

TITLE

Intron retention of an adhesion GPCR generates single transmembrane-helix isoforms to enable 7TM-adhesion GPCR function

AUTHORS

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SUMMARY

Adhesion G protein-coupled receptors (aGPCR) function as metabotropic mechanosensors in the nervous system and other organs. aGPCR are heavily spliced forecasting an extraordinary molecular structural diversity. Many predicted isoforms lack the transmembrane (7TM) signaling subunit, but to what extent these non-GPCR isoforms are produced and what physiological purpose they serve is unknown. Alternative splicing through intron retention of *ADGRL/Latrophilin/Cirl* mRNA in *Drosophila* generates transcripts encoding unconventional proteins with an extracellular domain anchored by a single transmembrane helix (Cirl^{1TM}). Here, we show that *Cirl*^{1TM} transcripts are translated *in vivo* and that Cirl^{1TM} binds Cirl^{7TM} N-terminal fragment-dependently. This interaction enables mechanosensory neurons to distinguish input intensities through Gα_o-dependent signaling. Similarly, a direct interaction was found for mammalian GPR126/ADGRG6 isoforms. Together, our findings define intron retention and isoform-specific heteromerization as extraordinary molecular strategies to adjust *Cirl*-dependent mechanosensation and demonstrate physiological relevance of versatile aGPCR isoform repertoire to tune cellular responsiveness.

Key words: Latrophilin, Cirl, aGPCR, isoforms, mechanobiology, sensory neurons, proprioception

INTRODUCTION

Mechanosensing is vital for nervous system development, morphology and function¹⁻⁴. Disturbances of the delicate mechanobiological system of nervous tissue are pathologically relevant and linked to traumatic brain injury, multiple sclerosis and brain cancer^{2,5-8}. Unfortunately, thus far, detailed knowledge about the molecular mechanisms at play remains scarce.

Cells of the nervous system can express several dozen G protein-coupled receptors (GPCR)⁹. Their signature layout with an extracellular N terminus, intracellular C terminus and a seven-transmembrane helix-spanning domain (ECD, ICD, 7TM domain respectively) is well-established and considered basic to transduce extracellular into intracellular signals. This way, GPCR assume roles in fundamental neurobiological processes in neurons and glia^{10,11}. On the pathological flip side, GPCR are considered one of *the* pharmaceutical targets to treat neurological disorders¹². In recent years, adhesion-type GPCR (aGPCR), a class within the GPCR superfamily, have emerged as metabotropic mechanosensors, relevant in several organ systems including the nervous system¹³⁻¹⁹. For example, in zebrafish, rapid electrical signal propagation depends on ADGRG6/GPR126 as it times the onset of peripheral neuron myelination likely by probing the matrix rigidity at the neuron-Schwann cell interface^{13,20}. In *Drosophila*, sensitivity towards sound, touch and proprioceptive cues hinges on ADGRL/Latrophilin/Cirl function^{21,22}. In mammals, vision and hearing are associated with ADGRV1/VLGR1 mediating mechanical stretch response through its association with focal adhesion contacts²³⁻²⁶.

aGPCR are encoded by large genomic loci and are known to be heavily alternatively spliced^{27,28}. However, there are only few examples where isoform-specific functions of aGPCR have been uncovered. For example, the S4 variant of ADGRG1/GPR56 with its shortened N-terminal fragment (NTF) was shown to be a vital regulator in synaptic pruning^{29,30}; an N-terminally truncated ADGRL3/LPHN3 isoform shapes the transmission of cone photoreceptor synapses³¹. Outside of the nervous system, different ADGRL3/LPHN3 isoforms (with a varying loop region) decrease insulin secretion from isolated pancreatic β cells³². Recent bioinformatic efforts helped to systematically catalog the aGPCR transcriptome²⁷. Fascinatingly, ~ 22% of all aGPCR genes produce transcripts that lack the sequence encoding a complete 7TM domain²⁷, rendering G protein coupling to 7TM-less receptor isoforms highly unlikely. This begs questions as to whether such transcript variants are translated into protein, especially *in vivo*, how their expression varies across time and space, if they contribute to physiologically relevant cellular functions, and if these extend beyond an adhesive role. To address these questions, we studied the ADGRL/Latrophilin homolog *Cirl* (Ca^{2+} -independent receptor of latrotoxin) from *Drosophila*. *Cirl* is predicted to undergo alternative splicing to generate transcripts encoding peculiar non-GPCR proteins with only NTF, the first transmembrane helix and an alternative C terminus (*Cirl*^{1TM}). *Cirl* was shown to be relevant in mechanosensing. It suppresses cAMP levels and likely modulates gating properties of ionotropic mechanosensors in proprioceptive sensory

neurons of the lateral pentascolopodial chordotonal (lch5) organ, a well-defined set of cells accessible to genetic, optical and functional approaches^{21,22}.

Here, we report that *Cir1*^{1TM} transcripts are produced *in vivo*, through intron retention and that they are co-expressed with *Cir1*^{7TM} transcripts throughout development. Contradictory to previous accounts suggesting degradation and/or physiological dispensability of intron retention-based molecules³³, we demonstrate the expression of the non-GPCR protein Cir1^{1TM} in a living animal. Moreover, we show that Cir1^{1TM} forms a heteromeric complex with Cir1^{7TM} through its NTF. Strikingly, our functional analyses provide evidence that these molecular Cir1^{1TM}-Cir1^{7TM} assemblies define the discriminative bandwidth of proprioceptive sensory neurons towards mechanical stimulation, a function conferred through Gα_o protein-dependent signaling. Importantly, we documented a similar complex formation for mammalian ADGRG6^{7TM} and unconventional ADGRG6^{1TM} proteins.

In sum, our findings uncover that alternative splicing of aGPCR enables cooperation between GPCR-like and non-GPCR isoforms to adjust cellular mechanophysiology providing a promising entry point for experimental and therapeutic approaches to interfere with aGPCR signals.

RESULTS

Intron retention produces *Cir1* transcripts encoding molecules that contain a single TM domain and alternative intracellular region

In *Drosophila*, Cir1/Latrophilin/ADGRL is encoded by a single gene²². Alternative splice events of *Cir1* pre-mRNA produce eight different transcripts (flybase.org) putatively encoding six unique proteins (Figures 1A, S1A,B). The majority of transcripts encode classical GPCR that contain a 7TM domain allowing G protein engagement, β-arrestin binding and canonical intracellular signaling (Cir1^{7TM} = *Cir1-G*, *-I*, *-B*, *-E*, *-H*; Figures 1A, S1A,E). However, a substantial fraction of transcripts encode proteins containing only the N terminus integrated in the plasma membrane through a single TM helix (Cir1^{1TM} = *Cir1-J*, *-F*, *-K*; Figures 1A, S1B,E). *Cir1*^{1TM} transcripts likely arise through intron retention, where a specific splice donor is ignored and intron 6 is retained. The resulting transcripts are either degraded or translated. In the latter case, translation would continue until an intronic in-frame stop codon is reached producing 207 alternative amino acid residues of the putative Cir1^{1TM} isoforms (Figure 1A).

To test for the presence of *Cir1*^{1TM} transcripts, we used *Cir1*^{1TM} and *Cir1*^{7TM}-specific primer sets to conduct RT-PCR analyses on cDNA libraries generated from adult and larval *w*¹¹¹⁸ *Drosophila*. We amplified *Cir1*^{1TM} and *Cir1*^{7TM}-specific DNA fragments (Figure 1A,B), implying co-expression of both in the developing and adult fly. To exclude the possibility that small amounts of yet to be degraded *Cir1*^{1TM} transcripts served as PCR template and to investigate whether *Cir1*^{1TM} and *Cir1*^{7TM} are consistently co-expressed, we quantified the amount of each *Cir1* transcript generated at consecutive time points during embryonic, larval, pupal, and adult development employing an RNA-sequencing data set (Figure S1C,D;³⁴). *Cir1*^{7TM} and *Cir1*^{1TM} transcripts show similar ontogenetic expression profiles

with peak abundances during embryonic and pupal stages (Figures 1C, S1C,D). *Cirl*^{1TM} and *Cirl*^{7TM} account for ~ 22.5% and ~ 77.5% of total *Cirl* transcript produced, respectively (Figure S1E). These data validate the expression of *Cirl*^{1TM} transcripts and suggest that *Cirl*^{1TM} and *Cirl*^{7TM} transcript expression is co-regulated. This implies the need of both transcript species during specific developmental periods.

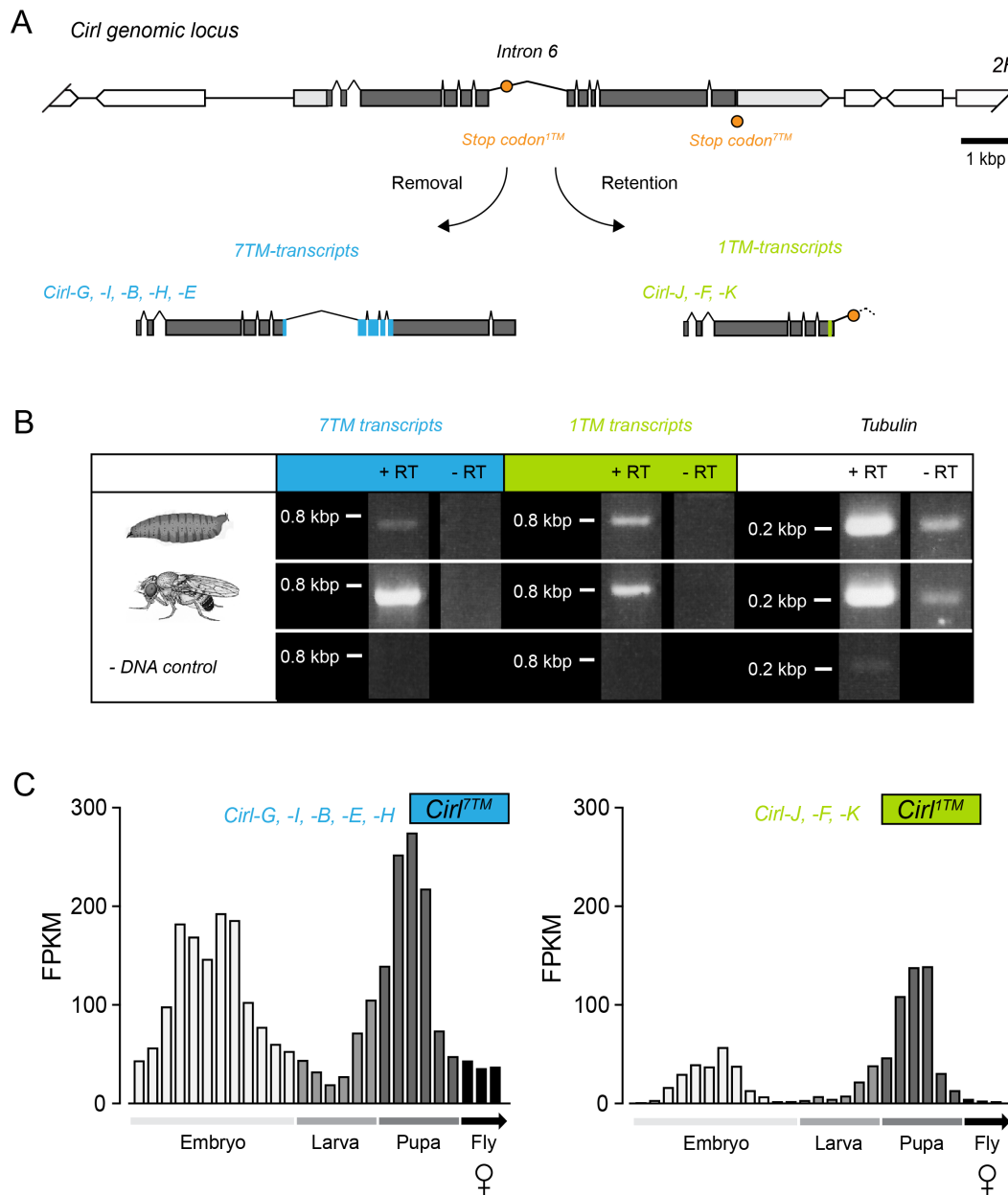


Figure 1. Intron retention produces *Cirl* transcripts encoding molecules with a single TM and alternative intracellular region

(A) Schematic of the *Cirl* gene locus. Removal of intron 6 gives rise to five *Cirl*^{7TM} transcripts: *Cirl*-G, -I, -B, -E, and -H (in blue). Retention of intron 6 generates three *Cirl*^{1TM} transcripts: *Cirl*-J, -F, -K (in green). Depicted are only protein coding portions of the transcripts. Resulting proteins are shown in Suppl. Figure S1.

(B) Validation of *Cir1^{1TM}* transcript in *Drosophila*. RT-PCR products amplified from larval and adult cDNAs for *Cir1^{7TM}* (blue column; 0.7 kbp) and *Cir1^{1TM}* transcripts (green column; 0.8 kbp). Tubulin served as loading and quality control of cDNA and RT-PCR; no DNA: specificity control.

(C) Cumulative fragments per kilobase million (FPKM) values of *Cir1^{7TM}* (left panel) and *Cir1^{1TM}* (right panel) at several developmental time points. While *Cir1^{7TM}* and *Cir1^{1TM}* have a similar ontogenetic expression profile, *Cir1^{7TM}* transcript quantity was higher than that of *Cir1^{1TM}* throughout development. Embryo (white): 12 time points equally distributed between 0 and 24 h; larval stages (light gray) L1, L2 and L3 (split across 4 time points) in days; pupa (dark gray): 6 time points across 4 days; female flies (black): 1, 5, 30 days old. The individual transcript quantity is given in Suppl. Figure S1.

Cir1^{1TM} protein is expressed *in vivo*

To ascertain if Cir1^{1TM} protein is expressed *in vivo*, we used the previously generated *Cir1^{KO}-attP* allele to knock-in *Cir1* variants that encode Cir1^{1TM} isoforms containing V5-tags at the C terminus and Cir1^{7TM} isoforms containing Flag-tags in the third intracellular loop, thus enabling their specific detection through matching antibodies. In addition, a monomeric RFP (mRFP) in the NTF of Cir1 (Figure 2A,B)²¹ was introduced to detect the entire isoform repertoire. This allele is referred to as *Cir1^{3x-tagged}*. Since RNA-sequencing data suggested elevated expression of *Cir1^{1TM}* during pupal stages (Figures 1C, S1C,D), pupal protein extracts were analyzed in Western blots using a monoclonal α-V5 antibody to detect a ~33 kDa fragment corresponding to the C-terminal fragment (CTF) of Cir1^{1TM} (Figure 2C).

GPCR autoproteolysis-inducing (GAIN) domain-dependent autoproteolytic processing presents one of the hallmark features of aGPCR and underlies their bipartite NTF/CTF structure. Autoproteolysis of Cir1 can be averted by mutating the GPCR proteolysis site (GPS)²¹, at which nascent receptors are self-cleaved. To further confirm the expression of Cir1^{1TM} proteins, we generated a cleavage-deficient version of *Cir1^{3x-tagged}* via a -2 GPS H>A mutation (*Cir1^{3x-tagged/H>A}*) and recovered full-length (FL) Cir1^{1TM} protein (~137 kDa; Figure 2C). Consistent with this finding, Western blot analysis of head homogenates from *Cir1^{3x-tagged}* and *Cir1^{3x-tagged/H>A}* animals showed Cir1-NTF (~110 kDa), FL Cir1^{1TM} (~135 kDa) and FL Cir1^{7TM} (~215 kDa; Figure S2) expression also in the central nervous system of adult flies. Thus, *Cir1^{1TM}* transcripts are translated to produce Cir1^{1TM} proteins *in vivo*.

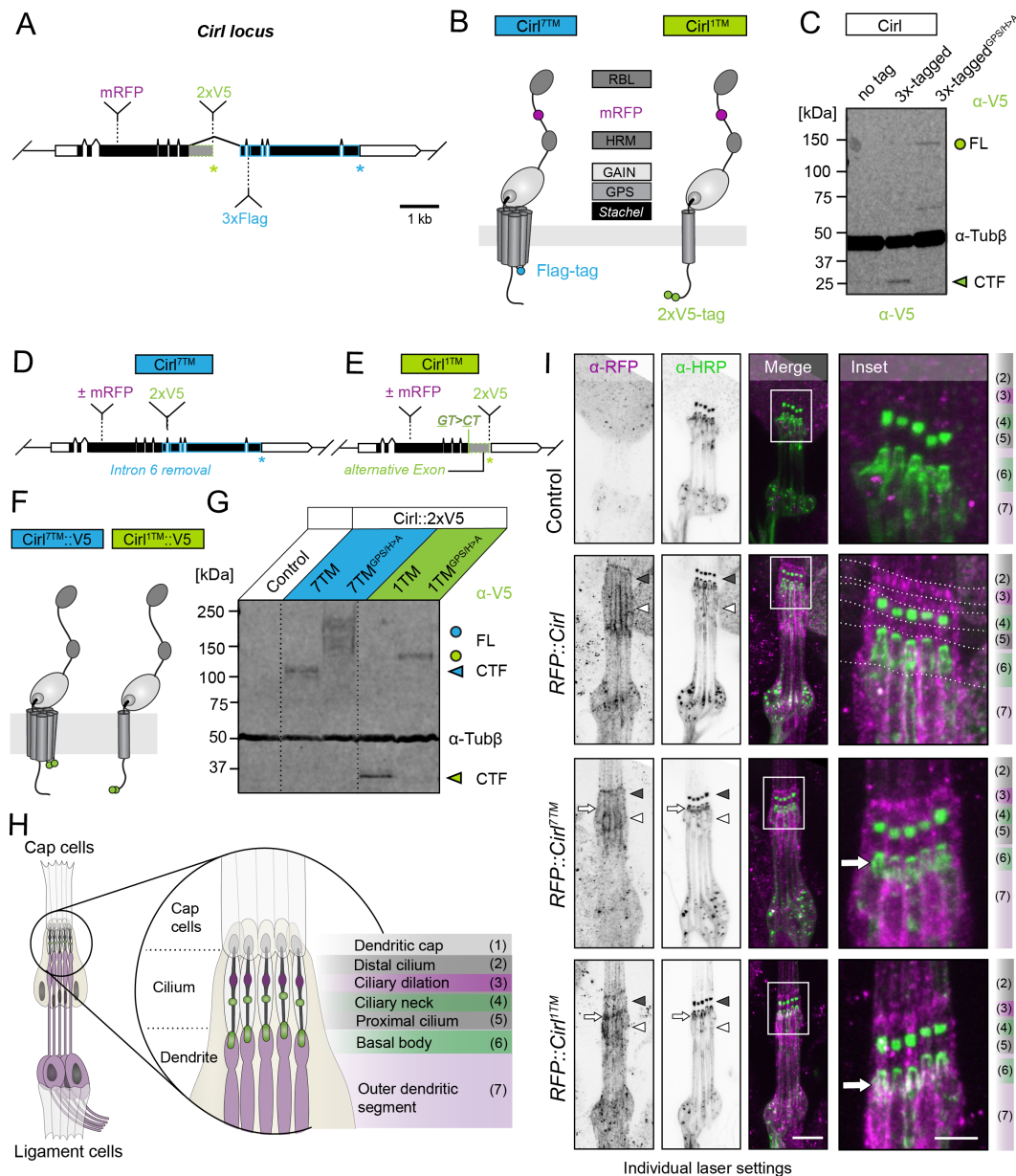


Figure 2. *Cirl*^{1TM} protein is expressed in mechanosensory neurons

(A) Schematic of *Cirl* locus. Insertion sites of an mRFP (indicated in magenta), a tandem V5 tag (2xV5, in green) and a triple Flag tag (3xFlag, in blue) are indicated.

