

Biolearn, an open-source library for biomarkers of aging

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Abstract

Identifying and validating biomarkers of aging is pivotal for understanding the aging process and testing longevity interventions. Despite the development of numerous aging biomarkers, their clinical validation remains elusive, largely due to the lack of cross-population validation, which is hampered by disparate biomarker designs and inconsistencies in dataset structures. To bridge this gap, we introduce Biolearn, an innovative open-source library dedicated to the implementation and application of aging biomarkers. Biolearn facilitates (1) harmonization of existing aging biomarkers, while presenting a structured framework for novel biomarkers in

24 standardized formats; (2) unification of public datasets, ensuring coherent structuring and
 25 formatting, thus simplifying cross-population validation studies; and (3) provision of
 26 computational methodologies to assess any harmonized biomarker against unified datasets. By
 27 furnishing a community-driven platform, Biolearn significantly augments the development,
 28 assessment, and validation trajectories of aging biomarkers, paving the way toward more
 29 rigorous clinical validation and, ultimately, application in clinical trials targeting healthy
 30 longevity. The Biolearn package is open-source and freely available at
 31 <https://Bio-Learn.github.io/>

32 Introduction

33 The quest for reliable biomarkers of aging (BoAs) is an essential focus in aging research, driven
 34 by a growing understanding of aging as a fundamental factor in chronic diseases and mortality. A
 35 plethora of biomarkers have been proposed to quantify biological age or the pace of aging and
 36 elucidate the underlying biological intricacies; however, their clinical validation remains a
 37 significant challenge due to varied formulations and disparate structures in validation datasets
 38 across populations. Since the introduction of composite omic biomarkers of aging, epitomized by
 39 Horvath's seminal work on DNA methylation aging clocks ¹, subsequent efforts have expanded
 40 the repertoire of aging biomarkers, which now span a wide array of omic modalities, including
 41 epigenomics, transcriptomics, and proteomics ²⁻⁷.

42 DNA methylation-based clocks are currently the most developed class of omic biomarkers of
 43 aging and represent robust tools for biological age estimation, offering insights into age-related
 44 changes at the molecular level and their implications for human health and longevity ^{1,2,4}. For
 45 instance, the Horvath multi-tissue clock and the subsequent DNA methylation-based clocks, such
 46 as DunedinPACE ³, GrimAge ⁸, causality-enriched DamAge/AdaptAge ⁹, PRC2 clock ¹⁰, and
 47 others, have shown promising correlations with various age-related conditions and mortality,
 48 reflecting the intricate interplay between epigenetic modifications and the aging trajectory ^{4,8,11,12}.
 49 However, the diverse formulation of these biomarkers and the inconsistency in structure across
 50 different validation datasets pose substantial hurdles for systematic cross-population validation
 51 and benchmarking of these biomarkers, critical steps towards their clinical translation.

Publicly available datasets, such as those from the Gene Expression Omnibus¹³, the National Health and Nutrition Examination Survey (NHANES), and the Framingham Heart Study (FHS) have the potential to greatly accelerate the validation of biomarkers of aging. However, their use for this purpose is complicated by the lack of a standardized framework that can accommodate the heterogeneous nature of their datasets. Thus, there is a clear need for a unified platform that can seamlessly integrate and analyze various biomarkers of aging across datasets with harmonized structures. Such a platform would transform the validation process, foster the discovery of novel biomarkers, and provide a structured avenue for community-driven efforts in advancing the field of aging biology.

To address this need, we introduce Biolearn, an open-source library designed to bridge these gaps by providing a unified framework for the harmonization and computational analysis of BoAs (Figure 1a). Biolearn serves as an innovative tool that harmonizes existing BoAs, structures, and formats human datasets and offers computational methodologies for assessing biomarkers against these datasets. By fostering a community-driven approach, Biolearn aims to propel the development and validation process of BoAs, ultimately contributing to a deeper understanding of the aging process, facilitating the discovery of interventions for age-related diseases, and bringing BoAs to the clinic.

