

The novel roles of bovine milk-derived exosomes on skin anti-aging

Lu Lu^{1, 2#}, Wei Bai^{1,2#}, Miao Wang^{1,2#}, Chunle Han^{1,2}, Huanqing Du^{1,2}, Na Wang^{1,2}, Mengya Gao^{1,2}, Dan Li^{1,2}, Fengwei Dong^{1,2}, Xiaohu Ge^{1,2*}

¹Tingo Exosomes Technology Co., Ltd, Tianjin, China

²Institute of TINGO Regenerative Medicine (Tianjin), Tianjin, China

[#]These authors contributed equally to this work.

*Correspondence to gexiaohu@tingocell.com (Xiaohu Ge)

Abstract

Exosomes are small vesicles released from cells and present in various mammal biological fluids, such as bovine milk, which worked for skin care for many years besides dairy. In addition, Exosomes were regarded as a vehicle for intercellular communication. Therefore, we aimed to investigate the novel roles of bovine milk-derived exosomes (MK-Exo) on human skin anti-aging. Purified MK-Exo can be directly uptake by the keratinocytes and fibroblast *in vitro* and upregulate the expression of the natural factors related to skin moisturizing, including Filaggrin (FLG), Aquaporin 3 (AQP3), CD44 in the keratinocytes and hyaluronidase (HAS2) in the fibroblast, and MK-Exo promoted the cell migration of the fibroblast, while rescue its expression of type I collagen (Col I), type III collagen (Col III) after ultraviolet radiation. Furthermore, the phototoxicity test, photoallergy test, repeated skin irritation test, skin allergy test, and patch test confirm the safety of MK-Exo on the skin. Finally, the roles of MK-Exo in preserving moisture and anti-wrinkle were also identified in humans. Then, MK-Exo was smeared on the facial skin of 31 female volunteers twice a day for 28 days, and the functions were evaluated following the safety assessment *in vivo*. These studies reveal the novel roles of bovine milk-derived exosomes in human skin aging, which opens a new way of skin care.

Keywords: Bovine milk derived Exosomes; Skin anti-aging; Moisture; Anti-wrinkle

Introduction

Bovine milk is known to be used as a raw material in the food industry. It is also widely used in cosmetic industries due to its considerable biological potential, mainly derived from casein and whey proteins [1, 2]. Milk-based products positively affect skin conditions, including improved wound healing, elasticity, and moisturizing when topically applied in creams, and ointments, et al. [3-5].

Skin is a protective layer of the body of any animal, including humans. As age progresses, specific changes occur in the skin, the most visible signs of which include

35 wrinkles, dryness, and loss of natural smoothness [6, 7]. Although skin aging is a
36 complex biological process, the mechanism is not yet completely understood; it is
37 commonly considered that intrinsic factors of free radical toxicity, hormonal
38 reduction, mitochondrial DNA damage, extrinsic factors of UV, and lifestyle are
39 responsible for skin aging [8-11]. Several anti-aging strategies are developed, such as
40 skin care, moisturizing preparation, and anti-wrinkling treatment. Wrinkles are mainly
41 caused by a lack of elastic features of the skin, so inhibition of elastic fiber
42 degradation is used for anti-wrinkling [12]. Hyaluronic acid and sericin were also
43 used to preserve the skin's hydration for anti-wrinkling [13, 14].

44 Exosomes (40–150 nm in diameter) are one subtype of extracellular vesicle
45 originating in the endocytotic compartment and released via biological membrane
46 fusion between the multi-vesicular bodies and the cell membranes from nearly all
47 kinds of cells. They contain proteins, lipids, nucleic acids, and other biological
48 molecules and work as intercellular communication tools to regulate the properties of
49 target cells [15-17]. The released exosomes widely exist in intercellular space and
50 various bodily fluids, including bovine milk [18].

51 Some studies have shown that exosomes distributed in the skin also play a role in skin
52 conditions through the intercellular crosstalk of various skin cells. For example, Hu et
53 al. found that human dermal fibroblast-derived exosomes could ameliorate skin
54 photoaging [19]. Liu et al. discovered that exosomes derived from keratinocytes could
55 regulate melanocyte pigmentation via loaded microRNA [20]. Besides exosomes
56 originally from the skin, Kim et al. found that ectopic exosomes derived from milk
57 can suppress melanogenesis [21].

58 Bovine milk is rich in exosomes. Although the application of bovine milk-derived
59 ingredients is widely accepted as functional cosmetics, there still needs to be more
60 studies on the roles of bovine milk-derived exosomes in skin conditions.

