

<sup>1</sup> AI reveals insights into link between CD33 and cognitive  
<sup>2</sup> impairment in Alzheimer's Disease

<sup>3</sup> Tamara Raschka<sup>1,2,3,\*</sup>, Meemansa Sood<sup>1,2</sup>, Bruce Schultz<sup>1</sup>, Aybuge Altay<sup>1,2,4</sup>,  
<sup>4</sup> Christian Ebeling<sup>1</sup>, and Holger Fröhlich<sup>1,2,\*</sup>

<sup>5</sup> <sup>1</sup>Department of Bioinformatics, Fraunhofer Institute for Algorithms and  
<sup>6</sup> Scientific Computing (SCAI), Sankt Augustin, Germany

<sup>7</sup> <sup>2</sup>Bonn-Aachen International Center for Information Technology (B-IT),  
<sup>8</sup> University of Bonn, Bonn, Germany

<sup>9</sup> <sup>3</sup>Fraunhofer Center for Machine Learning, Germany

<sup>10</sup> <sup>4</sup>Current Address: Department of Computational Molecular Biology, Max Planck  
<sup>11</sup> Institute for Molecular Genetics, Berlin, Germany

<sup>12</sup> <sup>\*</sup>Corresponding authors

<sup>13</sup> E-Mail: tamara.raschka@scai.fraunhofer.de (TR), holger.froehlich@scai.fraunhofer.de (HF)

<sup>14</sup> **Abstract**

<sup>15</sup> Modeling biological mechanisms is a key for disease understanding and drug-target identifica-  
<sup>16</sup> tion. However, formulating quantitative models in the field of Alzheimer's Disease is challenged  
<sup>17</sup> by a lack of detailed knowledge of relevant biochemical processes. Additionally, fitting dif-  
<sup>18</sup> ferential equation systems usually requires time resolved data and the possibility to perform  
<sup>19</sup> intervention experiments, which is difficult in neurological disorders. This work addresses these  
<sup>20</sup> challenges by employing the recently published Variational Autoencoder Modular Bayesian Net-  
<sup>21</sup> works (VAMBN) method, which we here trained on combined clinical and patient level gene  
<sup>22</sup> expression data while incorporating a disease focused knowledge graph. Our approach, called  
<sup>23</sup> iVAMBN, resulted in a quantitative model that allowed us to simulate a down-expression of  
<sup>24</sup> the putative drug target CD33, including potential impact on cognitive impairment and brain  
<sup>25</sup> pathophysiology. Experimental validation demonstrated a high overlap of molecular mechanism  
<sup>26</sup> predicted to be altered by CD33 perturbation with cell line data. Altogether, our modeling  
<sup>27</sup> approach may help to select promising drug targets.

## **28 Author Summary**

**29** For the last 20 years the field of Alzheimer's Disease (AD) is marked by a series of continuous  
**30** failures to deliver demonstrably effective medications to patients. This is also highlighted by the  
**31** highly controversial recent approval of Aduhelm (Biogen) by the FDA, which is now investigated  
**32** internally due to the lack of clear efficacy.

**33** One of the reasons for the continuous failure of trials in AD is the choice of the wrong target  
**34** mechanism. In essence there is a lack of understanding, how targeting a certain molecule would  
**35** affect cognitive impairment in human. One way to address this issue is the development of  
**36** quantitative system level models connecting the molecular level with the phenotype. However,  
**37** formulating such models in the field of Alzheimer's Disease is challenged by a lack of detailed  
**38** knowledge of relevant biochemical processes and the connection of molecular mechanisms to  
**39** cognitive impairment. Additionally, fitting of differential equation systems, which are often  
**40** used in systems biology, requires time resolved data and the possibility to perform intervention  
**41** experiments, which is difficult in neurological disorders due to the lack of realistic model systems.

**42** Our work addresses these challenges by employing a novel hybrid Artificial Intelligence (AI)  
**43** approach combining variational autoencoders with Bayesian Networks. Our proposed approach,  
**44** named Integrative Variational Autoencoder Modular Bayesian Networks (iVAMBN), was trained  
**45** on combined clinical and patient level gene expression data while incorporating a disease focused  
**46** knowledge graph. Our method resulted in an interpretable, quantitative model. It showed  
**47** connections between various biological mechanisms playing a role in AD. Furthermore, iVAMBN  
**48** directly connected the molecular level to the disease phenotype. Our model allowed us to simulate  
**49** a down-expression of the putative drug target CD33. Results showed a significantly increased  
**50** cognition and predicted perturbation of a number of biological mechanisms. We experimen-  
**51** tally validated these predictions using gene expression data from a knock-out THP-1 monocyte  
**52** cell line. This experiment confirmed our model predictions up to a very high extend. To our  
**53** knowledge we thus developed the first experimentally validated, quantitative, multi-scale model  
**54** connecting molecular mechanisms with clinical outcomes in the AD field.

## 55 Introduction

56 Alzheimer's Disease (AD) is a neurodegenerative disorder affecting about 50 million people world-  
57 wide, resulting in the inability to perform necessary, daily activities before leading to an often  
58 early death [1]. Despite decades of research and more than 2000 clinical studies listed on Clin-  
59 icalTrials.gov, to date there is no cure, and all existing treatments are purely symptomatic [1].  
60 New disease modifying treatments are urgently needed, but require a better mechanistic under-  
61 standing of the disease.

62 A common starting point in this context is to map out the existing knowledge landscape  
63 about the disease. In the past few decades, a large number of databases have been developed  
64 in the bioinformatics community, such as databases for biological pathways (like KEGG [2],  
65 PathwayCommons [3], WikiPathways [4], Reactome [5]), drug-target interactions (like Open-  
66 Targets [6], Therapeutic Targets Database [7]), disease-gene associations (like DisGeNET [8])  
67 or protein-protein interactions (like STRING [9], IntAct [10]). All these databases simplify the  
68 usage of the respective knowledge for algorithms and models, especially in the field of drug tar-  
69 get identification. Moreover, none of these databases have been compiled in a disease focused  
70 manner. The Biological Expression Language (BEL) provides this opportunity and can be used  
71 to represent literature-derived, disease focused knowledge in the form of attributed graphs in a  
72 precise manner. For AD a knowledge graph has been published in [11] and represents the manu-  
73 ally curated, disease focused mechanistic interplay between genetic variants, proteins, biological  
74 processes and pathways described in the literature, enabling the user to computationally query  
75 and integrate knowledge graphs into drug target identification algorithms.

76 One of the interesting molecules in the AD field is CD33, a transmembrane receptor protein  
77 expressed primarily in myeloid lineage cells. It has been associated with decreased risk of AD  
78 in GWAS studies [12–18] and discussed as a potential therapeutic target, for example via im-  
79 munotherapy [14]. In an AD mouse model, a knockout of CD33 mitigated amyloid- $\beta$  clearance  
80 and improved cognition [13, 17, 18]. Similarly, a positive effect on amyloid- $\beta$  phagocytosis could  
81 be observed in CD33 knock-out THP-1 macrophages [16]. In humans a correlation between  
82 CD33, cognition and amyloid clearance is known, however, the concrete underlying mechanisms  
83 are still not well understood. There is an ongoing clinical trial that is testing the effects of a CD33

84 inhibitor in patients with mild to moderate AD (NCT03822208). Along those lines, the EU-wide  
85 PHAGO project (<https://www.phago.eu>) funded via the Innovative Medicines Initiatives aimed  
86 to develop tools and methods to study the functioning of CD33 and related pathways in AD in  
87 order to facilitate decisions about potential drug development programs.

88 While graphs are useful for describing the disease focused knowledge landscape about AD, the  
89 principal incompleteness of disease focused biological knowledge may result in disagreements to  
90 observed data. Moreover, graphs do not allow to produce quantitative insights and predictions.  
91 For this purpose ordinary (ODEs) and partial differential equations (PDEs) are frequently used  
92 in systems biology and systems medicine, as they are able to describe biological mechanisms in  
93 a quantitative way. However, their formulation requires a detailed understanding of biochemical  
94 reactions, which in the AD field is only available for specific processes, like for example amyloid- $\beta$   
95 aggregation [19, 20]. Moreover, fitting differential equations usually requires time resolved data  
96 and the possibility to perform intervention experiments (as knock-downs or stimulation), which  
97 is challenged by the fact that cell lines and mouse models in the AD field can most likely only  
98 mimic specific aspects of the human disease [21–23].

99 A principle alternative to differential equation systems are probabilistic graphical models and  
100 in particular Bayesian Networks (BNs), which are quantitative as well. However, standard BN  
101 implementations require normally or multinomially distributed data, which is not the case in  
102 many applications. Furthermore, structure learning of BNs is an NP hard problem, where the  
103 number of possible network structures grows super-exponentially with the number of nodes in  
104 the network [24]. Hence, modeling higher dimensional data with a BN raises severe concerns  
105 regarding structure identifiability.

106 Altogether, these challenges lead to the fact that the AD field lacks a comprehensive quanti-  
107 tative model of the interplay between relevant molecules and biological processes, including the  
108 role of CD33, up to the phenotype level.

109 In this work, we developed a - to our knowledge - first quantitative, multi-scale model fo-  
110 cused on the multitude of mechanisms governing the CD33 molecule. Our model spans a variety  
111 of modalities, including gene expression, brain pathophysiology, demographic information and  
112 cognition scores. To address the challenges mentioned before, we started with a disease focused  
113 knowledge graph reconstruction, which we clustered into modules to significantly reduce dimen-

114 sionality. In the following we use the term "module" to denote a set of objects grouped together.  
115 Subsequently, we relied on our recently published Variational Autoencoder Modular Bayesian  
116 Network (VAMBN) algorithm [25], which is a hybrid Artificial Intelligence (AI) approach com-  
117 bining variational autoencoders [26] with modular Bayesian Networks [27], which is able to model  
118 arbitrary statistical distributions. We trained VAMBN on joint clinical and patient level gene ex-  
119 pression data while employing a clustered knowledge graph reflecting incomplete prior knowledge  
120 about disease mechanisms and their interplay. A simulated knock-down of CD33 and predicted  
121 downstream effects could be experimentally validated with gene expression data from a cell line.  
122 Overall, we believe that our work helps to move closer towards a systemic and quantitative un-  
123 derstanding of the disease, which is the prerequisite for finding urgently needed novel therapeutic  
124 options.