Results

Harmonization of Biomarkers of Aging

We successfully harmonized 22 well-established epigenetic aging clocks and biomarkers (Table 1), as well as two phenotypic biomarkers (Phenotypic Age and Mahalanobis distance metrics) and implemented these BoAs in Biolearn. The epigenetic biomarkers include chronological clocks like Horvath's multi-tissue clock, Hannum's blood clock^{1,14}; healthspan and mortality-related clocks like GrimAge, GrimAge2, PhenoAge, and Zhang clock^{8,15–17}; biomarkers of the rate of aging including DunedinPoAm38 and DunedinPACE^{3,18}; causality-enriched clocks including Ying's CausAge, DamAge, and AdaptAge⁹; as well as many other clocks and DNAm-based biomarkers (Table 1). All of the biomarkers were formatted into standardized input structures, allowing for consistent application across disparate datasets. This harmonization process involved collecting and unifying the annotation of clock specifications,

such as tissue type, predicted age range, and source references, ensuring transparent and reproducible analyses. To further support ongoing research in this field, we developed and implemented an open-source framework (<https://github.com/bio-learn/biolearn/blob/master/biolearn/model.py>). This framework is designed to provide a standardized format for epigenetic biomarkers, facilitating the seamless integration and comparison of any future aging clocks that are developed. This addition aims to foster collaboration and innovation in the study of aging biomarkers.

Integration of Diverse Human Datasets

To facilitate cross-population validation studies using publicly available data, we harnessed Biolearn's capabilities to integrate and structure multiple public datasets (Table 2). The structured datasets were refined to enable a shared analysis platform, addressing the challenges of data heterogeneity and formatting inconsistencies^{13,19}. With this capacity, the Biolearn is used as the backend of ClockBase for epigenetic age computation¹³, enabling the systemic harmonization of over 200,000 human samples from Gene Expression Omnibus (GEO) array data.

Computational Analysis and Biomarker Assessment

Using Biolearn's computational methodologies, we conducted extensive evaluations of the harmonized biomarkers of aging. As an example, we applied several models to two GEO dataset GSE41169 (N = 95) and GSE64495 (N = 113), with DNA methylation profiles across approximately 480,000 CpGs²⁰. We show that with a few lines of code, Biolearn is able to efficiently compute results for over 20 epigenetic biomarkers across hundreds of samples on the order of seconds (Figure 1b-c). Biolearn also allows rapid comparison of the performance of different biomarkers in independent datasets (Figure 1d). For example, in both testing datasets, the top 5 performing clocks, in terms of R^2 with chronological age, are the Zhang clock, Horvath v2 and v1, Hannum clock, and YingCausAge (Figure 1d).

Beyond aging biomarkers, we also integrated several common epigenetic predictors in Biolearn. For instance, sex can be inferred (Wang et al.²¹) from DNAm profiles with high accuracy (Figure 1e).

109 Clinical Data Harmonization

110 To provide further utility in handling clinical data, we implemented the blood-test-based
 111 Phenotypic Age ¹⁶, and Mahalanobis distance-based biomarker in Biolearn ²². These clinical
 112 biomarkers may be combined with the analysis tools built into Biolearn. For example, we
 113 calculated Phenotypic Ages for the NHANES 2010 dataset and illustrated the relationship
 114 between biological and chronological age using Biolearn. We further performed a survival
 115 analysis that distinguished between individuals with accelerated versus decelerated aging based
 116 on biological age discrepancies. The entire analysis was completed with only a few lines of code
 117 (Figure 2a-b) ¹⁶.

118 Discussion

119 Among the most significant challenges in aging biomarker research is the cross-population
 120 validation of proposed biomarkers ²³. To take steps to address this need and provide an
 121 open-source tool for validation efforts across the field, we built Biolearn to integrate a broad
 122 range of datasets, including those with varied formats and structures, and provide a suite of
 123 analysis tools. By standardizing data inputs and modeling approaches across multiple
 124 populations, Biolearn has demonstrated its potential to facilitate more extensive and robust
 125 validation processes that are essential for the clinical translation of biomarkers of aging ⁴.

