

1 **TITLE:**

2 **Stay clear and dry! How microstructure diversity can offset the hydrophobicity costs of transparency**  
3 **in clearwing Lepidoptera**

4

5 **AUTHORS:**

6 Doris Gomez<sup>1\*</sup>, Jonathan Pairraire<sup>2</sup>, Charline Pinna<sup>3</sup>, Monica Arias<sup>1,4</sup>, Céline Houssin<sup>3</sup>, Jérôme Barbut<sup>3</sup>,  
7 Serge Berthier<sup>2</sup>, Christine Andraud<sup>5</sup>, Thierry Ondarçuhu<sup>6</sup>, Marianne Elias<sup>3,7,8</sup>

8 <sup>1</sup> CEFÉ, CNRS, Univ Montpellier, EPHE, IRD, Montpellier, France

9 <sup>2</sup> INSP, Sorbonne Université, CNRS, Paris, France

10 <sup>3</sup> ISYEB, CNRS, MNHN, Sorbonne Université, EPHE, Université des Antilles, France

11 <sup>4</sup> PHIM, Univ Montpellier, CIRAD, INRAE, Institut Agro, IRD, Montpellier, France

12 <sup>5</sup> CRC, MNHN, Paris, France

13 <sup>6</sup> IMFT, Univ. Toulouse, CNRS, Toulouse, France

14 <sup>7</sup> Smithsonian Tropical Research Institute, Gamboa, Colón, Panama

15 <sup>8</sup> Centre Interdisciplinaire de Recherche en Biologie, CNRS, INSERM, Collège de France, Paris, Île-de-France France

17

18 **CORRESPONDING AUTHOR:** Doris Gomez, [doris.gomez@cefe.cnrs.fr](mailto:doris.gomez@cefe.cnrs.fr)

19

20 **ORCIDS**

21 Doris Gomez <http://orcid.org/0000-0002-9144-3426>

22 Charline Sophie Pinna <http://orcid.org/0000-0002-4947-0893>

23 Monica Arias: <http://orcid.org/0000-0003-1331-2604>

24 Christine Andraud <http://orcid.org/0000-0002-3112-9363>

25 Serge Berthier <http://orcid.org/0000-0002-9255-059X>

26 Thierry Ondarçuhu: <http://orcid.org/0000-0001-7657-5271>

27 Marianne Elias <http://orcid.org/0000-0002-1250-2353>

28

29 **Running title:** transparency and hydrophobicity in Lepidoptera

30 **ABSTRACT**

31 Living organisms are submitted to multiple developmental and selective constraints resulting in  
32 evolutionary compromises, one of the best examples being the integument (the outer protective layer  
33 of living organisms) which is fundamentally multifunctional. Integument anti-wetting or  
34 hydrophobicity – evolved in relation to complex and various structures – is a crucial property as it serves  
35 multiple functions like self-cleaning, locomotion, or defence against pathogens and may interfere with  
36 other functions like thermoregulation or communication. Elucidating the structure-property  
37 relationships and unravelling potential trade-offs is crucial to understand the evolution of the  
38 integument. In opaque Lepidoptera, wing scales actively contribute to anti-wetting. In clearwing  
39 Lepidoptera, wing scales are often reduced, raising the question of whether they can maintain similar  
40 hydrophobicity levels to those of opaque species and if not, whether wing microstructure (scale  
41 density, shape, insertion, and coloration) may mitigate the costs of a lower hydrophobicity. To answer  
42 these questions, we measure static contact angle (CA) of water droplets at different stages of  
43 evaporation in opaque and transparent patches of 23 Lepidoptera species that show a high diversity  
44 in wing microstructure. More specifically, we find that transparency is costly for hydrophobicity, and  
45 that such cost depends on wing microstructure. In general, transparent patches lose more  
46 hydrophobicity with water evaporation than opaque patches. Yet, this loss of hydrophobicity is  
47 attenuated for higher scale densities, erect scales compared to flat scales, coloured scales (for erect  
48 scales), multiple scale layers (for flat scales), or when combining two types of scales (piliform and  
49 lamellar) than having only one type of scale (piliform or lamellar). Nude membranes show the lowest  
50 hydrophobicity values. We find that wing hydrophobicity negatively relates to optical transparency,  
51 showing a trade-off between optics and hydrophobicity. Moreover, we find that tropical species have  
52 higher hydrophobicity in their transparent patches than temperate ones, suggesting transparent  
53 patches are under stronger selection for hydrophobicity in tropical than in temperate species. These  
54 novel findings, which are consistent with the physics of hydrophobicity, suggest that insect wings are  
55 evolutionary multifunctional compromises.

56

57 **Key words:**

58 hydrophobicity; Lepidoptera; multiscale roughness; trade-off ; transparency

59

60

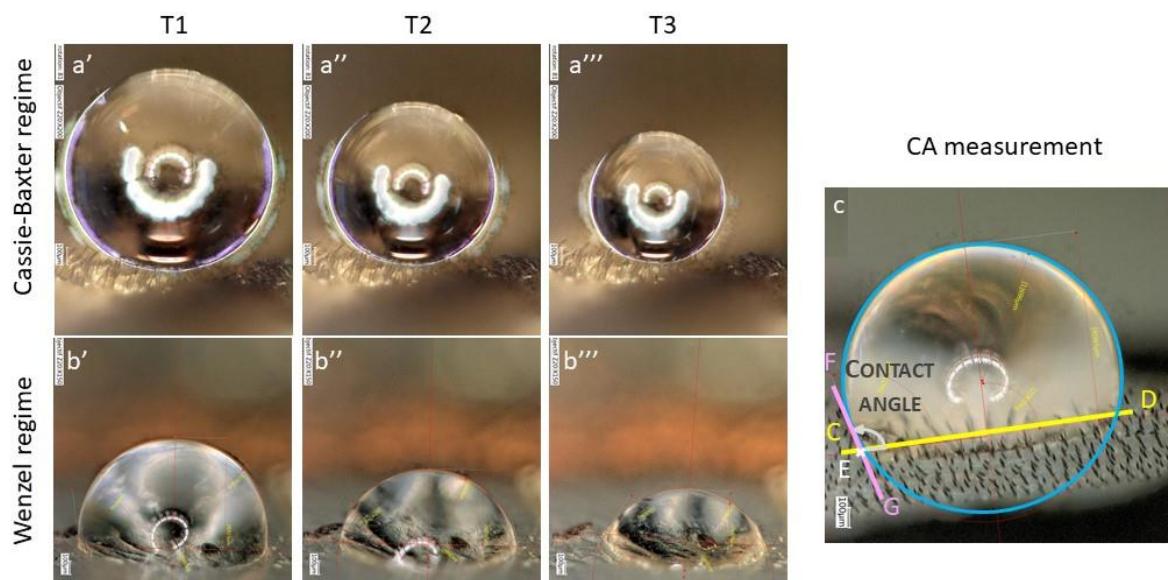
61 **INTRODUCTION**

62

63 Living organisms are submitted to multiple developmental and selective constraints resulting in  
64 evolutionary compromises. The integument, the outer protective layer of plants and animals, perfectly  
65 illustrates this concept as it is involved in multiple functions like mobility, thermoregulation,  
66 communication or camouflage, and defense against pathogens. Hydrophobicity is a crucial property of  
67 the integument in terrestrial organisms: avoiding unwanted water efficiently contributes to self-  
68 cleaning mechanisms and defense against pathogens, as water droplets that roll off remove potentially  
69 contaminating particles, like dust or bacteria (Wagner et al., 1996). Hydrophobic surfaces, by their  
70 structure, can have direct bactericidal effects, such as in geckos (Watson et al., 2015) and cicadas  
71 (Ivanova et al., 2012). Anti-wetting also contributes to locomotion: in geckos, the hydrophobicity of  
72 complex toepads allows adhesion and locomotion on any surface (Autumn et al., 2014); in flying  
73 insects, the hydrophobicity of wings – which reduces drag, removes weight, and limits wing damage –  
74 enhances flight ability, and extends it to rainy conditions (Watson et al., 2011). In water striders, the  
75 hydrophobicity of legs ensure water skating (Gao & Jiang, 2004) and the hydrophobicity of the body  
76 cuticle creates a plastron (air-film) which helps under-water respiration, provides buoyancy, and  
77 protects from submergence, thereby facilitating the colonization of the open ocean (Mahadik et al.,  
78 2020). Yet, hydrophobicity has its limits: for instance, water films, which help cooling through water  
79 evaporation, cannot form on hydrophobic surface; hence hydrophobicity can interfere with  
80 thermoregulation potentially limiting cooling mechanisms and efficiency. There can exist trade-offs  
81 between antagonistic needs and different evolutionary compromises can be selected in various  
82 environmental conditions. Finally, the hydrophobicity property has evolved in relation to integument  
83 structure: in geckos, hydrophobic toepads have been gained and lost multiple times independently,  
84 leading to a structural diversity at all scales (Gamble et al., 2012), which relates to differences in  
85 locomotion performance and habitat use between species (Elstrott & Irschick, 2004). Hence,  
86 deciphering the structure-property variations and unraveling potential trade-offs is crucial to  
87 understand the evolution of plants and animals.

88 As predicted by physics (Wenzel, 1936) and illustrated in plants (Barthlott & Neinhuis, 1997),  
89 a key parameter for hydrophobicity is surface texture or roughness. A water droplet on a textured  
90 hydrophobic surface can exhibit two different wetting states. In the Cassie-Baxter state (Figure 1, series  
91 a), the water droplet sits on top of the texture, with trapped air underneath and cavities filled with air  
92 (composite state, solid and air in contact with water under the drop), and hydrophobicity is at a  
93 maximum. If this state is thermodynamically metastable, the water droplet may undergo the so-called  
94 Cassie-Baxter to Wenzel transition, in which water penetrates the air-filled cavities by capillarity. In  
95 the Wenzel wetting state (Figure 1 series b), the water droplet fully fills all the cavities of the texture

96 and adheres to the surface (non-composite state, only solid in contact with water under the drop),  
97 decreasing the surface energy; hydrophobicity is then lost (Hasan et al., 2012). Compared to the  
98 Wenzel state, the Cassie-Baxter state is of high biological interest as it offers an incomplete water-  
99 surface contact and a weak water adhesion. Maintaining a stable Cassie-Baxter state is crucial to  
100 maintain high hydrophobicity under harsh environmental conditions, like rainfall.



101  
102  
103 **Figure 1.** Examples of water droplets dropped in the transparent zone: Cassie-Baxter regime (series a) for *Eutresis hypereia*  
104 combining erected coloured piliform and lamellar scales and Wenzel regime (series b) for *Neocarnegia basirei* with a nude  
105 membrane. Water droplet evolution is shown at different times: T1 (a', b'), T2 (a'', b''), and T3 (a''', b'''). Principle of contact  
106 angle (here called CA) measurement on a photo example from *Parantica sita* (c): first, we draw a theoretical circle on the  
107 droplet shape (blue). We then figure the wing surface (yellow segment CD). We define the point E (white) as the intersection  
108 of the water droplet and the segment CD and the segment FG (pink) as the tangent to the circle in E. We then compute the  
109 contact angle expressed in degrees as the angle between (CD) and the tangent (FG) of the water droplet at the point E. The  
110 rounder the droplet, the higher the CA value.

