

1 **Variation in albumin glycation rates in birds suggests**
2 **resistance to relative hyperglycaemia rather than**
3 **conformity to the pace of life syndrome hypothesis**

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25 ABSTRACT

26 The pace of life syndrome hypothesis (POLS) suggests that organisms' life history, physiological
27 and behavioural traits should co-evolve. In this framework, how glycaemia (i.e., blood glucose
28 levels) and its reaction with proteins and other compounds (i.e. glycation) covary with life history
29 traits remain relatively under-investigated, despite the well documented consequences of
30 glucose and glycation on ageing, and therefore potentially on life history evolution. Birds are
31 particularly relevant in this context given that they have the highest blood glucose levels within
32 vertebrates and still higher mass-adjusted longevity when compared to organisms with similar
33 physiology as mammals. We thus performed a comparative analysis on glucose and albumin
34 glycation rates of 88 bird species from 22 orders, in relation to life history traits (body mass,
35 clutch mass, maximum lifespan and developmental time) and diet. Glucose levels correlated
36 positively with albumin glycation rates in a non-linear fashion, suggesting resistance to glycation
37 in species with higher glucose levels. Plasma glucose levels decreased with increasing body mass
38 but, contrary to what is predicted to the POLS hypothesis, glucose levels increased with
39 maximum lifespan before reaching a plateau. Finally, terrestrial carnivores showed higher
40 albumin glycation compared to omnivores despite not showing higher glucose, which we discuss
41 may be related to additional factors as differential antioxidant levels or dietary composition in
42 terms of fibres or polyunsaturated fatty acids. These results increase our knowledge about the
43 diversity of glycaemia and glycation patterns across birds, pointing towards the existence of
44 glycation resistance mechanisms within comparatively high glycaemic birds.

45 INTRODUCTION

46 The pace of life (POL) hypothesis [1-3] postulates that organisms behavioural/physiological
47 characteristics and life histories have co-evolved in answer to specific ecological conditions
48 forming pace of life syndromes (POLS) along a fast-slow continuum (but see [4] for the more
49 recent consideration of other axes). According to the classical approach from life history theory,
50 slower organisms would have, e.g. large body mass, late maturation, slow growth rate, small
51 number of large offspring and high longevity, with faster ones representing the opposite trait
52 values (see [5-8]) within this continuum. Pace of life hypothesis added physiology to this
53 continuum, with studies testing its predictions focusing often on metabolic rate (i.e. higher or
54 lower metabolic rate corresponding to faster or slower pace of life, respectively; see e.g. [9-12]).
55 However, key energy substrates related to metabolic performance, such as glucose

56 concentrations in tissues, have been largely overlooked (but see [13-16]). Notably, glucose, a
57 central energy source for many organisms, plays a pivotal role in metabolism and there are some
58 indications that its circulating blood or plasma levels correlate positively with metabolic rate per
59 gram of mass at the interspecific level [13,17], and with whole body metabolism at the
60 intraspecific level ([18] for the case of non-diabetic humans).

61 Research focusing on plasma glucose becomes highly relevant as high glycaemia can entail costs
62 that accelerate ageing. Along with other reducing sugars, glucose can react non-enzymatically
63 with free amino groups of proteins, lipids or nucleic acids, a reaction known as glycation [19][20].
64 This reaction, after several molecular reorganizations, can lead to the formation of a plethora of
65 molecules called Advanced Glycation End-products (AGEs) [21]. AGEs can form aggregates in
66 tissues that are well known to contribute to several age-related diseases, such as diabetes (e.g.,
67 reviews by [22-25]). AGEs are also known to promote a proinflammatory state through their
68 action on specific receptors called RAGEs [26-27].

69 Remarkably, birds show circulating glucose levels much higher than other vertebrate groups, on
70 average almost twice as high as mammals [28]. These relatively high glucose levels might
71 support the higher metabolic rates [29, 11] and body temperatures observed in birds compared
72 to mammals [30-31]. In addition, elevated glycaemia is thought to be an adaptation to flight,
73 providing birds with rapid access to an easily oxidizable fuel during intense bursts of aerobic
74 exercise (see [32-33]), such as during take-off and short-term flapping flights [34-35]). However,
75 the presence of high glycaemia in birds is also paradoxical, given their remarkable longevity
76 compared to their mammalian counterparts — living up to three times longer than mammals of
77 equivalent body mass [36, 11].

78 Several non-excluding hypotheses have been proposed to explain how birds apparently resist
79 the pernicious effects of a high glycaemic state. One possibility is that birds might resist protein
80 glycation through a lower exposure of lysine residues at the surface of their proteins [37].
81 Another hypothesis suggests that increased protein turnover may play a role [38-39].
82 Additionally, birds might benefit from more effective antioxidant defences [40-42] (although
83 within Passeriformes, [16] shows no coevolution between glycaemia and antioxidant defences)
84 or even the presence of “glycation scavengers” (i. e. molecules that bind glucose avoiding it to
85 react with proteins) such as polyamines which circulate at high concentration [35]. Moreover, it
86 is generally considered that RAGEs are not present in avian tissues, suggesting possible adaptive
87 mechanisms that would allow birds to avoid the inflammatory consequences associated with
88 RAGE activation [43-44]. However, a putative candidate for RAGE and a case of AGE-induced

89 inflammation in birds were identified by Wein et al. [45], warranting further investigation on the
90 occurrence of RAGEs in birds.

91 To date, although protein glycation levels have been measured in birds in several species, these
92 results were mostly descriptive and concerned a small number of species [46-54]. Additionally,
93 many of these studies employ commercial kits that were not specifically designed for avian
94 species, making it challenging to interpret the results [53]. These limitations restrict our ability
95 to draw general conclusions from these studies. Here, we performed a comparative analysis on
96 primary data (for glycation and most of glucose values, see methods and ESM6) from 88 bird
97 species belonging to 22 orders (see ESM6) and assessed whether and how bird glycaemia and
98 glycation rates are linked to ecological and life-history traits (including body mass). While doing
99 this, we also checked whether glycation levels are influenced by circulating plasma glucose
100 concentrations. Finally, as diet can influence glycaemia (see e.g. [15, 55-56] but [14] found no
101 significant effects) and glycation, we included it in our analyses. We hypothesized that
102 carnivorous birds would exhibit higher glycaemia and glycation levels than omnivorous or
103 herbivorous species, as diets with low-carbohydrates, high-proteins and high-fat are associated
104 with comparatively high glycaemia and reduced insulin sensitivity in some vertebrates (e.g.; [57-
105 59]). Accordingly, a recent comparative study in birds showed higher blood glucose levels in
106 carnivorous species [56]. This may be attributed to high levels of constitutive gluconeogenesis,
107 which has been confirmed in certain raptors [60-61]. Conversely, bird species with high sugar
108 intake, such as frugivores or nectarivores, are expected to exhibit high glycaemia, as observed
109 in hummingbirds [49] and Passeriformes with such diet [15] (although the opposite was found
110 in [56]). In line with POLS hypothesis, we also hypothesized that species with a slower POL should
111 exhibit a lower glycaemia and higher glycation resistance (glycation levels for a given glycaemia)
112 compared to species with a faster POL. Hence, we included the following four parameters in our
113 analyses of glycaemia and glycation levels: body mass, maximum lifespan, clutch mass and
114 developmental time. Body mass is one of the main factors underlying life history, with higher
115 body mass associated with “slower” strategies and vice versa [62]. Previous studies have
116 reported a negative relationship between body mass and glycaemia [63, 34, 13-14, 56](but see
117 [49, 64] with non-significant trends in birds, and [15] only for temperate species within
118 Passeriformes). Therefore, we predicted that bird species that live longer, develop more slowly
119 and invest less per reproductive event (see ESM6 for further justification of the chosen variables)
120 should show lower plasma glucose levels and albumin glycation rates (after controlling for body
121 mass, phylogeny, diet and glucose in the case of glycation; see methods).

