

1 **Venom-inspired somatostatin receptor 4 (SSTR4) agonists as new drug leads** 2 **for peripheral pain conditions**

3
4 Walden E. Bjørn-Yoshimoto*¹, Iris Bea L. Ramiro¹, Thomas Lund Koch^{1,2}, Ebbe Engholm³, Ho
5 Yan Yeung^{1,2}, Kasper K. Sørensen³, Carolyn M. Goddard⁴, Kathrine L. Jensen⁴, Nicholas A.
6 Smith⁵, Laurent F. Martin⁶, Brian J. Smith⁵, Kenneth L. Madsen⁴, Knud J. Jensen³, Amol
7 Patwardhan^{6,7}, Helena Safavi-Hemami*^{1,2,8}

8
9 ¹Department of Biomedical Sciences, University of Copenhagen; Copenhagen-N, Denmark

10 ²Department of Biochemistry, University of Utah; Salt Lake City, UT, USA

11 ³Department of Chemistry, University of Copenhagen; Frederiksberg, Denmark

12 ⁴Department of Neuroscience, University of Copenhagen; Copenhagen-N, Denmark

13 ⁵La Trobe Institute for Molecular Science, La Trobe University; Melbourne, Australia

14 ⁶Department of Anesthesiology and Pharmacology, University of Arizona; Tucson, AZ, USA.

15 ⁷Department of Anesthesiology and Pain Management, Peter O'Donnell Jr. Brain Institute, UT
16 Southwestern Medical Center; Dallas, Texas, USA

17 ⁸School of Biological Sciences, University of Utah; Salt Lake City, UT, USA

18
19 *Corresponding authors: walden@sund.ku.dk, helena.safavi@utah.edu

21 **One Sentence Summary:**

22 Venom peptides from predatory marine mollusks provide new leads for treating peripheral pain
23 conditions through a non-opioid target.

25 **Abstract:**

26 Persistent pain affects one in five people worldwide, often with severely debilitating consequences.
27 Current treatment options, which can be effective for mild or acute pain, are ill-suited for
28 moderate-to-severe persistent pain, resulting in an urgent need for new therapeutics. In recent
29 years, the somatostatin receptor 4 (SSTR4), which is expressed in sensory neurons of the peripheral
30 nervous system, has emerged as a promising target for pain relief. However, the presence of several
31 closely related receptors with similar ligand-binding surfaces complicates the design of receptor-
32 specific agonists. In this study, we report the discovery of a potent and selective SSTR4 peptide,
33 consomatin Fj1, derived from extensive venom gene datasets from marine cone snails. Consomatin
34 Fj1 is a mimetic of the endogenous hormone somatostatin and contains a minimized binding motif
35 that provides stability and drives peptide selectivity. Peripheral administration of synthetic
36 consomatin Fj1 provided analgesia in mouse models of postoperative and neuropathic pain. Using
37 structure-activity studies, we designed and functionally evaluated several Fj1 analogs, resulting in
38 compounds with improved potency and selectivity. Our findings present a novel avenue for
39 addressing persistent pain through the design of venom-inspired SSTR4-selective pain
40 therapeutics.

42 **INTRODUCTION**

43
44 Persistent pain, also referred to as chronic pain, poses a significant global challenge, impacting
45 more than one in five individuals and exacting substantial personal, societal, and economic burdens
46 (1). Current therapeutic approaches depend on the type of pain and involve medications such as
47 acetaminophen (paracetamol), non-steroidal anti-inflammatory drugs (NSAIDs), anticonvulsants,
48 antidepressants, and opioids (2). Although generally effective for acute pain, these interventions
49 often fail to sufficiently address severe persistent pain and can be associated with serious dose-
50 limiting side effects, tolerance, and dependence, especially after prolonged use (3). In particular,
51 the misuse of opioids has resulted in an epidemic of addiction and abuse of unprecedented scale
52 (4), underscoring the urgent need for novel therapeutics and targets that act through opioid-
53 independent pathways.

54
55 One such promising alternative target is the somatostatin receptor 4 (SSTR4), a member of the G
56 protein-coupled receptor (GPCR) family activated by the peptide hormones somatostatin (SST)
57 and cortistatin. The human SSTR family comprises five receptor subtypes (SSTR₁₋₅) that share
58 46–63 % sequence identity and similar ligand-binding surfaces. Based on their similarities and
59 evolutionary origin, the five subtypes can be divided into two groups: Group 1 comprises the
60 SSTR_{2,3} and SSTR₅ with sequence identities ranging from 50–58 %, and group 2 comprises the
61 SSTR₁ and SSTR₄, sharing 63 % sequence identity (5). Activation of SSTRs, particularly group 1
62 SSTRs, is associated with inhibition of the secretion of various hormones (6). Different members
63 of the SSTR family share the same endogenous ligands and are coupled to the G $\alpha_{i/o}$ family, but
64 have distinct expression profiles that can be associated with different physiological effects. For
65 example, the SSTR₂ and SSTR₅ are highly expressed in neuroendocrine neoplasms, where their
66 activation inhibits mitogenic signaling and growth (6). Somatostatin-based drug agonists that
67 specifically activate these subtypes have long been used for the treatment of neuroendocrine
68 disorders (e.g., acromegaly) and as diagnostic and therapeutic agents for certain types of cancers
69 (7-9).

70
71 The first indication of the potential role of SSTR4-selective agonists for the treatment of pain came
72 from the SSTR_{1,4}-targeting peptide, TT-232, originally developed for the treatment of cancer but
73 was later found to exhibit anti-inflammatory and anti-nociceptive effects (10-12). These results, as
74 well as additional studies on the small molecule agonist J-2156 (13, 14), established the SSTR4 as
75 a novel target for pain relief. Although the molecular mechanisms of SSTR4-mediated
76 antinociception are not fully understood, several studies have suggested that SSTR4 activation in
77 nociceptive neurons of the dorsal root ganglia (DRG) and trigeminal ganglia leads to downstream
78 G $\beta\gamma$ -mediated activation of G protein-coupled inwardly rectifying potassium channels (GIRKs),
79 allowing an outward flux of potassium ions, thereby hyperpolarizing cells involved in pain sensing
80 and/or transmission (15). Furthermore, SSTR4 activation can reduce capsaicin-induced transient
81 receptor potential cation channel subfamily V member 1 (TRPV1) currents in DRG neurons
82 through G α_i signaling, further hyperpolarizing the cell (15). Thus, SSTR4-selective ligands provide
83 analgesia through the inhibition of peripheral pain signals and do not require central nervous
84 system (CNS) administration or penetration.

85

86 Our recent discovery of a toxin agonist of group 2 SSTRs from a venomous cone snail, consomatin
87 Ro1 (16), suggested that cone snail venoms may provide a unique source for the discovery of novel
88 SSTR₄ ligands. Consomatin Ro1 provided analgesia in two mouse models of pain, but only
89 exhibited micromolar potency at the SSTR₄ and was equipotent at the SSTR₁ (16). To investigate
90 whether cone snails evolved additional SSTR₄-targeting toxins with improved potency and
91 selectivity profiles, we computationally searched the venom gene datasets of 247 species, resulting
92 in the identification of 529 SST-like lead sequences. Computational selection, synthesis, and
93 receptor profiling of eight candidate toxins from this dataset led to the identification of consomatin
94 Fj1, a potent and selective SSTR₄ agonist that provides analgesia in postoperative and neuropathic
95 pain models. Structure-activity studies using molecular dynamics (MD) simulations in
96 combination with analog design revealed the peptide's minimized binding mode and opportunities
97 for further improvement of Fj1 for drug design and development.

98

99 RESULTS

100 Identification of candidate SSTR agonists from large venom gene library

101 We recently showed that cone snails have evolved peptide toxins that share sequence similarity
102 with SST and its related peptides, cortistatin, urotensin II, and urotensin-related peptide (17). These
103 toxins, referred to as “consomatins”, have evolved from an SST-like peptide used for endogenous
104 signaling in cone snails. Following recruitment into the venom, these peptides greatly diversified
105 to form a large family of SST-like toxins (17). To facilitate the selection of consomatin candidates
106 that are likely to activate vertebrate SSTRs, with potential selectivity for the SSTR₄, we extracted
107 consomatin sequences from exon capture datasets of 247 cons snail species. We identified
108 consomatin sequences in 169 species, in which we identified 529 sequences which subjected to
109 principal component analysis (PCA) (see Methods for details). In addition to exon-capture data,
110 consomatin Ro1 and consomatin Ro2, which were previously identified by transcriptome
111 sequencing and mass spectrometric analysis of the venom of *Conus rolani*, as well as consomatin
112 pG1 (previously named G1) from the transcriptome of *C. geographus*, were also included in the
113 PCA (16). According to their prey, cone snails can be classified into worm hunters, snail hunters,
114 and fish hunters. To enable comparative analysis of consomatins to endogenous SST-like
115 hormones, peptide sequences from vertebrates (27 sequences), annelid worms (29 sequences), and
116 mollusks (18 sequences) were included in the analysis. Consistent with our previous findings (17),
117 toxins from worm hunting cone snails closely grouped with endogenous SST-like sequences from
118 annelid worms, whereas many of the consomatins from fish hunters, including consomatin Ro1,
119 grouped with vertebrate peptides (**Fig. 1A**). Consomatins were absent from snail hunting species.
120 For the selection of toxin sequences from the PCA plot for chemical synthesis and SSTR receptor
121 profiling, sequences from known fish hunting species were prioritized, as the SSTRs present in
122 fish were highly similar to their human orthologs. Of these, sequences that grouped with vertebrate
123 SST and related peptides were selected as these are more likely to have activity at the human
124 receptors. The final list of sequences and their respective positions on the PCA is shown in **Fig.**
125 **1A-B).**

126

127 **Receptor profiling identifies potent and selective agonist of the SSTR₄**

128 The predicted mature peptide sequences selected from the PCA plot were synthesized to > 90 %
129 purity using standard solid-phase peptide synthesis and verified using reverse-phase high-
130 performance liquid chromatography and mass spectrometry. Post-translational modifications were
131 predicted based on modifications previously observed for the venom peptide consomatin Ro1 (16).
132 Modifications included γ -carboxylation of Glu, hydroxylation of Pro (Hyp), L-to-D epimerization
133 of a Trp positioned within the disulfide loop, and C-terminal amidation based on the presence of a
134 Gly-Arg/Lys motif (18). Disulfide bonds were predicted based on the presence of two cysteine
135 residues. We note that the predicted modifications and the proteolytic N- and C-terminal cleavage
136 sites may differ from those found in the native toxins. Future proteomics studies on collected
137 venom are needed to establish the exact chemical identity of the toxins studied here.

