

**Title:** A Genetic Variant of Fatty Acid Amide Hydrolase (FAAH) Exacerbates Hormone-Mediated Orexigenic Feeding in Mice

**Short Title:** FAAH exacerbates orexigenic feeding responses

Georgia Balsevich<sup>1\*</sup>, Gavin N Petrie<sup>1</sup>, Daniel E Heinz<sup>2</sup>, Arashdeep Singh<sup>3</sup>, Robert J Aukema<sup>1</sup>, Avery C Hunker<sup>4</sup>, Haley A Vecchiarelli<sup>1</sup>, Hiulan Yau<sup>1</sup>, Martin Sticht<sup>1</sup>, Roger J Thompson<sup>1</sup>, Francis S Lee<sup>5</sup>, Larry S Zweifel<sup>4</sup>, Prasanth K Chelikani<sup>6</sup>, Nils C Gassen<sup>2</sup>, Matthew N Hill<sup>1\*</sup>

<sup>1</sup>Hotchkiss Brain Institute, University of Calgary, Calgary, AB, Canada

<sup>2</sup>Neurohomeostasis Research Group, Department of Psychiatry and Psychotherapy, University Hospital Bonn, Bonn, Germany

<sup>3</sup>Monell Chemical Senses Center and Department of Neuroscience, University of Pennsylvania, Philadelphia, PA, USA

<sup>4</sup>Department of Psychiatry and Behavioral Sciences, University of Washington, Seattle, WA, USA

<sup>5</sup>Weill Cornell Medical College, Cornell University, New York, NY USA

<sup>6</sup>Texas Tech University School of Veterinary Medicine, Amarillo, TX, USA

\* Corresponding Authors:

Georgia Balsevich, Hotchkiss Brain Institute, University of Calgary, 3330 Hospital Drive NW, Calgary, AB Canada T2N4N1; Email: georgia.balsevich@ucalgary.ca

21 Matthew N. Hill, PhD, Hotchkiss Brain Institute, University of Calgary, 3330 Hospital Drive  
22 NW, Calgary, AB Canada T2N4N1; Email: mnhill@ucalgary.ca

23

## Abstract

Fatty acid amide hydrolase (FAAH) degrades the endocannabinoid anandamide. A polymorphism in *FAAH* (FAAH C385A) reduces FAAH expression, increases anandamide levels, and increases the risk of obesity. Nevertheless, some studies have found no association between FAAH C385A and obesity. We investigated whether the environmental context governs the impact of FAAH C385A on metabolic outcomes. Using a C385A knock-in mouse model, we found that under basal conditions, there was no effect of the FAAH C385A polymorphism on body weight or composition. By contrast, FAAH A/A mice are more susceptible to glucocorticoid-induced hyperphagia, weight gain, and activation of hypothalamic AMPK. AMPK inhibition occluded the amplified hyperphagic response to glucocorticoids in FAAH A/A mice. FAAH knockdown exclusively in AgRP neurons mimicked the exaggerated feeding response of FAAH A/A mice to glucocorticoids. FAAH A/A mice likewise presented exaggerated orexigenic responses to ghrelin, while FAAH knockdown in AgRP neurons blunted leptin anorectic responses. Together, the FAAH A/A genotype amplifies orexigenic responses and decreases anorexigenic responses, providing a putative mechanism explaining the diverging human findings.

## Introduction

Since 1975, the prevalence of worldwide obesity has nearly tripled and has reached epidemic proportions (World Health Organisation, 2020). An incomplete understanding surrounding the etiology of obesity has hindered the development of therapeutic interventions. Regardless, it is well known that both genetic and environmental factors contribute to its

development (Goodarzi, 2018). Therefore, it is important to identify how specific genetic and environmental factors interact to contribute to the mechanisms underlying the etiology of obesity.

The endocannabinoid system (ECS), comprised of two receptors (CB1 and CB2), endogenous cannabinoids (N-arachidonylethanolamide, [anandamide], AEA and 2-arachidonoylglycerol, 2-AG), and their synthetic and degradative enzymes, is recognized as a powerful regulator of energy homeostasis and body weight (Mazier et al., 2015). In general, activation of the ECS promotes energy storage, food intake, and increased weight gain. Therefore, components of the ECS are perfectly positioned to moderate individual susceptibility to obesity. In humans, there is a common genetic variant in the gene for fatty acid amide hydrolase (FAAH) (Sipe et al., 2002), the primary enzyme responsible for the inactivation of the endocannabinoid AEA. This missense mutation in FAAH (C385A; rs324420) destabilizes the FAAH protein, reduces AEA metabolism, and increases AEA signaling (Chiang et al., 2004; Sipe et al., 2002). Importantly, increased body mass index (BMI) has been associated with the low-expressing FAAH variant (A-allele carriers) (Durand et al., 2008; Monteleone et al., 2008; Sipe et al., 2005; Zhang et al., 2009). This agrees with the well-known role of AEA to stimulate feeding and enhance energy storage through the activation of CB1 receptor signaling (Mazier et al., 2015). However, the human data surrounding the C385A variant of FAAH and body weight regulation are oftentimes conflicting, with some studies showing no effect of FAAH C385A on metabolic outcomes (Jensen et al., 2007; Lieb et al., 2009). The conflicting findings may be attributed to differences in an individual's environmental context and resulting endocrine milieu. In this regard, gene x environment interactions could govern whether the FAAH C385A genotype affects obesity susceptibility under particular environmental contexts.



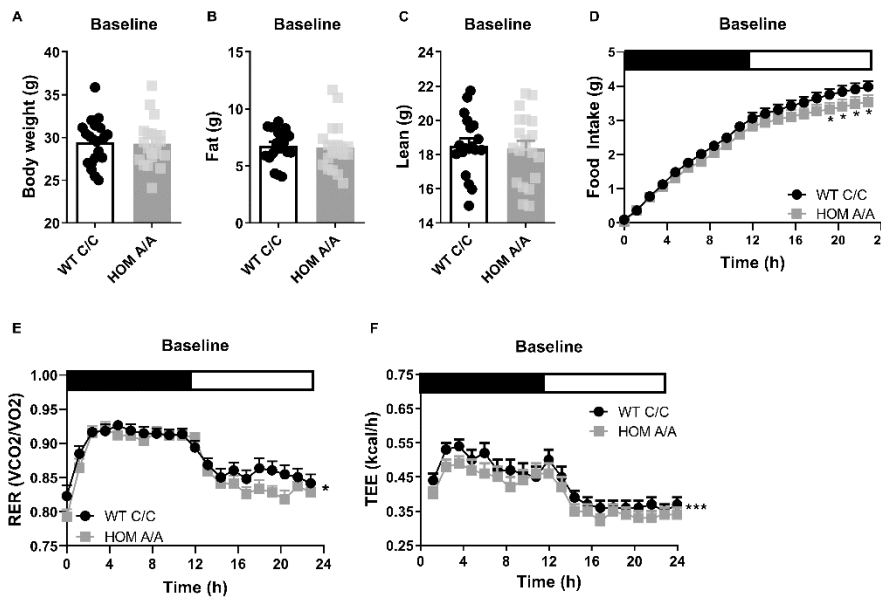
Environmental conditions dictate an individual's physiological endocrine state. For example, chronic stress and high fat diet feeding are associated with shifts in levels of circulating hormones that regulate appetite and energy homeostasis, including, but not limited to, glucocorticoids (GCs), ghrelin, and leptin (Bouassida et al., 2010; Schwarz et al., 2011; Ulrich-Lai & Ryan, 2014). Interestingly, the metabolic actions of orexigenic signals, such as GCs and ghrelin, increase endocannabinoid signaling (Balsevich et al., 2017; Edwards & Abizaid, 2016). By contrast, the anorectic peptide leptin suppresses endocannabinoid signaling to mediate its effects (Balsevich et al., 2018; Di Marzo et al., 2001). Therefore, differences in FAAH expression, due to distinct genetic FAAH variants, may influence the metabolic outcomes resulting from shifts in endocrine states. Specifically, we have previously shown that chronic exposure to GCs leads to obesity through an endocannabinoid-mediated mechanism (Bowles et al., 2015). It was likewise shown that GCs increase the activity of AMP-activated protein kinase (AMPK) through a CB1 receptor-dependent mechanism in the hypothalamus to promote weight gain (Scerif et al., 2013). Indeed hypothalamic AMPK plays an integral role in the regulation of feeding and body weight, where its activation leads to increased feeding (Andersson et al., 2004; Kim et al., 2004; Minokoshi et al., 2004). Similar to GCs, the orexigenic gastric hormone ghrelin increases the activity of hypothalamic AMPK and consequently feeding through an CB1 receptor-dependent mechanism (Kola et al., 2008). Taken together, we hypothesized that the discrepancies in the human literature for FAAH C385A and body weight outcomes are attributed to differences in their environmental context and ultimately their endocrine milieu. Therefore, we sought to characterize the behavioral, cellular, and molecular changes associated with the FAAH C385A variant in response to GC (Corticosterone, CORT) exposure using the FAAH C385A knock-in mouse model, recapitulating the common human mutation in the *FAAH* gene. We

furthermore assessed a role for FAAH in ghrelin- and leptin-dependent feeding outcomes to determine the generalizability of these effects.

## Results

### Body weight and body composition are similar between FAAH C/C and FAAH A/A mice under basal conditions

As a first step, we examined the metabolic outcomes arising in FAAH C/C (WT) and FAAH A/A (HOM) mice under baseline conditions. FAAH C/C and FAAH A/A mice presented similar body weights, fat mass, and lean mass at baseline (Figure 1). Yet despite no difference in body weight or composition, there was a genotype x time trend ( $p = 0.082$ ) for 24 h cumulative food intake, whereby *post hoc* analysis indicated that FAAH A/A mice present decreased food intake compared to FAAH C/C mice. In support of this finding, 24 h energy expenditure was lower in FAAH A/A mice compared to FAAH C/C mice. Furthermore, FAAH A/A mice presented an overall lower respiratory exchange ratio (RER), favoring fat utilization compared to FAAH C/C mice. Collectively, these data suggest that under baseline conditions, there is no effect of the C385A variant on body weight or composition despite differences in underlying metabolic parameters. The lower food intake in FAAH A/A mice matches their lower energy expenditure. These findings support the human studies reporting no association between A-allele carriers and obesity susceptibility under standard feeding conditions.

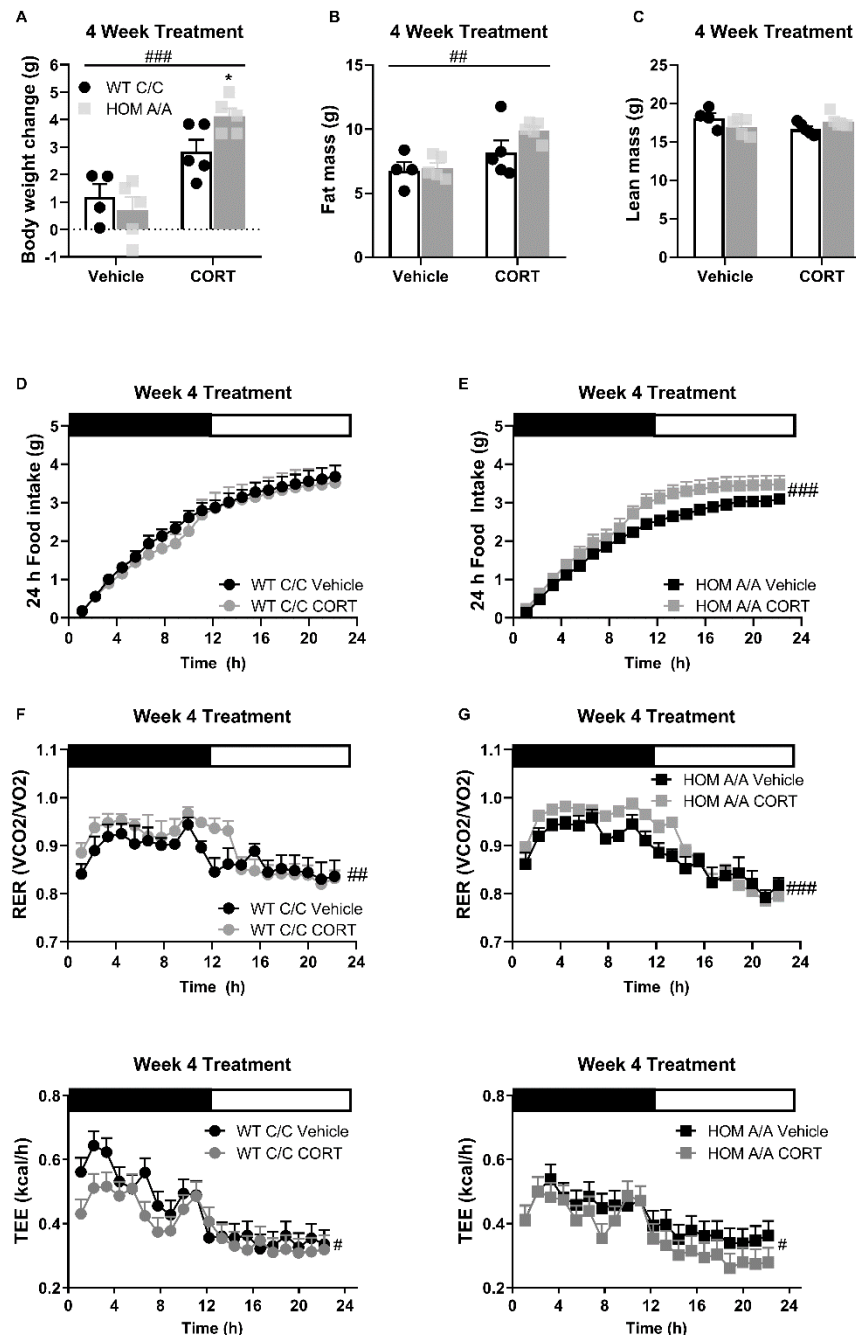


**Figure 1 – The FAAH C385A variant has no effect on baseline body weight or body composition.** (A – C) Under basal conditions, there is no effect of the FAAH C385A variant on body weight ( $T(39) = 0.1516$ ,  $p = 0.88$ ), fat mass ( $T(39) = 0.2640$ ,  $p = 0.79$ ), or lean mass ( $T(39) = 0.4219$ ,  $p = 0.6754$ ). (D – F) By contrast, the C385A variant modifies underlying metabolic parameters, wherein FAAH A/A (HOM) mice present reduced food intake (genotype  $\times$  time:  $F(76, 2360.107) = 1.238$ ,  $p = 0.082$  plus post hoc testing), decreased RER (genotype:  $F(1, 197.189) = 6.406$ ,  $p = 0.012$ ), and decreased energy expenditure (genotype:  $F(1, 387.547) = 22.949$ ,  $p < 0.001$ ) compared to FAAH C/C (WT) mice. For panels A – C,  $n = 20$  WT;  $n = 21$  HOM. For panels D – F,  $n = 17$  per genotype. Data are presented as means  $\pm$  SEM, and in panel F estimated means adjusted for body weight are presented. Panels A – C were analyzed by unpaired student's t-tests. Panels D – F were analyzed by linear mixed models with repeated measured. Asterisks denote significant genotype effect. \* $P < 0.05$ , \*\*\* $P < 0.001$ .

## FAAH A/A mice are more sensitive to the prolonged effects of glucocorticoids

To examine whether the FAAH C385A variant affects GC-mediated metabolic outcomes, we characterized the metabolic phenotypes arising in FAAH C/C (WT) and FAAH A/A (HOM) mice following four week exposure to CORT (25  $\mu$ g/ml) or vehicle (1% ethanol) delivered through their drinking water as previously described (Karatsoreos et al., 2010). At baseline, circulating CORT was comparable between FAAH C/C and FAAH A/A mice (Supplemental

Figure S1). Similarly, prolonged CORT exposure significantly elevated circulating CORT measured during the dark (active) stage regardless of genotype. There was furthermore a corresponding decrease in adrenal weights following prolonged CORT treatment, supporting previous reports (Karatsoreos et al., 2010). With regards to metabolic outcomes, prolonged CORT exposure led to significant body weight gain and increased fat mass regardless of genotype (Figure 2). Furthermore, a genotype x treatment trend ( $p = 0.0520$ ) followed by *post hoc* analysis indicated that FAAH A/A mice gained significantly more weight on account of CORT treatment compared to FAAH C/C mice. There was neither an effect of genotype nor treatment on lean mass. Importantly, prolonged CORT treatment significantly increased 24 h cumulative food intake exclusively in FAAH A/A mice (Figure 2). In accordance, food intake throughout the 4-week treatment was significantly elevated by CORT solely in FAAH A/A mice (Supplemental Figure S2). However, regardless of genotype, prolonged CORT exposure favoured carbohydrate utilization as reflected in the increased RER across genotypes as well as decreased total energy expenditure (TEE) (Figure 2). Therefore, FAAH A/A mice present a heightened sensitivity to GC-mediated hyperphagia and weight gain following prolonged exposure.



