

Forces directing the systemic correlations of cell migration

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Significance

Cellular migration is a cornerstone issue in many human physiological and pathological processes. For years, the scientific attention has been focused on the individualized study of the diverse molecular parts involved in directional motility; however, locomotion movements have never been regarded as a systemic process that operates at a global cellular scale. In our quantitative experimental analysis essential *systemic* properties underlying locomotion movements were detected. Such emergent systemic properties are not found specifically in any of the molecular parts, partial mechanisms, or individual processes of the cell. Cellular displacements seem to be regulated by integrative processes operating at *systemic* level.

Abstract

Directional motility is an essential property of cells. Despite its enormous relevance in many fundamental physiological and pathological processes, how cells control their locomotion movements remains an unresolved question. Here we have addressed the systemic processes driving the directed locomotion of cells. Specifically, we have performed an exhaustive study analyzing the trajectories of 700 individual cells belonging to three different species (*Amoeba proteus*, *Metamoeba leningradensis* and *Amoeba borokensis*) in four different scenarios: in absence of stimuli, under an electric field (galvanotaxis), in a chemotactic gradient (chemotaxis), and under simultaneous galvanotactic and chemotactic stimuli. All movements were analyzed using advanced quantitative tools. The results show that the trajectories are mainly characterized by coherent integrative responses that operate at the global cellular scale. These systemic migratory movements depend on the cooperative non-linear interaction of most, if not all, molecular components of cells.

Keywords

Cellular migration, systemic behavior, self-organization, quantitative analysis, galvanotaxis, chemotaxis, *Amoeba proteus*, *Metamoeba leningradensis*, *Amoeba borokensis*

Introduction

Self-locomotion is one of the most important complex behaviors of cells endowed with migratory responses. In the permanent struggle for survival free cells move efficiently to find food following adequate direction and speed, avoiding predators and adverse conditions. Cell motility is crucial for life in Metazoan organisms and fundamental to establish the appropriate organization of all multicellular organisms, playing a central role in a plethora of essential biological phenomena such as embryogenesis, morphogenesis, organogenesis, neural development, adult tissue remodeling, wound healing, immune responses, angiogenesis, tissue regeneration and repair, cell differentiation, etc. (1). Moreover, the deregulation of cell movements in humans is involved in many pathological processes such as chronic inflammatory diseases, atherosclerosis, vascular diseases, osteoporosis, congenital brain pathologies, hearing disorders, tumorigenesis and metastasis, among others (2–5).

Although cell locomotion was already observed in 1675 by van Leeuwenhoek in his respected pioneer microscopic studies (6), researchers and scholars have not yet come to unveil how cells move efficiently through diverse environments and migrate in the presence of complex cues.

Given its importance, a great attention has been focused on the study of the diverse molecular parts involved in directional motility. A relevant number of these experimental studies have unequivocally shown that locomotion movements are complex processes that involve practically all cellular components. So, directed movements are primarily driven by the cytoskeleton (the essential part of the locomotion system) which is a sophisticated dynamic structure formed by three main molecular components: actin microfilaments, microtubules, and intermediate filaments, all of them interacting in complex dynamic networks (7).

In particular, the activity of actin cytoskeleton networks is largely dependent on a wide variety of regulatory molecules such as small GTPases (8), integrins (9), and many post-translational modifications such as phosphorylation, acetylation, arginylation, oxidation, and others (10). In addition, the dynamic turnover of the actin filament networks is essential to regulate cell migration (11). Cytoskeleton networks are coupled with other complex regulated systems such as membrane surface receptors and signal transduction pathways which also participate in the control of locomotion movements (12). Energy is another essential element in cell motility; when cells move the cytoskeleton transforms chemical energy into mechanical forces (dynein cytoskeletal motor proteins) entailing considerable bioenergetic demands; for such a purpose the mitochondrial activity and the adenylate energy system are important regulators of directionality motion (13). Cell membrane activities are also necessary to implement an adequate migration (14, 15).

Recent studies have revealed the importance of autophagy (an intracellular process that controls protein and organelle degradation and recycling) in the control of locomotion (16). The turnover of focal adhesions also regulates cell spreading and migration (17). Calcium ions (Ca^{2+}), which impact globally on almost every aspect of cellular life, play an important role in the control of directed

movements (18). In this sense, the endoplasmic reticulum, a multifunctional signaling organelle which controls a wide range of cellular processes such as the entry and release of calcium ions, also participate in the regulation of cell locomotion (19, 20). Cell polarity is required for an adequate directionality motion and there are a lot of molecular processes that have been implicated in the intrinsic polarity status of cells; in this regard, the centrosomes positioning serves as a steering device for the directional movement (21, 22) and dynein together with other molecules regulates centrosomal orientation to establish and maintain cell polarity (23).

The Golgi apparatus (another important molecular processing center for modified proteins received from the endoplasmic reticulum) allows the remodeling of intracellular traffic processes towards the direction of movement; therefore, signals from the Golgi matrix play an important role in cell motility (24). The nucleus is very important for developing appropriate mechanical responses during cell migration, in fact this organelle behaves as a central mechanosensory structure, and its physical properties strongly connected to the cytoskeleton guarantee a proper cell migration (25, 26). Recently, it has been described that structural chromatin organization also has a key role in the cellular migration process (27). In addition, many molecules and corresponding processes are involved in directional movement of cells as, for instance, focal adhesion proteins (talin, paxillin, vinculin and others) (12), SCAR/WAVE proteins (28), actin-binding proteins (ABPs) (29), p21-activated kinases (PAKs, a family of serine/threonine kinases) (30), TORC2/PKB pathway (31), mitogen-activated protein kinases (MAPKs) (32), Arp2/3 complexes (33), WASP family proteins (34), Nck family of adaptor proteins (35), etc.

All this evidence suggests that cell migration is not a mere metabolic-molecular process which can be regulated by any of its individually considered components. Most, if not all, fundamental cellular physiological processes appear to be involved in cellular locomotion, which is indicative of the emergence of a global functional phenomenon in the cell. However, confirming the systemic nature of cellular locomotion represents a scientific challenge of great difficulty. Such verification requires multidisciplinary approaches that combine complex experimental studies with advanced quantitative methods.

Here, we have addressed the key question: Is cell migration a highly coordinated and integrated emergent process at the global cellular level? To answer this question, we have designed a large quantitative study to analyze the systemic trajectories of 700 individual cells belonging to three different species: *Amoeba proteus*, *Metamoeba leningradensis* and *Amoeba borokensis*. Such analysis has been performed under four different scenarios: in absence of stimuli, under chemotactic gradient (we have used a nFMLP peptide, which indicates to the amoebae the possible presence of food in their immediate environment), in an electric field (the electric membrane potential of cells enables predators like amoebas the detection of preys), and under complex external conditions such as simultaneous galvanotactic and chemotactic gradient stimuli.

To understand the forces driving the locomotion movement of the cell, all trajectories were analyzed using computational methods and advanced non-linear physical-mathematical tools rooted in Statistical Physics (Statistical Mechanics). These quantitative studies focused on some essential characteristics of the systemic dynamics underlying locomotion movements. The results indicate that a very complex dynamic structure emerges in the migratory movements of all the cells analyzed. Such structure is characterized by highly organized move-step sequences with very low entropy and high information, non-trivial long-range interdependence in the move-steps, strong anomalous super-diffusion dynamics, long-term memory effects with trend-reinforcing behavior, and efficient movements to explore the extracellular medium.

This outstanding cellular dynamic structure is a consequence of the emergent systemic dynamics occurring in the cell. The locomotion movements seem to depend on a complex integrated self-organized system carefully regulated at global level, arising from the cooperative non-linear interaction of most, if not all, cellular components. Such emergent systemic properties are not found specifically in any of the molecular parts, partial mechanisms, or individual processes of the cell.

The findings presented here open a new perspective to improve the conceptual framework of the cell, and to develop researches on the pathological deregulation of migratory movements combining systemic analysis with molecular studies.

Results

The migratory trajectories of 700 individual cells belonging to the three species, *A. proteus*, *M. leningradensis* and *A. borokensis*, were recorded in four different scenarios: in absence of stimuli, under chemotactic gradient, in an electric field, and under simultaneous galvanotactic and chemotactic stimuli. Amoebae show robust movement in response to an electric field in a range between 300 mV/mm and 600 mV/mm (galvanotaxis). Under such conditions, practically all amoebae migrate towards the cathode (36). Likewise, these cells also exhibit chemotactic movements. More specifically, the peptide nFMLP (N-Formylmethionyl-leucyl-phenylalanine) secreted by bacteria may indicate that food might be in the near environment, provoking a strong chemotactic response (37).

All our experiments were performed in a specific set-up consisting of two standard electrophoresis blocks (17.5 cm long), two agar bridges, a power supply, and in the middle of the experimental platform, a structure of standard glass slide and covers where the cells were located (see *SI Appendix*, Fig. S1 and *Materials and Methods*). One electrophoresis block was directly plugged into a normal power supply and the other was connected to the first one through two agar bridges, thus preventing the direct contact of the anode and cathode with the medium (Chalkley's simplified medium (36)) where the cells were placed. Specifically, the amoebae were arranged in the center of the structure of standard glass slide and covers (experimental chamber) and their migratory displacements were monitored. The glass experimental structure enabled the generation of a laminar flux allowing the electric current to pass through, on one hand, and generating an nFMLP peptide gradient, on the other (see *Materials and Methods* and *SI Appendix*, Figs. S1 and S2).

Prior to each experiment, all cells were starved for 24h. The individual migratory movements of each cell were recorded over periods of 30 minutes using a digital camera attached to a stereo microscope. The experiments on flat two-dimensional surfaces were always made with small groups of cells (no more than 9 cells per replication). The following basic experimental information data (BEID) is provided for each scenario: "Nr" the number of cells per replication, "Er" the number of experimental replications, and "N" is the total number of cells. Finally, the recorded trajectories were analyzed in the form of time series using advanced non-linear dynamic tools.

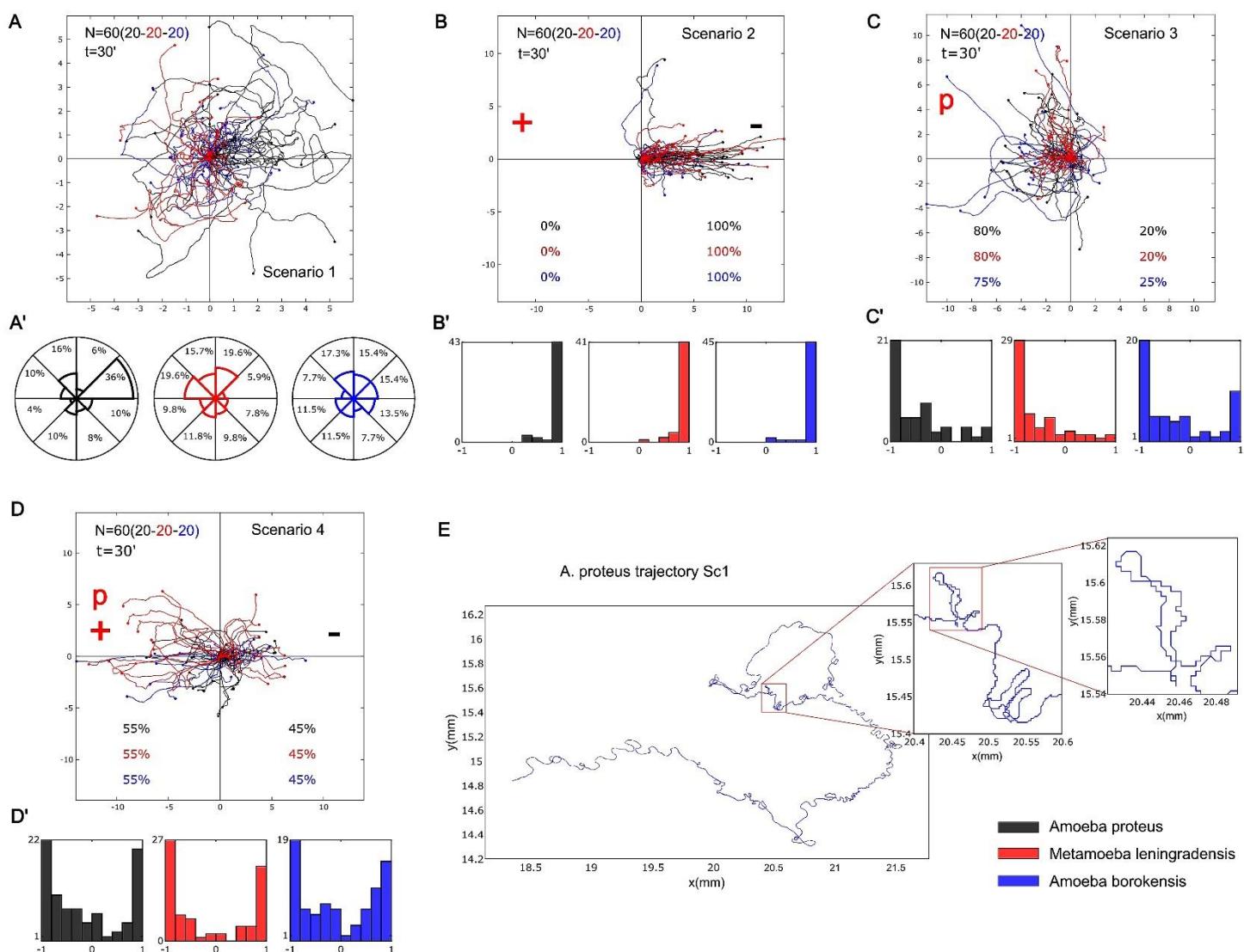


Fig. 1. Representative migration trajectories of the three species under four experimental scenarios.

(A-D) Migration behavior of the three species (in black *Amoeba proteus*, in red *Metamoeba leningradensis*, and in blue *Amoeba borokensis*) in the four experimental scenarios (“Scenario 1” absence of stimuli, “Scenario 2” presence of an electric field, “Scenario 3” presence of a chemotactic peptide gradient and “Scenario 4” simultaneous galvanotactic and chemotactic stimuli). (A') the percentage of cells moving in any specific sector of the experimental chamber is represented as a polar histogram divided into 8 areas of $\pi/4$ angle amplitude each. (B'-D') histograms of the displacement cosines from panels (B-D), respectively. (E) Digitized cell trajectory with two inserts highlighting displacements regions of interest. “N” is the total number of cells; “t”, is the time of galvanotaxis or chemotaxis; “p”, is the chemotactic peptide (nFMLP); “+”, represents the anode; “-”, represents the cathode; “Sc1” Scenario One (absence of stimuli). Both the x and y-axis show the distance in mm, and the initial location of each cell has been placed at the center of the diagram.

1. Cellular migratory movements without external stimulus.

First, we recorded the locomotion trajectories of 153 individual cells belonging to the three species considered in a medium without any external influence (BEID: *A. proteus*: N=50, Er=7, Nr=7-8; *M. leningradensis*: N=51, Er=7, Nr=5-8; *A. borokensis*: N=52, Er=7, Nr=6-8). In Fig. 1A, a representative example of these amoebae migratory movements in absence of stimuli is depicted (for clarity only 60 cells were randomly taken from the total). It can be observed that after 30 minutes, cells have explored practically all the directions of the experimentation chamber. To quantitatively analyze cell directionality, we calculated the displacement cosine for each trajectory, 153 cells in total (Fig. 1A'). Values close to -1 indicate a preference towards the left, while values close to 1 suggest a preference towards the right. Our analysis showed that values ranged between -1 and 1, with a median/IQR of 0.052/1.397. Median/IQR values for each species were 0.417/1.244 (*A. proteus*), -0.251/1.207 (*M. leningradensis*) and 0.126/1.232 (*A. borokensis*). These results indicate that in absence of stimuli cells moved randomly without any defined guidance.

2. Cell migration under galvanotaxis conditions.

The migratory trajectories of 147 cells belonging to the three species were recorded under an external controlled direct-current electric field of about 300-600 mV/mm. In Fig. 1B, a representative example of the migratory movements of 60 cells is depicted. They show an unequivocal systemic response consisting of the migration to the cathode which has been placed on the right side of the set-up. The overall median/IQR value of the displacement cosines of all 147 cells (Fig. 1B') was 0.989/0.0732. This finding confirmed that a fundamental behavior characterized by an unequivocal directionality towards the cathode had emerged under these galvanotactic conditions. The median/IQR values for each species were 0.993/0.023 (*A. proteus*), 0.978/0.098 (*M. leningradensis*) and 0.985/0.083 (*A. borokensis*). We compared the distributions of the values for the displacement cosines under galvanotaxis with the values obtained in the experiment without stimuli using the Wilcoxon rank-sum test. The results indicated that both behaviors were significantly different for the three species and that the galvanotactic cellular behavior is highly unlikely to be obtained by chance (p-values: 10^{-9} , 10^{-15} , and 10^{-12} ; Z: -5.854, -7.789, and -6.845 for *A. proteus*, *M. leningradensis*, and *A. borokensis*, respectively). BEID: *A. proteus*: N=49, Er=7, Nr=6-8; *M. leningradensis*: N=48, Er=7, Nr=6-8; *A. borokensis*: N=50, Er=8, Nr=3-9.

3. Cell locomotion under chemotaxis conditions.

The migratory behavior of 166 cells belonging to the three species considered was recorded under conditions of chemotactic gradient. All the amoebae were exposed for 30 minutes to an nFMLP peptide gradient which had been placed on the left side of the set-up. In Fig. 1C a representative example of these migratory trajectories with 60 cells is depicted. Under these conditions 78.31% of all studied amoebae showed locomotion movements towards the attractant peptide.

The displacement angle cosines of the 166 individual trajectories ranged from -1 to 1, with a median/IQR value of -0.672/0.874. Median/IQR values for each species were -0.652/0.745 (*A. proteus*), -0.774/0.676 (*M. leningradensis*) and -0.513/1.17 (*A. borokensis*), indicating that they exhibited a single fundamental behavior of movement towards the peptide (Fig. 1C'). The Wilcoxon rank-sum test showed significant differences between the cosine values obtained with and without chemotactic stimulus (p-values: 10^{-7} , 0.003, and 0.016; Z: 5.0841, 2.9379, and 2.403 for *A. proteus*, *M. leningradensis*, and *A. borokensis*, respectively), and between the cosine values with chemotactic

gradient and with the presence of an electric field (p-values: 10^{-15} , 10^{-17} , and 10^{-14} ; Z: 7.998, 8.554, and 7.677 for *A. proteus*, *M. leningradensis*, and *A. borokensis*, respectively). This confirmed that the systemic locomotion behavior under the chemotactic gradient was completely different from both the absence of stimuli and the presence of an electric field. BEID: *A. proteus*: N=51, Er=8, Nr=5-7; *M. leningradensis*: N=60, Er=9, Nr=5-8; *A. borokensis*: N=55, Er=10, Nr=5-7.

4. Cellular displacement under simultaneous galvanotactic and chemotactic stimuli.

Once the locomotion movements of the cells were recorded under the three previous independent experimental scenarios (without stimuli, under galvanotaxis and under chemotaxis), we studied the trajectories of 234 cells under simultaneous galvanotactic and chemotactic stimuli. For such a purpose, the nFMLP peptide was arranged on the left of the set-up (in the anode area) and the cathode was placed on the right. In Fig. 1D, a representative example of these locomotion movements (60 cells in total) is depicted. Under these complex external conditions, the results showed that 42% of the amoebae migrated towards the cathode while the remaining 58% moved towards the peptide (anode).

The displacement cosines of the 234 cells had an overall median/IQR value of -0.286/1.659. More specifically, the values for each species were (-0.315/1.591, median/IQR) for *A. proteus*, (-0.542/1.811, median/IQR) for *M. leningradensis*, and (-0.137/1.453, median/IQR) for *A. borokensis*. This analysis quantitatively verified that two main cellular migratory behaviors had emerged in the experiment, one towards the anode and another towards the cathode (Fig. 1D'). The statistical analysis (Wilcoxon rank-sum test) confirmed the presence of these two different behaviors for *A. proteus* (p-value= 10^{-14} ; Z=7.672), *M. leningradensis* (p-value= 10^{-13} ; Z=7.226) and *A. borokensis* (p-value= 10^{-14} ; Z=7.5549). BEID: *Amoeba proteus*: N=83, Er=12, Nr=6-8; *Metamoeba leningradensis*: N=73, Er=11, Nr=5-8; *Amoeba borokensis*: N=78, Er=12, Nr=4-8.

5. Long-range interdependence in the move-steps of cellular migratory displacements.

An essential characteristic of systemic behavior in complex systems is the presence of dynamics with strong long-range correlations (38). One of the most recognized tools to analyze the presence of these correlations in time series (migratory trajectories here) is the “root mean square fluctuation” (“rmsf” analysis), a classical method in Statistical Mechanics based on the ideas raised by Gibbs (39) and Einstein (40).

Long-range interdependence can be detected by a power-law relation such that $F(l) \sim l^\alpha$. Where l is the number of steps. For uncorrelated data, the fluctuation exponent α is about 0.5, whereas $\alpha > 0.5$ or $\alpha < 0.5$ indicate respectively the presence of positive or negative long-range correlations (*Materials and Methods*). In Fig. 2A, an illustrative “rmsf” analysis for the locomotion movements of three representative cells belonging to each species considered under simultaneous chemotactic and galvanotactic stimuli (*A. proteus* and *M. leningradensis*) and under chemotactic conditions (*A. borokensis*) is depicted.