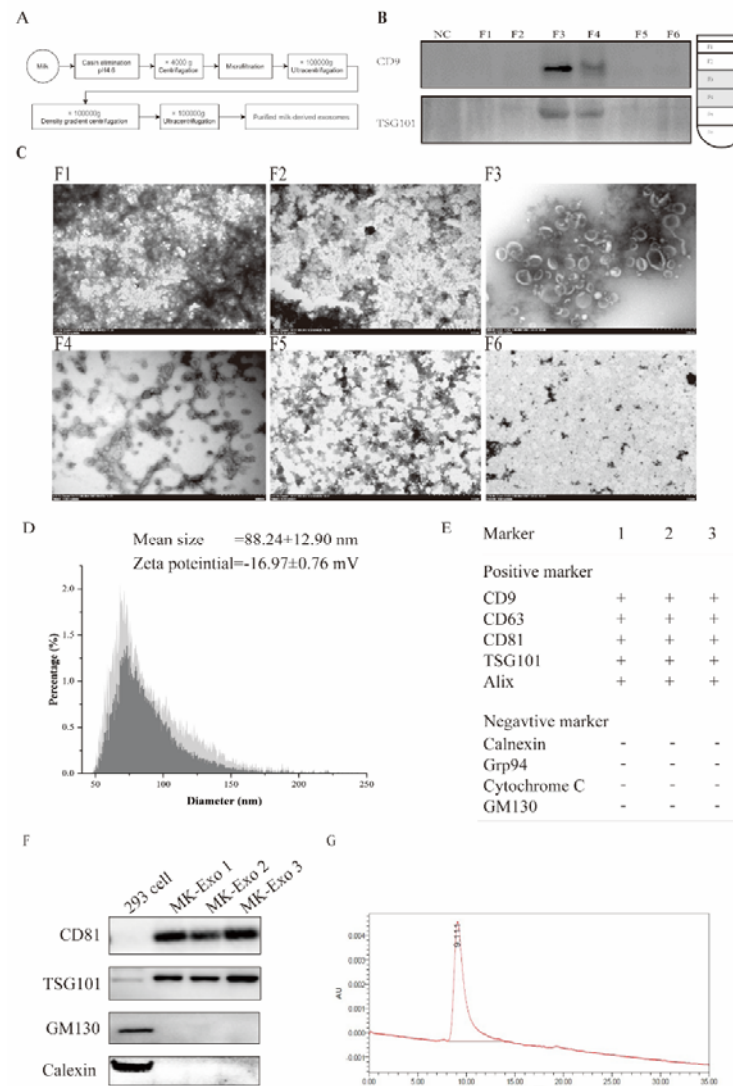
61 In this study, we found that bovine milk-derived exosomes(MK-Exo) can up-regulate
62 the expression of FLG and CD44 in keratinocytes and HAS2 in fibroblasts and arrest
63 the UV-triggered collagen reduction. Meanwhile, bovine milk-derived exosomes can
64 also significantly improve the cell migration of fibroblasts. Followed by the skin
65 toxicity tests in animals and humans, we further validated the functions of bovine
66 milk-derived exosomes moisturizing and anti-wrinkling in female volunteers. These
67 results indicate that the bovine milk-derived exosomes are safe and may work as a
68 novel ingredient for anti-aging skin.

69 **Results**

70 **Characterization and labeling of the exosomes isolated from bovine milk**

71 Owing to abundant non-EV proteins, sugars, milk fat, and other components, whole
 72 bovine milk is a highly complex material for isolating pure MK-Exo. Therefore, we
 73 combined acid precipitation and density gradient ultracentrifugation to isolate MK-
 74 Exo (Figure. 1A). We chose fresh bovine milk to avoid any unexpected risk from
 75 industrial food processing. Multiple steps of centrifugation clarified the supernatant of
 76 acid precipitation. We employed morphology and protein markers (CD9 and TSG101)
 77 to identify the MK-Exo existing fractions (Figure. 1B, C) following DC. The MK-Exo
 78 collection was further identified by size distribution and exosomal positive and
 79 negative markers (Figure. 1D, F). The slight peak of HPLC indicates the high purity
 80 of MK-Exo (Figure. 1G) in the collections.

81 The hydrophobic fluorescent dye is widely used in labeling exosomes for tracing *in*
 82 *vitro* and *in vivo*. Still, we found that the labeling efficiency of exosomes with
 83 different dyes is highly varied, from 15.7% to more than 90% (Supplementary Figure.
 84 1A, D). Therefore, to accurately trace the exosomes, we labeled MK-Exo using AIE,
 85 which has the highest efficiency for further study.



86

Fig. 1. Preparation and characterization of MK-Exo. (A) Flow chart of the preparation of MK-Exo by density gradient ultracentrifugation. Each fraction of density gradient ultracentrifugation was analyzed by Western blot of CD9, TSG101 (B), and the morphological characteristics of exosomes in MK-Exo were identified by TEM (C). (D) Mixed the fraction of F3 and F4, particle size was determined by NanoFCM. (E) LC-MS analyzed positive markers and negative markers of exosomes. (F) The expression of CD81, TSG101, GM130, and Calnexin was analyzed by Western blot. (G) HPLC identified the purity of MK-Exo.

The effects of MK-Exo on human keratinocytes

Keratinocytes are constructional skin cells and take part in moisturizing [22]. To investigate whether MK-Exo can influence the moisturizing functions of the

97

98 keratinocytes, we first incubated the MK-Exo with the HaCat cells. The cells could
 99 up-take the MK-Exo (Figure. 2A). The preliminary result indicated that MK-Exo
 100 might function across the species on the human keratinocytes.
 101 To further investigate whether MK-Exo could trigger gene expression change related
 102 to moisture in keratinocytes *in vitro*, we explored the mRNA and protein levels of a
 103 few relevant genes considered as the indicator for moisturizing. We found that MK-
 104 Exo was able to elevate the status of filaggrin (FLG), a natural moisturizer, up to
 105 around three times (Figure. 2B, C), and CD44, the receptor of HA, up to more than
 106 60% (Figure. 2B, E). Aquaporin 3 (AQP3) is the most abundant skin aquaporin that
 107 facilitates water and glycerin transport into SC to help keep it hydrated. However, we
 108 found that MK-Exo reduced the level of AQP3 at relatively high concentrations
 109 (Figure. 2B, D). The results indicate that MK-Exo may work as moisture by inducing
 110 the expression of FLG and CD44 in keratinocytes without a change in AQP3
 111 expression.