(B) Tagging strategy of *Cirl*^{7TM} and *Cirl*^{1TM} proteins. mRFP was inserted in the linker region between RBL and HRM domain²¹. Tandem V5 tag is located at the C terminus of *Cirl*^{1TM}, triple Flag tag is included in the third intracellular loop of *Cirl*^{7TM}.

(C) Western blot of lysates from pupae, which express V5-tagged *Cirl*^{1TM} protein. Bands at ~ 33 kDa (green arrowhead) and ~ 136 kDa (green circle) correspond to the expected molecular weight of CTF and FL *Cirl*^{1TM}, respectively. Additional band at ~ 70 kDa of unknown identity. Tubulin served as a loading control (α -Tub β). Related to Figure S2.

- (D) Schematic layout of *Cir1* allele exclusively expressing *Cir1*^{7TM}. Intron 6 encoding *Cir1*^{1TM}-C-terminus was removed.
- (E) Schematic layout of *Cir1* allele exclusively expressing *Cir1*^{1TM}. Splice donor at the exon 6-intron 6 boundary was mutated (GT > CT) and 7TM-specific exons and introns were deleted.
- (F) Schematic of *Cir1*^{7TM} and *Cir1*^{1TM} proteins fused to 2xV5 in the third intracellular loop (left) and the very C-terminal end (right), respectively.
- (G) Western blot analysis of head lysates from *Cir1*^{7TM}::V5 or *Cir1*^{1TM}::V5 showed CTFs with molecular mass of ~ 110 kDa and 33 kDa, respectively. Cleavage-deficient versions with predicted molecular masses of ~ 190 kDa (FL *Cir1*^{7TM-GPS/H>A}::V5) and ~136 kDa (FL *Cir1*^{1TM-GPS/H>A}::V5). *Cir1*^{Rescue} = untagged control. Tubulin served as a loading control (α -Tub β).
- (H) Schematic illustration of lch5 organ with 5 bipolar sensory neurons and typical anatomical features, numbered (1) – (7).
- (I) Subcellular localization of *Cir1* isoform species in the lch5 organ (intensity inverted α -RFP and α -HRP channels are shown in black and white). Maximum z-projections of confocal images show RFP::*Cir1*^{7TM} at the level of the cilium (gray arrowheads), in the ciliary dilation (3), outer dendritic segments [(7), white arrowheads]. RFP::*Cir1*^{1TM} location overlaps with RFP::*Cir1*^{7TM} at similar subcellular region. In addition RFP::*Cir1*^{1TM} is located at the basal bodies [(6), indicated by arrow]. *Cir1*^{Rescue} = Control (upper row). α -HRP recognizes neuronal membranes and was used to outline the lch5. Scale bar 10 μ m, inset: 5 μ m. Related to Figure S3.

***Cir1*^{1TM} protein localizes to mechanosensory neurons**

Previously, we reported on *Cir1*'s role in mechanosensitive lch5 neurons^{21,22} but the *Cir1*^{KO} allele used in those studies removes both *Cir1*^{7TM} and *Cir1*^{1TM} isoforms precluding analyses of their individual contributions. Consequently, we set out to decipher isoform-specific functions of *Cir1*, especially *Cir1*^{1TM} proteins.

To gain genetic control over *Cir1* isoform expression, we engineered *Cir1* alleles for exclusive expression of either *Cir1*^{1TM} or *Cir1*^{7TM} gene products under endogenous transcriptional control (Figure 2D,E). Exclusive *Cir1*^{7TM} expression was assured by removing the intron (intron 6 of isoform -I and intron 5 of -E, -B/H, -G) which otherwise could encode the *Cir1*^{1TM}-specific alternative intracellular tail (Figure 2D). Exclusive *Cir1*^{1TM} expression was achieved by a silent mutation of the splice donor (GT $\rightarrow$ CT) at the exon 6/intron 6 boundary and removal of the *Cir1*^{7TM}-specific genetic region (Figure 2E). To validate this genetic strategy, we conducted Western blot analyses from head homogenates of flies in which we inserted biochemical V5 tags at the C terminus of *Cir1*^{1TM} (wild-type: *Cir1*^{1TM}::V5; cleavage-deficient: *Cir1*^{1TM/H>A}::V5; Figure 2F,G) and the third intracellular loop *Cir1*^{7TM} proteins (wild-type:

Cir1^{7TM}::V5; cleavage-deficient: Cir1^{7TM/H>A}::V5; Figure 2F,G). We detected the expected FL and CTF bands of both Cir1^{1TM}::V5 and Cir1^{7TM}::V5 fusion proteins (Figure 2G).

α -V5 immunofluorescent detection via confocal imaging of Cir1^{1TM}::V5 in lch5 organs failed due to high signal background (data not shown). To locate Cir1^{1TM} in lch5 organs, we generated an independent set of Cir1^{1TM} or Cir1^{7TM} alleles carrying an additional mRFP within the NTF (referred to as RFP::Cir1^{1TM}::V5 and RFP::Cir1^{7TM}::V5)²¹ (Figure S3). α -RFP immunolabeling and microscopy of larval lch5 organs revealed specific RFP::Cir1^{1TM}::V5 and RFP::Cir1^{7TM}::V5 signals at the level of the ciliary and outer dendritic segments as well as at the ciliary dilation (Figure 2H,I), demonstrating that both Cir1 isoform species co-localize in these subcellular compartments. Moreover, in the outer dendritic segment, particularly around the basal body, we detected mostly RFP::Cir1^{1TM}::V5 (Figure 2I) showing exclusive localization of this isoform. Thus, mechanosensory lch5 neurons express both Cir1^{7TM} and Cir1^{1TM} proteins.

Cir1^{1TM} and Cir1^{7TM} proteins are co-required for neuronal discrimination of mechanical inputs

Drosophila larvae rely on *Cir1*-dependent proprioceptive feedback from lch5 function for locomotion (Figure 3A,B)^{22,35}. This begs the question whether Cir1^{1TM}, Cir1^{7TM} or both isoform species are involved in this behavior. Quantification of the average midpoint crawling speed of L3 larvae showed that sole expression of Cir1^{7TM} suffices to revert values back to those of the *Cir1^{Rescue}*. Interestingly, exclusive presence of Cir1^{1TM} even enhances movement velocity above *Cir1^{Rescue}* values (Figure 3A,B, Table S3). While this may implicate that Cir1^{7TM} suppresses Cir1^{1TM} function relevant for this particular behavior, coordinated locomotion results from numerous neurons and neuronal processes, some of which may require Cir1. To pinpoint the functional relevance of different Cir1 isoform species, we performed electrophysiological recordings from mechano-stimulated lch5 neurons. Previously, measurements from lch5 neurons showed that *Cir1^{KO}* larvae discriminate less robustly between different vibrational stimulation frequencies than *Cir1^{Rescue}*^{21,22}. The used direct vibration stimulation lacked control of stimulus amplitude^{21,22}. To increase stimulation precision in lch5 recordings and to emulate proprioceptive stretch-induced lch5 activity more adequately, we attached a miniature hook to a piezo element and pulled at the organ using defined solitary mechanical square pulses (Figure 3C,D). The hook was rapidly moved by defined distances between 0.03 and 3 μ m perpendicularly to the lch5 organ. This way the organ is stretched triggering action current (AC) frequencies with remarkably low variability.

The lch5 neurons of *Cir1^{Rescue}* larvae responded to a 0.03 μ m pull with a mean AC frequency of 140 Hz. The frequency increased with rising pull lengths to 325 Hz at a 3 μ m pull ($\Delta_{0.03-3\mu\text{m}} = 185$ Hz; Figure 3F-H, Table S4), which shows that lch5 neurons can differentiate between these two pull lengths, i.e., stimulation strengths. In contrast, lch5 neurons of *Cir1^{KO}* were much less capable of discriminating between different pull lengths ($\Delta_{0.03-3\mu\text{m}} = 26$ Hz; Figure 3E, Table S4). Hence, in *Cir1^{KO}* the lch5 output signal relayed to the central nervous system remains virtually unaltered although the actual stimulation

intensity has drastically changed. This finding is consistent with the discrimination deficits of *Cir1^{KO}* between different vibration frequencies in *Cir1^{KO}*²². Strikingly, lch5 responses from *Cir1^{7TM}* or *Cir1^{1TM}* neurons were similar to those of *Cir1^{KO}* animals in that each exhibited a discrimination deficit (Figure 3F,G, Table S4,5). Importantly, appending a biochemical tag at *Cir1^{1TM}* C-terminus (Figure S4, Table S5) or *Cir1^{7TM}* third intracellular loop had no impact²¹.

To test for a co-requirement and to rule out technical issues for the observed mechanosensation deficits in *Cir1^{7TM}* and *Cir1^{1TM}* mutants, we generated trans-heterozygous *Cir1^{1TM}/Cir1^{7TM}* (*Cir1^{1TM+7TM}*) animals reconstituting the natural *Cir1* isoform repertoire. Reassuringly, lch5 responses in these animals ($\Delta_{0.03-3\mu\text{m}} = 106$ Hz; Figure 3H, Tables S4,5) were comparable to *Cir1^{Rescue}* despite containing only a single allele copy of each splice species. Collectively, this shows that *Cir1^{7TM}* (GPCR-like) isoforms require *Cir1^{1TM}* (non-GPCR) isoforms to tell different mechanical input intensities apart.

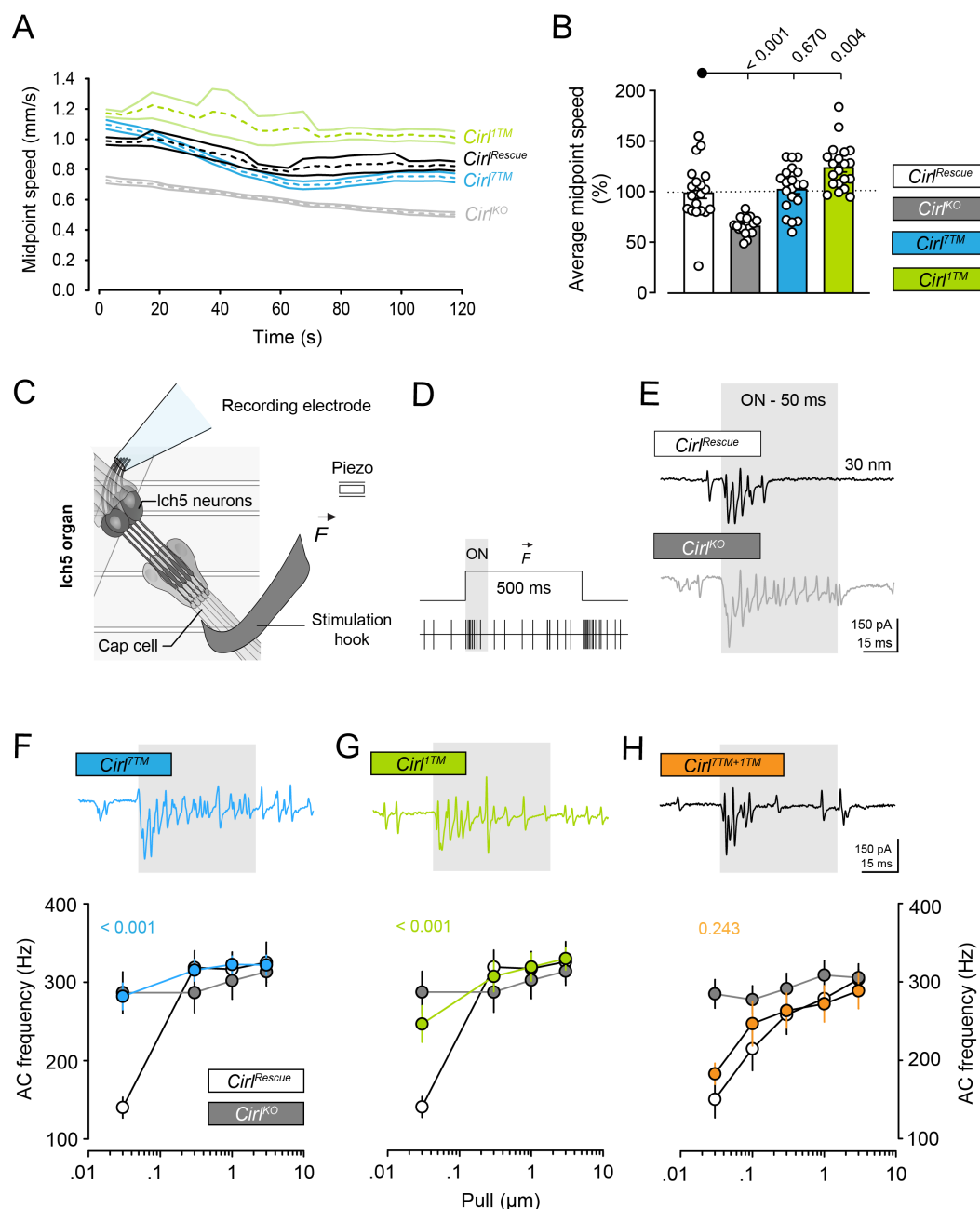


Figure 3. *Cir1^{1TM}* and *Cir1^{7TM}* proteins are co-required for neuronal discrimination of mechanical inputs

(A) Groups of *Cir1^{KO}*, *Cir1^{Rescue}*, *Cir1^{7TM}* and *Cir1^{1TM}* larvae were recorded in free locomotion. The average midpoint speed of each genotype in 5 s bins and the 95% confidence interval of the recording is displayed. *Cir1^{KO}* animals were slower than the *Cir1^{Rescue}* ²². This phenotype was rescued to and above control in *Cir1^{7TM}* and *Cir1^{1TM}* animals, respectively.

(B) Quantification of average midpoint speed (normalized to *Cir1^{Rescue}*) of each animal cohort over 2 min recordings. Related to Table S3.

(C) Schematic of action current recording configuration.

(D, E) Experimental protocol. Pull lengths (0.03, 0.1, 0.3, 1, and 3 μ m) were applied for 500 ms. Differential reactions of the *lch5* neurons as an increase of AC frequency after applying and releasing the pull are shown schematically (D) and in representative traces (E). ACs within the first 50 ms (gray box) after stimulation onset were quantified to calculate the AC frequency. F = Force.

(F-H) Quantification of AC frequencies from mechano-stimulated *lch5* neurons. Upper panels: Sample AC traces in response to a 30-nm pull length. The gray shades indicate the 50 ms time period from which the AC frequency was quantified. Lower panels: Quantification of *lch5* neuron activity at different pull lengths (in μ m: 0.03, 0.1, 0.3, 1, 3). Data are displayed as mean $\pm$ SEM. P-values denote statistical difference between *Cir1^{Rescue}* and *Cir1^{7TM}* (F), *Cir1^{1TM}* (G) and *Cir1^{7TM+1TM}* (H) at 30 nm pull length. Color code: *Cir1^{Rescue}* white, *Cir1^{KO}* gray, *Cir1^{7TM}* blue, *Cir1^{1TM}* green, *Cir1^{7TM+1TM}* orange. Related to Figure S4 and Tables S4,5.

