Drivers of dispersal and diversification in bromeliads

- 3 Igor M. Kessous*^{1,2,3,4}; Harith Farooq^{1,2,5}; Weston Testo^{1,2,6}; María Fernanda T. Jiménez^{2,7};
- 4 Beatriz Neves^{1,2,3}; Alessandra R. Pinto^{4,8}; Fabiano Salgueiro⁹; Andrea F. Costa³; Christine D.
- 5 Bacon^{1,2}

1

2

6

- 7 ORCID IDs
- 8 Igor M. Kessous (Orcid ID: 0000-0001-6280-4424)
- 9 Harith Farooq (Orcid ID: 0000-0001-9031-2785)
- 10 Weston Testo (Orcid ID: 0000-0003-3194-5763)
- 11 María Fernanda T. Jiménez (Orcid ID: 0000-0002-7177-4164)
- 12 Beatriz Neves (Orcid ID: 0000-0002-6469-7930)
- 13 Alessandra R. Pinto (Orcid ID: 0000-0003-4633-4930)
- 14 Fabiano Salgueiro (Orcid ID: 0000-0002-0352-0699)
- 15 Andrea F. Costa (Orcid ID: 0000-0002-9200-4222)
- 16 Christine D. Bacon (Orcid ID: 0000-0003-2341-2705)
- ¹Department of Biological and Environmental Sciences, University of Gothenburg, Carl
- 19 Skottsbergs Gata 22B, SE 41319, Gothenburg, Sweden;
- ²Gothenburg Global Biodiversity Centre, Carl Skottsbergs Gata 22B, SE 41319, Gothenburg,
- 21 Sweden;

- 22 ³Departamento de Botânica, Museu Nacional, Universidade Federal do Rio de Janeiro,
- 23 Quinta da Boa Vista s.n., São Cristóvão, 20949-040, Rio de Janeiro, Brazil;
- ⁴Instituto de Pesquisas Jardim Botânico do Rio de Janeiro, Rua Pacheco Leão, 915, 22460-
- 25 030, Rio de Janeiro, Brazil;
- ⁵ Faculty of Natural Sciences, Lúrio University, Cabo Delgado, Mozambique;
- 27 ⁶ Botanical Research Institute of Texas, 1700 University Drive, Fort Worth, TX 76107-3400,
- 28 USA;
- ⁷ Institute of Biosciences, Zoology department, Vilnius University, Saulėtekio al. 7, Vilnius,
- 30 Lithuania;
- 31 ⁸Programa de Pós-Graduação em Ecologia, Universidade Federal do Rio de Janeiro, Av.
- 32 Carlos Chagas Filho, 373, 21941-971, Rio de Janeiro, Brazil;
- 33 Departamento de Botânica, Universidade Federal do Estado do Rio de Janeiro, Av. Pasteur
- 34 458, 22290-255, Rio de Janeiro, Brazil

- 35 **Corresponding author:*
- 36 Igor M. Kessous

- 37 Instituto de Pesquisas Jardim Botânico do Rio de Janeiro, Rua Pacheco Leão, 915, 22460-
- 38 030, Rio de Janeiro, Brazil.
- 39 Email: <u>igor.kessous@gmail.com</u>
- 40 Phone: +5521977939779.

Summary

- Dispersal strategies strongly influence an array of plant traits, especially the shape and function of fruits and seeds, and can be important drivers of diversification dynamics. In this study we investigated how fruit morphology and habitat influence dispersal capacity and diversification rate in bromeliads. We hypothesize that (1) the evolution of berry fruits increased dispersal capacity and diversification rates; and (2) climatic factors contribute to increased dispersal capacity and diversification rates.
- To understand the influence of fruit and habitat traits on evolutionary dynamics, we generated a time-calibrated phylogeny including 1,268 species of bromeliads and integrated that evolutionary framework with distribution, habitat, and morphological trait data.
- We find that lineages with berry fruits have the highest rates of diversification. We
 also identify significant correlation between diversification rates and both elevation
 and forest canopy height. We demonstrate that dispersal capacity is not related to fruit
 morphology and covaries with forest canopy height and mean annual temperature.
- We show that factors influencing the dispersal capacity and diversification are
 heterogeneous among the subfamilies. These new insights into the rise and spread of
 bromeliads emphasize the importance of considering the plurality of morphological
 and ecological features to improve the understanding of the evolutionary dynamics.
- 61 Keywords: berry, canopy height, epiphytes, fruit dispersal, monocots, Neotropics,
- 62 phylogenetic comparative methods

63 Introduction 64 Dispersal capacity mediates species occupancy and drives biodiversity patterns through its 65 influence on reproductive isolation and speciation (Antonelli & Sanmartín, 2011; Smith et al., 66 2014; Steinbauer et al., 2016). In plants, dispersal-related traits such as adaptations in the 67 shape of fruits and seeds are closely associated with dispersal mechanisms associated with 68 wind, animals, amongst others (Seale & Nakayama, 2020). In addition to morphological 69 adaptions that facilitate dispersal, different climatic conditions may also favor different 70 dispersal strategies and thus, the evolution of different fruit and seed morphologies (Seale & 71 Nakayama, 2020). 72 73 There are higher levels of biotic interactions (Schemske et al., 2009; Antonelli & Sanmartín, 74 2011; Brown, 2014; Andresen et al., 2018) and higher dispersal limitations (Janzen, 1967) in 75 tropical compared to temperate regions. Both these patterns drive the current latitudinal 76 diversity gradient (e.g. Mittelbach et al., 2007). Tropical rainforests are characterized by 77 closed canopies and high temperatures and precipitation levels (Eiserhardt et al., 2017). 78 Species richness generally peaks at low latitudes, where forests present the highest canopies 79 (Zhang et al., 2015). Particularly for arboreal communities, such as epiphytes, higher 80 canopies favor the presence of higher niche variation, because of the vertical stratification 81 and microclimatic variation (de la Rosa-Manzano et al., 2014; Oliveira & Scheffers, 2019). In 82 addition to latitude and canopy height, elevation also has an effect on dispersal and 83 diversification, where high elevations have increased speciation rates (Madriñán et al., 2013; 84 Lagomarsino et al., 2016; Testo et al., 2019). 85 86 Tropical America (the Neotropics) harbors unparalleled species richness and includes several 87 biodiversity hotspots, with high levels of endemism in both animals and plants (Myers et al., 88 2000). Previous work has indicated that a unique combination of abiotic and biotic processes 89 interacting over millions of years has led to a unique biodiversity and the high rates of species 90 richness and endemism in the region (Antonelli & Sanmartín, 2011). Endemism is most 91 common at the species level, but there are 52 plant families that are endemic (or nearly so) to 92 the Americas, almost all of which are species-poor (fewer than 100 recognized species; Ulloa 93 Ulloa et al., 2017; Givnish, 2017). A remarkable exception is Bromeliaceae (bromeliads), a 94 hyperdiverse family of monocots that include air plants and pineapples. All but one of the ca. 95 3,700 recognized species of bromeliads are endemic to the "New World", most of them

96 restricted to the Neotropics (Smith & Downs, 1974; Ulloa Ulloa et al., 2017; Givnish, 2017; 97 Gouda et al., 2020 [cont. updated]). Bromeliads are ecologically diverse and occur from sea 98 level to ca. 4,000 m in elevation. This ecological diversity is linked to morphological, 99 ecological, and physiological adaptations, including a tank habit, epiphytism, Crassulacean 100 Acid Metabolism (CAM) photosynthesis, and a myriad of biotic interactions (Smith & 101 Downs, 1974; Benzing, 2000; Givnish et al., 2014). The uniqueness in the distribution, 102 ecology, and endemism of bromeliads provide an excellent opportunity to understand the 103 relationship between dispersal-related traits, diversity patterns, and spatial distribution in the 104 Neotropics. 105 106 There are two means of dispersal in bromeliads: (1) abiotic, present in capsule-fruited species 107 (with plumose or winged seeds and its varieties) dispersed by wind or gravity (Benzing, 108 2000; Smith & Downs, 1974) and (2) biotic, in berry-fruited species (Givnish et al., 2011, 109 2014; Silva et al., 2020; Fig. 1) with "naked" seeds, often with mucilaginous and sticky 110 appendages, dispersed by vertebrates or insects (Smith & Till, 1998; Siva et al., 2020; Leme 111 et al., 2021). Some berry-fruited plants have higher diversification rates than other fruit types 112 (Lagomarsino et al., 2016), since they are able to disperse at longer distances, which may 113 increase the opportunities for lineage diversification. In bromeliads, the relationship between 114 dispersal and morphological or habitat traits remains poorly known, and further, has not been 115 correlated with diversification. 116 117 In this study, we identify how fruit morphology and habitat influence dispersal capacity and 118 diversification rate. To do so, we compiled DNA sequence data for 1,268 species of 119 bromeliads, imputed unsampled taxa to produce a comprehensive species-level phylogeny, 120 assembled morphological, distributional, and ecological data sets, and conducted comparative 121 phylogenetic analyses. Specifically, we hypothesize that (1) the evolution of berry fruits 122 increased dispersal capacity and diversification rates. We also hypothesize that (2) climatic 123 factors inherent in certain habitat types, such as precipitation, canopy height, temperature and 124 elevation, contribute to increased dispersal capacity and diversification rates. Determining the 125 dynamics of dispersal and diversification is fundamental to understanding the economically-126 important bromeliads as well as the Neotropical environments where they are distributed. 127