126 With Biolearn, we have also harmonized and evaluated several well-established aging clocks,
 127 providing the opportunity for these biomarkers to be refined and potentially for new ones to be
 128 developed. The modular design of Biolearn encourages the addition of new models and datasets,
 129 making it a living library that will grow in tandem with the field itself. By centralizing resources
 130 and knowledge, Biolearn considerably reduces redundancy and accelerates biomarker
 131 development and validation efforts ^{3,7}.

132 Our approach emphasizes transparency and reproducibility, core tenets of open science. By
 133 making Biolearn publicly available and maintaining detailed documentation and development
 134 guidelines, we have established an ecosystem that supports open collaboration and knowledge
 135 sharing. This open-science framework ensures that findings and tools can be widely accessed,
 136 providing equitable opportunities for researchers globally to contribute to and benefit from the
 137 collective advances in aging research. Moreover, our hope is that the open-access nature of

138 Biolearn will promote cross-fertilization between aging researchers and scientists currently
139 outside of the field, incentivizing the development of novel and innovative biomarker models
140 and validation approaches.

141 Previous efforts to harmonize biomarkers of aging, notably methylCIPHER and BioAge^{24,25},
142 have been limited in scope, focusing on methylation or blood-based biomarkers only.
143 Furthermore, being R packages limits their scope of use in production. Biolearn supports
144 biomarkers based on multiple different biological data modalities and is written in Python, which
145 has a much broader reach. In comparison to PyAging²⁶, a preliminary contemporaneous Python
146 biomarker library, Biolearn is focused on ease of use and reproducibility through automated
147 testing against reference data.

148 While Biolearn represents a significant advance for the field, certain limitations remain.
149 Currently, the library is tailored to biomarkers derived from biological samples, predominantly
150 DNA methylation data. Moving forward, the scope of Biolearn will continue to expand to
151 encompass diverse biological modalities—such as proteomics, metabolomics, and
152 microbiomics—broadening its applicability^{5,10}. Moreover, integration with larger and more
153 diverse population datasets will be vital in advancing cross-population validation efforts. As new
154 datasets emerge, Biolearn will adapt to incorporate these resources, ensuring ongoing robustness
155 and scalability²⁷. Finally, bioinformatics tools, including Biolearn, depend on a user base
156 proficient in programming and data analysis. Efforts to make these tools more accessible to a
157 wider audience, including those with limited computational expertise, will be crucial. This could
158 involve the development of graphical user interfaces (GUIs) or web-based platforms to
159 streamline the user experience.

160 We anticipate that Biolearn will become a key resource for the field and will transform many
161 facets of aging biomarker studies. Our preliminary survival studies conducted using Biolearn
162 demonstrate not only the power of this new platform, but illuminate the real-world implications
163 of validated biomarkers. Biolearn's standardization and analysis capabilities stand to serve as
164 pivotal tools for researchers seeking to bridge the gap between biomarker discovery and clinical
165 implementation²³.

166 **Methods**

167 **Overview of Biolearn Library**

168 Biolearn is an open-source computational suite that facilitates the harmonization and analysis of
169 biomarkers of aging (BoAs). It is written in Python and is readily accessible through the Python
170 Package Index (PyPI). Biolearn is developed using modern software engineering practices,
171 including automated testing to ensure correctness and adherence to software design principles
172 that ensure the safe interchangeability of like components. The library is designed to be
173 user-friendly while offering robust functionalities for researchers across various disciplines
174 involved in aging studies.

175 **System Requirements and Installation**

176 Biolearn requires Python version 3.10 or newer. It can be installed using the Python package
177 manager, pip, with the command `pip install biolearn`. The successful installation of the
178 library can be verified through the import test of Biolearn's core classes. The library is
179 cross-platform and is compatible with major operating systems, including Windows, MacOS,
180 and Linux. The more detailed instructions can be found here:
181 <https://Biolearn.github.io/quickstart.html>

182 **Data Library and Model Gallery**

183 Biolearn incorporates a data library capable of loading and structuring datasets from a multitude
184 of public sources like Gene Expression Omnibus (GEO), National Health and Nutrition
185 Examination Survey (NHANES), and Framingham Heart Study. The model gallery within
186 Biolearn holds reference implementations for various aging clocks and biomarkers, presenting a
187 unified interface for users to apply these models to their datasets. All models were verified to be
188 correct by comparing the outputs on a reference data set against their original implementations
189 where available.