111  
112  
113 Roughness at nanoscale – the parameter most studied to date in animals and plants - increases  
114 hydrophobicity, as shown in cicadids and dragonflies (Byun et al., 2009; Oh et al., 2017; M. Sun et al.,  
115 2009). Yet, multiscale roughness –at nano and micro scale – is even more efficient: it increases  
116 hydrophobicity and its thermodynamic stability and it reduces water adhesion. This was shown in  
117 modelling studies (Bell et al., 2015; Su et al., 2010) and repeatedly illustrated in various biological  
118 examples: in the Lotus (so-called 'Lotus effect') and other plants (Barthlott et al., 2016) as well as in  
119 insects such as water striders (Gao & Jiang, 2004) and mosquitoes (Wu et al., 2007). Increasing  
120 thermodynamic stability allows maintaining hydrophobicity with water droplets of various sizes (dew,  
121 fog, rain) and increases anti-fogging properties, i. e. the resistance to tiny water droplets condensing  
122 on the surface. While the role of nanostructures in hydrophobicity has been extensively documented

123 (e.g. Patankar, 2004; Porcheron & Monson, 2006), the role of microstructures shape in determining  
124 hydrophobic properties has been limited to simple geometries (cylinders in Cansoy et al., 2011; cones  
125 in Ding et al., 2019; P. Tsai et al., 2010) and remains poorly investigated from an empirical perspective.  
126 The only existing empirical studies with such an approach either describe the variations in  
127 hydrophobicity between various micro-architectures but without invoking explanations (Sanchez-  
128 Monge et al., 2015) or focus on one type of micro-architecture (Fang et al., 2015; Oh et al., 2017; G.  
129 Sun & Fang, 2015), inescapably showing a major influence of nanostructures to explain the variations  
130 in hydrophobicity, since nanostructures are the main elements varying among these species.

131 Lepidoptera (from the ancient Greek λεπτός: scale and πτερόν: wing) – butterflies and moths –  
132 offer an outstanding study system to investigate this question. They are typically characterized by large  
133 wings entirely covered with flat and coloured lamellar microscopic scales (Ghiradella, 1998; Miaoulis  
134 & Heilman, 1998). Scales are in average around 100  $\mu\text{m}$  long and 50  $\mu\text{m}$  wide. Through their  
135 pigmentation and structure, scales are involved in multiple functions such as antipredator defences (e.  
136 g. camouflage, deflection, mimicry in Cuthill et al., 2005; Stevens et al., 2008), communication (Kemp,  
137 2007), thermoregulation (Berthier, 2005; Krishna et al., 2020; Miaoulis & Heilman, 1998; C.-C. Tsai et  
138 al., 2020) and flight enhancement (Nachtigall, 1967; Slegers et al., 2017). They also confer  
139 superhydrophobic properties to the wing, resulting in water repellency and self-cleaning (Wagner et  
140 al., 1996; Wanasekara & Chalivendra, 2011). Superhydrophobicity *sensu lato* is defined by water  
141 droplets making high contact angles ( $>150^\circ$ ) with a surface. Self-cleaning – superhydrophobicity *stricto*  
142 *sensu* (definition not taken here) – adds to this condition a weak water adhesion, estimated by a  
143 minimal tilt from the horizontal plane needed for water droplets to roll-off (roll-off angle of a few  
144 degrees) or a minimal hysteresis (difference between advancing and receding contact angles).  
145 Superhydrophobicity is thus a *sine qua non* condition for water repellency and self-cleaning. Opaque  
146 butterflies and moths typically have self-cleaning wings, as attested by small roll-off angles (Fang et  
147 al., 2015, 2017). Scarce relevant studies suggest that wing hydrophobicity may depend on wing  
148 microstructure (scale presence in Finet et al., 2023; scale type and insertion angle in Perez Goodwyn  
149 et al., 2009; presence and type of scale in Wagner et al., 1996), and on wing macrostructure: species  
150 with longer wings (Byun et al., 2009), or larger ratio of wing area to body mass (Wagner et al., 1996)  
151 show higher hydrophobicity and wing shape was invoked to explain natural variations in  
152 hydrophobicity (Byun et al., 2009).

153 While the vast majority of Lepidoptera species has opaque wings, some species from various  
154 lineages show transparent or translucent wings (Gomez et al., 2021). While the evolution of  
155 transparency in an order of insects that typically harbour large wings covered by coloured scales may  
156 appear puzzling, recent experiments have shown that transparency is beneficial to butterflies and  
157 moths because it reduces their detectability from visually-hunting predators (Arias, Elias, et al., 2020;

158 Arias et al., 2019; McClure et al., 2019). In Lepidoptera, wing transparency is involved in various anti-  
159 predator defences, spanning from camouflage (Arias, Elias, et al., 2020) to Batesian mimicry with  
160 Hymenoptera (Skowron Volponi et al., 2018) and masquerade (Arias, Barbut, et al., 2020; Costello et  
161 al., 2013). Transparency is associated with a broad microstructural diversity (i. e. scale diversity, see  
162 examples in ESM, Figure S1), the membrane being nude or covered with scales varying in type (piliform,  
163 i. e., hair-like, and/or lamellar), insertion on the membrane (flat or erect), and colouration (coloured  
164 or transparent) (Gomez et al., 2021). All combinations of scale type, insertion, and colouration (i. e.,  
165 structural strategies Gomez et al., 2021) can be found in nature (ESM, Figure S1), and they differ in  
166 their efficiency at transmitting light : the nude membrane are most efficient while flat coloured scales  
167 (lamellar alone or in combination with piliform) are least efficient (Gomez et al., 2021). Microstructures  
168 are complemented by nanostructures on the scales and on the wing membrane. Membrane  
169 nanostructures reduce reflection levels and increase light transmission (Pinna et al., 2021; Pomerantz  
170 et al., 2021; Siddique et al., 2015; Yoshida et al., 1997).

171 Because transparency often entails profound modifications – and, in the vast majority of cases,  
172 reduction - of scale dimensions and scale density (Gomez et al., 2021), it can be hypothesized that  
173 achieving optical transparency may come at a cost to hydrophobic performance, both in water  
174 repellency and self-cleaning properties. This cost manifests as a trade-off which arises when two  
175 functional requirements cannot be simultaneously optimized, since structural features that enhance  
176 one property may inherently compromise the other. Water repellency and self-cleaning are vital for  
177 butterflies and moths, which are large-winged insects: water repellency is crucial for flight and for  
178 preventing wings from sticking together, especially in tropical rainforest species with daily rain and  
179 high humidity. Likewise, self-cleaning helps removing dust contamination that impairs flight (Wagner  
180 et al., 1996). Among the lepidopteran species investigated so far for hydrophobicity (Fang et al., 2015;  
181 Perez Goodwyn et al., 2009; Wagner et al., 1996; Wanasekara & Chalivendra, 2011; Zheng et al., 2007),  
182 only four clearwing butterfly species have been included: *Parantica sita* (with lamellar titled scales)  
183 and *Parnassius glacialis* (with flat lamellar scales), with high or moderate hydrophobicity respectively  
184 (Fang et al., 2015; Perez Goodwyn et al., 2009), *Phanus vitreus* (with erect lamellar scales) with lower  
185 hydrophobicity in transparent than opaque areas (Finet et al., 2023), *Greta oto* (with piliform scales)  
186 with one of the lowest hydrophobicity values found in butterflies (Wanasekara & Chalivendra, 2011).  
187 Scarce data suggest that a greater reduction in scale dimensions or coverage on the wing membrane  
188 may entail higher costs in terms of hydrophobicity, as suggested by the lower hydrophobicity in  
189 transparent areas of *Phanus vitreus* with removed scales than with scales (Finet et al., 2023). However,  
190 large-scale comparative studies are currently lacking.

191 To fill that knowledge gap, we here explore to what extent anti-wetting ability is influenced by  
192 scale microstructure in species, whether it entails a trade-off with optical transparency and whether it

193 depends on climatic conditions, by selecting a subset of 23 species (ESM, Figure S2) from a broad study  
194 of 123 clearwing Lepidoptera species (Gomez et al., 2021) that show a large diversity in microstructure.  
195 In these species, we explored the links between microstructure, macrostructure, hydrophobicity and  
196 optics while controlling for phylogenetic relatedness between species. We measured the contact angle  
197 (CA) made by water droplets on the wing at different stages of water evaporation (thus droplet size).  
198 With these measurements, (i) we explored the relationships between hydrophobicity and wing  
199 macrostructure. (ii) We then explored the relationships between hydrophobicity and wing  
200 microstructure, i. e. structural strategy (presence or absence of scale, scale shape, scale insertion  
201 angle, coloration and density). (iii) To go beyond the associations beyond hydrophobicity and structural  
202 strategy, we explored the geometry of some structural strategies of particular interest (erect versus  
203 flat geometries, geometries involving two types of scales). We tested whether there existed consistent  
204 associations between geometrical scale features (scale dimensions, spacing, density) within the  
205 structural strategies that could explain the observed variations in hydrophobicity. (iv) To identify the  
206 selective pressures acting on hydrophobicity, we tested whether hydrophobicity and light transmission  
207 showed potential trade-off or synergy. A trade-off between hydrophobicity and light transmission  
208 would reveal a cost of transparency for water repellency. If microstructures play a dominant role in  
209 conferring hydrophobicity, species most efficient at transmitting light – which lack scales or have highly  
210 modified scales, resulting in low coverage of the wing surface – are expected to be less efficient at  
211 repelling water. (v) Finally, to identify whether hydrophobicity is influenced by climatic conditions, we  
212 tested the links between the latitude of species habitat and hydrophobicity: if repelling water is more  
213 important in the tropics where rain and humidity are inescapable, tropical species are expected to  
214 show higher hydrophobicity than temperate species.

215

216

217

## 218 METHODS

219

### 220 Species selection

221 Scale type and scale insertion have been suggested to influence hydrophobicity (Perez Goodwyn et al.,  
222 2009). Scale coloration, often involving melanin deposition which increases cuticle hardening in insects  
223 (Sugumaran, 2009), could increase scale stiffness and ability to repel water droplets. Hence, we  
224 selected a set of species varying in structural strategies – scale type (N=nude membrane, P=piliform  
225 bifid or monofid scales, L=shape different than piliform, hereafter called lamellar, or PL=association of  
226 piliform and lamellar scales), insertion (E=erect or F=flat), and colouration (C=coloured or  
227 T=transparent) – from the study of 123 species of clearwing Lepidoptera (Gomez et al., 2021). We

228 minimized the phylogenetic relatedness between species harbouring the same type of structural  
229 strategies to increase the power of comparative analyses. We selected a total of 23 species from 10  
230 families (Figure 1 & ESM, Figure S1, list in ESM Table S1), comprising 3 species for the structural  
231 strategies (N, PFC, PEC, LFC, LFT, LET), 2 species for PLEC and LEC, and 1 species for PLET, as for some  
232 species only a limited number of specimens were present in the collections. For each species, we  
233 selected three specimens in good condition either from Paris MNHN collections or from our own  
234 private collections. 54/69 specimens (all species but *Eutresis hypereia*) had labels with exact collect  
235 location that could be tracked down to GPS coordinates.