138 Synthetic peptides were screened at the five human SSTRs using the PRESTO-Tango β -arrestin
139 recruitment assay (19) (**Fig 1C** and **Fig S1**). Human somatostatin-14 (SST-14) and the previously
140 characterized peptides, consomatin Ro1 and pG1, were included for comparison. The eight new
141 peptides that were tested showed a range of SSTR activity profiles demonstrating that cone snail
142 venoms are a rich source for the discovery of novel ligands of the human SSTRs. For example,
143 Fj2 activates the SSTR₂ and SSTR₄ with EC₅₀ values of 0.20 μ M and 2.0 μ M, respectively, Pr1
144 activates the SSTR₁ and SSTR₂ with EC₅₀ values of 130 nM and 41 nM, respectively, and Oc3
145 shows activity at SSTR₂, SSTR₃, and SSTR₄ with EC₅₀ values ranging from 5.0 nM to 4.2 μ M.
146 Interestingly, although the sequence of Ro2 is distinct from the previously characterized Ro1, the
147 two peptides that are both derived from the same cone snail species, *Conus rolani*, have very
148 similar activity profiles at the human SSTR₁ and SSTR₄. Notably, none of the peptides tested
149 showed any significant activity at SSTR₅, although it should be noted that the peptides were tested
150 for agonism only; thus, we cannot rule out antagonist activity. Of particular interest to us was Fj1,
151 which activated the SSTR₄ with an EC₅₀ value of 22 nM, showing 173-fold selectivity over SSTR₁
152 (EC₅₀ = 3.8 μ M) and no measurable activity at the other three receptor subtypes when tested at
153 concentrations up to 100 μ M.

154

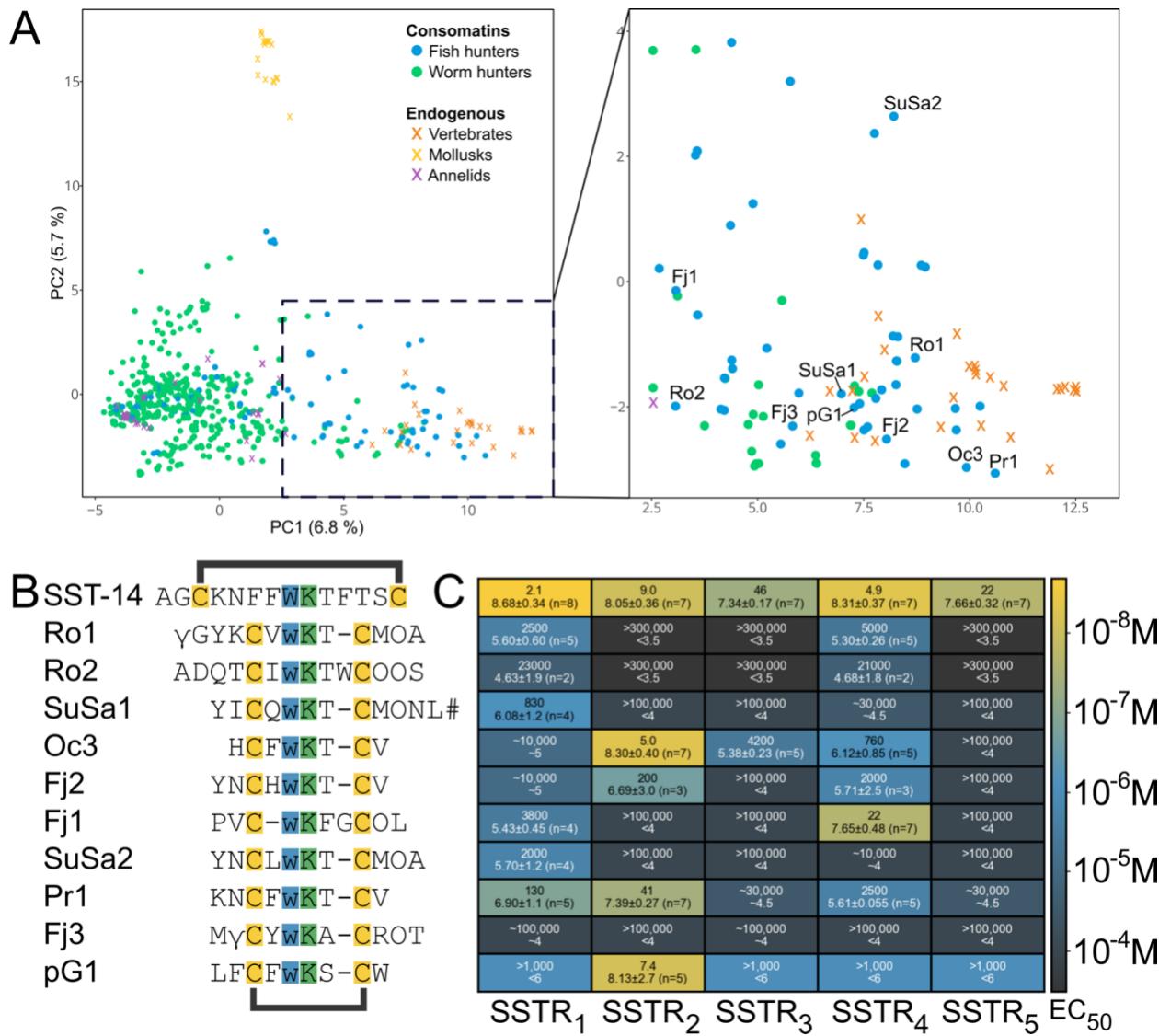


Fig. 1. Computational selection and SSTR profiling of consomatin sequences from a large toxin gene dataset.

(A) Principal component analysis (PCA) of 529 SST-like toxin sequences was used as the basis for selecting lead compounds for SSTR screening. Percentages refer to the proportion of the total variance in the data explained by the given principal component. A subset of the PCA plot, including vertebrate SST-like sequences and selected consomatin sequences, is shown (right box). (B) Synthesized consomatin sequences selected from the PCA plot. Cleavage sites and posttranslational modifications, such as D-Trp, were predicted based on the original somatostatin-like venom peptide, consomatin Ro1. Cysteines forming intramolecular disulfide bonds are shown in yellow and the essential Trp-Lys motif is highlighted in blue and green. Post-translational modifications are indicated by γ = γ -carboxyglutamate, w = D-Trp, O = hydroxyproline, and # = C-terminal amidation. SST-14 is human somatostatin-14. Seven of the listed sequences are from snails of the *Asprella* clade; Ro1 and Ro2 from *C. rolani*, Fj1, Fj2, and Fj3 from *C. fijisulcatus*, and SuSa1 and SuSa2 from *C. sulcatus samiae*, two are from the snails of the *Phasmoconus* clade; Oc3 from *C. ochroleucus*; Pr1 from *C. parius*, while pG1 was predicted from the transcriptome of *C. geographus* from the *Gastridium* clade (16). (C) Heatmap of activity values of selected consomatin sequences tested at the five human SSTRs using the PRESTO-Tango β -arrestin recruitment assay. Rows correspond to the sequences shown in (B). Data are represented as EC₅₀ (top line, nanomolar, two significant digits shown) and pEC₅₀ \pm 95 % confidence intervals (CI95) with the number of independent repeats in parentheses (bottom line, two significant decimals shown). Approximate values (~) or “higher than” (>) and “lower than” (<) are used when we did not obtain full curves within the concentrations tested (see Fig S1 for representative curves).

175

176 **Consomatin Fj1 is a somatostatin “evolog” with a minimized receptor binding motif**

177 Consomatin Fj1 is a ten amino acid long cyclic peptide from *Conus fijisulcatus* that contains four
178 residues inside a disulfide loop and two residues at both its N- and C-termini (Sequence: Pro1-
179 Val2-Cys3-D-Trp4-Lys5-Phe6-Gly7-Cys8-Hyp9-Leu10, **Fig. 2A-B**). The minimized disulfide
180 loop of Fj1 closely mirrors that of the SSTR₂-selective somatostatin drug analogs, such as
181 octreotide and lanreotide. Incorporation of a shortened disulfide loop and a D-Trp in these analogs
182 significantly prolonged their half-lives compared to the native human hormone (9). In contrast to
183 these analogs, which were developed using medicinal chemistry approaches, consomatin Fj1 and
184 other consomatins evolved from an endogenous SST-like signaling peptide following the
185 principles of natural selection. Thus, we refer to these naturally evolved analogs as evologs (16).
186 Notably, despite these distinctively different approaches, the final products exhibit a remarkably
187 high degree of similarity (**Fig. 2B**). One notable difference that distinguishes consomatin Fj1 from
188 other SST analogs and evologs is that the characteristic D-Trp-Lys motif essential for SSTR
189 activation (6) resides immediately following the first cysteine residue (**Fig. 2B**). As discussed in
190 more detail below, these differences partly contribute to the potency and selectivity of the peptide
191 for the SSTR₄.

192

193 **Consomatin Fj1 induces G protein dissociation at the SSTR₄**

194 The analgesic effect of SSTR₄ activation has been proposed to occur via G_{i/o} proteins (15), and is largely
195 mediated by the G $\beta\gamma$ subunits released from pertussis toxin-sensitive G $\alpha_{i/o}$ proteins (20).
196 Therefore, we assessed the activity of Fj1 at the SSTR₄ using a bioluminescence resonance energy
197 transfer (BRET)-based assay that measures GPCR activation by monitoring the association of
198 labeled G $\beta\gamma$ subunits and a G protein-coupled receptor kinase (GRK) fragment, following
199 dissociation of G $\beta\gamma$ subunits from the G α subunit (21). We tested the activity of Fj1 at the SSTR₄
200 using a range of different G α proteins in the G α_i family, including G α_{i1} , G α_{i2} , G α_{i3} , as well as the
201 two dominant splice variants of G α_o , and observed similar potencies for each G α protein (**Fig S2**).
202 As the G α_o type is the most highly expressed type in DRGs (22), the proposed site for SSTR₄-
203 mediated analgesia, we performed subsequent tests with the canonical G α_o sequence (G α_o A). Here,
204 we observed similar overall receptor activation profiles as in the PRESTO-Tango screening assay:
205 potent activation of SSTR₄ (EC₅₀ = 6.0 nM) with only limited activity at SSTR₁ (~30 % activation
206 at 1 μ M) and no activation of the SSTR_{2,3,5} when tested at up to 1 μ M (**Fig 2C**).
207