190 **Harmonization Process**

191 We used Biolearn to harmonize several aging clocks. Clock definitions were standardized,
192 specifying the name, publication year, applicable species, target tissue, and the biological aspect
193 they predict (e.g., age, mortality risk). We provided sources for both the original publications and
194 the coefficients necessary for clock applications. Coherence across biological modalities and

195 datasets was assured through Biolearn’s systematic approach to data preprocessing,
196 normalization, and imputation procedures.

197 **Integration with Public Datasets**

198 Biolearn’s ability to interface seamlessly with public datasets was tested by integrating and
199 formatting data from GEO and NHANES. Preprocessing pipelines were developed to convert
200 raw data into a harmonized format suitable for subsequent analysis. Particular attention was
201 given to metadata structures, variable normalization, and missing data treatment, ensuring
202 consistent input formats required by the aging models.

203 **Statistical Analysis**

204 All statistical analyses were performed using tools embedded within the Biolearn library or
205 through integration with renowned Python statistics libraries such as statsmodels and seaborn for
206 visualization. The robustness and reproducibility of the analysis were ensured through the use of
207 randomized cross-validation techniques for model assessment and bootstrapping methods for
208 estimating confidence intervals where applicable.

209 **Acknowledgments**

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213 Foundation, Methuselah Foundation, and VoLo Foundation.

214

215 Tables and Figures

216 **Table 1.** Harmonized biomarkers of aging in Biolearn.

