

# Incubation of palatable food craving is associated with brain-wide neuronal activation in mice

Rajtarun Madangopal<sup>1†</sup>, Eric R. Szelenyi<sup>3,4†</sup>, Joseph Nguyen<sup>1</sup>, Megan B. Brenner<sup>1</sup>, Olivia R. Drake<sup>1</sup>, Diana Pham<sup>1</sup>, Aniruddha Shekara<sup>1</sup>, Michelle Jin<sup>1</sup>, Jia Jie Choong<sup>2,3,4</sup>, Connor Heins<sup>1</sup>, Lauren E. Komer<sup>1</sup>, Sophia J. Weber<sup>1</sup>, Bruce T. Hope<sup>1</sup>, Yavin Shaham<sup>1</sup>, Sam A. Golden<sup>3,4,\*</sup>

<sup>1</sup> Intramural Research Program, National Institute on Drug Abuse, Baltimore, MD, USA.

<sup>2</sup> University of Washington, Department of Electrical and Computer Engineering, Seattle, Washington, USA

<sup>3</sup> Department of Biological Structure, University of Washington, Seattle, WA, USA.

<sup>4</sup> University of Washington, Center of Excellence in Neurobiology of Addiction, Pain, and Emotion (NAPE), Seattle, Washington, USA

† These authors contributed equally

**\*Correspondence to:** Sam A. Golden, Ph.D., Department of Biological Structure, University of Washington, Seattle, WA 98195

**Email:** sagolden@uw.edu

**Author Contributions:** RM, ES, YS, BTH and SAG designed the experiments; RM, MJ, CH, LEK, SJW, and SAG ran the behavioral experiments and collected data; RM, MJ, CH, LEK, SJW, MBB, ORD, AS, and DP analyzed behavioral data; RM, MBB, and SAG performed whole-brain immunohistochemistry and clearing; RM, ES, and SAG optimized imaging protocols, RM, ES, MJ, JN, JJC, and SAG established analysis pipelines; RM, ES, MBB, ORD, AS, DP and JJC analyzed data; RM, ES, YS, BTH and SAG wrote the paper. All authors reviewed and approved the final version prior to submission.

**Competing Interest Statement:** The authors declare that they do not have any conflicts of interest (financial or otherwise) related to the text of the paper. The research was supported by the NIDA Intramural Research Program funds to the labs of Yavin Shaham and Bruce Hope. SAG received funding from NIH PRAT 1FI2GM117583-01, NIDA R00DA045662, NIDA R01DA054317, NIDA 1UG3DA053802, NIDA P30DA048736 and NARSAD Young Investigator Award 27082. RM received funding from the NIH Center for Compulsive Behaviors. ES received funding from the Washington Research Foundation Fellowship Program. ORD and DP were supported by the NIDA IRP Scientific Director's Fellowship for Diversity in Research.

**Classification:** Biological Sciences

**Keywords:** addiction, incubation, whole-brain analysis, mice, Fos

1    **Abstract (245 words)**

2        Studies using rodent models have shown that relapse to drug or food seeking increases  
3        progressively during abstinence, a phenomenon termed 'incubation of craving'. Mechanistic  
4        studies of incubation of craving have focused on specific neurobiological targets within pre-  
5        selected brain areas. Recent methodological advances in whole-brain immunohistochemistry,  
6        clearing, and imaging now enable unbiased brain-wide cellular resolution mapping of regions and  
7        circuits engaged during learned behaviors. However, these whole brain imaging approaches were  
8        developed for mouse brains while incubation of drug craving has primarily been studied in rats  
9        and incubation of food craving has not been demonstrated in mice. Here, we established a  
10       mouse model of incubation of palatable food craving and examined food reward seeking after 1,  
11       15, and 60 abstinence days. We then used the neuronal activity marker Fos with intact brain  
12       mapping procedures to identify corresponding patterns of brain-wide activation. Relapse to food  
13       seeking was significantly higher after 60 abstinence days than after 1 or 15 days. Using unbiased  
14       ClearMap analysis, we identified increased activation of multiple brain regions, particularly  
15       corticostriatal structures, following 60, but not 15 abstinence days. We used orthogonal SMART2  
16       analysis to confirm these findings within corticostriatal and thalamocortical subvolumes and  
17       applied expert-guided registration to investigate subdivision and layer-specific activation patterns.  
18       Overall, we (1) identified novel brain-wide activity patterns during incubation of food seeking using  
19       complementary analytical approaches, and (2) provide a single-cell resolution whole-brain atlas  
20       that can be used to identify functional networks and global architecture underlying incubation of  
21       food craving.

22

23    **Significance Statement**

24        Relapse to reward seeking progressively increases during abstinence, a phenomenon  
25        termed incubation of craving. Mechanistic studies of incubation can lead to novel relapse  
26        treatments. However, previous studies have primarily used rat models and targeted region-by-

27 region analyses and a brain-wide functional atlas of incubation of reward seeking is lacking. We  
28 established a behavioral procedure for incubation of palatable food seeking in mice and applied  
29 whole-brain activity mapping with Fos as a neuronal activity marker to identify the functional  
30 connectome of this incubation. Like rats, mice showed incubation of food seeking during  
31 abstinence. Using two complementary activity mapping approaches, we identified a brain-wide  
32 pattern of increased neural activation that mirrored incubation of food seeking after 60, but not 15,  
33 days of abstinence.

34

### 35 **Main Text**

#### 36 **Introduction**

37 Studies using rodent models have shown that non-reinforced drug or food seeking  
38 progressively increases during abstinence in the home cage (1-3). This phenomenon, termed  
39 '*incubation of craving*', was first identified in rats after cocaine self-administration (4-7), and has  
40 since been demonstrated for other drugs such as heroin (8), methamphetamine (9), alcohol (10),  
41 and nicotine (11), as well as for non-drug rewards such as sucrose (12, 13), standard chow  
42 pellets (14), and high-carbohydrate pellets (15). These preclinical findings mirror reports of  
43 incubation of cue-induced drug craving and physiological responses in human drug-users (16-18)  
44 and could provide avenues to identify cellular and molecular mechanisms underlying persistent  
45 relapse vulnerability to both unhealthy palatable food (3, 19) and addictive drugs (1, 2, 20-22).

46 Immediate early gene (IEG, eg. Fos, Arc, Zif) expression serves as a proxy for strongly  
47 activated neurons, and quantification of Fos-positive (Fos+) cells is routinely used to identify  
48 changes in neural activation patterns after exposure to different unconditioned and conditioned  
49 stimuli (23-27). Previous activity-mapping studies of incubation of drug and food craving have  
50 identified several brain regions ([Supplementary Table S1](#)) relevant to (1) relapse (increased Fos  
51 expression during relapse tests vs. home cage controls), and/or (2) incubation (higher Fos  
52 expression during late abstinence test vs. early abstinence test) (1, 3, 5, 28, 29). These studies  
53 used targeted one-by-one regional quantification of Fos+ cell counts in thin sectioned tissue

54 samples and focused on changes in specific pre-determined brain areas. Thus, it is currently  
55 unknown whether brain-wide activity patterns, including multi-regional patterns, are altered during  
56 incubation of reward seeking during abstinence.

57 To address this knowledge gap, we leveraged recent developments in brain-wide activity  
58 mapping approaches, including whole mouse brain immunofluorescent staining and clearing (30-  
59 35), light sheet fluorescence microscopy (LSFM) (36, 37), and open-source analysis tools (38-41)  
60 to investigate changes in brain-wide activation patterns during incubation of palatable food  
61 seeking in mice.

62 We trained food-sated CD-1 male mice to self-administer palatable high-carbohydrate food  
63 pellets (42, 43) for 7 days and then tested them for relapse to food seeking after 1, 15, or 60  
64 abstinence days. We perfused and extracted their brains 90 min after the relapse tests (or directly  
65 from homecage as a baseline activity control), labeled 'active' Fos+ nuclei across intact mouse  
66 brains using an optimized iDISCO+ Fos immunofluorescent staining protocol (41), and imaged  
67 Fos immunofluorescence at single cell resolution using LSFM. We used the ClearMap pipeline  
68 (39, 40) for unbiased mapping of "incubation-associated" neural activation patterns across the  
69 entire anterior-posterior axis of the mouse brain. We also updated the SMART analysis package  
70 (41) to conduct targeted analysis of neural activation patterns within LSFM coronal subvolumes  
71 and used SMART2 to cross-validate and extend our ClearMap findings within a subset of  
72 corticostriatal and thalamocortical brain regions and subdivisions.

73

#### 74 **Materials and Methods**

#### 75 **Subjects**

76 We used male (n = 60) 4-6-month-old sexually experienced CD-1 mice (Charles River Lab,  
77 CRL), weighing ~40 g prior to food self-administration training. We confirmed with CRL animal-  
78 facility staff that all sexually experienced CD-1 males had equal access to receptive females. Per  
79 CRL's procedure male mice are pair-housed with several females from PD28 until purchase.  
80 Pregnant females are switched with new non-pregnant females, with no break between cycles

81 and male mice that do not successfully breed are removed from the breeding pool and not made  
82 available for purchase. We excluded 14 mice due to failure to acquire food self-administration.  
83 The mice had free access to food and water in the homecage and were maintained on a reverse  
84 12:12 h light-dark cycle (light off at 8 am). We only used male mice because the behavioral study  
85 was performed before the implementation of the NIH Sex as a Biological Variable Guideline.

86 **Apparatus**

87 We trained and tested all mice in standard Med Associates operant chambers, enclosed in  
88 ventilated sound-attenuating cubicles. Each chamber was equipped with a stainless-steel grid  
89 floor and two side-walls, each with three modular operant panels. We used a houselight located  
90 on one side of the chamber to illuminate the chamber during the training and test sessions. Two  
91 levers served as operant manipulanda – (1) a non-retractable lever on the same side as the  
92 houselight served as the inactive lever and (2) a retractable lever on the side opposite the  
93 houselight served as the active lever; both levers were positioned 2.4 cm above the grid floor. We  
94 placed a yellow LED light for the food-paired conditioned stimulus (CS) above the active lever  
95 and equipped the central panel on the same side with a pellet receptacle connected to a pellet  
96 dispenser. Presses on the active lever (only extended during food self-administration sessions or  
97 food-seeking tests) resulted in delivery of 20-mg food pellets and a 2-s light CS (bright yellow  
98 LED), while presses on the inactive lever had no programmed consequences.

99 **Behavioral procedures**

100 The experimental timeline is shown in [Figure 1A](#). Details of the food self-administration  
101 procedure, abstinence phase, and relapse test are provided below.

102 Food self-administration

103 The food self-administration procedure is based on our previous study (43). We gave mice  
104 free access to regular chow and water in their homecages during this phase and other phases of  
105 the experiment. Prior to the operant training sessions, we gave all mice one 30-min session of

106 food-magazine training. During this session mice received 15 evenly spaced (every 2 min)  
107 deliveries of a 20-mg palatable food pellet (TestDiet, Catalogue #1811142, 12.7% fat, 66.7%  
108 carbohydrate, and 20.6% protein) paired with a 2-s discrete cue light (Food-paired CS). Next, we  
109 trained the mice to lever press for palatable food reward during one 1-h session per day. The  
110 start of a session was signaled by the illumination of the house light followed 10-s later by the  
111 presentation of the central retractable active lever for 60 min. The houselight remained on for the  
112 duration of the session and served as a discriminative stimulus that signaled availability of the  
113 palatable food upon lever press.

114 Throughout the session, responses on the active lever were rewarded under a fixed-ratio-1  
115 (FR1), 20-s timeout (TO) reinforcement schedule – active lever presses resulted in illumination of  
116 the food-paired CS for 2-s followed by the delivery of a palatable food pellet. Additional active  
117 responses during the 20-s timeout had no programmed consequence. Responses on the inactive  
118 lever had no programmed consequences throughout the session. We recorded (1) the total  
119 number of active lever presses, (2) the total number of inactive lever presses and (3) the total  
120 number of food pellet rewards earned during the entire session. Lever-press data for initial  
121 training sessions was not recorded for 2 mice due to technical malfunction so we recorded their  
122 values as zero to include them in statistical analysis. We gave mice at least 7 training sessions to  
123 acquire stable food self-administration behavior before moving to the homecage forced  
124 abstinence phase.

125 We used the above mentioned TestDiet pellets, because both mice and rats prefer this pellet  
126 over other nutritional or flavor compositions and show reliable acquisition of food self-  
127 administration without any food deprivation (42, 44). Additionally, food-stated CD-1 male mice  
128 strongly preferred these food pellets over operant aggression self-administration (43), and food-  
129 sated male and female rats strongly prefer these pellets over methamphetamine, heroin, and  
130 fentanyl self-administration in rats (45, 46).

131 Forced abstinence

132 During the abstinence phase, we housed mice in individual cages in the animal facility for 1,  
133 15 or 60 days with no access to palatable food pellets. We gave mice ad libitum access to regular  
134 chow and water during this phase and handled them once per week.

135 **Relapse tests**

136 Following abstinence, we tested mice for non-reinforced food seeking during a 30-min  
137 relapse test. During the test, responses on the active lever resulted in presentation of the Food-  
138 paired CS on the same FR1 – 20-s TO reinforcement schedule but were not reinforced with food  
139 pellets (extinction conditions). After the test, we returned mice to their homecage for 60-min prior  
140 to perfusions and brain tissue collection (n=11 for Day 1, n=12 for Day 15 and n=10 for Day 60).  
141 At each incubation timepoint, we also collected brains from food-trained mice directly from the  
142 homecage (n=4 for Homecage Day 1, n=6 for Homecage Day 15, n=3 for Homecage Day 60)  
143 and collapsed them into a single group (Homecage, n=13) to serve as baseline controls for  
144 whole-brain analysis. We matched mice from the four groups for food-reinforced responding  
145 during the training phase.

146 **Whole brain Fos immunohistochemistry (IHC)**

147 We used a modified version of the iDISCO+ protocol for intact mouse brain Fos IHC (41).  
148 We processed 37 brains across the 4 groups following perfusions (n=11 for Homecage, n=9 for  
149 Day 1, n=9 for Day 15 and n=8 for Day 60) based on perfusion quality, level of intactness, and  
150 behavioral data. Details of the sample collection, pretreatment, immunolabeling and clearing  
151 steps are provided in the sections below.

152 **Sample collection**

153 We anesthetized the mice with isoflurane and perfused them transcardially with 200 ml of  
154 0.1 M phosphate buffered saline (PBS, pH 7.4) followed by 400 ml of 4% paraformaldehyde in  
155 PBS (4% PFA, pH 7.4). We extracted brains, post-fixed them for an additional 24-h in 4% PFA at  
156 4°C, and then stored them in PBS with 0.1% sodium azide at 4°C prior to processing.

157 Sample pretreatment with methanol

158 We used 15 ml conical tubes for sample pretreatment and gently mixed tubes on a rotating  
159 mixer (Daigger Scientific, EF24935). We first washed the brains in PBS (3 x 30 min) at room  
160 temperature (RT) to remove 4% PFA. We then dehydrated samples in ascending concentrations  
161 of methanol (MeOH) in deionized H<sub>2</sub>O (dH<sub>2</sub>O) – 0%, 20%, 40%, 60%, 80%, 100%, 100% MeOH  
162 (RT, 1.5 h each). Next, we incubated samples in 66% Dichloromethane (DCM)/33% MeOH (RT, 1  
163 x 8 h followed by overnight) for delipidation. We then washed samples in 100% MeOH (RT, 2 x 3  
164 h each), and bleached them by incubating in a chilled H<sub>2</sub>O<sub>2</sub> /H<sub>2</sub>O/MeOH solution (1 volume 30%  
165 H<sub>2</sub>O<sub>2</sub> to 5 volumes 100% MeOH) overnight at 4°C. Next, we rehydrated the samples in  
166 descending concentrations of MeOH in dH<sub>2</sub>O - 80%, 60%, 40%, 20%, 0% MeOH (RT, 1.5 h  
167 each). We then washed the samples first in PBS (RT, 1 x 1 h), then three times in a buffer  
168 containing PBS with 0.5% TritonX-100 (PTx0.5) at 37°C (2 x 1 h, followed by 1 x overnight).

169 Immunolabeling

170 We used 1.5 ml Nalgene cryotubes for immunolabeling and 15 ml conical centrifuge tubes  
171 for permeabilization, blocking and wash steps. We performed staining over 7 days – we started  
172 with a lower initial concentration on day 1 (1° - 1:1000; 2° - 1:500), stepped up the concentration  
173 of antibody using booster doses (1° - 1:1000; 2° - 1:500) over the next 4 days, and let the  
174 samples incubate at the final concentration (1° - 1:200; 2° - 1:100) for an additional 2 days. We  
175 gently mixed the sample containers by rotation and always filled to the top to prevent oxidation.  
176 We first incubated the samples in permeabilization buffer containing 78.6% PTx0.5, 1.4%  
177 Glycine, and 20% Dimethyl Sulfoxide (DMSO), and then in blocking buffer containing 84%  
178 PTx0.5, 6% Normal Donkey Serum (NDS), and 10% DMSO (37°C, 2 d each). Next, we incubated  
179 the samples in primary (1°) antibodies (anti-cFos: Phospho-c-Fos (Ser32) (D82C12) XP® Rabbit  
180 mAb, Cell Signaling Technology, #5348S Lot 1; RRID#: AB\_10557109) diluted in 1° antibody  
181 buffer containing 92% PTwH0.5, 3% NDS, and 5% DMSO (37°C, 7 d). We then performed  
182 washes in a buffer containing PBS with 0.5% Tween-20, and 10ug/ml Heparin (PTwH0.5) over 4

183 days (37°C, 4 x 12 h followed by 2 x 1 d). We then incubated the samples in secondary (2°)  
184 antibodies (Alexa Fluor® 647 AffiniPure F(ab')<sub>2</sub> Fragment Donkey Anti-Rabbit IgG (H+L), Jackson  
185 ImmunoResearch Labs, 711-606-152, Lot 128806, RRID:AB\_2340625; Alexa Fluor® 488  
186 AffiniPure F(ab')<sub>2</sub> Fragment Donkey Anti-Chicken IgY (IgG) (H+L), Jackson ImmunoResearch  
187 Labs, 703-546-155, Lot 127495, RRID:AB\_2340375) diluted in 2° antibody buffer containing 97%  
188 PTwH0.5, and 3% NDS (37°C, 7 d). Finally, we performed washes in PTwH0.5 over 4 days  
189 (37°C, 4 x 8-12 h followed by 2 x 1 d).

190 **Clearing**

191 We used 15 ml conical centrifuge tubes for dehydration and delipidation, followed by glass  
192 vials with Teflon (PTFE) caps for clearing and refractive index matching. We dehydrated samples  
193 using ascending concentrations of MeOH in dH<sub>2</sub>O – 0%, 20%, 40%, 60%, 80%, 100%, 100%  
194 MeOH (RT, 1.5 h each) and performed delipidation using 66% dichloromethane (DCM)+33%  
195 MeOH (RT, 2 x 3 h, followed by 1 x overnight). Next, we washed the samples in 100% DCM (RT,  
196 2 x 3 h) to remove MeOH and incubated them in 100% dibenzyl ether (DBE) for clearing and  
197 refractive index matching (RT, 2 x 3 h, followed by 1 x overnight).

198 **Whole-brain imaging and analysis**

199 We imaged stained and cleared intact mouse brains using LSFM. We used the 3D rendering  
200 software Arivis Vision 4D (3.0.0) to stitch image tiles, manually corrected coronal alignment where  
201 necessary, and exported the images as TIFF files for whole-brain analysis. We analyzed the data  
202 using ClearMap (40) and SMART (41) analysis pipelines as described below.

203 **Light-sheet fluorescent microscopy imaging (LSFM)**

204 We used a light-sheet microscope (UltraMicroscope II with Infinity Corrected Objective  
205 Lenses, Miltenyi Biotec) with an attached camera (Andor Zyla sCMOS), a 1.1x/0.1NA objective  
206 (MI PLAN; LaVision BioTec), and a non-corrected dipping cap. Imaging parameters and  
207 acquisition order were controlled through InspectorPro software (v 7.1.4). We mounted cleared

208 Fos-stained brains in coronal orientation (olfactory bulb side up) using a custom sample platform  
209 and imaged at 2.2x effective magnification (1.1x objective x 2x magnification slider) in DBE. We  
210 acquired images for autofluorescence (Excitation: 488 nm laser, Emission: 535/43 bandpass  
211 filter) and Fos-IHC (Excitation: 647 nm laser, Emission: 690/50 bandpass filter) in separate 2 x 1  
212 tiled scans (scan order: z-x-y). We used the following fixed parameters for acquisition: exposure =  
213 ~100 ms; sheet NA = 0.16; sheet thickness = 3.89 um; sheet width = 70%; zoom = 2x; dynamic  
214 horizontal focus = 5 (Fos channel only); dynamic horizontal focus processing = blend; merge  
215 light-sheet = blend; 488 nm laser power = 20%; 647 nm laser power = 50%. Final image pixel  
216 resolution was 2.956 um X x 2.956 um Y x 3 um Z. Resulting tiles were stitched into full size  
217 coronal planes using Arivis Vision 4D (3.0.0) and exported as TIFFs. During the analyses, we  
218 observed that illumination of the entire coronal plane by the light-sheet during the first imaged tile  
219 scan led to significant photo-bleaching of the second imaged tile in all samples. Therefore, we  
220 only used the first imaged hemisphere from each sample for analysis and mirrored outputs for all  
221 visualizations. We excluded 3 brains due to insufficient clearing and staining, and 2 brains due to  
222 technical issues during image acquisition. We analyzed LSFM data for 32 brains across the 4  
223 groups (n=11 for Homecage, n=8 for Day 1, n=8 for Day 15, and n=5 for Day 60).

224 ClearMap analysis

225 We used the open-source program ClearMap 1.0 (40) for whole-brain volumetric analysis on  
226 a dedicated machine (Intel Xeon® CPU E5-2650 v4 @ 2.20GHz x 48; 4 x GeForce GTX 1080  
227 Ti/PCIe/SSE2; 256GB RAM). We downsampled autofluorescence image stacks for each sample  
228 and registered them to a common 25  $\mu$ m isotropic serial two-photon (STP) tomography reference  
229 template (47). We manually validated registration for each sample by post-hoc inspection of  
230 overlaid reference template and post-transformation image stacks in ImageJ. Three brains failed  
231 registration and were excluded from further analysis in ClearMap.

232 Next, we used the spot-detection method in ClearMap to automate Fos+ cell detection  
233 across all images. We used the same spot detection filter parameters for Fos+ cell detection  
234 across all samples (illumination correction: mean scaling; cell shape detection: threshold (150);

235 Find intensity: Mean and size (3,3,3); Background removal: pixel size (5,5); DoG filter: pixel size  
236 (6,6,11); Detect cell shape: threshold (150) and then applied a voxel size threshold (50,200000)  
237 to constrain the size of detected Fos+ cells.

238 We validated this Fos+ cell detection procedure against ground truth manual cell detection  
239 performed by 2 expert raters across five separate 300 um x 300 um regions of interest (ROIs)  
240 from 3 sample image volumes. For each sample, we first isolated an 81 mm coronal image stack  
241 (26 image z-stack) at ~1.46 from bregma along the anterior-posterior axis, and within it selected 5  
242 ROIs encompassing a wide range of Fos+ cell density and background fluorescence signal. Two  
243 expert raters performed ground truth manual annotation of Fos+ cells in each FOV using ITK-  
244 SNAP software (48) version 3.8.0 (<http://www.itksnap.org>), resulting in ~3500 manually annotated  
245 Fos+ cells. We used the FIJI image analysis package (49) to overlay the automated ClearMap  
246 annotation over the expert annotation and used Analyze Objects and Image Calculator plugins to  
247 determine expert-rated Fos+ cell counts, ClearMap-rated Fos+ cell counts and overlap. We  
248 calculated precision (ratio of correctly predicted Fos+ cells to all predicted cells), recall (ratio of  
249 correctly predicted Fos+ cells to expert annotated Fos+ cells), and F-score (harmonic mean of  
250 precision and recall) in Microsoft Excel ([Supplementary Figure S1A](#)).

251 We warped all ClearMap detected Fos+ cells into the reference space by applying  
252 transformation coordinates from the registration step and obtained counts for individual brain  
253 regions based on the Allen Brain Institute atlas ontology provided with the ClearMap installation  
254 package. We extracted Fos+ cell counts for all 1205 annotations and used custom python scripts  
255 to generate summed counts within regions of interest (ROIs) for analysis of activity changes  
256 between groups.

257 SMART2 analysis

258 We used an updated version of the open-source R package SMART (41) (SMART2) for  
259 expert-guided registration and volumetric activity mapping within two coronal subvolumes  
260 selected from ClearMap analysis (subvolume 1: AP +1.55 to AP +1.75 relative to Bregma;  
261 subvolume 2: AP -1.08 to AP -1.28 relative to Bregma). SMART extends the WholeBrain

262 analytical framework (38) to volumetric analysis using mouse LSFM datasets and allows user-  
263 guided refinement of registration prior to automated Fos+ cell detection. We followed the steps  
264 outlined in the updated online tutorial (<https://github.com/sgoldenlab/SMART2>) for analysis. First,  
265 we set up sample information (animal ID, initials, paths, z spacing, registration step (space  
266 between z images), most anterior AP coordinate and z image number, most posterior AP  
267 coordinate and z image number), and file paths for each sample volume using the functions  
268 `setup_pl()`, `im_sort()`, and `get_savepaths()`. Next, we used the function `choice()` to align the entire  
269 sample volume to internal reference atlas plates along the anterior-posterior axis and used the  
270 function `interpolate()` to identify z-stack numbers corresponding to substacks of interest.

271 Next, we used the interpolated values to select 4 consecutive reference plates (100 mm  
272 spacing) for analysis of each coronal subvolume. We selected 2 'internal plates' to register the  
273 subvolume of interest (subvolume 1: ABA atlas plate numbers 36 and 37 corresponding to AP  
274 +1.6 and AP +1.7; subvolume 2: ABA atlas plate numbers 65 and 66 corresponding to AP -1.13  
275 and AP -1.23) and specified 2 additional 'outer plates' to enable identification of Fos+ cells  
276 corresponding to 50 mm on either side of the registered internal plates.

277 This resulted in a final analyzed subvolume of 200 mm thickness in atlas space. We used  
278 the functions `regi_loop()` and `filter_loop()` for user-guided registration correction between  
279 autofluorescence channel images and atlas reference plates. We used the same initial  
280 parameters for all samples [`alim = c(50, 50)`, `threshold.range = c(50000, 60000L)`, `eccentricity =`  
281 `999L`, `Max = 3025`, `Min = 0`, `brain.threshold = 400`, `resize = 0.25`, `blur = 7`, `downsample = 2`] and  
282 applied additional correspondence points to improve the automated registration output.

283 We then used the function `seg_loop()` for Fos+ cell detection throughout the subvolume of  
284 interest and the function `clean_duplicates()` to correct for duplicate detection of Fos+ cells across  
285 adjacent z-slices. We applied the same segmentation filter parameters for all samples [`alim = c(4,`  
286 `100)`, `threshold.range = c(500, 2200)`, `eccentricity = 300`, `Max = 2200`, `Min = 100`, `brain.threshold`  
287 `= 400`, `resize = 0.25`, `blur = 7`, `downsample = 2`]. We validated the automated Fos+ cell detection

288 against ground truth expert annotation as described in the previous section and calculated  
289 precision, recall, and F-score in Microsoft Excel ([Supplementary Figure S1B](#)).

290 Finally, we used the `forward_warp()` function to warp detected Fos+ cells into the reference  
291 atlas space and obtained counts for individual brain regions based on the Allen Mouse CCF  
292 ontology provided in the WholeBrain package. We extracted Fos+ cell counts and used the  
293 `get_rois()` function to generate summed counts by ROI. We also implemented additional  
294 `isolate_dataset()`, `cell_count_compilation()`, `get_groups()`, and `voxelize()` functions to generate  
295 Fos+ cell counts and Fos+ cell density heatmaps for individual samples, and to output summary  
296 counts by experimental group. We provide these additional scripts as part of the updated  
297 SMART2 package repository (<https://github.com/sgoldenlab/SMART2>). We also provide a Docker  
298 installation image for rapid and user-friendly installation  
299 ([https://hub.docker.com/repository/docker/goldenneurolab/wholebrain\\_smart2](https://hub.docker.com/repository/docker/goldenneurolab/wholebrain_smart2)).

300 **Statistical analyses**

301 We analyzed behavioral and whole-brain Fos data using IBM SPSS Statistics (version 28)  
302 and GraphPad Prism (version 9.3). For the behavioral and whole brain analysis, alpha  
303 (significance) level was set at 0.05, two-tailed. We tested the data for sphericity and homogeneity  
304 of variance when appropriate. When the sphericity assumption was not met, we adjusted the  
305 degrees of freedom using the Greenhouse-Geisser correction. Because our analyses yielded  
306 multiple main effects and interactions, we report only those that are critical for data interpretation.  
307 See [Supplementary Table S2](#) for a listing of number of subjects/samples included in each phase  
308 of the study, and [Supplementary Tables S3-S11](#) for statistical analyses and summary statistics.

309 Behavior

310 We analyzed two behavioral measures during food self-administration training - (1) the total  
311 number of lever presses on active and inactive lever (denoted as *lever presses*), and (2) the total  
312 number of food pellet rewards earned (denoted as *rewards*) during a 1-h self-administration  
313 session. Following training, we tested separate groups of mice for palatable food reward-seeking

314 after 1, 15 or 60 days of abstinence and analyzed non-reinforced responding (*lever presses*)  
315 during the 30-min food seeking test. We describe the within- and between-subjects factors in the  
316 mixed ANOVAs we used to analyze the behavioral data in the Results section and in  
317 [Supplementary Table S3](#).

318 ClearMap analysis

319 We processed the 32 brains using the ClearMap pipeline, of which 3 failed the registration to  
320 the reference atlas. We analyzed the remaining 29 brains (n=9 for Homecage, n=7 for Day 1, n=8  
321 for Day 15 and n=5 for Day 60) as described below. We computed Fos+ cell counts within  
322 regions at two levels of the atlas hierarchy - (1) 10 major anatomical divisions, and (2) 56  
323 subregions across the brain based on hierarchical relationships defined in the Allen Mouse  
324 Common Coordinate Framework (50). We selected regions within each level such that there were  
325 no parent-child relationships and/or overlapping spatial footprints between them. We transformed  
326 the raw Fos+ cell counts to Z-scores prior to the statistical analyses to normalize the data and  
327 account for differences in volume across regions of interest. We computed Z-scores for each  
328 region of interest relative to the homecage group's values using the formula  $z = (x - \mu) / \sigma$ , where  
329  $x$  is the sample Fos+ cell count,  $\mu$  is the mean Fos+ cell count of the homecage group, and  $\sigma$  is  
330 the standard deviation of the homecage group. For each level, we first used 2-way mixed ANOVA  
331 (GLM procedure in SPSS) with the within-subject factor of Region and the between-subjects  
332 factor of Group (Homecage, Day 1, Day 15, Day 60). We followed up on significant main effects  
333 and interactions with 1-way ANOVAs within each region and used Tukey HSD test for post-hoc  
334 comparisons between Groups. We provide the statistical outputs for all analysis pertaining to  
335 ClearMap in [Supplementary Tables S4 and S5](#).

336 SMART2 analysis

337 In addition to the 29 brains used for ClearMap analysis, we used manual registration  
338 correction within each subvolume to recover and analyze brains that failed ClearMap registration.  
339 We analyzed 32 brains across the 4 groups (n=11 for Homecage, n=8 for Day 1, n=8 for Day 15

340 and n=5 for Day 60) for subvolume 1 and 31 brains across the 4 groups (n=10 for Homecage,  
341 n=8 for Day 1, n=8 for Day 15 and n=5 for Day 60) for subvolume 2. We extracted Fos+ cell  
342 counts within regions of interest at three levels of the atlas hierarchy - (1) major anatomical  
343 regions (subvolume 1: 5 regions; subvolume 2: 8 regions), (2) main subregions (subvolume 1: 18  
344 subregions; subvolume 2: 28 regions), and (3) internal subdivisions within the subregions  
345 (subvolume 1: 43 subdivisions; subvolume 2: 64 subdivisions). Similar to ClearMap analysis, we  
346 selected regions within each level such that there were no parent-child relationships and/or  
347 overlapping spatial footprints between them. We transformed the raw Fos+ cell counts to Z-  
348 scores prior to statistical testing. For each level, we first used 2-way mixed ANOVA (GLM  
349 procedure in SPSS) with the within-subject factor of Region and the between-subjects factor of  
350 Group (Homecage, Day 1, Day 15, Day 60). We followed up on significant main effects and  
351 interactions with 1-way ANOVAs within each region and used Tukey HSD test for posthoc  
352 comparisons between Groups. We also generated spatial maps of Fos+ cell densities (mean  
353 across group) for each subvolume to aid visualization of differences in activation patterns  
354 between groups. We provide statistical outputs for all analysis pertaining to SMART2 subvolumes  
355 1 and 2 in [Supplementary Tables S6-S8](#) and [S9-S11](#), respectively.