Sound stimulation evokes activity in select *lch5* neurons that necessitates *Cir1^{7TM}* and *Cir1^{1TM}* proteins

Cir1 is involved in *lch5*-dependent sound detection²². With the tonotopic organization of the mammalian auditory system in mind, we asked if individual *lch5* neurons respond differently toward the same sound stimulus and whether different *Cir1* isoforms are relevant. Therefore, we performed Ca^{2+} imaging as a proxy for neuronal *lch5* activity employing the genetically encoded Ca^{2+} indicator *UAS-jGCaMP7³⁶* expressed under *iav-GAL4* control³⁷ (Figure 4A,B). Interestingly, in *Cir1^{Rescue}* 90 dB sound pressure level (SPL) stimuli repeatedly elicited the largest response in the most ventral neuron (= neuron 1; Figure 4C, Table S6), implying that *lch5* neurons are differentially tuned. Hence, to investigate *Cir1* isoform function on single-cell level, we quantified sound-induced Ca^{2+} signals in neuron 1. Consistent with the elevation in AC frequency measured electrophysiologically (Figure 3F-H), neuron 1 of *Cir1^{KO}* showed markedly increased Ca^{2+} signals when challenged with sound (Figure 4D-F, Table S7). Importantly, loss of either isoform species had similar effects corroborating the physiological necessity of the non-GPCR *Cir1^{1TM}* proteins (Figure 4D-E). Note that $\Delta F/F_0$ values from *Cir1^{7TM+1TM}* animals (containing only a single gene copy of *Cir1^{7TM}* and *Cir1^{1TM}* allele, respectively) ranged between those of

Cir1^{KO} and *Cir1^{Rescue}* (Figure 4F) suggesting that the amount of Cir1 protein is a limiting factor for the acoustic sensitivity of neuron 1. Ca^{2+} signals in neuron 2 and 4 seemed indistinguishable between genotypes (Figure 4G,H, Table S8). In sum, spatial analyses of mechano-dependent Ca^{2+} signals uncovered differential activity profiles of *lch5* neurons and further underpins that Cir1 function hinges on the presence of the Cir1^{ITM} non-GPCR isoforms.

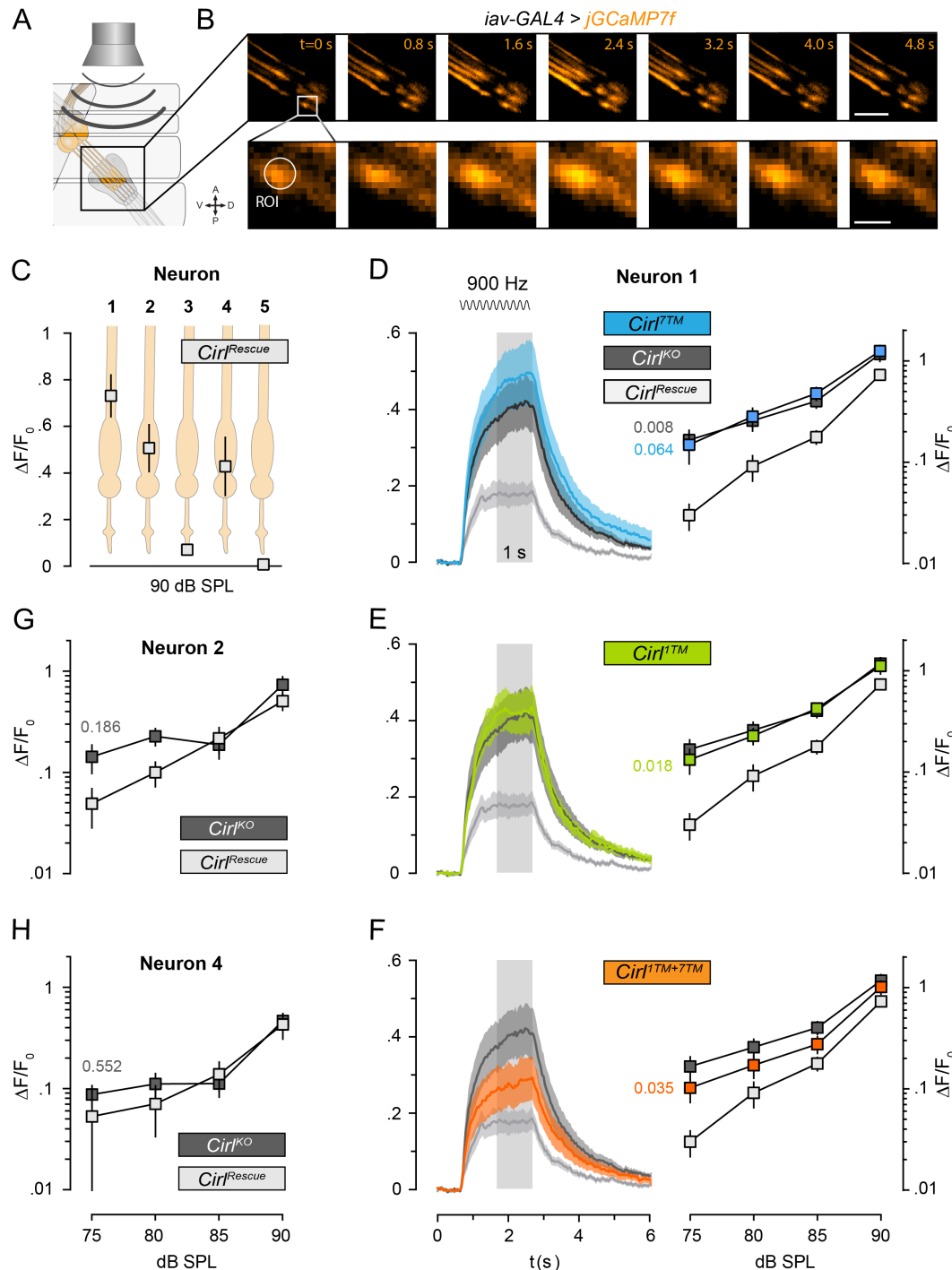


Figure 4. Sound stimulation evokes activity in select *lch5* neurons that necessitates Cir1^{7TM} and Cir1^{1TM} proteins

(A) Schematic of Ca^{2+} imaging configuration. Ca^{2+} indicator jGCaMP7f (orange) outlines *lch5* neurons.

(B) Representative images of a time series ($t = 0 - 4.8$ s) of Ca^{2+} signals visualized from larval *lch5* organs (*iav-GAL4 > jGCaMP7f*). A 900 Hz, 85 dB SPL sound was applied for 2 s (from 0.7 s to 2.7 s). To quantify Ca^{2+} signals, fluorescence intensities within a circular area with a 5-pixel diameter (~ 815 nm) were measured in the distal part of the dendrite. Region of interest (ROI) in neuron 1 shown exemplarily. Scale bars: 5 μm (upper), 1 μm (lower).

(C) Schematic of *lch5* neurons (orange) overlaid by Ca^{2+} signal average $\Delta F/F_0$ (square) of the plateau phase (1 s indicated by gray shade in (D)) measured from *Cir1^{Rescue}*, shown as mean $\pm$ SEM. Neuron 1 is the most ventral neuron. Stimulation intensity: 90 dB SPL. Related to Table S6.

(D-F) Quantification of Ca^{2+} signals measured within the dendrite of neuron 1. Left panels: Mean time course of $\Delta F/F_0 \pm$ SEM (each genotype $n = 10$). Stimulation period (2 s, 900 Hz, 85 dB SPL) is indicated by a sine wave. Right panel: To quantitatively compare sound evoked Ca^{2+} signals between genotypes, $\Delta F/F_0$ averages and SEMs were calculated from the plateau phase (1 s) at different stimulation intensities (75, 80, 85, 90 dB SPL). P-values shown are derived from statistical comparisons at 75 dB SPL between *Cir1^{Rescue}* and the three mutant genotypes ($n=10$). Color code: *Cir1^{Rescue}* white, *Cir1^{KO}* gray, *Cir1^{7TM}* blue, *Cir1^{1TM}* green, *Cir1^{7TM/1TM}* orange. Related to Table S7.

(G, H) Quantification of the Ca^{2+} signals in neurons 2 and 4. P-values from comparison of *Cir1^{Rescue}* and *Cir1^{KO}*. In (G), at 75 dB SPL, $n = 4$ and $n = 7$ for *Cir1^{Rescue}* and *Cir1^{KO}*, respectively. In (H), at 75 dB SPL, $n = 5$ for both *Cir1^{Rescue}* and *Cir1^{KO}*, respectively. Related to Table S8.

Cir1^{1TM} and Cir1^{7TM} form a heteromeric complex

The previous results collectively suggest that Cir1^{1TM} and Cir1^{7TM} are co-required to modulate the neuro-mechanical response of *lch5* neurons. Consequently, we asked whether Cir1^{1TM} and Cir1^{7TM} functions merely intersect or if Cir1 isoforms assemble signaling complexes to shape mechanosensation. To test for a direct Cir1^{1TM}-Cir1^{7TM} interaction, we used HEK293T cells as a heterologous expression system. First, we conducted ELISA to quantify the expression levels of specific Cir1 isoforms using an N-terminal hemagglutinin (HA) tag^{21,38} and found Cir1^{1TM} isoform (-F/K) more abundantly at the cell surface than Cir1^{7TM} isoforms (-B/H and -E) (Figure 5A). GPS-dependent autoproteolytic cleavage is relevant for trafficking of several aGPCR (Figure 5B)³⁹⁻⁴¹. Therefore, we wondered whether self-cleavage affects cell surface expression of different Cir1 isoforms and, if so, whether the position of the cleavage-disabling mutation ($\text{H} > \text{A}^{\text{GPS-2}}$ or $\text{T} > \text{A}^{\text{GPS+1}}$) is relevant. ELISA showed that cleavage-deficiency increased total and surface expression of Cir1^{7TM}-E receptors (Figure 5C). In contrast, the cell surface pool of Cir1^{1TM}-F/K isoforms remained unaltered irrespective of autoproteolytic cleavage competence (Figure 5C). Collectively, this shows that the faculty for self-cleavage and the layout of the CTF impacts cell surface trafficking of Cir1 isoforms.

To test whether Cir1^{7TM}-Cir1^{1TM} complexes assemble, we overexpressed HA::Cir1^{7TM}-E with and without C-terminally tagged Cir1^{1TM}-F/K proteins (Cir1^{1TM}-F/K::V5) under *CMV* promoter (*CMVp*) and

performed pull-downs. HA::Cirl^{7TM}-E served as bait to capture Cirl^{1TM}-F/K::6xV5 protein (Figure 5D). Western blot analyses of cell lysates from pull-down experiments showed Cirl^{7TM}-E-specific enrichment of Cirl^{1TM}-F/K-CTF (~ 38 kDa; Figure 5E) lending support for a direct interaction of Cirl^{7TM} and Cirl^{1TM} isoforms (input samples shown in Figure S4B-D). We repeated the experiment employing cleavage-deficient (T>A^{GPS+1}) isoform versions and were able to enrich FL Cirl^{1TM}/GPS-T>A-F/K (~ 117 kDa; Figure 5F). We detected faint background signals when Cirl^{1TM}-F/K::V5 was overexpressed alone or together with the class A GPCR P2Y12. However, the intensities of these signals were well below those of Cirl^{1TM}-F/K when enriched by Cirl^{7TM}-E (Figure 5E,G). Importantly, Cirl^{7TM}-E could also be captured in an inverse experimental set-up using Cirl^{1TM}-F/K as bait (Figure S4H-J).

Interestingly, when we placed *Cirl^{1TM}* under transcriptional control of a single *UAS*-promotor (Figure S4E) and induced Gal4 expression using the *actin5C*-promoter⁴², we detected Cirl^{1TM}-F/K in input samples only when Cirl^{7TM}-E was co-expressed (Figure S4F). Importantly, in this modified pull-down set-up Cirl^{1TM} also co-immunoprecipitated with Cirl^{7TM} cleavage-independently (Figure S4G). Note that we used ADGRE5/CD97 here as mock bait instead of P2Y12. This was done as an additional measure to ensure specificity of Cirl complex formation (Figure S4G).

Finally, to gain a first insight into Cirl isoform interaction interface(s), we expressed a soluble NTF-only version of Cirl^{1TM}-F/K *i.e.* lacking the CTF (Cirl^{1TM}-F/K^{ΔCTF}, Figure 5H). Cirl^{1TM}-F/K^{ΔCTF} also formed a complex with Cirl^{7TM}-E (Figure 5I,J), which suggests that Cirl^{1TM}-NTF is part of the binding interface between different Cirl isoforms. Reassuringly, Cirl^{1TM}-F/K^{ΔCTF} engaged in a homomeric interaction with membrane anchored Cirl^{1TM}-F/K, which was expected as it shares NTF layout with Cirl^{7TM}-E (Figure 5J). Collectively, these findings render homomeric interaction between Cirl isoforms possible and show that Cirl^{1TM} molecules can physically interact with Cirl^{7TM} receptor in an NTF-dependent and cleavage-independent fashion.

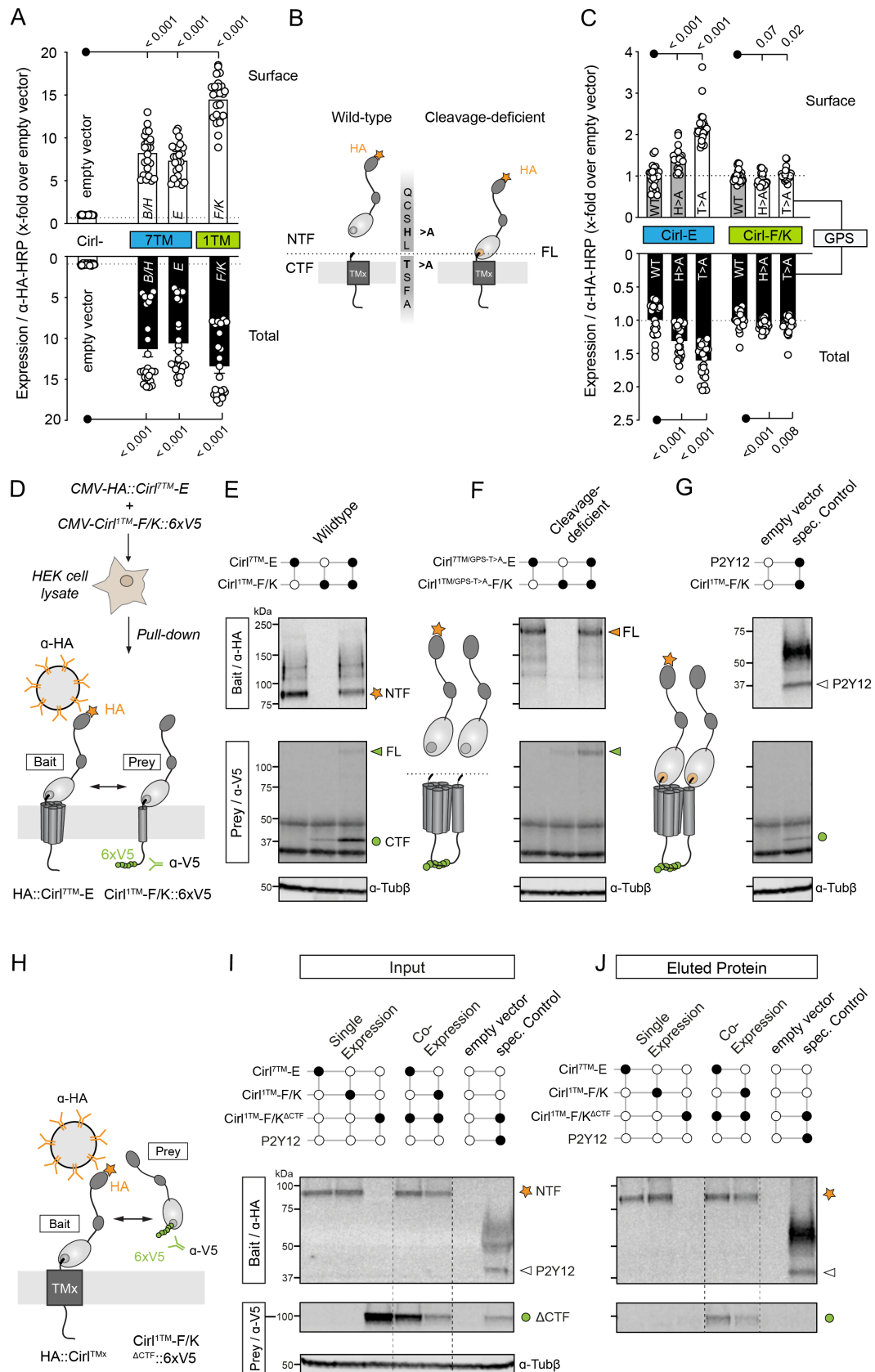


Figure 5. Cirl^{1TM} and Cirl^{7TM} form a heteromeric complex

(A) Quantification of total and cell surface expression of individual Cirl isoforms by ELISA. Cirl detection was done using a N-terminal HA tag (HA::Cirl-X). Data are shown as mean $\pm$ SEM. Related to Table S9.

(B) Schematic of HA-tagged wildtype and cleavage-disabled Cirl.

(C) ELISA of wildtype and cleavage-deficient Cirl-E and -F/K. Data of cleavage-deficient Cirl variants were normalized to respective wildtype receptor and are shown as mean $\pm$ SEM. Related to Table S9.

(D) Schematic of pull-down experiments shown in E-G.

(E-G) Immunoblot of a pull-down experiment using wildtype (E) and cleavage-deficient (F) Cirl. HA-tagged Cirl^{7TM} receptor (HA::Cirl-E) served as bait and was immobilized/detected using α -HA antibody (upper row); Cirl^{1TM} prey (Cirl^{1TM}-F/K::V5) was detected using α -V5 antibody (lower row). FL and NTF of Cirl^{7TM}-E are indicated by orange arrowhead and orange star, respectively. FL and CTF of Cirl^{1TM}-F/K are indicated by green arrowhead and green circle, respectively. All samples were run side-by-side on the same gel. (G) Empty vector: beads incubated with lysate from empty vector transfected cells. Specificity control: P2Y12 receptor (white arrowhead) + Cirl^{1TM}-F/K. Unspecific binding of Cirl^{1TM}-F/K::V5 to the beads returned background signal (green circle), which was below that of co-precipitated Cirl^{1TM}-F/K (E). Tubulin served as loading control (α -Tub β).

(H) Schematic of pull-down experiments shown in I and J. HA-tagged Cirl^{TMx} protein (HA::Cirl^{7TM}-E or HA::Cirl^{1TM}-F/K) served as bait for soluble Cirl^{1TM}-NTF(Cirl^{1TM}-F/K^{ACTF}::6xV5).