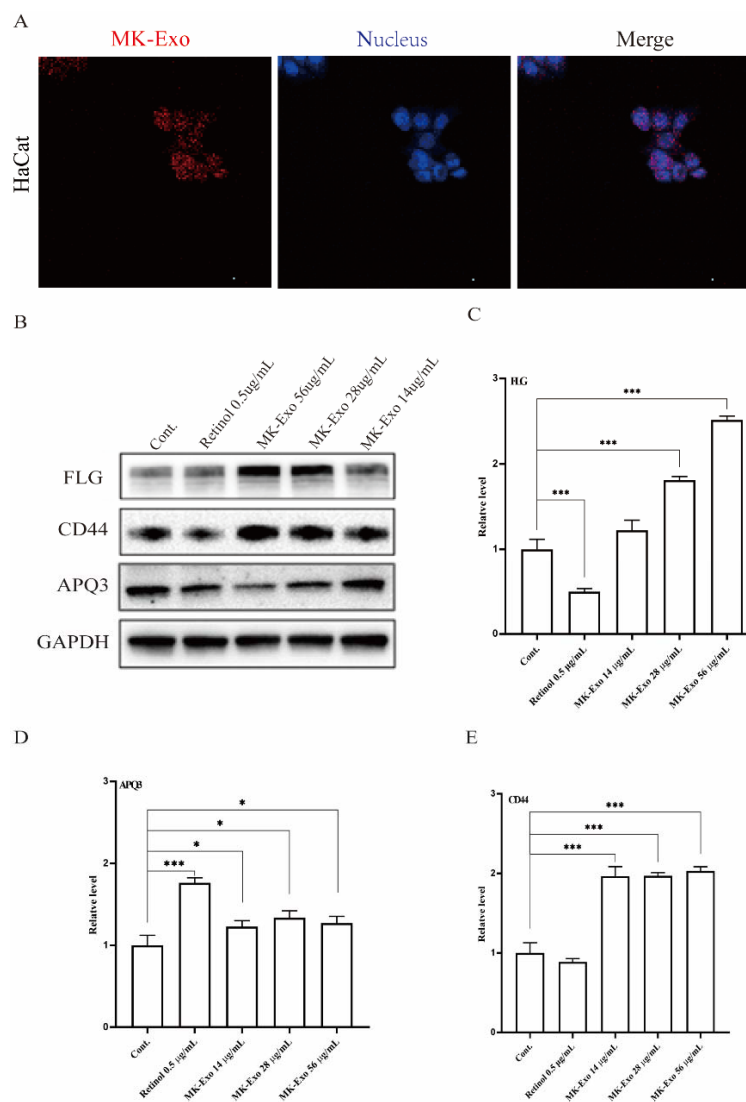


Fig. 2. The effect of MK-Exo on HaCat. (A) Representative fluorescence images of AIE-labeled MK-Exo uptake by HaCaT. (B) The protein expression levels of FLG, APQ3 and CD44 were determined by Western blot. The gene of FLG (C), APQ3 (D), and CD44 (E) transcript levels were determined by RT-qPCR. Asterisk (*) represented $P < 0.05$; a double asterisk (**) represented $P < 0.01$; a triple asterisk (***) represented $P < 0.001$.

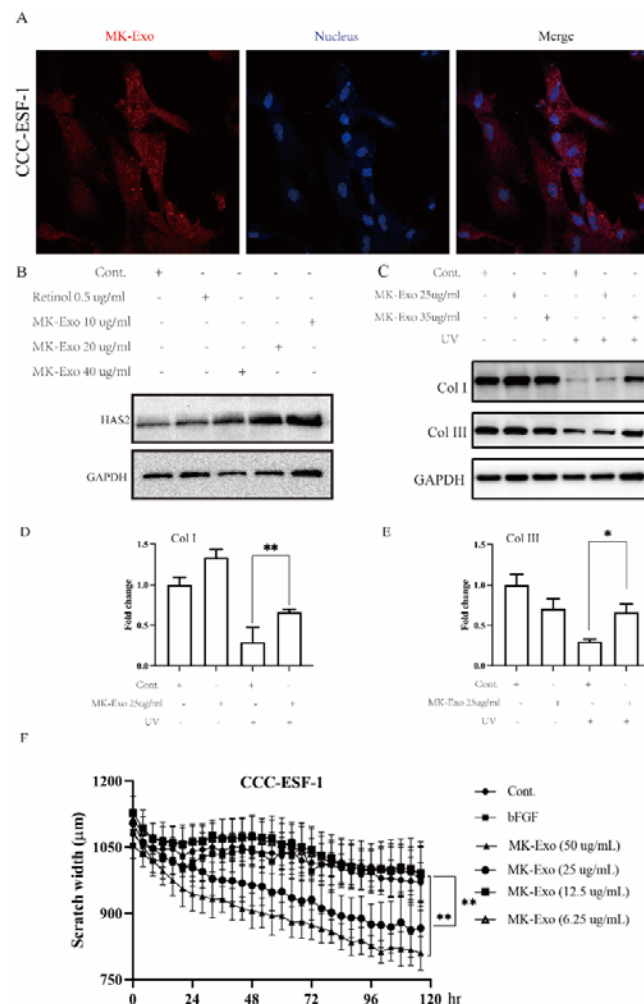
The effects of MK-Exo on human fibroblasts

Fibroblasts are constructional skin cells and participate in skin conditions, including moisturizing and anti-wrinkling. To investigate whether MK-Exo can influence the functions of the fibroblasts, we first incubated the MK-Exo with the CCC-ESF-1 cells. We found that the cells also could up-take the MK-Exo (Figure. 3A). The preliminary result suggested that MK-Exo also might function across the species on

125 the human skin fibroblasts.

126 To further investigate whether MK-Exo could influence the moisturizing skin
127 function of human fibroblasts *in vitro*, we explored the protein levels of hyaluronan
128 synthases (HAS2). We found that MK-Exo could elevate the level of HAS2 more than
129 two times (Figure. 3B). More HAS2 indicates that more hyaluronan acid will be
130 synthesized. The result suggests that MK-Exo also could play the role of moisturizing
131 by inducing the production of hyaluronan acid, which is also one kind of natural
132 moisture factor, by HAS2.