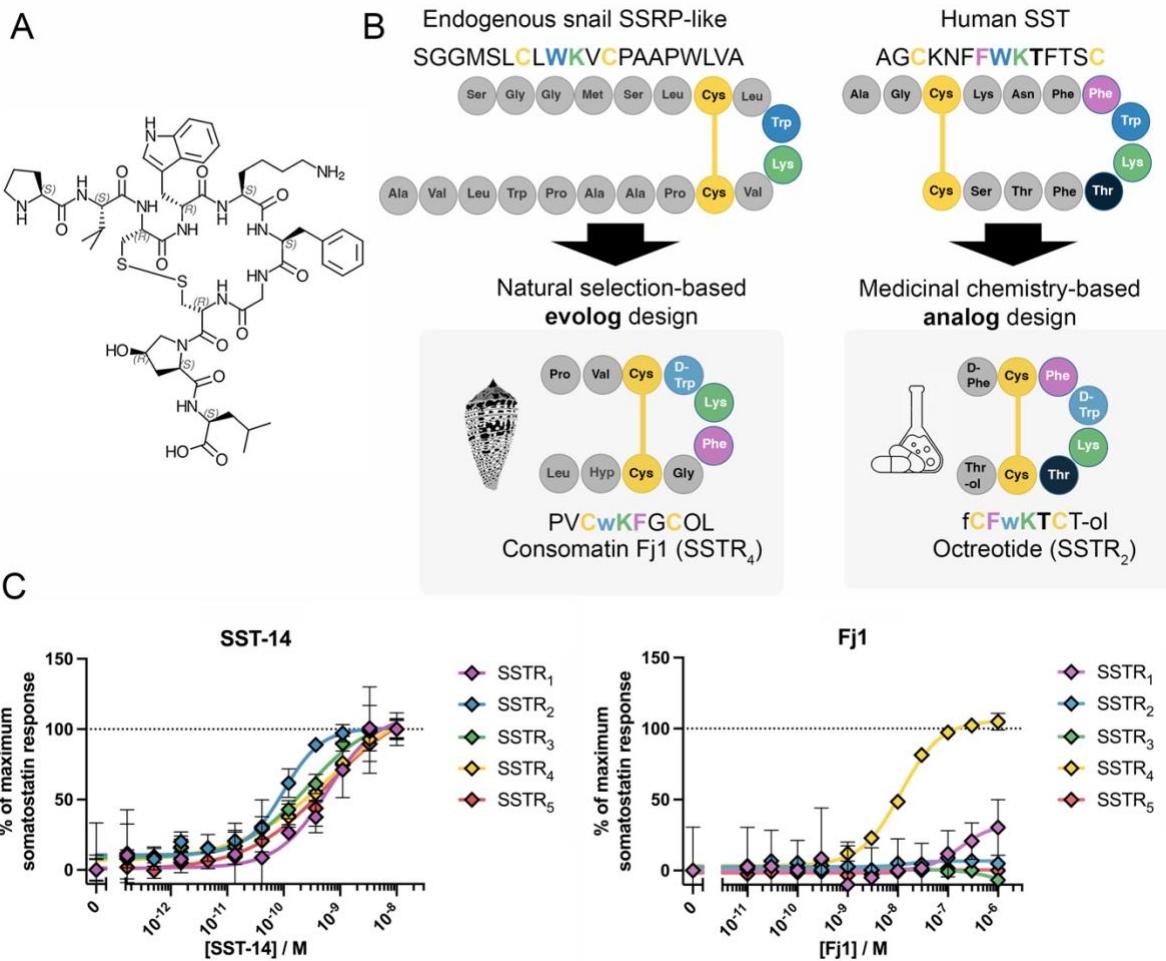


Fig. 2. Consomatin Fj1 is a naturally evolved SSTR4-selective somatostatin evolog.

(A) Chemical structure of consomatin Fj1, a ten-residue long cyclic peptide predicted from the venom gene-encoding exons of *Conus fijisulcatus*. Standard amino acid one or three letter abbreviations are used, extended with O/Hyp = 4-hydroxyproline, w/D-Trp = D-tryptophan, and -ol = C-terminal alcohol group in place of a carboxylic acid group. (B) Schematic comparison of the contrasting origins of consomatin Fj1 and the SST drug analog octreotide, both representing minimized SST scaffolds. Fj1 is a minimized SSTR4-selective evolog that originated from an endogenous SS-like signaling gene, while the SSTR2-selective drug analog octreotide was designed using medicinal chemistry approaches. (C) Representative concentration-response curves for G protein ($G\alpha_o$) dissociation of SST-14 and Fj1 at the five human SSTRs. Fj1 shows nM potency at the human SSTR4, with low potency at SSTR1 and no discernable activity at the other SSTR subtypes at 1 μ M. Error bars represent the standard deviation of two technical replicates.

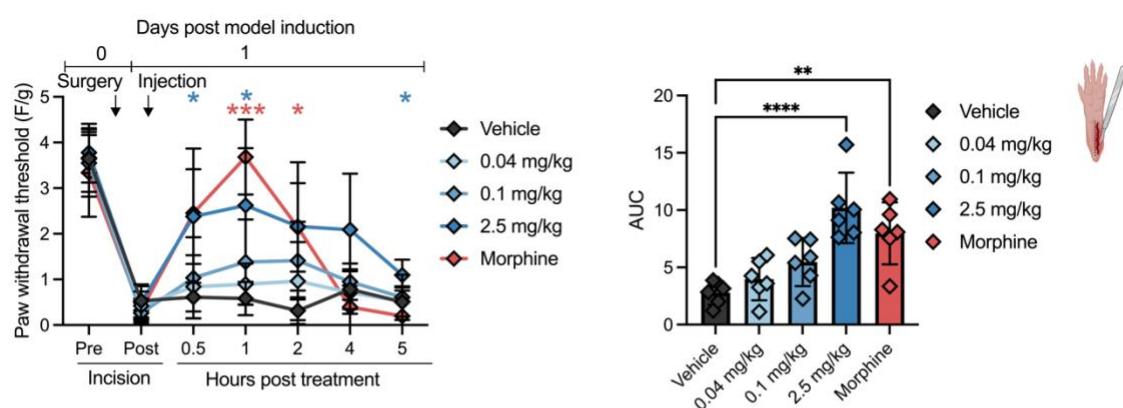
Peripheral consomatin Fj1 administration provides analgesia in a post-operative pain model

Having established that consomatin Fj1 selectively activates the SSTR4 in the PRESTO-Tango and G protein dissociation assays, we next performed a dose response study for potential analgesic action in a model of post-operative pain, the paw incision model (23). C57BL/6J male mice injected intraperitoneally (i.p.) with 0.04–2.5 mg/kg of Fj1 and its effect on post-incision mechanical hypersensitivity was evaluated. Fj1 dose-dependently reduced post-incisional mechanical hypersensitivity, as measured by paw withdrawal thresholds in response to von Frey filament stimulation, with maximal effect observed at the dose of 2.5 mg/kg, and a peak effect observed at 1 h. The positive control used was morphine sulfate (5 mg/kg). While morphine

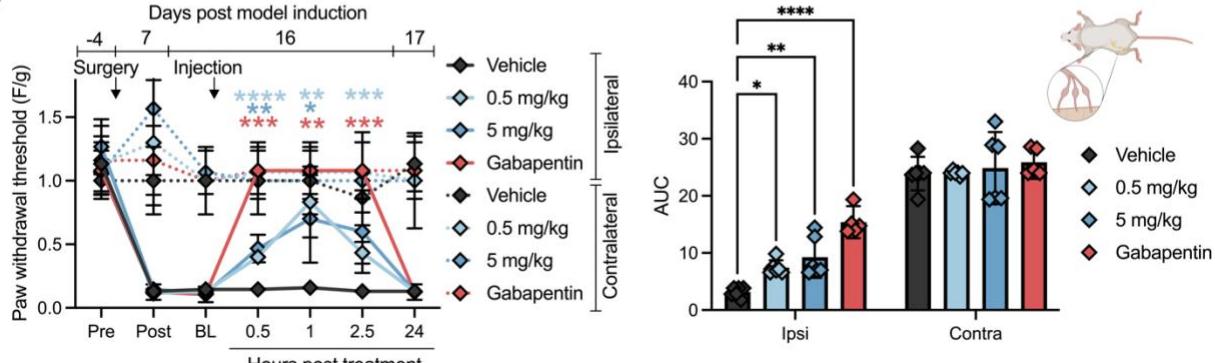
230 exhibited higher peak efficacy than Fj1, the duration of action was shorter. Even at the 5 h time
231 point, the mice treated with 2.5 mg/kg of Fj1 showed an increased paw withdrawal threshold of
232 about 28 % of maximum Fj1 response ($P = 0.0284$, as compared to vehicle). Both in the groups
233 treated with 0.04 mg/kg and with 0.1 mg/kg of Fj1, there was a similar trend as for 2.5 mg/kg,
234 though neither reached a statistically significant difference from the vehicle control at any time
235 point ($P = 0.2936$ and $P = 0.0565$, respectively, at 2 h).
236

237 **Peripheral consomatin Fj1 administration provides analgesia in a neuropathic pain model**
238 Activation of the SSTR4 has previously been shown to provide analgesia in rodent models of
239 neuropathic pain (14, 24-26). Drugs that are effective in post-operative pain may not be efficacious
240 in neuropathic pain states and vice a versa. In general, neuropathic pain is particularly difficult to
241 treat; many current treatment options are only effective in a limited subset of patients and can elicit
242 severe side effects (27-29). To assess whether Fj1 could alleviate the mechanical hypersensitivity
243 associated with neuropathic pain, the peptide was tested in the spared nerve injury (SNI) model of
244 peripheral neuropathic pain using male C57BL/6 mice. The sciatic nerve was injured on one side
245 of the mouse (ipsilateral side), whereas the other side (contralateral side) served as a control. On
246 day 16 post-injury, either 0.5 mg/kg or 5 mg/kg of Fj1, 30 mg/kg of gabapentin, or vehicle was
247 administered by i.p. injection (Fig. 3B). Both doses of Fj1 reduced mechanical hypersensitivity
248 30 min after injection, and this effect was more pronounced after 1 h. For the 0.5 mg/kg dose, the
249 effect persisted at 2.5 h, while for the 5 mg/kg dose, the tendency was the same, albeit not reaching
250 statistical significance ($P = 0.0548$ compared to vehicle). These results demonstrate that Fj1 can
251 provide potent and efficacious analgesia in this model, although with a slower onset and shorter
252 duration of action than 30 mg/kg of gabapentin.
253

A



B



255 **Fig. 3. Consomatin Fj1 provided analgesia in two mouse models of pain.**

256 (A) Post-surgical pain model (paw incision). Plantar incision surgery was performed 24 h post-surgery, and mice were
257 administered morphine sulfate (5 mg/kg), indicated doses of Fj1, or vehicle (normal saline) by intraperitoneal (i.p.)
258 injections, whereafter mechanical hypersensitivity was evaluated at indicated times (left). Values represent mean \pm
259 CI95 from six mice. A summary of the area under the curve is shown (right), where each data point represents one
260 mouse, and the error bars represent CI95. (B) Neuropathic pain model (spared nerve injury, SNI). The peroneal and
261 tibial branches of the sciatic nerve of the left hindleg were ligated, 16 days post-surgery, the mice were administered
262 gabapentin (30 mg/kg), indicated doses of Fj1, or vehicle by i.p. injections, whereafter mechanical hypersensitivity
263 was evaluated at indicated times. Values represent the mean \pm CI95 from five (gabapentin) or six (all other conditions)
264 mice. Statistical analysis was performed as described in the methods section. Asterisks indicate P-values < 0.05 (*),
265 0.01 (**), 0.001 (***)¹, and 0.0001 (****) as compared with the vehicle control.

266 **Molecular dynamics simulation reveals the binding mode of Fj1 at the SSTR₄**

267 To investigate the binding mode of Fj1 at the SSTR₄ and better understand the molecular basis for
268 the peptide's selectivity and potency, we performed molecular dynamics (MD) simulations in the
269 μ s range of Fj1 to the holo-SSTR₄ embedded within a 1-palmitoyl-2-oleoyl-*sn*-glycero-3-
270 phosphocholine (POPC) bilayer. The binding mode of consomatin Fj1 was stabilized and almost
271 exclusively characterized through the side chain interactions of D-Trp4 and Lys5 with receptor
272 residues (Fig. 4A). D-Trp4 buried deep within the hydrophobic core cavity formed by the side
273 chains of Leu123^{3x29}, Met130^{3x36}, Phe131^{3x37}, Ile181^{4x61}, Leu200^{45x52}, and Phe275^{6x51} (superscripts
274 denote the GPCRdb numbering scheme outlined by Isberg *et al.* (30)). Interestingly, D-Trp4
275 occupied the same binding pocket as the corresponding L-Trp of SST-14 observed in the recent
276 cryogenic electron microscopy (cryoEM) structure of SSTR₄ (PBD ID: 7XMS) (31) (Fig. 4B).
277 Lys5 forms a salient salt bridge with the side chain of Asp126^{3x32}, also mirroring the conformation
278 and interactions of the corresponding Lys9 in SST-14. Additionally, Lys5 in Fj1 also demonstrated
279 transient yet notable hydrogen bonding to the side chain of Ser300^{7x41} during simulation. On the
280 opposite face of the side chain, Lys5 maintains a side-by-side stacking arrangement against the
281 indole ring of D-Trp4, which is also present in the monomeric structure of Fj1, and is characteristic
282 of SST receptor binding and activation (6). Both D-Trp4 and Lys5 demonstrated limited sampling
283 of other conformations throughout the simulation.

