

Ocean acidification alters the transcriptomic response in the nervous system of
Aplysia californica during reflex behaviour

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1 Abstract

2 Ocean acidification (OA) has numerous impacts on marine organisms including behaviour.

3 While behaviours are controlled in the neuro system, its complexity makes linking behavioural

4 impairments to environmental change difficult. Here we use a neurological model *Aplysia*

5 *californica* with well-studied simple neuro system and behaviours. By exposing *Aplysia* to

6 current day ($\sim 500 \mu\text{atm}$) or near-future CO₂ conditions ($\sim 1100 \mu\text{atm}$), we test the effect of OA

7 on their tail withdrawal reflex (TWR) and the underlying neuromolecular response of the

8 pleural-pedal ganglia, responsible for the behaviour. Under OA, *Aplysia* relax tails faster due

9 to increased sensorin-A expression, an inhibitor of mechanosensory neurons. We further

10 investigate how OA affects habituation, which produced a “sensitization-like” behaviour and

11 affected vesicle transport and stress response, revealing an influence of OA on neuronal and

12 behavioural outputs associated with learning. Finally, we test whether GABA-mediated

13 neurotransmission is involved in impaired TWR, but exposure to gabazine did not restore

14 normal behaviour and provoked little molecular response, rejecting the involvement in TWR

15 impairment. Instead, vesicular transport and cellular signalling link other neurotransmitter

16 processes directly with TWR impairment. Our study shows effects of OA on neurological tissue

17 parts that control for behaviour revealing the neurological mechanisms when faced with OA.

18 *Keywords: pH, sea hare, carbon dioxide, climate change, brain, neuromolecular, ganglia*

19 Introduction
20 Ocean acidification (OA) can have impacts on both calcifying and non-calcifying organisms'
21 survival and development (1,2), physiology (3,4) and behaviour (5) observed in several marine
22 taxa (6). Fish exhibit behavioural impairments amongst others in antipredator response (7–9)
23 and learning (10,11). However, invertebrates such as snails and crabs have also displayed
24 behavioural impairments caused by OA (12–14). Given that behaviour can significantly impact
25 population dynamics (15–17), it is crucial to understand the drivers and underlying
26 mechanisms to predict the trajectories of behavioural responses in a rapidly changing
27 environment (18,19).

28 The alteration of behaviour is associated with the brain or neuronal cells with a change
29 occurring to the molecular drivers (such as gene expression). One proposed cause of
30 behavioural impairments in a reduced pH environment is altered GABA neurotransmission
31 (20). GABA is a major inhibitory neurotransmitter found across the animal kingdom (21–23)
32 and GABA_A receptors are ion channels whose activation in normal conditions results in neuron
33 inhibition. When the partial pressure of CO₂ (pCO₂) is elevated in seawater, the acidosis
34 response of marine animals modifies internal ion gradients and the subsequent activation of
35 GABA_A receptors results in neuron excitation (20). This hypothesis was built on the fact that
36 animals reared in elevated CO₂ conditions had behavioural impairments at least partially
37 restored when treated with the GABA_A receptor antagonist gabazine (13,24,25). Changes in
38 the expression of genes involved in GABAergic neurotransmission in response to OA have also
39 been found (26–29), supporting that elevated pCO₂ acts in neuronal cells at the molecular
40 level, affecting pathways throughout the brain which in turn can produce impairments at the
41 whole organism level. Nevertheless, the complexity of the vertebrate brain brings difficulties
42 in understanding how molecular alterations inside neurons caused by environmental changes
43 such as OA modify the course of information transmission across synapses and throughout
44 regions of the nervous system, *in fine* driving behavioural changes.

45 One way of linking environmental influence with nervous system functioning and its
46 behavioural output is through the study of a simple and well-studied neuro system, such as
47 that of the California sea hare (*Aplysia californica*). Contrary to vertebrate brains, the nervous
48 system of *Aplysia* is composed of only 20,000 large neurons (30). Several of its behaviours
49 have been characterized in neural networks that link the nervous system and sensory organs

50 to muscles (31,32). As a result, it is possible to obtain the gene expression profile of specific
51 nerve ganglia which control for specific behaviours. In addition, *Aplysia* displays a similar
52 “acid–base regulator” profile to that of fishes during OA by accumulating bicarbonate (HCO_3^-)
53 ions (33). Since invertebrate GABA also binds to HCO_3^- permeable ionotropic GABA receptors
54 (34), the hypothesis of OA-altered GABA neurotransmission causing behavioural impairments
55 should be applicable in this model species. One suitable behaviour performed by *Aplysia* to
56 test this hypothesis is the Tail Withdrawal Reflex (TWR): during the TWR, the tail is withdrawn
57 for protection after a tactile stimulus is applied to it (35,36). It is a behaviour for which
58 controlling neurons in the pleural-pedal ganglia have been extensively described (35,37,38),
59 with neuron populations sensitive to several specific neurotransmitters such as glutamate
60 (39), dopamine, FMRFamide (40) and GABA (41). The TWR was also shown to be impaired by
61 OA itself (33). Finally, TWR can be involved in non-associative learning, notably reflex
62 habituation (42), which is likely caused by a presynaptic mechanism preventing transmitters
63 release (43).

64 We investigate the molecular processes affected by OA in the nervous system of the California
65 sea hare (*Aplysia californica*) as it performs a “simple” behaviour. We observed the TWR
66 response when animals were reared in either control ($\sim 500 \mu\text{atm}$) or predicted near-future
67 ($\sim 1100 \mu\text{atm}$) CO_2 conditions. This further allows us to investigate whether OA influences
68 learning experiences and the involvement of GABAergic neurotransmission in behavioural
69 impairments. Our three experiments focused on (1) how the innate TWR behaviour changes
70 with elevated CO_2 and the underlying mechanisms in the nervous system are, (2) whether
71 habituation of the TWR is influenced by elevated CO_2 and what corresponding
72 neuromolecular changes take place in the reflex’s circuitry and (3) the potential role of
73 GABAergic neurotransmission in the OA-induced behavioural changes of *Aplysia* (Fig. 1).
74 Together, these experiments pinpoint the molecular basis driving behavioural modifications
75 when faced with future ocean acidification.