## 125 Results

126 In this work, we relied on AD patient data from the Religious Orders Study and Memory and  
127 Aging Project (ROSMAP) [28–30] for model training and the Mayo RNAseq Study (Mayo) [31]  
128 for external validation. An overview about clinical characteristics of the patients in these studies  
129 can be found in Table 1.

### 130 Overview about Modeling Strategy

131 Figure 1 shows an overview about our modeling strategy, which we call integrative VAMBN  
132 (iVAMBN), combining clinical and patient-level gene expression data with disease focused knowl-  
133 edge graphs. The first step of our workflow compiles an AD focused knowledge graph describing  
134 cause and effect relationships between biological processes, genes and pathologies. The generated  
135 graph consisted of 383 nodes and 607 edges. The graph was subsequently clustered into mod-  
136 ules with the help of the Markov Clustering algorithm [32] to significantly reduce the number of  
137 variables for subsequent modeling steps. Genes within modules were annotated with AD disease  
138 mechanisms coming from the NeuroMMSig gene set collection [33].

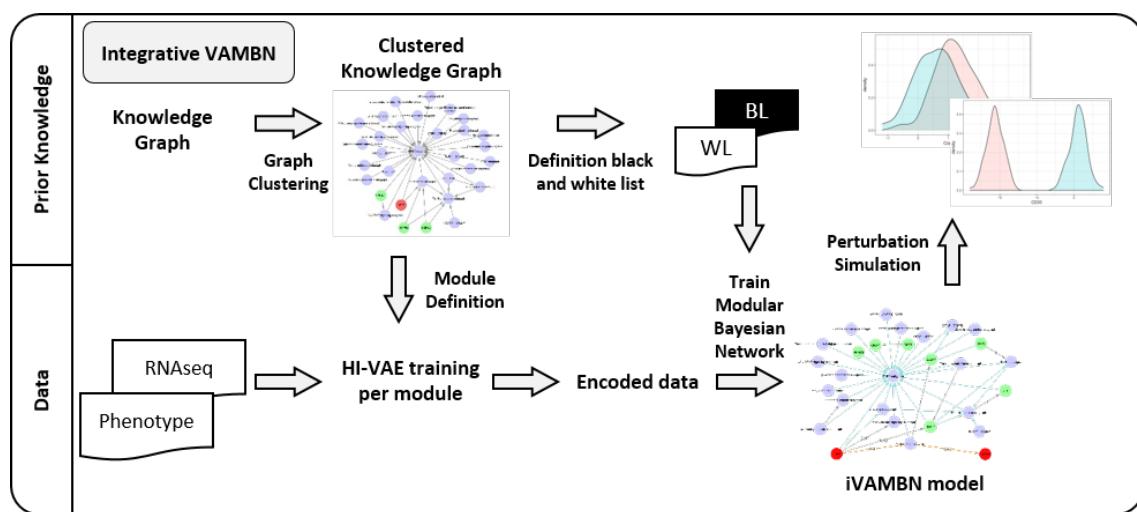
139 Using patient-level clinical and gene expression data from post-mortem cerebral cortex tis-  
140 sues, in a second step the VAMBN algorithm was employed to quantitatively model relation-

Table 1: **Patient statistics.** Shown are the number of patients, their age in years (with mean and standard deviation), sex, APOE genotype (binary encoding for at least one present E4 allele), MMSE score (with mean and sd) and Braak stage.

	ROSMAP	Mayo
no. patients	221	82
age	87.95 $\pm$ 3.38	82.66 $\pm$ 7.61
sex		
male	68	33
female	153	49
APOE		
0	138	39
1	83	43
MMSE	13.16 $\pm$ 8.38	-
Braak		
1	7	-
2	6	-
3	42	-
4	71	6
5	88	35
6	7	41

ships between gene modules as well as phenotype related scores (Mini-Mental State Examination (MMSE), Braak staging) and demographic features based on ROSMAP data. ROSMAP was chosen for training of the algorithm, because of the comparably large number of patients (more than 200) and available MMSE plus Braak scores. VAMBN takes as input patient-level data hierarchically organized into pre-defined modules (here: either gene modules or a phenotype related module including i.e. MMSE plus Braak stages), original features (here: demographic and clinical variables like age, sex, APOE genotype, and brain region) and prior knowledge regarding their possible connections. The output is a probabilistic graphical model describing connections between modules and original features. There is a per-patient score for each module, and each of these scores can be further decoded into feature-level gene expression and phenotype data, respectively.

In the third step of our strategy we evaluated, whether our iVAMBN model could also explain gene expression data from the Mayo study. Notably, at this step we only considered the Braak stage in the phenotype module, because the Mayo study does not report MMSE scores. For that purpose we first re-trained our iVAMBN model on ROSMAP while leaving out MMSE scores and then assessed the marginal log-likelihood of the modified model on the Mayo dataset. We



**Figure 1: The Integrative VAMB (iVAMB) approach.** The iVAMB approach integrates gene expression data, clinical and patho-physiological (phenotype) measures (bottom left) into a joint quantitative, probabilistic graphical model. The method initially uses a knowledge graph (top left) for defining modules and for informing about potential connections between them. In a second step, a representation of each module using a Heterogeneous Incomplete Variational Autoencoder (HI-VAE) is learned. In a third step a modular Bayesian Network between autoencoded modules is learned while taking into account the information derived from the knowledge graph. Finally, the iVAMB model is used to simulate gene perturbation (top right).

157 then tested the marginal log-likelihood of the true model against randomly permuted versions  
 158 of the learned probabilistic graph. This allowed us to assess, in how far the model learned on  
 159 ROSMAP could explain Mayo data better than expected by pure chance.

160 For the last step, we used our iVAMB model trained on ROSMAP to simulate several  
 161 therapeutic interventions, including a CD33 inhibition. Based on available data, we were able  
 162 to experimentally validate the predicted effects of a CD33 inhibition using CD33 knockout gene  
 163 expression data from a THP-1 monocyte cell line. More details about the entire iVAMB  
 164 approach can be found in the Methods section of this paper.

165 In the following we elaborate on the results obtained in each of these different steps, while  
 166 technical details are provided in the Methods part of this article.

## 167 Knowledge Graph Compilation

168 As outlined in the previous section, our modeling approach started with the compilation and  
169 Markov clustering of a knowledge graph. The Markov clustering resulted in 32 modules, including  
170 4 single gene modules, namely CD33, HSPB2, HSPB3, and MIR101-1. Most of the non-single  
171 gene modules comprised only two genes, while others had multiple combinations, like the GABA  
172 subgraph module with 289 genes. The exact number of genes clustered together as well as the  
173 result of a statistical over-representation analysis (hypergeometric test) using the AD focused  
174 gene set collection NeuroMMSig [33] can be found in Supplementary Table S1. A complete list  
175 of molecules within each module can be found in Supplementary Table S2. The modules were  
176 considered as nodes of a graph between them, where an edge was set between modules  $M_1, M_2$ ,  
177 if in the original knowledge graph there was at least one gene in  $M_1$  and one in  $M_2$  that was  
178 connected via a directed path. The resulting (acyclic) module graph is shown in Figure S1.

## 179 Integrative Variational Autoencoder Modular Bayesian Network Model

180 Integrative VAMBN combines the advantages of Bayesian Networks with the capabilities of  
181 variational autoencoders, more specifically Heterogeneous Incomplete Variational Autoencoders  
182 (HI-VAEs) [34]. Briefly, the idea is to initially learn a low dimensional Gaussian representation  
183 of features mapping to each of the defined modules. HI-VAEs differ from classical variational  
184 autoencoders in the sense that they can be applied to heterogeneous input data of different  
185 numerical scales, potentially containing missing values. In a second step a Bayesian Network  
186 structure is then learned over the low dimensional representations of modules, resulting in a  
187 modular Bayesian Network. More details are presented in the Methods part of this paper and  
188 in [25].

189 We here trained an iVAMBN model using the identified modules (i.e. feature groups in the  
190 original data) as - potentially multivariate - nodes of a probabilistic graphical model. Note-  
191 worthy exceptions are described in detail in Supplementary Note S1. In cases where multiple  
192 features map to one and the same module (i.e. the corresponding node / random variable in  
193 the probabilistic graphical model is multivariate), our method initially learns a low dimensional  
194 representation using a HI-VAE. Second, we learned the Bayesian Network structure connecting

195 these modules. At this stage it is possible to provide information about possible connections  
196 between modules given in the knowledge derived module graph (Supplementary Figure S1). We  
197 tested three different strategies to incorporate the information provided in the module graph:

198 • *completely data driven*: the entire Bayesian Network was only learned from data,  
199 • *knowledge informed*: the module graph was either used to only initialize Bayesian Network  
200 structure learning, to enforce / white list the existence of specific edges, or used for a  
201 combination of both, and  
202 • *completely knowledge driven*: strictly constrain edges between modules to those provided  
203 via the module graph, and additionally learned ones are only allowed to connect cognition  
204 scores, patho-physiological stages, and demographic features. All other possible edges are  
205 black listed, i.e. not allowed.

206 A systematic comparison of these strategies via a cross-validation yielded a better performance  
207 of the second strategy (knowledge informed), in which we used the module graph to white list  
208 edges and to initialize a greedy hill climbing based structure learning, see details in Methods  
209 Section and Supplementary Note S2. That means, iVAMBN was allowed to add additional  
210 edges, if the data provided according evidence.

211 We repeated the knowledge informed modular Bayesian Network learning 1000 times on  
212 random bootstrap sub-samples of the data drawn with replacement, hence allowing to quantify  
213 the statistical confidence of each inferred edge. The results of this analysis can be found in  
214 Supplementary Table S3.

215 In the following we only focus on the 130 edges that were found in at least 40% of the  
216 1000 modular Bayesian Network reconstructions (Figure 2). Notably, this threshold was only  
217 chosen for better visualization purposes and to limit the subsequent discussion. Edges with  
218 lower bootstrap probability might also exist in reality despite lower statistical confidence. Nodes  
219 corresponding to sex, APOE status, and brain region were not connected to any other nodes  
220 with sufficient statistical confidence, meaning that these features might have no impact on the  
221 rest of the network. Nodes with only outgoing edges in the network (i.e. source nodes) were:  
222 the years of education, the age, and the single gene NAV3. The GABA subgraph (containing

more than 280 genes) and the phenotype module were leaf nodes, meaning they had no outgoing edges. Only patient age had a direct influence on CD33. CD33 had eight directly influenced molecular mechanisms: the GABA subgraph, the Amyloidogenic subgraph (containing genes SRC and APBA2), the Acetylcholine signaling subgraph (containing genes ACHE and PRNP), the Prostaglandin subgraph, and the Chaperone subgraph (containing genes HSPB6, CXCL8, and CCR2). Also, the single gene module, TRAF1, was a child of CD33. Altogether, CD33 had a predicted causal influence on every node, except for the source nodes.