Materials and Methods

129 We used the Encyclopaedia of Bromeliads (Gouda et al., 2020 [cont. updated], 130 https://bromeliad.nl/encyclopedia/) to standardize the taxonomic classification used in the 131 molecular and spatial datasets (see Appendix S1). 132 133 *Taxon and sequence sampling* 134 We obtained sequence data for 13 chloroplast (agt1, ycf1, rps16-intron, rps16-trnK, rpl32, 135 matK, nadH, petD, trnL-trnF, rpoB, atpB-trnC, psbA-trnH, and trnC-petN) and three nuclear 136 (PHYC, PRK, and LEAFY) loci from Genbank for a total of 1,268 species of Bromeliaceae 137 (ca. 30% of species in the family) and five outgroups from Typhaceae and Rapateaceae (see 138 Appendix S2). We aligned each locus with MAFFT (Katoh & Toh, 2008) using default 139 parameters. We removed poorly aligned and divergent regions using Gblocks (Castresana, 140 2000), allowing gap positions in the final blocks and less strict flanking positions. Both 141 alignment analyses were performed in R v. 3.5.3 (R Core Team, 2019), using the package ips 142 0.0.11 (Heibl, 2019). We concatenated the alignments using Geneious Prime (version 143 2021.0.1, Biomatters, New Zealand) and Mesquite v. 3.61 (Maddison & Maddison, 2019; see 144 Appendix S3) under the criterion of mean pairwise identity over all pairs in the column of at 145 least 30%. 146 147 Phylogenetic analysis and molecular dating 148 To perform our dispersal and diversification analyses we used two different trees: (1) the 149 backbone tree (BB), output from the phylogenetic analysis including 1,268 species, and; (2) 150 the species level tree (SL) including the BB terminals and the remaining species of the 151 family. To converge multiple chains of the BB tree, we first generated an XML file with 152 BEAUTi v. 1.10.4 (Suchard et al., 2018) to run an exploratory analysis to identify the 153 parameters with disproportionately high effective sample sizes (ESS) and reduce their 154 operator weights in subsequent analyses. We then identified the parameters with 155 disproportionately high effective sample sizes (ESS) compared to others and reduced their 156 operator weights, under the assumption that the data was informative enough for their 157 estimation, which allowed us to speed up data analysis. We did not have calibrations in the 158 tree, however the large number of terminals required high performance computing resources. 159 Thus, to facilitate Markov chain Monte Carlo (MCMC) convergence, model complexity was 160 reduced by selecting the HKY and strict molecular clock (Yule, 1925; Gernhard, 2008). After 161 adjusting operator weights, we used BEAST v. 1.10.4 (Suchard et al., 2018) to run three 162 rounds of two independent chains, each with 300 million MCMC generations sampled every

163 30,000 generations (see Appendix S4). After every round, we selected the last tree of the 164 analysis with the best ESS values and used it as the starting tree of a subsequent round in 165 order to reach stationarity in both chains. To obtain the maximum clade credibility (MCC) 166 tree, we applied a 15% burn-in to the resulting tree distribution such that the remaining ESS 167 values were > 600. After all rounds, we reached 549,060,000 sampled generations. We 168 performed all phylogenetic analyses at the Swedish National Infrastructure for Computing 169 (https://www.snic.se/) and in the CIPRES Science Gateway V. 3.3 (https://www.phylo.org/). 170 171 After estimating the BB tree, we inferred a SL tree of Bromeliaceae by imputing missing taxa 172 using the R package V. Phylomaker 0.1.0 (Jin & Qian, 2019), after matching the taxa in the 173 BB tree with the species list in Gouda et al. (cont. updt.). We imputed unsampled species 174 only at the genus level. In this case, we used scenario 3 (S3) where new tips of an existing 175 genus were bound in the basal node of this genus (see Qian & Jin, 2016; Jin & Qian, 2019). 176 For the imputation, we used the build.nodes.2 function that uses backbone node information 177 based on our species list and the bind.relative function to include the taxa. 178 179 Both the BB and SL trees were dated a posteriori using penalized likelihood performed in 180 treePL (Smith & O'Meara, 2012). Because of the absence of reliable fossils of Bromeliaceae 181 (Kessous et al., 2021), we assigned age constraints based on secondary calibrations from 182 Givnish et al. (2018), with minimum and maximum age bounds set at 20% younger and older 183 than the median ages reported for the stem (96 -) 120 (-144) Ma and crown (16 -) 20 (-24)184 Ma of the family. We ran this analysis 100 times, used TreeAnnotator (Bouckaert et al., 185 2014) to generate a consensus tree and calculate the 95% highest posterior density (HPD) for 186 each node age. 187 188 Spatial data and dispersal capacity 189 We downloaded 121,978 records of bromeliads from GBIF (www.gbif.org; 03 February 190 2021): https://doi.org/10.15468/dl.ny8dnt). To remove duplicates and erroneous occurrence 191 records, we performed two sequential analyses with the R package CoordinateCleaner 2.0.18 192 (Zizka et al., 2019), flagging "capitals", "centroids", "equal", "institutions", "outliers" and 193 "zeros". We removed species with a single occurrence point, rounded all coordinates to two 194 decimals, and deleted duplicates. The final dataset consisted of 99,863 records of 2,720 195 species (75% of known species).

We derived a proxy for dispersal capacity using the mean distance between all pairwise occurrence combinations for each species with the function earth.dist of the R package fossil 0.4.0 (Vavrek & Vavrek, 2020), which returned a list in kilometers. Our proxy alleviates issues inherent in other metrics, such as range size and maximum distance between occurrences, which result in problematic inferences for widespread species or those with disjunct ranges. Collection bias may influence the dispersal metrics, since the distance between individuals may be affected by the number of points of each species. We opted to use a different proxy because in convex hull restricted species can be more sensitive to errors (Burgman & Fox, 2003). We also observed widespread species inferred with convex hull with overestimated areas. Biotic and abiotic traits Based on the literature and herbarium specimens, we scored each species for fruit type (according to Smith & Downs, 1974; Benzing, 2000; Givnish et al., 2011; Gouda et al., 2020 [cont. updated]): (1) berry and (2) capsule. Because of the heterogeneity of seeds in bromeliads, as in the case of "naked" seeds belonging to the subfamily Bromelioideae and Navioideae that have appendages with distinct anatomical origins (Silva et al., 2020, Leme et al., 2021), the great variation in winged seeds (Smith & Downs, 1974; Benzing, 2000), and non-homology of appendages in some plumose seeds (Palací et al., 2004), we only used the fruit morphology. We transformed the fruit type trait into binary presence-absence matrices for the regression analyses. In addition, we scored four continuous variables concerning habitat: canopy height, elevation, mean annual precipitation, and mean annual temperature. We downloaded a canopy height raster with 1 km resolution (https://landscape.jpl.nasa.gov/; accessed on 22 March 2021) and an elevation raster with 30 m resolution (https://www.usgs.gov/centers/eros/science/usgs-eros-archive-digital-elevation-shuttle-radartopography-mission-srtm?qt-science_center_objects=0#qt-science_center_objects; accessed on 22 March 2021). We downloaded the mean annual precipitation and mean annual temperature variables from CHELSA v1.2 (Karger & Zimmermann, 2019; https://envicloud.os.zhdk.cloud.switch.ch/chelsa/chelsa V1/climatologies/bio/; accessed on