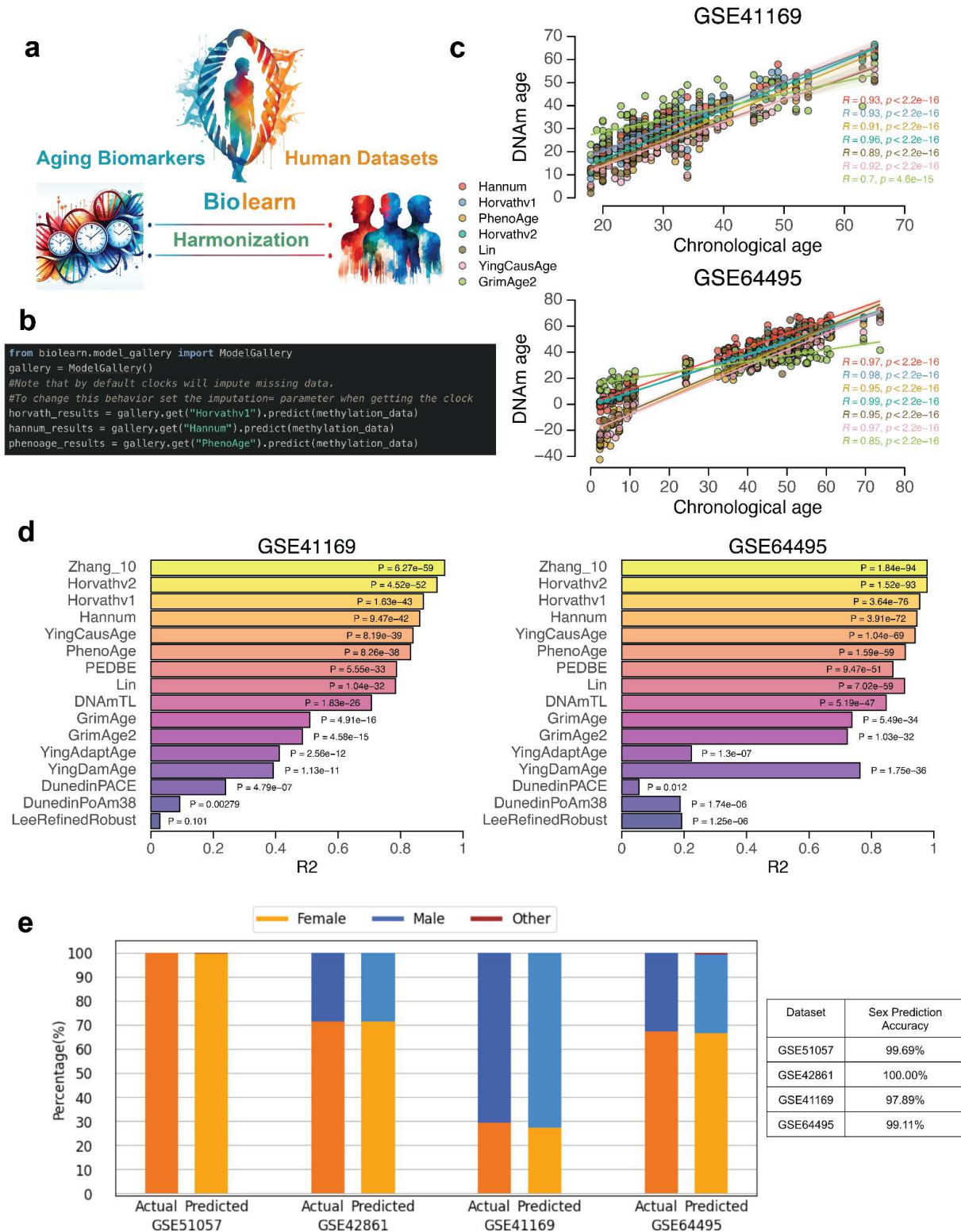
Name	Year	Species	Tissue	Predicts
Horvathv1 ¹	2013	Human	Multi-tissue	Age (Years)
Hannum ¹⁴	2013	Human	Blood	Age (Years)
Lin ²⁸	2016	Human	Blood	Age (Years)
PhenoAge ¹⁶	2018	Human	Blood	Age (Years)
Horvathv2 ²⁹	2018	Human	Skin + blood	Age (Years)
PEDBE ³⁰	2019	Human	Buccal	Age (Years)
Zhang_10 ¹⁷	2019	Human	Blood	Mortality Risk
GrimAge ⁸	2019	Human	Blood	Age Adjusted by Mortality Risk (Years)
GrimAge2 ¹⁵	2022	Human	Blood	Age Adjusted by Mortality Risk (Years)
DunedinPoAm38 ¹⁸	2020	Human	Blood	Aging Rate (Years/Year)
DunedinPACE ³	2022	Human	Unknown	Aging Rate (Years/Year)
AlcoholMcCartney ³¹	2018	Human	Blood	Alcohol Consumption
BMI_McCartney ³¹	2018	Human	Blood	BMI
DNAmTL ³²	2019	Human	Blood, Adipose	Telomere Length
Knight ³³	2016	Human	Cord Blood	Gestational Age
LeeControl ³⁴	2019	Human	Placenta	Gestational Age
LeeRefinedRobust ³⁴	2019	Human	Placenta	Gestational Age
LeeRobust ³⁴	2019	Human	Placenta	Gestational Age
YingCausAge/DamAge/AdaptAge ⁹	2022	Human	Blood	Age (Years)
SmokingMcCartney ³¹	2018	Human	Blood	Smoking Status