133 Skin aging is primarily due to alterations in the dermal extracellular matrix,
134 significantly decreasing collagen I content. Meanwhile, about 80% of facial aging is
135 attributed to ultraviolet radiation exposure from sunlight, which also could introduce
136 collagen degradation [23]. However, MK-Exo did not significantly change the
137 expression level of collagen I and III; instead, it can moderate collagen I and III
138 reductions in fibroblasts following UV exposure (Figure. 3C, E). In addition, we
139 found that MK-Exo also could improve fibroblast cell migration (Figure. 3F,
140 Supplementary Figure. 2) instead of keratinocytes (Data not shown). These results
141 suggested that MK-Exo may play the role of anti-wrinkling by reducing collagen
142 degradation and improving cell migration.



143

144 **Fig. 3. The effect of MK-Exo on CCC-ESF-1.** (A) Representative fluorescence
145 images of AIE-labeled MK-Exo uptake by CCC-ESF-1. (B) The protein expression
146 levels of HAS2 and GAPDH were analyzed by Western blot. The expression of Col I
147 and Col III were determined by Western blot (C) and RT-qPCR (D-E). (F) The varies
148 of scratch width at different time points. (Asterisk (*) represented $P < 0.05$; a double
149 asterisk (**)) meant $P < 0.01$.)

150 Primary safety evaluation of MK-Exo on skin

151 As the potential to be used as a cosmetic material, skin contact is one of the most
152 common routes to MK-Exo during their intended daily use. Nevertheless, the toxic
153 effects of cutaneous MK-Exo exposure on animal and human skin remain unexplored.
154 Therefore, first, we preliminary evaluated the toxicity of MK-Exo on the skin in
155 animals by combing skin allergy tests, skin photoallergy tests, repeated skin irritation
156 tests, and skin photo-irritation tests. No allergic reaction occurred following the
157 sensitization by MK-Exo in the skin allergy tests, skin photoallergy tests (Table. 1),

158 and repeated skin irritation tests (Supplementary Table. 2). Meanwhile, no skin
159 damage occurred on the skin in the skin photo-irritation tests (Supplementary Table.
160 3).
161 To further confirm the potential sensitization on human skin, we recruit 31 female
162 volunteers to evaluate MK-Exo sensitization by a patch test. None of the skin
163 reactions were observed at 0.5hr, 24hr, and 48hr (Supplementary Table. 4) in all the
164 volunteers.
165 Overall, the results indicated the absence of sensitization and irritant potential of MK-
166 Exo on animal and human skin, and there will be no risk in efficacy evaluation in
167 humans.

Table 1 Skin allergic test and skin photoallergy test

Group	Time (h)	Erythema					Edema				Quantity (Score ≥ 2)	Sensitization rate (%)
		0	1	2	3	4	0	1	2	3		
Skin allergic test												
NC	24	10/10	0	0	0	0	10/10	0	0	0	0	0
	48	10/10	0	0	0	0	10/10	0	0	0	0	0
PC	24	0	9/20	11/20	0	0	0	8/20	12/20	0	20	100
	48	2/20	13/20	5/20	0	0	1/20	13/20	6/20	0	17	85
MK-Exo	24	20/20	0	0	0	0	0	0	0	0	0	0
	48	20/20	0	0	0	0	0	0	0	0	0	0
Skin photoallergy test												
NC	24	10/10	0	0	0	0	10/10	0	0	0		0
	48	10/10	0	0	0	0	10/10	0	0	0		0
PC	24	0	7/10	3/10	0	0	2/10	8/10	0	0		80
	48	2/10	8/10	0	0	0	6/10	4/10	0	0		40
MK-Exo	24	10/10	0	0	0	0	10/10	0	0	0		0
	48	10/10	0	0	0	0	10/10	0	0	0		0

168
169 The proportion of responding animals to the number of tested animals in the erythema
170 and edema column of the skin reaction score (0, 1, 2, 3, 4), and the sensitization rate
171 is the percentage of the number of animals with a response score of 2 or more to the
172 total number of animals in that group.

173 The anti-aging effect of MK-Exo in human

174 To confirm the potential effect of MK-Exo in anti-aging through moisturizing and
175 anti-wrinkle in humans, we recruited 31 female volunteers aged 26 to 45 years old.
176 After the patch tests, everyone facial applied the MK-Exo twice daily for 28 days
177 without using other cosmetics. The skin conditions were detected on day 2, day 14,
178 and day 28 separately. The skin's moisture content increased by 4.64% on day 14 and
179 5.6% on day 28 in all the cohorts. Interestingly, the increase of moisture content is
180 higher in the volunteers aged 36-45, while the increase in volunteers aged 26-35 is not
181 statistically significant. (Table 2). The skin brightness also dramatically increased on
182 day 28. More than 90% satisfy the moisturizing effect of MK-Exo (Data not shown).

In addition, the photo of the volunteer's facial skin gloss increased on day 28. The results indicated that MK-Exo has the function of moisturizing. Skin elasticity was also detected on day 14 and day 28. We found that F3/F4 value and R2 value increased by 6.33% and 7.24% on day 28, with a higher increase of 10.74% in the cohort of age 36-45 (Table. 2). In addition, the wrinkle area and amount of wrinkles were detected and reduced by 9.37% and 5.27% on day 14 and by 9.59% and 4.99 % on day 28. The results suggested that MK-Exo could also play the role of anti-wrinkle in humans. All results indicated that MK-Exo was safe and may moisturize and anti-wrinkle by improving the expression of FLG, CD44 in keratinocytes, and HAS2 in fibroblasts, enhancing cell migration and inhibiting the UV-induced reduction of collagen in fibroblasts.