The results of “rmsf” analysis of the 700 experimental cell trajectories are shown in Fig. 2B (for more details see *SI Appendix*, Table S1). All the migratory trajectories exhibit non-trivial correlations in their cellular move-step migratory fluctuations. Specifically, we found that the scaling exponent α of the “rmsf” had a median/IQR value of 0.723/0.08 for *A. proteus*, 0.734/0.081 for *M. leningradensis*, and 0.707/0.076 for *A. borokensis*. The values of the “rmsf” analysis of the total experimental migratory trajectories analyzed ranged from 0.557 to 0.867, with a median/IQR of 0.723/0.08, whereas the values of the scaling exponent α of all shuffled trajectories ranged from 0.351 to 0.644, with a median/IQR value of 0.468/0.069 (see *SI Appendix*, Table S2 for more details). Moreover, a

Wilcoxon test comparing measured exponents to those from shuffled trajectories revealed highly significant long-range correlations in our data ($p\text{-value}\approx 0$, $Z = -32.312$), indicating the improbability of chance occurrence.

We also calculated the time duration of the correlations regime and found that all cells exhibited long-range coordination over periods ranging from 1.042 to 16.667 minutes with a median/IQR value of 9.375/7.292 minutes. These findings indicate strong dependences of past movements lasting approximately 1125/875 (median/IQR) move-steps (Fig. 2C and *SI Appendix*, Table S3). *A. proteus* cells exhibited long-range correlations up to a median/IQR duration of 10.417/6.25 minutes, *M. leningradensis* cells showed 9.375/7.292 minutes and *A. borokensis* cells 8.333/6.25 minutes, thus highlighting the influence of previous trajectory values on each cellular move-step. These results show the presence of non-trivial long-range correlations in all migration trajectories.

6. Strong anomalous migratory dynamics in cellular locomotion.

Another characteristic of the migratory movement of cells is their strong anomalous dynamics. This property is directly related to anomalous super-diffusion, a complex process with a high non-linear relationship to time which also corresponds to efficient systemic directional trajectories (41, 42). One of the best methods to determine such dynamic property is the Mean Square Displacement (MSD), a method proposed by Einstein (43) and later by Smoluchowski (44). This Statistical Mechanics tool allows to quantify the amount of space explored by the amoebae during their locomotion. According to this procedure (see *Materials and Methods*), the anomalous diffusion exponent β is commonly used to refer to whether normal (Brownian, $\beta = 1$) or anomalous diffusion ($\beta \neq 1$) is observed. The dynamics of sub-diffusion and super-diffusion correspond to $0 < \beta < 1$ and $\beta > 1$, respectively.

In Fig. 3A, we depicted an MSD analysis for the locomotion movements of three representative cells belonging to each species under absence of stimuli. The results of MSD analysis of the 700 experimental cells (see Fig. 3B, Fig. 3C and *SI Appendix*, Table S4) show that practically all trajectories exhibit strong anomalous migratory dynamics. For experimental trajectories, the variable β , which characterizes the behavior of the diffusion process, had a median/IQR value of 1.899/0.127 for *A. proteus* cells, of 1.878/0.168 for *M. leningradensis* cells, and of 1.845/0.146 for *A. borokensis* cells. These values suggest an anomalous super-diffusive process, a complex behavior which appears to govern the three groups of cell trajectories. The values for experimental trajectories of the anomalous diffusion exponent β ranged from 1.092 to 2.022, median/IQR value of 1.874/0.15, whereas the values for shuffled trajectories ranged from -0.007 to 0.006, with a median/IQR value of 10⁻⁴/0.002 (see Fig. 3B and *SI Appendix*, Table S5). A Wilcoxon test comparing anomalous diffusion exponents from shuffled showed that our results are extremely unlikely to be obtained by chance ($p\text{-value}\approx 0$, $Z=32.392$).

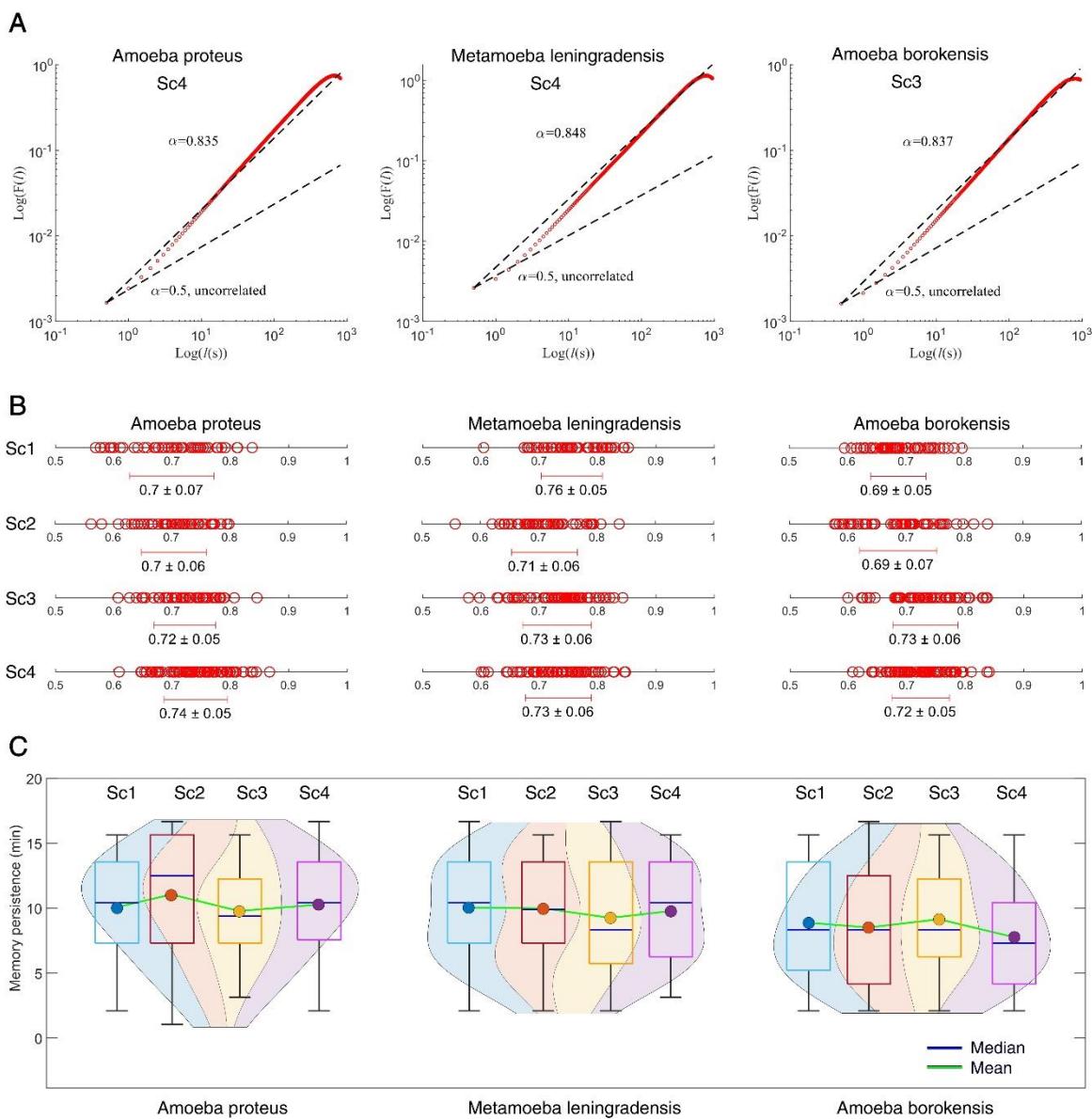


Fig. 2. Long-range interdependence in the move-steps of cellular migratory displacements.

(A) Log-log plot of RMSF F versus l step for a representative cell of each species. The slope was $\alpha=0.835$ for *A. proteus*, $\alpha=0.848$ for *M. leningradensis* and $\alpha=0.837$ for *A. borokensis*, indicating the presence of strong long-range interdependence in the move-steps of all of them. (B) Diagram representing the values (and the overall average \pm SD) of all the scaling exponents α from cells belonging to each species (*A. proteus*, *M. leningradensis* and *A. borokensis*) under each experimental scenario (Sc1-Sc4). (C) Violin plots showing the estimated distribution, median and average memory persistence values from cellular trajectories. “Sc1” Scenario One, absence of stimuli; “Sc2” Scenario Two, presence of an electric field; “Sc3” Scenario Three, presence of a chemotactic peptide gradient and “Sc4” Scenario Four, simultaneous galvanotactic and chemotactic stimuli.

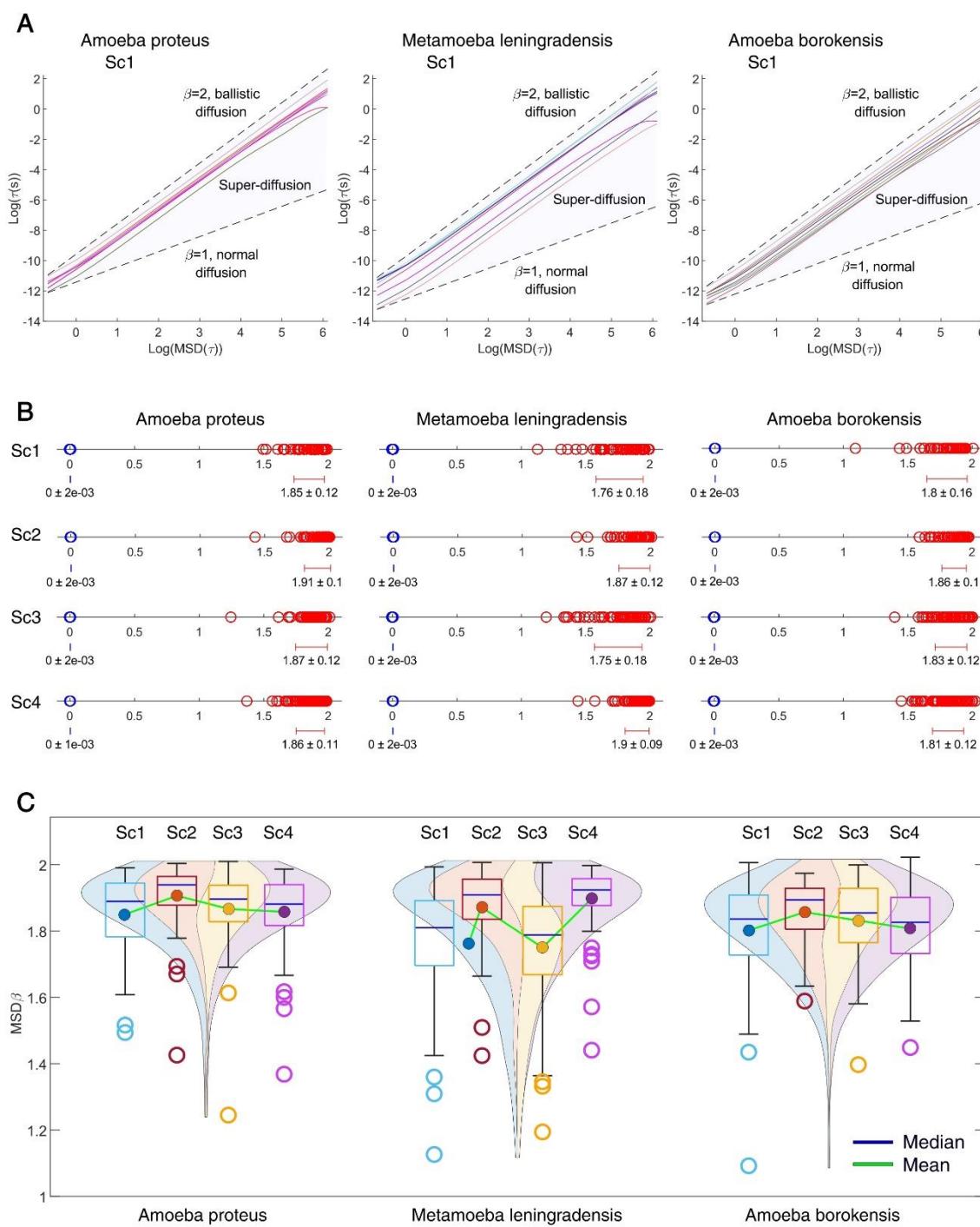


Fig. 3. Strong anomalous migratory dynamics in cellular locomotion.

(A) Graphics showing the value of the exponent β by fitting log-log plots of MSD as a function of the time interval τ , for 8 prototypic cells of each species (*A. proteus*, *M. leningradensis* and *A. borokensis*). $\beta=1$ indicates normal diffusion while $\beta=2$ indicates ballistic diffusion. The grey region defines the area of super-diffusion, which is a complex process with a high non-linear relationship to time, within which all the experimental values fall. (B) Diagram representing all values of the β exponents (and the overall average \pm SD) for all cells of the three species in each experimental scenario (Sc1-Sc4) experimental values in red and shuffled values in blue. The shuffling step extinguished the long-term correlation structure, causing the sharp division between the experimental and shuffled value distributions ($p\text{-value} \approx 0$, $Z = -32.3921$) for all species and experimental conditions. (C) Estimated distribution, median and mean MSD β exponent values from experimental trajectories are illustrated using violin plots. “Sc1” Scenario One, absence of stimuli; “Sc2” Scenario Two, presence of an electric field; “Sc3” Scenario Three, presence of a chemotactic peptide gradient and “Sc4” Scenario Four, simultaneous galvanotactic and chemotactic stimuli.

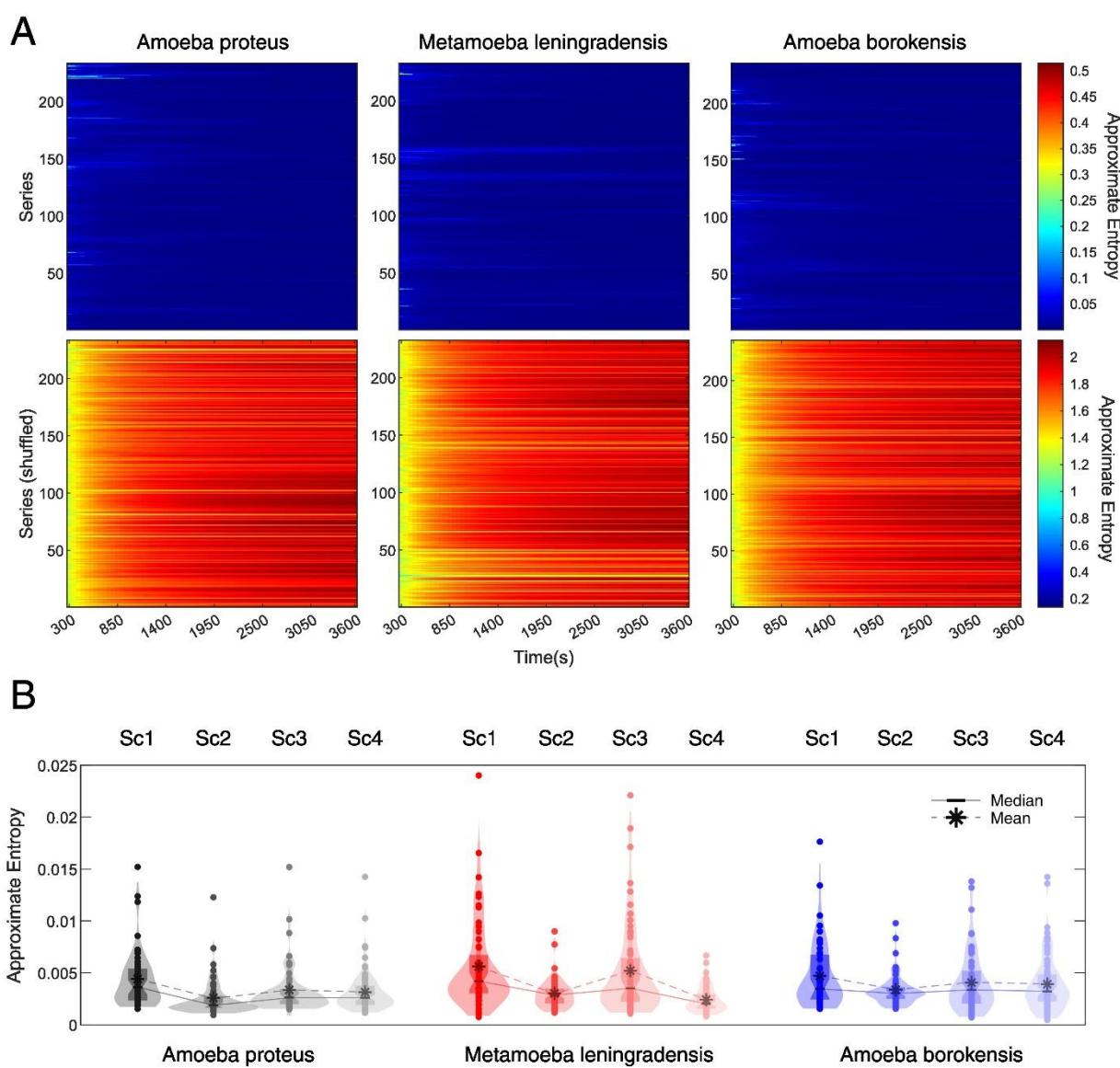


Fig. 4. Complexity and information in cellular migration.

(A) Heatmaps for the Approximate Entropy values of all 700 experimental (upper row panels) and shuffled (bottom row panels) cell trajectories from each species (*A. proteus*, *M. leningradensis* and *A. borokensis*). Each row in every panel corresponds to a single cell, while in the 72 columns the endpoint of the Approximate Entropy calculation is represented, increased in 25 seconds at every iteration. (B) Violin plots illustrate the estimated distribution, mean and median Approximate Entropy values for all experimental cell trajectories. “Sc1” Scenario One, absence of stimuli; “Sc2” Scenario Two, presence of an electric field; “Sc3” Scenario Three, presence of a chemotactic peptide gradient and “Sc4” Scenario Four, simultaneous galvanotactic and chemotactic stimuli.

7. Complexity and information in cellular migration.

To assess the information content within locomotion trajectories we implemented the Approximate Entropy (ApEn), a robust approximation of the Kolmogorov–Sinai (K-S) entropy (45, 46), providing insight into the complex migratory behavior that emerges from the cellular system.

In Fig. 4A (*SI Appendix*, Tables S6 and S7), the results of the ApEn estimation for the 700 cellular trajectories are shown. The heatmaps display the Approximate K-S entropy for all experimental (upper row) and shuffled trajectories (bottom row) from each species, calculated for 72 different time windows (intervals) of increasing length (interval duration was increased by 25 seconds at every iteration). Intervals present ApEn values that vary from 10^{-4} (in blue) to 0.515 (red) for experimental trajectories and from 0.141 (in blue) to 2.126 (in red) for shuffled trajectories. These findings allow to observe that practically all the experimental series exhibit extremely low entropy.

In Fig. 4B, (*SI Appendix*, Table S6), the decrease in entropy that occurs from SC1 to SC2-SC4 denotes how in absence of stimuli (SC1) cells maximize entropy, but when there is a more defined directionality cellular trajectories become more focused (less entropic displacements). Specifically, we found that the ApEn values of experimental trajectories exhibited a narrow range of low values displaying a median/IQR of 0.003/0.002 for *A. proteus*, 0.003/0.003 for *M. leningradensis* and 0.003/0.003 for *A. borokensis*. The ApEn analysis for all experimental migratory trajectories obtained under the four scenarios showed a median/IQR ApEn value of 0.003/0.002 with values ranging from 10^{-4} to 0.024. In Fig. 4B, it is also showed that the high regularity and order observed in the experimental migration series vanished after shuffling. The ApEn values for shuffled trajectories displayed a range of very high values (from 1.252 to 2.126, median/IQR equal to 1.966/0.156) relative to the values for experimental trajectories (*SI Appendix*, Table S7).

The whole analysis confirms the presence of a complex structure characterized by high information in the move-step sequences in the migration trajectories of all cells. Furthermore, the statistical analysis revealed that this complex dynamic structure observed in the move-step trajectories was highly unlikely to occur by chance, as indicated by a p -value ≈ 0 and $Z = -32.392$, results of a Wilcoxon test comparing the respective ApEn value distributions of experimental and shuffled trajectories.

8. Long-term memory effects in cellular migratory movements.

Long-range memory effects or persistence is another main characteristic of the systemic cellular migratory movements in unicellular organisms (47, 48). The Detrended Fluctuation Analysis (DFA) (see *Materials and Methods*) is a well-known technique for measuring persistent effects in physiological time series.

For a given observation scale ℓ , DFA calculates the function (ℓ) to quantify the fluctuations of the time series around the local trend. If the time series displays scaling properties, then $F(\ell) \sim \ell^\gamma$ asymptotically, where γ represents the scaling exponent. This exponent is commonly estimated as the slope of a linear fit in the $\log(F(n))$ versus $\log(\ell)$ plot. Thus, γ serves as a measure of the strength of long-term memory effects and characterizes the underlying dynamical system. Specifically, values close to 0.5 indicate the absence of long-range correlations, while when $1.5 < \gamma < 2$, the process exhibits positive long-range persistence (49) (Fig. 5A). Through the application of this quantitative method, we identified the presence of long-range persistence in all experimental trajectories (*SI Appendix*, Table S8), with a γ overall median/IQR value of 1.784/0.107. Specifically, the median/IQR DFA scaling parameter γ was found to be 1.795/0.087 for *A. proteus*, 1.785/0.138 for *M. leningradensis*, and 1.775/0.097 for *A. borokensis* (Fig. 5B and *SI Appendix*, Table S8), thus indicating that all the move-step trajectories exhibit trend-reinforcing memory.