**Figure 2 – FAAH A/A mice show exaggerated body weight gain and food intake on account of prolonged CORT.** (A) Body weight gain was increased following 4 weeks of CORT treatment (25  $\mu$ g/ml in drinking water) (treatment  $F(1,15) = 6.65$ ,  $p < 0.001$ ), where FAAH A/A (HOM) mice gained significantly more weight than FAAH C/C (WT) mice treated with CORT (treatment  $\times$  genotype =  $F(1,15) = 4.456$ ,  $p = 0.052$  plus post hoc testing). (B) Fat mass was increased from 4 weeks of CORT exposure independent of genotype (treatment  $F(1, 15) = 11.66$ ,  $p = 0.004$ ). (C) A significant interaction was detected for lean mass (treatment  $\times$  genotype:  $F(1, 15) = 5.111$ ,  $p = 0.0391$ ) but Bonferroni's post hoc testing revealed no effect of either genotype or

prolonged CORT exposure. (D – E) Whereas 4 weeks of CORT treatment did not affect cumulative food intake measured over 24 h in FAAH C/C mice (treatment:  $F(1, 20.758) = 0.820$ ,  $p = 0.320$ ), it significantly increased food intake in FAAH A/A mice (treatment:  $F(1, 24.770) = 24.538$ ,  $p < 0.001$ ). (F – G) Regardless of genotype, 4 weeks of CORT exposure increased RER (treatment: WT ( $F(1, 47.659) = 4.289$ ,  $p = 0.044$ ); HOM ( $F(1, 68.057) = 19.493$ ,  $p < 0.001$ ). (H – I) whereas decreased energy expenditure measure over 24 h (treatment: WT ( $F(1, 47.659) = 4.289$ ,  $p = 0.044$ ; HOM ( $F(1, 68.057) = 19.493$ ,  $p < 0.001$ ). For panels A – I,  $n = 4$  WT-vehicle;  $n = 5$  WT-CORT,  $n = 5$  HOM-vehicle;  $n = 5$  HOM-CORT. Data are presented as means  $\pm$  SEM, and in panels H & I estimated means adjusted for body weight are presented. Panels A – C were analyzed by 2-way ANOVAs. Panels D – I were analyzed by linear mixed models with repeated measures. Pound signs denote significant treatment effect. Asterisks denote significant genotype effect. \* $P < 0.05$ , # $P < 0.05$ , ## $P < 0.01$ , ### $P < 0.001$ .

## **FAAH A/A mice are more sensitive to the immediate effects of glucocorticoids**

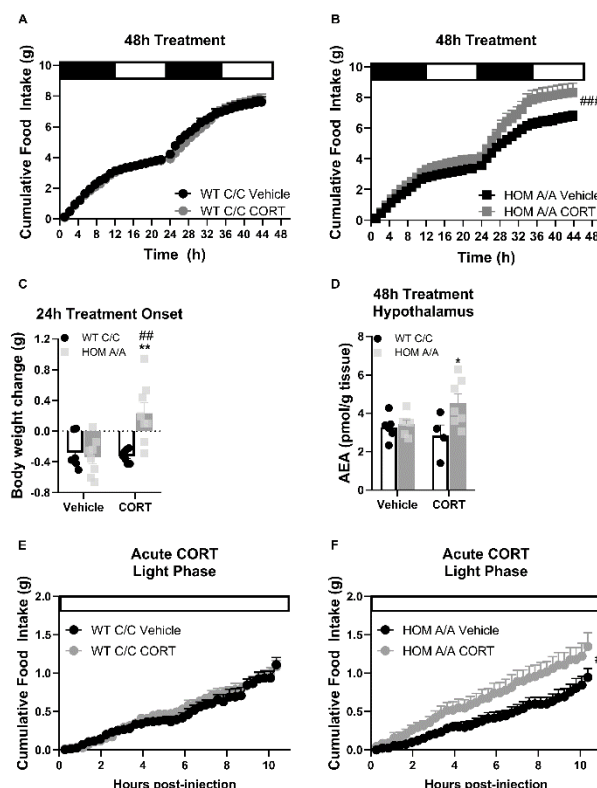
It is well known that short- and long-term exposures to GCs have distinct outcomes on the ECS (reviewed by (G Balsevich et al., 2017)). Therefore, we investigated whether the FAAH C385A variant likewise modifies the metabolic responses to CORT exposure within the initial 48 h of treatment onset. Similar to 30-day CORT exposure, FAAH A/A mice present heightened sensitivity to the immediate effects of GC exposure on food intake and weight gain. Specifically, within 48 h of treatment onset, food intake was increased on account of oral CORT exposure exclusively in FAAH A/A mice (Figure 3). Likewise, within the first 24 h of treatment onset, CORT exposure led to weight gain exclusively in FAAH A/A mice. However, regardless of genotype, RER was elevated on account of 48 h CORT exposure (Supplemental Figure S3). Furthermore, TEE was exclusively decreased in FAAH C/C mice on account of CORT exposure. Indeed CORT exposure is known to decrease energy expenditure (Hardwick et al., 1989; Ramage et al., 2016; Strack et al., 1995; Van Den Beukel et al., 2014). The lack of CORT-induced suppression of energy expenditure in FAAH A/A mice is consistent with their observed hyperphagic response to CORT, which is known to elevate energy expenditure on account of

diet-induced thermogenesis (Westerterp, 2017), thus possibly countering the CORT-mediated decrease in energy expenditure. Finally, we employed implanted telemeters to examine total activity under basal conditions and during the initial 24 h of CORT exposure. Neither CORT treatment nor genotype influenced home-cage activity (Supplemental Figure S4).

Given our findings that the FAAH C385A variant specifically modifies GC-mediated feeding effects, we next examined AEA content in brain regions known to play an integral role in endocannabinoid-mediated feeding (Ruiz de Azua & Lutz, 2019) to determine whether CORT exposure differentially mobilizes AEA in FAAH C/C and FAAH A/A mice. Remarkably, in the hypothalamus AEA content was exclusively elevated in FAAH A/A mice on account of 48 h CORT (Figure 3). By contrast, in the ventral tegmental area (VTA) and nucleus accumbens, there was no significant effect of either CORT treatment or genotype (Supplemental Figure S5). Furthermore, there was no effect of the FAAH C385A polymorphism on AEA levels in peripheral organ systems that are recognized as the primary mediators of endocannabinoid-mediated regulation of metabolism (Osei-Hyiaman et al., 2008; Pagotto et al., 2006; Ruiz de Azua et al., 2017), namely brown adipose tissue (BAT), inguinal white adipose tissue (iWAT), epididymal WAT (eWAT) and liver. However, in the liver, 48 h CORT significantly elevated AEA levels independent of genotype. Taken together, FAAH A/A mice are more sensitive to CORT-induced hyperphagia and weight gain following 48 h exposure, which is accompanied by increased hypothalamic AEA levels exclusively in CORT-treated FAAH A/A mice.

GCs are known to signal through genomic and non-genomic signaling mechanisms to elicit both delayed and rapid behavioural effects, respectively (Balsevich et al., 2017). In order to decipher the signaling mechanism through which the FAAH C385A variant impacts GC-mediated feeding outcomes, we examined the feeding responses of FAAH C/C and FAAH A/A

mice to an acute (single) injection of CORT (3 mg/kg). CORT was delivered at the onset of the light/inactive phase, a period of low feeding, to capture any hyperphagic effects of CORT. In agreement with our previous findings, an acute injection of CORT significantly elevated light phase feeding exclusively in the FAAH A/A mice within 10 h (Figure 3). The effects of CORT on cumulative food intake were no longer evident by 24 h post-injection (Supplemental Figure S6). Accordingly, there was no effect of acute CORT treatment on body weight gain examined 24 h post-injection. In terms of TEE and RER, acute CORT injections significantly reduced energy expenditure whereas promoted carbohydrate utilization regardless of genotype (Supplemental Figure S6). Our data indicate that the FAAH C385A variant selectively exaggerates the hyperphagic effects of GCs.



**Figure 3 FAAH A/A mice are hyperresponsive to the immediate orexigenic effects of CORT.** (A – B) Within the first 48 h of treatment onset, CORT exposure (25 µg/ml in drinking water) had no effect on cumulative food intake in FAAH C/C (WT) mice (treatment: F(1, 57.970)



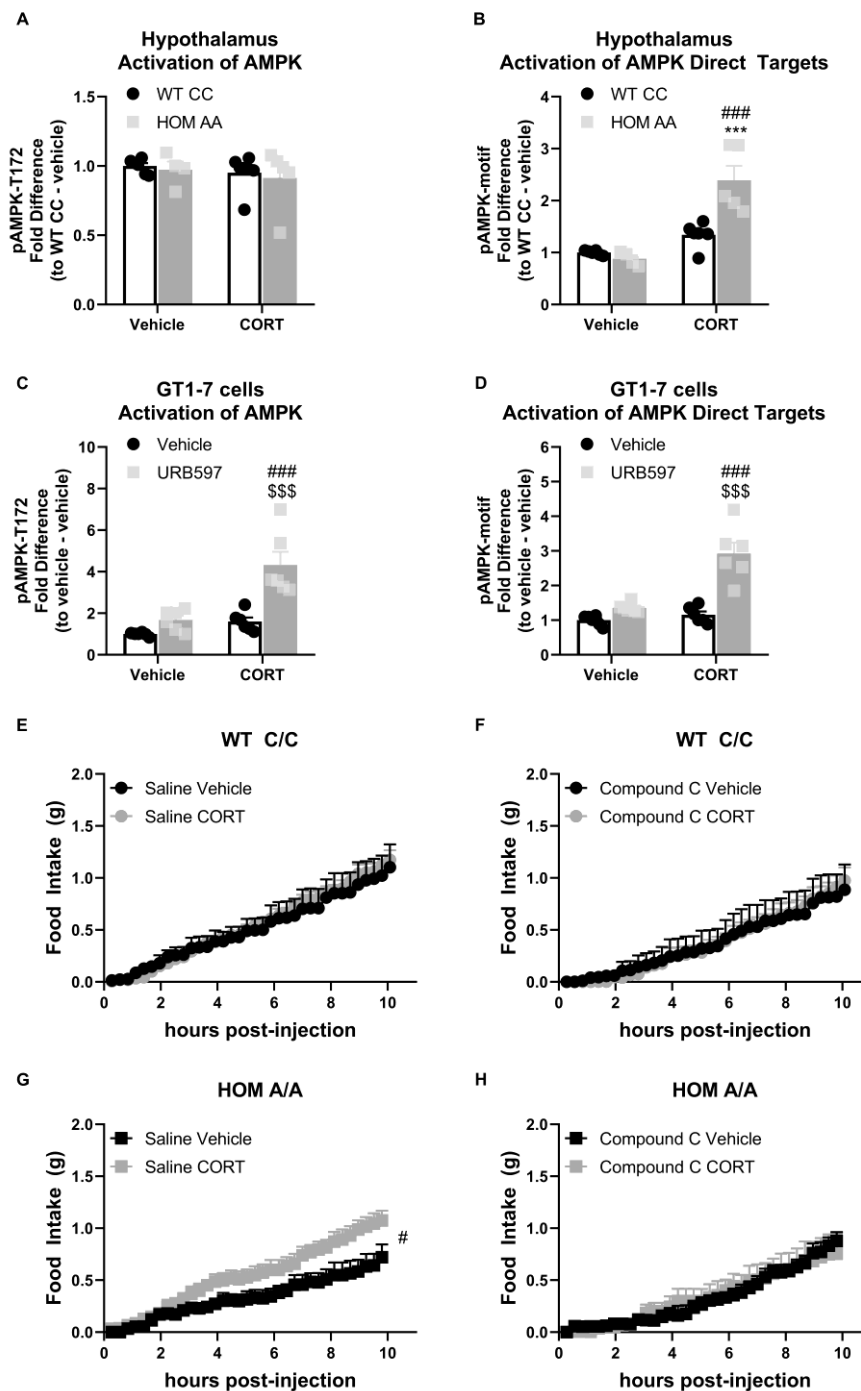
= 0.600,  $p = 0.442$ ) whereas it significantly increased food intake in FAAH A/A (HOM) mice (treatment:  $F(1, 55.289) = 17.202$ ,  $p < 0.001$ ). (C) Within 24 h of treatment onset, CORT exposure led to weight gain exclusively in FAAH A/A mice (treatment x genotype:  $F(1, 26) = 10.06$ ,  $p = 0.004$  plus Bonferroni post hoc testing). (D) Likewise, hypothalamic AEA levels were increased from 48 h CORT treatment exclusively in FAAH A/A mice (treatment x genotype:  $F(1, 20) = 4.177$ ,  $p = 0.054$  plus Bonferroni post hoc testing). (E – F) Acute CORT (3 mg/kg, single i.p. injection) again had no effect on cumulative food intake in FAAH C/C mice (treatment:  $F(1, 20.236) = 0.482$ ,  $p = 0.495$ ) yet increased light phase food intake in FAAH A/A mice (within the initial 10 h following CORT injections) (treatment:  $F(1, 14.720) = 4.729$ ,  $p = 0.046$ ). For panel A:  $n = 9$  per group. For panel B,  $n = 10$  HOM-vehicle;  $n = 11$  HOM-CORT. For panel C,  $n = 7$  WT-vehicle;  $n = 7$  WT-CORT;  $n = 8$  HOM-vehicle;  $n = 8$  HOM-CORT. For panel D,  $n = 7$  WT-vehicle;  $n = 4$  WT-CORT;  $n = 6$  HOM-vehicle;  $n = 7$  HOM-CORT. For panel E  $n = 8$  per group. For panel F:  $n = 8$  HOM-vehicle;  $n = 7$  HOM-CORT. Data are presented as means  $\pm$  SEM. Panels A – B and E – F were analyzed by linear mixed models with repeated measures. Panels C – D were analyzed by 2-way ANOVAs. Pound signs denote significant treatment effect. Asterisks denote significant genotype effect. \* $P < 0.05$ , \*\* $P < 0.01$ , # $P < 0.05$ , ## $P < 0.01$ , ### $P < 0.001$ .

## **Hypothalamic AMPK signaling is downstream of AEA-GC-mediated feeding effects**

As the effect of the FAAH C385A variant on GC-dependent outcomes was exclusive to feeding, we next addressed the underlying mechanism responsible for mediating this interaction. Given a previous study demonstrating that GCs increase hypothalamic AMPK activity through an endocannabinoid-dependent mechanism (Scerif et al., 2013), we examined the effects of the FAAH C385A variant on GC-mediated effects on hypothalamic AMPK signaling. We first examined the phosphorylation status of AMPK and the direct targets of AMPK as a proxy of AMPK signaling activation. For this purpose, FAAH C/C and A/A mice were injected with CORT (3 mg/kg) or vehicle, and exactly 1 h later, hypothalamic tissue was collected. We found that AMPK signaling was markedly increased exclusively in FAAH A/A mice following administration of CORT. Specifically, while we did not detect a significant increase in AMPK activity per se (as reflected through the phosphorylation status of AMPK), we observed a significant increase in the activation status of direct downstream AMPK targets (as reflected

through the increased phosphorylation status of targets expressing the AMPK-consensus sequence) in CORT-treated FAAH A/A mice (Figure 4). To complement these findings, we treated immortalized hypothalamic cells (GT1-7 cells) with the FAAH inhibitor URB597 to mimic the increased AEA signaling present in FAAH A/A mice. Following 4 h pretreatment with either URB597 (1  $\mu$ M) or vehicle, GT1-7 cells were treated for an additional 2 h with CORT (1  $\mu$ M) or vehicle in a 2x2 design. In support of our previous findings, CORT treatment significantly increased AMPK signaling activity exclusively in URB597-treated cells as reflected in increased AMPK phosphorylation and increased phosphorylation status of downstream targets of AMPK (Figure 4).

To assess the functional implications of these FAAH-dependent events, we examined the effects of AMPK inhibition on CORT-mediated feeding outcomes in the FAAH C/C and A/A mice. At 1 h prior to CORT/vehicle injections, FAAH C/C and A/A mice were treated with compound C (5 mg/kg, i.p.), an AMPK inhibitor, or saline. As expected, FAAH C/C mice did not show a feeding response to CORT (during the light phase) nor was there a CORT effect in the FAAH C/C mice when AMPK was inhibited (Figure 4). By contrast, FAAH A/A mice showed the expected hyperphagic response to CORT treatment during their light phase feeding, which was blocked by pretreatment with the AMPK inhibitor. After 24 h, feeding returned to baseline (Supplemental Figure S7). Taken together, our data demonstrates that the FAAH C385A variant amplifies the effects of CORT on feeding through an AMPK-dependent mechanism.



**Figure 4: The FAAH A/A genotype amplifies the GC-mediated orexigenic effects through an AMPK-dependent mechanism.** (A) Hypothalamic AMPK activity (as assessed by pAMPK protein expression) was unaffected by genotype ( $F(1, 17) = 0.249$ ,  $p = 0.624$ ) or CORT (3 mg/kg, i.p.) ( $F(1, 17) = 0.711$ ,  $p = 0.411$ ). (B) However, in the hypothalamus, downstream targets of AMPK were activated from CORT exposure exclusively in FAAH A/A (HOM) mice ( $F(1, 17) = 15.45$ ,  $p = 0.001$  plus post hoc testing). (C – D) In GT1-7 hypothalamic cells, AMPK activity was enhanced exclusively in CORT-treated cells pre-treated with URB597, assessed by

pAMPK ( $F(1, 20) = 8.807$ ,  $p = 0.008$  plus post hoc testing) and pAMPK-motif ( $F(1, 20) = 16.74$ ,  $p < 0.001$  plus post hoc testing) protein expression. (E – F). Acute CORT treatment had no effect on cumulative light phase food intake in FAAH C/C (WT) mice following administration of either saline (treatment:  $F(1, 11.155) = 0.45$ ,  $p = 0.836$ ) or an AMPK inhibitor (Compound C, 5 mg/kg, i.p.) (treatment:  $F(1, 8.755) = 0.001$ ,  $p = 0.982$ ). By contrast, (G – H) AMPK inhibition prevented CORT-mediated orexigenic effects in FAAH A/A mice measured during the light phase (treatment: saline ( $F(1, 14.344) = 6.112$ ,  $p = 0.027$ ); compound C ( $F(1, 8.793) = 0.009$ ,  $p = 0.927$ )). Panels A – B:  $n = 6$  WT-vehicle biological replicates;  $n = 6$  WT-CORT biological replicates;  $n = 4$  HOM-vehicle biological replicates;  $n = 5$  HOM-CORT biological replicates. Panels C – D:  $n = 6$  biological replicates per group. Panel E:  $n = 4$  WT-vehicle;  $n = 8$  WT-CORT. Panel F:  $n = 4$  HOM-vehicle;  $n = 6$  HOM-CORT. Panel G:  $n = 6$  WT-vehicle.  $N = 7$  WT-CORT. Panel H:  $n = 5$  per group. Protein expression data are expressed as relative fold change compared to control condition  $\pm$  SEM. Food intake data are presented as means  $\pm$  SEM. Panels A – D were analyzed by 2-way ANOVAs. Panels E – H were analyzed by linear mixed models with repeated measures. Pound signs denote significant treatment effect. Asterisks denote significant genotype effect. Dollar signs denote significant URB597 treatment effect. \*\*\* $P < 0.001$ , # $P < 0.05$ , ### $P < 0.001$ , \$\$\$ $P < 0.001$ .