122 **MATERIALS AND METHODS**

123 **Species and sample collection**

124 A total of 484 individuals from 88 species measured were included for this study (see **Table**
125 **ESM6. 1**). A detailed list with the provenance of the samples, including zoos, a laboratory
126 population, and both designated captures and samples provided by collaborators from wild
127 populations, is provided in ESM3, with a textual description in ESM6.
128 Blood samples were collected from the brachial or tarsal vein, or from the feet in the case of
129 swifts, using heparin-lithium capillaries or Microvette® (Sarstedt). As samples were collected
130 mostly by different collaborators, handling times were not always recorded and could not be
131 adjusted for, potentially rendering the results more conservative (for a more detailed discussion
132 on potential stress effects on glucose, see ESM6). Samples were centrifuged at 4°C, 3500 g for
133 10 min and plasma was aliquoted when a large volume was available. Subsequently, they were
134 transported on dry ice to the IPHC (Institut Pluridisciplinaire Hubert Curien) in Strasbourg and
135 stored at -80° C until analysis. Glycaemia was measured in the laboratory on the remaining
136 plasma after taking an aliquot for glycation assessment, using a Contour Plus® glucometer
137 (Ascensia diabetes solutions, Basel, Switzerland) and expressed in mg/dL. These of point-of-care
138 devices have previously been assessed for its usage in birds [65-66], with several examples of its
139 usage in recent literature related to the topic presented here (e.g. [14, 67-69]. We also
140 performed an assay with this particular brand on a subset of 46 samples from this study, coming
141 from nine species distributed across the whole range of glucose values (three species with "low"
142 values, three with "medium" and three with "high", from both captive and wild populations,
143 with five individuals per species, except one including six), confirming a positive linear
144 correlation ($R^2_{\text{marginal}}=0.66$; $R^2_{\text{conditional}}=0.84$, for a model including the species as random factor)
145 of this device with Randox GLUC-PAP colourimetry kits (P-value<0.001, unpublished data). Due
146 to technical issues, related to the incapability of the device to determine certain glucose values
147 (not because of the glucose concentration, but perhaps the particular composition of the plasma
148 samples from certain species), we could not determine glycaemia values of 95 individuals of
149 those that we sampled and in which glycation levels were assessed (belonging to 40 species
150 coming from different sources). For these species, if not a single individual had a glucose
151 measurement (which was the case in 13 species), we obtained mean plasma glucose values
152 reported for the species from the ZIMS database from Species360 (Species360 Zoological

153 Information Management System (ZIMS) (2023), zims.Species360.org). This database provides
154 plasma glucose data measured by colorimetric glucose kits on zoo specimens not necessary
155 corresponding to those in which we measured glycation values. The sample sizes for both
156 glucose and glycation measurements, either from our measured individuals or from the ones
157 from ZIMs, are reported on the ESM5.

158 **Glycation levels**

159 Glycation levels for each individual were determined using liquid chromatography coupled to
160 mass spectrometry (LC-MS), which is considered the gold standard for the assessment of protein
161 glycation levels (see e.g. [70]) and has previously been used for birds [52-53, 71]. Given the
162 relatively high time intrinsically taken by the employed methodology for analysing all the
163 samples, linked to constraints in the access to the mass spectrometry devices, the whole set of
164 samples of this study were analysed across several instances within a total timespan of less than
165 2 years (2021-2023). Only one sample from each individual was measured, given logistic
166 limitations on the total number of samples that could be processed. Briefly, 3 µl of plasma were
167 diluted with 22 µl of distilled water containing 0.1% of formic acid, followed by injection of 5 µl
168 into the system. The glycation values used in the analyses represent the total percentage of
169 glycated albumin, obtained by adding the percentages of singly and doubly glycated albumin.
170 These percentages were calculated by dividing the areas of the peaks corresponding to each
171 glycated molecule form (albumin plus one or two glucose) by the total albumin peak area (sum
172 of the areas of glycated plus non-glycated molecules) observed in the spectrograms obtained
173 from the mass spectrometry outcomes. These spectrograms represent the different intensities
174 of signal for the components differentiated in the sample by their TOF (Time Of Flight), which
175 depends on the mass to charge ratio (m/z) of the ionized molecules. More detailed information
176 regarding functioning of the method and data processing can be found in [53]. In cases where
177 albumin glycation values dropped below the limit of detection, resulting in missing data, these
178 individuals (10 individuals from 4 species) were excluded from the statistical analyses, as
179 outlined below.

180 **Data on ecology and life-history**

181 Diets of individual species were extracted from the AVONET dataset ([72], coming from [73], and
182 those adapted from [74]) and life history traits from the Amniote Database [75]. After

183 determining that AVONET diet classifications did not align with our research needs, minor
184 changes were made after consulting the original Wilman et al. [74] database (see ESM6).
185 For missing data on life history traits after this stage, we extracted values, in order of preference,
186 from the AnAge database [76] or the Birds of the World encyclopaedia [77], by calculating mean
187 values if an interval was given, and then averaging if male and female intervals were provided
188 separately. For European species, maximum lifespan records have always been checked against
189 the most recent Euring database [78], and from *Chionis minor* and *Eudyptes chrysophrys* (not
190 available from the previous sources) they were extracted from the Centre d'Etudes Biologiques
191 de Chizé, Centre National de la Recherche Scientifique (CNRS-CEBC) database. Several species
192 from the zoo had maximum lifespan values available on the ZIMs from Species 360 (Species360
193 Zoological Information Management System (ZIMS) (2023), zims.Species360.org), so these were
194 also compared with the data we had from other sources.
195 In the case of variables (other than maximum lifespan) for which two different sources provided
196 different records, an average value was calculated. For the maximum lifespan value, available
197 sources were cross-checked and the highest value was always used. In the absence of
198 satisfactory support from another source, values for maximum lifespan indicated as being
199 anecdotal and of poor quality have been excluded from the analyses. A table with the adequate
200 citations for each value is provided as part of the online available data (see Data accessibility
201 section). When the source is AVONET [72], this database is cited, and when it is Amniote
202 Database [75], we cite the sources provided by them, so the references can be checked in [75].
203 A list with all the additional references not coming from any of these databases or not provided
204 by their authors (and that are not already in the main text) is given as ESM4.

205 **Statistical analyses**

206 All analyses were performed in R v.4.3.2 [79]. Alpha (α) ≤ 0.05 was reported as significant, and
207 $0.05 < \alpha \leq 0.1$ as trends. General Linear Mixed Models with a Bayesian approach (*MCMCglmm*
208 function in R; [80]) were performed. All models were run with 6×10^6 iterations, with a thinning
209 interval of 100 and a burn-in of 1000. The models were simplified by eliminating quadratic terms
210 where they were not significant and selecting models with lower AIC (Akaike Information
211 Criterion) and BIC (Bayesian Information Criterion). Gaussian distribution of the response
212 variables was assumed (after \log_{10} conversion for glucose; see ESM6 for further discussions). The
213 priors were established assuming variance proportions of 0.2 for the G matrix and 0.8 for the R
214 matrix, with $v=2$. Less informative priors with equal variance proportions for each partition