284 In contrast to the persistent orientation adopted by D-Trp4 and Lys5, residues bordering and
285 outside the cyclic core, Pro1, Cys3, Phe6, Cys8, Hyp9, and Leu10 (i.e., with the exception of Val2)
286 displayed considerable mobility, demonstrating limited persistent contacts describing the ligand-
287 bound orientation (Fig. 4C-D). Likewise, residues outside of the conserved pharmacophore (Phe7-
288 Trp8-Lys9-Thr10) in SST-14 do not form significant side chain interactions with SSTR₄, as has
289 been observed with SSTR₂ (31), instead forming more backbone and intramolecular interactions
290 (most notably between Phe6-Phe11). The short cyclic core (four residues in Fj1, compared to ten
291 residues in SST-14) does not protrude significantly beyond the binding pocket and aligns well with
292 the smaller four-residue pseudo-core formed between Phe6 and Phe11 of the SSTR-bound
293 structure of SST-14. Val2 presents a somewhat smaller hydrophobic interface within the helical
294 core, packing between residues Val212^{5x40} of helix 5 and Leu283^{6x59} of helix 6, occupying a
295 position similar to that of Phe7 in SST-14. These simulations showed that Fj1 binds SSTR₄ in a
296 manner similar to SST-14, particularly in regard to the Trp-Lys motif, and that residues outside of
297 this could be amenable to modification while retaining SSTR₄ binding.

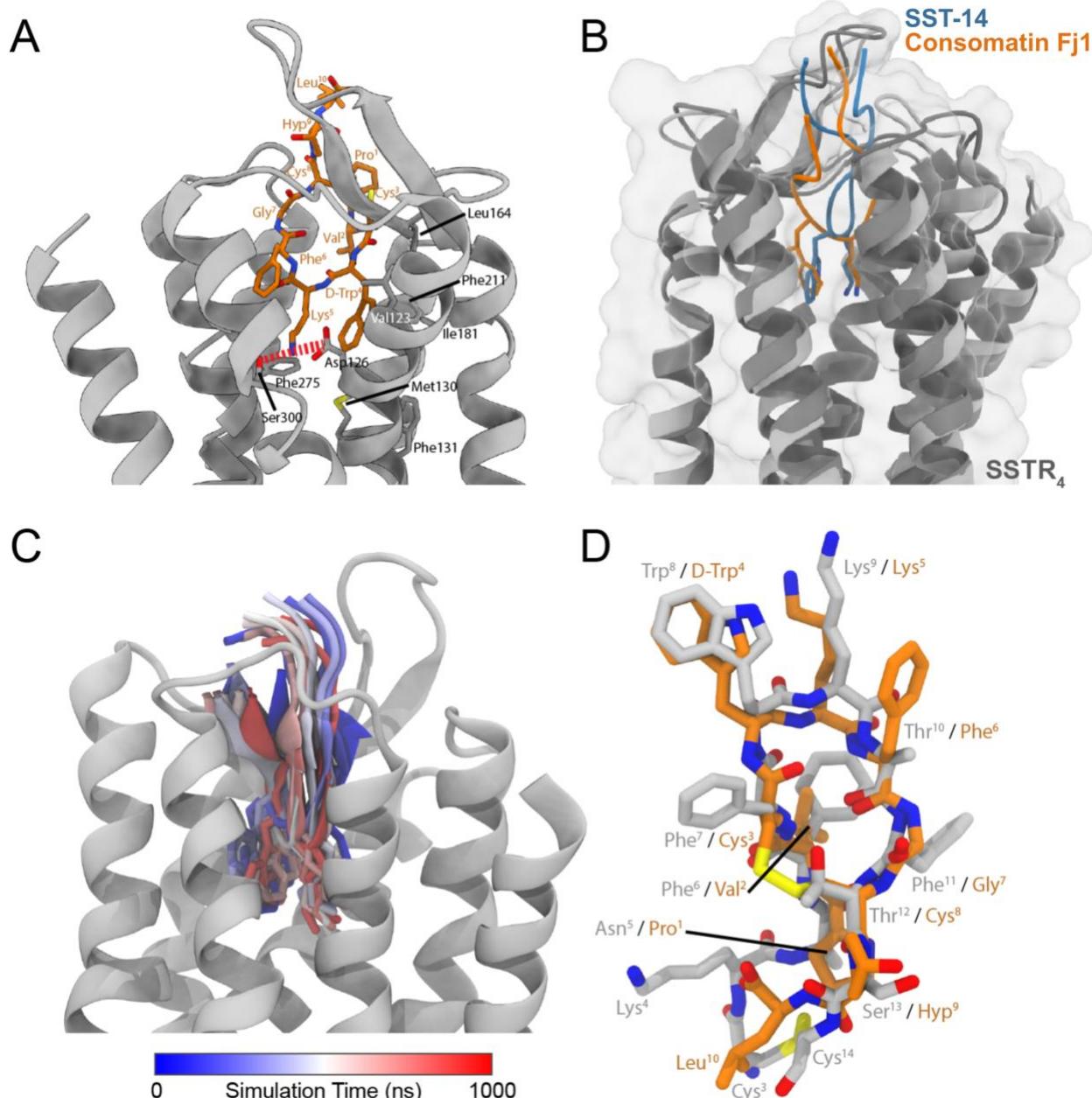


Fig. 4. Molecular Dynamics (MD) simulations reveal contacts between Fj1 and SSTR4.

(A) Representative orientation of Fj1 (orange stick representation) in complex with SSTR4 (gray ribbon representation). The membrane present within the simulation was removed for clarity. The interactions made by Fj1, which facilitated binding, are labelled. The representative binding mode was obtained by clustering all replicate trajectories, with a model representing the largest cluster displayed. (B) Overlay of Fj1 (orange) and SST-14 (blue, PDB 7XMS) in complex with SSTR4 (dark gray). (C) The orientation of Fj1 throughout a representative 1 μ s simulation is shown with 10 snapshots colored from blue to white to red. (D) Representative orientation of Fj1 (orange) from the simulation when in complex with SSTR4, overlaid with SST-14 in complex with the same receptor construct (gray, PDB 7XMS).

Analog design provides improved agonists and scaffolds for future drug development

To assess the molecular determinants of SSTR4 potency and selectivity in the G protein dissociation assay, we first performed alanine scanning of the four amino acids in the loop that

313 were buried in the binding pocket of the receptor (**Fig. 5**). As expected, mutating key residues
314 known to be important for the binding of somatostatin (6), Trp4 (Fj1A1) and Lys5 (Fj1A2), to Ala
315 markedly decreased potency by 5000-fold and 730-fold, respectively. The mutation of Phe6
316 (corresponding to Thr10 in SST-14, see **Fig. 4D**) to Ala (Fj1A3) also decreased the potency by
317 630-fold. However, mutating either Gly7 to an Ala (Fj1A4) or deleting it, thereby shortening the
318 loop (Fj1A5), resulted in a slight increase in potency at SSTR₄ (6.2-fold and 65-fold, respectively)
319 with no discernable increase in potency at other SSTRs. Next, we assessed the role of the predicted
320 posttranslational modifications. Changing D-Trp and L-Hyp, to their unmodified equivalents, L-
321 Trp and L-Pro (Fj1A6 and Fj1A7, respectively), had virtually no effect on the SSTR₄ potency,
322 although both showed an increase in SSTR₁ potency. Another epimerization, the substitution of L-
323 Phe6 to D-Phe (Fj1A8), on the other hand decreased the potency at SSTR₄ by 42-fold. Since the
324 MD simulations suggested that residues outside the cyclic core did not form significant and salient
325 interactions with the receptor, we tested the “minimal core” peptide Cys3-D-Trp4-Lys5-Phe6-
326 Gly7-Cys8 (Fj1A9). This decreased the potency by 47-fold, while deleting only the N-terminus
327 (Fj1A10) resulted in a 13-fold decrease in potency. However, deletion of only the C-terminus
328 (Fj1A11) resulted in an analog that was virtually equipotent at the SSTR₄.
329 Recently, we identified consomatotropin pG1 as a potent and selective agonist of the SSTR₂ (16). To
330 investigate whether determinants from pG1 could be used to improve the potency of Fj1, we
331 designed a hybrid analog combining the cyclic core residues of Fj1 with the amino acids flanking
332 the pG1 core (Fj1A12). This hybrid analog showed a 92-fold increase in potency at the SSTR₄,
333 although with a corresponding increase at the SSTR₁ and SSTR₅. Interestingly, this did not lead to
334 increased potency at the SSTR₂. Introducing only the N-terminus of pG1 (Fj1A13) increased
335 potency by 11-fold at the SSTR₄, with SSTR₁ activity similar to that of Fj1A12. With the
336 suggestion from Fj1A11, that the Fj1 C-terminus was not required, we next tested the effect of
337 adding the C-terminus from pG1, namely the single Trp residue, to Fj1. This analog (Fj1A14)
338 showed an 85-fold increase in potency at the SSTR₄, with no enhanced activity observed at the
339 other receptor subtypes, including the SSTR₁. Given the apparent redundancy of Gly7, we
340 evaluated whether replacing Gly7 with hydrophilic amino acids to decrease the hydrophobicity of
341 the peptide could be tolerated. Substituting Gly9 with Ser (Fj1A15) or Asn (Fj1A17) slightly
342 increased the potency at SSTR₄ (14-fold and 6.8-fold, respectively), whereas a Thr at this position
343 (Fj1A16) retained the SSTR₄ potency of Fj1, while showing a slight increase in SSTR₁ activity.
344 Based on these observations, we next assessed whether the increased potency of Fj1A14 could be
345 combined with the polar substitutions of Gly7. While a Ser in place of Gly7 (Fj1A18) was well
346 tolerated at the SSTR₄, both the Thr and Asn substitutions (Fj1A19 and Fj1A20, respectively)
347 showed a slight decrease in SSTR₄ potency compared to Fj1A14. However, all three double
348 substitutions increased SSTR₁ activity, thereby decreasing selectivity.
349 These results show that while Fj1 was the most potent and selective SSTR₄ peptide agonist
350 identified from venom-encoding genes, there are clear opportunities for further optimization of the
351 peptide’s potency, selectivity, and physiochemical properties such as hydrophilicity.
352