217

218 Table 2. Harmonized datasets in Biolearn.

ID	Title	Format	Samples	Age Present	Sex Present
GSE40279	Genome-wide Methylation Profiles Reveal Quantitative Views o...	Illumina450k	656	Yes	Yes
GSE19711	Genome wide DNA methylation profiling of United Kingdom Ovar...	Illumina27k	540	Yes	No
GSE51057	Methylome Analysis and Epigenetic Changes Associated with Me...	Illumina450k	329	Yes	Yes
GSE42861	Differential DNA methylation in Rheumatoid arthritis	Illumina450k	689	Yes	Yes
GSE41169	Blood DNA methylation profiles in a Dutch population	Illumina450k	95	Yes	Yes
GSE51032	EPIC-Italy at HuGeF	Illumina450k	845	Yes	No
GSE73103	Many obesity-associated SNPs strongly associate with DNA met...	Illumina450k	355	Yes	Yes
GSE69270	Aging-associated DNA methylation changes in middle-aged indi...	Illumina450k	184	Yes	No
GSE36054	Methylation Profiling of Blood DNA from Healthy Children	Illumina450k	192	No	No
GSE64495	DNA methylation profiles of human blood samples from a sever...	Illumina450k	113	Yes	Yes
GSE30870	DNA methylomes of Newborns and Nonagenarians	Illumina450k	40	Yes	No
NHANES	National Health and Nutrition Examination Survey	Phenotypic	2877	Yes	Yes
FHS	Framingham Heart Study	Phenotypic	4434	Yes	Yes

219



220

Figure 1. a. Overview of Biolearn's functionalities. **b.** The code snippet showing that the DNAm age can be calculated with a few lines of code using Biolearn library. **c.** Scatter plot of predicted

age (y-axis) vs. chronological age (x-axis) for the GSE41169 (N = 95) and GSE64495 (N = 113) datasets. The top 7 clocks with the smallest mean absolute errors are shown in the plot. Pearson's R and unadjusted P-values based on two-sided tests are provided. **d.** Bar graph showing the R-square (x-axis) of methylation biomarker prediction vs. chronological age. The color indicates the overall rank of the clock based on the R-square value. The unadjusted P-values based on two-sided tests are shown on the bar. **e.** Stacked bar graph shows the actual vs. predicted sex distribution in four different datasets: GSE51057, GSE42861, GSE41169, and GSE64495. The accompanying table provides the sex prediction accuracy for each dataset.

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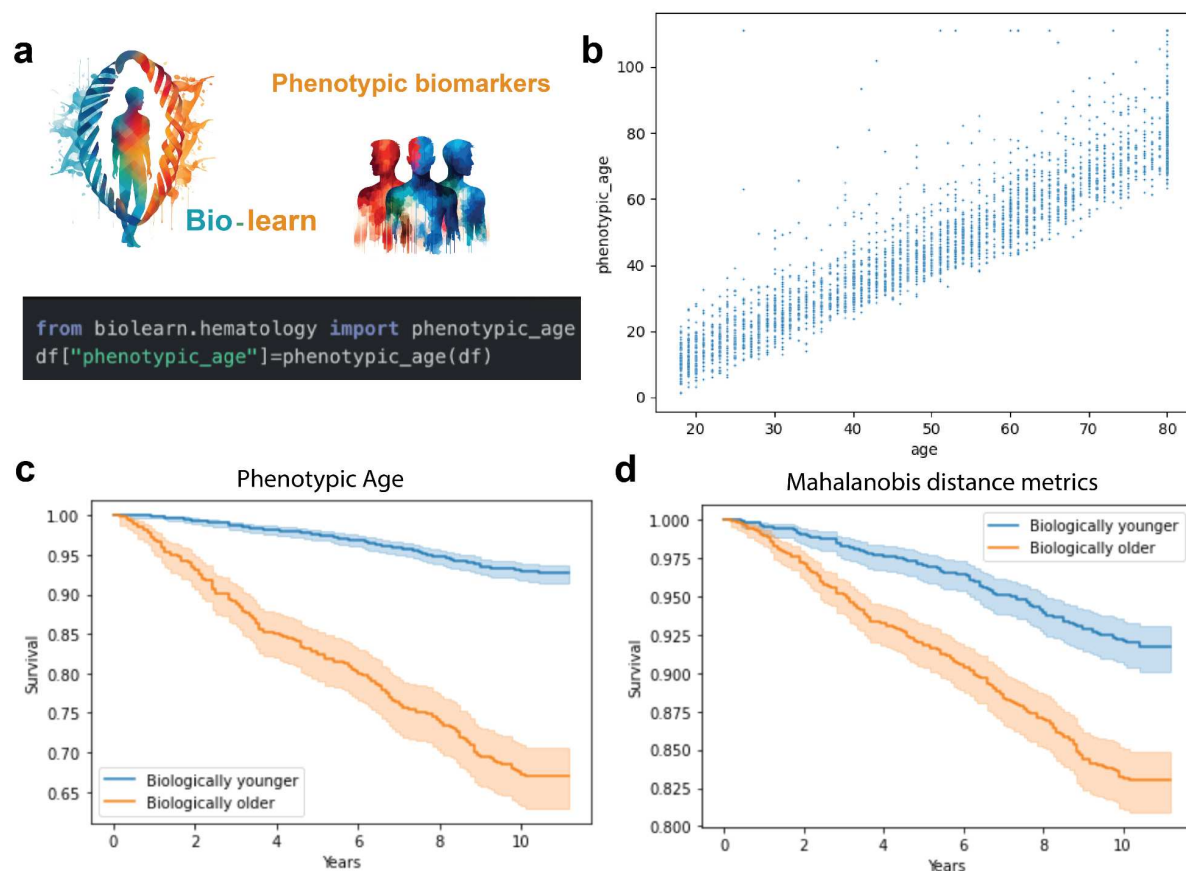