196

197

198

199

200

201

202

203

204

205

206

207208

209

210

211

212

213

214

215

216

217

218

219

220

221

222

223

224

225

226

22 March 2021). We extracted the values of each geographic occurrence point using the R package raster 3.4.13 (Hijmans et al., 2015), and calculated the mean for each species. Phylogenetic regression Using the SL tree, we determined the effect of traits on the dispersal capacity of each species by performing phylogenetic regressions using the pgls function of the R package caper 1.0.1 (Orme et al., 2013). For the phylogenetic regression, we log-transformed and scaled all variables to mean=0 and standard deviation=1. We estimated Pagel's lambda (λ) using the phylogenetic generalized linear square (PGLS) test, which varies from 0 to 1 (towards the Brownian motion expectation). We determined the most likely model to describe our data using the likelihood ratio test implemented through the function *lrtest* in the R package *lmtest* 0.9.38 (Hothorn et al., 2015). We tested all possible combinations of traits and used the Akaike information criterion (AIC) to select the best-fit model. Finally, we performed an ANOVA test with the function *anova.pgls* in *caper* to investigate the effect under each trait. We tested model assumptions for all models by visual inspection of the residuals after applying the *qqnorm* function. Diversification analyses We used state-dependent diversification models to identify the relationship of species traits and rates of diversification in the SL tree. We used BiSSE (binary state speciation and extinction; Maddison et al., 2007) to analyze the fruit type. We conducted all analyses in the R package diversitree 0.9.15 (FitzJohn, 2012) and evaluated six different models separately, from the simplest (constraining all parameters) to the most complex (allowing all parameters, in which all rates of speciation depend on the character state). We used the AIC to select the best-fit model, through an ANOVA, and then used the function make.prior.exponential to set exponential priors (FitzJohn, 2012). We ran a preliminary MCMC chain of 100 steps to set the control parameter (w) and sequentially subsampled it in the 0.05-0.95 interval. We then ran the final MCMC chain of 10,000 steps, sampling every 10 steps, applying a burn-in of 5% and calculated the net diversification rate for the fruit type trait. The reliability of SSE methods to account for diversification has been widely discussed in the literature (Davis et al., 2013; Rabosky & Goldberg, 2015; Maddison & FitzJohn, 2015; Herrera-Alsina et al., 2019). Because state-dependent analysis is sensitive to large differences

228

229

230231

232

233

234

235

236

237

238

239

240

241

242

243

244245

246

247

248

249

250

251

252

253

254

255

256

257258

259

261 between tip ratios and small-sized trees (Davis et al., 2013; Rabosky & Goldberg, 2015), we 262 used the fully sampled tree (SL) and the fully coded traits. It has been suggested that the state 263 tip ratio of 3:1 has high rates of power in large sampled (>500 taxa) asymmetrical simulations 264 and that a 10 % minimal threshold of tip ratio and 300 terminals should be used to better 265 interpret BiSSE results (Davis et al., 2013). We obtained a ratio of 26% (berry) and 74% 266 (capsule) and the analysis was based on 3,585 taxa (SL tree). 267 268 We then performed a time-dependent analysis to obtain the net diversification tip rates and 269 shifts using BAMM v. 2.5.0 and analyzed the outputs using the R package BAMMTools 270 2.1.7 (Rabosky et al., 2014). Unlike the previous analyses, in this case, we used the BB tree, 271 since null branch-lengths, such as some in the SL tree, can influence the measurement of 272 diversification across clades. However, to solve the incomplete sampling and heterogeneity 273 of the BB tree (totaling 30% for the whole family), we specified different sampling fractions 274 for each clade, based on the proportion of recognized species present in the phylogeny: 275 Bromelioideae + Puyoideae (0.42), Tillandsioideae (0.33), Navioideae (0.06), 276 Brocchinioideae (0.4), Lindmanioideae (0.07), and Pitcairnioideae (0.3). We ran four 277 reversible jump MCMC chains, each for twenty million generations, sampling each 20,000 278 generations. We discarded the first 20% generations as burn-in and checked the convergence 279 in the estimated sample size with a threshold >200. We then performed Spearman's 280 correlation tests and plotted the extracted diversification (lambda) and extinction (mu) tip 281 rates of each terminal, in addition to the dispersal capacity (mean distance), in order to 282 compare the influence of each numerical variable. 283 284 BAMM analyses do not allow negative or null branch-lengths and the higher rates of 285 diversification found in young lineages may be influenced by the "pull of present", those that 286 have had less time to become extinct (Eiserhardt et al., 2017; Helmstetter et al., 2022). Moore 287 et al. (2016) presented a critique of the BAMM method, mainly regarding the likelihood 288 function, rate-shifts, and diversification rates. However, Rabosky et al. (2017) concluded that 289 BAMM has high accuracy and consistency to account for diversification and rate shifts, and 290 argued that Moore et al. (2016) based their discussion on a set of low-power analyses. 291 Following Rabosky et al. (2017), we used this method and compared the distribution of the 292 morphological, habitat and diversification data with each tip in the trees we used the R 293 package *ggtree* 2.99.0 (Yu et al., 2017).

294 **Results** 295 Alignment and phylogenetic trees 296 Our dataset resulted in an alignment of 21,271 bp from 1,273 taxa (1,268 bromeliads + 5 297 outgroups; see Appendices S3 and S4). Major clades within the family (subfamilies, tribes, 298 and subtribes) were strongly supported (Fig. 1; see Appendix S5; Tab. S1). The relationships 299 of lineages within the family correspond to those reported by Givnish et al. (2011), except for 300 a strongly-supported, novel clade of *Bromelia* + (Bromelioideae + Puyoideae) (PP=1). Some 301 relationships at the generic and infrageneric level were poorly-supported, especially for the 302 taxonomically-problematic genera (see Appendix S5 for BB tree; and S6 for SL tree; 3,585 303 taxa). 304

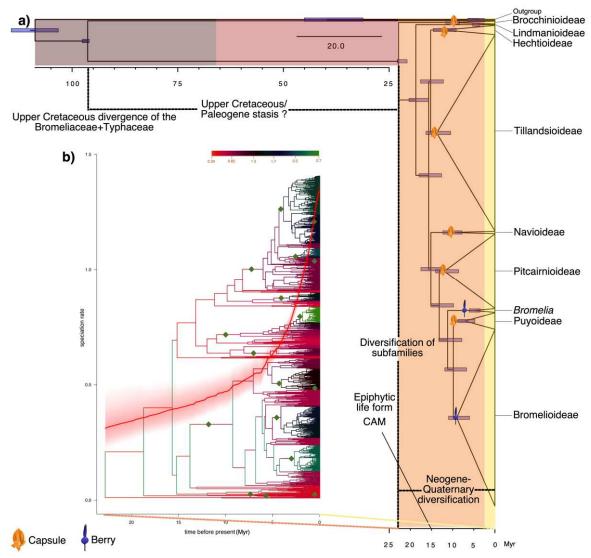


Fig. 1 Phylogeny and diversification of bromeliads. a) Maximum clade credibility (MCC) using the combined sequence data of 1,273 taxa. Colored vertical bars refer to geological periods: Quaternary (yellow); Neogene (orange); Paleogene (brownish red); Cretaceous (gray). Horizontal bars on the nodes indicate 95% CI resulted from the treePL analysis. The topology of this time-calibrated tree suggests a Cretaceous origin, Upper Cretaceous-Paleogene stasis and a Neogene-Quaternary diversification of the group; b) BAMM output tree overlapped in the LTT (Lineage Through Time) plot. Time axes are in Myr.

Divergence time estimate The divergence between Bromeliaceae and Typhaceae occurred during the Upper Cretaceous (~96 Ma; Fig. 1, Tab. S1) however, diversification in Bromeliaceae only started at the beginning of the Neogene (~22 Ma). Divergence of the major bromeliad groups occurred during the Miocene (~22 Ma), with Brocchinioideae and Lindmanioideae as the early divergent subfamilies. Despite the early divergence of these groups, the diversification of all subfamilies only started in the Mid-Upper Miocene, except Lindmainioideae, which diversified during the Pliocene (4.6 Ma). Phylogenetic regression The best-fit model for dispersal capacity included annual temperature and canopy height (AIC value of 5937.534; Tab. S2, Fig. S1), which was corroborated with significant ANOVA results (Tab. 1). Although annual temperature was significant in the full model ANOVA (Tab. S3), the slope was not significant (p=0.274). For the best-fit model, we conducted a phylogenetic regression to identify the effect of temperature and canopy height on dispersal capacity. Our analyses showed that annual temperature and canopy height were strongly significant (p<0.0001). The effect of annual temperature (0.180 \pm 0.028) was marginally stronger than the canopy height (0.137 \pm 0.028) and moderately supported (λ =0.75, 95% CI=0.64-0.82). All the effects of independently tested variables on dispersal capacity had high AIC values, and all of the trait variables were non-significant.

Tab. 1 ANOVA of the PGLS (Phylogenetic Generalized Least Squares) best-fit dispersal capacity model according to AIC (Akaike Information Criterion) values (MD~AT+CH). Fruit morphology is not a predictor for dispersal capacity.

Trait	df	Sum Sq	Mean Sq	F value	p-value
AT	1	1719	1718.63	42.612	8.273e-11
СН	1	921	921.30	22.843	1.875e-06
Residuals	2186	88166	40.33	-	-

Notes: Estimated value of lambda of 0.748 considering the influence of all variables under
 the mean distances. AT: mean annual temperature; CH: canopy height; df: degrees of
 freedom.

315316

317

318

319

320

321

322

323

324325

326

327

328

329

330

331

332

333

334

335

336

337

338

339

Diversification

Among the six models tested in the trait-dependent diversification analyses, the best fit was the model where character change (q) was constrained for fruit type. Our analyses indicated that berry-fruited species (λ =0.20, μ =0.000034) have higher net diversification rates than capsule-fruited species (λ =0.13; Fig. 2).