In order to assess the reliability of the DFA analysis, we conducted a random shuffling procedure on 700 time series. The results demonstrated that the strong correlation values observed in the experimental migration series vanished after shuffling (refer to Fig. 5B, Fig. 5C and *SI Appendix*, Table S9 for more information), with γ overall median/IQR of 0.477/0.135. This finding confirms that the complex locomotion structure, characterized by well-organized move-step sequences and persistent dynamics observed in the migration trajectories of the three cell groups, is not attributable to a random chance (p -value ≈ 0 , $Z = 32.392$).

9. Kinematic properties in cellular locomotion trajectories.

To quantify some kinematic properties of the cell migration trajectories, we studied the Intensity of the response (IR), the directionality ratio (DR) and the average speed (AS) of amoebae (Fig. 6A, B, and C).

The IR is associated to the space explored by the cell, and in particular, we quantified the module of the trajectories to represent the strength of the response. In this case, the median/IQR IR was 5.322/3.5 for *A. proteus*, 4.961/5.053 for *M. leningradensis* and 3.575/3.072 for *A. borokensis*. Next, we studied the DR, which quantifies the trajectory straightness, ranging between 0 (for fully curved trajectories) and 1 (for fully straight trajectories), by considering the start and end point of the trajectory. The values ranged between 0.052 and 0.875 (median/IQR 0.525/0.307) for *A. proteus*, 0.043 and 0.871 (median/IQR 0.508/0.315) for *M. leningradensis* and 0.013 and 0.949 for *A. borokensis* (median/IQR 0.469/0.296).

Finally, we calculated the average speed (AS) of the trajectories, which ranged between 0.002 and 0.011 mm/s (median/IQR 0.006/0.002) for *A. proteus*, 0.002 and 0.012 mm/s (median/IQR 0.006/0.003) for *M. leningradensis* and 0.001 and 0.009 mm/s (median/IQR 0.005/0.002) for *A. borokensis*.

As it can be observed from the p-values derived from Kruskal-Wallis analyses there is a remarkable variability regarding kinetic properties, both between species (for example, the p-values comparing the IR, DR and AS in Scenario 4 were 10^{-13} , 10^{-4} and 10^{-16} , respectively) and between scenarios (for example, the p-values of *A. proteus* for IR, DR and AS compared among all four scenarios were 10^{-6} , 10^{-9} and 10^{-10} , respectively); for more information, see Fig. 6 and *SI Appendix*, Table S10.

Fig. 6D-G shows a clustering analysis performed on all kinematic properties considered. Each cluster was characterized by the proportion of cell types and experimental condition present in each group. The performance of the obtained clustering solution was assessed using the Silhouette Coefficient, which estimates all the differences between intra-cluster points minus the distances between inter-cluster points. A higher Silhouette index indicates a model with better defined clusters. The implementation was achieved using the silhouette score implemented in Scikit-Learn. The three clusters identified in Fig. 6 yielded a Silhouette coefficient of 0.366. Similarly, the four clusters identified provided a Silhouette coefficient of 0.369. Another alternative clustering analysis by means of a hierarchical agglomerative method (*SI Appendix*, Fig. S5) showed that the distinction between cell-types or experimental conditions remains unchanged when varying the clustering strategy, which indicates the robustness of the findings.

The high variability of the statistics of the kinematic parameters, combined with the cluster analysis, indicates a high level of heterogeneity in any of the three metrics used. This heterogeneity exists both among cell types and among different experimental conditions and suggests that the behavior is individual in each one and hence they cannot be separated into groups.

10. Dynamic structure in migratory movements.

Finally, we have represented all the main metrics considered in our study, such as RMSF Alpha, RMSF correlation time (measured in move steps or in minutes), DFA Gamma, MSD Beta, and Approximate Entropy by comparing them with those data obtained in the corresponding shuffling procedures (Fig. 7A-C). As evident in each panel, the metrics' shuffled and non-shuffled values could be distinctly grouped and differentiated. This suggests that the inherent systemic information structure was disrupted during the shuffling process. The trajectories of the experimentally observed cells are completely differentiated from the cells whose trajectories lost systemic properties.

In Fig. 7G-J a clustering analysis of the main metrics was performed. The three clusters related to cell type identified in Fig. 7 yielded a Silhouette coefficient of 0.347. Similarly, the four clusters identified related to experimental conditions provided a Silhouette coefficient of 0.311. This unsupervised clustering analysis combined to the small variability of the results, appears to be quite homogeneous, regardless the cell type or scenario considered, since its quantitative aspects show no dependency on either cell type or experimental condition. Moreover, we utilized an alternative approach, specifically hierarchical agglomerative clustering (*SI Appendix*, Fig. S6), and the resulted profile remained consistent when the different clustering methods were applied, highlighting the robustness of the results.

These findings suggest the emergence of a highly intricate dynamic structure within the migratory patterns of all examined cell trajectories. Furthermore, this structure appears to be an inherent aspect of cell locomotion, irrespective of species or environmental conditions. Indeed, cluster analysis of all quantitative parameters revealed no reliance on either cell type or experimental context, suggesting the potential universality of this behavior.

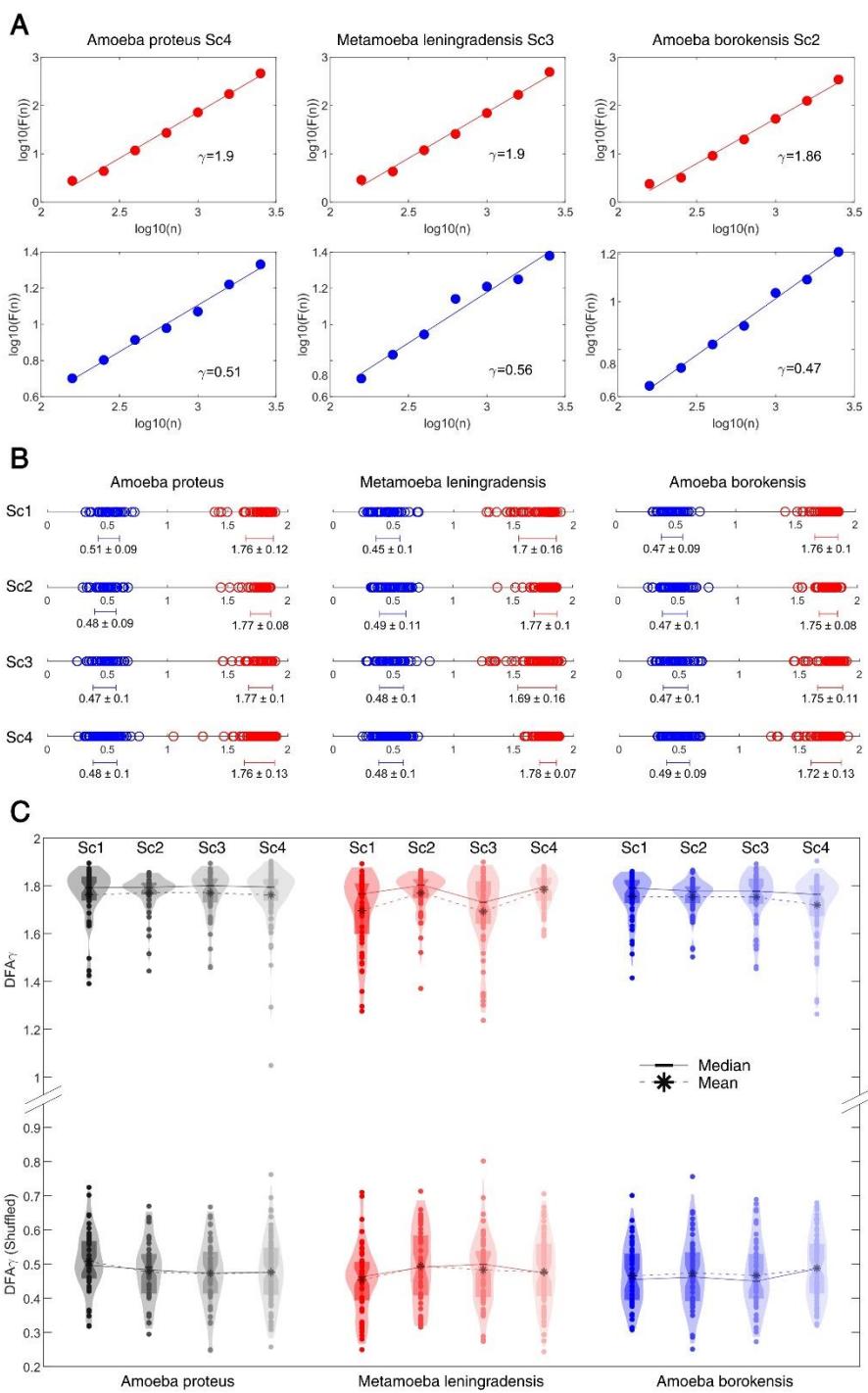


Fig. 5. Long-term memory effects in cellular migratory movements.

(A) Log-log plot of the detrended fluctuation parameter $F(n)$ versus window size n for a prototype cell from each species. The scaling exponent γ was $\gamma=1.9$ for the *A. proteus* cell, $\gamma=1.9$ for the *M. leningradensis* cell and $\gamma=1.86$ for the *A. borokensis* cell, indicating long-range memory effects in all the species. The shuffling procedure removed the memory information contained in the original trajectories, causing γ values to drop to $\gamma=0.51$ for the representative *A. proteus* cell, $\gamma=0.56$ for the *M. leningradensis* cell and $\gamma=0.47$ for the *A. borokensis* cell. (B) Diagram displaying all the values of the scaling exponent γ (and the overall average \pm SD) in all cells, separately for each species (*A. proteus*, *M. leningradensis* and *A. borokensis*) and experimental scenario (Sc1-Sc4). Values of the scaling exponent γ belonging to shuffled time series are depicted in blue, while exponents corresponding to experimental trajectories are depicted in red. (C) Violin plots illustrate the estimated distribution, mean and median scaling exponent γ values for all experimental (upper row) and shuffled (bottom row) cell trajectories. “Sc1” Scenario One, absence of stimuli; “Sc2” Scenario Two, presence of an electric field; “Sc3” Scenario Three, presence of a chemotactic peptide gradient and “Sc4” Scenario Four, simultaneous galvanotactic and chemotactic stimuli.

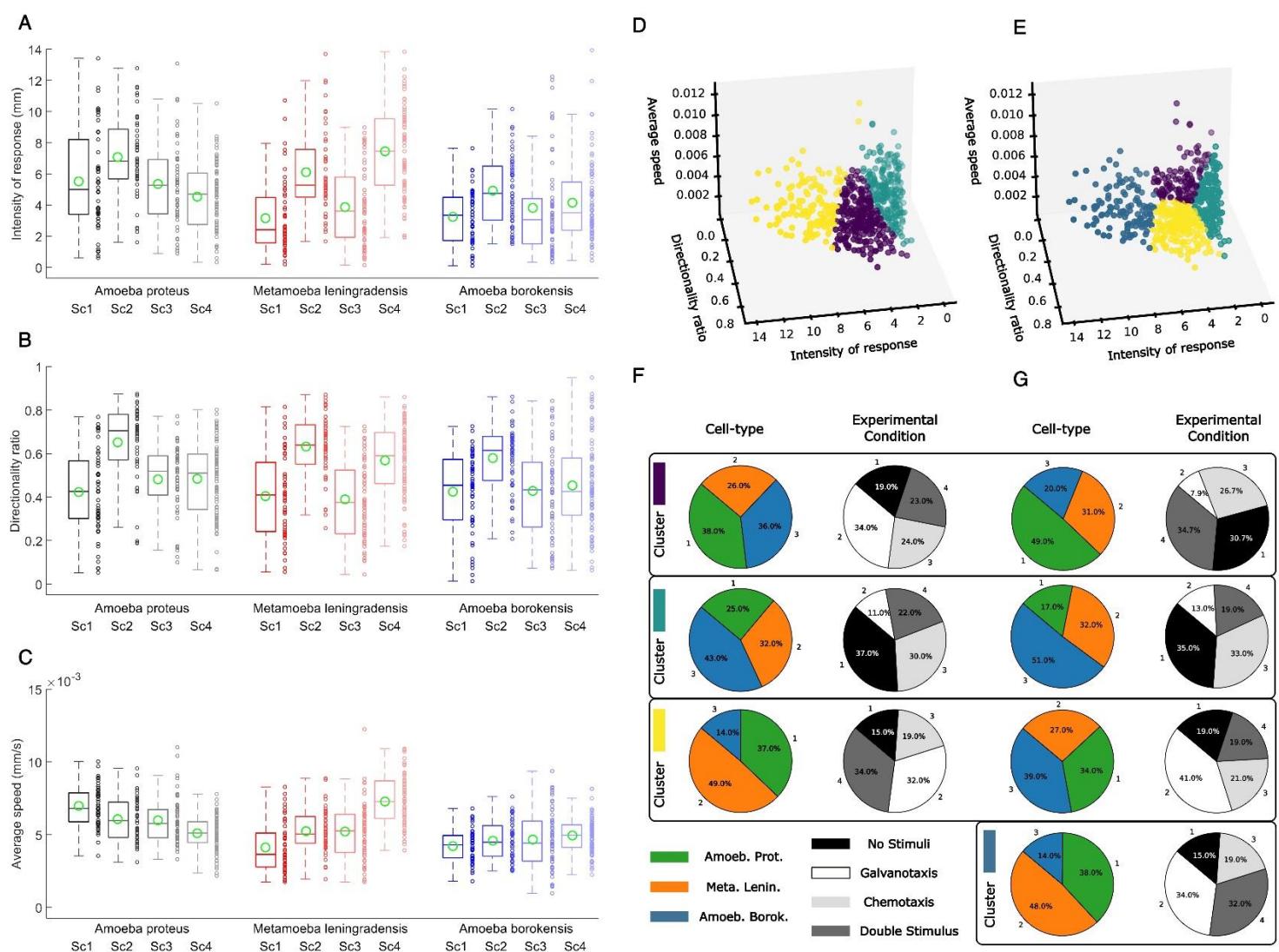


Fig. 6. Kinematic properties and clustering analysis of cellular locomotion trajectories.

(A-C) Group boxplots of the distributions of all experimental values for three main cytokinetic metrics, revealing basic data about the cell migration characteristics -(A) Intensity of the response, (B) Directionality ratio and (C) Average speed- of the three species used in this experiment (*A. proteus*, *M. leningradensis* and *A. borokensis*) under all four experimental scenarios (Sc1, Sc2, Sc3 and Sc4). (D-G) Unsupervised clustering was performed using *k-means* on experiments defined by 3D vectors of three metrics. (D) Three clusters are depicted (yellow, purple, and green), with each dot representing a different cellular motion experiment (combining three cell types and four experimental scenarios). (E) Similar to (D) but with four clusters. Note that the clusters in Panel (D) do not have the same colours as in Panel (E), although three of the four colours are the same. (F) Characterization of the three clusters (panel D) in terms of cell types and experimental conditions. (G) Similar to (F) but with four clusters (panel D).

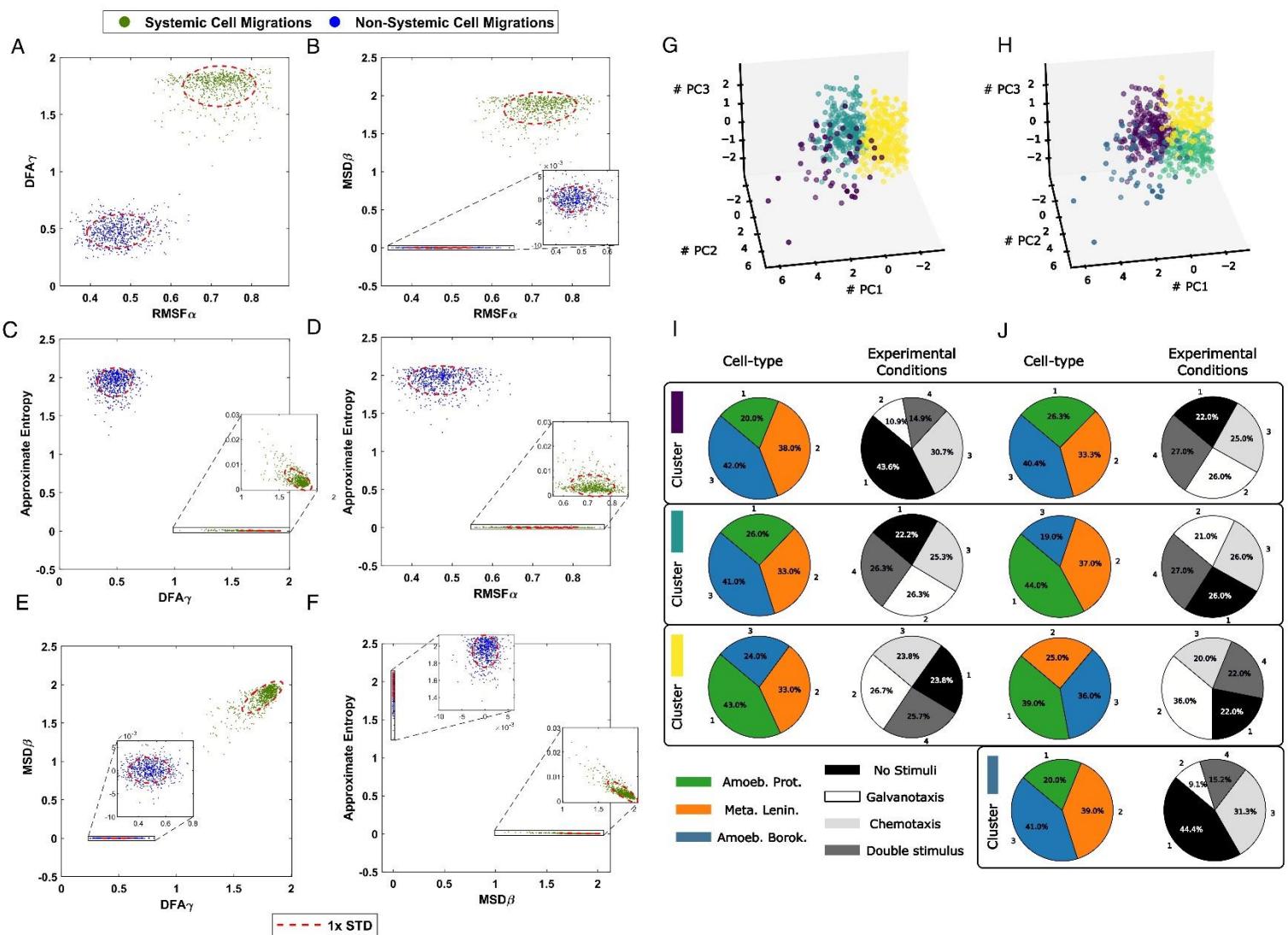


Fig. 7. Systemic metrics and clustering analysis of cellular migratory displacements.

(A-F) Analysis of the 700 experimental cell trajectories with the main metrics considered in our work such as RMSF Alpha, RMSF correlation time, DFA Gamma, MSD Beta, and Approximate Entropy is compared with corresponding shuffled data in which the systemic informational structure was completely lost. (G-J) Clustering analysis of cellular motion experiments does not distinguish between cell types and experimental conditions when using non-linear advanced movement metrics. Similar to Fig. 6 (D-G) but defining each experiment using the 5D main vectors of metrics. K-means clustering was performed using the first three principal components (PC1, PC2, and PC3), which accounted for 92.54% of the total variance. The interpretation of the different panels is the same as in Fig. 6, but the variables used for clustering are now different.

Discussion

Cellular migration is a cornerstone issue in many essential physiological and pathological processes. Despite its enormous relevance, how cells control their locomotion movements is a still unresolved question which represents a fundamental challenge in contemporary biology.

For years, the scientific attention has been focused on the individualized study of the diverse molecular parts involved in cell migration; however, locomotion movements have never been regarded as a systemic process that operates at a global cellular scale.

Here, we have addressed the integrative systemic dynamics involved in the regulation of directional motility. To this end, we have studied the migratory displacements of 700 single cells, belonging to three different species (*Amoeba proteus*, *Metamoeba leningradensis* and *Amoeba borokensis*), in four different scenarios: in absence of stimuli, under chemotactic gradient, in an electric field and under complex external conditions such as simultaneous galvanotactic and chemotactic gradient stimuli. The experimental trajectories, obtained on flat two-dimensional surfaces, have been quantitatively studied using a multidisciplinary approach to understand how integrative systemic forces drive the locomotion movement of cells.

First, we have analyzed the long-range interdependence in the move-steps using the “rmsf” method, a classical Statistical Mechanics approach based on the concepts developed by Gibbs and Einstein (39, 40), later developed and used to quantify biological processes (50–52). Our results demonstrate that each move-step ahead at a given point is strongly influenced by its preceding displacements, indicating that strong dependences of past movements lasting approximately 1,137 move-steps over periods averaging 9.5 minutes do exist. Practically, all the 700 unicellular organisms analyzed in the four experimental conditions exhibited non-trivial long-range correlations in their directional trajectories, which represents a key characteristic of the systemic dynamic movements emerging in the cell system (47).