(I, J) Immunoblots from pull-down experiments show that isoform assembly involves the NTF of Cirl. Input samples (I) and eluted protein (J). Cirl^{TMx} was immobilized/detected using α -HA antibody (upper row, orange star). Detection of Cirl^{1TM}-F/K^{ACTF} (green circle) was done using α -V5 antibody (lower row). Empty vector: beads incubated with lysate from empty vector transfected cells. Specificity control: P2Y12 receptor (white arrowhead) + Cirl^{1TM}-F/K^{ACTF}. Tubulin served as loading control (α -Tub β).

Cirl signals through the G α_o subunit to attenuate cAMP quantity

Previous work showed that Cirl affects lch5 mechanosensing through a cAMP-dependent signaling route²¹. Cirl^{1TM} proteins are ill-equipped to couple G proteins and thus are most likely unfit to act through inhibition of cAMP formation. This in combination with the isoform interaction led us to interrogate if aberrant lch5 mechanosensing in *Cirl*^{1TM} mutants can be reversed through pharmacological inhibition of adenylyl cyclases (ACyc), enzymes responsible for G protein-dependent cAMP production (Figure 6A). Similar to *Cirl*^{KO} and *Cirl*^{7TM} lch5 neurons, application of 100 μ M SQ22536, an ACyc inhibitor, indeed rescued mechanically-evoked AC frequencies in *Cirl*^{1TM} (Figures

6B-D, S5A, Table S10,11) suggesting a role of $Cir1^{1TM}$ - $Cir1^{7TM}$ complexes in the suppression of cAMP levels.

Next, we asked which $G\alpha$ protein species acts downstream of $Cir1$ to suppress neuronal cAMP levels. $G\alpha_i$ protein is expressed in developing $lch5^{43}$, inhibits ACycs and thus reduces cAMP⁴⁴. However, when we knocked-down the $G\alpha_i$ gene in $lch5$ using two different RNAi lines (*iav-GAL4* > *UAS- $G\alpha_i$ -RNAi-1* and -2), we found no effect on $lch5$ mechanoresponses (Figures 6E, S5B, Table S12,13,14). Despite the controversial view on how $G\alpha_o$ modulates cAMP levels, this subunit is known to play vital roles in different neuronal contexts⁴⁵⁻⁴⁹. We measured the mechano-response of $G\alpha_o$ -depleted $lch5$ s (*iav-GAL4* > *UAS- $G\alpha_o$ -RNAi*), which phenocopied the deficit of null and isoform-specific *Cir1* mutants (Figure 6F, Table S12,13). Affirmingly, $lch5$ -specific neuronal overexpression of $G\alpha_o$ in $lch5$ neurons of *Cir1^{KO}* animals (*Cir1^{KO}*, *iav-GAL4* > *UAS- $G\alpha_o$*)⁴⁸ rescued the AC frequency (Figure 6G, Table S15,16). Together these data show that $Cir1$ acts through $G\alpha_o$ but not $G\alpha_i$ and suggest that 7TM-isoforms cooperate with 1TM protein, presumably to decrease neuronal cAMP levels.

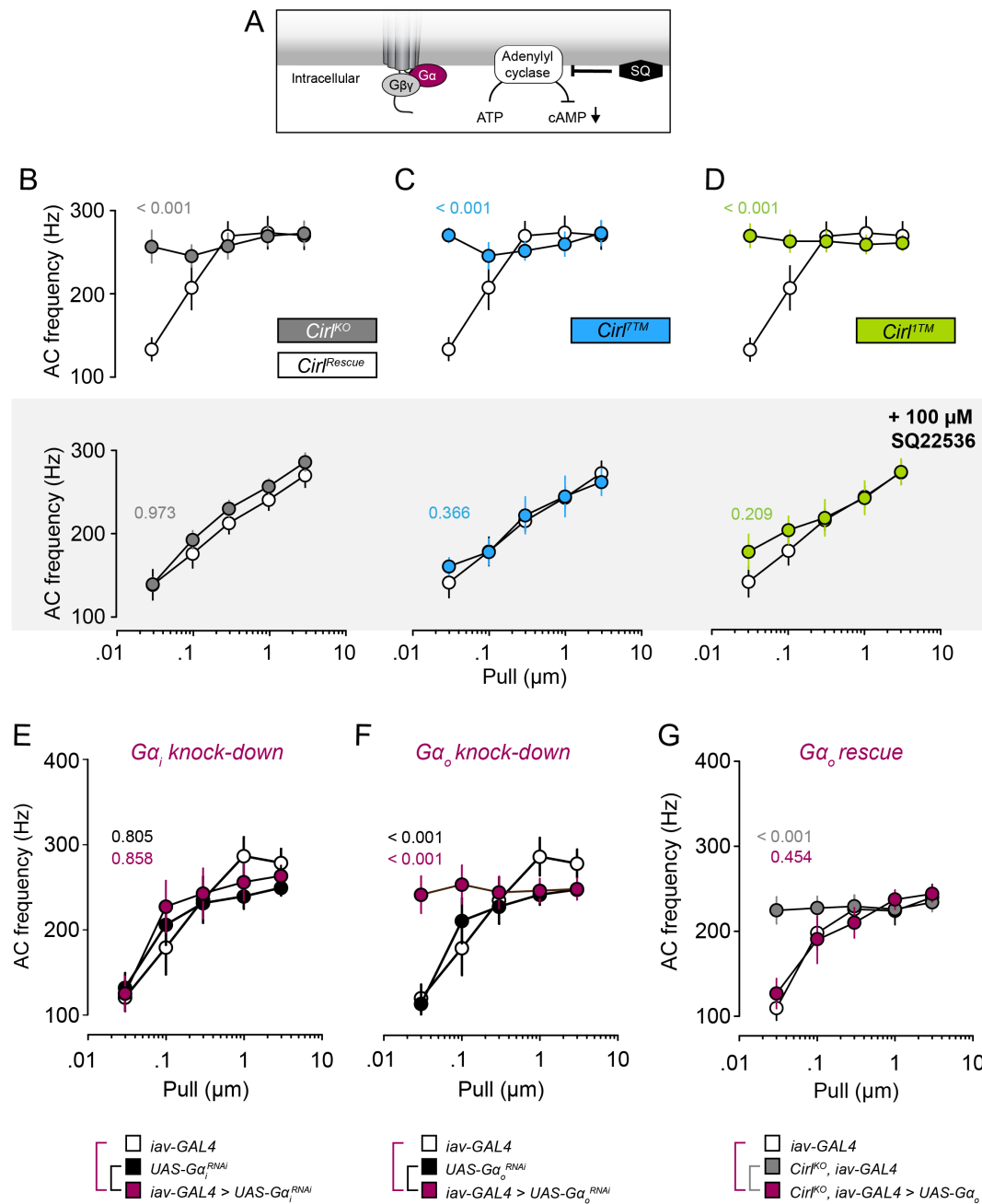


Figure 6. CirI signals through $G\alpha_o$ subunit to attenuate cAMP quantity

(A) Schematic of pharmacological inhibition of adenylyl cyclases (ACyc) by SQ22536, which decreases cAMP levels.

(B-D) Quantification of AC frequencies from mechanically stimulated *lch5* neurons before and after treatment with ACyc inhibitor. (B-D) Show the response of *CirI^{KO}* (B), *CirI^{7TM}* (C) and *CirI^{1TM}* (D) compared to *CirI^{Rescue}* before treatment with the ACyc blocker SQ22536. Gray-shaded area shows *lch5* responses of the three mutants 10 min after addition of 100 μM SQ22536. P-values denote statistical differences of AC frequencies at a pull length of 30 nm between *CirI^{Rescue}* and the three mutants. *CirI^{Rescue}*, white; *CirI^{KO}*, gray; *CirI^{7TM}*, blue; *CirI^{1TM}*, green. Related to Figure S5A and Table S10.

(E,F) Quantification of AC frequencies from mechanically stimulated $G\alpha_i$ or $G\alpha_o$ -deficient *lch5* neurons (*iav-GAL4* > *UAS-G α_i ^{RNAi}* or *UAS-G α_o ^{RNAi}*). (E) $G\alpha_i$ knock-down leaves AC frequencies unaffected. (F) $G\alpha_o$ knock-down leads to an increase in AC frequencies, phenocopying the *Cir1^{KO}*. P-values are derived from the statistical comparison of the AC frequencies at 30 nm pulls between *iav-GAL4* > *UAS-G α_i ^{RNAi}* or *iav-GAL4* > *UAS-G α_o ^{RNAi}* (in magenta) and *iav-GAL4* (in white) or *UAS-RNAi* controls (in black). Related to Figure S5B and Tables S12,13.

(G) Quantification of AC frequencies from mechanically stimulated *lch5* neurons overexpressing $G\alpha_o$ in the absence of *Cir1*. *lch5*-specific $G\alpha_o$ expression rescues elevated AC frequency of *Cir1^{KO}* back to control levels. P-values are derived from the statistical comparison of the AC frequencies at 30-nm pulls between *Cir1^{KO}*, *iav-GAL4* > *UAS-G α_o* , (in magenta) and *iav-GAL4* (in white) or *Cir1^{KO}*, *iav-GAL4* (in gray). Related to Figure S5C and Table S16.

(B-G) Data are shown as mean $\pm$ SEM (each genotype n = 10).

Mammalian 1TM and 7TM GPR126/G6 isoforms interact

Finally, we tested if isoform-specific complex assembly is a common phenomenon also observed for mammalian aGPCR. GPR126/G6 is an aGPCR with abundant expression in the nervous system, heart and bone⁵⁰, whose splice spectrum includes 1TM and 7TM-containing transcript variants²⁷. For co-immunoprecipitation experiments, we appended the N terminus of G6^{7TM} with an HA tag (HA::G6^{7TM}) and the C terminus of G6^{1TM} with V5 tags (G6^{1TM}::V5, Figure 7A). Both proteins were robustly expressed in HEK293T cells (Figure 7B,C) and an extracellular domain fragment (ECD^{fragment}) of HA::G6^{7TM} and an FL G6^{1TM}::V5 protein were found in the input sample (Figure 7B). Intriguingly, a FL and an intracellular fragment (ICD^{fragment}) of G6^{1TM}::V5 was immobilized by HA::G6^{7TM} bait when co-expressed. When GAIN domain cleavage of HA::G6^{7TM} and G6^{1TM}::V5 was disabled by mutation, co-immunoprecipitation still captured FL G6^{1TM}::V5 protein but, as expected, no ICD^{fragment} (Figure 7C). Importantly, G6^{1TM}::V5 using the ADP receptor P2Y12 receptor as bait was not detected confirming specificity of the G6^{1TM}-G6^{7TM} interaction (Figure 7C).

Collectively, our findings suggest that the formation of isoform complexes may be a general phenomenon shaping aGPCR biology in different species.

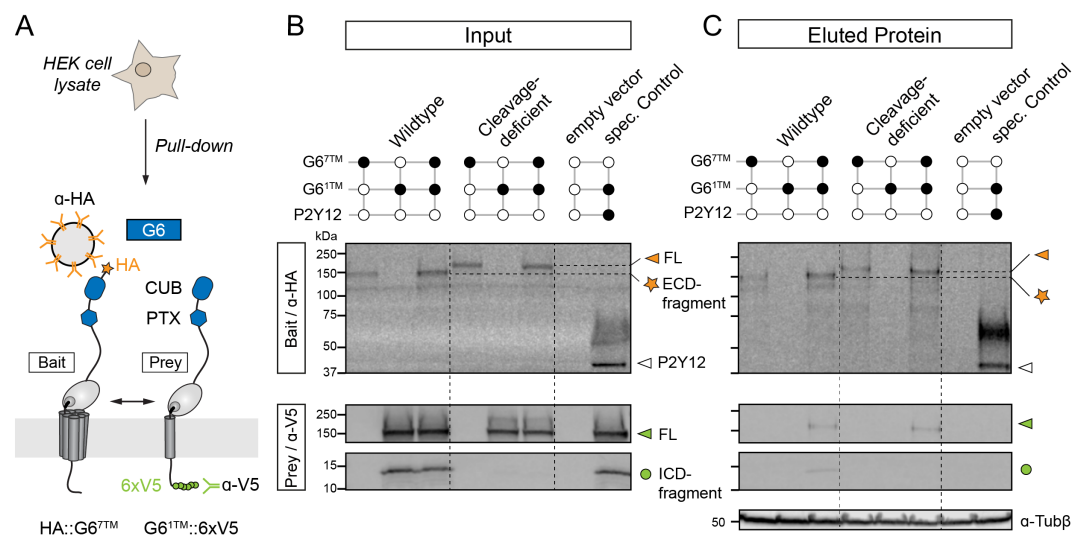


Figure 7. Mammalian 1TM and 7TM GPR126/G6 isoforms interact

(A) Schematic of the experimental configuration. N-terminally tagged HA::G6^{7TM} immobilized via α-HA functionalized magnetic beads served as bait to capture G6^{1TM}::V5.

(B, C) Immunoblots of G6 pull-down experiment. Input (B); eluted protein (C). Upper row (bait): α-HA antibody recognizes FL (orange arrowhead) and an ECD fragment of HA::G6^{7TM} (orange star). Lower row (prey): α-V5 antibody recognizes FL (green arrowhead) and an ICD fragment of G6^{1TM}::V5 (green circle). G6 isoforms were expressed under *CMVp*. HA-tagged P2Y12 was used as control bait to ensure specificity of the interaction between G6 isoform. Tubulin served as loading control (α-Tubβ).

DISCUSSION

While mechanosensing through ionotropic mechanosensors is well recognized¹⁶, it is largely unknown how metabotropic mechanosensors establish their sensitivity. Here, we show that *Cir1* uses intron retention as a physiological strategy to produce non-GPCR-coding Cir1^{1TM} isoforms. These form protein complexes with GPCR-like Cir1^{7TM} receptors to tune the mechanosensitive abilities of sensory neurons (Figures 3-5). At first glance, this is reminiscent of receptor modifying proteins (RAMP)⁵¹, which however, are encoded in genes separate from those GPCR that they modify.

Intron retention is a major alternative splicing mode⁵², which is poorly understood⁵³ and even often regarded a consequence of erroneous splicing counteracted by nonsense-mediated mRNA decay⁵⁴. Contrarily, intron retention was reported to control developmentally regulated genes⁵⁵ and to curtail the abundance of protein-coding transcripts⁵⁶. Importantly, intron retention rarely has been seen through to the level of predicted protein isoforms encoded in mature transcripts with retained introns⁵⁷. Alternative splicing of *Cir1* generates *Cir1*^{7TM} transcripts (removal of intron 6) and *Cir1*^{1TM} transcripts (retention of intron 6) that encode an alternative intracellular tail and a premature stop codon (Figures 1 and S1). We found both transcript species co-expressed throughout development (Figure 1). In sensory lch5 neurons

of *Drosophila* larvae, Cirl^{7TM} and Cirl^{1TM} isoforms are located in close proximity to the ciliary mechanosensing apparatus and are equally required for the detection of different mechanical stimulus qualities: sound and stretch at different intensities (Figures 2-4). Interestingly, exclusive Cirl^{7TM} or Cirl^{1TM} pools exist in some subcellular regions implying functions outside of the Cirl^{1TM}-Cirl^{7TM} complex (Figure 2). The finding that Cirl^{7TM} suffices to restore locomotion speed to control values lends support to this notion (Figure 3). The functional relevance of different isoforms may explain why previous rescue attempts of aGPCR knockout animal models with transgenes that only encoded GPCR-like 7TM proteins were futile, while rescue strategies with genomic transgenes (retaining the ability to produce GPCR-like and non-GPCR isoforms) were successful^{22,58,59}.

Dimer and oligomer formation between canonical 7TM-GPCR has been controversially discussed for many years⁶⁰⁻⁶³. While dimerization of GPCR such as GABA_B and mGlu receptors occurs between 7TM protomers and is required for membrane localization, ligand binding and signal transduction^{64,65}, we show in this study that complex formation manifests between 7TM and 1TM protomers of the aGPCR Cirl from *Drosophila* and ADGRG6 from mouse (Figures 5 and 7). This finding is in agreement with a previous intermolecular complementation experiment in which co-expression of a wildtype LAT-1 CTF (7TM protein) and a chimeric protein containing the ECD of the *C. elegans* latrophilin homolog LAT-1 fused to the transmembrane unit of PAT-3 (1TM protein) was sufficient to restore full receptor activity. Moreover, the LAT-1 ECD was found to form stable homodimers⁶⁶, which is in line with our finding that Cirl isoforms complex via their NTFs into heteromeric and potentially homomeric assemblies (Figure 5).

Recent structural evidence shows direct engagement of the tethered agonist (TA)/Stachel region with the 7TM domain for several aGPCR⁶⁷⁻⁷⁰. The newfound functionality of Cirl 7TM-1TM dimers implies that the GAIN domain of at least the 1TM protomer can exist in a steric configuration that does not require TA-7TM-interaction. Alternatively, TA-7TM engagement may occur between rather than within protomers as implied before⁶⁶.

We show that Cirl signals through Gα_o, but not Gα_i proteins to reduce cAMP levels and to adjust mechano-evoked AC frequencies of sensory lch5 neurons (Figures 6 and S5). Brain-derived rat Latrophilin-1 co-precipitates with Gα_o protein⁷¹, the most abundant yet puzzling Gα subunit in nervous tissue^{45,72}. Whether Gα_o protein couples to Cirl^{7TM} or the Cirl^{1TM}-Cirl^{7TM} complex is unclear, but the fact that a loss of either of the three proteins results in a similar phenotype indicates the necessity of Cirl^{1TM} to efficiently couple Gα_o to Cirl^{7TM} (Figure 6). Alternatively, Cirl^{1TM} may act through another Gα_o-coupled receptor in lch5, however, due to absence of additive effects in *Cirl*^{KO}, i.e. loss of both *Cirl*^{7TM} and *Cirl*^{1TM}, this scenario is less likely. Whether Gα_o impacts cAMP concentrations directly is currently debated⁴⁹, but it is tempting to speculate that Gα_o as a potential liberator of Gβγ subunits affects ion channel gating and indirectly cAMP levels⁷³. This model fits previous work, where modulation of mechanogating properties of transient potential receptors through Cirl was shown²¹.

