

1 Running head: Host space constrains symbiont abundance

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3 **Host space, not energy or symbiont size, constrains**
4 **feather mite abundance across passerine bird species**

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56 **Statement of authorship**

57 ML, DS, RJ, MG conceptualised the study. All authosrs except for MG, LZG, and JD
58 collected data. ML and RJ performed analyses with support from JD and LZG.
59 ML and RJ wrote the first draft of the manuscript, and all authors contributed to
60 revisions.

61

62 **Data accessibility statement**

63 Data and code used in this study are available in the Dryad Digital Repository:
64 <https://datadryad.org/stash/share/LsvyoRzttwfkX1f-tUi5Fi2bJKNwJxeu48SX9VoIU0>

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70 **Abstract**

71 Comprehending symbiont abundance among host species is a major ecological
72 endeavour, and the metabolic theory of ecology has been proposed to understand what
73 constraints symbiont populations. We parameterized metabolic theory equations to
74 predict how bird species' body size and the body size of their feather mites relate to
75 mite abundance according to four potential energy (microbial abundance, uropygial
76 gland size) and space constraints (wing area, number of feather barbs). Predictions were
77 compared with the empirical scaling of feather mite abundance from 26,604 birds of
78 106 passerine species, using phylogenetic modelling and quantile regression. Feather
79 mite populations were strongly constrained by host space (number of feather barbs) and
80 not energy. Moreover, feather mite species' body size was unrelated to their abundance
81 or to the body size of their host species. We discuss the implications of our results for
82 our understanding of the bird-feather mite system and for symbiont abundance in
83 general.

84

85 **Introduction**

86 A central scope in ecology is to describe abundance patterns, to comprehend the
87 processes that underlay these patterns, and to understand their ecological consequences.
88 These questions have been mainly studied in free-living organisms, while symbiont
89 abundance patterns have received less attention (Cunning & Baker 2004; Dobson et al.,
90 2008). Symbionts (including mutualists, commensals, and parasites) are the most
91 ubiquitous, abundant, and diverse organisms on Earth (Morand 2015; Larsen et al.,
92 2017). They are key components of ecosystems and influence nutrient cycles, food
93 webs, energy flows, and community structure (Hatcher et al., 2012), and their
94 abundance can shape individual host performance and the evolution of host species
95 (Poulin & George-Nascimento 2007). Indeed, the abundance of a given symbiont in or
96 on a given host may determine the nature of the host–symbiont interaction (Bronstein
97 1994; Holland et al., 2002), with the potential to shift the nature of this relationship
98 between mutualism and parasitism (Hopkins et al., 2017).

99 Studies on symbiont abundance have mainly focused on parasites rather than on
100 non-parasitic symbionts, and on understanding differences in symbiont abundance
101 among members of a single host species rather than interspecific differences among host
102 species (Turgeon et al., 2018; Mennerat et al., 2021). At the interspecific scale, several
103 studies have found support for Harrison’s Rule which postulates that there is a positive
104 covariation between host size and symbiont size. In contrast, when considering
105 symbiont abundance instead of symbiont size, mixed results have been found for its
106 correlation with the body size of either the hosts or the symbionts (Rózsa 1997a, b;
107 Poulin 1999; Clayton & Walther 2001; Presley & Willig 2008; Krasnov et al., 2013;
108 Galloway & Lamb 2017; Surkova et al., 2018; Lamb & Galloway 2019). At macro-
109 evolutionary scale, host body size largely explained the variation in feather lice

110 effective population size, which is expected to positively correlate with symbiont
111 abundance (Doña & Johnson 2022). Overall, we are still far from understanding why
112 some host species harbour many symbiont individuals of a given taxon, while others
113 carry only a few.

114 The study of the scaling of symbiont abundance with host body size is an
115 underexplored approach to understand symbiont abundance (Morand & Poulin 2002;
116 George-Nascimento et al., 2004; Poulin & George-Nascimento 2007; Hechinger 2013).
117 Hechinger (2013) developed a hypothesis-driven quantitative framework based on the
118 metabolic theory of ecology (*sensu* Brown et al., 2004) to disentangle how host and
119 symbiont traits shape symbiont abundance across host species. This framework tries to
120 explain symbiont abundance in different hosts through the comparison of theoretical vs.
121 empirical scaling exponents of host and symbiont body size according to energy (e.g.
122 blood or secretions) and space (e.g. surface) provided by the host and according to the
123 metabolic rate and space use of symbionts (see below). Hechinger et al. (2019) used this
124 approach to investigate the relationship between host body size and the abundance of
125 ectosymbiotic mites and lice of 263 bird individuals of 42 species. Their results
126 indicated that the numbers of mites and lice were limited by access to host energy and
127 not by space. However, Hechinger et al. (2019) did not distinguish among
128 ectosymbionts with different diets, e.g. blood-feeding mites were equivalent to non-
129 parasitic mites provided that mite body sizes were similar. Here, we implemented
130 Hechinger's (2013) framework by analysing an unprecedently large dataset and
131 parametrizing scaling equations using current knowledge of the biology of a particular
132 host–symbiont system: vane-dwelling feather mites (Acariformes: Astigmata:
133 Analgoidea and Pterolichoidea) from European passerine bird species.

134 Feather mites are ectosymbionts found on almost all birds (Walter & Proctor
135 2013). Their entire life cycle is spent on their living hosts, mainly on the wing and tail
136 flight feathers, where they are usually queuing between the feather barbs (i.e., the
137 primary branches of the feather rachis; Figure 1) or next to the rachis (Kelso & Nice
138 1963; Choe & Kim 1989; Yamasaki et al., 2018). They are often said to feed on the
139 preen gland secretions and organic material trapped in them (Dubinin 1951; OConnor
140 1982; Proctor 2003; Walter & Proctor 2013; Galván et al., 2008). Still, other evidence
141 suggests a lower relevance of preen waxes as food resources (Pap et al., 2010). Algae
142 are also potential food resources for mites (Blanco et al., 2001). However, Doña et al.
143 (2019) studied the gut contents of a large sample of mites using microscopy and DNA
144 metabarcoding, and found that bacteria and fungi were the main food resources for
145 feather mites, while algae and plant materials were rather anecdotic, and bird tissues
146 such as blood or skin were not found.

147 Bird species strongly differ in feather mite abundance even when accounting for
148 intraspecific variance between localities (Díaz-Real et al., 2014). For instance, species
149 such as *Phylloscopus collybita* and *Periparus ater* consistently have very few feather
150 mites on their wings, while similar-sized *Aegithalos caudatus* and *Acrocephalus*
151 *melanopogon* often have hundreds of feather mites (Díaz-Real et al., 2014).
152 Interspecific differences in feather mite abundance are partly explained by the ecology
153 and morphology of bird species, but a large proportion of the variance remains
154 unexplained after controlling for these traits (Galván et al., 2008; authors' unpublished
155 data). To date, only one interspecific study has related bird body size to feather mite
156 abundance (Rózsa 1997b). This study found a positive correlation, albeit based on a
157 relatively small number of host species (N = 17), small number of host individuals
158 within species (range of 3–138), and without quantitatively addressing the underlying

159 mechanisms generating the positive relationship between bird size and feather mite
160 abundance.

161 In this study, we apply Hechinger's (2013) quantitative framework to disentangle
162 hosts' energy and space constraints explaining differences in feather mite abundance
163 across passerine bird species. Here we follow Hechinger's (2013) use of the term size to
164 refer to the body mass of hosts and symbionts. According to Hechinger (2013), the
165 metabolic theory of ecology predicts that if energy provided by the host (h) imposes an
166 effective ceiling to the growth of symbiont (s) populations, the maximal or carrying-
167 capacity abundance (but also mean abundance) of the symbiont in a given host
168 individual (N_s) will scale with host body size (M_h) and symbiont size (M_s) as

$$169 \quad N_s \propto M_h^{\sigma_h - \gamma_h} M_s^{-\alpha_s}. \quad \text{Eq. 1}$$

170 $-\gamma_h$ is the scaling exponent for host mass-specific metabolic rate and equals to $\alpha - 1$,
171 where α is the scaling exponent for whole-organism metabolic rate to body size ($\sim 3/4$
172 across multicellular species; Hechinger 2013). Thus, $-\gamma_h = -1/4$, and $-\alpha_s = -3/4$.
173 σ_h is the spatial exponent for host body size, and is related to the host body part that is
174 metabolically relevant for the studied symbionts (i.e., the host body part that provides
175 the food resources to the symbionts; Hechinger 2013; Hechinger et al., 2019). Current
176 knowledge points to two main energy (food) resources for feather mites, and thus, there
177 are two σ_h potential values in our study:

178 (1) waxes produced by the uropygial gland that birds spread on feathers (Galván
179 et al., 2008; Doña et al., 2019). We used data on uropygial gland size (see below) to
180 parametrize σ_h in Eq. 1, given that uropygial gland size is positively correlated, at least
181 within bird species, with the amount of waxes produced (Møller et al., 2009; Pap et al.,
182 2010).