Table 2 Skin moisturizing test and skin wrinkle test

	Age	D2	D14	D28
Moisture content	26-45 (n=31)	+1.66%	+4.64%*	+5.60%*
	26-35 (n=14)	+3.23%	+4.04%	+4.66%
	36-45 (n=17)	+1.39%	+7.16%*	+7.96%*
F3/F4	26-45 (n=31)	-	+5.35%*	+6.33%*
	26-35 (n=14)	-	+6.15%	+3.66%
	36-45 (n=17)	-	+6.29%*	+10.74%**
R2	26-45 (n=31)	-	+2.76%	+7.24%**
	26-35 (n=14)	-	+0.12%	+2.62%
	36-45 (n=17)	-	+5.37%	+11.80%**
Amount of wrinkles	26-45 (n=31)	-	-5.27%***	-4.99%***
	26-35 (n=14)	-	-7.24%**	-6.77%**
	36-45 (n=17)	-	-3.65%***	-3.52%**
Wrinkle area	26-45 (n=31)	-	-9.37%***	-9.59%***
	26-35 (n=14)	-	-7.67%**	-10.01%**
	36-45 (n=17)	-	-10.59%***	-9.29%***

(Asterisk (*) represented $P < 0.05$; a double asterisk (**) meant $P < 0.01$; a triple asterisk (***) represented $P < 0.001$.)

Discussion

It's the first study to combine and evaluate the safety and efficacy of MK-Exo in the skins of animals and humans and the first evidence supporting that MK-Exo could participate in skin care by anti-aging. It is also the first discovery that MK-Exo could change gene expressions such as FLG, CD44, HAS2, and collagen. This study may provide shreds of more solid evidence for the potential of MK-Exo as cosmetic

material. However, it is not yet clear how MK-Exo works and what is the critical molecular player. MK-Exo consists of thousands of types of proteins and non-coding RNAs. Omics technology can be employed to explore the key players [24-26], the specific proteins or RNAs which induce the changes in the given biological processes of the skin, in further studies. The key players will be the critical quality attributes and stand at the center of the quality control system in the industrial process. Interestingly, we found the MK-Exo function differently on cohorts of different ages in some test items. MK-Exo may have a stronger position on skin anti-aging in older females. Indeed, more subjects may be needed for confirmation. However, the difference might suggest that different age cohorts have specific skin conditions and respond differently to the MK-Exo. The mechanism should be explored deeply. In addition, exosomes are a novel potential non-virus delivery system for small molecular compounds, peptides, proteins, and nuclear acids in the pharmaceutical industry [27-30]. Because of unlimited supply, low cost, and simple oral route, MK-Exo may be one optimal candidate for the drug delivery system. However, it is generally believed that large-scale production and quality control standards may hinder industry progress. Fortunately, we have erased the central problems in the production process. In addition to drugs, MK-Exo can also work as a delivery system in the cosmetic industry. Crossing the skin barrier is a critical issue for most functional cosmetic ingredients. Meanwhile, MK-Exo function in human skin indicates that it can cross the skin barrier, at least partially. Therefore, MK-Exo could also work as a delivery system for other available cosmetic materials crossing the skin and improving their efficacy.

Materials and Methods

2.1 Exosomes preparation

Fresh bovine milk was obtained from a local dairy factory. Exosomes were isolated by density gradient centrifugation. Briefly, the pH of milk was adjusted to pH 4.6 by hydrochloric acid (Merck) followed by centrifugation at 4000×rpm (Beckman) at 4°C for 30 min. Then, Sucrose density gradient centrifugation was performed as described previously[31], and the isolated MK-Exo were sterile filtered through a 0.22 µm filter and stored at -80 °C until further use.

2.2 Quantitative assessment of protein concentration

The protein concentration of MK-Exo was measured by BCA Kit (Thermo, Waltham Mass, USA) according to the manufacturer's instructions.

238 **2.3 Transmission electron microscope**

239 The MK-Exo (100 µg/mL) was fixed in 2% (w/v) paraformaldehyde at room
240 temperature for 15 min. The mixture (10 µL) was mounted on a formvar-carbon
241 coated grid (Beijing XXBR Technology, Beijing, China) at room temperature for 3
242 min and then stained by uranyl oxalate solution (4% uranyl acetate, 0.0075 M oxalic
243 acid, pH 7) for 1 min. After the samples were observed by TEM (Hitachi High-
244 Technologies Corporation, Tokyo, Japan).

245 **2.4 Size distribution and particle number**

246 The MK-Exo size distribution and number were determined by NanoFCM
247 (NanoFCM Inc., Xiamen, China) according to the user manual.

248 **2.5 Western blotting**

249 Western blot analyses were performed according to the previously described protocols
250 [32]. The primary antibodies used in this study were as follows: CD81 (Abcam),
251 TSG101 (BD, 612696), GM130 (Abcam), Calnexin (Abcam), FLG(Santa Cruz),
252 CD44(Thermo), AQP3(Abcam), Col I(Abcam), Col III(Abcam), HAS2(Santa Cruz),
253 GAPDH (Abcam). Depending on the primary antibody, the secondary antibodies were
254 either goat anti-rabbit (Abbsa) or goat anti-mouse antibodies (Protientech). The
255 immunoreactive protein was used with ECL (Thermo) to image immunoblots.

256 **2.6 Purity analysis**

257 The sample was analyzed using an SEC-1000 column (7.8*150 mm, 7 µm, Thermo).
258 The column was eluted at a 0.3 mL/min flow rate with 150 mM NaCl and 20 mM
259 phosphate buffer pH 7.2.

260 **2.7 Proteomic analysis**

261 To identify the proteomics of MK-Exo, liquid chromatography/mass spectroscopy
262 (LC-MS/MS, Thermo) analysis was performed as described before with modification
263 [33]. The LC-MS/MS using Easy NLC 1200-Q Exactive Orbitrap mass spectrometers
264 (Thermo). The nano-HPLC system was equipped with an Acclaim PepMap nano-trap
265 column (C18, 100 Å, 75 µm × 2 cm) and an Acclaim Pepmap RSLC analytical
266 column (C18, 100 Å, 75 µm × 25 cm). One µL of the peptide mix was typically
267 loaded onto the enrichment (trap) column. All spectra were collected in positive mode
268 using full-scan MS spectra scanning in the FT mode from m/z 300-1650 at resolutions
269 of 70000. For MSMS, the 15 most intense ions with charge states ≥2 were isolated
270 with an isolation window of 1.6 m/z and fragmented by HCD with a normalized
271 collision energy of 28. A dynamic exclusion of 30 seconds was applied.