Ca²⁺ imaging of acoustically stimulated lch5 neurons showed robust differences in neuronal activity across the five lch5 cells (Figure 4). Upon acoustic stimulation the highest Ca²⁺ signals were recorded from neuron 1, medium signals in neurons 2 and 4, while neurons 3 and 5 were unresponsive. This could stem from morphological variability of lch5 neurons or differing sensitivity thresholds across cells, ultimately contributing to the mechanical responsiveness of the neurons. While Ca²⁺ response of neuron 1 relied on the presence of both Cirl^{1TM} and Cirl^{7TM}, it is unclear if and how Cirl or which isoforms are involved in weaker signals recorded from neurons 2 and 4.

Alternative splicing is particularly prominent in nervous tissues^{74,75} and may be causally related to its tremendous information processing power and plasticity. Here, we present proof that alternative splicing of aGPCR is a physiological strategy that establishes sensitivity towards defined mechanical stimuli in individual neurons. This discovery is underpinned by the utility of aGPCR complex formation through the expression and unexpected signaling role of non-GPCR isoforms. This offers an explanation for the versatile splice repertoire of aGPCR genes as a source of modulators for metabotropic signals transduced by aGPCR proteins. Such interactions may prove useful as means to interfere with aGPCR signaling and mechanosensitivity in the nervous system and other organs, which may pave the way to novel pharmacological strategies.

EXPERIMENTAL PROCEDURES

Drosophila culture and stocks

Drosophila were cultured on standard cornmeal food at 25 °C. The following strains were generated in this study:

LAT403, w^{1118} ; $Cir1^{KO}$ $\{w^{+mC}=pNH103 [RFP::Cir1^{GPS-WT}::2xV5::3xFlag w^+]\}/CyoGFPw$;;
($Cir1::3x$ -tagged)
LAT475, w^{1118} ; $Cir1^{KO}$ $\{w^{+mC}=pNH177 [RFP::Cir1^{GPS-H>A}::2xV5::3xFlag w^+]\}/attP^{dCir1}loxP/CyO$;;
($Cir1::3x$ -tagged^{GPS-H>A})
LAT650, w^{1118} ; $Cir1^{KO}$ $\{w^{+mC}=pNH191 [Cir1^{7TM}::2xV5 w^-]\}/attP^{Cir1}/CyoGFPw$;;
($Cir1^{7TM}::V5$)
LAT651, w^{1118} ; $Cir1^{KO}$ $\{w^{+mC}=pNH190 [Cir1^{1TM}::2xV5 w^-]\}/attP^{Cir1}/CyoGFPw$;;
($Cir1^{1TM}::V5$)
LAT1115, w^{1118} ; $Cir1^{KO}$ $\{w^{+mC}=pNH311 [Cir1^{7TM-GPS/H>A}::2xV5 w^-]\}/attP^{Cir1}/CyoGFPw$;;
($Cir1^{7TM-GPS/H>A}::V5$)
LAT1111, w^{1118} ; $Cir1^{KO}$ $\{w^{+mC}=pNH310 [Cir1^{1TM-H>A}::2xV5 w^-]\}/attP^{Cir1}/CyoGFPw$;;
($Cir1^{1TM-GPS/H>A}::V5$)
LAT652, w^{1118} ; $Cir1^{KO}$ $\{w^{+mC}=pNH189 [RFP::Cir1^{7TM}::2xV5 w^-]\}/attP^{Cir1}/CyoGFPw$;;
($RFP::Cir1^{7TM}::V5$)
LAT653, w^{1118} ; $Cir1^{KO}$ $\{w^{+mC}=pNH188 [RFP::Cir1^{1TM}::2xV5 w^-]\}/attP^{Cir1}/CyoGFPw$;;
($RFP::Cir1^{1TM}::V5$)
LAT838, w^{1118} ; $Cir1^{KO}$ $\{w^{+mC}=pNH217 [Cir1^{1TM} w^-]\}/attP^{Cir1}/CyoGFPw$;;
($Cir1^{1TM-no V5}$)

The following strains were previously generated or generated elsewhere:

LAT163, w^{1118} ; $Cir1^{KO}$ $\{w^{+mC}=pTL370 [dCir1]\}/attP^{Cir1}loxP$;;²² ($Cir1^{Rescue}$)
LAT26, w^{1118} ; $Cir1^{KO}$ $attP^{Cir1}loxP$;;²² ($Cir1^{KO}$)
LAT159, w^{1118} ; $Cir1^{N-RFP}$;;²¹ (RFP^{Cir1})
LAT176, w^{1118} ; $RFP^{Cir1^{GPS-T>A}}$;;²¹ ($RFP^{Cir1^{GPS-T>A}}$)
LAT112, w^{1118} ; $P\{w^{mc}=iav-Gal4 w^+\}/attP2$;;²² ($iav-GAL4$)
GN393, w^* ; $UAS-dG\alpha_o$, $ZH-22A-3xP3-RFP/CyoGFPw$;;⁴⁸ ($UAS-dG\alpha_o$)

Bloomington Stock Center (NIH P40OD018537) or Vienna *Drosophila* Research Center⁷⁶.

w^{1118} ; $P\{20xUAS-IVS-jGCamp7f w^+\}/VK00005$;;³⁶ ($20xUAS-jGCAMP7f$)
 $P\{TRIP.JF01608\}/attP2$, BDSC #31133⁷⁷ ($UAS-G\alpha_iRNAi-1$)
 $P\{TRIP.GL00328\}/attP2$, BDSC #35407⁷⁷ ($UAS-G\alpha_iRNAi-2$)
 $P\{TRIP.JF02844\}/attP2$, BDSC #28010⁷⁷ ($UAS-G\alpha_oRNAi$)

Molecular cloning

Cir1 cDNAs were N-terminally complemented with a mammalian Kozak sequence (gccacc), followed by the start codon, signal peptide (SP) of mammalian ADGRG1/GPR56, an HA tag (*HA::Cir1-X*) and are contained in the *CMV* promoter-containing pcDNA3.1 backbone (Table S1). *Cir1*^{1TM} constructs used for immunoprecipitation experiments lacked HA tag and contained six V5 tags at their C termini (*Cir1*^{1TM}::V5). Primers used for cloning of *Cir1* and *ADGRG6/GPR126* cDNA constructs are listed in Table S2. Plasmid amplification was done via transformation in *E. coli* (XL1-Blue, DH5 α or EPI400) and DNA isolation via Midiprep (NucleoBond® Xtra Midi, Macherey & Nagel). The Ranger (Bioline, #BIO21121) or Q5 (New England Biolabs, #M0491S) high-fidelity proof-reading DNA polymerases were utilized for all PCR-based cloning steps. Initial insert verification was done by restriction analyses. To ensure absence of errors each PCR-amplified region was sequenced completely.

Construction of *Drosophila* transgenes was done as follows:

pNH103 (Cir1::3x-tagged)

Primers *nh_199F/259R* were used to amplify a 4.9-kbp fragment from *pMN21* (genomic 1.9 kbp *Cir1* subclone) and to introduce *SgfI* and *StuI* restriction sites at the 3' and 5' end, respectively.

2xV5 tag was generated using *nh_260F/261R*, which contained a *SgfI* and *StuI* sites for subsequent ligation with 4.9-kbp fragment (resultant subclone *pNH101*). *Cir1::2xV5* fragment was transplanted from *pNH101* into *pMN24* (genomic *RFP::Cir1::3xFlag* construct) via *PacI* and *SpeI* restriction sites resulting in *pNH103*.

pNH177 (Cir1::3x-tagged^{GPS/H>A})

Primers *mn_38F/39R* were used to substitute the histidine codon of the canonical GPS sequence for an alanine codon through site-directed mutagenesis of *pNH101*, which resulted in *pNH176*. *PacI/SpeI* fragment of *pNH176* was cloned into *pNH103* resulting in *pNH177*.

pNH188 (RFP::Cir1^{1TM}::2xV5)

Genomic sequence encoding *RFP::Cir1^{1TM}::2xV5* (~ 3.9 kbp) in pMK backbone was synthesized by ThermoFisher/GeneArt (resulting construct *pNH186*). A 3.9-kbp fragment was cloned into the expression vector using *MluI* and *BstBI* sites to generate *pNH188*.

pNH189 (RFP::Cir1^{7TM}::2xV5)

Genomic sequence encoding *RFP::Cir1^{7TM}::2xV5* (~ 5.7 kbp) in pMK backbone was synthesized by ThermoFisher/GeneArt (resulting construct *pNH187*). A 5.7-kbp fragment was cloned into expression vector using *MluI* and *AatII* sites to generate *pNH189*.

pNH190 (Cir1^{1TM}::2xV5)

AgeI-based removal of mRFP sequence and subsequent re-circularization of *pNH188* resulted in *pNH190*.

pNH191 (Cirl^{7TM}::2xV5)

AgeI-based removal of mRFP sequence and subsequent re-circularization of *pNH189* resulted in *pNH191*.

pNH217(Cirl^{1TM} no tags)

Engineered by GenScript.

pNH310 (Cirl^{1TM-GPS/H>A}::2xV5)

Engineered by GenScript.

pNH311 (Cirl^{7TM-GPS/H>A}::2xV5)

Engineered by GenScript.

Primers (5'-3' orientation)

nh_199F	tgattggcgatcgctgaaaaacagataaacacaatttt
nh_259R	aagcctccaccaccacttatattgtaag
nh_260F	cctggcaagcccatcccaacccccctgctggcctggattccaccggcaagcccatcccaacccccctgctggcctggatt ccacctgcgat
nh_261R	cgcaggtggaatccaggccagcagggggtggggatgggcttgcgggtggaatccaggccagcagggggtggggat gggcttgcagg
mn_38F	gcgtctgcagttgcaacgccctgacaaactttgcc
mn_39R	ggcaaagttgtcagggcgttgcaactgcagacgc

PhiC31-mediated integration of transgenes

Plasmids containing *w⁺* and an *attB* site were injected into *phiC31*[3xP3-RFP-3xP3-GFP-vas-*PhiC31*]; *Cirl^{KO} attP-loxP*; embryos (done by BestGene Inc.)²². *w⁺* served as the selection marker to identify recombinants and was subsequently removed by Cre-mediated excision (done by BestGene Inc.). Precise transgene insertion and removal of *w⁺ cassette* was validated by PCR genotyping.

RT-PCR

Total RNA was isolated from larvae and adult *w¹¹¹⁸* animals using RNeasy RNA Isolation Kit (Qiagen) and was directly used for reverse transcription using Superscript III Reverse Transcriptase according to manufacturer's protocol (Invitrogen). PCR-amplification on transcribed cDNA libraries was carried out with the following primers:

Primers (5'-3' orientation)

tl_5F:	tcacagggagcgcagcgtggtgca
tl_6R:	atgctggtatagatcgaggtgcgcg
tl_392R:	cgtgggtctggtcgcaatattgt
rk_16:	gtgaatttccttgcgcgtg
rk_17:	ctccagtctcgctgaagaag

RNA-sequencing analyses

RNA-seq datasets that document the transcriptome at 30 distinct developmental stages of the *yl; cn bw1 spl* *Drosophila melanogaster* strain³⁴ from NCBI Sequence Read Archive (SRA)⁷⁸ were chosen to analyze and quantify *Cirl* splice variants during ontogenesis. Reads were mapped to the reference *Drosophila melanogaster* genome (July 2014, BDGP6 (GCA_000001215.4)) with Ensembl 88 annotations using Tophat 2.0.14.⁷⁹, which aligns reads using Bowtie2 (version 2.1.0). Reads, which did not map uniquely to a genome position were excluded. After indexing with SAMtools (version 1.3.1)⁸⁰, the mapped reads were assembled to transcripts and quantified by StringTie (version 1.3.3)^{81,82}. For Tophat, we used the ‘default’ parameters, which are commonly used in most studies. StringTie parameters ‘read coverage’ (-c), ‘transcript length’ (-m) and ‘bases on both sides of a junction a spliced read has to cover’ (-a) were set to minimal values in order to avoid missing transcripts and generating a bias. The parameter ‘fraction of most abundant transcript at one locus’ (-f) was lowered from default (0.01) to 0 since correction for artifacts and incompletely processed mRNA with a 1% cutoff was performed after the comparative analysis. For all other StringTie parameters default values were used. Assembled transcripts were inspected with the Integrated Genome Viewer (Broad Institute) (version 2.3.91)^{83,84}, and samples showing a visible 3’ bias due to oligo-dT/poly-A primer selection were not included.

Cell culture

HEK293T cells (RRID:CVCL_0063) were cultivated in Dulbecco’s Modified Eagle Medium (DMEM; Gibco, #11995065) supplemented with 10% (v/v) fetal bovine serum (Sigma-Aldrich, #F7524 or Gibco, #10500) and 1% penicillin/streptomycin (Sigma-Aldrich, #P4333 or Gibco, #15140122) at 37 °C and a 5% CO₂ humidified atmosphere.

HEK293T cells were split into 6-well cell culture plates (4x10⁵ cells/well) for co-immunoprecipitation analysis and into 96-well cell culture plates (3x10⁴ cells/well) coated with 0.01% poly-L-lysine (Sigma-Aldrich, #P9404) for ELISA. After 24 h, cells were transiently transfected with 2.5 µg plasmid DNA per well (6-well plate) for co-immunoprecipitation, 25 ng receptor plasmid and 75 ng empty vector plasmid per well (96-well plate) for ELISA, respectively, using Lipofectamine 2000 (Invitrogen, #11668019) according to manufacturer’s recommendations. Cells were incubated 48 h for co-immunoprecipitation and 24 h for the cell surface ELISA in a 5% CO₂ humidified atmosphere at 37 °C. After 24 h, cell culture medium was exchanged against 2 ml DMEM for co-immunoprecipitation.

ELISA

For total and cell surface expression, transfected cells (equimolar amounts of DNA) were fixed with 50 µl of 4% (w/v) paraformaldehyde in PBS at RT for 10 min. Next, unspecific epitopes were blocked with 100 µl phosphate buffered saline (PBS) containing 5% (v/v) goat serum (Sigma-Aldrich, #G6767) at RT for 1 h. Then, cells were incubated for 1 h at RT in a 1:1000 dilution of α-HA-peroxidase (RRID:

AB_390917) in 100 μ l PBS containing 5% (v/v) goat serum. After two washing steps using PBS, the cells were incubated with a substrate solution (1 mg/ml *o*-phenylenediamine and 1 μ l/ml hydrogen peroxide solved in a 0.05 M citric acid and 0.05 M disodium phosphate solution; pH 5) for 5 – 10 min at RT. The reaction was stopped with 100 μ l 2.5 M sulfuric acid. 150 μ l of the supernatant was used for absorbance measurements at 490 nm with a multimode-reader (Molecular Devices, SpectraMax M). For total ELISA, PBT (PBS + 0.05% Triton-X100) was used instead of PBS throughout the experiment.

Immunofluorescence and imaging

Drosophila larvae

Third instar *Drosophila* larvae were dissected in ice-cold HL-3 (hemolymph-like solution)⁸⁵, fixed for 10 min using cold 4% paraformaldehyde in 0.1 M phosphate buffered saline (PBS) and blocked overnight in PBT (PBS with 1% Triton X-100, Sigma-Aldrich), containing 5% normal goat serum (NGS, Jackson ImmunoResearch). Primary antibody incubation was done at 4°C overnight. The next day, secondary antibody incubation was done for 24 h at 4 °C. Each antibody incubation step was followed by two short and three 20 min washing steps (PBS with 1 % Triton X-100). Samples were stored in Vectashield (Vector Laboratories) overnight at 4 °C before mounting. Confocal imaging was performed with a Leica SP8 confocal system. To ascertain comparability between different genotypes, larvae were imaged in one session and image acquisition of genotypes alternated. Image analysis was done using ImageJ (NIH). Antibodies and dyes were used at following dilutions: rabbit- α -dsRed (1:500, Takara, #632496; RRID:AB_10013483), α -HRP conjugated with Alexa Fluor-488 (1:250; Jackson Immuno Research, #123-545-021, RRID: 2338965), Cy5-conjugated goat- α -rabbit (1:250; Jackson Immuno Research, #111-175-144, RRID: AB_2338013).

Protein Isolation

HEK293T cells

Cells were lysed using 200 μ l M-PER buffer (ThermoFisher, #78503) supplemented with protease inhibitor (1:1000, Sigma-Aldrich, #P8340). After addition of lysis buffer, cells were detached from the culture dish bottom using a cell scraper and incubated on ice for 5 min. Transferred samples were then centrifuged for 5 min at 14,000 rpm (4 °C) and a sample of the supernatant diluted 4:1 with 4x sample buffer (Licor, #928-40004) containing 10% (v/v) β -mercaptoethanol. Total protein concentration was determined using the Pierce BCA Protein Assay Kit (ThermoFisher, #23227). Note that in intra-co-immunoprecipitation experiments the same amount of total protein was used for all samples.