183 (2) organic matter (mainly fungi and bacteria) available on feathers' surface
184 (Dubinin 1951; Doña et al., 2019; Labrador et al., 2022). This organic matter is not
185 produced by the host, and thus there is not a host body part that is metabolically
186 responsible for its production. We parametrized this alternative σ_h with data on how the
187 abundance of fungal and bacterial DNA (microbial abundance hereafter) on the wings
188 of passerine bird species scales with bird species body size.

189 Space provided by the host can also impose an effective ceiling on symbiont
190 populations, and then the maximal or carrying-capacity symbiont abundance in a given
191 host individual would scale with host and symbiont body size as

$$192 N_s \propto M_h^{\sigma_h} M_s^{-\sigma_s}. \quad \text{Eq. 2}$$

193 Here, σ_h indicates how the host body portion that the symbiont inhabits scales to
194 host body size (Hechinger 2013; Hechinger et al., 2019). Theoretical σ_h values are 1
195 when the studied symbionts use the host volumetrically, or 2/3 if symbionts inhabit the
196 host surface. Ideally, though, σ_h should be calculated empirically for each particular
197 study system (Hechinger 2013). We hypothesized that feather mite infracommunities
198 (all of the mite infrapopulations within a single host; Bush et al., 1997) could be
199 spatially constrained by wing area, which is the largest scale habitat for these mites.

200 Alternatively, feather mites could be constrained by the number of feather barbs on the
201 wing because they (except the genus *Trouessartia*) live in the corridors between feather
202 barbs in the ventral side of feathers (Figure 1; Mironov 2022). Moreover, *Trouessartia*
203 spp., despite living on the dorsal surface of feathers (where there are not such well-
204 defined corridors), they also queue along feather barbs (Figure 1 in Mironov &
205 González-Acuña 2013; authors' personal observation). Thus, we studied the scaling of
206 wing area and the number of barbs to bird species body size to parameterize σ_h in Eq. 2.
207 Similarly, $-\sigma_s$ is the relevant aspect of symbiont bodies that determines their spatial

208 packing on host bodies. Given that mites align in a single row along feather barbs,
209 feather mite length would be the most relevant aspect, and thus we parametrized $-\sigma_s$ as
210 $-1/3$ because this is how mite length scales to mite body size (in μg) (Supporting
211 Information).

212 In sum we used empirical data to complete the parametrization of Eqs. 1 and 2,
213 and then compared predicted scaling exponents with the empirical exponents obtained
214 by phylogenetic generalized least squares regressions and quantile regressions for the
215 abundance of feather mites across bird species, following Hechinger (2013). We show,
216 using a large dataset on feather mite abundance, how a biologically-informed
217 parametrization of the metabolic theory of ecology proposed by Hechinger (2013) is a
218 powerful approach to understanding why symbiont abundance differs between host
219 species.

220 **Materials and Methods**

221 *Feather mite morphometric data*

222 Body size in Hechinger's (2013) equations (M_h, M_s) refers to host and symbiont
223 species' body masses. Given that these data were available for only one of the mite
224 species studied here, we calculated them from feather mite species' biometry following
225 the equation provided by Edwards (1967) (Supporting Information). To do so, we
226 gathered data from adult female morphology because they are typically the largest (e.g.
227 Atyeo & Braasch 1966; Santana 1976) and more abundant life stage (e.g. Muzaffar &
228 Jones 2005; Marčanova & Janiga 2021). Feather mites ranged from 394 μm and 0.989
229 μg for *Scutulanayssus nuntiaeversis* [Berlese] to 1,121 μm and 22.85 μg for
230 *Joubertophyllodes modularis* [Berlese]. Then, to obtain a reliable measure of the mean
231 M_s on each bird species, we calculated the weighted mean body size (in μg) of the
232 feather mite species reported for each bird species. The weighted mean was calculated

233 using the number of records reported by Doña et al. (2016) for each mite species in each
234 bird species, using only the most reliable bird–mite associations (i.e., those with quality
235 score = 2; see Doña et al., 2016 for more details).

236 *Feather mite abundance data*

237 Data were obtained from *FeatherMites*, the largest dataset available on feather mite
238 abundances (see Díaz-Real et al., 2014 for details), where, for each bird individual, the
239 total number of vane-dwelling feather mites was counted (i.e., without differentiating
240 between mite species) on the 19 flight feathers (10 primaries, six secondaries, and three
241 tertials) of one wing. Because we aimed to understand the mechanisms setting the upper
242 limit for feather mite abundance, birds without feather mites were not included in the
243 analyses. Therefore, according to parasitological terminology, we analysed feather mite
244 intensity (or infracommunity size; Bush et al., 1997), i.e., the number of feather mites
245 counted in each individual bird with at least one mite, but we use the term ‘abundance’
246 hereafter due to its general use in the ecology literature. Since we could not find data on
247 the morphology of certain feather mite species in our dataset, some bird species were
248 not included in the analyses, leading to a final dataset of 26,604 individual birds from
249 106 passerine species.

250 Given the non-normal frequency distribution of feather mite abundance (Díaz-
251 Real et al., 2014), we used quantiles of mite counts at regular intervals from the 5th (Q5)
252 to the 95th quantile (Q95) to characterize feather mite abundance in each bird species.
253 Special relevance was given to Q95 as the best surrogate of the carrying capacity of
254 feather mite abundance of each bird species, following Hechinger et al. (2019).

255 *Microbial abundance data*

256 We used microbial abundance data from a recent study where the amount of fungi and
257 bacteria DNA available on feathers’ surfaces was quantified by qPCR (Labrador et al.,

258 2021). This is justified not only by current knowledge on feather mites' diet (see
259 above), but because Labrador et al. (2022) found that feather mites also occur on wing
260 flight feathers during the night, when they forage. In brief, microbial DNA was
261 extracted and amplified from the second secondary feather of the right wing of 133
262 individuals of 22 species. The amount of fungal and bacteria DNA were positively
263 correlated at the individual bird and bird species levels (Labrador et al., 2021). Hence,
264 here we combined fungal and bacterial values for each individual bird, and then
265 calculated the mean microbial DNA abundance for each bird species. This value was
266 used as a rough estimate of the microbial food resources available for their feather
267 mites.

268 *Bird morphology data*

269 Three morphological traits for the studied bird species were retrieved from the
270 literature: body size (in g), wing area, and uropygial gland size (see Supporting
271 Information for details). Moreover, the number of feather barbs was calculated for each
272 bird species combining original data on feather lengths for 40,346 birds (sample size:
273 mean = 917, min-max = 1-9,506 birds per species) captured from 1994 to 2015 at the
274 Manecorro Ringing Station (Doñana National Park, SW Spain), and feather barb density
275 reported in the literature (see Supporting Information for details, and Figure 2 for the
276 number of bird species for each morphological variable).

