

## Transcription factors ER $\alpha$ and Sox2 have differing multiphasic DNA and RNA binding mechanisms

*Wayne O. Hemphill<sup>1,2,†</sup>, Halley R. Steiner<sup>1,†</sup>, Jackson R. Kominsky<sup>1,2,†</sup>, Deborah S. Wuttke<sup>1,\*</sup>,  
Thomas R. Cech<sup>1,2,\*</sup>*

<sup>1</sup> Department of Biochemistry, University of Colorado Boulder, Boulder, Colorado 80303 USA

<sup>2</sup> Howard Hughes Medical Institute and BioFrontiers Institute, University of Colorado Boulder, Boulder, Colorado 80303 USA

<sup>†</sup> Authors contributed equally

\* Corresponding authors: [thomas.cech@colorado.edu](mailto:thomas.cech@colorado.edu) & [deborah.wuttke@colorado.edu](mailto:deborah.wuttke@colorado.edu)

**Running Title:** ER $\alpha$  and Sox2 exhibit multiphasic binding kinetics

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## ABSTRACT

Many transcription factors (TFs) have been shown to bind RNA, leading to open questions regarding the mechanism(s) of this RNA binding and its role in regulating TF activities. Here we use biophysical assays to interrogate the  $k_{on}$ ,  $k_{off}$ , and  $K_d$  for DNA and RNA binding of two model human transcription factors, ER $\alpha$  and Sox2. Unexpectedly, we found that both proteins exhibited multiphasic nucleic acid binding kinetics. We propose that Sox2 RNA and DNA multiphasic binding kinetics could be explained by a conventional model for sequential Sox2 monomer association and dissociation. In contrast, ER $\alpha$  nucleic acid binding exhibited biphasic dissociation paired with novel triphasic association behavior, where two apparent binding transitions are separated by a 10-20 min “lag” phase depending on protein concentration. We considered several conventional models for the observed kinetic behavior, none of which adequately explained all the ER $\alpha$  nucleic acid binding data. Instead, simulations with a model incorporating sequential ER $\alpha$  monomer association, ER $\alpha$  nucleic acid complex isomerization, and product “feedback” on isomerization rate recapitulated the general kinetic trends for both ER $\alpha$  DNA and RNA binding. Collectively, our findings reveal that Sox2 and ER $\alpha$  bind RNA and DNA with previously unappreciated multiphasic binding kinetics, and that their reaction mechanisms differ with ER $\alpha$  binding nucleic acids via a novel reaction mechanism.

## INTRODUCTION

The human genome encodes ~1500 transcription factors (TFs) (Ignatieva et al. 2015; Zhang et al. 2012; Wingender et al. 2013, 2015), which direct cell-type specificity and gene expression programs by interacting with a multitude of binding partners (Spitz and Furlong 2012). TFs modulate transcription by utilizing their DNA-binding domains to stably interact with DNA elements, such as promoters and enhancers, with sequence specificity, and subsequently recruit various coactivator and repressor proteins via their effector domains (Schwabe et al. 1993; Frietze and Farnham 2011). However, the site of transcription is immersed in more than DNA and protein – it's also crowded with RNA. Thousands of RNA species are produced at loci where TFs are bound, such as mRNA, enhancer RNAs, promoter antisense RNAs, and chromatin-enriched long noncoding RNAs (Han and Li 2022; Yang et al. 2021; Werner and Ruthenburg 2015). Additionally, long noncoding RNAs transcribed distally, even kilobases away, are capable of engaging in long-range interactions with chromatin (Mishra and Kanduri 2019; Rinn and Chang 2020). The prevalence of RNA at chromatin begs the question of whether RNA plays a direct role in regulating TFs.

Many TFs have been shown to bind RNA (G Hendrickson et al. 2016; Khalil et al. 2009; Skalska et al. 2021; Oksuz et al. 2023; Hudson and Ortlund 2014; Parsonnet et al. 2019), raising the question of whether RNA directly regulates TF function. In many cases, the TF's RNA binding domains are adjacent to their DNA binding domains (Oksuz et al. 2023). Estrogen receptor  $\alpha$  (ER $\alpha$ ) (Steiner et al. 2022) and sex determining region Y box 2 (Sox2) (Holmes et al. 2020) are two such TFs that bind DNA and RNA competitively with tight affinities, suggesting potential biological relevance for the RNA binding activity. ER $\alpha$  and Sox2 can therefore be used as model systems to study RNA regulation of TF activities. Although RNA-DNA competition experiments provide some useful information, a detailed investigation of the mechanism(s) and kinetics for TF polynucleotide association and dissociation is critical for understanding how TFs could be regulated by RNA binding.

ER $\alpha$  is a ligand-activated TF, which functions as the nuclear receptor for estrogen, a hormone that dictates reproductive development (Björnström and Sjöberg 2005; Deroo and Korach 2006) (mouse studies reviewed in ref (Hewitt and Korach 2018)). Abnormal ER $\alpha$  signaling leads to a variety of diseases such as metabolic and cardiovascular disease, neurodegeneration, and inflammation (Jia et al. 2015). Additionally,

ER $\alpha$  is aberrantly expressed in 80% of breast cancers, making it a recurrent therapeutic target (Alluri et al. 2014). ER $\alpha$  is a 595 amino acid polypeptide (~66 kDa) comprised of six domains, including DNA-binding, ligand-binding and transcriptional activation domains (Hewitt and Korach 2018; Ponglikitmongkol et al. 1988). Its DNA-binding domain (DBD) facilitates sequence-specific DNA binding to the palindromic estrogen response element (ERE) motif (GGTCAnnnTGACC) and binds as a dimer via its two zinc finger elements (Kuntz and Shapiro 1997; Helsen et al. 2012; Schwabe et al. 1993). The hinge region sits just C-terminal of the DBD, and recent work has demonstrated that part of the hinge region is critical for RNA binding (but dispensable for DNA binding). ER $\alpha$  uses a combination of the DBD and hinge elements to preferentially bind hairpin RNA (hRNA) with no apparent sequence specificity (Steiner et al. 2022; Xu et al. 2021). While *in vitro* experiments indicate that ER $\alpha$  RNA and DNA binding are competitive, and ER $\alpha$  has been shown to interact with RNA *in vivo* (Xu et al. 2021; Nassa et al. 2019), the question of how RNA may regulate ER $\alpha$ -DNA interactions *in vivo* remains an active area of investigation (Steiner et al. 2022).

Sox2, a member of the SoxB1 TF family, regulates pluripotency in embryonic stem cells via expression of the pluripotency-associated TFs Oct4 and Nanog and via repression of lineage-specific genes (Avilion et al. 2003; Zhang and Cui 2014; Chew et al. 2005). Additionally, Sox2 is critical for differentiating pluripotent stem cells to neural progenitors and maintaining the properties of neural progenitor stem cells (Zhang and Cui 2014). In mice, deletion of Sox2 is embryonic lethal (Avilion et al. 2003), while knockout in adult mice leads to the loss of hippocampal neurogenesis (Favaro et al. 2009). In humans, mutations in Sox2 have been associated with eye defects such as bilateral anophthalmia and microphthalmia (Fantes et al. 2003; Chassaing et al. 2014), as well as cognitive abnormalities (Sisodiya et al. 2006; Kelberman et al. 2006). Functional Sox2 contains 317 amino acids partitioned into two key domains (Nowling et al. 2000). The Sox2 high mobility group (HMG) domain binds DNA at the minor groove, and recognizes a species-specific sequence centered around four highly conserved nucleotides (CCCATTTGTTTC in humans) (Grosschedl et al. 1994; Dodonova et al. 2020; Schaefer and Lengerke 2020; Yesudhas et al. 2017; Hou et al. 2017; Weiss 2001). *In vivo* studies have suggested that lncRNAs interact directly with Sox2 to regulate its function(s) in stem cell pluripotency (Ng et al. 2012). Subsequent *in vitro* findings show that the Sox2 HMG domain preferentially binds the double-stranded RNA within hRNA with no apparent sequence specificity (Holmes et al. 2020). Like ER $\alpha$ , Sox2 HMG domain binding to RNA and DNA was found to be competitive (Holmes et al. 2020). However, another study suggests

that a novel RNA-binding module C-terminal of the HMG domain also contributes to Sox2 RNA binding, and that Sox2 can stably bind RNA and DNA simultaneously (Hou et al. 2020).

To determine the mechanisms of RNA and DNA binding on the Sox2 and ER $\alpha$  binding surfaces, we used fluorescence polarization (FP) and surface plasmon resonance (SPR) to measure their RNA and DNA association and dissociation kinetics. In contrast to the expectation for a simple binding reaction, both TFs exhibited complex multiphasic association and dissociation kinetics from RNA and DNA. We evaluated several common models for multiphasic association and/or dissociation to describe the observed kinetics for the two TF interactions with RNA and DNA. These findings reveal a previously unappreciated level of complexity in the ER $\alpha$  and Sox2 interactions with nucleic acids, and they suggest that the two TFs achieve multiphasic kinetics through different mechanisms.

## RESULTS

### ER $\alpha_{\text{DBD-Ext}}$ and Sox2 $_{\text{HMG}}$ equilibrium ligand binding

ER $\alpha_{180-280}$ , a region of the protein containing the canonical DBD and a set of basic residues from the hinge region (ER $\alpha_{\text{DBD-Ext}}$ ), and recombinant Sox2 $_{40-123}$ , the region of the protein containing the high mobility group (HMG) domain (Sox2 $_{\text{HMG}}$ ), were expressed and purified as previously described (Steiner et al. 2022; Holmes et al. 2020). We performed FP-based binding experiments with ER $\alpha_{\text{DBD-Ext}}$  and Sox2 $_{\text{HMG}}$  and a variety of dsDNA and RNA ligands to assess the binding affinities (Supplemental Figure 1) at the same experimental conditions used to measure binding kinetics.

For ER $\alpha_{\text{DBD-Ext}}$ , we tested an 18-bp dsDNA containing its palindromic ERE recognition sequence (ERE dsDNA), a 15-bp dsDNA containing only half of its palindromic recognition sequence ( $\Delta$ ERE dsDNA), and a 37-nt hairpin RNA (hRNA) derived from the X-box binding protein 1 (XBP1) mRNA sequence (XBP1 hRNA) (Steiner et al. 2022). We found that ER $\alpha_{\text{DBD-Ext}}$  bound ERE with high affinity ( $K_d^{\text{app}} \approx 11$  nM, see Table 1 for error) and positive cooperativity ( $n \approx 2.1$ ) (Supplemental Figure 1a and Table 1), while ER $\alpha_{\text{DBD-Ext}}$  bound  $\Delta$ ERE with comparable to higher affinity ( $K_d^{\text{app}} \approx 2.8$  nM) and less to no positive cooperativity ( $n \approx 1.5$ ) (Supplemental

Figure 1a and Table 1). ER $\alpha$ <sub>DBD-Ext</sub> bound the XBP1 hRNA with lower affinity ( $K_d^{app} \approx 370$  nM) and no apparent cooperativity ( $n \approx 0.91$ ) (Supplemental Figure 1a and Table 1). These findings are consistent with prior studies (Steiner et al. 2022). We note that the anisotropy dynamic range was less for the  $\Delta$ ERE versus ERE binding curve, consistent with a lower TF-DNA binding stoichiometry for  $\Delta$ ERE versus ERE, as expected from prior studies (Steiner et al. 2022; Schwabe et al. 1993).

For Sox2<sub>HMG</sub>, we tested a 10-bp dsDNA containing its cognate binding sequence (CBS dsDNA) (Holmes et al. 2020), and for comparison we also measured the binding affinities for a 19-bp dsDNA with a nonspecific binding sequence (NBS dsDNA), a 43-nt hairpin RNA (hRNA), a (G<sub>3</sub>A<sub>2</sub>)<sub>4</sub> RNA that adopts a G-quadruplex (G4) structure (rG4), and a 20-nt poly-A RNA (rA20) (Supplemental Figure 1b and Table 1). Our findings indicate that Sox2<sub>HMG</sub> binding to CBS dsDNA and G4 RNA were best described by a two-transition binding curve, while Sox2<sub>HMG</sub> binding to NBS dsDNA, hRNA, and poly-A RNA fit well to a standard Hill binding equation. The Sox2<sub>HMG</sub> CBS and rG4 high-affinity binding transitions both had  $K_d^{app} \leq 1$  nM, being limited by the ligand concentration in our assays, while their lower-affinity binding transitions had  $K_d^{app}$  of 420 and 110 nM, respectively. For CBS dsDNA, this was previously attributed to Sox2<sub>HMG</sub> initial sequence-specific binding versus subsequent nonspecific binding (Holmes et al. 2020; Hamilton et al. 2022). Relative to the high-affinity binding transition, Sox2<sub>HMG</sub> exhibited  $\geq 30$ x greater affinity for CBS dsDNA and G4 RNA than for NBS dsDNA, hRNA, and poly(A) RNA ( $K_d^{app} \approx 29$ -53 nM). We also note that the Sox2<sub>HMG</sub> NBS dsDNA and poly(A) RNA binding curves exhibited modest negative cooperativity ( $n \approx 0.57$ -0.68), while Sox2<sub>HMG</sub> bound the hRNA without apparent cooperativity (Supplemental Figure 1b and Table 1). All these findings are in agreement with prior studies (Hamilton et al. 2022; Holmes et al. 2020), validating the reagents and methods for the subsequent measurements below.