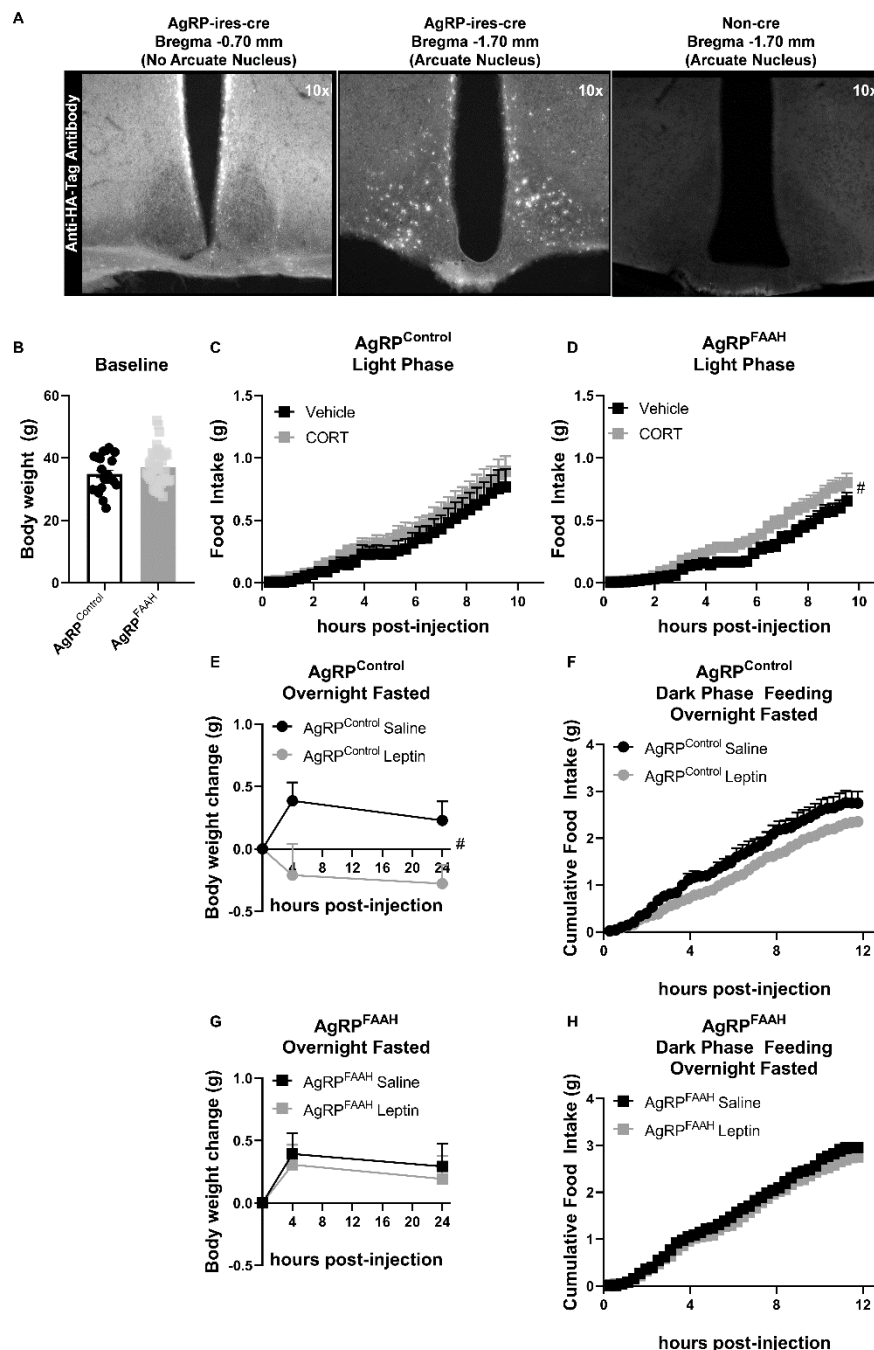
# **FAAH knockdown exclusively in hypothalamic AgRP neurons is sufficient to mediate the exaggerated hyperphagic response to glucocorticoids and ablate the anorexigenic effects of leptin**

Within the hypothalamus, different neuronal populations regulate feeding. For example, activation of agouti-related protein (AgRP) neurons is known to drive feeding (Krashes et al., 2011). Interestingly, it has been shown that glucocorticoids increase the activity of AgRP neurons to elicit feeding (Perry et al., 2019). Therefore, we sought to assess the impact of knocking down FAAH expression exclusively in AgRP neurons on CORT-mediated feeding responses. Here, we used a cre-dependent CRISPR/SaCas9 AAV-based system containing an HA-tagged SaCas9 with a sgRNA directed against FAAH (SaCas9-FAAH) or control (SaCas9-Control) vector (Hunker et al., 2020). To validate these constructs, we examined FAAH activity in *CaMKII $\alpha$ -cre* mice following intra-hippocampal delivery of the AAV vectors. CaMKII $\alpha$  drives cre expression in the forebrain, predominantly in CA1 pyramidal cells of the hippocampus (Tsien et al., 1996), where FAAH is highly expressed (Egertová et al., 1998; Gulyas et al., 2004;

Tsou et al., 1998). Using this strategy, we are able to assess changes in FAAH function on account of CRISPR/SaCas9 gene editing in the hippocampus of *CaMKIIα-cre* mice, where it is otherwise unfeasible in *AgRP-Ires-cre* mice on account of relatively low FAAH activity in the sparsely distributed AgRP neurons. We found that mutagenesis of FAAH (using SaCas9-FAAH) significantly decreased FAAH activity relative to the controls (using SaCas9-Control) in *CaMKIIα-cre* mice (Supplemental Figure S8). To directly examine the role of FAAH in AgRP neurons on CORT-mediated feeding, these AAV-CRISPR/SaCas9 constructs were stereotactically injected into the arcuate nucleus of the hypothalamus of *Agrp-Ires-cre* mice to facilitate mutagenesis exclusively in AgRP neurons. HA-tagged protein expression in the arcuate nucleus was probed to confirm cre-mediated DNA recombination. The HA-tag was detected exclusively in the ARC, in a pattern reminiscent of AgRP neurons (Figure 5A). Furthermore, no expression was detected in non-cre-expressing mice.

Following a minimum of 4 weeks recovery to allow for viral expression and gene knockout, we compared feeding responses to CORT in control mice ( $\text{AgRP}^{\text{control}}$ ) and in mice where FAAH had been knocked down in AgRP neurons ( $\text{AgRP}^{\text{FAAH}}$ ). Body weights were similar between  $\text{AgRP}^{\text{control}}$  and  $\text{AgRP}^{\text{FAAH}}$  mice at the onset of CORT treatment (Figure 5B). In  $\text{AgRP}^{\text{control}}$  mice, there was no hyperphagic effect from a single CORT (3 mg/kg) injection (Figure 5C). However, and consistent with GC's effects in FAAH A/A mice, CORT significantly increased light phase feeding in  $\text{AgRP}^{\text{FAAH}}$  mice (Figure 5D). Interestingly, we found lower basal food intake in  $\text{AgRP}^{\text{FAAH}}$  mice relative to  $\text{AgRP}^{\text{control}}$  mice under vehicle conditions, mimicking the phenotype observed in FAAH A/A mice under basal conditions (Figure 1). Taken together, suppression of FAAH exclusively in hypothalamic AgRP neurons recapitulates the exaggerated hyperphagic response to CORT observed in FAAH A/A mice.

We have previously shown that FAAH A/A mice are unresponsive to the anorectic effects of acute leptin exposure (Balsevich et al., 2018), which agrees with the literature indicating that leptin suppresses endocannabinoid signaling to mediate its feeding responses (Balsevich et al., 2018; Di Marzo et al., 2001). We therefore investigated whether FAAH knockdown exclusively in AgRP neurons is also sufficient to blunt leptin feeding responses. For this purpose, we allowed AgRP<sup>control</sup> and AgRP<sup>FAAH</sup> mice to recover for 5 weeks following the single CORT injection. Following recovery, AgRP<sup>control</sup> and AgRP<sup>FAAH</sup> mice were fasted overnight and then injected with either vehicle (saline) or leptin (2 mg/kg) immediately before receiving food. As expected, leptin reduced body weight gain and food intake following an overnight fast in AgRP<sup>control</sup> mice (Figure 5E – F). However, AgRP<sup>FAAH</sup> mice were insensitive to leptin's effects on weight gain or feeding (Figure G – H), indicating that FAAH knockdown exclusively in hypothalamic AgRP neuron is sufficient to block the anorexigenic effects of leptin. Similar to the effects of CORT, this demonstrates that knockdown of FAAH explicitly in AgRP neurons is sufficient to recapitulate the altered effects of leptin seen in the FAAH C385A mice (Balsevich et al., 2018).



**Figure 5 FAAH knockdown in hypothalamic AgRP neurons recapitulates the feeding responses to either acute CORT or leptin treatment of FAAH A/A mice.** (A) For validation of cre-dependent CRISPR/SaCas9 AAV-based knockdown of FAAH in AgRP neurons, HA-tag fluorescence was detected exclusively in the arcuate nucleus of AgRP-cre mice following bilateral injections of SaCas9-Control or SaCas9-FAAH into the hypothalamus of *AgRP-Ires-cre* mice. Non-cre mice showed no Anti-HA-tag fluorescence in the arcuate nucleus. (B) Under basal conditions, there was no effect of FAAH knockdown in AgRP neurons on body weight ( $T(61) = 1.406$ ,  $p = 0.1648$ ). (C) A single CORT injection (3 mg/kg) had no effect on cumulative light phase



food intake in AgRP<sup>control</sup> mice (treatment:  $F(1, 21.045) = 0.465$ ,  $p = 0.503$ ). (D) By contrast, acute  
CORT caused a hyperphagic feeding response in AgRP<sup>FAAH</sup> mice measured during the light phase  
(treatment:  $F(1, 44.286) = 4.257$ ,  $p = 0.045$ ). (E – F) In terms of leptin exposure, AgRP<sup>control</sup> mice  
were responsive to leptin-mediated reductions in weight gain (treatment:  $F(1, 22.253) = 8.282$ ,  $p$   
 $= 0.009$ ) and refeeding (treatment:  $F(1, 16.659) = 5.635$ ,  $p = 0.030$ ) following an overnight fast.  
(G–H) However, AgRP<sup>FAAH</sup> mice were unresponsive to leptin (weight gain:  $F(1, 32.278) = 0.181$ ,  
 $p = 0.673$ ; refeeding:  $F(1, 30.596) = 0.703$ ,  $p = 0.408$ ). For panel B,  $n = 21$  AgRP<sup>control</sup> mice ;  $n =$   
 $42$  AgRP<sup>FAAH</sup> mice. For panels C,  $n = 12$  AgRP<sup>control</sup> -vehicle;  $n = 10$  AgRP<sup>control</sup> -CORT. For panel  
D,  $n = 20$  AgRP<sup>FAAH</sup> -vehicle;  $n = 21$  AgRP<sup>FAAH</sup> -CORT. For panels E & F,  $n = 7$  AgRP<sup>control</sup> -saline;  
 $n = 7$  AgRP<sup>control</sup> -leptin. For panels G & H,  $n = 14$  AgRP<sup>FAAH</sup> -saline ;  $n = 15$  AgRP<sup>FAAH</sup> -leptin. Body  
weight and food intake data are presented as means  $\pm$  SEM. Panel B was analyzed by student's t-  
test. Panels C –H were analyzed by linear mixed models with repeated measures. Pound signs  
denote significant treatment effect. # $P < 0.05$ .

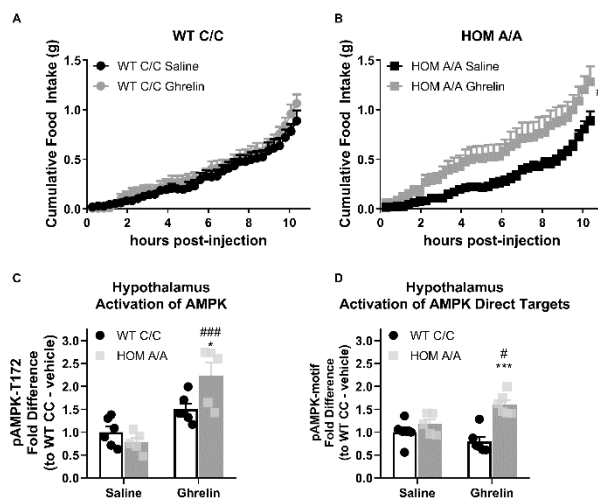
### **FAAH A/A mice are more sensitive to the acute hyperphagic effects of ghrelin**

Endocannabinoids not only play a central role in GC-mediated metabolic outcomes, but  
furthermore are integral to the orexigenic effects of ghrelin (Kola et al., 2008; Tucci et al., 2004).  
Therefore, we further investigated the generalizability of this phenomenon to other orexigenic  
signals beyond GC, and as such explored whether the FAAH C385A variant affects ghrelin-  
mediated feeding effects. To address this question, we examined the feeding responses of FAAH  
C/C and FAAH A/A mice to a single, subthreshold injection of ghrelin (1 mg/kg) or saline  
delivered during the light/inactive phase to capture the hyperphagic effects of ghrelin during a  
period of low feeding. Similar to acute GC exposure, a single exposure to ghrelin increased light  
phase feeding exclusively in FAAH A/A mice (Figure 6). By 24 h post-injection, the effects of  
ghrelin on cumulative food intake were no longer evident (Supplemental Figure S9). There was  
also no lasting effect of ghrelin on body weight gain measured at 24 h post-injection. Ghrelin  
likewise significantly increased RER during the light phase exclusively in FAAH A/A mice,  
which agrees with the hyperphagic effect (Supplemental Figure S9). The elevated RER tended ( $p$



= 0.068) to continue 24 h post ghrelin injections in FAAH A/A mice. By contrast, there was no effect of ghrelin on energy expenditure in either FAAH C/C or A/A mice.

Similar to hypothalamic GC signaling, ghrelin also activates AMPK signaling in the hypothalamus to increase feeding through an endocannabinoid-dependent manner (Kola et al., 2008). To determine whether hypothalamic AMPK is differentially affected by the FAAH C385A variant, we again examined the phosphorylation status of AMPK and the direct targets of AMPK as a proxy of AMPK signaling activation in FAAH C/C and A/A mice. Ghrelin (1 mg/kg) significantly increased hypothalamic levels of phosphorylated AMPK and direct downstream AMPK targets specifically and exclusively in FAAH A/A mice 1 h post-injection (Figure 6). Together, our data indicate that the effects of the FAAH C385A variant are not selective for GC-mediated effects. Rather, the FAAH C385A variant exaggerates both GC and ghrelin orexigenic effects, which may suggest a generalized sensitivity of the FAAH A/A genotype to orexigenic signals, coupled to an impairment in the sensitivity to anorectic signals, such as leptin.



**Figure 6 FAAH A/A mice are hyperresponsive to the orexigenic effects of ghrelin.** (A – B) Acute ghrelin (1 mg/kg, single i.p. injection) had no effect on cumulative food intake in FAAH

C/C mice (treatment:  $F(1, 19.753) = 0.494$ ,  $p = 0.490$ ) yet increased light phase food intake in FAAH A/A mice (within the initial 10 h following ghrelin injections) (treatment:  $F(1, 21.835) = 5.406$ ,  $p = 0.030$ ). (C) Hypothalamic AMPK activity (as assessed by pAMPK protein expression) was significantly increased by ghrelin treatment exclusively in FAAH A/A mice (treatment x genotype:  $F(1, 18) = 8.071$ ,  $p = 0.0108$  plus post hoc testing). (D) Similarly, downstream targets of AMPK (with a shared consensus sequence) were activated from ghrelin exposure exclusively in FAAH A/A mice measured 1 h following injections (treatment x genotype:  $F(1, 19) = 9.597$ ,  $p = 0.0059$  plus post hoc testing). For panel A,  $n = 8$  WT-saline;  $n = 10$  WT-ghrelin. For panel B,  $n = 11$  HOM-saline;  $n = 10$  HOM-CORT. For panel C,  $n = 6$  WT-saline biological replicates;  $n = 6$  WT-ghrelin biological replicates;  $n = 5$  HOM-saline biological replicates;  $n = 5$  HOM-ghrelin biological replicates. For panel D,  $n = 6$  WT-saline biological replicates;  $n = 6$  WT-ghrelin biological replicates;  $n = 5$  HOM-saline biological replicates;  $n = 6$  HOM-ghrelin biological replicates. Food intake data are presented as means  $\pm$  SEM. Protein expression data are expressed as relative fold change compared to FAAH C/C-vehicle condition  $\pm$  SEM. Panels A – B were analyzed by linear mixed models with repeated measures. Panels C – D were analyzed by 2-way ANOVAs. Pound signs denote significant treatment effect. Asterisks denote significant genotype effect. \* $P < 0.05$ , \*\* $P < 0.01$ , # $P < 0.05$ , ### $P < 0.001$ .