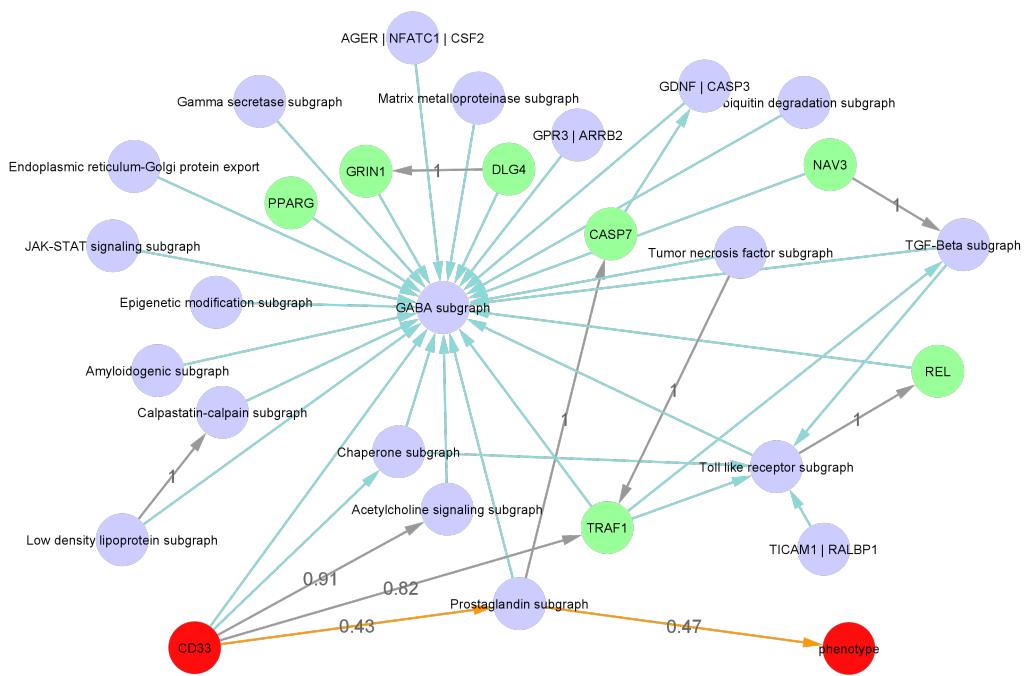


Figure 2: **Network representation of iVAMBN model for ROSMAP data.** Shown are the learned (grey) and knowledge-derived (green) edges between gene modules (purple nodes), single gene modules (green) and CD33 and phenotype module (red). All these edges appeared with bootstrap frequency  $> 0.4$ . The newly inferred shortest path between CD33 and phenotype is displayed in orange. Other edges with bootstrap frequency  $> 0.4$  have been removed for visualization purposes, except for those six edges which were trained with a bootstrap confidence of 1.

Model reveals path between CD33 and disease phenotype

As shown in Figure 2 the shortest path between CD33 and the disease phenotype was observed through the Prostaglandin subgraph. All the edges from this connection were newly learned

233 from data, meaning that they had not previously been identified in the knowledge graph. Nev-  
234 ertheless, these correlations have been previously reported in the literature: Prostaglandines are  
235 eicosanoides, which were found to play a role in memory learning and neuroinflammation [35,36].  
236 A major producer is activated microglia, which itself is activated through amyloid- $\beta$  and pro-  
237 duces inflammatory cytokines [37]. Currently, microglia and their effects on AD is a major focus  
238 in the field of research [38,39]. Also, PGD2, a prostaglandin mainly synthesized in neurons,  
239 was previously found to be upregulated in AD patients [40]. Pairwise correlation plots between  
240 the genes of the prostaglandin pathway and CD33 or phenotype can be found in Supplementary  
241 Figure S3.

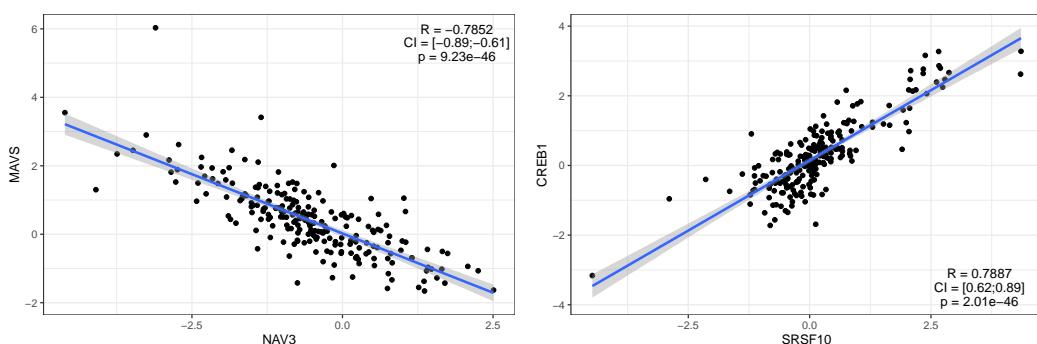
242 In total, 130 of the 162 edges of the bootstrapped iVAMBN model were newly learned from  
243 the data and had not been previously identified within the literature derived knowledge graph.  
244 Out of these 130 edges, six edges had a bootstrap confidence of 100%, meaning that they were  
245 learned consistently from 1000 random sub-samples of the data. A list of these edges can be  
246 found in Table 2.

Table 2: **Consistently newly learned edges in iVAMBN model.** All edges were found in each of 1000 network reconstructions from randomly subsampled data.

from	to
DLG4	GRIN1
Tumor necrosis factor subgraph	TRAF1
Toll like receptor subgraph	REL
Low density lipoprotein subgraph	Calpastatin-calpain subgraph
Prostaglandin subgraph	CASP7
NAV3	TGF-Beta subgraph

247 These high confidence edges demonstrated strong pairwise correlations between connected  
248 modules. NAV3, for example, had a strong negative correlation with MAVS, a member of the  
249 TGF-Beta subgraph module (Figure 3 left). In contrast to that SRSF10 and CREB1, members of  
250 the Low density lipoprotein subgraph and Calpastatin-calpain subgraph modules, were strongly  
251 positive correlated (Figure 3 right).

252 Although no direct correlation between NAV3 and MAVS is known, their effects are both  
253 linked to AD. NAV3, which is predominantly expressed in the nervous system, is increased in  
254 AD patients [41], while MAVS encodes a gene that is needed for the expression of beta interferon  
255 and thus contributes to antiviral innate immunity and may protect the cells from apoptosis [42].



**Figure 3: Quantitative relationships learned by iVAMBN.** Each correlation ( $R$ ) is shown along with its confidence interval (CI) and multiple testing adjusted p-value. Left: Correlation of NAV3 with TGF-Beta subgraph module member MAVS. Right: Correlation of Low density lipoprotein subgraph module member SRSF10 with CREB1, a member of the Calpastatin-calpain subgraph module. Further plots can be found in Supplementary Figures S3 and S4.

256 Together with the strong negative correlation seen in the data, one can hypothesize that the  
257 increased level of NAV3 in AD leads to a decreased level of MAVS, which elevates apoptosis of  
258 the cells.

259 The strong positive correlation between SRSF10 and CREB1 linked the Low density lipopro-  
260 tein (LDL) and Calpastatin-calpain subgraphs. LDL is a major APOE receptor, which is the  
261 strongest genetic factor for AD, where different alleles are either risk or protective alleles [43].  
262 APOE is also linked to amyloid- $\beta$ , whose production is increased with elevated activity of calpain  
263 due to the decreased levels of calpastatin. Calpastatin is also linked to synaptic dysfunction and  
264 to the tau pathology of AD [44, 45]. Tau is another protein that accumulates in the brains of  
265 AD patients. The exact underlying mechanisms here are still unknown, but regulatory mech-  
266 anisms of calpain are highly influenced by Calcium ( $Ca^{2+}$ ) influx and increased intracellular  
267 calcium levels are a main reason for the loss of neuronal function in AD [44–46]. Changes in the  
268 Calpastatin-calpain mechanism may therefore also lead to reduced amyloid- $\beta$  deposition.

## 269 **External Validation of iVAMBN model**

270 We assessed the ability of the model to explain normalized gene expression data from an inde-  
271 pendent study, Mayo. Notably, all gene expression data was from the same brain region, namely  
272 the cerebral cortex. However, Mayo does not contain MMSE scores. Therefore, we first trained

273 a modified version of our iVAMB model on ROSMAP, which only contained the Braak score in  
274 the phenotype module, but otherwise had the edges shown in Figure 2. The full list of edges of  
275 this model together with their corresponding bootstrap confidences can be found in Supplemen-  
276 tary Table S3. We then explored the marginal log-likelihood  $\log p(\text{data} | \text{graph})$  of the model on  
277 the Mayo dataset and subtracted the marginal log-likelihood obtained by 1000 random permu-  
278 tations of the network (Figure 4), resulting in an empirical p-value. This showed that our model  
279 could explain Mayo gene expression data significantly better than randomly permuted networks  
280 ( $p = 0.035$ ) despite the clinical differences between patients in both studies shown in Table 1.

281 In addition, we trained a separate iVAMB model on MSBB data and explored the overlap  
282 with the ROSMAP model at different thresholds of the bootstrap confidence (Supplementary  
283 Figure S5). At the previously chosen 40% threshold the overlap of the newly learned edges  
284 contained in the iVAMB models trained on ROSMAP and MSBB was statistically significant,  
285 even if edge directions were considered (hypergeometric test,  $p < 1e - 38$ ).