### **ER $\alpha$ <sub>DBD-Ext</sub> and Sox2<sub>HMG</sub> ligand dissociation are multiphasic**

We measured the ER $\alpha$ <sub>DBD-Ext</sub> and Sox2<sub>HMG</sub> ligand dissociation kinetics using FP-based competitive dissociation (FPCD) experiments. These involve pre-incubation of protein and fluorescently labeled nucleic acid followed by self-competition with unlabeled nucleic acid and observation of binding states by FP (Figure 1a). Contrary to the expectation for a simple binding scheme (i.e., protein + ligand  $\leftrightarrow$  protein-ligand), both

ER $\alpha$ <sub>DBD-Ext</sub> and Sox2<sub>HMG</sub> exhibited multiphasic dissociation from all ligands that we tested (Figure 1b-c). For ER $\alpha$ <sub>DBD-Ext</sub>, the ligand dissociation curves were well fit by bi-exponential regression and produced similar rate constants for all the nucleic acids tested. The rate constants and other relevant parameter values for these regressions are summarized in Table 1.

We then asked what could be producing the biphasic dissociation curves for our TF nucleic acid interactions. Previously, we demonstrated that direct transfer is used by multiple nucleic acid binding proteins to transfer between polynucleotide species through unstable ternary intermediates (Hemphill et al. 2023). Furthermore, at the competitor concentrations used in our FPCD experiments (Figure 1), protein-polynucleotide dissociation might occur via both direct transfer and intrinsic dissociation in comparable proportions (Hemphill et al. 2023). If the fast components of the ER $\alpha$ <sub>DBD-Ext</sub> biphasic dissociation curves were the result of ligand displacement via direct transfer, their dissociation curves should become monophasic slow in the absence of competitor. To test this hypothesis, we induced complex dissociation by dilution rather than competitor addition with FP-based jump dilution (FPJD) experiments (Figure 2).

For ER $\alpha$ <sub>DBD-Ext</sub>, we measured  $\Delta$ ERE dsDNA dissociation since it had the binding properties most compatible with the limitations of FPJD methodology. Notably,  $\Delta$ ERE dissociation was still biphasic in the absence of competitor (Figure 2), with no apparent reduction in the contribution of the fast component to the bi-exponential regression (FPCD  $\approx$  60%, FPJD  $\approx$  69%). In essence, the biphasic nature of the dissociation curves was not attributable to direct transfer. However, the presence of competitor appeared to greatly (4-5x) increase the rate of the fast component (FPCD<sub>fast</sub>  $\approx$   $8.9 \times 10^{-2} \text{ s}^{-1}$  vs FPJD<sub>fast</sub>  $\approx$   $1.9 \times 10^{-2} \text{ s}^{-1}$ ), but not slow component (FPCD<sub>slow</sub>  $\approx$   $5.9 \times 10^{-4} \text{ s}^{-1}$  vs FPJD<sub>slow</sub>  $\approx$   $4.2 \times 10^{-4} \text{ s}^{-1}$ ), of the bi-exponential regression. This suggests that the ER $\alpha$ <sub>DBD-Ext</sub>- $\Delta$ ERE complex state associated with fast dissociation is susceptible to direct transfer, while the state associated with slow dissociation is not. For Sox2<sub>HMG</sub>, we are unable to make a similar assessment, because the FPJD assay produces less signal-to-noise relative to the FPCD assay and the Sox2<sub>HMG</sub>-CBS interaction has a lower anisotropy dynamic range than the ER $\alpha$ <sub>DBD-Ext</sub>-ERE interaction. Thus, the Sox2<sub>HMG</sub> dissociation curves were too noisy for reliable analysis.

Ruling out direct transfer as the origin of the biphasic kinetics, we moved on to a second hypothesis. A prior *in vitro* study demonstrated that the RNA binding affinity of our ER $\alpha$ <sub>DBD-Ext</sub> construct is facilitated by the stretch of basic residues from the hinge region of the protein, while these residues don't significantly affect

binding affinity for ERE dsDNA (Steiner et al. 2022). Thus, we assessed whether these additional nucleic acid binding residues in the construct explain the biphasic nature of our  $ER\alpha_{DBD-Ext}$  dsDNA dissociation kinetics by providing a lower-affinity alternative binding site. We therefore compared the dsDNA dissociation kinetics for the  $ER\alpha_{DBD-Ext}$  construct to a construct lacking the basic hinge region residues ( $ER\alpha_{DBD}$ ) by FPCD (Figure 1a). Our findings indicated that  $ER\alpha$  dsDNA dissociation kinetics were still biphasic with  $ER\alpha_{DBD}$ , ruling out these additional basic residues as the cause for biphasic dissociation (Supplemental Figure 2).

### **$ER\alpha_{DBD-Ext}$ , but not Sox2<sub>HMG</sub>, exhibits multiphasic association to target DNA**

The observation of biphasic ligand dissociation kinetics implies the presence of multiple complex states. To probe this further, we performed FP-based ligand association experiments for  $ER\alpha_{DBD-Ext}$  binding to ERE,  $\Delta$ ERE, and XBP1, and for Sox2<sub>HMG</sub> binding to CBS and rG4 (Figure 3a-e). Sox2<sub>HMG</sub> binding to CBS dsDNA was strictly monophasic (Supplemental Figure 3d). Sox2<sub>HMG</sub> rG4 association appeared classically biphasic (i.e., fitting a bi-exponential) (Supplemental Figure 3e), where the second Sox2<sub>HMG</sub> rG4 association phases in the bi-exponential regressions were  $\sim 15$ x slower than the first phases. We note, however, that percent slow association phase contributions trended downward from  $\sim 35\%$  at  $1 \mu M$  protein to  $<10\%$  at  $8 nM$  protein (Supplemental Figure 3e), resulting in monophasic association at lower protein concentrations. This trend seemed to correlate to the second transition in the binding curve, and it suggests a monomer-dimer equilibrium (Supplemental Figure 1b).

In contrast,  $ER\alpha_{DBD-Ext}$  exhibited multiphasic association for both DNA and RNA (Figure 3a-c). For dsDNA association, we observed a highly unusual triphasic association that was protein concentration dependent. An approximately monophasic association phase was complete in  $\sim 2$  minutes (Supplemental Figure 3a-b), followed by a 10-20 minute “lag” phase, and ending in a second association phase that completes on a similar time scale as the first association phase (Figure 3a-b). Perplexingly, the “lag” phase was only evident at protein concentrations  $\leq 10 \times K_d$  of the DNA, while at higher concentrations  $ER\alpha_{DBD-Ext}$  DNA association appeared monophasic (Figure 3a-b). For  $ER\alpha_{DBD-Ext}$  hRNA association, we observed what appeared to be biphasic association, but regression with a bi-exponential equation revealed an inadequate fit (Supplemental Figure 3c). On closer inspection, the  $ER\alpha_{DBD-Ext}$  hRNA association curves are more like the



‘transition’ protein concentrations in the ER $\alpha_{\text{DBD-Ext}}$  dsDNA association curves (e.g., Figure 3a, [E] = 125 nM), suggesting similar association mechanisms.

We used the first phases of the association curves to determine apparent initial association rate constants (Figure 3f and Table 1). For ER $\alpha_{\text{DBD-Ext}}$ , we find that initial nucleic acid association is ~7,000-17,000x slower than diffusion-limited binding (Table 1) (Fersht 1985). For Sox2 $_{\text{HMG}}$ , we find that CBS dsDNA and G4 RNA initial associations are likewise slower than diffusion-limited binding (~1,000-1,300x) (Table 1). These rates are notably slower than some TFs, suggesting a potential conformational barrier during initial ER $\alpha_{\text{DBD-Ext}}$  nucleic acid binding (Halford and Marko 2004).

To gain insights into the underlying ER $\alpha_{\text{DBD-Ext}}$  nucleic acid binding mechanism(s), we compared our measured rate constants with the equilibrium binding data. We noted that ER $\alpha_{\text{DBD-Ext}}$  associates at a similar or modestly greater rate with hRNA versus dsDNA (Figure 3f), despite having lower affinity and similar dissociation rates (Figure 1b and Supplemental Figure 1a). Dividing the  $k_{\text{off}}^{\text{app}}$  (apparent dissociation rate constants) by the  $k_{\text{on}}^{\text{app}}$  for the respective ligands, which should yield the  $K_d^{\text{app}}$ , suggests that the ER $\alpha_{\text{DBD-Ext}}$  dsDNA  $K_d^{\text{app}}$  is mostly influenced by the slow bi-exponential dissociation phase ( $k_{\text{off}}^{\text{app}}_{\text{slow}} / k_{\text{on}}^{\text{app}}$ ; ERE = 13 nM,  $\Delta$ ERE = 6.8 nM, XBP1 = 4.7 nM) while the RNA  $K_d^{\text{app}}$  is mostly influenced by the fast bi-exponential dissociation phase ( $k_{\text{off}}^{\text{app}}_{\text{fast}} / k_{\text{on}}^{\text{app}}$ ; ERE = 0.89  $\mu$ M,  $\Delta$ ERE = 1.0  $\mu$ M, XBP1 = 400 nM). We made similar comparisons for the Sox2 $_{\text{HMG}}$  CBS and rG4 interactions ( $k_{\text{off}}^{\text{app}}_{\text{slow}} / k_{\text{on}}^{\text{app}}$  and  $k_{\text{off}}^{\text{app}}_{\text{fast}} / k_{\text{on}}^{\text{app}}$ ; CBS = 1.7 and 67 nM, rG4 = 0.87 and 49 nM). These values suggest that the Sox2 $_{\text{HMG}}$  CBS and rG4 fast versus slow dissociation phases (Figure 1c) could correspond to the complex states in the low versus high affinity binding curve transitions (Supplemental Figure 1b), respectively. We note for the Sox2 $_{\text{HMG}}$  G4 RNA interaction that association was biphasic (Supplemental Figure 3e), while the use of  $k_{\text{on}}^{\text{app}}$  in these calculations corresponds to the initial association phase only.

### ER $\alpha_{\text{DBD-Ext}}$ multiphasic dissociation is not due to a “locked” binding conformation

We then sought a molecular model to explain the multiphasic dissociation kinetics observed for ER $\alpha_{\text{DBD-Ext}}$ . An *in vitro* study of the full-length glucocorticoid receptor (GR), a nuclear hormone receptor with strong similarities to ER $\alpha$ , reported multiphasic dsDNA dissociation kinetics remarkably similar to the ER $\alpha_{\text{DBD-Ext}}$

dsDNA dissociation kinetics observed here (De Angelis et al. 2015). Those authors proposed a “locked” binding conformation model to explain GR multiphasic ligand dissociation kinetics (Supplemental Figure 4a). This model suggests that after initial GR-dsDNA association, the complex can slowly isomerize to an alternative state but must slowly isomerize back to the initial complex state before ligand dissociation can occur. A prediction of this model is that if a brief protein-ligand incubation period is allowed before complex dissociation is induced (e.g., by competitor addition), then the complex should not have time to isomerize to the more stable alternative state, and the slow phase of the dissociation kinetics should be ablated.

To test if  $ER\alpha_{DBD-Ext}$  ligand dissociation kinetics could be explained by the “locked” binding conformation model, we conducted FPCD experiments with variable protein-ligand incubation times. Our findings indicated that 2-min versus 60-min protein-ligand incubations produced similarly biphasic  $ER\alpha_{DBD-Ext}$  dsDNA dissociation, with slow phase contributions of 30-40% based on biexponential regression (Supplemental Figure 4a-b). In contrast, based on our Figure 1 data, the GR model predicts a ~5% slow phase contribution after a 2-min incubation. The behavior was somewhat different when RNA was the ligand – the shorter incubation time did affect  $ER\alpha_{DBD-Ext}$  hRNA dissociation, but by partially reducing the fast phase of the bi-exponential regression instead of the anticipated slow phase reduction (Supplemental Figure 4c). It’s notable that the  $ER\alpha_{DBD-Ext}$  hRNA association is incomplete after a 2-min incubation at the protein concentrations used (Supplemental Figure 3c), suggesting that the fast dissociation phase of the bi-exponential regression emerges during the second  $ER\alpha_{DBD-Ext}$  hRNA association phase (Figure 3a-c). Overall, despite the similarities in dissociation kinetics, our data indicate that the previously proposed model for GR multiphasic ligand dissociation does not apply to  $ER\alpha_{DBD-Ext}$  DNA or RNA biphasic dissociation.