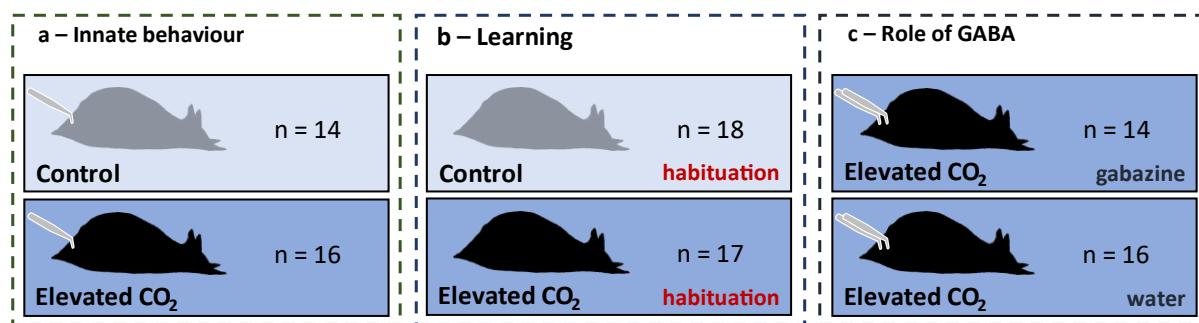


Figure 1: Experimental design for the study of Aplysia's Tail Withdrawal Reflex (TWR) response under elevated CO₂ conditions. The TWR is elicited by a tap on the tail with a needle. In the learning experiment (b), animals received a habituation training and TWR responses were observed pre- and post- training. In the GABA experiment (c), animals were either momentarily exposed to gabazine or to control water. The number of biological replicates is indicated by the n value for each group.

76 Methods

77 *Experimental design*

78 Adult sea hares weighing 52.8 ± 9.8 g were imported from the National Resource for Aplysia
79 (University of Miami, USA), and acclimated to their new environment for three days in
80 ambient control conditions. After this acclimation period they were exposed to either control
81 (pCO₂ ~ 450 μ atm; n = 36) or near-future CO₂ conditions (pCO₂ ~ 1200 μ atm; n = 54) for seven
82 to ten days at the University of Hong Kong. The elevated CO₂ level was chosen following the
83 RCP 8.5 scenario of the IPCC (44). Animals were housed in flow-through systems with natural
84 seawater from Hong Kong Island shores (Fig. S1). Exposure of animals to elevated CO₂ was
85 created through bubbling a mix of air and CO₂ gases at a rate of 15L/min, controlled by a
86 PEGAS 4000 MF Gas Mixer (Columbus Instruments). To reach the desired pCO₂, the mixing
87 parameters were set as follows: air flow was at 15 000 cc/min, CO₂ flow (vCO₂ in cc/min) was
88 calculated as a function of the CO₂ desired partial pressure (pCO₂ in %), the CO₂ ambient
89 partial pressure was at 0.04 % and the gas mix flow rate was 15L/min. The exposure to
90 elevated CO₂ was gradual and consisted in a first exposure to 700 μ atm for two days, then
91 1000 μ atm for four days, to then finally reach the target value of 1200 μ atm for seven to ten
92 days (depending on the experiment, see next section). The pH was measured daily in
93 randomly chosen tanks from both control and treatment groups using a WP-91 waterproof
94 pH meter (TPS) and a Seven2GO pH meter (Mettler Toledo) and the total alkalinity (TA) was
95 measured weekly using a G20S titrator (Mettler Toledo). Using the titrator's measurements,
96 pCO₂ values were calculated using the CO₂_{SYS} software (45). Following the recommendations

97 of the National Resource for *Aplysia*, temperature was set at 16°C using HC-1000A chillers
98 (Hailea) and measured daily in randomly chosen tanks from both control and treatment
99 groups using a WP-91 waterproof thermometer (TPS) and a Seven2GO thermometer (Mettler
100 Tolder). Salinity was monitored daily using a portable refractometer. Weekly measurements
101 of nitrate in randomly chosen tanks from both control and treatment groups were done using
102 a HI97728 nitrate photometer (Hanna Instruments) to ensure good water quality. The animals
103 were kept under a 12/12h light-dark cycle. *Aplysia* were fed *Agardhiella subulata* algae every
104 3 days as instructed by the National Resource for *Aplysia*.

105 Across all tanks, the salinity was at $32 \pm 1.6\text{‰}$ and the temperature was at $15.5 \pm 0.6^\circ\text{C}$ for
106 the whole experimental duration (Suppl. Table 1). The elevated pCO_2 conditions over the final
107 seven to ten days of experiment were $1100.3 \pm 59.9\text{ }\mu\text{atm}$, whereas the control pCO_2 was
108 $497.5 \pm 57.4\text{ }\mu\text{atm}$ (Fig. S2). pCO_2 was significantly different between treatment groups
109 (Wilcoxon rank sum test, p -value < 0.001). Additionally, there was no effect of the rearing
110 system identity on pCO_2 within the “treatment” group (ANOVA, p -value = 0.628).

111 *Behavioural assays*

112 All behavioural assays were performed by video recording using a Canon EOS M50 camera.
113 To test the innate reflex response to Tail Withdrawal Reflex (TWR) after elevated CO_2
114 exposure of seven days, animals reared either in control ($n = 14$) or elevated CO_2 conditions
115 ($\text{pCO}_2 \sim 1200\text{ }\mu\text{atm}$; $n = 16$) were acclimated for 30 min in the behavioural experimental tank
116 (Fig. 1a). The TWR was then elicited three times per animal, by pressing onto the tail with an
117 20G gauge needle and with each assay separated by ten minutes (Fig. 1a), as performed in
118 previous studies investigating the TWR (33,36).

119 To test if learning is impaired under OA conditions, *Aplysia* either reared in control ($n = 18$) or
120 elevated CO_2 conditions ($n = 17$) received a habituation training after a ten days exposure in
121 their rearing tanks (Fig. 1b). A period of 30 min for acclimation to the experimental setup was
122 given to individuals, and then the pre-training response was assessed three times the same
123 way as for the innate experiment, with a resting period of five min each. Training was then
124 provided by delivering 30 repeated tail stimuli, with 30s intervals between each. The post-
125 training TWR response was assessed 20, 25 and 30 min after training in the same way as pre-
126 training. This protocol is the same as that of another previous study investigating habituation
127 of the tail withdrawal reflex (46).

128 To test if GABA-ergic neurotransmission is involved in observed behavioural impairments,
129 *Aplysia* reared under near-future CO₂ conditions (pCO₂ ~ 1200 μ atm) for seven days were
130 either administered the GABA_A receptor antagonist gabazine (4 mg/L of SR-95531, Sigma; n =
131 14), or control water solution (n = 16; Fig. 1c). The administration route was dissolution
132 through the experimental tank water. The TWR behaviour was recorded after a 30 min
133 acclimation period similarly to the innate experiment.