## Discussion

The ECS, and specifically anandamide signaling, has an important role in the regulation of body weight. In particular, activation of anandamide signaling is known to promote a positive energy balance and weight gain (Mazier et al., 2015). Moreover, endocannabinoid signaling lies downstream of several metabolic signals. For example, it is known that GCs and ghrelin, both orexigenic signals, mobilize endocannabinoid signaling, whereas anorexigenic leptin is known to suppress endocannabinoid signaling (Balsevich et al., 2018; Bowles et al., 2015; Di Marzo et al., 2001; Kola et al., 2008; Tucci et al., 2004). Therefore, genetic factors that modify endocannabinoid signaling may affect individual responses to orexigenic and anorexigenic signals. FAAH, the primary catabolic enzyme for anandamide, regulates anandamide signaling by controlling its levels. In humans, the common missense mutation (C385A) in FAAH affects FAAH protein stability, with the A/A genotype associated with approximately half the FAAH protein expression and enzymatic activity compared with the C/C genotype (Chiang et al., 2004).

Here we fully characterized the effects of the functional FAAH C385A mutation on GC-dependent metabolic outcomes, establishing underlying molecular and cellular mechanisms that couple FAAH-AEA signaling to GC-induced feeding responses. We furthermore demonstrated that the FAAH C385A variant is not selective for GC-mediated metabolic responses, but rather governs an individual's sensitivity to additional orexigenic signals, namely ghrelin.

As a first step, we monitored the metabolic phenotype of FAAH C/C and A/A mice under basal conditions. In this condition, FAAH C/C and A/A mice did not present differences in body weight or body composition. Interestingly, the underlying metabolic parameters contributing to body weight regulation differed between FAAH C/C and A/A mice at baseline. Compared to FAAH C/C mice, FAAH A/A mice showed lower basal food intake and matching lower energy expenditures. This phenotype could be driven by basal elevations in AEA causing reduced basal metabolism, which in turn results in lower food intake to balance the alterations in energy expenditure. Alternately, in addition to AEA, FAAH also regulates the metabolism of fatty acid amides such as oleoylethanolamide, which is known to be a potent anorectic signal (Gaetani et al., 2003). Regardless of the exact mechanism of this subtle basal phenotype, this balance between food intake and energy expenditure offsets any differences in body weight or body composition at baseline. However, it may also predispose subjects with the FAAH A/A genotype to weight gain from overconsumption. Indeed, a low basal metabolic rate is a risk factor for weight gain in both humans and rodents (Astrup et al., 1996, 1999; Buscemi et al., 2005; Kunz et al., 2000; Lazzer et al., 2010; Sadowska et al., 2017). Lower basal energy expenditure relates to lower basal maintenance costs, and consequently an energy imbalance could easily arise from overconsumption in response to specific environmental contexts (abundant access to palatable food) or endocrine states (elevated levels of orexigenic signals). In fact, this is in line with our

findings that FAAH A/A mice show an amplified feeding response to GC and ghrelin orexigenic signals.

In terms of GC-endocannabinoid signaling interactions, it is recognized that the duration of GC exposure affects the interaction between them (reviewed by (Balsevich et al., 2017)). For example, GCs mobilize endocannabinoids through both rapid, non-genomic mechanisms and delayed, genomic mechanisms. Therefore, we examined 3 treatment durations to assess whether the FAAH C385A polymorphism is relevant to the chronic (4 weeks), immediate (48 h), and/or acute (single exposure) metabolic outcomes of GCs. For chronic and immediate GC treatments, we delivered a low dose of corticosterone (CORT; 25 µg/mL) in the drinking water, which maintains the diurnal rhythmicity of circulating GCs (Karatsoreos et al., 2010). We found that regardless of treatment duration, GC exposure increased food intake exclusively in FAAH A/A mice. Furthermore, continuous GC exposure delivered for 48 h, increased weight gain within the first 24 h and 4 weeks of continuous GC exposure exaggerated GC-dependent weight gain. By contrast, there was no body weight phenotype 24 h following acute GC injections, suggesting that continuous GC treatment is required to elicit weight gain. This agrees with the body of literature demonstrating that in the short-term, GC actions are adaptive, redirecting energy stores and increasing food intake without affecting body weight maintenance (Dallman et al., 2004). However, prolonged GC exposure becomes maladaptive, as seen with chronic stress, which is associated with hypercortisolism, fat accumulation, and central obesity. In this context, our findings indicate that individuals with the FAAH A/A genotype are more susceptible to weight gain when confronted with elevations in GC signaling, and therefore may be particularly vulnerable to the influence of stress (Pecoraro et al., 2004; Ulrich-Lai, 2016).

The effects of the FAAH C385A variant on GC-mediated metabolic outcomes were specific to feeding responses. In particular, whereas the FAAH A/A genotype amplified the hyperphagic effects of GCs, there was no genotype effect on GC-dependent changes to energy expenditure or substrate utilization. Specifically, GC exposure decreased energy expenditure and increased carbohydrate utilization in both FAAH C/C and A/A mice. This agrees with the known role of GCs to suppress energy expenditure and promote carbohydrate utilization (Dulloo et al., 1990; Flatt, 1993; Poggioli et al., 2013). Therefore, our data suggest that while GCs mobilize AEA signaling to mediate feeding responses, it is unclear whether AEA signaling lies downstream of GC actions to regulate energy expenditure or substrate utilization. Regardless, the data herein delineate how the FAAH C385A variant affects energy balance by demonstrating that the FAAH C385A polymorphism modifies GC-induced weight gain through its effect on feeding rather than energy expenditure.

The exaggerated hyperphagic response to GC exposure was associated with increased hypothalamic anandamide levels exclusively in FAAH A/A mice. The hypothalamus is certainly recognized as one of the major brain regions responsible for endocannabinoid-mediated effects on feeding. Therefore, it is likely that elevated hypothalamic anandamide in the FAAH A/A genotype drives the heightened feeding response to orexigenic signals. Interestingly, one human study found that the C385A SNP did not associate with increased BMI per se, but rather higher levels of anandamide were positively associated with both the FAAH A/A genotype and a higher BMI (De Moraes Martins et al., 2015). In agreement, we show that under basal conditions there is neither an effect of the A/A genotype on body weight nor anandamide content in brain regions central to endocannabinoid-mediated feeding. However, under the influence of specific endocrine drivers, such as GCs, anandamide levels increased exclusively in FAAH A/A mice

within the hypothalamus, which likewise associated with a hyperphagic response selectively in the FAAH A/A mice. Taken together, GCs elevate hypothalamic anandamide levels selectively in the FAAH A/A genotype to sensitize the FAAH A/A genotype to hyperphagic feeding outcomes.

We next sought to characterize the cellular population within the hypothalamus driving the AEA-dependent exaggerated feeding response to GC exposure. Within the arcuate nucleus of the hypothalamus, POMC- and AgRP-expressing neurons are considered the prototypic regulators of homeostatic food intake (Gautron et al., 2015). Whereas POMC neuronal activity promotes satiety, AgRP neuronal activity promotes feeding. Interestingly, GCs increase AgRP neuronal firing rates to induce hyperphagia (Perry et al., 2019). Therefore, we examined whether FAAH knock down exclusively in AgRP neurons is sufficient to recapitulate the exaggerated feeding response to GC exposure seen in FAAH A/A mice. For this purpose, we employed a recently described conditional single vector CRISPR/SaCas9 virus system (Hunker et al., 2020) for efficient cre-dependent mutagenesis of *Faah* (or control) into *AgRP-Ires-cre* mice. Remarkably decreasing FAAH expression selectively in AgRP neurons paralleled the feeding phenotype of FAAH A/A mice in response to acute CORT injections. Specifically, under vehicle conditions AgRP<sup>FAAH</sup> mice displayed reduced cumulative food intake despite no difference in basal body weights compared to AgRP<sup>Control</sup> mice. However, following a single injection of CORT, AgRP<sup>FAAH</sup> mice increased their light phase feeding, a period when feeding is typically low. By contrast, this hyperphagic response to acute CORT was absent in AgRP<sup>Control</sup> mice. Therefore, this study demonstrates that lowering FAAH expression exclusively in AgRP neurons is sufficient to recapitulate the hyperphagic feeding response of FAAH A/A mice to acute GC treatment.

Given the effects of FAAH knockdown in AgRP neurons on GC-dependent feeding responses, we investigated whether AgRP neurons are likewise important cellular mediators of FAAH's effects on leptin-dependent feeding outcomes. It is known that leptin suppresses hypothalamic endocannabinoid signaling to decrease feeding (Balsevich et al., 2018; Cardinal et al., 2012; Di Marzo et al., 2001; Jo et al., 2005). Furthermore we have specifically shown that elevated AEA signaling in FAAH A/A mice prevents the anorectic effects of leptin following 16 h food deprivation (Balsevich et al., 2018). Therefore, we hypothesized that FAAH deletion specifically in AgRP neurons would dampen the hypophagic responses to leptin, similar to what was seen in FAAH A/A mice. In agreement with this, AgRP<sup>Control</sup> mice exhibited weight loss and reduced feeding in response to acute leptin exposure. However, AgRP<sup>FAAH</sup> mice were unresponsive to acute leptin. Our results indicate that AgRP neurons are an important cellular population mediating the effects of FAAH-AEA signaling events of multiple upstream endocrine signals regulating feeding behavior.

We next explored whether the FAAH C385A variant modulates the metabolic outcomes of additional orexigenic signals. Specifically, we examined the metabolic phenotypes of FAAH C/C and A/A mice in response to ghrelin exposure based on the known interactions between ghrelin and endocannabinoid signaling pathways (Edwards & Abizaid, 2016). A single, subthreshold dose of ghrelin paralleled the outcomes arising from acute GC exposure, wherein ghrelin increased light phase food intake exclusively in FAAH A/A mice. Again, the effects were specific to feeding as ghrelin had no effect on energy expenditure across genotypes. Considering chronic stress elevates circulating GC and ghrelin levels (Abizaid, 2019), it is reasonable to suspect that individuals presenting the FAAH A/A genotype are more susceptible to stress-associated weight gain on account of amplified responses to these orexigenic signals. Follow-up



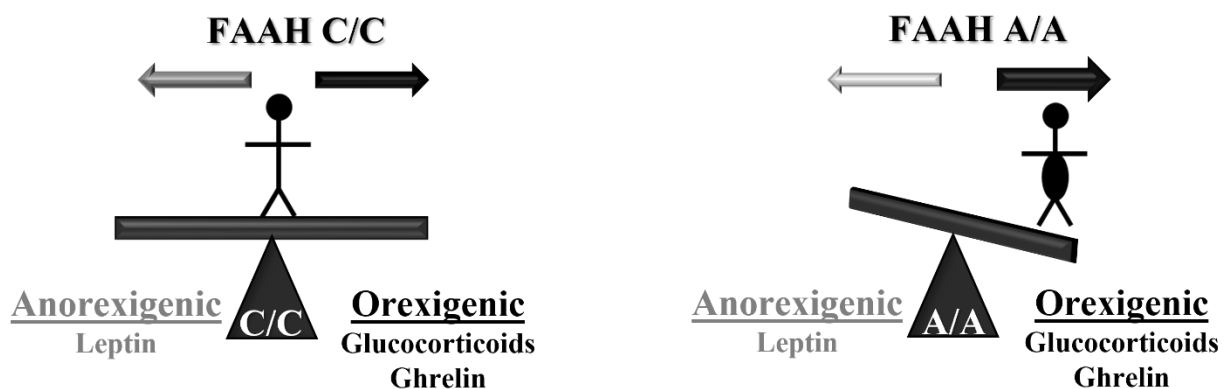
studies should address the feeding response of FAAH C/C and A/A mice to acute and chronic stress exposures

An important question raised by this study is how does the FAAH C385A variant coordinate the feeding responses to multiple orexigenic signals? For instance, is there a common pathway downstream of FAAH-AEA signaling driving the orexigenic feeding responses? Along these lines, it has previously been shown that GCs and ghrelin activate hypothalamic AMPK through a CB1R-dependent mechanism (Kola et al., 2008; Scerif et al., 2013). Indeed, hypothalamic AMPK activity elicits powerful feeding responses (Minokoshi et al., 2004). In agreement, we found that AMPK signaling is heightened in the hypothalamus following acute exposure to either GCs or ghrelin exclusively in FAAH A/A mice. Moreover, pretreatment with an AMPK inhibitor blocked the orexigenic response to acute GC treatment in FAAH A/A mice. The data herein provide unequivocal evidence that AMPK lies downstream of FAAH-AEA signaling to coordinate ghrelin and GC signaling.

The current study clearly demonstrates that the FAAH A/A genotype predisposes individuals to the feeding responses of orexigenic signals, CORT and ghrelin. Specifically, our working model is that orexigenic signals mobilize AEA in AgRP neurons of the hypothalamus. This effect is amplified in the A/A genotype (with low FAAH expression) because there is reduced enzymatic degradation of AEA. The exaggerated AEA levels lead to heightened CB1R activation and downstream AMPK activation to ultimately sensitize A/A individuals to orexigenic signaling outcomes. By contrast, our previous work determined that FAAH A/A mice are less responsive to the anorexigenic signal leptin (G Balsevich et al., 2018). Furthermore, the current study established that FAAH knockdown exclusively in AgRP neurons is sufficient to blunt feeding and body weight responses to leptin. Thus, based on the current study and our



previous findings, we demonstrate that the FAAH C385A variant shifts an individual's sensitivity to metabolic feeding signals, where individuals with the FAAH A/A genotype are more responsive to orexigenic signals at the expense of being less responsive to anorexigenic signals (Figure 7). Given orexigenic signals favor weight gain and anorexigenic signals favor weight loss, individuals with the minor A/A genotype may show exaggerated weight gain in response to environmental conditions that increase orexigenic signals (such as GCs and ghrelin) and decrease anorexigenic signals (such as leptin). Chronic stress for example, is known to increase levels of both GCs and ghrelin while also producing decreased levels of leptin (Bouillon-Minois et al., 2021; Lu et al., 2006). Collectively, our data indicate that human studies be conducted for the FAAH C385A polymorphism in a manner that considers external environmental contexts.



**Figure 7 Working model of how the FAAH C385 polymorphism influences individual susceptibility to body weight gain.** Individuals with the FAAH C/C genotype are able to respond to anorexigenic and orexigenic signals based on their metabolic needs. However, the FAAH A/A genotype shifts responses to feeding cues, sensitizing individuals to orexigenic signals, favoring weight gain, and desensitizing individuals to weight loss.

## Methods

**Animals** The FAAH C385A knockin mouse line (MGI # 5644434) had been previously generated and characterized (Dincheva et al., 2015). The *Agrp-Ires-cre* mice (JAX #012899) and the CamKII $\alpha$ -cre mice (JAX # 005359) were purchased from Jackson Laboratories (Bar Harbour, Maine). For all experiments, age-matched male mice between 3 and 6 months were used. All mice were group-housed until 1-2 weeks before the experiment onset at which point mice were individually housed. For the duration of the experiments, mice were maintained under controlled temperature (25 +/- 1°C) and humidity (30 +/- 5%) conditions. For acute studies with CORT and ghrelin, mice were maintained on a 12:12 h light/dark cycle with lights on at 08:00. For extended exposure studies, mice were maintained on a 12 h reverse dark/light schedule, with lights on at 22:00. For acute leptin study, mice were also maintained on a 12 h reverse dark/light schedule with lights on at 22:00. Mice received ad libitum access to tap water and standard lab chow. Although sample size calculations were not performed, sample sizes for each animal study were determined from prior studies in our laboratory examining metabolic outcomes (feeding and energy expenditure) in mice (Balsevich et al., 2018; Bowles et al., 2015). Mice of each genotype were randomly assigned to a treatment group counterbalanced for body weight. Blinding was not employed. The experimental unit for all studies was a single adult, male animal. All studies were carried out in compliance with the ARRIVE guidelines. All protocols had been approved by the University of Calgary Animal Care Committee and were carried out in accordance with Canadian Council on Animal Care.

**Chemical reagents** Corticosterone (Sigma, 27840), human ghrelin (Tocris, 1463), recombinant mouse leptin (Cedarlane Laboratories Ltd, CLCYT351-2), and Compound C (Selleck Chemicals,

S7306), were used in all in vitro and in vivo experiments. All reagents were prepared freshly immediately before the onset of use.

**Extended CORT Exposure** For 48-h and 30-day CORT exposure, mice were given CORT (25 µg/ml) or vehicle (1% ethanol) through their drinking water as previously described (Karatsoreos et al., 2010). Briefly, treatment began at the onset of their dark (active phase) and continued for either 48-h or 30-days. Metabolic measurements were monitored throughout the treatment period. For 30-day treatment, fresh CORT/Vehicle-treated water was supplied every 3-4 days. At the end of the experimental period, mice were sacrificed by rapid decapitation at the onset of the dark phase. Tissues were rapidly frozen and stored at -80 °C until further processed.