356

## 357 **Results**

358 The experimental design and timeline of behavioral training, intact mouse brain activity  
359 labeling, and brain-wide activity mapping is shown in [Figure 1A](#). See [Supplementary Table S2](#) for  
360 a listing of number of subjects/samples included in each phase of the study.

### 361 **Incubation of palatable food seeking in male CD1 mice**

362 We determined whether the time-dependent increases in food seeking during abstinence  
363 (incubation of food craving), previously observed in rats (3, 19), generalize to mice. The  
364 experimental timeline is shown in [Figure 1A-B](#). We trained food sated CD-1 male mice to lever  
365 press for high carbohydrate food pellets for 7 days. Next, we tested different groups for relapse to

366 food seeking in the presence of contextual (food self-administration chamber) and discrete (light  
367 cue paired with food delivery) cues after 1, 15, or 60 days of homecage forced abstinence.  
368 Statistical outputs for all analyses pertaining to training and relapse tests are provided in tabular  
369 format as [Supplementary Table S3](#).

370 Training phase: The mice showed reliable food self-administration as indicated by increased  
371 responding on the food-paired lever during the daily sessions ([Figure 1C](#)). The repeated-  
372 measures ANOVA of *rewards*, which included the within-subjects factor of Training session  
373 (sessions 1-7), showed a significant effect of this factor ( $F_{3,2,144.7}= 67.2$ ,  $p<0.001$ ). The repeated-  
374 measures ANOVA of *lever presses*, which included the within-subjects factors of Training session  
375 and Lever (inactive, active), showed a significant interaction between the two factors ( $F_{3,6,160.3}=$   
376  $28.9$ ,  $p<0.001$ ).

377 Relapse (incubation) tests: Non-reinforced presses on the previously active lever were  
378 significantly higher after 60 abstinence days than after 1 or 15 days ([Figure 1D](#)), indicating  
379 'incubation of food craving.' The mixed ANOVA of total lever presses, which included the  
380 between-subjects factor of Abstinence day (1, 15, 60) and Lever, showed a significant interaction  
381 between the two factors ( $F_{2,30}=3.4$ ,  $p=0.045$ ). This incubation effect was primarily due to higher  
382 lever presses in the day 60 group during the first 10 min of the test session ([Figure 1D](#) right  
383 panel). Post-hoc group differences (Tukey test) are depicted in [Figure 1D](#).

384 **Unbiased intact brain-wide activity mapping of incubation of palatable food-seeking**

385 We processed brains using the ClearMap pipeline and extracted Fos+ cell counts within  
386 regions of interest (ROIs) at two levels of the atlas hierarchy - (1) 10 major anatomical divisions  
387 ([Fig. 2A](#), [Fig. 2B](#) right panel), and (2) 56 subregions of interest ([Fig. 2C](#)) based on hierarchical  
388 relationships defined in the Allen Mouse Common Coordinate Framework (50). We selected  
389 regions within each level such that there were no parent-child relationships and/or overlapping  
390 spatial footprints between them and performed z-score normalization (relative to homecage group  
391 mean) for each ROI prior to statistical analyses ([Fig. 2C](#), center and left panels).

392        Main regions: For Level 1, the mixed ANOVA, using the between-subjects factor of Group  
393        (homecage, day 1, day 15, day 60) and the within-subjects factor of Region, showed significant  
394        main effects of Group ( $F_{3,25}=4.4$ ,  $p=0.013$ ) and Region ( $F_{1,9,47.7}=18.2$ ,  $p<0.001$ ), and an interaction  
395        between the two factors ( $F_{5.7,47.7}=6.1$ ,  $p<0.001$ ). Follow up one-way ANOVAs (between-subjects  
396        factor Group) were significant for 6 out of 10 tested regions (denoted by \* in Figure 2B, left panel).  
397        These data indicate increased brain-wide activation after the relapse test after 60 abstinence  
398        days, but not 1 or 15 days. Detailed statistical reporting of the ClearMap analysis at Level 1 are  
399        provided in [Supplementary Table S4](#).

400        Sub-regions: For Level 2, the mixed ANOVA showed significant main effects of Group  
401        ( $F_{3,25}=6.0$ ,  $p=0.003$ ), and Sub-region ( $F_{1,8,45.3}=18.7$ ,  $p<0.001$ ) and an interaction between the two  
402        factors ( $F_{5.4,45.3}=8.0$ ,  $p<0.001$ ). Follow up one-way ANOVAs (between-subjects factor Group)  
403        were significant for 33 out of 56 tested subregions, reflecting selective increased activation  
404        following relapse test after 60 but not 1 or 15 days of abstinence. The most activated areas were  
405        olfactory, cortical, cortical subplate, and striatal subregions - collectively designated as  
406        'corticostriatal', with sparse subregional activation in the retrohippocampal region of the  
407        hippocampus and medulla of the hindbrain. Increased Fos expression in the day 60 group was  
408        not observed in subregions of thalamus, hypothalamus, and midbrain structures. Detailed  
409        statistical reporting of ClearMap analysis at Level 2 are provided in [Supplementary Table S5](#).

410        **Targeted analysis of the corticostriatal coronal subvolume using SMART2**

411        We processed a 200  $\mu\text{m}$  thick coronal subvolume spanning AP +1.55 to AP +1.75 relative to  
412        Bregma using the SMART2 pipeline. Spatial maps of group-wise Fos+ cell densities within the  
413        subvolume are shown in [Figure 3A](#) to aid visualization of differences in activation patterns  
414        between groups. We extracted Fos+ cell counts within regions of interest (ROIs) at three levels  
415        of the atlas hierarchy - (1) 5 major anatomical regions ([Fig. 3B](#)), (2) 18 main subregions ([Fig. 3C](#))  
416        and (3) 43 internal subdivisions within the subregions ([Fig. 3D](#)). We performed z-score  
417        normalization (relative to homecage mean) for each ROI prior to statistical analysis. One-way

418 ANOVA and Tukey HSD p-values for all Level 2 ROIs and a subset of Level 3 ROIs are shown as  
419 heatmaps in [Figure 3C](#) and [Figure 3D](#).

420 Main regions: For Level 1, the mixed ANOVA showed significant main effects of Region  
421 ( $F_{1.6,4.7}=6.5$ ,  $p=0.006$ ) and Group ( $F_{3,28}=6.9$ ,  $p=0.001$ ), and an interaction between the two factors  
422 ( $F_{4.7,44.0}=4.7$ ,  $p=0.002$ ). Follow-up one-way ANOVAs were significant for all 5 tested regions,  
423 reflecting increased activation following the relapse test after 60 but not 1 or 15 days of  
424 abstinence. Detailed statistical reporting for SMART2 analysis at Level 1 are provided in  
425 [Supplementary Table S6](#).

426 Sub-regions: For Level 2, the mixed ANOVA showed significant main effects of Sub-region  
427 ( $F_{1.2,32.7}=9.5$ ,  $p=0.003$ ) and Group ( $F_{3,28}=7.2$ ,  $p=0.001$ ), and an interaction between the two factors  
428 ( $F_{3.5,32.7}=6.3$ ,  $p=0.001$ ). Follow-up one-way ANOVAs were significant for all 18 tested subregions,  
429 reflecting increased activation following the relapse test after 60 but not 1 or 15 days of  
430 abstinence. Detailed statistical reporting for SMART2 analysis at Level 2 are provided in  
431 [Supplementary Table S7](#).

432 Sub-divisions: For Level 3, the mixed ANOVA showed significant main effects of Sub-  
433 division ( $F_{1.2,32.2}=10.3$ ,  $p=0.002$ ) and Group ( $F_{3,28}=7.1$ ,  $p=0.001$ ), and an interaction between the  
434 two factors ( $F_{3.5,32.2}=6.5$ ,  $p=0.001$ ). Follow-up one-way ANOVAs were significant for all 43 tested  
435 subdivisions, reflecting increased activation following the relapse test after 60 but not 1 or 15  
436 days of abstinence. Detailed statistical reporting for SMART2 analysis at Level 3 are provided in  
437 [Supplementary Table S8](#).

438 **Targeted analysis of the thalamocortical coronal subvolume using SMART2**

439 We processed a 200  $\mu\text{m}$  thick coronal subvolume spanning AP -1.08 to AP -1.28 relative to  
440 Bregma using the SMART2 pipeline. Spatial maps of group-wise Fos+ cell densities within the  
441 subvolume are shown in [Figure 4A](#) to aid visualization of differences in activation patterns  
442 between groups. We extracted Fos+ cell counts within ROIs at three levels of the atlas hierarchy -  
443 (1) 8 major anatomical regions ([Fig. 4B](#)), (2) 28 main subregions ([Fig. 4C](#)), and (3) 64 internal

444 subdivisions within the subregions (Fig. 4D). One-way ANOVAs and Tukey HSD p-values for all  
445 Level 2 ROIs and a subset of Level 3 ROIs are shown as heatmaps in Figure 4C and Figure 4D.

446 Main regions: For Level 1, the mixed ANOVA showed significant main effects of Region  
447 ( $F_{1,2,32,3}=14.0$ ,  $p<0.001$ ) and Group ( $F_{3,27}=3.2$ ,  $p=0.040$ ), and an interaction between the two  
448 factors ( $F_{3,6,32,3}=5.5$ ,  $p=0.002$ ). Follow-up one-way ANOVAs were significant for 3 of 8 tested  
449 regions, reflecting increased activation following the relapse test after 60 but not 1 or 15 days of  
450 abstinence. Detailed statistical reporting for SMART2 analysis at Level 1 are provided in  
451 [Supplementary Table S9](#).

452 Sub-regions: For Level 2, two-way ANOVA showed significant main effects of Sub-region  
453 ( $F_{1,1,30,8}=9.5$ ,  $p=0.003$ ) and Group ( $F_{3,27}=3.4$ ,  $p=0.033$ ), and an interaction between the two factors  
454 ( $F_{3,4,30,8}=3.5$ ,  $p=0.024$ ). Follow-up one-way ANOVAs were significant for 12 of 18 tested  
455 subregions, reflecting increased activation following relapse test after 60 but not 1 or 15 days of  
456 abstinence. Detailed statistical reporting for SMART2 analysis at Level 2 are provided in  
457 [Supplementary Table S10](#).

458 Sub-divisions: For Level 3, the mixed ANOVA showed significant main effects of Region  
459 ( $F_{1,2,31,2}=7.8$ ,  $p=0.007$ ) and Group ( $F_{3,27}=3.1$ ,  $p=0.044$ ), and an interaction between the two factors  
460 ( $F_{3,5,31,2}=3.0$ ,  $p=0.038$ ). Follow-up one-way ANOVAs were significant for 20 of 64 tested  
461 subdivisions, reflecting increased activation following the relapse test after 60 but not 1 or 15  
462 days of abstinence. Detailed statistical reporting for SMART2 analysis at Level 3 are provided in  
463 [Supplementary Table S11](#).

464

## 465 **Discussion**

466 We used unbiased intact-brain mapping of Fos expression to investigate brain-wide  
467 activation patterns during incubation of palatable food seeking in mice. Relapse to food seeking  
468 was higher after 60 abstinence days than after 1 or 15 days, indicating incubation of food  
469 seeking. More importantly, unbiased whole-brain analysis of Fos expression using ClearMap

470 showed a strong induction of neural activity across multiple brain regions that mirrored incubation  
471 of food seeking after 60, but not 15, days of abstinence. Targeted coronal slice analysis of Fos  
472 expression using SMART2 replicated and validated the time-dependent increases in activation  
473 patterns within corticostriatal and thalamocortical subvolumes and enabled detailed analysis of  
474 subdivision and layer-specific changes during abstinence. Overall, our data indicate that  
475 incubation of palatable food craving in male mice correlates with widespread activation of many  
476 brain regions beyond those previously implicated in incubation of food or drug craving (see  
477 [Supplementary Table S1](#)).

478 Outbred mice show incubation of palatable food seeking similar to outbred rat models

479 Food-seeking in the male mice increased following 60, but not 15, days of abstinence from  
480 food self-administration training. Previous studies in rats reported incubation of food seeking but  
481 with a different time course; robust incubation of sucrose craving was observed in rats after 7, 15,  
482 and 30 abstinence days, but not 60 days (5). Incubation of food craving has also been reported  
483 in rats trained to self-administer standard chow pellets after 30 abstinence days (14), and high-  
484 carbohydrate pellets after 21 abstinence days (15). By extending the model to mice we were able  
485 to leverage mouse-optimized procedures to investigate brain-wide activation patterns of  
486 incubation of food craving. A question for future research is whether a similar timecourse of  
487 incubation of food craving (and pattern of brain activation) will be observed in CD-1 female mice.

488 Notably, we used outbred CD-1 mice in these experiments rather than more commonly used  
489 inbred C57BL/6J. In our experience, outbred and hybrid mice exhibit a more complex spectrum of  
490 behavior and acquire learned behavior more robustly than their inbred counterparts(51-54).  
491 Additionally, an inbred genetic background limits the generalizability of genotype-phenotype  
492 relationships(55) and display highly heritable stain-specific phenotypes in brain volume, scalar  
493 diffusion tensor imaging metrics, and quantitative connectomes(56). Similarly, contrary to the  
494 assumed relationship, meta-analysis of coefficients of variation do not find evidence of greater  
495 trait stability in inbred mice than outbred mice, and hybrid mice show enhanced properties

496 desired for neurobiological research such as reduced anxiety-like behavior, improved learning,  
497 and enhanced long-term spatial memory(57). Taken within the context of developing a resource  
498 whole brain atlas for incubation of food craving in mice, outbred populations likely provide a more  
499 generalizable and robust platform.

500 Brain-wide patterns of increased neural activation following incubation of food seeking during  
501 abstinence

502 Previous targeted region-by-region analyses in rats have identified several brain regions that  
503 are activated during incubation of food and drug craving during abstinence (1, 3, 21, 28, 29).  
504 However, an unbiased brain-wide interrogation of regions engaged during 'incubated' reward  
505 seeking has not previously been performed. We used a modified version of the iDISCO+  
506 procedure to label active (Fos+) neurons in intact mouse brains and employed light-sheet  
507 fluorescence microscopy to image these potentially behaviorally relevant neurons across the  
508 entire brain volume at single cell resolution. Our unbiased ClearMap analysis revealed a time-  
509 dependent increase in activation of multiple brain regions (e.g., prelimbic, infralimbic,  
510 orbitofrontal, and insular cortices, central amygdala and basolateral amygdala, ventral and dorsal  
511 striatum) similar to that shown in rats following incubation of food and drug seeking (58-64) (see  
512 [Supplementary Table S1](#)). However, this time-dependent increase of neural activity was not  
513 restricted to these previously identified incubation-related regions. Indeed, over half of the tested  
514 main regions (6 out of 10 main anatomical divisions, and 33 of 56 subregions), including most  
515 regions within the isocortex, olfactory areas, cortical subplate, and striatum, showed statistically  
516 significant increased activation after 60, but not 15 abstinence days. Additionally, compared to  
517 'resting state' activity in the homecage, most regions showed an initial dip in activation following  
518 food seeking tests on days 1 and 15, which preceded strong induction on day 60.

519 Of note, it is possible that a small number of the statistically significant activated regions  
520 reflect false positive results, because we did not correct the statistical analyses for multiple one-  
521 way ANOVA comparisons. On the other hand, the fact that statistically significant increases in

522 Fos expression in multiple brain regions were detected using a relatively small n per group (n=5  
523 to 8 for abstinence days 1, 15, and 60) speaks to the robustness of our results.

524 Two recent studies used a similar unbiased approach to investigate brain-wide activation  
525 (assessed by Fos) patterns of short-term alcohol abstinence after two-bottle choice and chronic  
526 intermittent ethanol vapor exposure (65) and acute withdrawal following experimenter-  
527 administered psychostimulants (66). They used hierarchical clustering techniques to demonstrate  
528 a strong decrease in modularity after abstinence/withdrawal compared to drug-naïve controls and  
529 employed graph theory approaches to identify hub regions that might drive this functional  
530 restructuring. It is possible that this observed decrease in modularity might be a result of the  
531 recruitment of networks of regions like that seen in our study following food seeking in mice.  
532 However, it is important to note that in these studies, brains were collected directly from  
533 homecage and not after behavioral testing. Thus, the observed changes likely reflect shifts in  
534 'resting-state' functional brain architecture and not behaviorally evoked differential network  
535 engagement during reward seeking.

536 Targeted subvolume analysis reveals variations in time-dependent brain activation patterns

537 We followed-up on our global volumetric ClearMap findings by analyzing activation patterns  
538 within two coronal volumes using an updated version of the SMART pipeline (41). This approach  
539 allowed us to use expert-guided registration and Fos-segmentation to (1) isolate ClearMap effects  
540 to specific coronal plates along the anterior-posterior axis, (2) validate our data against previous  
541 incubation studies that used selected coronal slices for Fos-mapping, (3) include brains that failed  
542 ClearMap due to physical damage during processing, and (4) extend our analysis to subdivisions  
543 and layers within these subvolumes for future mechanistic investigation. In agreement with the  
544 ClearMap analyses, SMART2 analysis identified several subregions with increased activation  
545 (following the relapse tests on abstinence day 60) within the isocortex, olfactory areas, cortical  
546 subplate, and striatum across the two selected subvolumes, several of which have been  
547 previously identified after incubation of food and drug seeking (e.g., prelimbic cortex, infralimbic

548 cortex, orbitofrontal cortex, basolateral amygdala, central amygdala nucleus, nucleus accumbens,  
549 somatosensory cortex) and drug seeking (e.g., prelimbic cortex, infralimbic cortex, agranular  
550 insular area, basolateral amygdala, central amygdalar nucleus, nucleus accumbens) in rat  
551 models (58-64, 67-69) (see [Supplementary Table S1](#)). However, while some identified subregions  
552 showed similar increases in activation across both subvolumes (e.g., somatomotor and  
553 somatosensory areas, claustrum, piriform area), others were only present in one subvolume (e.g.,  
554 prelimbic and infralimbic cortex, basolateral and central amygdala) or showed differential  
555 engagement along anterior-posterior axis (e.g., anterior cingulate and agranular insular cortices).  
556 Even within a subvolume and subregion, activation patterns were not uniform but sometimes  
557 graded across layers (e.g., layers of piriform area, dorsal peduncular area, infralimbic and  
558 prelimbic areas) or isolated to specific subdivisions (e.g., central but not medial or lateral  
559 subdivisions of central amygdala), suggesting different degrees of engagement across multiple  
560 brain regions and likely circuits after 60 abstinence days.

561 Conclusions

562 We demonstrated that incubation of palatable food reward seeking is accompanied by an  
563 induction of neural engagement in multiple brain regions, many of them extend beyond the  
564 traditional brain areas and circuits involved in incubation of food and drug craving. We extend the  
565 rat incubation of food seeking model to male CD1 mice and leverage mouse-specific unbiased  
566 whole-brain staining, clearing, and analysis pipelines to generate a single-cell resolution whole-  
567 brain atlas of incubation food seeking during prolonged abstinence. The results of our study  
568 suggest that the overarching neural mechanism underlying incubation of reward seeking is more  
569 anatomically widespread than suggested by the published literature and likely not localized to a  
570 particular brain area or circuit. Whatever neural mechanisms mediate incubation of reward  
571 seeking, our findings suggest that these mechanisms affect acute neural responses throughout  
572 the brain, either through widespread alterations in all these brain areas or in key brain areas that  
573 regulate brain-wide circuitry. And finally, the 'incubation atlas' here provides a mineable dataset

574 for better understanding system-level alterations related to incubation of food craving and  
575 relapse.

576

577 **Acknowledgments**

578 We want to thank members of the Hope, Shaham, and Golden labs for their support and insight  
579 during all stages of this study. We also thank Dr. Jennifer M. Bossert and Dr. Ida Fredriksson for  
580 assistance with whole-brain immunohistochemical assays, and Dr. Carlos A. Mejias-Aponte and  
581 Vadim Kashtelyan for technical assistance with sample processing and light sheet fluorescence  
582 microscopy.

583

584

585 **References**

- 586 1. C. L. Pickens *et al.*, Neurobiology of the incubation of drug craving. *Trends in*  
587 *neurosciences* **34**, 411-420 (2011).
- 588 2. M. E. Wolf, Synaptic mechanisms underlying persistent cocaine craving. *Nature Reviews*  
589 *Neuroscience* **17**, 351-365 (2016).
- 590 3. J. W. Grimm, Incubation of food craving in rats: A review. *Journal of the Experimental*  
591 *Analysis of Behavior* **113**, 37-47 (2020).
- 592 4. J. W. Grimm, B. T. Hope, R. A. Wise, Y. Shaham, Neuroadaptation. Incubation of cocaine  
593 craving after withdrawal. *Nature* **412**, 141-142 (2001).
- 594 5. L. Lu, J. W. Grimm, B. T. Hope, Y. Shaham, Incubation of cocaine craving after  
595 withdrawal: a review of preclinical data. *Neuropharmacology* **47 Suppl 1**, 214-226  
596 (2004).
- 597 6. J. L. Neisewander *et al.*, Fos protein expression and cocaine-seeking behavior in rats  
598 after exposure to a cocaine self-administration environment. *J Neurosci* **20**, 798-805  
599 (2000).
- 600 7. L. T. L. Tran-Nguyen *et al.*, Time-Dependent Changes in Cocaine-Seeking Behavior and  
601 Extracellular Dopamine Levels in the Amygdala during Cocaine Withdrawal.  
602 *Neuropsychopharmacology* **19**, 48-59 (1998).
- 603 8. U. Shalev, M. Morales, B. Hope, J. Yap, Y. Shaham, Time-dependent changes in  
604 extinction behavior and stress-induced reinstatement of drug seeking following  
605 withdrawal from heroin in rats. *Psychopharmacology* **156**, 98-107 (2001).
- 606 9. J. D. Shepard, J. M. Bossert, S. Y. Liu, Y. Shaham, The anxiogenic drug yohimbine  
607 reinstates methamphetamine seeking in a rat model of drug relapse. *Biol Psychiatry* **55**,  
608 1082-1089 (2004).
- 609 10. P. Bienkowski *et al.*, Time-dependent changes in alcohol-seeking behaviour during  
610 abstinence. *Eur Neuropsychopharmacol* **14**, 355-360 (2004).
- 611 11. A. Abdolahi, G. Acosta, F. J. Breslin, S. E. Hemby, W. J. Lynch, Incubation of nicotine  
612 seeking is associated with enhanced protein kinase A-regulated signaling of dopamine-  
613 and cAMP-regulated phosphoprotein of 32 kDa in the insular cortex. *Eur J Neurosci* **31**,  
614 733-741 (2010).
- 615 12. J. W. Grimm, Y. Shaham, B. T. Hope, Effect of cocaine and sucrose withdrawal period on  
616 extinction behavior, cue-induced reinstatement, and protein levels of the dopamine  
617 transporter and tyrosine hydroxylase in limbic and cortical areas in rats. *Behav*  
618 *Pharmacol* **13**, 379-388 (2002).
- 619 13. J. W. Grimm, A. M. Fyall, D. P. Osincup, Incubation of sucrose craving: effects of reduced  
620 training and sucrose pre-loading. *Physiology & Behavior* **84**, 73-79 (2005).
- 621 14. R. A. Darling, P. M. Dingess, K. C. Schlidt, E. M. Smith, T. E. Brown, Incubation of food  
622 craving is independent of macronutrient composition. *Scientific Reports* **6**, 30900 (2016).

623 15. I. N. Krasnova *et al.*, Incubation of Methamphetamine and Palatable Food Craving after  
624 Punishment-Induced Abstinence. *Neuropsychopharmacology* **39**, 2008-2016 (2014).

625 16. G. Bedi *et al.*, Incubation of cue-induced cigarette craving during abstinence in human  
626 smokers. *Biol Psychiatry* **69**, 708-711 (2011).

627 17. M. A. Parvaz, S. J. Moeller, R. Z. Goldstein, Incubation of cue-induced craving in adults  
628 addicted to cocaine measured by electroencephalography. *JAMA Psychiatry* **73**, 1127-  
629 1134 (2016).

630 18. G. Wang *et al.*, Effects of length of abstinence on decision-making and craving in  
631 methamphetamine abusers. *PLoS one* **8**, e68791 (2013).

632 19. S. G. Nair, T. Adams-Deutsch, D. H. Epstein, Y. Shaham, The neuropharmacology of  
633 relapse to food seeking: methodology, main findings, and comparison with relapse to  
634 drug seeking. *Prog Neurobiol* **89**, 18-45 (2009).

635 20. Y. Dong, J. R. Taylor, M. E. Wolf, Y. Shaham, Circuit and Synaptic Plasticity Mechanisms  
636 of Drug Relapse. *J Neurosci* **37**, 10867-10876 (2017).

637 21. I. Fredriksson *et al.*, Animal models of drug relapse and craving after voluntary  
638 abstinence: A review. *Pharmacol Rev* **73**, 1050-1083 (2021).

639 22. K. K. Szumlinski, C. B. Shin, Kinase interest you in treating incubated cocaine-craving? A  
640 hypothetical model for treatment intervention during protracted withdrawal from  
641 cocaine. *Genes Brain Behav* **17**, e12440 (2018).

642 23. H. Bito, K. Deisseroth, R. W. Tsien, CREB Phosphorylation and Dephosphorylation: A  
643 Ca<sup>2+</sup>- and Stimulus Duration-Dependent Switch for Hippocampal Gene Expression. *Cell*  
644 **87**, 1203-1214 (1996).

645 24. F. C. Cruz *et al.*, New technologies for examining the role of neuronal ensembles in drug  
646 addiction and fear. *Nat Rev Neurosci* **14**, 743-754 (2013).

647 25. F. C. Cruz, F. Javier Rubio, B. T. Hope, Using c-fos to study neuronal ensembles in  
648 corticostriatal circuitry of addiction. *Brain Research* **1628**, 157-173 (2015).

649 26. J. I. Morgan, T. Curran, Stimulus-Transcription Coupling in the Nervous System:  
650 Involvement of the Inducible Proto-Oncogenes fos and jun. *Annual Review of  
651 Neuroscience* **14**, 421-451 (1991).

652 27. E.-L. Yap, M. E. Greenberg, Activity-Regulated Transcription: Bridging the Gap between  
653 Neural Activity and Behavior. *Neuron* **100**, 330-348 (2018).

654 28. Y. Alonso-Caraballo, S. K. Guha, E. H. Chartoff, The neurobiology of abstinence-induced  
655 reward-seeking in males and females. *Pharmacology Biochemistry and Behavior* **200**,  
656 173088 (2021).

657 29. X. Li, D. Caprioli, N. J. Marchant, Recent updates on incubation of drug craving: a mini-  
658 review. *Addiction Biology* **20**, 872-876 (2015).

659 30. A. Erturk *et al.*, Three-dimensional imaging of solvent-cleared organs using 3DISCO. *Nat  
660 Protoc* **7**, 1983-1995 (2012).

661 31. A. Erturk *et al.*, Three-dimensional imaging of the unsectioned adult spinal cord to  
662 assess axon regeneration and glial responses after injury. *Nat Med* **18**, 166-171 (2011).

663 32. E. Lee *et al.*, ACT-PRESTO: Rapid and consistent tissue clearing and labeling method for  
664 3-dimensional (3D) imaging. *Sci Rep* **6**, 18631 (2016).

665 33. N. Renier *et al.*, iDISCO: a simple, rapid method to immunolabel large tissue samples for  
666 volume imaging. *Cell* **159**, 896-910 (2014).

667 34. E. A. Susaki *et al.*, Whole-brain imaging with single-cell resolution using chemical  
668 cocktails and computational analysis. *Cell* **157**, 726-739 (2014).

669 35. H. R. Ueda *et al.*, Tissue clearing and its applications in neuroscience. *Nat Rev Neurosci*  
670 **21**, 61-79 (2020).

671 36. R. M. Power, J. Huisken, A guide to light-sheet fluorescence microscopy for multiscale  
672 imaging. *Nat Methods* **14**, 360-373 (2017).

673 37. E. G. Reynaud, U. Krzic, K. Greger, E. H. Stelzer, Light sheet-based fluorescence  
674 microscopy: more dimensions, more photons, and less photodamage. *HFSP J* **2**, 266-275  
675 (2008).

676 38. D. Fürth *et al.*, An interactive framework for whole-brain maps at cellular resolution.  
677 *Nature Neuroscience* **21**, 139-149 (2018).

678 39. C. Kirst *et al.*, Mapping the Fine-Scale Organization and Plasticity of the Brain  
679 Vasculature. *Cell* **180**, 780-795.e725 (2020).

680 40. N. Renier *et al.*, Mapping of Brain Activity by Automated Volume Analysis of Immediate  
681 Early Genes. *Cell* **165**, 1789-1802 (2016).

682 41. M. Jin *et al.*, SMART: An Open-Source Extension of WholeBrain for Intact Mouse Brain  
683 Registration and Segmentation. *eneuro* **9**, ENEURO.0482-0421.2022 (2022).

684 42. D. J. Calu, Y.-W. Chen, A. B. Kawa, S. G. Nair, Y. Shaham, The use of the reinstatement  
685 model to study relapse to palatable food seeking during dieting. *Neuropharmacology* **76**,  
686 395-406 (2014).

687 43. S. A. Golden *et al.*, Compulsive Addiction-like Aggressive Behavior in Mice. *Biological  
688 Psychiatry* **82**, 239-248 (2017).

689 44. S. A. Golden *et al.*, Persistent conditioned place preference to aggression experience in  
690 adult male sexually-experienced CD-1 mice. *Genes, Brain and Behavior* **16**, 44-55 (2017).

691 45. M. Venniro, M. Zhang, Y. Shaham, D. Caprioli, Incubation of methamphetamine but not  
692 heroin craving after voluntary abstinence in male and female rats.  
693 *Neuropsychopharmacology* **42**, 1126-1135 (2017).

694 46. D. J. Reiner *et al.*, Role of projections between piriform cortex and orbitofrontal cortex  
695 in relapse to fentanyl seeking after palatable food choice-induced voluntary abstinence.  
696 *J Neurosci* **40**, 2485-2497 (2020).

697 47. Y. Kim *et al.*, Mapping Social Behavior-Induced Brain Activation at Cellular Resolution in  
698 the Mouse. *Cell Reports* **10**, 292-305 (2015).

699 48. P. A. Yushkevich *et al.*, User-guided 3D active contour segmentation of anatomical  
700 structures: Significantly improved efficiency and reliability. *NeuroImage* **31**, 1116-1128  
701 (2006).

702 49. J. Schindelin *et al.*, Fiji: an open-source platform for biological-image analysis. *Nature  
703 Methods* **9**, 676-682 (2012).

704 50. Q. Wang *et al.*, The Allen Mouse Brain Common Coordinate Framework: A 3D Reference  
705 Atlas. *Cell* **181**, 936-953.e920 (2020).

706 51. L. A. Ramsey, F. M. Holloman, B. T. Hope, Y. Shaham, M. Venniro, Waving Through the  
707 Window: A Model of Volitional Social Interaction in Female Mice. *Biol Psychiatry* **91**,  
708 988-997 (2022).

709 52. S. A. Golden, M. Jin, Y. Shaham, Animal Models of (or for) Aggression Reward, Addiction,  
710 and Relapse: Behavior and Circuits. *J Neurosci* **39**, 3996-4008 (2019).

711 53. S. A. Golden *et al.*, Nucleus Accumbens Drd1-Expressing Neurons Control Aggression  
712 Self-Administration and Aggression Seeking in Mice. *J Neurosci* **39**, 2482-2496 (2019).

713 54. S. A. Golden *et al.*, Persistent conditioned place preference to aggression experience in  
714 adult male sexually-experienced CD-1 mice. *Genes Brain Behav* **16**, 44-55 (2017).

715 55. L. J. Sittig *et al.*, Genetic Background Limits Generalizability of Genotype-Phenotype  
716 Relationships. *Neuron* **91**, 1253-1259 (2016).

717 56. N. Wang *et al.*, Variability and heritability of mouse brain structure: Microscopic MRI  
718 atlases and connectomes for diverse strains. *NeuroImage* **222**, 117274 (2020).

719 57. H. E. Sloin *et al.*, Hybrid offspring of C57BL/6J mice exhibit improved properties for  
720 neurobehavioral research. *bioRxiv* 10.1101/2022.05.03.490527, 2022.2005.2003.490527  
721 (2022).