134 The reflex response was measured from video recordings in terms of duration by an observer
135 blind to experimental conditions (Table S2). Measurements were discarded either if the tail
136 was not clearly visible after retraction, or if the TWR triggered inking, or if the reflex was a
137 Siphon Withdrawal Reflex. Mean durations were compared between control and treatment
138 groups for each experiment. All statistical were performed in R (R Core Team, 2018).

139 *RNA sequencing and gene expression analysis*

140 All individuals tested served for collection of RNA from the pleural-pedal ganglia, the nervous
141 tissue part known to be in control of the TWR's circuitry with sensory neurons located in the
142 pleural ganglia and motoneurons found in the pedal ganglia (31,47–49). The animals were
143 dissected immediately following their behavioural assay and for each animal the pleural-pedal
144 ganglia were snap frozen in liquid nitrogen and kept at -80°C until RNA extraction. The pedal-
145 pleural ganglia were then homogenized using sterile silicon beads for one minute at highest
146 speed in a Tissue Lyzer (Qiagen) and RNA was extracted following the TRIzol™ Plus RNA
147 Purification Kit (Thermo Fisher). The RNA concentration and quality of some samples was
148 measured using TapeStation (Agilent) to ensure good sample quality. Samples were
149 sequenced at 150bp paired end on an Illumina NovaSeq at the Centre for PanorOmic Sciences
150 (CPOS) of the University of Hong Kong. After sample sequencing, raw sequence data (on
151 average 38.5± 3.9M reads per sample; Table S3) were trimmed off adapters and filtered based
152 on read quality using Trimmomatic (50). Trimmomatic was ran using the following
153 parameters: “ILLUMINACLIP: all_PE.fa:2:30:10:8:TRUE LEADING:4 TRAILING:3
154 SLIDINGWINDOW:4:20 MINLEN:30”. Then, the filtered data (on average 36.5± 3.7M reads per
155 sample; Table S3) was mapped against the reference genome AplCal3.0 and its RefSeq
156 annotation available for *Aplysia* (51), using the program HISAT2 (52).

157 Differential expression (DE) analyses were led using DESeq2 v 1.38 (53) to investigate which
158 genes were differentially expressed between *Aplysia* reared either in control or elevated
159 pCO₂. A Likelihood Ratio Test (LRT) was used to ensure that neither the system nor the tank
160 identity were factors of influence on gene differential expression. Then, a Wald test was
161 performed to identify DE genes depending on the factor of interest. The design formula of
162 innate and learning experiments ("~ pCO₂") allowed pairwise comparisons of groups
163 depending on the two pCO₂ levels, whereas the design formula of the GABA experiment ("~
164 Gabazine") allowed pairwise comparisons between *Aplysia* either exposed to gabazine or
165 control water, all at elevated pCO₂. Differentially expressed genes with a baseMean under 10
166 and/or an absolute value of log2foldchange inferior to 0.3 were discarded to ensure that
167 differential expression was not an artifact of low counts, and to increase stringency.
168 Additionally, to identify which genes have their expression patterns possibly mediated by
169 either the TWR response, CO₂ and/or habituation, and could in turn play a role in the
170 behavioural change of *Aplysia*, a weighted gene co-expression network analysis (WGCNA) was
171 performed using the WGCNA v 1.72.1 package (54). Mean TWR duration of each individual
172 and final target pCO₂ were provided as trait data, as well as binary encoded information
173 regarding their habituation status ("0" = naïve, "1" = habituated) and gabazine exposure ("0"
174 = not exposed to gabazine; "1" = exposed to gabazine). The following parameters were used
175 to build the network: power = 7 (with R² > 0.90), TOMType = "signed", minModuleSize = 30,
176 reassignThreshold = 0, mergeCutHeight = 0.25, verbose = 3. Clusters of genes whose
177 expression patterns were correlated with either pCO₂ and/or habituation along with TWR
178 duration, or with TWR duration alone were identified. For significant differentially expressed
179 (DE) genes and for genes highlighted by the WGCNA analysis, functional enrichment analyses
180 were performed using OmicsBox v 1.4.11 (Fisher's Exact Test). The GO annotations used for
181 the enrichment analysis were retrieved from BioMart in OmicsBox, using the "Database of
182 genes from NCBI RefSeq genomes IDs". The Gene Ontology (GO) terms with an FDR adjusted
183 p-value below the 0.05 threshold were considered enriched among the DE genes, and the list
184 of GO terms was reduced to its most specific. Uncharacterized loci highlighted by the gene
185 expression analyses were searched in Rapid Ensembl release 109 (55) in the *Aplysia californica*
186 genome AplCal3.0 (assembly GCA_000002075.2) to predict their putative function.

187 Results

188 *Behavioural response to ocean acidification during TWR*

189 In the innate experiment, *Aplysia* exposed to elevated CO₂ conditions relaxed their tail after
190 withdrawal 29% faster with 12 ± 5 (mean \pm SD) seconds to relax, which was significantly
191 different from control condition which relaxed after 17 ± 6 seconds (Wilcoxon rank sum test,
192 p-value < 0.001; Figure 2a). In the habituation experiment, we saw the same significant effect
193 of CO₂ exposure on Tail Withdrawal Reflex (TWR) duration before habituation (Wilcoxon rank
194 sum test, p-value = 0.0216; Fig. S4), as *Aplysia* exposed to elevated pCO₂ before conditioning
195 took on average 12 ± 5 seconds to relax in elevated CO₂ but 15 ± 7 seconds in control
196 condition.