Anomalous behavior is another characteristic of the migratory movements that we have identified using the MSD method also proposed by Einstein (43). Migratory dynamics that do not result in a linear MSD can be considered as non-trivial. Specifically, the anomalous nature of cell migration can be detected by super-diffusion, a physical phenomenon detected in the trajectories of all the 700 cells analyzed. Likewise, the MSD is a proxy for the surface area explored by the cell over time and is a measure related to the overall migration efficiency. The cellular displacements analyzed correspond to efficient movements during the exploration of the extracellular medium (41, 42, 53). The strong manifestation of the anomalous nature of cell migration can be caused by temporal memory effects as a consequence of the non-trivial long-range correlations in the cellular move-steps (53–56).

We have also quantified the regularity and unpredictability of the fluctuations over the migratory displacements (45, 46). The obtained results show high levels of information in all the analyzed trajectories. This finding, together with the previous ones, confirms the presence of a very complex structure in the migratory move-step sequences. Entropy is directly related to the complexity of the system dynamics, and the very low level of entropy in the directional movements indicates that the migration patterns are organized on a level of complexity that is above the individual components of the cell system. Such complex

dynamic structure observed in the trajectories was highly unlikely to occur by chance (practically, p-value $\cong 0$).

In addition, we have verified the presence of long-term memory effects in the cellular migratory movements. The results of the DFA fluctuation analysis (57) show that the scaling exponent γ displays a total average (\pm SD) of 1.749 ± 0.117 , indicating that the move-steps trajectories exhibit a trend-reinforcing memory, that is, if the directional movements in the past show an increase in a set of their move-step values it is very likely to be followed by an increasing trend in the future; and vice versa, a decreasing trend in the past, is likely to be continued in the future. In other words, the evolution of the cell system trajectories is strongly influenced by previous system movements over long periods of time. Long-term memory is a key concept, closely related to long-term correlations, widely developed in Physics with a robust and formal Mathematical construction. Therefore, the results we have obtained with DFA also validate the presence of strong long-range correlations in the locomotion movements.

The quantitative studies carried out here unequivocally show that a very complex dynamic structure emerges in the migratory movements of all the analyzed cell systems. Such structure is characterized by highly organized move-step sequences with very low level of entropy and high information, non-trivial long-range interdependence in the move-steps, strong anomalous super-diffusion dynamics, long-term memory effects with trend-reinforcing behavior, and efficient movements to explore the extracellular medium. The outstanding detected dynamic structure underlies all the migration trajectories of 700 cells of three different species analyzed under the four experimental scenarios. On the other hand, the results of the two types of clustering analysis performed suggested the potential universality of this complex systemic structure in the cellular locomotion movements.

These essential characteristics of the locomotion movements are a consequence of the self-organized dynamics intrinsic to all unicellular organisms. Cells are sophisticated systems conformed by the mutual interactions of millions of molecules and hundreds of thousands of macromolecular subcellular structures, following an intricate interplay that challenges the human mind (58). They are open systems that operate far from the thermodynamic equilibrium and exchange energy-matter with the environment (59–61). Under these conditions, non-linear enzymatic interactions and irreversible metabolic processes allow the cell system to become spatially and temporally self-organized (58, 62–66). If cells reach the equilibrium, their sophisticated dynamic functionality and molecular order disappears and they die.

Briefly described, the essential energy-matter flow generates a negative entropy variation inside the cell which corresponds to an emergent positive increment in the information of the system (67). Such information increases the complexity, producing collective functional patterns, highly ordered macrostructures, and complex self-organized behaviors as for instance molecular-metabolic rhythms and spatial traveling waves (61, 68).

These emergent non-equilibrium molecular dynamics supported by permanent energy dissipation (continuously exporting entropy to the external medium) are known as self-organized dissipative structures (69, 70). The principles of self-organization through energy dissipation were conceived and developed by the Nobel Prize Laureate in Chemistry Ilya Prigogine (38).

Intensive studies over the last six decades have demonstrated that cells are very complex self-organized dissipative systems (58, 60, 66, 67, 71–73) in which integrated processes and systemic properties at different levels of organization and complexity do appear. At a basic level, dissipative molecular behaviors emerge, for example, in shaping actin polymerization waves involved in the cytoskeleton activities during cell migration (74, 75), in self-organized oscillations in actin networks (76), in myosin dynamics (77), in microtubular behavior (78), and in intracellular calcium rhythms (79). At the highest level, complex systemic properties such as directional mobility, integral growth, reproduction, sensitivity to the external medium, adaptive responses, and evolution do occur (58). To note, these strong emergent properties cannot be found in their individual molecular components or in their single molecular-metabolic processes (66).

Systemic dynamics are emergent integrative processes in all unicellular organisms, and such behaviors are a consequence of the self-organization of the biochemical system as a whole (80). From collective metabolic-molecular constituents, all of them interacting nonlinearly with each other, emerge basic coherent self-organized structures and functional ordered patterns which originate a cell system that increases at different levels its structural and functional complexity driven by energy dissipation and molecular information processing (58, 63, 64).

A critical attribute of these dissipative self-organized systems is the interacting dynamics exhibiting long-range correlations (81–83). In his Nobel Lecture dissertation, Ilya Prigogine stressed that the main feature of systemic dynamics in dissipative systems is the emergence of long-range correlations (38).

On the other hand, long-term correlations have also been showed in different metabolic processes such as calcium-activated potassium channels (84), intracellular transport pathway of Chlamydomonas (85), glycolytic studies (86-89), NADPH series (90), and metabolic networks (91,92), attractor dynamics (93), neural activity (94). In additions, complex emergent systemic behaviors have been observed in cellular locomotion movements (95,96).

Until now, cell migration has never been considered as a systemic property, but in this work we have shown evidences that indicate the existence of complex integrative dynamic processes at a global cellular scale involved in directional motility.

First, we have highlighted how a relevant number of experimental studies have verified that most of the fundamental cellular physiological-molecular activities in the cell are implicated in their locomotion movements, which is indicative of a phenomenon that operates at a systemic level.

We have also collected previous evidences found over the last decades showing that unicellular organisms constitute self-organized systems characterized by emergent molecular-metabolic processes and integrative responses operating at a systemic level. These self-organized processes are mainly characterized by exhibiting coherent behaviors with long-range correlations, a ubiquitous characteristic of all dissipative systems.

Our quantitative analysis has shown that all cell systems analyzed here display a kind of dynamic migration structure characterized by non-trivial long-range correlations, strong anomalous super-diffusion dynamics, long-term memory effects, highly organized move-step sequences with very low level of entropy and high information, and efficient movements

to explore the environment. Such characteristics correspond to critically self-organized systems. The locomotion trajectories change continuously, since they exhibit random magnitudes that vary over time, but these stochastic movements shape a dynamic structure whose defining characteristics are preserved in all the conditions analyzed. This movement structure corresponds to complex behavior belonging to a self-organized cell system, in which the emergent systemic dynamics drive the locomotion movement of cells.

Cellular locomotion seems to be regulated by complex integrated self-organized dynamics, carefully regulated at a systemic level, which depends on the cooperative non-linear interaction of most, if not all, cellular components. The systemic properties responsible of this locomotor behavior are not found specifically in any of their singular molecular parts, partial mechanisms, or individual processes in the cell.

Our quantitative experimental results together with the remarkable amount of evidences provided by other scientific analyses seem to indicate that migration is a systemic property of cells. This fact does not invalidate the importance of studying the influence of individual metabolic-molecular pathways on cell migration.

Cell migration is a central issue in many human physiological and pathological processes. We consider that new researches combining migratory systemic dynamics with molecular studies are crucial for the development of next-generation, efficient cellular therapies for migration disorders. In addition, the findings presented here open a new perspective for improving the conceptual framework of the cell, the most complex molecular system in Nature.

Materials and Methods

Cell cultures

The three species of free-living amoebae were cultured in Ø100 x 20 mm Petri dishes (Corning® CLS430167) at 21°C. *Amoeba proteus* (Carolina Biological Supply, #131306) and *Metamoeba leningradensis* (CCAP: Culture Collection of Algae and Protozoa, Oban, Scotland, United Kingdom, catalogue number 1503/6) were grown in Chalkley's simplified medium (36) (NaCl, 1.4 mM; KCl, 0.026 mM; CaCl₂, 0.01 mM) with heat-treated wheat grains. *Amoeba borokensis* (ACCIC (97): Amoebae Cultures Collection of Institute of Cytology in Saint Petersburg) were grown in Prescott & Carrier media (98) and fed every fourth day 0.5 ml of a mixed *Chilomonas* sp. (Carolina Biological Supply® #131734) and *Colpydium* sp. (ACCIC culture. *Chilomonas* sp. and *Colpydium* sp. were grown in Prescott & Carrier media as well.

Experimental set-up

A schematic drawing of the experimental set-up is shown in *SI Appendix*, Figs. S1 and S2. Two electrophoresis blocks (BIO-RAD Mini-Sub cell GT) were interconnected by two ~12 cm long agar bridges (2% agar in 0.5 N KCl in) and the first one was plugged into a BIO-

RAD Model 1000/500 power supply unit. Atop the central elevated platform of the second block, a custom-devised experimental glass chamber was placed. By using agar bridges and removing all the electrodes from the second electrophoresis block we avoided direct contact between the media, where the cells moved, and any metallic electrodes, thus preventing ionic contamination.

The experimental glass chamber consisted of a modified 25 x 75 x 1 mm standard glass slide and three additional, smaller glass pieces (one central piece measuring 24 x 3 x 0.17 mm and two flanking pieces of 24 x 40 x 0.17 mm each) crafted by carefully trimming three 24 x 60 x 0.17 mm cover glasses (SI Appendix, Figs. S1 and S2). To build the modified standard glass slide, two 24 x 60 x 0.1 mm cover glasses were stuck using silicone to a standard glass slide and let dry for 24h, whereupon the 20 x 60 x 0.17 mm overhanging parts from both cover glasses were cut off, leaving two 4 x 60 x 0.17 mm glass sheets as the chamber's sideway walls. All the experiments were conducted within the experimental glass chamber, which allowed to establish a laminar flow when closed and to place or extract the cells when opened.

Fresh agar bridges were used in every experimental replica to avoid contamination and conductivity loss (leak of KCL into the simplified Chalkley's medium, which has a much lower osmotic concentration than the agar bridges). Furthermore, one electrophoresis block was used exclusively for chemotaxis and another one for galvanotaxis, while the whole set-up (both the electrophoresis blocks and the glass chamber) was scrupulously cleaned and reassembled after each experimental replica.

Ahead of every experiment, the modified glass slide was adhered to the top of the central platform of the second electrophoresis block using a droplet of olive oil to prevent both the medium and the electric current from passing beneath the experimental glass chamber. Following this, the 24 x 3 x 0.17 mm central glass piece was gently placed atop the middle of the modified glass slide. Next, amoebae were washed in fresh simplified Chalkley's medium and placed beneath the central piece of the glass chamber, where they were left to attach to the surface of the modified glass slide for ~2 minutes. This step needs to be performed in less than 15 seconds, for the amoebae will immediately start attaching to the inner surface of the plastic micropipette tip, and any further pipetting will henceforth potentially damage their cellular membrane. *Metamoeba leningradensis* cells attached faster and stronger to the plastic micropipette tips than the other two species, thus being especially susceptible to being damaged. Subsequently, the two 24 x 40 x 0.17 mm lateral sliding glasses were laid flanking the central piece and slightly overhanging the electrophoresis block wells. Each well was carefully filled with 75 ml of clean simplified Chalkley's medium, and the two 24 x 40 x 0.1 mm lateral sliding glasses were gently poked down using a micropipette tip until they contacted the medium, which spread beneath them by surface tension. Finally, the two lateral glass pieces were longitudinally slid till they touched the central piece where the amoebae laid, thus closing the glass chamber, and establishing a connection between the media in both wells.

All experimental replicates were performed with a maximum 9 cells and lasted for exactly 30 minutes, during which cell behavior was recorded using a digital camera. In preparation for each experiment, amoebae were starved for 24 hours in fresh, nonnutritive simplified Chalkley's medium. Only cells deemed healthy (motile and rod-shaped) were chosen for the experiments. Deviations from optimal culture and experimental conditions, as well as mechanical issues in the recording system, rendered $\leq 9\%$ of the experimental replicates invalid. Those replicates have not been considered in this investigation.

Galvanotactic stimulus

The galvanotactic stimulus is traumatic for the amoebae, and thus can affect their response. Four key steps were taken to minimize those effects by ensuring optimal current intensity and voltage in all the experiments where a galvanotactic stimulus was applied: 1. The power supply unit was programmed to keep the voltage at 60V; 2. The flow sectional area of the experimental glass chamber was adjusted by modifying the amount of silicone used to glue the longitudinal walls of the modified glass slide, which determined their height; 3. A variable $1\text{ M}\Omega$ resistor and a microammeter were installed in series, in that order (*SI Appendix*, Fig. S3), and the intensity of the electric current was manually corrected when needed in real time by turning the variable resistor's screw to tune the global resistance of the set-up; thus, a stable 60V electric potential and optimal current intensity values of 70-74 μA (*A. proteus*), 70-80 μA (*M. leningradensis*) and 68-75 μA (*A. borokensis*) were kept throughout the galvanotactic experiments; 4. Lastly, the current was stopped immediately once the 30-minute image recording finished.

We found some amoebae populations displayed an anomalous, inverted or even null response to the galvanotactic stimulus. We therefore carried out a 5-minute galvanotactic test prior to any experiments where amoebae were exposed to galvanotaxis for the first time, using intensity and voltage values within the optimal ranges provided.

Chemotactic stimulus & gradient calculation

To establish the chemotactic peptide gradient, we added 750 μl of $2 \times 10^{-4}\text{ M}$ nFMLP (Sigma-Aldrich, #F3506) to one well of the second electrophoresis block to obtain a working peptide concentration of $2 \times 10^{-6}\text{ M}$. The medium in that well was stirred immediately after adding the peptide to properly mix the peptide until the amoebae began to respond to it.

To assess the nFMLP peptide gradient concentration, an experiment was carried where 60 μl of medium were sampled at 0, 2, 5, 10, 15, 20, and 30 minutes since the addition of the nFMLP peptide. Samples were taken from the center of the experimental glass chamber through a small gap opened between the central glass piece and one of its flanking glass pieces by slightly moving this sliding lateral glass. Known fluorescein-tagged peptide concentration values from a standard curve were used to extrapolate nFMLP concentration at each time point (*SI Appendix*, Fig. S4). Two samples were taken at each time point, and the whole procedure was replicated three times, yielding a total of 6 measurements for each time point. Fluorescence was measured at 460/528 excitation/emission wavelengths on 96 well glass bottom black plates (Cellvis, #P96-1.5H-N) using a SynergyHTX plate reader (BIOTEK) following the protocol established by Green and Sambrook (99).

Experiment recording and cell tracking

Experiments were recorded with a digital camera attached to an SM-2T stereomicroscope. Two frames were captured every second for 30 minutes (3600 frames). Individual cell movements (tracks) were manually digitized using the TrackMate (100) plugin from FIJI (ImageJ) and saved as a list of (x, y) coordinate tuples. Manual tracking was chosen over automated and semi-automated alternatives due to the well-known imprecision of such tools (101).

Approximation Entropy calculation

Approximation Entropy (ApEn) is a statistic developed by Pincus et al. in 1991 (45) as a measure of relative regularity in data series. In Fig. 4A, Approximation Entropy ApEn values corresponding to intervals of length <300 data points were not represented in the heatmaps,

as ApEn requires at least 10^m data points ($m=2$ in our calculations) to produce meaningful results (45), and these are known to be unreliable in time series of length ≤ 200 points (102).

Clustering numerical experiments

The clustering analyses were implemented using our custom code in Python 3.9.13 and Scikit Learn 1.0.2. Combining all experiments across the three cell types (*M. leningradensis*, *A. Proteus*, and *A. borokensis*) and the four experimental conditions (no stimuli, galvanotaxis, chemotaxis, and simultaneous galvanotactic and chemotactic stimuli), unsupervised clustering was performed on all experiments of cellular movement, each defined by a vector of different movement metrics (MM). In the first clustering analysis (Fig. 6), experiments were characterized by the 3D vector of MMs: Intensity of the response (mm), Directionality Ratio, and Average Speed (mm/s). In the second clustering analysis (Fig. 6), experiments were identified using the 5D vector of MMs: RMSF Alpha, RMSF correlation time (move-steps), DFA Gamma, MSD Beta, and Approximate Entropy. Before performing the clustering analysis, and to ensure a comparable visualization to the first analysis, we first applied Principal Component Analyses (PCA) and subsequently applied the clustering strategy to the first three components (PC1, PC2, and PC3), which explained the 92.54% of the total variance in the data. For the second analysis, the clustering results did not change when using RMSF correlation time measured in minutes instead of move steps, as both metrics are highly correlated. Although the best clustering solution (measured by the Silhouette Coefficient) was obtained in both analyses for the number of two clusters, Figs. 6 and 7 display 3 and 4 clusters with the purpose of exploring potential associations with, respectively, cell type and experimental condition. After normalizing the different values using z-scores across all possible experiments, we applied the *k-means* algorithm to obtain 3 and 4 clusters. Subsequently, each cluster was characterized by the proportion of cell types and experimental condition present in each cluster. The performance of the obtained clustering solution was assessed using the *Silhouette Coefficient*, which estimates all the differences between intra-cluster points minus the distances between inter-cluster points. A higher Silhouette index indicates a model with better defined clusters, while a smaller index suggests more heterogeneity in the data, and no well-separated groups. The implementation was achieved using the silhouette score implemented in Scikit-Learn library in Python. To show robustness of the clustering solution and that our results were not dependent on the clustering strategy employed, we repeated the same analyses as in Figs. 6 and 7 but using *hierarchical agglomerative* clustering and depicting the solutions of 3 and 4 clusters (SI Appendix, Figs. S5 and S6).

Statistical analysis

First, the normality of the distribution of our quantitative data was assessed using the Kolmogorov-Smirnov test for single samples. Given that normality was rejected, the significance of our quantitative results was estimated through the Kruskal-Wallis test for groups and the Wilcoxon rank-sum test for pairs. Since these two are non-parametric tests, results are represented as median/IQ instead of mean \pm SD. In addition to the p-values the Z statistics have been reported.

Data and code availability

The data and code generated by this study are publicly accessible from the Zenodo repository at <https://doi.org/10.5281/zenodo.10974258>.

Any additional information required to reanalyze the data reported in this paper is available from the lead contact upon request.

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Author contributions

JC-P, and MF: performed the experiments; CB, MF and IMDF: designed the setup; CB methodology with experimental glass chamber; MF and JC-P: cell cultures and cellular behavior advice; JC-P: performed the digitalization of trajectories; IM, JMC, IMDF: designed quantitative analysis; JC-P, IM, JCM and BC-P: performed the quantitative studies; GPY, JIL, AP-S, JC-P, IM, JMC and IMDF: analysis and design of the research mapping; all authors wrote the manuscript and agreed with its submission; IMDF: conceived, designed and directed the investigation.

Declaration of interests

The authors declare no competing interests.

Supporting information

Appendix 01 (PDF)

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Supplemental information

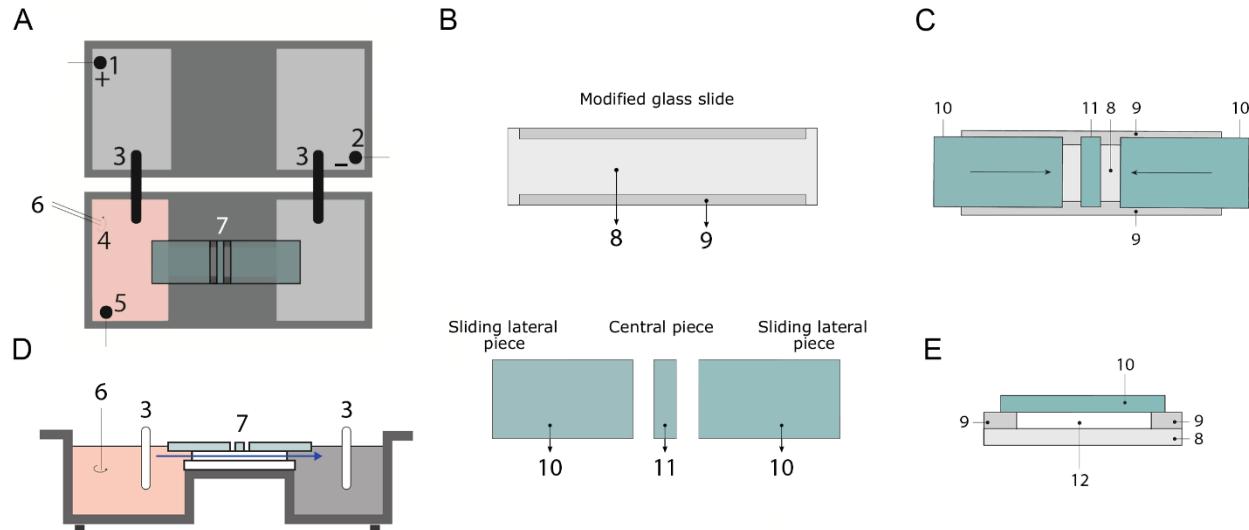


Figure S1. Experimental set-up layout.