Fly heads

Fly heads (25/genotype) were severed using fine scissors (Fine Science Tools, #15005-08), collected in a 0.5 ml Eppendorf tubes and immediately frozen in liquid nitrogen. Subsequently, the heads were homogenized in 2% SDS supplemented with a protease inhibitor cocktail (1:1000; Sigma-Aldrich) using a glass rod (4°C). Next, Triton X-100 was added (final concentration 1%) to the samples, which

were then centrifuged for 15 min (14,000 rpm) at 4 °C. The supernatant was collected in fresh, pre-cooled tubes and centrifuged twice again for 30 min at 14,000 rpm and 4 °C. Finally, the supernatant was supplemented with SDS-based sample buffer (Li-cor) and β -mercaptoethanol (Merck).

Pupae

One- to two-day-old pupae (~ 0.6 g/genotype) were collected, immediately frozen in liquid nitrogen and homogenized in a pre-cooled mortar using a pestle. The resulting protein powder was transferred into a pre-cooled tube and supplemented with homogenization buffer (50 mM Tris, 150 mM NaCl, 1% Triton X-100, Protease Inhibitor 1:1000, 1 mM DTT, 0.5 mM PMSF). Samples were further homogenized, first using an Ultra-Turrax (4 x 15 sec; IKA T10 Basic) and second using a glass homogenisator (squished 20x/sample). Next, samples were centrifuged at 4,000 rpm for 30 min, again at 13,000 rpm for 30 min and finally at 25,000 rpm for 30 min. The samples were kept at 4 °C throughout the procedure. Finally, the supernatant was supplemented with Laemmli buffer.

Pull down

Co-immunoprecipitation was performed using the Pierce Magnetic HA tag IP/Co-IP Kit (ThermoFisher, #88838). The manufacturer's Manual IP/Co-IP protocol and the Elution Protocol using sample buffer were applied.

Immunoblots

Protein samples were electrophoretically separated using 8%, 4-12% or 4-20% Tris-glycine gradient gels and blotted (protein from fly heads: 15 V, 7 min; protein from pupae: 20 V, 1 min; 23 V, 4 min; 25 V, 2 min; protein from HEK293T cells: 15 V, 6 min) onto nitrocellulose membrane (0.2 μ m pore diameter, iBlotTM Transfer stack, ThermoFisher, #IB23001/#IB23002) using the iBlot2 (Invitrogen). The membrane was blocked for 1h using Odyssey Blocking buffer (Li-cor, #927-40000) diluted 1:2 with PBS. Blots were probed with primary antisera at indicated concentrations overnight at 4 °C: rabbit- α -RFP (1:1000, Antibodies-Online, RRID:AB_10781500), mouse- α -V5 (1:1000, Invitrogen, RRID:AB_2556564), rabbit- α -HA (1:1000, C29F4, Cell signaling technology; RRID:AB_1549585), rabbit- α -Tubulin α (1:5000, Santa Cruz antibodies, #12462-R), mouse- α -Tubulin β (1:2000, Developmental Studies Hybridoma Bank e7, RRID: AB_528499).

After rinsing twice and 3 x 5 min washing steps using 0.1% Tween-20-containing PBS, membranes were incubated with secondary antibodies from Li-cor (1:15000): IRDye 680RD goat- α -mouse (RRID: AB_10956588), IRDye 680RD goat- α -rabbit (RRID: AB_10956166), 800CW goat- α -mouse (RRID: AB_621842), 800CW goat- α -rabbit (RRID: AB_621843) for 1 h at RT and again rinsed twice and washed 3x5 min. Western blots were imaged with an OdysseyFc 2800 (Li-cor).

Larval locomotion

Briefly, third instar larvae were positioned in the center of a 9-cm petri dish filled with 1% agarose (in ddH₂O) and their behavior was recorded for two minutes using a camera (Logitech C920 HD Pro). The recorded videos were tracked and analyzed with a custom-written software (for details, see⁸⁶). We calculated the speed of the midpoint of the larvae in mm/s. For the data displayed in Figure 3B, we calculated the average midpoint speed of all larvae on a given petri dish over 2 min – thus, the sample size represents the number of experimental repetitions. For the display in Figure 3A we binned the data in 5 s intervals, and calculated the average midpoint speed, as well as the 95% confidence interval of all data of a given genotype in each bin. Then, we smoothened the curve with a sliding average filter of 5 bins. Data were analyzed with Prism 7 (GraphPad).

Ca²⁺ Imaging

To compare the effects of different *Cir1* mutations on *lch5* function, the genetically encoded Ca²⁺ indicator jGCaMP7f³⁶ was expressed under *UAS* control, using the *iav-gal4* driver²² in homozygous *Cir1^{Rescue}*, *Cir1^{KO}*, *Cir1^{1TM}*, *Cir1^{7TM}* and *Cir1^{1TM+7TM}* backgrounds. Non-wandering male third instar larvae were pinned to a Sylgard pad in ice-cold, Ca²⁺ free HL-solution^{22,87} and cut open along the dorsal midline. The body muscle wall was gently stretched to the sides and pinned to the pad with fine dissection needles (#26002-10, Fine Science Tools). Innards and the CNS were gently removed. The Sylgard pad was transferred to the imaging chamber, filled with 2 mM CaCl₂-containing HL-solution. To stimulate the *lch5* mechanically, a 900-Hz tone (sine wave) of different intensities (75 – 90 dB SPL, starting with 75) was applied via a small suspended computer loud speaker. *lch5* activity pattern was unchanged even when stimulus direction was moved 180° from its initial position (data not shown). The sound frequency was controlled by Audacity software (V3.0.0, audacityteam.org). Stimulation intensity was verified by measuring sound pressure levels prior to every experiment. At small stimulation intensities, several measurements were averaged to increase the signal to noise ratio: 75 dB, 10 measurements; 80 dB, 5 measurements. Fluorescence imaging of the jGCaMP7f signal was performed on a DM 6FS (Leica) upright microscope equipped with a 40x, 0.8 N.A. dip-in objective and a GFP filter cube: excitation, 450 – 490 nm; dichroic 495 nm; emission, 500 – 550 nm. After pre-bleaching samples for 90 s to reduce bleaching rates during the experiment, samples were illuminated by blue LED light of the same intensity (LedHUB equipped with a LedH.465.4600, Omicron Lasers). jGCaMP7f fluorescence signals were video-recorded with a Hamamatsu Orca flash 4.0, V3 sCMOS camera at a 25-Hz frame rate and a 163 x 163 nm pixel size. Ten larvae of each genotype were measured. Videos were digitized and stored using the HOKAWO 3.0 software (Hamamatsu) and imported to Fiji ImageJ (NIH). A round, five-pixel diameter region of interest (ROI) was placed at the distal end of *lch5* dendrites. The time course of the mean fluorescence intensity (F) in the ROI was exported to GraphPad Prism 6. $\Delta F/F_0$ was determined by calculating F_0 , i.e. the mean intensity during the 0.36 s period before stimulation and normalizing ΔF (i.e. $F - F_0$) to F_0 . To quantitatively compare Ca²⁺ signals between

genotypes, the $\Delta F/F_0$ averages of the Ca^{2+} signal plateaus (time 1.68 – 2.68 s) were calculated. Videos, in which any movement was detected were discarded.

Electrophysiology

To compare the effects of different *Cir1* alleles on *lch5* function, non-wandering male third instar larvae were dissected as described in ‘ Ca^{2+} imaging’. After the innards and the CNS were removed, longitudinal muscles of the third, right abdominal hemi segment were cut cautiously. The *lch5* nerve bundle was cut near the ventral, anterior end of muscle 21. The muscles 21, 22 and 23 were carefully cut and removed. The Sylgard pad was transferred to the recording chamber, filled with 2 mM CaCl_2 -containing HL-solution⁸⁸. Micropipettes with 6 – 10 μm diameters were fabricated using the DMZ Universal Puller (Zeitz-Instrumente). To perform action current (AC) recordings, the pipette, which harbored the Ag/AgCl electrode, was filled with the bath solution. The *lch5* nerve bundle was sucked into the pipette, and its opening was moved close to the soma of the neurons. Electrical activity was measured in voltage-clamp mode at 0 V using the Axopatch 200B amplifier (Molecular Devices). The measured current was low-pass Bessel filtered at 1 kHz and digitized at 10 kHz with the Axon Digidata 1550B analog-digital converter and Clampex 10.2 Software (both from Molecular Devices). To stimulate the *lch5* neurons, cap cells were perpendicularly hooked with a miniature stimulation hook made from fine dissection needles (#26002-10, Fine Science Tools). The hook was attached to a piezo element (Physik Instrumente, P-840.30). Pulls of 0.03, 0.1, 0.3, 1, and 3 μm were administered by applying negative voltage jumps to the piezo using the E663 amplifier (Physik Instrumente), thus deflecting the *lch5* to the side. Pull lengths of 3 μm and 1 μm were confirmed optically. The voltages for the 0.3, 0.1, and 0.03 μm had to be extrapolated from the voltages of longer pulls. Although the setup was thoroughly mechanically uncoupled, the hook most probably moved relatively to the organ by a much larger distance than the shortest pull length. Therefore, a reliable 30 nm pull seems unrealistic at first glance. However, since the piezo movement is very fast, its stability was measured as ± 10 nm, and the organ reacts to stimulation in a differential manner, a meaningful reaction to the 30 nm pull could be easily detected. Each pull length, starting with 0.03 μm , was applied three times for 500 ms with a one-second break between the pulls. After a 7 s pause, the next pull length was applied three times. Action currents (AC), which occurred during the first 50 ms of the pull were manually counted. The AC frequency was calculated and evaluated in SigmaPlot 12 (SyStat). The mean of the three values of the same pull length was calculated to give $n = 1$. Ten measurements in ten different animals per genotype were performed. Treatment with the adenylyl cyclase inhibitor was carried out by adding 100 μM SQ22536 (Merck, #568500) to the bath solution and incubate it for 10 min before starting the electrophysiological measurement. For sham treatment, the adenylyl cyclase inhibitor was replaced with water.

Statistics

If not stated otherwise, Data are reported as mean $\pm$ SEM, n indicates the sample size, p-values are given as an estimate of statistical significance. Data were analyzed with Prism 6 and 7 (GraphPad) and Sigma Plot 12 (Systat). If not stated otherwise, group means were compared by two-tailed Student's t-tests, unless data was non-normally distributed. In this case, group means were compared by non-parametric tests, Mann-Whitney test for unpaired comparisons and Wilcoxon Signed Rank test for paired comparisons.

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AUTHOR CONTRIBUTIONS

N.S. and D.L. conceived and supervised the study; prepared figures and wrote the manuscript with consent from all co-authors.

Conceptualization	NS, DL, TL
Methodology	NS, DL, MT, MS, MB, MBK, JI, TL, JL, MSG, JA
Validation	NS, DL, MB, DCNH, MBK, AB, AKD
Formal analysis	NS, DL, MT, MS, MB, AB, MBK, JI, AKD, TL, MSG
Investigation	NS, DL, TS, VL, MB, AB, MBK, EK, JI, DCNH, AKD, TL, JL, MSG, JA
Resources	
Writing – Original draft	NS, DL
Writing – Review & Editing	NS, DL, TL, TS, JL, AB, MBK, AKD, JL
Visualization	NS, DL, AB
Supervision	NS, DL
Project administration	NS, TL
Funding Acquisition	NS, DL, TL, TS

DECLARATION OF INTERESTS

The authors declare no competing interests.

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SUPPLEMENTAL INFORMATION

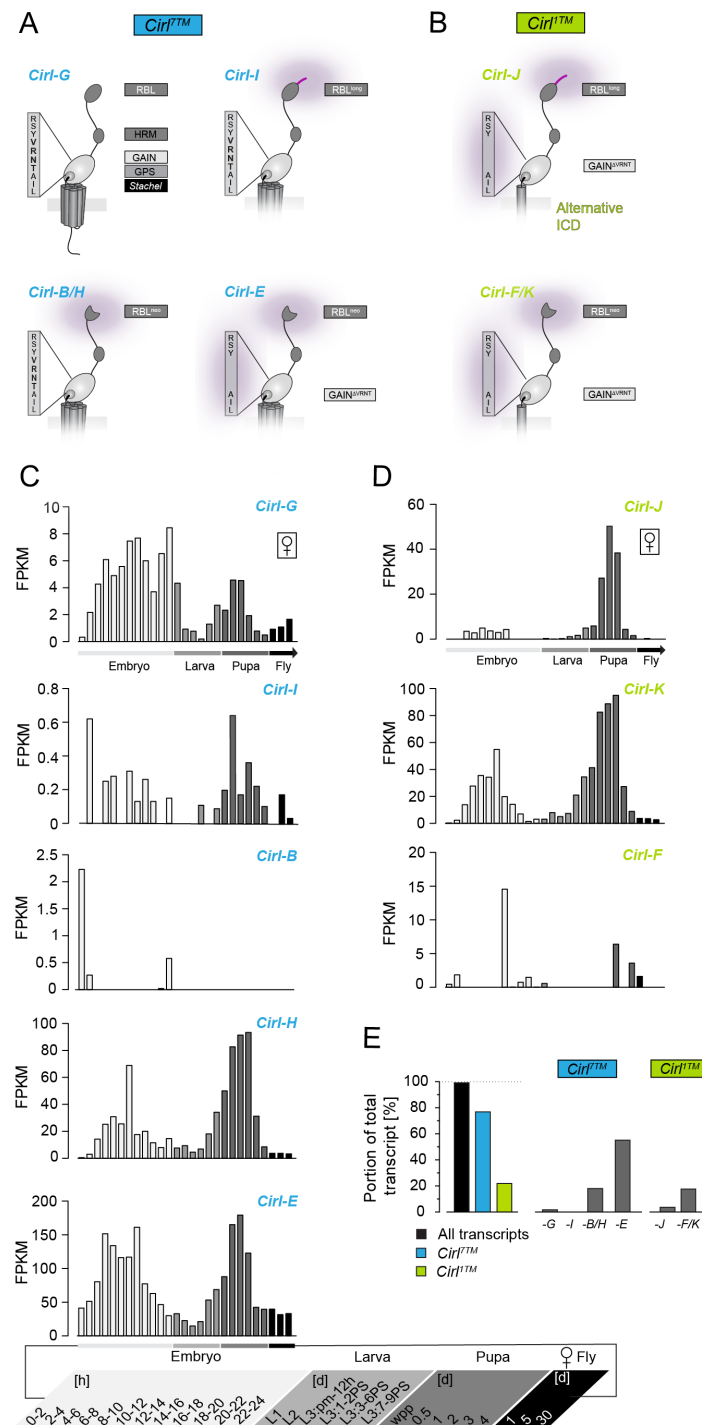


Figure S1. Ontogenetic analysis of the *Cirl* transcriptome

(A, B) Schematic of receptor isoforms encoded by the *Cirl* locus. Eight different *Cirl* transcripts are produced from a single locus. Five of them encode canonical receptors that contain 7TMs (indicated in blue) (A), while the rest encodes non-canonical protein with only 1TM (indicated in green) (B). *Cirl^{7TM}* transcripts -B and -H as well as *Cirl^{1TM}* transcripts -K and -F are translated into identical proteins, respectively, yielding six putative isoforms. Both species contain transcripts with a truncated N

terminus (RBL^{neo} domain) and/or a four amino acid deletion in the GAIN domain (GAIN^{ΔVRNT}). All transcripts contain an intact *Stachel* sequence^{41,89}.

(C, D) Bioinformatic analyses of mRNAs isolated from *yl; cn bw1 spl* *Drosophila* strain at different developmental stages³⁴ revealed peak expression of *Cir1*^{7TM} (left panel) and *Cir1*^{1TM} (right panel) transcript in embryo and pupae. Embryos were collected every two hours for one day; L1 larvae were harvested 42-44 h after egg-laying, L2 larvae 66-70 h after egg-laying and L3 larvae 83-85 h after egg-laying; PS: puff stages; wpp: white prepupae³⁴.

(E) *Cir1*^{7TM} and *Cir1*^{1TM} transcript quantification shows that ~ 80% of *Cir1* mRNA encodes *Cir1*^{7TM} protein, ~ 74.7% of which present *Cir1-B/H* and *-E* transcripts. The remaining *Cir1* mRNAs encode *Cir1*^{1TM} proteins (*-F/K*: 18.3%; *-I*: 4.2%).

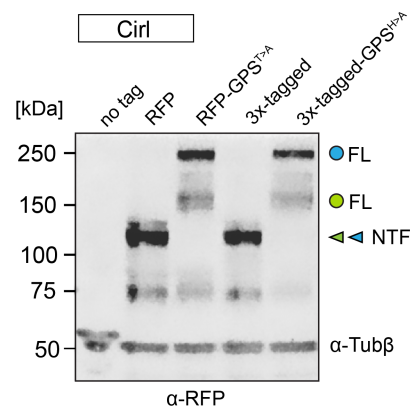


Figure S2. Cirl^{1TM} is expressed in the adult fly brain

Western blot of head homogenates shows expression of Cirl^{7TM} and Cirl^{1TM}. An α-RFP antiserum recognized the NTFs of RFP::Cirl²¹ and Cirl3x-tagged (mRFP also inserted in the NTF²¹; arrowheads green: Cirl^{1TM}; blue: Cirl^{7TM}) as well as the FL-1TM (indicated by green circle) and FL-7TM molecules of RFP::Cirl^{GPS/T>A} and Cirl3x-tagged^{GPS/H>A} animals (indicated by the blue circle). An additional Cirl band of unknown identity was detected (~ 70 kDa). Tubulin served as a loading control (α-Tubβ).

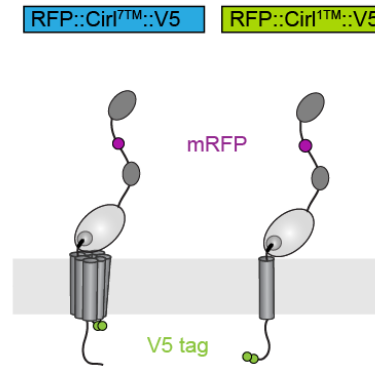


Figure S3. Scheme of RFP-tagged CirI isoform

Both CirI^{7TM} and CirI^{1TM} proteins contain an N-terminal mRFP. CirI^{7TM} contains a tandem V5-tag in the last intracellular loop (RFP::CirI^{7TM}::V5). CirI^{1TM} contains a tandem V5-tag at the C terminus (RFP::CirI^{1TM}::V5). Related to Figure 2.