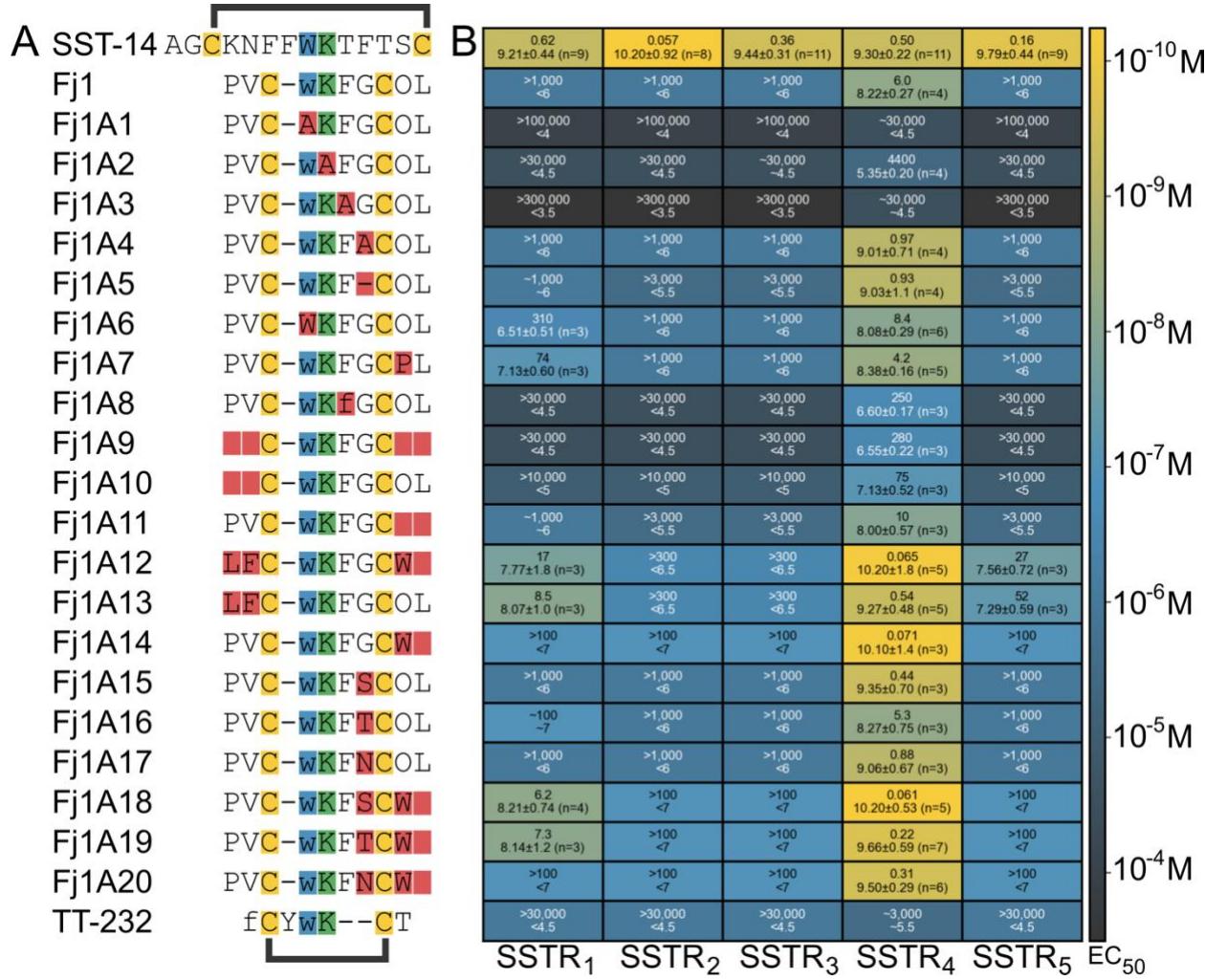


Fig. 5. Analog design and testing identified improved ligands for SSTR₄-selective drug development.

(A) Identifiers and sequences of SST-14, Fj1, and the analogs of Fj1. Sequences (left) show the Cys residues forming the intramolecular disulfide in Fj1 and its analogs highlighted in yellow. For all sequences, the Trp residue is shown in blue, the Lys residue in green, and all modifications in the analogs, as compared to Fj1, are highlighted in red. Sequences use standard amino acid one-letter abbreviations extended with O=hydroxyproline and lower-case letters denoting D-amino acids. (B) Heatmap showing the potency of Fj1 and its analogs at the five human SSTRs in the G protein dissociation assay using Gα_o. The text in each cell represents the EC₅₀ value (first line, nanomolar, two significant digits) and the pEC₅₀ ± CI95 (two significant decimals) with the number of independent repeats for each in parentheses (second line). Approximate values (~) or “higher than” (>) and “lower than” (<) are used when we did not obtain full curves within the concentrations tested (see Fig S3 for representative curves). The known SSTR₁ and SSTR₄ peptide agonist, TT-232 was also tested. However, we note that we could not dissolve it at sufficiently high concentrations to fully test this peptide in the assay used).

353
354

355
356
357
358
359
360
361
362
363
364
365
366

367

DISCUSSION

Cone snail venoms represent a rich source for the discovery of biomedical tools and drug leads. Among these, the cone snail peptide ω-MVIIA derived from the venom of the magician cone, *Conus magus*, is arguably the most prominent example (32). MVIIA, also known as ziconotide, inhibits pain signals by selectively blocking the N-type calcium channel, Ca_v2.2, expressed in the CNS (33). Ziconotide not only revealed the existence of the Ca_v2.2 channel, but also delineated its

374 role in pain signaling and became an approved drug for the treatment of chronic intractable pain
375 in 2004 (34). Another analgesic cone snail peptide toxin that modulates pain signals in the CNS
376 and has proven analgesic effects in humans is contulakin-G, also known as CGX-1160, derived
377 from the venom of the geographer cone, *Conus geographus* (35). Contulakin-G shares sequence
378 similarity with the human neuropeptide, neuropeptides. Although the exact mode of action of
379 contulakin-G remains unknown, recent studies suggest that, *in vivo*, this peptide acts as an agonist
380 of the neuropeptide 2 receptor (NTSR2), leading to the inhibition of the R-type calcium channel,
381 Ca_v2.3, expressed in the spinal cord (36). Contulakin-G entered a phase Ia clinical study for the
382 treatment of spinal cord injury related pain, one of the most difficult to treat type of pain conditions.
383 Despite its significant effects in reducing pain, the clinical development of contulakin-G was
384 discontinued when the company developing this peptide shut down (34).

385 Our recent discovery of consomatin Ro1, an analgesic toxin from *Conus rolani* that activates the
386 human SSTR_{1,4}, provided another lead compound for pain from cone snail venom. Unlike MVIIA
387 and contulakin-G, consomatin Ro1 targets receptors expressed in the peripheral nervous system
388 and has no analgesic effect when administered centrally in mice (16). This peripherally restricted
389 site of action can be advantageous in two different ways. First, the delivery of the drug will not
390 require invasive techniques such as intrathecal or epidural puncture. Next, the likelihood of CNS-
391 related side effects such as dependence or respiratory depression are significantly reduced. Here,
392 we describe the discovery and preclinical evaluation of another SSTR₄ agonist with superior
393 potency and selectivity, consomatin Fj1, derived from *Conus fijisulcatus*.

394 Based on their relatedness, cone snails can be grouped into approximately 44 distinct clades (37).
395 Snails belonging to the same clade typically share similar venom compositions and hunting
396 behaviors, although exceptions exist. Notably, *C. fijisulcatus* and *C. rolani* both belong to the
397 *Asprella* clade, one of eight known lineages of fish hunting cone snails (37). Unlike most other
398 cone snails that inhabit shallow coastal waters, *Asprella* snails live offshore at depths of 60–250 m
399 (38). *Asprella* snails use an unusual hunting strategy characterized by venom injection followed
400 by a slow onset of venom action. We previously observed predation by *Asprella* snails in captivity,
401 which took between 15 min and 3 h from envenomation until the prey was incapacitated¹⁶. We
402 hypothesized that this “ambush-and-assess” hunting strategy is accompanied by the evolution of
403 toxins that inhibit the prey’s escape response. Furthermore, the slow onset of action suggested that
404 the venom of *Asprella* snails may not be dominated by classical ion channel modulators but may
405 contain toxins that target GPCRs. Here, to discover novel ligands of human SSTRs, we used PCA
406 to identify consomatins that group with human SST and its related peptides and prioritized toxins
407 that belonged to species of the *Asprella* clade. In the small set of *Asprella* consomatins we
408 synthesized and tested, all had activity at the human SSTRs, with a notable bias towards activation
409 of the SSTRs in group 2 (SSTR₁ and SSTR₄). Only a single *Asprella* consomatin showed notable
410 activity at a group 1 SSTR at the concentrations tested (Fj2 at the SSTR₂). There is emerging
411 evidence for the role of agonists group 2 SSTRs in inhibiting pain and inflammation in rodent
412 models of disease, as well as recent clinical data from a phase II clinical trial showing that an
413 SSTR₄ agonist is effective for treating peripheral diabetic neuropathy (39). Given the observed
414 predominance of SSTR₁ and SSTR₄ activity of consomatins identified from only five *Asprella*
415 species of at least 18 known (40, 41), there are likely to be more consomatins of interest for
416 mammalian pain modulation in this clade of fish hunting cone snail (42).

417
418 Because of its high potency and selectivity for the SSTR₄, we focused our subsequent structure-
419 activity studies on consomatin Fj1. An unusual feature of consomatin Fj1 is its unique sequence

420 motif, with the canonical Trp-Lys dyad immediately following the first Cys residue. To better
421 understand the role of this dyad and the overall mode of action of the peptide, we performed MD
422 simulations combined with functional studies on select analogs. MD simulations of Fj1 and
423 comparison with the known structure of SST-14 bound to the SSTR₄ revealed a similar binding
424 mode between the toxin and the human peptide. Where SST-14 utilizes the two Phe residues in
425 positions 6 and 11 to essentially create a T-shaped π - π interaction with Phe7, Trp8, Lys9, Thr10
426 in between, in Fj1, these aromatic residues are replaced with Cys residues, creating a covalent
427 bond linking the corresponding residues, and retaining the four-residue “loop” that SST-14
428 transiently inhabits in its SSTR-bound form (see **Fig. 4B,D** as well as work by Robertson *et. al*
429 and Zhao *et al.* (31, 43)). The D-Trp-Lys motif remained in the lower binding pocket in the Fj1
430 simulation, where residues interacted largely with the same SSTR₄ side chains as the
431 corresponding residues in SST-14. Notably, the D-Trp of Fj1 inhabits the same binding pocket as
432 the corresponding L-Trp of SST-14, which likely explains why we observed little effect of
433 changing D-Trp to L-Trp in (Fj1A6). While we did observe a slight difference in SSTR₁ potency,
434 the L-to-D epimerization that was previously observed in consomatin Ro1, and predicted in other
435 consomatins, including Fj1, likely plays a larger role in stabilizing the minimized fold and
436 providing resistance to proteolytic cleavage, as this modification is known to evade recognition by
437 endogenous enzymes and has been introduced in other peptide drugs, including SST analogs (44).
438 To identify the minimum binding motif at the SSTR₄ we tested the “core” motif of Fj1 linked by
439 the disulfide loop (Cys-D-Trp-Lys-Phe-Gly-Cys; Fj1A9). This analog retained both activity and
440 selectivity, albeit being approximately 47-fold less potent at SSTR₄. In line with this observation,
441 residues outside this core did not show notable static side chain-side chain interactions with the
442 SSTR₄ throughout the simulation. However, analog screening suggested a differential role for the
443 N- and C-terminal residues in binding. Removing the two N-terminal residues which, due to the
444 “shifted” D-Trp-Lys motif in relation to the cysteines, are situated closer to the binding pocket
445 surrounded by TM5 and TM6 (Fj1A10), showed a potency loss of 13-fold. In comparison,
446 removing the two C-terminal residues (Fj1A11), which protrude from the binding pocket and are
447 surrounded by the flexible ECL2 and ECL3 in the MD simulation, barely changed the potency at
448 the SSTR₄. As most of the polar interactions with residues outside the core of Fj1 involve backbone
449 residues, these findings suggest the possibility of introducing additional modifications outside the
450 core. Indeed, adding the C-terminal Trp from consomatin pG1, a potent and selective agonist of
451 the SSTR₂, to the Fj1 scaffold resulted in a significant increase in potency while retaining
452 selectivity (Fj1A12). This surprising finding demonstrates that there is significant room for
453 improving the potency of Fj1 at the human SSTR₄ without compromising the selectivity of the
454 peptide. Additionally, analog screening demonstrated that Gly7, which is located within the
455 disulfide loop, is amenable to deletion (Fj1A5) and substitutions (Fj1A15-Fj1A20) that can retain
456 or even improve potency while introducing hydrophilic side chains. However, some of these come
457 at the expense of decreased selectivity for SSTR₄ over SSTR₁.
458