(A and D) Top and lateral views of the experimental setup. 1: anode; 2: cathode; 3: agar + KCl bridges; 4: chemotactic peptide; 5: electrode used to probe the electric field; 6: stirrer used to properly mix the peptide; 7: experimental glass chamber, the blue arrow signals the trajectory and direction of the laminar flow. (B) Top view of the glass pieces that compose the experimental glass chamber. 8: 75 × 25 mm standard glass slide; 9: longitudinal trimmed glasses; 10: sliding lateral glasses; 11: central piece of glass underneath which the cells are placed. (C) Top view of the experimental glass chamber. (E) Axial section of the experimental chamber. 12: flow sectional area. The experimental chamber can be opened and closed by longitudinally displacing #10, allowing to place or remove cells when open and establishing a laminar flow of medium through #12 when closed (See *Materials and Methods* for further details).

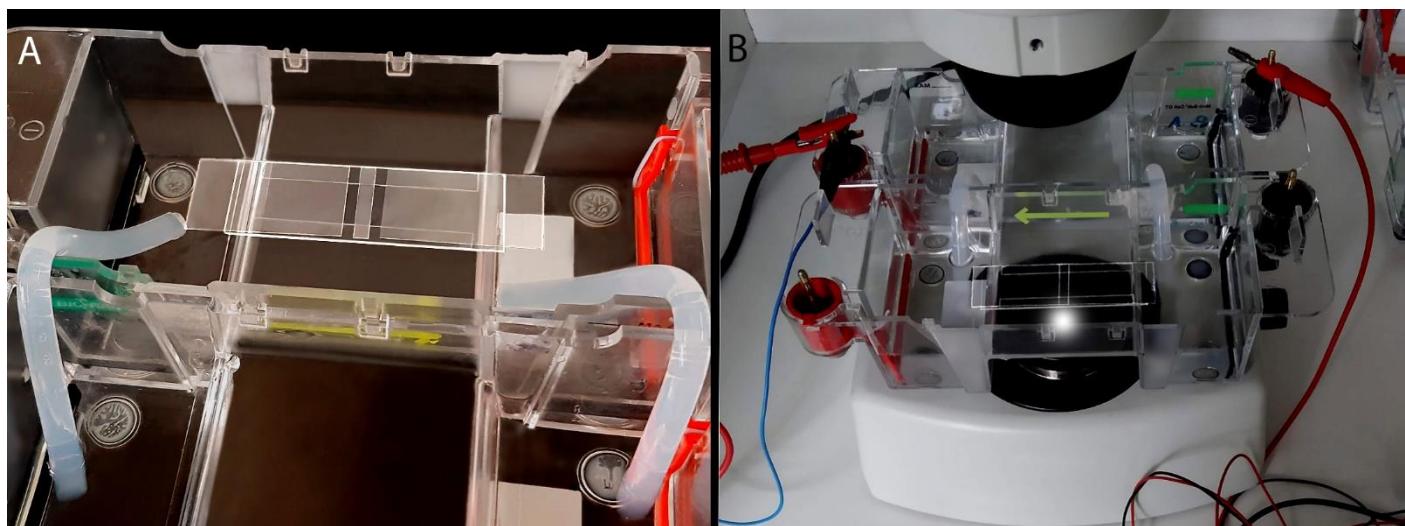


Figure S2. Experimental set-up, related to Figure S1.

(A and B) All elements sketched and described in Figure S1 are depicted here under experimental conditions.

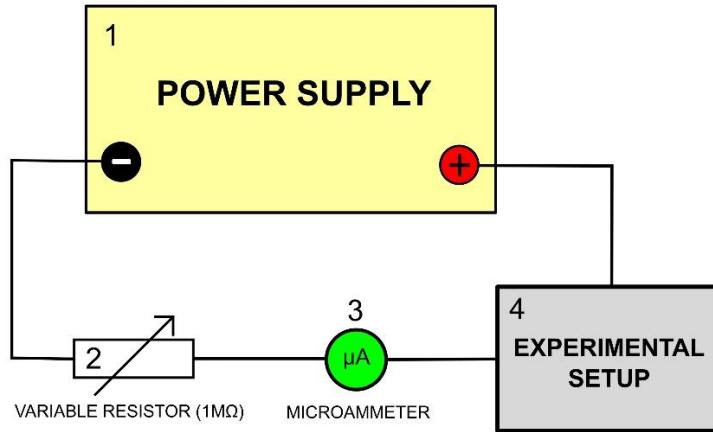


Figure S3. Auxiliary electric circuitry.

This circuit allows the electric current that circulates through the experimental system to be monitored and adjusted as necessary. 1: BIO-RAD model 1000/500 power supply unit; 2: $1\text{ M}\Omega$ variable resistor to adjust the intensity of the electric current; 3: Microammeter to monitor current intensity flowing through the system; 4: Experimental set-up (see Figures S1 and S2 for more detail).

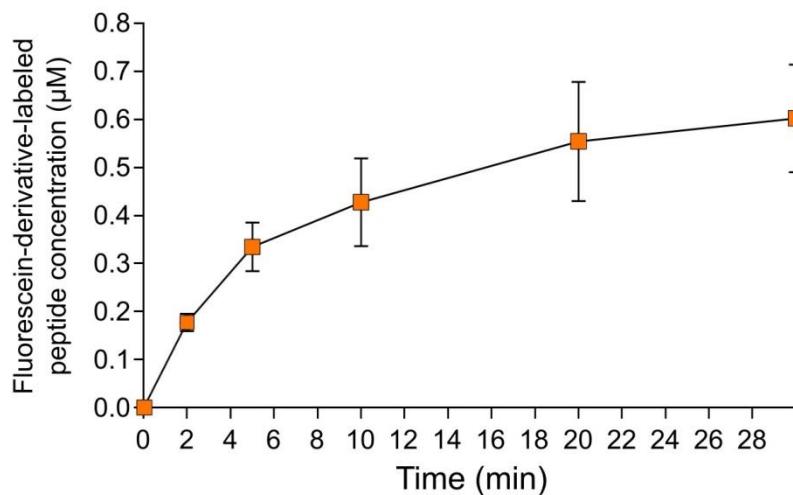


Figure S4. Peptide gradient concentration measurement.

Average fluorescein-tagged peptide concentration as a function of time, measured in the center of the experimental glass chamber at 0, 2, 5, 10, 20 and 30 min. Each data point represents the average ($\pm \text{SD}$) of six measurements (duplicate sampling in three separate experimental replicates) taken at the location where the amoebae were placed. Peptide concentration rises to $\sim 0.2\text{ }\mu\text{M}$ two minutes after the laminar flow is established, and further to $0.6\text{ }\mu\text{M}$ towards the end of the experiment (30 min since laminar flow is established).

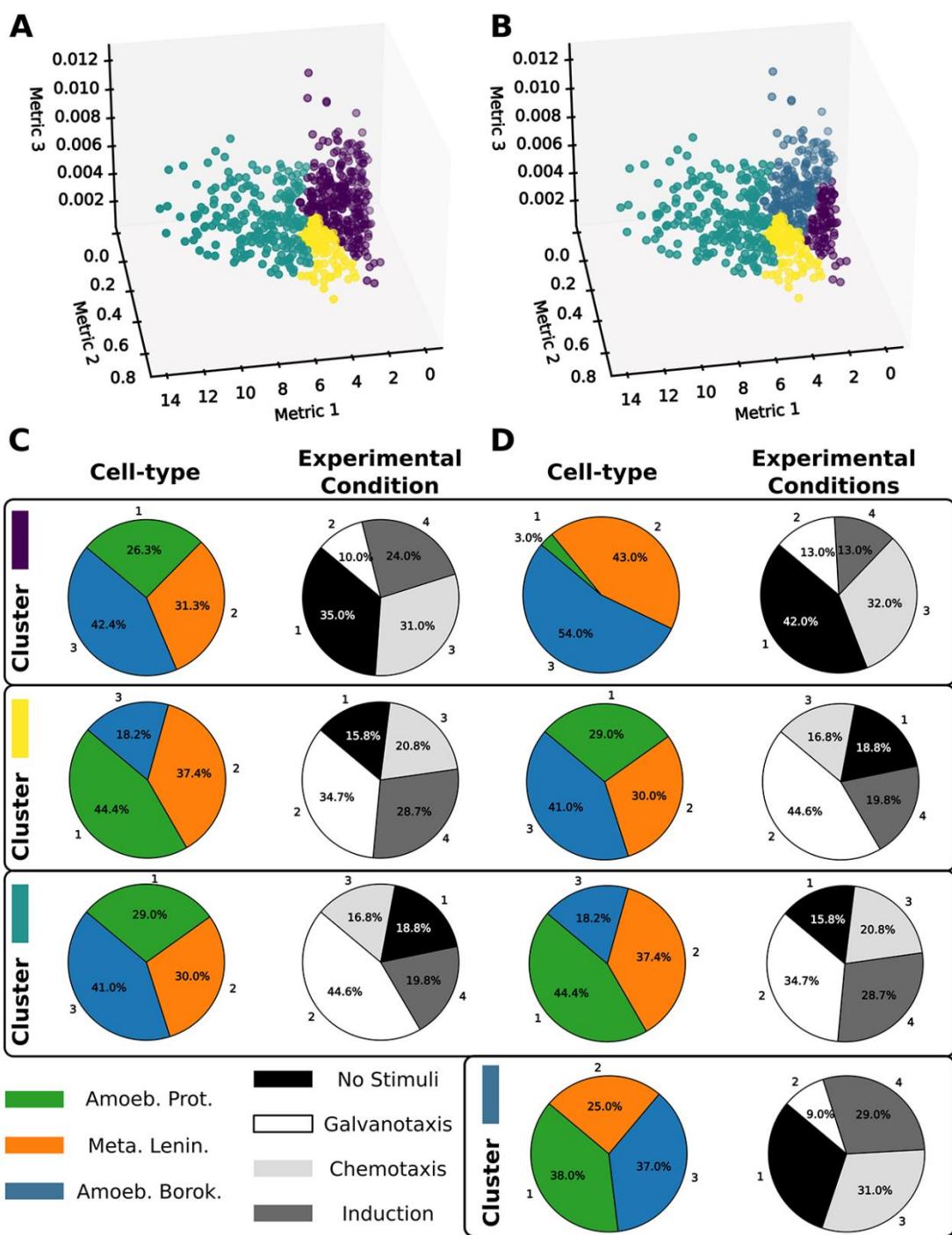


Figure S5: Similar to Fig. 6, but employing a different clustering strategy, namely hierarchical agglomerative clustering. The clustering characterization regarding the distinction between cell-types or experimental conditions remains unchanged when varying the clustering strategy, indicating the robustness of the findings.

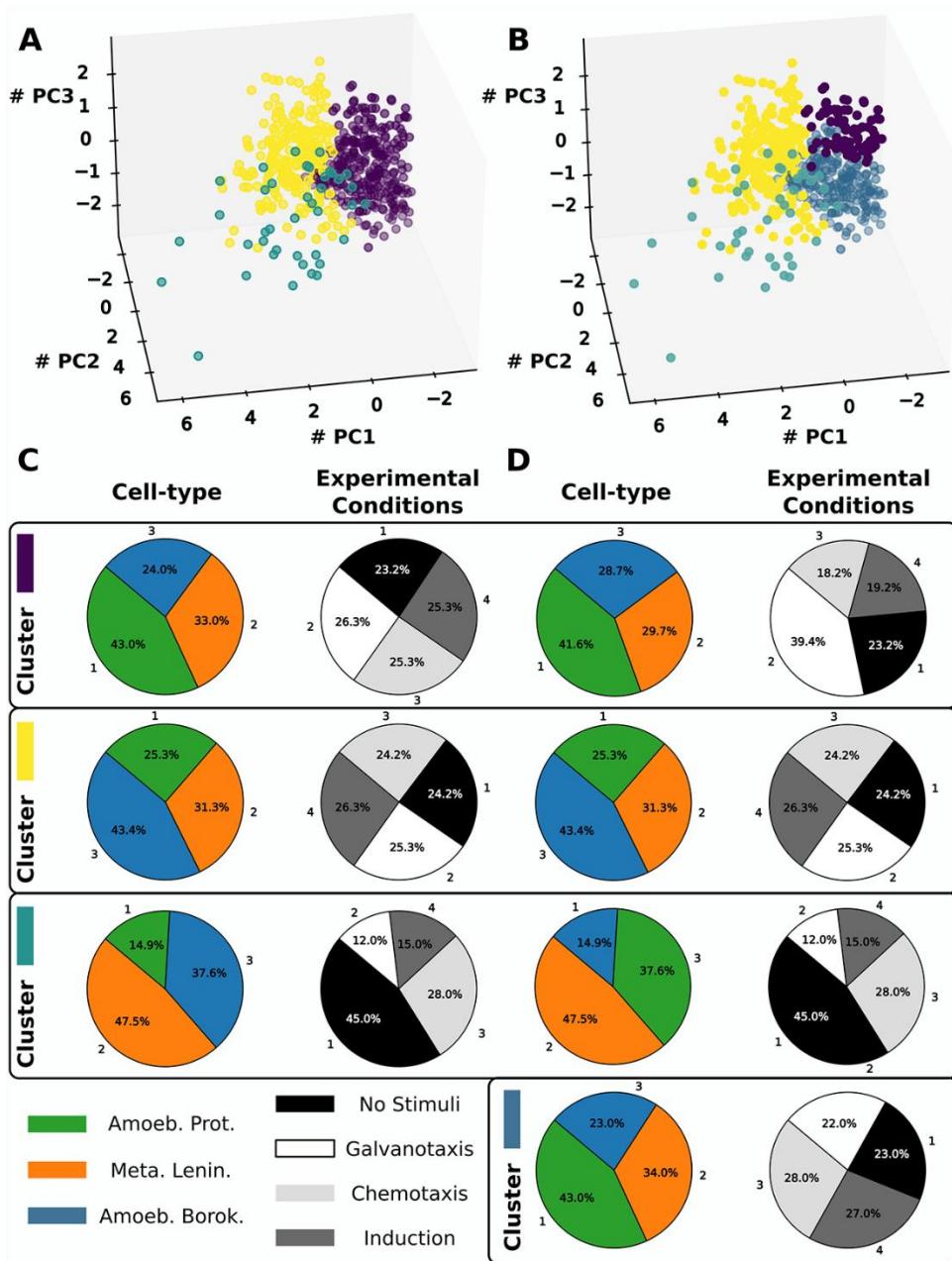


Figure S6: Similar to Fig. 7, but employing a different clustering strategy, namely hierarchical agglomerative clustering.

Table S1

Cell number	Amoeba proteus				Metamoeba leningradensis				Amoeba borokensis			
	Sc1	Sc2	Sc3	Sc4	Sc1	Sc2	Sc3	Sc4	Sc1	Sc2	Sc3	Sc4
1	0.791	0.718	0.783	0.682	0.808	0.653	0.797	0.755	0.684	0.645	0.645	0.670
2	0.748	0.774	0.693	0.733	0.853	0.788	0.731	0.812	0.648	0.757	0.636	0.691
3	0.748	0.677	0.846	0.666	0.708	0.679	0.647	0.796	0.658	0.681	0.747	0.746
4	0.709	0.562	0.753	0.723	0.737	0.666	0.763	0.778	0.631	0.641	0.679	0.781
5	0.837	0.747	0.707	0.706	0.736	0.620	0.755	0.846	0.758	0.629	0.686	0.694
6	0.740	0.668	0.607	0.800	0.844	0.684	0.727	0.717	0.663	0.769	0.622	0.750
7	0.672	0.621	0.650	0.750	0.695	0.759	0.714	0.730	0.739	0.765	0.710	0.684
8	0.811	0.720	0.707	0.835	0.817	0.721	0.814	0.715	0.639	0.759	0.758	0.782
9	0.719	0.707	0.708	0.787	0.756	0.642	0.828	0.766	0.724	0.737	0.722	0.731
10	0.812	0.666	0.715	0.653	0.686	0.711	0.787	0.761	0.782	0.734	0.743	0.733
11	0.703	0.693	0.568	0.804	0.721	0.633	0.685	0.771	0.689	0.731	0.732	0.784
12	0.611	0.714	0.699	0.710	0.749	0.676	0.783	0.741	0.642	0.691	0.705	0.670
13	0.715	0.691	0.773	0.756	0.748	0.691	0.653	0.604	0.750	0.606	0.713	0.775
14	0.698	0.686	0.790	0.782	0.674	0.771	0.757	0.729	0.682	0.698	0.689	0.758
15	0.599	0.772	0.757	0.682	0.784	0.807	0.747	0.643	0.714	0.585	0.632	0.723
16	0.736	0.796	0.638	0.700	0.707	0.739	0.803	0.738	0.738	0.709	0.688	0.740
17	0.652	0.653	0.782	0.799	0.604	0.557	0.710	0.801	0.677	0.762	0.743	0.680
18	0.635	0.727	0.677	0.771	0.681	0.734	0.733	0.605	0.689	0.746	0.707	0.607
19	0.560	0.637	0.757	0.721	0.799	0.685	0.714	0.612	0.721	0.734	0.684	0.644
20	0.757	0.693	0.719	0.738	0.763	0.663	0.732	0.715	0.606	0.683	0.687	0.746
21	0.593	0.700	0.806	0.664	0.796	0.699	0.787	0.723	0.628	0.590	0.774	0.680
22	0.753	0.785	0.748	0.764	0.822	0.640	0.768	0.690	0.628	0.685	0.732	0.796
23	0.785	0.690	0.668	0.714	0.753	0.708	0.660	0.695	0.648	0.707	0.624	0.707
24	0.641	0.609	0.648	0.731	0.781	0.790	0.674	0.696	0.680	0.710	0.684	0.655
25	0.776	0.713	0.775	0.785	0.748	0.648	0.760	0.756	0.661	0.618	0.721	0.716
26	0.669	0.691	0.761	0.664	0.739	0.783	0.773	0.729	0.622	0.797	0.719	0.696
27	0.615	0.690	0.679	0.771	0.764	0.666	0.656	0.780	0.663	0.641	0.735	0.764
28	0.778	0.722	0.754	0.656	0.824	0.685	0.754	0.716	0.674	0.714	0.599	0.617
29	0.749	0.799	0.722	0.759	0.708	0.732	0.749	0.692	0.657	0.646	0.728	0.688
30	0.693	0.754	0.707	0.804	0.682	0.838	0.726	0.789	0.615	0.705	0.743	0.671
31	0.595	0.734	0.732	0.726	0.746	0.713	0.665	0.666	0.704	0.593	0.704	0.685
32	0.599	0.580	0.752	0.774	0.767	0.682	0.597	0.731	0.796	0.632	0.721	0.777
33	0.568	0.642	0.723	0.703	0.720	0.782	0.578	0.669	0.676	0.580	0.830	0.674
34	0.744	0.740	0.718	0.746	0.679	0.633	0.721	0.737	0.685	0.712	0.718	0.746
35	0.745	0.710	0.687	0.769	0.764	0.732	0.627	0.730	0.666	0.610	0.681	0.688
36	0.683	0.770	0.731	0.749	0.806	0.725	0.657	0.710	0.743	0.577	0.710	0.664
37	0.737	0.768	0.729	0.726	0.807	0.792	0.645	0.692	0.772	0.694	0.768	0.666
38	0.740	0.784	0.781	0.676	0.714	0.764	0.843	0.721	0.694	0.815	0.771	0.729
39	0.707	0.732	0.729	0.844	0.793	0.695	0.761	0.646	0.746	0.646	0.807	0.736
40	0.732	0.636	0.708	0.721	0.809	0.742	0.739	0.681	0.715	0.700	0.730	0.694
41	0.716	0.774	0.728	0.749	0.756	0.794	0.783	0.703	0.765	0.786	0.781	0.783
42	0.778	0.745	0.789	0.700	0.699	0.696	0.759	0.662	0.595	0.601	0.728	0.702
43	0.657	0.660	0.627	0.695	0.743	0.715	0.802	0.600	0.639	0.604	0.782	0.697
44	0.577	0.688	0.742	0.762	0.755	0.689	0.782	0.768	0.669	0.626	0.837	0.726
45	0.599	0.631	0.707	0.805	0.785	0.677	0.810	0.747	0.694	0.632	0.765	0.640
46	0.583	0.677	0.705	0.795	0.736	0.683	0.767	0.761	0.720	0.678	0.789	0.755

47	0.688	0.718	0.770	0.729	0.814	0.735	0.694	0.778	0.686	0.674	0.740	0.705
48	0.740	0.647	0.690	0.763	0.830	0.728	0.821	0.792	0.663	0.730	0.829	0.837
49	0.762	0.758	0.654	0.704	0.728		0.745	0.705	0.701	0.839	0.775	0.670
50	0.676		0.741	0.725	0.845		0.710	0.679	0.733	0.693	0.806	0.774
51			0.745	0.678	0.722		0.730	0.754	0.668		0.766	0.730
52				0.646			0.752	0.764	0.649		0.751	0.641
53				0.703			0.707	0.692			0.786	0.723
54				0.609			0.770	0.757			0.835	0.727
55				0.666			0.683	0.848			0.801	0.795
56				0.674			0.734	0.756				0.702
57				0.748			0.685	0.828				0.752
58				0.794			0.664	0.772				0.733
59				0.747			0.737	0.772				0.695
60				0.744			0.630	0.654				0.728
61				0.810				0.722				0.684
62				0.799				0.692				0.687
63				0.671				0.746				0.678
64				0.867				0.806				0.768
65				0.783				0.775				0.758
66				0.664				0.747				0.732
67				0.713				0.691				0.810
68				0.713				0.689				0.737
69				0.760				0.690				0.707
70				0.647				0.812				0.776
71				0.735				0.827				0.794
72				0.823				0.766				0.700
73				0.663				0.802				0.751
74				0.733								0.794
75				0.821								0.775
76				0.838								0.840
77				0.808								0.701
78				0.755								0.736
79				0.794								
80				0.775								
81				0.727								
82				0.746								
83				0.793								

Table S1. Results of “rmsf” analysis (scaling exponent α) of the 700 experimental cell trajectories.