277 *Statistical analyses*

278 Phylogenetic generalized least squares regressions (PGLS; Symonds & Blomberg 2014)
279 were performed to retrieve (from the slope of the log-log regressions; following
280 Hechinger 2013) the scaling exponents between bird species' body size (\log_{10}
281 transformed) and the four variables (\log_{10} transformed) hypothesized to constrain
282 feather mite infracommunity sizes: wing feather microorganism amount, uropygial

283 gland size, wing area, and number of barbs of primary feathers. One multivariable
284 PGLS regression for each feather mite abundance quantile (dependent variable; \log_{10}
285 transformed) was used to calculate how it scaled with bird and mite body sizes
286 (independent variables; \log_{10} transformed). PGLS regression was also used to study the
287 relationship between bird size and the weighted mean body size of their feather mites.
288 We used the *gls* function of the *caper* R package (Orme et al., 2012) to perform the
289 PGLS regressions, which ensure the statistical independence of our samples, correcting
290 the model estimates by the phylogenetic relatedness of the studied species. We obtained
291 information on the phylogenetic relationship among bird species by downloading a
292 distribution of 1,000 trees from BirdTree (Jetz et al., 2012, <http://birdtree.org>) using the
293 Hackett backbone tree (only sequenced species; Hackett et al., 2008). Then, following
294 Rubolini et al. (2015), trees were summarized by computing a single 50% majority-rule
295 consensus tree in SumTrees v 4.5.1 in DendroPy (Sukumaran & Holder 2010, 2015).

296 In each PGLS model, we allowed the phylogenetic signal in the residuals (i.e.,
297 Pagel's lambda, λ) to be optimized towards its maximum likelihood value (Symonds &
298 Blomberg 2014). These models were also weighted by the sample size (\log_{10}
299 transformed) of each bird species to incorporate the higher uncertainty associated with
300 feather mite abundance data from host species with smaller sample sizes.

301 To further study the factors constraining feather mite infracommunities, we used a
302 multivariable quantile regression analysis on the $\log_{10}(Q95)$ of feather mite abundance
303 against $\log_{10}(\text{bird body size})$ and $\log_{10}(\text{feather mite body size})$ as independent variables
304 (Koenker & Bassett 1978; Cade & Noon 2003). We were especially interested in the
305 quantile regressions at the largest τ values because these would reflect the maximum
306 feather mite abundance that bird species can harbour, considering their body size and
307 that of their feather mites. However, we also explored the other τ values to obtain a

308 more complete picture of the scaling of feather mite abundance. We used the *quantreg*
309 R package (Koenker 2015), and assessed the slopes of the quantile regression models
310 for different τ values from 0.05 to 0.95. Quantile regression analyses were also weighted
311 by the sample size (\log_{10} transformed) of each bird species.

312 Estimated mean λ for Q95 in the PGLS regressions explained above was 0.413
313 (95% CI: 0.077–0.749). Thus, a phylogenetic modelling approach to the quantile
314 regression would require the phylogenetic scaling factor to be adjusted to $\lambda < 1$.

315 However, we were unaware of any tool able to perform such partial phylogenetic
316 correction in a quantile regression analysis (see Jovani et al., 2016). Consequently, we
317 present the results based on a non-phylogenetically corrected quantile regression and
318 assume that phylogeny is unlikely to be a confounding factor.

319 Current information on the annual cycle of European feather mites indicates that
320 their abundance peaks from winter until the onset of birds' reproductive season (Blanco
321 et al., 1997; Peet et al., 2022), when mites are transmitted from parents to offspring
322 birds, causing a lowering of feather mite abundance (Mironov & Malyshev 2002; Doña
323 et al., 2017). Thus, we tested the robustness of our conclusions by repeating all the
324 analyses on feather mite abundance for the subset of birds captured from the beginning
325 of October to the end of March (hereafter "winter"). This restriction reduced the sample
326 size to 8,066 individual birds of 77 species.

327 **Results**

328 *Predictions from Eq. 1 and Eq. 2*

329 Microbial abundance on feathers was not correlated with bird species' body size (Figure
330 2a, Table S1) which suggests that σ_h would be 0. Moreover, bacteria and fungi are not
331 produced by the host's metabolism (in contrast to uropygial gland waxes). Thus, we
332 removed $-\gamma_h$ because it refers to a host mass-specific metabolic rate scaling. In this

333 case, Eq. 1 predicts that feather mite abundance would only negatively scale with mite
334 body size as follows (note that $M_h^0 = 1$)

$$335 \quad N_s \propto M_s^{-3/4}. \quad \text{Eq. 3}$$

336 Uropygial gland size showed a strong allometric relationship with bird body
337 size, with a scaling exponent of 0.902 (Figure 2b, Table S1). Therefore, if energy
338 provided by the gland waxes of the hosts is the main constraint to feather mite
339 infracommunities, Eq. 1 would predict that the maximum feather mite abundance would
340 scale with bird and mite body size as follows

$$341 \quad N_s \propto M_h^{0.652} M_s^{-3/4}. \quad \text{Eq. 4}$$

342 Thus, the scaling exponent of uropygial gland size on bird body size in Eq. 4
343 predicts a positive effect upon feather mite abundance because larger birds would
344 provide more energy resources to mites. In contrast, the scaling exponent of feather
345 mites' body size is negative because (all else being equal) the higher energy
346 consumption of larger mites would lead to lower abundances.

347 Wing area scaled with bird species body size to 0.676 power in accordance with
348 the theoretical 2/3 scaling exponent for external host surfaces, while the number of
349 barbs scaled with a slope of 0.264 (Figures 2c and 2d, Table S1). Thus, if feather mite
350 infracommunities were limited by wing area, Eq. 2 would be

$$351 \quad N_s \propto M_h^{0.676} M_s^{-1/3}. \quad \text{Eq. 5}$$

352 However, if the number of barbs is the relevant spatial constraint for feather mite
353 infracommunities, Eq. 2 would read as

$$354 \quad N_s \propto M_h^{0.264} M_s^{-1/3}. \quad \text{Eq. 6}$$

355 Thus, Eq. 5 and Eq. 6 show the predicted positive effect of bird body size upon
356 feather mite abundance (larger birds provide more space to mites, depending on the

357 bird's body part relevant to the mites, i.e., wing area vs. barb amount), but that larger
358 mites would attain a lower abundance (fewer mites would fit on a host of a given size).

359 *Predicted vs. empirical scaling rules*

360 PGLS models showed a weak effect of feather mites' body size on their abundance
361 along all abundance quantiles (Table S2). For the few quantiles with slopes differing
362 from 0, the slopes were all positive, thus strongly departing from the predicted negative
363 slopes of $-3/4$ (Eqs. 3 and 4) and $-1/3$ (Eqs. 5 and 6) of mite abundance to mite body
364 size (Table S2). In contrast, we found a positive correlation between bird species' body
365 size and the abundance of their feather mites (Figure 3, Table S2), holding from the Q45
366 to Q95, with empirical slopes in close agreement with the slopes predicted by Eq. 6 for
367 the number of barbs (Figure 3a, Table S2). For the remaining host traits that might
368 constrain mite abundances, empirical slopes departed from those predicted by the
369 equations based on the scaling of microbial abundance (Eq. 3), uropygial gland size
370 (Eq. 4), and wing area (Eq. 5; Figure 3a, Table S2).

371 Quantile regression analyses (including both bird and mite body size as
372 independent variables) showed slopes for feather mite body size clearly departing from
373 the predicted $-1/3$ and $-3/4$ by Eqs. 3 to 6 (particularly for higher τ values; Figure S1).
374 However, we did find the expected positive relationship between Q95 feather mite
375 abundance and bird species' body size (Figure 4). Regression slopes for the highest τ
376 values were close to the one predicted by the scaling equation that considers the number
377 of barbs as a main spatial constraint (Eq. 6), but much larger than the slopes predicted
378 for microbial abundance (Eq. 3), and much lower than those predicted by the scaling
379 equations considering uropygial gland size (Eq. 4) or the wing area (Eq. 5) as the main
380 energetic or spatial constraint, respectively (Figure 4).

381 Feather mite's body size was uncorrelated with hosts' body size (Figure S2).
382 Thus, overall, these results show that larger birds hold larger feather mite abundances,
383 but this cannot be explained by feather mite size, i.e., larger birds do not carry larger
384 numbers of smaller mites.