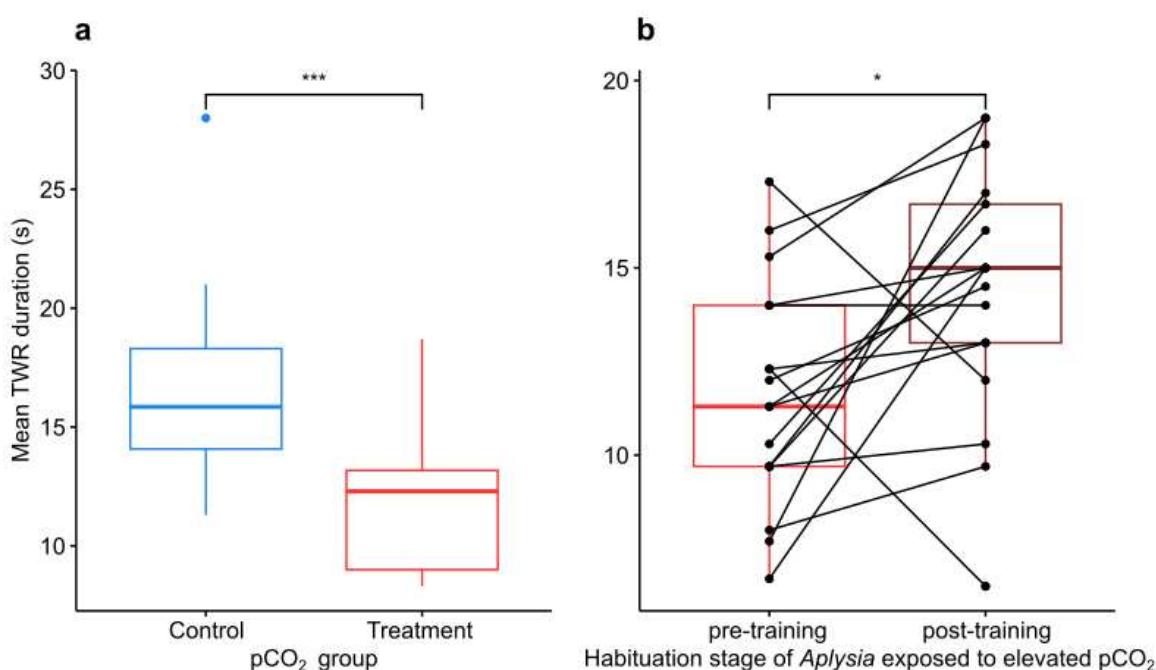


Figure 2: Tail Withdrawal Reflex (TWR) duration (s) of *Aplysia* in the innate experiment (a) as a function of their pCO₂ conditions (control ~500 μ atm or treatment ~1100 μ atm) and in the habituation experiment (b) reared at elevated pCO₂, before (pre-training) and after (post-training) habituation training; stars (*** or *) indicate the level of significant difference between the mean values

197 Habituation caused *Aplysia* to relax their tail in 14 ± 6 s for both control and elevated pCO₂
198 conditions. While it was not significantly different to pre-training for control individuals
199 (Wilcoxon rank sum test, p-value = 0.467; Fig. S5), habituation surprisingly increased TWR
200 duration for all but two individuals exposed to elevated CO₂ (p-value = 0.0238; Fig. 2b).
201 Elevated CO₂ caused the tail to relax approximately 17% slower after conditioning. Finally, in
202 the GABA experiment where all individuals were exposed to elevated CO₂, there was no

203 significant effect of gabazine on TWR duration (Wilcoxon rank sum test, p-value = 0.1857; Fig.
204 S3). Individuals who were administered control seawater took on average 12 ± 5 seconds to
205 relax from TWR whereas individuals who were administered gabazine relaxed after 14 ± 6
206 seconds.

207 *Molecular response to ocean acidification during TWR*

208 Acidification provoked a large transcriptomic response in the TWR-mediating parts of the
209 *Aplysia* nervous system, with a total of 1761 genes significantly differentially expressed
210 between naïve *Aplysia* exposed to different pCO₂ levels, while 725 genes were differentially
211 expressed among habituated *Aplysia*. Of these, 248 genes were commonly differentially
212 expressed due to acidification. This general response to ocean acidification affected calcium
213 ion (Ca²⁺) binding (Tables S4 & 5), through differential expression of receptors of Ca²⁺
214 important to nervous system functioning. Calcium transport exhibited downregulation of
215 calcyphosin-like gene possibly regulating ion transport and a trimeric intracellular cation
216 channel gene (*tric-1B.2*), which facilitates the active intracellular release of Ca²⁺ (Tables S6 &
217 7). Additionally, calmodulins, which are calcium-binding messengers involved in calcium
218 signal transduction and synaptic plasticity (Tables S6 & 7), and calcium-binding
219 protocadherins involved in cell-cell interactions were also differentially expressed (Tables S6
220 & 7). Related to calcium signalling, ocean acidification also provoked changes in excitatory
221 neurotransmission with the upregulation of neuronal acetylcholine (ACh) receptor subunits,
222 the Cerebral Peptide 2 precursor (CP2PP), the neuropeptide 15 receptor (*npr-15*), the
223 neuropeptide CCHamide-1 receptor and an ionotropic glutamate receptor (Fig. 4a, Table S5).
224 Furthermore, pCO₂ levels positively correlated with an ACh receptor subunit precursor (Table
225 S9). Another excitatory ionotropic glutamate receptor of the AMPA type was predicted for an
226 uncharacterized locus (*LOC101845288*) in the genome which had the highest expression level
227 among all transcripts yet was significantly downregulated due to acidification (Fig. 3b and 4a,
228 Tables S6 & 7) in both naïve and habituated *Aplysia*. Acidification specifically had an effect on
229 serotonergic neurotransmission in naïve *Aplysia* as the expression of two serotonin receptors
230 with excitatory activity on neurons (5-HT₂ and 5-HT₄) but also one inhibitory serotonin
231 receptor (5-HT_{1D}), was expressed at higher levels at elevated pCO₂ (Fig. 4b, Tables S5 & S9).
232 Another gene involved in inhibitory neurotransmission was the upregulation of BTB/POZ
233 domain-containing protein *KCTD12*, which is a GABA_B receptor subunit (Table S6). Finally,

234 acidification caused the upregulation of a calcium-binding inhibitory co-transmitter released
235 by mechanosensory neurons, the precursor mRNA of sensorin-A (*psc1*; Figure 3a & 4a; Table
236 S6).

237 Ocean acidification also affected components of the cytoskeletal network. Notably, the
238 expression of actin-related genes was significantly positively correlated with mean TWR
239 duration and negatively correlated with pCO₂ (Fig. S6; Table S8), including Actin related
240 protein (ARP) 2/3 complex subunits, ARPs, an F-actin-capping protein subunit, a myosin-10, a
241 FLII actin remodelling protein, a plastin-1 and one actin protein (Table S9). Furthermore,
242 genes involved in the actin-myosin interaction regulating cell morphology and cytoskeletal
243 organization were differentially expressed, such as myosin light chains coding genes (Tables
244 S6 & 7). Specifically in naïve *Aplysia*, an actin-interacting protein, ARP 2/3 complex subunits
245 and the PRKCA-binding protein (*pick1*) coding genes, which interact to regulate excitatory
246 synaptic plasticity, were downregulated (Table S6). Other genes associated with cytoskeletal
247 motion and intracellular transport were downregulated at-elevated pCO₂, such as genes
248 coding for tubulin chains as well as the centrin-3 and EF-hand domain-containing protein
249 coding genes which are involved in microtubule organization (Tables S6 & 7). Finally, genes
250 interacting with dynein also involved in cytoskeletal motion were consistently affected by
251 acidification, such as a parkin coregulated gene homolog, a dynein assembly factor, dynein
252 chains and dynein regulatory complex proteins (Tables S6 & 7).