Table S2

Cell number	Amoeba proteus				Metamoeba leningradensis				Amoeba borokensis			
	Sc1	Sc2	Sc3	Sc4	Sc1	Sc2	Sc3	Sc4	Sc1	Sc2	Sc3	Sc4
1	0.433	0.477	0.440	0.423	0.473	0.453	0.477	0.553	0.484	0.493	0.436	0.459
2	0.463	0.517	0.492	0.539	0.474	0.448	0.533	0.407	0.548	0.449	0.415	0.449
3	0.446	0.538	0.469	0.500	0.584	0.493	0.434	0.449	0.485	0.466	0.513	0.448
4	0.502	0.523	0.428	0.502	0.459	0.478	0.490	0.466	0.391	0.462	0.429	0.413
5	0.540	0.452	0.516	0.422	0.448	0.529	0.435	0.467	0.467	0.463	0.515	0.467
6	0.460	0.544	0.457	0.474	0.423	0.414	0.410	0.494	0.407	0.508	0.509	0.488
7	0.414	0.495	0.502	0.595	0.432	0.450	0.494	0.487	0.444	0.589	0.512	0.457
8	0.533	0.473	0.400	0.459	0.468	0.452	0.509	0.400	0.539	0.475	0.451	0.481
9	0.583	0.460	0.389	0.455	0.393	0.381	0.470	0.583	0.528	0.590	0.644	0.456
10	0.459	0.435	0.453	0.479	0.538	0.511	0.411	0.467	0.446	0.515	0.456	0.472
11	0.471	0.397	0.490	0.400	0.431	0.418	0.460	0.534	0.431	0.479	0.399	0.504
12	0.386	0.487	0.479	0.417	0.448	0.424	0.485	0.433	0.531	0.473	0.498	0.470
13	0.516	0.475	0.408	0.489	0.479	0.365	0.470	0.451	0.481	0.568	0.556	0.461
14	0.432	0.509	0.566	0.545	0.492	0.495	0.425	0.422	0.398	0.532	0.369	0.471
15	0.573	0.546	0.390	0.393	0.486	0.501	0.429	0.490	0.480	0.449	0.445	0.470
16	0.525	0.487	0.464	0.502	0.472	0.456	0.505	0.480	0.643	0.460	0.468	0.484
17	0.423	0.535	0.545	0.401	0.492	0.444	0.488	0.481	0.516	0.547	0.447	0.507
18	0.477	0.504	0.420	0.408	0.361	0.437	0.527	0.373	0.565	0.455	0.487	0.403
19	0.452	0.444	0.380	0.438	0.476	0.526	0.383	0.401	0.384	0.432	0.424	0.418
20	0.472	0.525	0.436	0.520	0.421	0.499	0.399	0.418	0.442	0.460	0.454	0.521
21	0.575	0.476	0.436	0.425	0.428	0.595	0.368	0.393	0.559	0.536	0.455	0.479
22	0.397	0.591	0.475	0.530	0.502	0.492	0.538	0.391	0.601	0.477	0.561	0.438
23	0.488	0.467	0.504	0.461	0.476	0.397	0.530	0.585	0.446	0.569	0.475	0.495
24	0.511	0.541	0.430	0.381	0.480	0.575	0.587	0.476	0.484	0.469	0.386	0.442
25	0.454	0.393	0.465	0.422	0.454	0.415	0.434	0.369	0.435	0.476	0.480	0.436
26	0.440	0.494	0.527	0.555	0.464	0.418	0.392	0.405	0.449	0.368	0.523	0.545
27	0.471	0.397	0.453	0.502	0.487	0.433	0.432	0.412	0.473	0.515	0.593	0.482
28	0.505	0.441	0.436	0.433	0.448	0.468	0.504	0.428	0.422	0.490	0.453	0.437
29	0.502	0.486	0.405	0.498	0.481	0.555	0.432	0.462	0.470	0.569	0.476	0.483
30	0.397	0.477	0.525	0.449	0.516	0.522	0.498	0.520	0.447	0.477	0.400	0.473
31	0.500	0.433	0.454	0.435	0.384	0.421	0.502	0.455	0.464	0.467	0.477	0.491
32	0.480	0.400	0.477	0.408	0.392	0.536	0.568	0.482	0.458	0.421	0.459	0.438
33	0.498	0.482	0.492	0.587	0.476	0.495	0.621	0.476	0.503	0.438	0.493	0.489
34	0.478	0.421	0.454	0.519	0.450	0.497	0.543	0.419	0.596	0.475	0.430	0.434
35	0.426	0.562	0.416	0.443	0.517	0.405	0.578	0.543	0.391	0.423	0.510	0.482
36	0.432	0.490	0.475	0.374	0.494	0.378	0.510	0.439	0.482	0.420	0.435	0.445
37	0.477	0.444	0.415	0.444	0.384	0.442	0.573	0.462	0.379	0.477	0.462	0.395
38	0.487	0.449	0.437	0.596	0.483	0.477	0.412	0.531	0.476	0.467	0.480	0.413
39	0.455	0.469	0.473	0.477	0.404	0.435	0.431	0.427	0.476	0.448	0.409	0.459
40	0.516	0.557	0.514	0.483	0.490	0.461	0.388	0.495	0.503	0.451	0.535	0.550
41	0.567	0.422	0.455	0.432	0.403	0.510	0.400	0.470	0.504	0.451	0.440	0.481
42	0.488	0.442	0.510	0.477	0.436	0.504	0.492	0.534	0.444	0.431	0.409	0.422
43	0.444	0.436	0.462	0.476	0.419	0.484	0.448	0.483	0.463	0.485	0.427	0.416
44	0.448	0.449	0.503	0.410	0.568	0.454	0.472	0.414	0.551	0.491	0.524	0.476
45	0.477	0.502	0.488	0.508	0.393	0.444	0.466	0.501	0.487	0.504	0.415	0.429
46	0.529	0.461	0.459	0.422	0.466	0.550	0.451	0.486	0.451	0.424	0.484	0.501

47	0.354	0.426	0.527	0.425	0.458	0.429	0.611	0.441	0.423	0.473	0.559	0.518
48	0.431	0.480	0.442	0.523	0.406	0.460	0.441	0.365	0.466	0.548	0.562	0.509
49	0.497	0.498	0.535	0.472	0.441		0.411	0.404	0.421	0.395	0.454	0.504
50	0.406		0.558	0.448	0.469		0.407	0.425	0.432	0.536	0.446	0.556
51			0.494	0.479	0.508		0.479	0.520	0.497		0.428	0.521
52				0.500			0.617	0.522	0.480		0.447	0.364
53				0.434			0.563	0.469			0.552	0.417
54				0.470			0.457	0.393			0.368	0.503
55				0.451			0.483	0.414			0.390	0.523
56				0.409			0.453	0.465				0.490
57				0.527			0.481	0.443				0.449
58				0.568			0.520	0.481				0.411
59				0.427			0.404	0.408				0.483
60				0.476			0.520	0.443				0.427
61				0.466				0.429				0.380
62				0.493				0.419				0.520
63				0.558				0.376				0.432
64				0.565				0.351				0.470
65				0.571				0.526				0.490
66				0.408				0.426				0.512
67				0.451				0.507				0.490
68				0.525				0.456				0.499
69				0.528				0.404				0.552
70				0.446				0.430				0.431
71				0.435				0.576				0.379
72				0.411				0.447				0.402
73				0.411				0.473				0.450
74				0.520								0.515
75				0.446								0.438
76				0.534								0.456
77				0.447								0.516
78				0.549								0.446
79				0.388								
80				0.457								
81				0.436								
82				0.535								
83				0.613								

Table S2. Results of “rmsf” analysis (scaling exponent α) of the 700 shuffled cell trajectories.

Table S3

Cell number	Amoeba proteus				Metamoeba leningradensis				Amoeba borokensis			
	Sc1	Sc2	Sc3	Sc4	Sc1	Sc2	Sc3	Sc4	Sc1	Sc2	Sc3	Sc4
1	2.083	14.583	11.458	10.417	13.542	15.625	15.625	4.167	14.583	10.417	14.583	5.208
2	13.542	15.625	6.250	5.208	12.500	7.292	7.292	10.417	12.500	8.333	8.333	5.208
3	14.583	11.458	14.583	4.167	5.208	7.292	16.667	14.583	9.375	11.458	10.417	4.167
4	8.333	7.292	8.333	3.125	7.292	12.500	5.208	9.375	6.250	3.125	13.542	2.083
5	9.375	15.625	5.208	14.583	6.250	3.125	13.542	13.542	14.583	4.167	2.083	11.458
6	10.417	15.625	11.458	5.208	11.458	4.167	8.333	10.417	4.167	14.583	4.167	3.125
7	7.292	16.667	15.625	4.167	4.167	15.625	6.250	13.542	13.542	3.125	10.417	11.458
8	12.500	15.625	9.375	13.542	13.542	6.250	6.250	6.250	10.417	4.167	7.292	3.125
9	5.208	8.333	3.125	7.292	4.167	5.208	6.250	8.333	9.375	10.417	8.333	4.167
10	5.208	16.667	8.333	14.583	8.333	7.292	12.500	8.333	15.625	13.542	6.250	6.250
11	3.125	13.542	8.333	14.583	11.458	14.583	6.250	6.250	7.292	15.625	7.292	7.292
12	13.542	13.542	10.417	12.500	5.208	7.292	14.583	6.250	14.583	6.250	15.625	3.125
13	5.208	5.208	11.458	12.500	4.167	7.292	11.458	7.292	13.542	6.250	6.250	4.167
14	5.208	12.500	8.333	14.583	14.583	7.292	3.125	6.250	15.625	10.417	4.167	4.167
15	13.542	10.417	6.250	10.417	14.583	11.458	13.542	3.125	2.083	8.333	13.542	4.167
16	2.083	10.417	12.500	7.292	8.333	10.417	2.083	5.208	12.500	3.125	6.250	9.375
17	14.583	7.292	9.375	12.500	10.417	15.625	4.167	5.208	3.125	7.292	8.333	7.292
18	3.125	7.292	13.542	3.125	12.500	5.208	3.125	10.417	2.083	4.167	12.500	11.458
19	9.375	15.625	12.500	11.458	11.458	13.542	4.167	14.583	6.250	9.375	11.458	14.583
20	13.542	14.583	9.375	3.125	5.208	13.542	5.208	4.167	8.333	12.500	7.292	12.500
21	12.500	2.083	11.458	11.458	7.292	11.458	15.625	6.250	6.250	2.083	7.292	14.583
22	15.625	11.458	7.292	10.417	10.417	13.542	3.125	14.583	6.250	12.500	6.250	3.125
23	13.542	2.083	5.208	10.417	12.500	7.292	3.125	13.542	3.125	4.167	12.500	9.375
24	15.625	2.083	4.167	8.333	10.417	15.625	13.542	6.250	4.167	5.208	5.208	5.208
25	10.417	9.375	10.417	13.542	2.083	14.583	11.458	7.292	5.208	13.542	8.333	10.417
26	6.250	5.208	10.417	16.667	8.333	9.375	5.208	6.250	14.583	14.583	5.208	8.333
27	14.583	14.583	9.375	4.167	13.542	8.333	13.542	14.583	9.375	2.083	15.625	15.625
28	14.583	1.042	13.542	11.458	13.542	9.375	8.333	13.542	2.083	11.458	10.417	9.375
29	14.583	11.458	14.583	12.500	15.625	10.417	5.208	6.250	7.292	7.292	6.250	8.333
30	10.417	12.500	5.208	13.542	16.667	14.583	6.250	10.417	8.333	8.333	7.292	5.208
31	7.292	13.542	7.292	13.542	9.375	10.417	12.500	12.500	4.167	12.500	8.333	7.292
32	3.125	15.625	12.500	13.542	12.500	8.333	14.583	12.500	15.625	4.167	4.167	3.125
33	14.583	11.458	12.500	10.417	15.625	7.292	9.375	10.417	2.083	5.208	10.417	11.458
34	13.542	15.625	6.250	12.500	15.625	7.292	15.625	12.500	6.250	14.583	8.333	9.375
35	9.375	6.250	9.375	10.417	8.333	2.083	4.167	5.208	11.458	16.667	6.250	13.542
36	8.333	13.542	8.333	9.375	15.625	11.458	12.500	3.125	14.583	11.458	12.500	12.500
37	9.375	8.333	11.458	13.542	9.375	15.625	4.167	6.250	5.208	9.375	7.292	5.208
38	9.375	15.625	10.417	10.417	8.333	11.458	4.167	5.208	14.583	14.583	13.542	10.417
39	9.375	13.542	11.458	10.417	9.375	14.583	12.500	10.417	6.250	13.542	11.458	10.417
40	7.292	11.458	7.292	10.417	10.417	6.250	13.542	10.417	3.125	3.125	7.292	5.208
41	15.625	15.625	9.375	9.375	6.250	13.542	7.292	6.250	11.458	8.333	4.167	10.417
42	6.250	2.083	8.333	4.167	3.125	13.542	11.458	4.167	7.292	8.333	6.250	8.333
43	15.625	4.167	11.458	2.083	5.208	6.250	9.375	5.208	6.250	7.292	12.500	3.125
44	15.625	5.208	7.292	6.250	8.333	6.250	13.542	3.125	9.375	7.292	15.625	13.542
45	10.417	6.250	15.625	6.250	9.375	4.167	14.583	12.500	5.208	7.292	7.292	8.333
46	2.083	15.625	11.458	9.375	10.417	11.458	10.417	14.583	2.083	3.125	10.417	4.167

47	7.292	13.542	5.208	10.417	7.292	12.500	7.292	14.583	15.625	4.167	5.208	10.417
48	10.417	15.625	12.500	12.500	14.583	9.375	15.625	13.542	13.542	7.292	11.458	3.125
49	14.583	15.625	14.583	7.292	6.250		15.625	15.625	14.583	12.500	15.625	3.125
50	10.417		12.500	10.417	15.625		8.333	13.542	11.458	6.250	8.333	3.125
51			4.167	3.125	15.625		6.250	15.625	7.292		11.458	12.500
52				9.375			14.583	11.458	10.417		12.500	6.250
53				12.500			10.417	13.542			3.125	8.333
54				15.625			7.292	6.250			14.583	8.333
55				11.458			6.250	15.625			11.458	9.375
56				5.208			8.333	15.625				3.125
57				11.458			7.292	14.583				12.500
58				13.542			7.292	15.625				2.083
59				10.417			3.125	15.625				7.292
60				10.417			15.625	10.417				5.208
61				10.417				14.583				7.292
62				11.458				11.458				7.292
63				3.125				13.542				3.125
64				14.583				12.500				3.125
65				11.458				7.292				9.375
66				11.458				6.250				10.417
67				4.167				6.250				11.458
68				14.583				12.500				7.292
69				9.375				6.250				12.500
70				15.625				13.542				11.458
71				8.333				5.208				5.208
72				11.458				4.167				14.583
73				13.542				5.208				4.167
74				15.625								7.292
75				14.583								4.167
76				15.625								14.583
77				15.625								4.167
78				8.333								12.500
79				6.250								
80				4.167								
81				13.542								
82				11.458								
83				14.583								

Table S3. Long-range correlation duration values for the 700 experimental cell trajectories.

Table S4

Cell number	Amoeba proteus				Metamoeba leningradensis				Amoeba borokensis			
	Sc1	Sc2	Sc3	Sc4	Sc1	Sc2	Sc3	Sc4	Sc1	Sc2	Sc3	Sc4
1	1.655	1.962	1.693	1.850	1.926	2.007	1.895	1.833	1.833	1.671	1.790	1.792
2	1.871	1.903	1.809	1.666	1.893	1.963	1.789	1.943	1.778	1.839	1.738	1.905
3	1.761	1.956	1.928	1.692	1.993	1.691	1.332	1.940	1.836	1.666	1.854	1.774
4	1.894	1.964	1.967	1.815	1.610	1.424	1.525	1.991	1.804	1.786	1.844	1.564
5	1.817	1.912	1.974	1.986	1.867	1.919	1.851	1.954	1.877	1.589	1.623	1.843
6	1.901	1.948	1.930	1.870	1.886	1.509	1.866	1.930	1.852	1.790	1.655	1.732
7	1.949	1.939	1.613	1.709	1.705	1.923	1.823	1.967	1.882	1.743	1.862	1.597
8	1.907	1.940	1.976	1.942	1.986	1.957	1.992	1.981	1.880	1.805	1.904	1.802
9	1.766	1.800	1.869	1.756	1.943	1.901	2.006	1.967	1.841	1.683	1.770	1.775
10	1.494	1.964	1.915	1.945	1.753	1.918	1.932	1.830	1.833	1.926	1.711	1.870
11	1.782	1.874	1.900	1.938	1.865	1.735	1.809	1.954	1.838	1.920	1.871	1.760
12	1.864	1.670	1.949	1.865	1.816	1.918	1.910	1.900	1.594	1.769	1.726	1.886
13	1.759	1.919	1.897	1.982	1.839	1.976	1.827	1.920	1.846	1.929	1.811	1.828
14	1.709	1.961	1.840	1.964	1.796	1.877	1.863	1.879	1.668	1.946	1.869	1.695
15	1.920	1.825	1.966	1.970	1.954	1.687	1.915	1.929	1.671	1.705	1.871	1.847
16	1.741	1.921	1.881	1.820	1.797	1.989	1.945	1.959	1.942	1.883	1.844	1.449
17	1.833	2.003	1.796	1.977	1.786	1.664	1.694	1.943	1.737	1.853	1.680	1.818
18	1.651	1.860	1.874	1.881	1.707	1.918	1.623	1.811	1.729	1.821	1.803	1.901
19	1.664	1.957	1.926	1.862	1.883	1.969	1.692	1.964	1.919	1.772	1.874	1.869
20	1.909	1.986	1.896	1.846	1.905	1.929	1.623	1.953	1.930	1.915	1.834	1.901
21	1.774	1.925	1.856	1.917	1.126	1.982	1.693	1.950	1.684	1.939	1.691	1.881
22	1.869	1.922	1.844	1.802	1.896	1.786	1.743	1.892	1.629	1.945	1.924	1.774
23	1.953	1.918	1.880	1.902	1.699	1.904	1.571	1.896	1.913	1.967	1.864	1.730
24	1.944	1.973	1.849	1.822	1.736	1.980	1.847	1.923	1.435	1.941	1.927	1.795
25	1.866	1.878	1.903	1.918	1.963	1.915	1.965	1.971	1.682	1.935	1.772	1.810
26	1.608	1.938	1.782	1.852	1.820	1.772	1.876	1.931	1.919	1.736	1.932	1.786

27	1.966	1.811	1.891	1.884	1.713	1.896	1.784	1.934	1.725	1.936	1.873	1.835
28	1.899	1.965	1.965	1.760	1.838	1.974	1.898	1.859	1.489	1.634	1.397	1.528
29	1.909	1.960	1.970	1.958	1.309	1.955	1.606	1.997	1.625	1.896	1.894	1.823
30	1.973	1.929	1.691	1.951	1.580	1.990	1.704	1.858	1.836	1.856	1.580	1.908
31	1.968	1.426	1.706	1.813	1.785	1.827	1.933	1.799	1.928	1.812	1.795	1.954
32	1.959	1.693	1.983	1.845	1.841	1.914	1.490	1.981	1.894	1.857	1.976	1.911
33	1.858	2.000	2.010	1.368	1.360	1.947	1.451	1.948	1.945	1.908	1.947	1.705
34	1.887	1.973	1.938	1.875	1.648	1.858	1.425	1.971	1.826	1.837	1.981	1.936
35	1.897	1.778	1.911	1.946	1.424	1.901	1.345	1.915	1.893	1.944	1.607	1.957
36	1.946	1.821	1.987	1.841	1.950	1.845	1.780	1.961	1.928	1.924	1.902	1.863
37	1.906	1.829	1.978	1.899	1.642	1.870	1.194	1.944	1.892	1.870	1.641	1.696
38	1.990	1.954	1.913	1.893	1.707	1.892	1.767	1.987	2.006	1.918	1.967	1.608
39	1.807	1.961	1.943	1.939	1.627	1.922	1.804	1.979	1.947	1.891	1.848	1.720
40	1.948	1.969	1.826	1.902	1.474	1.918	1.764	1.903	1.805	1.834	1.739	1.773
41	1.894	1.925	1.936	1.831	1.560	1.986	1.799	1.926	1.834	1.915	1.928	1.712
42	1.925	1.877	1.916	1.756	1.618	1.895	1.785	1.989	1.904	1.928	1.781	1.783
43	1.944	1.992	1.923	1.794	1.770	1.858	1.711	1.962	1.708	1.974	1.763	1.629
44	1.516	1.899	1.785	1.889	1.853	1.820	1.856	1.905	1.810	1.882	1.933	1.803
45	1.979	1.968	1.245	1.821	1.926	1.885	1.921	1.827	1.706	1.957	1.977	1.577
46	1.806	1.949	1.795	1.979	1.882	1.718	1.841	1.944	1.092	1.923	1.993	1.630
47	1.887	1.971	1.784	1.965	1.695	1.792	1.787	1.940	1.943	1.946	1.999	1.820
48	1.891	1.983	1.890	1.962	1.821	1.962	1.956	1.923	1.823	1.912	1.964	1.688
49	1.950	1.973	1.833	1.617	1.951		1.872	1.810	1.895	1.901	1.960	1.825
50	1.876		1.860	1.939	1.936		1.869	1.571	1.951	1.910	1.823	1.838
51			1.792	1.829	1.810		1.642	1.880	1.931		1.942	1.845
52				1.752			1.868	1.907	1.797		1.837	1.979
53				1.857			1.963	1.856			1.763	1.965
54				1.566			1.936	1.916			1.970	1.863

55				1.748			1.775	1.959			1.861	1.950
56				1.687			1.765	1.898				1.868
57				1.935			1.694	1.888				1.909
58				1.890			1.645	1.957				1.842
59				1.934			1.364	1.896				2.002
60				1.600			1.431	1.729				1.846
61				1.864				1.904				1.551
62				1.884				1.874				1.903
63				1.935				1.725				1.693
64				1.923				1.938				1.612
65				1.940				1.978				1.770
66				1.950				1.840				1.717
67				1.675				1.441				1.862
68				1.865				1.749				1.939
69				1.916				1.976				1.995
70				1.723				1.891				2.022
71				1.907				1.837				1.890
72				1.972				1.878				1.785
73				1.778				1.709				1.936
74				1.929								1.925
75				1.811								1.870
76				1.916								1.907
77				1.968								1.787
78				1.980								1.902
79				1.958								
80				1.849								
81				1.860								
82				1.870								

83					1.949							
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Table S4. Results of MSD analysis of the 700 experimental cell trajectories.