215 (residual and random) gave similar results. Lower v values (0.1, 0.2, 0.002) were also tested,
216 without success (i.e. simulations aborted before the established number of iterations). For the
217 phylogenetically controlled analyses, consensus trees (each with the species included in the
218 model) were obtained by using the *consensus.edges* function (from the *phytools* package from
219 R [81] from a total of 10,000 trees downloaded from Birdtree.org [82] (option “Hackett All
220 species” [83]). For such purpose, a list of species with the names adapted to the synonyms
221 available in such website was used (see ESM5), including the change of *Leucocarbo verrucosus*
222 for *Phalacrocorax campbelli*, as the former was not available and it was the only species from
223 the order Suliformes in our dataset, so that neither the position in the tree nor the branch length
224 would be affected by this change. The consensus trees were included as *pedigree* in the models.
225 We performed models with either glucose or glycation as dependent variables and with the diet,
226 body mass and life history traits as predicting variables (and glucose in the models with glycation
227 as a response to assess glycation resistance; see introduction; another set of models without
228 glucose were performed to test if there was a covariation of glycation itself, independently of
229 glucose levels, covaried with life-history; see ESM6). Generalized Variance Inflation Factors
230 (GVIFs) were calculated for all the models with more than one predictor variable to assess the
231 collinearity of them, as it may be expected for life history traits (see results on ESM1),
232 considering values above 1.6 as slightly concerning, above 2.2 as moderately concerning and
233 above 3.2 as severely concerning (i.e. indicating high collinearity; following [84]). Models testing
234 the effects of age and sex on glucose and glycation levels and the number of exposed lysine
235 residues on glycation were also carried out. Finally, we performed models on glucose and
236 glycation values controlling for the taxonomic orders included in the dataset (see ESM5). The
237 effects of phylogeny on all models was determined by calculating the ratio of the variance
238 estimated for the “animal” variable, representing the *pedigree* (see ESM6) by the total variance
239 (all random factors plus *units*). A thorough description of all of the models, including
240 transformations of the variables and other details is given on the ESM6.

241 **RESULTS**

242 **Plasma glucose and albumin glycation variability across the birds’ tree**

243 Plasma glucose and albumin glycation values varied considerably across species (**Figure 1**).
244 Considerable within species repeatability (see ESM6) was observed (Glucose: $R = 0.716$, $SE =$

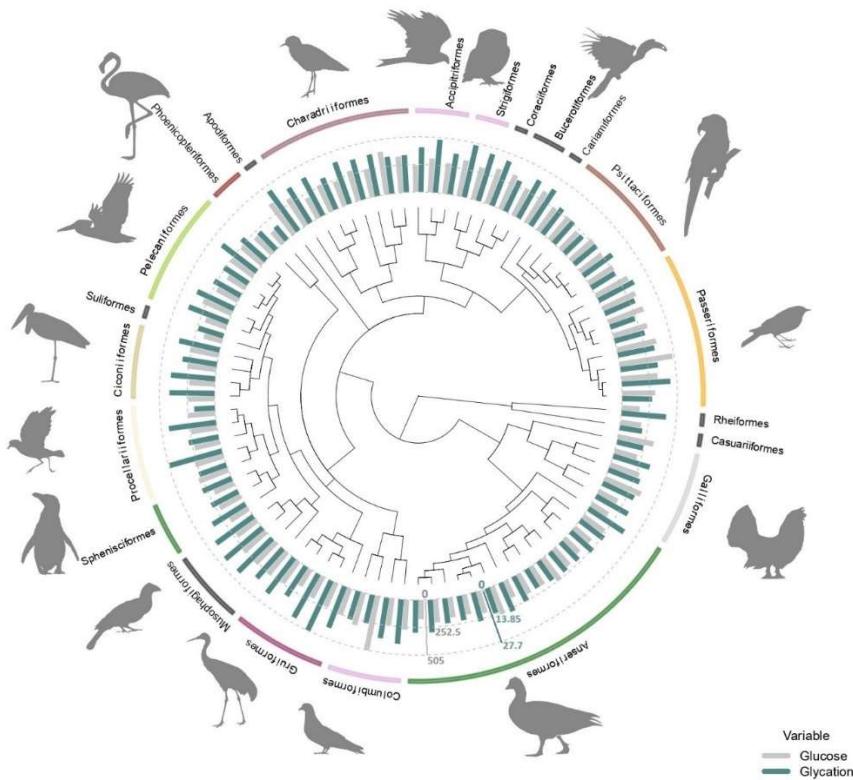
245 0.042, $CI_{95} = [0.619, 0.785]$, P-value <0.0001; Glycation: $R = 0.703$, $SE = 0.042$, $CI_{95} = [0.603,$
246 $0.768]$, P-value <0.0001).

247 We found significant differences between some orders for both glucose and albumin glycation.
248 ESM1 shows the raw outcomes of the models, showing such significant differences to the
249 intercept and the Credible Intervals to perform the pairwise comparisons across all groups. To
250 explore some of the details, we can see for example how Apodiformes showed the highest
251 average glucose values (species average model: Estimated mean=351.8 mg/dL, $CI_{95}[238.9,$
252 514.2]; model with individuals: Estimated mean=349.8 mg/dL, $CI_{95}[251.8, 476.7]$, n=39
253 individuals, 1 species), followed by Passeriformes (species average model: Estimated
254 mean=349.2 mg/dL, $CI_{95}[279.6, 438.6]$; model with individuals: Estimated mean=337.6 mg/dL,
255 $CI_{95}[274.5, 414.5]$, n=84 individuals, 8 species), while the Suliformes (species average model:
256 Estimated mean=172 mg/dL, $CI_{95}[117.6, 252.9]$; model with individuals: Estimated mean=169.5
257 mg/dL, $CI_{95}[120.9, 239.7]$, n=5 individuals, 1 species), Rheiformes (species average model:
258 Estimated mean=173.7 mg/dL, $CI_{95}[118, 254.7]$; model with individuals: Estimated mean=172
259 mg/dL, $CI_{95}[123.7, 239.8]$, n=9 individuals, 1 species) and Phoenicopteriformes (species average
260 model: Estimated mean=176 mg/dL, $CI_{95}[130, 241.1]$; model with individuals: Estimated
261 mean=141.2 mg/dL, $CI_{95}[100.4, 198.2]$, n=5 individuals, 1 species) show the lowest average
262 values (for the species average models, the pattern is
263 Suliformes<Rheiformes<Phoenicopteriformes, while for the individual models is
264 Phoenicopteriformes<Suliformes<Rheiformes).

265 For glycation, Strigiformes (species average model: Estimated mean=25.6 %, $CI_{95}[20, 31]$; model
266 with individuals: Estimated mean=25.3 %, $CI_{95}[19.1, 31.5]$, n=7 individuals, 2 species),
267 Apodiformes (species average model: Estimated mean=25.5 %, $CI_{95}[18.6, 32.4]$; model with
268 individuals: Estimated mean=24.8 %, $CI_{95}[17.9, 32]$, n=40 individuals, 1 species) and
269 Coraciiformes (species average model: Estimated mean=24.5 %, $CI_{95}[17.9, 31.6]$; model with
270 individuals: Estimated mean=23.9 %, $CI_{95}[15.8, 32]$, n=2 individuals, 1 species), all terrestrial
271 carnivores as by our sampled species (see discussion below), had the highest average.