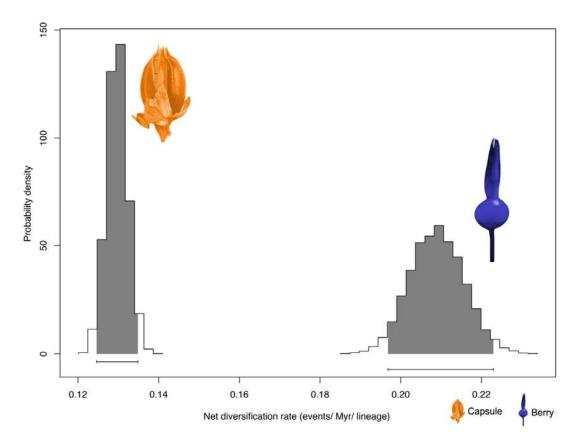


Fig. 2 BiSSE (Binary state speciation and extinction) indicating the effect of the fruit types on net diversification rate.

The BAMM diversification analysis identified heterogeneous rates across the tree (Fig. 1, see Appendix S7). Higher rates were observed in the clades of *Wittmackia* Mez, *Dyckia* Schult.f., and a group comprising North and Central American *Tillandsia* L. (see Appendix S7). Seventeen diversification rate shifts were identified in the Upper Cenozoic, six in the Miocene, five in the Pliocene and six in the Pleistocene, coinciding with the divergence and diversification of several lineages (Fig. 1).

Considering bromeliads at the family level, our results identified a significant negative correlation between net diversification rate with elevation (R=-0.15, p=2e-06) and canopy height (R=-0.11, p=0.00068). Annual precipitation (R=-0.011, p=0.72) was not significant and annual temperature (R=0.064, p=0.039) had a significant but low positive correlation with net diversification rate (Fig. 3, S2). Considering each subfamily separately (Tab. 2), the variables had different effects on the net diversification rate, and when significant, the slopes frequently had a stronger correlation compared to those at the family level.

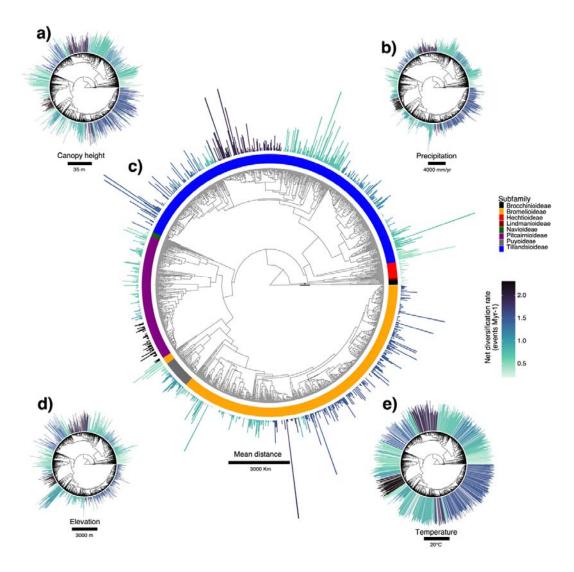


Fig. 3 Phylogeny of Bromeliaceae, habitat and diversification. Bar plots indicate habitat type in size and net diversification rate in color: a) Canopy height values; b) Precipitation values; c) Mean distance values; d) Elevation values; e) Temperature values.

Tab. 2 Correlation between the numeric variables and net diversification rate and mean distance for each bromeliad group.

Group	Temperati	ure	Precipitation		Canopy heigh	t	Elevation	
	NDR	MD	NDR	MD	NDR	MD	NDR	MD
Brocchinioideae ^a	R=-0.39,	R=-0.34,	R=-0.42,	R=0.49,	R=0.37,	R=0.31,	R=0.45,	R=0.37,
	p=0.34	p=0.17	p=0.3	p=0.04	p=0.37	p=0.21	p=0.26	p=0.14
	R=-	R=0.22,	R=0.21,	R=0.15,	R=0.24,	R=0.065,	R=-0.046,	R=-0.19,
Bromelioideae	0.014,	p=7.9e-08	p=5e-05	p=0.00027	p=2.5e-06	p=0.12	p=0.37	p=3.6e-06
	p=0.79							
Hechtioideae	R=-0.33,	R=-0.031,	R=-0.61,	R=-0.15,	R=-0.46,	R=-0.09,	R=0.14,	R=-0.0067,
	p=0.14	p=0.83	p=0.0026	p=0.3	p=0.032	p=0.53	p=0.53	p=0.96
Lindmanioideae ^a	R=0.5,	R=-0.045,	R=-1,	R=0.31,	R=0.5, p=1	R=0.1,	R=-0.5, p=1	R=-0.096,
	p=1	p=0.81	p=0.33	p=0.085		p=0.58		p=0.61
Navioideae ^a	R=0.14,	R=0.059,	R=0.68,	R=-0.13,	R=-0.14,	R=-0.24,	R=-0.14,	R=-0.057,
	p=0.78	p=0.61	p=0.11	p=0.26	p=0.78	p=0.038	p=0.78	p=0.62
Pitcairnioideae	R=0.067,	R=0.15,	R=0.082,	R=0.052,	R=-0.48,	R=0.064,	R=-0.28,	R=-0.064,
	p=0.4	p=0.003	p=0.3	p=0.3	p=1.4e-10	p=0.2	p=0.00031	p=0.2
Puyoideae	R=-0.31,	R=0.25,	R=0.48,	R=0.067,	R=0.63,	R=-0.069,	R=0.44,	R=-0.24,
	p=0.047	p=0.0042	p=0.0015	p=0.45	p=8.6e-06	p=0.44	p=0.0033	p=0.0058
Tillandsioideae	R=-	R=0.094,	R=-0.44,	R=0.1,	R=-0.38,	R=0.062,	R=0.049,	R=-0.092,
	0.093,	p=0.0045	p<2.2e-16	p=0.0019	p=6.3e-16	p=0.064	p=0.32	p =0.0055

	R=-0.066,	p=0.002	nples in
	R =-	0=d	N<25 san
	R=-0.15,	p=2e-06	ean distance; ^a
	R=0.057,	p=0.0081	n rate; MD= m
	R=-0.11,	p=0.00068	f correlation; NDR= net diversification rate; MD= mean distance; ^a N<25 samples in
	R=0.067,	p=0.0018	relation; NDR=1
	R=-0.011,	p=0.72	oefficient of cor
	R=0.064, R=-0.077,	p=0.039 p=0.00033	in bold. R= C
7	R=0.064,	p=0.039	o-values are
	A 11 Dromoling	All blomenaceae	Notes: Significant p-values are in bold. R=
			375

We investigated the effects of the variables in each group of fruit types. Among the variables we tested, canopy height and annual precipitation had significant correlations with net diversification rate (Fig. 4a–b), positive for the berry fruit type (R=0.24, p=2.5e-06 and R=0.21, p=5e-05, respectively) and negative for capsules (R=-0.28, p=1.4e-13 and R=-0.16, p=3.9e-05, respectively). For dispersal capacity, annual temperature and elevation had significant correlations with mean distance (Fig. 4c–d), with annual temperature positively correlated for both types of fruits (berry: R=0.22, p=7.9e-08; and capsule: R=0.056, p=0.024) and elevation negatively correlated (berry: R=-0.19, p=3.6e-08; and capsule: R=-0.054, p=0.03). In both cases, the correlation was stronger in berry-fruited species. Results on maps, data distribution and geographic information are in Fig. S3, see Appendices 8 and 9.

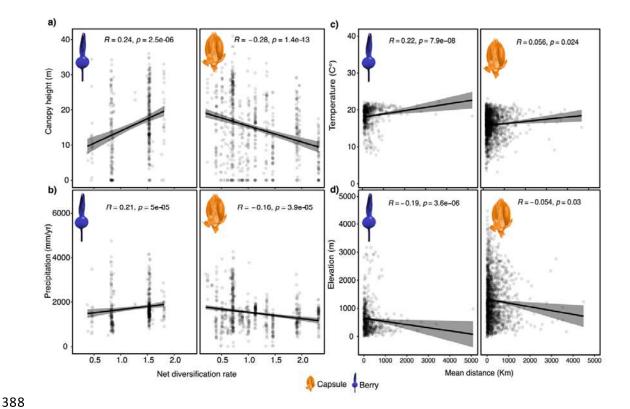


Fig. 4 Correlation plots of the habitat types and net diversification rates (NDR)/mean distances (MD) within different types of fruits. The analysis showed canopy height (a) and precipitation (b) correlated to the net diversification rate (positively in berry and negatively in capsule, in both) and temperature (c) and elevation (d) correlated to mean distance (positively in temperature and negatively in elevation, in both).