236

237

### 238 **Hydrophobicity measurements**

239 We measured the static contact angle of water droplets and wing surface in the transparent and  
240 opaque zones of the forewing of three museum specimens per species, and we monitored contact  
241 angle at three times, as water evaporated and droplet size decreased (Figure 1). For each specimen,  
242 we used a purpose-built water-droplet dispenser (a graduated pipette on a holder) and a Keyence VHX-  
243 5000 microscope (equipped with Z20 zoom) to image water droplets on butterfly wings. As a general  
244 procedure, we dropped a series of three 1  $\mu$ l water droplets (volume usually taken to assess  
245 hydrophobicity (Hasan et al., 2012; Perez Goodwyn et al., 2009)) at three locations of the transparent  
246 and opaque zones of the dorsal side of a wing. After the water droplet was dropped (time T1), we  
247 allowed its volume to be approximately divided by two (time T2) and by four (time T3) compared to its  
248 original volume. Since evaporation kinetics depended on droplet shape, time intervals elapsed  
249 between consecutive photos were not identical from one species to another. At each time, we took a  
250 photo in which we measured the static contact angle (measurement principle and examples in Figure  
251 1). The contact angle measured at time T1 can be considered as the advancing contact angle and thus  
252 serves as an indicator of surface hydrophobicity. The subsequent decrease in CA due to water  
253 evaporation, hereafter referred to as the "loss of hydrophobicity," reflects contact angle hysteresis  
254 and is indicative of the surface's self-cleaning ability. A smaller loss of hydrophobicity corresponds to  
255 a reduced contact angle of hysteresis, and an improved self-cleaning performance.

256 Our protocol only included the measurement of static contact angles which can potentially  
257 vary within a range of possible metastable values (Liu et al., 2019). Yet, statistical analyses showed that  
258 contact angle values were highly repeatable (i) for the same wing, zone and time, (ii) for both wings in  
259 the same zone, and (iii) for the same species. This ensured our protocol yielded reliable values (see  
260 detailed methods and results in ESM Table S2). We thus kept the same protocol, but we measured only  
261 the forewing.

262 We did all measurements on dry museum specimens, as widely done in comparative studies

263 of hydrophobicity (Fang et al., 2015; G. Sun & Fang, 2015; Wagner et al., 1996; Wanasekara &  
264 Chalivendra, 2011). Desiccation makes wings flatter and more comparable but it may alter the relative  
265 hydrophobic behaviour of the different species, i.e. the more/less hydrophobic species in dry  
266 conditions may not be the more/less hydrophobic species in humid conditions. In a restricted sample  
267 of 5 species showing the most common microstructures (see ESM for details), we measured the  
268 contact angle in the opaque zone and the transparent zone of a dry specimen and again on the same  
269 points, once the specimen rehydrated for 48h and showed that species ranking was conserved  
270 between dry and humid treatment, be it in the opaque or in the transparent zone, confirming the  
271 validity of our protocol (see ESM for details).

272

273

#### 274 **Measurements of wing macrostructure and microstructure**

275 To characterize wing macrostructure, we took photos of the three specimens of each species using a  
276 camera (D800E Nikon, 60mm lens, annular light). We analysed photos using ImageJ (Schneider et al.,  
277 2012). Given the role of wing length (Byun et al., 2009), wing shape (Byun et al., 2009), and ratio of  
278 total wing area to body mass (Watson et al., 2008) on hydrophobicity and self-cleaning ability, we  
279 computed wing length, length-to-width LW ratio and the ratio of total wing area to body volume, taking  
280 the volume as a proxy for mass for dry specimens, and assuming the body to be a cylinder, for which  
281 we measured length (thorax+abdomen) and width. Using the 'rptR' package (Stoffel et al., 2017), we  
282 found that all wing macrostructural measurements were repeatable, i.e. that a specimen was  
283 representative of its species for all wing macrostructural variables (ESM Table S2).

284 To characterize wing microstructure (i.e. scale characteristics, presence, type, insertion,  
285 coloration, density), we imaged the dorsal side of forewing transparent and opaque zones using  
286 microscopes (Zeiss Stereo Discovery V20 and Keyence VHX-5000). We did that in one specimen per  
287 species because scale dimensions and density were repeatable at zone by species level in Gomez et al.  
288 (2021). Using ImageJ and Keyence built-in tool, we measured scale density (per mm<sup>2</sup>), length and width  
289 (μm), scale surface (in μm<sup>2</sup>) as the product of length by width, and scale coverage as the product of  
290 scale surface (expressed in mm<sup>2</sup>) by scale density. We counted the number of different scale types:  
291 0=nude membrane, 1= lamellar or piliform, 2= lamellar and piliform. For flat lamellar scales, we also  
292 computed the density of scale top layer and computed the number of layers as the ratio between  
293 density and top layer density.

294 Not only the presence of multiscale roughness is important for hydrophobicity, but its spatial  
295 geometry is crucial for its stability (various fractal geometries tested in Bittoun & Marmur, 2012). For  
296 scale geometry (presence of one type of scales, either piliform or lamellar scales but not both), we  
297 investigated the variations in scale length or scale width with scale insertion on the membrane as this

298 was anticipated to greatly influence the spatial geometry of wing surface, and with scale coloration as  
299 melanin was a component of cuticle hardening in insects (Sugumaran, 2009). From Gomez et al.'s  
300 broad study (2021), we selected the 96 species with one type of scales only. When both piliform and  
301 lamellar scales were present, we investigated how these two types of scales were spatially associated  
302 as it was potentially the most complex micro-architecture. From Gomez et al.'s broad study (2021), we  
303 selected the 8 species that had a PL strategy, i.e. a combination of piliform scales and lamellar scales  
304 in the transparent zone. These species were *Athesis clearista*, *Diaphania unionalis*, *Dysschema*  
305 *boisduvalii*, *Eutresis hypereia*, *Macrosoma conifera*, *Methona curvifascia*, *Nagara vitrea*, *Praeamastus*  
306 *fulvizonata*. In these species, we computed (i) the ratio in length between piliform scales and lamellar  
307 scales, (ii) the ratio in density between piliform scales and lamellar scales, and (iii) the spatial  
308 association between piliform scales and lamellar scales.

309

### 310 **Optical measurements**

311 For one specimen per species, we measured specular transmittance from 300 to 700 nm as in Gomez  
312 et al. (2021), using a deuterium-halogen lamp (Avalight DHS), direct optic fibres (FC-UV200-2-1.5 x 100)  
313 and a spectrometer (Avaspec-2048 L, Avantes). Wing samples were placed perpendicular at equal  
314 distance between fibres aligned 5 mm apart (1 mm diameter spot). We took five measurements of the  
315 forewing in various points of the transparent zone. We computed the mean transmittance over [300-  
316 700] nm, which described the level of optical transparency. Optical measurements had been found  
317 highly repeatable at species level (ESM Table S2, and Gomez et al., 2021).

318

319

### 320 **Comparative analyses**

321 To explore the questions outlines below, we ran Bayesian mixed models with Markov Chain Monte  
322 Carlo, correcting for phylogenetic relatedness, using the R package MCMCglmm (Hadfield, 2010) and  
323 the maximum clade credibility (MCC) phylogeny obtained in Gomez et al. (2021) and pruned to  
324 targeted species. According to the analysis, we pruned it to 23 species (for CA analyses, Figure S2), to  
325 96 species (for P or L geometries) or to 8 species (for PL geometries). We tested different random  
326 factors (phylogeny, species, specimen) and retained the random assemblage that minimized DIC or if  
327 giving similar DIC values, the simplest in structure. Chain convergence was assessed visually and with  
328 Heidelberg's and Geweke's convergence and stationarity diagnostic functions from the R package coda  
329 (Plummer et al., 2006). We adjusted the number of iterations, the burn-in and the thinning to ensure  
330 best convergence and stationarity diagnostic and an effective sample size for all fixed and random  
331 parameters of at least 1000. Models were run with uninformative prior for random effect and residual

332 variances ( $V = 1$ ,  $\text{nu} = 0.002$ ) not to constrain the exploration of parameter values. We selected the  
333 best model with a backward selection of fixed parameters based on Bayesian P-value important (95%  
334 credibility intervals excluding zero) or less important (90% credibility interval excluding zero).

335 With Bayesian models, we analysed the variation in contact angle with (i) wing macrostructure  
336 descriptors – time, zone, forewing length, surface, LW ratio, the ratio of total wing area divided by  
337 body volume, and relevant interactions –; (ii) wing microstructure descriptors – time, wing length (to  
338 correct for variation in scale dimensions), scale length, width, density, scale type, number of different  
339 types, scale insertion, scale colouration, and the number of layers. (iii) we tested whether in some  
340 structural strategies of interest (erect versus flat geometries, involving one or two types of scales),  
341 there existed consistent associations between scale geometrical features (scale length and width, scale  
342 spacing and density) to quantify the geometrical bases of variations in hydrophobicity. For instance, in  
343 structural strategies based on both scale types (piliform and lamellar), we analyzed length ratio,  
344 density ratio and spatial association between the two scale types in relation to scale insertion (see ESM  
345 for details). (iv) We tested for a potential trade-off between optical transparency and wing  
346 hydrophobicity, considering all measurements of contact angle, individual mean values, or species  
347 mean values at T1. (v) Finally, we tested whether tropical species were more hydrophobic than  
348 temperate species. To do so, we related for each specimen its average CA value to its latitude to the  
349 equator, the proportion of wing area occupied by transparency and wing length, while taking species  
350 as random effect, for the opaque and transparent zone separately. Gomez et al (2021) found a positive  
351 relationship between optical transparency and wing surface area covered by transparency. Hence,  
352 finding a relationship between CA and optical transparency could be simply explained by latitudinal  
353 variations in the proportion of wing surface area covered by transparency, which we also tested.