Figure 2. **a.** Overview of Biolearn's phenotypic biomarker functionalities. The code snippet shows that the DNAm age can be calculated with a few lines of code using the Biolearn library. **b.** Scatter plot of chronological age vs. phenotypic age for the NHANES 2010 dataset. **c, d.** Survival analysis of the NHANES 2010 dataset (N = 2877), stratified by biological age discrepancies (marked by different colors) based on Phenotypic Age (c) and Mahalanobis distance metrics (d). Individuals with biological age higher than chronological age are marked as biologically older and vice versa. For the purpose of demonstration, the result is not adjusted by chronological age. The shadow shows the standard error.

242 References

- 243 1. Horvath, S. DNA methylation age of human tissues and cell types. *Genome Biology* **14**, R115 (2013).
- 244 2. Bell, C. G. *et al.* DNA methylation aging clocks: challenges and recommendations. *Genome Biol* **20**, 249 (2019).
- 245 3. Belsky, D. W. *et al.* DunedinPACE, a DNA methylation biomarker of the pace of aging. *Elife* **11**, e73420 (2022).
- 246 4. Chen, B. H. *et al.* DNA methylation-based measures of biological age: meta-analysis predicting time to death. *Aging* **8**, 1844–1865 (2016).
- 247 5. Galkin, F. *et al.* Biohorology and biomarkers of aging: Current state-of-the-art, challenges and opportunities. *Ageing Research Reviews* **60**,
248 101050 (2020).
- 249 6. Horvath, S. & Raj, K. DNA methylation-based biomarkers and the epigenetic clock theory of ageing. *Nat Rev Genet* **19**, 371–384 (2018).
- 250 7. Jylhävä, J., Pedersen, N. L. & Hägg, S. Biological Age Predictors. *EBioMedicine* **21**, 29–36 (2017).
- 251 8. Lu, A. T. *et al.* DNA methylation GrimAge strongly predicts lifespan and healthspan. *Aging* **11**, 303–327 (2019).
- 252 9. Ying, K. *et al.* Causal Epigenetic Age Uncouples Damage and Adaptation. 2022.10.07.511382 Preprint at
253 <https://doi.org/10.1101/2022.10.07.511382> (2022).
- 254 10. Moqri, M. *et al.* PRC2 clock: a universal epigenetic biomarker of aging and rejuvenation. 2022.06.03.494609 Preprint at
255 <https://doi.org/10.1101/2022.06.03.494609> (2022).
- 256 11. Marioni, R. E. *et al.* The epigenetic clock is correlated with physical and cognitive fitness in the Lothian Birth Cohort 1936. *Int. J.*
257 *Epidemiol.* **44**, 1388–1396 (2015).
- 258 12. Field, A. E. *et al.* DNA Methylation Clocks in Aging: Categories, Causes, and Consequences. *Molecular Cell* **71**, 882–895 (2018).
- 259 13. Ying, K. *et al.* ClockBase : a comprehensive platform for biological age profiling in human and mouse. 2023.02.28.530532 Preprint at
260 <https://doi.org/10.1101/2023.02.28.530532> (2023).
- 261 14. Hannum, G. *et al.* Genome-wide Methylation Profiles Reveal Quantitative Views of Human Aging Rates. *Molecular Cell* **49**, 359–367
262 (2013).
- 263 15. Lu, A. T. *et al.* DNA methylation GrimAge version 2. *Aging* **14**, 9484–9549 (2022).