272 The raw files were searched using Proteome Discover (version 2.4, Thermo) with
273 Sequest as the search engine. Fragment and peptide mass tolerances were set at 20
274 mDa and 10 ppm, respectively, allowing a maximum of 2 missed cleavage sites. The
275 false discovery rates of proteins and peptides were 1%. DAVID Bioinformatics
276 Resource 2021 (<http://david.abcc.ncifcrf.gov/>)² analyzed the differential expression
277 proteins with recommended analytical parameters to identify the most significantly
278 enriched signal transduction pathways in the data set.

279 **2.8 Cell lines**

280 The cells of CCC-ESF-1 and HaCaT were purchased From the National Institute of
281 Cell Resource, Beijing, China. CCC-ESF-1 was cultured in DMEM (Gibco, CA,
282 USA) supplemented with 10% FBS (Gibco), 100 µg/mL streptomycin. HaCaT were
283 cultured in MEM (Gibco) supplemented with 10% FBS, 100 µg/mL streptomycin.
284 Cells were maintained in a humidified incubator at 37°C under an atmosphere of 5%
285 CO₂.

286 **2.9 RNA isolation**

287 According to the manufacturer's instructions, total RNA was isolated from cells using
288 Trizol (Thermo).

289 **2.10 RT-qPCR**

290 RT-qPCR was performed using PrimeScriptTM RT Master Mix (Takara) and SYBR
291 Green Premix Pro Taq HS (Takara). The primers used in this study were listed in the
292 supplementary table 1.

293 **2.11 MK-Exo up-taking**

294 2×10^4 cells/well were seeded onto a chamber side and incubated overnight. Then,
295 the cells were treated with AIE labeled MK-Exo (10 µg/mL) for 4 h. after fixation
296 with 4% paraformaldehyde for 20 min. The nucleus was labeled with DAPI (Thermo)
297 and then observed by a confocal laser-scanning microscope.

298 **2.12 Cell migration assays**

299 The wound healing assay detected cell migration. Coculture experiments were
300 performed by seeding CCC-ESF-1 (2×10^4 cells/well) and culturing them with
301 different concentrations of MK-Exo for 24 h; the migration rate was measured by
302 quantifying the wound width automatically (CBM, Biotek).

303 **2.13 Skin allergy test**

304 The male Dunkin Hartley was randomized into three groups: NC (PBS, n=10), PC (2,
305 4-dinitrochlorobenzene, n=20), and MK-Exo (n=20), and were topically treated with

sample for six h induction at D0, D7 and D14 (NC: 0.2g PBS, PC: 0.2mL (10mg/mL) 2, 4-dinitrochlorobenzene, MK-Exo: 0.2g). Then, on day 28, the sample was applied to the untreated abdomen and the rib area for excitation (NC: 0.2g MK-Exo, PC: 0.2mL (5mg/mL) 2, 4-dinitrochlorobenzene, MK-Exo: 0.2g), and observed skin responses at different time points (24h, 48h).

2.14 Skin Photoallergy test

The male Dunkin Hartley was randomized into three groups: NC (PBS, n=10), PC (2, 4-Methylcoumarin, n=10), and MK-Exo (n=10). First, the light induction phase involved Dunkin Hartley injecting 0.1 mL sensitizer (Freund's Adjuvant Complete: Normal saline = 1:1) under the four corners of the neck hair removal area and applying samples in the hair removal area (NC: 0.1g PBS, PC: 0.1mL (10%) 2, 4-Methylcoumarin, MK-Exo: 0.1g), after 30 min, UVA (10.2J/cm²) was irradiated once daily for five times. Secondly, Dunkin Hartley divided the two sides of the spine into four acting sites (left 1, 3 and right 2, 4). Next, 3 and 4 were applied samples for 30 min (NC: 0.02g PBS, PC: 0.02 mL (10%) 2, 4-Methylcoumarin, MK-Exo: 0.02g). After that, necks 1 and 3 were covered with tin foil, and UVA irradiation was performed at 10.2 J/cm². Finally, the skin reaction was observed at 24h and 48h.

2.15 Skin moisturizing and wrinkle test

The healthy Chinese subjects of female (n=31, age: 26-45) and the MK-Exo was diluted with water to 60 ug/mL and applied morning and evening for 28 days. The efficacy of MK-Exo was measured by VISIA using skin hydration (Corneometer CM825), skin elasticity (Cutometer MPA580), skin wrinkle number, and area (PRIMOS CR).

2.16 Statistical analyses

The statistical methods used for each experiment are described in the relevant figure legends. Experiments were performed with at least three replicates, and results were considered statistically significant at $p < 0.05$.

Reference

- [1] K. Kazimierska, U. Kalinowska-Lis, Milk Proteins-Their Biological Activities and Use in Cosmetics and Dermatology, *Molecules* 26(11) (2021).
- [2] L. Jaiswal, M. Worku, Recent perspective on cow's milk allergy and dairy nutrition, *Crit Rev Food Sci Nutr* 62(27) (2022) 7503-7517.