Discussion The relationship between biological dispersal and distribution remains a large knowledge gap fundamental to the geographic distribution of species. It is also poorly understood how this relationship interacts with diversification, which together drive biodiversity patterns in time and space. Here, we use a large molecular phylogeny of bromeliads, fruit and climatic data to test how habitat and morphology influence dispersal capacity and lineage diversification. Berry-fruited species have higher diversification rates Our results demonstrate that berry-fruited species have higher speciation rates and lower extinction rates compared to the capsule-fruited species (Fig. 2). The evolution of fleshy fruits, including berries, is considered a key innovation in bromeliads (Givnish et al., 2014). In the Neotropical bellflowers, berries were strongly correlated to closed forest canopy habitats (Lagomarsino et al., 2014, 2016), as here observed for bromeliads (Fig. 4). Berries are primarily bird-dispersed, and the Neotropics has the highest species richness of frugivorous birds in the world (Kissling et al., 2012). Moreover, fleshy-fruited plants and their associated frugivores are commonly generalists (Eriksson, 2016; Bello & Barreto, 2021), meaning that seed dispersal does not rely on a single species and can promote diversification via increasing dispersal capacity (de Queiroz, 2002). Our BAMM results identify three bromeliad clades that had the highest diversification rates (see Appendix S7): (1) Wittmackia (berry fruit, Bromelioideae); (2) Dyckia (capsular fruit, Pitcairnioideae); (3) North and Central American *Tillandsia* (capsular fruit, Tillandsioideae) (Fig. 1). Aguirre-Santoro et al. (2020) also showed high diversification rates in Wittmackia, which was suggested to result from a recent long-dispersal event from the Brazilian Atlantic Forest to the Caribbean, causing a rapid radiation with a myriad of morphological adaptations. Central America (sensu Givnish et al., 2011) is considered an important biogeographic region for bromeliads (Givnish et al., 2011; Granados-Mendoza et al., 2017; Kessous et al., 2020), since it is one of the three species diversity centers of bromeliads (Smith & Downs, 1974; Givnish et al., 2011; Zizka et al., 2020). Also, events occurred in the Miocene-Pliocene, as the seasonality and aridity, were potential drivers of diversification of North and Central American Tillandsia (Granados-Mendoza et al., 2017). Our results indicate

395 396

397

398

399

400

401

402 403

404

405

406

407

408 409

410

411

412

413

414

415

416

417

418

419

420

421

422

423

424

425

that, considering bromeliads as a whole, berry-fruited species have higher diversification, although some individual capsule-fruited clades also have high diversification rates. Habitat drives dispersal and diversification Environmental conditions at high elevations contribute to diversification in mountain ecosystems (Rahbek et al., 2019), including Neotropical plants (Madriñán et al., 2013; Lagomarsino et al., 2016; Testo et al., 2019). Considering bromeliads as a whole, only temperature and canopy height influence in the dispersal capacity (Tab. 1). Fruit morphology was not found to be a predictor of dispersal capacity, probably because bromeliads have two different long-distance dispersal mechanisms (anemochory and zoochory) in their largest groups (see the distribution of the data in Fig. S4). Also, we observed an overall negative correlation between elevation and net diversification of bromeliads (Fig. S2a). In both cases, the relation among the traits, dispersal capacity and diversification are clearer considering case-by-case. Considering subfamilies common in high elevations such as the Andean Puyoideae, we found a strong positive correlation with net diversification rate (Tab. 2). Bromeliads have great morphological diversity, favoring the occurrence across broad elevation gradients ranging from coastal Brazilian restinga forests (e.g. Neoregelia cruenta (Graham) L.B.Sm.) to Andean highlands at almost 5,000 m (e.g. *Puya raimondii* Harms; Smith & Downs, 1974). According to our data, of the 2,718 species with an estimated mean elevation, only 15% occur higher than 2,000 m and 3% higher than 3,000 m. In contrast, among our 180 analyzed species of Puyoideae, 67% have a mean elevation higher than 2,000 m and 33% higher than 3,000 m. Thus, it is important to recognize that elevation-diversification dynamics differ considerably across clades and some patterns are driven largely by single groups. High elevation areas have high endemism, due to the geographic isolation of the species found there (Steinbauer et al., 2016), and to the lower species richness of frugivore dispersers (Sam et al., 2019). Our analysis shows that dispersal capacity is negatively correlated with elevation in Puyoideae (Tab. 2) and berry-fruited species (Fig. 4), suggesting that high elevation species tend to be endemic to smaller regions, probably owing to the specific attributes and adaptations to these habitats.

427

428

429 430

431

432

433

434

435

436

437

438

439

440

441

442

443

444

445

446

447

448

449

450

451

452 453

454

455

456

457

458

460 Plant lineages often have strong niche conservatism with climatic preferences such as 461 temperature and precipitation (Punyasena et al., 2008), and are therefore vulnerable to 462 climate change (Liu et al., 2020). Here, we show that bromeliad subfamilies have specific 463 climate preferences. As a whole, in bromeliads, there was no correlation between annual 464 temperature and net diversification rate (R=0.064, p=0.039, Tab. 2), but when considering 465 only Puyoideae we observe a negative correlation (R=-0.31, p=0.047, Tab. 2). In Puyoideae, 466 high rates of diversification are associated with lower temperatures, potentially suggesting 467 that a lack of competition in high elevation environments benefits the diversification in the 468 group. In contrast, we observed that high temperatures positively influence the dispersal 469 ability of Puyoideae (R=0.25, p=0.0042, Tab. 2), Bromelioideae (R=0.22, p=7.9e-08, Tab. 2) 470 and Pitcairnioideae (R=0.15, p=0.003, Tab. 2), indicating that for these groups, lower 471 temperatures lead to higher rates of endemism. 472 473 Annual precipitation is correlated with dispersal capacity and net diversification rate in some 474 bromeliad subfamilies. Low humidity regions may positively affect dispersal in capsule-475 fruited species (with wind-dispersed seeds; Heydel & Tackenberg, 2017). The diversification 476 in Tillandsioideae and Hechtioideae, both widely distributed in dry habitats, are negatively 477 correlated with precipitation, likely because of their seed morphology that is adapted to 478 dispersal by wind and/or gravity. On the other hand, precipitation positively influences the 479 diversification of Bromelioideae and Puyoideae, which despite their close phylogenetic 480 relationship (Fig. 1, Tab. 2), have differences in fruit and seed morphology (berry/naked 481 versus capsule/winged, respectively). Precipitation is positively correlated with dispersal 482 ability in Brocchinioideae (endemic to Amazonia), Bromelioideae, and Tillandsioideae (Tab. 483 2), the latter two being the most widely distributed subfamilies (Zizka et al., 2020). Co-484 occurrence may be reflected by similar habitat preferences among different groups 485 (Punyasena et al., 2008) as clearly observed among these niche-conserved bromeliads (see 486 Zizka et al., 2020). 487 488 Canopy height influences bromeliad dispersal and diversification 489 The net diversification rate of the family decreases with a higher canopy height (Tab. 1, Figs. 490 S2, 3a, 4), perhaps due to the predominance of species with capsular fruits (>70% of the 491 species), since in berry-fruited species (Bromelioideae, mostly epiphytic) the net 492 diversification rate increases with a higher canopy height and annual precipitation (Fig. 4a-

b). A higher canopy can behave as a physical barrier for the wind-dispersed seeds present in capsular fruits and also may positively influence the diversity of vertebrate dispersers, (Walter et al., 2017), favoring the dispersal in berry-fruited species. On the other hand, open canopies can favor the wind-dispersed seeds (Correa et al., 2022). In the Neotropics, the end of the Cretaceous (Maastrichtian: 72–66 Ma) was characterized by open canopied forests and the presence of several vascular plant groups. In contrast, early Paleocene forests (66–61 Ma) were denser, but with a lower diversity compared to the Maastrichtian (Carvalho et al., 2021). This different habitat composition is likely due to the (1) presence of large herbivores, mainly dinosaurs, that physically disturbed forests, (2) infertility of the soils in the Maastrichtian, and (3) the selective extinction of gymnosperms that occurred in the Cretaceous-Paleogene boundary (Carvalho et al., 2021). The reduction of 45% of the Paleocene plant diversity resulted from the slow recovery from the Cretaceous-Paleogene mass extinction caused by the Chicxulub impact and intense volcanic activity (Schulte et al., 2010; Hull et al., 2020), which suppressed sunlight, changed the atmosphere, and reduced global temperatures (Alvarez et al., 1980; Schulte et al., 2010; Vajda et al., 2014; Vajda et al., 2015; Hull et al., 2020). The increased complexity of Neotropical Paleocene forests provided opportunities that included vertical diversity, as in the case of the epiphytes, primarily because to changes in the water and light availability within closed canopies 512 (Carvalho et al., 2021). The first bromeliads were terrestrial and inhabited open environments (Givnish et al., 2014; Bouchenak-Khelladi et al., 2014), however, the evolution of other life forms and morphology, such as epiphytism, possibly behaved as a trigger for high and rapid diversification (Givnish et al., 2014; Givnish et al., 2017). The causes of the early divergence and late diversification of the bromeliads are still unknown (Kessous et al., 2021). According to our results we hypothesize that the open forests of the Upper Cretaceous favored the rise of the terrestrial/capsular bromeliads. However, after the mass extinction, during the Paleocene, the closed canopied forests negatively affected the bromeliad diversification, once the epiphytic life form of this group appeared for the first time in the Mid Miocene (Givnish et al., 2014). The Eocene-Oligocene extinction event resulted in semi-open canopies (Carvalho et al., 2021), which also may be one of the factors that triggered the diversification of bromeliads in the Miocene (Fig. 1). After the development of the epiphytic habitat (Fig. 1,