354

355

## 356 RESULTS

### 357 Variation in hydrophobicity and relation to wing macrostructure

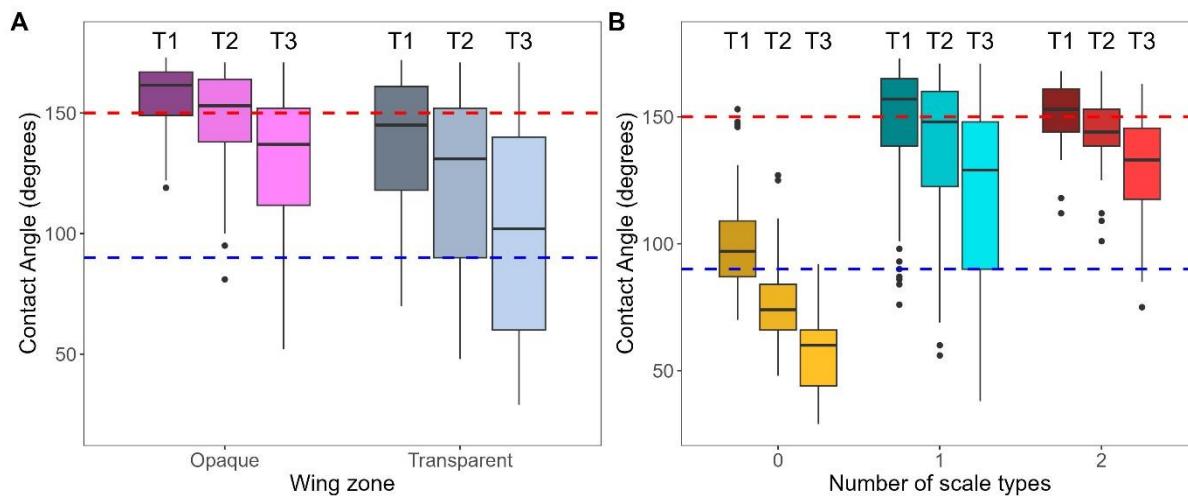
358 We observe a general decrease in hydrophobicity with water evaporation in the opaque zone and in  
359 the transparent zone (Figure 2). Transparency appears costly: (i) the transparent zone shows lower  
360 hydrophobicity than the opaque zone of the same wing, whatever the size of the water droplet  
361 considered (zone effect in Figure 2A and in ESM Table S3, see ESM Figure S4 for distribution of  
362 hydrophobicity levels). In addition, (ii) we observe a stronger decrease in hydrophobicity with water  
363 evaporation in the transparent zone compared to the opaque zone of the same wing (time x zone  
364 effect in Figure 2A, and ESM Table S3). CA does not correlate to wing length, to the wing area to body  
365 volume ratio, or to the elongation of the forewing (ESM Table S3). Yet, species with more elongated

366 wings, or with smaller wing area relative to their body or with shorter wings exhibit a greater loss of  
367 hydrophobicity with evaporation (negative forewing LW ratio x time, positive Wing Area to Body  
368 Volume x time, and positive wing length x time interaction effects in ESM Table S3).

369

370

371



372

373 **Figure 2.** (A) Variation in contact angle with wing zone and time. (B) Variation in contact angle (A) with evaporation time and  
374 number of different scale types (0=nude membrane, 1=piliform or lamellar scales, 2= piliform and lamellar scales). All  
375 measurements and both zones were included. Time corresponds to water droplet size (T1: droplet of 1 $\mu$ l, T2: diameter divided  
376 by 2 relative to T1, T3 diameter divided by 4 relative to T1). Superhydrophobic: >150° (above the red line), hydrophobic:  
377 <150° and >90°; hydrophilic: <90° (below the blue line). Results are presented in ESM Tables S4a and S4b.

378

379

### 380 Variation in hydrophobicity and relation to wing microstructure

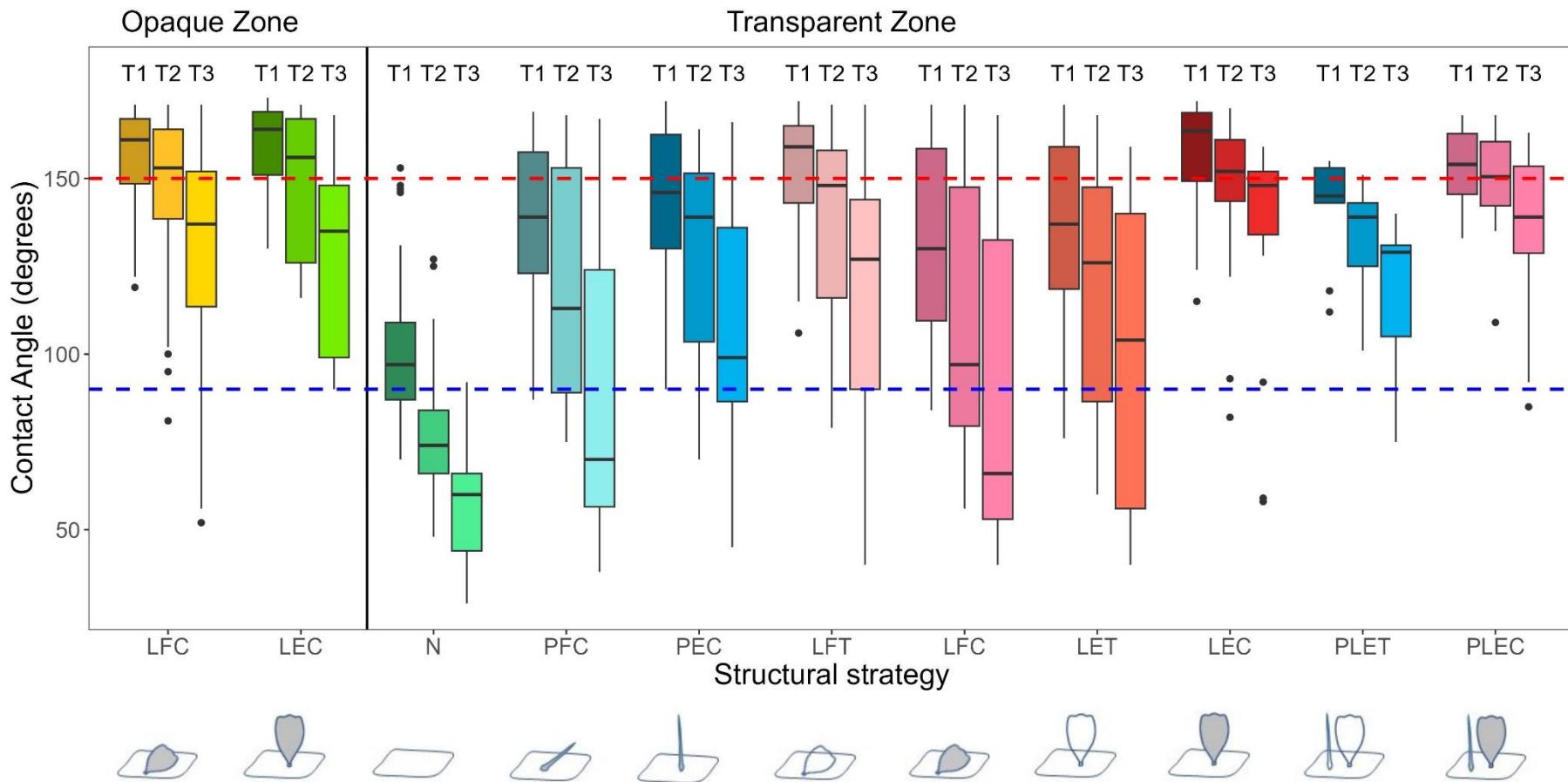
381 The influence of wing microstructure on hydrophobicity is pervasive in our results (Figure 3). First, we  
382 find a higher interspecific variance in contact angle values in the transparent than in the opaque zone  
383 (Figure 2A, Fligner-Killeen tests with all times together  $\chi^2=79.48$ ,  $p<0.001$  or separated at T1:  $\chi^2=49.57$ ,  
384  $p<0.001$ ; T2:  $\chi^2=29.24$ ,  $p<0.001$ ; T3  $\chi^2=26.47$ ,  $p<0.001$ ), likely in relation to the higher interspecific  
385 microstructural diversity of the transparent zone. Second, the nude membrane (N) yields a lower  
386 hydrophobicity (positive scale presence effect in Figure 2B and ESM Table S4a) and a higher loss of  
387 hydrophobicity with water evaporation compared to the structural strategies that involved scales  
388 (Figure 2B and 3, positive scale presence x time interaction effect in ESM Table S4a). Third, presenting  
389 two types of scales (piliform and lamellar) or only one (piliform or lamellar) yields comparable levels  
390 of hydrophobicity (non-significant Scale Nb effect in ESM Table S4b). Yet, combining two types of scales  
391 attenuates the loss of hydrophobicity with water evaporation and improves the self-cleaning ability  
392 more than having only one type of scales only (Figure 2B and Figure 3, positive scale nb x time

393 interaction effect in ESM Table S4b). Although erect and flat scales show comparable hydrophobicity  
394 (non-significant insertion effect in ESM Table S4b), erect scales limit more efficiently the loss of  
395 hydrophobicity with water evaporation than flat scales (Figure 3, negative insertion x time interaction  
396 effect in ESM Table S4b).

397 Considering all scales together, transparent scales are less hydrophobic than coloured scales  
398 which are highly hydrophobic (negative scale colour (T>C) effect in ESM Table S4b). Yet, in detail, the  
399 effect of scale coloration depends on scale insertion. In flat scales, hydrophobicity is higher in  
400 transparent scales than in coloured scales (positive colour effect in ESM Table S4d), but it shows similar  
401 loss with water evaporation in transparent and in coloured scales (colour x time effect not retained in  
402 ESM Table S4c). In erect scales, hydrophobicity is similar in transparent and coloured scales (non-  
403 significant colour effect in ESM Table S4c) but transparent scales lose more hydrophobicity with water  
404 evaporation than coloured scales (negative colour x time effect in ESM Table S4c).

405 Considering all scales together, increasing scale density increases hydrophobicity (positive  
406 density effect in ESM Table S4b). Again, in detail, the effect of density depends on scale insertion. This  
407 effect is seen in flat scales (positive density effect in ESM Table S4d) but not in erect scales (non-  
408 significant density effect in ESM Table S4c). In erect scales, increasing scale density attenuates the loss  
409 of hydrophobicity with water evaporation (positive density x time interaction effect in ESM Table S4c,  
410 ESM Figure S3A) but it is not the case in flat scales (ESM Figure S3A). In flat scales, increasing the  
411 number of layers of scales attenuates the loss in hydrophobicity with water evaporation (positive NL x  
412 Time interaction effect in ESM Table S4d, ESM Figure S3B). Finally, the gain in hydrophobicity with  
413 increasing scale density is higher for transparent than for coloured scales (positive scale colour x  
414 density interaction effect in ESM Table S4b).

415 Given that erect geometries (involving piliform and/or lamellar scales: PLE, PE, LE) seem to  
416 interact differently with water compared to flat geometries, we analysed scale dimensions and spacing  
417 (ESM Table S5). In species with only lamellar or only piliform scales, we find that after controlling for  
418 wing size, erect piliform scales are thinner than flat piliform scales (ESM Figure S5C), and both have  
419 similar length (ESM Table S5CD, Figure S5A). Lamellar scales are shorter when erect than when flat,  
420 especially when they are colored rather than transparent (ESM Figure S5B, Table S5A). Lamellar scales  
421



426 show similar width whether they are flat or erect on the wing membrane and transparent lamellar  
427 scales are larger than colored lamellar scales (ESM Table S5B). In species with erect lamellar and  
428 piliform scales, piliform and lamellar scales are in similar densities (density ratio close to 1 for the  
429 intercept in ESM Table S5F, Figure S6B), and closely associated in space (insertion effect lower for E  
430 for spacing in ESM Table S5G, Figure S6C). In species with flat scales, piliform scales are not as dense  
431 as lamellar scales (insertion effect negative in ESM Table S5F) and more distantly associated in space  
432 (ESM Table S5G). These relationships are found while controlling for phylogeny, suggesting that the  
433 tight spatial association between piliform scales and lamellar scales in hierarchical PLE geometries is  
434 likely the result of selection. In species with both lamellar and piliform scales, piliform scales are 2.6  
435 times longer than lamellar scales, creating a multi-hierarchical roughness at microscopic scales (ESM  
436 Figure S6A). In flat geometries, piliform scales are rare compared to lamellar scales, and both are  
437 more distantly spaced (ESM Table S5FG, Figure S6B); piliform scales are 5 times longer than lamellar  
438 scales (ESM Figure S6A).