272 On the other hand, Casuariiformes (species average model: Estimated mean=10.8 %, $CI_{95}[3.7,$
273 18.1]; model with individuals: Estimated mean=11.4 %, $CI_{95}[2.7, 20.6]$, n=1 individuals, 1
274 species), Phoenicopteriformes (species average model: Estimated mean=11.2 %, $CI_{95}[5.6, 16.7]$;
275 model with individuals: Estimated mean=10.5 %, $CI_{95}[4.6, 16.3]$, n=22 individuals, 2 species) and
276 Suliformes (species average model: Estimated mean=13.8 %, $CI_{95}[7, 20.7]$; model with

277 individuals: Estimated mean=13.2 %, CI₉₅[5.6, 20.5], n=5 individuals, 1 species) had the lowest
278 average levels for the species average models, the pattern is
279 Casuariiformes<Phoenicopteriformes<Suliformes, while for the individual models is
280 Phoenicopteriformes<Casuariiformes<Suliformes). Graphs with raw data on species average
281 and individual glucose and glycation values by order are shown on ESM2 (**Figure ESM2.3**).
282 Estimates of phylogeny effects on the residuals of the models on glucose and glycation values
283 differ if we consider the models with intraspecific variability or without it, but not so much
284 between the models with and without life history traits (within the previous categories). In the
285 case of species averages, the effect of the tree on residual glucose variation is lower and the
286 estimation is less precise than for residual glycation variation, while for the models considering
287 individual values, it is glycation residuals what shows lower levels of tree-related variance than
288 glucose residuals (see ESM1).



289
290 **Figure 1.** Average plasma glucose values in mg/dL (in grey) and average albumin glycation rate as a
291 percentage of total albumin (in blue) from all the species used in this study (some of them with glucose
292 values coming from ZIMs database; see methods) with the orders they belong to. Glucose and glycation
293 values are standardized in order to be compared, with the dotted lines representing half the maximum
294 and maximum values for each variable (as indicated by the axes in their corresponding colours), from
295 inside out. Tree from a consensus on 10,000 trees obtained from “Hackett All species” on Birdtree.org,
296 including 88 species from 22 orders (see methods).

297 **Bird glycemia appears related to body mass and maximum lifespan**

298 After controlling for intraspecific variation in our analyses, we found that variation in glucose
299 levels were significantly explained by variations in body mass and both the linear and quadratic
300 component of residual maximum lifespan (i.e. mass and phylogeny-adjusted maximum lifespan),
301 but not by variations in diet (**Figure 4.A** with predictions from the model without life history
302 traits and **Figure ESM2.2.A** with raw individual data from the same dataset), clutch mass and
303 developmental time (see **Table 1** for the model including life-history trait variables). Heavier
304 species had lower glucose levels (see **Figure 2.A**, drawn with the estimates from the model
305 without life history traits, which includes more species). Glucose levels increase with increasing
306 mass-adjusted lifespan until reaching a plateau (**Figure 2.B**). Models that did not consider
307 intraspecific variability show no significant effect of any of the aforementioned variables on
308 glucose levels (see ESM1). The provenance of the samples (wild versus captive) only showed a
309 trend to a higher glucose levels in the samples from captive individuals in the model without life
310 history traits (Estimate = 0.058; CI₉₅[-0.008, 0.125]; P-value=0.083).

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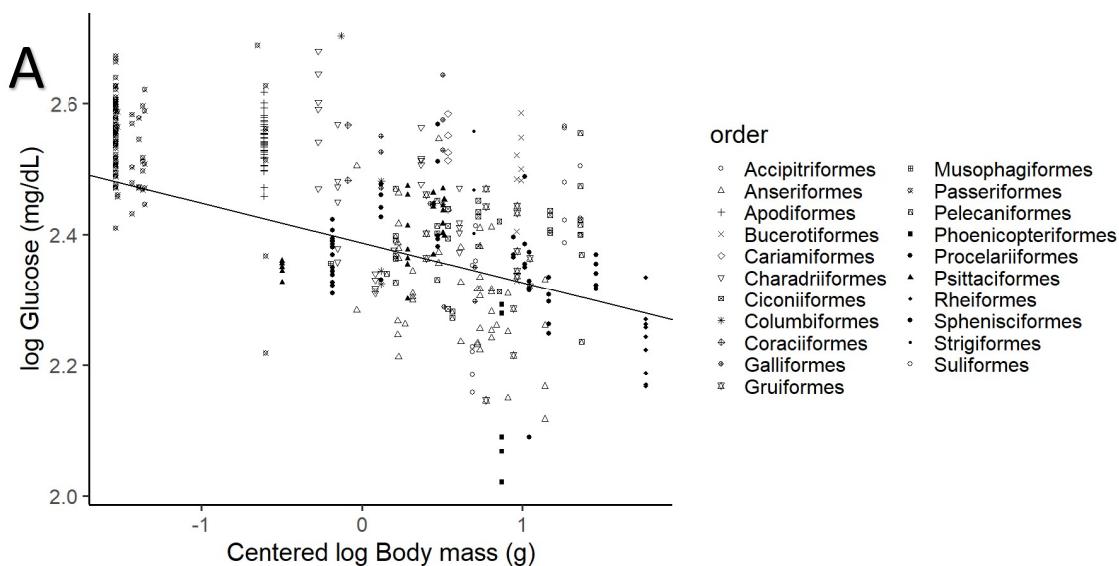
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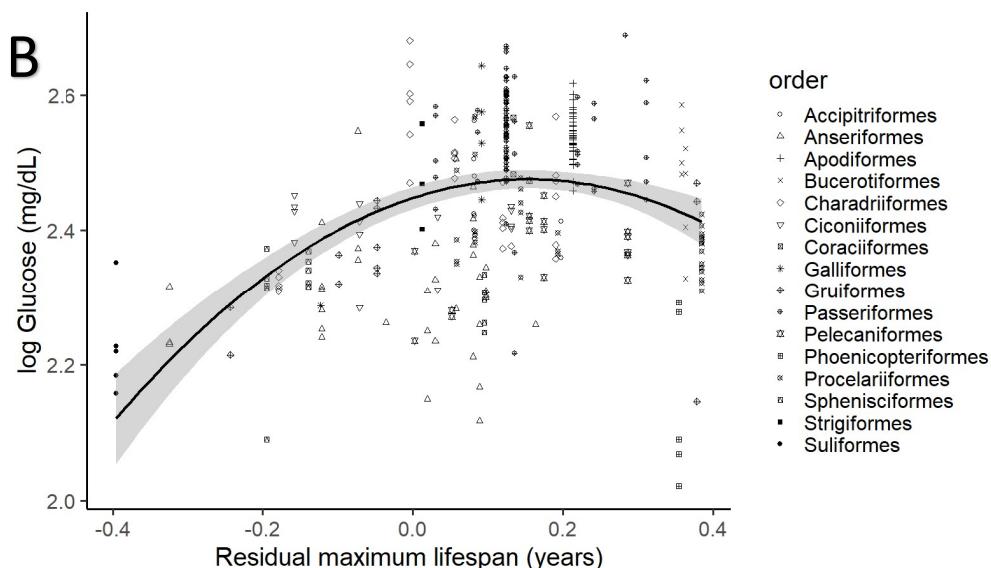
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322 **Table 1.** Final glucose model with diet, body mass, life history traits and sample provenance (wild versus
323 captive; see ESM6) as explanatory variables, including the significant quadratic effect of maximum
324 lifespan. Posterior means, CI_{95} and pMCMC from a phylogenetic MCMC GLMM model including n=326
325 individuals of 58 species. Both glucose and body mass are \log_{10} transformed and life history traits are
326 residuals from a pGLS model of \log_{10} body mass and \log_{10} of the trait in question (see ESM6). Body mass
327 was also centred to better explain the intercept, as 0 body mass would make no biological sense. The
328 intercept corresponds to the Omnivore diet, being used as the reference as it is considered the most
329 diverse and “neutral” group for this purpose. Significant predictors are indicated in bold.