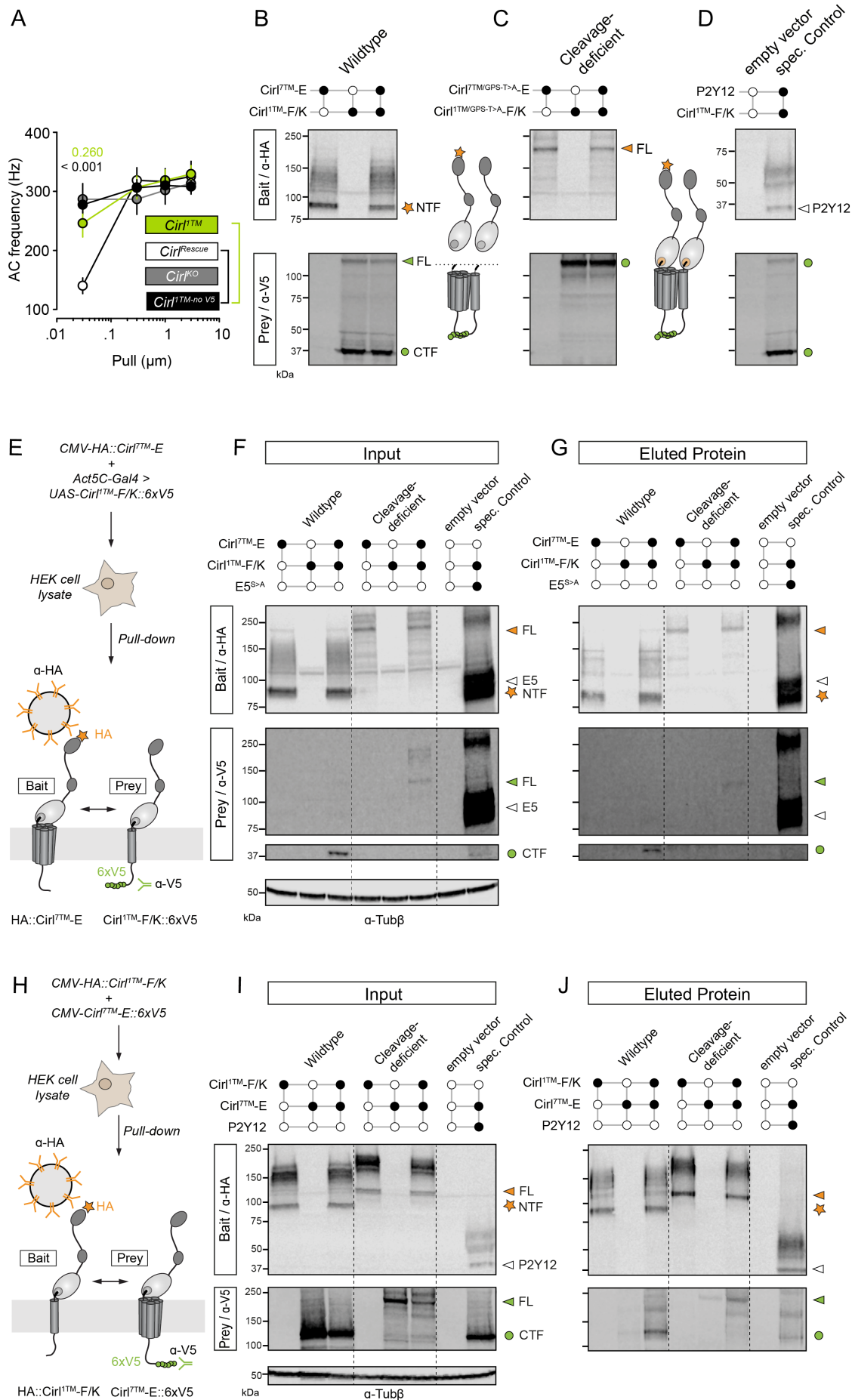


Figure S4. Cirl^{1TM} and Cirl^{7TM} co-immunoprecipitate in HEK cells when expressed using the UAS-GAL4 system

(A) Quantification of mechanically-evoked ICH5 AC frequencies (pull lengths 0.03 - 3 μ m). C-terminal addition of V5-tags has no impact on Cirl^{1TM} function. P-values denote statistical difference between *Cirl^{1TM-no V5}* and *Cirl^{Rescue}* (black) or *Cirl^{1TM}* (green) at 30 nm pull length. Data are shown as mean $\pm$ SEM (each genotype n = 10).

(B-D) Immunoblot input samples from the pull-down experiment shown in Figure 5E-G. Wildtype (B), cleavage-deficient Cirl (C) and Controls (D) were run on a gel side-by-side. Cirl^{7TM}-E bait (orange star/arrowhead, upper row). Cirl^{1TM}-F/K prey (green circle/arrowhead, lower row). HA-tagged P2Y12 served mock bait. Tubulin served as loading control (α -Tub β).

(E) Schematic of the co-immunoprecipitation experiment shown in (F and G). Note that Cirl^{1TM}-F/K::6xV5 prey is expressed using *UAS-Gal4* binary expression system⁴².

(F, G) Immunoblots of Cirl isoform input samples (F) and co-precipitation eluates (G). Cirl^{7TM}-E served as bait (upper row); FL protein is indicated by orange arrow, NTF with an orange star. Cirl^{1TM}-F/K prey (green circle/arrowhead, lower row). Cleavage-disabled HA-V5-tagged ADGRE5 served as mock bait (white arrowhead). Tubulin served as loading control (α -Tub β).

(H-J) Immunoblots of Cirl isoform input samples (I) and co-precipitation eluates (J). Cirl^{1TM}-F/K served as bait (upper row); FL protein is indicated by orange arrow, NTF with an orange star. Cirl^{7TM}-E prey (CTF = green circle; FL = arrowhead, lower row). HA-tagged P2Y12 served as mock bait (white arrowhead). Tubulin served as loading control (α -Tub β).

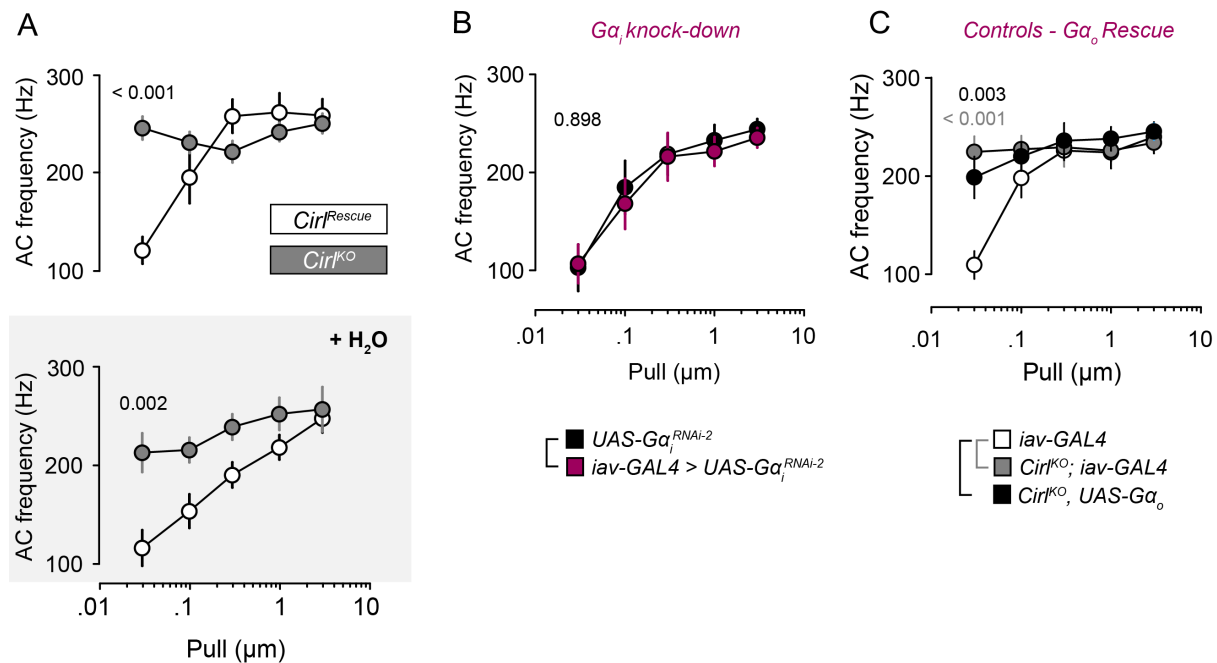


Figure S5. Sham treatment of $Cirr^{KO}$ lch5 leaves AC frequencies largely unaffected and overexpression of $G\alpha_o$ protein rescues mechano-discrimination deficit of $Cirr^{KO}$

(A) AC frequencies from mechanically stimulated lch5 neurons of $Cirr^{KO}$ animals before (A) and after sham treatment (+ H₂O) show no effect. Related to Figure 6B-D and to Table S11.

(B) AC frequencies from mechanically stimulated $G\alpha_i$ -deficient lch5 neurons ($iav-GAL4 > UAS-G\alpha_i^{RNAi-2}$). Lch5 neuron-specific $G\alpha_i$ knock-down leaves AC frequencies unaffected. P-value is derived from the statistical comparison of the AC frequencies (30 nm pull) between $iav-GAL4 > UAS-G\alpha_i^{RNAi-2}$ and $UAS-G\alpha_i^{RNAi-2}$. Related to Figure 6E and Table S14.

(C) AC frequencies from mechanically stimulated lch5 neurons of $Cirr^{KO}$ with a copy of $iav-GAL4$ (gray) or $UAS-G\alpha_o$ (black). Neither the presence of UAS nor GAL4 transgenes alone reduces AC frequencies. P-values are derived from the statistical comparisons of the AC frequencies (30 nm pull) between $iav-GAL4$ and $Cirr^{KO}$, $UAS-G\alpha_o$ (black) or $iav-GAL4$ and $Cirr^{KO}$, $iav-Gal4$ (gray). Data are shown as mean $\pm$ SEM (each genotype n = 10). Related to Figure 6G and Table S15.

Table S1. Collection of Cirl cDNA plasmids and their construction.

Receptor construct	Plasmid ID	Description	Tags	Construction/Reference	Primers
empty vector	pSA25	Empty pcDNA3.1 vector	-	-	-
empty vector	pNH366	Empty pHLSec vector	-	Gift from Björn Kieslich	
Cirl-B/H	pJT5	encodes isoform B/H	HA	-B/H ORF from pJT4 into pJT2 via <i>NheI</i> and <i>EcoRV</i>	-
Cirl-E	pJT2	encodes isoform E	HA	cloning of -E ORF (pJT1) in pcDNA3.1 (pSA25)	-
Cirl-F/K	pJT9	encodes isoform F/K	HA	SLIC: 7.6 kbp from pJT2 ORF + 1TM-specific alternative exon from pNH103	jt_11F/15R jt_14R/16F
Cirl-F/K ^{GPS/T>A}	pJT11	Cleavage-deficient isoform F/K	HA	site-directed mutagenesis of pJT9	mn_12F/13R
Cirl-E	pNH354	encodes isoform E	6xV5	Insertion of 6xV5 from pNH290 to -E ORF via <i>AgeI</i> and <i>EcoRI</i>	nh_588F/589R; nh_590F/591R
Cirl-E ^{GPS/T>A}	pNH356	Cleavage-deficient isoform E	6xV5	site-directed mutagenesis of pNH354	mn_12F/13R
Cirl-E ^{GPS/H>A}	pNH254	Cleavage-deficient isoform E	HA	site-directed mutagenesis of pJT2	mn_12F/13R
Cirl-E ^{GPS/H>A}	pNH226	Cleavage-deficient isoform E	HA	site-directed mutagenesis of pJT2	mn_38F/39R
Cirl-F/K	pAB24	encodes isoform F/K	6xV5	Removal of RBL domain fragment of pNH353 via <i>EcoRV</i>	-
Cirl-F/K ^{GPS/H>A}	pJT12	Cleavage-deficient isoform F/K	HA	site-directed mutagenesis of pJT9	mn_38F/39R
Cirl-F/K ^{GPS/T>A}	pAB25	Cleavage-deficient isoform F/K	6xV5	Removal of RBL domain fragment of pNH355 via <i>EcoRV</i>	-
UAS-Cirl-F/K	pNH288	Expression of Cirl-F/K under UAS control.	6xV5	GenScript synthesis	-
UAS-Cirl-F/K ^{GPS/T>A}	pNH322	Cleavage-deficient isoform F/K under UAS control.		site-directed mutagenesis of pNH288	mn_12F/13R
Cirl ^{1TM} -F/K ^{ΔCTF}	pAB23	Expression of Cirl ^{1TM} -F/K ^{ΔCTF} in pHLSec vector.	6xV5	Removal of RBL domain fragment of pAD22 via PCR and <i>AgeI</i> digest	ab_38R/39F
G6 ^{7TM}	pNH364	murine GPR126-7TM	HA	GenScript synthesis	-
G6 ^{1TM}	pNH363	murine GPR126-1TM	6xV5	GenScript synthesis	-
G6 ^{7TM} -GPS/T>A	pAB11	murine GPR126-7TM cleavage-deficient	HA	site-directed mutagenesis of pNH364	ab_10F/11R

G6^{1TM-GPS/T>A}	pAB9	murine GPR126-1TM cleavage deficient	6xV5	site-directed mutagenesis of pNH363	ab_10F/11R
P2Y12	pMIH57	human P2Y12	HA/FLAG	Gift from T. Schöneberg, transferred to pcDNA3.1	-
E5^{GPS/S>A}	pSA27	Cleavage-deficient human CD97/ADGRE5	HA/V5	Cloning of 1.3 kbp fragment of pSA14 into pSA26 via <i>EcoRV</i> and <i>SbfI</i>	-
Act5C-Gal4	-	Expression of GAL4 under Actin5C promoter.	-	Gift from Tobias Schwartz. Cloned into pUC18.	-

Table S2. Collection of primers used for cloning of cDNA constructs.

Name	Sequence (5'- 3' orientation)
nh_430F	ccgaccggtgatctctaccaaccatattgtcaatctctatgagcatatctgtagatctgtccaagtaccaaaccgctacgct
nh_431R	gggtgaccaacatgtcgccgtagagcggtt
nh_588F	ctagtctagatgaactagagggccctattctatag
nh_589R	ctagaccggtcttagccagtgggtccagataacat
nh_590F	ctagaccggtggaggcggtggcggggaaaaccca
nh_591R	ctagtctagatcacgtcgaatcgagtcgagcag
mn_12F	cagttgcaaccacctggcaactttgccatact
mn_13R	agtatggcaaagttgccaggtgggtgcaactg
jt_11F	tatcgccctgataacgctgaagctgttcaatggggtctttgtgaagg
jt_14R	cagcttcagcgttatcaggcgata
jt_15R	ctatagaataggccctctagttcaccaccaccacttatattgtaag
jt_16F	tgaactagagggccctattctatagtgac
ab_10F	gtctgtgcagccacttcgctcactttggag
ab_11R	ctccaaagtgcgaagtggctgcacagac
ab_38R	ctagaccggttcagctacgcaaccatc
ab_39F	ctagaccggtttccaagtcactcagcg

Table S3. Quantification of mean midpoint speed (N = 20, n=5/dish). Values represent the mean $\pm$ SEM, N denotes number of experiments. All groups were compared against *Cir1^{Rescue}* using an unpaired t-test.

Genotype	Mean midpoint speed (mm/s)	N	p-value
Related to Figure 3B			
<i>Cir1^{Rescue}</i>	0.90 $\pm$ 0.06	20	-
<i>Cir1^{KO}</i>	0.61 $\pm$ 0.02	20	< 0.001
<i>Cir1^{7TM}</i>	0.94 $\pm$ 0.05	20	0.670
<i>Cir1^{1TM}</i>	1.14 $\pm$ 0.05	20	0.004

Table S4. Quantification of AC frequencies of *lch5* neurons at different pull lengths (0.03-3 μ m).

Values represent the mean $\pm$ SEM response of 10 different *lch5* organs from 10 individual animals. AC frequencies at different pull lengths were compared using paired t-tests.

Genotype	Mean AC frequency at x μm pull in (Hz)					
Related to Figures 3F-H + S4						
	0.03 μm	0.1 μm	0.3 μm	1 μm	3 μm	p-value 0.03 vs 3 μm
<i>Cir1^{Rescue}</i>	140 ± 13	n.a.	319 ± 21	317 ± 22	325 ± 26	< 0.001
<i>Cir1^{KO}</i>	287 ± 26	n.a.	287 ± 26	302 ± 24	313 ± 18	0.251
<i>Cir1^{7TM}</i>	282 ± 17	n.a.	315 ± 14	323 ± 11	322 ± 12	0.020
<i>Cir1^{1TM}</i>	246 ± 23	n.a.	307 ± 19	319 ± 18	329 ± 14	0.012
<i>Cir1^{7TM+1TM}</i>	183 ± 14	247 ± 28	264 ± 22	273 ± 23	289 ± 23	0.002
<i>Cir1^{1TM} no V5</i>	277 ± 14	n.a.	306 ± 14	310 ± 16	308 ± 11	0.128

Table S5. Comparison of AC frequencies generated in response to 0.03 μ m pull. Values represent the mean response of 10 different *lch5* organs from 10 individual animals. All groups were compared using unpaired t-tests.