253 Several energy related processes were affected by acidification. Firstly, ATP metabolism
254 through proton transmembrane transport, proton-transporting ATPase activity and
255 acyltransferase activity (Table S4) were altered in naïve *Aplysia* with the downregulation of
256 ATP synthase, ATPase, and ATP-citrate synthase and citrate synthase coding genes (Table S6).
257 Biosynthesis of nucleic acids, which is energetically expensive, was suppressed (Table S4) and
258 protein metabolism was affected by acidification through the downregulation of all genes
259 involved in alpha-amino acid synthesis (Tables S4, 6 & 7) tRNA ligases, initiation factor
260 subunits and elongation factors (Tables S6 & 7) in both naïve and habituated *Aplysia*. Protein
261 synthesis itself was altered through the differential expression of ribosomal protein coding
262 genes (Table S6) as well as the post-translational modification glycosylation due to
263 downregulation of protein glycosyltransferase subunits (Tables S6 & 7). Finally,

264 downregulation of protein disulfide-isomerase coding genes (Table S6) impacted unfolded
265 protein binding in naïve *Aplysia* exposed to elevated pCO₂ (Table S4).

266 Acidification triggered changes in the expression of cellular stress response genes. Heat shock
267 proteins 70 (B2 subfamily) were upregulated in *Aplysia* exposed to elevated pCO₂ (Tables S6
268 & 7) and genes with antioxidant activity (Table S4) were downregulated such as glutathione
269 S-transferases, a glutathione reductase, a carbonyl reductase [NADPH] 1 isoforms,
270 glutaredoxin and peroxiredoxins (Tables S6 & 7).

271 Habituation training revealed on effect of acidification on learning experiences in the nervous
272 system of *Aplysia*. After *Aplysia* received habituation training, exposure to elevated pCO₂ did
273 not alter cell signalling and organization functions as seen in untrained *Aplysia*. Interestingly,
274 the precursor mRNA of sensorin-A (*psc1*) was upregulated due to acidification in naïve
275 *Aplysia*, but habituated *Aplysia* had elevated levels of this gene no matter the pCO₂ (Figure 3a
276 & 4a; Table S6), suggesting that overexpression of *psc1* could also be achieved through
277 habituation training. Similarly, only in naïve *Aplysia*, acidification provoked changes in the
278 expression of genes involved in cell signalling processes mediated by GTP binding and GTPase
279 activity (Table S4), with downregulation of G-proteins involved in vesicular traffic such as ADP
280 ribosylation factor (ARF), ARF-binding protein (*GGA1*) and two ADP-ribosylation factor-like
281 (ARL) proteins (Table S6), which were not altered by OA in habituated *Aplysia*. Further
282 processes seen for naïve *Aplysia* with OA, but not in habituated *Aplysia* despite the elevated
283 CO₂ levels were intracellular transport, notably vesicle-mediated and protein transport
284 (Fig.4a, Table S4). Expression in genes coding for vacuolar protein sorting-associated proteins,
285 YIPF proteins, coatomer subunits and exocyst complex components, as well as a vesicle-
286 associated membrane protein (VAMP)/synaptobrevin-binding protein, a synaptobrevin and a
287 synaptobrevin homolog, downregulated for naïve *Aplysia*, are at control levels in habituated
288 *Aplysia* (Table S6). Finally, otoferlin, involved in the Ca²⁺-triggered synaptic vesicle-plasma
289 membrane fusion and in the control of neurotransmitter release at output synapses, was
290 upregulated only in naïve *Aplysia* but not in habituated OA *Aplysia* (Table S6). Further
291 evidence that habituation and pCO₂ affect cell signalling differently was found as co-
292 expressed genes positively correlated with pCO₂ but negatively correlated with habituation
293 training participated in GTPase mediated signalling (Table S8) such as four guanine nucleotide
294 exchange factor coding genes and two genes coding for dedicators of cytokinesis (Table S9).

295 Furthermore, genes of the thioredoxin antioxidant system and further antioxidant enzymes
296 (glutathione peroxidase and superoxide dismutase) were at control levels for habituated
297 *Aplysia* but downregulated in naïve *Aplysia* (Table S6) and these genes were also positively
298 correlated with mean TWR duration and habituation, but negatively correlated with pCO₂ (Fig.
299 S6; Table S9). Overall, genes involved in cell signalling and cellular stress response whose
300 expression were altered by acidification in naïve *Aplysia* were not differentially expressed in
301 habituated *Aplysia*, indicating a possible “cancelling/antagonistic” effect of habituation
302 training on gene expression under acidification.