439

#### 440 **Variation in hydrophobicity in relation to optics**

441 Using spectrometric measurements of wing direct transmittance, we find a negative relationship  
442 between contact angle and mean transmittance over 300-700 nm (Figure 4, ESM Table S6). A 10%  
443 increase in transmittance results in a 4° loss in CA. The relationship is statistically significant when  
444 considering all measurements or mean individual values, and marginally significant when considering  
445 mean species values, likely because of weaker statistical power.

446

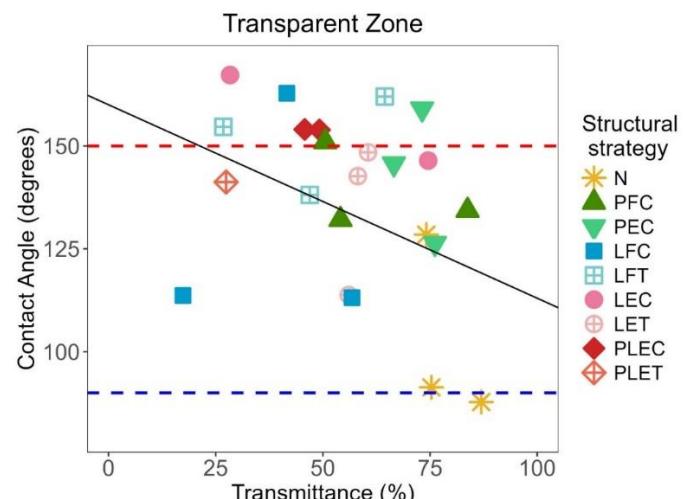
#### 447 **Variation in hydrophobicity in relation to the environment**

448 Finally, compared to their temperate counterparts, species living in the tropics have a higher  
449 hydrophobicity in their transparent zone – loss of 10° CA for 10° increase in latitude – but a similar  
450 hydrophobicity in their opaque zone (Figure 5, ESM Table S7). All species show superhydrophobic  
451 opaque patches (Figure 5B, intercept above 150° in ESM Table S7B). There is no relationship between  
452 the proportion of wing area occupied by transparency and latitude that can have explained the  
453 observed variations in CA (ESM Table S7C).

454

455

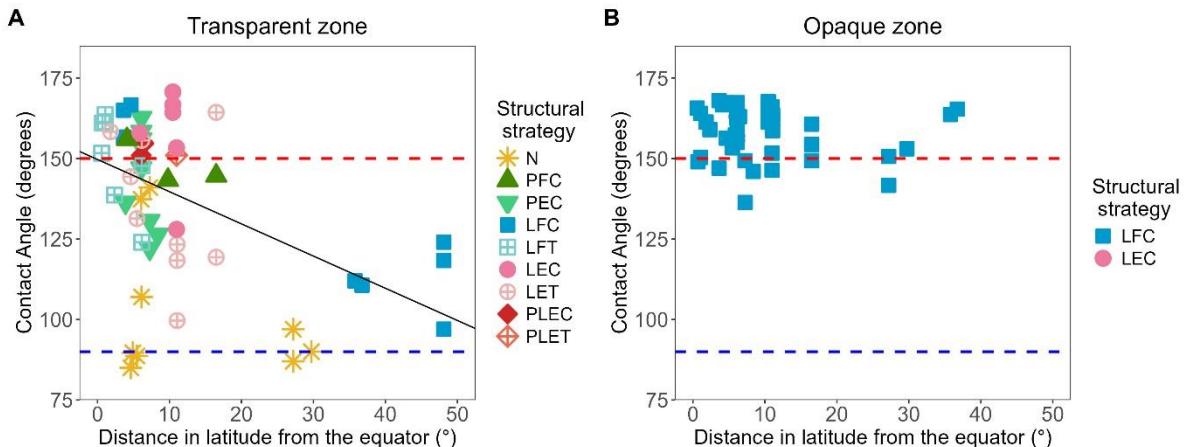
456



457 **Figure 4.** Variations of contact angle with wing transmittance for the different structural strategies. Structural strategy is a  
458 combination of scale type (N: no scales, P: piliform scales, L: lamellar scales, PL: combination of piliform scales and lamellar  
459 scales), scale insertion (E: erected, and F: flat), and scale colour (C: coloured, and T: transparent). Superhydrophobic:  $>150^\circ$   
460 (above the red line), hydrophobic:  $<150^\circ$  and  $>90^\circ$ ; hydrophilic:  $<90^\circ$  (below the blue line). We considered only the mean of  
461 CA for each species, for time T1, and for the transparent zone. The black plain line indicates the significant fitted regression  
462 line based on the Bayesian model. Results are presented in ESM Table S7.

463

464



465

466 **Figure 5.** Relationship between contact angle in the transparent (A) and in the opaque (B) zone and the distance in latitude  
467 to the equator. Structural strategy is a combination of scale type (N: no scales, P: piliform scales, L: lamellar scales, PL:  
468 combination of piliform scales and lamellar scales), scale insertion (E: erected, and F: flat), and scale colour (C: coloured, and  
469 T: transparent). Superhydrophobic:  $>150^\circ$  (above the red line), hydrophobic:  $<150^\circ$  and  $>90^\circ$ ; hydrophilic:  $<90^\circ$  (below the  
470 blue line). The black plain line in A indicates the significant fitted regression line based on the Bayesian model. Results are  
471 presented in ESM Table S8.

472

473

474

475 **DISCUSSION**

476 **Variation in hydrophobicity and relation to wing macrostructure**

477 We provide evidence for the first time at a broad taxonomic level that transparency is costly in terms  
478 of water repellency in Lepidoptera: transparent patches are less hydrophobic than opaque patches.  
479 We show that transparent patches have a lower hydrophobicity than opaque patches, and lose more  
480 hydrophobicity with water evaporation than opaque patches. Transparency thus entails important  
481 costs in terms of hydrophobicity in these two aspects. A loss of hydrophobicity with water evaporation  
482 has been commonly observed in hydrophobic human-made surfaces (McHale et al., 2005; Reyssat et  
483 al., 2007; P. Tsai et al., 2010) and in natural surfaces, as in the transparent-winged damselfly *Ischnura*  
484 *heterosticta* (Hasan et al., 2012). It has been interpreted as a loss of self-cleaning ability, especially  
485 when contact angle values get below the hydrophilicity threshold (Hasan et al., 2012).

486 We do not find any correlation between hydrophobicity and wing length, wing area to volume  
487 ratio, or forewing elongation, which at first sight partly contrasts with previous findings. In broad  
488 analyses covering many insect orders, Wagner et al. (1996) have found a positive correlation between  
489 CA and the ratio of wing area to body mass while Byun et al. (2009) have found a marginal positive  
490 correlation between CA and wing length and no correlation between CA and LW ratio (see ESM for  
491 analyses of their dataset). Yet, restricting their datasets to species more similar to Lepidoptera (with  
492 both wings involved in flight for Wagner, for wings with similar LW ratios in Byun), these relationships  
493 disappear (see ESM). More interestingly, we find that species with smaller wing area relative to their  
494 body or with shorter wings exhibit a greater loss of hydrophobicity with evaporation, hence lower self-  
495 cleaning ability. Species with a large body mass or short wings may move wings faster and small water  
496 droplets may roll off easily just through movement, attenuating selection for a high hydrophobicity  
497 towards small water droplets. The fact that species with more elongated wings exhibit a greater loss  
498 of hydrophobicity with evaporation (hence a lower self-cleaning ability) is more in contradiction with  
499 these results. A study of behaviour is needed to investigate further the relationships between wing  
500 macrostructure and water repellency ability.

501

502

503 **Variation in hydrophobicity and relation to wing microstructure**

504 Our results show close relationships between transparency and microstructure and demonstrate that  
505 variations in microstructure can mitigate the costs of transparency.

506 Going back to the physical theory behind hydrophobicity, several studies have shown that a  
507 single-level structure does not necessarily guarantee a low water adhesion, even in the Cassie-Baxter

508 state (see references in Su et al., 2010). Introducing higher levels of hierarchy increases the robustness  
509 of a surface hydrophobicity (Bell et al., 2015): it stabilizes the Cassie-Baxter state by dramatically  
510 decreasing the contact area fraction (ratio of contact area to the total surface area of the structure)  
511 and thus the adhesion force of water droplets, and by enlarging the energy barrier between the Cassie-  
512 Baxter and the Wenzel states. Hierarchical structures can be frequently found in plants and in animals.  
513 For instance, in the water strider *Gerris remigis*, leg water resistance is due to the hierarchical  
514 structures of nano-grooved microsetae, which prevented striders from being drowned under heavy  
515 rainfall (Gao & Jiang, 2004). A similar combination of micro- and nano-structuration has been found  
516 in the legs of mosquitoes, which ensured high hydrophobicity and high water-supporting ability; as a  
517 result, mosquitoes could stand effortlessly and walk easily on water (Wu et al., 2007). This likely  
518 explains why, in our study, the combination of erect piliform scales and lamellar scales yields a better  
519 self-cleaning property than piliform or lamellar scales alone. Interestingly, such geometries have a 3-  
520 level roughness: (1) erect piliform scales bending over lamellar scales (piliform scales are 2.6 times  
521 longer than lamellar scales and first in contact with water), (2) erect lamellar scales tightly associated  
522 in space with piliform scales (similar density and close spacing), and (3) nanostructures on scales and  
523 on the wing membrane. Hydrophobicity and self-cleaning likely results from the combination of the  
524 complex geometry of erect microstructures (which considerably reduces the proportion of the total  
525 surface in contact with water), and the gain in mechanical resistance (gain in elasticity and resistance  
526 against breakage) of piliform scales when bending against lamellar scales.

527 The importance of elasticity of bending hair-like microstructures has been found in several  
528 cases. In *Malacosoma castrensis* moths living by the sea, caterpillars withstood several hours to being  
529 flooded through a plastron protected by hairs (Kovalev et al., 2020). In the Lady's mantle plant  
530 (*Alchemilla vulgaris*), hairs were hydrophilic when measured individually, but they bended and  
531 coalesced into bundles when in contact with water droplets; their elasticity resulted in a repulsive  
532 interaction between the droplet and the plant surface, which maintained hydrophobicity (CA above  
533 90°) (Otten & Herminghaus, 2004). Likewise, in *Nasutitermes* termites, large bending hairs and small  
534 micrasters (micraster wavelength was around 11,7  $\mu\text{m}$  according to our measurements taken on  
535 Figure 4C from 41) enabled hydrophobicity (CA above 90°) in both rain and mist conditions (Watson  
536 et al., 2011). Finally, in mosquitoes, the huge buoyant force developed by the legs ensuring easy  
537 movement on water largely stemmed from their mechanical flexibility (Kong et al., 2015). The extent  
538 of scale elasticity in Lepidoptera and its contribution to hydrophobicity needs specific experimental  
539 study.