722 58. I. R. Davis, S. A. Coldren, X. Li, Methamphetamine seeking after prolonged abstinence is  
723 associated with activated projections from anterior intralaminar nucleus of thalamus to  
724 dorsolateral striatum in female rats. *Pharmacol Biochem Behav* **200**, 173087 (2021).

725 59. D. Funk *et al.*, Role of Central Amygdala Neuronal Ensembles in Incubation of Nicotine  
726 Craving. *J Neurosci* **36**, 8612-8623 (2016).

727 60. J. W. Grimm *et al.*, Effects of acute or chronic environmental enrichment on regional Fos  
728 protein expression following sucrose cue-reactivity testing in rats. *Brain Struct Funct*  
729 **221**, 2817-2830 (2016).

730 61. X. Li *et al.*, Role of Anterior Intralaminar Nuclei of Thalamus Projections to Dorsomedial  
731 Striatum in Incubation of Methamphetamine Craving. *J Neurosci* **38**, 2270-2282 (2018).

732 62. S. Fanous *et al.*, Role of orbitofrontal cortex neuronal ensembles in the expression of  
733 incubation of heroin craving. *J Neurosci* **32**, 11600-11609 (2012).

734 63. M. Venniro *et al.*, Abstinence-dependent dissociable central amygdala microcircuits  
735 control drug craving. *Proc Natl Acad Sci U S A* **117**, 8126-8134 (2020).

736 64. M. Venniro *et al.*, Volitional social interaction prevents drug addiction in rat models. *Nat  
737 Neurosci* **21**, 1520-1529 (2018).

738 65. A. Kimbrough *et al.*, Brain-wide functional architecture remodeling by alcohol  
739 dependence and abstinence. *Proceedings of the National Academy of Sciences* **117**,  
740 2149-2159 (2020).

741 66. A. Kimbrough *et al.*, Characterization of the Brain Functional Architecture of  
742 Psychostimulant Withdrawal Using Single-Cell Whole-Brain Imaging. *eneuro* **8**,  
743 ENEURO.0208-0219.2021 (2021).

744 67. R. D. Altshuler *et al.*, Role of orbitofrontal cortex in incubation of oxycodone craving in  
745 male rats. *Addict Biol* **26**, e12927 (2021).

746 68. C. A. Blackwood, M. Leary, A. Salisbury, M. T. McCoy, J. L. Cadet, Escalated Oxycodone  
747 Self-Administration Causes Differential Striatal mRNA Expression of FGFs and IEGs  
748 Following Abstinence-Associated Incubation of Oxycodone Craving. *Neuroscience* **415**,  
749 173-183 (2019).

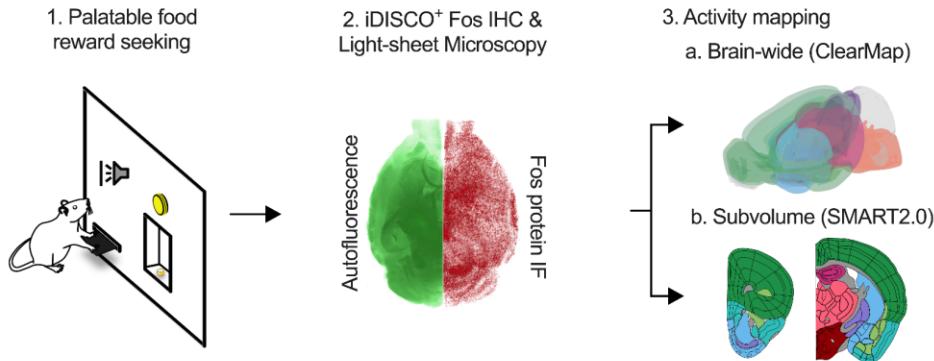
750 69. L. M. Rossi *et al.*, Role of nucleus accumbens core but not shell in incubation of  
751 methamphetamine craving after voluntary abstinence. *Neuropsychopharmacology* **45**,  
752 256-265 (2020).

753

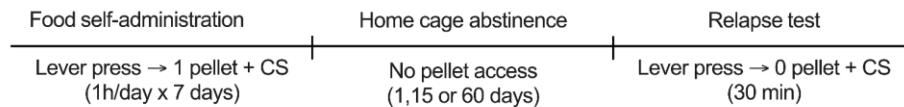
754 **Figures and Tables**

**Figure 1**

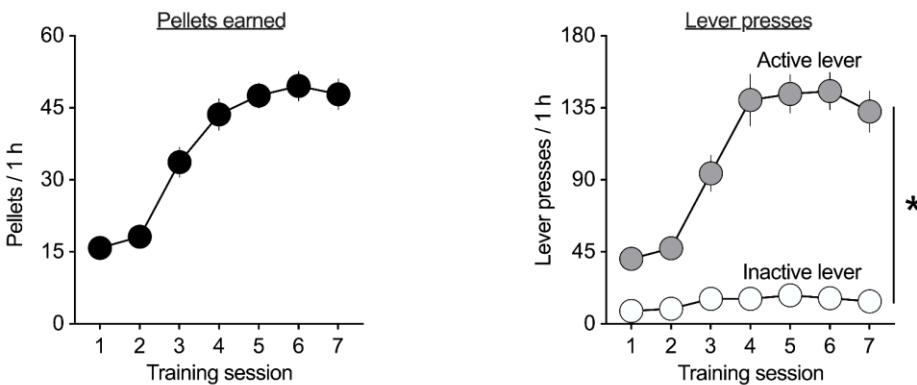
**A. Experimental overview**



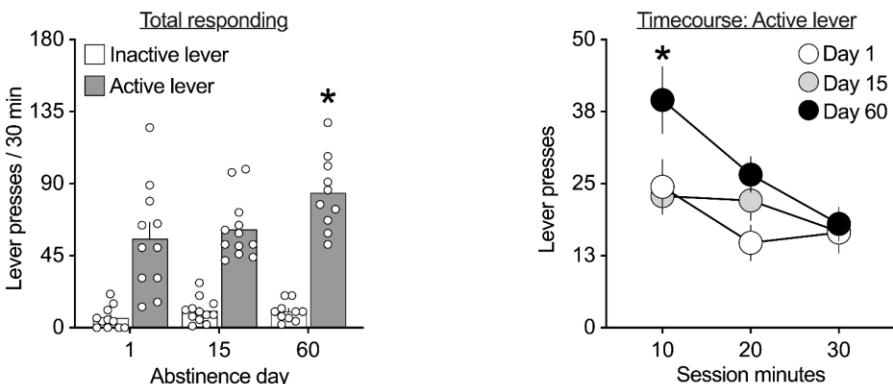
**B. Behavioral timeline**



**C. Food self-administration**



**D. Relapse (incubation) test**

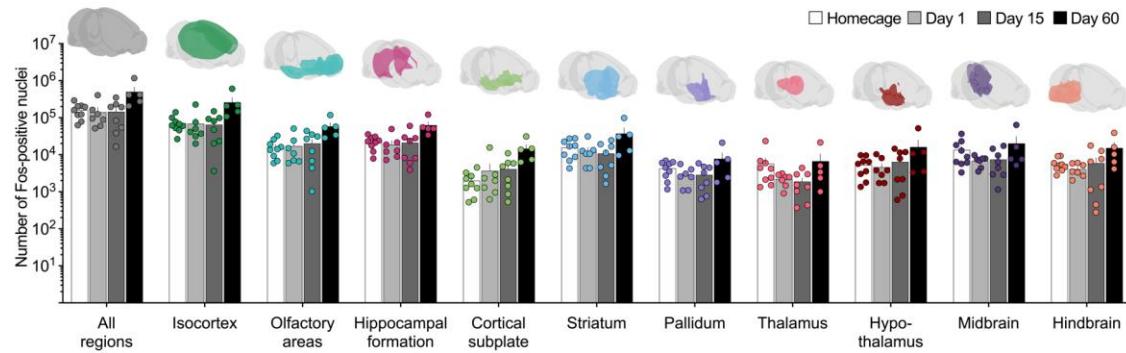


756 **Figure 1.** Incubation of palatable food seeking in male CD1 mice.

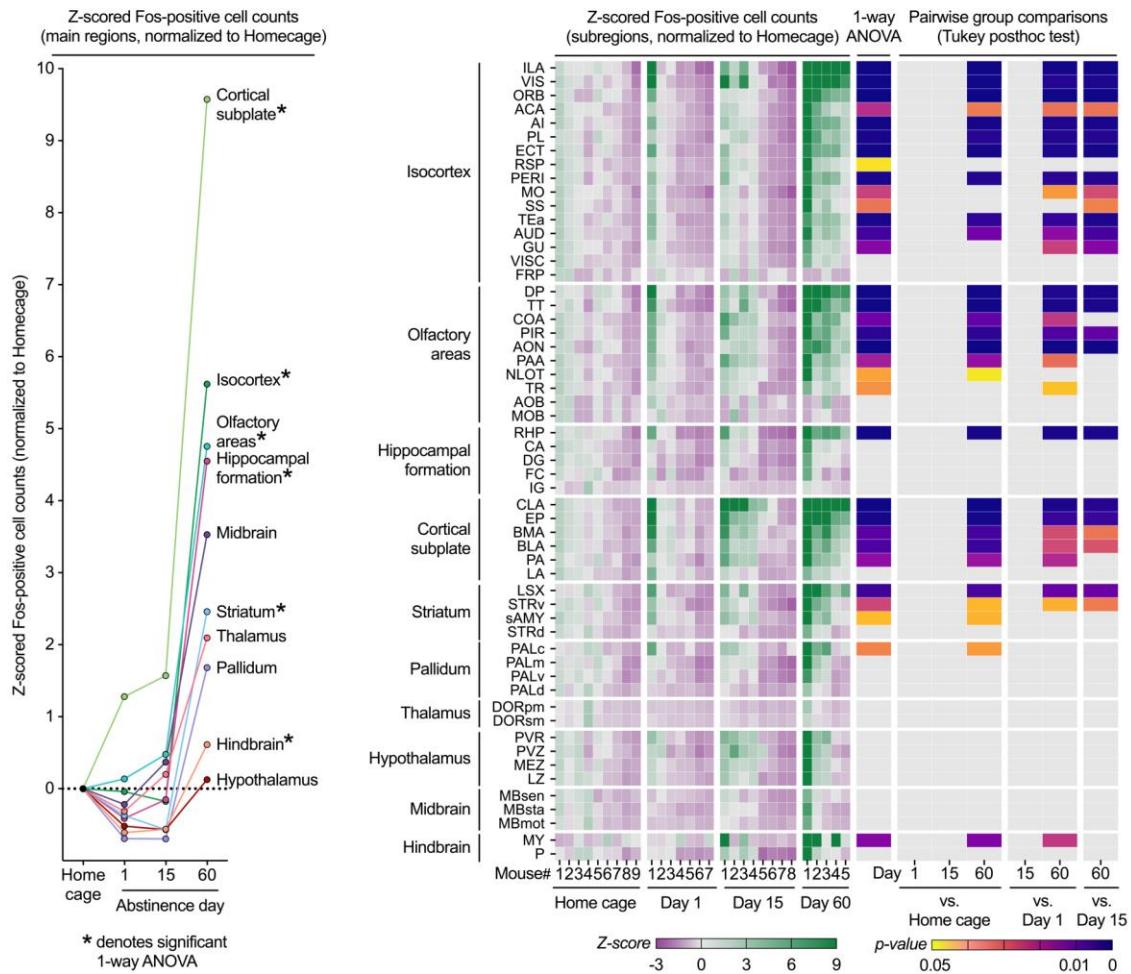
757 **(A)** Experimental overview. **(B)** Timeline of food self-administration training, abstinence, and  
758 relapse tests. **(C)** Food self-administration training. Mice learned to self-administer palatable food  
759 pellets over 7 sessions. Mean ( $\pm$ SEM) number of food pellets earned (left panel) and lever  
760 presses (right panel) during each 1-h session. \* Significant difference ( $p<0.05$ ) between active  
761 and inactive lever presses ( $n=46$ ). **(D)** Relapse (incubation) test. Responding on active but not  
762 inactive lever progressively increased during abstinence. Mean ( $\pm$ SEM) number of lever presses  
763 during the entire 30-min relapse test session (left panel) and binned 10-min timecourse of active  
764 lever presses (right panel). \* Significant difference ( $p<0.05$ ) from day 1. See [Supplementary](#)  
765 [Table S3](#) for a detailed listing of all statistical outputs relating to this figure.

Figure 2

A. Fos-positive cell counts across major anatomical sub-divisions



B. Brain-wide changes in activation during abstinence (counts z-scored normalized to Home cage distribution)



766

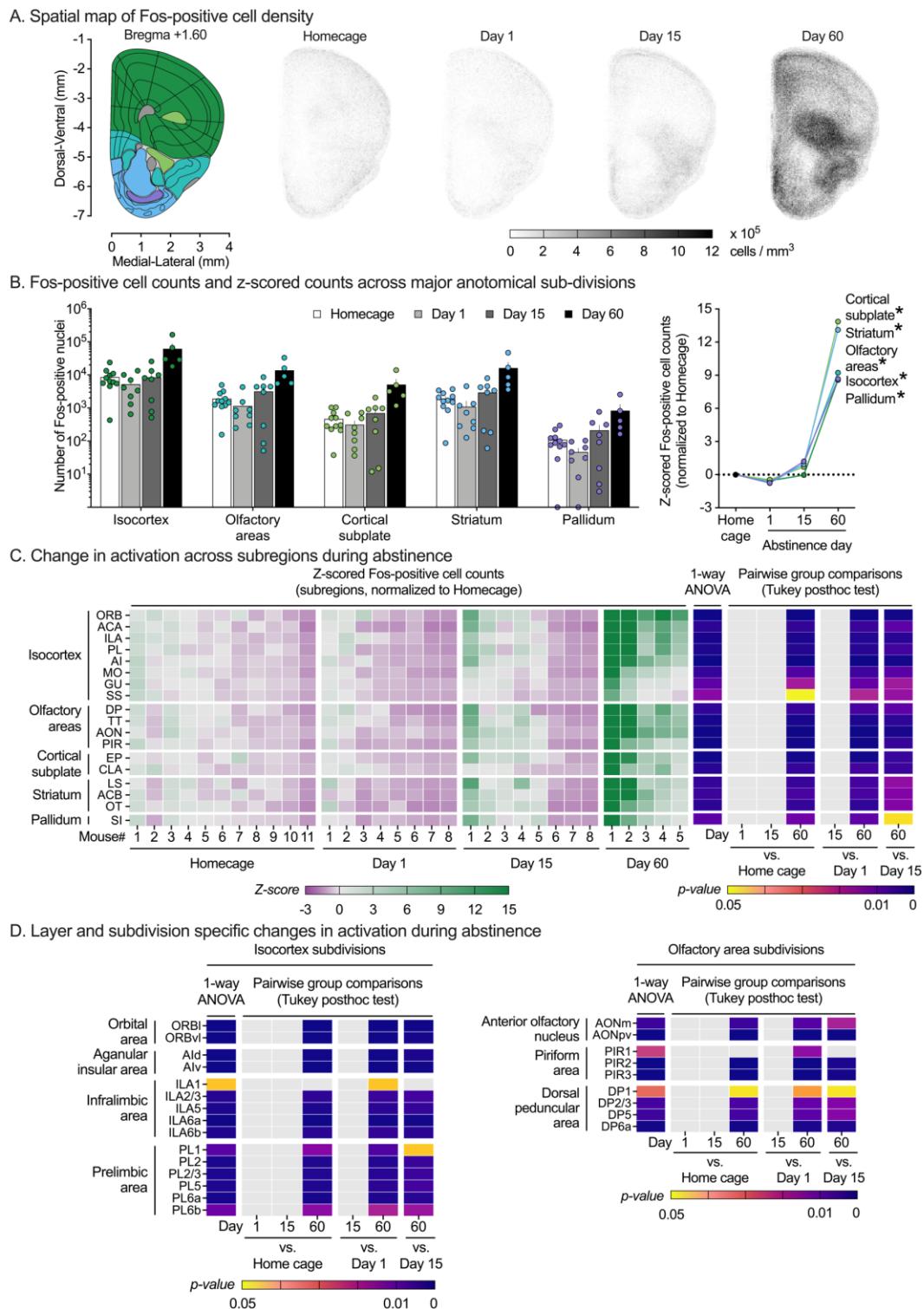
767

768 **Figure 2.** Unbiased brain-wide activity mapping of incubated relapse to palatable food-seeking  
769 using ClearMap.

770 (A) Mean ( $\pm$ SEM) Fos+ cell counts across the whole brain and for 10 major anatomical sub-  
771 divisions. (B) Brain-wide changes in activation during abstinence. Raw Fos+ cell counts for each  
772 region are z-score normalized to the homecage group distribution for statistical analysis. Mean z-  
773 scored cell counts for 10 major anatomical sub-divisions showing time-dependent changes in  
774 activation pattern in multiple brain regions induced by the relapse tests (left panel). \* Significant  
775 differences ( $p < 0.05$ ) 1-way ANOVA. Heatmap of individual z-scored Fos+ cell counts, 1-way  
776 ANOVA p-value and Tukey HSD pairwise group comparison p-values for 56 subregions across  
777 the analyzed brain volume (right panel). Individual data is sorted by group and ranked in  
778 descending order of activation level within each group. Subregions are organized by 10 parent  
779 anatomical sub-divisions and ranked in descending order of mean day 60 group activation level.  
780 See [Supplementary Tables S4-S5](#) for a detailed listing of all statistical results associated with this  
781 figure.

782

Figure 3

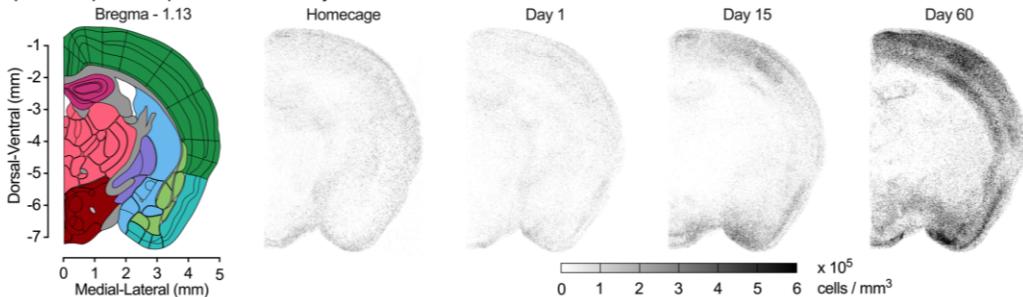


784 **Figure 3.** Targeted analysis of corticostriatal coronal subvolume (AP +1.55 to AP +1.75 relative to  
785 Bregma) using SMART2

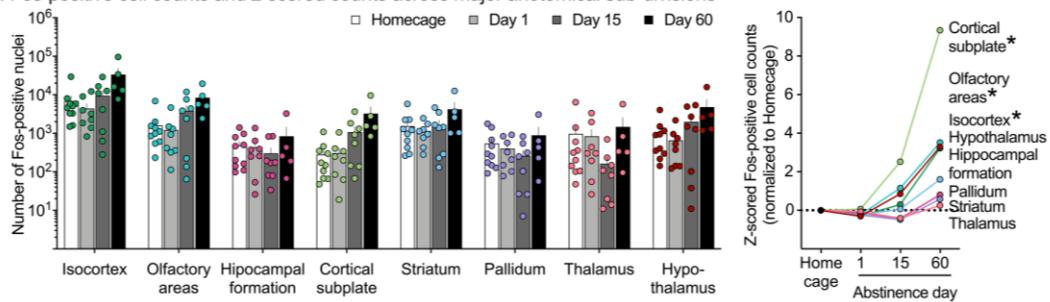
786 (A) Spatial map of Fos+ cell density. Grayscale intensity of individual points (20  $\mu\text{m}$  x 20  $\mu\text{m}$  x  
787 200  $\mu\text{m}$  voxel) represents mean cell density in cells/mm<sup>3</sup> within AP +1.55 to AP +1.75 coronal  
788 subvolume. (B, left) Fos+ cell counts for 5 major anatomical regions within the subvolume. (B,  
789 right) Z-score normalized counts (normalized to home cage group distribution) for 5 major  
790 anatomical regions within the subvolume. \* Significant differences (p<0.05) 1-way ANOVAs. (C)  
791 Changes in activation across the subvolume during abstinence. Fos+ cell counts for each region  
792 are z-score normalized to home cage group distribution for statistical analysis. Heatmap of  
793 individual z-scored Fos+ cell counts (left), 1-way ANOVA p-value (middle) and Tukey HSD  
794 pairwise group comparisons p-values (right) for 18 sub-regions within the analyzed coronal  
795 subvolume. Subregions are organized by 5 parent anatomical sub-divisions and ranked in  
796 descending order of mean day 60 group activation level. (D) Layer and subdivision specific  
797 changes in activation. Heatmap of 1-way ANOVA p-value and Tukey HSD pairwise group  
798 comparison p-values for selected sub-region layers or subdivisions within Isocortex (left) or  
799 Olfactory areas (right). See [Supplementary Tables S6-S8](#) for a detailed listing of all statistical  
800 results associated with this figure.

Figure 4

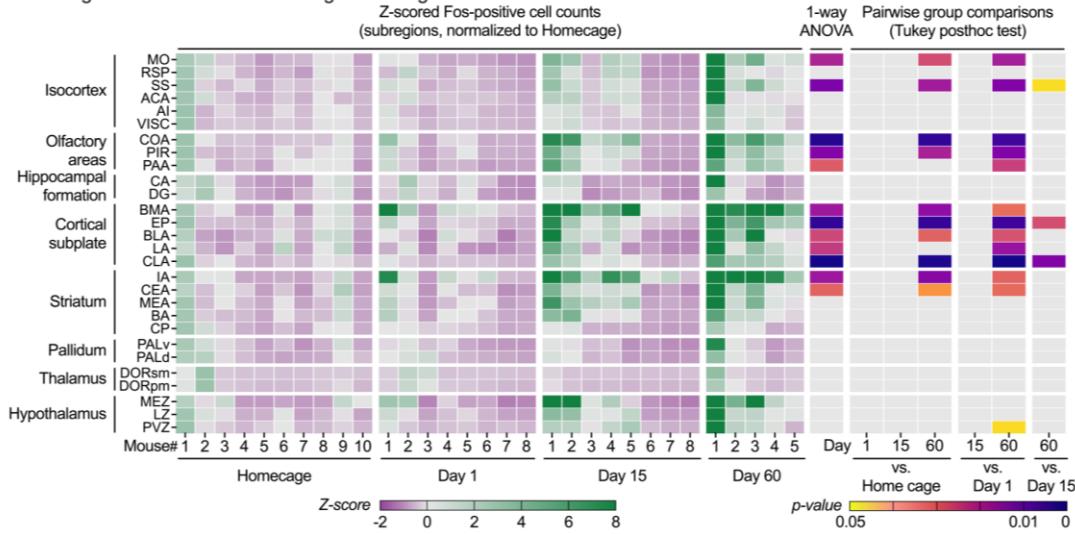
A. Spatial map of Fos-positive cell density



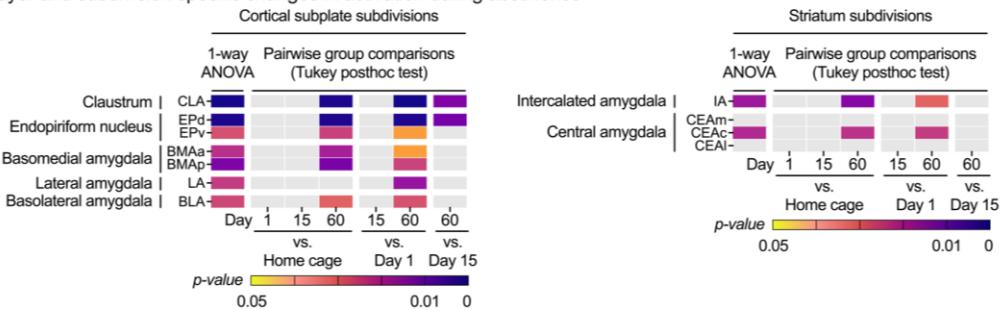
B. Fos-positive cell counts and z-scored counts across major anatomical sub-divisions



C. Change in activation across subregions during abstinence



D. Layer and subdivision specific changes in activation during abstinence



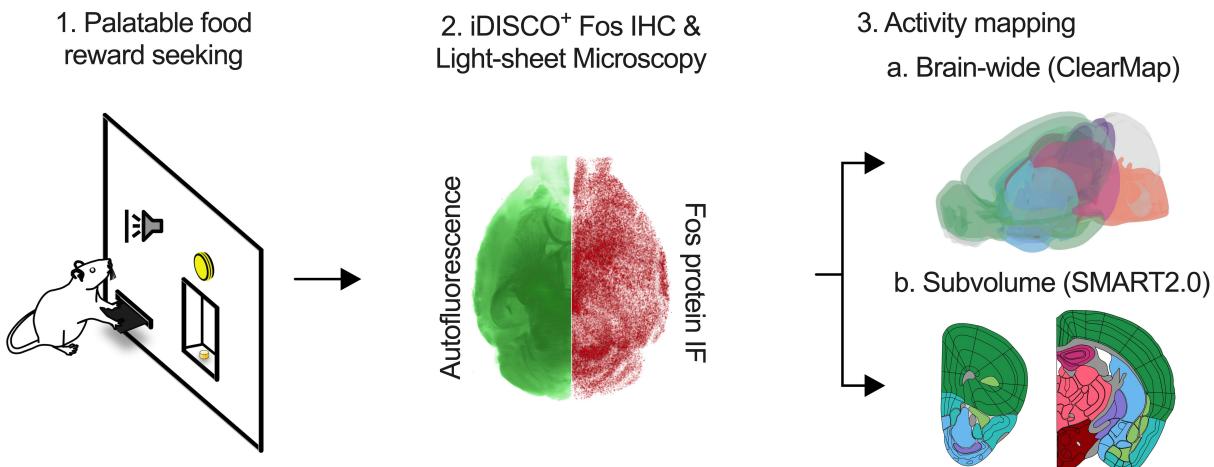
802 **Figure 4.** Targeted analysis of thalamocortical coronal subvolume (AP -1.08 to AP -1.28 relative  
803 to Bregma) using SMART2

804 (A) Spatial map of Fos+ cell density. Grayscale intensity of individual points (20  $\mu\text{m}$  x 20  $\mu\text{m}$  x  
805 200  $\mu\text{m}$  voxel) represents mean cell density in cells/mm<sup>3</sup> within AP -1.08 to AP -1.28 coronal  
806 subvolume. (B, left) Fos+ cell counts for 8 major anatomical regions within the subvolume. (B,  
807 right) Z-score normalized counts (normalized to home cage group distribution) for 5 major  
808 anatomical regions within the subvolume. \* Significant differences ( $p<0.05$ ) 1-way ANOVAs. (C)  
809 Changes in activation across the subvolume. Fos+ cell counts for each region are z-score  
810 normalized to home cage group distribution for statistical analysis. Heatmap of individual z-scored  
811 Fos+ cell counts (left), 1-way ANOVAs p-values (middle) and Tukey HSD pairwise group  
812 comparisons p-values (right) for 28 sub-regions within the analyzed coronal subvolume.  
813 Subregions are organized by 8 parent anatomical subdivisions and ranked in descending order of  
814 mean day 60 group activation level. (D) Layer and subdivision specific changes in activation.  
815 Heatmap of 1-way ANOVAs p-values and Tukey HSD pairwise group comparison p-values for  
816 selected subregion layers or subdivisions within Cortical subplate (left) or Striatum (right). See  
817 [Supplementary Tables S9-S11](#) for a detailed listing of all statistical results related to this figure.

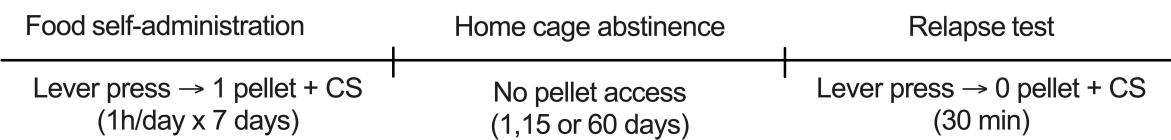
818

## Figure 1

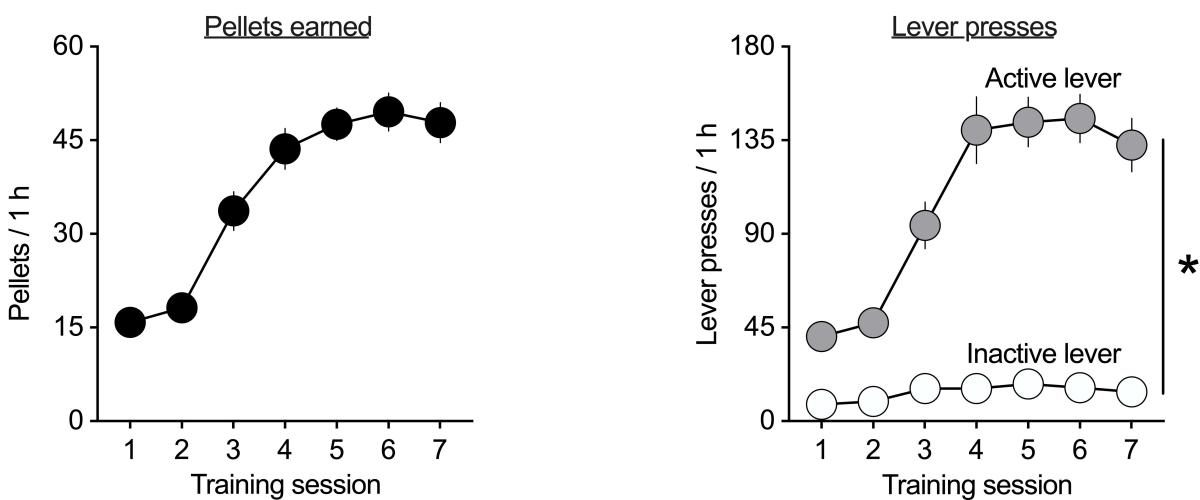
### A. Experimental overview



### B. Behavioral timeline



### C. Food self-administration



### D. Relapse (incubation) test

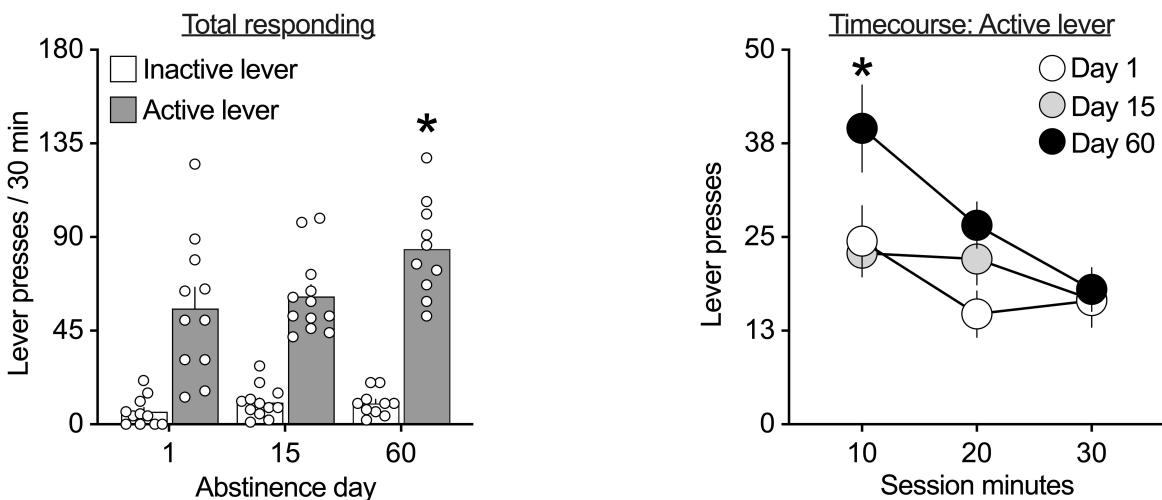
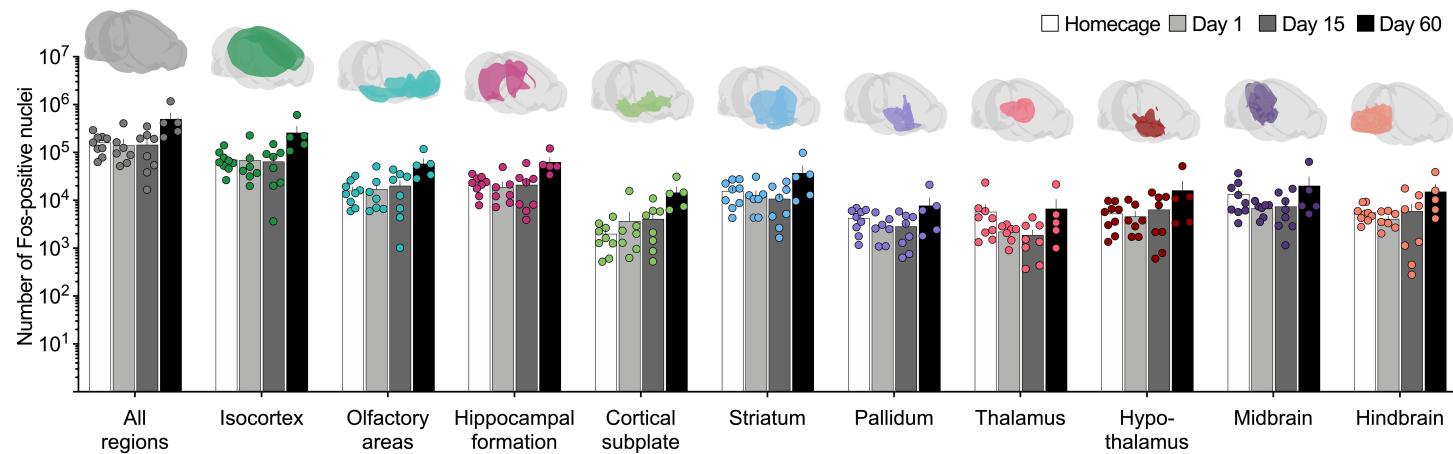