While these findings were sufficient to refute one model, we further investigated how  $ER\alpha_{DBD-Ext}$  dsDNA complex stability varied during its multiphasic association to provide insights into alternate models. The above dsDNA experiments used  $ER\alpha_{DBD-Ext}$  concentrations several fold above the ligand  $K_d$ , where nucleic acid association occurs in a single apparent step (Figure 3a). To determine if protein-ligand incubation time affects dissociation kinetics at lower protein concentrations, when association is multiphasic, we performed FPCD experiments under these conditions using ERE dsDNA (Figure 4). We selected protein-ligand incubation times (Figure 4a for reference) just after initial association at the beginning of the “lag” phase (2.5 min), at the end of the “lag” phase before secondary association (15 min), towards the end of secondary association (30 min), and

at binding equilibrium (60 min). These findings indicate that  $ER\alpha_{DBD-Ext}$  ERE dsDNA dissociation is slow and monophasic during the “lag” phase after initial association, but complex dissociation acquires a faster component and becomes biphasic during the second association phase (Figure 4b-e). Curiously, this suggests that the more stable complex state emerges first, followed by the less stable complex state, which contrasts with the positive cooperativity observed by the  $ER\alpha_{DBD-Ext}$  ERE binding curve (Supplemental Figure 1a). Notably, this is the same trend in dissociation behavior over multiphasic association that was observed for the  $ER\alpha_{DBD-Ext}$  XBP1 hRNA interaction above, consistent with the hypothesis that the two ligands could share an underlying mechanism.

### **$ER\alpha_{DBD-Ext}$ multiphasic DNA dissociation is conserved across methodology and temperature**

To ensure that our findings were not due to an unexpected feature of our FP experimental design, we used surface plasmon resonance (SPR) to independently measure  $ER\alpha_{DBD-Ext}$  dsDNA association and dissociation kinetics (Supplemental Figure 5). This also provided the opportunity to obtain data at a second temperature (25°C vs 4°C). Given our assay requirements and the limitations of SPR, we could only measure the association kinetics up to 5 minutes, which is before the secondary association phase that emerged in FP assays. Using SPR association curves to calculate  $ER\alpha_{DBD-Ext}$  apparent association rate constants for ERE and  $\Delta$ ERE dsDNA (Figure 5b), we infer that they are 2-3x higher than the respective values determined by FP (Table 1), which puts them in good agreement given the higher temperature for SPR experiments.

Based on the FP data, we predicted that SPR could be used to test the dissociation behavior at different protein concentrations. As noted above, SPR couldn’t accommodate the time range to fully repeat the FP experiments that revealed multiphasic association (Fig 3) or dsDNA dissociation kinetics over the time course of its multiphasic association (Fig 4). However, we estimated that complexes formed at lower  $ER\alpha_{DBD-Ext}$  concentrations should begin their dissociation curves in the “lag” phase of multiphasic association, while complexes formed at higher protein concentrations should begin their dissociation curves at equilibrium. As predicted, for both ERE and  $\Delta$ ERE dsDNA the  $ER\alpha_{DBD-Ext}$  dissociation curves were biphasic at high  $ER\alpha_{DBD-Ext}$  concentrations but monophasic slow at low  $ER\alpha_{DBD-Ext}$  concentrations (Figure 5c), suggesting that our SPR and FP findings are reporting the same phenomenon. The SPR-derived rate constants for the fast and slow phases

of the bi-exponential regressions (Fig. 5c) were 1-3x greater than their respective FP values (Table 1), which is in good agreement given the temperature differential. We note that the percent contribution of the fast phase to the bi-exponential regression was lower for SPR (32-36%) than for FP (60-71%). However, unlike the FP experiments (Supplemental Figure 1a), the SPR signal had not yet appeared to plateau at the highest protein concentrations, and the percent contribution of the fast phase to the bi-exponential regression still appeared to be increasing with protein concentration (Supplemental Figure 5) suggesting that these values may be more similar at saturating protein concentrations. Collectively, these SPR-based findings independently confirm the kinetic observations for our FP experiments.

## DISCUSSION

ER $\alpha$  and Sox2 have been previously demonstrated to bind RNA with structural specificity *in vitro*, and to associate with RNA *in vivo* (Holmes et al. 2020; Hamilton et al. 2022; Ng et al. 2012; Xu et al. 2021). In addition, their RNA and DNA interactions are reportedly competitive, but initial studies also suggest that the RNA and DNA binding surfaces do not perfectly overlap on the TFs (Steiner et al. 2022; Holmes et al. 2020; Hou et al. 2020). Our work expands on these *in vitro* findings by elucidating the timescales for ER $\alpha$  and Sox2 nucleic acid association and dissociation, and by interrogating their respective RNA versus DNA binding mechanisms.

### A model for Sox2<sub>HMG</sub> DNA and RNA binding

Our kinetic and thermodynamic data allow us to propose a minimum kinetic model for Sox2<sub>HMG</sub> binding to nucleic acids. Binding to target (CBS) dsDNA and G4 RNA exhibited two-transition equilibrium binding (Supplemental Figure 1b), biphasic dissociation (Figure 1c), and monophasic dsDNA association and biphasic G4 RNA association (Figure 3d-e, Supplemental Figure 3d-e). The second, lower affinity Sox2<sub>HMG</sub> CBS and rG4 binding transitions have similar affinities to non-target DNA and RNA. Prior data indicate that Sox2<sub>HMG</sub> can bind DNA and RNA at protein-ligand stoichiometries higher than 1:1 (Holmes et al. 2020; Hamilton et al. 2022; Moosa et al. 2018). We propose that the simplest model to sufficiently explain these findings is a sequential

protein binding model (Figure 6a). We also considered a ligand isomerization (but not protein isomerization) model as shown in Figure 6b that would be consistent with the data if the ligand states were in comparable proportions at equilibrium and had drastically different affinities for the protein. However, single dominant bands were observed via native-PAGE during nucleic acid preparation (see methods), no two-transition binding curves were produced for other dsDNA ligands (Supplemental Figure 1b), and G4 RNAs are normally quite stable *in vitro* under our 135 mM KCl conditions (Lane et al. 2008; Crenshaw et al. 2015). Consequently, we don't favor the Fig. 6b ligand isomerization model.

Under a sequential protein binding model (Figure 6a, see legend for inferred rate constants), Sox2<sub>HMG</sub> initially binds target dsDNA or G4 RNA with high affinity, then an additional Sox2<sub>HMG</sub> monomer (or more) binds with lower affinity. Prior studies attribute this to sequence-specific (or structure-specific) binding followed by nonspecific binding (Hamilton et al. 2022; Holmes et al. 2020). Upon addition of competitor, the fast dissociation phase results from the less stable monomer(s) dissociating followed by slow dissociation of the more stable monomer from the ligand. Since association appears approximately monophasic for DNA and biphasic for RNA (Supplemental Figure 3d-e), we infer that the DNA association rate constants for successive monomers are comparable, but RNA association rate constants decrease with sequential monomer association. Finally, we note that the single-transition Sox2<sub>HMG</sub> binding curves for non-target DNA and RNA support this model – without the tight binding, Sox2<sub>HMG</sub> would only bind weakly with increasing  $K_d$  for each subsequent monomer. Such behavior could produce the low Hill coefficients seen for nontarget DNA and RNA (Supplemental Figure 1b), and it would explain the similar affinities for the nontarget nucleic acids'  $K_d^{app}$  and the target DNA and RNA second transition  $K_d^{app}$ .

## A model for ER $\alpha$ <sub>DBD-Ext</sub> DNA and RNA binding

Synthesis of our thermodynamic and kinetic data allows us to consider several models for DNA and RNA binding by ER $\alpha$ <sub>DBD-Ext</sub>. ER $\alpha$ <sub>DBD-Ext</sub> equilibrium binding exhibited a single-transition with apparent positive cooperativity for ERE dsDNA but not for  $\Delta$ ERE dsDNA or XBP1 hRNA (Supplemental Figure 1a). Existing crystal structures of ER $\alpha$ <sub>DBD</sub> binding to ERE dsDNA reveal that one ER $\alpha$ <sub>DBD</sub> monomer binds each of the two repeats in the palindromic recognition sequence and the two monomers then stabilize one another on DNA

through protein-protein interactions (Schwabe et al. 1993), which explains the apparent positive cooperativity and suggests 2:1 protein-DNA stoichiometry. By extension, it seems reasonable that the loss of one recognition sequence repeat in the  $\Delta$ ERE dsDNA would reduce stoichiometry and cooperativity, consistent with our data (Supplemental Figure 1a). The ER $\alpha_{\text{DBD-Ext}}$  dsDNA and RNA interactions also exhibited biphasic dissociation with similar rate constants (Figure 1b), and for dsDNA this persisted even in the absence of competitor (Figure 2) or hinge residues (Supplemental Figure 2). ER $\alpha_{\text{DBD-Ext}}$  dsDNA association was apparently triphasic at lower protein concentrations, with a 10-20 min “lag” between two typical association phases, while RNA association seemed biphasic but did not adequately fit a standard bi-exponential (Figure 3a-b). Most notably, while the ER $\alpha_{\text{DBD-Ext}}$  dsDNA  $K_d^{\text{app}}$  was primarily predicted by the  $k_{\text{off}}^{\text{app}}_{\text{slow}}$ , the RNA  $K_d^{\text{app}}$  was more influenced by the  $k_{\text{off}}^{\text{app}}_{\text{fast}}$ .

This raises the question of what model best explains the ER $\alpha$  binding data. First, a standard sequential protein binding model for biphasic dissociation (Figure 6a), which was the favored model for Sox2<sub>HMG</sub> binding, is inconsistent with our data. Specifically, we note that this model can only explain biphasic dissociation from a state of saturated ligand binding if subsequent ER $\alpha_{\text{DBD-Ext}}$  monomers bind DNA/RNA much less stably than the first monomer, but ER $\alpha_{\text{DBD-Ext}}$  binding exhibits neither two-transition binding nor a low Hill coefficient (i.e.,  $n < 1$ ) (Supplemental Figure 1a). Furthermore, such a model would not be able to produce the “lagged” triphasic association curves that we observed, even if a slow isomerization step was included between sequential monomer binding events is Figure 6a (this would produce classic biphasic association). Next, the “locked” binding conformation model (Supplemental Figure 4a), which was previously proposed to explain biphasic DNA dissociation for GR (De Angelis et al. 2015), was specifically tested and discounted by our studies (Supplemental Figure 4). We also note that this model could explain monophasic association, or biphasic association if the “locked” complex conformation has significantly altered anisotropy, but it cannot explain our “lagged” triphasic association data (Figure 3). Finally, protein or ligand isomerization models as shown in Figure 6b would require significantly different complex state stabilities to explain our data, but ER $\alpha_{\text{DBD-Ext}}$  DNA/RNA binding exhibits neither two-transition binding nor a low Hill coefficient (Supplemental Figure 1a). In addition, the Fig. 6b models could be reconciled with monophasic or biphasic association, but they cannot explain the “lagged” triphasic association we observed (Figure 3). We considered if heterogeneity in ligand or protein could explain the data, but the RNA/DNA had a single dominant band via native-PAGE (see Methods)

and no protein heterogeneity was observed during size-exclusion chromatography or SDS-PAGE (see Methods).

None of the above models (Figure 6a-b, Supplemental Figure 4a) could explain the “lagged” triphasic association we observed (Figure 3). In contrast, one kinetic phenomenon that can produce an apparent association lag followed by seemingly rapid/spontaneous association involves sequential reactions where a downstream product has a “feedback” effect to catalyze an earlier step in the reaction. In the case of  $ER\alpha_{DBD-Ext}$  dsDNA binding, such a minimum kinetic model might resemble that shown in Figure 6c. In this model,  $ER\alpha_{DBD-Ext}$  initially binds dsDNA with high affinity to produce a stable complex. We note that it’s possible for the initial complex to have 2:1 instead of the 1:1 protein-ligand stoichiometry shown without drastically altering apparent kinetics. Additional  $ER\alpha_{DBD-Ext}$  monomers would be capable of inefficiently associating with the initial stable complex, but the initial complex could also slowly isomerize to an alternate complex state that better accommodates additional  $ER\alpha_{DBD-Ext}$  monomer binding. Critically, this complex isomerization would have to be susceptible to acceleration by already isomerized (and/or higher order stoichiometry) complex. Our simulations with the Figure 6c model suggest that it can recapitulate the kinetic trends observed for  $ER\alpha_{DBD-Ext}$  dsDNA binding (Supplemental Figure 6, see legend). Notably, accelerating the reverse isomerization (i.e., increasing  $k_{-2}$ ) in simulations made the data look more like the  $ER\alpha_{DBD-Ext}$  RNA binding kinetics (Supplemental Figure 7, see legend). In essence, our observations that  $ER\alpha_{DBD-Ext}$  DNA and RNA binding had similar dissociation rates and initial association rates despite differing  $K_d^{app}$ , Hill coefficients, and association curve shapes could be recapitulated by this model by changing a single rate constant value.