385 Dashed lines in Figure 4b (and Figure S3b) were drawn to cross the actual Q95
386 feather mite abundance (356 mites) for *Regulus ignicapilla*, the second smallest bird
387 species in our sample (5.6 g). Thus, the dashed lines extrapolate the Q95 feather mite
388 abundance expected for larger bird species, given the actual abundance of feather mites
389 for smaller ones. Strikingly, this predicted that the largest bird species in our sample
390 (*Pyrrhocorax pyrrhocorax*; 287.5 g) would have 5,098 and 4,635 Q95 feather mite
391 abundance, according to the allometry of the wing area (Eq. 5) or the uropygial gland
392 size (Eq. 4), respectively. However, the actual abundance was four times lower (1,155),
393 and in close agreement with Eq. 6 (1,005) that involves the number of barbs. Also, the
394 extrapolation according to Eq. 3 involving microbial abundance yielded a clear
395 underestimation (356) of the actual abundance of mites. In summary, the rather flat
396 slope of the quantile regression for the largest τ values (slope, 95% CI = 0.336, 0.272–
397 0.336) shows the strong ceiling that the number of barbs imposes on feather mites'
398 abundance, precluding larger birds from holding as many mites as expected based on
399 other bird features.

400 When analysing only data from birds sampled in winter, the smaller sample size
401 led to an increase in the uncertainty of the estimates, but similar qualitative results were
402 found (see Supporting Information).

403 **Discussion**

404 The carrying capacity of birds to support feather mite populations increases with bird
405 species' body size, with a scaling exponent close to that predicted by space (but not

406 energy) constraints. Specifically, the empirical scaling we found fits closely the scaling
407 exponents predicted by the equation involving the number of feather barbs, but not wing
408 area as spatial constraint or microbial abundance or uropygial gland size as energetic
409 constraints. The size of feather mites inhabiting bird species was not correlated with the
410 abundance of mites or with the size of their hosts.

411 This space constraint seems to be in conflict with the fact that birds with many
412 feather mites typically have large sections of each flight feather, or even entire feathers,
413 devoid of feather mites (e.g. Jovani & Serrano 2004). However, feather mites show
414 strong preferences for certain feathers and feather sections (e.g. Figure 1), and these
415 preferences differ among feather mite species (Bridge 2003; Jovani & Serrano 2004;
416 Mestre et al., 2011; Fernández-González et al., 2015; Stefan et al., 2015), feather mite
417 life stages (Labrador et al., 2022), and according to environmental conditions (Wiles et
418 al., 2000) or even to time of the day (Labrador et al., 2022). Therefore, our results,
419 complemented with previous knowledge about the bird–feather mite system, show that
420 feather mite populations are spatially limited, likely because of some negative density
421 dependence acting well before the entire feather surfaces are fully occupied.

422 Our results simultaneously show a strong ceiling for the maximum feather mite
423 abundance, and manifold differences in the abundance of feather mites among bird
424 species with similar body sizes (note the logarithmic y-axis of Figures 3 and 4). For
425 instance, in the Q95 abundance of feather mites of well-sampled bird species under 10 g
426 there is an 8-fold difference from 47.9 mites per bird in *Phylloscopus collybita* to 389.5
427 mites in *Aegithalos caudatus*. Further comparative studies (as the one by Galván et al.,
428 2008) are needed to understand which traits of birds and traits of feather mites are
429 responsible for the large differences in feather mite abundances across bird species
430 (Díaz-Real et al., 2014). Our results reject the role of uropygial gland size as an

431 important constraint to feather mite populations and provide a new evidence (in addition
432 to other studies, e.g. Pap et al., 2010; Doña et al., 2019) against preen waxes being
433 important food resources for feather mites. Lastly, the role of bacteria and fungi as
434 important food resources for feather mites would need further study because their
435 potential limiting role should not be fully discarded given the small number of host
436 species analyzed here in this regard (N = 21, Figure 2a).

437 Our results can be compared with those reported by Hechinger et al. (2019), who
438 also studied the allometry of bird ectosymbionts' abundance. While they found
439 energetic constraints to be more relevant for arthropod ectosymbionts of birds, we have
440 not found this energetic constraint. This disagreement may be because Hechinger et al.
441 (2019) mainly studied non-passerine birds, and here we studied only passernes.
442 Moreover, Hechinger et al. (2019) studied a more complete arthropod ectosymbiont
443 community (lice and mites, including a few ticks), rather than focusing on a more
444 taxonomically and ecologically restricted group as in our study (only feather mites).
445 While there may be constraints shaping the whole community of ectosymbionts (thus
446 supporting Hechinger et al.'s approach), it is also likely that different symbiont groups
447 are constrained by different host traits, or by the same host traits but in different ways.
448 For instance, unlike lice, blood-feeding mites and ticks, feather mites consume fungi,
449 bacteria and other organic matter present on feathers' surface, which are not produced
450 by the host's metabolism. Thus, this demands a different parameterization of the
451 metabolic theory equations. Interestingly, Hechinger (2013) suggested that space
452 constraints may be more relevant than energy in metabolically inactive symbiont stages
453 that do not use the energy resources provided by their hosts (e.g. because they are
454 trophically transmitted cyst stages waiting for an ultimate host to predate their current
455 intermediate host). Our findings support this view as feather mites may not consume

456 bird metabolic products, but organic material that they find on the feathers' surface
457 (Doña et al., 2019; Labrador et al., 2022). Thus, it is necessary to nurture the framework
458 proposed by Hechinger (2013) and Hechinger et al. (2019) with more knowledge about
459 the ecology and biology of the studied symbionts, and to integrate this with interspecific
460 comparative analyses to understand the relevant processes regulating symbiont
461 abundances and energy fluxes in host–symbiont systems.

462 The lack of correlation between the body size of the bird species studied here and
463 the size of their feather mites goes against the Harrison's Rule (Harrison 1915). This
464 may be the result of feather mite species showing a complex co-evolutionary history
465 with their hosts, with host-switching being as frequent as cospeciation (Doña et al.,
466 2017, 2019). In other words, mites currently found on one bird species may have
467 speciated on another host species (typically from the same genus or family). This may
468 partly explain why the smallest (*Regulus regulus*; 5.6 g) and the largest (*Pyrrhocorax*
469 *pyrrhocorax*; 287.5 g) bird species in our study have similarly sized mites (i.e., similar
470 weighted mean size of their mite species): 3.82 μ g and 2.61 μ g, respectively (Figure S2).

471 Besides the relevance of the number of barbs for mite abundance, the allometry of
472 other host traits may also have interesting implications for our understanding of the
473 entire symbiont community composed of all organisms living on bird feathers, the so-
474 called pterosphere (*sensu* Labrador et al., 2021). For instance, we showed that feather
475 mite abundance scaled with bird species's body size with a much shallower slope than
476 the wing area did (Figures 3 and 4, Table S2). Consequently, although absolute feather
477 mite abundance increased with host body size, the maximum density of feather mites
478 (i.e., Q95 feather mite abundance/cm² wing area) decreased sharply with increasing bird
479 species body size (PGLS: $t = -3.083$, $df = 86$, $p = 0.003$; Figures 5 and S5). This raises
480 the question of (1) whether a lower density of feather mites in larger bird species

481 implies a lower cleaning service provided by mites to their hosts; or (2) whether this
482 lower density is the result of a potential competition between feather mites and feather
483 lice, as numeric dominance of lice relative to mites has been observed in larger-bodied
484 bird species (Hechinger et al., 2019).

485 Overall, our study shows the potential of the theoretical and quantitative
486 framework proposed by Hechinger (2013) using the metabolic theory of ecology to
487 disentangle the mechanisms behind symbiont abundance across host species. It also
488 shows the necessity to fully integrate the biology of the studied species to make
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490

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507 TE-2021-0502).

508 **References**

509 Billerman, S.M., Keeney, B.K., Rodewald, P.G. & Schulenberg, T.S. (Eds.) (2020).

510 *Birds of the World. Cornell Laboratory of Ornithology*, Ithaca, NY, USA.

511 <https://birdsoftheworld.org/bow/home>

512 Blanco, G., Tella, J.L. & Potti, J. (1997). Feather mites on group-living red-billed
513 choughs: A non-parasitic interaction? *J. Avian Biol.*, 28, 197-206.