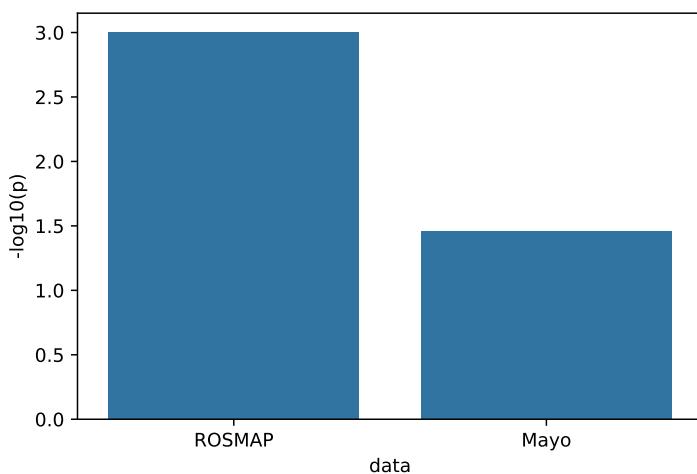


Figure 4: **External model validation.** Statistical significance  $-\log_{10}(p)$  value of the marginal log-likelihood of the model when evaluated on the training data (ROSMAP) and external validation data (Mayo).

## 286 CD33 Down-expression Simulation

287 To understand the potential systemic consequences of a therapeutic intervention into CD33 we  
288 simulated its down-expression. This was achieved by a counterfactual down-expression (here: 9-  
289 fold) of CD33 in every patient (Figure 5 (top left)). Due to the fact that iVAMBN is a quantitative  
290 model, associated downstream consequences on biological mechanisms and phenotype could be  
291 predicted in every patient (see examples in Figure 5). CD33 down-expression simulation (left)  
292 results in higher scores of the prostaglandin pathway module (right).

293 In addition, iVAMBN predicted a significant increase of MMSE scores ( $p < 0.001$ , Figure 6  
294 (left)), and also a significant decrease of Braak stages ( $p < 0.001$ , Figure 6 (right)). That means  
295 patients are not only predicted to improve the specific cognitive abilities tested by MMSE, but  
296 are also predicted to improve brain pathophysiology.

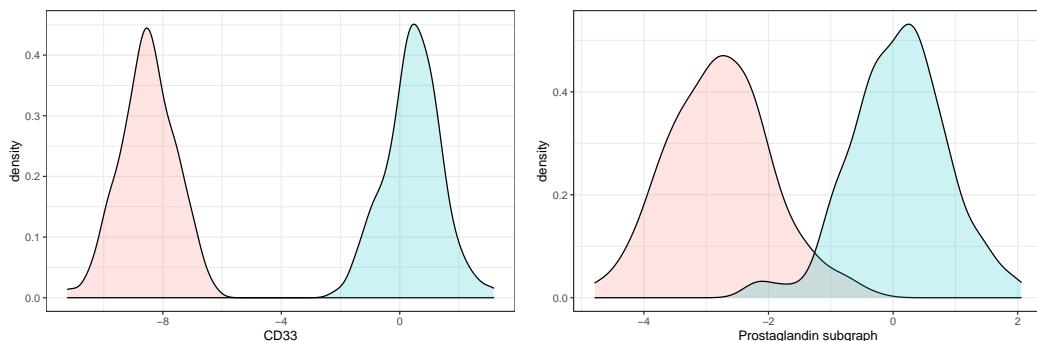
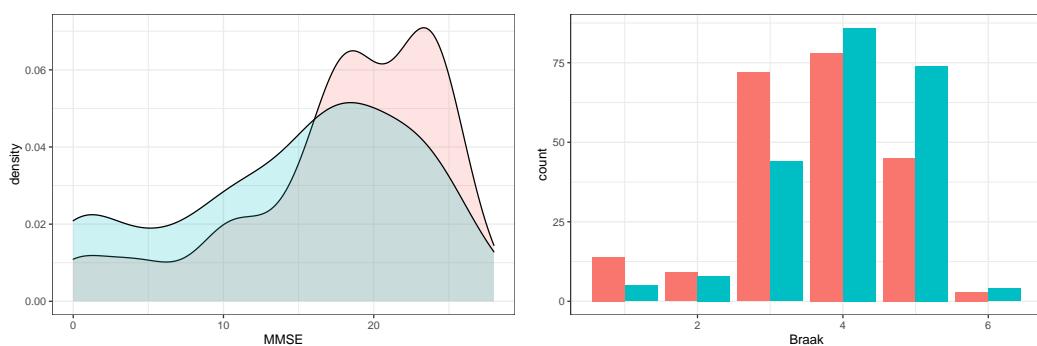


Figure 5: **Module distributions in original and simulated CD33 down-expression.** The blue curve describes the original distribution, while the red one describes the CD33 down-expression scenario. CD33 down-expression simulation (left) results in lower scores of the prostaglandin pathway module (right).

## 297 CD33 down-expression reveals significant changes in many mechanisms

298 Our iVAMBN model predicted significant effects on gene expression of 28 mechanisms and in-  
299 dividual genes, respectively (Table 3). Significant changes were, for example, predicted for the  
300 genes CASP7 and TRAF7, and the prostaglandin and calpastatin-calpain mechanisms. But  
301 also the amyloidogenic mechanism is significantly differential expressed in a CD33 knock-down  
302 scenario.

303 Decreased expression of the amyloidogenic mechanism will thus result in patients with less



**Figure 6: Predicted changes on phenotype (MMSE and Braak stages) as a consequence of CD33 down-expression.** Distribution of MMSE and Braak stages in CD33 original (blue) and down-expressed (red) patients shows a significant improvement of scores and thus cognition as well as brain pathophysiology.

304 amyloid- $\beta$  deposition. While this connection of the amyloidogenic mechanism and AD is clear,  
305 others need to be further explored.

306 The link between Calpastatin-calpain mechanism and AD was already described earlier. The  
307 key aspect is its negative influence on amyloid- $\beta$  deposition.

308 PGD2, a prostaglandin mainly synthesized in neurons, was previously found to be upregulated  
309 in AD patients [40]. Prostaglandines are eicosanoide, which were found to play a role in memory  
310 learning and neuroinflammation [35, 36]. A major producer is activated microglia, which itself  
311 is activated through amyloid- $\beta$  and produces inflammatory cytokines [37]. Currently, microglia  
312 and their effects on AD is a major focus in the field of research [38, 39]. Again, down-expression  
313 of the prostaglandin may result in reduced amyloid- $\beta$  deposition. Altogether, the vast majority  
314 of significantly differential expressed gene sets was highly linked to AD through the amyloid- $\beta$   
315 cascade.

### 316 Experimental validation with cell line data

317 We checked whether our iVAMBN based predictions experimentally agreed with cell line gene  
318 expression data, specifically reflecting wild type (WT) and CD33 knock-out (KO). Our analysis  
319 (see details in Methods part) revealed significant changes of 23 AD associated mechanisms and  
320 genes in KO versus WT. Interestingly, 19 out of these 23 mechanisms overlapped with those  
321 predicted by iVAMBN (Table 3). Likewise, iVAMBN predicted significant changes of 22 genes

Table 3: **Statistical significance of gene modules.** The table shows results of a global test [47], assessing the differential gene set expression of each gene module between WT and down-expression/KO of CD33. P-values of the test within simulated scenario, as well as, p-values from cell line KO are reported and corrected for multiple testing using the Benjamini-Hochberg method. The agreement of both tests is described in the last column, meaning if both tests are either significant or non-significant (+) or if they don't show same direction of significance (-). For GRIN1 no p-value could be computed, as that gene is not present in the cell line data.

Gene module	p-value simulated KD	p-value cell line KO	agreement significance
GABA subgraph	2.75e-04	3.60e-15	+
Toll like receptor subgraph	1.05e-26	1.05e-13	+
Prostaglandin subgraph	6.99e-109	1.02e-09	+
TGF-Beta subgraph	0.592	8.79e-11	-
Calpastatin-calpain subgraph	3.14e-91	5.41e-09	+
JAK-STAT signaling subgraph	0.454	2.91e-11	-
AGER / NFATC1 / CSF2	5.78e-41	0.0129	+
Chaperone subgraph	2.84e-75	2.02e-09	+
REL	4.45e-18	9.96e-11	+
Ubiquitin degradation subgraph	5.15e-20	1.06e-06	+
GRIN1	1.92e-132	NA	
PPARG	2.20e-04	1.78e-03	+
GDNF / CASP3	1.06e-17	2.98e-11	+
Gamma secretase subgraph	4.36e-10	1.93e-03	+
Epigenetic modification subgraph	6.90e-58	7.64e-03	+
TICAM1 / RALBP1	1.46e-16	0.0561	-
Amyloidogenic subgraph	4.54e-69	9.11e-10	+
Tumor necrosis factor subgraph	0.0997	0.769	+
Acetylcholine signaling subgraph	6.74e-04	0.337	-
Matrix metalloproteinase subgraph	0.0708	2.74e-10	-
NAV3	0.176	3.81e-07	-
TRAF1	1.66e-95	2.26e-08	+
CASP7	1.75e-138	0.151	-
GPR3 / ARRB2	4.87e-04	8.02e-04	+
Endoplasmic reticulum-Golgi protein export	5.19e-29	1.78e-11	+
Low density lipoprotein subgraph	0.891	8.11e-06	-
DLG4	5.85e-93	3.44e-07	+
CD33	3.33e-307	8.06e-08	+

322 and gene sets, respectively, out of which only 3 were false positives at a false discovery rate  
 323 threshold of 5%. Notably one of the false positive predictions (TICAM1 / RALBP1) had an  
 324 adjusted p-value of 5.6% in the experimental data.

325 Overall, we thus observed a high degree of overlap between the dysregulated mechanisms and  
 326 those predicted by the iVAMBN model, indicating that our model aligns well with the cell line  
 327 data.

**328    Simulation of the perturbation of other candidate targets**

**329** For comparison reasons, we further simulated the effect on the phenotype of a 9-fold up- or  
**330** down-regulation of all other genes in our model, which showed a directed path to the phenotype  
**331** module. Genes belonging to modules which were not an ancestor of the phenotype module were  
**332** excluded, because they could not have any effect on the phenotype according to our model. We  
**333** simulated for each candidate target an up- as well as a down-regulation.

**334** The simulated dys-regulations showed that none of the candidate targets had a predicted  
**335** effect on the phenotype stronger than CD33 (Figure S5). Only TRAF6 and TGFB3 down-  
**336** regulation as well as up-regulation of APBA2, TRAF5 and SALL1 were predicted to increase  
**337** the mean MMSE score by more than two points, compared to a predicted increase by almost  
**338** five points via CD33 perturbation.