Overall, we present evidence that  $ER\alpha_{DBD-Ext}$  DNA and RNA binding cannot be adequately explained by traditional reaction schemes. Instead, we provide a ‘framework’ model that can generally recapitulate  $ER\alpha_{DBD-Ext}$  DNA and RNA kinetic trends. We note that our few simulations with this model are certainly not a perfect fit to the experimental data herein. Indeed, the “flexibility” of the model made it difficult to exhaustively fit our experimental data via iterative numerical integration and regression. Thus, it’s likely that our Figure 6c model does not completely capture the mechanism(s) of  $ER\alpha_{DBD-Ext}$  DNA and RNA binding. Rather, the model represents a starting point for insights into the ‘true’ mechanism. First, the model suggests that, despite the seemingly disparate  $ER\alpha_{DBD-Ext}$  DNA versus RNA association behaviors and affinities, they could share a reaction mechanism. Second, the hallmark “lagged” triphasic association we observed is critically dependent



on reaction mechanism “feedback” and complex isomerization in our simulations. While it’s not hard to imagine a TF like ER $\alpha$  that binds gene targets as a homodimer having more than one conformational state after target binding, the novel implication that some ER $\alpha$ <sub>DBD-Ext</sub> nucleic acid binding states can influence the stability of other complex states warrants further investigation. Such a mechanism could be especially relevant in situations like ER $\alpha$  nucleic acid condensates where multiple complexes are crowded together (Nair et al. 2019), since overall condensate stability and architecture could be impacted if certain complex states influence the stabilities of other complex states and their ability to accommodate additional protein monomers.

## Concluding remarks

In a cell, hundreds to thousands of transcription factors search for their unique target DNA sequences to perform critical regulation of gene expression. During this target search, the TFs not only coordinate with many protein binding partners, but they are also inundated with numerous other nucleic acids like the nontarget DNA in surrounding chromatin and free and nascent RNA. Consequently, TFs likely experience a variety of transitory nucleic acid binding events on the way to their target DNA sites. This is likely to be even more pronounced in the dense environment of biological condensates. Discerning the relevance of a TF’s numerous nucleic acid interactions to its biological function requires careful consideration of the prevalence, lifetimes, reaction mechanism(s), and inter-ligand influences of these varied binding events. Our biophysical studies herein estimate the timescales of association and dissociation events for the RNA and DNA interactions of two model TFs, and they also provide insight into the reaction mechanism(s) for these TF nucleic acid interactions. Thus, our findings represent a valuable ‘touchstone’ for considerations of how ER $\alpha$ , Sox2, and other related TFs have their target site searches influenced by competing nucleic acids like RNA.



## MATERIALS & METHODS

### *Protein Expression and Purification*

For recombinant Sox2<sub>HMG</sub>, a plasmid encoding Sox2<sub>HMG</sub> as an N-terminal octa-histidine and maltose binding protein (MBP) fusion with rhinovirus 3C protease cleavage site in a pET30b vector was generously provided by Desmond Hamilton (Batey lab, University of Colorado Boulder) (Hamilton et al. 2022). The plasmid was transformed into BL21(DE3) *E. coli* bacterial cells, inoculated into 20 mL media (LB broth with 100 µg/mL kanamycin), then the starter culture incubated overnight at 37°C/200rpm until  $A_{600} \approx 5.0$ . The culture was then induced with 0.5 mM Isopropyl β-D- thiogalactopyranoside (IPTG) and incubated at 37°C/200rpm for an additional 4 h. Following induction, the culture was pelleted by centrifugation (4,000x g/4°C/20min) and resuspended in 50 mL Amylose A buffer (20 mM TRIS pH 7.5 at 25°C, 200 mM NaCl, 1 mM EDTA) with 1 Pierce Protease Inhibitor Tablet (Thermo Scientific #A32965) and 50 mg lysozyme (Sigma Aldrich #L6876), then lysed with an Emulsiflex C3 homogenizer (Avestin) at 15,000-18,000 psi. Lysate was clarified by centrifugation (27,000x g/4°C/30min) and the supernatant collected. A 4°C AKTA Pure FPLC system (Cytiva) was prepared with a 10 mL amylose column (NEB #E8021S) and 2 mL/min flow rate, equilibrated with Amylose A buffer, supernatant applied, washed with Amylose A buffer, and eluted with Amylose B buffer (20 mM TRIS pH 7.5 at 25°C, 200 mM NaCl, 1 mM EDTA, 10 mM maltose). To the eluent was added 1.0 mg of Prescission Protease, and I was then loaded into 10 kDa-cutoff SnakeSkin Dialysis Tubing (Thermo Scientific #68100) and dialyzed overnight at 4°C in P-cell A buffer (50 mM TRIS pH 7.5 at 25°C, 1 mM EDTA) with 50 mM NaCl. The FPLC system was prepared with a 10 mL P11-phosphocellulose column (Whatman) and 2 mL/min flow rate, equilibrated with P-cell A buffer, dialyzed eluent applied, washed with P-cell A buffer, and eluted with a 100 mL 0-100% P-cell B buffer (50 mM TRIS pH 7.5 at 25°C, 1 mM EDTA, 1 M NaCl) gradient. Protein-containing (via  $A_{280}$ ) fractions were reconciled and concentrated with a 10 kDa-cutoff centrifugal filter unit. The FPLC system was prepared with a Superose 6 size-exclusion column and 0.25 mL/min flow rate, then equilibrated with Sizing Buffer (10 mM TRIS pH 7.5 at 25°C, 250 mM NaCl, 1mM EDTA), concentrated eluent applied, and followed with Sizing Buffer. Protein-containing (via  $A_{280}$ ) eluent fractions were reconciled for final product, then flash frozen in liquid nitrogen and stored at -80°C. SDS-PAGE indicated ≥95% purity, and

protomer concentrations were determined by spectroscopy with  $\epsilon_{280} = 13,980 \text{ M}^{-1}\text{cm}^{-1}$ . One liter of culture typically yielded ~4 mg of final protein.

For recombinant ER $\alpha_{\text{DBD}}$  and ER $\alpha_{\text{DBD-Ext}}$  (residues 180-262 for ER $\alpha_{\text{DBD}}$ , 180-280 for ER $\alpha_{\text{DBD-Ext}}$ ), proteins were expressed with a thrombin-cleavable N-terminal hexahistidine tag using a pET28a (EMD Biosciences) vector. Protein expression and purification methods were described previously (Steiner et al. 2022). Starting with a single transformed colony of BL21(DE3)pLysS *E. coli*, expression cultures were grown at 37°C (with 50  $\mu\text{g/mL}$  kanamycin and 50  $\mu\text{g/mL}$  chloramphenicol) using 2x YT rich media to an OD<sub>600</sub> of 0.8-1.0 and cold shocked on ice for 20 min. IPTG was added to a final concentration of 0.4 mM, along with 50  $\mu\text{M}$  ZnCl<sub>2</sub>, to induce protein expression and cultures were grown for 3 h at 37°C in a shaker. Cells were harvested by centrifugation (5,000 x g) and pellets stored at -20°C. Cell pellets were thawed and resuspended in 50 ml lysis buffer (20 mM Tris pH 7.5 at 25°C, 1 M NaCl, 10 mM imidazole pH 7.5, 5% glycerol) per 1 L of cells with one EDTA-free protease inhibitor cocktail tablet (Roche). Cells were lysed using a Misonix Sonicator 3000 (110 W for 2 min total ON-time, pulse: 15 s ON/45 s OFF, ½ inch tip) and the lysate cleared by centrifugation (15,000x g, 30 min). Cleared lysate was loaded onto lysis buffer-equilibrated Ni-NTA resin (GoldBio, 5 mL resin per 50 mL lysate) and rocked gently for 1 h at 4°C. The bead slurry was loaded onto a gravity flow column and washed twice with increasing concentrations of imidazole in lysis buffer (wash 1: 20 mM imidazole, wash 2: 30 mM imidazole), then eluted with 300 mM imidazole in lysis buffer. Bovine  $\alpha$ -Thrombin (Haematologic Technologies Incorporated) was added (10 U/mg protein) to the eluate to remove the hexahistidine tag. The eluate solution was transferred to 6-8 kDa MWCO dialysis tubing (Spectra/Por – Spectrum Labs) and dialyzed overnight at 4°C in 4 L of column buffer (20 mM Tris pH 7.5 at 25°C, 100 mM NaCl, 5% glycerol, 1 mM DTT). Dialyzed eluate was filtered to 0.2  $\mu\text{m}$  and concentrated using 5 kDa MWCO spin concentrators (Vivaspin Turbo). The sample was again filtered to 0.2  $\mu\text{m}$  and loaded onto a HiLoad 16/600 Superdex 75 column (GE Healthcare) and eluted as a monomer. Pooled fractions containing recombinant ER $\alpha$  were assessed for purity, aliquoted, flash-frozen, and stored at -70°C. One liter of culture typically yielded 2 mg of purified protein as measured by absorption ( $\epsilon_{280} = 14,440 \text{ M}^{-1}\text{cm}^{-1}$ ). All experiments used ER $\alpha_{\text{DBD-Ext}}$  protein, unless otherwise indicated.

### *Preparation of Oligonucleotides*

All oligonucleotides except the XBP1 hRNA were ordered from IDT (Coralville, IA), and their sequences in IDT syntax are provided (Supplemental Table 1). For Sox2<sub>HMG</sub> dsDNA ligands, complementary oligonucleotides ordered from IDT were mixed at 100  $\mu$ M each in annealing buffer (50 mM TRIS pH 7.5 at 25°C, 200 mM NaCl) and subjected to a thermocycler program (95°C for 10 min, 95→4°C at 0.5 °C/min, hold at 4°C) for annealing. For ER $\alpha$  dsDNA, the complementary strands were combined at 1  $\mu$ M labeled and 5  $\mu$ M unlabeled (ligand prep) or 100  $\mu$ M each unlabeled (competitor prep) in annealing buffer (20 mM Tris pH 7.5 at 25°C, 50 mM NaCl), then annealed by bench-top slow cooling (heated 95°C for 1 min, then cooled to room temperature for 3 h). Complete annealing for all oligonucleotides was confirmed via native-PAGE. Concentrations of all ligands were confirmed spectroscopically using manufacturer-provided extinction coefficients.

According to prior methodology (Steiner et al. 2022), XBP1 hRNA was prepared by *in vitro* transcription (IVT) with T7 RNA polymerase, using dsDNA templates containing a T7 polymerase promoter sequence, which were created via PCR with IDT-synthesized oligonucleotides. Briefly, full PCR amplification was confirmed on 2% agarose gel, and subsequent IVTs were performed for 3 h at 37°. Successful transcription was confirmed via 10-18% denaturing PAGE. After IVT, RNAs were precipitated in 1/10<sup>th</sup> volume 3M sodium acetate and 2.5 volumes ice cold ethanol overnight. The following day ethanol precipitated RNA was pelleted and dried, then resuspended and purified by denaturing urea-polyacrylamide gel electrophoresis followed by buffer exchange and concentration in a 5 kDa MWCO spin concentrator (Vivaspin Turbo). Purified RNA oligonucleotides were 3'-end labeled with pCp-AF488 (Alexa Fluor). 200 pmol of RNA and 2400 pmol fluorophore were combined in labeling buffer (1mM ATP, 10% DMSO, 50% PEG, 40 U T4 ligase and 1x T4 ligase buffer) overnight at 16° C. The labeled RNA was purified using RNA Clean & Concentrator kit (Zymo #4060), passed through a G-25 spin column, and stored at -20°C. Concentration was determined by A<sub>260</sub> and total RNA yield was typically 10–50% with ~70% labeling efficiency. Purity of the final sample was assessed by 10–15% denaturing PAGE and imaged by fluorescence (Ex = blue wavelength filter, Em = green wavelength filter). RNA samples were prepared for binding assays by fast refolding at 1  $\mu$ M by snap-cooling (95°C for 1 min, ice for >5 min).