514 Blanco, G., Tella, J.L., Potti, J. & Baz, A. (2001). Feather mites on birds: costs of
515 parasitism or conditional outcomes? *J. Avian Biol.*, 32, 271-274.

516 Bridge, E.S. (2003). Densities and distributions of commensal feather mites
517 (*Zachvatkinia caspica*) among the primaries of Caspian Terns. *Int. J. Acarology*, 29,
518 389-398.

519 Bronstein, J.L. (1994). Conditional outcomes in mutualistic interactions. *Trends Ecol.*
520 *Evol.*, 9, 214-217.

521 Brown, J.H., Gillooly, J.F., Allen, A.P., Savage, V.M. & West, G.B. (2004). Toward a
522 metabolic theory of ecology. *Ecology*, 85, 1771-1789.

523 Bush, A.O., Lafferty, K.D., Lotz, J.M. & Shostak, A.W. (1997). Parasitology meets
524 ecology on its own terms: Margolis *et al.* revisited. *J. Parasitol.*, 83, 575-583.

525 Cade, B.S. & Noon, B.R. (2003). A gentle introduction to quantile regression for
526 ecologists. *Front. Ecol. Environ.*, 1, 412-420.

527 Choe, J.C. & Kim, K.C. (1989). Microhabitat selection and coexistence in feather mites
528 (Acari: Analgoidea) on Alaskan seabirds. *Oecologia*, 79, 10-14.

- 529 Clayton, D.H. & Walther, B.A. (2001). Influence of host ecology and morphology on
530 the diversity of Neotropical bird lice. *Oikos*, 94, 455-467.
- 531 Cunning, R. & Baker, A.C. (2014). Not just who, but how many: the importance of
532 partner abundance in reef coral symbioses. *Front. Microbiol.*, 5:400.
- 533 Díaz-Real, J., Serrano, D., Pérez-Tris, J., Fernández-González, S., Bermejo, A., Calleja,
534 J.A. *et al.* (2014). Repeatability of feather mite prevalence and intensity in passerine
535 birds. *PLoS One*, 9:e107341.
- 536 Dobson, A., Lafferty, K.D., Kuris, A.M., Hechinger, R.F. & Jetz, W. (2008). Homage to
537 Linnaeus: How many parasites? How many hosts? *Proc. Natl. Acad. Sci. U.S.A.*, 105,
538 11482-11489.
- 539 Doña, J. & Johnson, K. P. (2022). Host body size, not host population size, predicts
540 genome-wide effective population size of parasites. *bioRxiv*.
- 541 Doña, J., Potti, J., De la Hera, I., Blanco, G., Frías, Ó. & Jovani, R. (2017). Vertical
542 transmission in feather mites: insights into its adaptive value. *Ecol. Entomol.*, 42,
543 492-499.
- 544 Doña, J., Proctor, H.C., Mironov, S.V., Serrano, D. & Jovani, R. (2016). Global
545 associations between birds and vane-dwelling feather mites. *Ecology*, 97:3242.
- 546 Doña, J., Proctor, H.C., Serrano, D., Johnson, K.P., Oploo, A.O., Huguet-Tapia, J.C. *et*
547 *al.* (2019). Feather mites play a role in cleaning host feathers: New insights from
548 DNA metabarcoding and microscopy. *Mol. Ecol.*, 28, 203-218.
- 549 Doña, J., Serrano, D., Mironov, S.V., Montesinos-Navarro, A. & Jovani, R. (2018).
550 Unexpected bird-feather mite associations revealed by DNA metabarcoding uncovers
551 a dynamic ecoevolutionary scenario. *Mol. Ecol.*, 28, 379-390.

- 552 Dubinin, V.B. (1951). Feather mites (Analgesoidea). Part I. Introduction to their study.
- 553 *Fauna SSSR Paukoobraznye*, 6, 1-363 [In Russian].
- 554 Edwards, C.A. (1967). Relationships between weights, volumes and numbers of soil
555 animals. In: Graff, O., Satchell, J.E. editors. *Progress in soil biology*. North-Holland
556 Publishing Company, Amsterdam, (pp. 585-594).
- 557 Fernández-González, S., Pérez-Rodríguez, A., de la Hera, I., Proctor, H. C. & Pérez-
558 Tris, J. (2015). Different space preferences and within-host competition promote
559 niche partitioning between symbiotic feather mite species. *Int. J. Parasitol.*, 45, 655-
560 662.
- 561 Gaede, K. & Knüll, W. (1987). Water vapour uptake from the atmosphere and critical
562 equilibrium humidity of a feather mite. *Exp. Appl. Acarol.*, 3, 45-52.
- 563 Galván, I., Barba, E., Piculo, R., Cantó, J.L., Cortés, V., Monrós, J.S. *et al.* (2008).
564 Feather mites and birds: an interaction mediated by uropygial gland size? *J. Evol.*
565 *Biol.*, 21, 133-144.
- 566 Galloway, T.D. & Lamb, R.J. (2017). Abundance of chewing lice (Phthiraptera:
567 Amblycera and Ischnocera) increases with the body size of their host woodpeckers
568 and sapsuckers (Aves: Piciformes: Picidae). *Can. Entomol.*, 149, 473-481.
- 569 George-Nascimento, M., Muñoz, G., Marquet, P.A. & Poulin, R. (2004). Testing the
570 energetic equivalence rule with helminth endoparasites of vertebrates. *Ecol. Lett.*, 7,
571 527-531.
- 572 Hackett, S.J., Kimball, R.T., Reddy, S., Bowie, R.C., Braun, E.L., Braun, M.J. *et al.*
573 (2008). A phylogenomic study of birds reveals their evolutionary
574 history. *Science*, 320, 1763-1768.

- 575 Harrison, L. (1915). Mallophaga from *Apteryx*, and their significance; with a note on the
576 genus *Rallicola*. *Parasitology*, 8, 88-100.
- 577 Hatcher, M.J., Dick, J.T. & Dunn, A.M. (2012). Diverse effects of parasites in
578 ecosystems: linking interdependent processes. *Front. Ecol. Environ.*, 10, 186-194.
- 579 Hechinger, R.F. (2013). A metabolic and body-size scaling framework for parasite
580 within-host abundance, biomass, and energy flux. *Am. Nat.*, 182, 234-248.
- 581 Hechinger, R.F., Sheehan, K. L. & Turner, A.V. (2019). Metabolic theory of ecology
582 successfully predicts distinct scaling of ectoparasite load on hosts. *Proc. R. Soc.
583 Lond., B, Biol. Sci.*, 286:20191777.
- 584 Henson, S.M., Galusha, J.G., Hayward, J.L. & Cushing, J.M. (2007). Modeling territory
585 attendance and preening behavior in a seabird colony as functions of environmental
586 conditions. *J. Biol. Dyn.*, 1, 95-107.
- 587 Holland, J.N., Deangelis, D.L. & Bronstein, J.L. (2002). Population dynamics and
588 mutualism: functional responses of benefits and costs. *Am. Nat.*, 159, 231-244.
- 589 Hopkins, S.R., Wojdak, J.M. & Belden, L.K. (2017). Defensive symbionts mediate
590 host-parasite interactions at multiple scales. *Trends Parasitol.*, 33, 53-64.
- 591 Jetz, W., Thomas, G.H., Joy, J.B., Hartmann, K. & Mooers, A.O. (2012). The global
592 diversity of birds in space and time. *Nature*, 491, 444-448.
- 593 Johnson, K.P., Bush, S.E. & Clayton, D.H. (2005). Correlated evolution of host and
594 parasite body size: tests of Harrison's rule using birds and lice. *Evolution*, 59, 1744-
595 1753.
- 596 Jovani, R., Lascelles, B., Garamszegi, L.Z., Mavor, R., Thaxter, C.B. & Oro, D. (2016).
597 Colony size and foraging range in seabirds. *Oikos*, 125, 968-974.