**339** APBA2 is known to interact with APP and therefore plays a role in the amyloidogenic  
**340** pathway [48, 49]. TRAF6 was identified in multiple experiments as target of miR-146a which is  
**341** a key regulator of innate immunity that is up regulated in AD pathology affected brain regions  
**342** and might also has an effect on amyloid- $\beta$  metabolism [50]. It was found that treatment with  
**343** a miR-146a agomir inhibits TRAF6 expression and reduced the cognitive impairment in AD  
**344** mice [51].

**345    Discussion**

**346** The here presented work is the first to demonstrate, to our knowledge, that one can integrate  
**347** gene expression and clinical data together with qualitative knowledge about cause-and-effect  
**348** relationships into a quantitative, system medical model of AD. This was achieved via an AI  
**349** based method, which we combined with a knowledge graph representation of AD. We could  
**350** show that a simulated CD33 down-expression agrees well with experimental gene expression KO  
**351** data from a THP-1 cell line. Overall, our model could thus help to understand and quantify  
**352** intervention effects on a multi-scale biological system level and thus aid the identification of novel  
**353** therapeutic targets, which are urgently needed in the AD field.

**354** Our model predicted that CD33 down-regulation would yield a significant effect on cogni-  
**355** tion (MMSE) and brain pathophysiology (Braak scores) through the prostaglandin pathway.

356 Although the role of prostaglandins is known to play a role in memory, learning and neuroin-  
357flammation [35, 36], the exact mechanism by which cognition is affected remains unknown, but  
358 seems to be coupled to amyloid- $\beta$  deposition through microglia. In AD mice, a knockout of CD33  
359 mitigated amyloid- $\beta$  clearance and improved cognition [17, 18]. A positive effect on amyloid- $\beta$   
360 phagocytosis could also be observed in CD33 KO THP-1 macrophages [16].

361 Despite the evidence for a positive effect on cognition, we should mention that CD33 as  
362 a possible drug target has possible caveats that have been discussed in the literature [14]: i)  
363 It is not clear whether the genetic association of CD33 to AD is causal or just due to linkage  
364 disequilibrium with the true causal variant. ii) It is so far not entirely clear, how to therapeutically  
365 manipulate the expression level of CD33 in an optimal manner. iii) There might be safety issues  
366 due to the fact that CD33 is important for inhibiting immune responses and mediating self-  
367 tolerance. Systemic CD33 inhibition could potentially induce inflammatory autoimmune diseases.  
368 We therefore see the investigation of CD33 conducted in this paper more as a showcase for our  
369 iVAMBN approach rather than making any specific recommendation regarding the therapeutic  
370 value of CD33. Integrating known side effects of approved drugs targeting specific proteins in our  
371 model's graph structure could provide hints on possible side effects and is an interesting point  
372 for further research.

373 Altogether we see the impact of our work two-fold: first, we have introduced a novel multi-  
374 scale, quantitative modeling approach (iVAMBN), which is widely applicable in systems medicine,  
375 specifically in situations, where only a partial mechanistic understanding of biological phenom-  
376 ena is given. Secondly, our developed model can be further explored by the AD field and could  
377 aid a better understanding of the disease as well as identification of novel therapeutic options.

## 378 Methods

### 379 AD Knowledge Graph

380 A major part of this study is a BEL (<https://bel.bio/>) encoded, knowledge graph, which was  
381 initially compiled via text mining and later on manually curated via literature. In general, the  
382 BEL language helps to build a computer-process-able cause-and-effect relationship model. Each

383 BEL statement consists of a subject and an object, connected through a relation. Subjects and  
384 objects could be many different entities, like genes, proteins or RNA, but also biological processes,  
385 pathologies or even chemicals. Therefore, the relations have many different facets, as well. These  
386 could be relations like *increases*, *decreases* or *association*, describing the interaction between  
387 subject and object. But there are also relationships describing something like a membership of  
388 subject and object, for example *hasComponent* and *isA*. The BEL model used here, is an enriched  
389 version of the AD cause-and-effect relationship model published in [11] and can be found in the  
390 github repository. The enrichment was done around the two genes CD33 and TREM2, such that  
391 detailed knowledge about these two genes was gathered in the context of AD.

392 A filtering step was necessary, in order to get only entities measured in the gene expression  
393 data. In this case only gene and protein entities from the knowledge graph can be used. Addition-  
394 ally, the knowledge graph was filtered for only causal interactions, such as *increases*, *decreases*,  
395 or *regulates*, resulting in a network with 431 nodes and 673 edges. From that we only took the  
396 largest connected component to reduce the dimensionality. Hence, the used graph during our  
397 study consisted of 383 nodes and 607 edges, in which any two nodes were connected through  
398 some path.

399 **Clustering of Filtered Knowledge Graph** One of the key aspects of iVAMBN is grouping  
400 of input features (genes, pathophysiological and clinical features) into modules in order to allow  
401 for a statistically stable identification of a Bayesian Network structure in a subsequent step. For  
402 identifying modules of genes we clustered the knowledge graph with the help of different graph  
403 clustering algorithms:

404 • the Markov Cluster algorithm [32, 52] implemented in the *MCL* package in R [53].  
405 • edge betweenness [54] community detection implemented in the R package *igraph* [55]  
406 • infomap [56] community finding method implemented in the R package *igraph* [55]

407 After clustering, genes being part of a single cluster were assigned to a corresponding module.  
408 Genes being not clustered but only connected to one cluster, were merged into that cluster.  
409 Genes being connected to multiple clusters were kept as single gene modules (modules consisting  
410 of a single feature) for further analysis. We selected the best clustering algorithm according

411 to multiple metrics described in [57] including internal density, number of edges inside clusters,  
412 average degree, expansion, cut ratio, conductance, and norm cut. Based on these metrics the  
413 average ranking of each graph clustering algorithm was computed with the rational in mind, that  
414 each cluster should have an high internal density and sparse connections across clusters. This  
415 resulted in choosing the markov clustering algorithm for further analyses. The metrics for each  
416 clustering algorithm can be found in Supplementary Table S4.

417 **Annotation of Modules with AD Disease Mechanisms** For each module, an over-  
418 representation analysis for AD associated disease mechanisms was conducted. AD associated  
419 mechanisms were retrieved from the NeuroMMSig database [33]. For that purpose, the *enricher*  
420 function from the *clusterProfiler* package in R was used, which allows to use user-defined gene  
421 set annotations for a hypergeometric test [58]. We annotated each module with the most signif-  
422 icant NeuroMMSig gene set after multiple testing correction via control of false discovery rate  
423 (Benjamini-Hochberg method).

## 424 Gene Expression Data Analysis

425 RNAseq data from several observational clinical studies, as well as RNAseq data from a cell line  
426 knockout experiment, were used in this work. The patient data were from i) the Religious Orders  
427 Study and Memory and Aging Project (ROSMAP) [28–30], and ii) the Mayo RNAseq Study  
428 (Mayo) [31]. The last one contains two separate datasets referring to separate brain regions,  
429 namely cerebellum (CBE) and temporal cortex (TCX). Both studies were accessed through the  
430 AMP-AD Knowledge Portal at Synapse using the data deposited in the RNAseq Harmonization  
431 Study. The used data are gene counts provided as gene count matrices that had been generated  
432 using STAR [59]. Gene counts were normalized to log counts per Million (logCPMs) and counts  
433 from AD patients were scaled against the healthy control data within each study. That means  
434 for each AD sample and gene the corresponding mean expression value of the same gene in  
435 cognitively normal subjects was subtracted. Subsequently we divided the values by the standard  
436 deviation of the gene in healthy controls. That means raw expression values were converted into  
437 abnormality scores.

438 After that, the datasets were filtered for AD patients only, resulting in 221 samples for

439 ROSMAP, and 62 samples in each of the two Mayo studies. Further filtering was done based  
440 on the brain region the samples were taken from. While all brain regions in ROSMAP could be  
441 mapped to the cerebral cortex via the Uber-anatomy ontology (UBERON) [60], that could only  
442 be done for the temporal cortex part of the Mayo study, meaning that the cerebellum samples  
443 were excluded. For making the expression data across studies comparable, a batch correction with  
444 ComBat [61] was applied to the scaled AD data. This normalized, scaled, and batch corrected  
445 data was then used for further analysis steps.

446 The cell line RNAseq data used during this study is from a THP-1 monocyte cell line with  
447 two different genetic backgrounds and two treatments. It can be found under GEO accession  
448 GSE155567. A sample could have either wild-type CD33 or a knocked out CD33 gene, plus  
449 either a control vector or a SHP-1 knock-down vector, resulting in four different conditions:  
450 i) *wild-type with control*, ii) *wild-type with SHP-1 knock-down vector*, iii) *CD33 knockout with*  
451 *control vector*, and iv) *CD33 knockout with SHP-1 knock-down vector*. There were 6 biological  
452 replicates per condition. Within the here presented study, only samples containing the control  
453 vector were used, resulting in twelve used samples. Therefore samples from condition 1 were  
454 called as *wild-type (WT)* samples and samples from condition 3 as *knockout (KO)* samples.  
455 Reads were aligned with STAR and gene counts were generated via the *featureCounts* function  
456 of the *Rsubread* package [62]. More details about the data can be found in [16] and under GEO  
457 accession GSE155567.

## 458 Variational Autoencoders (VAE)

459 Variational autoencoders [26] are one of the most frequently used type of unsupervised neural  
460 network techniques. They can be interpreted as a special type of probabilistic graphical model,  
461 which has the form  $Z \rightarrow X$ , where  $Z$  is a latent, usually multivariate standard Gaussian, and  $X$   
462 a multivariate random variable describing the input data. Moreover, for any sample  $(x, z)$ , we  
463 have  $p(x | z) = N(\mu(z), \sigma(z))$ . One of the key ideas behind VAEs is to variationally approximate

$$\log q(z|x) = \log N(z | \mu(x), \sigma(x)) \quad (1)$$

<sup>464</sup> This means that  $\mu(x)$  and  $\sigma(x)$  are the multivariate mean and standard deviation of the approxi-  
<sup>465</sup> mate posterior  $q(z | x)$  and are outputs of a multi-layer perceptron neural network (the encoder)  
<sup>466</sup> that is trained to minimize for each data point  $x$  the criterion

$$\log(x) \geq \frac{1}{2} \sum_{j=1}^D (1 + \log \sigma_j(x)^2 - \mu_j(x)^2 - \sigma_j(x)^2) + \frac{1}{L} \sum_l \log p(x|z^{(l)}) \quad (2)$$

<sup>467</sup> Here the index  $j$  runs over the  $D$  dimensions of the input  $x$ , and  $z = \mu(x) + \sigma(x) \odot \epsilon^{(l)}$  with  
<sup>468</sup>  $\epsilon^{(l)} \sim N(0, I)$  being the  $l$ th random sample drawn from a standard multivariate Gaussian, and  
<sup>469</sup>  $\odot$  denotes an element-wise multiplication. Notably, the right summand corresponds to the re-  
<sup>470</sup> construction error of data point  $x$  by the model, whereas the first term imposes a regularization.  
<sup>471</sup> We refer to [26] for more details.