Figure 2

A. Fos-positive cell counts across major anatomical sub-divisions



B. Brain-wide changes in activation during abstinence (counts z-scored normalized to Home cage distribution)

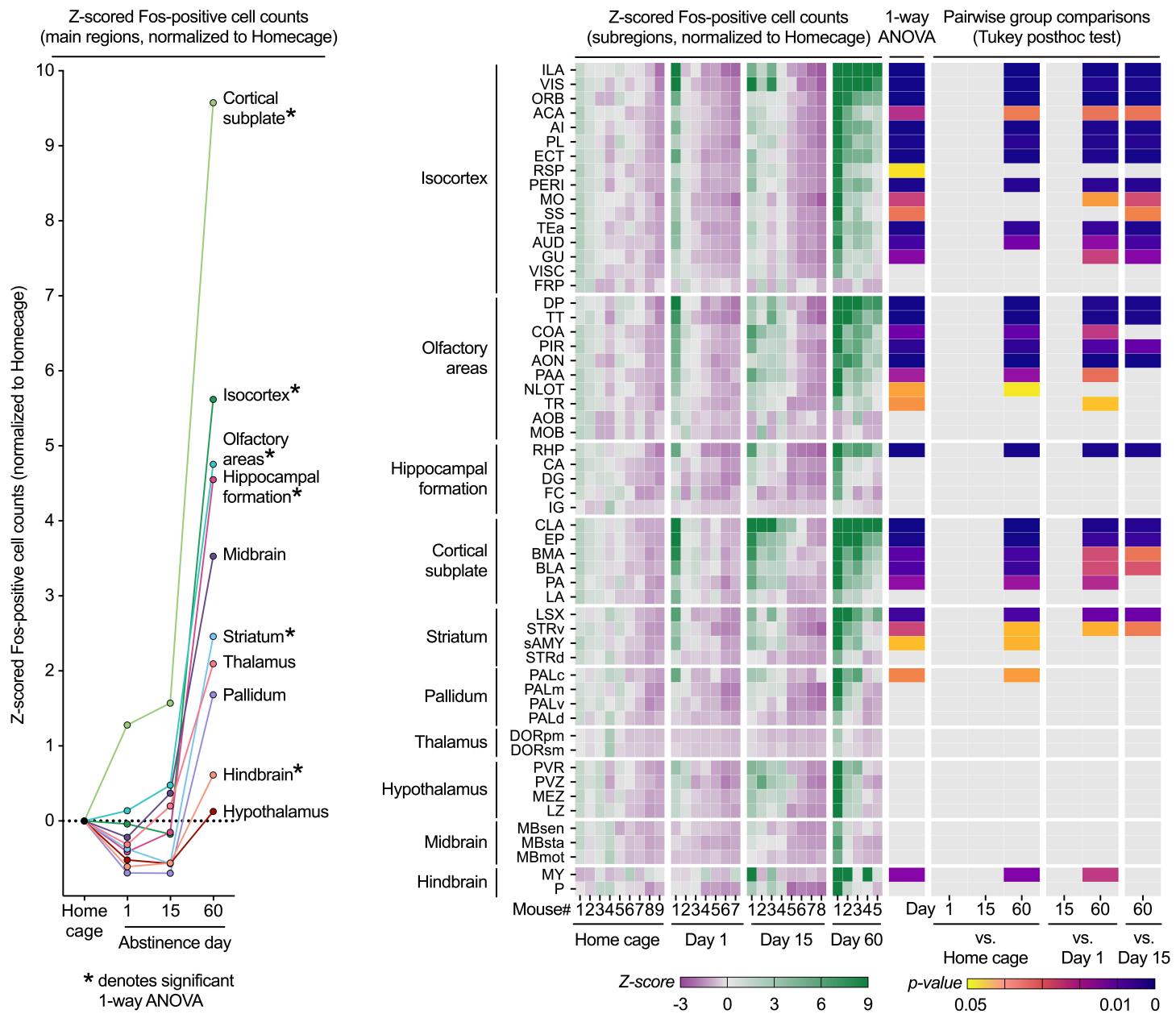
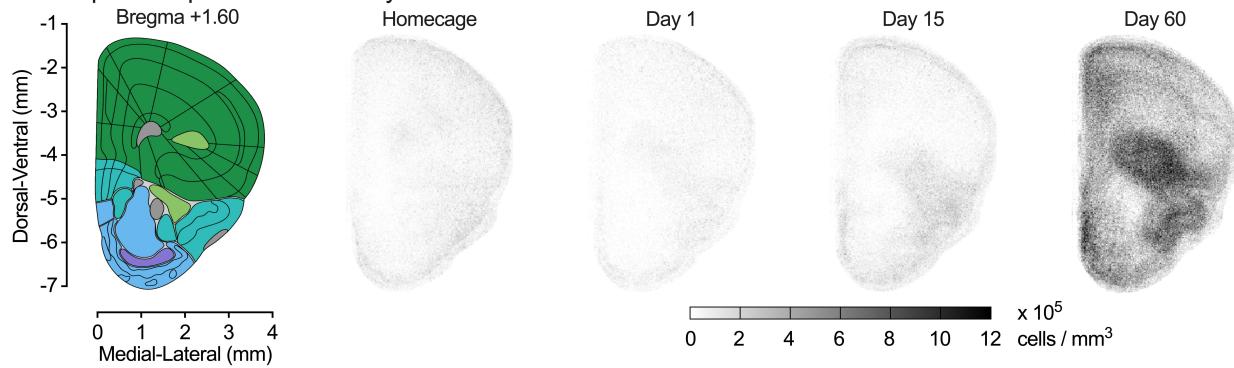
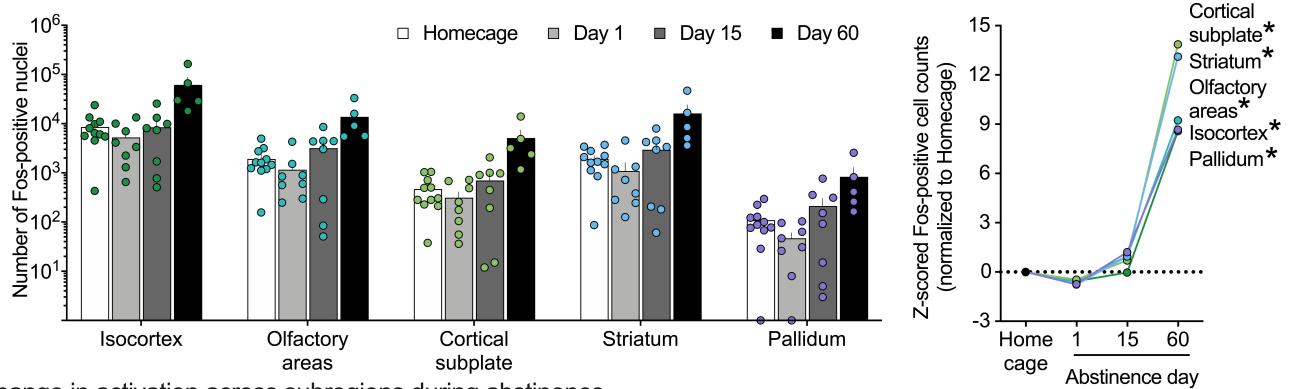


Figure 3

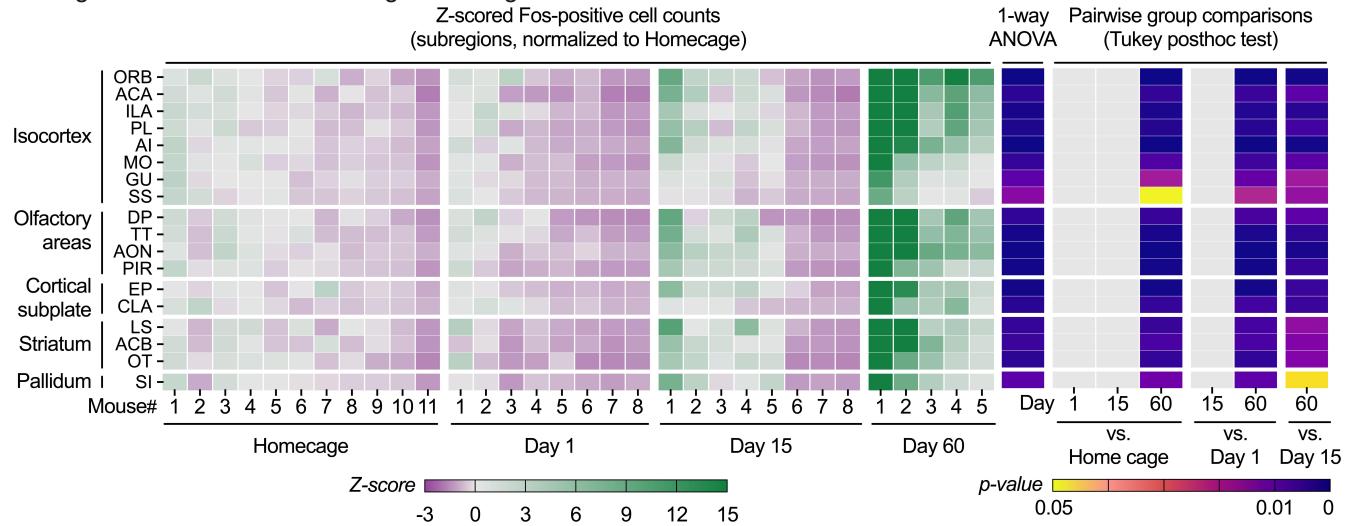
A. Spatial map of Fos-positive cell density



B. Fos-positive cell counts and z-scored counts across major anatomical sub-divisions



C. Change in activation across subregions during abstinence



D. Layer and subdivision specific changes in activation during abstinence

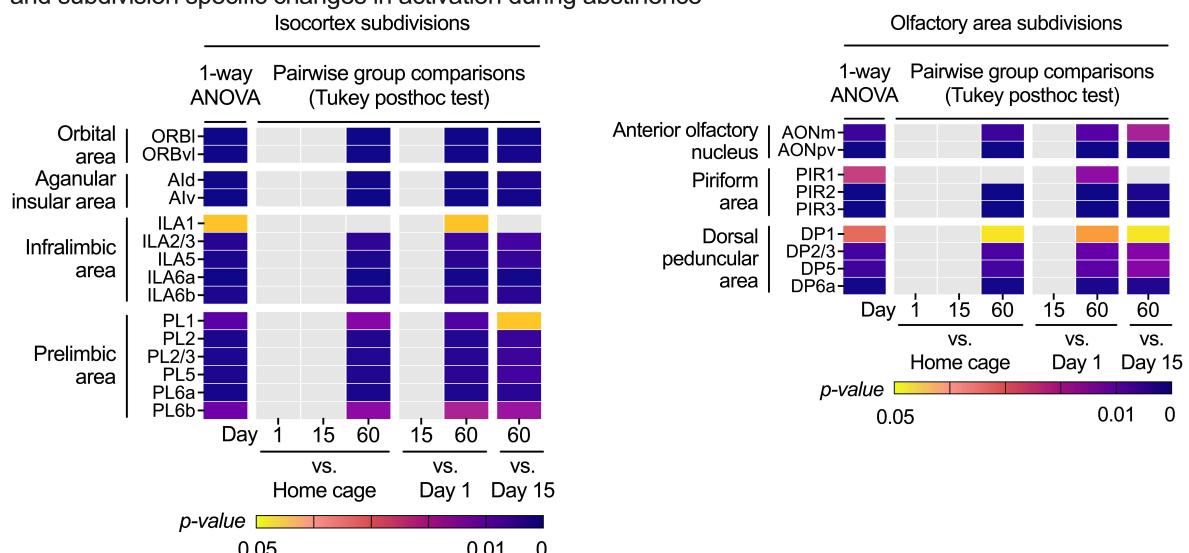
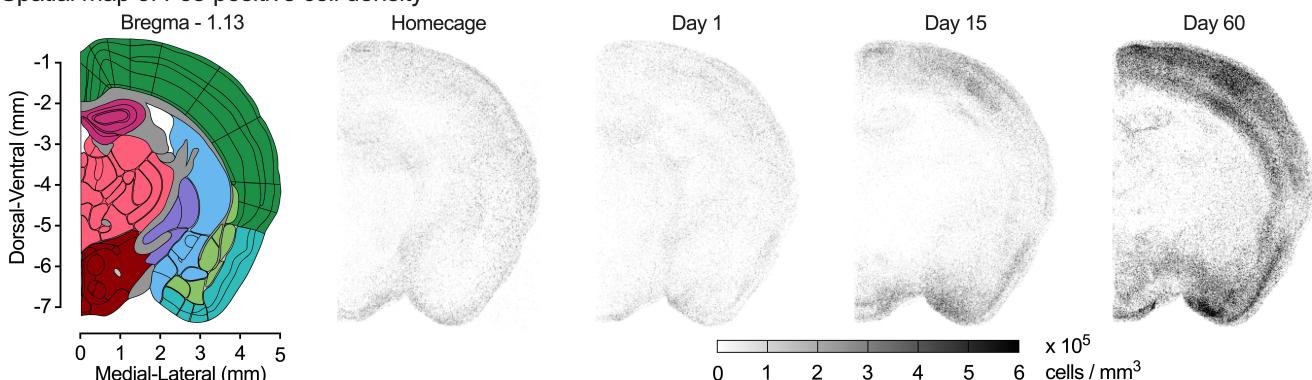
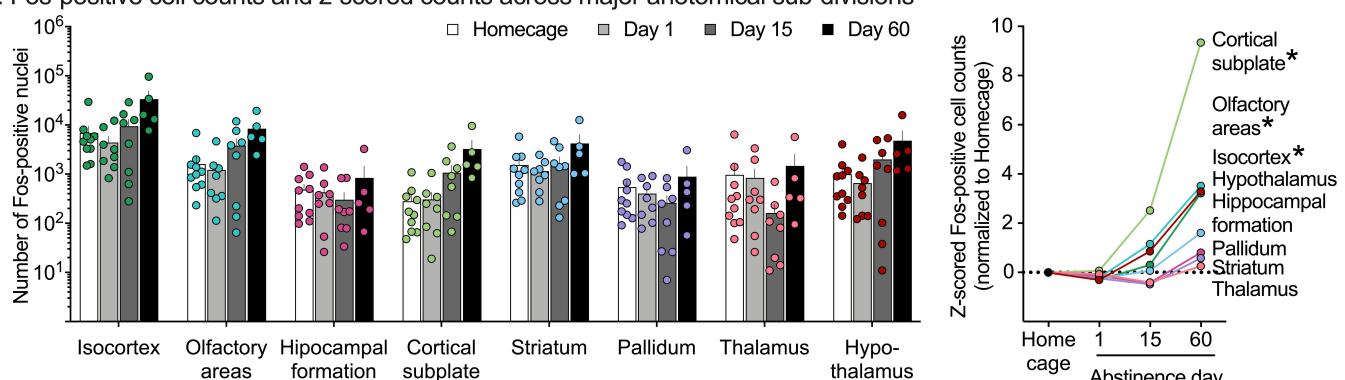


Figure 4

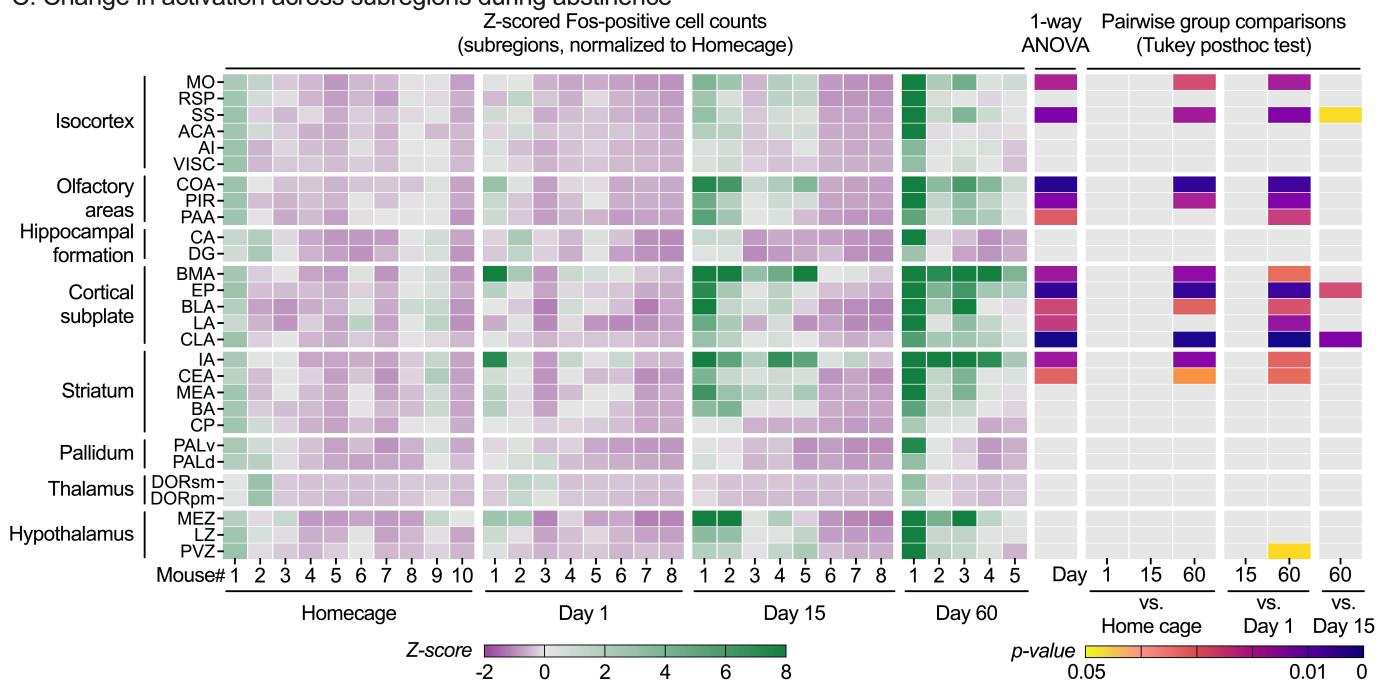
A. Spatial map of Fos-positive cell density



B. Fos-positive cell counts and z-scored counts across major anatomical sub-divisions



C. Change in activation across subregions during abstinence



D. Layer and subdivision specific changes in activation during abstinence

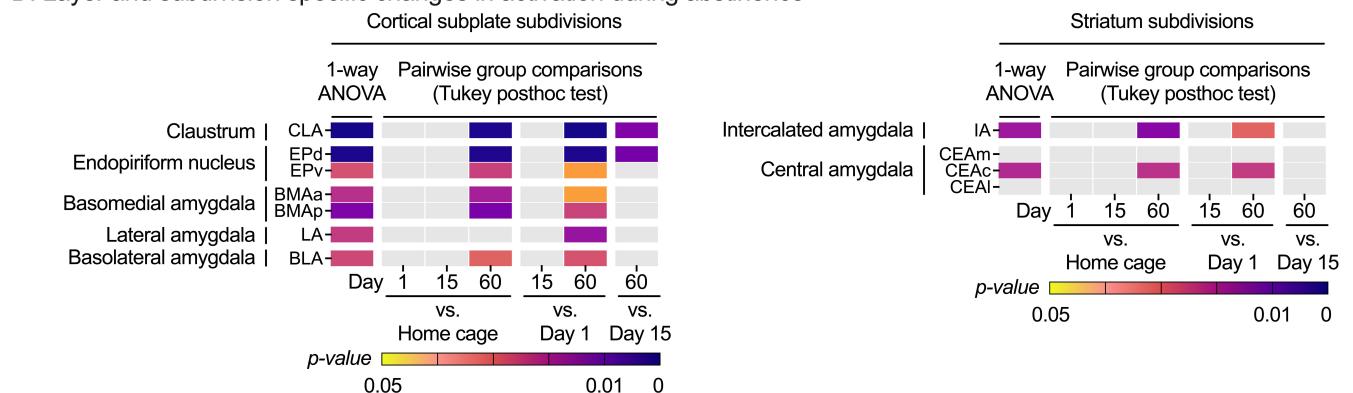
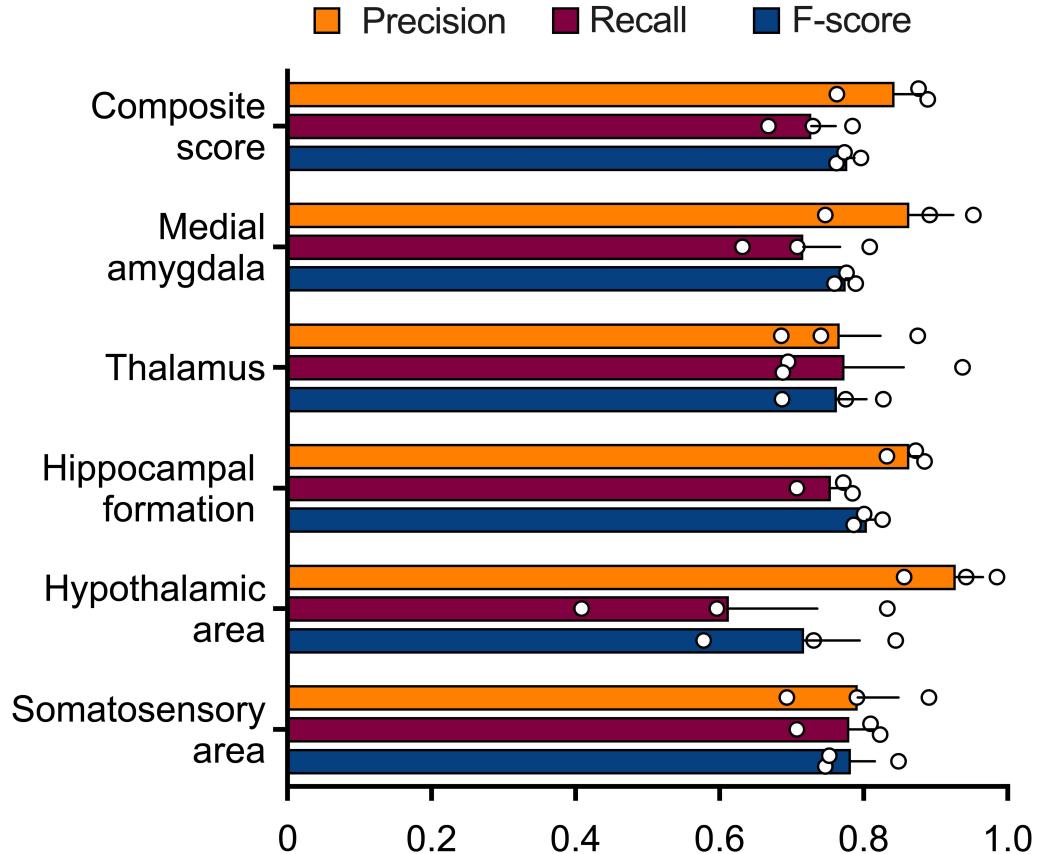
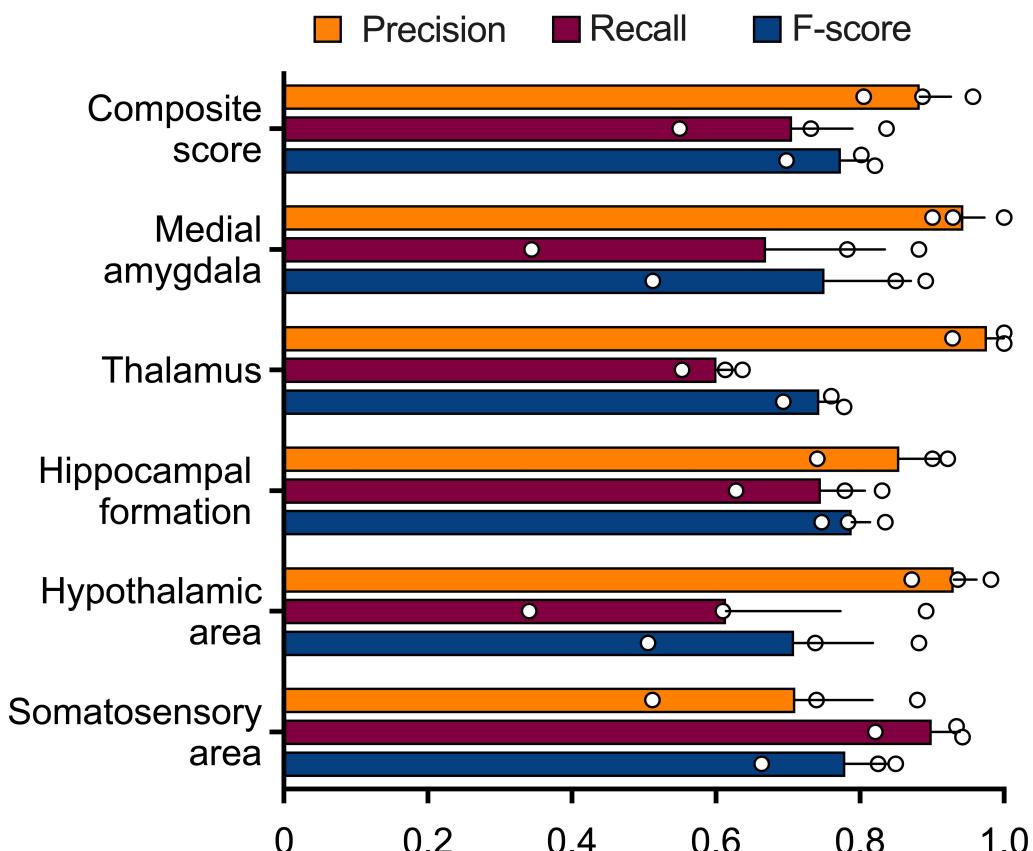


Figure S1

A. Ground-truth validation of ClearMap segmentation parameters



B. Ground-truth validation of SMART2.0 segmentation parameters



## Supplementary Information for

# Incubation of palatable food craving is associated with brain-wide neuronal activation in mice

Rajtarun Madangopal<sup>1†</sup>, Eric R. Szelenyi<sup>3,4†</sup>, Joseph Nguyen<sup>1</sup>, Megan B. Brenner<sup>1</sup>, Olivia R. Drake<sup>1</sup>, Diana Pham<sup>1</sup>, Aniruddha Shekara<sup>1</sup>, Michelle Jin<sup>1</sup>, Jia Jie Choong<sup>2,3,4</sup>, Connor Heins<sup>1</sup>, Lauren E. Komer<sup>1</sup>, Sophia J. Weber<sup>1</sup>, Bruce T. Hope<sup>1</sup>, Yavin Shaham<sup>1</sup>, Sam A. Golden<sup>3,4,\*</sup>

<sup>1</sup> Intramural Research Program, National Institute on Drug Abuse, Baltimore, MD, USA.

<sup>2</sup> University of Washington, Department of Electrical and Computer Engineering, Seattle, Washington, USA

<sup>3</sup> Department of Biological Structure, University of Washington, Seattle, WA, USA.