- 598 Jovani, R. & Serrano, D. (2004). Fine-tuned distribution of feather mites (Astigmata) on
599 the wing of birds: the case of blackcaps *Sylvia atricapilla*. *J. Avian Biol.*, 35, 16-20.
- 600 Kelso, L. & Nice, M.M. (1963). A Russian contribution to anting and feather mites.
601 *Wilson Bull.*, 75, 23-26.
- 602 Klimov, P.B., Mironov, S.V. & OConnor, B.M. (2017). Detecting ancient codispersals
603 and host shifts by double dating of host and parasite phylogenies: Application in
604 proctophyllodid feather mites associated with passerine birds. *Evolution*, 71, 2381-
605 2397.
- 606 Koenker, R. (2015). quantreg: Quantile Regression. R package version 5.86.
- 607 Koenker, R. & Bassett, G. (1978). Regression quantiles. *Econometrica*, 46, 33-50.
- 608 Krasnov, B.R., Vinarski, M.V., Korallo-Vinarskaya, N.P. & Khokhlova, I.S. (2013).
609 Ecological correlates of body size in gamasid mites parasitic on small mammals:
610 abundance and niche breadth. *Ecography*, 36, 1042-1050.
- 611 Labrador, M.D.M., Doña, J., Serrano, D. & Jovani, R. (2021). Quantitative interspecific
612 approach to the stylosphere: Patterns of bacteria and fungi abundance on passerine
613 bird feathers. *Microb. Ecol.*, 81, 1088-1097. Erratum in: *Microb. Ecol.* 2021. PMID:
614 33225409.
- 615 Labrador, M.D.M., Doña, J., Serrano, D. & Jovani, R. (2022). Feather mites at night: an
616 exploration of their feeding, reproduction, and spatial ecology. *Ecology*, 103:e03550.
- 617 Lamb, R.J. & Galloway, T.D. (2019). Host body size and the abundance of chewing lice
618 (Phthiraptera: Amblycera, Ischnocera) infesting eight owl species (Aves:
619 Strigiformes) in Manitoba, Canada. *Can. Entomol.*, 151, 621-628.

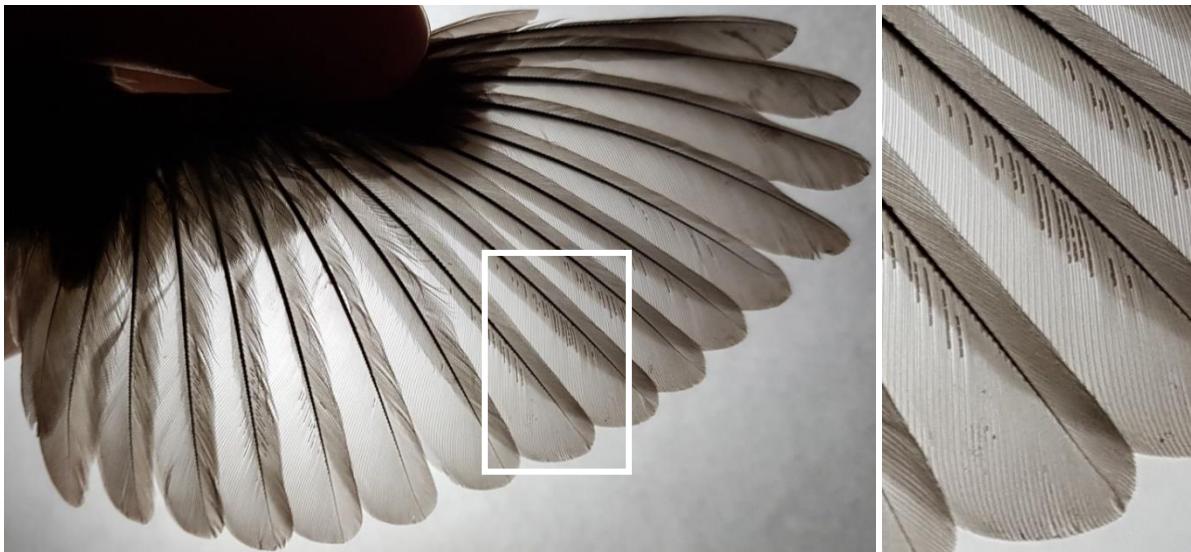
- 620 Larsen, B.B., Miller, E.C., Rhodes, M.K. & Wiens, J.J. (2017). Inordinate fondness
621 multiplied and redistributed: the number of species on earth and the new pie of life.
622 *Q. Rev. Biol.*, 92, 229-265.
- 623 Marčanova, N. & Janiga, M. (2021). Survival strategies and seasonal size variations of
624 feather mites *Proctophyllodes megaphyllus* on their host Alpine Accentor *Prunella*
625 *collaris*. *Pol. J. Ecol.*, 69, 25-34.
- 626 Maestri, R., Fiedler, M.S., Shenbrot, G.I., Surkova, E.N., Medvedev, S.G., Khokhlova,
627 I.S. *et al.* (2020). Harrison's rule scales up to entire parasite assemblages but is
628 determined by environmental factors. *J. Anim. Ecol.*, 89, 2888-2895.
- 629 Mennerat, A., Charmantier, A., Perret, P., Hurtrez-Boussès, S. & Lambrechts, M.M.
630 (2021). Parasite intensity is driven by temperature in a wild passerine bird. *Peer*
631 *Community Journal*, 1:e60.
- 632 Mestre, A., Mesquita-Joanes, F., Proctor, H.C. & Monrós, J.S. (2011). Different scales
633 of spatial segregation of two species of feather mites on the wings of a passerine
634 bird. *J. Parasitol.*, 97, 237-244.
- 635 Mironov, S.V. (2022). Notes on the systematics of the feather mite genus *Trouessartia*
636 Canestrini, 1899 (Acariformes: Trouessartiidae) with an updated world checklist.
637 *Acarina*, 30, 157-180.
- 638 Mironov, S.V. & Malyshev, L.L. (2002). Dynamics of infection of *Fringilla coelebs*
639 chaffinch nestlings with feather mites (Acari: Analgoidea). *Parazitologiya*, 36:356-
640 374 [In Russian with English abstract].
- 641 Møller, A.P., Czirjak, G.Á. & Heeb, P. (2009). Feather micro-organisms and uropygial
642 antimicrobial defences in a colonial passerine bird. *Funct. Ecol.*, 23, 1097-1102.

- 643 Morand, S. (2015). (macro-) Evolutionary ecology of parasite diversity: From
644 determinants of parasite species richness to host diversification. *Int. J. Parasitol.*
645 *Parasites Wildl.*, 4, 80-87.
- 646 Morand, S. & Poulin, R. (2002). Body size-density relationships and species diversity in
647 parasitic nematodes: patterns and likely processes. *Evol. Ecol. Res.*, 4, 951-961.
- 648 Muzaffar, S.B. & Jones, I.L. (2005). Population structure, distribution patterns and
649 precopulatory mate-guarding in the feather mite *Alloptes* Canestrini, 1879 (Acari:
650 Anagoidea: Alloptidae) on auks (Charadriiformes: Alcidae) at the Gannet Islands,
651 Labrador, Canada. *Int. J. Acarology*, 31, 407-416.
- 652 OConnor, B.M. (1982). Evolutionary ecology of astigmatid mites. *Annu. Rev. Entomol.*,
653 27, 385-409.
- 654 Orme, D., Freckleton, R., Thomas, G., Petzoldt, T., Fritz, S., Isaac, N. *et al.* (2013).
655 caper: Comparative Analysis of Phylogenetics and Evolution in R. *R package*
656 *version 5*, 1-36.
- 657 Pap, P.L., Vágási, C.I., Osváth, G., Mureşan, C. & Barta, Z. (2010). Seasonality in the
658 uropygial gland size and feather mite abundance in house sparrows *Passer*
659 *domesticus*: natural covariation and an experiment. *J. Avian Biol.*, 41, 653-661.
- 660 Peet, R., Kirk, A. & Behnke, J.M. (2022). A long-term study of temporal variation in
661 wing feather mite (Acari: Astigmata) infestations on robins, *Erithacus rubecula*, in
662 Nottinghamshire, U.K. *J. Zool.*, 316, 296-306.
- 663 Peters, R.H. (1986). *The Ecological Implications of Body Size* (Cambridge Studies in
664 Ecology). Cambridge University Press, Cambridge, UK.
- 665 Poulin, R. (1999). Body size vs abundance among parasite species: positive
666 relationships? *Ecography*, 22, 246-250.