## <sup>472</sup> Heterogeneous Incomplete Variational Autoencoders (HI-VAE)

<sup>473</sup> Variational autoencoders were originally developed for homogeneous, continuous data. However,  
<sup>474</sup> in our case variables grouped into the phenotype module do not fulfill this assumption, because  
<sup>475</sup> Braak stages and MMSE scores are discrete ordinal. In agreement to our earlier work [25] we  
<sup>476</sup> thus employed the HI-VAE [34], which is an extension of variational autoencoders and allows for  
<sup>477</sup> various heterogeneous data types, even within the same module. More specifically, the authors  
<sup>478</sup> suggest to parameterize the decoder distribution as

$$p(x_j | z) = p(x_j | \gamma_j = h_j(z)) \quad (3)$$

<sup>479</sup> where  $h_j(\cdot)$  is a function learned by the neural network, and  $\gamma_j$  accordingly models data modality  
<sup>480</sup> specific parameters. For example, for real-valued data we have  $\gamma_j = (\mu(z), \sigma_j(z)^2)$ , while for  
<sup>481</sup> ordinal discrete data we use a thermometer encoding, where the probability of each ordinal  
<sup>482</sup> category can be computed as

$$p(x_j = r | \gamma_j) = p(x_j \leq r | \gamma_j) - p(x_j \leq r - 1 | \gamma_j) \quad (4)$$

483 with

$$p(x_j \leq r | z) = \frac{1}{1 + \exp(-(\theta_j(z) - h_j(z)))} \quad (5)$$

484 The thresholds  $\theta_j(z)$  divide the real line into  $R$  regions, and  $h_j(z)$  indicates, in which region  $z$   
 485 falls. The data modality specific parameters are thus  $\gamma_j = \{h_j(z), \theta_1(z), \dots, \theta_{R-1}(z)\}$  and are  
 486 modeled as output of a feed forward neural network.

487 According to [34] we use batch normalization to account for differences in numerical ranges  
 488 between different data modalities.

489 For multi-modal data and in particular discrete data a single Gaussian distribution may not  
 490 be a sufficiently rich representation in latent space. Hence, the authors propose to replace the  
 491 standard Gaussian prior distribution imposed for  $z$  in VAEs by a Gaussian mixture prior with  
 492  $K$  components:

$$s \sim \text{Categorical}(\pi) \quad (6)$$

$$z | s \sim N(\mu(s), I_K) \quad (7)$$

493 where  $\pi_k = \frac{1}{K}$  for  $k = 1, 2, \dots, K$  and  $s$  is a one-hot vector encoding of the mixture compo-  
 494 nent. We evaluated different choices of  $K$  using a 3-fold cross-validation, while employing the  
 495 reconstruction error  $\frac{1}{L} \sum_l \log p(x|z^{(l)})$  as an objective. In conclusion it turned out that  $K = 1$   
 496 component was an optimal choice for all modules in our iVAMBN model.

## 497 Modular Bayesian Networks

498 Let  $X = (X_v)_{v \in V}$  be a set of random variables indexed by nodes  $V$  in a directed acyclic graph  
 499 (DAG)  $G = (V, E)$ . In our case each of these nodes corresponds either to lower dimensional  
 500 embedding of a group of variables (i.e. module) in the original data, or to an original features  
 501 (e.g. biological sex) in the dataset. According to the definition of a Bayesian Network (BN), the  
 502 joint distribution  $p(X_1, X_2, \dots, X_n)$  factorizes according to

$$p(X_1, X_2, \dots, X_n) = \prod_{v \in V} p(X_v | X_{pa(v)}) \quad (8)$$

503 where  $pa(v)$  denotes the parent set of node  $v$  [27]. In our case random variables follow either  
504 a Gaussian or a multinomial distribution, i.e. the BN is hybrid. Notably, no discrete random  
505 variable was allowed to be a child of a Gaussian one.

506 Since the BN in our case is defined over low dimensional representations of groups of variables,  
507 we call the structure Modular Bayesian Network (MBN). Notably, a MBN is a special instance  
508 of a hierarchical BN over a structured input domain [63–66].

509 A typical assumption in (M)BNs is that the set of parameters  $(\theta_v)_{v \in V}$  associated to nodes  
510  $V$  are statistically independent. For a Gaussian node  $v$  parameters can thus be estimated by  
511 fitting a linear regression function with parents of  $v$  being predictor variables [27]. Similarly, for  
512 a discrete node  $\tilde{v}$  having only discrete parents, parameters can be estimated by counting relative  
513 frequencies of variable configurations, resulting into a conditional probability table.

## 514 Quantitative Modeling Across Biological Scales via iVAMBN

### 515 Model Training

516 The here presented *Integrative Variational Autoencoder Modular Bayesian Network (iVAMBN)*  
517 approach (Figure 1), integrates different biological scales together with a knowledge graph into  
518 the previously published Variational Autoencoder Modular Bayesian Network (VAMBN) ap-  
519 proach [25]. More precisely, there are four steps to build an iVAMBN model: i) Definition of  
520 modules of variables, ii) Training of a HI-VAE for each module, iii) Definition of logical con-  
521 straints for possible edges in the MBN, and iv) Structure and parameter learning of the MBN  
522 using encoded values for each module. These four steps result from the fact that HI-VAEs (as  
523 well as any other variants of variational autoencoders) themselves can be interpreted as specific  
524 types of BNs and thus the overall log-likelihood of an iVAMBN model can be decomposed accord-  
525 ingly. That means the overall iVAMBN model can be interpreted as a special type of Bayesian  
526 Network, see [25] for details.

527 The four model building steps were followed in the application of the iVAMBN approach  
528 in this work as well. The modules of variables were mainly defined through the previously  
529 explained Markov clustering of the knowledge graph, plus an additional module summarizing  
530 MMSE (Mini–Mental State Examination) and Braak stages into one *phenotype* module. MMSE

531 measures cognitive impairment by testing the orientation in time and space, recall, language,  
532 and attention, while Braak stages refer to the degree of biological brain pathology [67]. Some  
533 non-assigned genes, were directly treated as nodes in the MBN construction and thus also called  
534 gene modules. The same was done for demographic features, like sex, age, years of education  
535 and the APOE genotype.

536 For training the HI-VAEs for each module a hyperparameter optimization (grid search) was  
537 implemented over learning rate (learning rate  $\in \{0.001, 0.01\}$ ) and minibatch size (minibatch  
538 size  $\in \{16, 32\}$ ) as in [25]. Each parameter combination was evaluated with the reconstruction  
539 loss as objective function in a 3-fold cross-validation scenario.

540 In general the number of possible MBN DAG structures for  $n$  nodes grows super-exponentially  
541 with  $n$  [24], making identification of the true graph structure highly challenging. Therefore, our  
542 aim was to restrict the set of possible DAGs a priori as much as possible via knowledge based  
543 logical constraints. More specifically we imposed the following causal restrictions:

- 544 • Nodes defined by demographic or clinical features (like age, gender, APOE genotype, and  
545 brain region) can only have outgoing edges.
- 546 • The phenotype module (= clinical outcome measures) can only have incoming edges.
- 547 • Genes and gene modules can not influence demographic or clinical features, except the age.

548 To additionally integrate prior knowledge defined through the knowledge graph, we tested three  
549 different strategies while building a MBN:

- 550 1) **Completely data driven:** The knowledge graph is completely ignored for structure learn-  
551 ing.
- 552 2) **Knowledge informed:** The knowledge graph is used in the greedy hill climbing algorithm  
553 for structure learning i) as starting point, ii) as white list (intending that those edges were  
554 defined as pre-existing), or iii) as both.
- 555 3) **Completely knowledge driven:** The knowledge graph provides the structure of the  
556 MBN and additional connections are only allowed for demographics or the phenotype mod-  
557 ule.

558 Structure learning of the MBN was always performed via a greedy hill climber using the Bayesian  
559 Information Criterion for model selection. We employed the implementation provided in R-  
560 package *bnlearn* [68].

561 **Evaluating the Model Fit**

562 To evaluate the fit of the overall iVAMBN model we employed the generative nature of our model:  
563 Following a topological sorting of the nodes of the DAG of the MBN we first sampled from the  
564 distribution of each node conditional on its parent. Notably, for MBN nodes representing modules  
565 this amounted to sample from the posterior of the according HI-VAE, which in practice can be  
566 realized via injection of normally distributed noise, see Section Variational Autoencoders, Eq.  
567 (2). Subsequently, the random sample was then decoded via the HI-VAE. Altogether we thus  
568 generated as many synthetic subjects as real ones. We then compared the marginal distribution  
569 of each variable based on the synthetic and the real data. Results, including summary statistics  
570 and Kullback-Leibler divergences are shown in the supplementary material. Furthermore, we  
571 compared the correlation matrices of synthetic and real data.

572 **CD33 down-expression simulation and analysis**

573 To be able to simulate a down-expression of CD33, we first shifted the distribution of CD33  
574 such that it reflects a 9-fold down-expression of CD33. In agreement to the theory of Bayesian  
575 Networks this operation made CD33 conditionally independent of its parents in the MBN, which  
576 amounts to deleting any of its incoming edges and resulted into a mutilated MBN. Afterwards we  
577 exploited the fact that iVAMBN is a generative model. That means we first drew samples from  
578 the conditional densities of the mutilated MBN. Practically this amounted to first topologically  
579 sort the nodes in the MBN, hence exploiting the fact that the underlying graph structure cannot  
580 have cycles. Subsequently, samples were drawn from the statistical distribution of each node  
581 while conditioning on the value of its parents. The result was a per-sample module activity  
582 scores, which we then decoded through our HI-VAE models into single gene scores.