## FP-based $K_d^{app}$ and Association Rate Determination

Pre-reaction mix was prepared with 5 nM ligand in ER $\alpha$  binding buffer (20 mM TRIS pH 7.5 at 25°C, 100 mM NaCl, 5% glycerol, 0.01% IGEPAL) or Sox2 binding buffer (10 mM TRIS pH 7.5 at 25°C, 135 mM KCl, 15 mM NaCl, 0.1 mg/mL nonacetylated BSA, 4% Ficoll, 0.05% NP-40, 1 mM DTT), then dispensed in 36  $\mu$ L volumes into the wells of a 384-well black microplate (Corning #3575). A range of protein concentrations was prepared at 10x the final reaction concentrations via serial dilution in respective binding buffer. Protein and microplates were then thermally equilibrated at 4°C for 30 min. After thermal equilibration, reactions were initiated by addition of 4  $\mu$ L of the respective 10x protein concentration to the corresponding microplate well, then incubated for  $\geq$ 60 min at 4°C. Fluorescence anisotropy readings were taken over the course of incubation immediately after protein addition (in  $<10$  s intervals) with a TECAN Spark microplate reader (Ex =  $481 \pm 20$  nm, Em =  $526 \pm 20$  nm). Each experiment had 1 reaction (well) per protein concentration, and 1-3 independent experiments were performed per protein-polynucleotide interaction.

For equilibrium dissociation constant calculations, the last 10 min of data points from the anisotropy versus time data of each reaction were averaged to give equilibrium values, then equilibrium anisotropy values versus protein concentration data were regressed with Eq. 1.2 (Sox2<sub>HMG</sub>-CBS and Sox2<sub>HMG</sub>-rG4) or 1.1 (all other interactions) to determine  $K_d^{app}$  and  $n$ . For the displayed Sox2<sub>HMG</sub> binding curves (Supplemental Figure 1b), traces were normalized to the respective maximum and minimum signals determined by regression. Mean and error (50% range) are reported in Table 1.

For initial association rate constant calculations, anisotropy versus time data for each protein concentration were fit with a smoothing spline and the initial slope of the regression was divided by the dynamic range in anisotropy for the regression to calculate apparent association rates. Apparent association rate versus protein concentration data were pruned to include only the initial linear phases, and to exclude the higher protein concentrations with incomplete curves. Then, pruned data were regressed with 0-intercept linear regression to determine  $k_{on}^{app}$ . Mean and error (50% range) are reported in Table 1.

For the monophasic or biphasic association fits in Supplemental Figure 3, anisotropy versus time data for each protein concentration were pruned to only include the initial data points shown in respective graphs in

Supplemental Figure 3, then pruned data were regressed with Eq. 2.2 ( $ER\alpha_{DBD-Ext}$ -XBP1, Sox2<sub>HMG</sub>-rG4) or 2.1 (all others). Analyses were performed in R v4.3.1.

### *FP-based Competitive Dissociation (FPCD)*

Pre-reaction mix was prepared with 5 nM ligand in  $ER\alpha$  (20 mM TRIS pH 7.5 at 25°C, 100 mM NaCl, 5% glycerol, 0.01% IGEPAL) or Sox2 (10 mM TRIS pH 7.5 at 25°C, 135 mM KCl, 15 mM NaCl, 0.1 mg/mL nonacetylated BSA, 4% Ficoll, 0.05% NP-40, 1 mM DTT) binding buffer, then dispensed in 32  $\mu$ L volumes into the wells of a 384-well black microplate (Corning #3575). Protein was prepared at 10x the reaction concentrations of 30 nM ( $ER\alpha_{DBD-Ext}$ -dsDNA Figure 4), 100 nM ( $ER\alpha_{DBD-Ext}$ -dsDNA Figure 1), or 500 nM ( $ER\alpha_{DBD-Ext}$ -RNA and Sox2<sub>HMG</sub>). Competitor was prepared at 10x the reaction concentration of 10  $\mu$ M (i.e., 100  $\mu$ M). Microplates (ligand), protein, and competitor were then thermally equilibrated at 4°C for 30-60 min. After thermal equilibration, 4  $\mu$ L of 10x protein or binding buffer (baseline control) was added to the microplate wells, then the reactions incubated at 4°C for 1 h or indicated (Figs. 4-5) shorter times. After protein-ligand incubation, 4  $\mu$ L of 10x competitor or binding buffer (max signal control) was added to the microplate wells, then incubated for  $\geq 60$  min at 4°C. Fluorescence anisotropy readings were taken over the course of incubation immediately after protein addition (in  $< 10$  s intervals) with a TECAN Spark microplate reader ( $Ex = 481 \pm 20$  nm,  $Em = 526 \pm 20$  nm). Figure 1a provides helpful clarification of the methodology. Two internal controls were employed: control-1, which used buffer controls for protein addition in step-1 (Figure 1a) and competitor addition in step-2 (Figure 1a), and control-2, which used a buffer control for competitor addition in step-2 (Figure 1a). Each experiment had 2-3 reaction replicates per condition/control, and 2 independent experiments were performed per protein-polynucleotide interaction.

Anisotropy versus time data for the experimental reaction was normalized to the data for the two internal controls to give fraction bound versus time. For reactions with shorter ( $< 1$  h) protein-ligand incubation times, an additional internal max signal control was always included where protein-ligand incubation was  $\geq 1$  hr, and this was used for normalization to calculate fraction bound. Fraction bound versus time data were regressed with Eq. 3 to determine  $k_{fast}$ ,  $k_{slow}$ , and  $\beta_{fast}$ . For regression, the initial fraction bound ( $A_{max}$ ) was constrained to the initial value calculated for the internal control with identical protein-ligand incubation that

omitted competitor addition. In cases where dissociation was approximately slow monophasic, the regression was additionally constrained such that  $k_{\text{fast}} = 0$  and  $\beta_{\text{fast}} = 0$ . Analyses were performed in R v4.3.1. Across experiments, percent contributions of the bi-exponential typically varied <15%.

### *FP-based Jump Dilution (FPJD)*

Pre-reaction mix was prepared with 50 nM ligand in binding buffer (20 mM TRIS pH 7.5 at 25°C, 100 mM NaCl, 5% glycerol, 0.01% IGEPAL), and with (experimental reaction) or without (baseline control) 50 nM protein. Wells of a 384-well black microplate (Corning #3575) were filled with 79  $\mu\text{L}$  binding buffer (experimental reaction and baseline control) or 50 nM protein (max signal control), then the microplates and pre-reaction mixes were incubated at 4°C for 1 hr. After incubation, 1  $\mu\text{L}$  of protein-ligand mix was diluted in the buffer-only (experimental reaction) or 50 nM protein (max signal control) microplate wells, and ligand-only mix was diluted in buffer-only microplate wells (baseline control), then the reactions incubated at 4°C for 1 h. Fluorescence anisotropy readings were taken over the course of incubation immediately after dilutions (in <10 s intervals) with a TECAN Spark microplate reader ( $\text{Ex} = 481 \pm 20 \text{ nm}$ ,  $\text{Em} = 526 \pm 20 \text{ nm}$ ). Two controls are employed: control-1, which uses a buffer control for protein addition, and control-2, which uses an equimolar protein control for buffer dilution. Each experiment had 1 reaction (well) per condition/control, and 3 independent experiments were performed.

Anisotropy versus time data for the experimental reaction were normalized to the data for the two internal controls to give fraction bound versus time. Fraction bound versus time data were regressed with Eq. 3 to determine  $k_{\text{fast}}$ ,  $k_{\text{slow}}$ , and  $\beta_{\text{fast}}$ . For regression, the initial fraction bound ( $A_{\text{max}}$ ) was constrained to a value of 1. Analyses were performed in R v4.3.1. Across experiments, percent contributions of the bi-exponential typically varied <15%.

### *Surface Plasmon Resonance*

Streptavidin-coated S-series chips were purchased commercially (Xantec #SCBS-SAD200M) and docked into a Biacore T200 SPR instrument (Cytiva). Before first-time use, all four chip flow cells (FCs) were

washed (25  $\mu$ L/min for 1 min) 5 times with Activation Buffer (50 mM NaOH, 1 M NaCl) to remove any unbound streptavidin, then washed (25  $\mu$ L/min for 10 min) with Running Buffer (20 mM TRIS pH 7.5 at 25°C, 100 mM NaCl, 5% glycerol, 0.01% IGEPAL) to ensure surface stability. Ligand immobilization of biotin-labeled ERE (FC-2) or  $\Delta$ ERE (FC-4) dsDNA was performed by flowing 20 nM ligand solutions over the respective FC at 1  $\mu$ L/min until a  $\Delta$ RU of 300 was achieved, then priming the system with Running Buffer three times. FCs 1 and 3 were used as background controls for FC 2 and 4 experiments, respectively.

For kinetic experiments, the indicated (Supplemental Figure 4)  $ER\alpha_{DBD-Ext}$  concentrations were flowed sequentially over the control and ligand FCs at 70  $\mu$ L/min for 5 min to monitor association, followed by a 30-60 min wash phase (70  $\mu$ L/min) with Running Buffer to monitor dissociation.  $ER\alpha_{DBD-Ext}$  concentrations were tested in increasing order, and the flow cells were washed (70  $\mu$ L/min for 1 min) between  $ER\alpha_{DBD-Ext}$  concentrations 1 time with Regeneration Buffer (1 M NaCl) and 3 times with Running Buffer. Control FC signal was subtracted from experimental FC signal to generate adjusted signal versus time data, then adjusted data was exported from the instrument.

Adjusted data for each protein concentration were subjected to baseline subtraction to generate  $\Delta$ RU versus time data. For initial association rate constant calculations,  $\Delta$ RU versus time data were pruned to include only the association phases, then time points adjusted to start at zero time. For each protein concentration, pruned association data were used to calculate the change in  $\Delta$ RU over the first 3 s of association, then this  $\Delta\Delta$ RU was normalized to the dynamic range in  $\Delta$ RU for each protein concentration to calculate apparent association rates. Apparent association rate versus protein concentration data were regressed with 0-intercept linear regression to determine  $k_{on}^{app}$ . For dissociation rate calculations,  $\Delta$ RU versus time data were pruned to include only the dissociation phases, then time points adjusted to start at zero time. Pruned dissociation data for each protein concentration were regressed with Eq. 3 to determine  $k_{fast}$ ,  $k_{slow}$ , and  $\beta_{fast}$ . For regression, the initial signal ( $A_{max}$ ) was constrained to the final  $\Delta$ RU observed at the end of the preceding SPR association phase. In cases where dissociation was approximately slow monophasic, regression was additionally constrained such that  $k_{fast} = 0$  and  $\beta_{fast} = 0$ . Analyses were performed in R v4.3.1. We note that our association data have prolonged linear phases (Figure 5a), suggesting that, despite our low ligand seeding densities and high flow rates during analyte injection, mass transfer effects could be deflating the apparent association rates to some unknown degree.



## ERa Reaction Scheme Simulations

Reactions (Supplemental Figs. 6-7) were simulated and analyzed in R v4.3.1 using `deSolve::ode` (package::function) and the `lsoda` integrator (Soetaert et al. 2010). Numerical integration of the Eq. 4.1-5 system of differential equations was performed with a given integration time-step ( $\Delta t$ ) in two phases with fixed rate constant values from Figure 6c. In phase 1 (the association phase), for initial conditions the total protein ( $[E_T]$ ) and ligand ( $[L_T]$ ) were included as free protein and ligand and all other reactants concentrations were set to zero, then the reactions were simulated for a given association time ( $t_{on}$ ). In phase 2, the initial conditions were set to the final reactant concentrations from phase 1 divided by a given dilution factor ( $d_f$ ), then the reactions were simulated for a given dissociation time ( $t_{off}$ ). Next, relative predicted anisotropy over time was calculated from reactant concentrations over time via Eq. 4.6. Equilibrium values were taken from endpoints in phase 1 simulations. Supplemental Figure 6 simulations used  $\Delta t = 25$  ms,  $t_{on} = 1.5$  h,  $t_{off} = 1$  h,  $d_f = 10^6$ ,  $[E_T] = 2^{12.0}$   $\mu$ M,  $[L_T] = 5$  nM,  $k_1 = 10^6$   $M^{-1}s^{-1}$ ,  $k_{-1} = 1.6 \times 10^{-3}$   $s^{-1}$ ,  $k_2 = 10^{-5}$   $M^{-1}s^{-1}$ ,  $k_{-2} = 3.2 \times 10^{-3}$   $s^{-1}$ ,  $k_3 = 10^9$   $M^{-1}s^{-1}$ ,  $k_{-3} = 2 \times 10^{-2}$   $s^{-1}$ ,  $k_{3b} = 10^4$   $M^{-1}s^{-1}$ ,  $k_\alpha = 2 \times 10^7$   $M^{-1}s^{-1}$ ,  $k_\beta = 2 \times 10^6$   $M^{-1}s^{-1}$ . Supplemental Figure 7 simulations were identical, except  $k_{-2} = 10$   $s^{-1}$ .