- 264 16. Levine, M. E. *et al.* An epigenetic biomarker of aging for lifespan and healthspan. *Aging* **10**, 573–591 (2018).
- 265 17. Zhang, Q. *et al.* Improved precision of epigenetic clock estimates across tissues and its implication for biological ageing. *Genome Med* **11**,
266 54 (2019).
- 267 18. Belsky, D. W. *et al.* Quantification of the pace of biological aging in humans through a blood test, the DunedinPoAm DNA methylation
268 algorithm. *eLife* **9**, e54870 (2020).
- 269 19. Thompson, M. J. *et al.* A multi-tissue full lifespan epigenetic clock for mice. *Aging* **10**, 2832–2854 (2018).
- 270 20. Horvath, S. *et al.* Aging effects on DNA methylation modules in human brain and blood tissue. *Genome Biol* **13**, R97 (2012).
- 271 21. Wang, Y. *et al.* DNA methylation-based sex classifier to predict sex and identify sex chromosome aneuploidy. *BMC Genomics* **22**, 484
272 (2021).
- 273 22. Li, Q. *et al.* An objective metric of individual health and aging for population surveys. *Popul Health Metr* **20**, 11 (2022).
- 274 23. Moqri, M. *et al.* Biomarkers of aging for the identification and evaluation of longevity interventions. *Cell* **186**, 3758–3775 (2023).
- 275 24. Thrush, K. L., Higgins-Chen, A. T., Liu, Z. & Levine, M. E. R methylCIPHER: A Methylation Clock Investigational Package for
276 Hypothesis-Driven Evaluation & Research. 2022.07.13.499978 Preprint at <https://doi.org/10.1101/2022.07.13.499978> (2022).

25. Kwon, D. & Belsky, D. W. A toolkit for quantification of biological age from blood chemistry and organ function test data: BioAge. *GeroScience* **43**, 2795–2808 (2021).
26. Camillo, L. P. de L. pyaging: a Python-based compendium of GPU-optimized aging clocks. 2023.11.28.569069 Preprint at <https://doi.org/10.1101/2023.11.28.569069> (2023).
27. Yang, J. *et al.* Prevalence of comorbidities and its effects in patients infected with SARS-CoV-2: a systematic review and meta-analysis. *International Journal of Infectious Diseases* **94**, 91–95 (2020).
28. Lin, Q. *et al.* DNA methylation levels at individual age-associated CpG sites can be indicative for life expectancy. *Aging* **8**, 394–401 (2016).
29. Horvath, S. *et al.* Epigenetic clock for skin and blood cells applied to Hutchinson Gilford Progeria Syndrome and ex vivo studies. *Aging (Albany NY)* **10**, 1758–1775 (2018).
30. McEwen, L. M. *et al.* The PedBE clock accurately estimates DNA methylation age in pediatric buccal cells. *Proc. Natl. Acad. Sci. U.S.A.* **117**, 23329–23335 (2020).
31. McCartney, D. L. *et al.* Epigenetic prediction of complex traits and death. *Genome Biol* **19**, 136 (2018).
32. Lu, A. T. *et al.* DNA methylation-based estimator of telomere length. *Aging* **11**, 5895–5923 (2019).
33. Knight, A. K. *et al.* An epigenetic clock for gestational age at birth based on blood methylation data. *Genome Biol* **17**, 206 (2016).
34. Lee, Y. *et al.* Placental epigenetic clocks: estimating gestational age using placental DNA methylation levels. *Aging* **11**, 4238–4253 (2019).