**Acute Exposure Studies** For acute exposure, mice were administered with CORT (3 mg/kg) or vehicle (Saline/DMSO/Tween-80, 18:1:1) by intraperitoneal injection (i.p.). Similarly, ghrelin (1 mg/kg) or vehicle (saline) and leptin (2 mg/kg) or vehicle (saline) were administered by i.p. injection. For CORT and ghrelin studies, drug administration occurred 2h after the onset of the light (inactive) phase to capture hyperphagic responses in light-phase food consumption. For leptin study, drug administration occurred at the onset of the dark phase following an overnight fast (~16h). After 24 h of metabolic measurements, mice were allowed to recover in their home cages for at least one week before sacrifice. On the day of sacrifice, mice were administered with their respective drug 1h prior to rapid decapitation and tissue collection. Harvested tissues were stored at -80°C until further processing.

**Pharmacological Blockade of AMPK** Compound C (5 mg/kg), an AMPK inhibitor, or vehicle (saline) were administered by i.p. injections 1 h before injecting CORT (3 mg/kg) or vehicle to mice according to the ‘Acute Exposure Studies’ section above. Briefly, mice received compound

C or saline at the onset of the light phase. 1 h later, mice received a second injection of either  
CORT or vehicle in a 2x2 design. Metabolic measurements were monitored over the next 24 h.

**Generation of AAV1-CMV-FLEX-HA-Tag, SaCas9-U6-sgRNA** The single guide (sg)RNA for targeting exon 1 of *Faah* was designed as previously described (Hunker et al., 2020). Briefly, the sgRNA sequence targeting exon 1 of *Faah* (5'- CTGCAGGCTAGGCAAACC-3') and a control sgRNA sequence (5'- CTGCAGGCTAGGCAAACCTTT-3' were synthesized (Invitrogen) and cloned into the shuttle plasmid for adeno-associated viral (pAAV-FLEX-SaCas9-U6-sgRNA; Addgene #124844) to generate a plasmid for cre-dependent expression of HA-tagged *Staphylococcus aureus* Cas9 (SaCas9) and selective mutagenesis of *Faah* (SaCas9-FAAH) (Hunker et al., 2020). The control AAV vector (SaCas9-Control) had an identical guide except the 3 terminal base pairs in the seed region of the guide sequence were mutated. Following sequence validation, AAV serotype 1 (AAV1) vectors were generated for SaCas9-FAAH and SaCas9-Control using the packaging plasmid pDG1 and transient transfection of HEK293T cells, as described (Hunker et al., 2020).

**Stereotaxic Surgical Procedure** Mice were anaesthetized with isofluorane and placed on a stereotaxic frame. For FAAH knockdown in AgRP neurons, a cre-dependent CRISPR/Cas9 AAV vector targeting *Faah* (SaCas9-FAAH) or control (SaCas9-Control) was delivered bilaterally (210 nl per side) into the arcuate nucleus of the hypothalamus (AP: -1.6 mm, ML +/- 0.15 mm, DV -5.8 mm). For FAAH knockdown in CamKII $\alpha$ -cre mice, SaCas9-FAAH or SaCas9-Control was bilaterally injected (400 nl per side) into the dorsal hippocampus (AP: -2.1 mm, ML: +/-1.3 mm, DV: -1.4mm). Virus was injected at a rate of 0.1  $\mu$ l min<sup>-1</sup>. Metacam (3 mg/kg) was given as a postoperative analgesic. Animals recovered for a minimum of 4 weeks in their home cage to allow for viral expression before the onset of experimentation.

**FAAH Activity** To assess FAAH activity, hippocampi were collected from CamKII $\alpha$  mice that either received SaCas9-FAAH or SaCas9-Control vector. Briefly, hippocampi were excised on ice and immediately snap frozen and stored at  $-80^{\circ}\text{C}$ . Hippocampi were homogenized and membrane fractions were isolated as previously described (Gray et al., 2015). Enzymatic activity of FAAH was measured as the conversion of [3H]-labeled AEA (in the ethanolamine portion of the molecule) to [3H]ethanolamine as previously described (Maccarrone et al., 1999). Membranes were incubated in TME buffer (50 mM Tris-HCl, 3.0 mM  $\text{MgCl}_2$ , and 1.0 mM EDTA, pH 7.4) containing 1.0 mg/ml fatty acid-free bovine serum albumin and 0.2 nM [3H]AEA. Calibration curves were prepared using eight concentrations of AEA at concentrations between 10 nM and 10  $\mu\text{M}$ . Incubations were kept at  $37^{\circ}\text{C}$  and were stopped by adding ice-cold chloroform/methanol (1:2). Samples were incubated at room temperature for 30 min before the addition of chloroform and water. Aqueous and organic phases were separated by centrifugation at 1,000 rpm for 10 min. The concentration of [3H] in the aqueous phase was determined by liquid scintillation counting and the conversion of [3H]AEA to [3H]ethanolamine was calculated. The maximal hydrolytic activity ( $V_{\text{max}}$ ) for this conversion was determined by fitting the data to the Michaelis-Menten equation in Prism v8 (GraphPad, San Diego, CA, USA).

**Metabolic Measurements** Energy expenditure (heat), food intake, and respiratory exchange ratio ( $\text{RER} = \text{CO}_2 \text{ produced} / \text{O}_2 \text{ consumed}$ ) were measured by indirect calorimetry in Comprehensive Lab Animal Monitoring System (CLAMS) metabolic chambers (Columbus Instruments; Columbus, OH, USA) as previously described (Pezeshki et al., 2015). Following 2-4 days of acclimatization to the chambers, metabolic measurements were performed every 14 minutes over the experimental period (flow rate  $0.5 \text{ L} \cdot \text{min}^{-1}$ ). Total energy expenditure was calculated using  $\text{O}_2$  consumption ( $\text{VO}_2$ ,  $[\text{ml h}^{-1}]$ ) and  $\text{CO}_2$  production ( $\text{VCO}_2$ ,  $[\text{ml h}^{-1}]$ ) in the

following equation following a previously published protocol (Pezeshki et al., 2015): Calorific Value =  $VO_2 \times [3.815 + (1.232 \times RQ)]$ , with data collection and processing using Oxymax v5.2 and CLAX v2.2.2. Data are represented as  $\text{kcal} \cdot \text{h}^{-1}$ . When mice spilled their food, their food intake was excluded from analyses (see Supplementary Source Data). Body composition was examined by quantitative magnetic resonance using the Minispec LF-110 NMR Analyzer (Bruker Optics, Milton, ON, Canada) on awake mice.

**Home-cage activity** Mice were surgically implanted with a telemetric transponder (Data Sciences International, DSI, St. Paul, USA) to examine activity counts in their home-cages. Briefly, under isoflurane anesthesia, a small incision was made along the midline of the abdomen. The abdominal wall was opened and the transponder was implanted into the abdominal cavity of the mice. The incision was closed with absorbable sutures and the mice were allowed to recover for one week before home-cage activity counts were measured under basal conditions and during 24 h CORT exposure (delivered in their drinking water). Metacam (3 mg/kg) was given as a postoperative analgesic.

**Endocannabinoid Quantification** Lipids were extracted from harvested tissues as described previously (Hill et al., 2009). Briefly, weighed tissue samples were homogenized in borosilicate glass culture tubes containing 2 ml of acetonitrile with 84 pmol of [2H8]anandamide. Tissues were subsequently sonicated for 30 min. To precipitate proteins, samples were incubated overnight at  $-20^{\circ}\text{C}$  proteins, followed by centrifugation at  $1,500 \times g$  to remove particulates. The supernatants were collected and evaporated to dryness using  $\text{N}_2$  gas. Next, samples were suspended in methanol and dried using  $\text{N}_2$  gas. The resulting lipid extracts were suspended in methanol and stored at  $-80^{\circ}\text{C}$  until further processed. AEA tissue levels were subsequently

determined using isotope-dilution, liquid chromatography-mass spectrometry as described previously (Patel et al., 2005).

**Corticosterone Quantification** Plasma corticosterone levels were determined using a commercially available enzyme immunoassay kit (Arbor Assays, Ann Arbor, USA, Catalogue Number K014-H5; sensitivity 20.9 pg/ml) according to the manufacturer's instructions.

**Cell lines and culture conditions** The GT1-7 hypothalamic cell line (RRID:CVCL\_0281) was kindly provided by Dr. Pamela Mellon (Salk Institute, La Jolla, CA). Cells were grown in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% fetal bovine serum (FBS), 25 mM glucose and 1% penicillin/streptomycin at 37°C in 95% O<sub>2</sub> and 5% CO<sub>2</sub>. Once cells reached ~80% confluence, experiments were initiated. Briefly, cells were starved for 4 h in DMEM supplemented with 1 mM glucose. During this incubation period, FAAH inhibitor URB597 (1 μM for 4h), or vehicle (DMSO for 4h) was added to the media. To examine the effects of CORT and CORTxFAAH interactions on AMPK activity, CORT (1 μM) or vehicle (DMSO) was added to the media for an additional 2 h. After 6 h, cells were immediately trypsinized, centrifuged, and stored as pellets at -80 °C until further processed.

**Protein Quantification** Tissues or cells were homogenized in Pierce IP Lysis buffer supplemented with protease inhibitor cocktail 3 (Merck Millipore, Darmstadt, Germany) and phosphatase inhibitor PhosSTOP (Roche, Penzberg, Germany). Following centrifugation, the supernatant (total protein extract) was collected. Samples were subsequently prepared for the automated capillary western blot system, WES System (ProteinSimple, Bio-Techne, San Jose, CA, USA) according to the manufacturer's instructions. Target proteins were probed with the following antibodies: AMPKα (1:1,00, Cell Signaling Technology, #2532, RRID:AB\_330331), pAMPK (T172) (1:50, Cell Signaling Technology, #2531, RRID:AB\_330330), pAMPK

substrate motif (1:50, Cell Signaling Technology, #5759, RRID:AB\_10949320), and secondary antibody anti-rabbit IgG, HRP-linked antibody (1:10,000, Cell Signaling Technology, #7074, RRID:AB\_2099233). Total protein was quantified using the protein normalization module (ProteinSimple, Bio-Techne, San Jose, CA, USA, #DM-PN02). Signal intensities of phosphorylated proteins were normalized to their respective non-phosphorylated proteins. Quantitative analysis was performed using the Compass software (Protein Simple, Bio-Techne, San Jose, CA, USA) and data expressed as fold change relative to control group..

**Immunohistochemistry** For validation of cre-dependent CRISPR/SaCas9 AAV-vector expression, HA-tag fluorescence HA-tagged SaCas9 was visualized using the following antibodies: rabbit anti-HA-tag (Sigma-Aldrich, #H6908, RRID:AB\_260070) and secondary antibody anti-rabbit Alexa Fluor 488 (1:1000 ThermoFisher, # 32731, AB\_2633280). Briefly, mice were deeply anaesthetised with pentobarbital before undergoing intracardial perfusion. Mice were perfused with PBS before being fixed with a 4% paraformaldehyde solution (PFA). Brains were then removed and dropped into a 4% PFA solution for 24h. Following drop fixing, brains were placed into a 30% sucrose solution for 48h for cryoprotection. Fixed brains were frozen and sliced at 40µm thickness on a microtome and stored in an antifreeze solution until they were ready to be stained. Before staining for HA, slices were washed three times in PBS and three times in PBST. Following washing steps, slices were stained in a 1:1500 dilution of anti-HA for 24 h at 4 °C. Following staining, slices were washed in PBST 3 more times, then stained with a 1:200 dilution of Alexa fluor 488 for 1 hour. Slices then underwent three more washes in PBST before being mounted on glass slides and covered with fluoroshield<sup>TM</sup> with DAPI stain (Sigma F6057). Mounted slices were imaged using a fluorescent microscope (Leica



DM4000 B LED), and CRISPR expression was imaged under an L5 filter cube. Brightness and contrast were adjusted equally across all images in ImageJ.

**Statistical Analysis** Data were analyzed using IBM SPSS Statistics 26 software (IBM SPSS Statistics, IBM, Chicago, IL, USA). Independent t-tests were used to compare genotype effects. Two-way ANOVAs (analysis of variance) were used to examine studies with a 2x2 design. Where the initial test yielded a significant interaction, independent Student's t-tests (two-tailed) were conducted to locate the interaction effect using simple comparisons. Repeated measures on CLAMS data (energy expenditure, respiratory quotient, and food intake) were analyzed by linear mixed models. The fixed effects of treatment, genotype, time, and their interactions were included in the model. Animal nested in the group was the random variable on which repeated measures were taken and covariance structures modeled (Pezeshki et al., 2015). Body weight was included as a covariate in the analyses of energy expenditure (Tschöp et al., 2011). *Post-hoc* analyses were undertaken using Bonferroni's multiple comparison *post-hoc* test where appropriate. Statistical analyses of CLAMS data were performed using individual data points (every 15 minutes) collected over the experimental period. For 24-h measurements, CLAMS data were graphed using the 4-point averages of individual data points. Outlier testing was performed using Grubb's test for outliers, and when an outlier was detected ( $p < 0.05$ ), it was omitted from analysis. For the 48-h CORT exposure study, select metabolic chambers (see Supplementary source data) had dysfunctional O<sub>2</sub>/CO<sub>2</sub> sensors and were excluded from RER and TEE analyses. Statistical significance was set at  $p < 0.05$ ; a statistical tendency was set at  $p < 0.1$ . For interactions at  $p < 0.1$ , we also examined lower order main effects. Data are presented as the mean  $\pm$  SEM.

# References

- Abizaid, A. (2019). Stress and obesity: The ghrelin connection. *Journal of Neuroendocrinology*, 31(7), 1–8. <https://doi.org/10.1111/jne.12693>
- Andersson, U., Filipsson, K., Abbott, C. R., Woods, A., Smith, K., Bloom, S. R., Carling, D., & Small, C. J. (2004). AMP-activated Protein Kinase Plays a Role in the Control of Food Intake. *Journal of Biological Chemistry*, 279(13), 12005–12008. <https://doi.org/10.1074/jbc.C300557200>
- Astrup, A., Buemann, B., Toubro, S., Ranneries, C., & Raben, A. (1996). Low resting metabolic rate in subjects predisposed to obesity: a role for thyroid status. *The American Journal of Clinical Nutrition*, 63(6), 879–883. <https://doi.org/10.1093/ajcn/63.6.879>
- Astrup, A., Gøtzsche, P. C., van de Werken, K., Ranneries, C., Toubro, S., Raben, A., & Buemann, B. (1999). Meta-analysis of resting metabolic rate in formerly obese subjects. *The American Journal of Clinical Nutrition*, 69(6), 1117–1122. <https://doi.org/10.1093/ajcn/69.6.1117>
- Balsevich, G, Petrie, G. N., & Hill, M. N. (2017). Endocannabinoids: Effectors of glucocorticoid signaling. *Frontiers in Neuroendocrinology*, 47. <https://doi.org/10.1016/j.yfrne.2017.07.005>
- Balsevich, G, Sticht, M., Bowles, N. P., Singh, A., Lee, T. T. Y., Li, Z., Chelikani, P. K., Lee, F. S., Borgland, S. L., Hillard, C. J., McEwen, B. S., & Hill, M. N. (2018). Role for fatty acid amide hydrolase (faah) in the leptin-mediated effects on feeding and energy balance. *Proceedings of the National Academy of Sciences of the United States of America*, 115(29). <https://doi.org/10.1073/pnas.1802251115>

781 Balsevich, G., Petrie, G. N., & Hill, M. N. (2017). Endocannabinoids: Effectors of glucocorticoid  
782 signaling. *Frontiers in Neuroendocrinology*, 47, 86–108.  
783 <https://doi.org/10.1016/j.yfrne.2017.07.005>

784 Balsevich, G., Sticht, M., Bowles, N. P., Singh, A., Lee, T. T. Y., & Li, Z. (2018). Role for fatty  
785 acid amide hydrolase (FAAH) in the leptin-mediated effects on feeding and energy balance  
786 [Neuroscience]. *Proceedings of the National Academy of Sciences*, 115(29), 7605–7610.  
787 <https://doi.org/10.1073/pnas.1802251115>

788 Bouassida, A., Chamari, K., Zaouali, M., Feki, Y., Zbidi, A., & Tabka, Z. (2010). Review on  
789 leptin and adiponectin responses and adaptations to acute and chronic exercise. *British*  
790 *Journal of Sports Medicine*, 44(9), 620–630. <https://doi.org/10.1136/bjsm.2008.046151>

791 Bouillon-Minois, J.-B., Trousselard, M., Thivel, D., Benson, A. C., Schmidt, J., Moustafa, F.,  
792 Bouvier, D., & Dutheil, F. (2021). Leptin as a Biomarker of Stress: A Systematic Review  
793 and Meta-Analysis. *Nutrients*, 13(10), 3350. <https://doi.org/10.3390/nu13103350>

794 Bowles, N. P., Karatsoreos, I. N., Li, X., Vemuri, V. K., Wood, J., Li, Z., Tamashiro, K. L. K.,  
795 Schwartz, G. J., Makriyannis, A. M., Kunos, G., Hillard, C. J., McEwen, B. S., & Hill, M.  
796 N. (2015). A peripheral endocannabinoid mechanism contributes to glucocorticoid-  
797 mediated metabolic syndrome. *Proceedings of the National Academy of Sciences of the*  
798 *United States of America*, 112(1), 285–290. <https://doi.org/10.1073/pnas.1421420112>

799 Buscemi, S., Verga, S., Caimi, G., & Cerasola, G. (2005). Low relative resting metabolic rate  
800 and body weight gain in adult Caucasian Italians. *International Journal of Obesity*, 29(3),  
801 287–291. <https://doi.org/10.1038/sj.ijo.0802888>

802 Cardinal, P., Bellocchio, L., Clark, S., Cannich, A., Klugmann, M., Lutz, B., Marsicano, G., &

803 Cota, D. (2012). Hypothalamic CB1 cannabinoid receptors regulate energy balance in mice.  
804 *Endocrinology*, 153(9), 4136–4143. <https://doi.org/10.1210/en.2012-1405>

805 Chiang, K. P., Gerber, A. L., Sipe, J. C., & Cravatt, B. F. (2004). Reduced cellular expression  
806 and activity of the P129T mutant of human fatty acid amide hydrolase: Evidence for a link  
807 between defects in the endocannabinoid system and problem drug use. *Human Molecular*  
808 *Genetics*, 13(18), 2113–2119. <https://doi.org/10.1093/hmg/ddh216>

809 Dallman, M. F., la Fleur, S. E., Pecoraro, N. C., Gomez, F., Houshyar, H., & Akana, S. F.  
810 (2004). Minireview: Glucocorticoids—Food Intake, Abdominal Obesity, and Wealthy  
811 Nations in 2004. *Endocrinology*, 145(6), 2633–2638. <https://doi.org/10.1210/en.2004-0037>

812 De Moraes Martins, C. J., Genelhu, V., Pimentel, M. M. G., Celoria, B. M. J., Mangia, R. F.,  
813 Aveta, T., Silvestri, C., Di Marzo, V., & Francischetti, E. A. (2015). Circulating  
814 endocannabinoids and the polymorphism 385C>A in fatty acid amide hydrolase (FAAH)  
815 gene may identify the obesity phenotype related to cardiometabolic risk: A study conducted  
816 in a Brazilian population of complex interethnic admixture. *PLoS ONE*, 10(11), 1–15.  
817 <https://doi.org/10.1371/journal.pone.0142728>

818 Di Marzo, V., Goparaju, S. K., Wang, L., Liu, J., B tkai, S., J rai, Z., Fezza, F., Miura, G. I.,  
819 Palmiter, R. D., Sugiura, T., & Kunos, G. (2001). Leptin-regulated endocannabinoids are  
820 involved in maintaining food intake. *Nature*, 410(6830), 822–825.  
821 <https://doi.org/10.1038/35071088>

822 Dincheva, I., Drysdale, A. T., Hartley, C. A., Johnson, D. C., Jing, D., King, E. C., Ra, S., Gray,  
823 J. M., Yang, R., DeGruccio, A. M., Huang, C., Cravatt, B. F., Glatt, C. E., Hill, M. N.,  
824 Casey, B. J., & Lee, F. S. (2015). FAAH genetic variation enhances fronto-amygdala

825 function in mouse and human. *Nature Communications*, 6, 6395.