Table S5

Cell number	Amoeba proteus				Metamoeba leningradensis				Amoeba borokensis			
	Sc1	Sc2	Sc3	Sc4	Sc1	Sc2	Sc3	Sc4	Sc1	Sc2	Sc3	Sc4
1	-	0.001	0.001	0.001	-	0.001	0.001	0.002	0.001	0.001	0.003	0.004
2	0.005	0.000	0.001	0.002	0.002	0.001	0.002	0.001	0.003	0.002	0.003	0.000
3	-	0.004	0.001	0.002	0.002	0.001	0.001	0.003	0.002	0.001	0.001	0.002
4	0.002	-	0.001	0.000	0.001	0.003	0.000	0.002	0.000	0.005	0.003	0.000
5	0.000	0.001	0.001	0.001	0.001	0.001	0.001	0.000	0.000	0.001	0.005	0.000
6	-	0.002	0.002	0.002	0.003	0.000	0.000	0.002	0.000	0.001	0.001	0.001
7	-	0.002	0.003	0.001	0.001	0.000	0.002	0.001	0.001	0.003	0.000	0.001
8	-	0.001	0.001	0.001	0.000	-	0.002	0.001	0.001	0.000	0.001	0.003
9	-	0.001	0.002	0.001	0.001	0.001	0.002	0.001	-	0.002	0.000	0.001
10	0.002	0.002	0.000	0.002	0.000	0.002	0.001	0.000	0.003	0.002	0.001	0.001
11	0.001	-	0.001	0.002	0.001	0.001	-	0.002	0.000	0.001	0.000	0.001
12	-	0.001	0.003	0.001	0.001	0.000	0.002	0.001	0.001	0.001	0.001	0.001
13	-	0.003	0.001	0.001	0.000	0.002	0.000	0.002	0.003	0.001	0.001	0.002
14	0.003	-	0.004	0.001	0.001	0.003	0.002	0.001	0.002	0.003	0.000	0.000
15	0.002	0.003	0.002	0.001	0.001	0.003	-	0.000	0.001	0.000	0.002	0.001
16	-	0.001	0.002	0.000	0.002	-	0.002	0.002	0.003	0.003	0.000	0.000
17	-	0.001	0.001	0.000	0.000	-	-	0.000	0.002	0.002	0.000	0.001
18	-	0.001	0.000	0.001	0.002	0.002	0.000	0.000	-	0.002	0.000	0.001
19	0.002	0.000	0.000	0.002	0.003	0.004	-	0.002	0.001	0.001	0.001	0.000
20	-	0.004	0.001	0.001	0.003	0.003	0.002	0.002	0.001	0.003	0.001	0.003
21	0.002	-	0.004	0.001	0.002	0.004	0.001	0.002	0.001	0.002	0.001	0.003
22	0.002	0.000	0.001	0.002	0.002	0.000	0.002	0.002	0.002	0.002	0.001	0.001
23	0.000	-	0.001	0.000	-	0.003	0.000	0.001	0.001	0.003	0.001	0.002

24	- 0.001	0.000	0.001	- 0.002	0.000	0.001	- 0.001	0.005	0.000	0.001	0.002	- 0.003
25	0.001	0.001	- 0.001	0.001	0.000	0.002	0.000	0.002	- 0.001	0.003	0.002	0.000
26	0.005	0.001	0.003	0.000	0.001	0.001	- 0.001	0.001	0.001	0.002	0.000	0.002
27	0.003	0.001	- 0.001	0.000	0.006	0.001	0.003	0.001	0.002	0.001	0.001	0.000
28	0.001	0.000	- 0.004	0.002	0.001	0.001	0.001	0.000	0.002	0.000	0.001	0.003
29	- 0.002	0.001	0.001	- 0.001	0.001	0.000	- 0.001	0.002	0.003	0.004	0.000	0.001
30	0.000	- 0.001	0.002	0.001	0.002	0.000	- 0.002	0.003	0.000	0.003	0.002	0.003
31	0.000	0.001	0.000	0.001	0.002	0.003	0.001	0.002	0.000	0.001	0.003	0.000
32	- 0.001	0.000	0.001	- 0.001	0.001	0.002	0.004	0.002	0.003	0.001	0.000	0.004
33	0.001	- 0.001	0.001	- 0.003	0.002	0.002	0.000	0.002	0.000	0.002	0.001	0.002
34	- 0.002	0.000	- 0.003	0.001	0.001	- 0.002	0.000	0.000	- 0.001	0.001	0.001	- 0.001
35	0.006	0.000	0.002	0.001	0.006	0.001	0.003	0.002	- 0.001	0.000	0.005	0.002
36	0.000	0.002	0.002	0.001	0.000	0.007	0.000	0.000	0.002	0.002	0.002	0.000
37	- 0.001	0.002	0.001	0.001	0.001	0.001	- 0.001	0.001	0.001	0.001	0.001	0.000
38	0.000	- 0.004	0.003	0.000	0.000	0.001	0.000	0.001	0.000	0.001	0.000	0.003
39	- 0.001	0.002	0.004	0.000	- 0.001	0.000	0.002	0.001	0.001	0.000	0.001	0.000
40	- 0.002	0.001	0.000	- 0.001	0.001	- 0.001	0.003	0.002	0.003	0.003	0.000	- 0.001
41	0.004	0.001	0.002	0.001	0.001	0.001	0.001	0.002	0.001	0.002	0.003	0.002
42	0.001	- 0.001	0.001	0.001	0.006	0.001	0.000	0.000	- 0.001	0.000	0.002	0.001
43	0.001	0.000	0.001	0.003	0.001	0.001	- 0.007	0.001	- 0.001	0.000	0.001	- 0.001
44	0.002	0.001	0.002	0.002	0.001	0.001	0.001	0.000	0.003	0.000	0.000	0.001
45	- 0.004	0.002	- 0.001	0.001	0.000	- 0.001	0.000	- 0.002	0.002	0.002	0.001	- 0.001
46	0.001	- 0.001	0.000	- 0.002	0.002	0.001	- 0.002	0.000	0.001	0.000	0.001	- 0.001
47	0.000	0.001	- 0.002	0.002	0.003	0.002	0.001	0.002	0.002	0.001	0.002	0.000
48	- 0.001	0.001	0.000	0.000	0.002	0.000	0.000	0.003	- 0.002	0.001	0.001	- 0.002

49	0.001	0.001	0.001	-	0.001	0.000		-	0.002	0.001	-	0.001	0.000	-	0.004	-	0.002	
50	0.001		-	0.001	0.000	0.002		0.001	0.002	0.001	0.000	0.000	-	0.003	-	0.002		
51			-	0.000	0.001	0.001		0.003	0.000	0.001		-	-	0.001	0.003	-	-	
52				0.000				0.004	0.003	0.002		-		0.002	0.002	-	-	
53					0.001			0.002	0.003			-	0.001	0.002	-	-	-	-
54					0.000			-	-	0.001	0.001			0.001	0.001			
55				-	0.001			0.001	0.001	-			0.001	0.000				
56				-	0.002			0.001	0.002					0.001	0.001			
57					0.001			-	0.002	0.001				0.002				
58					0.001			-	0.002	0.003				0.001				
59					0.001			-	0.002	0.001				0.000				
60					0.002			-	0.002	0.001				0.000				
61					0.000					0.002	-				0.003			
62					0.000					-	0.001				-	0.001		
63					0.001					0.001					0.002			
64					0.000					0.000					-	0.003		
65				-	0.001					0.000					0.001			
66				-	0.002					-	0.003				-	0.003		
67				-	0.001					-	0.001				0.001			
68					0.002					-	0.001				0.002			
69					0.000					0.000					-	0.001		
70				-	0.002					-	0.001				0.000			
71				-	0.002					0.002					0.001			
72					0.000					0.000					-	0.003		
73				-	0.002					-	0.001				0.001			
74					0.001										-	0.001		

75				- 0.003								0.003
76				0.000								0.003
77				0.000								0.003
78				0.000								- 0.004
79				0.000								
80				- 0.001								
81				- 0.002								
82				0.001								
83				0.000								

Table S5. Results of MSD analysis of the 700 shuffled cell trajectories.

Table S6

Cell number	Amoeba proteus				Metamoeba leningradensis				Amoeba borokensis			
	Sc1	Sc2	Sc3	Sc4	Sc1	Sc2	Sc3	Sc4	Sc1	Sc2	Sc3	Sc4
1	0.012	0.002	0.006	0.003	0.003	0.001	0.002	0.003	0.003	0.006	0.005	0.004
2	0.002	0.002	0.004	0.010	0.003	0.002	0.002	0.002	0.004	0.004	0.004	0.002
3	0.006	0.001	0.003	0.006	0.001	0.004	0.022	0.003	0.002	0.005	0.004	0.003
4	0.002	0.001	0.002	0.004	0.012	0.009	0.012	0.001	0.007	0.004	0.003	0.008
5	0.005	0.002	0.002	0.002	0.003	0.003	0.002	0.001	0.002	0.007	0.011	0.003
6	0.002	0.002	0.003	0.002	0.002	0.008	0.006	0.002	0.007	0.008	0.009	0.004
7	0.006	0.003	0.010	0.004	0.004	0.003	0.003	0.001	0.003	0.003	0.003	0.007
8	0.002	0.003	0.002	0.002	0.001	0.003	0.002	0.001	0.004	0.002	0.002	0.004
9	0.005	0.004	0.004	0.004	0.003	0.003	0.001	0.001	0.005	0.004	0.003	0.003
10	0.012	0.003	0.003	0.002	0.005	0.003	0.003	0.005	0.004	0.002	0.003	0.008
11	0.009	0.003	0.003	0.003	0.003	0.004	0.005	0.002	0.004	0.002	0.003	0.004
12	0.002	0.007	0.002	0.003	0.005	0.002	0.002	0.002	0.008	0.003	0.008	0.003
13	0.003	0.003	0.002	0.001	0.003	0.002	0.005	0.002	0.003	0.002	0.007	0.008
14	0.005	0.002	0.002	0.002	0.003	0.003	0.002	0.004	0.004	0.002	0.004	0.007
15	0.003	0.005	0.002	0.002	0.002	0.004	0.001	0.003	0.005	0.005	0.003	0.003
16	0.007	0.002	0.002	0.003	0.004	0.002	0.002	0.004	0.002	0.003	0.003	0.014
17	0.004	0.002	0.002	0.002	0.003	0.005	0.005	0.001	0.011	0.002	0.005	0.004
18	0.006	0.004	0.003	0.003	0.007	0.003	0.004	0.003	0.009	0.003	0.003	0.002
19	0.007	0.003	0.004	0.005	0.002	0.001	0.005	0.002	0.002	0.003	0.002	0.003
20	0.004	0.002	0.003	0.003	0.004	0.001	0.009	0.001	0.003	0.004	0.003	0.003
21	0.005	0.002	0.003	0.002	0.024	0.002	0.009	0.001	0.007	0.003	0.006	0.002
22	0.005	0.002	0.003	0.004	0.003	0.003	0.007	0.003	0.007	0.003	0.002	0.004
23	0.002	0.002	0.002	0.002	0.003	0.003	0.007	0.002	0.003	0.002	0.004	0.004
24	0.003	0.002	0.003	0.004	0.008	0.001	0.002	0.001	0.013	0.003	0.002	0.003
25	0.002	0.002	0.002	0.002	0.003	0.003	0.001	0.002	0.004	0.003	0.007	0.003
26	0.007	0.002	0.006	0.003	0.004	0.005	0.003	0.001	0.003	0.005	0.001	0.003
27	0.002	0.002	0.005	0.004	0.004	0.003	0.004	0.001	0.008	0.003	0.003	0.004
28	0.006	0.002	0.002	0.004	0.003	0.002	0.003	0.004	0.008	0.010	0.013	0.006

29	0.004	0.002	0.002	0.002	0.014	0.002	0.004	0.001	0.006	0.003	0.003	0.003
30	0.003	0.002	0.009	0.001	0.009	0.002	0.004	0.004	0.003	0.003	0.014	0.003
31	0.003	0.012	0.006	0.003	0.008	0.004	0.001	0.004	0.002	0.004	0.003	0.001
32	0.003	0.005	0.002	0.002	0.004	0.003	0.017	0.001	0.003	0.003	0.001	0.002
33	0.005	0.002	0.001	0.007	0.017	0.002	0.014	0.002	0.002	0.003	0.002	0.007
34	0.006	0.002	0.002	0.002	0.012	0.003	0.009	0.002	0.007	0.004	0.001	0.001
35	0.003	0.004	0.003	0.002	0.009	0.002	0.010	0.002	0.002	0.003	0.009	0.001
36	0.002	0.006	0.002	0.003	0.003	0.005	0.003	0.001	0.002	0.003	0.006	0.004
37	0.002	0.005	0.003	0.003	0.006	0.002	0.019	0.001	0.002	0.004	0.004	0.006
38	0.002	0.002	0.002	0.003	0.005	0.002	0.003	0.002	0.002	0.003	0.002	0.006
39	0.005	0.002	0.002	0.003	0.011	0.002	0.003	0.002	0.002	0.003	0.006	0.005
40	0.003	0.002	0.003	0.002	0.013	0.002	0.003	0.003	0.004	0.003	0.007	0.003
41	0.004	0.001	0.002	0.003	0.010	0.002	0.007	0.003	0.003	0.002	0.002	0.005
42	0.002	0.002	0.002	0.005	0.006	0.003	0.008	0.002	0.002	0.003	0.007	0.004
43	0.002	0.001	0.002	0.003	0.004	0.003	0.005	0.002	0.005	0.002	0.003	0.005
44	0.015	0.002	0.003	0.002	0.006	0.003	0.003	0.002	0.010	0.003	0.002	0.003
45	0.002	0.001	0.015	0.003	0.002	0.003	0.002	0.004	0.008	0.002	0.001	0.014
46	0.004	0.001	0.004	0.001	0.005	0.004	0.002	0.002	0.018	0.002	0.001	0.005
47	0.003	0.001	0.007	0.002	0.006	0.004	0.002	0.002	0.003	0.002	0.001	0.003
48	0.006	0.001	0.003	0.002	0.004	0.001	0.002	0.002	0.004	0.002	0.001	0.008
49	0.003	0.001	0.004	0.005	0.001		0.003	0.004	0.003	0.003	0.002	0.003
50	0.005		0.003	0.002	0.002		0.003	0.006	0.002	0.002	0.002	0.002
51			0.003	0.004	0.003		0.005	0.003	0.002		0.001	0.003
52				0.004			0.002	0.002	0.003		0.006	0.001
53				0.003			0.002	0.002			0.005	0.001
54				0.014			0.002	0.002			0.002	0.003
55				0.004			0.005	0.001			0.004	0.002
56				0.006			0.004	0.002				0.003
57				0.003			0.006	0.002				0.002
58				0.004			0.006	0.001				0.005
59				0.003			0.013	0.003				0.001
60				0.006			0.011	0.004				0.003
61				0.008				0.003				0.009
62				0.002				0.004				0.002
63				0.002				0.004				0.004
64				0.002				0.003				0.009
65				0.001				0.002				0.007
66				0.002				0.003				0.006
67				0.004				0.007				0.005
68				0.002				0.004				0.001
69				0.002				0.002				0.000
70				0.005				0.003				0.001
71				0.003				0.002				0.001
72				0.003				0.002				0.005
73				0.004				0.003				0.001
74				0.002								0.001
75				0.004								0.002
76				0.001								0.001
77				0.002								0.002
78				0.001								0.001

79				0.002								
80				0.002								
81				0.003								
82				0.003								
83				0.002								

Table S6. Results of Approximate Entropy estimation for the 700 experimental cell trajectories.

Table S7

Cell number	Amoeba proteus				Metamoeba leningradensis				Amoeba borokensis			
	Sc1	Sc2	Sc3	Sc4	Sc1	Sc2	Sc3	Sc4	Sc1	Sc2	Sc3	Sc4
1	2.071	1.964	1.853	1.857	1.968	2.067	1.613	1.927	1.688	1.896	1.933	2.045
2	1.849	1.901	2.004	1.952	1.762	1.892	1.963	2.062	1.922	1.879	1.840	2.049
3	1.740	2.097	1.683	1.826	2.090	1.974	2.006	1.901	1.890	1.669	1.605	1.970
4	2.009	2.077	1.947	1.897	1.830	1.894	2.000	1.886	1.988	2.033	1.871	1.995
5	1.892	2.031	2.079	2.015	1.914	2.079	1.770	1.656	1.747	1.960	2.057	1.761
6	1.979	1.943	1.953	1.928	1.574	1.992	1.982	1.864	2.041	1.788	1.836	1.957
7	2.053	1.986	2.076	2.015	1.960	2.004	1.998	2.016	1.979	1.807	1.681	1.813
8	1.700	2.036	2.028	1.650	2.007	2.023	1.991	2.070	1.908	1.996	1.806	2.076
9	1.938	1.856	1.993	1.763	1.960	2.074	1.916	2.003	1.941	1.911	1.752	1.993
10	2.003	2.033	2.032	1.991	1.996	2.021	1.823	1.989	1.608	1.917	1.789	2.055
11	2.006	2.034	2.115	1.762	1.832	1.931	1.977	1.962	1.891	1.984	1.826	1.859
12	1.931	1.732	2.027	1.814	1.958	1.999	2.003	1.878	1.734	2.008	1.660	2.088
13	1.936	1.969	1.955	2.046	1.996	2.069	1.984	1.904	2.010	2.102	1.997	2.003
14	2.028	2.074	2.008	1.973	1.636	2.022	2.024	1.606	2.026	2.099	1.881	1.976
15	2.093	1.815	2.026	1.976	2.085	1.595	2.008	2.079	1.895	1.807	2.090	1.830
16	2.013	2.036	2.050	1.833	1.699	2.049	2.065	2.019	1.891	1.950	1.945	1.928
17	2.083	2.046	1.992	1.849	2.023	1.978	1.918	2.059	2.031	1.983	1.867	2.013
18	1.865	1.922	1.957	1.984	1.894	2.080	1.981	1.972	2.032	2.037	1.888	2.079
19	1.809	2.043	1.960	1.745	1.893	2.081	1.953	2.096	2.039	1.960	1.944	1.942
20	1.866	2.036	1.973	1.963	1.981	2.108	2.039	2.108	2.086	1.912	2.005	2.018
21	1.885	2.104	1.851	2.037	1.766	2.041	1.969	2.082	1.859	2.093	1.804	1.925
22	1.857	1.745	1.945	1.734	1.809	1.865	2.049	2.081	1.912	2.097	2.032	1.920
23	1.912	2.097	1.960	2.075	1.252	2.106	1.821	1.971	2.051	2.042	1.956	1.968
24	1.901	2.098	1.862	1.862	1.954	2.051	1.919	2.065	1.879	2.014	2.076	2.066
25	1.923	2.053	1.895	1.826	2.055	2.062	1.982	1.953	1.897	2.101	1.999	1.870
26	1.969	2.115	1.747	2.053	1.976	1.852	1.999	2.072	1.952	1.818	2.106	1.929
27	2.081	1.834	1.991	1.989	1.428	2.088	1.856	2.052	1.869	2.070	1.821	1.613
28	2.052	2.066	2.007	1.862	1.357	2.039	1.877	1.985	1.974	1.898	1.972	2.053
29	1.918	1.888	2.022	2.001	1.728	1.969	1.924	2.081	1.797	2.085	1.956	1.979
30	2.001	2.006	1.899	1.855	1.605	2.014	1.759	1.960	1.858	2.066	1.896	2.037
31	2.051	1.589	1.803	1.860	1.881	2.018	1.970	1.830	1.965	1.973	1.995	2.048
32	2.074	1.922	1.899	1.941	1.885	2.030	1.999	2.055	1.653	2.062	2.055	2.054
33	2.037	2.036	1.990	1.671	1.835	2.007	2.015	1.963	2.077	2.100	1.800	1.952
34	1.947	2.066	2.088	1.904	1.988	2.098	1.942	1.901	2.006	1.860	2.021	2.041
35	2.039	1.961	1.890	1.879	1.579	2.072	1.890	1.957	1.903	2.091	1.738	1.997
36	2.085	1.956	2.013	1.898	1.749	1.897	1.819	2.094	1.916	2.054	2.081	1.980
37	2.080	2.034	1.872	1.896	1.754	1.697	2.016	2.075	1.964	2.088	1.773	1.707
38	2.013	2.054	1.894	2.080	1.844	1.990	1.782	2.003	2.016	1.951	1.913	1.786
39	1.827	2.028	1.969	1.677	1.919	2.078	1.811	2.036	1.981	2.079	1.920	1.901
40	2.029	2.087	1.910	1.826	1.945	1.975	1.543	1.999	1.834	2.057	2.011	1.968
41	1.843	2.077	1.986	1.780	1.890	1.980	1.912	1.830	1.977	2.077	1.879	1.778
42	1.927	2.092	1.822	1.934	1.610	2.018	1.998	2.026	2.126	1.921	2.059	1.896
43	2.053	2.064	2.043	2.049	1.714	1.963	1.835	2.008	2.059	2.068	1.749	1.928
44	1.952	2.057	1.862	1.810	1.935	2.012	1.651	2.001	2.082	2.072	1.711	2.032
45	2.075	2.085	1.967	1.899	1.884	2.017	1.763	1.699	1.953	2.089	2.075	1.841
46	1.931	2.075	1.999	1.967	1.921	1.916	1.824	1.915	1.995	2.065	1.968	2.011

47	2.102	2.053	1.809	1.909	1.743	1.927	1.994	1.942	1.884	2.026	2.061	2.093
48	2.007	2.019	1.791	1.857	1.514	2.050	1.904	1.916	1.986	1.848	1.968	1.999
49	2.011	2.050	2.060	1.971	2.055		1.820	1.934	1.967	1.880	1.950	1.986
50	1.831		1.870	1.824	1.628		1.896	1.762	1.893	2.070	1.672	1.919
51			1.833	1.974	1.913		1.799	1.984	2.044		2.045	1.821
52				1.873			1.934	1.996	2.033		1.809	2.045
53				1.689			2.059	2.004			1.906	2.015
54				1.940			1.805	2.059			1.671	1.929
55				1.772			1.905	1.925			1.963	1.831
56				1.963			1.796	2.009				1.985
57				2.060			1.780	1.811				1.943
58				1.799			1.630	2.038				2.016
59				1.992			2.046	1.950				2.040
60				2.003			1.943	1.979				1.922
61				1.921				1.904				1.860
62				1.993				1.958				2.058
63				2.092				1.832				1.973
64				1.606				1.922				1.894
65				1.924				1.956				2.013
66				2.003				1.982				1.925
67				1.849				2.057				1.941
68				1.816				1.937				1.987
69				1.974				1.998				2.084
70				1.908				1.840				2.009
71				1.993				1.979				2.082
72				1.731				2.083				1.898
73				1.694				1.962				2.088
74				1.973								2.007
75				1.552								1.972
76				1.668								1.854
77				1.980								1.928
78				2.037								1.994
79				1.993								
80				1.769								
81				1.863								
82				1.822								
83				1.804								

Table S7. Results of Approximate Entropy estimation for the 700 shuffled cell trajectories.