459 The small molecule SSTR₄ agonist, J-2156, which has been reported to be about 360-fold selective
460 for SSTR₄ over SSTR₁ and 390-fold over SSTR₅ in binding assays (45), has been widely used in
461 the literature (14, 15, 24, 26, 45, 46). However, peptide agonists that can distinguish between
462 SSTRs in group 2 (SSTR₁ and SSTR₄) have not been previously reported. The most widely used
463 peptide agonist at the SSTR₄ is the heptapeptide, TT-232, which suffers from low potency and a
464 6.5-fold reported selectivity over SSTR₁ (47). Using the venom peptide Fj1 as an inspiration, this

465 study provides several analogs with significantly improved potency and selectivity over previously
466 described peptide agonists and suggests additional opportunities for further optimization.
467 Moreover, having a potent and selective peptide agonist of the SSTR₄ expands the repertoire of
468 existing drug leads for this target from small molecules to peptides. Peptides, especially those that
469 have evolved by nature, often display better selectivity and fewer off-target effects or
470 complications from hepatic metabolism (48). While the *in vivo* half-life of peptides is often limited
471 by rapid renal clearance, the design of peptide conjugates can significantly increase the duration
472 of action, as seen with many recent peptide therapeutics that now have weekly, instead of daily,
473 dosing regimens (49). The importance of these advantages is reflected by the rapidly increasing
474 number of approved peptide therapeutics in recent years (50, 51). Here, we show that the small
475 cyclic peptide Fj1 can efficiently alleviate the mechanical hypersensitivity associated with two
476 types of pain in rodent models of the disease.

477
478 The incision model of postoperative pain gives rise to a complex pain response dependent on
479 multiple components, sensitizing both C- and A δ -fibers that innervate the area around the injury
480 (52). The expression pattern of the SSTR₄ in C- and A δ -fibers is not well established, but
481 differences here, along with the central effects of opioids, could help explain the difference in the
482 maximal response observed between Fj1 and morphine. The expression of SSTR₄ is upregulated
483 in inflammatory models (53), and activation can inhibit inflammation induced by either mustard
484 oil or lipopolysaccharide (10, 54, 55). In response to injury, inflammatory responses are generally
485 observed within 2 h (56, 57). SSTR₄-mediated inhibition of inflammation could help explain the
486 seemingly longer-lasting effect of Fj1 compared to morphine, where the later stage effect might,
487 in part, be due to a reduction in the post-injury inflammatory response that would otherwise
488 hypersensitize the pain response (58). However, it is also possible that the effects are due to a
489 slower off-rate at the receptor, or simple pharmacokinetic effects. A thorough examination of these
490 effects is outside the scope of this study.

491 Similar to the effect of Fj1 in the paw incision model, the maximum observed effect of a single
492 i.p. dose of Fj1 in the SNI model was also observed at the 1 h time point for both doses used. At
493 this time point, the 5.0 mg/kg dose did not elicit a larger effect than the 0.5 mg/kg, suggesting that
494 0.5 mg/kg was a saturating dose in this model. At the 2.5 h timepoint there was a tendency for a
495 more pronounced effect for the higher dose, which is likely a pharmacokinetic effect reflecting
496 clearance. Our preliminary investigations into the analgesic effects of Fj1 demonstrated that this
497 peptide can effectively alleviate pain like behavior in two mechanistically distinct mouse models.
498 Future studies on Fj1 and its analogs in additional pain models and species will provide insights
499 into the versatility of these new SSTR₄ agonists and likely inform on the design of additional
500 analogs with improved *in vivo* potency and efficacy profiles.

501
502 Our study has several limitations. While the MD simulations of Fj1 confirmed our experimental
503 data and supported our analog screen, an experimental structure is needed to further verify the exact
504 nature of the binding determinants. Furthermore, although we report various potent and selective
505 SSTR₄ agonist these compounds will likely require optimization of their pharmacokinetic profiles
506 before advancing to clinical approval. Finally, while Fj1 and other SSTR₄ agonist are capable of
507 alleviating pain in rodent models, clinical validation of the SSTR₄ is incomplete. Recent phase 2
508 clinical trials on a small molecule SSTR₄ agonist provided preliminary target validation for
509 diabetic neuropathy, while trials for osteoarthritis and chronic lower back pain showed no effect
510 over placebo (39). No clinical trials for therapeutics targeting the SSTR₄ for postoperative pain

511 have been done. As such, which animal models will ultimately translate to humans when targeting
512 the SSTR₄ is yet unknown.

513
514 In conclusion, the discovery of Fj1 and other cone snail peptides with distinct activity profiles at
515 the human SSTRs expands our knowledge of the chemical and pharmacological diversity of the
516 compounds evolved by these marine predators. Given the diversity of somatostatin evolgs
517 identified, we strongly anticipate that future functional characterization of these peptides will
518 continue to provide a stream of novel ligands of the SSTRs and potentially other GPCRs with
519 unique selectivity and potency profiles. Our findings regarding the broad use of somatostatin
520 evolgs in diverse cone snail species further suggest the existence of similar peptides in other
521 venomous animals. Finally, the apparent richness of group 2 SSTR-targeting peptides in *Asprella*
522 snails highlights the usefulness of this clade in the discovery of novel SSTR₄-targeting analgesic
523 leads.

524

525 MATERIALS AND METHODS

526 Study design

527 This study consists of preclinical call-based and rodent models, as well as computational
528 approaches, designed to investigate the potential for identifying lead compounds targeting the
529 human somatostatin system in the venom of marine cone snails, as well as the suitability of an
530 identified peptide as a lead for postoperative and neuropathic pain conditions. The number of
531 replicates and repeats are indicated and all statistical tests, curve fitting, and other data analysis are
532 described in the Materials and Methods section. All animals randomly assigned to their respective
533 treatment groups animal and the experimenters performing measurements were blinded to the
534 treatment. The number of animals was chosen by the group performing the experiments, based on
535 their experience, to ensure sufficient statistical power.

536

537 G protein dissociation assay

538 HEK293 cells (ATCC) maintained in DMEM (Gibco) supplemented with 10% FBS (Biowes,
539 Nuillé, France) and 100 U/mL penicillin / 100 mg/mL streptomycin (Gibco) (growth medium)
540 were maintained in a water jacketed 5 % CO₂ incubator and passaged every 2-3 days using
541 trypsin/EDTA (Gibco). 1E6 cells were seeded into 6 well plates. The following day, the medium
542 was changed, and the cells transfected by adding 0.33 µg of SSTR construct, 0.66 µg of G α
543 protein, 0.33 µg Venus 156-239-G β , 0.33 µg Venus 1-155-G γ , and 0.33 µg masGRK3ct-Nluc
544 DNA into 27 µL of 150 mM NaCl, and 12 µL of 100 mg/mL linear polyethyleneimine (PEI) at
545 molecular weight 25 kDa (Polysciences, Warrington, PA, USA) into 38 µL of a 150 mM NaCl
546 solution. The two tubes were mixed and incubated for 10-15 min before addition to the cells. The
547 following day, 5E4 cells per well were seeded in 100 µL of growth medium into poly-D-lysine-
548 coated white 96 well OptiPlates (PerkinElmer). The following day, the cells were washed in 80 µL
549 of HBSS supplemented with 20 mM HEPES, 1 mM CaCl₂, and 1 mM mgCl₂ (assay buffer), and
550 80 µL of assay buffer supplemented with 0.1 % BSA (stimulation buffer) was added. 10 µL of a
551 1:250 dilution of furimazine (Promega) in stimulation buffer was added, and after 120 s, 10 µL of
552 a 10x solution of the compounds in stimulation buffer was added. 180 s after furimazine addition,
553 the plate was placed in a SpectraMax iD5 and read using a program consisting of 60 s of shaking,
554 followed by 500 ms integration reads with a 485/20 nm filter, followed by a 535/25 nm filter.

555

556 **Animal ethics**

557 All animal studies were approved by the Institutional Animal Care and Use Committee (IACUC)
558 at the University of Arizona and were conform to the guidelines for the use of laboratory animals
559 of the National Institutes of Health (NIH publication no. 80–23, 1966), or in accordance with the
560 guidelines of the Danish Animal Experimentation Inspectorate (permission number 2021-15-
561 0201-01036) in a fully AAALAC-accredited facility, under the supervision of the local animal
562 welfare committee at the University of Copenhagen.

563

564 **Paw incision model of post-operative pain**

565 Male C57Bl/6j mice (6–8 weeks old, The Jackson Laboratory, Bar Harbor, ME) were housed one
566 to six per box (single housing due to fighting) with standard bedding and ad libitum access to
567 standard chow and tap water. Mice were kept in 12 h/12 h light/dark cycles and left to habituate
568 for one week prior to testing. To analyze post-surgery acute incisional hypersensitivity, a plantar
569 incision model was used as described by Brennan *et al.* (59). Briefly, a 0.5 cm long incision, from
570 the heel toward the toes, was made through the skin and fascia of the plantar aspect of the left hind
571 paw, including the underlying muscle. The plantaris muscle was then elevated and longitudinally
572 incised, leaving the origin and insertion intact. After hemostasis with gentle pressure, the skin was
573 closed with two mattress sutures of 5-0 nylon on a curved needle.

574

575 **Measurement of tactile sensory thresholds for post-operative pain model**

576 Tactile sensory thresholds were assessed by measuring the withdrawal response to probing the
577 plantar surface of the hind paw with a series of calibrated fine filaments (von Frey). Each filament
578 was applied perpendicular to the plantar surface of the paw of mice held in suspended wire mesh
579 cages. The “up and down” method was used to identify the mechanical force required for a paw
580 withdrawal response. The size range of stimuli was between 2.44 (0.4 mN) and 4.56 (39.2 mN).
581 The starting filament was 3.61 (3.9 mN). The filament was placed perpendicular to the skin with
582 a slowly increasing force until it bent; it remained bent for approximately 1 s and was then
583 removed. Data were analyzed using the nonparametric method of Dixon, as described by Chaplan
584 *et al.* (60). The results are expressed as the mean withdrawal threshold that induced a paw
585 withdrawal response in 50 % of the animals.