540 Increasing scale density helps water repellency, regardless of the type of scales. In erect scales,

541 increasing scale density does not influence CA but it attenuates the loss of hydrophobicity, thus  
542 improving self-cleaning ability. This latter result contradicts a previous finding that increasing the  
543 density of erect pillars increased the loss of hydrophobicity (Reyssat & Quéré, 2009). However, it is  
544 likely that increasing density stabilises the structure and increases the energy barrier between Cassie-  
545 Baxter and Wenzel regimes, resulting in maintaining high hydrophobicity despite water evaporation.  
546 In flat scales, increasing scale density increases hydrophobicity. In addition, increasing scale  
547 overlapping (number of layers) attenuates the loss of hydrophobicity with evaporation and improves  
548 self-cleaning. In the literature, scale overlap was assumed to help anisotropy in hydrophobicity (Bixler  
549 & Bhushan, 2014). In other words, during droplet evaporation, the Cassie-Baxter regime is more  
550 robust for denser microstructures. The mechanism by which hydrophobicity is maintained even for  
551 small droplets in multiple scale layers is still puzzling, but it may be related to scale arrangement, more  
552 specifically to scale bending, or to scale fine ridge ultrastructure (Burdin et al., 2025). Further  
553 experimental and modelling research is needed to clarify this density effect.

554 Not only scale architecture but also coloration can contribute to hydrophobicity. Erect scales  
555 show a lower loss of hydrophobicity (hence a greater self-cleaning ability) when pigmented than when  
556 transparent. In the transparent zone, coloured scales exhibit colours ranging from pale yellow to  
557 brown and black. They are likely impregnated by melanin pigments, which are known to be involved  
558 – for some biochemical forms – in cuticle sclerotization (hardening) (Sugumaran, 2009). Hence, the  
559 additional hardening conferred by pigments may increase their mechanical resistance to deformation  
560 and may contribute to maintaining hydrophobicity, even when evaporation occurs. The fact that in  
561 the flat geometry coloured and transparent scales lead to similar properties indicates that the role of  
562 colouration on hydrophobicity is more likely related to a change of elastic properties of the scales than  
563 a change of their surface chemistry.

564 Wing mechanical resistance is crucial for flight and geometries that limit protrusion height are  
565 more resistant to breakage while maintaining hydrophobicity (Bittoun & Marmur, 2012). Several of  
566 our results suggest scale height may be limited: (i) when piliform scales are alone, they have similar  
567 height, be they flat or erect, likely because they bend easily, which may limit their sensitivity to  
568 breakage. (ii) Erect lamellar scales are shortened and widened compared to flat lamellar scales, which  
569 likely increases their resistance to breakage. (iii) Erect transparent lamellar scales are densely packed,  
570 as shown in Gomez et al. (2021), which can also increase their mechanical resistance.

571 Our results bring novel evidence for a major role of microstructures in explaining large  
572 variations in hydrophobicity when diverse microstructures are considered. The rare existing studies  
573 on the subject suggest a synergistic effect of scale nanostructures and microstructures on enhancing

574 surface hydrophobicity (experiments on one type of microstructure, namely flat lamellar scales in  
575 opaque butterflies (Fang et al., 2015; Aideo & Mohanta, 2021) or hairs in the wing of the housefly  
576 *Musca domestica* (Wan et al., 2019); theoretical modelling on one type of microstructure (Sajadnia &  
577 Sharif, 2010)), or even a major role of nanostructures in the overall variation (Fang et al., 2015; Wan  
578 et al., 2019). Yet, these two latter analyses only examined one type of microstructure, thereby  
579 potentially underestimating their contribution relative to that of nanostructures that were the only  
580 parameters that varied between their study species. Further experiments are needed to elucidate  
581 these aspects, and clarify the role of nanostructures, as not only their presence, but their topography  
582 and its randomness have been recently suggested to play a role in determining antiwetting properties  
583 (Li et al., 2020). Our study also shows that the elastic properties of the microstructures plays a  
584 significant role.

585

586

### 587 **Trade-off between hydrophobicity and optical transparency**

588 In agreement with our prediction that microstructures play a major role in hydrophobicity, we find a  
589 negative relationship between hydrophobicity and transparency, a condition associated with major  
590 modifications in scale shape and density. This trade-off can be seen from the literature: the nymphalid  
591 butterfly *Greta oto* has been shown to exhibit a high transparency resulting from a low density of erect  
592 piliform scales and highly antireflective nanostructures (Pomerantz et al., 2021; Siddique et al., 2015)  
593 but a weak hydrophobicity (Wanasekara & Chalivendra, 2011). Likewise, the trade-off can be seen in  
594 the dragonfly *Gynacantha dravida* (which has micro and nanospikes), in which distal wing parts show  
595 higher hydrophobicity but lower transmittance compared to proximal wing parts (Aideo & Mohanta,  
596 2016).

597 Despite the trade-off between optical transparency and water repellency, the nude  
598 membrane, which shows the highest optical transparency and the lowest hydrophobicity, maintains a  
599 weak hydrophobicity or is hydrophilic. In such species, where wings are deprived of scales, membrane  
600 nanostructures are at full play. Membrane nanostructures reduce reflection, but their efficiency at  
601 reducing water adhesion depends on the species. The nipple array maintains a highly hydrophobic  
602 surface in the cicada *Aleeta curvicosta* (CA=144° in Watson et al., 2008) but it fails at maintaining  
603 hydrophobicity in the hesperiid *Phanus vitreus* once erect transparent scales are removed (CA=92.8°  
604 in Finet et al., 2023). The variable efficiency of nanostructures at repelling water may depend on their  
605 other parameters - density, shape, spatial disorder – calling for detailed study of those features.  
606 Questions are still open regarding the role of randomness in nanostructures, shown to improve optical

607 transparency (Siddique et al., 2015) but suggested to impair hydrophobicity (M. Sun et al., 2012; but  
608 see Li et al., 2020).

609 Revealing a trade-off between different properties and functions shows that species are  
610 submitted to antagonistic needs but can mitigate the ecological costs of clear wings. Several  
611 microstructural strategies – involving piliform and or lamellar scales, flat or erect, coloured or  
612 transparent – can show similar optical properties and levels of light transmission through the wings  
613 (drawing a horizontal line in Figure 6 in Gomez et al., 2021). Yet, these similarly optically-efficient  
614 microstructures differ in their hydrophobic properties, the combination of piliform and lamellar scales  
615 being most efficient. Hence, the high microstructural diversity (in scale presence, type, insertion,  
616 coloration, and density) allows species to offset some costs linked to transparency and tune functions  
617 separately, to a certain extent.

618

619

## 620 **Hydrophobicity and latitude**

621 We find that tropical species have more hydrophobic transparent patches than temperate species,  
622 suggesting microstructural features are under selection. This result is consistent with the prediction  
623 that in tropical climates where species face more humid conditions, and where rainfall can happen  
624 daily, there is a stronger selective pressure for increased hydrophobicity. While the opaque zone  
625 allows maximizing hydrophobicity in all environmental conditions, the differential in environmental  
626 conditions reveals the costs of transparency. To our knowledge, this is the first evidence for a higher  
627 hydrophobicity in more humid conditions. Scarce relevant studies have explored the link between  
628 habitat humidity and species hydrophobicity: at local geographical scale, all four cicada species studied  
629 by (Oh et al., 2017) showed superhydrophobicity regardless of whether they live in dry or more humid  
630 habitats, but annual species were more hydrophobic than the species that emerges in large swarms  
631 every 17 years. Likewise, Goodwyn et al. (2009) suggested that in transparent butterflies  
632 hydrophobicity may depend on lifespan and migration ability.

633 Further studies are needed to elucidate the links between hydrophobicity and species ecology  
634 and disentangle the relative contributions of micro and nanostructures to wing hydrophobicity in  
635 Lepidoptera and exploring novel questions, like the role of randomness in structural organization.

636

637

638

639

640 **ACKNOWLEDGEMENTS AND FUNDING**

641 We warmly thank Jacques Pierre, Rodolphe Rougerie, Thibaud Decaëns, Daniel Herbin, and Claude  
642 Tautel who helped with species choice and identification, and Edgar Attivissimo for additional Keyence  
643 imaging. This work was funded by Clearwing ANR project (ANR-16-CE02-0012), HFSP project on  
644 transparency (RGP0014/2016) and a France-Berkeley fund grant (FBF #2015-58).

645

646

647 **CONFLICT OF INTEREST DISCLOSURE**

648 The authors declare that they comply with the PCI rule of having no financial conflicts of interest in  
649 relation to the content of the article

650

651 **DATA AVAILABILITY STATEMENT**

652 Data and R code are available on an OSF public repository at:

653 <https://doi.org/10.17605/OSF.IO/GB49E>

654

655 **SUPPORTING INFORMATION**

656 It includes one file called Gomez et al\_SupportingInformation.docx

657

658

659 **REFERENCES**

660 Aideo, S. N., & Mohanta, D. (2016). Limiting hydrophobic behavior and reflectance response of  
661 dragonfly and damselfly wings. *Applied Surface Science*, 387, 609–616.  
662 <https://doi.org/10.1016/j.apsusc.2016.06.049>

663 Aideo, S. N., & Mohanta, D. (2021). Unusually diverse surface-wettability features found in the wings  
664 of butterflies across Lepidoptera order and evaluation of generic and vertical gibbosity-based  
665 models. *Physica Scripta*, 96(8), 085004. <https://doi.org/10.1088/1402-4896/abe82e>

666 Arias, M., Barbut, J., Rougerie, R., Dutry, M., Kohler, M., Laulan, B., Paillard, C., Berthier, S., Andraud,  
667 C., Elias, M., & Gomez, D. (2020). Ambient light and mimicry as drivers of wing transparency  
668 in Lepidoptera. *bioRxiv*, 2020.06.26.172932. <https://doi.org/10.1101/2020.06.26.172932v3>

669 Arias, M., Elias, M., Andraud, C., Berthier, S., & Gomez, D. (2020). Transparency improves concealment  
670 in cryptically coloured moths. *Journal of Evolutionary Biology*, 33(2), 247–252.  
671 <https://doi.org/10.1111/jeb.13560>

672 Arias, M., Mappes, J., Desbois, C., Gordon, S., McClure, M., Elias, M., Nokelainen, O., & Gomez, D.  
673 (2019). Transparency reduces predator detection in mimetic clearwing butterflies. *Functional  
674 Ecology*, 33(6), 1110–1119. <https://doi.org/10.1111/1365-2435.13315>

675 Autumn, K., Niewiarowski, P. H., & Puthoff, J. B. (2014). Gecko Adhesion as a Model System for  
676 Integrative Biology, Interdisciplinary Science, and Bioinspired Engineering. *Annual Review of  
677 Ecology, Evolution, and Systematics*, 45(1), 445–470. [https://doi.org/10.1146/annurev-ecols-120213-091839](https://doi.org/10.1146/annurev-<br/>678 ecols-120213-091839)

679 Barthlott, W., Mail, M., & Neinhuis, C. (2016). Superhydrophobic hierarchically structured surfaces in  
680 biology: Evolution, structural principles and biomimetic applications. *Philosophical  
681 Transactions of the Royal Society A-Mathematical Physical and Engineering Sciences*,  
682 374(2073), 20160191. <https://doi.org/10.1098/rsta.2016.0191>