	Estimates	Lower 95% CI	Upper 95% CI	Samplin g effort	pMCMC
Intercept (Omnivore)	2.387	2.268	2.5	59900	<0.001
Diet: Carnivore terrestrial	0.02	-0.056	0.099	59900	0.592
Diet: Aquatic predator	0.034	-0.047	0.112	59900	0.393
Diet: Herbivore	-0.053	-0.177	0.07	59900	0.393
Diet: Frugivore/granivore	-0.055	-0.239	0.135	59900	0.558
Centred \log_{10} Body mass	-0.061	-0.106	-0.015	59900	0.009
Maximum lifespan	0.107	-0.035	0.253	59900	0.142
Maximum lifespan ²	-0.616	-1.166	-0.095	59900	0.026
Clutch mass	-0.095	-0.265	0.069	59900	0.258
Developmental time	0.011	-0.185	0.212	59900	0.916
Provenance: Captive	0.034	-0.039	0.106	59900	0.346



332



333

334 **Figure 2.** Plasma glucose levels (in mg/dL) variation as a function of **A** species mean centred body mass
 335 and **B** residual maximum lifespan. Both glucose and body mass are log transformed. Maximum lifespan
 336 (in years) is given as the residues of a pGLS model with body mass (in grams), both \log_{10} transformed, so
 337 the effects of body mass on longevity are factored out (see ESM6). Different bird orders, are indicated by
 338 symbols, as specified on the legends at the right side of the graphs. **Figure 2.A** uses the values and
 339 estimates from the glucose model without life history traits (n=389 individuals from 75 species), while
 340 **Figure 2.B** uses only the data points employed on the complete model (n=326 individuals of 58 species).

341

342

343 **Bird albumin glycation is related to glycemia and diet**

344 After controlling for intraspecific variability, we found that diet was affecting variation in
345 albumin glycation rates, with terrestrial carnivorous species having higher glycation levels than
346 omnivorous species (**Table 2** for complete model; see **Figure 4.B** with predictions of the model
347 without life history traits and **Figure ESM2.2.B** with raw individual data from the same dataset).
348 However, for the models for species averages, there was only a trend on this pattern in the one
349 including life history traits, and no effect in the other (see ESM1). The relation between glycation
350 and glucose levels was positive and significant in all but the model that included life history traits
351 but not intraspecific variation (see **Table 2** for the outcome of the model with individual values
352 and life history traits and ESM1 for the rest; see **Figure 3** with estimates from the model including
353 individual variation but no life history traits, as it contains more species and the estimates are
354 similar). Given the logarithmic relationship between glycation and glucose (see ESM6), the slope
355 lower than one (see **Table 2**) implies that birds with higher glucose levels have relatively lower
356 albumin glycation rates for their glucose, fact that we would be referring to as higher glycation
357 resistance. The glycation models excluding glucose levels, and therefore testing for covariates
358 of life-history with glycation itself, without considering resistance, rendered similar results, with
359 only the abovementioned dietary effects being significant (see ESM1).

360 Additional analyses looking at the number of exposed lysines in the albumin sequence of a
361 species show no effect of this variable on albumin glycation rates ($\alpha=10.39$: $Cl_{95} [-3.713, 24.993]$,
362 $\beta=0.246$: $Cl_{95} [-0.144, 0.629]$, $P_{MCMC}=0.196$; ESM1).

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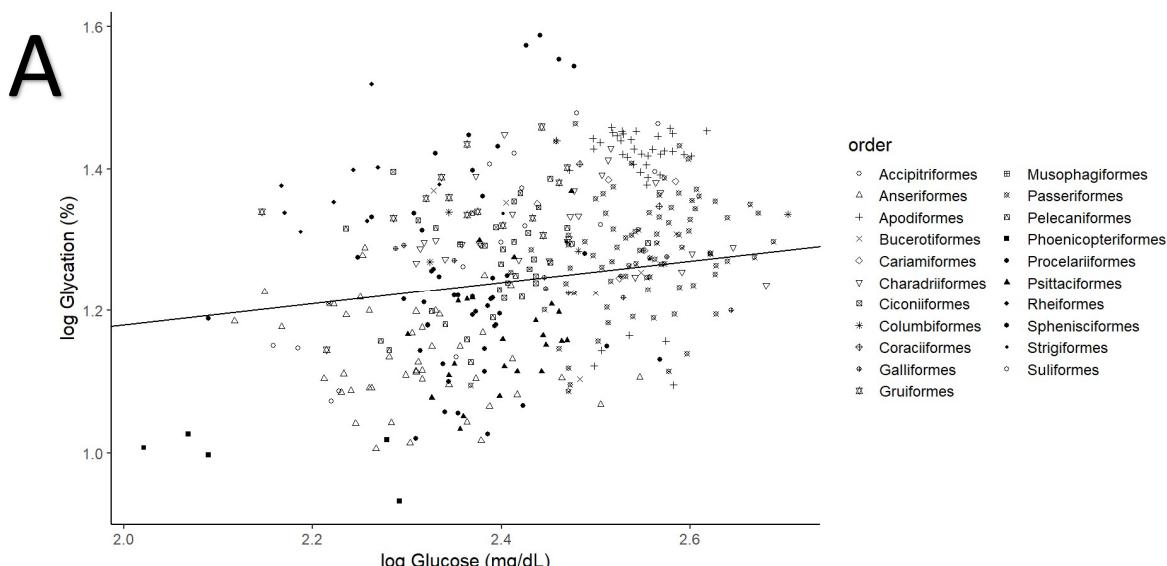
372 **Table 2.** Final glycation model with diet, body mass, glucose and life history traits as explanatory variables.
373 Posterior means, CI_{95} and pMCMC from a phylogenetic MCMC GLMM model including n=316 individuals
374 of 58 species. Glycation, glucose and body mass are \log_{10} transformed and life history traits are residuals
375 from a linear model of \log_{10} body mass and \log_{10} of the trait in question (see ESM6). Body mass and glucose
376 were also centred to better explain the intercept. The intercept corresponds to the Omnivore diet, being
377 used as the reference as it is considered the most diverse and “neutral” group for this purpose. Significant
378 predictors are indicated in bold and the credible intervals are considered for making pairwise comparisons
379 between the groups.

	Estimates	Lower 95% CI	Upper 95% CI	Sampling effort	pMCMC
Intercept	1.232	1.112	1.351	59900	<0.001
Diet: Carnivore terrestrial	0.101	0.017	0.187	59900	0.021
Diet: Aquatic predator	0.027	-0.062	0.118	59900	0.549
Diet: Herbivore	-0.019	-0.161	0.119	59936	0.781
Diet: Frugivore/granivore	0.095	-0.098	0.292	59900	0.329
Centred \log_{10} Body mass	0.004	-0.043	0.05	58765	0.876
Log₁₀Glucose	0.137	0.012	0.255	59900	0.027
Maximum lifespan	0.037	-0.122	0.195	59043	0.648
Clutch mass	0.151	-0.03	0.346	59900	0.114
Developmental time	0.04	-0.188	0.266	59303	0.725