826 <https://doi.org/10.1038/ncomms7395>

827 Dulloo, A. G., Seydoux, J., & Girardier, L. (1990). Role of corticosterone in adaptive changes in

828 energy expenditure during refeeding after low calorie intake. *American Journal of*

829 *Physiology-Endocrinology and Metabolism*, 259(5), E658–E664.

830 <https://doi.org/10.1152/ajpendo.1990.259.5.E658>

831 Durand, E., Lecoecur, C., Delplanque, J., Benzinou, M., Degraeve, F., Boutin, P., Marre, M.,

832 Balkau, B., Charpentier, G., Froguel, P., & Meyre, D. (2008). Evaluating the association of

833 FAAH common gene variation with childhood, adult severe obesity and type 2 diabetes in

834 the French population. *Obesity Facts*, 1(6), 305–309. <https://doi.org/10.1159/000178157>

835 Edwards, A., & Abizaid, A. (2016). Driving the need to feed: Insight into the collaborative

836 interaction between ghrelin and endocannabinoid systems in modulating brain reward

837 systems. *Neuroscience and Biobehavioral Reviews*, 66, 33–53.

838 <https://doi.org/10.1016/j.neubiorev.2016.03.032>

839 Egertová, M., Giang, D. K., Cravatt, B. F., & Elphick, M. R. (1998). A new perspective on

840 cannabinoid signalling: complimentary localization of fatty acid amide hydrolase and the

841 CB1 receptor in rat brain. *Proceedings of the Royal Society of London. Series B: Biological*

842 *Sciences*, 265(1410), 2081–2085. <https://doi.org/10.1098/rspb.1998.0543>

843 Flatt, J. P. (1993). Dietary Fat, Carbohydrate Balance, and Weight Maintenance. *Annals of the*

844 *New York Academy of Sciences*, 683(1 Dietary Lipid), 122–140.

845 <https://doi.org/10.1111/j.1749-6632.1993.tb35699.x>

846 Gaetani, S., Oveisi, F., & Piomelli, D. (2003). Modulation of Meal Pattern in the Rat by the

Anorexic Lipid Mediator Oleoylethanolamide. *Neuropsychopharmacology*, 28(7), 1311–1316. <https://doi.org/10.1038/sj.npp.1300166>

Gautron, L., Elmquist, J. K., & Williams, K. W. (2015). Neural Control of Energy Balance: Translating Circuits to Therapies. *Cell*, 161(1), 133–145. <https://doi.org/10.1016/j.cell.2015.02.023>

Goodarzi, M. O. (2018). Genetics of obesity: what genetic association studies have taught us about the biology of obesity and its complications. *The Lancet Diabetes & Endocrinology*, 6(3), 223–236. [https://doi.org/10.1016/S2213-8587\(17\)30200-0](https://doi.org/10.1016/S2213-8587(17)30200-0)

Gray, J. M., Vecchiarelli, H. A., Morena, M., Lee, T. T. Y., Hermanson, D. J., Kim, A. B., McLaughlin, R. J., Hassan, K. I., Kühne, C., Wotjak, C. T., Deussing, J. M., Patel, S., & Hill, M. N. (2015). Corticotropin-releasing hormone drives anandamide hydrolysis in the amygdala to promote anxiety. *The Journal of Neuroscience : The Official Journal of the Society for Neuroscience*, 35(9), 3879–3892. <https://doi.org/10.1523/JNEUROSCI.2737-14.2015>

Gulyas, A. I., Cravatt, B. F., Bracey, M. H., Dinh, T. P., Piomelli, D., Boscia, F., & Freund, T. F. (2004). Segregation of two endocannabinoid-hydrolyzing enzymes into pre- and postsynaptic compartments in the rat hippocampus, cerebellum and amygdala. *The European Journal of Neuroscience*, 20(2), 441–458. <https://doi.org/10.1111/j.1460-9568.2004.03428.x>

Hardwick, A. J., Linton, E. A., & Rothwell, N. J. (1989). Thermogenic Effects of the Antigluocorticoid RU-486 in the Rat: Involvement of Corticotropin-Releasing Factor and Sympathetic Activation of Brown Adipose Tissue\*. *Endocrinology*, 124(4), 1684–1688.

869 <https://doi.org/10.1210/endo-124-4-1684>

870 Hill, M. N., McLaughlin, R. J., Morrish, A. C., Viau, V., Floresco, S. B., Hillard, C. J., &  
871 Gorzalka, B. B. (2009). Suppression of Amygdalar Endocannabinoid Signaling by Stress  
872 Contributes to Activation of the Hypothalamic-Pituitary- Adrenal Axis HHS Public Access.  
873 *Neuropsychopharmacology*, 34(13), 2733–2745. <https://doi.org/10.1038/npp.2009.114>

874 Hunker, A. C., Soden, M. E., Krayushkina, D., Heymann, G., Awatramani, R., & Zweifel, L. S.  
875 (2020). Conditional Single Vector CRISPR/SaCas9 Viruses for Efficient Mutagenesis in the  
876 Adult Mouse Nervous System. *Cell Reports*, 30(12), 4303-4316.e6.  
877 <https://doi.org/10.1016/j.celrep.2020.02.092>

878 Jensen, D. P., Andreasen, C. H., Andersen, M. K., Hansen, L., Eiberg, H., Borch-Johnsen, K.,  
879 Jørgensen, T., Hansen, T., & Pedersen, O. (2007). The functional Pro129Thr variant of the  
880 FAAH gene is not associated with various fat accumulation phenotypes in a population-  
881 based cohort of 5,801 whites. *Journal of Molecular Medicine*, 85(5), 445–449.  
882 <https://doi.org/10.1007/s00109-006-0139-0>

883 Jo, Y. H., Chen, Y. J. J., Chua, S. C., Talmage, D. A., & Role, L. W. (2005). Integration of  
884 endocannabinoid and leptin signaling in an appetite-related neural circuit. *Neuron*, 48(6),  
885 1055–1066. <https://doi.org/10.1016/j.neuron.2005.10.021>

886 Karatsoreos, I. N., Bhagat, S. M., Bowles, N. P., Weil, Z. M., Pfaff, D. W., & McEwen, B. S.  
887 (2010). Endocrine and physiological changes in response to chronic corticosterone: A  
888 potential model of the metabolic syndrome in mouse. *Endocrinology*, 151(5), 2117–2127.  
889 <https://doi.org/10.1210/en.2009-1436>

890 Kim, E.-K., Miller, I., Aja, S., Landree, L. E., Pinn, M., McFadden, J., Kuhajda, F. P., Moran, T.

H., & Ronnett, G. V. (2004). C75, a Fatty Acid Synthase Inhibitor, Reduces Food Intake via Hypothalamic AMP-activated Protein Kinase. *Journal of Biological Chemistry*, 279(19), 19970–19976. <https://doi.org/10.1074/jbc.M402165200>

Kola, B., Farkas, I., Christ-Crain, M., Wittmann, G., Lolli, F., Amin, F., Harvey-White, J., Liposits, Z., Kunos, G., Grossman, A. B., Fekete, C., & Korbonits, M. (2008). The orexigenic effect of ghrelin is mediated through central activation of the endogenous cannabinoid system. *PLoS ONE*, 3(3). <https://doi.org/10.1371/journal.pone.0001797>

Krashes, M. J., Koda, S., Ye, C., Rogan, S. C., Adams, A. C., Cusher, D. S., Maratos-Flier, E., Roth, B. L., & Lowell, B. B. (2011). Rapid, reversible activation of AgRP neurons drives feeding behavior in mice. *Journal of Clinical Investigation*, 121(4), 1424–1428. <https://doi.org/10.1172/JCI46229>

Kunz, I., Schorr, U., Klaus, S., & Sharma, A. M. (2000). Resting metabolic rate and substrate use in obesity hypertension. *Hypertension (Dallas, Tex. : 1979)*, 36(1), 26–32. <https://doi.org/10.1161/01.hyp.36.1.26>

Lazzer, S., Bedogni, G., Lafortuna, C. L., Marazzi, N., Busti, C., Galli, R., de Col, A., Agosti, F., & Sartorio, A. (2010). Relationship Between Basal Metabolic Rate, Gender, Age, and Body Composition in 8,780 White Obese Subjects. *Obesity*, 18(1), 71–78. <https://doi.org/10.1038/oby.2009.162>

Lieb, W., Manning, A. K., Florez, J. C., Dupuis, J., Cupples, L. A., McAteer, J. B., Vasan, R. S., Hoffmann, U., O'Donnell, C. J., Meigs, J. B., & Fox, C. S. (2009). Variants in the CNR1 and the FAAH Genes and Adiposity Traits in the Community. *Obesity*, 17(4), 755–760. <https://doi.org/10.1038/oby.2008.608>



913 Lu, X. Y., Kim, C. S., Frazer, A., & Zhang, W. (2006). Leptin: a potential novel antidepressant.  
914 *Proc.Natl.Acad.Sci.U.S.A*, 103(0027-8424 (Print)), 1593–1598.  
915 <https://doi.org/10.1073/pnas.0508901103>

916 Maccarrone, M., Bari, M., & Agrò, A. F. (1999). A Sensitive and Specific  
917 Radiochromatographic Assay of Fatty Acid Amide Hydrolase Activity. *Analytical*  
918 *Biochemistry*, 267(2), 314–318. <https://doi.org/10.1006/abio.1998.2964>

919 Mazier, W., Saucisse, N., Gatta-Cherifi, B., & Cota, D. (2015). The Endocannabinoid System:  
920 Pivotal Orchestrator of Obesity and Metabolic Disease. *Trends in Endocrinology and*  
921 *Metabolism*, 26(10), 524–537. <https://doi.org/10.1016/j.tem.2015.07.007>

922 Minokoshi, Y., Alquier, T., Furukawa, H., Kim, Y. B., Lee, A., Xue, B., Mu, J., Foufelle, F.,  
923 Ferré, P., Birnbaum, M. J., Stuck, B. J., & Kahn, B. B. (2004). AMP-kinase regulates food  
924 intake by responding to hormonal and nutrient signals in the hypothalamus. *Nature*,  
925 428(6982), 569–574. <https://doi.org/10.1038/nature02440>

926 Monteleone, P., Tortorella, A., Martiadis, V., Di Filippo, C., Canestrelli, B., & Maj, M. (2008).  
927 The cDNA 385C to A missense polymorphism of the endocannabinoid degrading enzyme  
928 fatty acid amide hydrolase (FAAH) is associated with overweight/obesity but not with binge  
929 eating disorder in overweight/obese women. *Psychoneuroendocrinology*, 33(4), 546–550.  
930 <https://doi.org/10.1016/j.psyneuen.2008.01.004>

931 Osei-Hyiaman, D., Liu, J., Zhou, L., Godlewski, G., Harvey-White, J., Jeong, W., Bátkai, S.,  
932 Marsicano, G., Lutz, B., Buettner, C., & Kunos, G. (2008). Hepatic CB1 receptor is  
933 required for development of diet-induced steatosis, dyslipidemia, and insulin and leptin  
934 resistance in mice. *The Journal of Clinical Investigation*, 118(9), 3160–3169.

<https://doi.org/10.1172/JCI34827>

Pagotto, U., Cervino, C., Vicennati, V., Marsicano, G., Lutz, B., & Pasquali, R. (2006). How many sites of action for endocannabinoids to control energy metabolism? *International Journal of Obesity*, 30(S1), S39–S43. <https://doi.org/10.1038/sj.ijo.0803277>

Patel, S., Carrier, E. J., Ho, W.-S. V., Rademacher, D. J., Cunningham, S., Reddy, D. S., Falck, J. R., Cravatt, B. F., & Hillard, C. J. (2005). The postmortal accumulation of brain N-arachidonylethanolamine (anandamide) is dependent upon fatty acid amide hydrolase activity. *Journal of Lipid Research*, 46(2), 342–349. <https://doi.org/10.1194/jlr.M400377-JLR200>

Pecoraro, N., Reyes, F., Gomez, F., Bhargava, A., & Dallman, M. F. (2004). Chronic stress promotes palatable feeding, which reduces signs of stress: feedforward and feedback effects of chronic stress. *Endocrinology*, 145(0013-7227 (Print)), 3754–3762. <https://doi.org/10.1210/en.2004-0305>

Perry, R. J., Resch, J. M., Douglass, A. M., Madara, J. C., Rabin-Court, A., Kucukdereli, H., Wu, C., Song, J. D., Lowell, B. B., & Shulman, G. I. (2019). Leptin’s hunger-suppressing effects are mediated by the hypothalamic–pituitary–adrenocortical axis in rodents. *Proceedings of the National Academy of Sciences of the United States of America*, 116(27), 13670–13679. <https://doi.org/10.1073/pnas.1901795116>

Pezeshki, A., Fahim, A., & Chelikani, P. K. (2015). Dietary Whey and Casein Differentially Affect Energy Balance, Gut Hormones, Glucose Metabolism, and Taste Preference in Diet-Induced Obese Rats. *The Journal of Nutrition*, 145(10), 2236–2244. <https://doi.org/10.3945/jn.115.213843>

957 Poggioli, R., Ueta, C. B., e Drigo, R. A., Castillo, M., Fonseca, T. L., & Bianco, A. C. (2013).  
958 Dexamethasone reduces energy expenditure and increases susceptibility to diet-induced  
959 obesity in mice. *Obesity*, n/a-n/a. <https://doi.org/10.1002/oby.20338>

960 Ramage, L. E., Akyol, M., Fletcher, A. M., Forsythe, J., Nixon, M., Carter, R. N., van Beek, E. J.  
961 R., Morton, N. M., Walker, B. R., & Stimson, R. H. (2016). Glucocorticoids Acutely  
962 Increase Brown Adipose Tissue Activity in Humans, Revealing Species-Specific  
963 Differences in UCP-1 Regulation. *Cell Metabolism*, 24(1), 130–141.  
964 <https://doi.org/10.1016/j.cmet.2016.06.011>

965 Ruiz de Azua, I., & Lutz, B. (2019). Multiple endocannabinoid-mediated mechanisms in the  
966 regulation of energy homeostasis in brain and peripheral tissues. *Cellular and Molecular*  
967 *Life Sciences*, 76(7), 1341–1363. <https://doi.org/10.1007/s00018-018-2994-6>

968 Ruiz de Azua, I., Mancini, G., Srivastava, R. K., Rey, A. A., Cardinal, P., Tedesco, L.,  
969 Zingaretti, C. M., Sassmann, A., Quarta, C., Schwitter, C., Conrad, A., Wettschureck, N.,  
970 Vemuri, V. K., Makriyannis, A., Hartwig, J., Mendez-Lago, M., Bindila, L., Monory, K.,  
971 Giordano, A., ... Lutz, B. (2017). Adipocyte cannabinoid receptor CB1 regulates energy  
972 homeostasis and alternatively activated macrophages. *Journal of Clinical Investigation*, 1–  
973 15. <https://doi.org/10.1172/JCI83626>

974 Sadowska, J., Gębczyński, A. K., & Konarzewski, M. (2017). Metabolic risk factors in mice  
975 divergently selected for BMR fed high fat and high carb diets. *PLOS ONE*, 12(2),  
976 e0172892. <https://doi.org/10.1371/journal.pone.0172892>

977 Scerif, M., Füzesi, T., Thomas, J. D., Kola, B., Grossman, A. B., Fekete, C., & Korbonits, M.  
978 (2013). CB1 receptor mediates the effects of glucocorticoids on AMPK activity in the