583 Differences between the wild-type and simulated down-expression samples were investigated  
584 afterwards via multiple statistical hypothesis tests: First, a linear regression was used to model

585 the down-expression effect on gene expression and on the different phenotype scores. Second, the  
586 *globaltest* package in R was used to test the differential expression of specific gene sets between  
587 the wild-type and simulated down-expression group [47]. Those tested gene sets were here defined  
588 through the modules' genes used in the MBN, meaning that we tested for differential expression  
589 of MBN's gene modules. P-values were adjusted for multiple test scenario with the help of the  
590 *subsets* option of *globaltest* and via calculating the false discovery rate. The *globaltest* for gene  
591 sets, as well as the fold change analysis, was also applied to the cell line WT and KO data to be  
592 able to validate the results.

593 Effects of the perturbation of other candidate targets were simulated similarly as the CD33  
594 knock-down. Again, the distribution of the respective target was shifted such that it reflected  
595 a 9-fold down- or up-regulation. The module was identified to which the candidate target had  
596 been assigned, and all variables (including the perturbed target) mapping to that module were  
597 encoded via the previously trained HI-VAE for the module. Subsequently, the effects on the  
598 phenotype could be predicted in the same way as described for CD33.

## 599 Acknowledgements

600 This project has received partial funding from the Innovative Medicines Initiative 2 Joint  
601 Undertaking under grant agreement No 115976. This Joint Undertaking receives support from the  
602 European Union's Horizon 2020 research and innovation programme and EFPIA.

603 This work was partially developed in the Fraunhofer Cluster of Excellence "Cognitive Internet  
604 Technologies" and partially supported via the Fraunhofer Center for Machine Learning.

605 The results published here are in whole or in part based on data obtained from the AD  
606 Knowledge Portal ( <https://adknowledgeportal.org> ). Data generation was supported by the fol-  
607 lowing NIH grants: P30AG10161, P30AG72975, R01AG15819, R01AG17917, R01AG036836,  
608 U01AG46152, U01AG61356, U01AG046139, P50 AG016574, R01 AG032990, U01AG046139,  
609 R01AG018023, U01AG006576, U01AG006786, R01AG025711, R01AG017216, R01AG003949,  
610 R01NS080820, U24NS072026, P30AG19610, U01AG046170, RF1AG057440, and U24AG061340,  
611 and the Cure PSP, Mayo and Michael J Fox foundations, Arizona Department of Health Services  
612 and the Arizona Biomedical Research Commission. We thank the participants of the Reli-

613 gious Order Study and Memory and Aging projects for the generous donation, the Sun Health  
614 Research Institute Brain and Body Donation Program, the Mayo Clinic Brain Bank, and the  
615 Mount Sinai/JJ Peters VA Medical Center NIH Brain and Tissue Repository. Data and analysis  
616 contributing investigators include Nilüfer Ertekin-Taner, Steven Younkin (Mayo Clinic, Jack-  
617 sonville, FL), Todd Golde (University of Florida), Nathan Price (Institute for Systems Biology),  
618 David Bennett, Christopher Gaiteri (Rush University), Philip De Jager (Columbia University),  
619 Bin Zhang, Eric Schadt, Michelle Ehrlich, Vahram Haroutunian, Sam Gandy (Icahn School of  
620 Medicine at Mount Sinai), Koichi Iijima (National Center for Geriatrics and Gerontology, Japan),  
621 Scott Noggle (New York Stem Cell Foundation), Lara Mangravite (Sage Bionetworks).

## 622 References

623 [1] Hodson R. Alzheimer's disease. *Nature*. 2018 7;559:S1-1.

624 [2] Kanehisa M, Furumichi M, Tanabe M, Sato Y, Morishima K. KEGG: new perspectives on  
625 genomes, pathways, diseases and drugs. *Nucleic Acids Research*. 2017 1;45.

626 [3] Cerami EG, Gross BE, Demir E, Rodchenkov I, Babur O, Anwar N, et al. Pathway Com-  
627 mons, a web resource for biological pathway data. *Nucleic Acids Research*. 2011 1;39.

628 [4] Slenter DN, Kutmon M, Hanspers K, Ruitta A, Windsor J, Nunes N, et al. WikiPathways:  
629 a multifaceted pathway database bridging metabolomics to other omics research. *Nucleic  
630 Acids Research*. 2018 1;46.

631 [5] Fabregat A, Jupe S, Matthews L, Sidiropoulos K, Gillespie M, Garapati P, et al. The  
632 Reactome Pathway Knowledgebase. *Nucleic Acids Research*. 2018 1;46.

633 [6] Carvalho-Silva D, Pierleoni A, Pignatelli M, Ong C, Fumis L, Karamanis N, et al. Open  
634 Targets Platform: new developments and updates two years on. *Nucleic Acids Research*.  
635 2019 1;47.

636 [7] Wang Y, Zhang S, Li F, Zhou Y, Zhang Y, Wang Z, et al. Therapeutic target database 2020:  
637 enriched resource for facilitating research and early development of targeted therapeutics.  
638 *Nucleic Acids Research*. 2019 11.

639 [8] Piñero J, Ramírez-Anguita JM, Saüch-Pitarch J, Ronzano F, Centeno E, Sanz F, et al. The  
640 DisGeNET knowledge platform for disease genomics: 2019 update. *Nucleic Acids Research*.  
641 2019 11.

642 [9] Szklarczyk D, Gable AL, Lyon D, Junge A, Wyder S, Huerta-Cepas J, et al. STRING  
643 v11: protein–protein association networks with increased coverage, supporting functional  
644 discovery in genome-wide experimental datasets. *Nucleic Acids Research*. 2019 1;47.

645 [10] Orchard S, Ammari M, Aranda B, Breuza L, Brigandt L, Broackes-Carter F, et al. The  
646 MIntAct project—IntAct as a common curation platform for 11 molecular interaction  
647 databases. *Nucleic Acids Research*. 2014 1;42.

648 [11] Kodamullil AT, Younesi E, Naz M, Bagewadi S, Hofmann-Apitius M. Computable cause-  
649 and-effect models of healthy and Alzheimer’s disease states and their mechanistic differential  
650 analysis. *Alzheimer’s and Dementia*. 2015 11;11:1329-39.

651 [12] Estus S, Shaw BC, Devanney N, Katsumata Y, Press EE, Fardo DW. Evaluation of CD33  
652 as a genetic risk factor for Alzheimer’s disease. *Acta Neuropathologica*. 2019 8;138.

653 [13] Jiang T, Yu JT, Hu N, Tan MS, Zhu XC, Tan L. CD33 in Alzheimer’s Disease. *Molecular*  
654 *Neurobiology*. 2014 2;49.

655 [14] Zhao L. CD33 in Alzheimer’s Disease – Biology, Pathogenesis, and Therapeutics: A Mini-  
656 Review. *Gerontology*. 2019;65.

657 [15] Siddiqui SS, Springer SA, Verhagen A, Sundaramurthy V, Alisson-Silva F, Jiang W, et al.  
658 The Alzheimer’s disease-protective CD33 splice variant mediates adaptive loss of function  
659 via diversion to an intracellular pool. *Journal of Biological Chemistry*. 2017 9;292.

660 [16] Wißfeld J, Nozaki I, Mathews M, Raschka T, Ebeling C, Hornung V, et al. Deletion of  
661 Alzheimer’s disease-associated CD33 results in an inflammatory human microglia phenotype.  
662 GLIA. 2021.

663 [17] Griciuc A, Patel S, Federico AN, Choi SH, Innes BJ, Oram MK, et al. TREM2 Acts  
664 Downstream of CD33 in Modulating Microglial Pathology in Alzheimer’s Disease. *Neuron*.  
665 2019 9;103.

666 [18] Griciuc A, Serrano-Pozo A, Parrado A, Lesinski A, Asselin C, Mullin K, et al. Alzheimer's  
667 Disease Risk Gene CD33 Inhibits Microglial Uptake of Amyloid Beta. *Neuron*. 2013 5;78.

668 [19] Steckmann T, Awan Z, Gerstman BS, Chapagain PP. Kinetics of peptide secondary structure  
669 conversion during amyloid B-protein fibrillogenesis. *Journal of Theoretical Biology*. 2012  
670 5;301:95-102.

671 [20] Proctor CJ, Pienaar IS, Elson JL, Kirkwood TB. Aggregation, impaired degradation and im-  
672 munization targeting of amyloid-beta dimers in Alzheimers disease: A stochastic modelling  
673 approach. *Molecular Neurodegeneration*. 2012;7:32.

674 [21] Oblak AL, Forner S, Territo PR, Sasner M, Carter GW, Howell GR, et al. Model organism  
675 development and evaluation for late-onset Alzheimer's disease: MODEL-AD. *Alzheimer's  
& Dementia: Translational Research & Clinical Interventions*. 2020 1;6.

677 [22] Jankowsky JL, Zheng H. Practical considerations for choosing a mouse model of Alzheimer's  
678 disease. *Molecular Neurodegeneration*. 2017 12;12:1-22.

679 [23] Arber C, Lovejoy C, Wray S. Stem cell models of Alzheimer's disease: Progress and chal-  
680 lenges. *Alzheimer's Research and Therapy*. 2017 6;9:1-17.

681 [24] Chickering DM, Meek C, Heckerman D. Large-Sample Learning of Bayesian Networks is  
682 NP-Hard. 2012 10.

683 [25] Gootjes-Dreesbach L, Sood M, Sahay A, Hofmann-Apitius M, Fröhlich H. Variational Au-  
684 toencoder Modular Bayesian Networks for Simulation of Heterogeneous Clinical Study Data.  
685 *Frontiers in Big Data*. 2020 5;3:16.

686 [26] Kingma DP, Welling M. Auto-Encoding Variational Bayes. 2013 12.

687 [27] Heckerman D. In: Holmes DE, Jain LC, editors. *A Tutorial on Learning with Bayesian  
688 Networks*. Berlin, Heidelberg: Springer Berlin Heidelberg; 2008. p. 33-82.

689 [28] Mostafavi S, Gaiteri C, Sullivan SE, White CC, Tasaki S, Xu J, et al. A molecular network  
690 of the aging human brain provides insights into the pathology and cognitive decline of  
691 Alzheimer's disease. *Nature Neuroscience*. 2018 6;21:811-9.

692 [29] Bennett DA, Buchman AS, Boyle PA, Barnes LL, Wilson RS, Schneider JA. Religious  
693 Orders Study and Rush Memory and Aging Project. *Journal of Alzheimer's Disease*. 2018  
694 6;64.