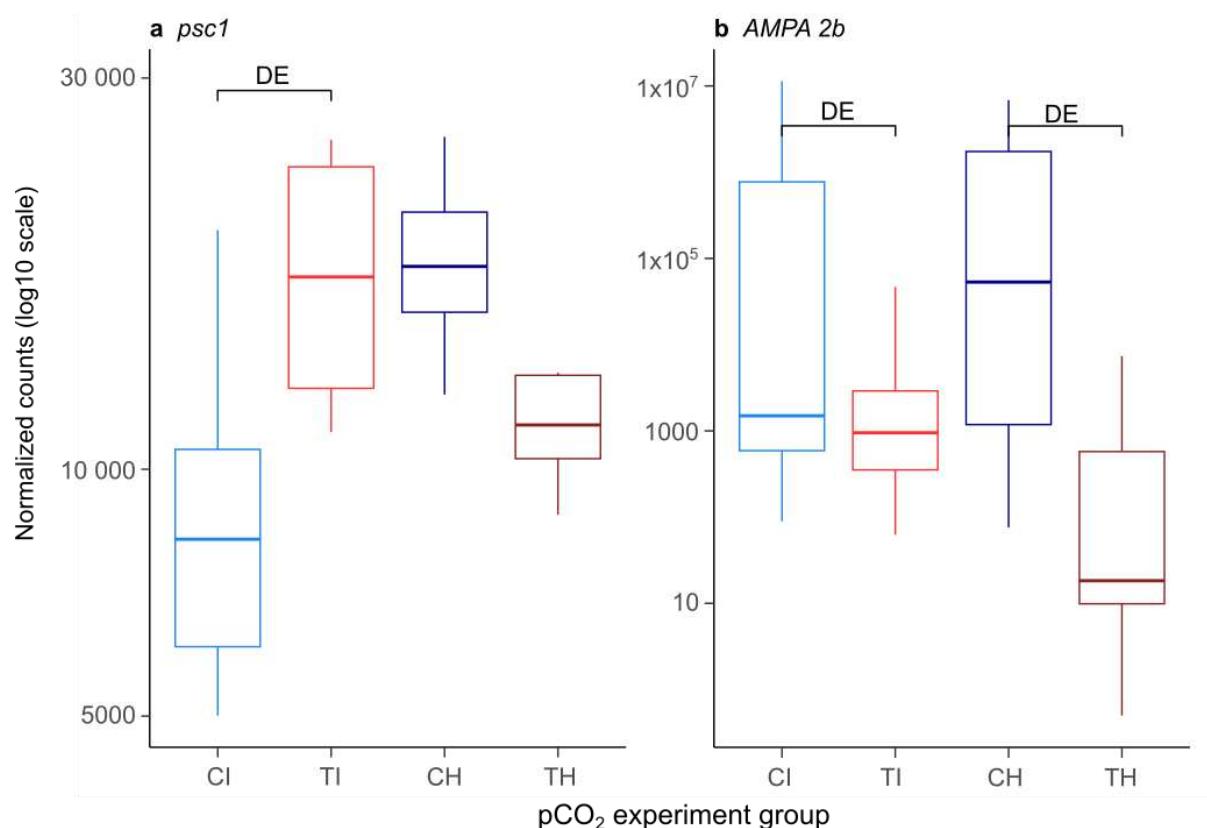


Figure 3: Normalized counts (\log_{10}) in all pCO_2 -experiment groups for the gene *psc1* (a) and the *AMPA 2b* predicted coding gene (b); *Aplysia* were grouped as a function of the pCO_2 they were reared at and the experiment they participated in: CI = Control ($\sim 500 \mu\text{atm}$) Innate, TI = Treatment ($\sim 1100 \mu\text{atm}$) Innate, CH = Control Habituated, TH = Treatment Habituated; “DE” shows which pairwise comparison revealed the gene as differentially expressed

303 Habituation training, however, also caused specific effects associated with acidification in the
304 nervous system of *Aplysia*. Signalling and vesicle transport were affected by the pCO_2 level,
305 but the genes involved in these functions were different after they experienced habituation
306 training. For instance, regucalcin, which is a suppressor protein of cell signalling, was
307 downregulated by acidification only in trained *Aplysia* (Table S7). Moreover, five genes
308 involved in cellular transport were upregulated with elevated pCO_2 only in habituated *Aplysia*,

309 notably two *vps-41* isoforms, one VAMP coding gene, one *VT11B* homolog and one gene
310 coding for an ARL protein 6-interacting protein (Fig. 4b, Table S7). GTP-mediated signal
311 transduction was also differently affected by pCO₂ after habituation, as the small GTPase *Rap*,
312 a Ras-related protein (*Rab-21*) involved in membrane trafficking, the *IQGAP1* gene regulating
313 the dynamics of the actin cytoskeleton and the Ras Guanine nucleotide exchange factor (GEF)
314 1B were all upregulated in trained *Aplysia* (Table S7). Furthermore, genes involved in
315 neuromodulation were downregulated at elevated pCO₂ only in habituated *Aplysia*, such as
316 two metabotropic glutamate receptors involved in the regulation of neurotransmitter release
317 notably by preventing cAMP production (Fig. 4b, Table S7). Further cAMP-responsive pathway
318 genes including CREB-regulated transcription coactivator and CREB3 regulatory factor were
319 upregulated specifically after habituation training (Table S7) and the CREB3 regulatory factor
320 was also positively correlated with both pCO₂ and habituation, alongside the cAMP response
321 element-binding protein (CREB) coding gene (Table S9). Both pCO₂ levels and habituation
322 training positively also correlated with the expression of serotonin and octopamine receptors
323 involved in neuromodulation (Fig. 4b, Table S9). Finally, genes involved in oxidoreduction and
324 cellular stress response were differentially expressed depending on pCO₂ only in habituated
325 *Aplysia* such as the cytochrome b, the cytochrome c oxidase and nitric oxide synthase coding
326 genes (Table S7). Therefore, habituation training triggers molecular changes specifically when
327 individuals are experiencing acidification, such as modified expression of genes performing
328 cell signalling, neuromodulation and oxidoreduction functions.

329 The potential role of GABA-ergic neurotransmission in the observed behavioural impairments
330 due to acidification was investigated by administering gabazine to *Aplysia* reared under near-
331 future CO₂ conditions. Only 20 genes were differentially expressed due to the administration
332 of gabazine in comparison to control water (Table S10). All except one (uncharacterized) were
333 upregulated in *Aplysia* exposed to gabazine. They coded for hydrolases, proteases, kinases, a
334 junction-mediating and regulatory protein (JMY), a cytochrome P450 2D26-like protein and a
335 betaine--homocysteine S-methyltransferase 1 (Table S10).