- 667 Poulin, R. & George-Nascimento, M. (2007). The scaling of total parasite biomass with
668 host body mass. *Int. J. Parasitol.*, 37, 359-64.
- 669 Presley, S.J. & Willig, M.R. (2008). Intraspecific patterns of ectoparasite abundances on
670 Paraguayan bats: effects of host sex and body size. *J. Trop. Ecol.*, 24, 75-83.
- 671 Proctor, H.C. (2003). Feather mites (Acari: Astigmata): Ecology, behavior, and
672 evolution. *Annu. Rev. Entomol.*, 48, 185-209.
- 673 Rózsa, L. (1997a). Patterns in the abundance of avian lice (Phthiraptera: Amblycera,
674 Ischnocera). *J. Avian Biol.*, 28, 249-254.
- 675 Rózsa, L. (1997b). Wing-feather mite (Acari: Proctophyllodidae) abundance correlates
676 with body mass of passerine hosts: a comparative study. *Can. J. Zool.*, 75, 1535-
677 1539.
- 678 Rubolini, D., Liker, A., Garamszegi, L.Z., Møller, A.P. & Saino, N. (2015). Using the
679 BirdTree.org website to obtain robust phylogenies for avian comparative studies: A
680 primer. *Curr. Zool.*, 61, 959-965.
- 681 Stefan, L.M., Gómez-Díaz, E., Elguero, E., Proctor, H.C., McCoy, K.D., González-
682 Solís, J. (2015). Niche partitioning of feather mites within a seabird host, *Calonectris*
683 *borealis*. *PLoS One*, 10:e0144728.
- 684 Sukumaran, J. & Holder, M.T. (2010). DendroPy: A Python library for phylogenetic
685 computing. *Bioinformatics*, 26, 1569-1571.
- 686 Sukumaran, J. & Holder, M.T. (2015). *SumTrees*: Phylogenetic Tree Summarization
687 and Annotation 4.5.1.

- 688 Surkova, E.N., Warburton, E.M., van der Mescht, L., Khokhlova, I.S. & Krasnov, B.R.
- 689 (2018). Body size and ecological traits in fleas parasitic on small mammals in the
- 690 Palearctic: larger species attain higher abundance. *Oecologia*, 188, 559-569.
- 691 Symonds, R.E. & Blomberg, S.P. (2014). A primer on phylogenetic generalised least
- 692 squares (PGLS). In: Garamszegi, L.Z. editor. *Modern Phylogenetic Comparative*
- 693 *Methods and Their Application in Evolutionary Biology: Concepts and Practice*.
- 694 Springer, Berlin. (pp. 105-130).
- 695 Turgeon, G., Kutz, S.J., Lejeune, M., St-Laurent, M.-H. & Pelletier, F. (2018). Parasite
- 696 prevalence, infection intensity and richness in an endangered population, the
- 697 Atlantic-Gaspésie caribou. *Int. J. Parasitol. Parasites Wildl.*, 7, 90-94.
- 698 Walter, D.E. & Proctor, H.C. (2013). *Mites: Ecology, Evolution and Behavior. Life at a*
- 699 *Microscale*. Second edition. Springer, Dordrecht, UK.
- 700 Wiles, R., Cameron, J., Behnke, J.M., Hartley, I.R., Gilbert, F.S. & McGregor, P.K.
- 701 (2000). Season and ambient air temperature influence the distribution of mites
- 702 (*Proctophyllodes stylifer*) across the wings of blue tits (*Parus caeruleus*). *Can. J.*
- 703 *Zool.*, 78, 1397-1407.
- 704 Yamasaki, Y.K., Graves, E.E., Houston, R.S., OConnor, B.M., Kysar, P.E., Straub,
- 705 M.H. *et al.* (2018). Evaluation of *Proctophyllodes huizilopochtlii* on feathers from
- 706 Anna's (*Calypte anna*) and Black-chinned (*Archilochus alexandri*) Hummingbirds:
- 707 Prevalence assessment and imaging analysis using light and tabletop scanning
- 708 electron microscopy. *PLoS One*, 13:e0191323.
- 709
- 710
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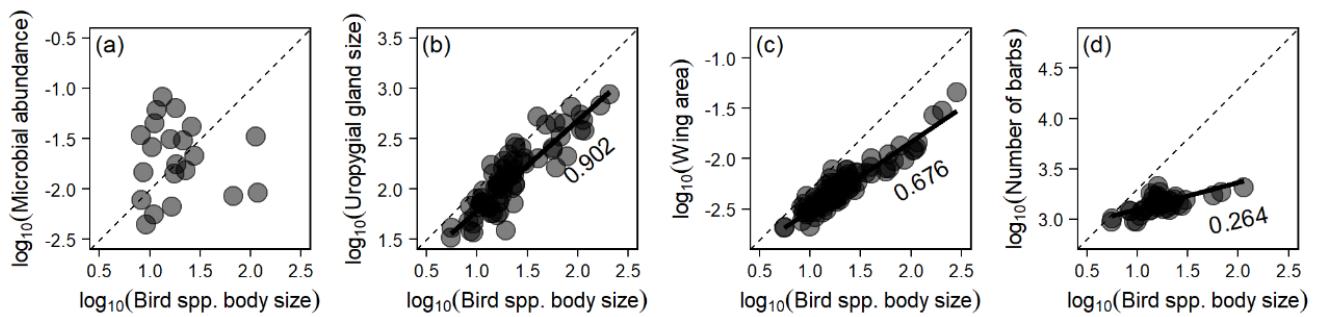
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720 **Figure 1:** Feather mites (*Proctophyllodes sylviae*) on the wing of a *Sylvia atricapilla*.

721 Note their strong aggregation in certain feathers along the wing and some sections
722 within those feathers, and their queuing along feather barbs.

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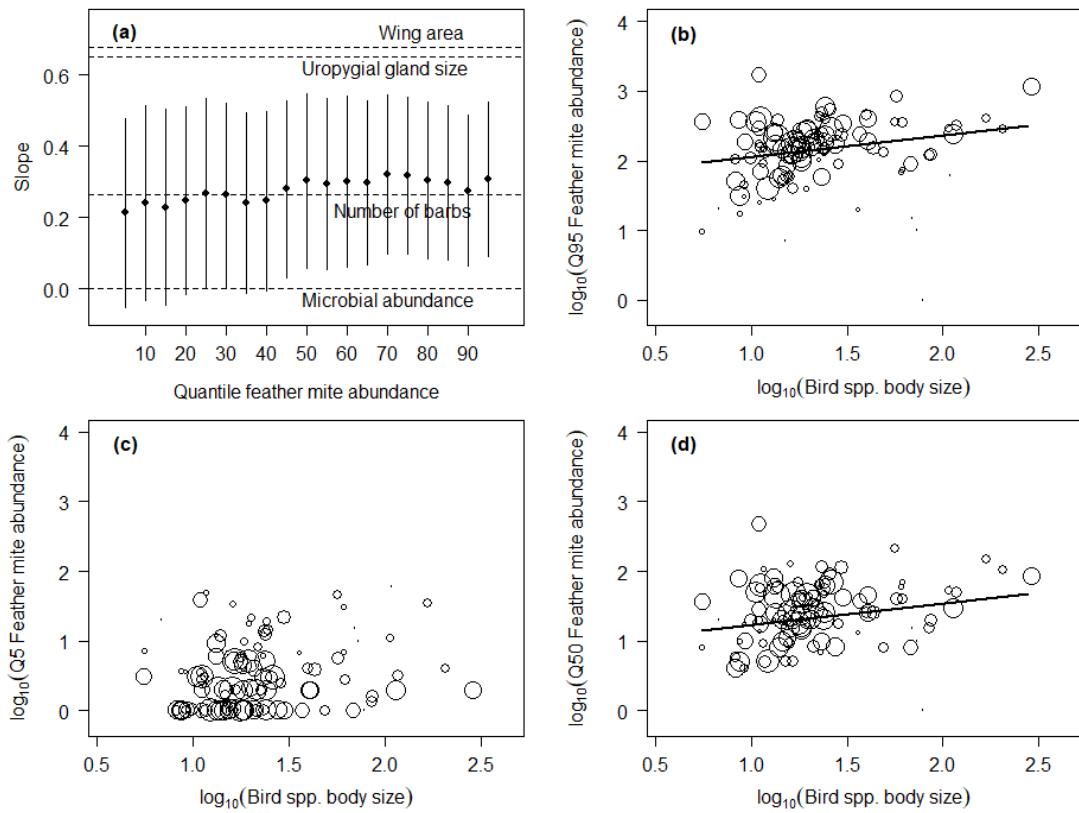
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725

726 **Figure 2:** Relationships between potential energetic (microbial abundance N = 21
727 species, uropygial gland size N = 76) and spatial (wing area N = 88, or number of barbs
728 N = 44) constraints against bird species body size (in g). Dashed lines show slope = 1.
729 Only slopes that departed from 0 (p-value < 0.05) are drawn (black line) and its
730 estimated value is shown.