695 [30] Bennett DA, Schneider JA, Arvanitakis Z, Wilson RS. Overview and Findings from the  
696 Religious Orders Study. *Current Alzheimer Research*. 2012 6;9.

697 [31] Allen M, Carrasquillo MM, Funk C, Heavner BD, Zou F, Younkin CS, et al. Human  
698 whole genome genotype and transcriptome data for Alzheimer's and other neurodegenerative  
699 diseases. *Scientific Data*. 2016 12;3:160089.

700 [32] Dongen SV. Graph Clustering Via a Discrete Uncoupling Process. *SIAM Journal on Matrix  
701 Analysis and Applications*. 2008 1;30.

702 [33] Domingo-Fernández D, Kodamullil AT, Iyappan A, Naz M, Emon MA, Raschka T, et al.  
703 Multimodal mechanistic signatures for neurodegenerative diseases (NeuroMMSig): A web  
704 server for mechanism enrichment. *Bioinformatics*. 2017;33.

705 [34] Nazábal A, Olmos PM, Ghahramani Z, Valera I. Handling incomplete heterogeneous data  
706 using VAEs. *Pattern Recognition*. 2020 11;107:107501.

707 [35] Bsci DT, Msc GK, Weisinger RS, Sinclair AJ. The role of eicosanoids in the brain. *Asia  
708 Pac J Clin Nutr*. 2008;17:220-8.

709 [36] Ardura-Fabregat A, Boddeke EWGM, Boza-Serrano A, Brioschi S, Castro-Gomez S,  
710 Ceyzériat K, et al. Targeting Neuroinflammation to Treat Alzheimer's Disease. *CNS Drugs*.  
711 2017 12;31:1057-82.

712 [37] Biringer RG. The role of eicosanoids in alzheimer's disease. *International Journal of Envi-  
713 ronmental Research and Public Health*. 2019 7;16.

714 [38] Candlish M, Hefendehl JK. Microglia Phenotypes Converge in Aging and Neurodegenerative  
715 Disease. *Frontiers in Neurology*. 2021 5;12.

716 [39] Schwabe T, Srinivasan K, Rhinn H. Shifting paradigms: The central role of microglia in  
717 Alzheimer's disease. *Neurobiology of Disease*. 2020 sep;143:104962.

718 [40] Iwamoto N, Kobayashi K, Kosaka K. The formation of prostaglandins in the postmortem  
719 cerebral cortex of Alzheimer-type dementia patients. *Journal of Neurology*. 1989 2;236:80-4.

720 [41] Shioya M, Obayashi S, Tabunoki H, Arima K, Saito Y, Ishida T, et al. Aberrant microRNA  
721 expression in the brains of neurodegenerative diseases: miR-29a decreased in Alzheimer  
722 disease brains targets neurone navigator 3. *Neuropathology and Applied Neurobiology*.  
723 2010 feb;36(4):320-30.

724 [42] Seth RB, Sun L, Ea CK, Chen ZJ. Identification and Characterization of MAVS, a Mitochon-  
725 drial Antiviral Signaling Protein that Activates NF- $\kappa$ B and IRF3. *Cell*. 2005 sep;122(5):669-  
726 82.

727 [43] Serrano-Pozo A, Das S, Hyman BT. APOE and Alzheimer's disease: advances in genetics,  
728 pathophysiology, and therapeutic approaches. *The Lancet Neurology*. 2021 jan;20(1):68-80.

729 [44] Ferreira A. Calpain Dysregulation in Alzheimer's Disease. *ISRN Biochemistry*. 2012  
730 10;2012:1-12.

731 [45] Vosler PS, Brennan CS, Chen J. Calpain-mediated signaling mechanisms in neuronal injury  
732 and neurodegeneration. *Molecular Neurobiology*. 2008;38:78-100.

733 [46] McCarty MF, Dinicolantonio JJ, Lerner A. A fundamental role for oxidants and intracellular  
734 calcium signals in Alzheimer's pathogenesis—and how a comprehensive antioxidant strategy  
735 may aid prevention of this disorder. *International Journal of Molecular Sciences*. 2021 2;22:1-  
736 27.

737 [47] Goeman JJ, van de Geer SA, de Kort F, van Houwelingen HC. A global test for groups of  
738 genes: testing association with a clinical outcome. *Bioinformatics*. 2004 1;20.

739 [48] Abou-Fadel J, Vasquez M, Grajeda B, Ellis C, Zhang J. Systems-wide analysis unravels the  
740 new roles of CCM signal complex (CSC). *Heliyon*. 2019 12;5:e02899.

741 [49] Jensen TMT, Albertsen L, Bartling CRO, Haugaard-Kedström LM, Strømgaard K. Probing  
742 the Mint2 Protein-Protein Interaction Network Relevant to the Pathophysiology of  
743 Alzheimer's Disease. *Chembiochem : a European journal of chemical biology*. 2018 3.

744 [50] Goodall EF, Heath PR, Bandmann O, Kirby J, Shaw PJ. Neuronal dark matter: the  
745 emerging role of microRNAs in neurodegeneration. *Frontiers in cellular neuroscience*. 2013  
746 10;7:178.

747 [51] Mai H, Fan W, Wang Y, Cai Y, Li X, Chen F, et al. Intranasal Administration of miR-  
748 146a Agomir Rescued the Pathological Process and Cognitive Impairment in an AD Mouse  
749 Model. *Molecular therapy Nucleic acids*. 2019 12;18:681-95.

750 [52] Dongen SV. Graph clustering by flow simulation. *Proefschrift Universiteit Utrecht*; 2000.

751 [53] Jäger ML. MCL: Markov Cluster Algorithm.. <https://CRAN.R-project.org/package=MCL>;  
752 2015.

753 [54] Newman MEJ, Girvan M. Finding and evaluating community structure in networks. *Physical Review E*. 2004 2;69:026113.

755 [55] Csardi G, Nepusz T, et al. The igraph software package for complex network research.  
756 *InterJournal, complex systems*. 2006;1695(5):1-9.

757 [56] Rosvall M, Bergstrom CT. Maps of random walks on complex networks reveal community  
758 structure. *Proceedings of the National Academy of Sciences*. 2008 1;105:1118-23.

759 [57] Arratia A, Mirambell MR. Clustering assessment in weighted networks. *PeerJ Computer  
760 Science*. 2021;7:1-27.

761 [58] Yu G, Wang LG, Han Y, He QY. clusterProfiler: an R Package for Comparing Biological  
762 Themes Among Gene Clusters. *OMICS: A Journal of Integrative Biology*. 2012 5;16.

763 [59] Dobin A, Davis CA, Schlesinger F, Drenkow J, Zaleski C, Jha S, et al. STAR: Ultrafast  
764 universal RNA-seq aligner. *Bioinformatics*. 2013 1;29:15-21.

765 [60] Mungall CJ, Torniai C, Gkoutos GV, Lewis SE, Haendel MA. Uberon, an integrative multi-  
766 species anatomy ontology. *Genome Biology*. 2012;13.

767 [61] Johnson WE, Li C, Rabinovic A. Adjusting batch effects in microarray expression data  
768 using empirical Bayes methods. *Biostatistics*. 2007 1;8.

769 [62] Liao Y, Smyth GK, Shi W. The R package Rsubread is easier, faster, cheaper and better  
770 for alignment and quantification of RNA sequencing reads. Nucleic Acids Research. 2019  
771 5;47.

772 [63] Parviainen P, Kaski S. Bayesian Networks for Variable Groups; 2016.

773 [64] Parviainen P, Kaski S. Learning structures of Bayesian networks for variable groups. International  
774 Journal of Approximate Reasoning. 2017 9;88:110-27.

775 [65] Becker AK, Dörr M, Felix SB, Frost F, Grabe HJ, Lerch MM, et al. From heterogeneous  
776 healthcare data to disease-specific biomarker networks: A hierarchical Bayesian network  
777 approach. PLOS Computational Biology. 2021 2;17:e1008735.

778 [66] Gyftodimos E, Flach PA. Hierarchical Bayesian Networks: An Approach to Classification  
779 and Learning for Structured Data. In: Vouros GA, Panayiotopoulos T, editors. Methods and  
780 Applications of Artificial Intelligence. Berlin, Heidelberg: Springer Berlin Heidelberg;  
781 2004. p. 291-300.

782 [67] Braak H, Braak E. Neuropathological stageing of Alzheimer-related changes. Acta Neu-  
783 ropathologica. 1991 9;82:239-59.

784 [68] Scutari M. Learning Bayesian Networks with the bnlearn R Package. Journal of Statistical  
785 Software. 2010;35.

786 **Author contributions**

787 HF designed and supervised the project. TR analysed the data. MS and AA helped with  
788 the analysis of the data. BS and CE generated the knowledge graph. TR and HF wrote the  
789 manuscript. All authors have read and approved the manuscript.

790 **Supporting information**

791 **Supplementary Table S1: Module enrichment analysis**  
792 **Supplementary Table S2: Module assignment**

793 **Supplementary Figure S1: Clustered knowledge graph.**

794 **Supplementary Note S1: iVAMBNs Module Definition**

795 **Supplementary Note S2: iVAMBNs Knowledge Integration**

796 **Supplementary Figure S2: Log likelihood loss (i.e. negative log-likelihood) of differ-**

797 **ent knowledge integration strategies.**

798 **Supplementary Figure S3: Quantitative effect between modules of shortest path.**

799 **Supplementary Figure S4: Quantitative effect between modules of newly trained**

800 **edges with confidence 1**

801 **Supplementary Table S3: Bootstrap confidence results**

802 **Supplementary Figure S5: Overlap of ROSMAP and Mayo network structures**

803 **Supplementary Figure S6: Effects on phenotype scores of up- and down-regulation**

804 **simulations.**

805 **Supplementary Table S4: Graph Clustering Metrics**

806 **Supplementary Note S3: Evaluating the Model Fit**

807 **Supplementary Figure S7: Distribution of single feature in real data versus simulated**

808 **data.**

809 **Supplementary Figure S8: Correlation matrix of real and drawn data.**

810 **Supplementary Appendix S1: Github repository of the code for this analysis <https://github.com/traschka/iVAMBN>**

## 812 **Additional information**

### 813 **Data Availability Statement**

814 • All code can be found at: <https://github.com/traschka/iVAMBN>

815 • ROSMAP and Mayo: <https://adknowledgeportal.synapse.org/Explore/Studies/DetailsPage?Study=syn21241740>

817 • CD33 KO cell line: <https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE155567>