493

494

495

496

497 498

499

500

501

502

503

504

505

506

507

508

509

510

511

513 514

515

516

517

518

519

520

521

522

523

524

3a), the higher canopy height may have also favored diversification, as for the berry-fruited species. We hypothesize that both higher and lower canopies were triggers of bromeliad diversification, but in different time frames and for different fruit morphologies. Taken together and according to our results, we offer two hypotheses: (1) capsular fruits have seeds that are primarily wind-dispersed, and closed canopies serve as a physical barrier to dispersal and subsequent allopatric speciation (2) the abundance of frugivores (such as large birds and bats) can be higher in dense forests (Walter et al., 2017) and benefit the dispersal of berry-fruited species. Although the first documented frugivore interactions are from the Upper Cretaceous, fleshy fruits evolved in several plant groups in the Miocene (Eriksson, 2016), as observed here in bromeliads. Conclusion Here we test drivers of dispersal and diversification in a hyperdiverse plant family in a phylogenetic framework. We hypothesize that both lower and higher canopy heights influenced the dispersal and diversification of bromeliads, and that the low canopies heights of paleoforests prompted the origin and diversification of the family. We also demonstrate that berry fruits promoted diversification in bromeliads. Fruit morphology is not a predictor for dispersal capacity and factors influencing fruit types are heterogeneous among the subfamilies. Our analyses show the importance of evaluating different groups as separate entities together with different effects of climatic variables in each group. Furthermore, we show that the evolutionary success of bromeliads does not rely on a single key innovation, but rather on a complex network of strategies on different temporal scales and that are linked to dispersal, colonization, and adaptation. We highlight the importance of investigating the influence of traits and variables on dispersal and diversification, while accounting for the ecological and morphological diversity of its subgroups. Data availability The data underlying this article are included in the Supporting Information. Acknowledgements This work was supported by Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES Foundation), CAPES-STINT (Project 88881.304776/2018-01; 8887.477452/2020-

526

527

528

529

530

531

532

533534

535

536

537538

539

540

541

542

543

544

545

546

547

548

549

550

551

552

553

554

555556

557

559 00); the Helge Ax:son Johnsons stiftelse (F21-0212); the Sven and Dagmar Saléns 560 Foundation; the Wilhelm and Martina Lundgrens Foundation (2020-3489); the Swedish 561 Research Council (2017-04980); and the Biodiversity and Ecosystems Services in a 562 Changing Climate (BECC) Strategic Research Area at the University of Gothenburg. We 563 thank Søren Faurby and the members of the Gothenburg Global Biodiversity Centre for 564 valuable discussions; and the Swedish National Infrastructure for Computing (SNIC) for 565 computational support (SNIC 2020/9-216; SNIC 2020/10-114). 566 567 **Conflict of interest** 568 The authors declare no conflict of interest. 569 570 **Author contributions** 571 IMK, AFC, FS and CDB planned and designed the study. IMK, HF, WT, MFT, BN and ARP 572 ran the statistical and phylogenetic analyses. IMK, AFC and CDB interpreted the results. 573 IMK wrote the manuscript with suggestions from all remaining authors. 574 575 576 References 577 Aguirre-Santoro J., Salinas N.R., Michelangeli F.A. 2020. The influence of floral variation 578 and geographic disjunction on the evolutionary dynamics of Ronnbergia and 579 Wittmackia (Bromeliaceae: Bromelioideae). Bot. J. Linn. Soc. 192(4): 609-624. 580 Alvarez L.W., Alvarez W., Asaro, F., Michel, H.V. 1980. Extraterrestrial cause for the 581 Cretaceous-Tertiary extinction. Science 208(4448):1095–1108. 582 Andresen E., Arroyo-Rodríguez V., Escobar F. 2018. Tropical biodiversity: the importance of 583 biotic interactions for its origin, maintenance, function, and conservation. In Ecological 584 networks in the tropics (pp. 1–13). Springer, Cham. 585 Antonelli A., Sanmartín, I. 2011. Why are there so many plant species in the Neotropics? 586 Taxon 60(2): 403–414. 587 Benzing D.H. 2000. Bromeliaceae: profile of an adaptive radiation. Cambridge: Cambridge 588 University Press. 589 Bello C., Barreto E. 2021. The footprint of evolution in seed dispersal interactions. Science 590 372(6543): 682–683.

591 Bouchenak-Khelladi Y., Muasya A.M., Linder H.P. 2014. A revised evolutionary history of 592 Poales: origins and diversification. Bot. J. Linn. Soc. 175(1): 4–16. 593 Bouckaert R., Heled J., Kühnert D., Vaughan T., Wu C.H., Xie D., Suchard M.A., Rambaut 594 A., Drummond A.J. 2014. BEAST 2: a software platform for Bayesian evolutionary 595 analysis. PLoS Comput. Biol. 10(4): e1003537. 596 Brown J.H. 2014. Why are there so many species in the tropics? J. Biogeogr. 41(1): 8–22. 597 Burgman M.A., Fox J.C. 2003. Bias in species range estimates from minimum convex 598 polygons: implications for conservation and options for improved planning. Anim. 599 Conserv. 6(1): 19–28. 600 Carvalho M.R., Jaramillo C., de la Parra F., Caballero-Rodríguez D., Herrera F., Wing S., 601 Turner B.L., D'Apolito C., Romero-Báez M., Narváez P., Martínez C. 2021. Extinction 602 at the end-Cretaceous and the origin of modern Neotropical rainforests. Science 603 372(6537): 63–68. 604 Castresana J. 2000. Selection of conserved blocks from multiple alignments for their use in 605 phylogenetic analysis. Mol. Biol. Evol. 17: 540–552. 606 Correa, D.F., Stevenson, P.R., Umaña, M.N., Coelho, L.D.S., Lima Filho, D.D.A., Salomão, 607 R.P., Amaral, I.L.D., Wittmann, F., Matos, F.D.D.A., Castilho, C.V., Phillips, O.L., 608 2022. Geographic patterns of tree dispersal modes in Amazonia and their ecological 609 correlates. Glob. Ecol. Biogeogr. 00: 1–21. 610 Davis M.P., Midford P.E., Maddison W. 2013. Exploring power and parameter estimation of 611 the BiSSE method for analyzing species diversification. BMC Evol. Biol. 13(1): 1–11. 612 Eiserhardt W.L., Couvreur T.L., Baker W.J. 2017. Plant phylogeny as a window on the 613 evolution of hyperdiversity in the tropical rainforest biome. New Phytol. 214(4): 1408– 614 1422. 615 Eriksson O. 2016. Evolution of angiosperm seed disperser mutualisms: the timing of origins 616 and their consequences for coevolutionary interactions between angiosperms and 617 frugivores. Biol. 91(1): 168–186. 618 FitzJohn R.G. 2012. Diversitree: comparative phylogenetic analyses of diversification in R. 619 Methods Ecol. Evol. 3(6): 1084–1092. 620 Givnish T.J., Barfuss M.H., Van Ee B., Riina R., Schulte K., Horres R., Gonsiska P.A., 621 Jabaily R.S., Crayn D.M., Smith J.A.C., Winter K. 2011. Phylogeny, adaptive 622 radiation, and historical biogeography in Bromeliaceae: insights from an eight-locus 623 plastid phylogeny. Am. J. Bot. 98(5): 872–895.