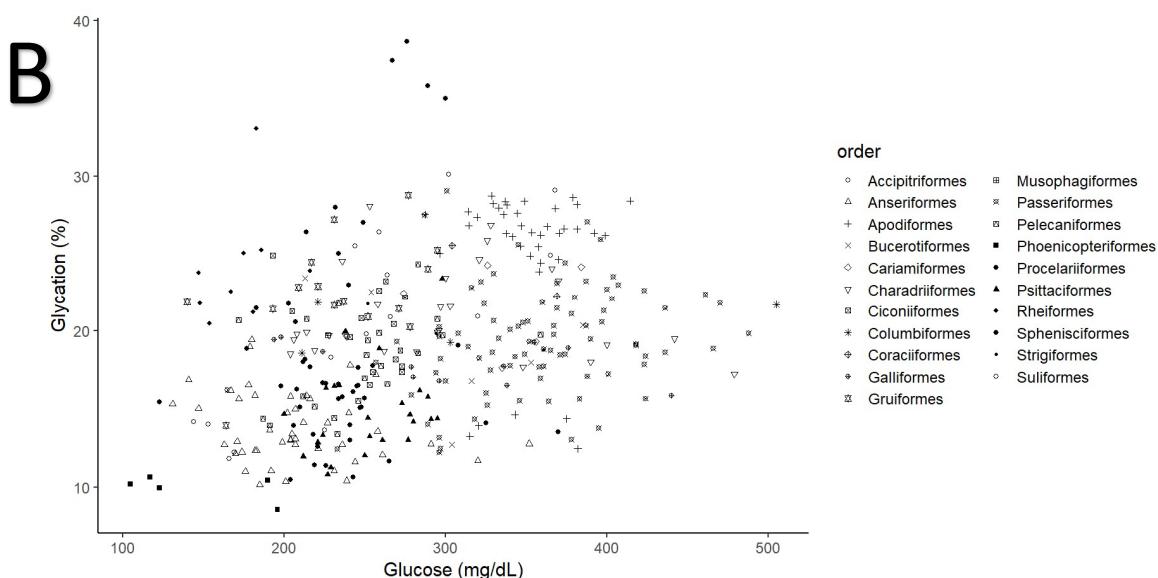
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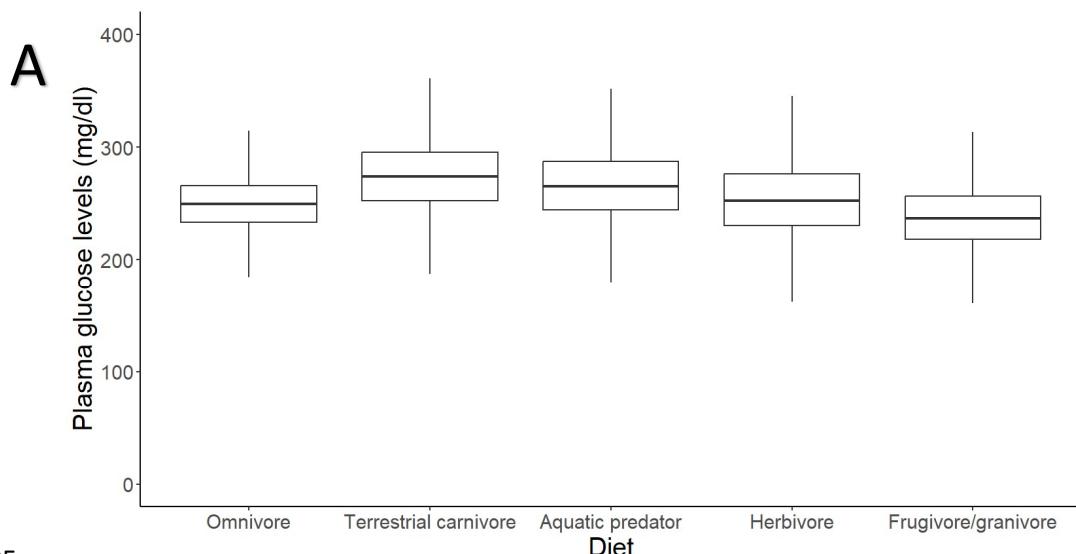


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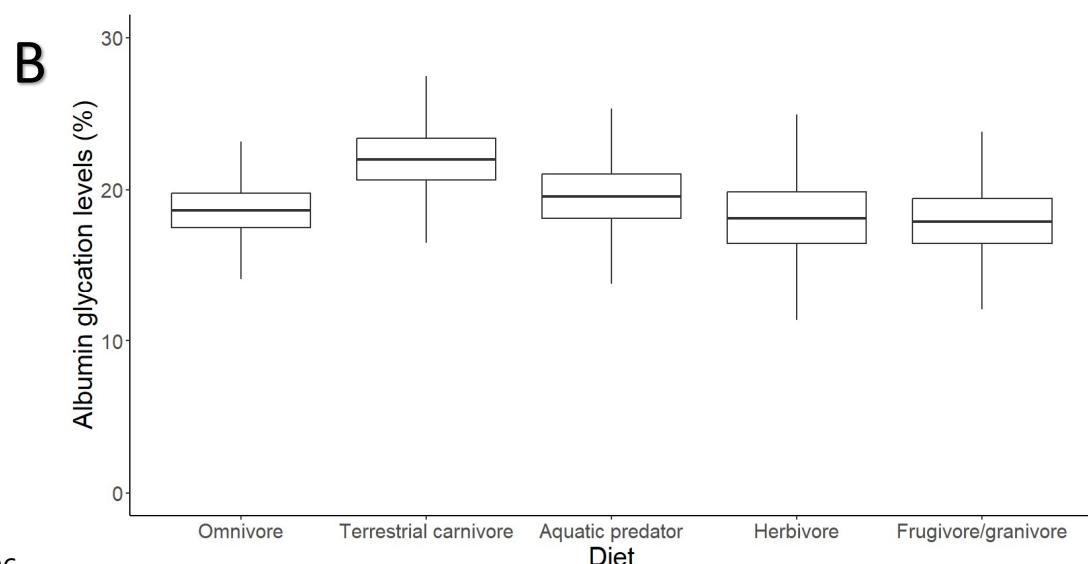
385 **Figure 3.** Individual albumin glycation rates (as a percentage of total albumin) variation as a function of
 386 individual plasma glucose values (mg/dL). **A.** Both variables \log_{10} transformed, as in the model, including
 387 the line representing the predicted relationship). **B.** Both variables in a linear form, to more explicitly
 388 illustrate the phenomenon referred to as higher albumin glycation resistance in birds with higher plasma
 389 glucose levels, inferred from the faster increase in glucose than albumin glycation, i.e. the negative
 390 curvature of the relationship. Different bird orders are indicated by symbols, as specified on the legends
 391 at the right side of the graphs. The values and estimates used are from the glycation model without life
 392 history traits (n=379 individuals from 75 species).

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397 **Figure 4.** Outcomes of the models (estimates with interquartile ranges from the posterior distributions
398 and whiskers representing credible intervals) on individual data on effects of diet on **(A)** plasma glucose
399 levels and **(B)** albumin glycation in birds. Glucose levels are given in mg/dL, while glycation levels are a
400 percentage of total plasma albumin which is found to be glycated. Terrestrial carnivores showed
401 significantly higher glycation levels than omnivores (Estimate=21.62 %, CI₉₅[18, 25.95], p_{MCMC}=0.049).
402 Models without life history traits, including more individuals, are represented, but the models with life
403 history traits do not show differences in their qualitative predictions (i.e. higher albumin glycation in
404 terrestrial carnivores than in omnivores; see ESM1).

405 We observed no significant effect of age relative to maximum lifespan nor sex on either
406 glycaemia or glycation (see ESM1). In some models, GVIF for body mass and/or clutch mass were
407 higher than 1.6, and in one case body mass is slightly above 2.2 (2.26, see ESM1). This indicates

408 that there may be moderate collinearity for these variables, but the fact that we used life-history
409 trait covariates that are excluding body mass associated variation, this small effects remain
410 mysterious, but nevertheless probably not worrying.