<sup>4</sup> University of Washington, Center of Excellence in Neurobiology of Addiction, Pain, and Emotion (NAPE), Seattle, Washington, USA

† These authors contributed equally

**\*Correspondence to:** Sam A. Golden, Ph.D., Department of Biological Structure, University of Washington, Seattle, WA 98195

**Email:** sagolden@uw.edu

### This PDF file includes:

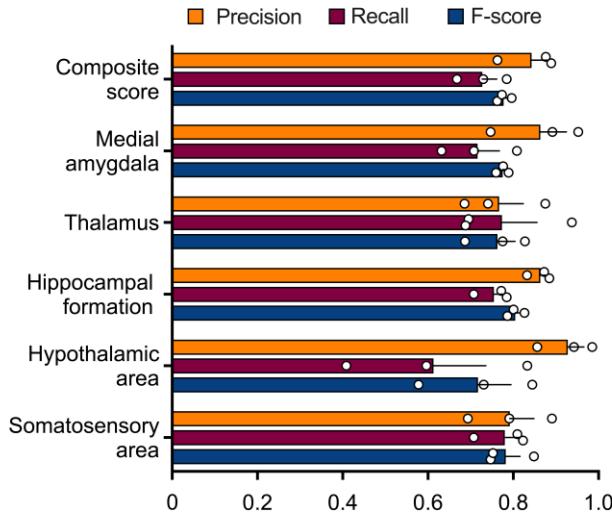
Figure S1

Tables S1 to S11

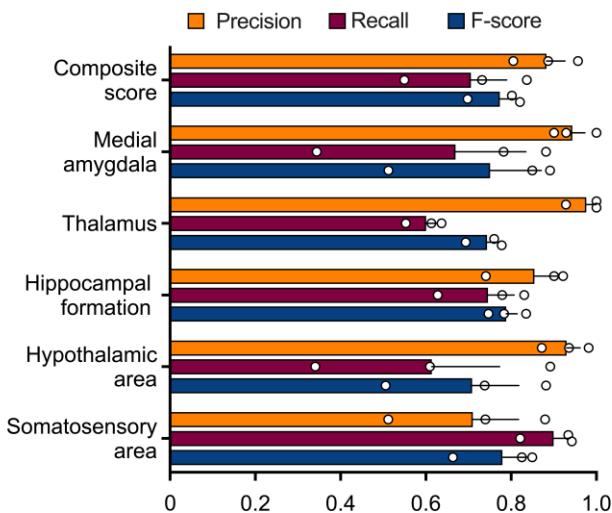
SI References

Figure S1

A. Ground-truth validation of ClearMap segmentation parameters



B. Ground-truth validation of SMART2.0 segmentation parameters



**Fig. S1.** Validation of ClearMap (A) and SMART2 (B) segmentation parameters against expert Fos+ cell annotation

**Table S1. Incubation-relevant regions identified using targeted section-based activity mapping approaches**

Brain Region	Abbreviation	Reward	Animal	Model	Abstinence duration (days)	Change in activation (Fos)			Citation
						Incubation	Seeking	Inactivation	
Ventral medial prefrontal cortex	vmPFC	Nicotine	Rat	SA	1, 7, 14, 28	—	↑	↑	(1)
	vmPFC	Methamphetamine	Rat	SA	1, 15	↑	↑	↑	(2)
	vmPFC	Methamphetamine	Rat	SA	1, 30	—	↑	↑	(3)
Prelimbic cortex	PL	Sucrose	Rat	SA	1, 30	↑	↑(CM), ↑(SMART)	↑(CM), ↑(SMART)	(4)
	PL	Pallatable food	Mouse	SA	1, 15, 60	↑(CM), ↑(SMART)	↑(CM), ↑(SMART)	↑(CM), ↑(SMART)	Current study
Infralimbic cortex	IL	Sucrose	Rat	SA	1, 30	↑	↑(CM), ↑(SMART)	↑(CM), ↑(SMART)	(4)
	IL	Pallatable food	Mouse	SA	1, 15, 60	↑(CM), ↑(SMART)	↑(CM), ↑(SMART)	↑(CM), ↑(SMART)	Current study
Dorsal medial prefrontal cortex	dmPFC	Nicotine	Rat	SA	1, 7, 14, 28	↑	↑	↑	(1)
	dmPFC	Methamphetamine	Rat	SA	1, 15	↑	↑	↑	(2)
	dmPFC	Methamphetamine	Rat	SA	1, 30	—	↑	↑	(3)
Orbitofrontal cortex	OFC	Oxycodone	Rat	SA	1, 15	—	↑	↑	(5)
	OFC	Methamphetamine	Rat	SA	1, 15	↑	↑	↑	(2)
	OFC	Methamphetamine	Rat	SA	1, 26	—	↑	↑	(6)
	OFC	Heroin	Rat	SA	1, 14	—	↑	↑	(7)
	OFC	Nicotine	Rat	SA	1, 7, 14, 28	↑	↑	↑	(1)
	OFC	Sucrose	Rat	SA	1, 30	—	↑(CM), ↑(SMART)	↑(CM), ↑(SMART)	(4)
	OFC	Pallatable food	Mouse	SA	1, 15, 60	↑(CM), ↑(SMART)	↑(CM), ↑(SMART)	↑(CM), ↑(SMART)	Current study
Anterior cingulate cortex	ACC	Methamphetamine	Rat	SA	1, 26	—	↑	↑	(6)
	ACC	Methamphetamine	Rat	SA	1, 15	↑	↑	↑	(2)
	ACC	Sucrose	Rat	SA	1, 30	↑	↑	↑	(4)
	ACC	Pallatable food	Mouse	SA	1, 15, 60	↑(CM), ↑(SMART)	↑(CM), ↑(SMART)	↑(CM), ↑(SMART)	Current study
Dorsal striatum	DS	Oxycodone	Rat	SA	5, 31	—	↑	↑	(8)
	DS	Methamphetamine	Rat	SA	2, 35	—	↑	↑	(9)
	DS	Pallatable food	Mouse	SA	1, 15, 60	—(CM)	—(CM)	—(CM)	Current study
Dorsomedial striatum	DMS	Methamphetamine	Rat	SA	1, 21	↑	↑	↑	(10)
Dorsolateral striatum	DLS	Methamphetamine	Rat	SA	1, 21	—	—	—	(10)
	DLS	Methamphetamine	Rat	SA	1, 26	—	↑	↑	(6)
	DLS	Sucrose	Rat	SA	1, 30	↑	↑	↑	(4)
	AI	Methamphetamine	Rat	SA	1, 26	—	↑	↑	(6)

Anterior insula cortex	AIV	Methamphetamine	Rat	SA	1, 15	↑	↑		(2)
	AI	Methamphetamine	Rat	SA	1, 30		↑		(3)
	AI	Pallatable food	Mouse	SA	1, 15, 60	↑(CM), ↑(SMART)	↑(CM), ↑(SMART)		Current study
Nucleus accumbens	NAc	Oxycodone	Rat	SA	5, 31		↓		(8)
	NAc	Pallatable food	Mouse	SA	1, 15, 60	↑(SMART)	↑(SMART)		Current study
Nucleus accumbens core	NAcC	Nicotine	Rat	SA	1, 7, 14, 28	↑			(1)
	NAcC	Sucrose	Rat	SA	1, 30	↑			(4)
	NAcC	Methamphetamine	Rat	SA	1, 15		↑	↓	(11)
Nucleus accumbens shell	NAcS	Nicotine	Rat	SA	1, 7, 14, 28	—			(1)
	NAcS	Sucrose	Rat	SA	1, 30	↑			(4)
	NAcS	Methamphetamine	Rat	SA	1, 15		—	—	(11)
Somatosensory cortex	SSC	Sucrose	Rat	SA	1, 30	↑			(4)
	SSC	Pallatable food	Mouse	SA	1, 15, 60	—(CM), ↑(SMART)	—(CM), ↑(SMART)		Current study
Medial anterior intralaminar nuclei of thalamus	AIT-M	Methamphetamine	Rat	SA	1, 26		↑		(6)
	AIT-M	Methamphetamine	Rat	SA	1, 30		—	—	(3)
Lateral anterior intralaminar nuclei of thalamus	AIT-L	Methamphetamine	Rat	SA	1, 26		↑		(6)
	AIT-L	Methamphetamine	Rat	SA	1, 30		↑	↓	(3)
Parafascicular nucleus	Pf	Methamphetamine	Rat	SA	1, 26		↑		(6)
	Pf	Methamphetamine	Rat	SA	1, 30		—		(3)
Basolateral amygdala	BLA	Methamphetamine	Rat	SA	1, 26		—	—	(6)
	BLA	Methamphetamine	Rat	SA	1, 15	↑	↑		(2)
	BLA	Nicotine	Rat	SA	1, 7, 14, 28	↑			(1)
	BLA	Sucrose	Rat	SA	1, 30	—			(4)
	BLA	Methamphetamine	Rat	SA	1, 30		↑		(3)
Central amygdala	BLA	Pallatable food	Mouse	SA	1, 15, 60	↑(CM), ↑(SMART)	↑(CM), ↑(SMART)		Current study
	CeA	Nicotine	Rat	SA	1, 7, 14, 28	↑		↓	(1)
	CeA	Methamphetamine	Rat	SA	1, 15	↑	↑		(2, 12)
	CeA	Sucrose	Rat	SA	1, 30	↑			(4)
Dentate Gyrus of hippocampus	DG	Sucrose	Rat	SA	1, 30	—	—(CM), —(SMART)	—(CM), —(SMART)	Current study
	DG	Pallatable food	Mouse	SA	1, 15, 60	—(CM), —(SMART)	—(CM), —(SMART)		(4)
									Current study

**Table S2. Number of subjects that completed each phase of the study**

<b>Experimental phase</b>	<b>Experimental group</b>			<b>Day 60</b>	<b>Total</b>	<b>Figure</b>
	<b>Homecage</b>	<b>Day 1</b>	<b>Day 15</b>			
Naïve (before start of training)	-	-	-	-	60	-
Food self-administration	-	-	-	-	46	1C
Food seeking & tissue collection	13	11	12	10	46	1D
iDISCO+ Fos IHC	11	9	9	8	37	-
Light sheet microscopy	11	8	8	5	32	-
ClearMap pipeline	9	7	8	5	29	2
SMART2.0 subvolume 1	11	8	8	5	32	3
SMART2.0 subvolume 2	10	8	8	5	31	4

**Table S3. Statistical output for Figure 1: (analyses shown in figure 1 are highlighted in grey)**

Experimental phase	Behavioral measure	Analysis method & factors	Statistical output			Figure	
Food self-administration training	Pellets earned over 1 h (n=46)	RM-ANOVA (Session x Group)	F-value	P-value	Partial $\eta^2$	1C, left	
		Session (within-subjects)	$F(3.079, 129.330) = 70.576$	<0.001	0.627		
		Group (between-subjects)	$F(3,42) = 1.165$	0.334	0.077		
		Session x Group	$F(9.238, 129.330) = 1.785$	0.075	0.133		
		RM-ANOVA (Session)	F-value	P-value	Partial $\eta^2$		
	Lever presses over 1 h (n=46)	Session (within-subjects)	$F(3.215, 144.687) = 67.173$	<0.001	0.599		
		RM-ANOVA (Lever x Session x Group)	F-value	P-value	Partial $\eta^2$	1C, right	
		Lever (within-subjects)	$F(1,42) = 136.117$	<0.001	0.764		
		Session (within-subjects)	$F(3.606, 151.460) = 38.849$	<0.001	0.481		
		Group (between-subjects)	$F(3,42) = 1.263$	0.299	0.083		
	Relapse (incubation) tests	Lever x Group	$F(3,42) = 0.564$	0.642	0.039		
		Session x Group	$F(10.819, 151.460) = 1.765$	0.066	0.112		
		Lever x Session	$F(3.5, 147.017) = 29.225$	<0.001	0.410		
		Lever x Session x Group	$F(10.501, 147.017) = 1.558$	0.121	0.100		
		RM-ANOVA (Lever x Session)	F-value	P-value	Partial $\eta^2$		
Relapse (incubation) tests	Lever presses over 30 min (n=33)	Lever (within-subjects)	$F(1,45) = 142.935$	<0.001	0.761	1D, left	
		Session (within-subjects)	$F(3.670, 165.134) = 37.188$	<0.001	0.452		
		Lever x Session	$F(3.561, 160.248) = 28.849$	<0.001	0.391		
		RM-ANOVA (Lever x Day)	F-value	P-value	Partial $\eta^2$		
		Lever (within-subjects)	$F(1,30) = 195.173$	<0.001	0.867		
	Binned (3 x 10 min) lever presses (n=33)	Day (between-subjects)	$F(2,30) = 3.300$	0.051	0.180	1D, right	
		Lever x Day	$F(2,30) = 3.438$	0.045	0.186		
		1-way-ANOVA (Lever)	F-value	P-value	$\eta^2$		
		Active lever	$F(2,30) = 3.491$	0.043	0.189		
		Inactive lever	$F(2,30) = 1.458$	0.249	0.089		
Relapse (incubation) tests	Binned (3 x 10 min) lever presses (n=33)	Tukey HSD (Active lever)	Mean Difference	Std. Error	P-value	1D, right	
		Day 1 vs. Day 15	-5.856	10.830	0.852		
		Day 1 vs. Day 60	-28.473	11.336	0.045		
		Day 15 vs. Day 60	-22.617	11.109	0.121		
		RM-ANOVA (Lever x Bin x Day)	F-value	P-value	Partial $\eta^2$		
		Lever (within-subjects)	$F(1,30) = 198.809$	<0.001	0.867		
		Bin (within-subjects)	$F(1.701, 51.033) = 16.618$	<0.001	0.356		
		Day (between-subjects)	$F(2,30) = 3.252$	0.053	0.178		
		Lever x Day	$F(2,30) = 3.432$	0.045	0.186		
		Bin x Day	$F(3.401, 51.033) = 2.230$	0.088	0.129		
Relapse (incubation) tests	Lever presses in first 10 min bin (n=33)	Lever x Bin	$F(1.882, 56.464) = 5.967$	0.005	0.166		
		Lever x Bin x Day	$F(3.764, 56.464) = 2.512$	0.055	0.143		
		RM-ANOVA (Lever x Day)	F-value	P-value	Partial $\eta^2$		
		Lever (within-subjects)	$F(1,30) = 104.334$	<0.001	0.777		
		Day (between-subjects)	$F(2,30) = 4.215$	0.024	0.219		
		Lever x Day	$F(2,30) = 4.892$	0.014	0.246		
		1-way-ANOVA (Lever)	F-value	P-value	$\eta^2$		
		Active lever	$F(2,30) = 4.578$	0.018	0.234		
		Inactive lever	$F(2,30) = 3.624$	0.039	0.195		

		Tukey HSD (Active lever)		P-value	
		Mean Difference			
		Day 1 vs. Day 15	1.621	5.812	0.958
		Day 1 vs. Day 60	-15.045	6.084	0.049
		Day 15 vs. Day 60	-16.667	5.962	0.024
		Tukey HSD (Inactive lever)			
		Mean Difference		P-value	
		Day 1 vs. Day 15	-3.962	1.491	0.032
		Day 1 vs. Day 60	-2.645	1.560	0.223
		Day 15 vs. Day 60	1.317	1.529	0.668

**Table S4. Statistical output for Figure 2B, left panel : Analysis of Z-scored counts from 10 major anatomical regions (n=29, analyses shown in figure 2B are highlighted in grey)**

Factors in analysis	Statistical output			
RM-ANOVA (Region x Group)	F-value	P-value	Partial n2	
Region (within-subjects)	F(1.908,47.688) = 18.157	<0.001	0.421	
Group (between-subjects)	F(3,25) = 4.414	0.013	0.346	
Region x Group	F(5.723,47.688) = 6.099	<0.001	0.423	
1-way-ANOVA (Region)	F-value (ndf=3,ddf=25)	P-value	n2	
Isocortex	5.690	0.004	0.406	
Olfactory areas	6.337	0.002	0.432	
Hippocampal formation	6.719	0.002	0.446	
Cortical subplate	7.040	0.001	0.458	
Striatum	3.102	0.045	0.271	
Pallidum	2.389	0.093	0.223	
Thalamus	1.501	0.238	0.153	
Hypothalamus	2.053	0.132	0.198	
Midbrain	1.568	0.222	0.158	
Hindbrain	3.051	0.047	0.268	
Tukey HSD	Comparison	Mean Difference	Std. Error	P-value
Isocortex	Home cage vs. Day 1	0.041	1.412	1.000
	Home cage vs. Day 15	0.175	1.362	0.999
	Home cage vs. Day 60	-5.619	1.563	0.007
	Day 1 vs. Day 15	0.133	1.451	1.000
	Day 1 vs. Day 60	-5.661	1.641	0.010
	Day 15 vs. Day 60	-5.794	1.598	0.007
	Home cage vs. Day 1	-0.138	1.077	0.999
Olfactory areas	Home cage vs. Day 15	-0.476	1.039	0.967
	Home cage vs. Day 60	-4.755	1.193	0.003
	Day 1 vs. Day 15	-0.338	1.107	0.990
	Day 1 vs. Day 60	-4.617	1.252	0.006
	Day 15 vs. Day 60	-4.279	1.219	0.009
	Home cage vs. Day 1	0.412	1.082	0.981
Hippocampal formation	Home cage vs. Day 15	0.151	1.043	0.999
	Home cage vs. Day 60	-4.551	1.197	0.004
	Day 1 vs. Day 15	-0.261	1.111	0.995
	Day 1 vs. Day 60	-4.963	1.257	0.003
	Day 15 vs. Day 60	-4.702	1.224	0.004
	Home cage vs. Day 1	-1.279	1.972	0.915
Cortical subplate	Home cage vs. Day 15	-1.571	1.902	0.842

	Homecage vs. Day 60	-9.575	2.183	0.001
	Day 1 vs. Day 15	-0.291	2.026	0.999
	Day 1 vs. Day 60	-8.296	2.292	0.007
	Day 15 vs. Day 60	-8.005	2.231	0.007
Striatum	Home cage vs. Day 1	0.374	0.948	0.979
	Home cage vs. Day 15	0.569	0.914	0.924
	Homecage vs. Day 60	-2.460	1.049	0.115
	Day 1 vs. Day 15	0.194	0.973	0.997
	Day 1 vs. Day 60	-2.834	1.101	0.073
	Day 15 vs. Day 60	-3.028	1.072	0.043
Pallidum	Home cage vs. Day 1	0.692	0.867	0.854
	Home cage vs. Day 15	0.695	0.836	0.839
	Homecage vs. Day 60	-1.681	0.959	0.319
	Day 1 vs. Day 15	0.002	0.890	1.000
	Day 1 vs. Day 60	-2.374	1.007	0.112
	Day 15 vs. Day 60	-2.376	0.981	0.099
Thalamus	Home cage vs. Day 1	0.520	0.382	0.533
	Home cage vs. Day 15	0.566	0.368	0.430
	Homecage vs. Day 60	-0.128	0.422	0.990
	Day 1 vs. Day 15	0.046	0.392	0.999
	Day 1 vs. Day 60	-0.648	0.443	0.475
	Day 15 vs. Day 60	-0.694	0.432	0.393
Hypothalamus	Home cage vs. Day 1	0.218	1.453	0.999
	Home cage vs. Day 15	-0.370	1.401	0.993
	Homecage vs. Day 60	-3.531	1.608	0.152
	Day 1 vs. Day 15	-0.587	1.492	0.979
	Day 1 vs. Day 60	-3.749	1.688	0.145
	Day 15 vs. Day 60	-3.161	1.644	0.244
Midbrain	Home cage vs. Day 1	0.611	0.563	0.701
	Home cage vs. Day 15	0.559	0.542	0.733
	Homecage vs. Day 60	-0.611	0.623	0.761
	Day 1 vs. Day 15	-0.051	0.578	1.000
	Day 1 vs. Day 60	-1.222	0.654	0.266
	Day 15 vs. Day 60	-1.171	0.636	0.279
Hindbrain	Home cage vs. Day 1	0.667	1.505	0.970
	Home cage vs. Day 15	-0.162	1.451	0.999
	Homecage vs. Day 60	-4.222	1.665	0.079
	Day 1 vs. Day 15	-0.829	1.545	0.949
	Day 1 vs. Day 60	-4.889	1.748	0.045
	Day 15 vs. Day 60	-4.060	1.702	0.106

**Table S5. Statistical output for Figure 2B, right panel: Analysis of Z-scored counts from 56 subdivisions of 10 major anatomical regions (n=29)**

Factors in analysis	Statistical Output		
RM-ANOVA (Region x Group)	F-value	P-value	Partial n2
Region (within-subjects)	F(1.813,45.313) = 18.664	<0.001	0.427
Group (between-subjects)	F(3,25) = 5.962	0.003	0.417
Region x Group	F(5.438,45.313) = 7.981	<0.001	0.489
1-way-ANOVA (Region)	F-value (ndf=3,ddf=25)	P-value	n2
Infralimbic area (ILA)	10.835	<0.001	0.565
Visual areas (VIS)	9.141	<0.001	0.523
Orbital area (ORB)	12.209	<0.001	0.594
Anterior cingulate area (ACA)	3.884	0.021	0.318
Agranular insular area (AI)	8.880	<0.001	0.516
Prelimbic area (PL)	7.764	<0.001	0.482
Ectorhinal area (ECT)	9.158	<0.001	0.524
Retrosplenial area (RSP)	3.044	0.047	0.268
Perirhinal area (PERI)	7.569	<0.001	0.476
Somatomotor areas (MO)	3.731	0.024	0.309
Somatosensory areas (SS)	3.419	0.033	0.291
Temporal association areas (TEa)	7.361	0.001	0.469
Auditory areas (AUD)	5.471	0.005	0.396
Gustatory areas (GU)	4.325	0.014	0.342
Visceral area (VISC)	2.905	0.055	0.259
Frontal pole, cerebral cortex (FRP)	0.574	0.638	0.064
Dorsal peduncular area (DP)	9.128	<0.001	0.523
Taenia tecta (TT)	9.403	<0.001	0.530
Cortical amygdalar area (COA)	4.597	0.011	0.355
Piriform area (PIR)	6.227	0.003	0.428
Anterior olfactory nucleus (AON)	11.283	<0.001	0.575
Piriform-amygdalar area (PAA)	4.054	0.018	0.327
Nucleus of the lateral olfactory tract (NLOT)	3.233	0.039	0.280
Postpiriform transition area (TR)	3.296	0.037	0.283
Accessory olfactory bulb (AOB)	0.089	0.966	0.011
Main olfactory bulb (MOB)	0.271	0.846	0.031
Retrohippocampal region (RHP)	9.113	<0.001	0.522
Ammon's horn (CA)	2.356	0.096	0.220
Dentate gyrus (DG)	2.470	0.085	0.229
Fasciola cinerea (FC)	1.094	0.370	0.116
Induseum griseum (IG)	1.124	0.358	0.119
Clastrum (CLA)	10.132	<0.001	0.549
Endopiriform nucleus (EP)	7.998	<0.001	0.490
Basomedial amygdalar nucleus (BMA)	4.967	0.008	0.373
Basolateral amygdalar nucleus (BLA)	5.178	0.006	0.383
Posterior amygdalar nucleus (PA)	4.254	0.015	0.338
Lateral amygdalar nucleus (LA)	2.122	0.123	0.203
Lateral septal complex (LSX)	5.580	0.005	0.401
Striatum ventral region (STRv)	3.698	0.025	0.307
Striatum-like amygdalar nuclei (sAMY)	3.146	0.043	0.274
Striatum dorsal region (STRd)	2.341	0.098	0.219
Pallidum, caudal region (PALc)	3.350	0.035	0.287
Pallidum, medial region (PALm)	2.585	0.076	0.237
Pallidum, ventral region (PALv)	2.382	0.093	0.222
Pallidum, dorsal region (PALd)	1.924	0.152	0.188
Thalamus, polymodal association cortex related (DORpm)	1.728	0.187	0.172
Thalamus, sensory-motor cortex related (DORsm)	1.236	0.318	0.129
Periventricular region (PVR)	2.619	0.073	0.239
Periventricular zone (PVZ)	1.849	0.164	0.182
Hypothalamic medial zone (MEZ)	2.273	0.105	0.214
Hypothalamic lateral zone (LZ)	2.033	0.135	0.196

Midbrain, sensory related (MBsen)	2.101	0.126	0.201		
Midbrain, behavioral state related (MBsta)	1.308	0.294	0.136		
Midbrain, motor related (MBmot)	1.414	0.262	0.145		
Medulla (MY)	4.341	0.014	0.343		
Pons (P)	2.064	0.131	0.199		
<b>Tukey HSD</b>	<b>Comparison</b>	<b>Mean Difference</b>	<b>Std. Error</b>	<b>P-value</b>	
Infralimbic area (ILA)	Home cage vs. Day 1	-0.389	3.823	1.000	
	Home cage vs. Day 15	-0.507	3.686	0.999	
	Homecage vs. Day 60	-21.537	4.231	<0.001	
	Day 1 vs. Day 15	-0.117	3.926	1.000	
	Day 1 vs. Day 60	-21.147	4.442	<0.001	
	Day 15 vs. Day 60	-21.030	4.325	<0.001	
Visual areas (VIS)	Home cage vs. Day 1	-1.796	3.236	0.944	
	Home cage vs. Day 15	-1.781	3.120	0.940	
	Homecage vs. Day 60	-17.515	3.581	<0.001	
	Day 1 vs. Day 15	0.015	3.323	1.000	
	Day 1 vs. Day 60	-15.719	3.760	0.002	
	Day 15 vs. Day 60	-15.734	3.661	0.001	
Orbital area (ORB)	Home cage vs. Day 1	-0.058	1.362	1.000	
	Home cage vs. Day 15	0.290	1.313	0.996	
	Homecage vs. Day 60	-7.954	1.508	<0.001	
	Day 1 vs. Day 15	0.348	1.399	0.994	
	Day 1 vs. Day 60	-7.896	1.583	<0.001	
	Day 15 vs. Day 60	-8.244	1.541	<0.001	
Anterior cingulate area (ACA)	Home cage vs. Day 1	0.405	2.254	0.998	
	Home cage vs. Day 15	0.195	2.173	1.000	
	Homecage vs. Day 60	-7.311	2.494	0.034	
	Day 1 vs. Day 15	-0.210	2.315	1.000	
	Day 1 vs. Day 60	-7.716	2.619	0.033	
	Day 15 vs. Day 60	-7.506	2.550	0.033	
Agranular insular area (AI)	Home cage vs. Day 1	-0.102	1.216	1.000	
	Home cage vs. Day 15	-0.017	1.172	1.000	
	Homecage vs. Day 60	-6.155	1.345	<0.001	
	Day 1 vs. Day 15	0.085	1.248	1.000	
	Day 1 vs. Day 60	-6.052	1.412	0.001	
	Day 15 vs. Day 60	-6.138	1.375	<0.001	
Prelimbic area (PL)	Home cage vs. Day 1	0.471	1.318	0.984	
	Home cage vs. Day 15	0.231	1.271	0.998	
	Homecage vs. Day 60	-5.976	1.459	0.002	
	Day 1 vs. Day 15	-0.240	1.354	0.998	
	Day 1 vs. Day 60	-6.447	1.532	0.002	
	Day 15 vs. Day 60	-6.207	1.491	0.002	
Ectorhinal area (ECT)	Home cage vs. Day 1	-0.108	0.946	0.999	
	Home cage vs. Day 15	0.114	0.912	0.999	
	Homecage vs. Day 60	-4.827	1.047	<0.001	
	Day 1 vs. Day 15	0.223	0.972	0.996	
	Day 1 vs. Day 60	-4.719	1.099	0.001	
	Day 15 vs. Day 60	-4.942	1.070	<0.001	
Retrosplenial area (RSP)	Home cage vs. Day 1	0.302	1.546	0.997	
	Home cage vs. Day 15	0.000	1.491	1.000	
	Homecage vs. Day 60	-4.458	1.711	0.068	
	Day 1 vs. Day 15	-0.302	1.588	0.998	
	Day 1 vs. Day 60	-4.760	1.797	0.062	
	Day 15 vs. Day 60	-4.458	1.749	0.076	
Perirhinal area (PERI)	Home cage vs. Day 1	0.053	0.873	1.000	
	Home cage vs. Day 15	0.038	0.842	1.000	
	Homecage vs. Day 60	-4.031	0.966	0.002	
	Day 1 vs. Day 15	-0.015	0.897	1.000	
	Day 1 vs. Day 60	-4.084	1.015	0.002	

	Day 15 vs. Day 60	-4.068	0.988	0.002
Somatomotor areas (MO)	Home cage vs. Day 1	0.565	1.308	0.972
	Home cage vs. Day 15	0.695	1.261	0.945
	Homecage vs. Day 60	-3.806	1.448	0.065
	Day 1 vs. Day 15	0.130	1.343	1.000
	Day 1 vs. Day 60	-4.371	1.520	0.038
	Day 15 vs. Day 60	-4.501	1.480	0.026
Somatosensory areas (SS)	Home cage vs. Day 1	0.211	1.155	0.998
	Home cage vs. Day 15	0.436	1.114	0.979
	Homecage vs. Day 60	-3.374	1.278	0.063
	Day 1 vs. Day 15	0.225	1.186	0.998
	Day 1 vs. Day 60	-3.586	1.342	0.059
	Day 15 vs. Day 60	-3.811	1.307	0.035
Temporal association areas (TEa)	Home cage vs. Day 1	0.088	0.767	0.999
	Home cage vs. Day 15	0.356	0.740	0.963
	Homecage vs. Day 60	-3.354	0.849	0.003
	Day 1 vs. Day 15	0.268	0.788	0.986
	Day 1 vs. Day 60	-3.442	0.892	0.004
	Day 15 vs. Day 60	-3.710	0.868	0.001
Auditory areas (AUD)	Home cage vs. Day 1	0.051	0.846	1.000
	Home cage vs. Day 15	0.361	0.816	0.970
	Homecage vs. Day 60	-3.186	0.936	0.011
	Day 1 vs. Day 15	0.310	0.869	0.984
	Day 1 vs. Day 60	-3.237	0.983	0.015
	Day 15 vs. Day 60	-3.547	0.957	0.005
Gustatory areas (GU)	Home cage vs. Day 1	0.422	0.643	0.912
	Home cage vs. Day 15	0.541	0.620	0.818
	Homecage vs. Day 60	-1.880	0.712	0.063
	Day 1 vs. Day 15	0.119	0.660	0.998
	Day 1 vs. Day 60	-2.303	0.747	0.024
	Day 15 vs. Day 60	-2.422	0.727	0.013
Visceral area (VISC)	Home cage vs. Day 1	0.212	0.578	0.983
	Home cage vs. Day 15	0.329	0.557	0.934
	Homecage vs. Day 60	-1.458	0.640	0.130
	Day 1 vs. Day 15	0.117	0.594	0.997
	Day 1 vs. Day 60	-1.671	0.672	0.087
	Day 15 vs. Day 60	-1.787	0.654	0.052
Frontal pole, cerebral cortex (FRP)	Home cage vs. Day 1	0.037	0.461	1.000
	Home cage vs. Day 15	-0.058	0.445	0.999
	Homecage vs. Day 60	0.574	0.510	0.678
	Day 1 vs. Day 15	-0.095	0.474	0.997
	Day 1 vs. Day 60	0.537	0.536	0.749
	Day 15 vs. Day 60	0.633	0.522	0.625
Dorsal peduncular area (DP)	Home cage vs. Day 1	-0.922	3.029	0.990
	Home cage vs. Day 15	-0.511	2.920	0.998
	Homecage vs. Day 60	-15.873	3.352	<0.001
	Day 1 vs. Day 15	0.412	3.110	0.999
	Day 1 vs. Day 60	-14.950	3.519	0.001
	Day 15 vs. Day 60	-15.362	3.426	<0.001
Taenia tecta (TT)	Home cage vs. Day 1	-0.156	1.645	1.000
	Home cage vs. Day 15	-0.524	1.586	0.987
	Homecage vs. Day 60	-8.727	1.821	<0.001
	Day 1 vs. Day 15	-0.368	1.690	0.996
	Day 1 vs. Day 60	-8.571	1.912	<0.001
	Day 15 vs. Day 60	-8.203	1.861	<0.001
Cortical amygdalar area (COA)	Home cage vs. Day 1	-0.463	1.844	0.994
	Home cage vs. Day 15	-1.448	1.778	0.847
	Homecage vs. Day 60	-7.131	2.041	0.009
	Day 1 vs. Day 15	-0.985	1.894	0.953
	Day 1 vs. Day 60	-6.668	2.143	0.022

	Day 15 vs. Day 60	-5.683	2.086	0.053
Piriform area (PIR)	Home cage vs. Day 1	-0.240	1.305	0.998
	Home cage vs. Day 15	-0.574	1.259	0.968
	Homecage vs. Day 60	-5.734	1.445	0.003
	Day 1 vs. Day 15	-0.334	1.341	0.994
	Day 1 vs. Day 60	-5.494	1.517	0.007
	Day 15 vs. Day 60	-5.160	1.477	0.009
Anterior olfactory nucleus (AON)	Home cage vs. Day 1	-0.095	0.860	0.999
	Home cage vs. Day 15	-0.280	0.829	0.986
	Homecage vs. Day 60	-4.991	0.951	<0.001
	Day 1 vs. Day 15	-0.185	0.883	0.997
	Day 1 vs. Day 60	-4.896	0.999	<0.001
	Day 15 vs. Day 60	-4.711	0.972	<0.001
Piriform-amyg达尔 area (PAA)	Home cage vs. Day 1	-0.233	1.280	0.998
	Home cage vs. Day 15	-1.101	1.234	0.809
	Homecage vs. Day 60	-4.634	1.417	0.015
	Day 1 vs. Day 15	-0.868	1.315	0.911
	Day 1 vs. Day 60	-4.401	1.487	0.032
	Day 15 vs. Day 60	-3.533	1.448	0.095
Nucleus of the lateral olfactory tract (NLOT)	Home cage vs. Day 1	0.006	1.259	1.000
	Home cage vs. Day 15	-0.120	1.214	1.000
	Homecage vs. Day 60	-3.862	1.394	0.048
	Day 1 vs. Day 15	-0.126	1.293	1.000
	Day 1 vs. Day 60	-3.868	1.463	0.063
	Day 15 vs. Day 60	-3.742	1.425	0.065
Postpiriform transition area (TR)	Home cage vs. Day 1	0.304	0.613	0.959
	Home cage vs. Day 15	0.195	0.591	0.987
	Homecage vs. Day 60	-1.702	0.679	0.083
	Day 1 vs. Day 15	-0.109	0.630	0.998
	Day 1 vs. Day 60	-2.006	0.712	0.043
	Day 15 vs. Day 60	-1.897	0.694	0.052
Accessory olfactory bulb (AOB)	Home cage vs. Day 1	0.027	0.588	1.000
	Home cage vs. Day 15	0.260	0.567	0.967
	Homecage vs. Day 60	0.010	0.651	1.000
	Day 1 vs. Day 15	0.233	0.604	0.980
	Day 1 vs. Day 60	-0.017	0.683	1.000
	Day 15 vs. Day 60	-0.250	0.665	0.982
Main olfactory bulb (MOB)	Home cage vs. Day 1	0.215	0.560	0.980
	Home cage vs. Day 15	0.049	0.540	1.000
	Homecage vs. Day 60	0.522	0.620	0.834
	Day 1 vs. Day 15	-0.167	0.575	0.991
	Day 1 vs. Day 60	0.306	0.651	0.965
	Day 15 vs. Day 60	0.473	0.634	0.877
Retrohippocampal region (RHP)	Home cage vs. Day 1	0.146	1.372	1.000
	Home cage vs. Day 15	-0.129	1.323	1.000
	Homecage vs. Day 60	-6.992	1.518	<0.001
	Day 1 vs. Day 15	-0.274	1.409	0.997
	Day 1 vs. Day 60	-7.138	1.594	<0.001
	Day 15 vs. Day 60	-6.863	1.552	<0.001
Ammon's horn (CA)	Home cage vs. Day 1	0.449	0.753	0.932
	Home cage vs. Day 15	0.400	0.726	0.945
	Homecage vs. Day 60	-1.625	0.834	0.234
	Day 1 vs. Day 15	-0.049	0.774	1.000
	Day 1 vs. Day 60	-2.075	0.875	0.109
	Day 15 vs. Day 60	-2.025	0.852	0.108
Dentate gyrus (DG)	Home cage vs. Day 1	0.914	0.658	0.518
	Home cage vs. Day 15	0.589	0.635	0.790
	Homecage vs. Day 60	-1.025	0.729	0.507
	Day 1 vs. Day 15	-0.325	0.676	0.963
	Day 1 vs. Day 60	-1.939	0.765	0.079