624 Givnish T.J., Barfuss M.H., Van Ee B., Riina R., Schulte K., Horres R., Gonsiska P.A., 625 Jabaily R.S., Crayn D.M., Smith J.A.C., Winter K., 2014. Adaptive radiation, correlated 626 and contingent evolution, and net species diversification in Bromeliaceae. Mol. 627 Phylogenet. Evol. 71: 55-78. 628 Givnish T.J. 2017. A New World of plants. Science 358(6370): 1535–1536. 629 Givnish T.J., Zuluaga A., Spalink D., Soto Gomez M., Lam V.K., Saarela J.M., Sass C., Iles 630 W.J., De Sousa D.J.L., Leebens-Mack J., Chris Pires J. 2018. Monocot plastid 631 phylogenomics, timeline, net rates of species diversification, the power of multi-gene 632 analyses, and a functional model for the origin of monocots. Am. J. Bot. 105(11): 633 1888-1910. 634 Gouda E.J., Butcher D., Gouda, C.S. 2020 [cont.updated]. Encyclopaedia of Bromeliads, 635 Version 4. University Botanic Gardens, Utrecht. http://bromeliad.nl/encyclopedia/ 636 Granados-Mendoza C., Granados-Aguilar X., Donadío S., Salazar G.A., Flores-Cruz M., 637 Hágsater E., Starr J.R., Ibarra-Manríquez G., Fragoso-Martínez I., Magallón S. 2017. 638 Geographic structure in two highly diverse lineages of *Tillandsia* (Bromeliaceae). 639 Botany 95(7): 641-651. 640 Heibl C. 2019. Package 'ips'. https://cran.r-project.org/web/packages/ips/ips.pdf 641 Helmstetter A.J., Glemin S., Käfer J., Zenil-Ferguson R., Sauquet H., de Boer H., Dagallier 642 L.P.M., Mazet N., Reboud E.L., Couvreur T.L. and Condamine F.L., 2022. Pulled 643 diversification rates, lineages-through-time plots and modern macroevolutionary 644 modelling. Syst. Biol. 71(3):758–773. 645 Herrera-Alsina L., van Els P., Etienne R.S., 2019. Detecting the dependence of diversification 646 on multiple traits from phylogenetic trees and trait data. Syst. Biol. 68(2): 317–328. 647 Heydel F., Tackenberg O. 2017. How are the phenologies of ripening and seed release 648 affected by species' ecology and evolution? Oikos 126(5): 738–747. 649 Hijmans R.J., Van Etten J., Cheng J., Mattiuzzi M., Sumner M., Greenberg J.A., Lamigueiro 650 O.P., Bevan A., Racine E.B., Shortridge A., Hijmans M.R.J. 2015. Package 'raster'. 651 https://cran.r-project.org/web/packages/raster/raster.pdf 652 Hothorn T., Zeileis A., Farebrother R.W., Cummins C., Millo G., Mitchell D., Zeileis, M.A. 653 2015. Package 'Imtest'. Testing linear regression models. https://cran. r-project. 654 org/web/packages/lmtest/lmtest.pdf 655 Hull P.M., Bornemann A., Penman D.E., Henehan M.J., Norris R.D., Wilson P.A., Blum P., 656 Alegret L., Batenburg S.J., Bown P.R., Bralower T.J. 2020. On impact and volcanism

across the Cretaceous-Paleogene boundary. Science 367(6475): 266–272.

26

658 Janzen D.H. 1967. Why mountain passes are higher in the tropics. Am. Nat. 101(919): 233– 659 249. 660 Jin Y., Qian H. 2019. V. PhyloMaker: an R package that can generate very large phylogenies 661 for vascular plants. Ecography 42(8): 1353–1359. 662 Karger D.N., Zimmermann N.E. 2019. Climatologies at high resolution for the earth land 663 surface areas CHELSA V1. 2: Technical specification. Swiss Federal Research Institute 664 WSL, Switzerland. 665 Katoh K., Toh H. 2008. Recent developments in the MAFFT multiple sequence alignment 666 program. Brief. Bioinform. 9(4): 286–298. 667 Kessous I.M., Neves B., Couto D.R., Paixão-Souza B., Pederneiras L.C., Moura R.L., 668 Barfuss M.H., Salgueiro F., Costa, A.F. 2020. Historical biogeography of a Brazilian 669 lineage of Tillandsioideae (subtribe Vrieseinae, Bromeliaceae): the Paranaean Sea 670 hypothesized as the main vicariant event. Bot. J. Linn. Soc. 192(4): 625–641. 671 Kessous I.M., Neves B., Salgueiro F., da Costa A.F. 2021. A 100-Million-Year Gap in the 672 Knowledge of the Evolutionary History of Bromeliaceae: A Brief Review of Fossil 673 Records. Feddes Repert. 132(1): 20–27. 674 Kissling W.D., Sekercioglu C.H., Jetz W. 2012. Bird dietary guild richness across latitudes, 675 environments and biogeographic regions. Glob. Ecol. Biogeogr. 21(3): 328–340. 676 Lagomarsino L.P., Antonelli A., Muchhala N., Timmermann A., Mathews S., Davis, C.C. 677 2014. Phylogeny, classification, and fruit evolution of the species-rich Neotropical 678 bellflowers (Campanulaceae: Lobelioideae). Am. J. Bot. 101(12): 2097–2112. 679 Lagomarsino L.P., Condamine F.L., Antonelli A., Mulch A., Davis C.C. 2016. The abiotic 680 and biotic drivers of rapid diversification in Andean bellflowers (Campanulaceae). New 681 Phytol. 210(4): 1430-1442. 682 Leme E.M., Zizka G., Paule J., Aguirre-Santoro J., Heller S., Ramirez-Morillo I.M., 683 Halbritter H., Mariath J.E., de Carvalho J.D., Forzza R.C. 2021. Re-evaluation of the 684 Amazonian Hylaeaicum (Bromeliaceae: Bromelioideae) based on neglected 685 morphological traits and molecular evidence. Phytotaxa 499(1): 1–60. 686 Liu B., Tan X., Gan T.Y., Chen X., Lin K., Lu M., Liu Z. 2020. Global atmospheric moisture 687 transport associated with precipitation extremes: Mechanisms and climate change 688 impacts. Wiley Interdiscip. Rev.: Water 7(2): e1412. 689 Maddison W.P., Midford P.E., Otto S.P. 2007. Estimating a binary character's effect on 690 speciation and extinction. Syst. Biol. 56(5): 701–710.

- 691 Maddison W.P., FitzJohn R.G. 2015. The unsolved challenge to phylogenetic correlation
- tests for categorical characters. Syst. Biol. 64(1): 127–136.
- 693 Maddison W.P., Maddison D.R. 2019. Mesquite: a modular system for evolutionary analysis.
- Version 3.61 http://www.mesquiteproject.org
- 695 Mittelbach G.G., Schemske D.W., Cornell H.V., Allen A.P., Brown J.M., Bush M.B.,
- Harrison S.P., Hurlbert A.H., Knowlton N., Lessios H.A., McCain C.M. 2007.
- 697 Evolution and the latitudinal diversity gradient: speciation, extinction and
- 698 biogeography. Ecol. Lett. 10(4): 315–331.
- 699 Moore B.R., Höhna S., May M.R., Rannala B., Huelsenbeck J.P. 2016. Critically evaluating
- 700 the theory and performance of Bayesian analysis of macroevolutionary mixtures. Proc.
- 701 Natl. Acad. Sci. USA 113(34): 9569–9574.
- Myers N., Mittermeier R. A., Mittermeier C. G., Da Fonseca G. A., Kent J. 2000.
- Biodiversity hotspots for conservation priorities. Nature 403(6772): 853–858.
- 704 Oliveira B.F., Scheffers B.R. 2019. Vertical stratification influences global patterns of
- 705 biodiversity. Ecography 42(2): 249–249.
- 706 Orme D., Freckleton R., Thomas G., Petzoldt T., Fritz S., Isaac N., Pearse W. 2013. The
- caper package: comparative analysis of phylogenetics and evolution in R. https://cran.r-
- 708 project.org/web/packages/caper/vignettes/caper.pdf
- 709 Palací C.A., Brown G.K., Tuthill D.E. 2004. The seeds of *Catopsis* (Bromeliaceae:
- 710 Tillandsioideae). Syst. Bot. 29(3): 518–527.
- 711 Punyasena S.W., Eshel G., McElwain J.C. 2008. The influence of climate on the spatial
- patterning of Neotropical plant families. J. Biogeogr. 35(1): 117–130.
- 713 de Queiroz A. 2002. Contingent predictability in evolution: key traits and diversification.
- 714 Syst. Biol. 51(6): 917–929.
- 715 Qian H., Jin Y. 2016. An updated megaphylogeny of plants, a tool for generating plant
- phylogenies and an analysis of phylogenetic community structure. J. Plant Ecol. 9(2):
- 717 233–239.
- 718 Rabosky D.L., Grundler M., Anderson C., Title P., Shi J.J., Brown J.W., Huang H., Larson
- J.G. 2014. BAMM tools: an R package for the analysis of evolutionary dynamics on
- phylogenetic trees. Methods Ecol. Evol. 5(7): 701–707.
- 721 Rabosky D.L., Goldberg E.E. 2015. Model inadequacy and mistaken inferences of trait-
- 722 dependent speciation. Syst. Biol. 64(2): 340–355.