Table S8

Cell number	Amoeba proteus				Metamoeba leningradensis				Amoeba borokensis			
	Sc1	Sc2	Sc3	Sc4	Sc1	Sc2	Sc3	Sc4	Sc1	Sc2	Sc3	Sc4
1	1.644	1.794	1.715	1.746	1.704	1.844	1.830	1.783	1.827	1.539	1.803	1.736
2	1.805	1.670	1.645	1.688	1.562	1.840	1.755	1.766	1.568	1.674	1.748	1.771
3	1.778	1.781	1.868	1.468	1.814	1.671	1.350	1.873	1.818	1.714	1.869	1.784
4	1.800	1.809	1.856	1.809	1.518	1.521	1.518	1.862	1.618	1.741	1.799	1.639
5	1.736	1.716	1.818	1.802	1.828	1.820	1.799	1.881	1.832	1.535	1.551	1.800
6	1.686	1.857	1.826	1.790	1.793	1.582	1.860	1.824	1.799	1.707	1.770	1.723
7	1.895	1.820	1.536	1.626	1.589	1.762	1.787	1.845	1.745	1.747	1.873	1.314
8	1.826	1.764	1.828	1.693	1.846	1.832	1.853	1.821	1.827	1.703	1.875	1.776
9	1.750	1.745	1.668	1.770	1.839	1.796	1.870	1.840	1.772	1.502	1.764	1.776
10	1.426	1.818	1.784	1.781	1.753	1.823	1.900	1.787	1.821	1.747	1.677	1.736
11	1.739	1.816	1.780	1.757	1.836	1.680	1.856	1.813	1.781	1.767	1.842	1.703
12	1.820	1.723	1.848	1.735	1.786	1.807	1.761	1.831	1.738	1.770	1.802	1.748
13	1.633	1.842	1.801	1.830	1.793	1.832	1.816	1.839	1.794	1.798	1.644	1.672
14	1.497	1.799	1.745	1.755	1.799	1.775	1.692	1.870	1.707	1.793	1.846	1.824
15	1.772	1.689	1.836	1.875	1.827	1.370	1.737	1.780	1.633	1.782	1.785	1.822
16	1.805	1.791	1.732	1.851	1.776	1.858	1.816	1.881	1.849	1.822	1.775	1.324
17	1.740	1.854	1.662	1.805	1.767	1.688	1.691	1.814	1.659	1.771	1.761	1.789
18	1.650	1.780	1.654	1.820	1.690	1.816	1.665	1.717	1.701	1.657	1.743	1.801
19	1.445	1.814	1.872	1.857	1.607	1.828	1.627	1.794	1.824	1.716	1.836	1.672
20	1.851	1.827	1.823	1.605	1.842	1.809	1.656	1.792	1.762	1.867	1.818	1.815
21	1.792	1.789	1.597	1.825	1.276	1.866	1.728	1.795	1.515	1.802	1.574	1.824
22	1.815	1.850	1.753	1.763	1.807	1.647	1.575	1.792	1.747	1.782	1.800	1.729
23	1.873	1.782	1.825	1.746	1.358	1.767	1.744	1.800	1.812	1.831	1.838	1.702
24	1.866	1.811	1.852	1.826	1.714	1.777	1.834	1.817	1.556	1.791	1.778	1.775
25	1.715	1.724	1.862	1.722	1.823	1.826	1.876	1.834	1.728	1.786	1.722	1.559
26	1.634	1.777	1.751	1.793	1.803	1.784	1.675	1.802	1.828	1.785	1.780	1.649
27	1.787	1.741	1.815	1.872	1.766	1.795	1.827	1.744	1.751	1.803	1.833	1.813
28	1.865	1.805	1.821	1.707	1.760	1.832	1.706	1.769	1.672	1.647	1.536	1.590
29	1.861	1.842	1.776	1.806	1.445	1.836	1.674	1.827	1.766	1.782	1.764	1.795
30	1.853	1.801	1.459	1.765	1.685	1.861	1.679	1.713	1.799	1.775	1.550	1.827
31	1.846	1.590	1.670	1.662	1.730	1.771	1.797	1.834	1.847	1.755	1.722	1.819
32	1.843	1.516	1.823	1.662	1.820	1.833	1.656	1.833	1.841	1.798	1.822	1.797
33	1.790	1.848	1.855	1.049	1.296	1.840	1.439	1.851	1.806	1.795	1.873	1.493
34	1.849	1.794	1.803	1.827	1.577	1.734	1.638	1.878	1.729	1.812	1.844	1.763
35	1.793	1.443	1.768	1.821	1.473	1.744	1.343	1.830	1.856	1.778	1.725	1.805
36	1.782	1.618	1.847	1.818	1.893	1.850	1.822	1.796	1.837	1.804	1.840	1.806
37	1.781	1.808	1.894	1.855	1.507	1.808	1.237	1.825	1.796	1.753	1.454	1.762
38	1.855	1.767	1.858	1.827	1.673	1.644	1.734	1.864	1.839	1.726	1.862	1.264
39	1.764	1.804	1.856	1.901	1.500	1.787	1.838	1.853	1.807	1.731	1.745	1.545
40	1.821	1.793	1.768	1.804	1.479	1.806	1.839	1.757	1.826	1.764	1.751	1.790
41	1.862	1.759	1.816	1.835	1.442	1.821	1.696	1.871	1.793	1.726	1.849	1.478
42	1.784	1.764	1.801	1.755	1.678	1.824	1.659	1.849	1.791	1.857	1.644	1.673
43	1.826	1.846	1.824	1.725	1.805	1.816	1.590	1.842	1.744	1.820	1.462	1.708
44	1.391	1.797	1.788	1.846	1.804	1.726	1.781	1.837	1.616	1.747	1.657	1.790
45	1.820	1.803	1.462	1.816	1.814	1.802	1.831	1.612	1.655	1.801	1.805	1.656
46	1.717	1.776	1.782	1.829	1.791	1.678	1.831	1.786	1.414	1.785	1.808	1.636

47	1.747	1.815	1.862	1.884	1.628	1.754	1.760	1.815	1.862	1.827	1.828	1.739
48	1.838	1.822	1.728	1.903	1.767	1.804	1.701	1.821	1.765	1.861	1.735	1.649
49	1.841	1.773	1.717	1.712	1.815		1.859	1.784	1.858	1.667	1.794	1.808
50	1.831		1.720	1.892	1.862		1.757	1.721	1.837	1.783	1.791	1.778
51			1.823	1.852	1.757		1.714	1.704	1.814		1.728	1.825
52				1.664			1.626	1.748	1.759		1.724	1.841
53				1.861			1.793	1.734			1.762	1.837
54				1.543			1.801	1.777			1.892	1.810
55				1.799			1.644	1.732			1.569	1.905
56				1.720			1.576	1.721				1.826
57				1.815			1.486	1.598				1.776
58				1.818			1.301	1.790				1.796
59				1.765			1.319	1.687				1.842
60				1.624			1.338	1.661				1.830
61				1.762				1.773				1.600
62				1.745				1.737				1.771
63				1.795				1.590				1.766
64				1.551				1.821				1.601
65				1.747				1.870				1.499
66				1.845				1.738				1.474
67				1.714				1.616				1.741
68				1.803				1.771				1.791
69				1.796				1.839				1.805
70				1.740				1.659				1.833
71				1.836				1.736				1.757
72				1.894				1.738				1.726
73				1.767				1.673				1.800
74				1.770								1.749
75				1.293								1.738
76				1.633								1.594
77				1.794								1.698
78				1.818								1.687
79				1.841								
80				1.843								
81				1.768								
82				1.867								
83				1.847								

Table S8. DFA (Detrended Fluctuation Analysis) scaling exponent γ values for the 700 experimental cell trajectories.

Table S9

Cell number	Amoeba proteus				Metamoeba leningradensis				Amoeba borokensis			
	Sc1	Sc2	Sc3	Sc4	Sc1	Sc2	Sc3	Sc4	Sc1	Sc2	Sc3	Sc4
1	0.582	0.412	0.344	0.615	0.325	0.660	0.531	0.350	0.410	0.474	0.310	0.659
2	0.570	0.670	0.577	0.505	0.477	0.624	0.507	0.426	0.312	0.389	0.519	0.351
3	0.318	0.470	0.472	0.482	0.291	0.329	0.506	0.507	0.500	0.349	0.563	0.443
4	0.496	0.346	0.421	0.411	0.529	0.510	0.694	0.293	0.512	0.393	0.351	0.410
5	0.577	0.579	0.568	0.350	0.280	0.603	0.499	0.610	0.435	0.456	0.497	0.536
6	0.435	0.435	0.483	0.383	0.462	0.323	0.454	0.448	0.352	0.373	0.627	0.372
7	0.493	0.411	0.472	0.464	0.699	0.591	0.357	0.539	0.509	0.467	0.424	0.509
8	0.582	0.467	0.570	0.592	0.566	0.563	0.421	0.500	0.393	0.633	0.621	0.417
9	0.725	0.491	0.249	0.330	0.402	0.496	0.400	0.508	0.628	0.519	0.620	0.520
10	0.515	0.637	0.615	0.615	0.322	0.413	0.560	0.439	0.421	0.545	0.397	0.347
11	0.320	0.637	0.618	0.493	0.476	0.501	0.491	0.514	0.380	0.417	0.376	0.555
12	0.492	0.631	0.420	0.331	0.510	0.640	0.585	0.275	0.580	0.450	0.314	0.584
13	0.479	0.326	0.488	0.505	0.397	0.628	0.274	0.667	0.520	0.554	0.522	0.680
14	0.591	0.435	0.542	0.382	0.443	0.314	0.396	0.431	0.434	0.447	0.390	0.561
15	0.480	0.353	0.456	0.546	0.444	0.433	0.401	0.358	0.607	0.461	0.337	0.460
16	0.559	0.500	0.517	0.460	0.330	0.322	0.439	0.375	0.567	0.538	0.493	0.432
17	0.498	0.523	0.641	0.525	0.315	0.590	0.288	0.566	0.460	0.462	0.419	0.583
18	0.618	0.530	0.499	0.450	0.362	0.578	0.500	0.560	0.515	0.616	0.335	0.446
19	0.480	0.476	0.375	0.434	0.492	0.339	0.523	0.654	0.458	0.453	0.473	0.437
20	0.565	0.440	0.504	0.563	0.369	0.616	0.385	0.330	0.411	0.372	0.544	0.517
21	0.573	0.505	0.409	0.564	0.470	0.567	0.401	0.395	0.701	0.290	0.435	0.599
22	0.361	0.581	0.346	0.397	0.507	0.384	0.538	0.407	0.566	0.375	0.496	0.492
23	0.448	0.507	0.451	0.381	0.430	0.346	0.572	0.534	0.471	0.541	0.450	0.558
24	0.605	0.530	0.403	0.526	0.631	0.368	0.278	0.584	0.388	0.485	0.444	0.552
25	0.348	0.491	0.545	0.344	0.302	0.405	0.597	0.602	0.536	0.531	0.587	0.443
26	0.427	0.401	0.455	0.481	0.368	0.474	0.567	0.464	0.563	0.429	0.472	0.530
27	0.644	0.328	0.413	0.409	0.390	0.437	0.445	0.507	0.456	0.249	0.689	0.542
28	0.495	0.504	0.412	0.566	0.448	0.509	0.560	0.452	0.557	0.284	0.548	0.403
29	0.494	0.526	0.414	0.657	0.466	0.645	0.420	0.417	0.452	0.560	0.449	0.447
30	0.564	0.293	0.514	0.446	0.445	0.481	0.348	0.446	0.313	0.380	0.508	0.645
31	0.430	0.465	0.473	0.399	0.500	0.464	0.410	0.400	0.395	0.361	0.301	0.411
32	0.506	0.352	0.442	0.257	0.477	0.539	0.523	0.345	0.544	0.649	0.449	0.345
33	0.420	0.530	0.329	0.534	0.506	0.635	0.802	0.522	0.578	0.386	0.452	0.540
34	0.557	0.509	0.569	0.410	0.481	0.714	0.510	0.565	0.431	0.449	0.400	0.560
35	0.454	0.572	0.415	0.417	0.545	0.446	0.389	0.351	0.486	0.587	0.466	0.619
36	0.451	0.542	0.471	0.417	0.547	0.410	0.368	0.454	0.389	0.757	0.421	0.393
37	0.584	0.623	0.370	0.476	0.470	0.401	0.487	0.413	0.382	0.603	0.401	0.494
38	0.365	0.495	0.549	0.411	0.561	0.486	0.393	0.488	0.348	0.495	0.431	0.371
39	0.503	0.435	0.591	0.513	0.710	0.570	0.535	0.373	0.500	0.619	0.529	0.514
40	0.514	0.561	0.505	0.445	0.454	0.339	0.404	0.320	0.337	0.500	0.385	0.490
41	0.702	0.413	0.502	0.441	0.507	0.464	0.646	0.438	0.453	0.400	0.530	0.669
42	0.456	0.405	0.366	0.579	0.534	0.646	0.589	0.626	0.525	0.654	0.471	0.416
43	0.459	0.509	0.531	0.556	0.249	0.461	0.511	0.472	0.375	0.433	0.571	0.484
44	0.483	0.516	0.247	0.391	0.446	0.483	0.494	0.465	0.393	0.378	0.549	0.394
45	0.542	0.411	0.480	0.762	0.442	0.337	0.516	0.350	0.306	0.507	0.655	0.415
46	0.488	0.460	0.489	0.321	0.595	0.563	0.419	0.521	0.447	0.486	0.273	0.412

47	0.642	0.461	0.458	0.316	0.471	0.571	0.555	0.592	0.396	0.511	0.378	0.571
48	0.525	0.425	0.495	0.521	0.311	0.517	0.516	0.577	0.537	0.473	0.660	0.476
49	0.525	0.352	0.636	0.463	0.467		0.508	0.640	0.597	0.349	0.424	0.633
50	0.462		0.667	0.441	0.462		0.539	0.706	0.515	0.526	0.418	0.431
51			0.326	0.488	0.427		0.488	0.478	0.472		0.341	0.447
52				0.481			0.407	0.509	0.401		0.479	0.373
53				0.480			0.586	0.646			0.677	0.445
54				0.377			0.527	0.243			0.374	0.533
55				0.308			0.626	0.545			0.403	0.366
56				0.355			0.509	0.408				0.361
57				0.491			0.396	0.458				0.633
58				0.556			0.570	0.603				0.460
59				0.695			0.469	0.445				0.369
60				0.560			0.351	0.333				0.493
61				0.426				0.562				0.351
62				0.585				0.464				0.628
63				0.475				0.562				0.604
64				0.603				0.564				0.544
65				0.449				0.361				0.537
66				0.349				0.527				0.417
67				0.638				0.512				0.586
68				0.578				0.474				0.384
69				0.478				0.394				0.565
70				0.469				0.369				0.325
71				0.331				0.480				0.568
72				0.470				0.551				0.320
73				0.510				0.559				0.628
74				0.510								0.472
75				0.659								0.632
76				0.599								0.414
77				0.323								0.426
78				0.503								0.492
79				0.442								
80				0.549								
81				0.431								
82				0.511								
83				0.550								

Table S9. DFA (Detrended Fluctuation Analysis) scaling exponent γ values for the 700 shuffled cell trajectories.

Table S10

Species	Scenario	Intensity of Response	Directionality Ratio	Average Speed
Amoeba Proteus	Sc1-Sc2	0.006	10^{-8}	0.002
	Sc1-Sc3	0.959	0.080	10^{-4}
	Sc1-Sc4	0.155	0.084	10^{-12}
	Sc2-Sc3	10^{-4}	10^{-7}	0.544
	Sc2-Sc4	10^{-8}	10^{-7}	0.001
	Sc3-Sc4	0.100	0.865	0.005
	All	10^{-6}	10^{-9}	10^{-10}
Metamoeba leningradensis	Sc1-Sc2	10^{-8}	10^{-8}	10^{-4}
	Sc1-Sc3	0.098	0.734	0.004
	Sc1-Sc4	10^{-13}	10^{-6}	10^{-13}
	Sc2-Sc3	10^{-5}	10^{-9}	0.899
	Sc2-Sc4	0.010	0.050	10^{-8}
	Sc3-Sc4	10^{-11}	10^{-7}	10^{-8}
	All	10^{-16}	10^{-12}	10^{-15}
Amoeba borokensis	Sc1-Sc2	10^{-4}	10^{-5}	0.160
	Sc1-Sc3	0.884	0.918	0.326
	Sc1-Sc4	0.099	0.862	10^{-4}
	Sc2-Sc3	0.003	10^{-4}	0.982
	Sc2-Sc4	0.029	10^{-4}	0.119
	Sc3-Sc4	0.133	0.659	0.230
	All	0.002	10^{-5}	0.022
Amoeba proteus Vs. Metamoeba leningradensis	Sc1-Sc2	0.263	10^{-7}	10^{-8}
	Sc1-Sc3	0.006	0.318	10^{-7}
	Sc1-Sc4	10^{-4}	10^{-5}	0.417
	Sc2-Sc3	10^{-8}	10^{-10}	0.010
	Sc2-Sc4	0.573	0.002	10^{-4}
	Sc3-Sc4	10^{-4}	0.003	10^{-5}
	All	0.293	0.793	0.065
Amoeba proteus Vs. Amoeba borokensis	Sc1-Sc2	0.537	10^{-5}	10^{-11}
	Sc1-Sc3	0.002	0.895	10^{-8}
	Sc1-Sc4	0.009	0.559	10^{-12}
	Sc2-Sc3	10^{-8}	10^{-7}	10^{-4}
	Sc2-Sc4	10^{-9}	10^{-7}	10^{-5}
	Sc3-Sc4	0.004	0.106	10^{-4}
	All	10^{-10}	0.032	10^{-17}
Metamoeba leningradensis Vs. Amoeba borokensis	Sc1-Sc2	10^{-5}	10^{-6}	0.064
	Sc1-Sc3	0.383	0.515	0.131
	Sc1-Sc4	0.005	0.228	0.001
	Sc2-Sc3	10^{-6}	10^{-7}	0.085
	Sc2-Sc4	10^{-5}	10^{-7}	0.375
	Sc3-Sc4	0.529	0.117	0.401
	All	10^{-5}	0.067	10^{-7}

Table S10. Kruskal-Wallis and Wilcoxon analyses to assess the variability in kinetic properties.