	Day 15 vs. Day 60	-1.614	0.745	0.160
Fasciola cinerea (FC)	Home cage vs. Day 1	0.424	0.713	0.933
	Home cage vs. Day 15	0.814	0.688	0.643
	Homecage vs. Day 60	-0.558	0.789	0.893
	Day 1 vs. Day 15	0.390	0.732	0.950
	Day 1 vs. Day 60	-0.982	0.829	0.641
	Day 15 vs. Day 60	-1.372	0.807	0.344
Induseum griseum (IG)	Home cage vs. Day 1	0.425	0.313	0.537
	Home cage vs. Day 15	0.475	0.302	0.412
	Homecage vs. Day 60	0.095	0.347	0.993
	Day 1 vs. Day 15	0.050	0.322	0.999
	Day 1 vs. Day 60	-0.331	0.364	0.801
	Day 15 vs. Day 60	-0.381	0.354	0.708
Clastrum (CLA)	Home cage vs. Day 1	-4.323	4.982	0.821
	Home cage vs. Day 15	-6.156	4.804	0.583
	Homecage vs. Day 60	-29.328	5.515	<0.001
	Day 1 vs. Day 15	-1.833	5.117	0.984
	Day 1 vs. Day 60	-25.006	5.789	0.001
	Day 15 vs. Day 60	-23.173	5.636	0.002
Endopiriform nucleus (EP)	Home cage vs. Day 1	-1.798	2.489	0.887
	Home cage vs. Day 15	-2.106	2.399	0.816
	Homecage vs. Day 60	-12.891	2.754	<0.001
	Day 1 vs. Day 15	-0.308	2.556	0.999
	Day 1 vs. Day 60	-11.093	2.891	0.004
	Day 15 vs. Day 60	-10.785	2.815	0.004
Basomedial amygdalar nucleus (BMA)	Home cage vs. Day 1	-1.148	2.059	0.944
	Home cage vs. Day 15	-1.584	1.986	0.855
	Homecage vs. Day 60	-8.437	2.279	0.005
	Day 1 vs. Day 15	-0.436	2.115	0.997
	Day 1 vs. Day 60	-7.289	2.393	0.026
	Day 15 vs. Day 60	-6.853	2.330	0.033
Basolateral amygdalar nucleus (BLA)	Home cage vs. Day 1	-0.778	1.181	0.911
	Home cage vs. Day 15	-0.924	1.139	0.849
	Homecage vs. Day 60	-4.961	1.307	0.004
	Day 1 vs. Day 15	-0.146	1.213	0.999
	Day 1 vs. Day 60	-4.183	1.372	0.026
	Day 15 vs. Day 60	-4.038	1.336	0.027
Posterior amygdalar nucleus (PA)	Home cage vs. Day 1	0.103	1.353	1.000
	Home cage vs. Day 15	-1.104	1.304	0.832
	Homecage vs. Day 60	-4.866	1.497	0.016
	Day 1 vs. Day 15	-1.207	1.389	0.821
	Day 1 vs. Day 60	-4.969	1.572	0.020
	Day 15 vs. Day 60	-3.762	1.530	0.092
Lateral amygdalar nucleus (LA)	Home cage vs. Day 1	-0.428	0.678	0.921
	Home cage vs. Day 15	0.302	0.653	0.966
	Homecage vs. Day 60	-1.540	0.750	0.196
	Day 1 vs. Day 15	0.731	0.696	0.722
	Day 1 vs. Day 60	-1.112	0.787	0.504
	Day 15 vs. Day 60	-1.842	0.767	0.102
Lateral septal complex (LSX)	Home cage vs. Day 1	-0.136	2.983	1.000
	Home cage vs. Day 15	-0.490	2.877	0.998
	Homecage vs. Day 60	-12.099	3.302	0.006
	Day 1 vs. Day 15	-0.353	3.064	0.999
	Day 1 vs. Day 60	-11.963	3.466	0.010
	Day 15 vs. Day 60	-11.610	3.375	0.010
Striatum ventral region (STRv)	Home cage vs. Day 1	0.246	1.452	0.998
	Home cage vs. Day 15	0.265	1.400	0.998
	Homecage vs. Day 60	-4.546	1.607	0.042
	Day 1 vs. Day 15	0.019	1.491	1.000
	Day 1 vs. Day 60	-4.793	1.687	0.041

	Day 15 vs. Day 60	-4.811	1.642	0.034
Striatum-like amygdalar nuclei (sAMY)	Home cage vs. Day 1	-0.193	1.447	0.999
	Home cage vs. Day 15	-0.558	1.395	0.978
	Homecage vs. Day 60	-4.541	1.601	0.042
	Day 1 vs. Day 15	-0.365	1.486	0.995
	Day 1 vs. Day 60	-4.347	1.681	0.071
	Day 15 vs. Day 60	-3.983	1.637	0.096
Striatum dorsal region (STRd)	Home cage vs. Day 1	0.457	0.566	0.851
	Home cage vs. Day 15	0.757	0.546	0.519
	Homecage vs. Day 60	-0.856	0.626	0.531
	Day 1 vs. Day 15	0.300	0.581	0.954
	Day 1 vs. Day 60	-1.312	0.657	0.216
	Day 15 vs. Day 60	-1.613	0.640	0.081
Pallidum, caudal region (PALc)	Home cage vs. Day 1	-0.198	2.328	1.000
	Home cage vs. Day 15	-0.487	2.245	0.996
	Homecage vs. Day 60	-7.402	2.577	0.038
	Day 1 vs. Day 15	-0.289	2.391	0.999
	Day 1 vs. Day 60	-7.204	2.705	0.060
	Day 15 vs. Day 60	-6.915	2.634	0.065
Pallidum, medial region (PALm)	Home cage vs. Day 1	0.720	1.136	0.920
	Home cage vs. Day 15	0.450	1.095	0.976
	Homecage vs. Day 60	-2.640	1.257	0.180
	Day 1 vs. Day 15	-0.270	1.166	0.995
	Day 1 vs. Day 60	-3.360	1.319	0.077
	Day 15 vs. Day 60	-3.090	1.285	0.102
Pallidum, ventral region (PALv)	Home cage vs. Day 1	0.656	0.796	0.842
	Home cage vs. Day 15	0.755	0.767	0.760
	Homecage vs. Day 60	-1.458	0.881	0.367
	Day 1 vs. Day 15	0.099	0.817	0.999
	Day 1 vs. Day 60	-2.114	0.924	0.128
	Day 15 vs. Day 60	-2.213	0.900	0.092
Pallidum, dorsal region (PALd)	Home cage vs. Day 1	0.683	0.453	0.449
	Home cage vs. Day 15	0.754	0.437	0.332
	Homecage vs. Day 60	-0.195	0.502	0.980
	Day 1 vs. Day 15	0.071	0.465	0.999
	Day 1 vs. Day 60	-0.878	0.526	0.361
	Day 15 vs. Day 60	-0.949	0.513	0.274
Thalamus, polymodal association cortex related (DORpm)	Home cage vs. Day 1	0.575	0.405	0.500
	Home cage vs. Day 15	0.651	0.390	0.360
	Homecage vs. Day 60	-0.152	0.448	0.986
	Day 1 vs. Day 15	0.077	0.416	0.998
	Day 1 vs. Day 60	-0.727	0.470	0.427
	Day 15 vs. Day 60	-0.804	0.458	0.318
Thalamus, sensory-motor cortex related (DORsm)	Home cage vs. Day 1	0.479	0.334	0.491
	Home cage vs. Day 15	0.494	0.323	0.435
	Homecage vs. Day 60	0.036	0.370	1.000
	Day 1 vs. Day 15	0.015	0.343	1.000
	Day 1 vs. Day 60	-0.443	0.389	0.669
	Day 15 vs. Day 60	-0.458	0.378	0.627
Periventricular region (PVR)	Home cage vs. Day 1	-0.035	1.965	1.000
	Home cage vs. Day 15	-0.922	1.895	0.961
	Homecage vs. Day 60	-5.591	2.175	0.073
	Day 1 vs. Day 15	-0.887	2.018	0.971
	Day 1 vs. Day 60	-5.556	2.283	0.096
	Day 15 vs. Day 60	-4.669	2.223	0.181
Periventricular zone (PVZ)	Home cage vs. Day 1	0.386	1.838	0.997
	Home cage vs. Day 15	-1.274	1.772	0.889
	Homecage vs. Day 60	-4.186	2.034	0.195
	Day 1 vs. Day 15	-1.660	1.888	0.816
	Day 1 vs. Day 60	-4.571	2.136	0.168

	Day 15 vs. Day 60	-2.912	2.079	0.511
Hypothalamic medial zone (MEZ)	Home cage vs. Day 1	0.318	1.568	0.997
	Home cage vs. Day 15	-0.477	1.512	0.989
	Homecage vs. Day 60	-3.987	1.736	0.126
	Day 1 vs. Day 15	-0.796	1.610	0.960
	Day 1 vs. Day 60	-4.305	1.822	0.111
	Day 15 vs. Day 60	-3.509	1.774	0.223
Hypothalamic lateral zone (LZ)	Home cage vs. Day 1	0.279	1.176	0.995
	Home cage vs. Day 15	-0.058	1.134	1.000
	Homecage vs. Day 60	-2.751	1.302	0.177
	Day 1 vs. Day 15	-0.337	1.208	0.992
	Day 1 vs. Day 60	-3.030	1.367	0.146
	Day 15 vs. Day 60	-2.694	1.331	0.206
Midbrain, sensory related (MBsen)	Home cage vs. Day 1	0.542	0.575	0.782
	Home cage vs. Day 15	0.508	0.554	0.796
	Homecage vs. Day 60	-0.939	0.636	0.466
	Day 1 vs. Day 15	-0.034	0.590	1.000
	Day 1 vs. Day 60	-1.482	0.668	0.146
	Day 15 vs. Day 60	-1.448	0.650	0.143
Midbrain, behavioral state related (MBsta)	Home cage vs. Day 1	0.756	0.750	0.746
	Home cage vs. Day 15	0.467	0.723	0.916
	Homecage vs. Day 60	-0.862	0.830	0.729
	Day 1 vs. Day 15	-0.289	0.770	0.982
	Day 1 vs. Day 60	-1.618	0.872	0.272
	Day 15 vs. Day 60	-1.329	0.849	0.415
Midbrain, motor related (MBmot)	Home cage vs. Day 1	0.640	0.588	0.700
	Home cage vs. Day 15	0.590	0.567	0.728
	Homecage vs. Day 60	-0.557	0.651	0.828
	Day 1 vs. Day 15	-0.050	0.604	1.000
	Day 1 vs. Day 60	-1.197	0.684	0.320
	Day 15 vs. Day 60	-1.147	0.666	0.334
Medulla (MY)	Home cage vs. Day 1	-0.203	2.017	1.000
	Home cage vs. Day 15	-1.732	1.945	0.810
	Homecage vs. Day 60	-7.485	2.233	0.013
	Day 1 vs. Day 15	-1.528	2.072	0.881
	Day 1 vs. Day 60	-7.282	2.344	0.023
	Day 15 vs. Day 60	-5.753	2.282	0.081
Pons (P)	Home cage vs. Day 1	0.692	1.280	0.948
	Home cage vs. Day 15	0.158	1.234	0.999
	Homecage vs. Day 60	-2.773	1.416	0.231
	Day 1 vs. Day 15	-0.534	1.314	0.977
	Day 1 vs. Day 60	-3.465	1.487	0.118
	Day 15 vs. Day 60	-2.931	1.447	0.206

**Table S6. Statistical output for Figure 3B, right panel : Analysis of Z-scored counts from 5 major anatomical regions within AP +1.55 to AP +1.75 coronal subvolume (n=31, analyses shown in figure 3B are highlighted in grey)**

Factors in analysis	Statistical output			
<u>RM-ANOVA (Region x Group)</u>	F-value	P-value	Partial n2	
Region (within-subjects)	F(1.573,4.720) = 6.533	0.006	0.189	
Group (between-subjects)	F(3,28) = 6.853	0.001	0.423	
Region x Group	F(4.720,44.049) = 4.689	0.002	0.334	
<u>1-way-ANOVA (Region)</u>	F-value (ndf=3,ddf=28)	P-value	n2	
Isocortex	7.388	0.001	0.442	
Olfactory areas	8.949	<0.001	0.489	
Cortical subplate	7.689	<0.001	0.452	
Striatum	5.953	0.003	0.389	
Pallidum	4.790	0.008	0.339	
<u>Tukey HSD</u>	Comparison	Mean Difference	Std. Error	P-value
Isocortex	Home cage vs. Day 1	0.545	1.778	0.990
	Home cage vs. Day 15	0.031	1.778	1.000
	Home cage vs. Day 60	-8.578	2.064	0.001
	Day 1 vs. Day 15	-0.514	1.913	0.993
	Day 1 vs. Day 60	-9.123	2.181	0.001
	Day 15 vs. Day 60	-8.609	2.181	0.003
Olfactory areas	Home cage vs. Day 1	0.595	1.703	0.985
	Home cage vs. Day 15	-0.957	1.703	0.942
	Home cage vs. Day 60	-9.228	1.977	0.000
	Day 1 vs. Day 15	-1.552	1.833	0.832
	Day 1 vs. Day 60	-9.822	2.090	0.000
	Day 15 vs. Day 60	-8.271	2.090	0.003
Cortical subplate	Home cage vs. Day 1	0.459	2.752	0.998
	Home cage vs. Day 15	-0.700	2.752	0.994
	Home cage vs. Day 60	-13.872	3.194	0.001
	Day 1 vs. Day 15	-1.159	2.961	0.979
	Day 1 vs. Day 60	-14.331	3.376	0.001
	Day 15 vs. Day 60	-13.172	3.376	0.003
Striatum	Home cage vs. Day 1	0.754	2.972	0.994
	Home cage vs. Day 15	-0.941	2.972	0.989
	Home cage vs. Day 60	-13.111	3.450	0.004
	Day 1 vs. Day 15	-1.696	3.198	0.951
	Day 1 vs. Day 60	-13.866	3.646	0.004
	Day 15 vs. Day 60	-12.170	3.646	0.012
Pallidum	Home cage vs. Day 1	0.750	2.204	0.986
	Home cage vs. Day 15	-1.217	2.204	0.945
	Home cage vs. Day 60	-8.677	2.559	0.011
	Day 1 vs. Day 15	-1.966	2.372	0.840
	Day 1 vs. Day 60	-9.427	2.704	0.008
	Day 15 vs. Day 60	-7.460	2.704	0.047

**Table S7. Statistical output for Figure 3C: Analysis of Z-scored counts from 18 subregions within AP +1.55 to AP +1.75 coronal subvolume (n=31)**

Factors in analysis	Statistical Output			
RM-ANOVA (Region x Group)	F-value	P-value	Partial $\eta^2$	
Region (within-subjects)	F(1.168,32.698) = 9.502	0.003	0.253	
Group (between-subjects)	F(3,28) = 7.240	0.001	0.437	
Region x Group	F(3.503,32.698) = 6.321	0.001	0.404	
1-way-ANOVA (Region)	F-value (ndf=3,ddf=28)	P-value	$\eta^2$	
Orbital area (ORB)	11.349	<0.001	0.549	
Anterior cingulate area (ACA)	6.074	0.003	0.394	
Infralimbic area (ILA)	7.624	<0.001	0.450	
Prelimbic area (PL)	6.868	0.001	0.424	
Agranular insular area (AI)	11.030	<0.001	0.542	
Somatomotor areas (MO)	5.737	0.003	0.381	
Gustatory areas (GU)	4.764	0.008	0.338	
Somatosensory areas (SS)	4.215	0.014	0.311	
Dorsal peduncular area (DP)	5.876	0.003	0.386	
Taenia tecta (TT)	7.782	<0.001	0.455	
Anterior olfactory nucleus (AON)	10.154	<0.001	0.521	
Piriform area (PIR)	8.909	<0.001	0.488	
Endopiriform nucleus (EP)	8.681	<0.001	0.482	
Clastrum (CLA)	6.220	0.002	0.400	
Lateral septum (LS)	5.714	0.004	0.380	
Nucleus accumbens (ACB)	5.509	0.004	0.371	
Olfactory tubercle (OT)	6.070	0.003	0.394	
Substantia innominata (SI)	4.790	0.008	0.339	
Tukey HSD	Comparison	Mean Difference	Std. Error	P-value
Orbital area (ORB)	Home cage vs. Day 1	0.018	4.269	1.000
	Home cage vs. Day 15	-1.379	4.269	0.988
	Home cage vs. Day 60	-26.455	4.956	<0.001
	Day 1 vs. Day 15	-1.397	4.594	0.990
	Day 1 vs. Day 60	-26.473	5.238	<0.001
	Day 15 vs. Day 60	-25.076	5.238	<0.001
Anterior cingulate area (ACA)	Home cage vs. Day 1	0.885	5.763	0.999
	Home cage vs. Day 15	-1.098	5.763	0.997
	Home cage vs. Day 60	-25.764	6.689	0.003
	Day 1 vs. Day 15	-1.982	6.201	0.988
	Day 1 vs. Day 60	-26.648	7.070	0.004
	Day 15 vs. Day 60	-24.666	7.070	0.008
Infralimbic area (ILA)	Home cage vs. Day 1	0.147	4.793	1.000
	Home cage vs. Day 15	-0.367	4.793	1.000
	Home cage vs. Day 60	-24.079	5.564	<0.001
	Day 1 vs. Day 15	-0.514	5.158	1.000
	Day 1 vs. Day 60	-24.226	5.881	0.002
	Day 15 vs. Day 60	-23.713	5.881	0.002
Prelimbic area (PL)	Home cage vs. Day 1	0.578	4.348	0.999
	Home cage vs. Day 15	-1.044	4.348	0.995
	Home cage vs. Day 60	-20.753	5.047	0.002
	Day 1 vs. Day 15	-1.623	4.678	0.985
	Day 1 vs. Day 60	-21.332	5.334	0.002
	Day 15 vs. Day 60	-19.709	5.334	0.005
Agranular insular area (AI)	Home cage vs. Day 1	0.349	1.931	0.998
	Home cage vs. Day 15	-0.879	1.931	0.968
	Home cage vs. Day 60	-11.727	2.241	<0.001
	Day 1 vs. Day 15	-1.228	2.077	0.934
	Day 1 vs. Day 60	-12.076	2.369	<0.001
	Day 15 vs. Day 60	-10.848	2.369	<0.001
Somatomotor areas (MO)	Home cage vs. Day 1	0.662	1.516	0.972

	Home cage vs. Day 15	0.255	1.516	0.998
	Home cage vs. Day 60	-6.283	1.760	0.007
	Day 1 vs. Day 15	-0.406	1.632	0.994
	Day 1 vs. Day 60	-6.945	1.860	0.004
	Day 15 vs. Day 60	-6.539	1.860	0.008
Gustatory areas (GU)	Home cage vs. Day 1	0.481	0.908	0.951
	Home cage vs. Day 15	0.201	0.908	0.996
	Home cage vs. Day 60	-3.361	1.054	0.017
	Day 1 vs. Day 15	-0.281	0.977	0.992
	Day 1 vs. Day 60	-3.842	1.115	0.009
	Day 15 vs. Day 60	-3.561	1.115	0.017
Somatosensory areas (SS)	Home cage vs. Day 1	0.503	0.744	0.905
	Home cage vs. Day 15	0.587	0.744	0.859
	Home cage vs. Day 60	-2.363	0.863	0.049
	Day 1 vs. Day 15	0.084	0.800	1.000
	Day 1 vs. Day 60	-2.866	0.912	0.019
	Day 15 vs. Day 60	-2.950	0.912	0.016
Dorsal peduncular area (DP)	Home cage vs. Day 1	0.309	6.916	1.000
	Home cage vs. Day 15	-1.005	6.916	0.999
	Home cage vs. Day 60	-30.601	8.027	0.004
	Day 1 vs. Day 15	-1.314	7.442	0.998
	Day 1 vs. Day 60	-30.910	8.485	0.006
	Day 15 vs. Day 60	-29.596	8.485	0.008
Taenia tecta (TT)	Home cage vs. Day 1	0.297	4.496	1.000
	Home cage vs. Day 15	-1.455	4.496	0.988
	Home cage vs. Day 60	-23.030	5.219	<0.001
	Day 1 vs. Day 15	-1.752	4.838	0.983
	Day 1 vs. Day 60	-23.327	5.516	0.001
	Day 15 vs. Day 60	-21.575	5.516	0.003
Anterior olfactory nucleus (AON)	Home cage vs. Day 1	0.447	2.981	0.999
	Home cage vs. Day 15	-1.445	2.981	0.962
	Home cage vs. Day 60	-17.425	3.460	<0.001
	Day 1 vs. Day 15	-1.892	3.208	0.934
	Day 1 vs. Day 60	-17.872	3.657	<0.001
	Day 15 vs. Day 60	-15.980	3.657	<0.001
Piriform area (PIR)	Home cage vs. Day 1	0.608	1.229	0.960
	Home cage vs. Day 15	-0.835	1.229	0.904
	Home cage vs. Day 60	-6.574	1.427	<0.001
	Day 1 vs. Day 15	-1.442	1.323	0.698
	Day 1 vs. Day 60	-7.182	1.508	<0.001
	Day 15 vs. Day 60	-5.739	1.508	0.004
Endopiriform nucleus (EP)	Home cage vs. Day 1	0.502	1.817	0.992
	Home cage vs. Day 15	-1.348	1.817	0.879
	Home cage vs. Day 60	-9.782	2.109	<0.001
	Day 1 vs. Day 15	-1.850	1.955	0.780
	Day 1 vs. Day 60	-10.284	2.229	<0.001
	Day 15 vs. Day 60	-8.434	2.229	0.004
Clastrum (CLA)	Home cage vs. Day 1	0.145	2.131	1.000
	Home cage vs. Day 15	0.332	2.131	0.999
	Home cage vs. Day 60	-9.499	2.474	0.003
	Day 1 vs. Day 15	0.187	2.293	1.000
	Day 1 vs. Day 60	-9.644	2.615	0.005
	Day 15 vs. Day 60	-9.831	2.615	0.004
Lateral septum (LS)	Home cage vs. Day 1	0.373	4.426	1.000
	Home cage vs. Day 15	-1.852	4.426	0.975
	Home cage vs. Day 60	-19.497	5.138	0.004
	Day 1 vs. Day 15	-2.225	4.763	0.966
	Day 1 vs. Day 60	-19.869	5.431	0.005
	Day 15 vs. Day 60	-17.644	5.431	0.015
Nucleus accumbens (ACB)	Home cage vs. Day 1	0.920	4.006	0.996

	Home cage vs. Day 15	-0.662	4.006	0.998
	Home cage vs. Day 60	-16.919	4.650	0.006
	Day 1 vs. Day 15	-1.582	4.311	0.983
	Day 1 vs. Day 60	-17.839	4.915	0.006
	Day 15 vs. Day 60	-16.257	4.915	0.013
Olfactory tubercle (OT)	Home cage vs. Day 1	0.638	2.081	0.990
	Home cage vs. Day 15	-0.817	2.081	0.979
	Home cage vs. Day 60	-9.247	2.415	0.004
	Day 1 vs. Day 15	-1.455	2.239	0.915
	Day 1 vs. Day 60	-9.885	2.553	0.003
	Day 15 vs. Day 60	-8.430	2.553	0.013
Substantia innominata (SI)	Home cage vs. Day 1	0.750	2.204	0.986
	Home cage vs. Day 15	-1.217	2.204	0.945
	Home cage vs. Day 60	-8.677	2.559	0.011
	Day 1 vs. Day 15	-1.966	2.372	0.840
	Day 1 vs. Day 60	-9.427	2.704	0.008
	Day 15 vs. Day 60	-7.460	2.704	0.047

**Table S8. Statistical output for Figure 3D: Analysis of Z-scored counts from 43 subdivisions within AP +1.55 to AP +1.75 coronal subvolume (n=31)**

Factors in analysis	Statistical Output			
RM-ANOVA (Region x Group)	F-value	P-value	Partial $\eta^2$	
Region (within-subjects)	F(1.151,32.235) = 10.299	0.002	0.269	
Group (between-subjects)	F(3,28) = 7.124	0.001	0.433	
Region x Group	F(3.454,32.235) = 6.518	0.001	0.411	
1-way-ANOVA (Region)	F-value (ndf=3,ddf=28)	P-value	$\eta^2$	
Prelimbic area, layer 5 (PL5)	6.890	0.001	0.425	
Infralimbic area, layer 6a (ILA6a)	9.712	<0.001	0.510	
Infralimbic area, layer 5 (ILA5)	7.170	0.001	0.434	
Orbital area, lateral part (ORBI)	12.128	<0.001	0.565	
Prelimbic area, layer 6a (PL6a)	7.815	<0.001	0.456	
Anterior cingulate area (ACA)	6.074	0.003	0.394	
Orbital area, ventrolateral part (ORBvl)	9.708	<0.001	0.510	
Prelimbic area, layer 2/3 (PL2/3)	6.974	0.001	0.428	
Agranular insular area, ventral part (Alv)	12.000	<0.001	0.563	
Prelimbic area, layer 2 (PL2)	7.211	<0.001	0.436	
Infralimbic area, layer 2/3 (ILA2/3)	6.352	0.002	0.405	
Gustatory areas, layer 6a (GU6a)	5.163	0.006	0.356	
Secondary motor area (MOs)	6.852	0.001	0.423	
Agranular insular area, dorsal part (Ald)	9.063	<0.001	0.493	
Infralimbic area, layer 6b (ILA6b)	6.884	0.001	0.424	
Prelimbic area, layer 1 (PL1)	4.814	0.008	0.340	
Prelimbic area, layer 6b (PL6b)	4.534	0.010	0.327	
Gustatory areas, layer 5 (GU5)	4.795	0.008	0.339	
Primary motor area (MOp)	4.837	0.008	0.341	
Infralimbic area, layer 1 (ILA1)	3.078	0.044	0.248	
Gustatory areas, layer 2/3 (GU2/3)	4.394	0.012	0.320	
Gustatory areas, layer 4 (GU4)	5.697	0.004	0.379	
Somatosensory areas (SS)	4.215	0.014	0.311	
Gustatory areas, layer 1 (GU1)	1.780	0.174	0.160	
Dorsal peduncular area, layer 6a (DP6a)	8.272	<0.001	0.470	
Anterior olfactory nucleus, medial part (AONm)	5.459	0.004	0.369	
Dorsal peduncular area, layer 5 (DP5)	5.389	0.005	0.366	
Dorsal peduncular area, layer 2/3 (DP2/3)	5.265	0.005	0.361	
Taenia tecta (TT)	7.782	<0.001	0.455	
Anterior olfactory nucleus, posteroventral part (AONpv)	14.286	<0.001	0.605	
Piriform area, pyramidal layer (PIR2)	11.287	<0.001	0.547	
Piriform area, polymorph layer (PIR3)	13.079	<0.001	0.584	
Dorsal peduncular area, layer 1 (DP1)	3.411	0.031	0.268	
Piriform area, molecular layer (PIR1)	3.680	0.024	0.283	
Endopiriform nucleus (EP)	8.681	<0.001	0.482	
Clastrum (CLA)	6.220	0.002	0.400	
Lateral septal nucleus (LS)	5.714	0.004	0.380	
Nucleus accumbens (ACB)	5.509	0.004	0.371	
Olfactory tubercle, pyramidal layer (OT2)	6.123	0.002	0.396	
Olfactory tubercle, polymorph layer (OT3)	5.647	0.004	0.377	
Islands of Calleja (isl)	5.863	0.003	0.386	
Olfactory tubercle, molecular layer (OT1)	5.797	0.003	0.383	
Substantia innominata (SI)	4.790	0.008	0.339	
Tukey HSD	Comparison	Mean Difference	Std. Error	P-value
Prelimbic area, layer 5 (PL5)	Home cage vs. Day 1	0.173	8.195	1.000
	Home cage vs. Day 15	-2.553	8.195	0.989
	Home cage vs. Day 60	-39.626	9.512	0.001
	Day 1 vs. Day 15	-2.726	8.818	0.990
	Day 1 vs. Day 60	-39.799	10.054	0.003
	Day 15 vs. Day 60	-37.073	10.054	0.005

Infralimbic area, layer 6a (ILA6a)	Home cage vs. Day 1	0.006	6.467	1.000
	Home cage vs. Day 15	-0.628	6.467	1.000
	Homecage vs. Day 60	-36.751	7.506	<0.001
	Day 1 vs. Day 15	-0.635	6.959	1.000
	Day 1 vs. Day 60	-36.758	7.934	<0.001
	Day 15 vs. Day 60	-36.123	7.934	<0.001
Infralimbic area, layer 5 (ILA5)	Home cage vs. Day 1	-0.164	7.212	1.000
	Home cage vs. Day 15	-1.514	7.212	0.997
	Homecage vs. Day 60	-35.505	8.372	0.001
	Day 1 vs. Day 15	-1.350	7.761	0.998
	Day 1 vs. Day 60	-35.341	8.849	0.002
	Day 15 vs. Day 60	-33.992	8.849	0.003
Orbital area, lateral part (ORBl)	Home cage vs. Day 1	-0.033	4.174	1.000
	Home cage vs. Day 15	-1.622	4.174	0.980
	Homecage vs. Day 60	-26.803	4.845	<0.001
	Day 1 vs. Day 15	-1.589	4.491	0.984
	Day 1 vs. Day 60	-26.770	5.121	<0.001
	Day 15 vs. Day 60	-25.181	5.121	<0.001
Prelimbic area, layer 6a (PL6a)	Home cage vs. Day 1	0.289	5.125	1.000
	Home cage vs. Day 15	-1.004	5.125	0.997
	Homecage vs. Day 60	-26.178	5.949	<0.001
	Day 1 vs. Day 15	-1.293	5.515	0.995
	Day 1 vs. Day 60	-26.467	6.288	0.001
	Day 15 vs. Day 60	-25.174	6.288	0.002
Anterior cingulate area (ACA)	Home cage vs. Day 1	0.885	5.763	0.999
	Home cage vs. Day 15	-1.098	5.763	0.997
	Homecage vs. Day 60	-25.764	6.689	0.003
	Day 1 vs. Day 15	-1.982	6.201	0.988
	Day 1 vs. Day 60	-26.648	7.070	0.004
	Day 15 vs. Day 60	-24.666	7.070	0.008
Orbital area, ventrolateral part (ORBvl)	Home cage vs. Day 1	0.108	3.902	1.000
	Home cage vs. Day 15	-0.749	3.902	0.997
	Homecage vs. Day 60	-22.237	4.530	<0.001
	Day 1 vs. Day 15	-0.857	4.199	0.997
	Day 1 vs. Day 60	-22.345	4.788	<0.001
	Day 15 vs. Day 60	-21.488	4.788	<0.001
Prelimbic area, layer 2/3 (PL2/3)	Home cage vs. Day 1	0.436	3.311	0.999
	Home cage vs. Day 15	-0.640	3.311	0.997
	Homecage vs. Day 60	-15.893	3.843	0.002
	Day 1 vs. Day 15	-1.076	3.563	0.990
	Day 1 vs. Day 60	-16.329	4.062	0.002
	Day 15 vs. Day 60	-15.253	4.062	0.004
Agranular insular area, ventral part (Alv)	Home cage vs. Day 1	0.259	2.367	1.000
	Home cage vs. Day 15	-1.416	2.367	0.932
	Homecage vs. Day 60	-15.115	2.747	<0.001
	Day 1 vs. Day 15	-1.674	2.547	0.912
	Day 1 vs. Day 60	-15.374	2.904	<0.001
	Day 15 vs. Day 60	-13.699	2.904	<0.001
Prelimbic area, layer 2 (PL2)	Home cage vs. Day 1	0.641	3.082	0.997
	Home cage vs. Day 15	-0.639	3.082	0.997
	Homecage vs. Day 60	-14.970	3.578	0.001
	Day 1 vs. Day 15	-1.280	3.317	0.980
	Day 1 vs. Day 60	-15.611	3.782	0.002
	Day 15 vs. Day 60	-14.331	3.782	0.004
Infralimbic area, layer 2/3 (ILA2/3)	Home cage vs. Day 1	0.140	3.022	1.000
	Home cage vs. Day 15	-0.099	3.022	1.000
	Homecage vs. Day 60	-13.806	3.507	0.003
	Day 1 vs. Day 15	-0.239	3.251	1.000
	Day 1 vs. Day 60	-13.947	3.707	0.004
	Day 15 vs. Day 60	-13.708	3.707	0.005