- 723 Rabosky D.L., Mitchell J.S., Chang J. 2017. Is BAMM flawed? Theoretical and practical
- 724 concerns in the analysis of multi-rate diversification models. Syst. Biol. 66(4): 477–
- 725 498.
- Rahbek C., Borregaard M.K., Antonelli A., Colwell R.K., Holt B.G., Nogues-Bravo D.,
- Rasmussen C.M., Richardson K., Rosing M.T., Whittaker R.J., Fjeldså J. 2019.
- 728 Building mountain biodiversity: Geological and evolutionary processes. Science
- 729 365(6458): 1114–1119.
- 730 R Core Team 2019. R: A language and environment for statistical computing. R Foundation
- 731 for Statistical Computing.
- de la Rosa-Manzano E., Andrade J.L., Zotz G., Reyes-García C. 2014. Epiphytic orchids in
- 733 tropical dry forests of Yucatan, Mexico–Species occurrence, abundance and
- correlations with host tree characteristics and environmental conditions. Flora 209(2):
- 735 100–109.
- 736 Sam K., Koane B., Bardos D.C., Jeppy S., Novotny V. 2019. Species richness of birds along
- a complete rain forest elevational gradient in the tropics: Habitat complexity and food
- 738 resources matter. J. Biogeogr. 46(2): 279–290.
- 739 Schemske D.W., Mittelbach G.G., Cornell H.V., Sobel J.M., Roy K. 2009. Is there a
- 740 latitudinal gradient in the importance of biotic interactions? Annu. Rev. Ecol. Evol.
- 741 Syst. 40: 245–269.
- 742 Schulte P., Alegret L., Arenillas I., Arz J.A., Barton P.J., Bown P.R., Bralower T.J.,
- 743 Christeson G.L., Claeys P., Cockell C.S., Collins G.S. 2010. The Chicxulub asteroid
- impact and mass extinction at the Cretaceous-Paleogene boundary. Science 327(5970):
- 745 1214–1218.
- 746 Seale M., Nakayama N. 2020. From passive to informed: mechanical mechanisms of seed
- 747 dispersal. New Phytol. 225(2): 653–658.
- 748 Silva K.R., Stützel T., Oriani A. 2020. Seed development and its relationship to fruit structure
- 749 in species of Bromelioideae (Bromeliaceae) with fleshy fruits. Bot. J. Linn. Soc.
- 750 192(4): 868–886.
- 751 Smith L.B., Downs R.J. 1974. Flora Neotropica, Monograph No. 14, Part 1, Pitcairnioideae
- 752 (Bromeliaceae). New York: The New York Botanical Garden.
- 753 Smith L.B., Till W. 1998. Bromeliaceae (pp. 74–99). In Flowering Plants Monocotyledons.
- 754 Berlin: Springer.
- 755 Smith S.A., O'Meara B.C. 2012. treePL: divergence time estimation using penalized
- 756 likelihood for large phylogenies. Bioinformatics 28(20): 2689–2690.

- 757 Smith B.T., McCormack J.E., Cuervo A.M., Hickerson M.J., Aleixo A., Cadena C.D., Perez-
- 758 Eman J., Burney C.W., Xie X., Harvey M.G., Faircloth B.C., 2014. The drivers of
- 759 tropical speciation. Nature: 515(7527): 406–409.
- 760 Steinbauer M.J., Field R., Grytnes J.A., Trigas P., Ah Peng C., Attorre F., Birks H.J.B.,
- 761 Borges P.A., Cardoso P., Chou C.H., De Sanctis, M. 2016. Topography-driven
- isolation, speciation and a global increase of endemism with elevation. Glob. Ecol.
- 763 Biogeogr. 25(9): 1097–1107.
- Suchard M.A., Lemey P., Baele G., Ayres D.L., Drummond A.J., Rambaut A. 2018.
- Bayesian phylogenetic and phylodynamic data integration using BEAST 1.10 Virus
- 766 Evol. 4: vey016.
- 767 Testo W.L., Sessa E., Barrington D.S. 2019. The rise of the Andes promoted rapid
- diversification in Neotropical *Phlegmariurus* (Lycopodiaceae). New Phytol. 222(1):
- 769 604–613.
- 770 Ulloa Ulloa C, Acevedo-Rodríguez P., Beck S., Belgrano M.J., Bernal R., Berry P.E., Brako
- L., Celis M., Davidse G., Forzza R.C., Gradstein S.R. 2017. An integrated assessment
- of the vascular plant species of the Americas. Science 358(6370): 1614–1617.
- 773 Vajda V., Bercovici A. 2014. The global vegetation pattern across the Cretaceous–Paleogene
- mass extinction interval: A template for other extinction events. Glob. Planet. Change
- 775 122: 29–49.
- 776 Vajda V., Ocampo A., Ferrow E., Koch C.B. 2015. Nano particles as the primary cause for
- 777 long-term sunlight suppression at high southern latitudes following the Chicxulub
- impact—evidence from ejecta deposits in Belize and Mexico. Gondwana Res. 27(3):
- 779 1079–1088.
- 780 Vavrek M.J., Vavrek M.M.J., 2020. Package 'fossil'. https://cran.r-
- 781 project.org/web/packages/fossil/fossil.pdf
- Walter S.T., Browne L., Freile J., Olivo J., González M., Karubian J. 2017. Landscape-level
- 783 tree cover predicts species richness of large-bodied frugivorous birds in forest
- 784 fragments. Biotropica 49(6): 838–847.
- 785 Yu G., Smith D.K., Zhu H., Guan Y., Lam T.T.Y. 2017. ggtree: an R package for
- visualization and annotation of phylogenetic trees with their covariates and other
- associated data. Methods Ecol. Evol. 8(1): 28–36.
- 788 Yule G.U. 1925. II.—A mathematical theory of evolution, based on the conclusions of Dr. JC
- 789 Willis, FR S. Philos. Trans. R. Soc. Lond., B, Biol. Sci. 213(402-410): 21–87.

790 Zhang J., Nielsen S.E., Mao L., Chen S., Svenning, J.C. 2016. Regional and historical factors 791 supplement current climate in shaping global forest canopy height. J. Ecol. 104(2): 792 469-478. 793 Zizka A., Silvestro D., Andermann T., Azevedo J., Duarte Ritter C., Edler D., Farooq H., 794 Herdean A., Ariza M., Scharn R., Svantesson S. 2019. CoordinateCleaner: 795 Standardized cleaning of occurrence records from biological collection databases. 796 Methods Ecol. Evol. 10(5): 744-751. 797 Zizka A., Azevedo J., Leme E., Neves B., da Costa A.F., Caceres D., Zizka G. 798 2020.Biogeography and conservation status of the pineapple family (Bromeliaceae). 799 Divers. Distrib. 26(2): 183-1 800

Supporting Information App.S1 List of accepted species names of Bromeliaceae according Gouda et al., 2020 [cont. updated], https://bromeliad.nl/encyclopedia/ (December 2020). **App. S2** List of the sequences of the sampled taxa. Available in GenBank (ncbi.nlm.nih.gov/genbank/). **App. S3** Concatenated matrix totaling 21,271 bp from 1,273 taxa. Sequence data for 13 chloroplast (agt1, ycf1, rps16-intron, rps16-trnK, rpl32, matK, nadH, petD, trnL-trnF, rpoB, atpB-trnC, psbA-trnH, and trnC-petN) and three nuclear (PHYC, PRK, and LEAFY) loci. **App. S4** XML final file generated with BEAUTi v. 1.10.4 (Suchard et al., 2018). **App. S5** Backbone tree (BB tree). Maximum clade credibility (MCC) using the combined sequence data of 1,273 taxa resulted from a 21,271 bp alignment. **App.S6** Species-level tree (SL tree). All-species bromeliad tree obtained from the BB tree by imputing missing taxa using the R package V. Phylomaker 0.1.0 (Jin & Qian, 2019). **App. S7** Tip rates out of net diversification obtained from BAMM v. 2.5.0 diversification analysis. **App.S8** Records, geographical, morphological and habitat information. **App.S9** Species and transformed data (log-transformed, scale, dummy variables), habitat and morphological information. **Tab. S1** Main clade support and timing of crown and stem ages. **Tab. S2** Comparison of trait-based best-fit models of dispersal represented by mean distance (MD).

801 802

803 804

805

806 807

808

809 810

811

812

813814

815816

817

818819

820

821 822

823

824 825

826827

828

829 830

831 832

Tab. S3 ANOVA of the PGLS complete model. Estimated value of lambda of 0.736 (0.620, 0.815), kappa and delta fixed in 1, considering the influence of all variables under the mean distances. The levels of berry (FT) and winged (ST) were considered points of comparison, thus not estimated in the analysis. All variables were scale 0 modified. Fig. S1 Residuals of the PGLS (Phylogenetic Generalized Least Squares) best-fit model according to AIC (Akaike Information Criterion). 842 Fig. S2 Correlation plots of the habitat types and net diversification rates (NDR). Elevation and Canopy height showed negative correlation with NDR, considering the whole family. a) Elevation; b) Temperature; c) Precipitation; d) Canopy height. Marginal lines represent the density distribution of the data. **Fig. S3** Maps, tree and distribution of the data. a) Precipitation maps and density plots; b) Temperature maps and density plots; c) Canopy height maps and density plots; d) Elevation maps and density plots; e) Species-level tree (SL tree) with bar plots of the habitat data. Fig. S4 Violin plots representing the distribution of the mean distance (proxy for dispersal capacity) between the two fruit types.

834

835

836

837

838

839 840

841

843

844

845

846

847 848

849

850

851

852