411 **DISCUSSION**

412 **Plasma glucose level and albumin glycation rate (co)variation patterns suggests**
413 **resistance mechanisms**

414 Our findings show that glucose levels vary widely among different bird groups. Interspecific
415 differences are partly explained by the allometric relationship of glycaemia with body mass
416 (**Figure 2.A**), which has already been reported in previous studies [56, 63, 34, 13, 14]. Indeed,
417 Passeriformes, Apodiformes and some Columbiformes (e.g. *Nesoenas mayeri* holds the highest
418 value in our dataset) are found at the higher end of the glucose level continuum, in accordance
419 with their relatively small body mass and powered flight, while groups of larger birds such as
420 Phoenicopteriformes, Anseriformes, Rheiformes and Suliformes tend to show low glycaemia
421 levels. This pattern is similar for glycation, with some groups of large birds (such as
422 Phoenicopteriformes, Anseriformes and Suliformes) showing the lowest levels of glycated
423 albumin, while small birds (such as Apodiformes) had the highest values. Nevertheless, glycation
424 remains high in some birds in relation to their glucose levels, as in Rheiformes, or low as in
425 Psittaciformes or Passeriformes. The case of Procellariiformes, which are typically long-lived
426 birds, is particularly striking, with some species exhibiting some of the highest glycation levels
427 (*Calonectris diomedea* and *Macronectes giganteus*) and others some of the lowest (*Procellaria*
428 *aequinoctialis*). This suggests that, if birds are protected against the deleterious effects of high
429 glycaemia, certain species may have evolved mechanisms to efficiently prevent proteins to be
430 glycated at high rates while others may have evolved mechanisms to resist the consequences of
431 protein glycation. We should also bear in mind that some taxonomic groups may be
432 underrepresented in our study, or biases in species selection due to availability contingencies
433 (e.g. species common in zoos or present in European countries) may exist, so further studies
434 should target this underrepresented groups in order to confirm our predictions.

435 As expected for a non-enzymatic reaction, just by the law of mass action, bird albumin glycation
436 rates increase with plasma glucose concentration. However, the logarithmic nature of the
437 relationship, and the fact that the slope is lower than one, suggest that species with higher
438 plasma glucose levels exhibit relatively greater resistance to glycation. This finding aligns with

439 previous research indicating that *in vitro* glycation levels of chicken albumin increase at a slower
440 rate than those of human albumin when glucose concentration is elevated [83]. Moreover, these
441 levels are consistently lower than those of bovine serum albumin regardless of glucose
442 concentration and exposition time [37]. As discussed in previous studies comparing chicken to
443 bovine serum albumin [37], or zebra finch to human haemoglobin [53], the lower glycation rates
444 observed in bird proteins may result from a lower number of glycatable amino acids (e.g. lysines)
445 in their sequence and/or their lesser exposure at the protein surface.
446 Our analyses do not succeed in indicating a significant positive relationship between average
447 glycation levels and the number of glycatable lysines in the albumin sequence. This may be
448 attributed to the limited number of species employed or the weak variation in the number of
449 glycatable lysine residues, which are mostly ranging from 33 to 39. An interesting exception are
450 the 18 glycatable lysine residues of flamingos (*Phoenicopterus ruber*), which also shows very low
451 glycation levels (mean=10.1%). However, the exceptions of 44 in zebra finches and 20 in godwits
452 (*Limosa lapponica*, used in place of *Limosa limosa*) are nevertheless associated with very similar
453 average glycation levels of 20.2% and 20.7% respectively.

454 **Plasma glucose relates with longevity and may be influenced by reproductive**
455 **strategies**

456 Our results were only in minor agreement with predictions from POLS theory: it holds for body
457 mass, but not for the other life history variables tested. The only previous comparative study of
458 glycaemia and life history traits to our knowledge [14] shows no relationship with mass-adjusted
459 maximum lifespan in passerines. However, our study over 88 bird species on 22 orders revealed
460 an increase in glucose with mass-adjusted longevity up to a plateau (see **Figure 2.B**). Thus, the
461 relationship between glucose and maximum lifespan may depend on differences between bird
462 orders or be tied to specific species particularities not explored in our study. Such species
463 particularities might involve additional undetermined ecological factors that modify the
464 relationship of glycaemia with longevity. Further exploration of glucose metabolism in relation
465 with lifestyle will bring further light on species-specific life-history adaptations concerning
466 glucose. For example, the species with the lower mass-adjusted maximum lifespan here was a
467 cormorant (*Leucocarbo verrucosus*), which have quite low glucose values for birds.
468 Regarding reproductive investment (i.e. clutch mass), our results show no relationship with
469 glycation (see **Table 2**), while previous studies reported positive relationships with glycaemia in
470 passerines [14, 83]. Interestingly, most of the species with high clutch mass included in our study

471 belong to the Anseriformes (**Figure ESM2.1**). While these species exhibit very low glycaemia and
472 albumin glycation rates, they are also characterized by a particular reproductive strategy
473 compared to passerines, for whom clutch mass does not imply the same in terms of parental
474 investment. For instance, Anseriformes, unlike Passeriformes, are precocial and their offspring
475 are not very dependent on parental care. Furthermore, they are capital breeders, accumulating
476 the energetic resources required for reproduction in advance rather than collecting them during
477 reproduction. These species typically have large clutch sizes and incubation is usually carried out
478 by females, who store internal fat resources to support this main part of the parental
479 investment. In addition, ducklings are born highly developed, reducing the amount of care
480 required post-hatching (see e.g. [85-86]). Consequently, their dependence on glucose as a rapid
481 energy source for reproduction may be lower, with investment in this activity likely more closely
482 linked to lipid accumulation. This could explain why we did not detect previously reported
483 effects of clutch mass on glucose levels.

484

485 **Terrestrial carnivores show a paradoxically increased albumin glycation rate without**
486 **increased plasma glucose levels**

487 Contrary to our expectation of finding differences across dietary groups, plasma glucose did not
488 significantly vary with species diet. This aligns with results previously reported by [14] for
489 Passeriformes or by [55] for 160 species of vertebrates among which 48 were birds, but not with
490 [15], that showed glycaemia to be increased with the proportion of fruits/seeds in the diet, or
491 with [56], that showed higher (mass-adjusted) glucose levels for terrestrial carnivores and
492 insectivores (those included in our terrestrial carnivores' category) and lower for
493 frugivorous/nectarivorous. On the other hand, intraspecific data indicates that changes in diet
494 composition hardly affects glycaemia in birds [87]. These studies suggest that glycaemia is tightly
495 regulated independently of dietary composition within species, while it probably varies across
496 species depending on the diet they are adapted to, although depending on the bird groups
497 included in the analyses and the way of assessing it.

498 Although terrestrial carnivore species did not have significantly higher glycaemia levels in our
499 study, they nonetheless demonstrated significantly higher albumin glycation rates, suggesting a
500 potential susceptibility to protein glycation. Besides unique structural features, this
501 phenomenon could be due to lower albumin turnover rates in terrestrial carnivores. Studies in
502 humans have linked higher oxidative damage to albumin with lower turnover rates (reviewed in