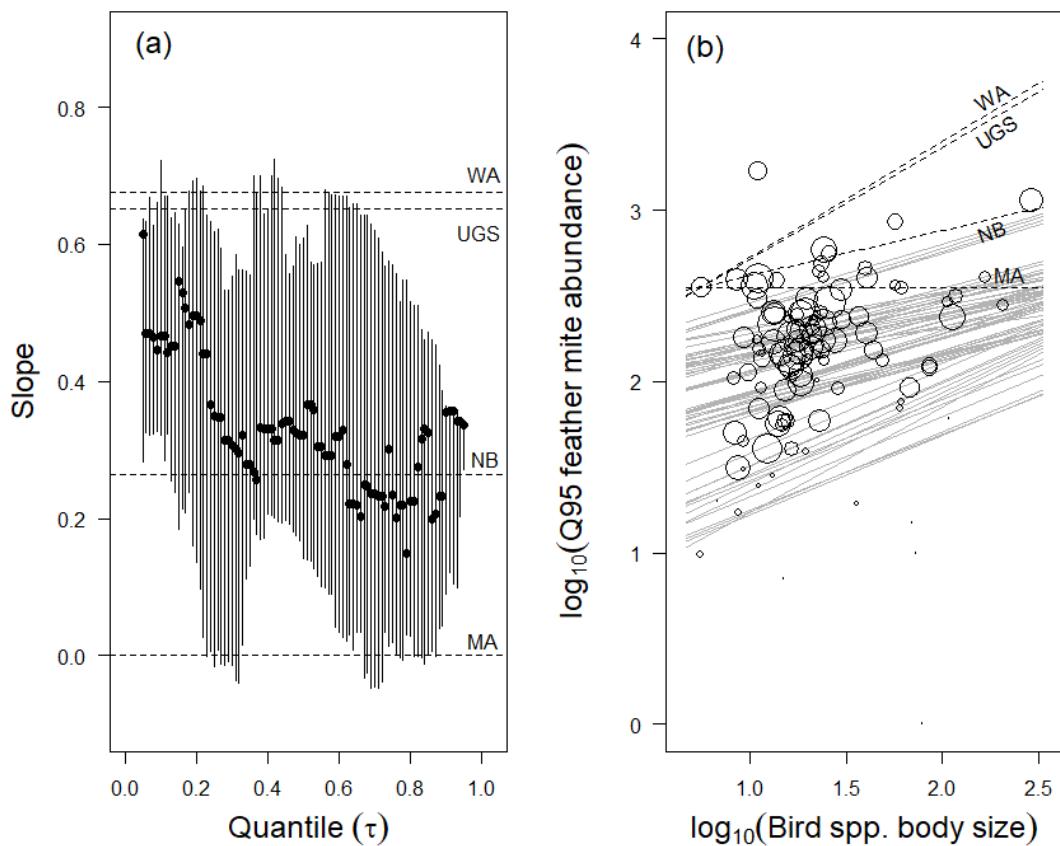
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732

733 **Figure 3:** PGLS models for the relationship between 19 quantiles (from Q5 to Q95) of
734 feather mite abundance in each bird species as dependent variable and \log_{10} (bird species
735 body size) and \log_{10} (feather mite body size) as independent variables. (a) Slopes ($\pm 95\%$
736 CI) for \log_{10} (bird species body size) are shown as dots and whiskers. Dashed lines show
737 slope predictions according to Eq. 3 (microbial abundance), Eq. 4 (uropygial gland
738 size), Eq. 5 (wing area), and Eq. 6 (number of barbs). (b) to (d) three examples of the
739 relationship between bird body size (in g) and feather mite abundance at different
740 quantiles (Q95, Q5, and Q50, respectively) from which slopes for plot (a) were
741 obtained. Dot size is proportional to the \log_{10} (sample size) for each bird species. Only
742 regression lines with slopes differing from 0 are shown.

743

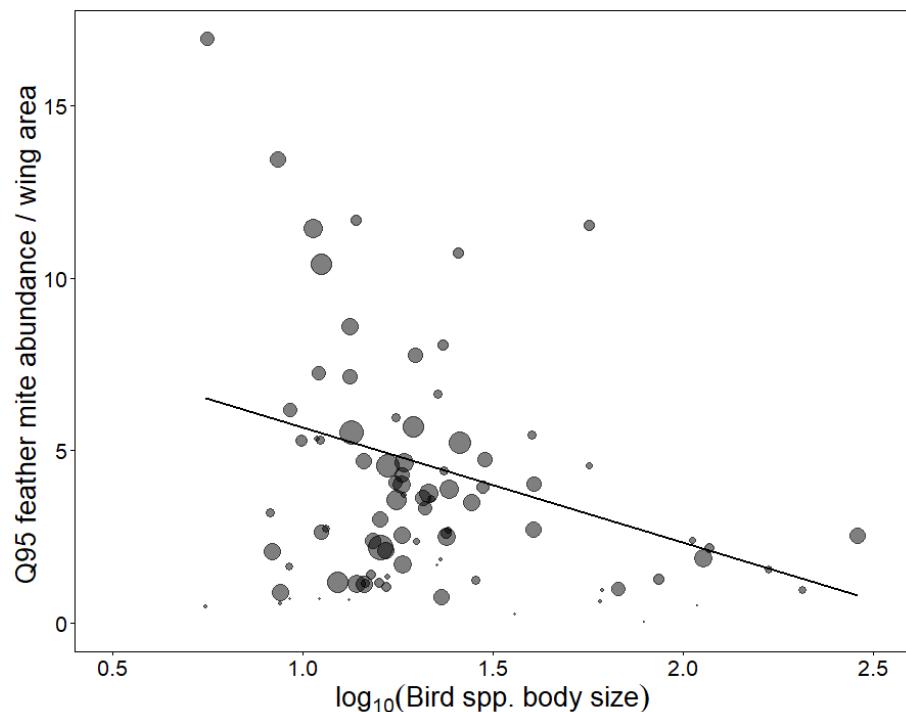


744

745 **Figure 4:** Multivariable quantile regression on $\log_{10}(\text{Q95 feather mite abundance})$ with
746 $\log_{10}(\text{bird species body size})$ and $\log_{10}(\text{feather mite body size})$ as independent variables.
747 Dashed lines show slope predictions according to Eq. 3 (microbial abundance, MA), Eq.
748 4 (uropygial gland size, UGS), Eq. 5 (wing area, WA), and Eq. 6 (number of barbs,
749 NB). (a) Bird species body size slopes ($\pm 95\%$ CI) for each tau (τ) value. (b) Quantile
750 regression of the allometry of Q95 feather mite abundance (continuous gray lines) and
751 predicted slopes (black dashed lines). Dot size is proportional to the $\log_{10}(\text{sample size})$
752 for each bird species.

753

754



755

756

757 **Figure 5:** Relationship between $\log_{10}(\text{bird species body size})$ (in g) and the maximum
758 density of their feather mites (Q95 feather mite abundance/cm² of wing area). Dot size
759 is proportional to the $\log_{10}(\text{sample size})$ for each bird species.