- 338 [3] S. Shanbhag, A. Nayak, R. Narayan, U.Y. Nayak, Anti-aging and Sunscreens:
339 Paradigm Shift in Cosmetics, *Adv Pharm Bull* 9(3) (2019) 348-359.
- 340 [4] C. Yan, J. Chen, C. Wang, M. Yuan, Y. Kang, Z. Wu, W. Li, G. Zhang, H.G.
341 Machens, Y. Rinkevich, Z. Chen, X. Yang, X. Xu, Milk exosomes-mediated miR-31-
342 5p delivery accelerates diabetic wound healing through promoting angiogenesis, *Drug*
343 *Deliv* 29(1) (2022) 214-228.
- 344 [5] H. Kim, D.E. Kim, G. Han, N.R. Lim, E.H. Kim, Y. Jang, H. Cho, H. Jang, K.H.
345 Kim, S.H. Kim, Y. Yang, Harnessing the Natural Healing Power of Colostrum: Bovine
346 Milk-Derived Extracellular Vesicles from Colostrum Facilitating the Transition from
347 Inflammation to Tissue Regeneration for Accelerating Cutaneous Wound Healing,
348 *Adv Healthc Mater* 11(6) (2022) e2102027.
- 349 [6] Z. Zou, X. Long, Q. Zhao, Y. Zheng, M. Song, S. Ma, Y. Jing, S. Wang, Y. He,
350 C.R. Esteban, N. Yu, J. Huang, P. Chan, T. Chen, J.C. Izpisua Belmonte, W. Zhang, J.
351 Qu, G.H. Liu, A Single-Cell Transcriptomic Atlas of Human Skin Aging, *Dev Cell*
352 56(3) (2021) 383-397 e8.
- 353 [7] J.C. Kim, T.J. Park, H.Y. Kang, Skin-Aging Pigmentation: Who Is the Real
354 Enemy?, *Cells* 11(16) (2022).
- 355 [8] S. Zhang, E. Duan, Fighting against Skin Aging: The Way from Bench to Bedside,
356 *Cell Transplant* 27(5) (2018) 729-738.
- 357 [9] C. Cao, Z. Xiao, Y. Wu, C. Ge, Diet and Skin Aging-From the Perspective of Food
358 Nutrition, *Nutrients* 12(3) (2020).
- 359 [10] D. Melzer, L.C. Pilling, L. Ferrucci, The genetics of human ageing, *Nat Rev*
360 *Genet* 21(2) (2020) 88-101.
- 361 [11] J. Campisi, P. Kapahi, G.J. Lithgow, S. Melov, J.C. Newman, E. Verdin, From
362 discoveries in ageing research to therapeutics for healthy ageing, *Nature* 571(7764)
363 (2019) 183-192.
- 364 [12] J.W. Shin, S.H. Kwon, J.Y. Choi, J.I. Na, C.H. Huh, H.R. Choi, K.C. Park,
365 Molecular Mechanisms of Dermal Aging and Antiaging Approaches, *Int J Mol Sci*
366 20(9) (2019).
- 367 [13] T.F. Hsu, Z.R. Su, Y.H. Hsieh, M.F. Wang, M. Oe, R. Matsuoka, Y. Masuda, Oral
368 Hyaluronan Relieves Wrinkles and Improves Dry Skin: A 12-Week Double-Blinded,

369 Placebo-Controlled Study, *Nutrients* 13(7) (2021).

370 [14] M. Essendoubi, C. Gobinet, R. Reynaud, J.F. Angiboust, M. Manfait, O. Piot,
371 Human skin penetration of hyaluronic acid of different molecular weights as probed
372 by Raman spectroscopy, *Skin Res Technol* 22(1) (2016) 55-62.

373 [15] R. Kalluri, V.S. LeBleu, The biology, function, and biomedical applications of
374 exosomes, *Science* 367(6478) (2020).

375 [16] X. Zhang, J. Tang, X. Kou, W. Huang, Y. Zhu, Y. Jiang, K. Yang, C. Li, M. Hao,
376 Y. Qu, L. Ma, C. Chen, S. Shi, Y. Zhou, Proteomic analysis of MSC-derived apoptotic
377 vesicles identifies Fas inheritance to ameliorate haemophilia a via activating platelet
378 functions, *J Extracell Vesicles* 11(7) (2022) e12240.

379 [17] Y.J. Lee, K.J. Shin, H.J. Jang, J.S. Ryu, C.Y. Lee, J.H. Yoon, J.K. Seo, S. Park, S.
380 Lee, A.R. Je, Y.H. Huh, S.Y. Kong, T. Kwon, P.G. Suh, Y.C. Chae, GPR143 controls
381 ESCRT-dependent exosome biogenesis and promotes cancer metastasis, *Dev Cell*
382 58(4) (2023) 320-334.e8.

383 [18] F. Aqil, R. Munagala, J. Jeyabalan, A.K. Agrawal, A.H. Kyakulaga, S.A. Wilcher,
384 R.C. Gupta, Milk exosomes - Natural nanoparticles for siRNA delivery, *Cancer Lett*
385 449 (2019) 186-195.

386 [19] S. Hu, Z. Li, J. Cores, K. Huang, T. Su, P.U. Dinh, K. Cheng, Needle-Free
387 Injection of Exosomes Derived from Human Dermal Fibroblast Spheroids
388 Ameliorates Skin Photoaging, *ACS Nano* 13(10) (2019) 11273-11282.

389 [20] Y. Liu, L. Xue, H. Gao, L. Chang, X. Yu, Z. Zhu, X. He, J. Geng, Y. Dong, H. Li,
390 L. Zhang, H. Wang, Exosomal miRNA derived from keratinocytes regulates
391 pigmentation in melanocytes, *J Dermatol Sci* 93(3) (2019) 159-167.

392 [21] I.S. Bae, S.H. Kim, Milk Exosome-Derived MicroRNA-2478 Suppresses
393 Melanogenesis through the Akt-GSK3beta Pathway, *Cells* 10(11) (2021).

394 [22] L.A. Beck, M.J. Cork, M. Amagai, A. De Benedetto, K. Kabashima, J.D.
395 Hamilton, A.B. Rossi, Type 2 Inflammation Contributes to Skin Barrier Dysfunction
396 in Atopic Dermatitis, *JID Innov* 2(5) (2022) 100131.

397 [23] H. Jo, S. Brito, B.M. Kwak, S. Park, M.G. Lee, B.H. Bin, Applications of
398 Mesenchymal Stem Cells in Skin Regeneration and Rejuvenation, *Int J Mol Sci* 22(5)
399 (2021).