Genotype	Comparison of mean AC frequencies at 0.03 μm pull			
Related to Figure 3F				
		<i>Cir1^{Rescue}</i>	<i>Cir1^{KO}</i>	<i>Cir1^{7TM}</i>
		140 Hz	287 Hz	282 Hz
<i>Cir1^{Rescue}</i>	140 Hz	-	< 0.001	< 0.001
<i>Cir1^{KO}</i>	287 Hz	-	-	0.883
<i>Cir1^{7TM}</i>	282 Hz	-	-	-

Related to Figures 3G + S4					
		<i>Cir1^{Rescue}</i>	<i>Cir1^{KO}</i>	<i>Cir1^{1TM}</i>	<i>Cir1^{1TM} no V5</i>
		140 Hz	287 Hz	246 Hz	277 Hz
<i>Cir1^{Rescue}</i>	140 Hz	-	< 0.001	< 0.001	< 0.001
<i>Cir1^{KO}</i>	287 Hz	-	-	0.263	0.758
<i>Cir1^{1TM}</i>	246 Hz	-	-	-	0.260
<i>Cir1^{1TM} no V5</i>	277 Hz	-	-	-	-
Related to Figure 3H					
		<i>Cir1^{Rescue}</i>	<i>Cir1^{KO}</i>	<i>Cir1^{7TM+1TM}</i>	
		151 Hz	285 Hz	183 Hz	
<i>Cir1^{Rescue}</i>	151 Hz	-	< 0.001	0.243	
<i>Cir1^{KO}</i>	285 Hz	-	-	< 0.001	
<i>Cir1^{7TM+1TM}</i>	183 Hz	-	-	-	

Table S6. Quantification of Ca²⁺ signals from the five larval *lch5* neurons of *Cir1^{Rescue}* at different sound-pressure levels (75 – 90 dB). Values represent the mean $\pm$ SEM response of different neurons from N individual animals. Ca²⁺ signals below $\Delta F/F_0 < 0.01$ are not shown.

Ca ²⁺ signal as $\Delta F/F_0$										
Related to Figure 4C										
<i>Cir^l</i> ^{Rescue}	Neuron 1		Neuron 2		Neuron 3		Neuron 4		Neuron 5	
	$\Delta F/F_0$	N	$\Delta F/F_0$	N	$\Delta F/F_0$	N	$\Delta F/F_0$	N	$\Delta F/F_0$	N
75 dB	0.03 ± 0.01	10	0.05 ± 0.02	4	< 0.01	-	0.05 ± 0.05	5	< 0.01	-
80 dB	0.09 ± 0.03	10	0.10 ± 0.03	6	0.01 ± 0.01	8	0.07 ± 0.04	5	< 0.01	-
85 dB	0.18 ± 0.03	10	0.22 ± 0.06	6	0.04 ± 0.02	8	0.14 ± 0.05	5	< 0.01	-
90 dB	0.73 ± 0.09	10	0.51 ± 0.10	6	0.07 ± 0.02	8	0.43 ± 0.13	5	< 0.01	-

Table S7. Quantification of Ca²⁺ signals from neuron 1 of larval *lch5* in different genotypes, at different sound-pressure levels (75 – 90 dB). Values represent mean responses ± SEM of 10 *lch5* organs from 10 individual animals. Mean $\Delta F/F_0$ of *Cir^l*^{Rescue} and mutant genotypes at 75 dB were compared using unpaired t-tests, unless stated otherwise (*).

Neuron 1	Ca ²⁺ signal as ΔF/F ₀				
Related to Figure 4D,E,F					
Genotype	75 dB	p-value	80 dB	85 dB	90 dB
<i>Cir^l</i> ^{Rescue}	0.03 ± 0.01	-	0.09 ± 0.03	0.18 ± 0.03	0.73 ± 0.09
<i>Cir^l</i> ^{KO}	0.17 ± 0.05	0.008	0.26 ± 0.06	0.40 ± 0.07	1.17 ± 0.20
<i>Cir^l</i> ^{7TM}	0.15 ± 0.05	0.064*	0.28 ± 0.07	0.48 ± 0.08	1.26 ± 0.19
<i>Cir^l</i> ^{1TM}	0.13 ± 0.04	0.018	0.23 ± 0.04	0.42 ± 0.05	1.11 ± 0.20
<i>Cir^l</i> ^{7TM+1TM}	0.10 ± 0.03	0.035	0.17 ± 0.05	0.28 ± 0.06	1.01 ± 0.17
*Mann-Whitney test					

Table S8. Quantification of Ca²⁺ signals from *lch5* neurons 2 and 4 in *Cir^l*^{Rescue} and *Cir^l*^{KO} larvae at different sound-pressure levels (75 – 90 dB). Values represent the mean ± SEM response of different neurons from N individual animals. Mean $\Delta F/F_0$ of *Cir^l*^{Rescue} and *Cir^l*^{KO} at 75 dB were compared using unpaired t-tests, unless stated otherwise (*).

Ca ²⁺ signal as $\Delta F/F_0$									
Related to Figure 4G,H									
Neuron 2	75 dB	N	p-value	80 dB	N	85 dB	N	90 dB	N
<i>Cir^l</i> ^{Rescue}	0.05 ± 0.02	4	-	0.10 ± 0.03	6	0.22 ± 0.07	6	0.51 ± 0.10	6
<i>Cir^l</i> ^{KO}	0.14 ± 0.05	7	0.186	0.23 ± 0.05	6	0.19 ± 0.05	7	0.74 ± 0.16	7
Neuron 4									
<i>Cir^l</i> ^{Rescue}	0.05 ± 0.05	5	-	0.07 ± 0.04	5	0.14 ± 0.05	5	0.43 ± 0.13	5
<i>Cir^l</i> ^{KO}	0.09 ± 0.02	5	0.151*	0.11 ± 0.03	5	0.11 ± 0.03	5	0.46 ± 0.09	5
*Mann-Whitney test									

Table S9. Quantification of expression of certain *Cir^l* isoforms via ELISA. Optical density was measured at 490 nm. P-values derived from statistical comparison to empty vector sample. Data represent the mean ± SEM (N=3, n=8) and were compared using unpaired t-tests, unless stated otherwise (*).

Genotype	Cell surface expression	n	p-value
Related to Figure 5A			
<i>Empty vector/pcDNA3.1</i>	0.043 ± 0.001	24	-
<i>HA::Cirl-B/H</i>	0.36 ± 0.020	24	< 0.001
<i>HA::Cirl-E</i>	0.32 ± 0.018	24	< 0.001
<i>HA::Cirl-F/K</i>	0.63 ± 0.022	24	< 0.001

Genotype	Total expression	n	p-value
Related to Figure 5A			
<i>Empty vector/pcDNA3.1</i>	0.052 ± 0.001	24	-
<i>HA::Cirl-B/H</i>	0.59 ± 0.048	24	< 0.001*
<i>HA::Cirl-E</i>	0.32 ± 0.042	24	< 0.001*
<i>HA::Cirl-F/K</i>	0.70 ± 0.041	24	< 0.001*

*Mann-Whitney test

Genotype	Cell surface expression	n	p-value
Related to Figure 5C			
<i>HA::Cirl-E</i>	0.196 ± 0.011	24	-
<i>HA::Cirl-E^{GPS-H>A}</i>	0.281 ± 0.010	24	< 0.001
<i>HA::Cirl-E^{GPS-T>A}</i>	0.426 ± 0.017	24	< 0.001*
Related to Figure 5C			
<i>HA::Cirl-F/K</i>	0.570 ± 0.017	24	-
<i>HA::Cirl-F/K^{GPS-H>A}</i>	0.525 ± 0.017	24	0.070
<i>HA::Cirl-F/K^{GPS-T>A}</i>	0.593 ± 0.017	24	0.200*

*Mann-Whitney test

Genotype	Total expression	n	p-value
Related to Figure 5C			
<i>HA::Cirl-E</i>	0.376 ± 0.020	24	-
<i>HA::Cirl-E^{GPS-H>A}</i>	0.495 ± 0.019	24	< 0.001
<i>HA::Cirl-E^{GPS-T>A}</i>	0.606 ± 0.020	24	< 0.001
Related to Figure 5C			
<i>HA::Cirl-F/K</i>	0.664 ± 0.018	24	-
<i>HA::Cirl-F/K^{GPS-H>A}</i>	0.726 ± 0.011	24	< 0.001*
<i>HA::Cirl-F/K^{GPS-T>A}</i>	0.721 ± 0.017	24	0.008*

*Mann-Whitney test

Table S10. Pharmacological inhibition of ACyc. Quantification of lch5 AC frequencies at different pull lengths (0.03 - 3 μm) before and after a 10 min treatment with 100 μM adenylyl cyclase inhibitor SQ22536. Values represent mean responses ± SEM of 10 different lch5 organs from 10 individual animals. Mean frequencies of mutant genotypes at 0.03 μm pulls were compared to mean frequencies at the same pull length in *Cirl^{Rescue}* using unpaired t-tests.

Genotype	Mean AC frequency at x pull in (Hz)					
Related to Figure 6B-D						
	0.03 μm	p-value	0.1 μm	0.3 μm	1 μm	3 μm
	0 min					
<i>Cir1^{Rescue}</i>	133 ± 14	-	207 ± 27	269 ± 17	273 ± 19	270 ± 17
<i>Cir1^{KO}</i>	257 ± 20	< 0.001	245 ± 13	257 ± 15	269 ± 11	273 ± 14
<i>Cir1^{7TM}</i>	270 ± 9	< 0.001	245 ± 16	251 ± 11	259 ± 14	273 ± 15
<i>Cir1^{1TM}</i>	270 ± 14	< 0.001	263 ± 13	263 ± 13	259 ± 11	261 ± 9
	10 min after addition of adenylyl cyclase inhibitor					
<i>Cir1^{Rescue}</i>	139 ± 18	-	176 ± 17	213 ± 13	241 ± 13	270 ± 14
<i>Cir1^{KO}</i>	139 ± 8	0.973	193 ± 11	230 ± 10	257 ± 9	286 ± 11
<i>Cir1^{7TM}</i>	158 ± 10	0.366	175 ± 16	219 ± 22	242 ± 24	259 ± 16
<i>Cir1^{1TM}</i>	175 ± 21	0.209	201 ± 17	215 ± 22	239 ± 20	271 ± 15

Table S11. Control experiment for pharmacological inhibition of ACyc. Quantification of *lch5* AC frequencies at different pull lengths (0.03 - 3 μm) before and after a 10 min sham treatment to exclude an unspecific effect of the 10 min incubation. Values represent mean responses $\pm$ SEM of 10 *lch5* organs from 10 individual animals. Mean frequencies of *Cir^{KO}* and *Cir^{Rescue}* at 0.03 μm pulls before and after treatment (sham treatment and 100 μM adenylyl cyclase inhibitor SQ22536 treatment, respectively) were compared using unpaired t-tests. Mean frequencies of *Cir^{KO}* at 0.03 μm pulls before and after sham treatment were due to non-normal data distribution compared using a Wilcoxon Signed Rank test (*).

Genotype	Mean AC frequency at X pull in (Hz)						
Related to Figure S5A							
	0.03 μm	p-value <i>Cir^{lKO}</i> vs. <i>Cir^{lRescue}</i>	0.1 μm	0.3 μm	1 μm	3 μm	p-value 0.03 μm t ₀ vs. t _{10min}
	0 min						
<i>Cir^{lKO}</i>	257 ± 12	< 0.001	243 ± 11	233 ± 11	253 ± 9	262 ± 10	-
	10 min after adding sham control						
<i>Cir^{lKO}</i>	235 ± 20	0.002	238 ± 13	261 ± 13	275 ± 16	279 ± 23	0.652

Table S12. Mean AC frequency at different pull lengths in G protein knock-down. Quantification of AC frequencies of *lch5* neurons at different pull lengths (0.03-3 μm). Values represent the mean $\pm$ SEM response of 10 *lch5* organs from 10 individual animals. AC frequencies at different pull lengths were compared using paired t-tests.

Genotype	Mean AC frequency at x μm pull in (Hz)					
Related to Figure 6E, F						
	0.03 μm	0.1 μm	0.3 μm	1 μm	3 μm	p-value 0.03 vs 3 μm
<i>iav-Gal4</i>	121 $\pm$ 16	179 $\pm$ 32	235 $\pm$ 27	287 $\pm$ 22	279 $\pm$ 16	< 0.001
<i>UAS-Gα^{RNAi}</i>	114 $\pm$ 13	211 $\pm$ 27	228 $\pm$ 20	242 $\pm$ 12	248 $\pm$ 7	< 0.001

<i>UAS-Gα_i^{RNAi}</i>	132 ± 17	206 ± 24	231 ± 20	239 ± 15	249 ± 9	< 0.001
<i>iav-Gal4>UAS-Gα_o^{RNAi}</i>	242 ± 22	254 ± 23	245 ± 19	247 ± 14	249 ± 13	0.755
<i>iav-Gal4>UAS-Gα_i^{RNAi}</i>	125 ± 20	227 ± 30	243 ± 30	256 ± 22	263 ± 12	< 0.001

Table S13. Mean AC frequencies at 0.03 μm pull lengths in G protein knock-down. Comparison of AC frequencies generated by a 0.03 μm pull. Values represent the mean response of 10 lch5 organs from 10 individual animals. Unless stated otherwise (*), all groups were compared using unpaired t-tests.

Genotype		Comparison of mean AC frequencies at 0.03 μ m pull				
Related to Figure 6E, F						
		<i>iav-Gal4/+</i>	<i>UAS-Gα_o^{RNAi}</i>	<i>UAS-Gα_i^{RNAi}</i>	<i>iav-Gal4>UAS-Gα_o^{RNAi}</i>	<i>iav-Gal4>UAS-Gα_i^{RNAi}</i>
		121 Hz	114 Hz	132 Hz	242 Hz	125 Hz
<i>iav-Gal4</i>	121 Hz	-	0.747	0.638	< 0.001	0.858
<i>UAS-Gα_o^{RNAi}</i>	114 Hz	-	-	0.495*	< 0.001	0.638
<i>UAS-Gα_i^{RNAi}</i>	132 Hz	-	-	-	< 0.001	0.805
<i>iav-Gal4>UAS-Gα_o^{RNAi}</i>	242 Hz	-	-	-	-	< 0.001
<i>iav-Gal4>UAS-Gα_i^{RNAi}</i>	125 Hz	-	-	-	-	-
*Mann-Whitney test						

Table S14. Mean AC frequency at different pull lengths in a second Gα_i protein knock-down. Quantification of AC frequencies of lch5 neurons at different pull lengths (0.03-3 μm). Values represent the mean ± SEM response of 10 lch5 organs from 10 individual animals. AC frequencies of both genotypes at 0.03 μm pulls were compared using an unpaired t-test (third column). AC frequencies at 0.03 and 3 μm in the same genotype were compared using paired t-tests (last column).

Genotype	Mean AC frequency at x μm pull in (Hz)						
Related to Figure S5B							
	0.03 μm	p-value	0.1 μm	0.3 μm	1 μm	3 μm	p-value 0.03 vs 3 μm
<i>UAS-Gα_i^{RNAi-2}</i>	103 ± 24	-	185 ± 27	219 ± 21	233 ± 16	244 ± 11	< 0.001
<i>iav-Gal4>UAS-Gα_i^{RNAi-2}</i>	107 ± 20	0.898	168 ± 26	216 ± 24	221 ± 15	235 ± 10	< 0.001

Table S15. Cell-specific $G\alpha_o$ rescue. Quantification of AC frequencies of *lch5* neurons at different pull lengths (0.03-3 μm). Values represent the mean $\pm$ SEM response of 10 *lch5* organs from 10 individual animals. Unless stated otherwise (*), AC frequencies at 0.03 and 3 μm in the same genotype were compared using paired t-tests.

Genotype	Mean AC frequency at x μm pull in (Hz)					
Related to Figures 6G + S5C						
	0.03 μm	0.1 μm	0.3 μm	1 μm	3 μm	p-value 0.03 vs 3 μm
<i>;; iav-Gal4</i>	109 $\pm$ 14	198 $\pm$ 20	226 $\pm$ 16	224 $\pm$ 16	240 $\pm$ 15	< 0.001
<i>;Cirl^{KO}; iav-Gal4</i>	225 $\pm$ 16	227 $\pm$ 14	229 $\pm$ 13	226 $\pm$ 11	234 $\pm$ 11	0.609
<i>;Cirl^{KO}, UAS-Gα_o;</i>	199 $\pm$ 21	220 $\pm$ 15	236 $\pm$ 18	238 $\pm$ 12	245 $\pm$ 8	0.074*
<i>Cirl^{KO}, UAS-Gα_o; iav-Gal4</i>	127 $\pm$ 18	190 $\pm$ 28	210 $\pm$ 18	237 $\pm$ 11	244 $\pm$ 9	< 0.001
*Wilcoxon Signed Rank test						

Table S16. Cell-specific $G\alpha_o$ rescue. Comparison of AC frequencies generated at 0.03 μm pull. Values represent mean responses of 10 *lch5* organs from 10 individual animals. All groups were compared using unpaired t-tests.

Genotype		Comparison of mean AC frequencies at 0.03 μm pull			
Related to Figures 6G + S5C					
		<i>;; iav-Gal4</i>	<i>Cirl^{KO}; iav-Gal4</i>	<i>;Cirl^{KO}, UAS-Gα_o;</i>	<i>;Cirl^{KO}, UAS-Gα_o; iav-Gal4</i>
		109 Hz	225 Hz	199 Hz	127 Hz
<i>;; iav-Gal4</i>	109 Hz	-	< 0.001	0.003	0.454
<i>;Cirl^{KO}; iav-Gal4</i>	225 Hz	-	-	0.339	< 0.001
<i>;Cirl^{KO}, UAS-Gα_o;</i>	199 Hz	-	-	-	0.018
<i>;Cirl^{KO}, UAS-Gα_o; iav-Gal4</i>	127 Hz	-	-	-	-