586

587 **Spared nerve injury (SNI) model of neuropathic pain**

588 C57BL/6Nrj male mice, approximately 10 weeks of age (Janvier, France), were housed 1–6 per
589 box (single housing due to fighting) with standard bedding and ad libitum access to standard chow
590 (Altromin 1342, Brogaarden, Denmark) and tap water. Mice were kept in 12 h/12 h light/dark
591 cycles (experiments performed during their inactive phase) and left to habituate to the facility for
592 one week prior to testing. Mice were anesthetized with 2 % isoflurane gas followed by
593 subcutaneous injection of buprenorphine (0.1 mg/kg, Department of Experimental Medicine,
594 University of Copenhagen (UCPH)), and SNI surgery was performed on the left hind leg as
595 follows. The skin on the lateral surface was incised for approximately 0.5 cm between the hip and
596 knee, followed by cutaneous application of a single drop of a mixture of 10 mg/kg lidocaine and
597 5 mg/kg bupivacaine (Department of Experimental Medicine, UCPH) before lengthwise division
598 of the biceps femoris muscle, leading to exposure of the three branches of the sciatic nerve. The
599 sural branch was left intact, whereas the peroneal and tibial branches were ligated with a single
600 surgical knot and axotomized distal to the ligation. Wounds were closed with surgical glue, and

601 animals were monitored daily for signs of stress or discomfort, but in all cases, recovered
602 uneventfully. Immediately post-surgery, the mice were administered carprofen (5 mg/kg,
603 Department of Experimental Medicine, UCPH). On day 16 post-surgery, mice were injected i.p.
604 with a volume of 10 μ L/g of the indicated doses of Fj1, 30 mg/kg of gabapentin (Sigma Aldrich),
605 or vehicle (phosphate buffered saline).

606

607 **Measurement of tactile sensory thresholds for SNI model**

608 For the SNI experiments, von Frey filaments ranging from 0.04 to 2 g (g = gram-forces) (0.04,
609 0.07, 0.16, 0.4, 0.6, 1.0, 1.4, 2.0) were used to determine the mechanical paw withdrawal threshold
610 (PWT). Filaments were applied in ascending order to the frontolateral plantar surface of the hind
611 paws innervated by the sural nerve. Mice were placed in red PVC plastic cylinders (8 (\varnothing) \times 7.5
612 (h) cm) on a wire mesh and allowed to habituate for a minimum of 20 min prior to the experiment
613 initiation. Each von Frey hair was applied five times with adequate resting periods between each
614 application, and the number of withdrawals was recorded. The withdrawal threshold was
615 determined as the von Frey filament eliciting at least three positive trials out of five applications
616 in two consecutive filaments. A positive trial was defined as a sudden paw withdrawal, flinching,
617 and/or paw licking induced by the filament. The animals were habituated to the experimental room
618 for a minimum of 60 min before the initiation of the experiment, and the experimenter (female)
619 was blinded to the treatment groups.

620

621 **Structural Preparation**

622 Modelling of consomatin Fj1 at the SSTR4 made use of the X-ray crystal structure of consomatin
623 Ro1 (PDB: 7SMU) (16), and the cryoEM structure of the SSTR4 in complex with the G_{i1} subunit
624 and SST-14 ligand (PDB: 7XMS) (31) as modelling templates. Because of the sequence similarity
625 of consomatin ligands and SST-14, specifically the core Trp-Lys receptor-binding motif
626 mimicking the broader conserved Phe-Trp-Lys-Thr residues present within SST ligands, Fj1 was
627 assumed to bind the same orthosteric binding pocket as the endogenous ligands. Modeller (v.10)
628 utility was used to create complexes of ligands bound to the SSTR4 binding pocket, generating 50
629 models with the top model assessed using the Modeller objective function to seed subsequent
630 simulations. Due to the absence of N-terminal SSTR4 residues present within the cryoEM
631 structure, only residues Met47-Phe322 were included within the final complex, with the G_{i1}
632 subunit removed. Non-standard amino acids hydroxyproline (Hyp9) and D-tryptophan (D-Trp4)
633 were included, with parameters assigned from the CHARMM36m force field. The resulting
634 complex was inserted into a model POPC membrane using the CHARMM-GUI server and
635 solvated in a box of CHARMM-modified TIP3P (mTIP3P) water, extending $> 10 \text{ \AA}$ beyond all
636 protein atoms. The resulting bilayer spanned an area of $100 \times 100 \text{ \AA}^2$, with a total of 130 lipids
637 each in the upper and lower leaflets. Sodium and chloride ions were included to neutralize the
638 system and attain an ionic strength of 0.1 M.

639

640 **Molecular Dynamics Simulations**

641 Simulations were performed using the GROMACS v2022 (61) package with the CHARMM36m
642 force field (62, 63). Temperature coupling was achieved using velocity rescaling applying a
643 coupling time of 0.1 ps with the protein, membrane and water/ions coupled separately at 310 K.
644 Semi-isotropic pressure coupling was maintained using the Parrinello-Rahman method with a
645 coupling time of 2.0 ps. All simulations were performed with a single non-bonded cut-off of 12 \AA ,
646 with van der Waals interactions switched at 1.0 \AA . The Verlet neighbor searching cut-off scheme

647 was applied with a neighbor-list update frequency of 25 steps (50 fs); the time step used in all the
648 simulations was 2 fs. Periodic boundary conditions were applied using the particle-mesh Ewald
649 method to account for long-range electrostatic interactions. The bond lengths were constrained
650 using the P-LINCS algorithm (64). Initial minimization and equilibration of all simulations
651 followed established procedures outlined by the CHARMM-GUI via a steepest descent protocol,
652 followed by short positionally restrained equilibration in the NVT (canonical) and NPT ensemble.
653 The systems were then allowed to progress for 1 μ s, with each system simulated in 10 replicas, for
654 a total simulation time of 10 μ s per consomatin analog.

655

656 **Statistical analysis**

657 All statistical analyses, fittings, and plotting of curves were performed using GraphPad Prism 10.
658 For the paw incision model, the effects at different time points were analyzed using two-way
659 ANOVA with Geisser-Greenhouse correction and Dunnett's post-test for multiple comparisons
660 (all compared to vehicle), while the area under the curve plot was analyzed using one-way
661 ANOVA with Dunnett's post-test for multiple comparisons (all compared to vehicle). For the SNI
662 model, the effect over time was analyzed as described for the paw incision model (all compared to
663 the vehicle on the ipsilateral side), while the area under the curve was analyzed using a two-way
664 ANOVA with Dunnett's post-test for multiple comparisons (all compared to the vehicle on the
665 ipsilateral side).

666

667

668 **List of Supplementary Materials**

669 **Supplemental file 1**

670 Methods

671 Figs. S1 to S3

672 References (65–70)

673

674 **Supplemental file 2**

675 Code to extract parameters for PCA.

676

677 **References and Notes**

- 678 1. M. Z. Murphy, T. P. Jackson, P. Mishra, *Hospitalized Chronic Pain Patient : A*
679 *Multidisciplinary Treatment Guide*. D. A. Edwards, P. Gulur, C. M. Sobey, Eds.,
680 (Springer International Publishing, 2022).
- 681 2. S. P. Cohen, L. Vase, W. M. Hooten, Chronic pain: an update on burden, best practices,
682 and new advances. *Lancet* **397**, 2082-2097 (2021).
- 683 3. D. A. Queremel Milani, D. D. Davis, *Pain Management Medications*. (StatPearls,
684 Treasure Island (FL), 2023).
- 685 4. CDC. (2023).
- 686 5. H. Tostivint *et al.*, Molecular evolution of GPCRs: Somatostatin/urotensin II receptors.
687 *Journal of molecular endocrinology* **52**, T61-86 (2014).

688 6. L. N. Møller, C. E. Stidsen, B. Hartmann, J. J. Holst, Somatostatin receptors. *Biochimica
689 et biophysica acta* **1616**, 1-84 (2003).

690 7. M. Chalabi *et al.*, Somatostatin analogs: does pharmacology impact antitumor efficacy?
691 *Trends in endocrinology and metabolism: TEM* **25**, 115-127 (2014).

692 8. L. J. Hofland, S. W. Lamberts, The pathophysiological consequences of somatostatin
693 receptor internalization and resistance. *Endocrine reviews* **24**, 28-47 (2003).

694 9. J. Pless, From somatostatin to Sandostatin®: History and chemistry. *Metabolism: clinical
695 and experimental* **41**, 5-6 (1992).

696 10. K. Elekes *et al.*, Inhibitory effects of synthetic somatostatin receptor subtype 4 agonists
697 on acute and chronic airway inflammation and hyperreactivity in the mouse. *Eur J
698 Pharmacol* **578**, 313-322 (2008).

699 11. E. Pinter, Z. Helyes, J. Szolcsanyi, Inhibitory effect of somatostatin on inflammation and
700 nociception. *Pharmacology & therapeutics* **112**, 440-456 (2006).

701 12. J. Szolcsanyi *et al.*, Analgesic effect of TT-232, a heptapeptide somatostatin analogue, in
702 acute pain models of the rat and the mouse and in streptozotocin-induced diabetic
703 mechanical allodynia. *Eur J Pharmacol* **498**, 103-109 (2004).

704 13. M. Engström, J. M. Savola, S. Wurster, Differential efficacies of somatostatin receptor
705 agonists for G-protein activation and desensitization of somatostatin receptor subtype 4-
706 mediated responses. *J Pharmacol Exp Ther* **316**, 1262-1268 (2006).

707 14. K. Sándor *et al.*, Analgesic effects of the somatostatin sst4 receptor selective agonist J-
708 2156 in acute and chronic pain models. *European Journal of Pharmacology* **539**, 71-75
709 (2006).

710 15. L. Gorham, S. Just, H. Doods, Somatostatin 4 receptor activation modulates
711 TRPV1[correction of TPRV1] currents in dorsal root ganglion neurons. *Neurosci Lett*
712 **573**, 35-39 (2014).

713 16. I. B. L. Ramiro *et al.*, Somatostatin venom analogs evolved by fish-hunting cone snails:
714 From prey capture behavior to identifying drug leads. *Science Advances* **8**, eabk1410
715 (2022).

716 17. T. L. Koch *et al.*, Reconstructing the Origins of the Somatostatin and Allatostatin-C
717 Signaling Systems Using the Accelerated Evolution of Biodiverse Cone Snail Venoms.
718 *Mol Biol Evol* **39**, (2022).

719 18. O. Buczek, G. Bulaj, B. M. Olivera, Conotoxins and the posttranslational modification of
720 secreted gene products. *Cellular and molecular life sciences : CMLS* **62**, 3067-3079
721 (2005).

722 19. W. K. Kroeze *et al.*, PRESTO-Tango as an open-source resource for interrogation of the
723 druggable human GPCRM. *Nature Structural & Molecular Biology* **22**, 362-369
724 (2015).

725 20. C. Lüscher, P. A. Slesinger, Emerging roles for G protein-gated inwardly rectifying
726 potassium (GIRK) channels in health and disease. *Nature Reviews Neuroscience* **11**, 301-
727 315 (2010).

728 21. I. Masuho, K. A. Martemyanov, N. A. Lambert, Monitoring G Protein Activation in Cells
729 with BRET. *Methods Mol Biol* **1335**, 107-113 (2015).