Gustatory areas, layer 6a (GU6a)	Home cage vs. Day 1	-0.031	3.305	1.000
	Home cage vs. Day 15	0.316	3.305	1.000
	Homecage vs. Day 60	-13.541	3.837	0.008
	Day 1 vs. Day 15	0.346	3.557	1.000
	Day 1 vs. Day 60	-13.510	4.055	0.012
	Day 15 vs. Day 60	-13.856	4.055	0.010
Secondary motor area (MOs)	Home cage vs. Day 1	0.804	2.360	0.986
	Home cage vs. Day 15	-0.225	2.360	1.000
	Homecage vs. Day 60	-10.987	2.739	0.002
	Day 1 vs. Day 15	-1.029	2.539	0.977
	Day 1 vs. Day 60	-11.791	2.895	0.002
	Day 15 vs. Day 60	-10.762	2.895	0.005
Agranular insular area, dorsal part (Ald)	Home cage vs. Day 1	0.444	1.445	0.990
	Home cage vs. Day 15	-0.273	1.445	0.998
	Homecage vs. Day 60	-7.813	1.677	<0.001
	Day 1 vs. Day 15	-0.717	1.555	0.967
	Day 1 vs. Day 60	-8.257	1.773	<0.001
	Day 15 vs. Day 60	-7.540	1.773	0.001
Infralimbic area, layer 6b (ILA6b)	Home cage vs. Day 1	0.115	1.549	1.000
	Home cage vs. Day 15	0.345	1.549	0.996
	Homecage vs. Day 60	-7.230	1.798	0.002
	Day 1 vs. Day 15	0.230	1.667	0.999
	Day 1 vs. Day 60	-7.345	1.900	0.003
	Day 15 vs. Day 60	-7.575	1.900	0.002
Prelimbic area, layer 1 (PL1)	Home cage vs. Day 1	0.904	1.622	0.944
	Home cage vs. Day 15	-0.659	1.622	0.977
	Homecage vs. Day 60	-6.202	1.883	0.013
	Day 1 vs. Day 15	-1.563	1.745	0.807
	Day 1 vs. Day 60	-7.107	1.990	0.007
	Day 15 vs. Day 60	-5.544	1.990	0.044
Prelimbic area, layer 6b (PL6b)	Home cage vs. Day 1	0.139	1.618	1.000
	Home cage vs. Day 15	0.253	1.618	0.999
	Homecage vs. Day 60	-6.130	1.878	0.014
	Day 1 vs. Day 15	0.115	1.741	1.000
	Day 1 vs. Day 60	-6.268	1.985	0.019
	Day 15 vs. Day 60	-6.383	1.985	0.016
Gustatory areas, layer 5 (GU5)	Home cage vs. Day 1	0.482	1.274	0.981
	Home cage vs. Day 15	0.475	1.274	0.982
	Homecage vs. Day 60	-4.746	1.479	0.017
	Day 1 vs. Day 15	-0.007	1.371	1.000
	Day 1 vs. Day 60	-5.228	1.564	0.012
	Day 15 vs. Day 60	-5.221	1.564	0.012
Primary motor area (MOp)	Home cage vs. Day 1	0.569	1.115	0.956
	Home cage vs. Day 15	0.427	1.115	0.981
	Homecage vs. Day 60	-4.109	1.294	0.018
	Day 1 vs. Day 15	-0.143	1.199	0.999
	Day 1 vs. Day 60	-4.678	1.368	0.010
	Day 15 vs. Day 60	-4.535	1.368	0.013
Infralimbic area, layer 1 (ILA1)	Home cage vs. Day 1	0.595	1.348	0.971
	Home cage vs. Day 15	0.167	1.348	0.999
	Homecage vs. Day 60	-4.021	1.565	0.071
	Day 1 vs. Day 15	-0.428	1.451	0.991
	Day 1 vs. Day 60	-4.616	1.654	0.044
	Day 15 vs. Day 60	-4.188	1.654	0.077
Gustatory areas, layer 2/3 (GU2/3)	Home cage vs. Day 1	0.450	0.933	0.962
	Home cage vs. Day 15	-0.060	0.933	1.000
	Homecage vs. Day 60	-3.391	1.083	0.020
	Day 1 vs. Day 15	-0.510	1.004	0.957
	Day 1 vs. Day 60	-3.841	1.145	0.012
	Day 15 vs. Day 60	-3.331	1.145	0.033

Gustatory areas, layer 4 (GU4)	Home cage vs. Day 1	0.424	0.612	0.899
	Home cage vs. Day 15	0.307	0.612	0.958
	Homecage vs. Day 60	-2.392	0.710	0.011
	Day 1 vs. Day 15	-0.118	0.658	0.998
	Day 1 vs. Day 60	-2.817	0.751	0.004
	Day 15 vs. Day 60	-2.699	0.751	0.006
Somatosensory areas (SS)	Home cage vs. Day 1	0.503	0.744	0.905
	Home cage vs. Day 15	0.587	0.744	0.859
	Homecage vs. Day 60	-2.363	0.863	0.049
	Day 1 vs. Day 15	0.084	0.800	1.000
	Day 1 vs. Day 60	-2.866	0.912	0.019
	Day 15 vs. Day 60	-2.950	0.912	0.016
Gustatory areas, layer 1 (GU1)	Home cage vs. Day 1	0.600	0.407	0.466
	Home cage vs. Day 15	0.199	0.407	0.961
	Homecage vs. Day 60	-0.520	0.472	0.693
	Day 1 vs. Day 15	-0.401	0.438	0.797
	Day 1 vs. Day 60	-1.120	0.499	0.137
	Day 15 vs. Day 60	-0.719	0.499	0.487
Dorsal peduncular area, layer 6a (DP6a)	Home cage vs. Day 1	-0.241	8.398	1.000
	Home cage vs. Day 15	-1.628	8.398	0.997
	Homecage vs. Day 60	-44.350	9.748	<0.001
	Day 1 vs. Day 15	-1.388	9.037	0.999
	Day 1 vs. Day 60	-44.110	10.303	0.001
	Day 15 vs. Day 60	-42.722	10.303	0.002
Anterior olfactory nucleus, medial part (AONm)	Home cage vs. Day 1	-0.032	10.103	1.000
	Home cage vs. Day 15	-4.662	10.103	0.967
	Homecage vs. Day 60	-43.890	11.727	0.004
	Day 1 vs. Day 15	-4.630	10.871	0.974
	Day 1 vs. Day 60	-43.858	12.395	0.007
	Day 15 vs. Day 60	-39.228	12.395	0.018
Dorsal peduncular area, layer 5 (DP5)	Home cage vs. Day 1	0.457	8.203	1.000
	Home cage vs. Day 15	-1.710	8.203	0.997
	Homecage vs. Day 60	-34.858	9.522	0.005
	Day 1 vs. Day 15	-2.167	8.827	0.995
	Day 1 vs. Day 60	-35.315	10.064	0.008
	Day 15 vs. Day 60	-33.147	10.064	0.013
Dorsal peduncular area, layer 2/3 (DP2/3)	Home cage vs. Day 1	0.106	6.575	1.000
	Home cage vs. Day 15	-0.988	6.575	0.999
	Homecage vs. Day 60	-27.615	7.632	0.006
	Day 1 vs. Day 15	-1.095	7.075	0.999
	Day 1 vs. Day 60	-27.722	8.067	0.009
	Day 15 vs. Day 60	-26.627	8.067	0.013
Taenia tecta (TT)	Home cage vs. Day 1	0.297	4.496	1.000
	Home cage vs. Day 15	-1.455	4.496	0.988
	Homecage vs. Day 60	-23.030	5.219	<0.001
	Day 1 vs. Day 15	-1.752	4.838	0.983
	Day 1 vs. Day 60	-23.327	5.516	0.001
	Day 15 vs. Day 60	-21.575	5.516	0.003
Anterior olfactory nucleus, posteroventral part (AONpv)	Home cage vs. Day 1	0.505	1.855	0.993
	Home cage vs. Day 15	-0.902	1.855	0.962
	Homecage vs. Day 60	-12.766	2.153	<0.001
	Day 1 vs. Day 15	-1.407	1.996	0.894
	Day 1 vs. Day 60	-13.271	2.276	<0.001
	Day 15 vs. Day 60	-11.864	2.276	<0.001
Piriform area, pyramidal layer (PIR2)	Home cage vs. Day 1	0.300	1.761	0.998
	Home cage vs. Day 15	-1.671	1.761	0.779
	Homecage vs. Day 60	-10.933	2.044	<0.001
	Day 1 vs. Day 15	-1.971	1.894	0.727
	Day 1 vs. Day 60	-11.233	2.160	<0.001
	Day 15 vs. Day 60	-9.262	2.160	0.001

Piriform area, polymorph layer (PIR3)	Home cage vs. Day 1	0.455	1.583	0.992
	Home cage vs. Day 15	-1.758	1.583	0.686
	Homecage vs. Day 60	-10.521	1.838	<0.001
	Day 1 vs. Day 15	-2.213	1.704	0.571
	Day 1 vs. Day 60	-10.976	1.943	<0.001
	Day 15 vs. Day 60	-8.762	1.943	<0.001
Dorsal peduncular area, layer 1 (DP1)	Home cage vs. Day 1	0.736	2.473	0.991
	Home cage vs. Day 15	0.445	2.473	0.998
	Homecage vs. Day 60	-7.903	2.871	0.048
	Day 1 vs. Day 15	-0.291	2.661	1.000
	Day 1 vs. Day 60	-8.640	3.035	0.039
	Day 15 vs. Day 60	-8.348	3.035	0.048
Piriform area, molecular layer (PIR1)	Home cage vs. Day 1	0.785	0.754	0.726
	Home cage vs. Day 15	0.020	0.754	1.000
	Homecage vs. Day 60	-2.226	0.875	0.074
	Day 1 vs. Day 15	-0.765	0.811	0.782
	Day 1 vs. Day 60	-3.012	0.925	0.015
	Day 15 vs. Day 60	-2.247	0.925	0.094
Endopiriform nucleus (EP)	Home cage vs. Day 1	0.502	1.817	0.992
	Home cage vs. Day 15	-1.348	1.817	0.879
	Homecage vs. Day 60	-9.782	2.109	<0.001
	Day 1 vs. Day 15	-1.850	1.955	0.780
	Day 1 vs. Day 60	-10.284	2.229	<0.001
	Day 15 vs. Day 60	-8.434	2.229	0.004
Clastrum (CLA)	Home cage vs. Day 1	0.145	2.131	1.000
	Home cage vs. Day 15	0.332	2.131	0.999
	Homecage vs. Day 60	-9.499	2.474	0.003
	Day 1 vs. Day 15	0.187	2.293	1.000
	Day 1 vs. Day 60	-9.644	2.615	0.005
	Day 15 vs. Day 60	-9.831	2.615	0.004
Lateral septal nucleus (LS)	Home cage vs. Day 1	0.373	4.426	1.000
	Home cage vs. Day 15	-1.852	4.426	0.975
	Homecage vs. Day 60	-19.497	5.138	0.004
	Day 1 vs. Day 15	-2.225	4.763	0.966
	Day 1 vs. Day 60	-19.869	5.431	0.005
	Day 15 vs. Day 60	-17.644	5.431	0.015
Nucleus accumbens (ACB)	Home cage vs. Day 1	0.920	4.006	0.996
	Home cage vs. Day 15	-0.662	4.006	0.998
	Homecage vs. Day 60	-16.919	4.650	0.006
	Day 1 vs. Day 15	-1.582	4.311	0.983
	Day 1 vs. Day 60	-17.839	4.915	0.006
	Day 15 vs. Day 60	-16.257	4.915	0.013
Olfactory tubercle, pyramidal layer (OT2)	Home cage vs. Day 1	0.594	3.094	0.997
	Home cage vs. Day 15	-1.463	3.094	0.964
	Homecage vs. Day 60	-13.995	3.591	0.003
	Day 1 vs. Day 15	-2.057	3.329	0.925
	Day 1 vs. Day 60	-14.589	3.796	0.003
	Day 15 vs. Day 60	-12.532	3.796	0.013
Olfactory tubercle, polymorph layer (OT3)	Home cage vs. Day 1	0.689	3.130	0.996
	Home cage vs. Day 15	-1.222	3.130	0.979
	Homecage vs. Day 60	-13.525	3.634	0.005
	Day 1 vs. Day 15	-1.912	3.368	0.941
	Day 1 vs. Day 60	-14.215	3.841	0.005
	Day 15 vs. Day 60	-12.303	3.841	0.017
Islands of Calleja (isl)	Home cage vs. Day 1	0.637	1.457	0.971
	Home cage vs. Day 15	-0.384	1.457	0.993
	Homecage vs. Day 60	-6.247	1.691	0.005
	Day 1 vs. Day 15	-1.022	1.568	0.914
	Day 1 vs. Day 60	-6.884	1.787	0.003
	Day 15 vs. Day 60	-5.862	1.787	0.014

Olfactory tubercle, molecular layer (OT1)	Home cage vs. Day 1	0.589	1.366	0.973
	Home cage vs. Day 15	-0.442	1.366	0.988
	Homecage vs. Day 60	-5.842	1.586	0.005
	Day 1 vs. Day 15	-1.030	1.470	0.896
	Day 1 vs. Day 60	-6.430	1.676	0.003
	Day 15 vs. Day 60	-5.400	1.676	0.016
Substantia innominata (SI)	Home cage vs. Day 1	0.750	2.204	0.986
	Home cage vs. Day 15	-1.217	2.204	0.945
	Homecage vs. Day 60	-8.677	2.559	0.011
	Day 1 vs. Day 15	-1.966	2.372	0.840
	Day 1 vs. Day 60	-9.427	2.704	0.008
	Day 15 vs. Day 60	-7.460	2.704	0.047

**Table S9. Statistical output for Figure 4B, right panel : Analysis of Z-scored counts from 8 major anatomical regions within AP -1.08 to AP -1.28 coronal subvolume (n=30, analyses shown in figure 3B are highlighted in grey)**

Factors in analysis	Statistical output			
RM-ANOVA (Region x Group)	F-value	P-value	Partial n2	
Region (within-subjects)	F(1.198,32.349) = 14.028	<0.001	0.342	
Group (between-subjects)	F(3,27) = 3.189	0.040	0.262	
Region x Group	F(3.594,32.349) = 5.476	0.002	0.378	
1-way-ANOVA (Region)	F-value (ndf=3,ddf=27)	P-value	n2	
Isocortex	4.078	0.016	0.312	
Olfactory areas	5.049	0.007	0.359	
Hippocampal formation	0.754	0.530	0.077	
Cortical subplate	4.772	0.009	0.346	
Striatum	2.085	0.126	0.188	
Pallidum	1.098	0.367	0.109	
Thalamus	0.761	0.526	0.078	
Hypothalamus	2.723	0.064	0.232	
Tukey HSD	Comparison	Mean Difference	Std. Error	P-value
Isocortex	Home cage vs. Day 1	0.309	0.910	0.986
	Home cage vs. Day 15	-0.304	0.910	0.987
	Home cage vs. Day 60	-3.219	1.051	0.024
	Day 1 vs. Day 15	-0.613	0.959	0.918
	Day 1 vs. Day 60	-3.528	1.093	0.016
	Day 15 vs. Day 60	-2.915	1.093	0.058
Olfactory areas	Home cage vs. Day 1	0.215	0.883	0.995
	Home cage vs. Day 15	-1.152	0.883	0.568
	Home cage vs. Day 60	-3.513	1.019	0.010
	Day 1 vs. Day 15	-1.367	0.930	0.469
	Day 1 vs. Day 60	-3.728	1.061	0.008
	Day 15 vs. Day 60	-2.361	1.061	0.142
Hippocampal formation	Home cage vs. Day 1	0.143	0.692	0.997
	Home cage vs. Day 15	0.417	0.692	0.930
	Home cage vs. Day 60	-0.808	0.799	0.744
	Day 1 vs. Day 15	0.273	0.729	0.982
	Day 1 vs. Day 60	-0.951	0.831	0.666
	Day 15 vs. Day 60	-1.224	0.831	0.467
Cortical subplate	Home cage vs. Day 1	-0.071	2.315	1.000
	Home cage vs. Day 15	-2.517	2.315	0.700
	Home cage vs. Day 60	-9.349	2.673	0.008
	Day 1 vs. Day 15	-2.446	2.440	0.749
	Day 1 vs. Day 60	-9.278	2.782	0.013
	Day 15 vs. Day 60	-6.832	2.782	0.090
Striatum	Home cage vs. Day 1	0.245	0.655	0.982
	Home cage vs. Day 15	-0.075	0.655	0.999
	Home cage vs. Day 60	-1.602	0.757	0.173
	Day 1 vs. Day 15	-0.320	0.691	0.966
	Day 1 vs. Day 60	-1.847	0.787	0.113
	Day 15 vs. Day 60	-1.527	0.787	0.236

Pallidum	Home cage vs. Day 1	0.248	0.507	0.961
	Home cage vs. Day 15	0.478	0.507	0.782
	Homecage vs. Day 60	-0.585	0.586	0.751
	Day 1 vs. Day 15	0.231	0.535	0.973
	Day 1 vs. Day 60	-0.833	0.610	0.531
	Day 15 vs. Day 60	-1.064	0.610	0.321
Thalamus	Home cage vs. Day 1	0.071	0.385	0.998
	Home cage vs. Day 15	0.412	0.385	0.710
	Homecage vs. Day 60	-0.253	0.444	0.940
	Day 1 vs. Day 15	0.341	0.405	0.834
	Day 1 vs. Day 60	-0.324	0.462	0.896
	Day 15 vs. Day 60	-0.665	0.462	0.487
Hypothalamus	Home cage vs. Day 1	0.301	1.133	0.993
	Home cage vs. Day 15	-0.862	1.133	0.871
	Homecage vs. Day 60	-3.289	1.308	0.080
	Day 1 vs. Day 15	-1.162	1.194	0.766
	Day 1 vs. Day 60	-3.589	1.362	0.062
	Day 15 vs. Day 60	-2.427	1.362	0.303

**Table S10. Statistical output for Figure 4C: Analysis of Z-scored counts from 28 subregions within AP -1.08 to AP -1.28 coronal subvolume (n=30)**

Factors in analysis	Statistical Output			
RM-ANOVA (Region x Group)	F-value	P-value	Partial $\eta^2$	
Region (within-subjects)	F(1.140,30.793) = 9.528	0.003	0.261	
Group (between-subjects)	F(3,27) = 3.373	0.033	0.273	
Region x Group	F(3.421,30.793) = 3.453	0.024	0.277	
1-way-ANOVA (Region)	F-value (ndf=3,ddf=27)	P-value	$\eta^2$	
Somatomotor areas (MO)	3.936	0.019	0.304	
Retrosplenial area (RSP)	1.904	0.153	0.175	
Somatosensory areas (SS)	4.377	0.012	0.327	
Anterior cingulate area (ACA)	1.580	0.217	0.149	
Agranular insular area (AI)	2.042	0.132	0.185	
Visceral area (VISC)	1.943	0.146	0.178	
Cortical amygdalar area (COA)	6.456	0.002	0.418	
Piriform area (PIR)	4.306	0.013	0.324	
Piriform-amygdalar area (PAA)	3.508	0.029	0.280	
Ammon's horn (CA)	0.832	0.488	0.085	
Dentate gyrus (DG)	0.683	0.570	0.071	
Basomedial amygdalar nucleus (BMA)	4.061	0.017	0.311	
Endopiriform nucleus (EP)	5.828	0.003	0.393	
Basolateral amygdalar nucleus (BLA)	3.638	0.025	0.288	
Lateral amygdalar nucleus (LA)	3.728	0.023	0.293	
Clastrum (CLA)	7.956	<0.001	0.469	
Intercalated amygdalar nucleus (IA)	4.051	0.017	0.310	
Central amygdalar nucleus (CEA)	3.465	0.030	0.278	
Medial amygdalar nucleus (MEA)	2.867	0.055	0.242	
Bed nucleus of the accessory olfactory tract (BA)	1.560	0.222	0.148	
Caudoputamen (CP)	1.126	0.356	0.111	
Pallidum, ventral region (PALv)	1.535	0.228	0.146	
Pallidum, dorsal region (PALd)	0.835	0.487	0.085	
Thalamus, sensory-motor cortex related (DORsm)	0.726	0.545	0.075	
Thalamus, polymodal association cortex related (DORpm)	0.786	0.512	0.080	
Hypothalamic medial zone (MEZ)	2.509	0.080	0.218	
Hypothalamic lateral zone (LZ)	2.677	0.067	0.229	
Periventricular zone (PVZ)	2.933	0.051	0.246	
Tukey HSD	Comparison	Mean Difference	Std. Error	P-value
Somatomotor areas (MO)	Home cage vs. Day 1	0.523	1.555	0.987
	Home cage vs. Day 15	-0.803	1.555	0.954
	Home cage vs. Day 60	-5.428	1.795	0.026
	Day 1 vs. Day 15	-1.326	1.639	0.849
	Day 1 vs. Day 60	-5.951	1.868	0.018
	Day 15 vs. Day 60	-4.625	1.868	0.087
Retrosplenial area (RSP)	Home cage vs. Day 1	0.264	1.817	0.999
	Home cage vs. Day 15	-0.350	1.817	0.997
	Home cage vs. Day 60	-4.456	2.098	0.171
	Day 1 vs. Day 15	-0.614	1.915	0.988
	Day 1 vs. Day 60	-4.720	2.184	0.160
	Day 15 vs. Day 60	-4.106	2.184	0.260
Somatosensory areas (SS)	Home cage vs. Day 1	0.274	0.928	0.991
	Home cage vs. Day 15	-0.343	0.928	0.982
	Home cage vs. Day 60	-3.431	1.072	0.017
	Day 1 vs. Day 15	-0.617	0.978	0.921
	Day 1 vs. Day 60	-3.705	1.116	0.013
	Day 15 vs. Day 60	-3.088	1.116	0.047
Anterior cingulate area (ACA)	Home cage vs. Day 1	0.238	1.115	0.996
	Home cage vs. Day 15	-0.347	1.115	0.989

	Home cage vs. Day 60	-2.465	1.287	0.246
	Day 1 vs. Day 15	-0.585	1.175	0.959
	Day 1 vs. Day 60	-2.703	1.340	0.207
	Day 15 vs. Day 60	-2.118	1.340	0.406
Granular insular area (AI)	Home cage vs. Day 1	0.354	0.425	0.838
	Home cage vs. Day 15	0.148	0.425	0.985
	Home cage vs. Day 60	-0.864	0.490	0.313
	Day 1 vs. Day 15	-0.206	0.448	0.967
	Day 1 vs. Day 60	-1.218	0.510	0.104
	Day 15 vs. Day 60	-1.012	0.510	0.219
Visceral area (VISC)	Home cage vs. Day 1	0.342	0.400	0.828
	Home cage vs. Day 15	0.162	0.400	0.977
	Home cage vs. Day 60	-0.776	0.462	0.355
	Day 1 vs. Day 15	-0.181	0.422	0.973
	Day 1 vs. Day 60	-1.118	0.481	0.118
	Day 15 vs. Day 60	-0.937	0.481	0.233
Cortical amygdalar area (COA)	Home cage vs. Day 1	-0.030	1.080	1.000
	Home cage vs. Day 15	-2.262	1.080	0.180
	Home cage vs. Day 60	-4.858	1.247	0.003
	Day 1 vs. Day 15	-2.232	1.138	0.227
	Day 1 vs. Day 60	-4.828	1.297	0.005
	Day 15 vs. Day 60	-2.596	1.297	0.213
Piriform area (PIR)	Home cage vs. Day 1	0.278	0.852	0.988
	Home cage vs. Day 15	-0.735	0.852	0.824
	Home cage vs. Day 60	-3.122	0.984	0.018
	Day 1 vs. Day 15	-1.013	0.898	0.676
	Day 1 vs. Day 60	-3.400	1.024	0.013
	Day 15 vs. Day 60	-2.387	1.024	0.116
Piriform-amyg达尔 area (PAA)	Home cage vs. Day 1	0.434	0.709	0.927
	Home cage vs. Day 15	-0.660	0.709	0.789
	Home cage vs. Day 60	-2.178	0.819	0.059
	Day 1 vs. Day 15	-1.094	0.747	0.472
	Day 1 vs. Day 60	-2.612	0.852	0.024
	Day 15 vs. Day 60	-1.518	0.852	0.304
Ammon's horn (CA)	Home cage vs. Day 1	0.068	0.820	1.000
	Home cage vs. Day 15	0.345	0.820	0.975
	Home cage vs. Day 60	-1.155	0.947	0.620
	Day 1 vs. Day 15	0.277	0.865	0.988
	Day 1 vs. Day 60	-1.223	0.986	0.607
	Day 15 vs. Day 60	-1.500	0.986	0.439
Dentate gyrus (DG)	Home cage vs. Day 1	0.288	0.453	0.920
	Home cage vs. Day 15	0.547	0.453	0.628
	Home cage vs. Day 60	-0.098	0.523	0.998
	Day 1 vs. Day 15	0.259	0.478	0.948
	Day 1 vs. Day 60	-0.386	0.545	0.893
	Day 15 vs. Day 60	-0.645	0.545	0.641
Basomedial amygdalar nucleus (BMA)	Home cage vs. Day 1	-1.346	5.743	0.995
	Home cage vs. Day 15	-7.829	5.743	0.532
	Home cage vs. Day 60	-21.664	6.632	0.015
	Day 1 vs. Day 15	-6.483	6.054	0.710
	Day 1 vs. Day 60	-20.318	6.903	0.031
	Day 15 vs. Day 60	-13.835	6.903	0.211
Endopiriform nucleus (EP)	Home cage vs. Day 1	0.136	1.601	1.000
	Home cage vs. Day 15	-1.252	1.601	0.862
	Home cage vs. Day 60	-7.064	1.848	0.004
	Day 1 vs. Day 15	-1.388	1.687	0.843
	Day 1 vs. Day 60	-7.200	1.924	0.005
	Day 15 vs. Day 60	-5.812	1.924	0.026

Basolateral amygdalar nucleus (BLA)	Home cage vs. Day 1	0.386	2.051	0.998
	Home cage vs. Day 15	-1.199	2.051	0.936
	Homecage vs. Day 60	-7.035	2.369	0.030
	Day 1 vs. Day 15	-1.585	2.162	0.883
	Day 1 vs. Day 60	-7.421	2.465	0.027
	Day 15 vs. Day 60	-5.836	2.465	0.108
Lateral amygdalar nucleus (LA)	Home cage vs. Day 1	0.642	0.867	0.880
	Home cage vs. Day 15	-0.399	0.867	0.967
	Homecage vs. Day 60	-2.729	1.002	0.051
	Day 1 vs. Day 15	-1.041	0.914	0.670
	Day 1 vs. Day 60	-3.371	1.043	0.016
	Day 15 vs. Day 60	-2.330	1.043	0.139
Clastrum (CLA)	Home cage vs. Day 1	0.332	0.529	0.922
	Home cage vs. Day 15	-0.473	0.529	0.808
	Homecage vs. Day 60	-2.585	0.611	0.001
	Day 1 vs. Day 15	-0.805	0.558	0.485
	Day 1 vs. Day 60	-2.917	0.636	<0.001
	Day 15 vs. Day 60	-2.112	0.636	0.013
Intercalated amygdalar nucleus (IA)	Home cage vs. Day 1	-1.038	4.303	0.995
	Home cage vs. Day 15	-4.797	4.303	0.684
	Homecage vs. Day 60	-16.372	4.968	0.014
	Day 1 vs. Day 15	-3.758	4.535	0.840
	Day 1 vs. Day 60	-15.334	5.171	0.030
	Day 15 vs. Day 60	-11.576	5.171	0.138
Central amygdalar nucleus (CEA)	Home cage vs. Day 1	0.306	1.325	0.996
	Home cage vs. Day 15	-0.553	1.325	0.975
	Homecage vs. Day 60	-4.391	1.530	0.037
	Day 1 vs. Day 15	-0.859	1.397	0.926
	Day 1 vs. Day 60	-4.698	1.593	0.031
	Day 15 vs. Day 60	-3.838	1.593	0.099
Medial amygdalar nucleus (MEA)	Home cage vs. Day 1	0.156	1.389	0.999
	Home cage vs. Day 15	-1.515	1.389	0.698
	Homecage vs. Day 60	-4.193	1.604	0.065
	Day 1 vs. Day 15	-1.671	1.464	0.668
	Day 1 vs. Day 60	-4.349	1.669	0.066
	Day 15 vs. Day 60	-2.677	1.669	0.393
Bed nucleus of the accessory olfactory tract (BA)	Home cage vs. Day 1	0.077	0.695	0.999
	Home cage vs. Day 15	-0.702	0.695	0.745
	Homecage vs. Day 60	-1.485	0.802	0.272
	Day 1 vs. Day 15	-0.779	0.732	0.714
	Day 1 vs. Day 60	-1.562	0.835	0.264
	Day 15 vs. Day 60	-0.783	0.835	0.785
Caudoputamen (CP)	Home cage vs. Day 1	0.277	0.369	0.875
	Home cage vs. Day 15	0.447	0.369	0.625
	Homecage vs. Day 60	-0.294	0.426	0.900
	Day 1 vs. Day 15	0.170	0.389	0.971
	Day 1 vs. Day 60	-0.571	0.443	0.578
	Day 15 vs. Day 60	-0.741	0.443	0.358
Pallidum, ventral region (PALv)	Home cage vs. Day 1	0.452	0.706	0.918
	Home cage vs. Day 15	0.546	0.706	0.865
	Homecage vs. Day 60	-1.127	0.815	0.520
	Day 1 vs. Day 15	0.095	0.744	0.999
	Day 1 vs. Day 60	-1.579	0.848	0.268
	Day 15 vs. Day 60	-1.673	0.848	0.223
Pallidum, dorsal region (PALd)	Home cage vs. Day 1	0.167	0.443	0.981
	Home cage vs. Day 15	0.441	0.443	0.753
	Homecage vs. Day 60	-0.373	0.512	0.885
	Day 1 vs. Day 15	0.275	0.467	0.935
	Day 1 vs. Day 60	-0.540	0.533	0.743
	Day 15 vs. Day 60	-0.814	0.533	0.435

Thalamus, sensory-motor cortex related (DORsm)	Home cage vs. Day 1	-0.044	0.433	1.000
	Home cage vs. Day 15	0.367	0.433	0.831
	Homecage vs. Day 60	-0.384	0.500	0.868
	Day 1 vs. Day 15	0.411	0.456	0.805
	Day 1 vs. Day 60	-0.340	0.520	0.914
	Day 15 vs. Day 60	-0.750	0.520	0.485
Thalamus, polymodal association cortex related (DORpm)	Home cage vs. Day 1	0.115	0.368	0.989
	Home cage vs. Day 15	0.429	0.368	0.654
	Homecage vs. Day 60	-0.203	0.425	0.964
	Day 1 vs. Day 15	0.314	0.388	0.850
	Day 1 vs. Day 60	-0.317	0.443	0.890
	Day 15 vs. Day 60	-0.631	0.443	0.495
Hypothalamic medial zone (MEZ)	Home cage vs. Day 1	0.011	5.940	1.000
	Home cage vs. Day 15	-4.111	5.940	0.899
	Homecage vs. Day 60	-17.359	6.859	0.078
	Day 1 vs. Day 15	-4.122	6.261	0.912
	Day 1 vs. Day 60	-17.369	7.139	0.095
	Day 15 vs. Day 60	-13.248	7.139	0.271
Hypothalamic lateral zone (LZ)	Home cage vs. Day 1	0.303	0.979	0.989
	Home cage vs. Day 15	-0.737	0.979	0.875
	Homecage vs. Day 60	-2.790	1.131	0.088
	Day 1 vs. Day 15	-1.040	1.032	0.746
	Day 1 vs. Day 60	-3.093	1.177	0.063
	Day 15 vs. Day 60	-2.053	1.177	0.321
Periventricular zone (PVZ)	Home cage vs. Day 1	0.329	0.763	0.973
	Home cage vs. Day 15	-0.667	0.763	0.818
	Homecage vs. Day 60	-2.211	0.881	0.081
	Day 1 vs. Day 15	-0.996	0.804	0.609
	Day 1 vs. Day 60	-2.540	0.917	0.046
	Day 15 vs. Day 60	-1.545	0.917	0.351

**Table S11. Statistical output for Figure 4D: Analysis of Z-scored counts from 64 subdivisions within AP -1.08 to AP -1.28 coronal subvolume (n=30)**