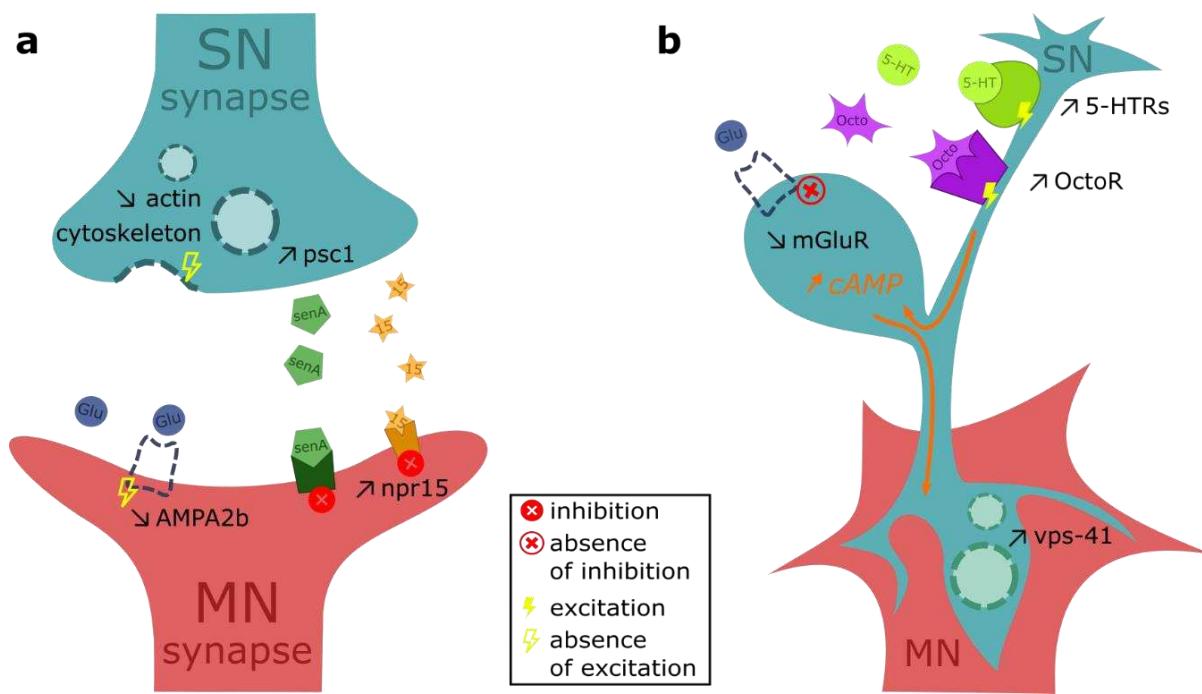


Figure 4: Molecular mechanisms in the TWR neural circuitry of *Aplysia* when exposed to elevated pCO_2 (a) during innate elicitation of the reflex and (b) after habituation of the reflex. SN = sensory neuron; MN = motoneuron; *npr15* = neuropeptide 15 receptor; *psc1* = pleural sensory cluster 1 gene, precursor of *senA* = sensorin A; Glu = glutamate; AMPA2b = AMPA 2b receptor gene; 5-HT = serotonin; 5-HTR = serotonin receptor; Octo = octopamine; OctoR = octopamine receptor; mGluR = metabotropic glutamate receptor; cAMP = cyclic adenosine monophosphate; *vps-41* = vacuolar protein sorting protein 41 gene

336 Discussion

337 Our study connected the behavioural impairment caused by OA in *Aplysia californica* with the
338 molecular changes occurring in the relevant part in the nervous system and thereby
339 pinpointing the underlying mechanisms of OA-driven behavioural alterations. Acidification
340 significantly decreased Tail Withdrawal Reflex (TWR) duration consistent with a previous
341 study (33) and provoked large changes in gene expression inside the TWR mediating parts of
342 the nervous system, the pleural-pedal ganglia. Several of those observed changes constitute
343 possible molecular mechanisms causing the behavioural impairment, such as the
344 upregulation of the peptide sensorin-A (*psc1*) coding gene. As *psc1* is an inhibitory transmitter
345 in *Aplysia* mechanosensory neurons (56), CO_2 -mediated increases in sensorin-A mRNA
346 expression can cause downstream neurons of the circuit to be inhibited, therefore not
347 transmitting the stimulus information to the end of the reflex circuitry which is responsible
348 for prolonged contraction. Hence, by leveraging the intricate knowledge of the TWR circuit in

349 the simple neuro system of *Aplysia* we are able to connect the behavioural impairment to
350 underlying mechanisms.

351 Further neuronal signalling inhibition may be at play, however, as we find a neuropeptide
352 receptor (*npr-15*) and an AMPA receptor changed expression levels when exposed to elevated
353 CO₂. These alterations could lead to an increase in inhibitory response, such as seen in an
354 octopaminergic manner for *C. elegans* in the case of *npr-15* (57) with increased inhibition of
355 neurons inside the reflex circuitry. Furthermore, reduced glutamatergic excitatory
356 transmission via downregulation of the AMPA receptor may lead to decreased neuron
357 excitability in *Aplysia* (36,58) resulting in less firing of motoneurons for prolonged foot muscle
358 contraction. Differential expression of receptors involved in neurotransmission can therefore
359 lead to the decreased TWR duration caused by acidification.

360 Apart from neurotransmission, cellular organisation and signalling are altered by elevated CO₂
361 exposure and could be involved in the decreased TWR duration, notably genes involved in
362 actin dynamics were correlated with TWR duration. The actin cytoskeleton regulates synaptic
363 areas morphology and synaptic vesicle pools (59,60) and it is sensitive to ocean acidification
364 in other molluscs (61,62). Synaptic vesicle mobilization in particular was found to be sensitive
365 to acidification in nerve terminals of vertebrate motoneurons (63), as an increase in
366 intracellular proton concentration suppresses synaptic vesicle delivery. In the case of *Aplysia*
367 and fishes, even though acid-base regulation occurs following exposure to elevated CO₂,
368 maintaining acid-base balance is achieved through retention and/or uptake of HCO₃⁻ to
369 maintain the internal pH despite elevated proton concentrations (33). Since internal proton
370 concentration is maintained at a high level, synaptic vesicle delivery could be suppressed
371 through the observed reduced expression of actin-related genes involved in the mobilisation
372 of synaptic vesicles. As a result, this reduced vesicle activity would *in fine* inhibit
373 neurotransmission and decrease the reflex response and duration. Furthermore, other
374 cytoskeletal motors, such as tubulin, were also downregulated by elevated pCO₂. This same
375 pattern has been found in tubeworm and oyster larvae (64,65). Evidence of changes in
376 cytoskeleton components during exposure to elevated pCO₂ suggests that acidification not
377 only interferes with normal vesicle functioning at the synapse, causing behavioural
378 impairment, but might also alter numerous other functions performed by the cytoskeleton.
379 For example, in blood clams changes in the cytoskeleton due to acidification may have

380 implications on the immune response because cytoskeletal components perform
381 phagocytosis (66). Hence, cytoskeletal changes may reduce synaptic mobilization which is a
382 further mechanism for TWR changes under acidification conditions, but modified
383 cytoskeleton organization could also cause other downstream effects at the whole organism
384 level.

385 The decreased synaptic activity among neurons of the reflex circuitry with acidification may
386 have led to compensatory mechanisms through differential expression of genes involved in
387 neurotransmission. One potential compensation is the upregulation of acetylcholine
388 receptors in *Aplysia* pedal-pleural ganglia, also observed in pteropod nervous systems under
389 ocean acidification (67,68). The upregulation of neuronal acetylcholine could increase Ca^{2+}
390 levels and intensify calcium-mediated exocytosis of synaptic vesicles, which can be an
391 alternative way of restoring neurotransmitter delivery impaired by the downregulation of
392 genes involved in synaptic mobilization in *Aplysia*. Additionally, with otoferlin placing synaptic
393 vesicles near calcium channels to allow fast exocytosis of neurotransmitters (69), the
394 observed upregulation of otoferlin in *Aplysia* at elevated pCO_2 could also facilitate Ca^{2+} -
395 mediated release of synaptic vesicles from the sensory neurons and restore a flow of
396 neurotransmitters in the synaptic area. Furthermore, the upregulation of genes involved in
397 the recycling of synaptic vesicles could be a compensatory mechanism in *Aplysia* when faced
398 with acidification. Maintenance of neurotransmission by recycling existing receptors could
399 counteract decreased gene expression of such receptors. For example, the upregulation of
400 the huntingtin-interacting protein and AP180 coding genes in *Aplysia* neurons at elevated
401 pCO_2 could maintain neurotransmission by enhancing the recycling of already existing AMPA
402 receptors (70,71), despite the downregulation of the AMPA receptor coding gene that would
403 limit its neobiosynthesis. Finally, increased production of neuropeptides, such as those
404 derived from the Cerebral Peptide 2 precursor (CP2PP) could also be a way to compensate for
405 impaired neurotransmission. CP2PP is a precursor to ten bioactive neuropeptides in *Aplysia*,
406 among which some act on the foot muscle (72,73), therefore its upregulation could lead to
407 increased production of neuropeptides which could activate tail motoneurons. Overall,
408 *Aplysia* shows signs of transcriptional reprogramming to compensate for reduced
409 neurotransmission when exposed to acidification in the nervous system.