683 Barthlott, W., & Neinhuis, C. (1997). Purity of the sacred lotus, or escape from contamination in  
684 biological surfaces. *Planta*, 202(1), 1–8. <https://doi.org/10.1007/s004250050096>

685 Bell, M. S., Shahraz, A., Fichthorn, K. A., & Borhan, A. (2015). Effects of Hierarchical Surface Roughness  
686 on Droplet Contact Angle. *Langmuir*, 31(24), 6752–6762.  
687 <https://doi.org/10.1021/acs.langmuir.5b01051>

688 Berthier, S. (2005). Thermoregulation and spectral selectivity of the tropical butterfly *Prepona  
689 meander*: A remarkable example of temperature auto-regulation. *Applied Physics A-Materials  
690 Science & Processing*, 80(7), 1397–1400. <https://doi.org/10.1007/s00339-004-3185-x>

691 Bittoun, E., & Marmur, A. (2012). The Role of Multiscale Roughness in the Lotus Effect: Is It Essential  
692 for Super-Hydrophobicity? *Langmuir*, 28(39), 13933–13942.  
693 <https://doi.org/10.1021/la3029512>

694 Bixler, D., & Bhushan, B. (2014). Rice- and butterfly-wing effect inspired self-cleaning and low drag  
695 micro/nanopatterned surfaces in water, oil, and air flow. *Nanoscale*, 6(1), 76–96.  
696 <https://doi.org/10.1039/C3NR04755E>

697 Burdin, L., Brulez, A.-C., Mazurczyk, R., Leclercq, J.-L., & Benayoun, S. (2025). How the structure and  
698 wettability properties of *Morpho peleides* butterfly wings can be a source of Inspiration.  
699 *Biomimetics*, 10(2), 89. <https://doi.org/10.3390/biomimetics10020089>

700 Byun, D., Hong, J., Saputra, Ko, J. H., Lee, Y. J., Park, H. C., Byun, B. K., & Lukes, J. R. (2009). Wetting  
701 Characteristics of Insect Wing Surfaces. *Journal of Bionic Engineering*, 6(1), 63–70.  
702 [https://doi.org/10.1016/s1672-6529\(08\)60092-x](https://doi.org/10.1016/s1672-6529(08)60092-x)

703 Cansoy, C. E., Erbil, H. Y., Akar, O., & Akin, T. (2011). Effect of pattern size and geometry on the use of  
704 Cassie–Baxter equation for superhydrophobic surfaces. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 386(1–3), 116–124.  
705 <https://doi.org/10.1016/j.colsurfa.2011.07.005>

706 Costello, M. J., Brennan, L. A., Basu, S., Chauss, D., Mohamed, A., Gilliland, K. O., Johnsen, S., Menko,  
707 A. S., & Kantorow, M. (2013). Autophagy and mitophagy participate in ocular lens organelle  
708 degradation. *Experimental Eye Research*, 116, 141–150.  
709 <https://doi.org/10.1016/j.exer.2013.08.017>

710 Cuthill, I. C., Stevens, M., Sheppard, J., Maddocks, T., Parraga, C. A., & Troscianko, T. S. (2005).  
711 Disruptive coloration and background pattern matching. *Nature*, 434(7029), 72–74.  
712 <https://doi.org/10.1038/nature03312>

713 Ding, W., Fernandino, M., & Dorao, C. A. (2019). Conical micro-structures as a route for achieving  
714 super-repellency in surfaces with intrinsic hydrophobic properties. *Applied Physics Letters*,  
715 115(5), 053703. <https://doi.org/10.1063/1.5096776>

716 Elstrott, J., & Irschick, D. J. (2004). Evolutionary correlations among morphology, habitat use and  
717 clinging performance in Caribbean *Anolis* lizards: Comparative clinging. *Biological Journal of  
718 the Linnean Society*, 83(3), 389–398. <https://doi.org/10.1111/j.1095-8312.2004.00402.x>

719 Fang, Y., Sun, G., Bi, Y. H., & Zhi, H. (2015). Multiple-dimensional micro/nano structural models for  
720 hydrophobicity of butterfly wing surfaces and coupling mechanism. *Science Bulletin*, 60(2),  
721 256–263. <https://doi.org/10.1007/s11434-014-0653-3>

722 Fang, Y., Sun, G., Wang, J., Hou, Y., Jin, D., & Bai, L. (2017). Micro-morphological Models for the Special  
723 Wettability of Locust and Moth Wing. In M. Wang & X. Zhou (Eds.), *Proceedings of the 2017  
724 5th International Conference on Mechatronics, Materials, Chemistry and Computer*

726 *Engineering (icmmcce 2017)* (Vol. 141, pp. 330-333. DOI: 10.2991/icmmcce-17.2017.69).  
727 Atlantis Press.

728 Finet, C., Ruan, Q., Bei, Y. Y., Chan, J. Y. E., Saranathan, V., Yang, J. K. W., & Monteiro, A. (2023). Multi-  
729 scale dissection of wing transparency in the clearwing butterfly *Phanus vitreus*. *Journal of The*  
730 *Royal Society Interface*, 20(202), 20230135. <https://doi.org/10.1098/rsif.2023.0135>

731 Gamble, T., Greenbaum, E., Jackman, T. R., Russell, A. P., & Bauer, A. M. (2012). Repeated Origin and  
732 Loss of Adhesive Toepads in Geckos. *PLoS ONE*, 7(6), e39429.  
733 <https://doi.org/10.1371/journal.pone.0039429>

734 Gao, X., & Jiang, L. (2004). Water-repellent legs of water striders. *Nature*, 432(7013), 36–36.  
735 <https://doi.org/10.1038/432036a>

736 Ghiradella, H. (1998). Hairs, bristles, and scales. In F. Harrison & M. Locke (Eds.), *Microscopic Anatomy*  
737 *of Invertebrates* (Vol. 11A, pp. 257–287). Wiley-Liss.

738 Gomez, D., Pinna, C., Pairraire, J., Arias, M., Barbut, J., Pomerantz, A., Daney de Marcillac, W., Berthier,  
739 S., Patel, N., Andraud, C., & Elias, M. (2021). Wing transparency in butterflies and moths:  
740 Structural diversity, optical properties and ecological relevance. *Ecological Monographs*,  
741 91(4), e01475. <https://doi.org/10.1002/ecm.1475>

742 Hadfield, J. D. (2010). MCMC Methods for Multi-Response Generalized Linear Mixed Models: The  
743 **MCMCglmm** R Package. *Journal of Statistical Software*, 33(2).  
744 <https://doi.org/10.18637/jss.v033.i02>

745 Hasan, J., Webb, H. K., Truong, V. K., Watson, G. S., Watson, J. A., Tobin, M. J., Gervinskas, G., Juodkazis,  
746 S., Wang, J. Y., Crawford, R. J., & Ivanova, E. P. (2012). Spatial Variations and Temporal  
747 Metastability of the Self-Cleaning and Superhydrophobic Properties of Damselfly Wings.  
748 *Langmuir*, 28(50), 17404–17409. <https://doi.org/10.1021/la303560w>

749 Ivanova, E. P., Hasan, J., Webb, H. K., Truong, V. K., Watson, G. S., Watson, J. A., Baulin, V. A., Pogodin,  
750 S., Wang, J. Y., Tobin, M. J., Löbbecke, C., & Crawford, R. J. (2012). Natural bactericidal surfaces:  
751 Mechanical rupture of *Pseudomonas aeruginosa* cells by cicada wings. *Small*, 8(16), 2489–  
752 2494. <https://doi.org/10.1002/smll.201200528>

753 Kemp, D. J. (2007). Female butterflies prefer males bearing bright iridescent ornamentation.  
754 *Proceedings of the Royal Society B-Biological Sciences*, 274(1613), 1043–1047.  
755 <https://doi.org/10.1098/rspb.2006.0043>

756 Kong, X. Q., Liu, J. L., Zhang, W. J., & Qu, Y. D. (2015). Load-bearing ability of the mosquito tarsus on  
757 water surfaces arising from its flexibility. *AIP Advances*, 5(3), 037101.  
758 <https://doi.org/10.1063/1.4908027>

759 Kovalev, A., Rebora, M., Salerno, G., & Gorb, S. (2020). Air-entrapping capacity in the hair coverage of  
760 *Malacosoma castrensis* (Lasiocampidae: Lepidoptera) caterpillar: a case study. *Journal of*  
761 *Experimental Biology*, jeb.225029. <https://doi.org/10.1242/jeb.225029>

762 Krishna, A., Nie, X., Warren, A. D., Llorente-Bousquets, J. E., Briscoe, A. D., & Lee, J. (2020). Infrared  
763 optical and thermal properties of microstructures in butterfly wings. *Proceedings of the*  
764 *National Academy of Sciences*, 117(3), 1566–1572. <https://doi.org/10.1073/pnas.1906356117>

765 Li, Y., Li, X., Liu, X., Zhu, B., Muzammil, I., Lei, M., & Lakhtakia, A. (2020). Biomimetic Random Arrays of  
766 Nanopillars and Nanocones with Robust Antiwetting Characteristics. *Journal of Physical*  
767 *Chemistry C*, 124(31), 17095–17102. <https://doi.org/10.1021/acs.jpcc.0c04804>

768 Liu, K., Vuckovac, M., Latikka, M., Huhtamäki, T., & Ras, R. H. A. (2019). Improving surface-wetting  
769 characterization. *Science*, 363(6432), 1147–1148. <https://doi.org/10.1126/science.aav5388>

770 Mahadik, G. A., Hernandez-Sanchez, J. F., Arunachalam, S., Gallo, A., Cheng, L., Farinha, A. S.,  
771 Thoroddsen, S. T., Mishra, H., & Duarte, C. M. (2020). Superhydrophobicity and size reduction  
772 enabled *Halobates* (Insecta: Heteroptera, Gerridae) to colonize the open ocean. *Scientific*  
773 *Reports*, 10(1), 7785. <https://doi.org/10.1038/s41598-020-64563-7>

774 McClure, M., Clerc, C., Desbois, C., Meichanetzoglou, A., Cau, M., Bastin-Héline, L., Bacigalupo, J.,  
775 Houssin, C., Pinna, C., Nay, B., Llaurens, V., Berthier, S., Andraud, C., Gomez, D., & Elias, M.  
776 (2019). Why has transparency evolved in aposematic butterflies? Insights from the largest  
777 radiation of aposematic butterflies, the Ithomiini. *Proceedings of the Royal Society B: Biological Sciences*, 286(1901), 20182769. <https://doi.org/10.1098/rspb.2018.2769>

779 McHale, G., Aqil, S., Shirtcliffe, N. J., Newton, M. I., & Erbil, H. Y. (2005). Analysis of droplet evaporation  
780 on a superhydrophobic surface. *Langmuir*, 21(24), 11053–11060.  
781 <https://doi.org/10.1021/la0518795>

782 Miaoulis, I. N., & Heilman, B. D. (1998). Butterfly thin films serve as solar collectors. *Annals of the*  
783 *Entomological Society of America*, 91(1), 122–127. <https://doi.org/10.1093/aesa/91.1.122>

784 Nachtingall, W. (1967). Aerodynamische Messungen am Tragfluegelsystem segeinder Schmetterlinge.  
785 *Journal of Comparative Physiology A*, 54, 210–231.