## Equations

For Eq. 1.1,  $A$  is the signal (anisotropy,  $\Delta RU$ , fraction bound, etc.),  $A_{min}$  is the minimum signal,  $A_{max}$  is the maximum signal,  $E_T$  is the total protein concentration,  $K_d$  is the (apparent) equilibrium dissociation constant, and  $n$  is the Hill coefficient. For Eq. 1.2,  $L_T$  is the total ligand concentration,  $K_{d1}$  is the equilibrium dissociation constant of the first binding state,  $K_{d2}$  is the equilibrium dissociation constant of the second binding state,  $\alpha$  is the proportion of the signal dynamic range attributable to the first binding state, and remaining parameters are as defined for Eq. 1.1.

$$(Eq. 1.1) \quad A = A_{min} + (A_{max} - A_{min}) \frac{E_T^n}{E_T^n + K_d^n}$$



$$(Eq. 1.2) \quad A = A_{\min} + (A_{\max} - A_{\min}) \left( \alpha \frac{E_T + L_T + K_{d1} - \sqrt{(E_T + L_T + K_{d1})^2 - 4 E_T L_T}}{2 L_T} + (1 - \alpha) \frac{E_T}{E_T + K_{d2}} \right)$$

For Eq. 2.1,  $A_t$  is the signal (anisotropy,  $\Delta RU$ , fraction bound, etc.) at a given time ( $t$ ),  $A_{\min}$  is the minimum signal,  $A_{\max}$  is the maximum signal,  $k_{on}$  is the rate constant for the association curve, and  $t$  is time. For Eq. 2.2,  $k_{fast}$  is the rate constant for the fast phase of the association curve,  $k_{slow}$  is the rate constant for the slow phase of the association curve,  $\beta_{fast}$  is the proportion of the signal dynamic range attributable to the fast phase of the association curve, and remaining parameters are as defined for Eq. 2.1.

$$(Eq. 2.1) \quad A_t = A_{\min} + (A_{\max} - A_{\min})(1 - e^{-k_{on} t})$$

$$(Eq. 2.2) \quad A_t = A_{\min} + (A_{\max} - A_{\min})(1 - \beta_{fast} e^{-k_{fast} t} - (1 - \beta_{fast}) e^{-k_{slow} t})$$

For Eq. 3,  $A_t$  is the signal (anisotropy,  $\Delta RU$ , fraction bound, etc.) at a given time ( $t$ ),  $A_{\min}$  is the minimum signal,  $A_{\max}$  is the maximum signal,  $t$  is time,  $k_{fast}$  is the rate constant for the fast phase of the dissociation curve,  $k_{slow}$  is the rate constant for the slow phase of the dissociation curve, and  $\beta_{fast}$  is the proportion of the signal dynamic range attributable to the fast phase of the dissociation curve.

$$(Eq. 3) \quad A_t = A_{\min} + (A_{\max} - A_{\min})(\beta_{fast} e^{-k_{fast} t} + (1 - \beta_{fast}) e^{-k_{slow} t})$$

For Eq. 4, rate constants are defined in Figure 6c,  $E$  is protein,  $L$  is ligand, conjugations of these reactants are complexes, equations give rates of change for indicated reactants as a function of time ( $t$ ), bracketed terms indicate concentrations, and  $A_{rel}$  is relative predicted anisotropy.

$$(Eq. 4.1) \quad [E]_t' = k_{-1} [EL]_t + k_{-3} [E_2L]_t - [E]_t (k_1 [L]_t + k_3 [EL^*]_t)$$

$$(Eq. 4.2) \quad [L]_t' = k_{-1} [EL]_t - k_1 [E]_t [L]_t$$

$$(Eq. 4.3) \quad [EL]_t' = k_1 [E]_t [L]_t + k_{-2} [EL^*]_t - [EL]_t (k_{-1} + k_2 + k_{\alpha} [EL^*]_t + k_{\beta} [E_2L]_t + k_{3b} [E]_t)$$

$$(Eq. 4.4) \quad [EL^*]_t' = (k_2 + k_{\alpha} [EL^*]_t + k_{\beta} [E_2L]_t) [EL]_t + k_{-3} [E_2L]_t - [EL^*]_t (k_{-2} + k_3 [E]_t)$$

$$(Eq. 4.5) \quad [E_2L]_t' = k_3 [E]_t [EL^*]_t + k_{3b} [E]_t [EL]_t - k_{-3} [E_2L]_t$$

$$(Eq. 4.6) \quad A_{rel} = \frac{[E_2L] + \frac{1}{2}([EL^*] + [EL])}{[E_2L] + [EL^*] + [EL] + [L]}$$

### *Diagram, Reaction Scheme, and Figure Generation*

All diagrams and reaction schemes were prepared on BioRender.com, tables were prepared with Word (Microsoft), graphs were prepared with R v4.3.1, and figures were assembled in PowerPoint (Microsoft).

### *Data, Materials, and Software Availability*

The bacterial expression plasmid for Sox2<sub>HMG</sub> is available from the lab of Robert Batey (University of Colorado Boulder, Department of Biochemistry). Our bacterial expression plasmids for ERα<sub>DBD(-Ext)</sub> are available upon request (contact D.S.W.). Our R script for simulating the Figure 6c reactions is available on GitHub ([github.com/whemphil/ER-Sox2\\_Manuscript](https://github.com/whemphil/ER-Sox2_Manuscript)).

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## **COMPETING INTERESTS STATEMENT**

T.R.C declares consulting status for Storm Therapeutics, Eikon Therapeutics, and SomaLogic. The other authors have no competing interests to declare.

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Protein	Ligand	$K_d^{app}$ (nM)	n (Hill)	$k_{on}^{app}$ ( $M^{-1}s^{-1}$ )	$k_{off}^{app_{fast}}$ ( $\times 10^{-2} s^{-1}$ )	$k_{off}^{app_{slow}}$ ( $\times 10^{-4} s^{-1}$ )
<i>ER<math>\alpha</math><sub>DBD-Ext</sub></i>	ERE dsDNA	11 $\pm$ 2.1	2.1 $\pm$ 0.30	5.8 $\pm$ 3.1 $\times 10^4$	4.7 (71%)	7.3 (29%)
	$\Delta$ ERE dsDNA	<sup>¥</sup> 2.8 $\pm$ 0.6	<sup>¥</sup> 1.5 $\pm$ 0.22	8.7 $\pm$ 4.6 $\times 10^4$	8.9 $\pm$ 4.4 (60%)	5.9 $\pm$ 1.1 (40%)
	XBP1 hRNA	370	0.91	14 $\times 10^4$	5.6 $\pm$ 2.7 (64%)	6.6 $\pm$ 1.2 (36%)
<i>Sox2<sub>HMG</sub></i>	CBS dsDNA	<sup>†</sup> $\leq$ 1 (54%), 420 $\pm$ 120	<sup>†</sup> n/a	0.75 $\times 10^6$	5.0 $\pm$ 2.3 (67%)	13 $\pm$ 2.1 (33%)
	G4 RNA	<sup>†</sup> $\leq$ 1 (44%), 110 $\pm$ 32	<sup>†</sup> n/a	1.1 $\times 10^6$	5.4 (73%)	9.6 (27%)
	NBS dsDNA	29 $\pm$ 9.1	0.57 $\pm$ 0.06	n.d.	n.d.	n.d.
	hRNA	53 $\pm$ 16	1.1 $\pm$ 0.21	n.d.	n.d.	n.d.
	poly-A RNA	40 $\pm$ 13	0.68 $\pm$ 0.11	n.d.	n.d.	n.d.

n/a = not applicable

n.d. = not determined

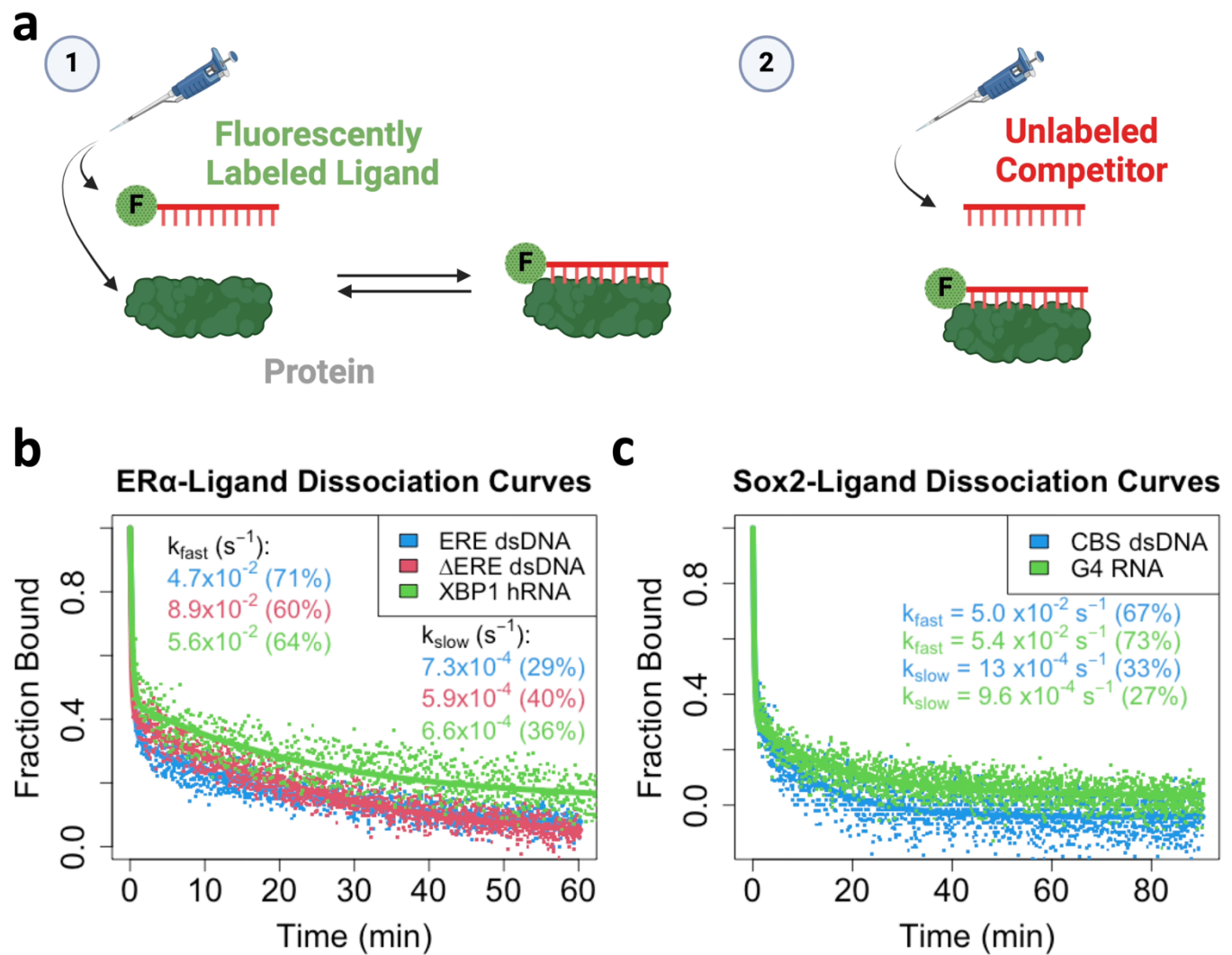
<sup>¥</sup>  $K_d^{app}$  is close to ligand concentration; it's possible that  $K_d < K_d^{app}$  and n (Hill coefficient) is artificially inflated

<sup>†</sup> Binding curves were fit with a two-transition equation, not Hill equation

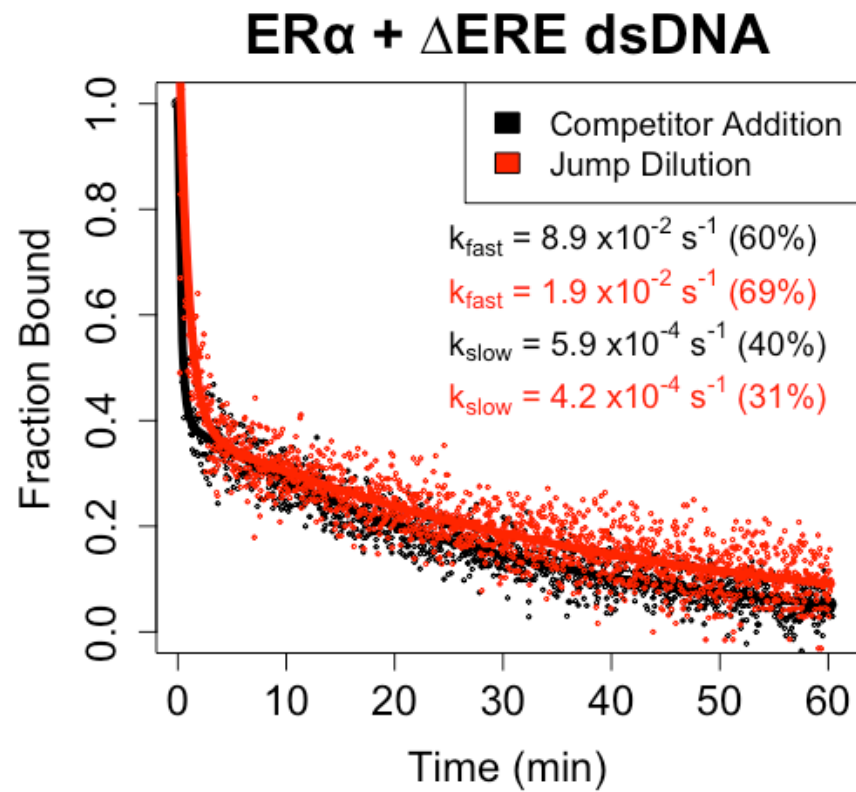


**Table 1: Kinetic Constant Values from FP-based ERα<sub>DBD-Ext</sub> and Sox2<sub>HMG</sub> Binding Experiments.** Table