410 The molecular response to acidification in the pedal-pleural ganglia also comprised several
411 important functions such as ATP, nucleic acid and protein metabolism. Downregulation of
412 metabolism genes in *Aplysia* could be a sign of metabolic suppression as commonly seen in
413 other invertebrates when exposed to elevated pCO₂ (74–77), implying that downregulation
414 of oxidative metabolism genes in sensitive organisms might compromise the cellular stress
415 response (78). However, in *Aplysia*, the downregulation of oxidative metabolism genes under
416 OA is not sufficient to imply that the nervous system may be more susceptible to cellular
417 stress, because the upregulation of heat shock protein genes 70 was also observed. This type
418 of heat shock protein is expressed when cellular protection is required, notably but not
419 restricted to oxidative stress conditions (79). Their upregulation in *Aplysia* to protect cells
420 against OA-mediated stress has also been documented in Sydney rock oysters (80). Hence,
421 instead of metabolic suppression, the metabolic downregulation may rather point to
422 reallocation of energy towards homeostasis. This has been suggested as a cellular strategy to
423 redirect the energy in the most effective way possible towards immediate essential processes
424 at the expense of other functions (77). *Aplysia* exposed to 1200 µatm of pCO₂ are able to
425 maintain their haemolymph pH_e levels similar to that of control by accumulating HCO₃[−] (33).
426 It is therefore possible that the downregulation of genes involved in expensive processes is a
427 means to ensure that sufficient resources are allocated to the uptake of HCO₃[−] inside the
428 haemolymph to maintain acid-base homeostasis at the whole organism level.

429 When *Aplysia* were trained under OA, contrary to our expectations, the TWR duration
430 increased. Increased duration is consistent with short-term sensitization rather than
431 habituation (42). The cellular mechanism underlying short-term sensitization training in
432 *Aplysia* consist of the activation of serotonergic neurons, which act on the sensory neurons
433 of the reflex circuitry by increasing cAMP intracellular levels (81,82). Hence, increased
434 expression of serotonin and octopamine receptors in the neural circuitry at elevated CO₂
435 following habituation could cause a sensitization and produce the increased TWR duration.
436 Furthermore, sensitization-like responses may also be caused by the alteration of
437 metabotropic glutamate receptors. A downregulation of *MGR2* activation, as exhibited in
438 habituated *Aplysia* with OA, could increase production of cAMP in the presynaptic area
439 following habituation training and produce changes in the nervous system consistent with
440 sensitization (83). Trained *Aplysia* exposed to elevated pCO₂ resulted in specific molecular

441 changes related to the habituation process, for example of *vps-41* coding gene expression.
442 Expression of circular mRNA of *vps-41* improves synaptic plasticity and learning functions by
443 acting as a “sponge” delivering miRNAs to regulate the transcriptome (84,85). Its upregulation
444 under OA and after habituation training could therefore facilitate synaptic changes in the
445 reflex circuitry. Upregulation of *vps-41* could strengthen synaptic connections between
446 sensory neurons and motoneurons through decreased presynaptic transmitter release (86),
447 leading to a sensitization-like behavioural response after elicitation of the reflex. Overall,
448 habituation training while experiencing OA provokes molecular changes that could act on the
449 nervous system in a similar way to that of a sensitization experience in normal conditions,
450 hence modulating the behaviour differently under the influence of changes pH conditions.

451 *Aplysia* exposed to elevated CO₂ and administered gabazine did not show restoration of TWR
452 duration consistent with that of control individuals and few changes were observed on the
453 molecular level. Previous studies in molluscs have demonstrated that gabazine could restore
454 impaired behaviours such as burrowing and predator escape (13,87), therefore it is unlikely
455 that the absence of behavioural change is due to the fact that gabazine would not act as a
456 correct antagonist of GABA_A receptors. A possible explanation would be that contrary to other
457 behaviours previously investigated in the context of acidification (13,87), the TWR
458 impairment under OA is not caused by interfering with GABA_A neurotransmission. This was
459 also seen for another invertebrate which altered behaviour in acidified conditions was not
460 restored with gabazine administration (88). Instead, it is possible that other ligand-gated ion
461 channels (LGICs) performing neurotransmission could have their function altered by the acid-
462 base regulation response in *Aplysia*, such as glutamate or acetylcholine-gated chloride
463 channels which are affected by OA in *Aplysia* (89). Hence, our results suggest that different
464 mechanisms than changes GABAergic neurotransmission may be responsible for TWR
465 impairment.

466 Overall, our study reveals links between behavioural alterations and molecular changes
467 occurring in the model neurosystem of *Aplysia* as it is experiencing ocean acidification (OA).
468 Our findings show that the TWR and its modulation through learning is impaired during OA
469 through alterations in neurotransmission between mechanosensory neurons and neurons
470 downstream of the reflex circuitry. Gabazine did not restore TWR duration revealing GABA-
471 ergic neurotransmission not to be the main mechanism responsible for impaired behaviour.

472 By leveraging the simple nervous system of *Aplysia*, we show direct links between near-future
473 predicted ocean conditions and the molecular processes occurring in neurological systems
474 that control for animal behaviour.

475 Data availability

476 The raw sequencing data can be found in BioProject PRJNA1031344. The reviewer link to the
477 data is:

478 <https://dataview.ncbi.nlm.nih.gov/object/PRJNA1031344?reviewer=hf09nmkmqj8ebd5b7fa6cu6gb>

480 Author contributions

481 JMS conceived and carried out the experiments with input from CS. JMS analysed the data
482 under the supervision of CS. JMS wrote the paper with input from CS.

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