Factors in analysis	Statistical Output		
RM-ANOVA (Region x Group)	F-value	P-value	Partial $\eta^2$
Region (within-subjects)	F(1.157,31.231) = 7.762	0.007	0.223
Group (between-subjects)	F(3,27) = 3.085	0.044	0.255
Region x Group	F(3.470,31.231) = 3.030	0.038	0.252
1-way-ANOVA (Region)	F-value (ndf=3,ddf=27)	P-value	$\eta^2$
Primary motor area (MOp)	4.339	0.013	0.325
Secondary motor area (MOs)	3.221	0.038	0.264
Retrosplenial area, dorsal part (RSPd)	2.218	0.109	0.198
Primary somatosensory area (SSp)	4.814	0.008	0.348
Retrosplenial area, ventral part (RSPv)	1.483	0.241	0.142
Anterior cingulate area, dorsal part (ACAd)	1.843	0.163	0.170
Lateral visual area, layer 6a (VISC6a)	4.728	0.009	0.344
Supplemental somatosensory area (SSs)	2.948	0.051	0.247
Lateral visual area, layer 5 (VISC5)	2.624	0.071	0.226
Anterior cingulate area, ventral part (ACAv)	0.815	0.497	0.083
Agranular insular area (AI)	2.042	0.132	0.185
Lateral visual area, layer 4 (VISC4)	1.378	0.271	0.133
Lateral visual area, layer 2/3 (VISC23)	0.994	0.411	0.099
Lateral visual area, layer 1 (VISC1)	0.959	0.426	0.096
Cortical amygdalar area, anterior part (COAa)	6.442	0.002	0.417
Piriform area, pyramidal layer (PIR2)	6.051	0.003	0.402
Piriform area, polymorph layer (PIR3)	3.758	0.022	0.295
Cortical amygdalar area, posterior part (COAp)	5.892	0.003	0.396
Piriform-amygdalar area, pyramidal layer (PAA2)	5.601	0.004	0.384
Piriform-amygdalar area, polymorph layer (PAA3)	3.050	0.046	0.253
Piriform-amygdalar area, molecular layer (PAA1)	2.201	0.111	0.197
Piriform area, molecular layer (PIR1)	2.858	0.056	0.241
Ammon's horn (CA)	0.832	0.488	0.085
Dentate gyrus (DG)	0.683	0.570	0.071
Basolateral amygdalar nucleus, anterior part (BMAa)	3.825	0.021	0.298
Basolateral amygdalar nucleus, posterior part (BMAp)	4.353	0.013	0.326
Endopiriform nucleus, ventral part (EPv)	3.584	0.027	0.285
Basolateral amygdalar nucleus (BLA)	3.638	0.025	0.288
Endopiriform nucleus, dorsal part (EPd)	7.269	0.001	0.447
Lateral amygdalar nucleus (LA)	3.728	0.023	0.293
Clastrum (CLA)	7.956	<0.001	0.469
Intercalated amygdalar nucleus (IA)	4.051	0.017	0.310
Medial amygdalar nucleus, anterodorsal part (MEAad)	2.909	0.053	0.244
Central amygdalar nucleus, capsular part (CEAc)	3.893	0.020	0.302
Central amygdalar nucleus, medial part (CEAm)	2.632	0.070	0.226
Medial amygdalar nucleus, anteroventral part (MEAav)	2.158	0.116	0.193
Bed nucleus of the accessory olfactory tract (BA)	1.560	0.222	0.148
Central amygdalar nucleus, lateral part (CEAl)	2.627	0.071	0.226
Caudoputamen (CP)	1.126	0.356	0.111
Pallidum, ventral region (PALv)	1.535	0.228	0.146
Globus pallidus, external segment (GPe)	0.890	0.459	0.090
Globus pallidus, internal segment (GPi)	0.789	0.511	0.081
Central medial nucleus of the thalamus (CM)	1.667	0.197	0.156
Anteroventral nucleus of thalamus (AV)	1.514	0.233	0.144
Ventral medial nucleus of the thalamus (VM)	1.298	0.295	0.126
Lateral habenula (LH)	1.481	0.242	0.141
Anteromedial nucleus (AM)	1.094	0.369	0.108
Submedial nucleus of the thalamus (SMT)	0.806	0.501	0.082
Paracentral nucleus (PCN)	0.990	0.412	0.099
Mediodorsal nucleus of thalamus (MD)	0.739	0.538	0.076
Ventral anterior-lateral complex of the thalamus (VAL)	0.686	0.568	0.071

Central lateral nucleus of the thalamus (CL)	0.793	0.508	0.081		
Ventral posterior complex of the thalamus (VP)	0.581	0.633	0.061		
Medial habenula (MH)	1.102	0.366	0.109		
Reticular nucleus of the thalamus (RT)	0.572	0.638	0.060		
Anterodorsal nucleus (AD)	1.295	0.296	0.126		
Lateral dorsal nucleus of thalamus (LD)	0.829	0.490	0.084		
Anterior hypothalamic nucleus (AHN)	2.324	0.097	0.205		
Hypothalamic medial zone (MEZ)	2.509	0.080	0.218		
Paraventricular hypothalamic nucleus (PVH)	1.827	0.166	0.169		
Lateral hypothalamic area (LHA)	2.356	0.094	0.207		
Arcuate hypothalamic nucleus (ARH)	2.970	0.049	0.248		
Tuberous nucleus (TU)	2.406	0.089	0.211		
Supraoptic nucleus (SO)	0.554	0.650	0.058		
<b>Tukey HSD</b>					
Primary motor area (MOp)					
Home cage vs. Day 1					
Home cage vs. Day 15					
Home cage vs. Day 60					
Day 1 vs. Day 15					
Day 1 vs. Day 60					
Day 15 vs. Day 60					
Secondary motor area (MOs)					
Home cage vs. Day 1					
Home cage vs. Day 15					
Home cage vs. Day 60					
Day 1 vs. Day 15					
Day 1 vs. Day 60					
Day 15 vs. Day 60					
Retrosplenial area, dorsal part (RSPd)					
Home cage vs. Day 1					
Home cage vs. Day 15					
Home cage vs. Day 60					
Day 1 vs. Day 15					
Day 1 vs. Day 60					
Day 15 vs. Day 60					
Primary somatosensory area (SSp)					
Home cage vs. Day 1					
Home cage vs. Day 15					
Home cage vs. Day 60					
Day 1 vs. Day 15					
Day 1 vs. Day 60					
Day 15 vs. Day 60					
Retrosplenial area, ventral part (RSPv)					
Home cage vs. Day 1					
Home cage vs. Day 15					
Home cage vs. Day 60					
Day 1 vs. Day 15					
Day 1 vs. Day 60					
Day 15 vs. Day 60					
Anterior cingulate area, dorsal part (ACAd)					
Home cage vs. Day 1					
Home cage vs. Day 15					
Home cage vs. Day 60					
Day 1 vs. Day 15					
Day 1 vs. Day 60					
Day 15 vs. Day 60					
Lateral visual area, layer 6a (VISC6a)					
Home cage vs. Day 1					
Home cage vs. Day 15					
Home cage vs. Day 60					
Day 1 vs. Day 15					
Day 1 vs. Day 60					
Day 15 vs. Day 60					
Supplemental somatosensory area (SSs)					
Home cage vs. Day 1					
Home cage vs. Day 15					
Home cage vs. Day 60					
Day 1 vs. Day 15					

	Day 1 vs. Day 60	-2.091	0.782	0.057
	Day 15 vs. Day 60	-2.056	0.782	0.063
Lateral visual area, layer 5 (VISC5)	Home cage vs. Day 1	0.252	0.579	0.972
	Home cage vs. Day 15	0.179	0.579	0.990
	Homecage vs. Day 60	-1.519	0.669	0.130
	Day 1 vs. Day 15	-0.073	0.611	0.999
	Day 1 vs. Day 60	-1.771	0.696	0.076
	Day 15 vs. Day 60	-1.698	0.696	0.094
Anterior cingulate area, ventral part (ACAv)	Home cage vs. Day 1	0.061	0.651	1.000
	Home cage vs. Day 15	0.141	0.651	0.996
	Homecage vs. Day 60	-0.975	0.752	0.572
	Day 1 vs. Day 15	0.080	0.686	0.999
	Day 1 vs. Day 60	-1.036	0.782	0.556
	Day 15 vs. Day 60	-1.116	0.782	0.494
Agranular insular area (AI)	Home cage vs. Day 1	0.354	0.425	0.838
	Home cage vs. Day 15	0.148	0.425	0.985
	Homecage vs. Day 60	-0.864	0.490	0.313
	Day 1 vs. Day 15	-0.206	0.448	0.967
	Day 1 vs. Day 60	-1.218	0.510	0.104
	Day 15 vs. Day 60	-1.012	0.510	0.219
Lateral visual area, layer 4 (VISC4)	Home cage vs. Day 1	0.274	0.370	0.880
	Home cage vs. Day 15	0.178	0.370	0.963
	Homecage vs. Day 60	-0.579	0.427	0.536
	Day 1 vs. Day 15	-0.096	0.390	0.995
	Day 1 vs. Day 60	-0.853	0.445	0.244
	Day 15 vs. Day 60	-0.757	0.445	0.342
Lateral visual area, layer 2/3 (VISC23)	Home cage vs. Day 1	0.369	0.365	0.744
	Home cage vs. Day 15	0.125	0.365	0.986
	Homecage vs. Day 60	-0.371	0.421	0.814
	Day 1 vs. Day 15	-0.244	0.384	0.919
	Day 1 vs. Day 60	-0.740	0.438	0.349
	Day 15 vs. Day 60	-0.496	0.438	0.674
Lateral visual area, layer 1 (VISC1)	Home cage vs. Day 1	0.426	0.312	0.532
	Home cage vs. Day 15	0.262	0.312	0.835
	Homecage vs. Day 60	-0.101	0.360	0.992
	Day 1 vs. Day 15	-0.163	0.329	0.959
	Day 1 vs. Day 60	-0.527	0.375	0.507
	Day 15 vs. Day 60	-0.364	0.375	0.768
Cortical amygdalar area, anterior part (COAa)	Home cage vs. Day 1	-0.106	1.128	1.000
	Home cage vs. Day 15	-2.405	1.128	0.169
	Homecage vs. Day 60	-5.100	1.303	0.003
	Day 1 vs. Day 15	-2.299	1.189	0.239
	Day 1 vs. Day 60	-4.994	1.356	0.005
	Day 15 vs. Day 60	-2.696	1.356	0.218
Piriform area, pyramidal layer (PIR2)	Home cage vs. Day 1	-0.037	1.015	1.000
	Home cage vs. Day 15	-1.509	1.015	0.459
	Homecage vs. Day 60	-4.583	1.172	0.003
	Day 1 vs. Day 15	-1.472	1.070	0.525
	Day 1 vs. Day 60	-4.546	1.220	0.005
	Day 15 vs. Day 60	-3.074	1.220	0.079
Piriform area, polymorph layer (PIR3)	Home cage vs. Day 1	0.257	1.240	0.997
	Home cage vs. Day 15	-1.021	1.240	0.843
	Homecage vs. Day 60	-4.309	1.432	0.027
	Day 1 vs. Day 15	-1.278	1.307	0.763
	Day 1 vs. Day 60	-4.566	1.490	0.024
	Day 15 vs. Day 60	-3.288	1.490	0.147
Cortical amygdalar area, posterior part (COAp)	Home cage vs. Day 1	0.357	0.851	0.975
	Home cage vs. Day 15	-1.475	0.851	0.326
	Homecage vs. Day 60	-3.489	0.982	0.007
	Day 1 vs. Day 15	-1.833	0.897	0.197
	Day 1 vs. Day 60	-3.847	1.022	0.004

Piriform-amyg达尔 area, pyramidal layer (PAA2)	Day 15 vs. Day 60	-2.014	1.022	0.224
	Home cage vs. Day 1	0.361	0.715	0.957
	Home cage vs. Day 15	-0.878	0.715	0.615
	Homecage vs. Day 60	-2.904	0.825	0.008
	Day 1 vs. Day 15	-1.239	0.753	0.371
	Day 1 vs. Day 60	-3.265	0.859	0.004
	Day 15 vs. Day 60	-2.025	0.859	0.110
Piriform-amyg达尔 area, polymorph layer (PAA3)	Home cage vs. Day 1	0.207	0.727	0.992
	Home cage vs. Day 15	-0.835	0.727	0.663
	Homecage vs. Day 60	-2.178	0.839	0.068
	Day 1 vs. Day 15	-1.042	0.766	0.534
	Day 1 vs. Day 60	-2.386	0.873	0.051
	Day 15 vs. Day 60	-1.343	0.873	0.430
Piriform-amyg达尔 area, molecular layer (PAA1)	Home cage vs. Day 1	0.614	0.694	0.813
	Home cage vs. Day 15	-0.322	0.694	0.966
	Homecage vs. Day 60	-1.493	0.801	0.267
	Day 1 vs. Day 15	-0.936	0.732	0.584
	Day 1 vs. Day 60	-2.107	0.834	0.078
	Day 15 vs. Day 60	-1.171	0.834	0.508
Piriform area, molecular layer (PIR1)	Home cage vs. Day 1	0.472	0.543	0.821
	Home cage vs. Day 15	-0.080	0.543	0.999
	Homecage vs. Day 60	-1.407	0.627	0.137
	Day 1 vs. Day 15	-0.552	0.573	0.771
	Day 1 vs. Day 60	-1.880	0.653	0.036
	Day 15 vs. Day 60	-1.328	0.653	0.201
Ammon's horn (CA)	Home cage vs. Day 1	0.068	0.820	1.000
	Home cage vs. Day 15	0.345	0.820	0.975
	Homecage vs. Day 60	-1.155	0.947	0.620
	Day 1 vs. Day 15	0.277	0.865	0.988
	Day 1 vs. Day 60	-1.223	0.986	0.607
	Day 15 vs. Day 60	-1.500	0.986	0.439
Dentate gyrus (DG)	Home cage vs. Day 1	0.288	0.453	0.920
	Home cage vs. Day 15	0.547	0.453	0.628
	Homecage vs. Day 60	-0.098	0.523	0.998
	Day 1 vs. Day 15	0.259	0.478	0.948
	Day 1 vs. Day 60	-0.386	0.545	0.893
	Day 15 vs. Day 60	-0.645	0.545	0.641
Basolateral amygdalar nucleus, anterior part (BMAa)	Home cage vs. Day 1	-1.473	5.942	0.995
	Home cage vs. Day 15	-7.660	5.942	0.577
	Homecage vs. Day 60	-21.841	6.861	0.018
	Day 1 vs. Day 15	-6.186	6.263	0.758
	Day 1 vs. Day 60	-20.368	7.141	0.039
	Day 15 vs. Day 60	-14.181	7.141	0.218
Basolateral amygdalar nucleus, posterior part (BMAp)	Home cage vs. Day 1	-1.094	5.286	0.997
	Home cage vs. Day 15	-7.763	5.286	0.470
	Homecage vs. Day 60	-20.491	6.104	0.012
	Day 1 vs. Day 15	-6.669	5.572	0.634
	Day 1 vs. Day 60	-19.396	6.353	0.024
	Day 15 vs. Day 60	-12.728	6.353	0.212
Endopiriform nucleus, ventral part (EPv)	Home cage vs. Day 1	-0.359	3.557	1.000
	Home cage vs. Day 15	-2.447	3.557	0.901
	Homecage vs. Day 60	-12.575	4.108	0.024
	Day 1 vs. Day 15	-2.088	3.750	0.944
	Day 1 vs. Day 60	-12.215	4.275	0.038
	Day 15 vs. Day 60	-10.127	4.275	0.108
Basolateral amygdalar nucleus (BLA)	Home cage vs. Day 1	0.386	2.051	0.998
	Home cage vs. Day 15	-1.199	2.051	0.936
	Homecage vs. Day 60	-7.035	2.369	0.030
	Day 1 vs. Day 15	-1.585	2.162	0.883
	Day 1 vs. Day 60	-7.421	2.465	0.027
	Day 15 vs. Day 60	-5.836	2.465	0.108

Endopiriform nucleus, dorsal part (EPd)	Home cage vs. Day 1	0.239	1.168	0.997
	Home cage vs. Day 15	-0.963	1.168	0.843
	Homecage vs. Day 60	-5.697	1.349	0.001
	Day 1 vs. Day 15	-1.201	1.231	0.764
	Day 1 vs. Day 60	-5.936	1.404	0.001
	Day 15 vs. Day 60	-4.734	1.404	0.011
Lateral amygdalar nucleus (LA)	Home cage vs. Day 1	0.642	0.867	0.880
	Home cage vs. Day 15	-0.399	0.867	0.967
	Homecage vs. Day 60	-2.729	1.002	0.051
	Day 1 vs. Day 15	-1.041	0.914	0.670
	Day 1 vs. Day 60	-3.371	1.043	0.016
	Day 15 vs. Day 60	-2.330	1.043	0.139
Clastrum (CLA)	Home cage vs. Day 1	0.332	0.529	0.922
	Home cage vs. Day 15	-0.473	0.529	0.808
	Homecage vs. Day 60	-2.585	0.611	0.001
	Day 1 vs. Day 15	-0.805	0.558	0.485
	Day 1 vs. Day 60	-2.917	0.636	<0.001
	Day 15 vs. Day 60	-2.112	0.636	0.013
Intercalated amygdalar nucleus (IA)	Home cage vs. Day 1	-1.038	4.303	0.995
	Home cage vs. Day 15	-4.797	4.303	0.684
	Homecage vs. Day 60	-16.372	4.968	0.014
	Day 1 vs. Day 15	-3.758	4.535	0.840
	Day 1 vs. Day 60	-15.334	5.171	0.030
	Day 15 vs. Day 60	-11.576	5.171	0.138
Medial amygdalar nucleus, anterodorsal part (MEAad)	Home cage vs. Day 1	0.407	2.621	0.999
	Home cage vs. Day 15	-1.729	2.621	0.911
	Homecage vs. Day 60	-8.060	3.027	0.059
	Day 1 vs. Day 15	-2.136	2.763	0.866
	Day 1 vs. Day 60	-8.466	3.150	0.056
	Day 15 vs. Day 60	-6.331	3.150	0.209
Central amygdalar nucleus, capsular part (CEAc)	Home cage vs. Day 1	0.192	1.745	1.000
	Home cage vs. Day 15	-1.233	1.745	0.894
	Homecage vs. Day 60	-6.266	2.015	0.021
	Day 1 vs. Day 15	-1.426	1.839	0.865
	Day 1 vs. Day 60	-6.459	2.097	0.023
	Day 15 vs. Day 60	-5.033	2.097	0.101
Central amygdalar nucleus, medial part (CEAm)	Home cage vs. Day 1	0.352	0.996	0.985
	Home cage vs. Day 15	0.062	0.996	1.000
	Homecage vs. Day 60	-2.730	1.150	0.107
	Day 1 vs. Day 15	-0.290	1.050	0.992
	Day 1 vs. Day 60	-3.081	1.197	0.071
	Day 15 vs. Day 60	-2.792	1.197	0.116
Medial amygdalar nucleus, anteroventral part (MEAav)	Home cage vs. Day 1	0.007	0.757	1.000
	Home cage vs. Day 15	-1.266	0.757	0.357
	Homecage vs. Day 60	-1.731	0.874	0.220
	Day 1 vs. Day 15	-1.273	0.798	0.398
	Day 1 vs. Day 60	-1.738	0.910	0.248
	Day 15 vs. Day 60	-0.465	0.910	0.956
Bed nucleus of the accessory olfactory tract (BA)	Home cage vs. Day 1	0.077	0.695	0.999
	Home cage vs. Day 15	-0.702	0.695	0.745
	Homecage vs. Day 60	-1.485	0.802	0.272
	Day 1 vs. Day 15	-0.779	0.732	0.714
	Day 1 vs. Day 60	-1.562	0.835	0.264
	Day 15 vs. Day 60	-0.783	0.835	0.785
Central amygdalar nucleus, lateral part (CEAl)	Home cage vs. Day 1	0.524	0.542	0.770
	Home cage vs. Day 15	0.391	0.542	0.888
	Homecage vs. Day 60	-1.179	0.626	0.259
	Day 1 vs. Day 15	-0.133	0.572	0.995
	Day 1 vs. Day 60	-1.703	0.652	0.065
	Day 15 vs. Day 60	-1.570	0.652	0.099
Caudoputamen (CP)	Home cage vs. Day 1	0.277	0.369	0.875

	Home cage vs. Day 15	0.447	0.369	0.625
	Home cage vs. Day 60	-0.294	0.426	0.900
	Day 1 vs. Day 15	0.170	0.389	0.971
	Day 1 vs. Day 60	-0.571	0.443	0.578
	Day 15 vs. Day 60	-0.741	0.443	0.358
Pallidum, ventral region (PALv)	Home cage vs. Day 1	0.452	0.706	0.918
	Home cage vs. Day 15	0.546	0.706	0.865
	Home cage vs. Day 60	-1.127	0.815	0.520
	Day 1 vs. Day 15	0.095	0.744	0.999
	Day 1 vs. Day 60	-1.579	0.848	0.268
	Day 15 vs. Day 60	-1.673	0.848	0.223
Globus pallidus, external segment (GPe)	Home cage vs. Day 1	0.136	0.441	0.990
	Home cage vs. Day 15	0.368	0.441	0.837
	Home cage vs. Day 60	-0.482	0.509	0.780
	Day 1 vs. Day 15	0.232	0.465	0.958
	Day 1 vs. Day 60	-0.618	0.530	0.653
	Day 15 vs. Day 60	-0.850	0.530	0.393
Globus pallidus, internal segment (GPi)	Home cage vs. Day 1	0.197	0.451	0.971
	Home cage vs. Day 15	0.513	0.451	0.669
	Home cage vs. Day 60	-0.260	0.521	0.958
	Day 1 vs. Day 15	0.316	0.475	0.909
	Day 1 vs. Day 60	-0.457	0.542	0.833
	Day 15 vs. Day 60	-0.774	0.542	0.494
Central medial nucleus of the thalamus (CM)	Home cage vs. Day 1	0.013	1.224	1.000
	Home cage vs. Day 15	0.288	1.224	0.995
	Home cage vs. Day 60	-2.706	1.413	0.246
	Day 1 vs. Day 15	0.275	1.290	0.996
	Day 1 vs. Day 60	-2.719	1.471	0.273
	Day 15 vs. Day 60	-2.995	1.471	0.200
Anteroventral nucleus of thalamus (AV)	Home cage vs. Day 1	-0.088	0.699	0.999
	Home cage vs. Day 15	0.486	0.699	0.898
	Home cage vs. Day 60	-1.284	0.807	0.400
	Day 1 vs. Day 15	0.574	0.736	0.863
	Day 1 vs. Day 60	-1.196	0.840	0.496
	Day 15 vs. Day 60	-1.770	0.840	0.176
Ventral medial nucleus of the thalamus (VM)	Home cage vs. Day 1	0.101	0.676	0.999
	Home cage vs. Day 15	0.354	0.676	0.953
	Home cage vs. Day 60	-1.182	0.780	0.443
	Day 1 vs. Day 15	0.253	0.712	0.984
	Day 1 vs. Day 60	-1.283	0.812	0.406
	Day 15 vs. Day 60	-1.536	0.812	0.256
Lateral habenula (LH)	Home cage vs. Day 1	0.279	0.454	0.927
	Home cage vs. Day 15	0.479	0.454	0.720
	Home cage vs. Day 60	-0.618	0.525	0.646
	Day 1 vs. Day 15	0.200	0.479	0.975
	Day 1 vs. Day 60	-0.896	0.546	0.374
	Day 15 vs. Day 60	-1.097	0.546	0.210
Anteromedial nucleus (AM)	Home cage vs. Day 1	-0.044	0.404	1.000
	Home cage vs. Day 15	0.347	0.404	0.826
	Home cage vs. Day 60	-0.530	0.466	0.671
	Day 1 vs. Day 15	0.391	0.426	0.795
	Day 1 vs. Day 60	-0.486	0.485	0.751
	Day 15 vs. Day 60	-0.877	0.485	0.293
Submedial nucleus of the thalamus (SMT)	Home cage vs. Day 1	0.161	0.447	0.984
	Home cage vs. Day 15	0.329	0.447	0.882
	Home cage vs. Day 60	-0.479	0.516	0.790
	Day 1 vs. Day 15	0.168	0.471	0.984
	Day 1 vs. Day 60	-0.640	0.537	0.637
	Day 15 vs. Day 60	-0.808	0.537	0.449
Paracentral nucleus (PCN)	Home cage vs. Day 1	0.076	0.364	0.997

	Home cage vs. Day 15	0.330	0.364	0.801
	Home cage vs. Day 60	-0.418	0.420	0.754
	Day 1 vs. Day 15	0.254	0.384	0.910
	Day 1 vs. Day 60	-0.494	0.438	0.675
	Day 15 vs. Day 60	-0.748	0.438	0.338
Mediodorsal nucleus of thalamus (MD)	Home cage vs. Day 1	0.070	0.410	0.998
	Home cage vs. Day 15	0.363	0.410	0.812
	Home cage vs. Day 60	-0.361	0.474	0.871
	Day 1 vs. Day 15	0.293	0.433	0.905
	Day 1 vs. Day 60	-0.432	0.493	0.818
	Day 15 vs. Day 60	-0.725	0.493	0.469
Ventral anterior-lateral complex of the thalamus (VAL)	Home cage vs. Day 1	0.085	0.356	0.995
	Home cage vs. Day 15	0.386	0.356	0.702
	Home cage vs. Day 60	-0.185	0.411	0.969
	Day 1 vs. Day 15	0.301	0.375	0.853
	Day 1 vs. Day 60	-0.270	0.428	0.921
	Day 15 vs. Day 60	-0.571	0.428	0.550
Central lateral nucleus of the thalamus (CL)	Home cage vs. Day 1	0.198	0.330	0.931
	Home cage vs. Day 15	0.405	0.330	0.616
	Home cage vs. Day 60	-0.132	0.381	0.985
	Day 1 vs. Day 15	0.206	0.348	0.933
	Day 1 vs. Day 60	-0.330	0.397	0.839
	Day 15 vs. Day 60	-0.537	0.397	0.539
Ventral posterior complex of the thalamus (VP)	Home cage vs. Day 1	-0.170	0.405	0.974
	Home cage vs. Day 15	0.364	0.405	0.806
	Home cage vs. Day 60	-0.073	0.468	0.999
	Day 1 vs. Day 15	0.534	0.427	0.601
	Day 1 vs. Day 60	0.097	0.487	0.997
	Day 15 vs. Day 60	-0.437	0.487	0.806
Medial habenula (MH)	Home cage vs. Day 1	0.271	0.367	0.880
	Home cage vs. Day 15	0.603	0.367	0.373
	Home cage vs. Day 60	-0.025	0.424	1.000
	Day 1 vs. Day 15	0.331	0.387	0.827
	Day 1 vs. Day 60	-0.297	0.441	0.906
	Day 15 vs. Day 60	-0.628	0.441	0.496
Reticular nucleus of the thalamus (RT)	Home cage vs. Day 1	-0.001	0.338	1.000
	Home cage vs. Day 15	0.391	0.338	0.659
	Home cage vs. Day 60	0.047	0.390	0.999
	Day 1 vs. Day 15	0.392	0.356	0.692
	Day 1 vs. Day 60	0.048	0.406	0.999
	Day 15 vs. Day 60	-0.344	0.406	0.831
Anterodorsal nucleus (AD)	Home cage vs. Day 1	0.314	0.313	0.749
	Home cage vs. Day 15	0.579	0.313	0.273
	Home cage vs. Day 60	0.066	0.362	0.998
	Day 1 vs. Day 15	0.265	0.330	0.852
	Day 1 vs. Day 60	-0.248	0.376	0.911
	Day 15 vs. Day 60	-0.514	0.376	0.531
Lateral dorsal nucleus of thalamus (LD)	Home cage vs. Day 1	0.211	0.300	0.895
	Home cage vs. Day 15	0.473	0.300	0.408
	Home cage vs. Day 60	0.222	0.346	0.918
	Day 1 vs. Day 15	0.262	0.316	0.841
	Day 1 vs. Day 60	0.011	0.360	1.000
	Day 15 vs. Day 60	-0.251	0.360	0.897
Anterior hypothalamic nucleus (AHN)	Home cage vs. Day 1	0.281	6.307	1.000
	Home cage vs. Day 15	-2.952	6.307	0.965
	Home cage vs. Day 60	-17.598	7.283	0.098
	Day 1 vs. Day 15	-3.232	6.649	0.962
	Day 1 vs. Day 60	-17.878	7.581	0.110
	Day 15 vs. Day 60	-14.646	7.581	0.239
Hypothalamic medial zone (MEZ)	Home cage vs. Day 1	0.011	5.940	1.000

	Home cage vs. Day 15	-4.111	5.940	0.899
	Home cage vs. Day 60	-17.359	6.859	0.078
	Day 1 vs. Day 15	-4.122	6.261	0.912
	Day 1 vs. Day 60	-17.369	7.139	0.095
	Day 15 vs. Day 60	-13.248	7.139	0.271
Paraventricular hypothalamic nucleus (PVH)	Home cage vs. Day 1	-0.083	5.886	1.000
	Home cage vs. Day 15	-1.804	5.886	0.990
	Home cage vs. Day 60	-14.620	6.797	0.163
	Day 1 vs. Day 15	-1.721	6.204	0.992
	Day 1 vs. Day 60	-14.537	7.074	0.194
	Day 15 vs. Day 60	-12.816	7.074	0.290
Lateral hypothalamic area (LHA)	Home cage vs. Day 1	0.337	1.425	0.995
	Home cage vs. Day 15	-0.964	1.425	0.905
	Home cage vs. Day 60	-3.861	1.646	0.112
	Day 1 vs. Day 15	-1.300	1.502	0.822
	Day 1 vs. Day 60	-4.198	1.713	0.091
	Day 15 vs. Day 60	-2.898	1.713	0.347
Arcuate hypothalamic nucleus (ARH)	Home cage vs. Day 1	0.323	0.509	0.920
	Home cage vs. Day 15	-0.423	0.509	0.839
	Home cage vs. Day 60	-1.414	0.588	0.100
	Day 1 vs. Day 15	-0.746	0.537	0.516
	Day 1 vs. Day 60	-1.737	0.612	0.040
	Day 15 vs. Day 60	-0.991	0.612	0.385
Tuberal nucleus (TU)	Home cage vs. Day 1	0.256	0.533	0.963
	Home cage vs. Day 15	-0.563	0.533	0.718
	Home cage vs. Day 60	-1.319	0.615	0.165
	Day 1 vs. Day 15	-0.818	0.562	0.476
	Day 1 vs. Day 60	-1.574	0.640	0.090
	Day 15 vs. Day 60	-0.756	0.640	0.644
Supraoptic nucleus (SO)	Home cage vs. Day 1	0.302	0.662	0.968
	Home cage vs. Day 15	-0.103	0.662	0.999
	Home cage vs. Day 60	-0.713	0.764	0.787
	Day 1 vs. Day 15	-0.405	0.697	0.937
	Day 1 vs. Day 60	-1.015	0.795	0.585
	Day 15 vs. Day 60	-0.610	0.795	0.868

## SI References

1. D. Funk *et al.*, Role of Central Amygdala Neuronal Ensembles in Incubation of Nicotine Craving. *J Neurosci* 36, 8612-8623 (2016).
2. M. Venniro *et al.*, Volitional social interaction prevents drug addiction in rat models. *Nat Neurosci* 21, 1520-1529 (2018).
3. X. Li *et al.*, Role of Anterior Intralaminar Nuclei of Thalamus Projections to Dorsomedial Striatum in Incubation of Methamphetamine Craving. *J Neurosci* 38, 2270-2282 (2018).
4. J. W. Grimm *et al.*, Effects of acute or chronic environmental enrichment on regional Fos protein expression following sucrose cue-reactivity testing in rats. *Brain Struct Funct* 221, 2817-2830 (2016).
5. R. D. Altshuler *et al.*, Role of orbitofrontal cortex in incubation of oxycodone craving in male rats. *Addict Biol* 26, e12927 (2021).
6. I. R. Davis, S. A. Coldren, X. Li, Methamphetamine seeking after prolonged abstinence is associated with activated projections from anterior intralaminar nucleus of thalamus to dorsolateral striatum in female rats. *Pharmacol Biochem Behav* 200, 173087 (2021).
7. S. Fanous *et al.*, Role of orbitofrontal cortex neuronal ensembles in the expression of incubation of heroin craving. *J Neurosci* 32, 11600-11609 (2012).
8. C. A. Blackwood, M. Leary, A. Salisbury, M. T. McCoy, J. L. Cadet, Escalated Oxycodone Self-Administration Causes Differential Striatal mRNA Expression of FGFs and IEGs Following Abstinence-Associated Incubation of Oxycodone Craving. *Neuroscience* 415, 173-183 (2019).
9. X. Li *et al.*, Incubation of methamphetamine craving is associated with selective increases in expression of Bdnf and trkB, glutamate receptors, and epigenetic enzymes in cue-activated fos-expressing dorsal striatal neurons. *J Neurosci* 35, 8232-8244 (2015).
10. D. Caprioli *et al.*, Role of dorsomedial striatum neuronal ensembles in incubation of methamphetamine craving after voluntary abstinence. *J. Neurosci.* 37, 1014-1027 (2017).
11. L. M. Rossi *et al.*, Role of nucleus accumbens core but not shell in incubation of methamphetamine craving after voluntary abstinence. *Neuropsychopharmacology* 45, 256-265 (2020).
12. M. Venniro *et al.*, Abstinence-dependent dissociable central amygdala microcircuits control drug craving. *Proc Natl Acad Sci U S A* 117, 8126-8134 (2020).