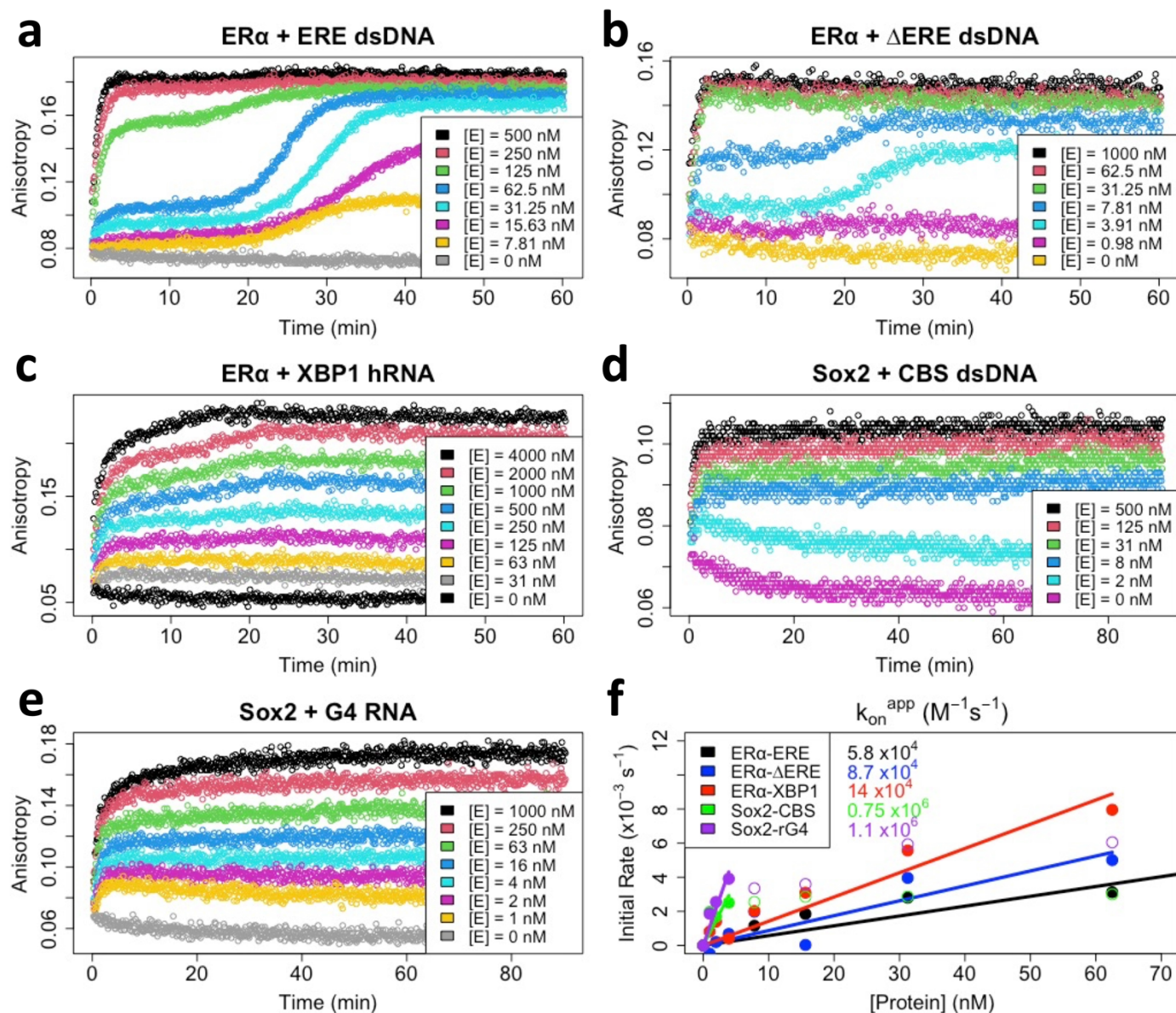
includes apparent equilibrium dissociation constants ( $K_d^{app}$ ) and Hill coefficients ( $n$ ) from Supplemental Figure 1 (fit with Eq. 1.1-2), apparent initial association rate constants ( $k_{on}^{app}$ ) from Figure 3, and apparent dissociation rate constants ( $k_{off}^{app}$ ) from Figure 1 (fit with Eq. 3). Values are mean  $\pm$  1/2 range (across 2-3 independent experiments). Values without error are based on a single experiment. For  $K_d^{app}$ , values in parentheses are the percent signal contributions of the first transitions in two-transition binding regression, and for  $k_{off}^{app}$ , values in parentheses are the percent contributions of the fast or slow components in bi-exponential regression. Percentages are the averages across 2-3 independent experiments, or values from a single experiment if the associated rate constants have no error.



**Figure 1: ER $\alpha$ <sub>DBD-Ext</sub> and Sox2<sub>HMG</sub> Exhibit Multiphasic Ligand Dissociation. [a]** *Graphical summary of Fluorescence Polarization-based Competitive Dissociation (FPCD) experiments.* (1) Fluorescently labeled polynucleotide is mixed with protein and incubated at 4°C for a variable amount of time, then (2) an excess of unlabeled polynucleotide (i.e., competitor) is added to the protein-ligand reaction and polarization is monitored over time (at 4°C) to observe protein-ligand dissociation kinetics. **[b-c]** *Dissociation curves from FPCD experiments.* FPCD experiments (panel a) were performed using 5 nM ligand, 10  $\mu$ M competitor, and 100-500 nM protein; protein-nucleic acid binding reactions were incubated long enough to reach equilibrium before competitor addition. Anisotropy traces were normalized to the internal controls to give 'Fraction Bound' over time, then normalized data fit with bi-exponential regression (Eq. 3) to determine rate constants. Dots are data points and solid lines are regression fits from a single experiment for each ligand. Rate constants and (in parenthesis) the percent contributions of fast versus slow components to the bi-exponential regression are reported with error in Table 1.

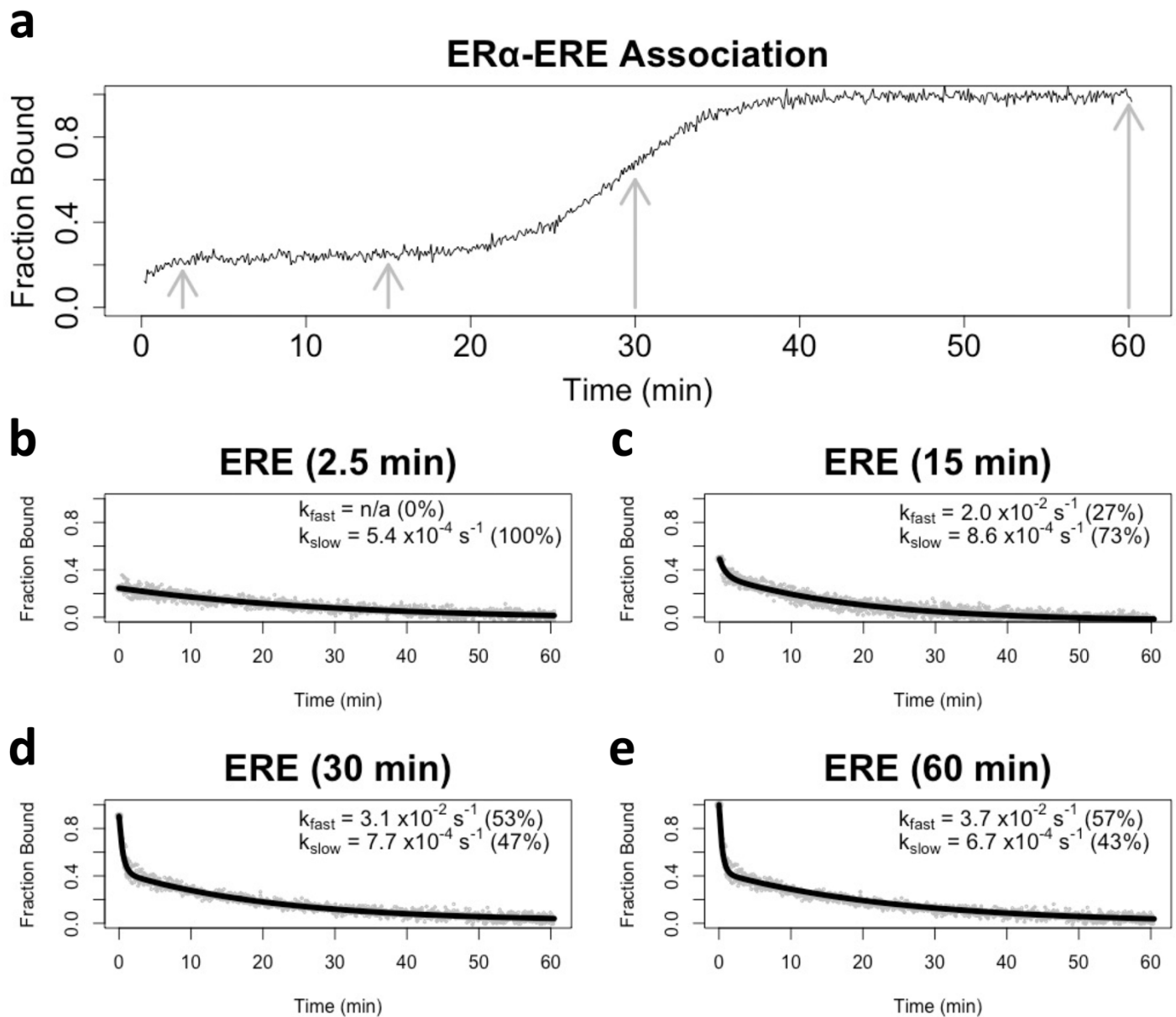


**Figure 2: ER $\alpha$ <sub>DBD-Ext</sub> Multiphasic Ligand Dissociation is Independent of Competitor.** *Dissociation curves from FPCD versus FPJD experiments.* FPJD experiments were performed for the ER $\alpha$ <sub>DBD-Ext</sub>- $\Delta$ ERE interaction using 50 nM ligand and 50 nM protein (pre-dilution), with protein-ligand reactions being incubated to equilibrium before dilution. The protein-polynucleotide reaction was then diluted ~80-fold in buffer (at 4°C) and polarization was monitored over time post-dilution (at 4°C) to quantify protein-ligand dissociation kinetics. FPCD experiments were performed as described for Figure 1. Anisotropy traces were normalized to the controls to give ‘Fraction Bound’ over time, then normalized data were fit with bi-exponential regression to determine rate constants. Dots are data points and solid lines are regression fits (Eq. 3) from single experiments. Rate constants are the average values from all independent experiments (2 for FPCD, 3 for FPJD), with percent contributions of fast and slow components to the bi-exponential curve in parentheses.