503 [88]), which may extend to other species and post-translational modifications of albumin, such
504 as bird's albumin glycation. However, the contrary outcome would be anticipated if protein
505 intake were high as seen in carnivorous species [88-89].
506 Interestingly, the contrast between terrestrial and aquatic predators, which did not show such
507 high glycation rates, suggests the beneficial influence of a substantial difference in food
508 composition or the pre-eminence of another ecological factor, yet to be determined, which may
509 explain lower glycation levels in aquatic predators. In terms of food composition, the proportion
510 of essential n-3 PUFA (polyunsaturated fatty acids), particularly long-chain ones such as DHA and
511 EPA compared with n-6 PUFA and short-chain n-3 PUFA, is different in aquatic and terrestrial
512 environments [90] and therefore in the diet of predators in each environment (e.g. [91]). For
513 instance, low n-6 PUFA/n3- PUFA ratio have been shown to decrease insulin levels and improve
514 insulin resistance in humans [92]. On the other hand, while some studies report that an increase
515 in dietary n3-PUFA reduces the levels of glycated haemoglobin (HbA1c) in humans, this effect
516 was not detected in many other studies and therefore remains inconclusive (reviewed in [93]).
517 The detailed study of the fat composition of the diets of terrestrial and aquatic predators, in
518 particular the ratio between different types of PUFA, merits more attention. In addition,
519 micronutrient content, such as circulating antioxidant defences, should also be taken into
520 consideration. Indeed, a positive relationship between oxidative stress and glycated levels of
521 bovine serum albumin has been reported [94]. One hypothesis is that terrestrial predators have
522 higher systemic oxidative stress levels compared to other species, which may be explained by
523 defects in their antioxidant defences. Uric acid is one of the main non-enzymatic antioxidants in
524 birds [95-97], and uric acid levels are especially high in carnivore species that have a rich protein
525 diet [96, 98-101]. We should therefore take into account other important antioxidants such as
526 vitamin E, vitamin A and carotenoids, which may be less abundant in the diets of terrestrial
527 carnivores (e.g. [102], but see [52]). The question of whether the diet of aquatic carnivores
528 provides a better intake of antioxidants would therefore requires a more detailed description of
529 dietary habits. The herbivorous diet, meanwhile, despite the expected possibility contributing
530 to lower glycaemia and glycation levels due to higher levels of PUFA compared to SFA (saturated
531 fatty acids) (effects reviewed in [93] for humans) and higher fibre content (see [103]), did not
532 lead to significantly lower levels of either of these two parameters. However, the relatively low
533 sample size in that group or the presence of outliers as *Nesoenas mayeri*, with the highest
534 glucose levels of this dataset, makes the interpretation of the obtained results somehow limited.

535 Therefore, further research should be carried out on these species to determine if the expected
536 pattern would emerge with a better sampling.

537 Finally, differences between captive and wild populations could be considered as a source of
538 variation in glucose or glycation levels, due to a more sedentary lifestyle in captivity, with lower
539 activity levels and a higher food intake likely to lead to increased circulating glucose levels in
540 captive individuals. Additionally, differences in nutrient intake, such as antioxidants, specific
541 fatty acids or amino acids, between captive and wild populations could contribute to variations
542 in glycation levels, as we saw above. Nevertheless, we think that this factor is unlikely to
543 significantly affect our findings regarding diet categories, because our study encompasses
544 species from both captive and wild populations across various diet groups, particularly those
545 exhibiting significant differences (e.g., omnivores versus terrestrial carnivores).

546 **CONCLUSION AND PERSPECTIVES**

547 In conclusion, the avian plasma glucose levels measured here are generally higher or in the high
548 range of values recorded for mammalian species [28]. Our study also concludes that there is
549 considerable variation in plasma glucose levels and albumin glycation rates among bird species,
550 with those with the highest glucose levels showing greater resistance to glycation. The
551 correlation between plasma glucose and life history traits are primarily influenced by its inverse
552 association with body mass, along with non-linear, mass-independent effects of longevity.
553 Finally, although diet does not explain plasma glucose levels in our study, terrestrial carnivores
554 have higher albumin glycation rates than omnivores. Whether these intriguing results are
555 explained by specific effects of certain dietary components, such as PUFAs and/or antioxidants,
556 remains to be determined. Differences in plasma glucose levels, albumin glycation rates and
557 glycation resistance (as glycation levels adjusting for plasma glucose concentration) across bird
558 species do not seem consistent with predictions according to the POL hypothesis, except for
559 body mass. Further investigation is needed to elucidate the correlation between these traits and
560 specific life conditions, such as reproductive strategy, migration patterns, flight mode or more
561 detailed diet composition. In addition, more in-depth exploration of glycation levels within high
562 glycaemic groups such as the Passeriformes and other small birds, which make up a significant
563 proportion of avian species, could provide valuable new insights. Similarly, investigating groups
564 such as the Phoenicopteriformes or Anseriformes, which are at the other end of the glycaemic-
565 glycation spectrum, could shed light on the origin of differences between avian orders.
566 Furthermore, notable variations were observed between species within orders, as in the

567 Procellariiformes, which with their high mass-adjusted longevity, and particularly given that
568 some of the age known individuals in our dataset (indeed, two specimens of snow petrels,
569 *Pagodroma nivea*, showed ages higher than the maximum lifespan reported in the sources we
570 explored, determining a new record) have still relatively low levels of glycation, may suggest that
571 some species (the ones that do show higher glycation levels) are not intrinsically resistant to
572 glycation, but rather to its adverse consequences for health. As the main limitations from this
573 study, the usage of individuals from both wild and captive populations, sampled at different
574 periods of the year, and the low sample size for some species, due to logistic constraints, may
575 have introduced noise in the values reported that should be addressed in future studies by
576 implementing a stricter sampling protocol. Also, a more thorough and accurate report and
577 compilation of life history traits from multiple species would allow to increase the number of
578 species included in this kind of analyses. Future research should also focus on specific species to
579 unravel the physiological mechanisms mediating the effects of blood glucose and protein
580 glycation on life-history trade-offs, in particular mechanisms that may vary between taxa and
581 contribute to characteristic adaptations in birds to mitigate the potentially negative effects of
582 comparatively high glycaemia.

583 **CONFLICTS OF INTEREST**

584 None declared.

585 **DATA AND CODE ACCESSIBILITY**

586 All data used, including references for the values taken from bibliography or databases, will be
587 available as Electronic Supplementary Material (ESM5). Code is made available as Electronic
588 Supplementary Material (ESM7).

589 **ETHICS STATEMENT**

590 This study followed all the legal considerations, with the ethic authorisations from the French
591 Ministry of Secondary Education and Research, n°32475 for the zebra finches sampling, the
592 Swiss Veterinary Office (FSVO) n° 34497 for the Alpine swifts, the Ethics Committee of the
593 University of Extremadura (licenses112//2020 and 202//2020) and the Government of
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595 Spain, and Sampling in Terres Australes et Antarctic Françaises was approved by a Regional

596 Animal Experimentation Ethical Committee (French Ministry of Secondary Education and
597 Research permit APAFIS #31773-2019112519421390 v4 and APAFIS#16465–
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599 The samples from the Mulhouse zoo were taken by its licensed veterinary with capacity number:
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618 **AUTHORS' CONTRIBUTIONS**

619 F. Criscuolo and F. Bertile conceived the idea, directed most of the sample collection and logistics
620 and contributed significantly to the writing. A. Moreno-Borrallo contributed to the development
621 of the questions, gathered the diet and life-history data, performed the statistical analyses, and
622 led the writing. S. Jaramillo Ortiz and C. Schaeffer performed the mass spectrometry analyses
623 for protein glycation measurements, and S. Jaramillo Ortiz contributed to the glucose measures
624 and commented on the manuscript. T. Boulinier. and V. A. Viblanc coordinated the ECONERGY
625 and ECOPATH polar programs, organised the collection of samples on subantarctic seabirds, and

626 commented on the manuscript. Olivier Chastel collaborated collecting marine bird samples and
627 commented on the manuscript. B. Rey contributed with the collection of samples from Parc des
628 Oiseaux (Villars-les-Dombes, France) and commented on the manuscript. P. Bize leads the
629 monitoring of Alpine swifts' populations from which the samples were obtained, he helped
630 collecting them and commented on the manuscript. J. S Gutiérrez and J. A. Masero contributed
631 with samples from Spain and part of the statistic scripts and commented on the manuscript. B.
632 Quintard leads the health monitoring of Mulhouse zoo bird collection, organized and realized
633 most of Mulhouse zoo samplings and commented on the manuscript. All authors gave their
634 approval for publication.

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