- 400 [24] H.T. Luo, Y.Y. Zheng, J. Tang, L.J. Shao, Y.H. Mao, W. Yang, X.F. Yang, Y. Li,
401 R.J. Tian, F.R. Li, Dissecting the multi-omics atlas of the exosomes released by
402 human lung adenocarcinoma stem-like cells, *NPJ Genom Med* 6(1) (2021) 48.
- 403 [25] J.W. Song, S.M. Lam, X. Fan, W.J. Cao, S.Y. Wang, H. Tian, G.H. Chua, C.
404 Zhang, F.P. Meng, Z. Xu, J.L. Fu, L. Huang, P. Xia, T. Yang, S. Zhang, B. Li, T.J.
405 Jiang, R. Wang, Z. Wang, M. Shi, J.Y. Zhang, F.S. Wang, G. Shui, Omics-Driven
406 Systems Interrogation of Metabolic Dysregulation in COVID-19 Pathogenesis, *Cell*
407 *Metab* 32(2) (2020) 188-202 e5.
- 408 [26] W. Cohn, M. Melnik, C. Huang, B. Teter, S. Chandra, C. Zhu, L.B. McIntire, V.
409 John, K.H. Gylys, T. Bilousova, Multi-Omics Analysis of Microglial Extracellular
410 Vesicles From Human Alzheimer's Disease Brain Tissue Reveals Disease-Associated
411 Signatures, *Front Pharmacol* 12 (2021) 766082.
- 412 [27] G. Liang, Y. Zhu, D.J. Ali, T. Tian, H. Xu, K. Si, B. Sun, B. Chen, Z. Xiao,
413 Engineered exosomes for targeted co-delivery of miR-21 inhibitor and
414 chemotherapeutics to reverse drug resistance in colon cancer, *J Nanobiotechnology*
415 18(1) (2020) 10.
- 416 [28] J. Wang, W. Tang, M. Yang, Y. Yin, H. Li, F. Hu, L. Tang, X. Ma, Y. Zhang, Y.
417 Wang, Inflammatory tumor microenvironment responsive neutrophil exosomes-based
418 drug delivery system for targeted glioma therapy, *Biomaterials* 273 (2021) 120784.
- 419 [29] L. Del Pozo-Acebo, M.L.L. Hazas, J. Tome-Carneiro, P. Gil-Cabrerizo, R. San-
420 Cristobal, R. Busto, A. Garcia-Ruiz, A. Davalos, Bovine Milk-Derived Exosomes as a
421 Drug Delivery Vehicle for miRNA-Based Therapy, *Int J Mol Sci* 22(3) (2021).
- 422 [30] T. Tian, H.X. Zhang, C.P. He, S. Fan, Y.L. Zhu, C. Qi, N.P. Huang, Z.D. Xiao,
423 Z.H. Lu, B.A. Tannous, J. Gao, Surface functionalized exosomes as targeted drug
424 delivery vehicles for cerebral ischemia therapy, *Biomaterials* 150 (2018) 137-149.
- 425 [31] T. Hata, K. Murakami, H. Nakatani, Y. Yamamoto, T. Matsuda, N. Aoki, Isolation
426 of bovine milk-derived microvesicles carrying mRNAs and microRNAs, *Biochem*
427 *Biophys Res Commun* 396(2) (2010) 528-33.
- 428 [32] L. Lu, X. Zhang, B. Zhang, J. Wu, X. Zhang, Synaptic acetylcholinesterase
429 targeted by microRNA-212 functions as a tumor suppressor in non-small cell lung
430 cancer, *Int J Biochem Cell Biol* 45(11) (2013) 2530-40.

[33] B.A. Brown, X. Zeng, A.R. Todd, L.F. Barnes, J.M.A. Winstone, J.C. Trinidad, M.V. Novotny, M.F. Jarrold, D.E. Clemmer, Charge Detection Mass Spectrometry Measurements of Exosomes and other Extracellular Particles Enriched from Bovine Milk, *Anal Chem* 92(4) (2020) 3285-3292.

435

436 **Acknowledgments:**

437 We thank Professor Dan Ding of Nankai University for the gift of AIE. We also thank
438 Ganggang Zhao, Jianxin Yin, Ning Chen, Few li, Zhijun Wen, and Quan Zhang for
439 their excellent technical assistance.

440 **Funding:**

441 Funding was provided by Tingo Exosomes Technology Co., Ltd, Tianjin, China.

442 **Author contributions:**

443 All authors reviewed the manuscript; X.H.G. funding acquisition, conceptualization,
444 supervision; F.W.D. funding acquisition; L.L. conceptualization, validation, formal
445 analysis, writing - review & editing, project administration; W.B. and M.W.
446 methodology, resources, investigation, data curation, visualization, writing an original
447 draft; C.L.H. investigation, formal analysis, data curation, visualization; M.Y.G., and
448 N.W. investigation, data curation; H.Q.D. Project administration, investigation, D.L.
449 visualization, writing an original draft. All authors contributed to the article and
450 approved the submitted version.

451 **Conflict of interest:** The authors declare that the research was conducted in without
452 any commercial or financial relationships that could be construed as a potential
453 conflict of interest.

454 **Data and materials availability:** All data are in the main text or the supplementary
455 materials. Further inquiries can be directed to the corresponding author.

456 **Ethics statement:**

457 The animal study was reviewed and approved by the Experimental Animal Ethics
458 Committee of the CAST (Tianjin) Inspection and Test Co., Ltd. (2021010401).

459 The research on volunteers was in line with the basic principles of the international
460 declaration of Helsinki. Therefore, it was approved by the local Institutional Review
461 Board and Ethical Committee of the Centre Testing International (Hangzhou) Co.,
462 LTD (project identification code: A22200825411010102CR1).