979 hypothalamus. *Journal of Endocrinology*, 219(1), 79–88. <https://doi.org/10.1530/JOE-13->  
980 0192

981 Schwarz, N. A., Rigby, B. R., La Bounty, P., Shelmadine, B., & Bowden, R. G. (2011). A  
982 Review of Weight Control Strategies and Their Effects on the Regulation of Hormonal  
983 Balance. *Journal of Nutrition and Metabolism*, 2011, 1–15.  
984 <https://doi.org/10.1155/2011/237932>

985 Sipe, J C, Waalen, J., Gerber, A., & Beutler, E. (2005). Overweight and obesity associated with a  
986 missense polymorphism in fatty acid amide hydrolase (FAAH). *International Journal of*  
987 *Obesity* (2005), 29(7), 755–759. <https://doi.org/10.1038/sj.ijo.0802954>

988 Sipe, J C, Chiang, K., Gerber, A. L., Beutler, E., & Cravatt, B. F. (2002). A missense mutation in  
989 human fatty acid amide hydrolase associated with problem drug use. *Proceedings of the*  
990 *National Academy of Sciences of the United States of America*, 99(12), 8394–8399.  
991 <https://doi.org/10.1073/pnas.082235799>

992 Strack, A. M., Bradbury, M. J., & Dallman, M. F. (1995). Corticosterone decreases nonshivering  
993 thermogenesis and increases lipid storage in brown adipose tissue. *American Journal of*  
994 *Physiology-Regulatory, Integrative and Comparative Physiology*, 268(1), R183–R191.  
995 <https://doi.org/10.1152/ajpregu.1995.268.1.R183>

996 Tschöp, M. H., Speakman, J. R., Arch, J. R. S., Auwerx, J., Brüning, J. C., Chan, L., Eckel, R.  
997 H., Farese, R. V, Galgani, J. E., Hambly, C., Herman, M. a, Horvath, T. L., Kahn, B. B.,  
998 Kozma, S. C., Maratos-Flier, E., Müller, T. D., Münzberg, H., Pfluger, P. T., Plum, L., ...  
999 Ravussin, E. (2011). A guide to analysis of mouse energy metabolism. *Nature Methods*,  
1000 9(1), 57–63. <https://doi.org/10.1038/nmeth.1806>

1001 Tsien, J. Z., Chen, D. F., Gerber, D., Tom, C., Mercer, E. H., Anderson, D. J., Mayford, M.,  
1002 Kandel, E. R., & Tonegawa, S. (1996). Subregion- and Cell Type-Restricted Gene  
1003 Knockout in Mouse Brain. *Cell*, 87(7), 1317–1326. <https://doi.org/10.1016/S0092->  
1004 8674(00)81826-7

1005 Tsou, K., Nogueron, M. I., Muthian, S., Sañudo-Peña, M. C., Hillard, C. J., Deutsch, D. G., &  
1006 Walker, J. M. (1998). Fatty acid amide hydrolase is located preferentially in large neurons  
1007 in the rat central nervous system as revealed by immunohistochemistry. *Neuroscience*  
1008 *Letters*, 254(3), 137–140. [https://doi.org/10.1016/S0304-3940\(98\)00700-9](https://doi.org/10.1016/S0304-3940(98)00700-9)

1009 Tucci, S. A., Rogers, E. K., Korbonits, M., & Kirkham, T. C. (2004). The cannabinoid CB1  
1010 receptor antagonist SR141716 blocks the orexigenic effects of intrahypothalamic ghrelin.  
1011 *British Journal of Pharmacology*, 143(5), 520–523. <https://doi.org/10.1038/sj.bjp.0705968>

1012 Ulrich-Lai, Y. M. (2016). Self-medication with sucrose. *Current Opinion in Behavioral*  
1013 *Sciences*, 9, 78–83. <https://doi.org/10.1016/j.cobeha.2016.02.015>

1014 Ulrich-Lai, Y. M., & Ryan, K. K. (2014). Neuroendocrine circuits governing energy balance and  
1015 stress regulation: Functional overlap and therapeutic implications. *Cell Metabolism*, 19(6),  
1016 910–925. <https://doi.org/10.1016/j.cmet.2014.01.020>

1017 Van Den Beukel, J. C., Grefhorst, A., Quarta, C., Steenbergen, J., Mastroberardino, P. G.,  
1018 Lombés, M., Delhanty, P. J., Mazza, R., Pagotto, U., Van Der Lely, A. J., & Themmen, A.  
1019 P. N. (2014). Direct activating effects of adrenocorticotrophic hormone (ACTH) on brown  
1020 adipose tissue are attenuated by corticosterone. *FASEB Journal*, 28(11), 4857–4867.  
1021 <https://doi.org/10.1096/fj.14-254839>

1022 Westerterp, K. R. (2017). Control of energy expenditure in humans. *European Journal of*

- 1023        *Clinical Nutrition*, 71(3), 340–344. <https://doi.org/10.1038/ejcn.2016.237>
- 1024    World Health Organisation. (2020). *Obesity and Overweight*. [https://www.who.int/news-](https://www.who.int/news-room/fact-sheets/detail/obesity-and-overweight)
- 1025        [room/fact-sheets/detail/obesity-and-overweight](https://www.who.int/news-room/fact-sheets/detail/obesity-and-overweight)
- 1026    Zhang, Y., Sonnenberg, G. E., Baye, T. M., Littrell, J., Gunnell, J., DeLaForest, A., MacKinney,
- 1027        E., Hillard, C. J., Kissebah, A. H., Olivier, M., & Wilke, R. A. (2009). Obesity-related
- 1028        dyslipidemia associated with FAAH, independent of insulin response, in multigenerational
- 1029        families of Northern European descent. *Pharmacogenomics*, 10(12), 1929–1939.
- 1030        <https://doi.org/10.2217/pgs.09.122>
- 1031

## 1032 **Acknowledgements**

1033 The authors thank Dr. Frank Visser for his genotyping support. We acknowledge Dr. Pamela  
1034 Mellon for donating the GT1-7 cells. We thank the animal care staff at the University of Calgary  
1035 for providing technical support.

## 1036 **Funding**

1037 This work was supported by funding from the Natural Sciences and Engineering Research  
1038 Council of Canada to MNH; American Heart Association Grant # 953881 to PKC. GB received  
1039 salary support from a Banting Postdoctoral Fellowship provided by the Canadian Institutes of  
1040 Health Research (CIHR) and an Alberta Innovates Scholarship.

## 1041 **Author Contributions**

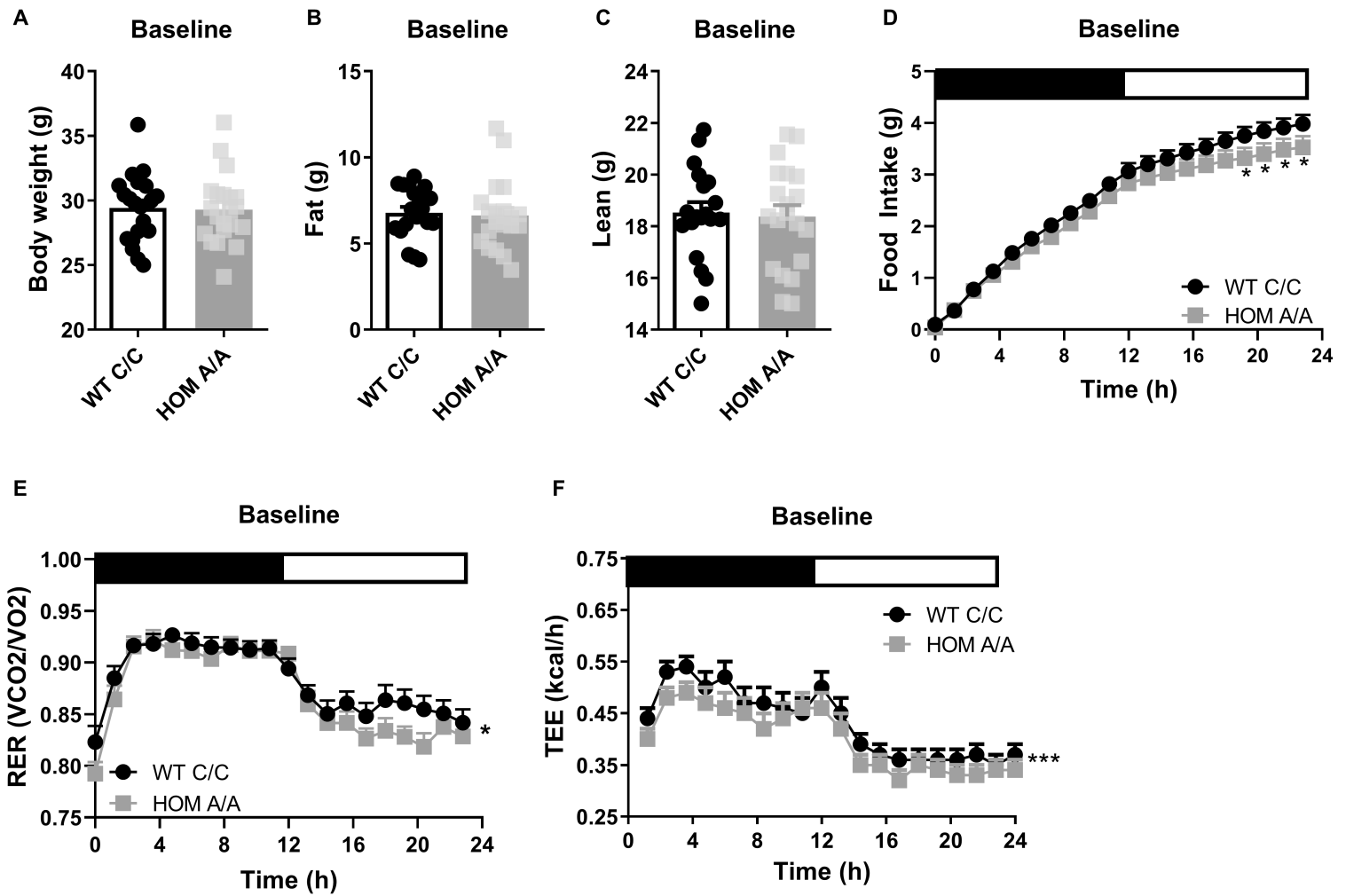
1042 G.B. and M.N.H. conceived the project and designed the experiments. G.B. managed the mouse  
1043 lines and genotyping. G.B., G.N.P., A.S., R.A., H.A.V., H.Y. and M.S. performed animal studies  
1044 and ran enzyme/hormone/endocannabinoid assays. D.E.H. and N.C.G. performed and analyzed  
1045 protein experiments. A.C.H. and L.S.Z. designed and contributed CRISPR/Cas9 vectors. R.J.T.,  
1046 P.K.C., and F.S.L. contributed analytical/genetic tools. G.B. and M.N.H. analyzed the data. G.B.  
1047 wrote the initial version of the manuscript and all authors revised the manuscript.

## 1048 **Competing Interests**

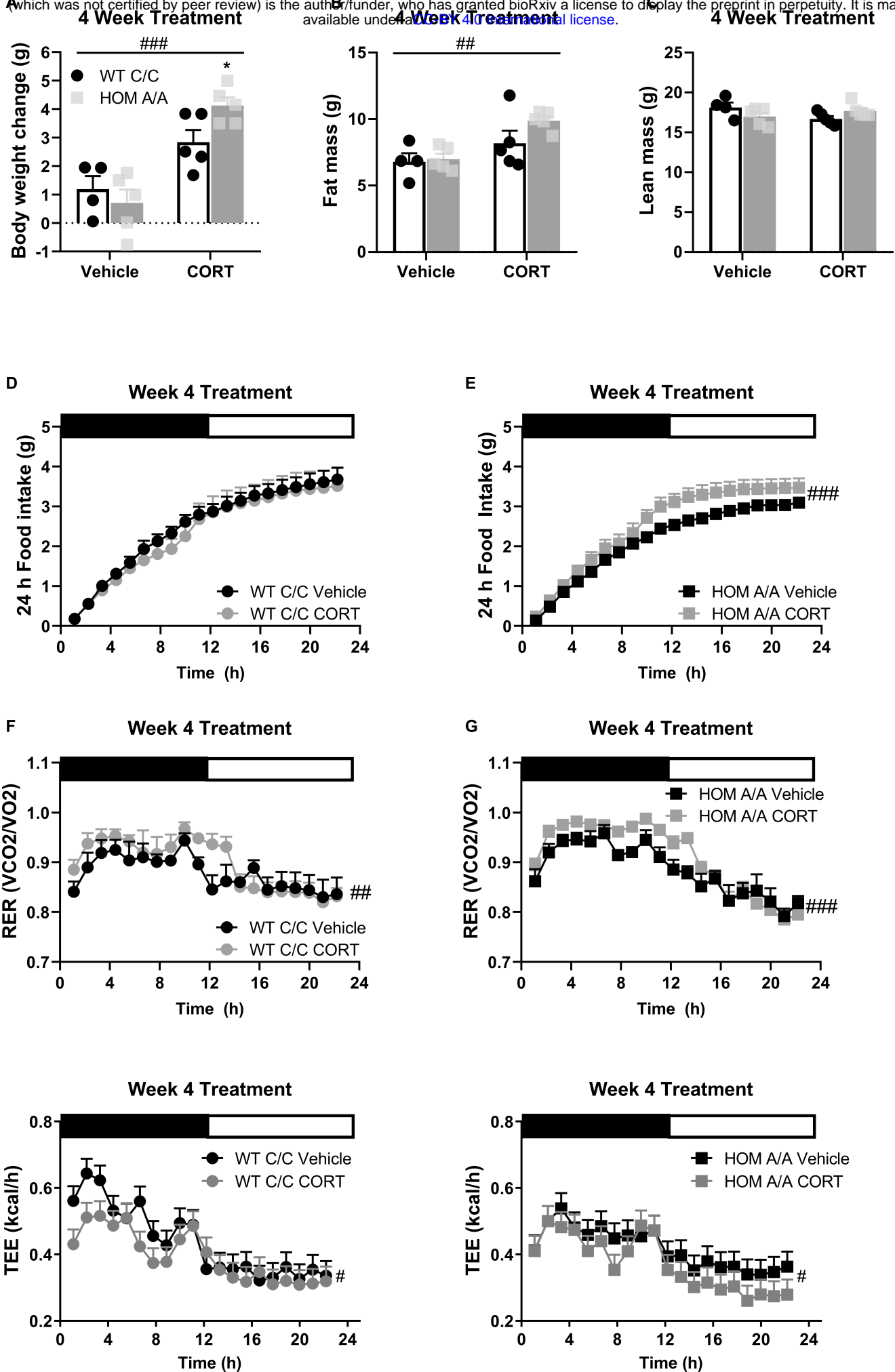
1049 The authors declare no conflict of interest.

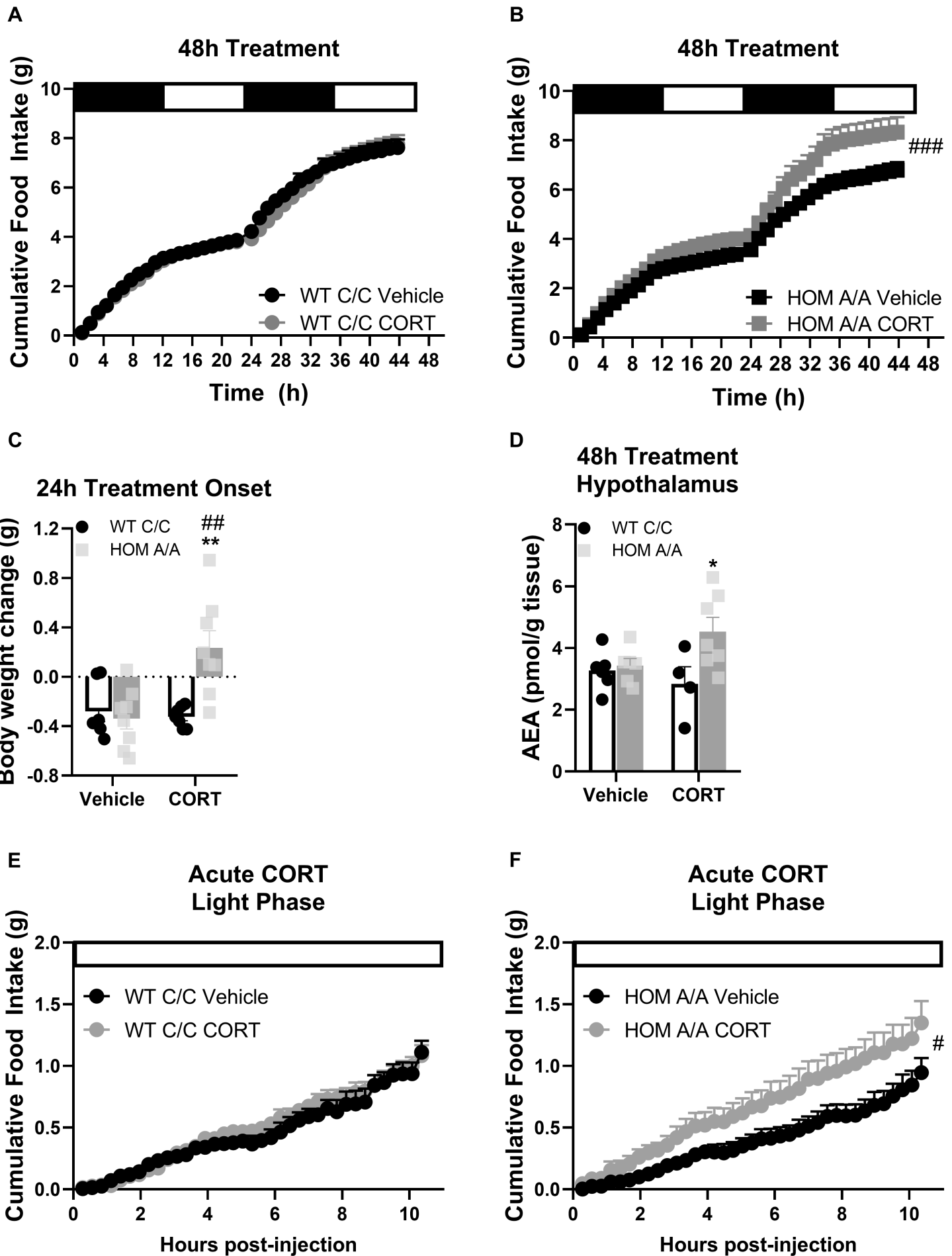
## 1050 **Data and Materials Availability**

1051 All data needed to evaluate the conclusions in the paper are present in the paper and/or the  
1052 Supplementary Materials including the Source Data supplementary file.



