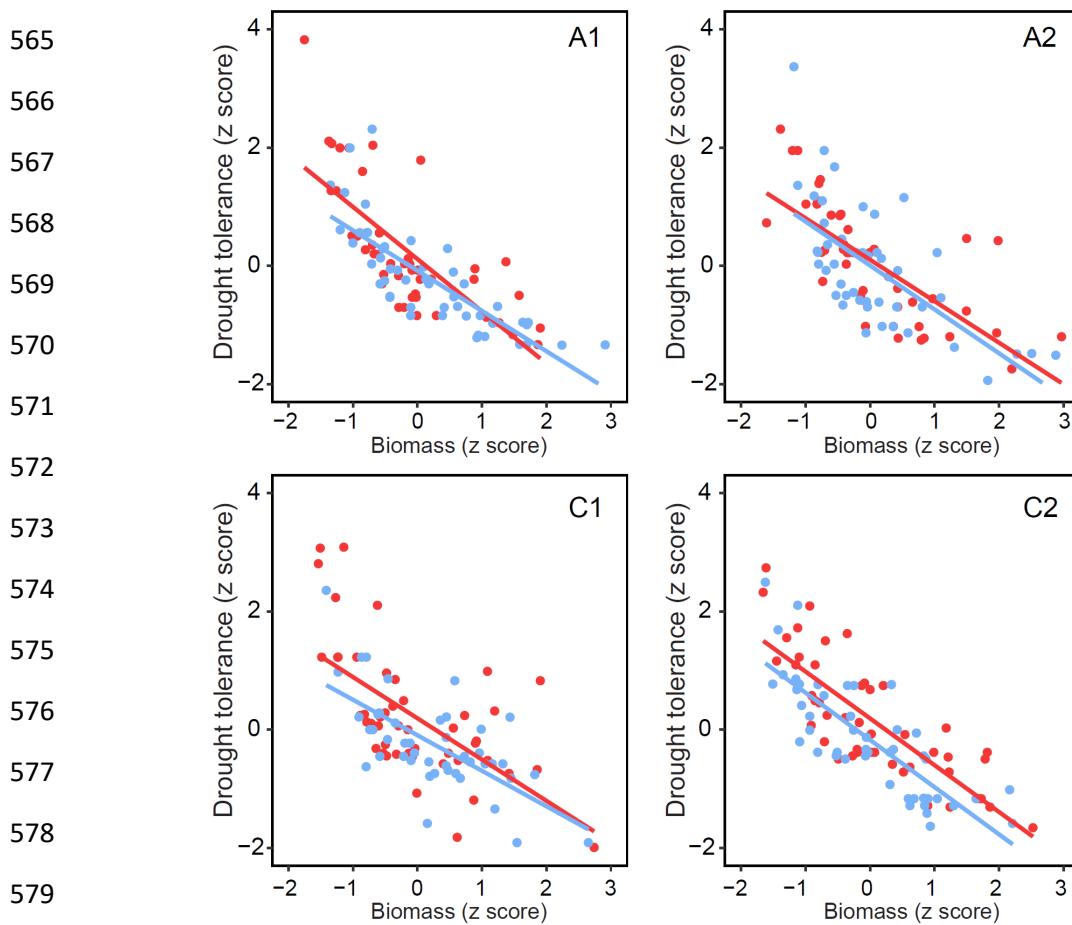
550

551
552
553
554
555



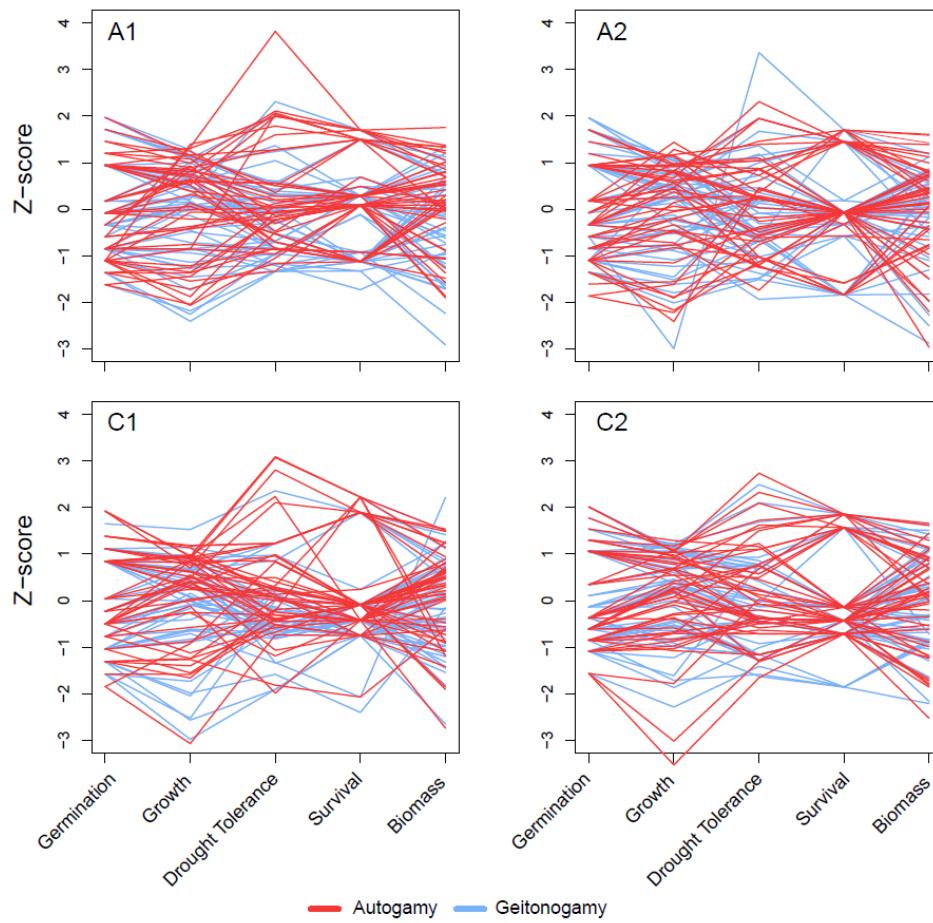
556
557
558
559
560
561
562
563
564

Figure S2. The relationship between drought tolerance and early seedling growth rate across the four units. Red points correspond to seedlings derived from autogamy and blue points correspond to seedlings derived from geitonogamy. Trendlines are derived from linear models testing the effect of drought tolerance against growth rate, separately for each pollination treatment. Fitness values are standardized to z-scores, with a mean of zero and standard deviation of 1.



581 Figure S3. The relationship between drought tolerance and biomass across the four units. Red
582 points correspond to seedlings derived from autogamy and blue points correspond to seedlings
583 derived from geitonogamy. Trendlines are derived from linear models testing the effect of
584 drought tolerance against biomass, separately for each pollination treatment. Fitness values are
585 standardized to z-scores, with a mean of zero and standard deviation of 1.
586

587
588
589



590
591 Figure S4. Line plots connecting fitness estimates for individual seedlings derived from either
592 autogamy (red) or geitonogamy (blue) across the four units. Fitness values are standardized to z-
593 scores, with a mean of zero and standard deviation of 1.