786 Oh, J., Dana, C. E., Hong, S., Roman, J. K., Jo, K. D., Hong, J. W., Nguyen, J., Cropek, D. M., Alleyne, M.,  
787 & Miljkovic, N. (2017). Exploring the Role of Habitat on the Wettability of Cicada Wings. *Acsl Applied Materials & Interfaces*, 9(32), 27173–27184.  
788 <https://doi.org/10.1021/acsami.7b07060>

790 Otten, A., & Herminghaus, S. (2004). How plants keep dry: A physicist's point of view. *Langmuir*, 20(6),  
791 2405–2408. <https://doi.org/10.1021/la034961d>

792 Patankar, N. A. (2004). Mimicking the lotus effect: Influence of double roughness structures and  
793 slender pillars. *Langmuir*, 20(19), 8209–8213. <https://doi.org/10.1021/la048629t>

794 Perez Goodwyn, P., Maezono, Y., Hosoda, N., & Fujisaki, K. (2009). Waterproof and translucent wings  
795 at the same time: Problems and solutions in butterflies. *Naturwissenschaften*, 96(7), 781–787.  
796 <https://doi.org/10.1007/s00114-009-0531-z>

797 Pinna, C., Vilbert, M., Borensztajn, S., Marcillac, W. D. de, Piron-Prunier, F., Pomerantz, A., Patel, N.,  
798 Berthier, S., Andraud, C., Gomez, D., & Elias, M. (2021). Convergence in light transmission  
799 properties of transparent wing areas in clearwing mimetic butterflies. *eLife*, 10, e69080.  
800 <https://doi.org/10.7554/eLife.69080>

801 Plummer, M., Best, N., Cowles, K., & Vines, K. (2006). CODA: *Convergence Diagnosis and Output*  
802 *Analysis for MCMC: Vol. 6(1)* (R News, pp. 7–11). <https://journal.r-project.org/archive/>

803 Pomerantz, A. F., Siddique, R. H., Cash, E. I., Kishi, Y., Pinna, C., Hammar, K., Gomez, D., Elias, M., &  
804 Patel, N. H. (2021). Developmental, cellular, and biochemical basis of transparency in the  
805 glasswing butterfly *Greta oto*. *Journal of Experimental Biology*, 224, eb237917.  
806 <https://doi.org/10.1242/jeb.237917>

807 Porcheron, F., & Monson, P. A. (2006). Mean-Field Theory of Liquid Droplets on Roughened Solid  
808 Surfaces: Application to Superhydrophobicity. *Langmuir*, 22(4), 1595–1601.  
809 <https://doi.org/10.1021/la051946v>

810 Reyssat, M., & Quéré, D. (2009). Contact Angle Hysteresis Generated by Strong Dilute Defects. *The*  
811 *Journal of Physical Chemistry B*, 113(12), 3906–3909. <https://doi.org/10.1021/jp8066876>

812 Reyssat, M., Yeomans, J. M., & Quéré, D. (2007). Impalement of fakir drops. *EPL (Europhysics Letters)*,  
813 81(2), 26006. <https://doi.org/10.1209/0295-5075/81/26006>

814 Sajadinia, S. H., & Sharif, F. (2010). Thermodynamic analysis of the wetting behavior of dual scale  
815 patterned hydrophobic surfaces. *Journal of Colloid and Interface Science*, 344(2), 575–583.  
816 <https://doi.org/10.1016/j.jcis.2009.12.058>

817 Sanchez-Monge, A., Rodriguez Arrieta, J., Jimenez-Chavarria, M., & Retana-Salazar, A. (2015).  
818 Observations on the Ultrastructure and Hydrophobicity of the Wings of Thirteen Neotropical  
819 Families of Diptera (insecta) with Comments on Their Flight. *Acta Microscopica*, 24(2), 111–  
820 117.

821 Schneider, C. A., Rasband, W. S., & Eliceiri, K. W. (2012). NIH Image to ImageJ: 25 years of image  
822 analysis. *Nature Methods*, 9(7), 671–675. <https://doi.org/10.1038/nmeth.2089>

823 Siddique, R. H., Gomard, G., & Hölscher, H. (2015). The role of random nanostructures for the  
824 omnidirectional anti-reflection properties of the glasswing butterfly. *Nature Communications*,  
825 6, 6909. <https://doi.org/10.1038/ncomms7909>

826 Skowron Volponi, M. A., McLean, D. J., Volponi, P., & Dudley, R. (2018). Moving like a model: Mimicry  
827 of hymenopteran flight trajectories by clearwing moths of Southeast Asian rainforests. *Biology  
828 Letters*, 14(5), 20180152. <https://doi.org/10.1098/rsbl.2018.0152>

829 Slegers, N., Heilman, M., Cranford, J., Lang, A., Yoder, J., & Habegger, M. L. (2017). Beneficial  
830 aerodynamic effect of wing scales on the climbing flight of butterflies. *Bioinspiration &  
831 Biomimetics*, 12(1), 016013. <https://doi.org/10.1088/1748-3190/aa551d>

832 Stevens, M., Stubbins, C. L., & Hardman, C. J. (2008). The anti-predator function of “eyespots” on  
833 camouflaged and conspicuous prey. *Behavioral Ecology and Sociobiology*, 62(11), 1787–1793.  
834 <https://doi.org/10.1007/s00265-008-0607-3>

835 Stoffel, M. A., Nakagawa, S., & Schielzeth, H. (2017). rptR: Repeatability estimation and variance  
836 decomposition by generalized linear mixed-effects models. *Methods in Ecology and Evolution*,  
837 8(11), 1639–1644. <https://doi.org/10.1111/2041-210X.12797>

838 Su, Y., Ji, B., Zhang, K., Gao, H., Huang, Y., & Hwang, K. (2010). Nano to Micro Structural Hierarchy Is  
839 Crucial for Stable Superhydrophobic and Water-Repellent Surfaces. *Langmuir*, 26(7), 4984–  
840 4989. <https://doi.org/10.1021/la9036452>

841 Sugumaran, M. (2009). Complexities of cuticular pigmentation in insects. *Pigment Cell & Melanoma  
842 Research*, 22(5), 523–525. <https://doi.org/10.1111/j.1755-148X.2009.00608.x>

843 Sun, G., & Fang, Y. (2015). Anisotropic Characteristic of Insect (Lepidoptera) wing Surfaces. In S.  
844 Yingying, C. Guiran, & L. Zhen (Eds.), *Roceedings of the international conference on logistics,  
845 engineering, management, and computer science (LEMCS 2015)* (Vol. 117, pp. 633–635).  
846 Atlantis Press.

847 Sun, M., Liang, A., Watson, G. S., Watson, J. A., Zheng, Y., Ju, J., & Jiang, L. (2012). Influence of Cuticle  
848 Nanostructuring on the Wetting Behaviour/States on Cicada Wings. *PLOS ONE*, 7(4), e35056.  
849 <https://doi.org/10.1371/journal.pone.0035056>

850 Sun, M., Watson, G. S., Zheng, Y., Watson, J. A., & Liang, A. (2009). Wetting properties on  
851 nanostructured surfaces of cicada wings. *Journal of Experimental Biology*, 212(19), 3148–  
852 3155. <https://doi.org/10.1242/jeb.033373>

853 Tsai, C.-C., Childers, R. A., Nan Shi, N., Ren, C., Pelaez, J. N., Bernard, G. D., Pierce, N. E., & Yu, N. (2020).  
854 Physical and behavioral adaptations to prevent overheating of the living wings of butterflies.  
855 *Nature Communications*, 11(1), 551. <https://doi.org/10.1038/s41467-020-14408-8>

856 Tsai, P., Lammertink, R. G. H., Wessling, M., & Lohse, D. (2010). Evaporation-Triggered Wetting  
857 Transition for Water Droplets upon Hydrophobic Microstructures. *Physical Review Letters*,  
858 104(11), 116102. <https://doi.org/10.1103/PhysRevLett.104.116102>

859 Wagner, T., Neinhuis, C., & Barthlott, W. (1996). Wettability and contaminability of insect wings as a  
860 function of their surface sculptures. *Acta Zoologica*, 77(3), 213–225.  
861 <https://doi.org/10.1111/j.1463-6395.1996.tb01265.x>

862 Wan, Q., Li, H., Zhang, S., Wang, C., Su, S., Long, S., & Pan, B. (2019). Combination of active behaviors  
863 and passive structures contributes to the cleanliness of housefly wing surfaces: A new insight  
864 for the design of cleaning materials. *Colloids and Surfaces B-Biointerfaces*, 180, 473–480.  
865 <https://doi.org/10.1016/j.colsurfb.2019.05.010>

866 Wanasekara, N. D., & Chalivendra, V. B. (2011). Role of surface roughness on wettability and  
867 coefficient of restitution in butterfly wings. *Soft Matter*, 7(2), 373–379.  
868 <https://doi.org/10.1039/c0sm00548g>

869 Watson, G. S., Cribb, B. W., & Watson, J. A. (2011). Contrasting micro/nano architecture on termite  
870 wings: Two divergent strategies for optimising success of colonisation flights. *PLOS ONE*, 6(9),  
871 e24368. <https://doi.org/10.1371/journal.pone.0024368>

872 Watson, G. S., Green, D. W., Schwarzkopf, L., Li, X., Cribb, B. W., Myhra, S., & Watson, J. A. (2015). A  
873 gecko skin micro/nano structure – A low adhesion, superhydrophobic, anti-wetting, self-  
874 cleaning, biocompatible, antibacterial surface. *Acta Biomaterialia*, 21, 109–122.  
875 <https://doi.org/10.1016/j.actbio.2015.03.007>

876 Watson, G. S., Myhra, S., Cribb, B. W., & Watson, J. A. (2008). Putative functions and functional  
877 efficiency of ordered cuticular nanoarrays on insect wings. *Biophysical Journal*, 94(8), 3352–  
878 3360. <https://doi.org/10.1529/biophysj.107.109348>

879 Wenzel, R. N. (1936). Resistance of solid surfaces to wetting by water. *Industrial & Engineering  
880 Chemistry*, 28(8), 988–994. <https://doi.org/10.1021/ie50320a024>

881 Wu, C. W., Kong, X. Q., & Wu, D. (2007). Micronanostructures of the scales on a mosquito's legs and  
882 their role in weight support. *Physical Review E*, 76(1), 017301.  
883 <https://doi.org/10.1103/PhysRevE.76.017301>

884 Yoshida, A., Motoyama, M., Kosaku, A., & Miyamoto, K. (1997). Antireflective nanoprotuberance array  
885 in the transparent wing of a hawkmoth, *Cephonodes hylas*. *Zoological Science*, 14(5), 737–741.  
886 <https://doi.org/10.2108/zsj.14.737>

887 Zheng, Y. M., Gao, X. F., & Jiang, L. (2007). Directional adhesion of superhydrophobic butterfly wings.  
888 *Soft Matter*, 3(2), 178–182. <https://doi.org/10.1039/B612667G>

889

890