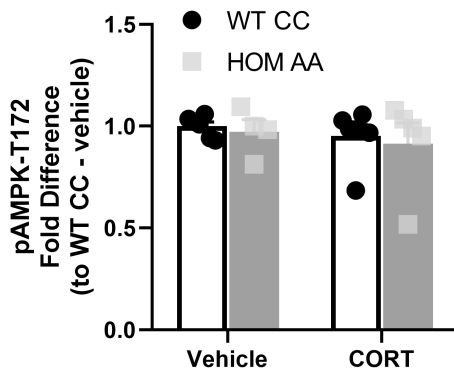






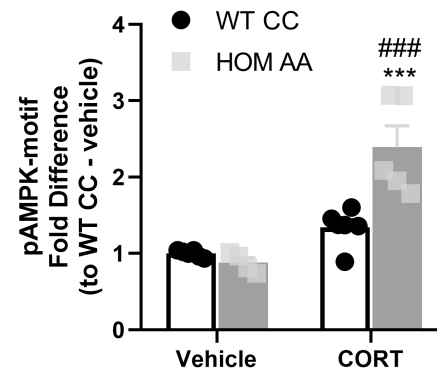
## Hypothalamus

### Activation of AMPK



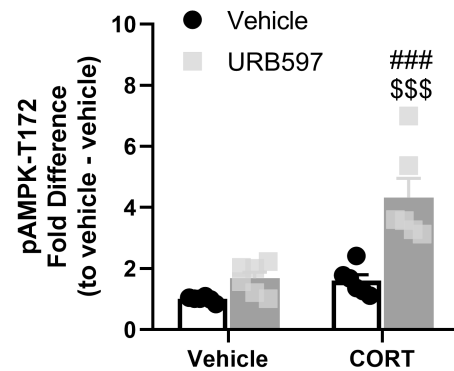
## Hypothalamus

### Activation of AMPK Direct Targets



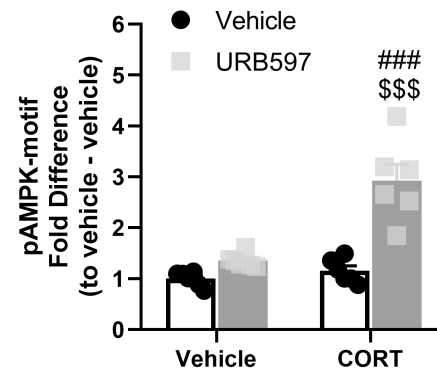
## GT1-7 cells

### Activation of AMPK

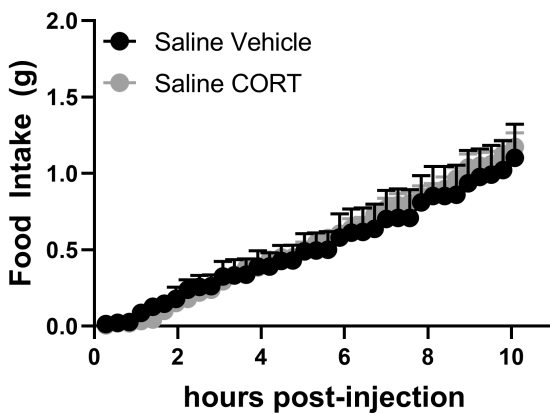


## GT1-7 cells

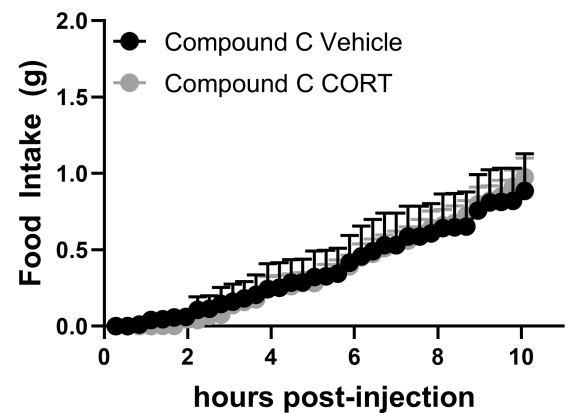
### Activation of AMPK Direct Targets



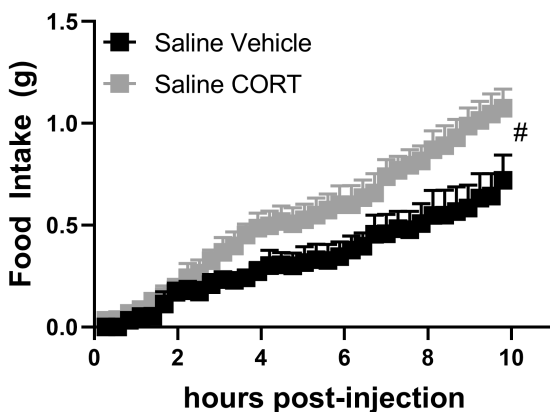
## WT C/C



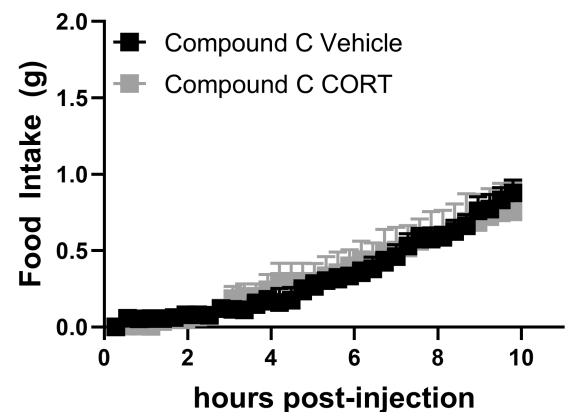
## WT C/C

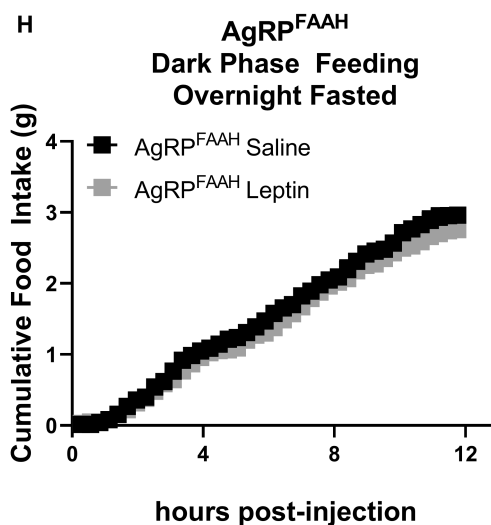
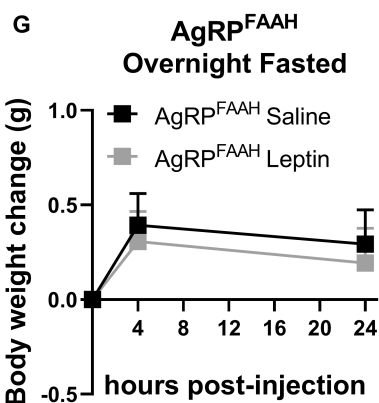
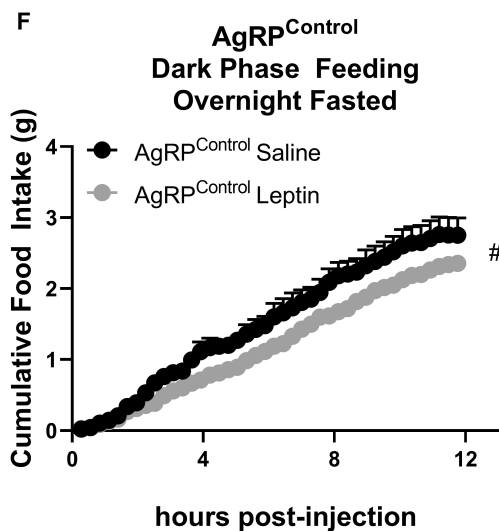
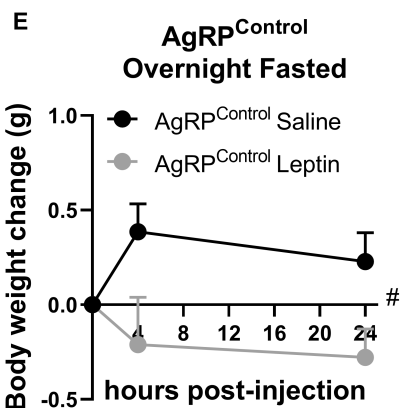
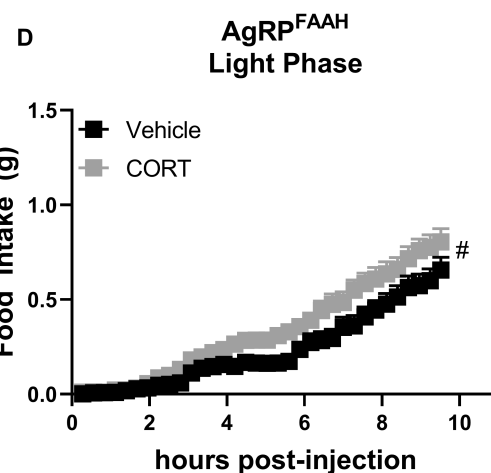
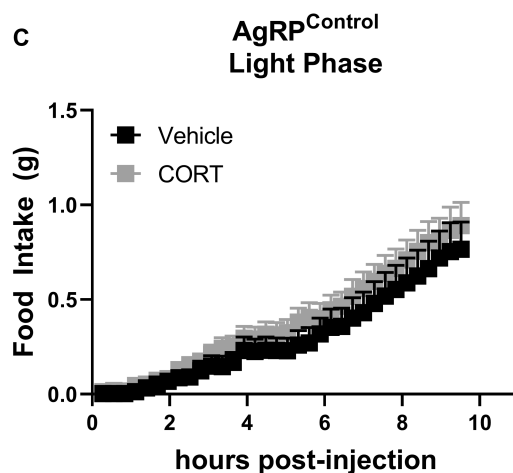
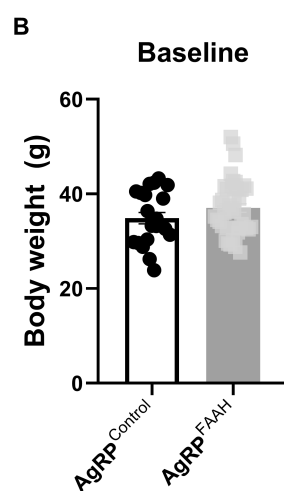
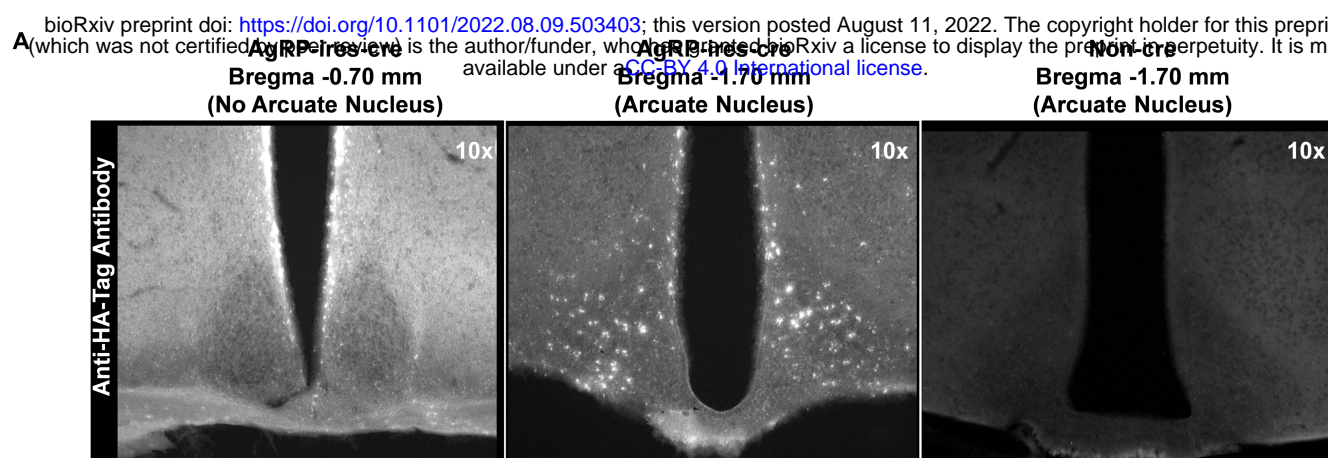


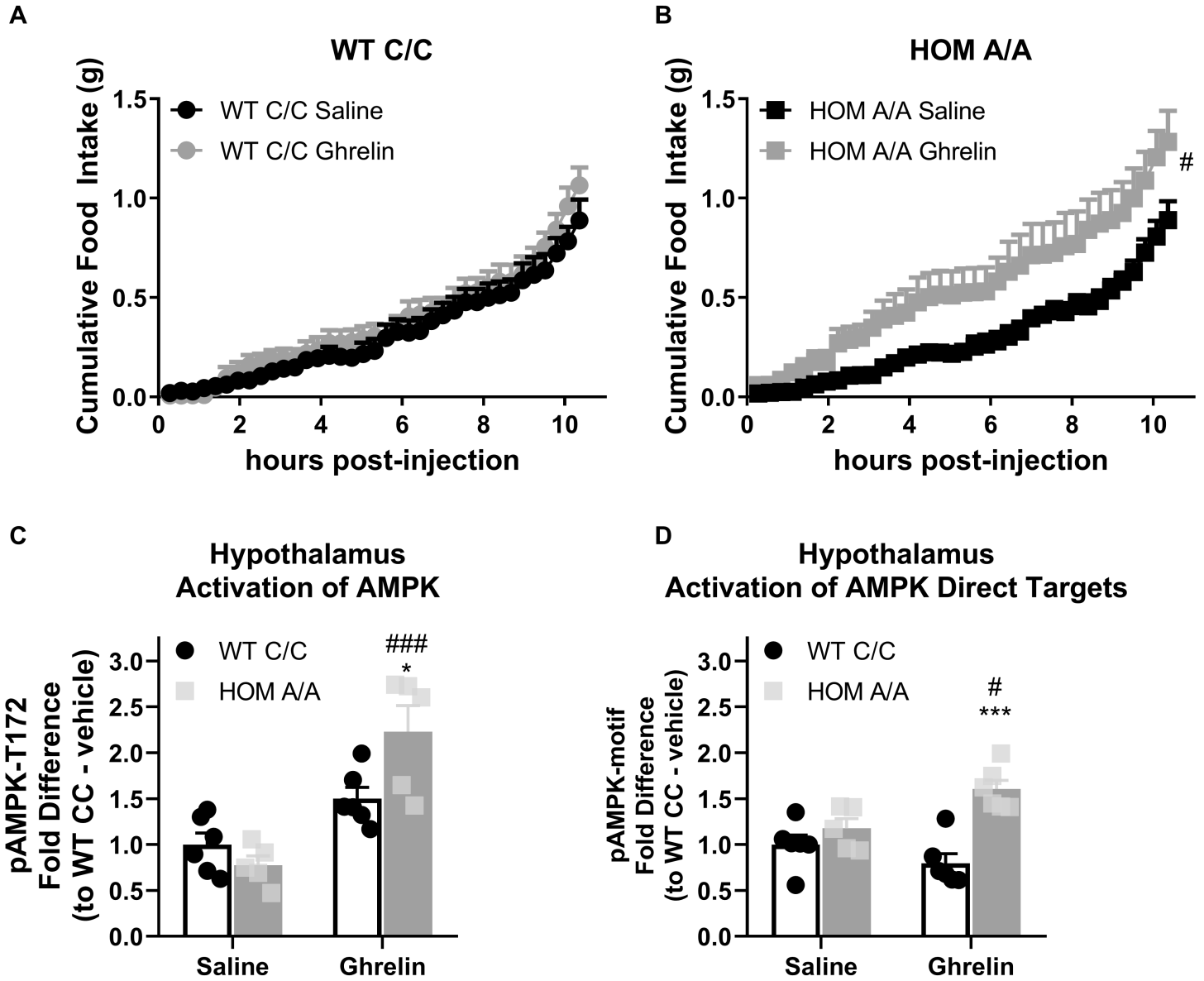
## HOM A/A



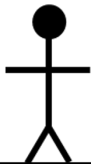
## HOM A/A







**FAAH C/C**



Anorexigenic  
Leptin

Orexigenic  
Glucocorticoids  
Ghrelin

**FAAH A/A**



Anorexigenic  
Leptin

Orexigenic  
Glucocorticoids  
Ghrelin