**Figure 3: ER $\alpha$ <sub>DBD-Ext</sub>, but not Sox2<sub>HMG</sub>, Exhibits Multiphasic Target DNA Association. [a-e] *FP-based association curves.* Protein-ligand reactions were prepared after thermal equilibration (4°C), and anisotropy monitored over time immediately after protein addition to quantify association kinetics. Ligand concentrations were 5 nM, and protein concentrations ([E]) are indicated. Data are from single representative experiments (of 1-3 per protein-ligand interaction). The first 10-45 min of association data were subjected to regression with an equation for monophasic (Eq. 2.1) or biphasic (Eq. 2.2) association; the regression fits are shown in Supplemental Figure 3. [f] *Association rate constant analysis.* Apparent initial association rates were determined with smoothing spline regression (see Methods) and are plotted as a function of protein concentration. Each interaction has an initial linear component, followed by a plateau in apparent association rate at higher protein concentrations, which corresponds to the incomplete association curves seen at higher protein concentrations due to methodological limitations. Apparent association rates from these linear stages were used for 0-intercept linear regression to calculate apparent association rate constants ( $k_{on}^{app}$ ). Filled circles are data points used for linear regression, open circles are data points excluded from linear regression, and solid lines are linear regression fits. Rate constants are reported with error in Table 1.**

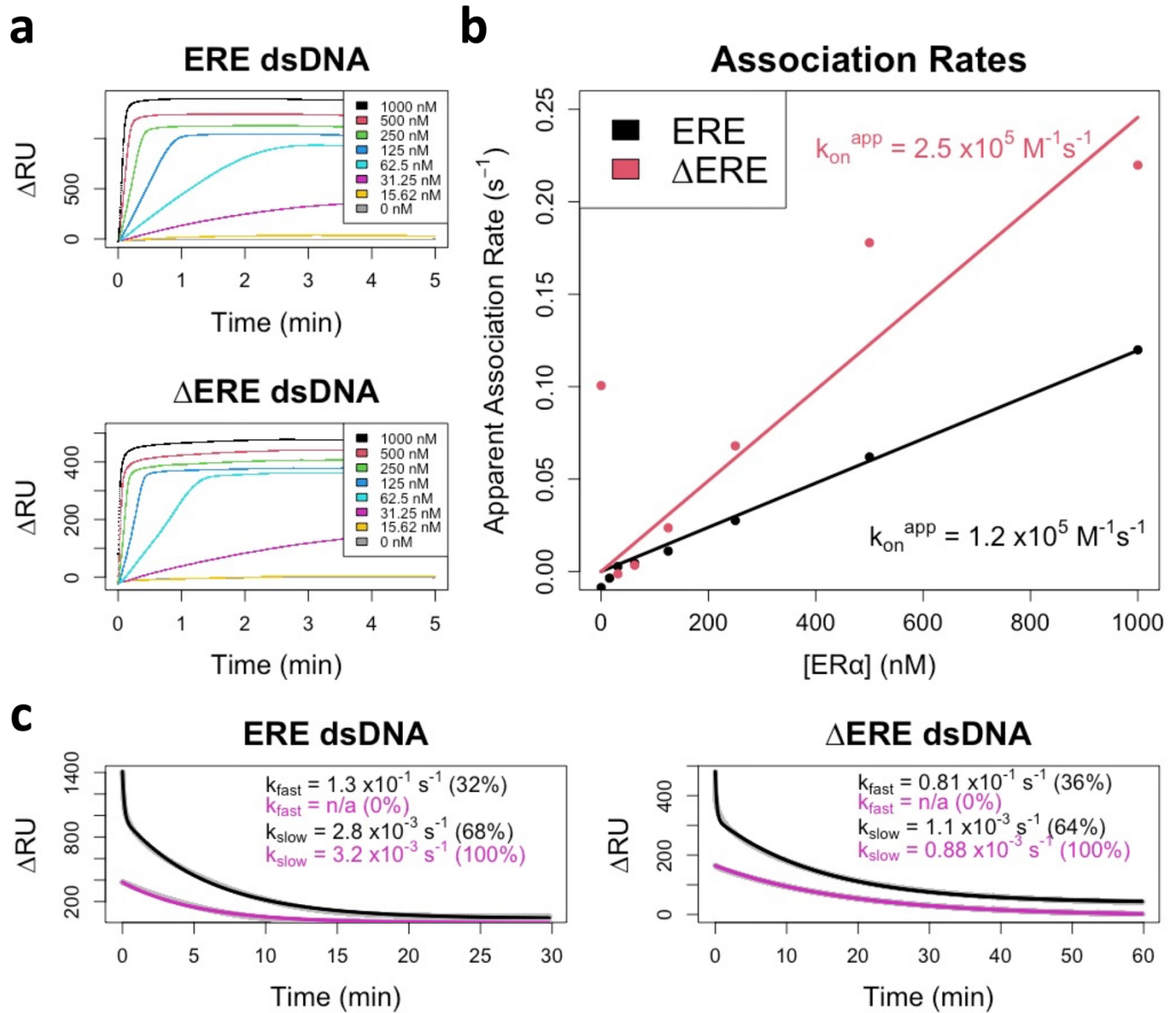






#### Figure 4: The More Stable Complex State Forms First During Multiphasic ER $\alpha$ <sub>DBD-Ext</sub> Ligand

**Association. [a]** *Normalized ER $\alpha$ <sub>DBD-Ext</sub> association curve.* Normalized association curve of 30 nM ER $\alpha$ <sub>DBD-Ext</sub> and 2 nM ERE dsDNA, taken from Figure 3a. Grey arrows correspond to incubation times prior to competitor addition in panel b-e experiments. **[b-e]** *Dissociation curves from FPCD experiments where competitor was added after variable protein-ligand incubation times.* FPCD experiments (see Figure 1a) were performed for the ER $\alpha$ <sub>DBD-Ext</sub>-ERE interaction using 2 nM ligand, 10  $\mu$ M competitor, and 30 nM protein; protein-ligand reactions were incubated for 2.5 min (b), 15 min (c), 30 min (d), or 60 min (e) before competitor addition. Anisotropy traces were normalized to the internal controls to give 'Fraction Bound' over time, then normalized data were fit with bi-exponential regression (Eq. 3) to determine rate constants. Grey dots are data points and solid black lines are regression fits from single experiments. Percent contributions of fast versus slow components to the bi-exponential curve are in parentheses.



## Figure 5: Surface Plasmon Resonance Confirms Multiphasic ER $\alpha$ <sub>DBD-Ext</sub>-dsDNA Binding Kinetics. [a]

*ER $\alpha$ <sub>DBD-Ext</sub> ligand association curves.* SPR association phase curves, taken from Supplemental Figure 5.

Legends indicate the concentration of ER $\alpha$ <sub>DBD-Ext</sub> used during protein injection. Lines are data, not regression

fits. [b] *Association rate analysis.* Association curves from panel a had their initial slopes normalized to their

signal dynamic range to calculate their apparent association rates (see Methods). Plots of apparent initial

association rates versus protein concentrations were fit with 0-intercept linear regression to calculate apparent

initial association rate constants ( $k_{on}^{app}$ ). Dots are data and solid lines are linear regression fits. [c] *Dissociation*

*rate analysis.* SPR dissociation phase curves, taken from Supplemental Figure 3. Two key protein

concentrations (1  $\mu$ M and 30 nM) are shown, corresponding to the color scheme in panel a. Data were fit with

bi-exponential regression (Eq. 3) to determine rate constants. Grey dots are data and solid black/purple lines

are regression fits; data points are mostly obscured by regression lines. Percent contributions of fast versus

slow components to the bi-exponential curve are in parentheses. All SPR data in this figure are from a single

experiment per ligand.

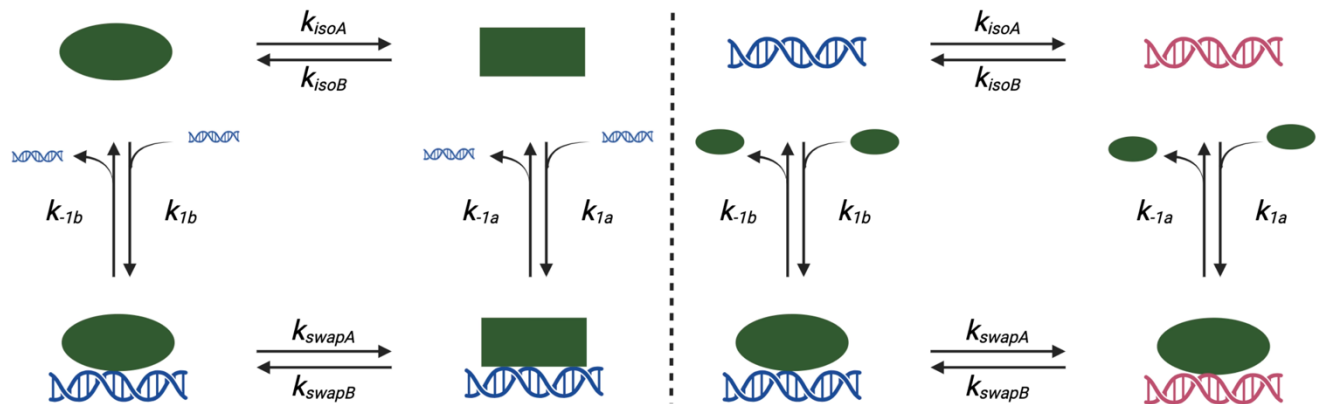
**a**

### Sequential Protein Binding Model



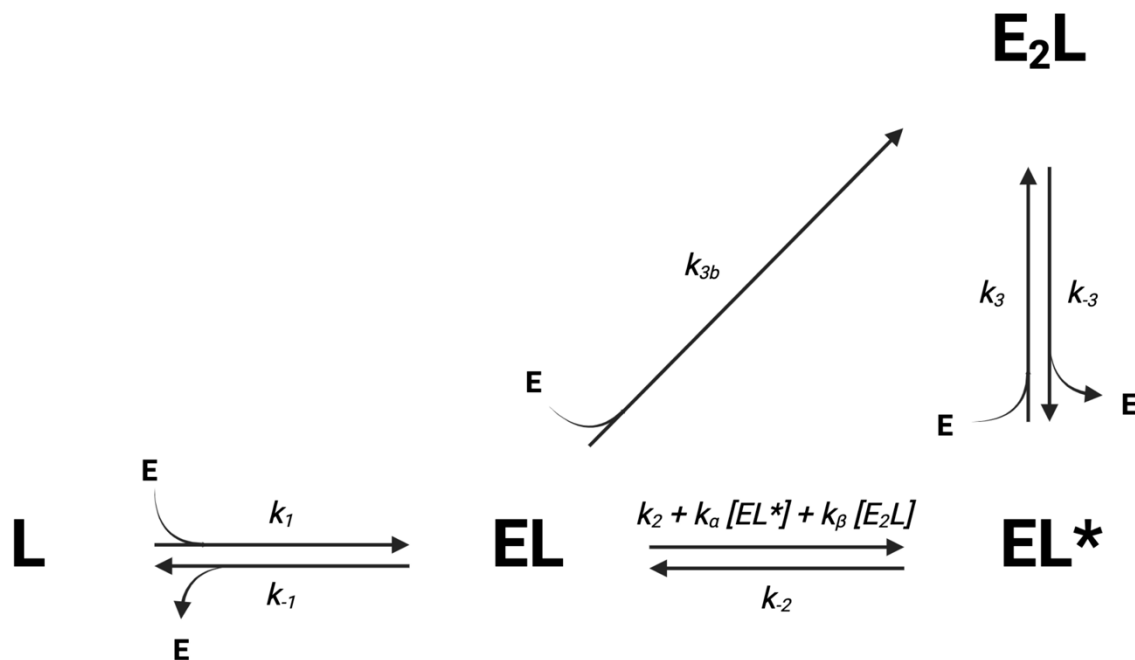
**b**

### Protein or Ligand Isomer Model



**c**

### Isomer-Limited Sequential Binding with Feedback Isomerization



**Figure 6: Various Reaction Schemes that Predict Biphasic Protein-Ligand Dissociation.** **[a] Sequential Protein Binding Model.** After initial protein-ligand association, additional protein can associate with the complex to form higher stoichiometry complexes. If the complex states with differing stoichiometries also have differing stabilities, multiphasic dissociation kinetics can be produced. If protein associates at differing rates with ligand versus existing complex, multiphasic association kinetics can be produced. Based on this model, rate constants for Sox2<sub>HMG</sub> CBS dsDNA binding are inferred to be  $k_{1,2} \approx 8 \times 10^5 \text{ M}^{-1} \text{ s}^{-1}$ ,  $k_{-1} \approx 1 \times 10^{-3} \text{ s}^{-1}$ , and  $k_{-2} \approx 5 \times 10^{-2} \text{ s}^{-1}$ . The rate constants for Sox2<sub>HMG</sub> G4 RNA binding are inferred to be  $k_1 \approx 1 \times 10^6 \text{ M}^{-1} \text{ s}^{-1}$ ,  $k_2 \approx 7 \times 10^4 \text{ M}^{-1} \text{ s}^{-1}$ ,  $k_{-1} \approx 1 \times 10^{-3} \text{ s}^{-1}$ , and  $k_{-2} \approx 5 \times 10^{-2} \text{ s}^{-1}$ . **[b] Protein or Ligand Isomer Model.** The protein (left) or ligand (right) may isomerize to an alternative state, which produces different protein-ligand association and/or dissociation rates. **[c] Isomer-Limited Sequential Binding with Feedback Isomerization.** After initial protein-ligand association to form a stable complex (EL), protein can inefficiently associate with the initial complex to form higher-order stoichiometry complex (E<sub>2</sub>L), or the initial complex can isomerize to an alternative complex state (EL\*) that can more readily accommodate additional protein monomers. Complex isomerization is intrinsically slow but may be accelerated by “feedback” from isomerized complex or higher-order stoichiometry complex. Reaction scheme could produce monophasic, biphasic, or “lagged” triphasic association, and monophasic, biphasic, or triphasic dissociation, depending on specific values of rate constants.