730 22. P. Ray *et al.*, Comparative transcriptome profiling of the human and mouse dorsal root
731 ganglia: an RNA-seq-based resource for pain and sensory neuroscience research. *PAIN*
732 **159**, 1325-1345 (2018).

733 23. A. M. Cowie, C. L. Stucky, A Mouse Model of Postoperative Pain. *Bio Protoc* **9**, (2019).

734 24. P. A. Shenoy *et al.*, The Somatostatin Receptor-4 Agonist J-2156 Alleviates Mechanical
735 Hypersensitivity in a Rat Model of Breast Cancer Induced Bone Pain. *Frontiers in*
736 *Pharmacology* **9**, (2018).

737 25. B. Kántás *et al.*, Novel Drug-Like Somatostatin Receptor 4 Agonists are Potential
738 Analgesics for Neuropathic Pain. *International Journal of Molecular Sciences* **20**, 6245
739 (2019).

740 26. É. Szőke *et al.*, Small molecule somatostatin receptor subtype 4 (sst4) agonists are novel
741 anti-inflammatory and analgesic drug candidates. *Neuropharmacology* **178**, 108198
742 (2020).

743 27. S. H. Sindrup, M. Otto, N. B. Finnerup, T. S. Jensen, Antidepressants in the Treatment of
744 Neuropathic Pain. *Basic & Clinical Pharmacology & Toxicology* **96**, 399-409 (2005).

745 28. G. C. Quintero, Review about gabapentin misuse, interactions, contraindications and side
746 effects. *Journal of Experimental Pharmacology* **9**, 13-21 (2017).

747 29. M. van Velzen, A. Dahan, M. Niesters, Neuropathic Pain: Challenges and Opportunities.
748 *Frontiers in Pain Research* **1**, (2020).

749 30. V. Isberg *et al.*, Generic GPCR residue numbers – aligning topology maps while minding
750 the gaps. *Trends in Pharmacological Sciences* **36**, 22-31 (2015).

751 31. W. Zhao *et al.*, Structural insights into ligand recognition and selectivity of somatostatin
752 receptors. *Cell Res* **32**, 761-772 (2022).

753 32. B. M. Olivera *et al.*, Peptide neurotoxins from fish-hunting cone snails. *Science* **230**,
754 1338-1343 (1985).

755 33. G. P. Miljanich, Ziconotide: neuronal calcium channel blocker for treating severe chronic
756 pain. *Curr Med Chem* **11**, 3029-3040 (2004).

757 34. H. Safavi-Hemami, S. E. Brogan, B. M. Olivera, Pain therapeutics from cone snail
758 venoms: From Ziconotide to novel non-opioid pathways. *J Proteomics* **190**, 12-20
759 (2019).

760 35. A. G. Craig *et al.*, An O-glycosylated neuroexcitatory conus peptide. *Biochemistry* **37**,
761 16019-16025 (1998).

762 36. L. Martin *et al.*, Conotoxin contulakin-G engages a neuropeptidin receptor 2/R-type
763 calcium channel (Cav2.3) pathway to mediate spinal antinociception. *Pain* **163**, 1751-
764 1762 (2022).

765 37. N. Puillandre *et al.*, Molecular phylogeny and evolution of the cone snails (Gastropoda,
766 Conoidea). *Mol Phylogenet Evol* **78**, 290-303 (2014).

767 38. B. Olivera, M. M. Watkins, N. Puillandre, M. Tenorio, J. Hidden diversity in the Asprella
768 clade: description of *Conus* (Asprella) neocostatus sp. nov. (Gastropoda, Conidae).
769 *Xenophora Taxonomy* **33**, 22-29 (2021).

770 39. ClinicalTrials.gov. (2023), Trial ID: NCT04707157.

771 40. MolluscaBase.org. (2024), <https://molluscabase.org/aphia.php?p=taxdetails&id=428949>.

772 41. N. Kaliss, D. Pressman, Plasma and Blood Volumes of Mouse Organs, As Determined
773 with Radioactive Iodoproteins. *Proceedings of the Society for Experimental Biology and*
774 *Medicine* **75**, 16-20 (1950).

775 42. B. M. Olivera, J. Seger, M. P. Horvath, A. E. Fedosov, Prey-Capture Strategies of Fish-
776 Hunting Cone Snails: Behavior, Neurobiology and Evolution. *Brain Behavior and*
777 *Evolution* **86**, 58-74 (2015).

778 43. M. J. Robertson, J. G. Meyerowitz, O. Panova, K. Borrelli, G. Skiniotis, Plasticity in
779 ligand recognition at somatostatin receptors. *Nat Struct Mol Biol* **29**, 210-217 (2022).

780 44. Z. Feng, B. Xu, Inspiration from the mirror: D-amino acid containing peptides in
781 biomedical approaches. *Biomol Concepts* **7**, 179-187 (2016).

782 45. M. Engström *et al.*, Superagonism at the human somatostatin receptor subtype 4. *J*
783 *Pharmacol Exp Ther* **312**, 332-338 (2005).

784 46. N. Schuelert *et al.*, The somatostatin receptor 4 agonist J-2156 reduces
785 mechanosensitivity of peripheral nerve afferents and spinal neurons in an inflammatory
786 pain model. *European Journal of Pharmacology* **746**, 274-281 (2015).

787 47. Z. Helyes, E. Pinter, J. Szolcsanyi, TT-232. *Drugs of the Future* **30**, (2005).

788 48. C. Lamers, Overcoming the shortcomings of peptide-based therapeutics. *Future Drug*
789 *Discovery* **4**, FDD75 (2022).

790 49. P. Kurtzhals, S. Østergaard, E. Nishimura, T. Kjeldsen, Derivatization with fatty acids in
791 peptide and protein drug discovery. *Nature Reviews Drug Discovery* **22**, 59-80 (2023).

792 50. M. Muttenthaler, G. F. King, D. J. Adams, P. F. Alewood, Trends in peptide drug
793 discovery. *Nature Reviews Drug Discovery* **20**, 309-325 (2021).

794 51. L. Wang *et al.*, Therapeutic peptides: current applications and future directions. *Signal*
795 *Transduction and Targeted Therapy* **7**, 48 (2022).

796 52. S. Kang, T. J. Brennan, Mechanisms of postoperative pain. *Anesth Pain Med* **11**, 236-248
797 (2016).

798 53. Z. Varecza *et al.*, Expression of the Somatostatin Receptor Subtype 4 in Intact and
799 Inflamed Pulmonary Tissues. *Journal of Histochemistry & Cytochemistry* **57**, 1127-1137
800 (2009).

801 54. Z. Helyes *et al.*, Effects of the somatostatin receptor subtype 4 selective agonist J-2156
802 on sensory neuropeptide release and inflammatory reactions in rodents. *British Journal of*
803 *Pharmacology* **149**, 405-415 (2006).

804 55. A. Silwal *et al.*, Novel Somatostatin Receptor-4 Agonist SM-I-26 Mitigates
805 Lipopolysaccharide-Induced Inflammatory Gene Expression in Microglia.
806 *Neurochemical Research* **47**, 768-780 (2022).

807 56. E. U. Carreira *et al.*, Neutrophils recruited by CXCR1/2 signalling mediate post-
808 incisional pain. *European Journal of Pain* **17**, 654-663 (2013).

809 57. R. Kumar, S. Gupta, M. Gautam, S. K. Jhajhria, S. B. Ray, Diverse characters of
810 Brennan's paw incision model regarding certain parameters in the rat. *Korean J Pain* **32**,
811 168-177 (2019).

812 58. E. M. Pogatzki-Zahn, D. Segelcke, S. A. Schug, Postoperative pain—from mechanisms
813 to treatment. *PAIN Reports* **2**, e588 (2017).

814 59. T. J. Brennan, P. K. Zahn, E. M. Pogatzki-Zahn, Mechanisms of incisional pain.
815 *Anesthesiology clinics of North America* **23**, 1-20 (2005).

816 60. S. R. Chaplan, F. W. Bach, J. W. Pogrel, J. M. Chung, T. L. Yaksh, Quantitative
817 assessment of tactile allodynia in the rat paw. *Journal of neuroscience methods* **53**, 55-63
818 (1994).

819 61. M. J. Abraham *et al.*, GROMACS: High performance molecular simulations through
820 multi-level parallelism from laptops to supercomputers. *SoftwareX* **1-2**, 19-25 (2015).

821 62. R. B. Best *et al.*, Optimization of the additive CHARMM all-atom protein force field
822 targeting improved sampling of the backbone phi, psi and side-chain chi(1) and chi(2)
823 dihedral angles. *Journal of chemical theory and computation* **8**, 3257-3273 (2012).

824 63. O. Guvench *et al.*, CHARMM additive all-atom force field for carbohydrate derivatives
825 and its utility in polysaccharide and carbohydrate-protein modeling. *Journal of chemical*
826 *theory and computation* **7**, 3162-3180 (2011).

827 64. B. Hess, P-LINCS: A Parallel Linear Constraint Solver for Molecular Simulation.
828 *Journal of chemical theory and computation* **4**, 116-122 (2008).

829

830 **Acknowledgments:**

831 Prof. Hans Bräuner-Osborne for help with establishing GPCR receptor assays.
832 Dr. Joanna Gajewiak for illustrations of the chemical structure of consomatin Fj1.
833 Dr. Paula Flórez Salcedo for illustration of the cone snail shell.

834 **Funding:**

835 Villum Foundation Young Investigator grant 19063 (HS-H)
836 Lundbeck Foundation Experiment grant R400-2022-509 to WEB)
837 Independent Research Fund Denmark grant 3102-00006 (TLK)
838 Lundbeck Foundation Ascending Investigator grant R344-2020-1063 (KLM)
839 Lundbeck Postdoc grant R322-2019-1816 (KLJ)
840 National Institutes of Health R01NS116694 (AP)
841 National Institutes of Health K08NS104272 (AP)
842 Part of this work was undertaken with the assistance of resources from the National Computational
843 Infrastructure (NCI), which is supported by the Australian Government and provided through
844 Intersect Australia Ltd.

846 **Author contributions:**

847 Conceptualization: WEB, HS-H
848 Methodology: WEB, IBLR, TLK, EE, HYY, CMG, KLJ, NAS, LFM
849 Software: WEB, TLK, NAS
850 Formal Analysis: WEB, TLK, NAS
851 Resources, The NCI (supported by the Australian Government and provided through Intersect
852 Australia Ltd)
853 Investigation: WEB, IBLR, TLK, EE, HYY, CMG, KLJ, NAS, and LFM
854 Data curation: WEB
855 Writing – Original Draft: WEB, HS-H
856 Writing – Review and Editing: WEB, IBLR, TLK, EE, HYY, KLJ, NAS, LFM, BJS, KLM, AP,
857 HS-H
858 Visualization: WEB, TLK, HS-H
859 Supervision: WEB, KKS, BJS, KLM, AP, HS-H
860 Project Administration: WEB and HS
861 Funding Acquisition: WEB, TLK, KLJ, KLM, HS-H, AP

863 **Competing interests:**

864 WEB, IBLR, TLK, and HS-H are inventors of a patent application for Fj1 and its analogs (patent
865 application # WO2023180125A1).

868 **Data and materials availability:**

869 All data and code can be found either in databases as described in the text, or in the main text of
870 supplemental materials. All materials are commercially available.

871